

PHYSIO-HISTOLOGICAL STUDIES ON THE
PHYSIOLOGICAL OBESITY OF THE MEAT PIGS (REPORT
I), ESPECIALLY ON THE OCCURRENCE OF THE
GLYCOGEN WITHIN THE NUCLEI AND CYTOPLASMS OF
THE ZONA RETICULARIS IN THE ADRENALS

著者	ITIKAWA Osamu, ISHIDA Kazuo, HOSHINO Tadahiko, TAMATE Hideo, YONEYA Sadamitsu, GOTO Kiko
journal or publication title	Tohoku journal of agricultural research
volume	14
number	4
page range	277-305
year	1963
URL	http://hdl.handle.net/10097/29409

PHYSIO-HISTOLOGICAL STUDIES ON THE PHYSIOLOGICAL
OBESITY OF THE MEAT PIGS (REPORT I), ESPECIALLY
ON THE OCCURRENCE OF THE GLYCOGEN WITHIN
THE NUCLEI AND CYTOPLASMS OF THE ZONA
RETICULARIS IN THE ADRENALS

By

Osamu ITIKAWA, Kazuo ISHIDA, Tadahiko HOSHINO, Hideo TAMATE,
Sadamitsu YONEYA and Kiko GOTO

*Department of Animal Husbandry, Faculty of Agriculture,
Tohoku University, Sendai, Japan*

(Received February 26, 1964)

Introduction

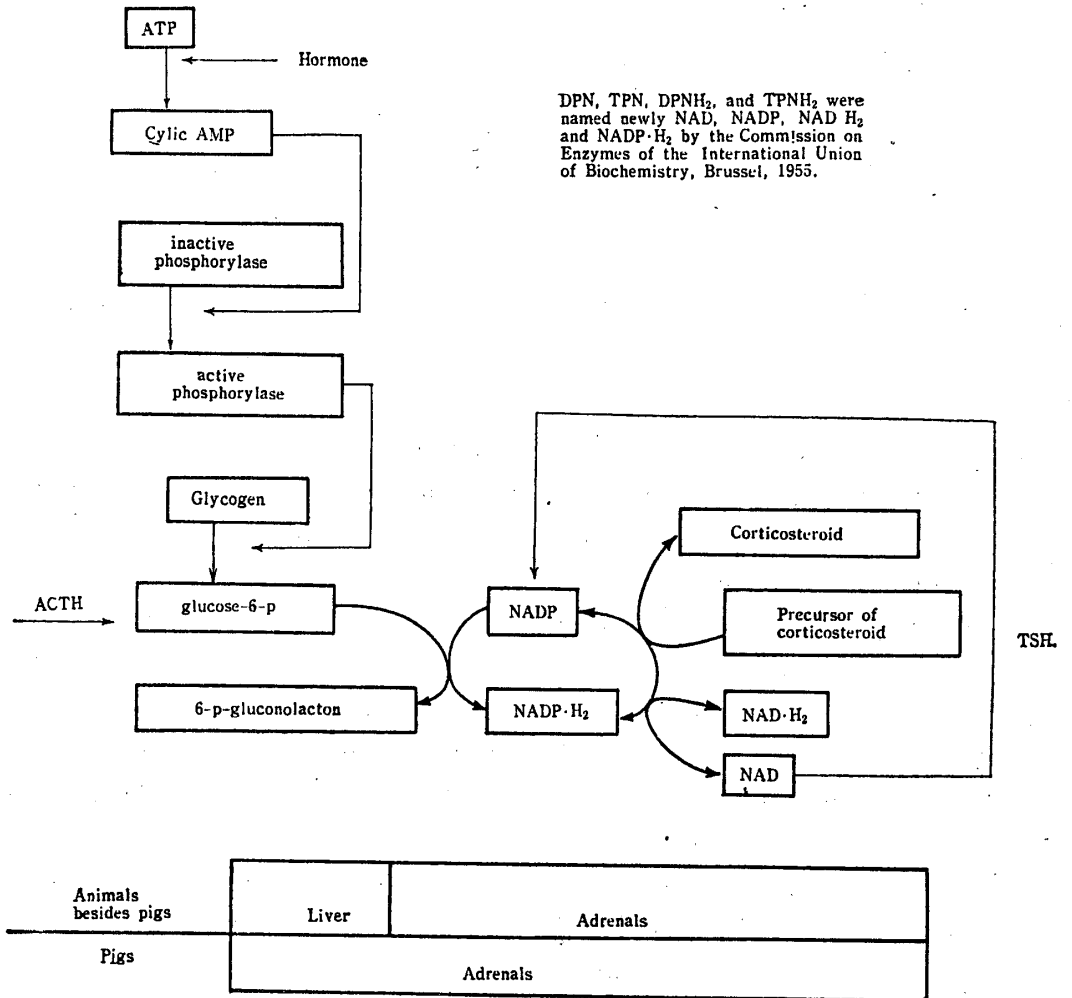
Physio-histological studies on the mechanism of adipositas and the effect of fattening has been done histochemically on the various organs of Yorkshire pigs used for experiment of the feeding standard on the meat pigs. Through these results, meat pigs were considered to have different structure in comparison with the other domestic animals. Accordingly, the pigs might be interesting animals from the point of physiology. Hereafter the authors will report on some problems in connection with the physiological adipositas (1).

By the present study there was found the occurrence of glycogen in the nuclei and cytoplasm of the zona reticularis in the adrenals, and it was planned to examine the mode of glycogen-bearing nuclei, the relationships between glycogen and fat deposition related to the production of ketosteroid, the shape of intranuclear glycogen and the intracytoplasmic one, and the vacuolization in the nuclei.

No one has found glycogen in the nuclei of the adrenals. In the present investigation special attention was also paid to the presence of glycogen in the nuclei and also in the cytoplasm of the adrenal cells of the pigs, because the deposition of glycogen in the cells was demonstrated. With regard to this physiological meaning, it is expected that it will become clarified in the future. However, it has been shown that the working hypothesis as the regulation of the intracellular reaction by the hormone (SUTHERLAND (2-3), 1960 and KONDO (4) 1963) might be established. According to SUTHERLAND's opinion (2-3), there

were found the following metabolic systems: glycolysis from the glycogen to glucose-6-P by the active phosphorylase, transformation of NADP to NADP·H₂ accelerated by the changes of glucose-6-P to 6-P-gluconolactone, and the production system of NADP to NADP·H₂, accordingly these relations are closely connected with glycolysis, coenzymes and steroid hormone (Table 1).

Table 1. Our opinion based on KONDO's hypothesis (1963) with regard to regulation of cellular reactions by the hormone. (ITIKAWA et al 1964)



The chemical reactions shown in Table 1 are related to each other. In the animals besides pigs the glycolysis was done in the liver and the production of corticosteroids in the adrenals. The presence of glycogen in the nuclei and cytoplasm of the swine adrenals found recently by us, might be effective to produce the corticosteroids.

According to WILLIAMS (5) (1962), recent investigations have helped to clarify the mechanisms involved in the synthesis and release of corticosteroids, as follows: the adrenal has a relatively high concentration of glycogen in the

zona fasciculata and zona reticularis, but very little in the zona glomerulosa. The following sequence of events seemed significant in syntheses and secretion of corticosteroids formed in the adrenal. ACTH stimulated formation of cyclic adenylic acid, which, with various cofactors, led to the activation of phosphorylase. Phosphorylase accelerated glycogenolysis, thereby increasing glucose-6-phosphate. Since a very active hexosemonophosphate shunt was present in the adrenal tissue, the amount of glucose-6-phosphate metabolized via this way is increased, with the result that more TPNH was produced. This co-factor accelerated splitting of the cholesterol side-chain and reduction of certain steroids, leading to increased synthesis of corticosteroids. As the food intake increased, the β -cells of the pancreas were stimulated, producing more insulin. This hormone was highly important in lipogenesis, and when it was present in excess it promoted excess deposit of fat, particularly with excess food ingestion. Insulin produced an increase in lipids in each and enlargement of each. Insulin was known to increase the output of glucosteroids, which in turn played an important role in stimulating increased production of insulin antagonists. Obesity tended to produce hyperinsulinism and hyperadrenocorticism, which could in turn significantly further the obesity.

Accordingly, it has been clarified on the mechanism of obesity that hyperadrenocorticism accelerated hyperinsulinism to increase the fat metabolism. To the effect that the pigs were considered as the physiological adipositas, it might be important to observe the relationship between the occurrence of glycogen in the nuclei and cytoplasm of the adrenal cells and the deposition of fat in the cytoplasm of the adrenal cells. These fat-containing cells corresponded to the ketosteroid-producing cells, but the certification of ketosteroid in detail will be described at another occasion.

Materials and Methods for Studies

Eleven sows and nine hogs of Yorkshire were used. Boars were castrated at the age of 23 to 24 days after birth. These hogs and sows were used for 123 days. Animals were studied on the feeding standard by the administration with various feeds such as medium protein-high energetic group (E and F), medium protein-medium energetic group (AC and M) and low protein-medium energetic group (B) in the Miyagi Prefectural Agricultural Experiment Station in Sendai in 1963. The rations of the feed are shown in Table 2:

Pigs in each lot were fed with the ration above given in Table 2. Rations were administered under the dry lot-feeding conditions and the pigs were set for about two hours each day. All the pigs showed an increase of body weight and rapid growth and good appetite. Autopsy was done at the age of 196 days after birth, and the body-weight and feed intake according to sex and experimental periods are shown in Table 3:

Table 2. Ration used for the experiment

Exp. groups		Period	Former period					Later period				
		Feeding	E	F	AC	B	M	E	F	AC	B	M
Administered Feeds	Concentrated combined feed : Barley feed		6 : 4	6 : 4	7 : 3	7 : 3	7.2:2.8	6 : 4	6 : 4	7 : 3	7 : 3	7.2:2.8
	Basal ration	yellow corn	36	37	16	18	25.2	40	39	20	20	28.0
		Bran	6	2	20	11	4.2	8	2	22	11	5.6
		Defattend rice-bran	2	1	7	8	1.4	2	1	7	8	1.4
		Soybean oil meal	7	3	6	3	4.9	3	1	2	1	2.1
		Fish meal	3	3	4	3	2.1	1	1	2	1	0.7
		Starch meal	—	8	11	21	—	—	10	11	23	—
		Others	6	6	6	6	4.2	6	6	6	6	4.2
Barley	40	40	30	30	28	40	40	30	30	28		
Alphalpa meal					30						30	

Table 3. Body weight and feed intake of the pigs according to sex and experimental periods.

Lots	Sex (Cases)	Age (days)			Body weight (kg)				Feed intake(kg) during all periods			Remarks
		Initial	Middle period	Final period	Initial	Former period	Middle period	Final period	Former period	Final period	Whole period	
E	♀ (2)	70	140	196	22.5	50.7	73.2	114.7	318	307	625	Medium protein-High energetic ration
	♂ (2)	70	140	196	20.6	49.7	70.2	109.4	307	360	667	
F	♀ (2)	70	140	196	23.0	44.3	67.3	97.4	306	283	589	Medium protein-High energetic ration
	♂ (2)	70	140	196	21.5	53.5	75.0	114.8	331	338	669	
AC	♀ (2)	70	140	196	21.0	44.1	65.1	100.6	297	343	640	Medium protein-Medium energetic ration
	♂ (2)	70	140	196	22.7	52.8	75.5	120.6	357	443	800	
B	♀ (2)	70	140	196	21.8	39.8	61.6	98.1	286	360	646	Medium protein-Medium energetic ration
	♂ (2)	70	140	196	22.3	48.6	70.9	112.5	322	385	707	
M	♀ (2)	70	140	196	21.3	44.9	66.2	108.1	531	478	1009	Low protein-Medium energetic ration with alphalpa-meal
	♂ (3)	70	140	196	20.2	43.0	63.2	99.7	514	452	966	

The total adrenals from these sows and hogs were fixed either in buffered formol or in CARNOY's fluid, embedded in paraffine and cut into 6 μ sections, and cut with frozen microtome for fat-staining. The stains employed were as follows: Hematoxylin eosin stain and CROSSMAN's stain for general histology; PAS-hematoxylin stain with or without saliva digestion for the glycogen; and FEULGEN's nuclear reaction counterstained with light green for th DNA.

Results

1. Glycogen occurred in the cytoplasm and nuclei, and vacuolized giant nuclei.

According to OKAMOTO (6) (1958), there are no reports on the presence of glycogen in the cortex of adrenals. However, WILLIAMS (5) (1962) described that the recent investigations have helped in the mechanisms involved in the synthesis and the release of corticosteroids as follows: relatively high concentration of glycogen in the zona fasciculata and zona reticularis, plentiful presence of glucose-6-P dehydrogenase in the zona fasciculata and zona reticularis and etc.

It was noticed that a large amount of the nuclei and cytoplasm of the swine adrenals showed a strong positive reaction to PAS stain. This reaction

Table 4. The occurrence of glycogen-laden nuclei and cytoplasm, and that of the vacuolized giant nuclei in the zona reticularis of swine adrenals

Sex	Cortex	Feeding			Medium protein-High energetic group				Medium protein-High energetic group					Low protein, Medium energetic	
		Cytoplasmic Glycogen	Nuclear Glycogen	Group	E		F		AC		M			B	
					Name	22	24	11	27	21	7	2	14	25	1
Sows	Z. reticulares	+	+	-	-	○1	-	○1	-	-	○1	○6	○7	◎29	-
		-	+	-	○5	○9	○14	○3	◎35	○12	○3	◎592	◎26	◎52	○
		+	-	+	-	-	-	-	-	-	○	○	○	○	○
		+	-	-	◎467	◎792	○65	○76	○84	○92	◎134	○63	○72	○69	◎225
	-	-	+	-	-	○6	-	○2	○	○14	◎97	○16	○16	○4	
	Zona fasciculata*	-	-	-	-	-	-	-	-	-	◎	-	◎	-	
Zona glomerularis*	-	-	-	-	-	-	-	-	-	-	-	-	-		
					28	29	4	5	12	23	13	15	3	26	6
Hogs	Z. reticularis	+	+	-	-	-	-	○4	-	○8	○8	-	-	◎140	-
		-	+	-	○14	◎22	◎478	◎29	-	◎99	◎63	○1	◎118	◎39	
		+	-	+	-	-	○	○	missing	○	○	missing	○	○16	○
		+	-	-	◎468	○28	○21	◎1872	missing	◎196	◎639	missing	◎53	◎80	◎397
	-	-	+	○12	○1	◎62	○1	missing	○7	○12	missing	○3	◎115	○7	
Zona fasciculata*	-	-	-	-	-	-	-	-	○	◎	-	-	-		
Zona glomerularis*	-	-	-	-	-	-	-	-	-	-	-	-	-		

Remarks: * indicated the presence of intranuclear glycogen, ◎, ○, ○ in the table showed the degree, and the numbers added to right corner showed the cell numbers, and—no glycogen.

became negative in the sections previously treated with saliva for less than one hour at 37°C. This indicated that the PAS-positive substance in the nuclei was glycogen. Eleven sows and nine hogs contained the glycogen-laden nuclei of the cells in the zona reticularis or zona fasciculata in 100 percentage. There were found various types as follows: glycogen in both cytoplasm and nuclei; no glycogen in the cytoplasm and glycogen in the nuclei; glycogen in the cytoplasm and no glycogen in the nuclei; no glycogen in both cytoplasm and nuclei, and vacuolized giant nuclei with no glycogen. Usually the glycogen deposition in the nuclei was shown in zona reticularis at 100 percent, but in zona fasciculata at 18 percent of 11 sows and at 22 percent of nine hogs. No intranuclear glycogen was found in zona glomerularis. These results were indicated as shown in Table 4, and the cell counts of the glycogen-laden nuclei in one section are shown in the table. The presence of glycogen in the cytoplasm and nuclei seemed to have no relation to the difference with the feeding ration, but it might be related to the sex from the point of more intensive appearance in the hogs than that in the sows.

2. The shape of intranuclear and intracytoplasmic glycogen deposition.

Up to this time it was known that there were no glycogen besides DNA, RNA and histon in the ordinary nuclei. With the exception of the hepatic cells or pancreatic β -cells in diabetes melitus the presence of glycogen was found in these cells. However, recently TORYU *et al* (7) (1960) of Institute reported that a large amount of glycogen was demonstrated histochemically in the cells of the anterior lingual salivary glands of adult laying hens, but not in adult cocks. The occurrence of the glycogen-laden nuclei in the glands was also investigated in growing female chicks and adult female chicks during laying and moulting (TORYU *et al*) (8-9) (1961). They also investigated the presence of glycogen-laden nuclei of the male chicks of from 2 to 180 days of age (TORYU *et al*) (8-9) (1961).

The presence of glycogen in the nuclei of swine adrenals were found to be remarkably stronger or more intensive in the hogs than in the sows. Almost all of the intranuclear glycogen present in the juxta medullary zone of zona reticularis adjoins the medullary, and also a large amount of glycogen in the nuclei existed in the boundary zone (temporary cortex or central body). In these regions lipids deposited remarkably.

In severe cases there occurred a large amount of glycogen in the nuclei, this extended to the deep zone of zona fasciculata, and also to the spongy zone. Generally the cells in the zona reticularis were arranged in an irregular network and they possessed fat droplets and glycogen in the cytoplasm. The cells indicated a network of round chain-like appearance in the juxta medullary

zone, and that of strand-like appearance in the boundary zone. A large part of the glycogen-laden nuclei were giant in size, but some nuclei were normal.

The size of the normal and abnormal nuclei with or without glycogen in the zona reticularis was various types as shown in Table 5.

Table 5. The size of glycogen-laden nuclei and no glycogen-laden nuclei in some cells of zona reticularis in the swine adrenals

cyto- plasms	Nuclei				Size (length and width, square microns)						
	Glycogen	Glycogen	Vacuolization	Normal	(3.97-5.0) × (5.0-7.15) μ	(6.35-7.15) × (5.0-7.15) μ	(7.95-8.7) × (4.76-8.7) μ	(9.5-10.3) × (4.76-10.3) μ	(11.0-12) × (8.7-12) μ	(13-13.5) × (9.5-13.5) μ	(14-19.2) × (16-19.2) μ
-	+	+	-	-	•	•••	••	•		•R	
-	-	+	-	-		•••			•••	•	
-	+	-	+	-				•	•••		
-	+	-	-	+	••••••	••••••	••••		•		
-	-	-	-	+	••	••••	•••				
-	+	+	-	-	••I••••	••	••	••••	••	••	•R
-	+	-	+	-			•••G••G••R••	•••	••R•R	••R	•
-	-	-	+	-					••	••	
-	+	-	-	+	•	•			•••	•	
-	-	-	-	+	•	•	•				

Remarks : I. indicated internal bodies ; R. ring form ; and G. granule form.

Giant nuclei with glycogen were four-five times as large as the normal nuclei. There were various types of intranuclear glycogen as follows: large entire bodies (glycogen occupied entirely in the nuclei), ring form (glycogen deposition near nuclear membrane and empty cavity), multiple granule form (spreading diffusely as the granular bodies in mixture to chromatin), and internal bodies (round bodies, bottle form, and funnel form in the nuclei). The development of these types of intranuclear glycogen deposition will be described in another paper. There existed hyperchromic or hypochromic nuclei which reacted to PAS with glycogen.

The nuclei had hydrops recognized as a simple vacuole in the hematoxylin-eosin stain and these glycogenic vacuolization was similar to that in the diabetes. The vacuolized nuclei without glycogen in the PAS-stain were called "vacuolization in the giant nuclei. Glycogen might be lost from the ring form of nuclei which contained a small amount of glycogen around the nuclear membrane. According to TORJU *et al* (7) (1960) the process of the nuclear vacuolation and the migration of the chromatin granules towards the nuclear membrane, were also ascertained in the sections stained by FEULGEN's technique or by basic dyes.

Various forms of the nuclei were arranged in an order according to their shape and the amount of the intranuclear glycogen. The nucleus, which might

be taken as "normal", was approximately $6.0\ \mu$ in diameter, containing the chromatin granules and nucleolus. They contained no glycogen. When the vacuoles became larger, they nearly occupied the entire space of the nucleus, pushing aside all chromatin granules towards the nuclear membrane. Consequently, the chromatin granules were found attached to the membrane. Glycogen deposition occurred in such nuclei. They consisted of the one called the entire bodies with glycogen and the other called the ring form which glycogen demonstrated by PAS stain was limited in the largest vacuoles occupying most of the nuclear space. They consisted of the two forms called the large entire bodies and the small entire ones. A few glycogen granules, small in size, appeared in the periphery of the vacuoles close to the chromatin granules attached to the nuclear membrane or near the dispersed chromatin granules in the nuclei. They were called the multiple granular form of glycogen-laden nuclei. The size of the glycogen granules remarkably increased, united with each other, and they formed internal bodies in the whole vacuolar cavity. And then the whole vacuolar cavity was filled with glycogen. They are called the large and small entire bodies.

The occurrence of the glycogen-laden nuclei was examined in zona reticularis (sometimes in zona fasciculata) of the adrenals. Variation of the nuclei containing glycogen and that of the glycogen-laden cytoplasms are shown in Table 6. Whole cells in the zona reticularis were counted in the sections stained by PAS-hematoxylin stain using the square-meter

The variation of the intranuclear or intracytoplasmic glycogen deposition and the glycogen-free vacuolization in the nuclei, was divided into the under-mentioned six types by means of observing the combination of intracytoplasmic and intranuclear glycogen deposition and the vacuolized giant nuclei without glycogen:

- a) The cells contained glycogen in both cytoplasms and nuclei.
- b) The cells contained the glycogen-laden nuclei and glycogen-free cytoplasms.
- c) The cells contained the glycogen-laden cytoplasms and the vacuolized nuclei without glycogen.
- d) The cells contained the glycogen-laden cytoplasms and normal nuclei without glycogen and vacuoles.
- e) The cells contained the vacuolized giant nuclei and the glycogen-free cytoplasms and nuclei.
- f) The cells seemed to be normal and contained no glycogen-laden cytoplasms and nuclei and no vacuolized giant nuclei.

Type a happen at the same time in the severe cases of type b (M-14, B-1 in sows and F-5, AC-21, B-26, M-13 in hogs, see in Table 6). The appearance of nuclear glycogen in the sixth type was found in all of cases. In the inten-

sive cases there occurred 548/1 section of the adrenal (M 14 ♀) or 478/1 section (F4 ♂) at maximum and in the slight cases 4-5/1 section or only 1/1 section (M1 ♂) at the minimum. On the average there occurred 67/1 section in sows and 113/1 section in hogs. These types had a tendency to have no glycogen in the cytoplasm. Almost all of type c did not appear, but only one (B 26 ♂) of 20 cases existed. In a word the cells contained the vacuolized giant nuclei had no glycogen in the cytoplasm. On the contrary in type a occurred the glycogen-laden cytoplasm and nuclei, was found in the six cases of 11 sows (54%) and in the five cases of nine hogs (55%). It is of interest to find that M-3 (♂) seemed to be a resting type which occurred in only type a.

Type a consisted of a small amount of large and small entire bodies and internal bodies, but type b contained a large amount of internal bodies and multiple granular bodies and a small amount of large and small entire bodies. It might be related to various phases in the carbohydrate metabolism.

Intracytoplasmic glycogen deposition was divided into the following two forms: the cells contained a large amount of glycogen in the whole cytoplasm, called the entire glycogen; the cells contained a small amount of glycogen in the locus near the nuclear membrane as the form of granular aggregates or large globule (it might be related to GOLGI apparatus), called the bodies. Diffuse form stained lightly with PAS or occurred as a very minute granules, was omitted from the cell-count. The adrenals contained entire glycogen (E 24 ♀, F 5 ♂, E 28 ♂, M 2 ♀ and B 6 ♂) had a few glycogen-bodies, but the adrenals contained a large amount of glycogen-bodies (E 22 ♀ and B 28 ♂) and had a few entire glycogen. However the intermediate forms contained both entire glycogen and glycogen-bodies, and occupied a large part of the adrenals.

In the cells occurred the vacuolized giant cells which were indicated as the large vacuole. Some cases (M 14 ♀, F 4 ♂ and B 26 ♂) contained a large amount of these cells, but the other cases only a small amount of them. In the cases in which there occurred a large amount of the vacuolized giant nuclei, there was found the severe occurrence of the intranuclear glycogen deposition. Generally, the vacuolized giant nuclei were indicated in the 63 per cent of 11 sows and in the 100 per cent of nine hogs, but the intensity of the degree was parallel to the occurrence of the intranuclear glycogen.

The degree of occurrence of Type a, b, c, d and e to type f (normal cells contained on glycogen) is shown in Table 6. In short, there were found at the average in one section as follows: nuclear glycogen 67, cytoplasmic glycogen 200, vacuolized giant nuclei 10 and glycogen-free nuclei and cytoplasm 12230 in the adrenal of the sow; nuclear glycogen 113, cytoplasmic glycogen 424, vacuolized giant nuclei 24 and glycogen-free nuclei and cytoplasm 17540 in the adrenal of the hog. From this results there might have occurred more intensive

glycogen in the cells of the sows than of the hogs.

3. The relationships between the fat-stored cells and glycogen-laden cells in zona reticularis.

With regards to the histochemistry on the corticosteroids of the adrenals many papers have been published, but they seemed to be not valuable because of coloring under the unpurified states of corticosteroids as the tissue sections. According to many reports that demonstrated the presence or localization of steroids up to now, the nature of the products in the staining reactions consisted of the following substances: plasmal substance (LISON 1953) (10), pseudo-plasmal substance (CHU 1950) (11), peroxide (CAIN 1949) (12), unsaturated fatty acids (BAYLEY 1945) (21), enzyme (GOMORI 1950) (13), unknown unsaponified substance (UI 1957) (14) and cholesterol (SCHULTZ 1924) (15).

MAYEDA (1962) (20) described on some problems in the corticosteroids staining of adrenals as follows: it is indirectly possible to presume the presence of steroids in the sudanophilic granules by means of experiments. According to YOSHIMURA (1962) (21) who mentioned doubt as to the ketosteroid-staining from the side of cytology, he asserted that it is dangerous to consider the barrier of steroids in the fat granules, and it is important to observe the hormonal barrier in the protein granules with the so-called masked fat as the precursor. NAKAO (1962) (19) found a new method which he called $SbCl_3$ -reaction to demonstrate corticosteroid, and he stated the antimony-granules to be reactive substance in the sections. However, there remained some doubt in this histochemical regions. However, owing to the correspondence to the sudanophilic part and $SbCl_3$ -reactive part, there were considered sudanophilic cells which contained the ketosteroid, and indicated the significance and localization of the fat-stored cells shown in this report. In future the relationships between the sudanophilic granules, $SbCl_3$ -granules, and glycogen-laden nuclei and glycogen bearing cytoplasm, may be discussed in other papers.

In the present investigation the fat-stored cells are shown in the tables 7, 8 and 9). The following opinions are summarized from these tables:

a) When the cells of zona reticularis contained a large amount of glycogen laden nuclei and cytoplasm, this zone tended to enlarge by means of the increase of cells.

b) There were divided into two types of rich fat-stored cells and poor fat-stored cells in the zona reticularis with no enlargement.

c) The ratio of nuclear glycogen to cytoplasmic glycogen was various, but they were divided into three types such as 1:1 (0.03-1), 1:50 (2-49) and 1:100 (53-93). In the zona reticularis of the ratio 1:1, the cell number of glycogen-laden nuclei was more abundant than that of glycogen-stored cytoplasm. In the ratio of 1:100, the cell number of glycogen-laden nuclei was

Table 9. Relationships between glycogen- and fat-metabolism in the cells of zona reticularis of swine adrenals

Sex	Changes	Medium Protein-Feeding						Medium Protein-Medium Energetic Feeding						Low Protein Med. Energetic Feeding		
		High Energetic Feeding			F			AC			M			B		
		E	24	11	27	21	7	2	14	25	1	8				
♀	Nuclear glycogen	5	10	14	4	35	12	4	548	33	71	6				
	Cytoplasmic glycogen.	467	792	65	76	84	92	134	63	72	69	225				
	Vacuol. giant nuc lens.			6		2		14	47	16	16	4				
	Free glyc. in nuc. and cytopl.	6709	4935	8325	10179	8930	19267	6762	32770	17200	10800	8680				
	Fat-stored cells	8851	4586	4370	2409	7919	6656	2271	201	321	1832	1021				
	Total cells in z. retic.	16032	10323	12780	12668	16960	26028	9185	33629	17642	12788	9936				
	Enlargement of z. retic.	⊙	⊙	⊙	⊙	⊙	⊙	⊙	⊙	⊙	⊙	⊙				
	Nucl.-Glyc. : Cyt-Glyc	1:93	1:79	1:5	1:19	1:2	1:8	1:34	1:0.1	1:0.1	1:1	1:38				
	Nucl.-Glyc. : Free Glyc. Vac.	1:0	1:0	1:0.4	1:0	1:0.1	1:0	1:4	1:0.1	1:0.5	1:0.2	1:0.7				
	Nucl.-Cyt. Glic. : Free Glyc.	1:14	1:6	1:105	1:127	1:75	1:183	1:50	1:53	1:164	1:78	1:37				
	Fat stored cells	8851	4586	4730	2409	7919	6656	2271	201	321	1832	1021				
	Free Fat Stor. Cells	7181	5737	8410	10259	9051	19371	6914	33428	17321	10956	8915				
	Fat Cells : Free Fat Cells	1:0.8	1:1.2	1:2	1:4	1:1.1	1:3	1:3	1:166	1:54	1:6	1:8				
	Appearance of fat strage	⊙	⊙	⊙	⊙	⊙	⊙	⊙	⊙	⊙	⊙	⊙				
Sex	Changer	28	29	4	5	12	23	13	15	3	26	6				
♂	Nuclear glycogen	14	22	478	33	107	71	71	258	1	34					
	Cytoplasm. glycogen	468	28	21	1872	196	689	689	87	53	397					
	Vacuol. giant cells	12	1	62	1	7	12	12	115	3	7					
	Free glycogen in nuc. cytopl.	13220	12027	25922	22457	28737	13289	13289	17408	12497	16800					
	Fat. stored cells	1020	4996	7157	6025	174	2876	2876	5331	2872	641					
	Total cells in z. retic.	14734	17074	33140	38388	29221	16937	16937	23199	15426	17879					
	Enlargement of z. retic.	⊙	⊙	⊙	⊙	⊙	⊙	⊙	⊙	⊙	⊙					
	Nucl.-glyc. : cyto. glyc.	1:33	1:1	1:0.04	1:57	1:2	1:10	1:10	1:0.3	1:53	1:0.3	1:12				
	Nucl.-glyc. : free glyc.	1:1	1:0.05	1:0.1	1:0.03	1:0.07	1:0.2	1:0.2	1:0.4	1:3	1:0.4	1:0.2				
	Nucl.-cyt. glic. : Free glyc.	1:27	1:40	1:51	1:12	1:94	1:17	1:17	1:52	1:230	1:39	1:39				
	Fat stored cells	1020	4996	7157	6025	174	2876	2876	5331	2872	641	641				
	Free fat stored cells	13714	12078	25983	24363	29047	14061	14061	17868	12554	17238	17238				
	Fat cells : free fat cells	1:13	1:2	1:3	1:4	1:170	1:5	1:5	1:3	1:4	1:3	1:27				
	Appearance of fat storage	⊙	⊙	⊙	⊙	⊙	⊙	⊙	⊙	⊙	⊙	⊙				

Remarks : Nucl. indicated Nucleus or nuclear ; cytopl. or cyt, cytoplasm or cytoplasmic ; vac. or vacuol., vacuolized ; z. retic., zona reticularis ; glyc, glycogen ; ⊙ high or strong degree ; ⊕ medium degree ; ○ low or weak degree.

more scanty than that of glycogen-contained cytoplasm.

d) The ratio of the nuclei with glycogen to the vacuolized nuclei without glycogen was various, but there were divided into three types such as 1:0 (0~0.03~0.7), 1:1 and 1:4 (3-4). In the ratio of 1:0 were indicated no appearance of vacuolized giant nuclei without glycogen, and the ratio of 1:4 showed a small amount of nuclear glycogen in comparison with a large amount of cytoplasmic glycogen.

e) The ratio of glycogen in the cells (nuclei and cytoplasm) to no glycogen in the cells was various, but they were divided into three types such as 1:20 (6-17); 1:50 (21-94) and 1:100-200 (105-240). In the ratio of 1:20 were indicated the appearance of a large amount of nuclear and cytoplasmic glycogen in comparison with the whole cells.

f) The adrenals contained a large amount of the fat-stored cells which tended to have a few glycogen-bearing cells, and on the contrary the ones contained a few fat-stored cells tended to have a large amount of glycogen-bearing cells.

From these result it is important to find that the relation between nuclear and cytoplasmic glycogen was close and also the relation between cytoplasmic glycogen and fat-storage existed.

In coparison with the cell counts of the fat-stored cells and the free fat-stored cells, the difference was as shown in the following Table 7, 8 and 9. These ratio was various as 1:0.8~1:166 (average 1:23) in the sows, but it was 1:2~1:170 (average 1:14) in the hogs. Accordingly the hogs might contain more plentiful fat-stored cells than the hogs. There existed at the average value of 2766 of the fat-stored cells and that of 11689 of the free fat-stored cells in the zona reticularis of the sows; and 3452 of the fat-stored cells and 15838 of the free fat-stored cells in the hogs.

The combination of the fat-stored cells, glycogen in the cytoplasm and nuclei, and vacuolized giant nuclei are shown in Tables 8 and 9 and the cells in the zona reticularis were divided into 12 types as Table 10:

Table 10. Various types of the cells in the zona reticularis of the sows and hogs

Type	Cytoplasmic Fat	Cytoplasmic Glycogen	Nuclear glycogen	Free glycogen in cytoplasm	Normal nuclei	Vaccuolized giant nuclei
a	+	+	+	-	-	-
b	+	-	+	+	-	-
c	+	+	-	-	-	+
d	+	-	-	+	-	+
e	+	+	-	-	+	-
f	+	-	-	+	+	-
g	-	+	+	-	-	-
h	-	-	+	+	-	-
i	-	+	-	-	-	+
j	-	-	-	+	-	+
k	-	+	-	-	+	-
l	-	-	-	+	+	-

In the sows there were found a large amount of type l and f, and secondly abundant in types e and k; but in the hogs a large amount of types e, f and k, and next abundant in types h and g. Accordingly the function in the hogs might be accelerated in comparison with that in sows.

Summary and Discussion

Physio-histological studies on the mechanism of adipositas and the effect of fattening has been investigated histochemically on the various organs of Yorkshire pigs used for experiment of the feeding standard on the meat pigs. During these investigations there has been found the occurrence of glycogen in the nuclei and cytoplasm of the zona reticularis in the adrenals of the pigs. No one has found glycogen in the nuclei of the adrenals.

The present study described the frequency of occurrence of glycogen-bearing cytoplasm and nuclei, their shape and form, and the relationship between the glycogen-laden cells and fat stored cells. The results investigated are summarized as follows:

1) It was noticed that a large amount of the nuclei and cytoplasm of the swine adrenals showed a strong positive reaction to PAS-stain. This reaction became negative in the sections previously treated with saliva digestion. Accordingly this indicated that the PAS-positive substance in the nuclei and cytoplasm was glycogen. Eleven sows and nine hogs contained the glycogen-laden nuclei of the cells in the zona reticularis in the 100 per cent. In the animals which contained remarkable large amounts of the glycogen-laden nuclei there were found also in the zona fasciculata 18 percent of the sows and 22 percent of the hogs and none in the zona glomerulosa.

2) The appearance of intranuclear glycogen seemed to be not related to the feeding, but it was clear that it is related to the sex. The intranuclear glycogen tended to appear more intensive in the hogs than in the sows.

3) The nuclei contained glycogen generally were giant in size, but some glycogen-laden nuclei were small. Usually the size of the normal nuclei of the cells in the zona reticularis was $5.3 \times 5.3 \mu$ in diameter, but that of glycogen-laden nuclei arranged in the range of $8.0 \times 8.0 \mu$ to $19.0 \times 19.0 \mu$ in diameter as the giant form. At first there occurred glycogen like form of granules or bodies in the normal-sized nuclei, and then gradually they developed to large nuclear bodies increasing the storage of glycogen. Afterwards there appeared the internal bodies, funnel like bodies and ring form of glycogen in the vacuolized giant nuclei. At last the vacuolized giant nuclei without glycogen appeared by the decrease of glycogen.

4) The modes of the intranuclear glycogen deposition were various as follows: glycogen occupied in the whole large or small nuclei (large entire bodies or small entire bodies), the glycogen attached to the nuclear membrane

and the internal space was empty (ring form), round internal bodies in the vacuolized nuclei (internal bodies), the flask-like bodies, club-like bodies, dumb-bell-like bodies or funnel-like bodies attached to the one part of nuclear membrane within the vacuolized giant nuclei (one kind of internal bodies), several granules or rods contained glycogen near the chromatin dispersed within the nuclei (multiple granular form), hyperchromic or hypochromic glycogen-laden nuclei by the stainability of PAS-stain.

The vacuolized nuclei with free glycogen divided to giant nuclei and small nuclei. It was possible to presume that the vacuolized nuclei lost glycogen from the ring form.

5) The modus of the intracytoplasmic glycogen were divided into three types as follows: a large amount of glycogen in the entire cytoplasm (entire glycogenic form), glycogen deposition formed the round solitary islet near the nuclear membrane (glycogenic bodies), and a small quantity of glycogen in the cytoplasm (diffuse form).

6) On an average in the section of one zona reticularis of the sows, 67 of intranuclear glycogen deposition, 200 of intracytoplasmic glycogen deposition, 10 of vacuolized glycogen-free giant nuclei, 12230 of no glycogen deposition in both nuclei and cytoplasm were found. On the contrary, in the hogs, 113 of intranuclear glycogen, 424 of intracytoplasmic glycogen, 24 of vacuolized nuclei, and 17540 of free glycogen in both nuclei and cytoplasm were existed. From these results the zona reticularis of the hogs seemed to contain more plentiful glycogen than that of the sows.

7) Sudanophilic granules existed in the cytoplasm of zona reticularis of adrenals. The authors desired to consider the sudanophilic cells as a ketosteroid-bearing cells because of the correspondence to the stainability of sudanophilia and antimony granules (NAKAO) in the cells of zona reticularis. Accordingly in the present investigation the numbers of the fat-stored cells were calculated, but this problem will be diversified in another paper. On the average, in one section of adrenal of the sow there were existed 2766 of the fat-stored cells and 1538 of the free fat-stored cells. From this point the zona reticularis of the hogs seemed to contain fat-storage more plentiful than that of the sows.

8) From the points of the numbers of cells in the whole zona reticularis, the enlarged zone contained a large amount of cell-numbers with a large quantity of glycogen-laden nuclei and cytoplasm. The non-enlarged zone was divided into two types such as a rich type and a poor type of the fat-stored cells. The rich type of the fat-stored cells tended to have a few glycogen-laden cells, and in opposition the poor types of fat-stored cells tended to have many glycogen-laden cells.

In short, it is important to observe that the intranuclear glycogen related to the cytoplasmic glycogen, and the cellular glycogen (in both nuclei and

Table 11. Relationships between cellular glycogen and fats in the zona reticularis of the pigs

sex	Cytoplasm		Nuclei	Vacuol- ized giant nuclei	Medium Protein High Energic Feeding				Medium Protein Med. Energic Feeding					Low Protein Medium Energic	
	Fats	Glyco- gen	Glyco- gen		E		F		AC		M			B	
					22	24	11	27	21	7	2	14	25	1	8
Sows		+	+	-	-	○	-	○	-	○	○	○	○	○	○
		-	+	+	○	○	○	○	○	○	○	○	○	○	○
		+	-	-	◎	◎	◎	◎	◎	◎	◎	◎	◎	◎	◎
		-	-	-	◎	◎	◎	◎	◎	◎	◎	◎	◎	◎	◎
	+	-	-	-	◎	◎	◎	◎	◎	◎	◎	◎	◎	◎	◎
					28	29	4	5	12	23	13	15	3	26	6
Hogs		+	+	-	-	○	-	○	○	○	○	○	○	○	○
		-	+	+	○	○	○	○	○	○	○	○	○	○	○
		+	-	-	◎	◎	◎	◎	◎	◎	◎	◎	◎	◎	◎
		-	-	-	◎	◎	◎	◎	◎	◎	◎	◎	◎	◎	◎
	+	-	-	-	◎	◎	◎	◎	◎	◎	◎	◎	◎	◎	◎

Remarks, ○ 1-20, ◎ 21-100, ⊙, 101-200-400 of cells.

cytoplasm) were related to the fat-stored cells in inverse proportion.

As the recent biochemical theories of the regulation of cellular reaction by the hormone owing to SUTHERLAND (1960), WILLIAMS (1962) and KONDO (1963) have published, they state that the production of ketosteroid and glycogenolysis in the adrenals have been becoming the interesting problems to study the mechanism of fattening or physiological obesity.

WILLIAMS (1962) described that the recent investigations have helped the mechanisms involved in the synthesis and release of corticosteroid as follows: relatively high concentration of glycogen in the zona fasciculata and zona reticularis, plentiful presence of glucose-6-P dehydrogenase in the zona fasciculata and zona reticularis, and the others. The following sequence of events seemed significant in synthesis and secretion of corticosteroids from the adrenal. ACTH stimulated the formation of cyclic adenylic acid, which, with various co-factors, led to the activation of phosphorylase. Phosphorylase accelerated glycogenolysis, thereby increasing glucose-6-phosphate. Since a very active hexosemonophosphate shunt was present in the adrenal tissue, the amount of glucose-6-phosphate metabolized via this way was increased, with the result that TPNH (same as NADH₂) was produced. This co-factor accelerated splitting of the cholesterol side-chain and the reduction of certain steroids, leading to increased synthesis of corticosteroids. As the food intake increased, the β-cells of the pancreas were stimulated, producing more insulin. This hormone was highly important in lipogenesis, and when it was present in excess it pro-

moted an excess deposit of fat, particularly with the excess food ingestion. Insulin produced an increase in lipids in each and an enlargement of each. Insulin was known to increase the output of glucosteroids, which in turn played an important role in stimulating the increased production of insulin antagonists. Obesity tended to produce hyperinsulinism and hyperadrenocorticism, which can in turn significantly further the obesity.

Our present report described the facts that played a role of understanding morphologically for biochemical studies on the mechanisms of regulation in the cellular reaction by the hormone as the above mentioned.

The relationships between glycogen in the cytoplasm and nuclei and fat in the cytoplasm, in the cells of zona reticularis of the pigs are shown in Table 11.

In the cases of 11 sows and 9 hogs there were observed the presence of glycogen in the cytoplasm and nuclei, glycogenolysis to glucose-6-phosphate, and the increase of fat related to the ketosteroids, so it seemed to be closely related to the physiological obesity. The production of insulin was accelerated by the production of ketosteroid. If the obesity would develop by the hyperinsulinism and hyperadrenocorticism, the glycogen deposition in the cytoplasm and nuclei might play an important role to hyperadrenocorticism. In this meaning, it seemed to be important to have the glycogen-deposition in the cytoplasm and nuclei of the cells in the zona reticularis of the swine adrenals. Moreover it is very important to solve histochemically the problems of co-factor and corticosteroid, and this way become the subject for future investigation.

Acknowledgement

The present authors wish to express their thanks to Mr. T. YOSHIMOTO and others of the Miyagi Prefectural Agriculture Experiment Station for giving them the materials used in this study, to Mr. A. ITIKAWA, Faculty of Pharmacology of the Tokyo University, for his valuable advice during the course of this study, and to Miss. Y. KAMIOKA of their laboratory for her technical assistance.

References

- 1) Itikawa, O. (1963). *Bulletin of Tohoku Branch of Jap. Soc. Zootechn. Sci.*, 14(1), 2 (in Japanese)
- 2) Haynes, Jr. R.C., Sutherland, E.W. Rall (1960). *Recent Progress in Hormone Research*, 16, 121.
- 3) Ratt T.W., Sutherland, E.W. (1961). *Symposia on quantitative Biology*, 26, 347.
- 4) Kondo, Y. (1963). *Science(KAGAKU)*, 33(5), 242—248 (in Japanese).
- 5) Willams, R.H. (1962). *Textbook of Endocrinology*, Third ed., P. 968, W.B. Saunders Co., Philadelphia and London, 1962.

- 6) Okamoto, K. (1958). *Histochemistry of Endocrine Glands*, Kyodo Isho Publisher, Tokyo, P. 42—48 (in Jappense).
- 7) Toryu, Y., Hoshino, T., and H. Tamate (1960). *Tohoku J. Agr. Res.*, **11** (4), 309—317.
- 8) Toryu, Y., Hoshino, T., and H. Tamate (1961). *Archiv. hist. jap.*, **21** (4—5), 463—468.
- 9) Toryu, Y., Hoshino, T., and H. Tamate (1961). *Jap. J. Zootechn. Sci.*, **32**(3), 164—171.
- 10) Lisos, L. (1960). *Histochemie et cytochimie animales*, 3 ed., Gauthier-Villars and Co., Paris.
- 11) Chu, C. H. U. (1950). *Anat. Rec.* **108**, 723.
- 12) Cain, A. J. (1949). *Quart. J. Mci.*, **90**, 75.
- 13) Gomori, G. (1950). *Ann N. Y. Acad. Sci.*, **50**, 968.
- 14) Ur, H., Kawamura, M, and M. Inaba (1957). On the prosperity and decay of unsaponified substance in some organs of the rat, Pharmacological Laboratory of Jikei Medical College, 1957. citrated NAKAO 's papers (16) (in Japanese)
- 15) Schultz, A (1924). *Zbl. Path.*, **35**, 314.
- 16) Nakao, T., and M. Hirai (1959). *Jikei Med J.*, **6**, 25, (in Japanese).
- 17) Nakao, T. and M. Hirai (1959). *Jikei Med. J.*, **6**, 43 (in Japanese).
- 18) Nakao, T. and M. Hirai (1960). *Jikei Med. J.*, **7**, 31 (in Japaneie).
- 19) Nakao, T. (1962). *Advances to Recent Medicine (Saishin Igaku)*, **17**(2), 344—356. (in Japanese)
- 20) Maeda, R. (1962). *Advances to Recent Medicine (Saishin Igaku)*, **17**(2), 323—330. (in Japanese)
- 21) Yoshimura, F. (1962). *Advances to Recent Medicine (Saishin Igaku)*, **17**(2), 339—343 (in Japanese).

Plate 1**Explanation of Figures**

Fig. 1. Small entire bodies of glycogen-laden nuclei, and vacuolized giant nuclei with the intracytoplasmic glycogen Hematoxylin stain, $\times 400$.

Small entire bodies (mark : s), vacuolized giant nuclei (v), vacuolized giant nuclei with the intracytoplasmic glycogen (v-w), and fat-stored cells (f).

Fig. 2. Fat-stored cells in the wide regions of zona reticularis. Hematoxylin-eosin stain, $\times 400$.

Fat stored cells(f) with a large amount of fat and no glycogen.

Fig. 3. Glycogen-laden body in the cytoplasm of the cells of the zona reticularis. Hematoxylin-eosin stain, $\times 400$.

Glycogenic bodies were similar to GOLGI's apparatus(B).

Fig. 4. Intracytoplasmic glycogen-laden cells and fat-stored cells. Hematoxylin-eosin, $\times 400$.

Intracytoplasmic glycogen-laden cells(G) with no glycogen in the nuclei.

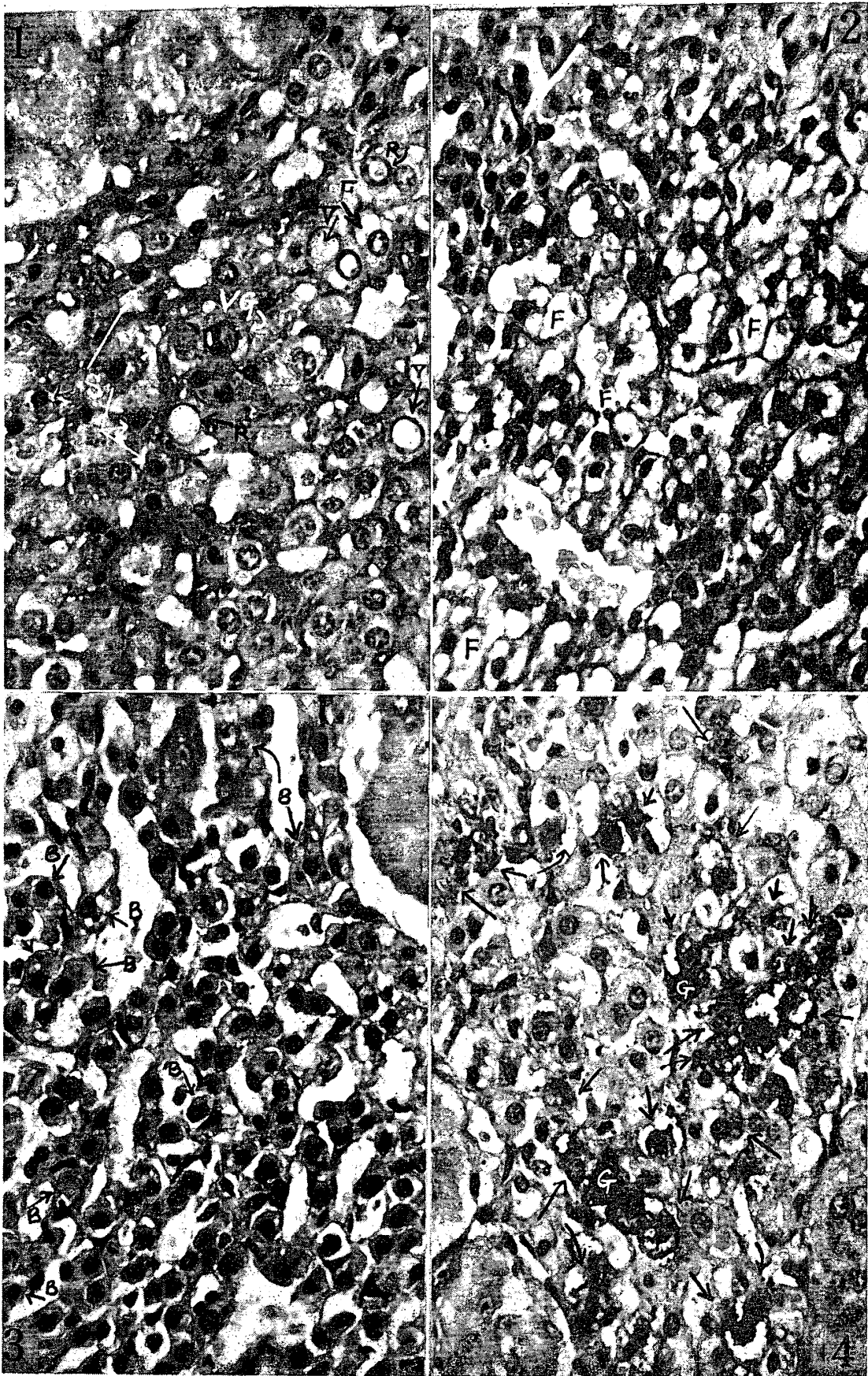


Plate 2**Explanation of Figures**

Fig. 5. Inner bodies of the glycogen-laden nuclei. Hematoxylin-eosin, stain, $\times 400$.

The inner bodies (I), multiple granules (MG), small entire bodies (S) in the glycogen-laden nuclei. The cells with the inner bodies of nuclei, contained the glycogen-laden cytoplasms.

Fig. 6. Multiple granules of the glycogen-laden nuclei. Hematoxylin-eosin stain, $\times 400$.

Multiple granules of the glycogen-laden nuclei (MG) appeared in large amount.

Fig. 7. Cytoplasmic glycogen deposition. Hematoxyline-eosen strin, $\times 40$.

The glycogen deposition in the cytoplasms. These glycogen-bealing cells(G) are indicated by the arrow.

Fig. 8. Cytoplasmic glycoqdn deposition in the zona reticularis. Hematoxylin eosin, $\times 40$.

The glycogen deposition (G) in the cytoplasm and inner bodies of glycogen-laden nuclei (I), fat-stored cells with the cytoplasmic glycogen (FG), glycogen deposition in both nuclei and cytoplasm (NCG), fat-stored cells (F).

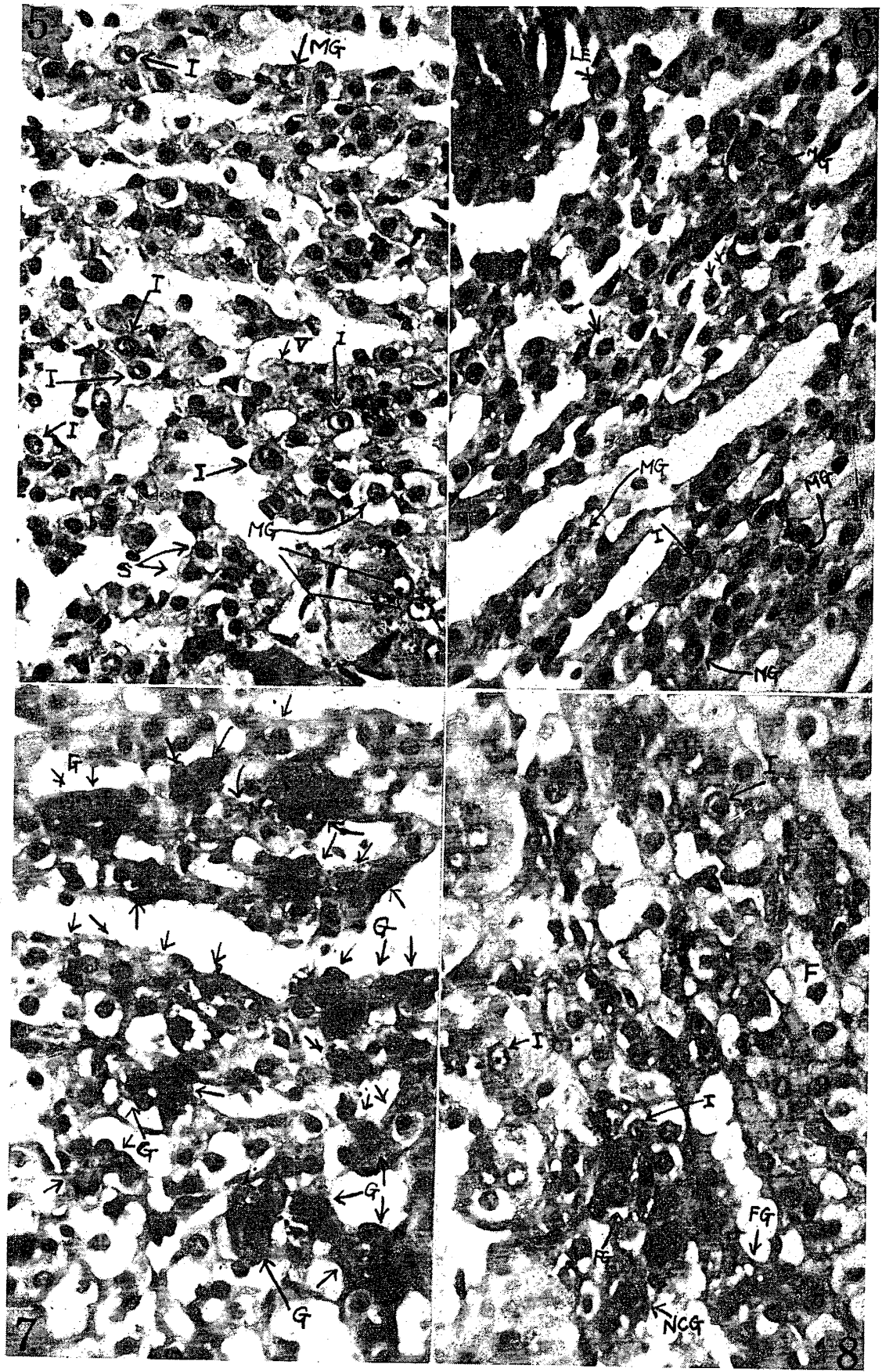


Plate 3**Explanation of Figures**

- Fig. 9. Small entire bodies or small vacuolized nuclei. PAS-Hematoxylin-stain $\times 900$.
Small entire bodies with glycogen(S), small vacuolized nuclei and intracytoplasmic glycogen(SV-G), and multiple granular forms with nuclear glycogen(NG).
- Fig. 10. Multiple granular forms with nuclear glycogen. PAS-Hematoxylin-stain, $\times 900$.
This type of intranuclear glycogen is rodlike(R). Some cells contained glycogen in both nuclei and cytoplasm(NC).
- Fig. 11. Multiple granular forms with nuclear glycogen. PAS-hematoxylin stain, $\times 900$.
Multiple granular forms(MG) with nuclear glycogen were various. Some nuclei consisted of granular glycogen(MGCH) near chromatin, and others formed irregular confluent masses with glycogen(MGC). The cells with these nuclei contained no glycogen in the cytoplasm.
- Fig. 12. Multiple granular forms and small entire bodies with nuclear glycogen. PAS-hematoxylin stain, $\times 900$.
In the cells of multiple granular forms(MG) and small entire bodies(S) with nuclear glycogen.
- Fig. 13. Ring forms with nuclear glycogen near the nuclear membrane. PAS-hematoxylin stain, $\times 900$.
Glycogen attached around the nuclear membrane(R) like ring form, and the intranuclear spaces enlarged in the form of vacuole. Those cells contained fat. Some nuclei contained minute glycogen granules near the chromatin and formed vacuoles in the center. One of the ring form showed a flask-like process within the vacuole from the membrane(F).
- Fig. 14. Large entire bodies with band and vacuole(LE), and fat deposition in the cytoplasm(F).
- Fig. 15. Inner bodies of nuclear glycogen with cytoplasmic glycogen. PAS-hematoxylin stain, $\times 900$.
Three cells contained glycogen in both nuclei and cytoplasm, and formed inner bodies in the nuclei. Two cells also contained fats.
- Fig. 16. Small entire bodies of nuclear glycogen. PAS-hematoxylin stain, $\times 900$.
The small entire bodies(S), ring form(R), and the small entire bodies of nuclear glycogen with fat-storage(SF).
- Fig. 17. Large entire bodies of nuclear glycogen and multiple granular forms of nuclear glycogen. PAS-hematoxylin stain, $\times 900$.
Large entire bodies(L) and multiple granules(MG) with nuclear glycogen which contained fat droplets.
- Fig. 18. Large entire bodies, ring form and flask-like process with ring form. PAS-Hematoxylin stain. $\times 900$.
Large entire bodies(L), ring form(R) and flask-like process with ring form(FR).
- Fig. 19. Large entire bodies, multiple granular forms and cytoplasmic bodies. PAS-Hematoxylin stain, $\times 900$.
Large entire bodies(L), multiple granular form(MG) and intranuclear glycogen-cytoplasmic bodies(B).
- Fig. 20. Large bodies in the cytoplasm and ring form of nuclei. PAS-Hematoxylin stain, $\times 900$.
Large bodies in the cytoplasm(LB) and ring form of nuclear glycogen(R).

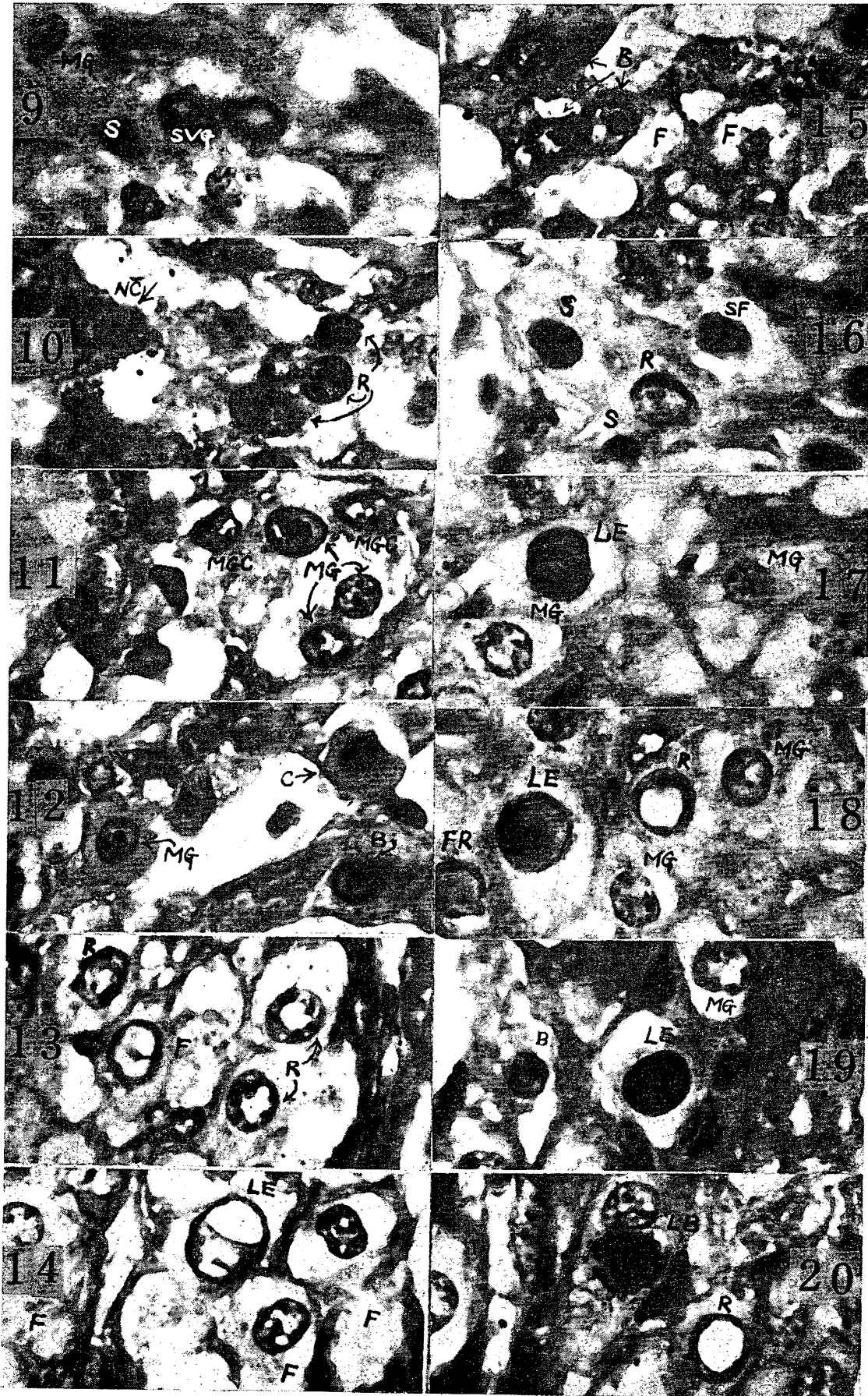


Plate 4**Explanation of Figures**

Fig. 21. Intranuclear glycogen deposition and fat or glycogen-storage in the cells of zona reticularis. PAS-Hematoxylin stain, $\times 200$.

Intranuclear glycogen deposition (shown by arrow), fat-storage(F) or glycogen-storage(G).

Fig. 22. Intranuclear glycogen deposition and fat-storage in the cells of zona reticularis. PAS-Hematoxylin-stain $\times 900$.

Intranuclear glycogen deposition (NG), fat-storage (F) and fat-glycogen-storage with nuclear glycogen(FGN), and cellular bodies in the cytoplasm(B).

Fig. 23. Ring form of nuclear glycogen and fat-storage. PAS-Hematoxylin, $\times 900$.

Ring form of nuclear glycogen(R) and fat-storage (F).

Fig. 24. Polysaccharide-bodies in the cytoplasm of the fat-stored cells. PAS-Hematoxylin stain, $\times 900$.

Polysaccharide-bodies (PB) in the cytoplasm of the fat-stored cells (F).

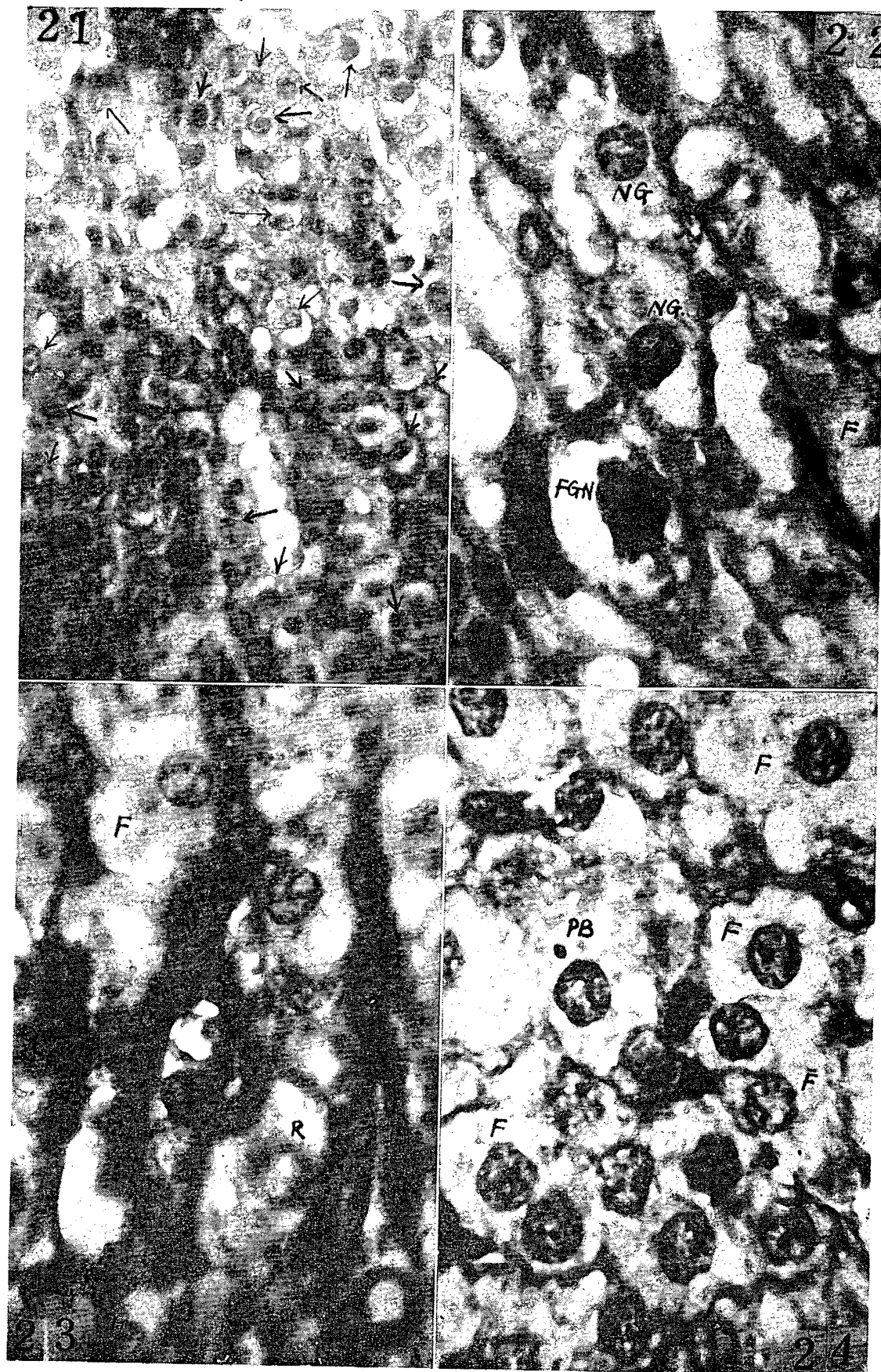


Plate 5**Explanation of Figures.**

Fig. 25. A large amount of fat-stored cells in the zona reticularis and zona fasciculata. Sudan III-Hematoxylin stain, $\times 200$.
Blackish cellular strangs with fat-stored cells.

Fig. 26. Fat-stored cells and vacuolized giant nuclei. Sudan III-Hematoxylin stain, $\times 300$.
Fat-stored cells shown by arrow-mark and the vacuolized giant nuclei (V).

Fig. 27. A little quantity of fat-stored cells in the zona reticularis. Sudan III-Hematoxylin stain, $\times 200$.
This microphoto showed no fat-stored cells.

Fig. 28. Free fat-stored cells in the zona reticularis. Sudan III-Hematoxylin stain, $\times 400$.
A few cells contained a little quantity of fat granules in the cells.

