Journal of Ecobiotechnology 2/4: 06-10, 2010 ISSN 2077-0464



Effect of Gamma Radiation on Survival and Fertility of Male *Anopheles stephensi* Liston, Irradiated as Pharate Adults

Kavita Yadav, Sunil Dhiman* Indra Baruah and Lokendra Singh

Defence Research Laboratory, Tezpur, India

*Corresponding author, Email: dhiman_81@ indiatimes.com

Keywords	Abstract
Anopheles stephensi Pharate adults Fecundity Fertility	Effect of radiations on the survival and fertility of male <i>Anopheles stephensi</i> Liston was evaluated in the laboratory with 70 and 100 Gy radiation dose. Irradiation of pharate adults with 100 Gy dose resulted in reduced emergence and longevity as compared to control (0 Gy) and 70 Gy dose. Fecundity and fertility of female <i>An. stephensi</i> mated with male irradiated as pharate adults was determined upto four blood meals. Significant decline was observed in fecundity after each blood meal at 100 Gy aradiation. Similarly, the fertility was also decreased after each blood meal at 100 Gy irradiation. The present study indicates the suitability of 100 Gy dose for irradiation of pharate adult stage of mosquito as a part of mosquito management.

1. Introduction

Across the developing world, the public health burden due to vector borne diseases continues to multiply, as current control measures fail to cope with the present scenario. There is an urgent need to identify improved control strategies that could remain effective even in the face of growing insecticide and drug resistance [1]. Ecocompatible biological based control approaches are gaining tremendous attention to tackle mosquito problem at present [2]. One such approach is the Sterile Insect Technique (SIT), which relies on the sterilization of insects, either by chemosterilization [3-5], irradiation [6] or modern biotechnological approaches [7-8]. The biotechnological approaches based on transgenic organisms are promising but are at an early stage of development and no legal framework yet exists to facilitate the introduction of such organisms in the wild [9-10]. Sterilization of mosquitoes by irradiation or chemosterilants has not been researched extensively for about last thirty years [11]. Promising results were obtained with chemosterilants in terms of the level of sterility induced and competitiveness but these have disadvantage of being mutagenic agents. Hence, they present a potential hazard to humans during the treatment process and non-target organisms if residues persist in the released individuals [5].

Malaria is the most important insect transmitted disease affecting 300-500 million people worldwide each year mainly in Africa and south of Sahara. About 2.7 million people are killed either with malaria alone or in combination with other diseases, and over 2400 million remain at risk. In India around 2 million malaria cases at average are reported annually, but the real picture is grossly underestimated due to various logistic lacunae [1]. In India, out of 58 Anopheles described, six namely An. culicifacies, An. dirus, An. fluviatilis, An. minimus, An. sundaicus and An. stephensi have been implicated to be main malaria vectors [12]. An. stephensi, a sub-tropical species is an important urban malaria vector in India, Pakistan and Iran [13]. Many international efforts [14-15] have been launched using SIT despite of scepticism in parts of India's scientific community, with the aim of eradicating An. stephensi. SIT relies on the fact that female mosquitoes mate only once and if this single mating occurs with sterilized males, it results in eggs that do not hatch. So, releasing sterilized males in numbers far greater than those of the wild male population could eradicate the targeted vector mosquitoes provided the area is not re-infested by mosquitoes from the surrounding region. The SIT projects have been developed for An. arabiensis Patton, the malaria vector in Reunion Island and along the Nile in Sudan [15], and might be appropriate for suppression of ecologically isolated urban island populations of Anopheles in India and Nigeria [16]. Control of mosquito vectors using SIT can be achieved more effectively in the small towns or islands, where only one or two kinds of vectors are present [14]. Although the long-term efficacy of SIT in eradicating mosquitoes has not been demonstrated till now, however the success in controlling screw worm, tsetse and mediterranean fruit-flies utilising SIT [17-18] can play a leading role in deploying this technology in malaria control.

The present investigation was aimed to understand the effects of gamma radiation on

2. Materials and Methods

Mosquito culture for experimental studies was obtained from the laboratory colony of *An. Stephensi*, maintained at the NIMR (National Institute of Malaria Research), Delhi, India. The culture was maintained at optimum environment conditions $(27 \pm 1^{\circ}C$ temperature and $75 \pm 5\%$ relative humidity). Irradiation of insects was performed in the Radiobiological unit of INMAS (Institute of Nuclear Medicine and Allied Sciences), Delhi using the ⁶⁰Co source (Gamma - 5000 irradiator, BRIT, BARC, Trombay). The dose rate of radiation was approximately 2.37 KGy/h. Seventy and 100 Gy radiation doses were selected on the basis of earlier work on *Anopheles* sp pupal stage [3, 11].

Male pharate adults (pupae 24 h before emergence) were sexed microscopically, placed on wet cotton in petriplates and transported to radiation source in the ice chest for irradiation dose [11]. After irradiation male pupae were kept in 30 cm³ cages. The mosquitoes that emerged successfully were counted whereas semi-emerged adults and dead adults were scored as non-emerged. The emerged adults were fed on 10% glucose solution in soaked cotton swabs. Longevity and mortality record of the adults was maintained daily till all the adults died.

For assessment of fecundity and fertility irradiated male and non-irradiated female pharate

adults were kept together in 30 cm³ cages after emergence and were provided with 10% glucose solution soaked in cotton swabs along with raisins. Mates were introduced into experimental cages in a 1:1 ratio. Each treatment was repeated at least thrice and 20 males and females each were taken for each replicate. For each cage, one egg bowl filled with water and lined with wet filter paper was offered two days after blood meal. The eggs laid by female mosquito were removed daily and kept in larger trays for hatching after counting [3]. Eggs hatchability was checked by counting first instar larvae (L₁) to monitor the fertility. Fecundity was calculated by dividing the number of eggs laid after blood meal by the number of female alive.

Data analysis: Comparison for fecundity and fertility among 0 Gy, 70 Gy and 100 Gy irradiation dosages and blood meals were carried out using ANOVA. Similarly longevity and emergence percentage among the three dosages were compared using ANOVA. The values in the table are expressed as mean± standard error of mean.

3. Results and Discussion

The fecundity of female mated with 100 Gy irradiated male observed after first, second, third and fourth blood meal was 35.6 ± 3.7 , 29.8 ± 3.9 , 24.7 ± 4.7 and 30.7 ± 4.6 (mean \pm SEM) respectively (table-1). The variation in fecundity among the four blood meals at 100 Gy and 70 Gy was found non significant (p= 0.404, df= 3, F= 1.098 and p= 0.066, df= 3, F= 3.585). On the other hand the fecundity after every blood meal was significantly reduced when irradiated with 100 Gy as compared to control (0 Gy) and 70 Gy (p≤ 0.026, F ≥ 7.113).

Dose (Gy)	After 1 st blood meal (Mean± SEM)	After 2 nd blood meal (Mean± SEM)	After 3 rd blood meal (Mean± SEM)	After 4 th blood meal (Mean± SEM)	F -stat
0 (95% CI)	$56.9 \pm 4.9 \\ (35.9-77.8)$	55.0 ± 5.7 (30.5- 79.5)	55.6 ± 4.6 (35.8- 75.4)	64.2 ± 1.6 (57.0- 71.4)	p= 0.479, F= 0.906
70 (95% CI)	46.1 ± 3.3 (32.0- 60.1)	32.3 ± 2.8 (20.0- 44.6)	44.7 ± 3.5 (29.5- 59.8)	32.7 ± 5.5 (8.8-56.5)	p= 0.066, F= 3.585
100 (95% CI)	35.6 ± 3.7 (19.7- 51.4)	29.8 ± 3.9 (13.1-46.5)	24.7± 4.7 (4.3- 45.1)	30.7 ± 4.6 (10.9-50.5)	p= 0.404, F= 1.098
F-stat	p= 0.026, F=7.113	p= 0.011, F=10.363	p= 0.006, F=13.145	p= 0.002, F=19.404	

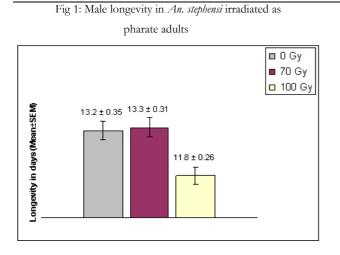
Table: 1. Effect of gamma radiation on fecundity of female An. stephensi mated with male irradiated as pharate adults

SEM = standard error of mean; F = Fisher's coefficient

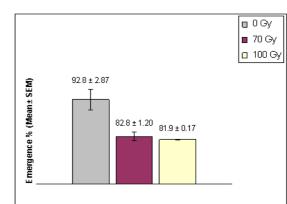
Male longevity (days) recorded at 100 Gy was 11.8 ± 0.264 , which was significantly less than that recorded in control and 70 Gy (p= 0.024, df= 2, F= 7.447) (fig 1). Similarly, male emergence (%) was also reduced significantly after 100 Gy as

compared to control and 70 Gy irradiation (p=0.009, df= 2, F= 11.259) (fig-2).

Fig 2: Male emergence in *An. stephensi* irradiated as pharate adult



The fertility of female mated with 100 Gy irradiated male decreased significantly after each blood meal (p< 0.0001, df= 2, F= 79.188 for first, p< 0.0001, df= 2, F= 126.42 for third, p< 0.0001, df= 2, F= 136.83 for fourth blood meals respectively) as compared to control and 70 Gy irradiation (table-2). Similarly,



fertility after fourth blood meals at 70 Gy was less as compare to other three blood meal (p= 0.026, df= 3, F= 5.312). However no difference was observed in fertility among four blood meals at control and 100 Gy irradiation (p= 0.865, df = 3, F= 0.241 and p= 0.086, df= 3, F= 3.147).

 Table: 2. Effect of gamma radiation on fertility of An. stephensi when normal females were mated with males irradiated as pharate adults

Dose (Gy)	After 1 st blood meal (Mean± SEM)	After 2 nd blood meal (Mean± SEM)	After 3 rd blood meal (Mean± SEM)	After 4 th blood meal (Mean± SEM)	F -stat
0 (95% CI)	74.4± 4.4 (55.64- 93.82)	72.4± 4.6 (52.72- 92.01)	69.6± 4.2 (51.31- 87.81)	72.6± 3.9 (55.41-89.79)	p= 0.865, F= 0.241
70 (95% CI)	22.9± 4.7 (2.57- 43.16)	24.6± 3.2 (10.77- 38.36)	9.1± 3.6 (-6.46- 24.66)	6.96± 4.1 (10.81- 24.75)	p= 0.026, F= 5.312
100 (95% CI)	7.5± 2.3 (-2.29- 12.22)	6.90± 2.0 (-1.75- 15.55)	1.9± 1.3 (-3.72- 7.52)	$\begin{array}{c} 1.9 \pm 1.0 \\ (-2.61 - 6.35) \end{array}$	p= 0.086, F= 3.147
F-stat	P< 0.000, F= 79.188	P< 0.000, F= 97.863	P< 0.000, F= 126.420	P< 0.000, F=136.830	

SEM = standard error of mean; F = Fisher's coefficient

Mosquito control using SIT methods have received very little attention however various SIT based control programmes at small scale have produced good results. In Aedes aegypti radiations have affected the normal insemination and fecundity by inhibiting normal accessory gland activity [19]. The decline in fecundity potential of female mosquito is important as less number of egg are laid, which ultimately decrease the vector density in a particular area of interest. Our results were nearly similar to those obtained elsewhere [11] in which pupal irradiation produced 83.4 % sterility at 70 Gy and 98.6 % sterility at 100 Gy in An. arabiensis. In An. Stephensi, Sharma et al. (1978)³ reported that male pupal irradiation with 80 Gy induced 97.2% male sterility whereas, irradiation of 120 Gy produced 99.1% sterility and reduced fitness of males for mating and survival.

Over the past few years, ionizing radiations have been proved to be a reliable way to induce sterility in variety of insects [6] which strongly favour the exploration and development of sterilization protocols for possible use of SIT against malaria vectors. Most of the earlier work on *Anopheles* irradiation has been focused on the pupal stage [21, 22, 20, 3]. The irradiation of pupae is preferably performed on older (> 15 h) pupae since irradiation of young pupae may result in reduced emergence [21, 23] and shorter survival of the adult [24].

Helinski and co workers (2006)11 have reported that emergence and survival rates of An. arabiensis mosquitoes irradiated as 22-26 h old pupae were similar or reduced as compared to unirradiated control mosquitoes of same species. Most of the studies carried out on mosquito longevity reveal the decrease in male longevity after irradiation at doses of 100-130 Gy [21] and 80 Gy [3]. Likewise, we have observed a considerable decline in both emergence and survival rates. In contrast, a few studies indicate the non significant increase in the longevity of mosquito after irradiation [25]. Irradiations are mainly intended to target the germ cells of subject organism but the irradiation process is highly non-specific and sometime somatic damage may occur. One of the commonest effects of somatic damage seen after irradiation is reduced longevity [21, 23].

4. Conclusion

While discussing SIT for mosquito control programmes, the irradiation is most favourite option for sterilization. The emergence, longevity was decreased significantly, which indicate the possibility to irradiate the pupal stage of mosquito. Further the fecundity and fertility was also reduced considerably using radiation at 100 Gy. The present study suggests the use of a dose range 70-100 Gy to sterilize *An. stephensi* pharate adults in integrated malaria control programmes. However, extensive study results produced in field conditions are necessary to determine the optimum dose and suitable developmental stage for irradiation.

Acknowledgement

The authors gratefully acknowledge NIMR, Delhi for providing quality culture of *An. stephensi* and INMAS, Delhi for providing the irradiation facilities. The help rendered by technical and supporting staff during the irradiation is also acknowledged.

References

- Dhiman, S., Baruah, I. and Singh, L., 2010. Military malaria in northeast region of India. Def. Sci. J., 60 (2): 213-218.
- Yadav, K., Baruah, I. and Goswami, D., 2010. Efficacy of *Bacillus Sphaericus* strain isolated from north east region of India as potential mosquito larvicide. J. Cell Tissue Research, 10(2): 2251-2256.
- 3. Sharma, V.P., Razdan, R.K. and Ansari, M.A., 1978. *Anopheles stephensi:* effect of gamma-radiation and chemosterilants on the fertility

and fitness for sterile male releases. J. Econ. Entomol., 71: 449-452.

- Dame, D.A., Lowe, R.E. and Williamson, D.L., 1981. Assessment of released sterile *Anopheles albimanus* and *Glossina morsitans morsitans*. *Cytogenetics and genetics of vectors*, Editors: Kitzmiller, J.B. and Kanda, T., Amsterdam, Netherlands: Elsevier Biomedical, 231-248.
- Dame, D.A., 1985. Genetic control by sterilized mosquitoes. *Biological Control of Mosquitoes*, 6, Editors: Chapman, R., Barr, R., Weidhaas, D.E. and Laird, M., Am. Mosquito Control Assoc. Bull., 159-172.
- 6. Dyck, A., Hendrichs, J. and Robinson, A.S., 2005. The Sterile Insect Technique: principles and practice in area-wide integrated pest management. Heidelberg, Germany: Springer.
- Thomas, D.D., Donnelly, C.A., Wood, R.J. and Alphey, L.S., 2000. Insect population control using a dominant, repressible, lethal genetic system. Science, 287: 2474-2476.
- Alphey, L., 2002. Re-engineering the sterile insect technique. Insect Biochem. Mol. Biol., 32: 1243-1247.
- Scott, T.W., Takken, W., Knols, B.G. and Boete, C., 2002. The ecology of genetically modified mosquitoes. Science, 298: 117-119.
- Knols, B.G.J. and Louis, C., 2005. Bridging laboratory and field research for genetic control of disease vectors. In Proceedings of the joint WHO/TDR, NIAID, IAEA and Frontis workshop on bridging Laboratory and field research for genetic control of disease vectors. 14–16 July 2004, Nairobi, Kenya.
- Helinski, M.E.H., Parker, A.G. and Knols, B.G.J., 2006. Radiation-induced sterility for pupal and adult stages of the malaria mosquitoes *Anopheles arabiensis*. Malaria J., 5: 41.
- Dhiman, S., Gopalakrishnan, R., Goswami, D., Das, N.G., Baruah, I., Rabha, B., Talukdar, P.K. and Singh, L., 2010. Diversity, spatiotemporal distribution and biting activity of mosquitoes in Tripura State, India. Entomon, 34(4): 1–10.
- Dash, A.P., Adak, T., Raghavendra, K. and Singh, O.P., 2007. The biology and control of malaria vectors in India. Curr. Sci., 92(1): 1571-1578.
- 14. Jayaraman, K. S., 1997. Consortium aims to revive sterile-mosquito project. Nature, 389: 6.
- 15. IAEA, 2001. Feasibility of Sterile Insect Technique for Control of Malaria Vectors

(draft). International Atomic Energy Agency, Vienna.

- Curtis, C.F. and Andreasen, M.H., 2000. Large scale control of mosquito vectors of disease. Area- Wide Control of Fruit Flies and Other Insect Pests (Editor, K.H. Tan), 135-142. Penerbit Universiti Sains, Penang, Malaysia.
- IAEA, 1991. Radiation Induced F₁ Sterility in Lepidoptera for Area-Wide Control. Proceedings of the Final Research Coordination Meeting, Phoenix, Arizona, 9-13 September, Vienna, 162.
- Follett, P. A., 2004. Irradiation to control insects in fruits and vegetables for export from Hawaii. Rad. Phy. Chem., 71: 163-166.
- Leahy, M.G. and Craig, G.B. Jr., 1965. Accessory gland substance as a stimulant for oviposition in *Aedes aegypti* and *Aedes albopictus*. Mosq. News, 25: 448-452.
- Ali, S.R. and Rozenboom, L.E., 1972. Observations on sterilization of *Anopheles (C.) albimanus* Wiedemann by X-irradiation. Mosq. News, 32: 574-579.
- 21. Abdel-Malek, A.A., Tantawy, A.O. and Wakid, A.M., 1967. Studies on the eradication of

Anopheles pharoensis Theobald by the sterile male technique using cobalt-60. III. Determination of the sterile dose and its biological effects on different characters related to fitness components. J. Econ. Entomol., 60(1): 20-23.

- 22. Tantawy, A.O., Abdel-Malek, A.A. and Wakid, A.M., 1967. Studies on the eradication of *Anopheles pharoensis* Theobald by the sterile male technique using cobalt-60. IV. Mating behavior and its frequency in the sterilized mosquitoes. J. Econ. Entomol., 60(1): 23-26.
- Curtis, C.F., 1976. Radiation sterilization. Report on mosquito research. Ross Institute of Tropical Hygiene, 01.01.76-31.12.77.
- Davis, A.N., Gahan, J.B., Weidhaas, D.E. and Smith, C.N., 1959. Exploratory studies on gamma radiation for the sterilization and control of *Anopheles quadrimaculatus*. J. Econ. Entomol., 52: 868-870.
- Abdel-Malek, A.A., Tantawy, A.O. and Wakid, A.M., 1966. Studies on the eradication of *Anopheles pharoensis* Theobald by the sterile male technique using cobalt-60. I. Biological effect of gamma radiation on different developmental stages. J. Econ. Entomol., 59(3): 672-678.

Please Cite This Article As:

Kavita Yadav, Sunil Dhiman Indra Baruah and Lokendra Singh. 2010. Effect of Gamma Radiation on Survival and Fertility of Male *Anopheles stephensi* Liston, Irradiated as Pharate Adults. J. Ecobiotechnol. 2/4:06-10.