

Studies on the mycoflora associated with the leaves of some plants

Dhale D. A.*

Post-Graduate Department of Botany, SSVPS's, L.K.Dr.P.R.Ghogrey Science College, Dhule -424005 (M.S.), India

Abstract

About twenty four diseased plants were collected from around Aurangabad city of Maharashtra State. The mycoflora on the leaf surfaces was observed by various methods such as direct observation on stained leaf, leaf wash method, spore fall method, leaf print method. About fifteen genera of different pathogenic fungi have been recorded. Potato dextrose agar media was used for growing the fungi. The some fungi which recorded are *Alternaria* spp., *Aspergillus niger, Aspergillus flavus, Cladosporium* spp., *Fusarium* spp., *Helminthosporium* spp., *Phytopthora* spp., *Pythium* spp., *Phyllactinia* spp, *Uncinulla* spp., *Urosystis* spp., *uromyces* spp., etc. The maximum infection is by *Fussarium* spp. and *Alternaria* spp. to 13 plant species. The pathogen *Fussarium* spp. and *Alternaria* spp. are found causing disease both on same host on up to 9 different plant species.

Keywords: Mycoflora, leaf surface, pathogen, fungi

INTRODUCTION

Phylospere –(Last -1955) introduced term phyllospere to denote the leaf surface of the plants. He further advocated that phylloshere region is similar to rhizosphere in that it is also nutritionally rich, microhabitat and provide suitable (host) substrate for the colonization and multiplication of micro-organism. The report of the intensive investigations on leaf surface mycoflora has been reported by Last and Deighton (1965).

The existence of active mycelium in the phyllosphere is a fundamental and much-discussed question (Kerling, 1958; Last & Dcighton, 1965; Dickinson, 1965; 1967; Fokkema, 1968). The results of this work, as well as photographs previously published (Diem, 1970), confirm the presence on green leaves and in natural conditions of a living mycoflora (Warnock 1973).

So, many important plants like some of horticultural plants (*Abelmoscus esculantus*), Garden plants (*Jasminum officinales, Rosa demascana*), Wild plants (*Xanthium strumarium, Cassia tora*), Medicinaly very important plants (*Azadirecta indica, Wthania somnifera*) are studied for there phylospere. Because it is the best way to affect plant with pathogen.

MATERIAL AND METHODS

The leaves of about twenty four plants were collected from the Dr. Babasaheb Ambedkar Marathwada University and Government institute of Science campus, Aurangabad. The plant materials (leaves, flowers) were identified using the Flora of Marathwada (Naik, 1998) at Department of Botany, Government institute of Science,

*Corresponding Author

Dhale D. A. Post-Graduate Department of Botany, SSVPS's, L.K.Dr.P.R.Ghogrey Science College, Dhule -424005 (M.S.) India

Tel-Fax:02562 – 271340 Email: datta.dhale@yahoo.com Caves Road, Aurangabad (M.S.).

Mycoflora of these different twenty four plants are grown on potato dextrose agar medium in Petri plate and observed under microscope. For study of mycoflora different methods are used like, i) direct observation on stained leaf ii) leaf wash method iii) spore fall method iv) leaf print method. (Last, 1955; Ruinen, 1961; Daft & Leben, 1966; Dickinson, 1967; Rusch & Leben, 1968).

RESULTS AND DISCUSSIONS

About twenty four important diseased plants were observed (Table 1). The microscopic examination of the phyllosphere gave valuable information on the distribution and sequence of the natural mycoflora of the leaf surface. Different results were obtained by the direct observation on stained leaf, leaf washing method, spore fall method and leaf print method have been noted (Table 2). The some important fungi which recorded are *Alternaria* spp., *Aspergillus niger, Aspergillus flavus, Cladosporium* spp., *Fusarium* spp., *Helminthosporium* spp., *Phytopthora* spp., *Pythium* spp., *Phyllactinia* spp, *Uncinulla* spp., *Urosystis* spp., *Uromyces* spp., etc.

By different observation of leaves, the data indicates that the maximum infection is by *Fussarium* spp. and *Alternaria* spp. to 13 plant species (Table 3, 4) while *Curvularia* spp. and *Cladospora* spp. have infect to 8 and 7 plant spp. respectively. The minimum infections causes by *Pythium* spp, *Collatotricum* spp., *Phyllactinia* spp., *Urocystis* spp., *Uromyces* spp., *Cercospora* spp. *Penicillium* spp. and *Uncinulla* spp. to one plant species (Table 3). The pathogen *Fussarium* spp. and *Alternaria* spp. are found causing disease both on same host on up to 9 different plant species (Table 5).

Received: July 05, 2011; Revised September 01, 2011; Accepted September 01, 2011.

Table 1: Plants Studied for the Phyllosphere

Sr. No.	Name of the Plant	Sr. No.	Name of the Plant	
1.	Abesmoschus esculantus	13.	Euphorbia hirta	
2.	Acacia leucophloea	14.	Jasminum officinale	
3.	Achyranthus aspera Linn.	15.	Lantana camara	
4.	Ailanthus excelsa	16.	Launaea procumbens	
5.	Azadirecta indica A. Huss.	17.	Leea macrophylla	
6.	Balanites aegyptiaca	18.	Matrynia annua	
7.	Boerhavia repens	19.	Rosa damascene	
8.	Butea monosperma	20.	Solanum virginianum	
9.	Calatripis procera	21.	Tectona grandis	
10.	Cassia tora	22.	Tinospora cordifolia	
11.	Cyamopsis tetragonoloba	23.	Withania Somnifera	
12.	Dalbergia sissoo	24.	Xanthium strumarium	

Table 2: Mycoflora associated with the leaves of some plants.

Methods used for	Study Ph	yllospere
------------------	----------	-----------

Direct Observation on Stained Leaf	Leaf Wash Method	Spore Fall Method	Leaf Print Method
Alternaria spp.	Alternaria spp.	Alternaria spp.	Alternaria spp.
Cercospora spp.	Aspergillus spp.	Aspergillus spp.	Aspergillus spp.
Curvularia spp.	Cladospora spp.	Cladospora spp.	Cladospora spp.
Helminthospore spp	Curvularia spp.	Collatotricum spp.	Curvularia spp.
Phyllactinia spp.	Fussarium spp.	Curvularia spp.	Helminthospore spp
Uncinulla spp.	Helminthospore spp	Fussarium spp.	Penicillium spp.
Urocystis spp.	Pythium spp.	Helminthospore spp	Phytopthora spp.
Uromyces spp.	, II	Pythium spp.	Pythium spp.

Table 3. Total Fleduelicy of Fully Found II Flightbore Stad	Table 3: Total Free	uency of Funai	Found in Pl	nvllospere Stud
---	---------------------	----------------	-------------	-----------------

Sr. No.	Name of the Fungi spp.	Out of 24 Medicinal plants	
1.	Fussarium spp.	13	
2.	Alternaria spp.	13	
3.	Curvularia spp.	8	
4.	Cladospora spp.	7	
5.	Aspergillus spp.	5	
6.	Phytopthora spp.	3	
7.	Helminthospore spp	2	
8.	Pythium spp	1	
9.	Collatotricum spp.	1	
10.	Phyllactinia spp.	1	
11.	Urocystis spp.	1	
12.	Uromyces spp.	1	
13.	Cercospora spp.	1	
14.	Penicillium spp.	1	
15.	Uncinulla spp.	1	

Table 4: Fungal spp. causing disease to following plants

Alternaria spp.		<i>Fusarium</i> spp.		
Sr. No.	Name of the Plant	Sr. No.	Name of the Plant	
1.	Abesmoschus esculantus	1.	Abesmoschus esculantus	
2.	Acacia leucophloea	2.	Acacia leucophloea	
3.	Ailanthus excelsa	3.	Ailanthus excelsa	
4.	Azadirecta indica A. Huss.	4.	Balanites aegyptiaca	
5.	Balanites aegyptiaca	5.	Boerhavia repens	
6.	Boerhavia repens	6.	Butea monosperma	
7.	Cassia tora	7.	Calatripis procera	
8.	Matrynia annua	8.	Cyamopsis tetragonoloba	
9.	Lantana camara	9.	Lantana camara	
10.	Launaea procumbens	10.	Launaea procumbens	
11.	Leea macrophylla	11.	Solanum virginianum	
12.	Withania Somnifera	12.	Withania Somnifera	
13.	Xanthium strumarium	13.	Xanthium strumarium	

Table V: Alternaria spp.	and Fusarium spp.	causing disease	both on same host

Sr. No.	Name of the Plant	
1.	Abesmoschus esculantus	
2.	Acacia leucophloea	
3.	Ailanthus excelsa	
4.	Balanites aegyptiaca	
5.	Boerhavia repens	
6.	Lantana camara	
7.	Launaea procumbens	
8.	Withania Somnifera	
9.	Xanthium strumarium	

ACKNOWLEDGEMENT

The authors wish to thank Head of Botany Department, Government Institute of Science, Aurangabad for providing the necessary laboratory facilities.

REFERENCES

- Daft, G. C. and Lebenc, (1966). A method for bleaching leaves for microscope investigation of microflora on the leaf surface. *Plant Disease Reporter 50*, 493.
- Dickimonc, H. (1967). Fungal colonization of pisum leaves. *Canadian Journal of Botany* 45, 915-927.
- Dickinsonc, H. (1965). The microflora associated with *Halimione portulacoides*. 111. Fungi on green and moribund leaves. *Transactions of the British Mycological Society* 48, 603-610.
- Diemh, G. (1970). Influence de l'huniidite atmospherique sur la survie de quelques champignons en cours de germination. *Comptes rendirs hebdomudaire des sPances de l'Academie des sciences 270*, 2922-2924.

- Fokkemna, J. (1968). The influence of pollen on the development of *Cladosporium* in the phyllosphere of rye. *Netherland Joirrnal of Plant Pathology* 74, I 59- I 65.
- Kerlipigl, C. P. (1958). De Microflora op het Blad van *Beta vulgaris. Tijdschriff over Planteziekfen* 64, 402-4 10.
- Last, F.T. (1955). The spore content of air within and above mildew infested cereal crops. Trans.Br. Mycol. Appl. 44: 289-294.
- Last, F.T. and Deighton, F.C. (1965). The non parasitic microflora on the surface of living leaves. Transactions of the British Mycological Society, 48: 83-99.
- Naik V.N. 1998."Flora of Marathwada", Amrut Prakashan Aurangabad.
- Rusch V. and Leben C, (1968). Epiphytic microflora : the balloon print isolation technique. *Canadian Joirrnal of Microbiology* 14, 486-487.
- Rutnen J., (1961). The phyllosphere. I. An ecologically neglected milieu. *Plant and Soil* 15, 81-109.
- Waryockd, W. (1973). Origin and development of fungal mycelium in grains of barley before harvest. *Transactions of the British Mycological Society* 61, 49-56.