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Differential utilization of proteins in Anabas testudineus(Bloch) exposed to brief and prolonged fasting

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Abstract

In the present study total proteins were investigated and estimated in *Anabas testudineus* (Bloch) which was exposed to brief (15days) and prolonged fasting (60days).Method developed by Lowry *et.al* (1951) was adopted to estimate the total proteins. Six tissues were selected for the study. The results showed differential utilization of protein in selected tissues. (liver, kidney, brain, accessory respiratory organ, pectoral and lateral line muscle).There was an overall decline observed in the protein levels in most of tissues, with the exception being liver, brain and accessory respiratory organ which showed an increase in protein levels. This increase also was not found to be uniform. Liver and brain showed an elevation only during long term fasting and accessory respiratory organ showed an insignificant increase during both the fasting regimes. This significant depletion of total proteins in most of the tissues of *Anabas* during both the fasting regimes, suggests protein utilization to satisfy the energy demands and to combat the stressful condition and thus prolonging the longevity of the starving *Anabas*.

Keywords: Proteins, Anabas, starvation stress, gluconeogenesis, fasting, hormones

INTRODUCTION

It is interesting and fascinating to note that fishes have an extra ordinary ability to with stand prolonged periods of starvation .Many species of teleosts demonstrate the ability to withstand prolonged periods of starvation (Larson and Lewander, 1973; Loughna and GoldSpink; Machado et.al, 1988). Starvation is known to be tolerated by many species of fish both in and their natural environment during migrations reproduction (Hinch et.al, 2005, Miller et.al, 2009) and fish farming (Miriam furne et.al, 2011). Starvation has certainly been a major factor in natural selection, since the beginning of life. The existing human and animal species is the result of the principle of survival of fittest which survived through countless episodes of food deprivation and shortage (Keyes et.al, 1950).When the crises of starvation hits the body of the fish, there seems to be differential utilization of three energy sources such as carbohydrates, proteins and also lipids so as to overcome the stressful situation. Starvation studies on some species of fish such as Acipenser and Oncorynchus, muscle protein was the last reserve the fish utilized (Navarro and Gutierrez, 1995) and in some other species of fish proteins would be degraded for gluconeogenesis and lipid and or /protein being mobilized as energy substrates.(Sheridan and Mommsen, 1991; Navarro and Gutierrez, 1995; Gillis and Ballantyne, 1996). Starvation studies on Penguins by Mario Spee et.al, (2011) have suggested the existence of three distinct phases during prolonged fasting characterized by lipid utilization during phase one, maintenance of

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Tel:+91-9059681804 Email: anjanipadmavathi@gmail.com constant rate of body mass and protein sparing in phase two and in phase three where animal depends on protein reserves. In another study by P.Vujonic *et.al*, (2011) on starving rats have also expressed a similar opinion stating that as and when the animal approaches the final phase of starvation, it is the proteolytic phase and is marked by simultaneous increase of both blood urea and corticosterone (CORT).A recent study on *Channapunctatus* (Sunnap Namrate *et.al*, 2011) has demonstrated a marked depletion in the protein content in fish which was subjected to starvation stress.

Based upon these studies, it is quite evident that proteins are known to play an important role in satisfying the energy demands of the starving animal .The present study is an attempt to understand if proteins were the preferred fuel over carbohydrates and lipids for *Anabas* during short-term and long term food deprivation. In this context we selected six tissues such as liver, kidney, brain, accessory respiratory organ pectoral and lateral line muscle so as to know if there was a tissue based selective mobilization of proteins during food deprivation.

MATERIAL AND METHODS

Fish weighing of 20-25gm were obtained from Kolleru lake of Eluru. Care was taken to ensure quick transport to the laboratory. Overcrowding was avoided during packing to minimize the mortality rate. They were carefully transferred into Durex storage tank of capacity 500 lits. made of material corrosive resistant polyproplene. The closed plastic lid of the tank was replaced by a grill lid made of iron. This helped in proper ventilation and aeration of the tank. Fish which were injured or dead were removed from the tank from time to time. Disinfectant (KMnO4) was used to avoid infection. They were given boiled egg, rice bran meal and commercial fish feed *ad. libitum.* Any leftover feed and fecal matter were removed daily. Water in the tank was changed every day. Fish were brought to the laboratory and sufficient time was allowed for acclimatization. Experimentation was done thereafter. Fish measuring about 3'-4" in

length and in the same range of weight were selected carefully. They were grouped together and kept in circular tubs made of plastic. The mouth of these tubs was covered with fine mesh and appropriately placed such that they were properly ventilated and well aerated. Two types of experimental set up were designed. In the first set up, fishes were allowed to starve for 15 days and parallel control was also maintained. The control animals were fed regularly both in the morning and evening. On the 16th day both experimental and control animals were sacrificed by concussion, and the tissues were removed for biochemical analysis. The II experimental set up consisted of fishes which were allowed to starve for 60 days (2 months; (long-term). Experimental group and a corresponding control group were maintained. Control group was fed regularly as in the case of short term. On the 61st day, the animals were sacrificed, for experimentation. Six animals from the control group and six from the experimental group were killed by concussion and the tissues were transferred on to ice blocks, weighed, homogenized, and experimented. The tissues selected for the experimentation were liver, kidney, brain, accessory respiratory organ, pectoral and, lateral line muscle.

Estimation of Proteins

Total protein content of the tissue was estimated by the method of Lowry, Farr and Randall (1951).

Procedure: Tissue homogenate was prepared with 10% TCA and centrifuged at 2000 rpm. for 15 minutes. Supernatant was discarded, while the precipitate was dissolved in 0.1 N NaOH(Sodium Hydroxide).To 1ml of sample, 4 ml of reagent "C" (alkaline copper

solution) was added and allowed to stand for 10 minutes at room temperature. Then 0.4 ml of reagent "D" (Follin's reagent) was added rapidly with immediate shaking. After 30 minutes the colour intensity was read in a speckol colorimeter at 540 nm against the distilled water blank. The protein concentration was determined with the help of working standard prepared using albumin and expressed as milli grams of protein / gm wt. of tissue.

RESULTS

Short term starvation stress, for 15 days, lead to a significant depletion in protein levels of all the tissues, but for accessory respiratory organ which showed an increase in the protein content.

Liver showed a decrease of 35.6% (P<0.001). In kidney the decrease was 13.9% (P<0.001). Depletion in brain was 1.8% (NS). In pectoral muscle the decrease was 20.5% (P<0.001). Lateral line muscle showed a decrease of 20.1% (P<0.001) (Tables- I, II&III Figs: 1, 2&3).

Long term starvation stress for 60 days showed a decline in protein levels in tissues like kidney, pectoral muscle and lateral line muscle. Other tissues like liver, accessory respiratory organ and brain showed an elevation in the protein concentration. The decrease in kidney tissue was found to be 25.7% (P<0.001). Pectoral muscle showed a decline of 34.5% (P<0.001). The decrease in lateral line muscle was 27.7% (P<0.001). Liver showed an increase of 40.7% (P<0.01). The increase in brain tissue was 112.8% (P<0.001).Accessory respiratory organ showed an elevation of 1% (NS) (Tables-I, II, III Figs.-1, 2&3).

Table I. Total Protein Levels in Liver and Kidney tissues during Short Term and Long Term starvation in A.testudineus

S.No.	Tissue	Short Term (15 Days)		Long Term (60 Days)	
5.NU.	analyzed	Control	Expt.	Control	Expt.
1.	Liver	6.729	4.33**	3.38	4.758*
		SE = <u>+</u> 0.155	SE = <u>+</u> 0.024	SE = <u>+</u> 0.060	SE = <u>+</u> 0.302
		% Variation = -35.651% Variation			
2.	Kidney	3.117	2.683**	2.124	1.578**
		SE = <u>+</u> 0.044	SE = <u>+</u> 0.04	SE = <u>+</u> 0.064	SE = <u>+</u> 0.048
			% Variation = -13.923	% Variation = -25.706	

Values expressed in mg of protein / gm wt. of tissue Each value is mean of SE \pm of 6 individual observations P < 0.01^{*}, P < 0.001^{**}

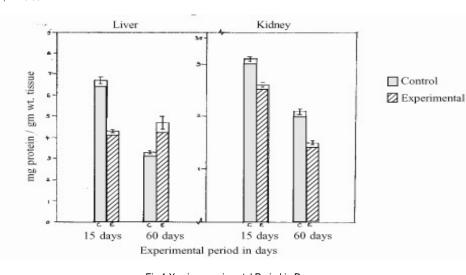


Fig 1.X axis: experimental Period in Days Y axis: mg of protein/gm wt of tissue

Table II. Total Proteins Levels in Brain and Accessory Respiratory	Organ during Short Term and Long Term starvation in A.testudineus

S.No	Tissue analysed	Short Term (15 Days)		Long Term (60 Days)	
		Control	Expt.	Control	Expt.
1.	Brain	2.269	NS	0.745	1.586*
		SE = + 0.051	2.228	$SE = \pm 0.0469$	$SE = \pm 0.0293$
			$SE = \pm 0.036$		
			% Variation = -1.805%	Variation = $+112.88$	5
2.	Accessory	1.055	1.353*	0.753	NS
	Respiratory	$SE = \pm 0.0316$	$SE = \pm 0.0256$	$SE = \pm 0.033$	0.761
	Organ				$SE = \pm 0.024$
	-		% Variation = +28.246	% Variation = +1.062	

Values expressed in mg of protein / gm wt. of tissue

Each value is mean of SE \pm of 6 individual observations

P < 0.001*, NS = Not Significant

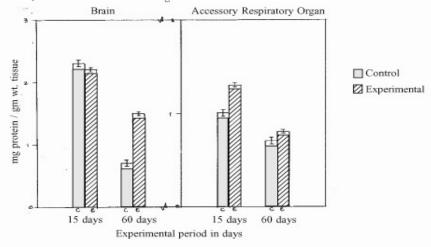
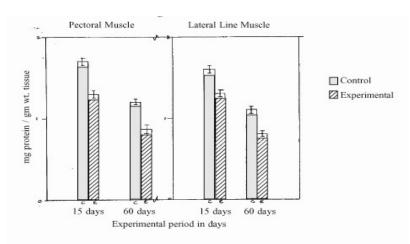


Fig 2.

Table III. Total Protein Levels in Pectoral and Lateral Line Muscles during , short Term and long Term starvation in A.testudineus

S.No.	Tissue	Short Term (15 Days)		Long Term (60 Days)	
3.NO.	analysed	Control	Expt.	Control	Expt.
1.	Pectoral Muscle	1.723 SE = <u>+</u> 0.039	1.369*** SE = <u>+</u> 0.003	1.297 SE = <u>+</u> 0.028	0.849*** SE = <u>+</u> 0.053
		% Variation = -20.545%	Variation = - 34.541		
2.	Lateral Line Muscle	1.681 SE = <u>+</u> 0.045	1.343*** SE = <u>+</u> 0.039	1.183 SE = <u>+</u> 0.048	0.855*** SE = <u>+</u> 0.046
		% Variation = -20.107	% Variation = -27.726		

Values expressed in mg of protein / gm wt. of tissue Each value is mean of SE \pm of 6 individual observations P < 0.001***



DISCUSSION

In the present study we had observed an overall depletion in protein content during both the short-term and long term starvation regimes. Most of the tissues showed a significant decline in protein levels. Tissues such as kidney, and muscle (pectoral and lateral line) showed a significant depletion during both the fasting periods. Accessory respiratory organ showed an increase during both the periods of starvation stress. Strangely, liver and brain showed an increase in protein levels only during long-term starvation.

Liver: When Anabas, was subjected to short term fasting of 15 days, it resulted in the depletion of protein levels. This observation coincides with that of Dave et.al (1975), Larson and Lewender, 1973 and Collins and Anderson, 1995 who have all observed depletion in protein content during starvation in Anguilla anguilla and Golden perch respectively. Starvation studies by different authors such as Storer, (1967), Mann, (1972) Creach and Serfaty, (1974), Larson Lewander (1973), Shantha Vijaya Raghavan, (1988) have all observed an increase in amino acid degrading enzymes such as aminotransferase and asparartate aminotransferase and also elevation in gluconeogenic enzymes such as G6Pase,FDPase and PEPCK was observed by R.Premakumari.(1988) in her starvation experiments in Anabas. All of these observations point to the occurrence of gluconeogenesis so as to provide glucose to the starving animal and the precursors for this process are provided by the protein. Protein catabolism is of prime importance during fasting and conservation of amino acids for protein synthesis becomes secondary (Harper, 1965). In this context of gluconeogenesis, certain hormones such as cortisol, insulin and glucagon are known impact this process. It has been suggested that cortisol excess, insulin deficiency (Bellamy et.al, 1968; Exton, 1972) and altered balance between glucagon and insulin (Cahill, 1966) all are known to enhance gluconeogenesis. Apart from hormones, cyclic AMP is also known to have an impact on glycogenolysis and gluconeogenesis (Jefferson et.al, 1968).

With reference to hormones, insulin in particular, we (P.Godavarthy and Y.Sunila Kumari, 2011) have observed a significant decline in insulin, during both the fasting regimes and perhaps this insulin deficiency may have lead to enhanced gluconeogenesis resulting in decline in the protein content in liver so as to satisfy the energy demands of the starving animal.

However during the long-term food deprivation, the results were different and strangely showed an elevation in protein content .This in accordance with the observations of Bouche and Villas (1975) on starving Cyprinus carpio, which showed a protein elevation, and suggested a simultaneous synthesis occurring along degradation. An increase in protein content during starvation may also be due to conservation of proteins taking place and the starving animal surviving at the expense of glycogen as suggested by Kamara, (1966), Inui and Oshima, (1966) and Patent, (1970), Kerr et.al, (1978). The increase in protein concentration in the present study may be partially due to conservation of protein and the fasting Anabas, deriving its energy from other energy reserves such as glycogen or lipid or a simultaneous synthesis of proteins occurring along degradtaion such as storage proteins which may be needed for gonad maturation as suggested by Brad ford, (1993) and Iles, (1984) in Atlantic herring.

Kidney: Brief and prolonged fasting of *Anabas*, showed a significant depletion in proteins in this tissue. Similar observations have been reported by Premakumari (1988). Her studies on *Anabas*, revealed high rates of protein catabolism with subsequent rise in the amino acid levels. It has been suggested by Exton, (1972) and Krebs (1964) that during prolonged starvation renal gluconeogenesis increases and hepatic gluconeogenesis decreases.

So it may be said that the decline of protein levels in this tissue may be attributed to enhanced gluconeogenesis and thus supplying energy to the starving animal.

Brain: In this tissue during brief fasting (of 15days) we observed, an insignificant decline in the protein levels. However the long term (60days) starvation showed an increase in the protein levels.

Short-term fasting in *Anabas*, led to a significant depletion in the protein levels of brain. This depletion may be attributed to their utilization for the metabolic activity of the brain. According to the studies of Walton and Cowey, (1982) body proteins in fish are in a continual state of turnover, being broken down and re synthesised in varying degrees. The depleted protein levels indicate protein utilization as brain is known to contain all enzymes of major metabolic pathways (Abood *et.al*, (1952); Berl *et.al*, (1962).

Long-term fasting showed an increase which may be explained as a simultaneous synthesis taking place along degradation as suggested by Walton and Cowey,1982. The synthesis of proteins may be for the synthesis of certain important enzymes like glutamine synthetase and glutamate decarboxylase. Glutamine synthetase is known to play a protective role in fish brain and Walton and Cowey, (1982).

Accessory Respiratory Organ: The results from this tissue are surprising, as we have observed an increase both the periods of starvation. The present starvation study on *Anabas*, resulted in elevated levels of proteins, which was not very significant. This slight increase in the protein levels may be due to slow mobilization of proteins. It may also be said that there may be also a synthesis of proteins occurring along their degradation. The enzymes required for amino acid catabolism may be synthesized during starvation. Since enzymes are also proteins, hence there is an elevation in the protein level in this tissue.

Muscle: Both the pectoral and lateral line muscle showed very significant decline in the protein levels during both the fasting regimes. These results coincide with that of Lim and Ip (1989) who have studied the effect of fasting on glycogen metabolism and various glycolytic and gluconeogenic enzymes in the mudskipper (Boleophthalmus boddaerti) and observed a reduction in muscle protein. Similarly, protein depletion is Anabas during food deprivation suggests its utilization for energy needs as protein is known to be the primary energy source during starvation as seen in most of the fish species (Cahn and Woo, 1978a, Larson and lewander, 1973 and Lorven, 1939, Creach and Serfaty, 1974). Starvation causes utilization of proteins via gluconeogenesis after being mobilized to gluconeogenic organs,((Moon, and Johnson, 1980; Black and Love,1986 and Sheridan and Momeson,1991), and both of these processes are influenced by hormones such as glucocorticoids (Cannon et.al, 1956)

CONCLUSIONS

The present study revealed a non uniform depletion in certain tissues and in some tissues a mild elevation in protein content. There was a tissue based selectivity observed in the mobilization and utilization of proteins. In conclusion it may be said that during the short-term and long-term starvation stress of *Anabas*, proteins are being utilized *via* gluconeogenesis after being mobilized to gluconeogenic organs along with other energy sources such as glycogen and lipids. There may be enhanced gluconeogenesis occurring due to insulin deficiency and cortisol excess. The mild increase in proteins in some tissues may be attributed to a simultaneous synthesis of proteins occurring along degradation which may be to produce certain protective enzymes or proteins needed for gonad maturation.

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