

Distribution of Recent Benthic Foraminiferal Faunas in the Pacific off Southwest Japan and Around Hachijojima Island

著者	Akimoto Kazumi
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Kazumi Akimoto

ABSTRACT

The distribution of benthic foraminifera in the Northwest Pacific Ocean off the coast of southwest Japan is investigated in order to establish a tool for analyzing paleoenvironments of Miocene strata over the Japanese Islands.

In this study, four areas of the Pacific are investigated; these are Tanegashima, Kumanonada, Enshunada, and Hachijojima areas, and are located in those oceanic regions which are currently under the influence of Kuroshio Current.

The method of Q-mode cluster analysis is applied to the foraminiferal distribution in each area in order to categorize foraminiferal assemblages characteristic of these four areas.

The results obtained are as follows:

1) The Tanegashima and Kumanonada areas each have eight types of benthic foraminiferal assemblages, whereas the Enshunada has five. Six types of assemblages also characterize the Hachijojima area.

2) The distribution of these assemblages is basically controlled by the oceanographic condition of water masses overlying each area.

3) The distribution of 13 species, which are familiar elements among the Miocene foraminiferal fauna of the South Fossa Magna region, is delineated.

Key words: benthic foraminifera, Recent, Pacific, Japan, distribution

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INTRODUCTION

This study intends to establish faunal assemblages and relate their distribution with various environments in order to obtain basic data for zonation and paleoenvironmental analysis by means of fossil benthic foraminifera.

Investigations were made in four areas under the influence of the Kuroshio

Current: off Enshunada (EN), off Kumanonada (KU), off Tanegashima Island (TA), and around Hachijojima Island (HA) (Fig. 1). Systematic research has been hitherto made by three workers. Ishiwada (1964) first studied Recent benthic foraminifera around Japan including Tosa Bay. Aoshima

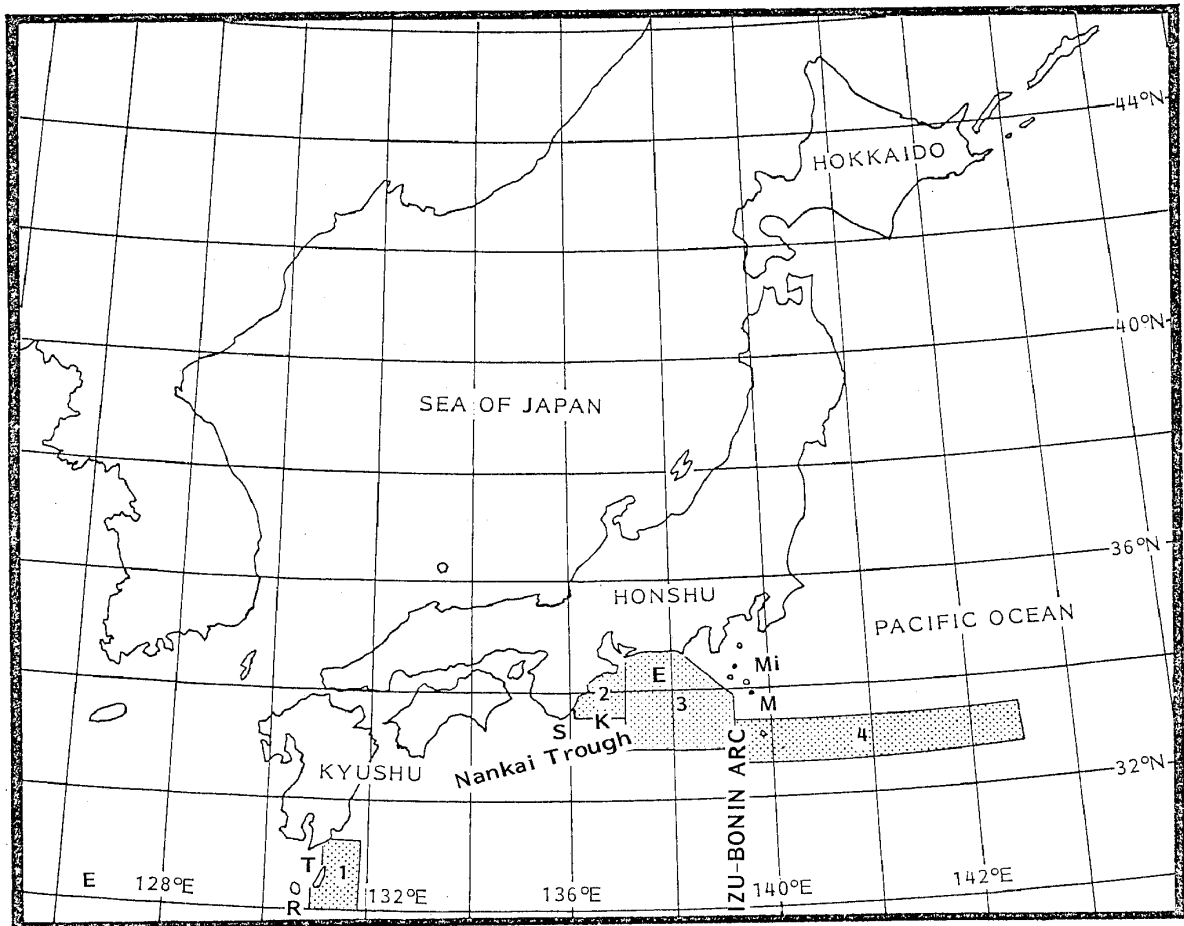


Fig. 1. Index map of study areas. (1, Tanegashima area; 2, Kumanonada area; 3, Enshunada area; 4, Hachijojima area; E, East China Sea; R, Ryukyu Arc; T, Tanegashima Island; S, Shionomisaki Cape; K, Kumanonada, En, Enshunada; M, Mikurujima Island; Mi, Miyakejima Island).

(1978) revealed the distribution of Recent benthic foraminiferal species in five traverses off southwest Japan, and recognized four assemblages on the basis of their depth range. Inoue (1986) reported 11 benthic foraminiferal assemblages at depths from 40 m to 2,000 m in Suruga Bay. Those previous studies, dealing with material taken from up to the middle bathyal zone, do not provide an enough basis for analysis of bathyal

faunas. Uchio (1952a) described several species in Recent beach sand on Hachijojima Island, but did not fulfill systematic studies in a traverse. Therefore, it is necessary to make clear the distribution of Recent benthic foraminiferal assemblages in traverses across the Izu-Mariana Arc and southwest Japan for performing a paleoenvironmental study in the South Fossa Magna region.

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Samples used in this study were provided from the collections of the Ocean Research Institute of the University of Tokyo and the Geological Survey of Japan.

Cluster analysis was processed with NEAC-ACOS Model 77/700 computer in the Tohoku University Computer Center.

METHOD OF STUDY

Field work

Sediment samples were collected at 25 stations in the Tanegashima area (Fig. 2), 14 in the Kumanonada area (Fig. 3), 16 in the Enshunada area (Fig. 4), and 18 in the Hachijojima area (Fig. 5). All samples were taken by various gears such as grab sampler, including Smith-McIntyre sampler piston corer and dredger. The examined samples represent the uppermost 2 cm of collected sediment columns except dredged one. In the Tanegashima, Enshunada (mostly KT sample series), and Hachijojima areas (KT sample series), an approximately 5% solution of neutralized formalin was used. A small amount of sodium carbonate was added to each sample to maintain alkalinity.

Laboratory work

All the samples were washed over a screen with an average opening of 0.063 mm (250-mesh) and rose Bengal stained to detect living specimens (Walton,

1952). The rose Bengal solution was prepared after Matoba (1970).

Kuwano (1956) verified the validity of the rose Bengal staining method, and clarified relations among the concentration of the rose Bengal solution, the time required for staining and conditions of stained specimens. For counting living specimens, he proposed to use a dry method instead of Walton's wet method. After staining, the sediment samples were dried before counting living specimens. Although there may be difficulties and errors in distinguishing between living and dead specimens, dried samples are more handy.

The dried sample was divided by a splitter into aliquot parts of workable size from which approximately 200 specimens of both living and dead benthic foraminifera were counted. All the specimens including planktonic foraminifera were picked from the split-part and counted.

SUBMARINE TOPOGRAPHY

1. Tanegashima area

Figure 2 shows submarine topography in this area. This figure delineates that the continental shelf is widely distributed, extending 30–50 km eastward in the north, but narrow off the southern part of Tanegashima Island. The continental slope develops off the eastern coast of Tanegashima Island, and extends to the Nankai Trough in the east. There are small continental breaks on the slope.

Two channel systems such as the Nakatane and Minamitane canyons also develop on the continental slope off Tanegashima Island. Their roots are at the shelf edge, and their tips reach to the Nankai Trough. Terrigenous and shallow water sediments have been displaced down to deep areas through those channels. The Osumi Strait is situated between the Osumi Peninsula and Tanegashima Island, and has a maxi-

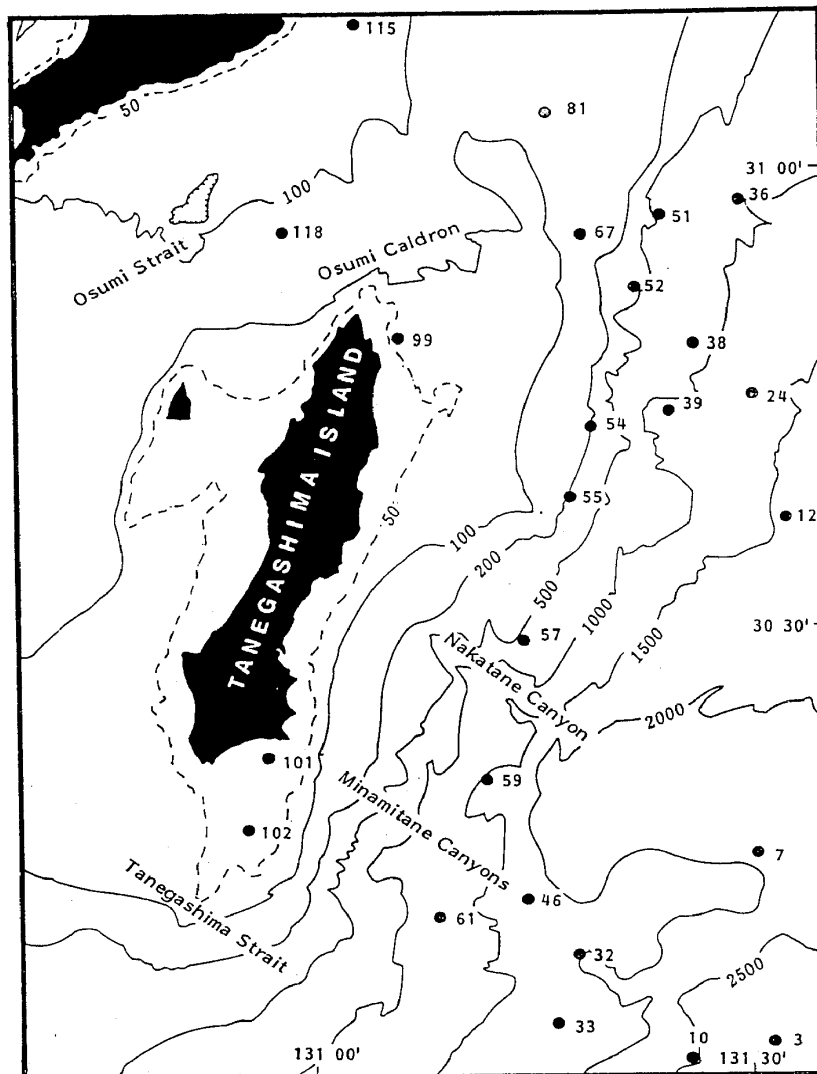


Fig. 2. Stations in the Tanegashima area of foraminiferal samples used in this study. Closed circle, Smith-McIntyre type sampler.

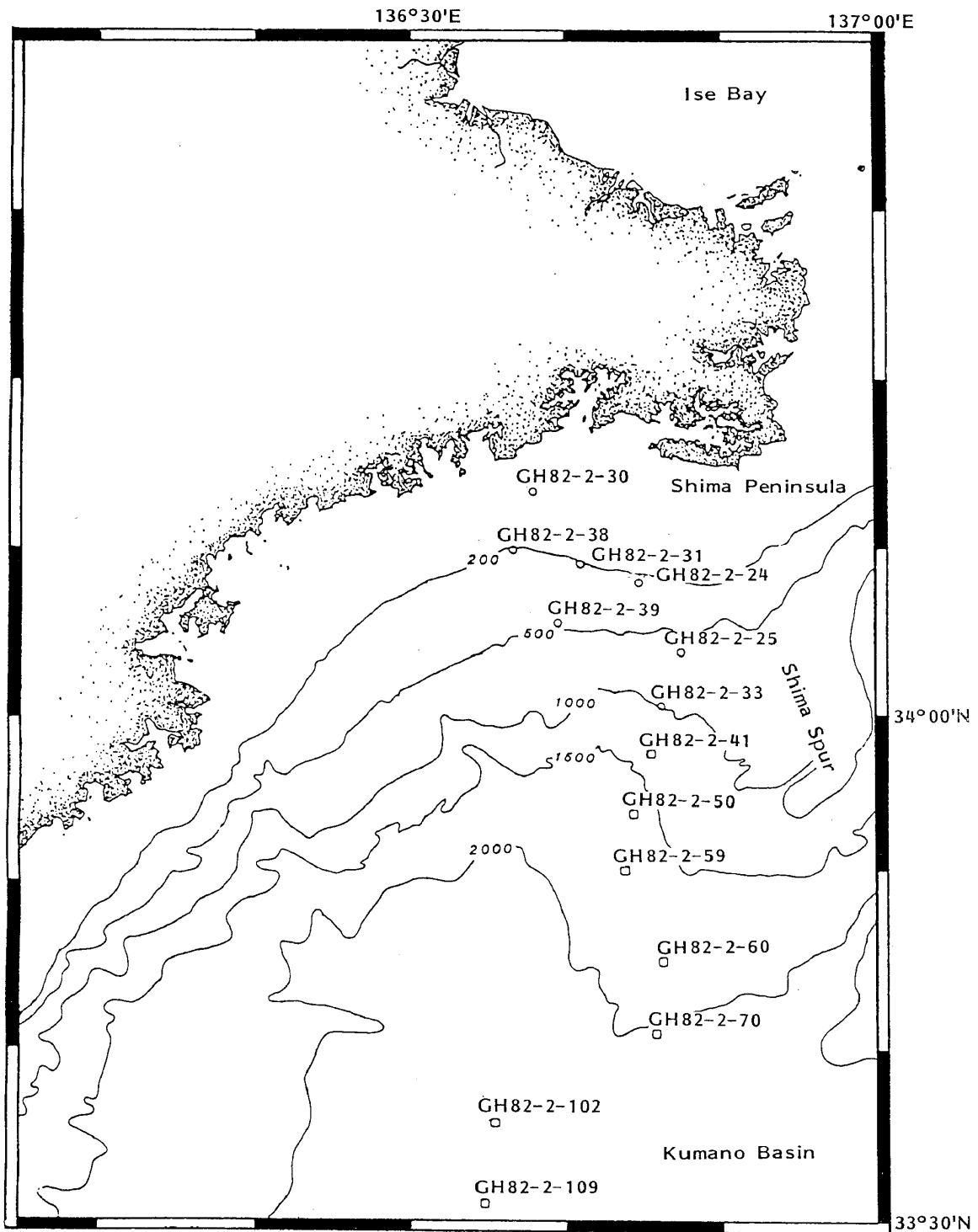


Fig. 3. Stations in the Kumanonada area of foraminiferal samples used in this study. Circle, Smith-McIntyre type sampler; square, piston corer.

mum depth of about 120 m. The Tanegashima Strait is present in the south off that island.

2. Kumano area

The outline of submarine topography is illustrated in Fig. 3. A remarkable feature is depicted by the Kumano Basin

which takes on the character of a forearc basin. This basin extends approximately 100 km in length and about 30 km in width, and its bottom is situated at a depth of about 2,000 m. Thick sedi-

ment fills in this basin are revealed on the seismic profiles (GH82-2 Cruise, the Geological Survey of Japan, 1983). The shelf is narrow or absent along the coast between Shima and Owase. The Shima

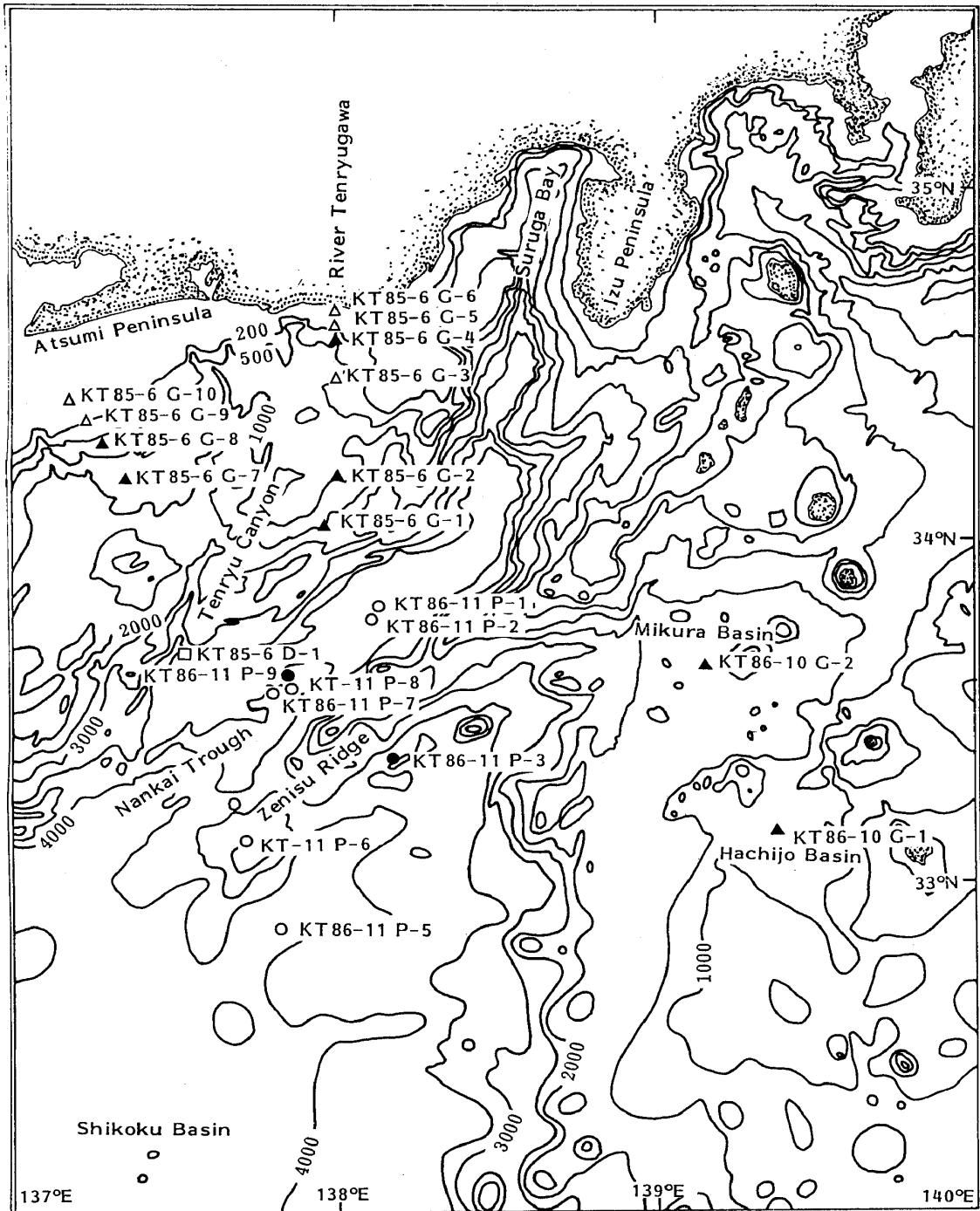


Fig. 4. Stations in the Enshunada area of foraminiferal samples used in this study. Triangle, grab sampler; circle, piston corer; square, dredge; closed mark, material stained.

Spur extends southward from the Shima Peninsula. The ridge deepens southward down to about 1,000 m at its extremity. There are two types of channels as distinguished by their course: one flows directly from its root off the Shima Peninsula into the Kumano Basin; the other runs from the shelf edge off Ise Bay to the Kumano Basin through the foot of the Shima Spur.

3. Enshunada area

The topographic outline of this area is shown in Fig. 4. This figure delineates that the study area is divided into such submarine topography as the Nankai Trough, the Shikoku Basin, and the continental slope. The continental shelf is well developed continuing from the Shima Spur to off west of the River Tenryugawa. The water depth of the shelf edge is between 140 m and 250 m, and the width is between 7 km and 30 km (Nagao *et al.*, 1977). In the western area off the River Tenryugawa, the depth of the shelf edge is deepest and the width is narrowest. The continental shelf is almost obscure along the coast from the River Tenryugawa to east of the Omaezaki Spur. The continental slope off Enshunada corresponds with the inner wall of the Nankai Trough. The slope is divided into the upper and lower slopes by the existence of a forearc basin. Those basins are located along the 2,000 m contour line. A basin in the area around the upper to middle part of the Tenryu Deep Sea Canyon is called the Enshu Trough by Sakurai and Sato (1983). There are nine small submarine canyons on the uppermost continental slope off Enshunada. These submarine canyons meet one another in a small basin at a depth of ca. 1,400 m off Hamamatsu, and come to form the Tenryu Deep Sea Canyon in a downstream direction. The most characteristic topography within the Nankai Trough is the Tenryu Deep Sea Fan.

The trough floor is almost flat, with a depth of a little more than 4,000 m in the studied area. The ocean floor is characterized by the Zenisu Ridge and the Shikoku Basin. The Zenisu Ridge is a unique topography, especially from the viewpoint of convergent plate boundary processes (Shimamura, 1988). Although this ridge is located on an outer swell of the Nankai Trough, it attains to relative heights between 1,500 m and 3,000 m. The northern slope of the Zenisu Ridge is smooth, but the southern one is steep (Figs. 4, 27). The Shikoku Basin is wide, and has an average depth of about 4,000 m.

4. Hachijojima area

The Hachijojima Island traverses (A-A' and B-B') are located between the Northwest Pacific and the Philippine Sea, and extend from the Izu-Bonin Trench to the Shikoku Basin through near Hachijojima Island across the Shichito Ridge (Fig. 5). The Shinkurose and Kurose Banks are shallow banks on the Shichito Ridge. The summit of the Shinkurose Bank is at about 200 m water depth. A rather large channel extends from near this summit to the Izu-Bonin Trench. The eastern continental slope shows a sharp inflection of inclination, without any slope break. The inflection is recognized on the eastern continental slope at about 5,000 m water depth: below this depth the slope is steeper (Tamaki *et al.*, 1981). A knoll exists at the same depth near GH79-4-D372. This depth corresponds with the boundary of sedimentary rocks: Quaternary sediments lie above and Paleogene-Early Miocene sedimentary rocks below (Yuasa *et al.*, 1982).

The Izu-Bonin Trench has a north-south trend, and attains to water depths of more than 9,000 m. The trench axis is shallower to the north of 32°30'N than in the remaining area.

On the contrary, large basins are pres-

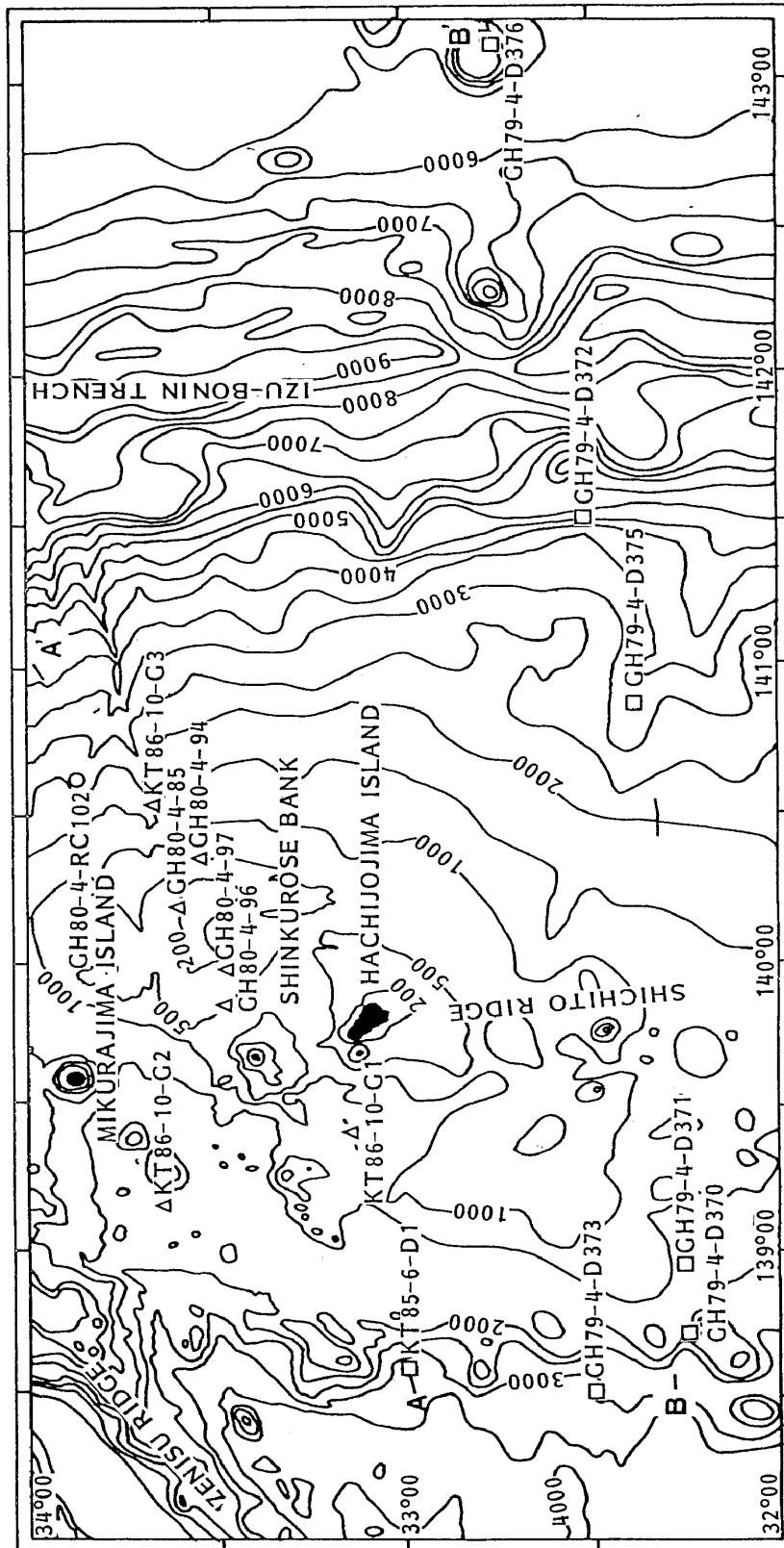


Fig. 5. Stations in the Hachijojima area of foraminiferal samples used in this study. Triangle, grab sampler; square, dredge.

ent in the backarc region of the Izu-Bonin Arc. The Hachijo and Mikura Basins are located on the western slope of the Shichito Ridge. The Mikura Basin is square and extends approximately 100 km in dimension, and its bottom is at about 1,500 m water depth. The Hachijojima Basin is rather small,

but the same in shape as the Mikura Basin, and its bottom is located at a depth of 1,000 m. Numerous peaks extend from north to south at about 2,000 m depth where the above-mentioned basins extend southward. The continental slope is steeper between those basins and the Shikoku Basin.

OCEANOGRAPHY

The study area is a portion of the Northwest Pacific off Enshunada, Kumanonada, Tanegashima Island, southwest Japan and around Hachijojima Island on the Izu-Bonin Arc.

The warm Kuroshio Current runs through those areas. The Current flows eastward from the East China Sea to the Pacific across the Ryukyu Arc, around Tanegashima Island. Off Tanegashima Island, this current turns northeastward, and runs along the Japanese coast to Shionomisaki, where it meanders southward (Kuroshio meandering). Consequently, its direction changes to the north and runs along the west side of the Izu-Bonin Arc. Reaching near Mikura Island, this current flows to the east through between this and Miyake islands. On the other hand, the coastal water mass is distributed off Enshunada and Kumanonada.

Illustrated are temperature, salinity, dissolved oxygen, pH, PO_4 , SiO_2 , $\text{NO}_2\text{-N}$, and $\text{NO}_3\text{-N}$ at Tanegashima Island (Figs. 6, 7), Enshunada (Figs. 8, 9) and Nankai Trough (Fig. 10). Figures 6, 7, 8 and 9 are basic oceanographic data published by the Japan Meteorological Agency. Figure 10 has been constructed on the basis of cruise reports of the *R.V. Bosei II* of Tokai University (Hirota *et al.*, 1986).

1. Tanegashima area

This area lies under the direct influence of the Kuroshio Current and is free from coastal water.

a. Temperature (Fig. 6)

The temperature is constant at the boundary between surface waters and the thermocline, being about 19°C, while surface water (seasonal layer) temperature ranges from 20°C in winter to 27°C in summer. Water temperature rapidly decreases downward from the thermocline, and reaches about 3°C at 1,000 m water depth. The boundary between the thermocline and the underlying deep waters coincides with this depth. The temperature of deep waters is 2°C, being invariable through the water column.

b. Salinity (Fig. 6)

Surface waters have salinities ranging from 33.8‰ in summer to 34.4‰ in winter. Salinities slightly increase downward to 150 m depth. Below this, there is a regular decrease in salinity within the main thermocline down to a little less than 34.3‰ at 800 m depth.

c. Dissolved oxygen (Fig. 6)

Surface waters have an average dissolved oxygen of about 4.9 ml/l in summer. This value regularly decreases downward, and reaches about 1.5 ml/l at 800 m water depth, but slightly increases below 1,000 m. Thus, the upper and lower boundaries of the oxygen minimum zone in this region are at approximately 800 m and 1,000 m water depth, respectively.

d. pH (Fig. 7)

Surface waters have a pH value of 8.4. This value regularly decreases downward to 1,000 m, and is about 7.9 at 1,000 m water depth.

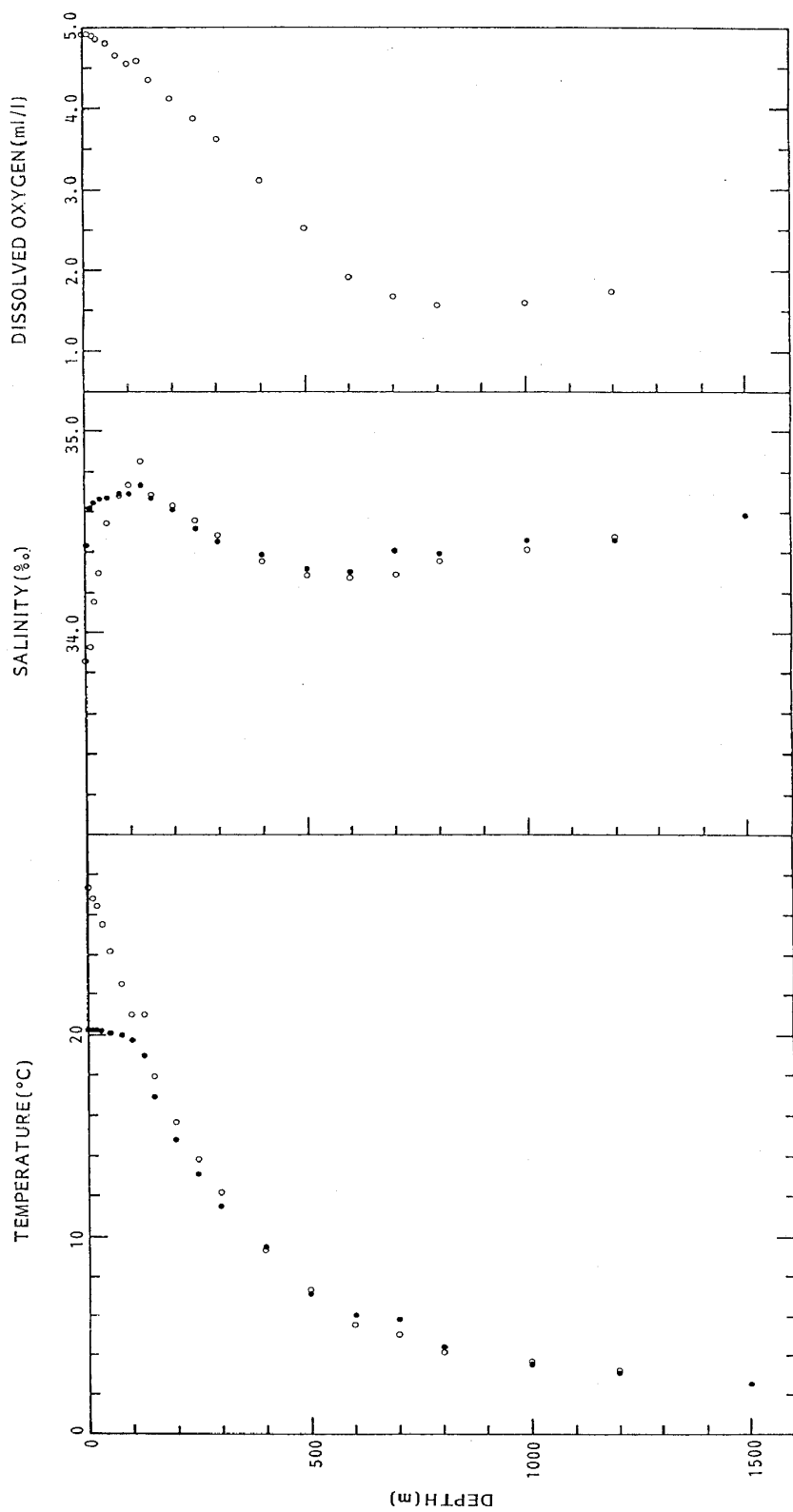


Fig. 6. Vertical distribution of temperature, salinity and dissolved oxygen in the Tanegashima area. (Data from the results of marine meteorological and oceanographical observations, 1906-1981).

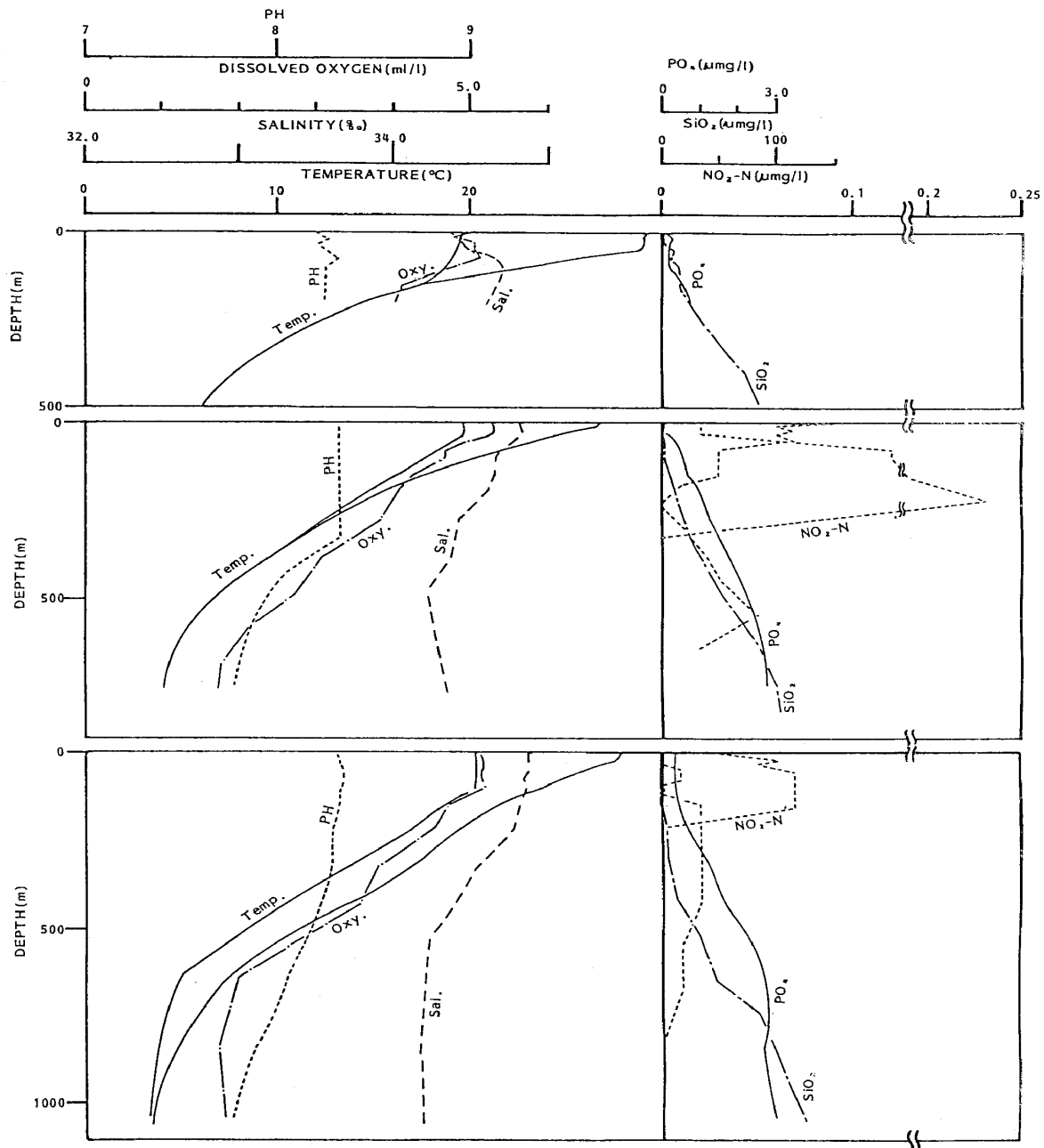


Fig. 7. Vertical distribution of temperature, salinity, dissolved oxygen and nitrogen in the Tanegashima area. (Data from the results of marine meteorological and oceanographical observations, February 14 and August, 1966, and February 9, June 30, July 20 and 30, 1969).

e. Other elements (Fig. 7)

PO_4 is about $0.5 \mu\text{g/l}$ and regularly increases up to about $2.5 \mu\text{g/l}$ at 700 m depth. This value at 800 m water depth is a little less than at that depth.

$NO_2\text{-N}$ varies with seasons. In summer, the surface water has a maximum

$NO_2\text{-N}$ value of about $0.08 \mu\text{g/l}$ below 50 m water depth, which rapidly decreases in the thermocline between 150 m and 200 m. The $NO_2\text{-N}$ is absent below 200 m water depth. On the other hand, two characteristics are recognized in winter. The maximum value in winter is 0.025

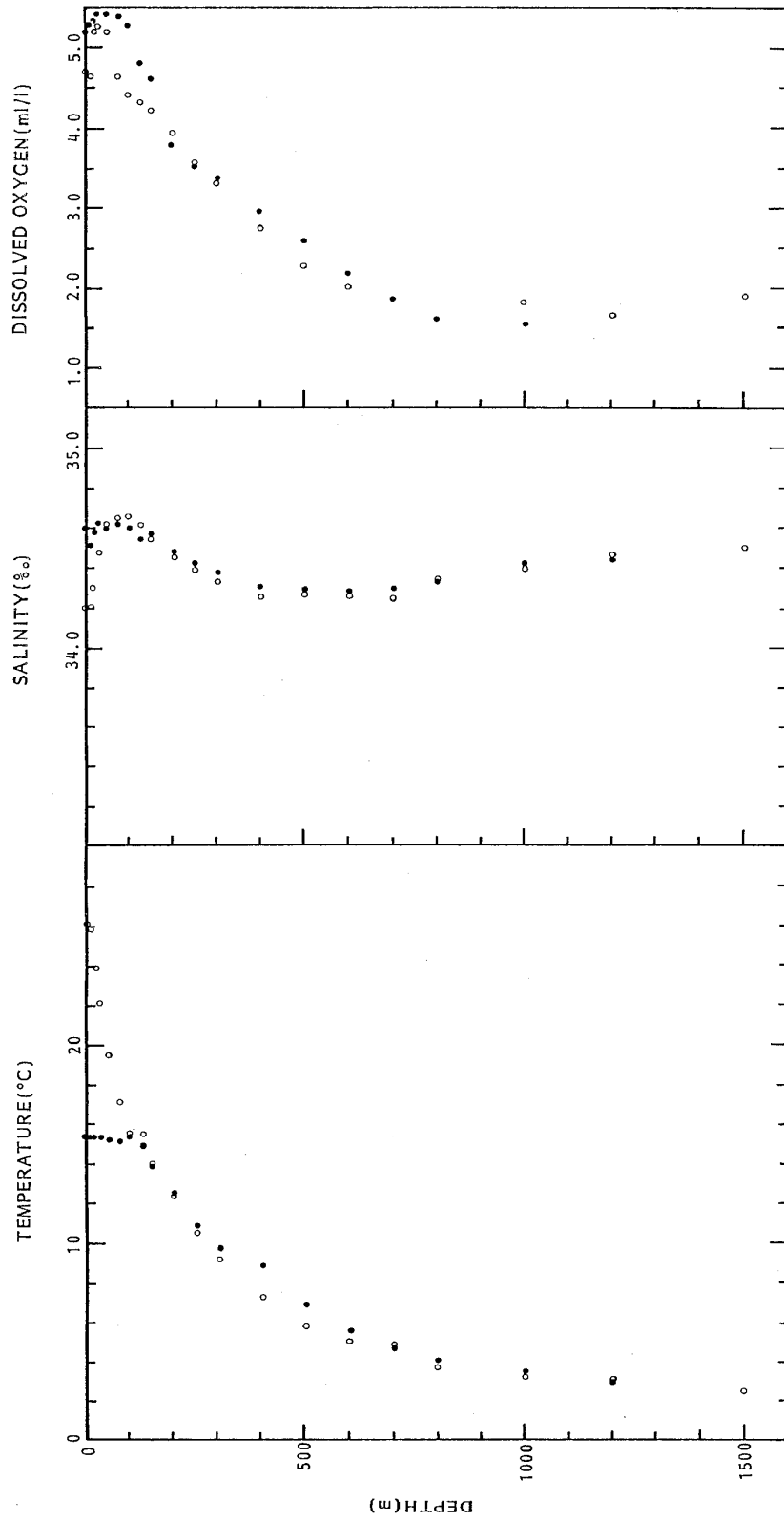


Fig. 8. Vertical distribution of temperature, salinity and dissolved oxygen in the Enshunada area. (Data from the results of marine meteorological and oceanographical observations, 1906-1981).

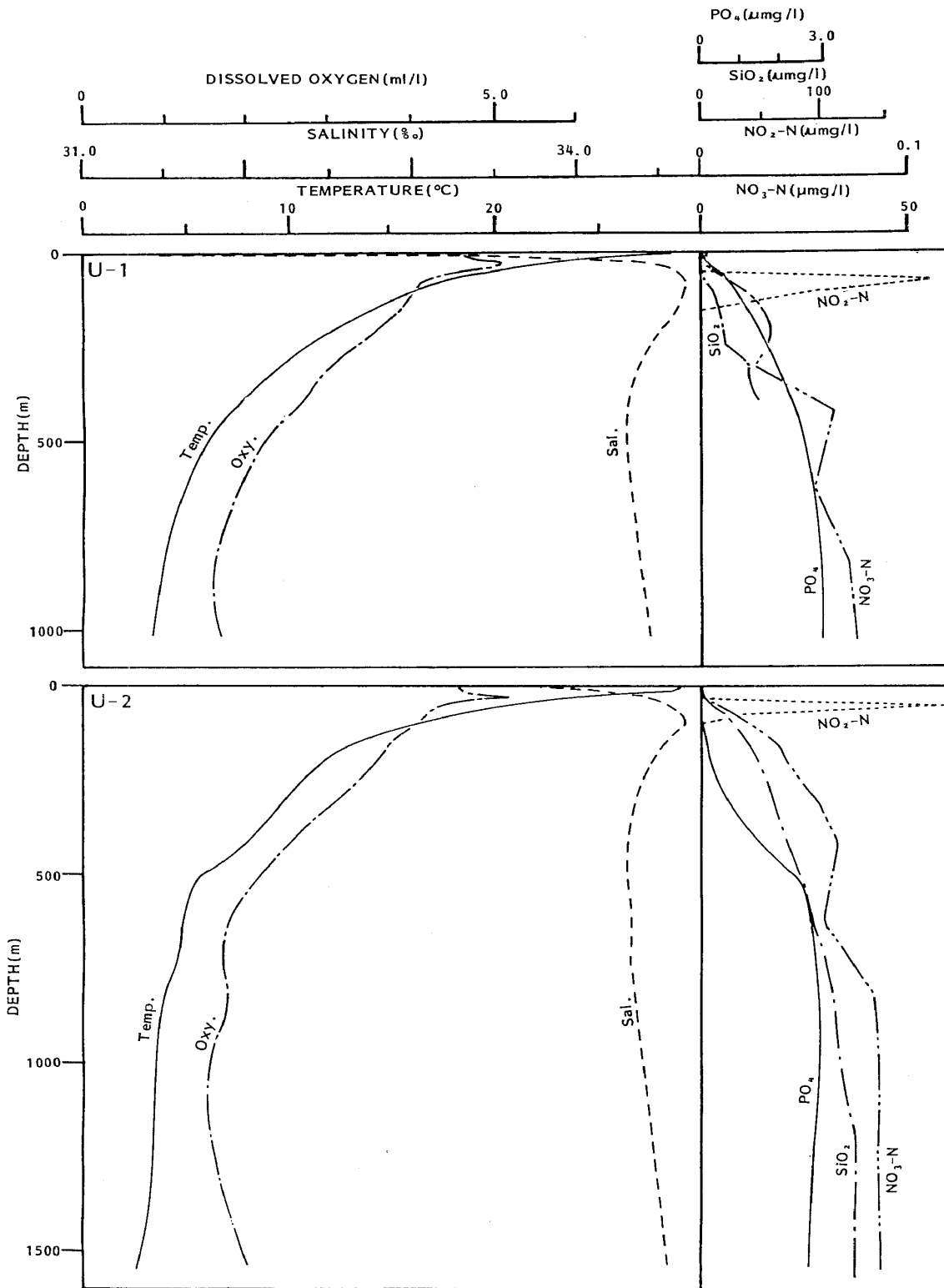


Fig. 9. Vertical distribution of temperature, salinity, dissolved oxygen and nitrogen off Atsumi. (Data from the results of marine meteorological and oceanographical observations, August 3, 1967 and July 31, 1969).

$\mu\text{g/l}$ at 150 m depth; in winter, $\text{NO}_2\text{-N}$ is recognizable in a deeper portion of the water column, extending down to 800 m water depth, but its maximum value is confined to the upper part of the thermocline.

SiO_2 is a little in surface waters. This value slightly increases between 100 m and 400 m, and rapidly increases below 400 m water depth.

2. Enshunada area

The coastal water mass is largely distributed in this and the Kumanonada areas. A great volume of fresh water and terrigenous materials flow into Enshunada from the mouth of the River Tenryugawa. Figure 8 shows hydrographic features in the Enshunada area, and Figure 9 illustrates those at two observation points (U-1, U-2) off the Atsumi Peninsula, as constructed on the basis of oceanographic data published in 1971 by the Japan Meteorological Agency. U-1 ($137^\circ 30' \text{E}$, $34^\circ 19' \text{N}$) is located more landward than U-2 ($137^\circ 27' \text{E}$, $33^\circ 59' \text{N}$).

a. Temperature (Fig. 8)

The temperature range between winter and summer affects only the upper layer of water above approximately 150 m depth. The average temperature is about 27°C in summer and 18°C in winter. The main thermocline extends to 1,000 m depth, where temperature is approximately 3°C . Deep water slightly decreases in temperature with water depth.

b. Salinity (Figs. 8, 9)

Figure 8 shows surface salinities ranging from 34.2‰ to about more than 34.7‰ , and a marked increase of salinity with depth. The main thermocline has minimum salinity values of less than 34.3‰ at about 600 m depth. The measured salinities are comparable between U-1 and U-2 (Fig. 9). Surface salinities at U-1 are lower than U-2. Considering locations of two points, it is apparent that a considerable amount of

runoff freshens coastal marine water in this region.

c. Dissolved oxygen (Fig. 8)

Surface waters have a dissolved oxygen of about 4.8 ml/l , which slightly increases downward up to 5.0 ml/l at 50 m depth. Below this depth, there is a regular decrease in dissolved oxygen within the main thermocline down to a little less than 1.9 ml/l at 900 m depth. This value slightly increases again below 1,200 m. The oxygen minimum zone in this region is approximately between 900 m and 1,200 m water depth.

d. Other elements (Fig. 9)

PO_4 differs between two observation points. At U-1, PO_4 is a little at the surface, and increases downward. On the other hand, it is absent in surface waters, and rapidly increases between 100 and 500 m water depth at U-2. At this point, the maximum value is approximately $2.8 \mu\text{g/l}$ at about 1,000 m water depth, which slightly decreases with water depth.

The presence of $\text{NO}_2\text{-N}$ slightly differs between the two points. At U-1, the maximum value is $0.12 \mu\text{g/l}$ at 100 m water depth. $\text{NO}_2\text{-N}$ is entirely absent at about 200 m water depth. The maximum value at U-2 is $0.13 \mu\text{g/l}$ at 50 m and the lowest extant value is situated at about 100 m.

$\text{NO}_3\text{-N}$ increases downward within the water column in this area, but the minimum values were recognized at 300 m and 600 m at U-1, and at 600 m at U-2.

SiO_2 is a little in surface waters. This value regularly increases with water depth.

3. Nankai Trough

Hydrographic observations during October 22–28, 1960 are shown in Fig. 10.

a. Temperature

The upper boundary of the main thermocline in this region is at approximately 100 m to 120 m water depth. However, there is no adequate information on

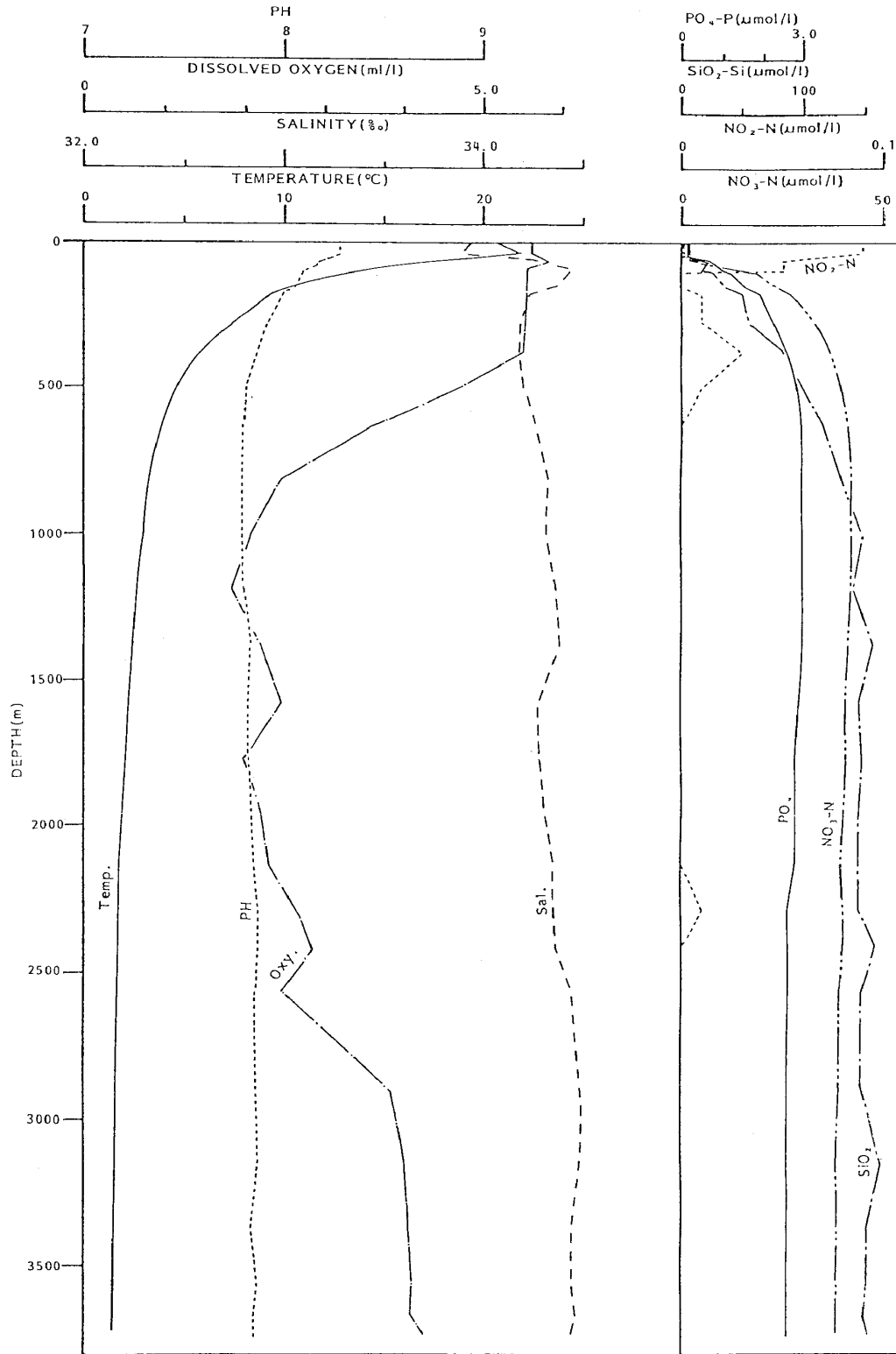


Fig. 10. Vertical distribution of temperature, salinity, dissolved oxygen, pH and nitrogen in Nankai Trough. (Data from the *Bosei II* of Tokai University, October 22-28, 1960).

vertical temperature distribution in this area through years. Thus, seasonal and annual variations of surface water temperature are unknown. The thermocline extends down to 800–1,000 m depth, with the sharpest decline between 100 m and 250 m. The water below 1,000 m is

approximately 2°C, and there may be a slight decrease in temperature with water depth.

b. Salinity

Surface waters have a salinity of about 33.9‰ at the surface, which slightly increases downward to 150 m depth.

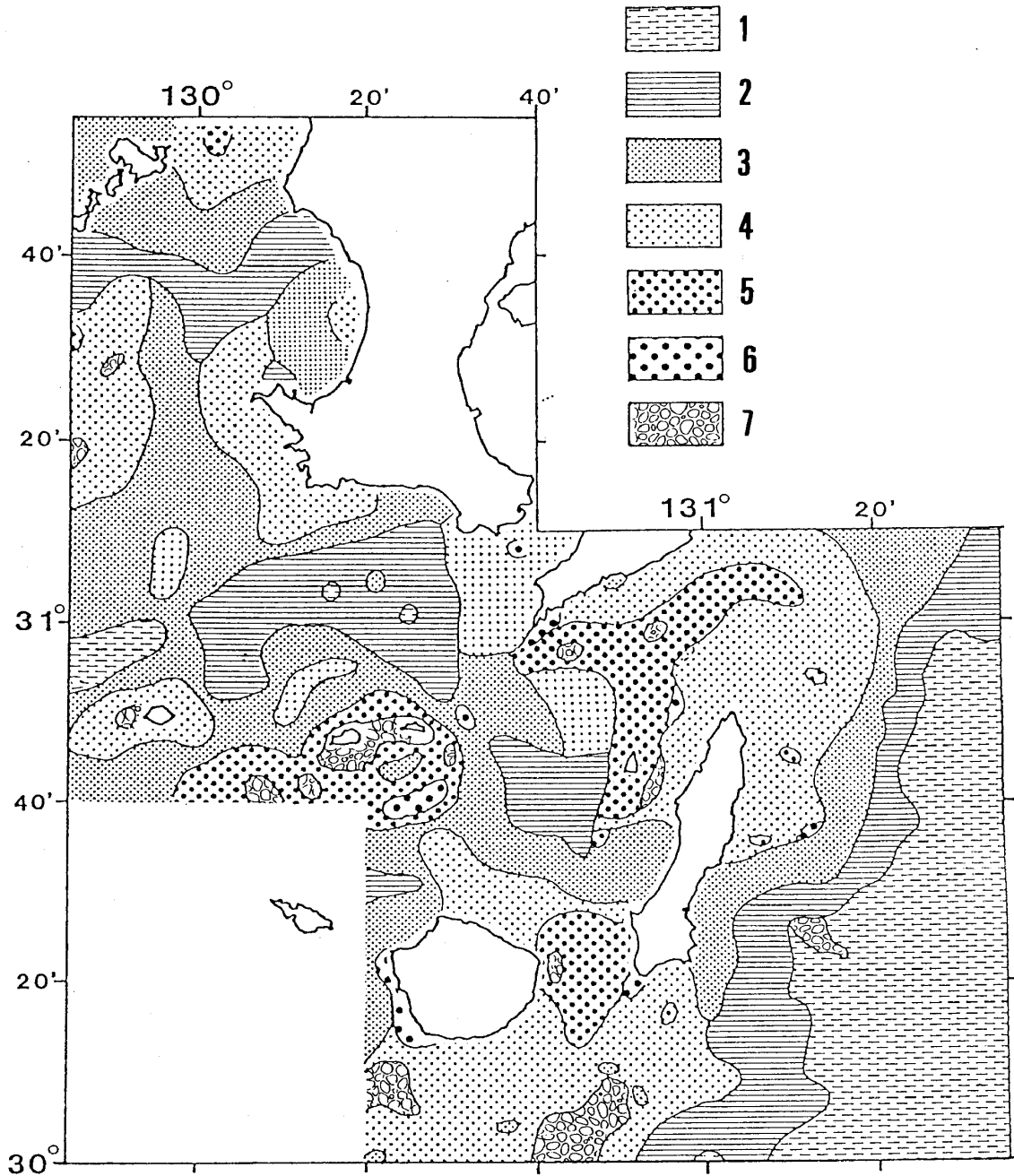


Fig. 11. Distribution of bottom materials around Tanegashima Island. 1, silt; 2, very fine sand; 3, fine sand; 4, medium sand; 5, coarse sand; 6, very coarse sand; 7, gravels or rocks. (after Ikehara, 1985).

The underlying water has a rather constant salinity which ranges from 34.3‰ to 34.4‰ below 200 m depth.

c. Dissolved oxygen

Surface waters have a dissolved oxygen of about 5.6 ml/l at the surface, which is nearly constant down to 250 m water depth. This value regularly decreases downward in the water column, reaching about 1.85 ml/l at 1,200 m water depth, and rapidly increases below 2,500 m, reaching 3.83 ml/l at about 3,000 m water depth. Below 3,000 m water depth, this value slightly increases. The upper and lower boundaries of the oxygen minimum zone in this region are at approximately 1,200 m and 2,500 m water depth, respectively. Dissolved oxygen in this oxygen minimum zone ranges from 1.85 ml/l to more than 2.8 ml/l.

d. pH

Surface waters are pH 8.21. This val-

ue regularly decreases downward to 500 m water depth, and is approximately pH 7.80 at 500 m water depth. Below 500 m there may be a slight increase in pH with water depth.

e. Other elements

A small amount of PO_4 exists at the surface 50 m of surface waters. Below 50 m water depth this value increases down to about 550 m water depth. The thermocline and deep waters contain a constant PO_4 value of about $2.7 \mu\text{mol/l}$.

$\text{NO}_2\text{-N}$ rapidly decreases in surface waters, and remarkably changes in the upper thermocline, with a maximum value of about $0.03 \mu\text{mol/l}$ at 370 m water depth.

$\text{NO}_3\text{-N}$ rapidly increases down to 600 m water depth, and is constant below this depth.

SiO_2 also increases down to 1,000 m water depth, and nearly constant, ranging from $149 \mu\text{mol/l}$ to $160 \mu\text{mol/l}$.

SEDIMENTS

Sediments bear upon the distribution of marine benthic organisms chiefly in ways that provide different types of substrate for organisms and their hosts, varied amounts and types of food that sediments contain, and various types of shell-constructing material for agglutinated foraminifers.

1. Tanegashima Island

Ikehara (1985) reported the distribution of surface sediments around Tanegashima Island. He showed that medium sand is widely distributed around the Osumi Strait and on the continental shelf off both the northeast part and southern extremity of this island. Coarser grains are recognized within the Osumi and the Tanegashima straits. Those areas where are distributed grains coarser than medium sand are characterized by abundant occurrences of derived fossils. The boundary between areas of

fine and medium sand corresponds to the contour of 120 m water depth. This boundary also coincides with the lower limit of erosion by the Kuroshio Current. The lower boundary of fine sand is expressed by mud contents of more than 50%. This boundary roughly corresponds with the 1,000 m contour, and is slightly deeper southward.

2. Kumanonada area

The distribution of surface sediments off Kumanonada is revealed by Honza *et al.* (1983). Figure 12 shows a general tendency for sediments to become finer with water depth, except around the Shima Spur. Sediments on the western slope at the root and the southern extremity of the Shima Spur consist mostly of medium sand. This kind of grains is largely composed of planktonic foraminiferal tests.

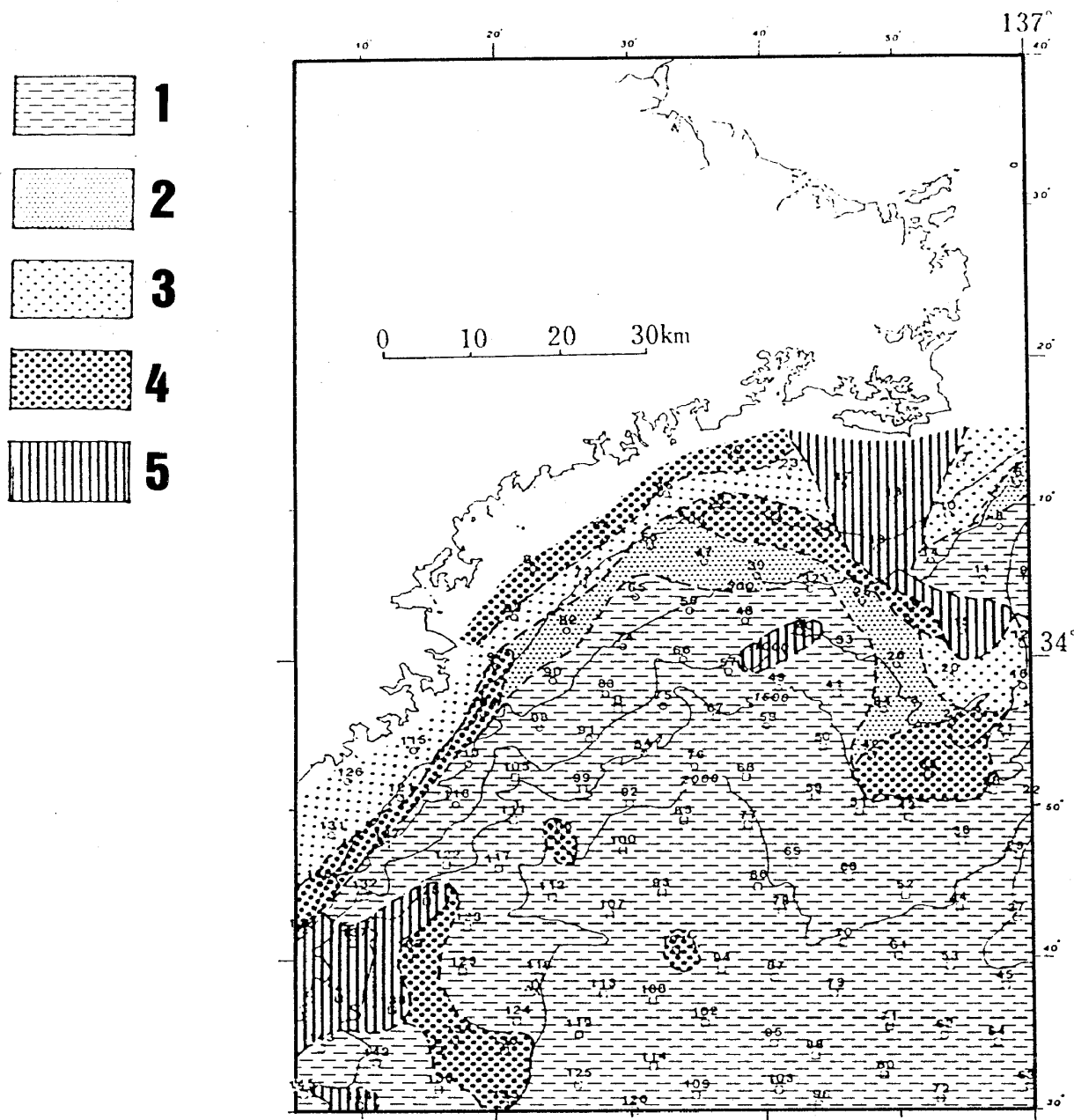


Fig 12. Distribution of bottom materials in the Kumanonada area. 1, silt ; 2, very fine sand ; 3, fine sand ; 4, medium sand ; 5, basements. (after Honza *et al.*, 1983).

3. Hachijojima area

Inouchi (1981) reported the distribution of surface sediments around the Shinkurose Bank. He divided the sea bottom into five types such as rocky, gravelly, sandy, muddy sand and muddy bottoms, and showed their distribution. According to him, the rocky bottom is exposed largely at the top of the Shin-

kurose, Kitakurose and Nakakurose banks, east of the "Kurose Hole" and north and east of Hachijojima Island. The areas of rocky bottoms around the Shinkurose and Nakakurose banks and to the east of Hachijojima Island are surrounded by gravelly bottoms. Two types of gravelly bottom are chiefly composed of calcareous and volcanic gravels.

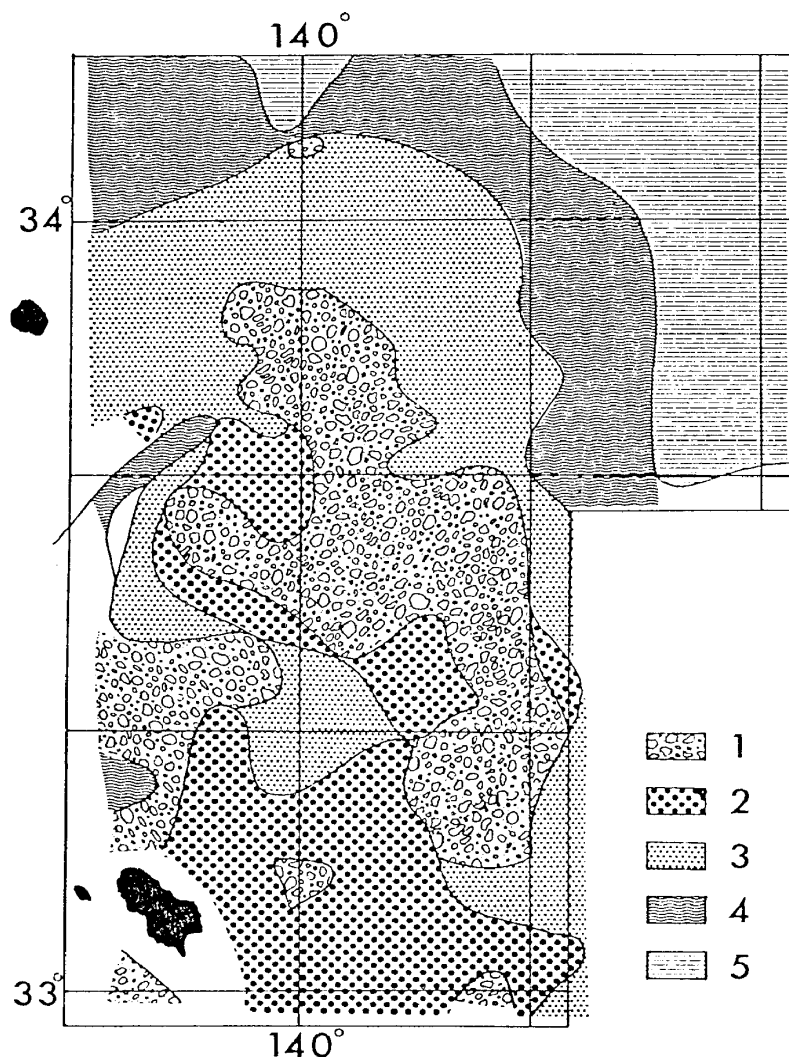


Fig. 13. Distribution of bottom materials around the Shinkurose Bank in the Hachijojima area. 1, rocks; 2, gravels; 3, sand; 4, muddy sand; 5, mud. (after Inouchi, 1981).

Around rocky and gravelly areas of the banks are fine to coarse-grained sandy areas. The sediments are well sorted in general. Sand is chiefly of volcanic and biogenic origin with subordinate quantities of clastic grains. Muddy sand areas are distributed around the sandy area. Muddy bottoms are widely distributed on the continental slope of the Shichito Ridge.

The distribution of sediments is delineated in the "Geological map of the northern Ogasawara Arc" (Honza *et al.*, 1982). This map shows that the geologic ages and characteristics of sedi-

ments differ between the east and west sides of the Shichito Ridge. In the west are Quaternary sediments and volcanic rocks dominant. Basaltic rocks are distributed on the western slope of the Shichito Ridge. On the other hand, the sediments on the eastern continental slope of this ridge are mainly of three different geologic ages. Pliocene sediments are distributed at a depth ranging from about 3,000 m to 5,000 m. The continental slope below 5,000 m is covered by sediments of Paleogene and Early Miocene ages.

MUD CONTENTS

Mud contents were measured by a dried weight percentage of particles smaller than 0.063 mm in diameter relative to the total treated sample.

1. Tanegashima area

The mud contents roughly show a zonal distribution in this area. The values are more than 50% at most points

below 1,000 m water depth. At points between 500 m and 1,000 m the values range from 30% to 50%. A point with less than 10% was recognized in an area under the influence of the Kuroshio Current (Fig. 14).

2. Kumanonada area

In two areas on the summit of the

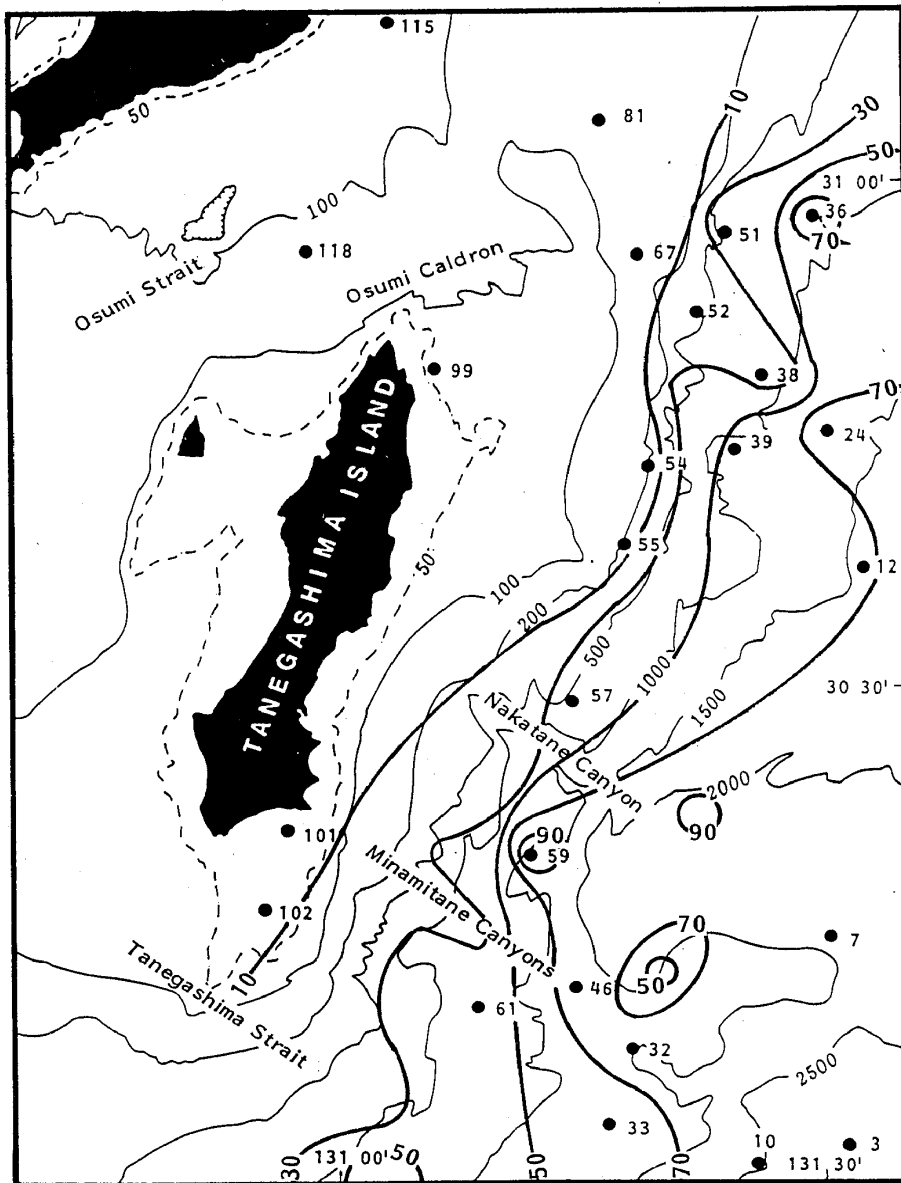


Fig. 14. Distribution of mud contents (%) off Tanegashima Island.

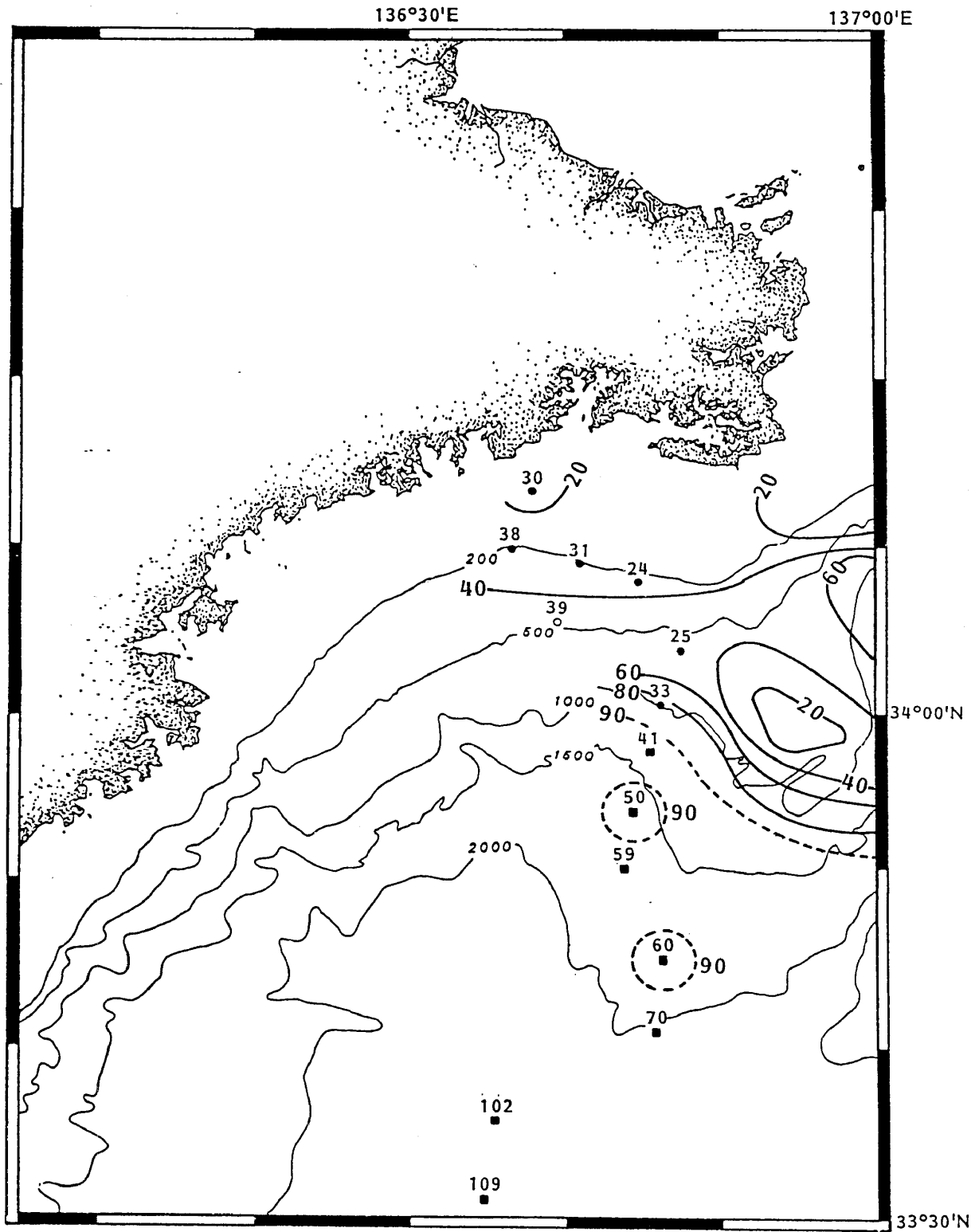


Fig. 15. Distribution of mud contents (%) in the Kumanonada area.

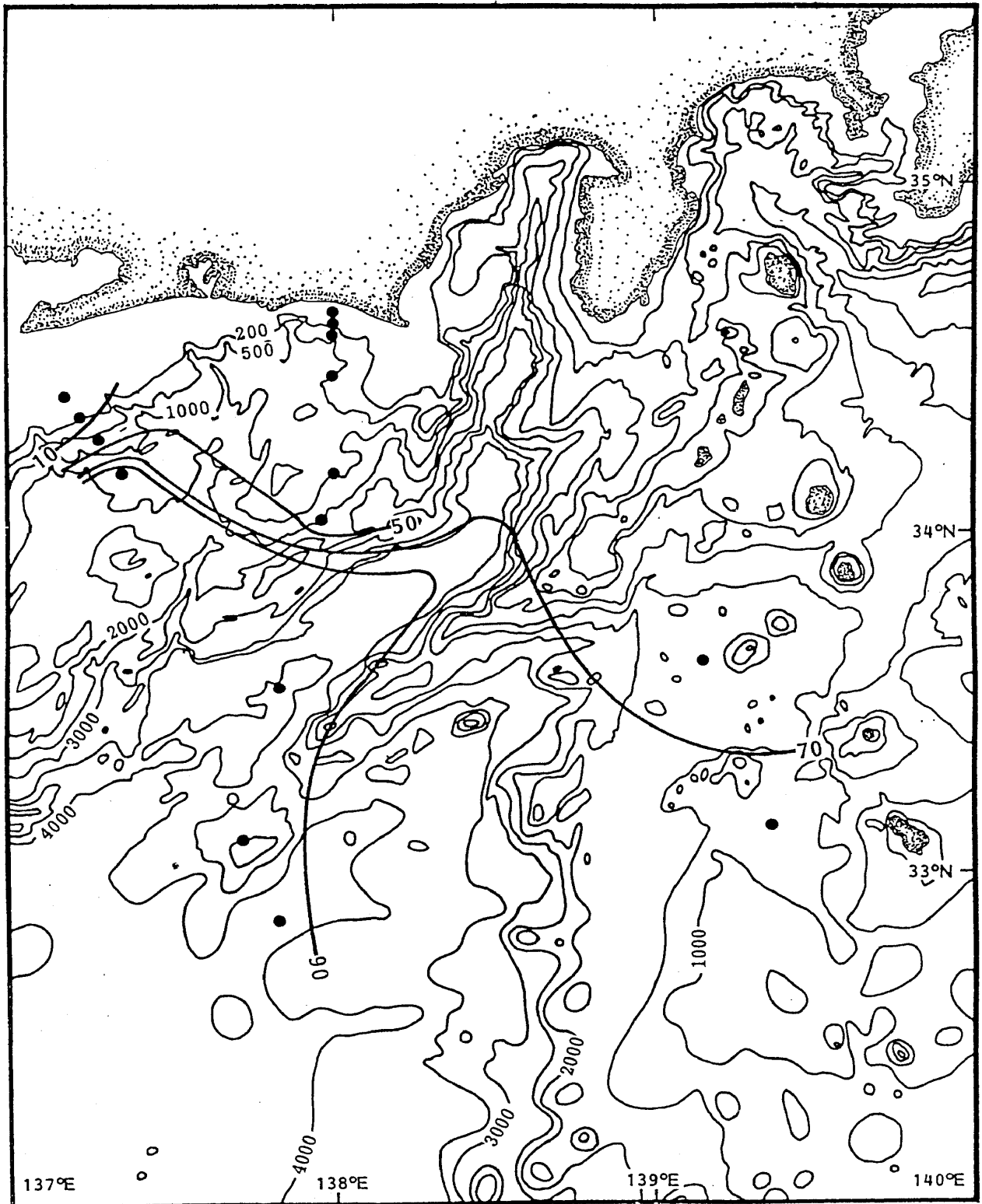


Fig. 16. Distribution of mud contents (%) in the Enshunada area.

Shima Spur and the coastal area around the Shima Peninsula, the mud contents are less than 20%. On the contrary, the highest value was recognized at points on the bottom floor of the Kumano Basin (more than 90%). The isoplethes are mostly parallel to the contour lines of submarine topography (Fig. 15).

3. Enshunada area

Sediments in this area are divided into two groups by mud contents. On the continental slope and shelf are rather low frequencies (less than 70%) except in KT85-6-G7. Off the mouth of the River Tenryugawa is a higher percentage of mud content than off Atsumi. On the contrary, in the Nankai Trough, Shikoku Basin and Zenisu Ridge frequencies are more than 70% (Fig. 16).

4. Hachijojima area

The frequency generally increases with depth in this area. Rather low values (less than 10%) are distributed on the Shinkurose Bank. Silty sediments containing more than 50% of mud are present on the continental slope. Frequencies over 90% were only recognized at GH80-4-RC102 on the eastern continental slope of the Shichito Ridge (Figs. 30 and 31).

In a summary, 1) mud contents are rather low on the continental slope and shelf; 2) highest frequencies over 90% are distributed in the Nankai Trough, Shikoku Basin and Kumano Basin; 3) rather coarse sediments accumulated on a projected topography. On the contrary, finer grains settled in topographic depression.

FAUNAL STATISTICS

This study intends to delineate faunal statistics in the Tanegashima, Kumanonada, Enshunada and Hachijojima areas. Planktonic foraminiferal numbers and benthic foraminiferal numbers stand respectively for the number of individuals of planktonic and benthic foraminifera contained in 1g of dry sediments. A/T and P/T ratios show the ratio of agglutinated foraminifera to the total benthic foraminifera and the ratio of planktonic foraminifera to the total foraminifera, respectively.

1. Tanegashima area

a. Planktonic foraminiferal numbers (Fig. 17)

The numbers regularly increase with depth down to about 900 m water depth, and slightly decrease below 1,700 m water depth. Large numbers (more than 1,000) are generally confined to a depth ranging from 1,000 m to 1,700 m. The maximum value is at GH84-3-61, running up to more than 4,000 individuals. The number is less than 100 in the

sublittoral zone. In the lower bathyal zone (below 2,000 m), the value ranges from 100 to a little more than 1,000.

b. Benthic foraminiferal numbers (Figs. 17, 18)

The numbers regularly increase with depth down to about 900 m water depth, and decrease below 1,000 m water depth. They are rather small (less than 1,000) at most points, with the maximum value at GH84-3-38 (Fig. 18), and numbers range from 10 to 100 below 2,000 m water depth (Fig. 17). Sample points with small numbers less than 10 are in the surface water region around Tanegashima Island (GH84-3-57, 81, 99, 102, 118). These points are mostly within the distribution of low mud contents and under the erosive influence of the Kuroshio Current. Points with rather small values on the continental slope are located within the canyon (GH84-3-24, 32, 59).

c. A/T ratio (Fig. 19)

This value regularly increases with depth in this area. Most of the points

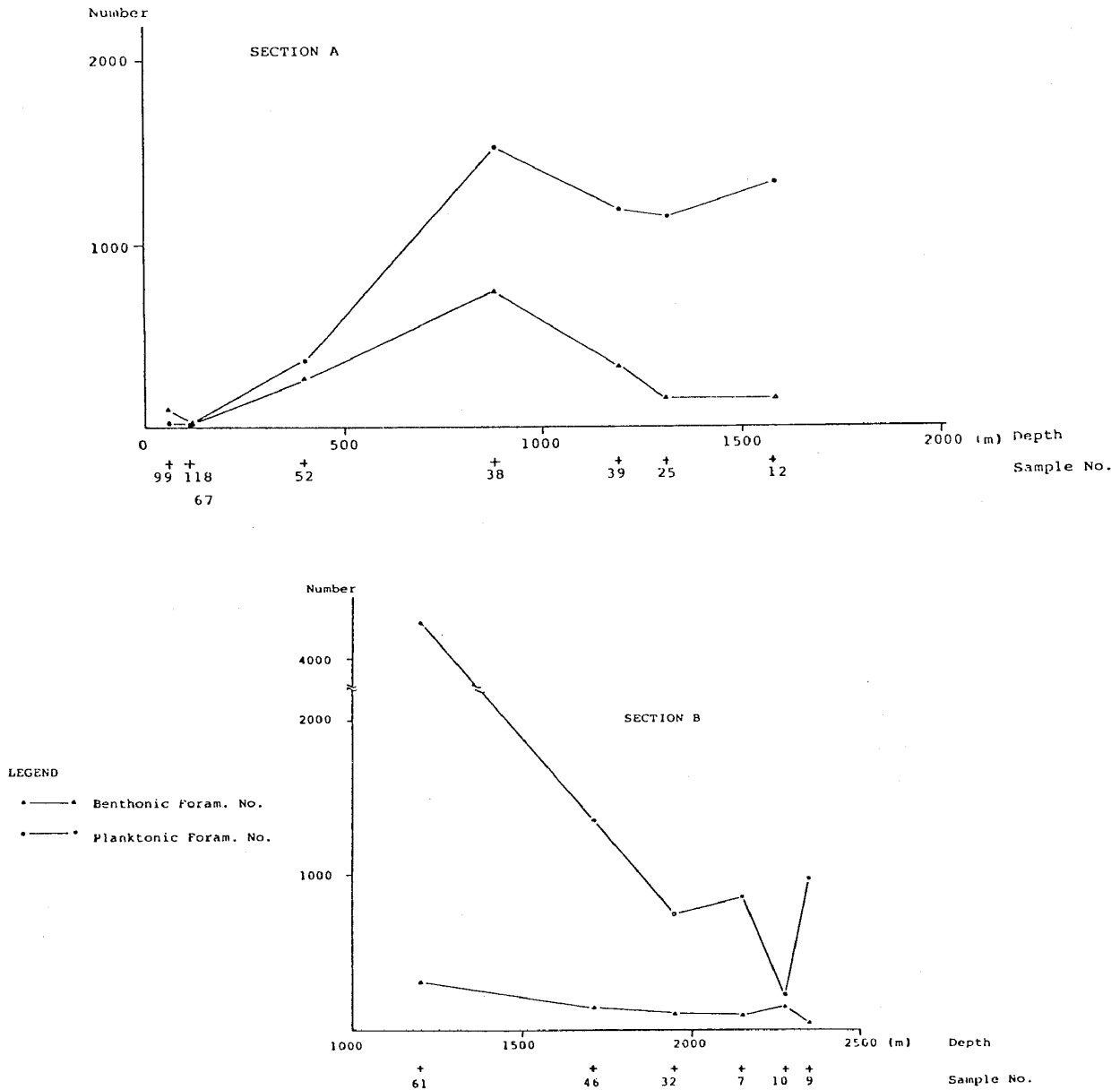


Fig. 17. Distribution of planktonic foraminiferal number and benthic foraminiferal number around Tanegashima Island.

on the continental slope below 1,500 m show values of more than 10%. Below 2,000 m water depth, the values are over 30%. In a shallow water area, the value is 0 or very small.

d. P/T ratio (Fig. 20)

This value rapidly increases with depth down to about 1,200 m water depth, and is constant down to 2,200 m and then slightly decreases deepward. Points marked over 50% are below about

400 m water depth. Sample points with low values of less than 10% are under surface waters around Tanegashima Island.

2. Kumanonada area

a. Planktonic foraminiferal numbers (Fig. 21)

The largest numbers range from 200 m to 1,000 m water depth. The number basically increases with depth down to

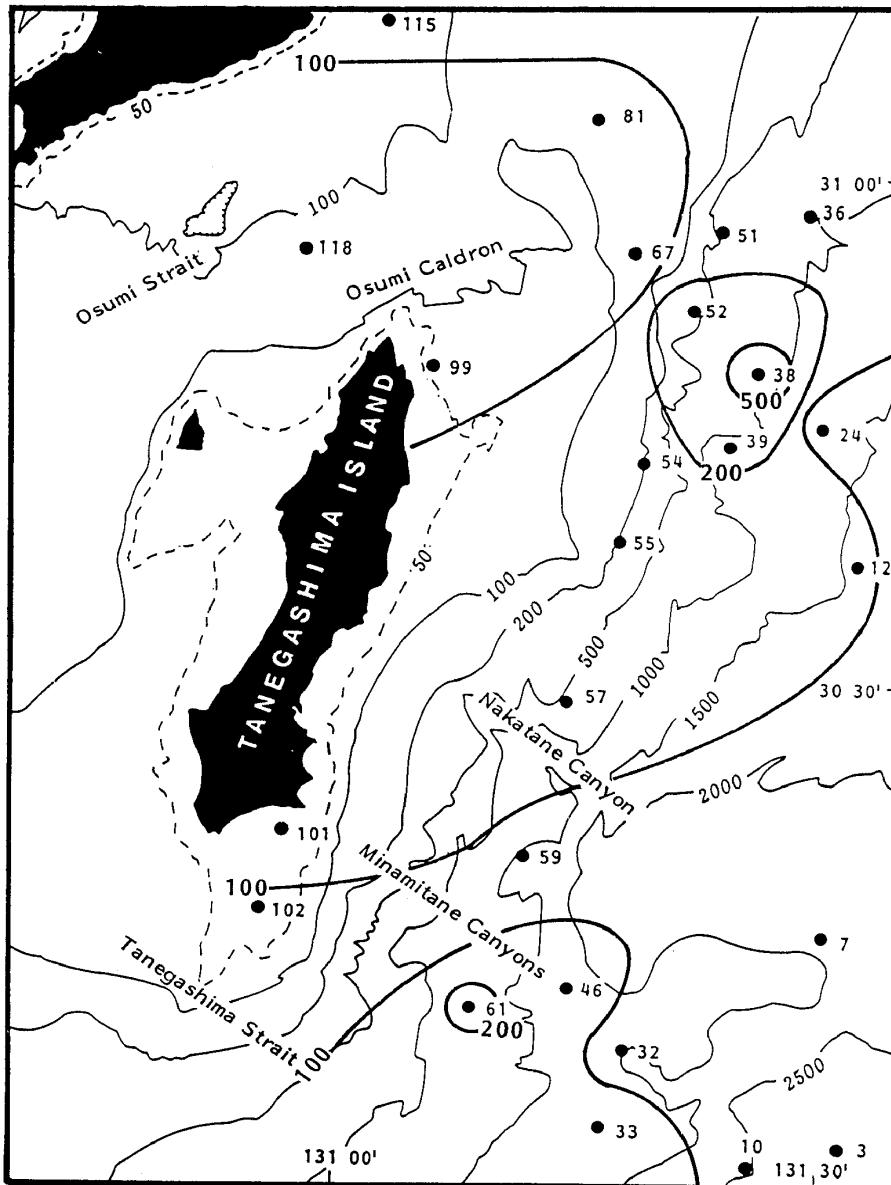


Fig. 18. Distribution of benthic foraminiferal number around Tanegashima Island.

about 1,000 m water depth, except for GH82-2-50. This point, located beneath the west side of the Shima Spur, has maximum values of more than 4,200. On the bottom of the Kumano Basin are rather low numbers, ranging from 50 to 100.

b. Benthic foraminiferal numbers (Fig. 22)

The numbers fundamentally decrease with depth below 250 m water depth. Points with large numbers are located on

a narrow shelf. The maximum number is 2,313 at GH82-2-31. The point beneath the southern extremity of the Shima Spur bore a rather large number of 655. The points on the bottom of the Kumano Basin yielded numbers less than 100.

c. A/T ratio (Fig. 23)

This ratio in this area is generally low. Most of the points show values of less than 10%, except for GH82-2-30 and 41.

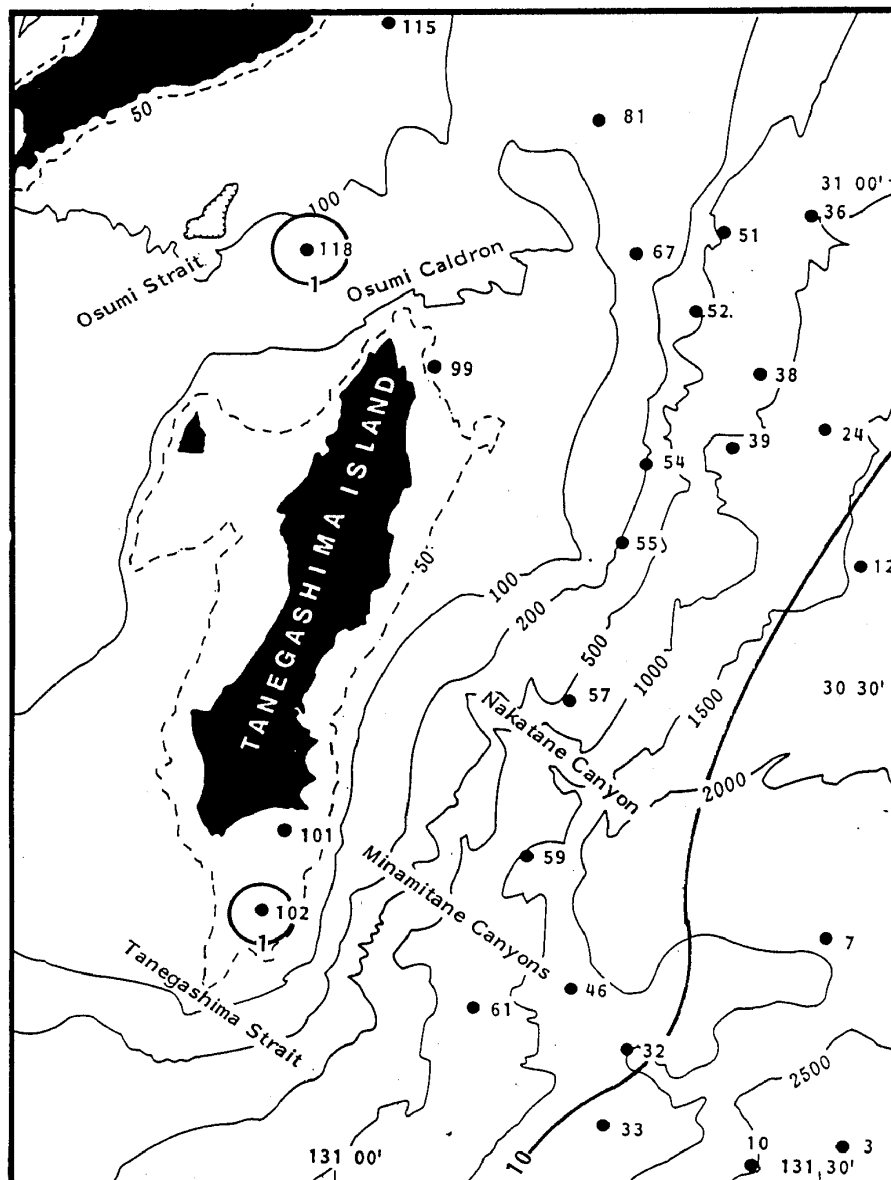


Fig. 19. Distribution of agglutinated foraminifer/total benthic foraminifer ratio (%) around Tanegashima Island.

d. P/T ratio (Fig. 24)

This value regularly increases with depth down to about 500 m water depth. A rather high value was recognized at the points on the continental slope which ranges in water depth from 500 m to 1,600 m. The points on the bottom of the Kumano Basin show values of more than 30%.

3. Enshunada area

a. Planktonic foraminiferal numbers (Fig. 29)

The points on the continental slope bear rather large numbers (KT85-6-G1, 3). The numbers are mostly null in three areas such as off the mouth of the River Tenryugawa, within the Nankai Trough and the Shikoku Basin. One of the peaks in this number is on the Zenisu Ridge.

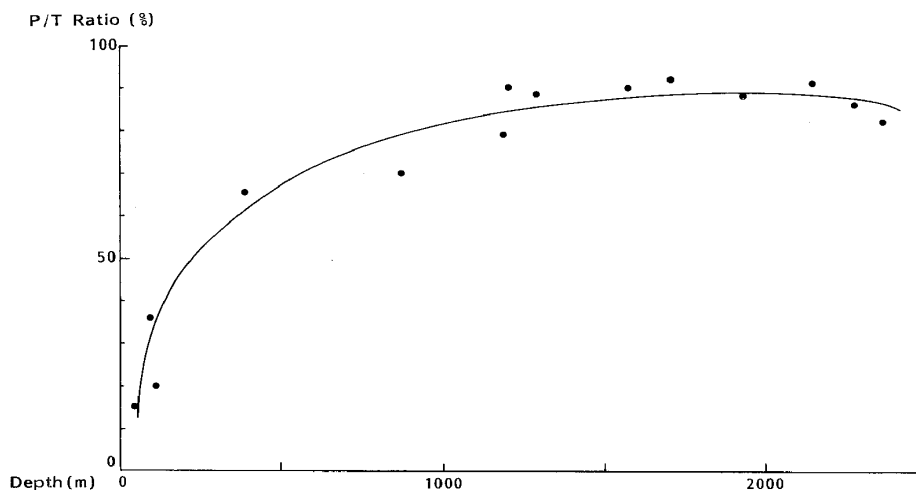


Fig. 20. Distribution of planktonic foraminifer/total foraminifer ratio(%) around Tanegashima Island.

b. Benthic foraminiferal numbers (Figs. 25, 29)

The numbers are small (less than 10) at samples from the Nankai Trough. However, points with rather large numbers are on the continental slope and shelf off Enshunada. For example, the points off the mouth of the River Tenryugawa contained an almost constant number ranging from about 20 to a little over 100, and the points on the continental slope off the Atsumi Peninsula yielded large numbers of more than 100. Two points in the Shikoku Basin and on the Zenisu Ridge bore less than 50.

c. A/T ratio (Fig. 26)

The value fundamentally increases deepward on the continental slope. The points on the Zenisu Ridge and bottom of the Shikoku Basin generally represent high values, especially more than 60% at KT86-11-PP5.

d. P/T ratio (Fig. 27)

This value is 0% on the shelf, and regularly increases on the continental slope. No planktonic foraminifera occur in surface sediments within the Nankai Trough and on the bottom of the Shikoku Basin below about 4,000 m water depth. On the other hand, at the

summit of the Zenisu Ridge the value is about 60%.

4. Hachijojima area

a. Planktonic foraminiferal numbers (Fig. 31)

Samples on the Shichito Ridge are marked by rather high numbers, which gradually decrease deepward. For instance, two western points on the Shichito Ridge yielded more than 4,000 individuals. On the contrary, samples near the summit of the Shinkurose Bank contained a rather low number, ranging from 170 to 530, except for GH80-4-94 (a little less than 2,800). These differences in number among samples are attributed to water depth.

b. Benthic foraminiferal numbers (Figs. 30, 31)

The points in the bathyal zone yielded rather large numbers of individuals. The maximum value is about 2,100 at KT86-10-G3. Surface sediments under surface waters and deep water below 3,200 m contained rather small numbers (less than 100).

c. A/T ratio (Figs. 30, 31)

At the points on the Shichito Ridge below 3,200 m water depth the values are more than 20%. This value runs up to

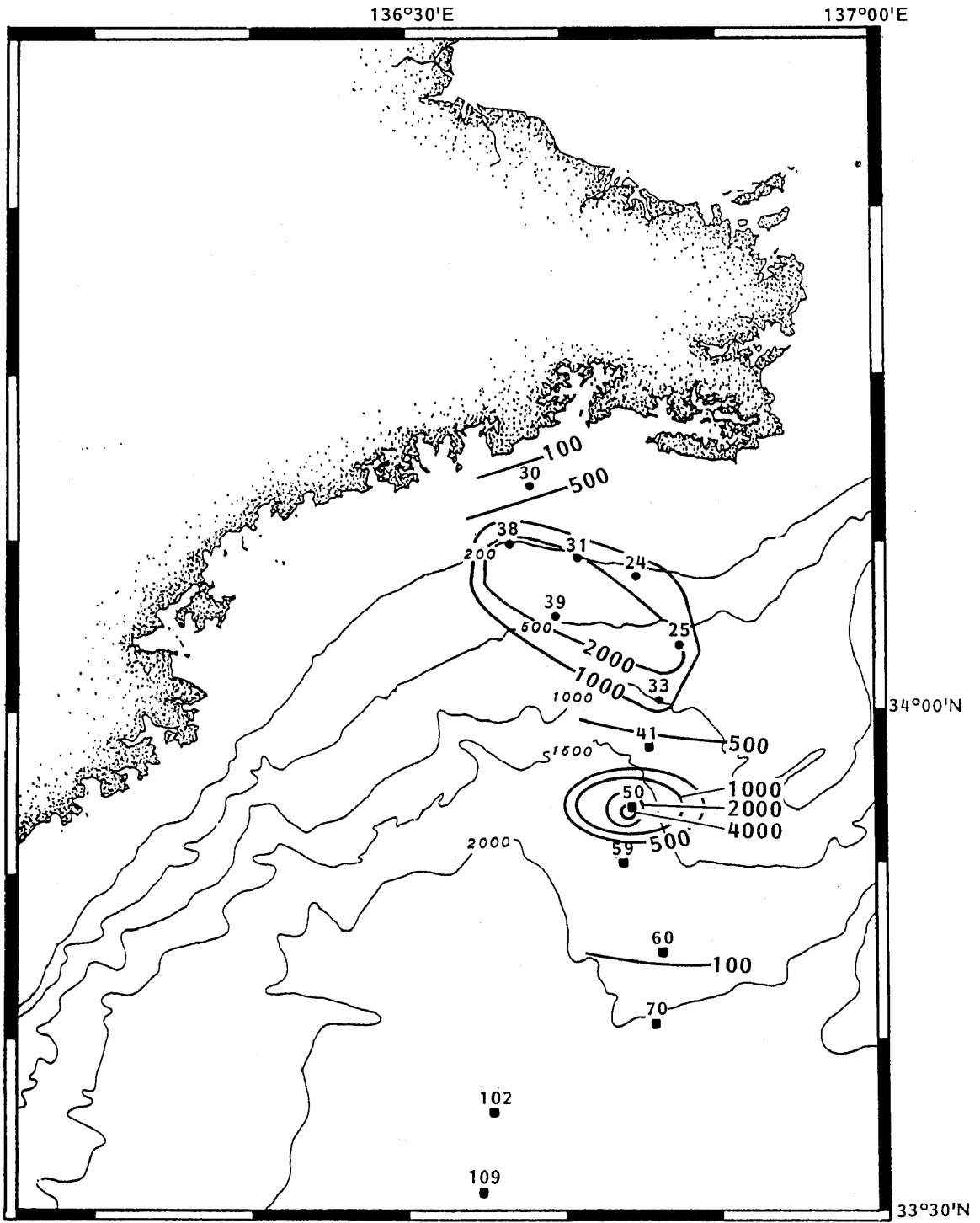


Fig. 21. Distribution of planktonic foraminiferal number in the Kumanonada area.

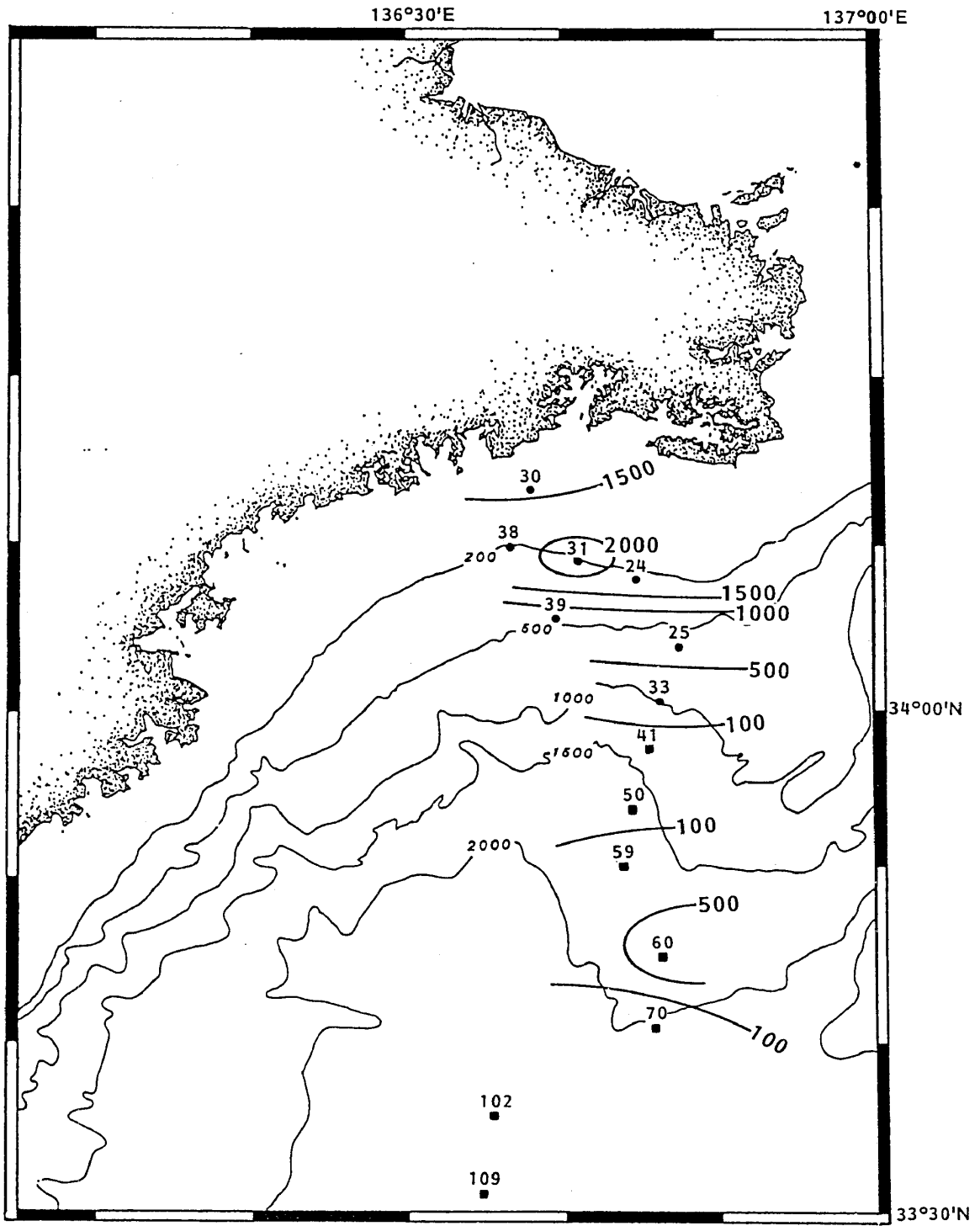


Fig. 22. Distribution of benthic foraminiferal number in the Kumanonada area.

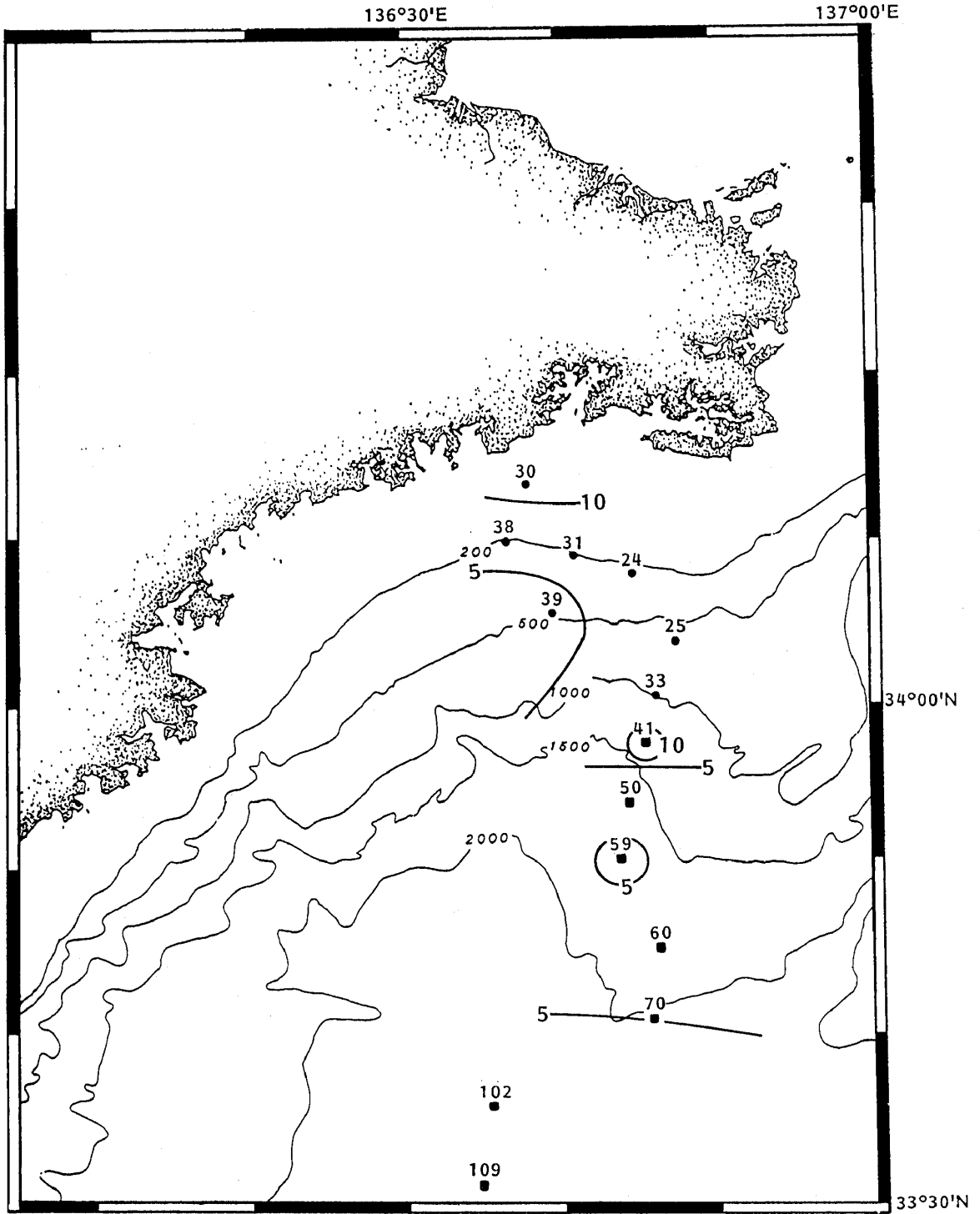


Fig. 23. Distribution of agglutinated foraminifer/total benthic foraminifer ratio (%) in the Kumanonada area.

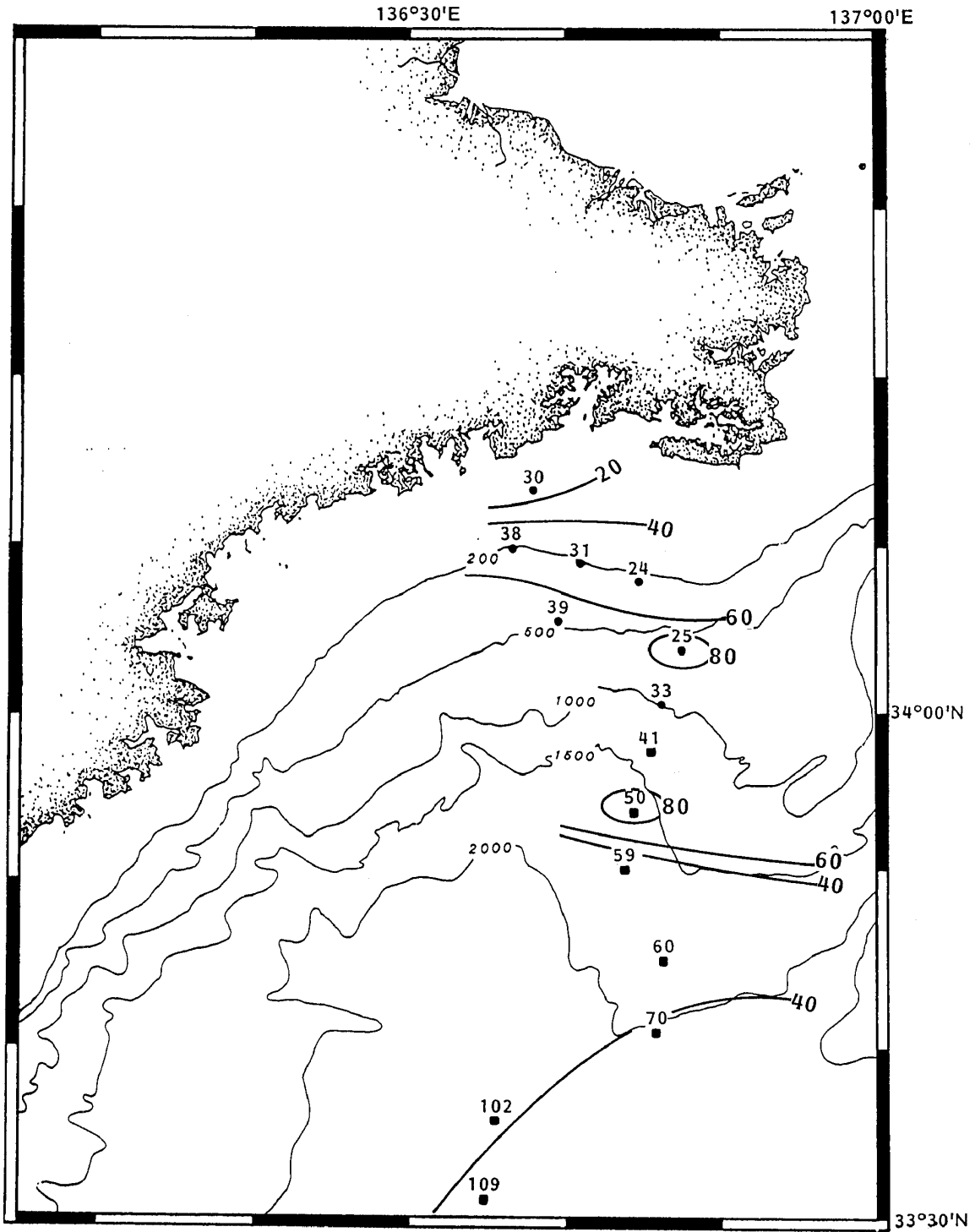


Fig. 24. Distribution of planktonic foraminifer/total foraminifer ratio (%) in the Kumanonada area.

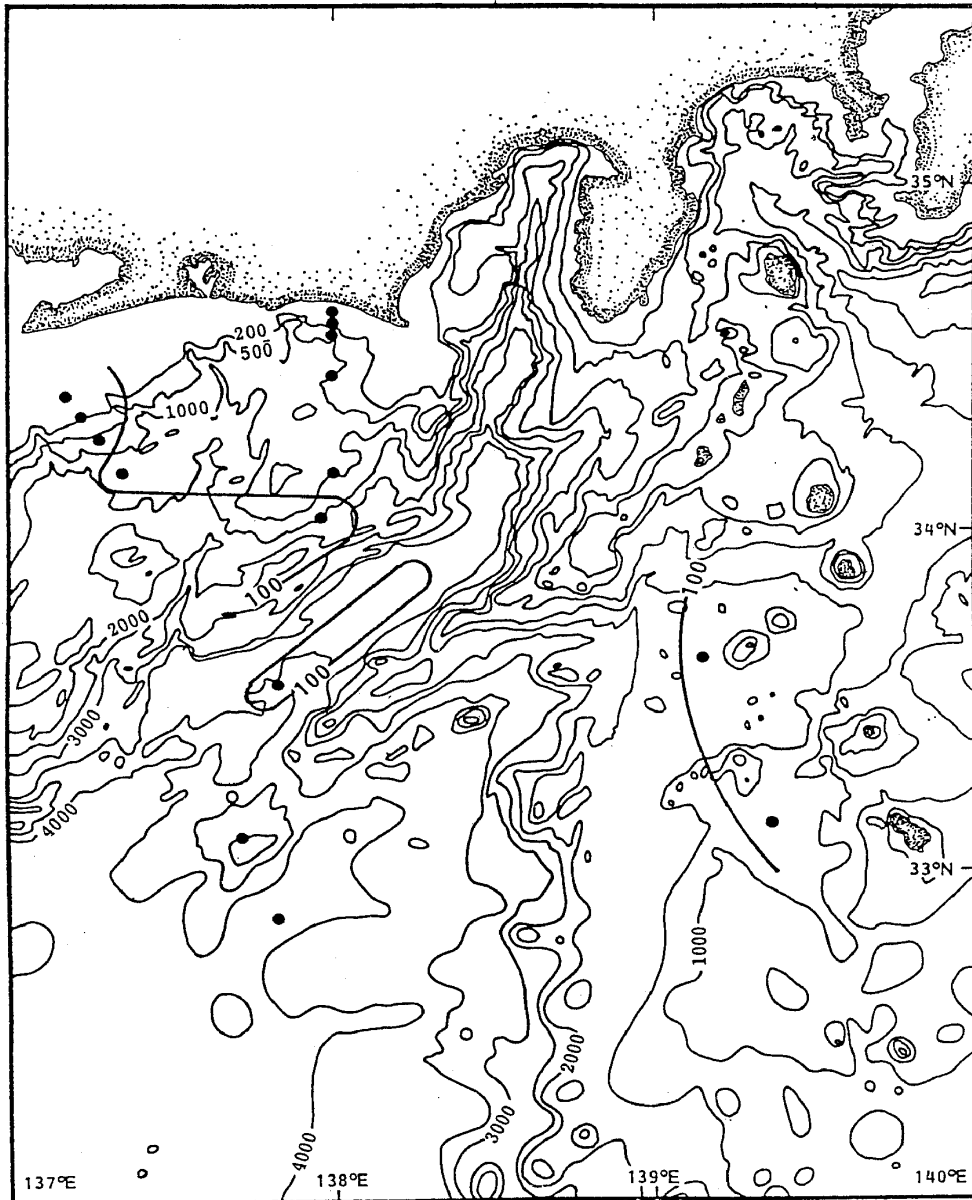


Fig. 25. Distribution of benthic foraminiferal number in the Enshunada area.

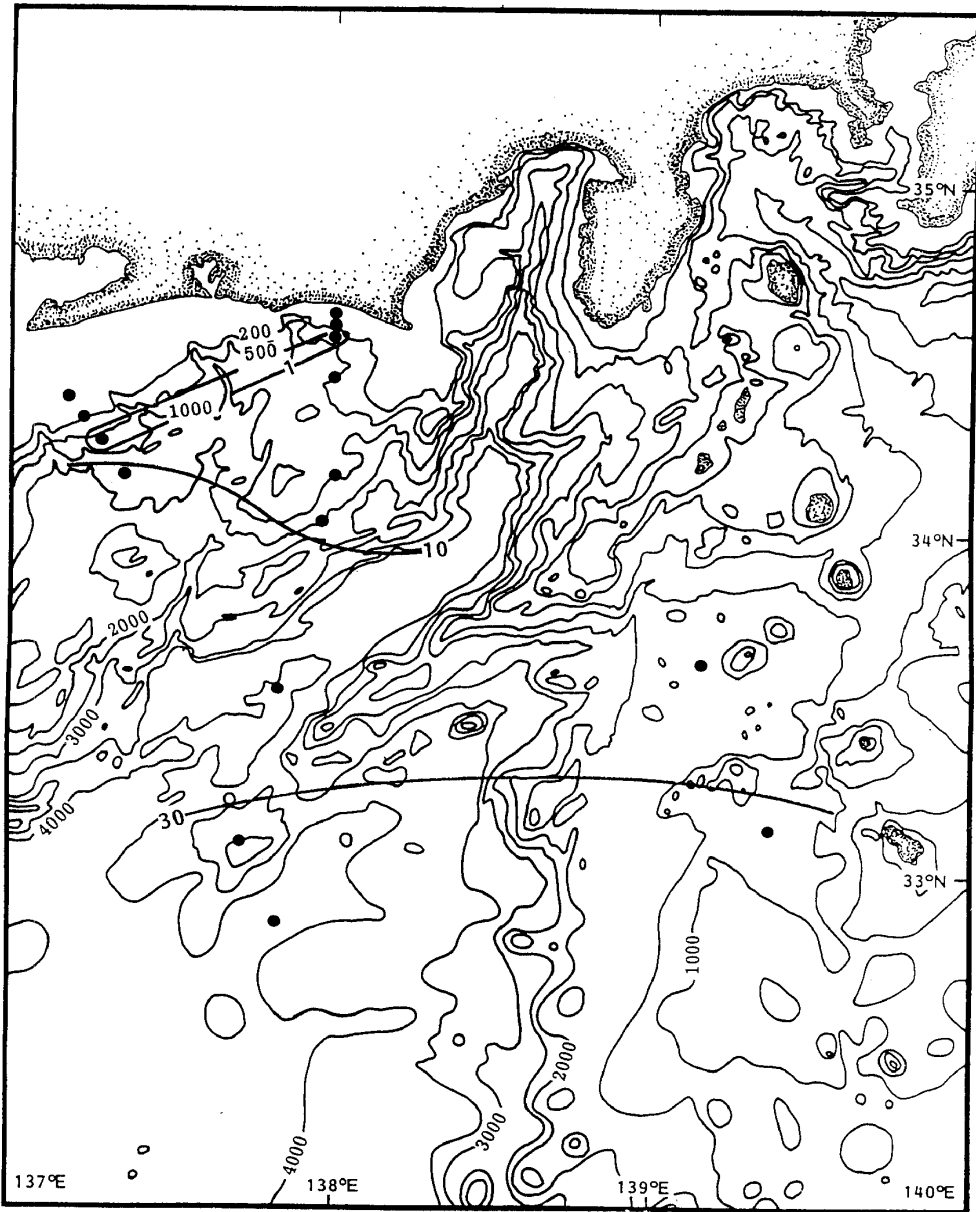


Fig. 26. Distribution of agglutinated foraminifer/total benthic foraminifer ratio (%) in the Enshunada area.

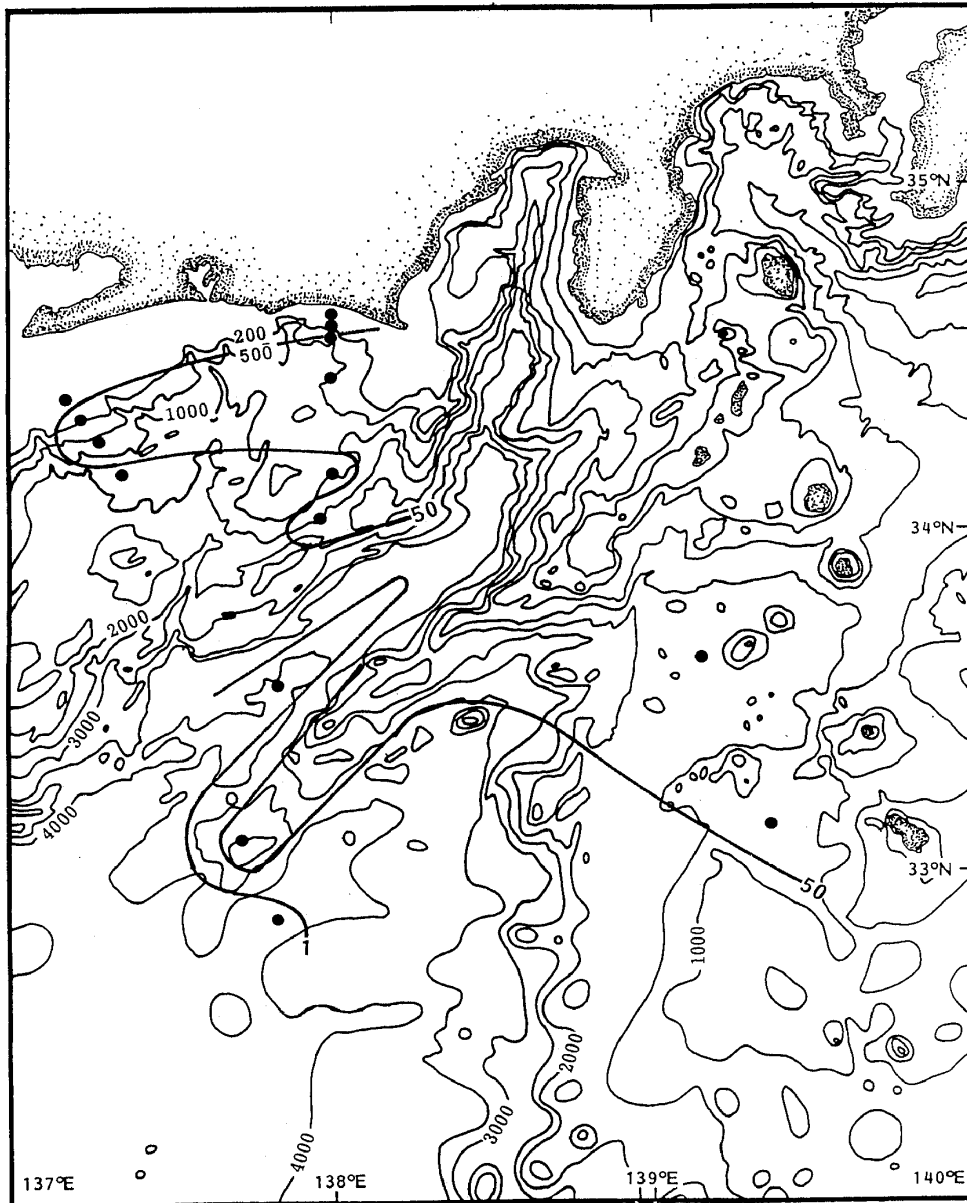


Fig. 27. Distribution of planktonic foraminifer/total foraminifer ratio (%) in the Enshunada area.

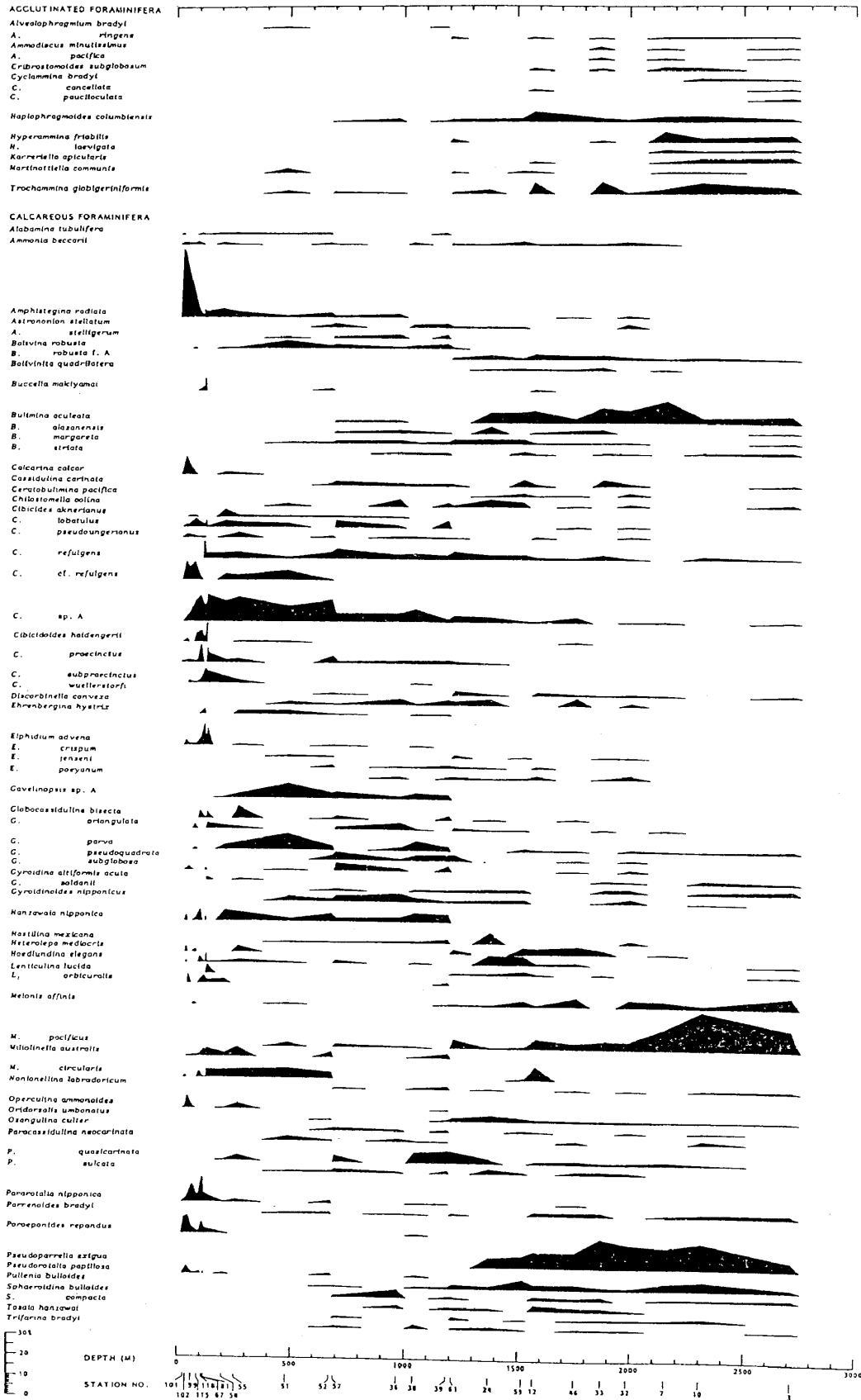


Fig. 28. Depth distribution of important benthic foraminifera off Tanegashima Island.

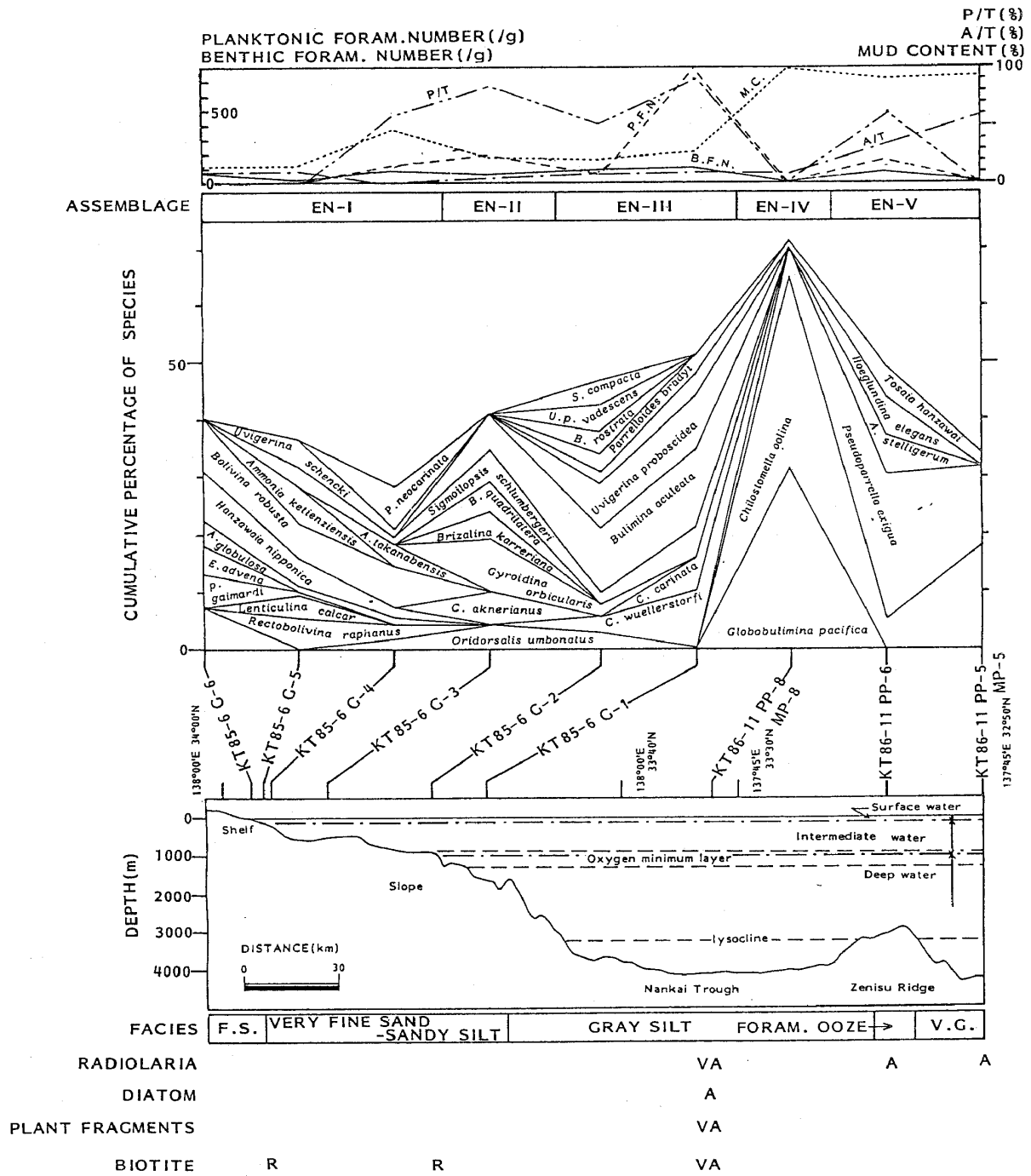


Fig. 29. Bathymetry along 138° E in the Enshunada area, with projected sample locations, cumulative frequency of benthic foraminiferal species, microfaunal statistics (PFN, planktonic foraminifer numbers; BFN, benthic foraminifer numbers; A/T, agglutinated foraminifer/ total benthic foraminifer ratio; and P/T, planktonic foraminifer/total foraminifer ratio), substrata (F.S., fine sand; V.G., volcanic glass), M.C., mud contents, selected grains (R, rare; A, abundant; VA, very abundant).

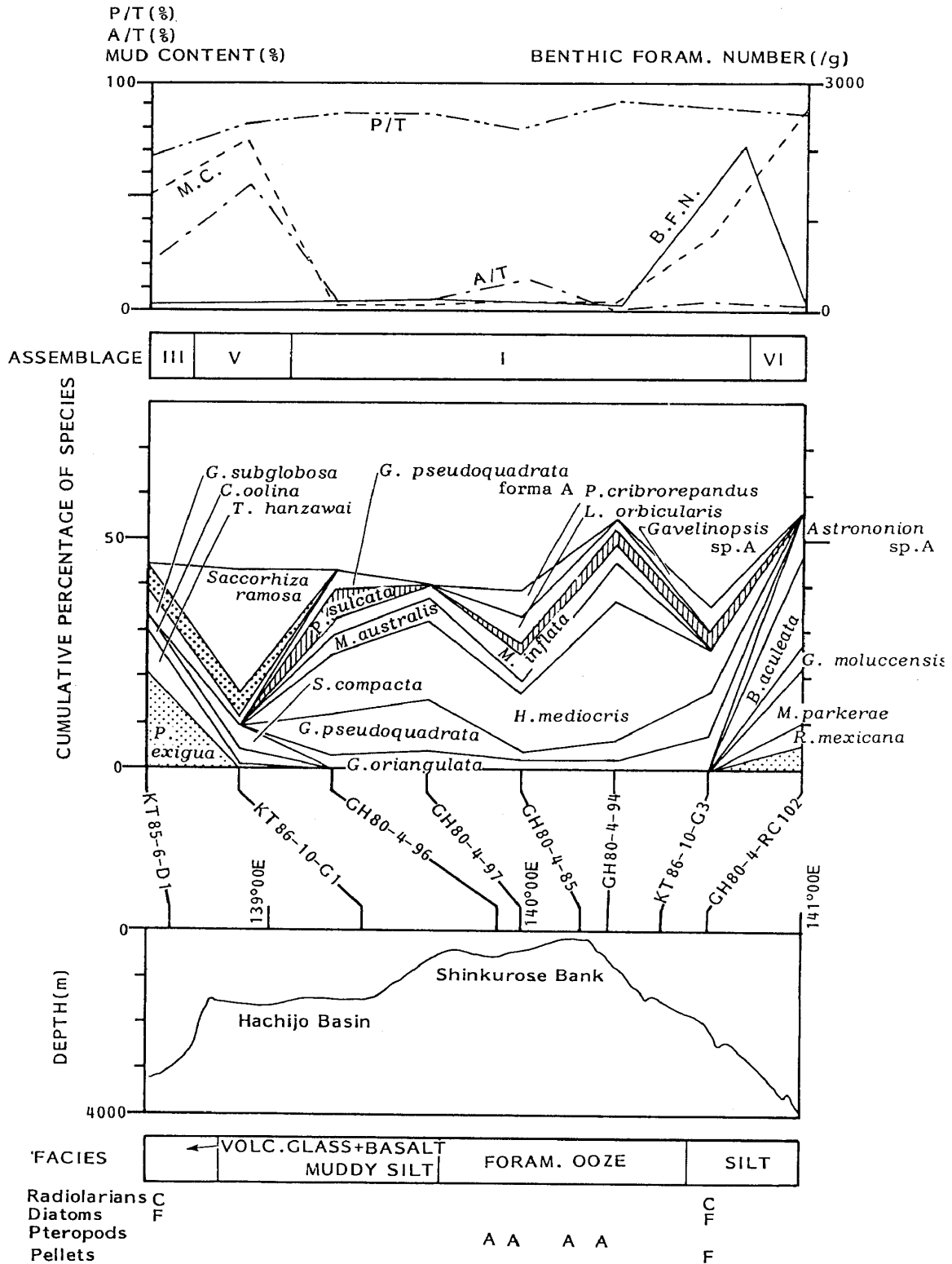


Fig. 30. Bathymetry across the Shinkurose Bank, with projected sample locations, cumulative frequency of benthic foraminiferal species, microfaunal statistics (PFN, planktonic foraminifer numbers; BFN, benthic foraminifer numbers; A/T, agglutinated foraminifer/total benthic foraminifer ratio; P/T, planktonic foraminifer/total foraminifer ratio), substrates, M.C., mud contents, and selected grains (F, few; C, common; A, abundant).

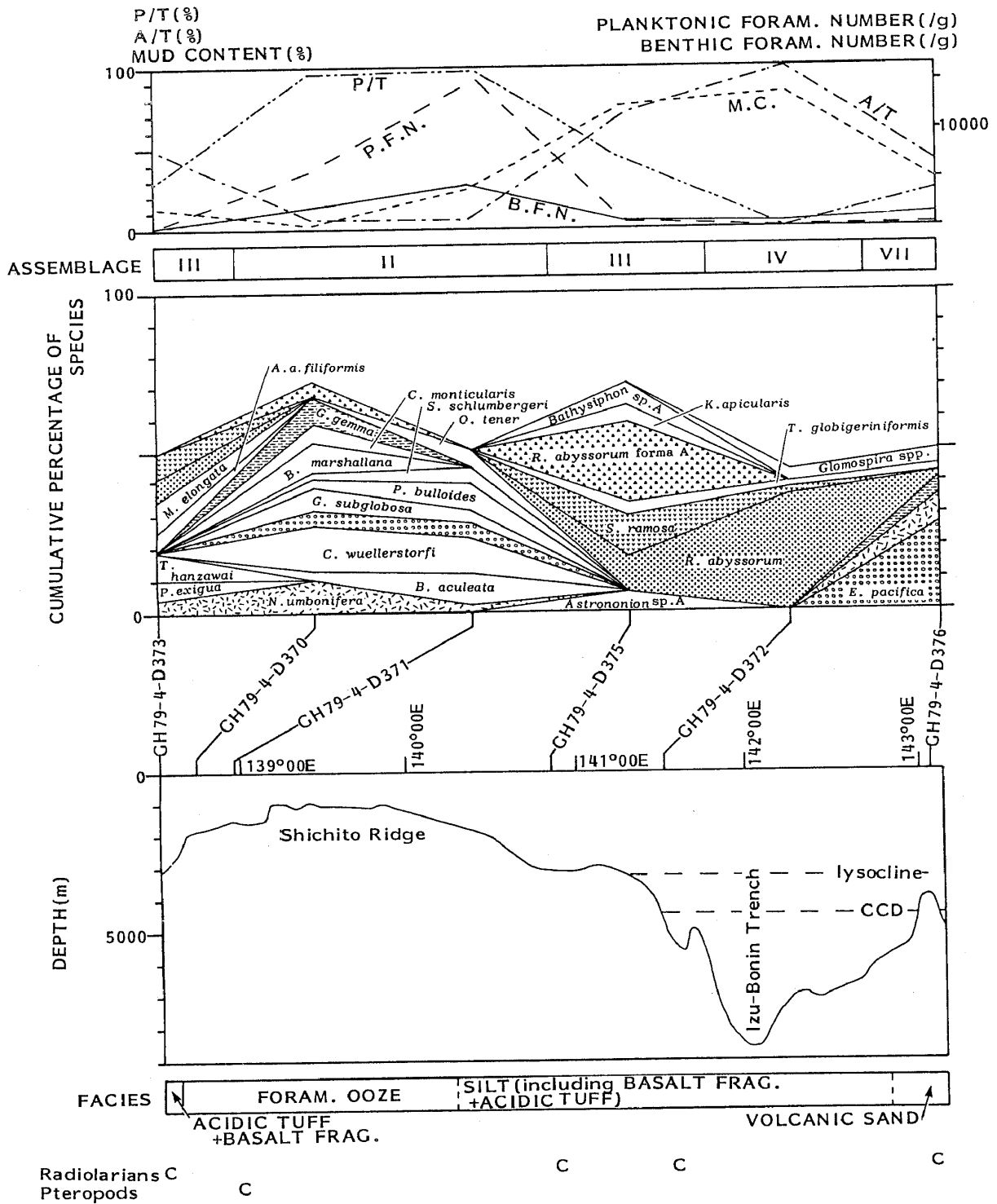


Fig. 31. Bathymetry along 33°N around Hachijojima Island, with projected sample locations, cumulative frequency of benthic foraminiferal species, microfaunal statistics (PFN, planktonic foraminiferal numbers; BFN, benthic foraminiferal numbers; A/T, agglutinated foraminifer/total benthic foraminifer ratio; and P/T, planktonic foraminifer/total foraminifer ratio), substrates, M.C., mud contents, and selected grains (C, common).

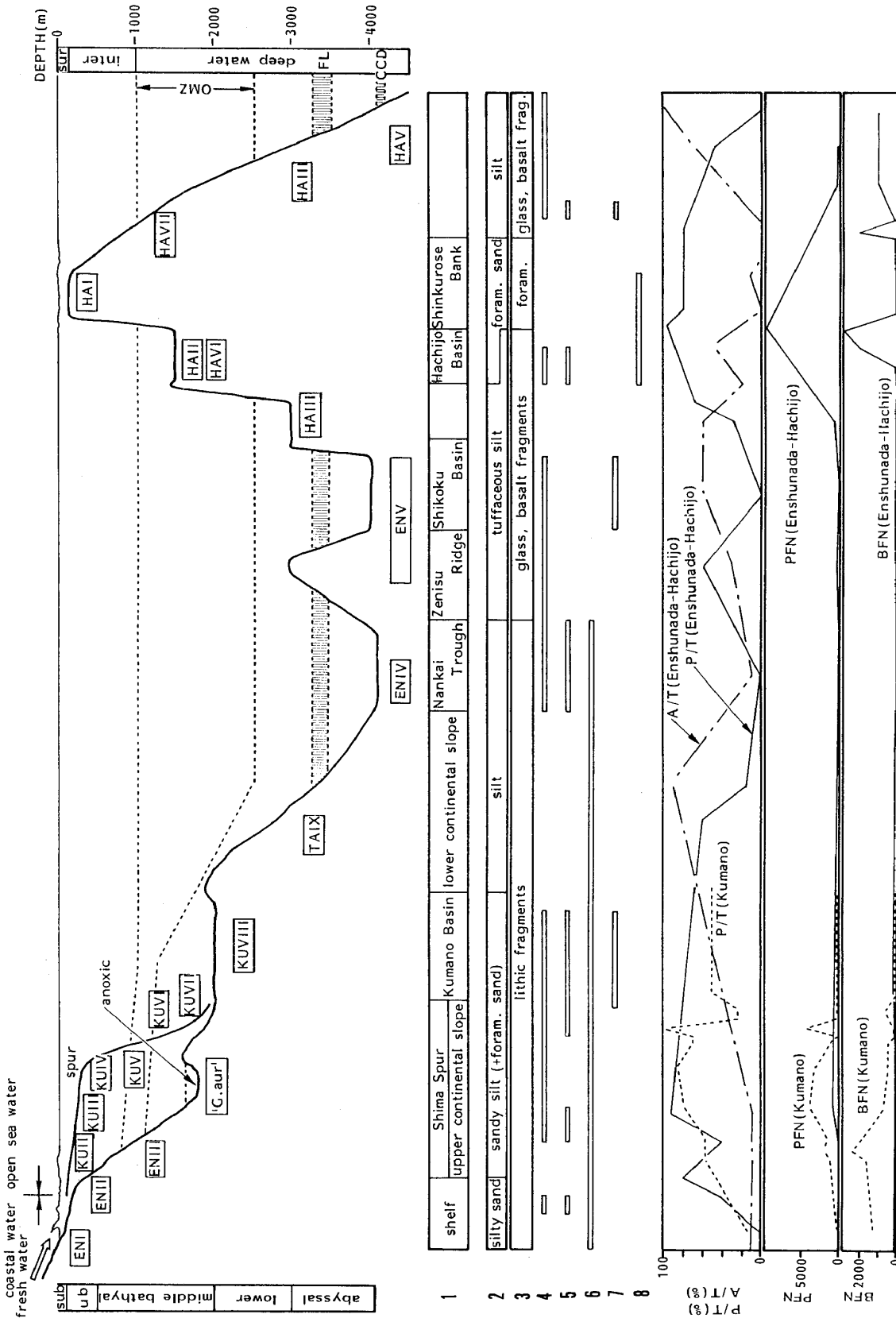


Fig. 32. Significant microfaunal and sedimentologic trends across a schematic littoral to abyssal bathymetric gradient along with associated major physical oceanographic boundaries which are based on the Kumanonada, Enshunada and Hachijojima areas. Letters in blocks below the surface of profile stand for Recent benthic foraminiferal assemblages defined in this paper. sub, sublittoral zone; ub, upper bathyal zone; lower, lower bathyal zone; sur, surface water; OMZ, oxygen minimum zone; FL, foraminiferal lysocline; 'G. aur', *Globobulimina auriculata* assemblage; 1, marine topographic division; 2, sediment types; 3, major grains; 4, radiolarians; 5, diatoms; 6, plant fragments; 7, fecal pellets; 8, pteropods; A/T, agglutinated foraminifer/total benthic foraminifer ratio; P/T, planktonic foraminifer/total foraminifer ratio; PFN, planktonic foraminifer numbers; BFN, benthic foraminifer numbers.

100% between 4,000 m and 5,000 m.

d. P/T ratio (Figs. 30, 31)

Samples around Hachijojima Island are marked by rather high values, which gradually decrease deepward. Sample points with large values (more than 80%) are on the continental slope of the Shichito Ridge above 2,000 m water depth.

In a summary, the above mentioned facts lead to the following lines: 1) Planktonic foraminiferal numbers regularly increase with depth under intermediate waters and decrease below about 1,500 m water depth. The maximum value runs up to more than 4,000 individuals off Enshunada and 15,000 around Hachijojima Island. The number is less than 100 in samples under surface waters. 2) Benthic foraminiferal num-

bers regularly increase with depth down to about 900 m water depth, and decrease below 1,000 m water depth. They are rather small (less than 1,000) at most points. 3) The A/T ratio fundamentally increases deepward on the continental slope. Two peaks of this ratio were recognized at about 3,200 m and 4,100 m. This value becomes 100% at about 4,100 m in the Hachijojima and Enshunada areas. From the A/T ratio, the author estimates the CCD to be at about 4,100 m water depth. 4) The P/T ratio rapidly decreases between 3,000 m and 3,500 m water depth, and becomes 0 at approximately 3,500 m in the Hachijojima and Enshunada areas. The foraminiferal lysocline was estimated at about 3,200 m water depth.

DESCRIPTION OF BENTHIC FORAMINIFERAL ASSEMBLAGE

Foraminiferal assemblages in the four studied areas were determined by Q-mode cluster analysis based on the distribution of foraminifera in each area.

The computer program used in this study has been written by Dr. Shiro Hasegawa to process the similarities of Horn's index of overlap (Horn, 1966) through a weighted pair group method.

1. Tanegashima area

The author examined 25 samples collected by the GH84-3 cruise from off Tanegashima Island. Eight assemblages were recognized and named TA I to TA VIII in accordance to the depth range in this area. The constituent species and distribution of those assemblages are as follows:

1) TA-I assemblage

Abundant species: *Amphistegina radiata*, *Pararotalia nipponica*, *Cibicidoides mediocris*.

Subordinate species: *Calcarina calcar*, *Elphidium advena*.

Depth range: 28-65 m.

Water depth zone: surface water.

Topographic setting: continental shelf at the eastern and southern margins of Tanegashima Island.

Sediment: poorly-sorted calcareous coarse sand, consisting mainly of coral fragments.

Occurrence: GH84-3-99, GH84-3-101, GH84-3-102.

This assemblage contains a considerable number of ill-preserved *Elphidium crispum* and *Miliolinella circularis*. Those specimens may have been winnowed out by the Kuroshio Current, and then were excluded from further analysis.

2) TA-II assemblage

Abundant species: *Cibicidoides mediocris*, *Hanzawaia nipponica* and *Miliolinella circularis*.

Depth range: 88-119 m.

Water depth zone: surface water.

Topographic setting: continental shelf off the Osumi Peninsula and the Osumi strait.

Sediment: well-sorted medium sand composed mostly of quartz grains.

Occurrence: GH84-3-67, GH84-3-81, GH84-3-115 and GH84-3-118.

Apparently derived specimens are absent in those samples. However, foraminiferal tests have been sorted into medium grain size.

3) TA-III assemblage

Abundant species: *Cibicidoides mediocris* and *Miliolinella circularis*.

Subordinate species: *Miliolinella australis*.

Depth range: 196–254 m.

Water depth zone: intermediate water.

Topographic setting: continental slope edge.

Sediment: fine sand.

Occurrence: GH84-3-54 and GH84-3-55.

4) TA-IV assemblage

Abundant species: *Cibicidoides mediocris*, *Gavelinopsis* sp. A and *Globocassidulina parva*.

Subordinate species: *Nonionellina labradorica*.

Depth: 477 m.

Water depth zone: intermediate water.

Topographic setting: continental slope.

Sediment: sandy silt.

Occurrence: GH84-3-51.

5) TA-V assemblage

Abundant species: *Cibicidoides mediocris*.

Subordinate species: *Paracassidulina quasicarinata*.

Depth range: 676–1,185 m.

Water depth zone: intermediate water to deep water.

Topographic setting: small slope break.

Sediment: sandy silt.

Occurrence: GH84-3-38, GH84-3-39, GH84-3-52 and GH84-3-57.

6) TA-VI assemblage

Abundant species: *Paracassidulina quasicarinata*.

Subordinate species: *Cibicidoides*

mediocris.

Depth range: 970–1,213 m.

Water depth zone: deep water (oxygen minimum zone).

Topographic setting: within valley on continental slope.

Sediment: sandy silt.

Occurrence: GH84-3-36 and GH84-3-61.

7) TA-VII assemblage

Abundant species: *Bulimina aculeata*, *Hoeglundina elegans* and *Pseudoparrella exigua*.

Subordinate species: *Haplophragmoides columbiensis* and *Trochammina globigeriniformis*.

Depth range: 1,371–1,520 m.

Water depth zone: deep water (oxygen minimum zone).

Topographic setting: continental slope.

Sediment: sandy silt.

Occurrence: GH84-3-24 and GH84-3-59.

8) TA-VIII assemblage

Abundant species: *Melonis pacificus* and *Pseudoparrella exigua*.

Subordinate species: *Bulimina aculeata*, *Rhabdammina abyssorum* and *Trochammina globigeriniformis*.

Depth range: 1,500–2,710 m.

Water depth zone: deep water (above foraminiferal lysocline).

Topographic setting: continental slope with a small slope break.

Sediment: sandy silt.

Occurrence: GH84-3-3, GH84-3-7, GH84-3-10, GH84-3-12, GH84-3-32, GH84-3-33 and GH84-3-36.

This assemblage is recognized in the lower middle bathyal to abyssal zone. *Bulimina aculeata* rapidly decreases under the foraminiferal lysocline.

9) TA-IX assemblage

Abundant species: *Rhabdammina abyssorum*.

Subordinate species: *Reophax difflugiformis*, *Rhizammina algaeformis* and *Saccorhiza ramosa*.

Depth : ca. 3,200 m.

Water depth zone : deep water (below foraminiferal lysocline).

Topographic setting : lower continental slope off Tosa Bay.

Sediment : silt.

Occurrence : KT87-10-PP5.

This assemblage was recognized offshore Tosa Bay, where are additional occurrences of several calcareous benthic foraminifera, mostly of *Tosaiia hanzawai*.

2. Kumanonada area

The author examined 14 samples collected by the GH82-2 cruise from off Kumanonada. Eight assemblages were recognized and named KU-I to KU-VIII in accordance to the depth range in this area. The constituent species and distribution of those assemblages are as follows :

1) KU-I assemblage

Abundant species : *Gyroidinoides nipponicus*, *Hanzawaia nipponica*, *Miliolinella circularis* and *Quinqueloculina akneriana*.

Subordinate species : *Textularia* spp., *Bulimina marginata*, *Cibicides subhaidingerii*, *Gavelinopsis* sp. A, *Gyroidina cushmani*, *Hoeglundina elegans* and *Rosalina vilardeboana*.

Depth : 100 m.

Water depth zone : surface water.

Topographic setting : top of continental slope (rarely on continental shelf).

Sediment : medium sand.

Occurrence : GH82-2-24.

2) KU-II assemblage

Abundant species : *Bolivina robusta*, *Bulimina marginata*, *Cibicides aknerianus*, *Gavelinopsis* sp. A, *Globocassidulina subglobosa*, *Gyroidinoides nipponicus*, *Cibicidoides mediocris*, *Hoeglundina elegans* and *Planulina ariminensis*.

Subordinate species : *Spiroplectamina higuchii*, *Elphidium advena* and *Uvigerina vadescens* var.

Depth range : 186-238 m.

Water depth zone : intermediate water.

Topographic setting : continental slope at western root of Shima Spur.

Sediment : fine sand.

Occurrence : GH82-2-24, GH82-2-31 and GH82-2-38.

3) KU-III assemblage

Dominant species : *Gyroidinoides nipponicus*, *Bolivina robusta* and *Uvigerina hispidocostata*.

Depth : 465 m.

Water depth zone : intermediate water.

Topographic setting : continental slope at western root of Shima Spur.

Sediment : fine sand.

Occurrence : GH82-2-39.

4) KU-IV assemblage

Abundant species : *Bulimina aculeata* and *Bulimina marginata*.

Subordinate species : *Bolivina robusta* forma A and *Uvigerina hispidocostata*.

Depth : 607 m.

Water depth zone : intermediate water.

Topographic setting : at western root of Shima Spur.

Sediment : coarse sand (foraminiferal sand).

Occurrence : GH82-2-25.

5) KU-V assemblage

Abundant species : *Bulimina rostrata*, *Chilostomella oolina*, *Nonionellina labradorica*, *Pseudoparrella exigua*, *Rutherfordooides mexicanus* and *Sphaeroidina compacta*.

Subordinate species : *Bulimina aculeata*, *Bulimina striata*, *Oridorsalis umbonatus*, *Pullenia salisburyi*, *Uvigerina hispidocostata* and *Uvigerina vadescens*.

Depth : 982 m.

Water depth zone : intermediate water (oxygen minimum zone).

Topographic setting : continental slope beneath western root of Shima Spur.

Sediment : silt.

Occurrence : GH82-2-33.

6) KU-VI assemblage

Abundant species : *Bulimina aculeata* and *Chilostomella oolina*.

Subordinate species : *Alveolophragmium subglobosum*, *Pseudoparrella exigua*, *Rutherfordoides mexicanus* and *Uvigerina hispidocostata*.

Depth : 1,492 m.

Water depth zone : deep water.

Topographic setting : western slope of Shima Spur.

Sediment : silt.

Occurrence : GH82-2-41.

7) KU-VII assemblage

Abundant species : *Bulimina aculeata* and *Cassidulina carinata*.

Subordinate species : *Bolivinita quadrilatera*.

Depth : 1,642 m.

Water depth zone : deep water.

Topographic setting : western slope of Shima Spur.

Sediment : silt.

Occurrence : GH82-2-50.

8) KU-VIII assemblage

Abundant species : *Pseudoparrella exigua*, *Uvigerina hispidocostata* and *Bulimina aculeata*.

Subordinate species : *Martinottiella communis*, *Chilostomella oolina*, *Melonis parkerae* and *Uvigerina proboscidea*.

Depth range : 1,872-2,082 m.

Water depth zone : deep water (above foraminiferal lysocline).

Topographic setting : basin floor of Kumano Basin.

Sediment : sandy silt.

Occurrence : GH82-2-59, GH82-2-60, GH82-2-70, GH82-2-102 and GH82-2-103.

3. Enshunada area

The author examined 10 samples collected by the KT85-6 cruise from off Enshunada and six samples by the KT86-11 cruise in the Nankai Trough. Five assemblages were recognized and

named EN-I to EN-V in accordance to the depth range in this area. The constituent species and distribution of those assemblages are as follows :

1) EN-I assemblage

Abundant species : *Bolivina robusta*, *Hanzawaia nipponica* and *Cibicidoides mediocris*.

Subordinate species : *Ammonia ketienziensis*, *Ammonia takanabensis*, *Rectobolivina raphanus* and *Uvigerina schencki*.

Depth range : 48-198 m.

Water depth zone : surface water.

Topographic setting : on continental shelf.

Sediment : silty fine sand.

Occurrence : KT85-6-G4, KT85-6-G5, KT85-6-G6, KT85-6-G9 and KT85-6-G10.

This assemblage is subdivided into two groups: one is distributed off the mouth of the River Tenryugawa, and includes abundant *A. ketienziensis*; the other is off Ise Bay, and consists mainly of *Cibicidoides mediocris*.

2) EN-II assemblage

Abundant species : *Gyroidina orbicularis* and *Bulimina aculeata*.

Subordinate species : *Bolivinita quadrilatera*, *Cibicides aknerianus* and *Oridorsalis umbonatus*.

Depth range : 495-587 m.

Water depth range : intermediate water.

Topographic setting : on continental slope.

Sediment : sandy silt.

Occurrence : KT85-6-G3 and KT85-6-G8.

3) EN-III assemblage

Abundant species : *Bulimina aculeata* and *Uvigerina proboscidea*.

Subordinate species : *Cibicides wuellerstorfi* and *Parrelloides bradyi*.

Depth range : 1,000-1,500 m.

Water depth zone : intermediate water (oxygen minimum zone).

Topographic setting : continental

slope.

Sediment: sandy silt.

Occurrence: KT85-6-G1, KT85-6-G2 and KT85-6-G7.

4) EN-IV assemblage

Abundant species: *Globobulimina pacifica* and *Chilostomella oolina*.

Depth: 4,047 m.

Water depth zone: deep water (above CCD).

Topographic setting: Nankai Trough.

Sediment: silt.

Occurrence: KT86-11-P8.

5) EN-V assemblage

Abundant species: *Pseudoparrella exigua* and *Chilostomella oolina*.

Subordinate species: *Tosaia hanzawai*.

Depth range: 3,000–4,050 m.

Water depth zone: deep water (around foraminiferal lysocline).

Topographic setting: bottom floor of Shikoku Basin and summit of Zenisu Ridge.

Sediment: silt, volcanic glass and basalt fragments.

Occurrence: KT86-11-PP5 and KT86-11-PP6.

4. Hachijojima Island area

The author examined 18 samples collected by the GH79-4, GH80-4, KT86-10 and 11 cruises from around Hachijojima Island. Six assemblages were recognized and named HA-I to HA-VI in accordance to the depth range in this area. The constituent species and distribution of those assemblages are as follows:

1) HA-I assemblage

Abundant species: *Cibicidoides mediacris*.

Subordinate species: *Globocassidulina pseudoquadrata*.

Depth range: 150–500 m.

Water depth zone: intermediate water.

Topographic setting: summit of Shin-kurose Bank.

Sediment: calcareous sand containing foraminiferal tests and molluscan valves.

Occurrence: GH80-4-85, GH80-4-94, GH80-4-96, GH80-4-97 and KT86-10-G3.

This assemblage includes *Miliolinella australis*, *Miliolinella circularis*, *Poroponides cribrorepandus* and *Robulus orbicularis* at shallower depths. The constituent species of this assemblage resemble those of TA-II or III, but no *Amphistegina* spp. are present in this assemblage.

2) HA-II assemblage

Abundant species: *Cibicides wuellerstorfi*.

Subordinate species: *Rhabdammina abyssorum*.

Depth range: 1,500–1,700 m.

Water depth zone: intermediate water (oxygen minimum zone).

Topographic setting: continental slope on back arc side of Shichito Ridge.

Sediment: calcareous sand (mainly foraminiferal sand).

Occurrence: GH79-4-D370 and GH79-4-D371.

This assemblage includes *Bulimina aculeata*, *Burserolina marshallana*, *Globocassidulina gemma*, *Globocassidulina subglobosa* and *Ehrenbergina pacifica*. *Nuttallides umbonifera* is abundant at only three points on the western continental slope.

Similar assemblages were also recognized at the same depth in the Tanegashima and Kumanonada areas. Comparison shows that the constituent *Cassidulina* species of the assemblages differ among areas: *Paracassidulina quasicarinata* is present in TA: *Cassidulina carinata* in KU.

3) HA-III assemblage

Abundant species: *Rhabdammina abyssorum* and *Pseudoparrella exigua*.

Subordinate species: *Saccorhiza ramosa*, *Chilostomella oolina*,

Globocassidulina subglobosa and *Tosaia hanzawai*.

Depth range: ca. 2,900–3,600 m.

Water depth zone: deep water (about foraminiferal lysocline).

Topographic setting: small basin on continental slope on backarc side of Shichito Ridge.

Sediment: silt, volcanic glass and basalt fragments.

Occurrence: GH79-4-D373, GH79-4-D375 and KT85-6-D1.

The assemblage at KT85-6-D1 resembles EN-V in the Enshunada area, but the latter assemblage does not include *Rhabdammina abyssorum*. The last two samples, collected at about the same depth on the east and west sides of the Shichito Ridge, contain nearly the same species composition, which implies that the deep water mass does not fundamentally differ on both sides of the ridge.

4) HA-IV assemblage

Abundant species: *Rhabdammina abyssorum*.

Subordinate species: *Glomospira* spp.

Depth: 5,200 m.

Water depth zone: deep water (below CCD).

Topographic setting: small basin on continental slope of Izu-Bonin Arc.

Sediment: silt including basalt fragments and acidic tuff.

Occurrence: GH79-4-D371.

5) HA-V assemblage

Abundant species: *Saccorhiza ramosa*

Subordinate species: *Rhabdammina abyssorum* and *Sphaeroidina compacta*.

Depth: 1,500 m.

Water depth zone: intermediate water (oxygen minimum zone).

Topographic setting: basin bottom of Hachijo Basin.

Sediment: silt.

Occurrence: KT86-10-G1.

This assemblage is characterized by an abundant occurrence of *Saccorhiza ramosa*. This species occurs in abundance at KT85-6-7 off Atsumi in the

Enshunada area, while *Uvigerina proboscidea* is abundant at KT85-6-G7, even though both points are situated at the same depth.

6) HA-VI assemblage

Abundant species: *Bulimina aculeata* and *Melonis parkerae*

Subordinate species: *Rutherfordoides mexicanus* and *Pseudoparrella exigua*.

Depth: 2,169 m.

Water depth zone: deep water (oxygen minimum zone).

Topographic setting: small basin on continental slope of Izu-Bonin Arc.

Sediment: silt including basalt fragments and acidic tuff.

Occurrence: GH80-4-RC102.

Although *Pseudoparrella exigua* is rather meager, this assemblage is almost identical with TA-VIII in the Tanegashima area. The species composition of KU-VIII in the Kumanonada area is likely what *Uvigerina hispidocostata* is added on to HA-VI.

The sum of the above descriptions is as follows:

1) Although water depth is equivalent, the assemblages of the Nankai Trough and Shikoku Basin differ from each other, as expressed by EN-V and EN-VI. This difference may be attributed to dissimilarities in the nature of sediments, resulted from distinctive submarine topography.

2) Where wide hinterlands exist, the following forms become dominant with water depth: *Ammonia ketienziensis*, *Bolivina robusta*, *Uvigerina* spp. (*U. proboscidea* and *U. hispidocostata*) and *Rhabdammina abyssorum*.

3) Where narrow or no hinterlands are present, assemblages are dominated by *Miliolinella circularis* — *M. australis*, *Cibicidoides mediocris*, *Globocassidulina* spp. — *Paracassidulina* spp., *Pseudoparrella exigua* — *Bulimina aculeata* and *Rhabdammina abyssorum* by turns in deepening order.

4) *Rhabdammina abyssorum* usually

occurs in abundance below the foraminiferal lysocline, but subordinate species differ among assemblages: for example, *Bulimina aculeata* and *Melonis pacificus* in TA-VIII, *Reophax*

diffugiiformis in TA-IX and *Glomospira* spp. in HA-V.

5) The deep water assemblage does not fundamentally differ on both the east and west sides of the Shichito Ridge.

DISTRIBUTION OF SELECTED SPECIES

The following 16 species, each can be traced back to the Neogene, were selected on the basis of principal components scores for facilitating paleoenvironmental analysis. Figure 33 shows their distribution in the water column off southwest Japan on the basis of Figs. 28-31.

Bathysiphon sp.: In all traverses, usually less than 3% of the fauna; only occurs below 1,000 m.

Alveolophragmium subglobosum (G.O. Sars): In all traverses, usually less than 3% of the fauna; occurs below 1,000 m. A sample off Ito, under anaerobic conditions, yielded a few specimens of this species, and no other agglutinated species.

Rhabdammina abyssorum M. Sars: Present in all traverses, running up to more than 9% only on the continental slope below the foraminiferal lysocline. This species is absent within the Nankai Trough and Shikoku Basin, located below the lysocline. The mud content of sediments containing this species is approximately 70%.

Trochammmina globigeriniformis (Parker and Jones): Present in all traverses, being usually less than 3% of the fauna. The mud content of sediments containing this species is approximately 70%.

Ammonia ketienziensis (Ishizaki): Sporadic down to over 200 m, most characteristic at depths between 80 m and 200 m; abundant in shallow water and fine sandy sediments off the mouth of the River Tenryugawa.

Ammonia takanabensis (Ishizaki): Most characteristic in a depth range nar-

rower than *A. ketienziensis*, spanning from 100 m and 200 m. The distribution of this species is also regional. It is abundant in shallow water and fine sandy sediments off the mouth of the River Tenryugawa.

Cibicides aknerianus (d'Orbigny): Occurs from a few points in two traverses (6% or less); in the traverse off Kumanonada approximately 6% at 215 m; in the traverse off the River Tenryugawa 6% at 150 m; ranges from 80 to 1,000 m in water depth.

Cibicides wuellerstorfi (Schwager): Recorded at a depth ranging from 200 m to 4,000 m; in the traverse off the River Tenryugawa approximately 10% at about 1,400 m water depth; occurs most frequently in the traverse around Hachijojima, between 1,500 m and 1,700 m, being as high as 15%. The mud content of samples containing large individuals is about 70%. Not widely distributed in other traverses.

Chilostomella oolina Schwager: Present in all traverses in abundance below 800 m water depth. Numerous living individuals were found within the Nankai Trough. Aside from this trough, it varies in abundance from sample to sample within limited ranges of water depth and area. The mud content of samples containing large numbers is more than 90%.

Cibicidoides mediocris (Finlay): At many stations in all traverses more than 3% of the total population; commonly over 9% between 100 m and 700 m, with a maximum of 20% at about 500 m water depth; no records

from more than about 2,000 m water depth.

Elphidium crispum (Linné): At a few stations shallower than 250 m in several traverses it is less than 1% of the total population.

Globobulimina auriculata (Bailey): Occurs in low abundance between 1,000 m and 2,200 m water depth in the oxygen minimum zone. A unique sample off Ito, apparently under anoxic conditions, yielded abundant specimens of this species (more than 90%). Thus, this species is related with dissolved oxygen content.

Gyroidina orbicularis d'Orbigny: At many stations in all traverses it is less than 1% of the total population.

Melonis parkerae (Uchio): Occurs in most samples from all traverses and at all depths above the CCD, with highest frequencies (commonly about 3%) between 1,000 m and 2,200 m.

Robulus orbicularis d'Orbigny: At a few stations in all traverses its frequency is usually 3% or less; In the traverse around Hachijojima Island approximately 6% at 195 m. The lower depth limit is probably 500 m.

Uvigerina proboscidea Schwager: Occurs between 1,000 m and 2,200 m water depth off southwest Japan; abundant in samples of sandy silt bottom under deep waters; Nothing was found on the continental slope around Hachijojima and Tanegashima islands.

COMPARISON WITH PREVIOUS STUDY

Several previous workers who investigated modern NW Pacific faunas off southwest Japan mentioned the relationship between the faunas and oceanographic conditions (Ishiwada, 1964; Aoshima, 1978; Inoue, 1986). The characteristics of assemblages and their mutual relations reported by previous workers from the NW Pacific are briefly mentioned below. Papers referred to here are those by Ishiwada (*op. cit.*) and Aoshima (*op. cit.*) in Tosa Bay; Aoshima (*op. cit.*) off Ogasa, Atsumi, Kumano and Miyazaki; and Inoue (*op. cit.*) in Suruga Bay, all in the NW Pacific off southwest Japan. Figure 33 shows the benthic foraminiferal assemblages which were distinguished by previous workers, assemblages in the four areas mentioned above, and relations between those assemblages and water depth. Comparison between the assemblages established by previous studies and those of the present author is also shown in this figure.

Inoue (1986) described 12 assemblages based on dominant species, including three assemblages recognized in the En-

shunada area. The *Bolivinita quadrilatera* Assemblage is distributed at 245 m to 485 m water depth off the Oigawa, and includes *Cassidulina carinata* and *Hyalinea balthica*. This assemblage is similar to EN-II assemblage in this study. The *Bolivina robusta* Assemblage is characterized by abundant occurrences of *B. robusta* and *Cassidulina carinata*, and is confined to 265 m to 660 m water depth on the western slope of Suruga Bay. The *Rectobolivina raphanus* Assemblage contains *Brizalina karrerianum*, *Quinqueloculina* spp. and *Hanzawaia nipponica* in shallow water off Omaezaki. The both assemblages correspond to EN-I off the mouth of the River Tenryugawa in this study.

Aoshima (1978) reported four assemblages in relation to water masses off southwest Japan. The species association of those four assemblages has not been described in his paper, but among the species six types of distribution are recognized relative to water masses. Those types almost coincide with the result of the present author.

Ishiwada (1964) described a benthic

assemblage off Tosa together with assemblages from other areas off the Pacific coast of Japan. The Tosa fauna includes four biofacies: *Bolivina robusta*, *Rectobolivina raphanus* and *Pseudorotalia papillosa*, accompanied by *Ammonia ketienziensis* in shallow water (Facies IA; 50-80 m water depth); *Bolivina robusta*, *Bulimina marginata* and *Uvigerina proboscidea vadescens* (Facies IB; 100-280 m water depth); *Bulimina nipponica* with *Uvigerina peregrina disrupta* (Facies IIA; 400 to 680 m); and *Bulimina nipponica* associated with

Martinottiella communis and *Cibicides wuellerstorfi* (IIB; 900 m water depth). Although his studied area differs geographically, Facies IA and IB are similar to assemblage EN-1 off the River Tenryugawa.

Thus, it is considered that assemblages assorted by the present author in the Enshunada and Kumanonada areas are widely distributed off southwest Japan. On the contrary, assemblages around Hachijojima Island and the Tanegashima area differ from those off southwest Japan, but no comparison is made

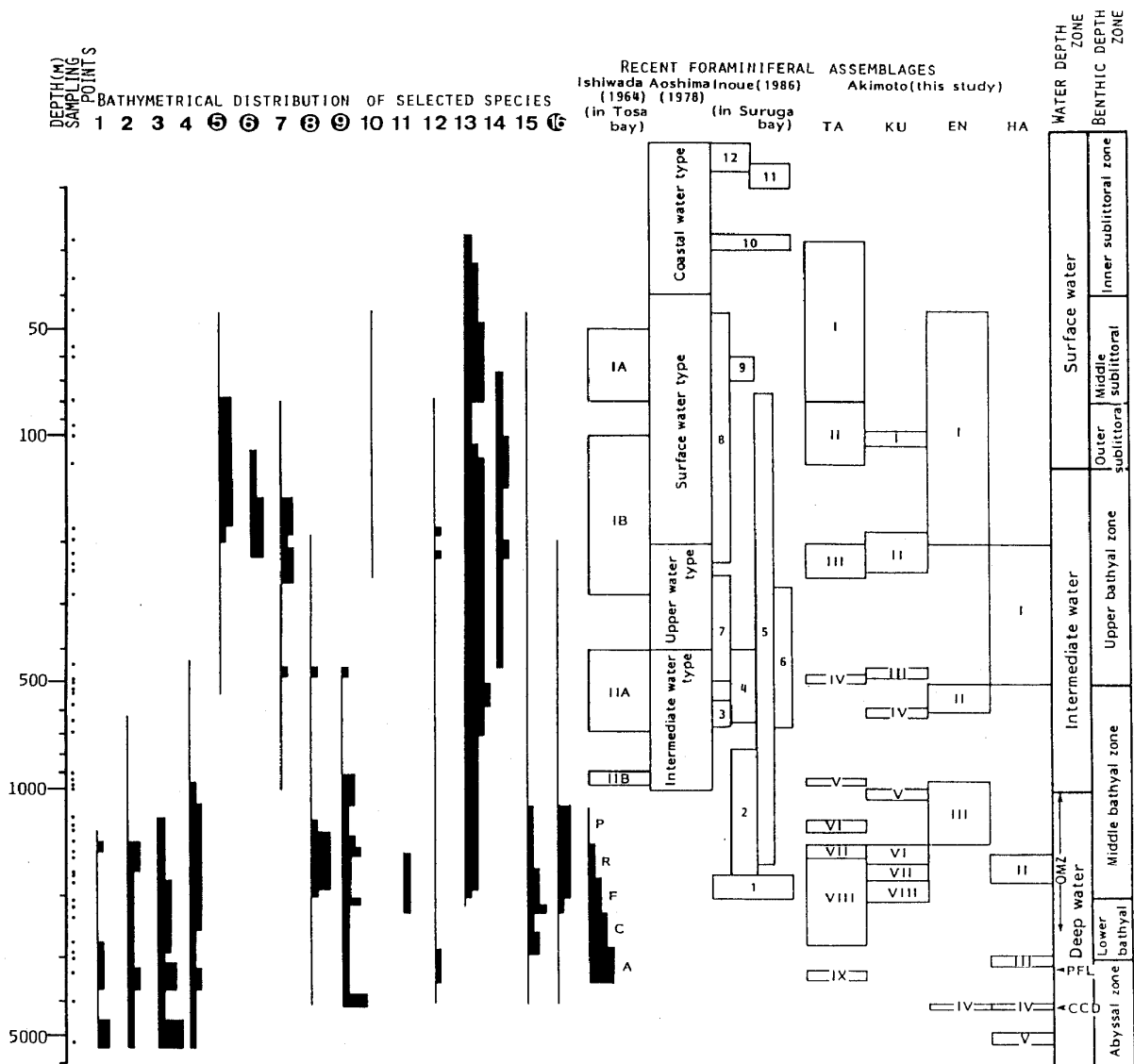


Fig. 33.

because of lack of data for deeper water assemblages.

FAUNAL STRUCTURE

Gibson and Buzas (1973) studied diversity of Recent benthic foraminifera distributed off along the eastern margin of North America, and revealed paleoenvironments of the Miocene sequence distributed along the east coast by comparison with Recent assemblages.

The present author incidentally investigated diversity of Recent benthic foraminiferal assemblages distributed off southwest Japan, in connection with water masses, sediment characteristics,

and submarine topography.

Benthic foraminiferal diversity and equitability are expressed by Shannon-Weaver's information function as somewhat revised by Buzas and Gibson (1969). Those two parameters are given in the following formulae :

$$H(s) = - \sum_{i=1}^s p_i \ln p_i, \text{ and } E = e^{H(s)}/S$$

where p_i stands for the proportion of the i th species in a sample, and S the num-

Fig. 33. Bathymetric distribution and frequency of selected species, and comparison of established assemblages by Ishiwada (1964), Aoshima (1978) and Inoue (1986) with those of Recent benthic foraminifera in the Pacific Ocean off southwest Japan, determined by the present author. Numbers indicate the following selected species :

1. *Bathysiphon* sp.
2. *Alveolophragmium subglobosa* (G.O. Sars)
3. *Rhabdammina abyssorum* M. Sars
4. *Trochammina globigeriniformis* (Parker and Jones)
5. *Ammonia ketienziensis* (Ishizaki)
6. *Ammonia takanabensis* (Ishizaki)
7. *Cibicides aknerianus* (d'Orbigny)
8. *Cibicides wuellerstorfi* (Schwager)
9. *Chilostomella oolina* Schwager
10. *Elphidium crispum* (Linné)
11. *Globobulimina auriculata* (Bailey)
12. *Gyroldina orbicularis* d'Orbigny
13. *Cibicidoides mediocris* (Finlay)
14. *Lenticulina orbicularis* (d'Orbigny)
15. *Melonis parkerae* (Uchio)
16. *Uvigerina proboscidea* Schwager

White numbers in solid circles indicate species whose frequencies vary from area to area. TA, off Tanegashima Island; KU, off Kumanonada; EN; off Enshunada; HA, around Hachijojima Island.

Numbers for Recent assemblages established by Inoue (1986) in Suruga Bay stand for as follows :

1. *Bulimina aculeata* Assemblage
2. *Nonionellina labradorica* Assemblage
3. *Stainforthia exilis tenuata* Assemblage
4. *Bulimina aculeata-Stainforthia exilis tenuata* Assemblage
5. *Eggerella scabra* Assemblage
6. *Bolivinita quadrilatera* Assemblage
7. *Bolivina robusta* Assemblage
8. *Cibicides* spp. Assemblage
9. *Siphogenerina raphanus* Assemblage
10. *Pararotalia nipponica* Assemblage
11. *Ammobaculites exiguus-Ammonia japonica* Assemblage
12. *Pseudononion japonicum-Ammonia beccarii* Assemblage

ber of species. The latter equation equals the formulation, $H(s) = \ln(S \cdot E)$. By this formula, the relationships among the three parameters, diversity, equitability and number of species in a sample can be expressed simultaneously.

1. Tanegashima area

The above-mentioned three parameters are shown in Fig. 34 for 24 samples. Six clusters of points may be interpreted in relation to water depth and topographic features. Clusters of the bathyal zone are characterized by high values of diversity, equitability and number of species. On the contrary, clusters of the sublittoral zone show moderate values of diversity, equitability and number of species.

2. Kumanonada area

Figure 35 shows the three parameters for all 16 samples. All points are grouped into six clusters based on their topographic features and water depth. Groups 1 and 2 (sublittoral zone), group 3 (upper bathyal zone) and group 4 (upper middle bathyal zone) are relative-

ly high in number of species, moderate in diversity and equitability. Samples of the lower bathyal zone are grouped under two categories based on equitability: groups 5 and 6 are represented by rather low equitability, diversity and number of species; group 6 is characterized by much less equitability values. Those two groups stand for different topography in this area: group 5 is located on the upper continental slope and group 6 on the bottom floor of the Kumano Basin.

3. Enshunada area and Shinkurose Bank

Figure 36 shows the three parameters for all 18 samples. These points are grouped into seven clusters based on their topographic features and water depth. Group 1 on the Shinkurose Bank bears large numbers of species, moderate in diversity and equitability. On the other hand, group 4 off Enshunada in the same water depth as group 1 is characterized by moderate values of diversity, equitability and

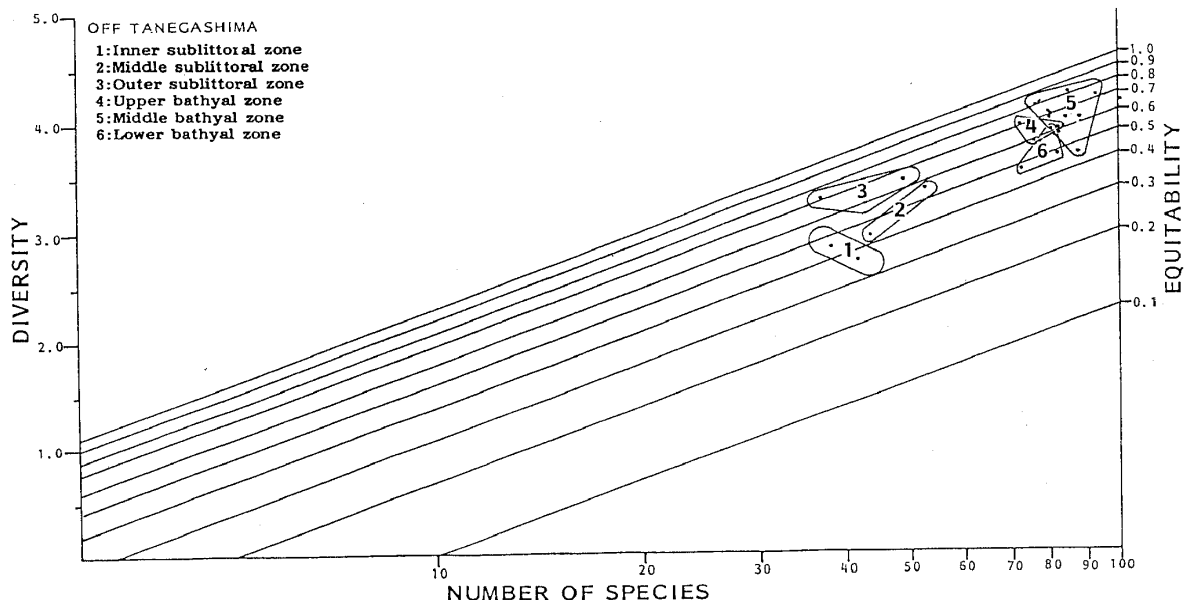


Fig. 34. Diagram showing species diversity, equitability, and number of species of samples from off Tanegashima Island.

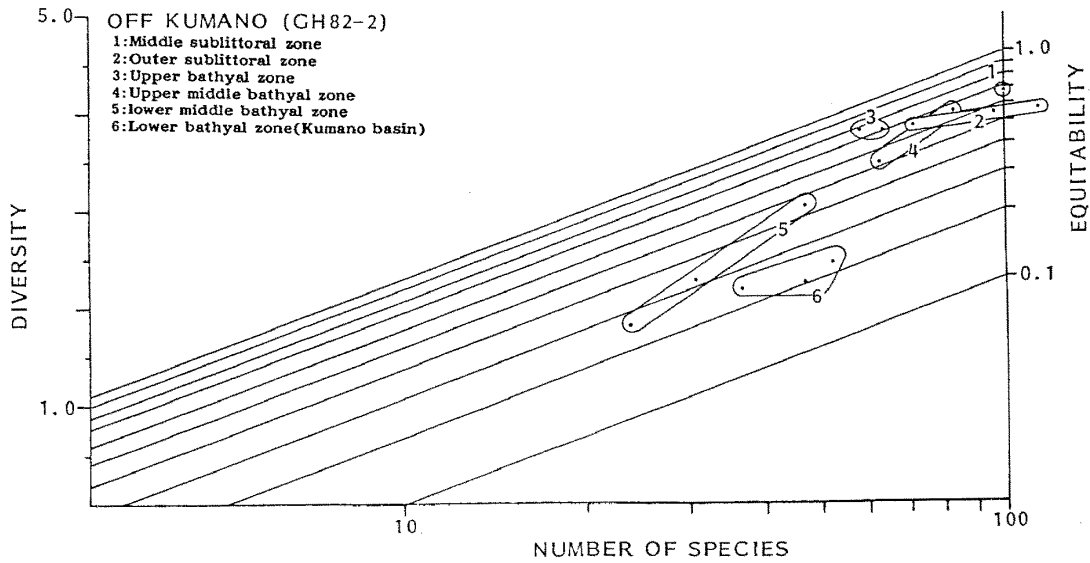


Fig. 35. Diagram showing species diversity, equitability, and number of species of samples in the Kumanonada area.

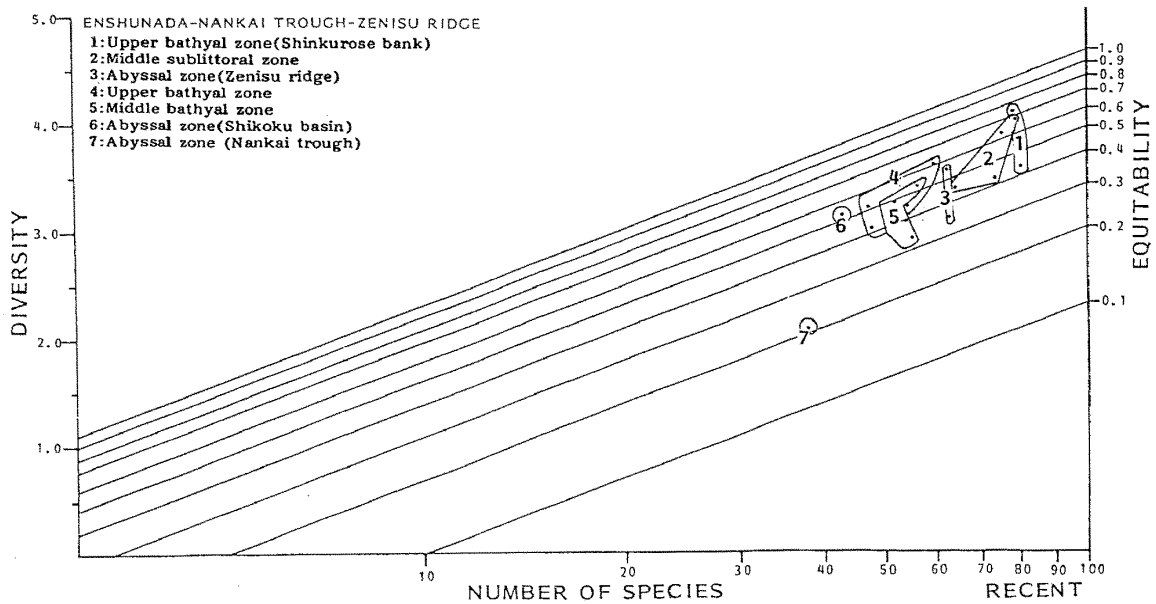


Fig. 36. Diagram showing species diversity, equitability, and number of species of samples in the Enshunada area.

number of species. Points belonging to group 5 are located in the middle bathyal zone, and exhibit the same values as in group 4. Two groups (group 2 in the middle sublittoral zone off Enshunada and group 3 in the abyssal zone on the

Zenusu Ridge) take intermediate numbers of species between group 1 and groups 4-5, with moderate other parameters. Points in the abyssal zone are grouped under two clusters. Group 6 on the bottom floor of the Shikoku Basin is

represented by moderate values of diversity, equitability and number of species. A point of group 7, which represents the Nankai Trough, shows a lower value of equitability than that in the abyssal zone.

In a summary, these figures (Fig. 34-36) lead to the following lines: 1) Points in the Nankai Trough and Kumano Basin are characterized by a rather low value of equitability (about

0.2). 2) The bathyal zone sample is marked by moderate values of diversity, equitability and number of species. 3) The three parameters of points in the sublittoral zone vary from locality to locality. Especially, those values in the Tanegashima area are extraordinarily low in comparison with other areas. This may be attributed to the influence of the Kuroshio Current.

FAUNAL REFERENCE LIST

Benthic foraminiferal species from the Enshunada, Kumanonada, Tanegashima and Hachijojima areas are alphabetically listed below. Selected species are illustrated with micrographs taken with the scanning electron microscope (Plates 11 to 24). The original references are given for each of the species.

- Adercotryma glomeratum* (Brady)
.....Pl. 11, fig. 1
- Lituola glomerata* Brady, 1878, p. 433, pl. 20, fig. 1. (*vide* Ellis and Messina, 1940 *et seq.*)
- Adercotryma glomeratum* (Brady).
Loeblich and Tappan, 1952, p. 141, figs. 1-4. (*vide* Loeblich and Tappan, 1953).
- Alabamina tubulifera* (Heron-Allen and Earland) ..Pl. 15, fig. 17; Pl. 24, fig. 2
- Truncatulina tubulifera* Heron-Allen and Earland, 1915, p. 710, pl. 52, figs. 37-40.
- Alabamina tubulifera* (Heron-Allen and Earland). Hofker, 1951, p. 392, figs. 271-273.
- Allomorphina pacifica* Hofker
Allomorphina pacifica Hofker, 1951, p. 139, fig. 86.
- Alveolophragmium bradyi* (Cushman)
Cribrostomoides bradyi Cushman, 1910, p. 108, text-fig. 167.
- Alveolophragmium bradyi* (Cushman).
Barker, 1960, p. 70, pl. 34, fig. 9.
- Alveolophragmium kosterense* (Höglund)
Labrospira kosterensis Höglund, 1947,
p. 139, 147, pl. 11, fig. 4, text-figs. 130, 131.
- Alveolophragmium kosterense* (Höglund).
Graham and Militante, 1959, p. 24, pl. 1, fig. 11.
- Alveolophragmium ringens* (Brady)
.....Pl. 11, fig. 9
- Trochammina ringens* Brady, 1879, p. 57, pl. 5, fig. 12.
- Alveolophragmium ringens* (Brady).
Parker, 1954, p. 487.
- Alveolophragmium scitulum* (Brady)
Haplophragmium scitulum Brady,
1881, p. 20.
- Alveolophragmium scitulum* (Brady).
Parker, 1954, p. 487.
- Alveolophragmium subglobosum* (G.O. Sars)
Pl. 11, fig. 3; Pl. 13, fig. 10; Pl. 14, fig. 2
- Haplophragmium subglobosa* G.O. Sars, 1868, p. 250. (*vide* Ellis and Messina, 1940 *et seq.*)
- Alveolophragmium subglobosa* (G.O. Sars).
Barker, 1960, p. 70, pl. 34, figs. 7, 8, 10 (not 14).
- Alveolophragmium wiesneri* (Parr)
Labrospira wiesneri Parr, 1950, p. 272.
(*vide* Ellis and Messina, 1940 *et seq.*)
- Alveolophragmium wiesneri* (Parr).
Barker, 1960, p. 82, pl. 40, figs. 14, 15.
- Ammobaculites agglutinans* (d'Orbigny)
.....Pl. 12, fig. 4
- Spirolina agglutinans* d'Orbigny, 1846,

- p. 137, pl. 7, figs. 10-12.
Ammobaculites agglutinans filiformis
 EarlandPl. 12, fig. 3
Ammobaculites agglutinans filiformis
 Earland, 1934, p. 92, figs. 11-13.
Ammobaculites americanus Cushman
Ammobaculites americanus Cushman,
 1910, p. 117, text-figs. 184, 185.
Ammodiscus minutissimus Cushman and
 McCulloch
Ammodiscus minutissimus Cushman
 and McCulloch, 1939, p. 70, pl. 5,
 figs. 3, 4.
Ammodiscus pacifica Cushman and
 ValentinePl. 13, fig. 4
Ammodiscus pacifica Cushman and
 Valentine, 1930, p. 7, pl. 1, fig. 1.
Ammolagena clavata (Parker and Jones)
Trochammina irregularis (d'Orbigny)
 var. *clavata* Parker and Jones,
 1860, p. 304. (*vide* Ellis and Messina,
 1940 *et seq.*).
Ammolagena clavata (Parker and
 Jones). Cushman, 1910, p. 68,
 text-figs. 86-89.
Ammonia beccarii (Linné)
Nautilus beccarii Linné, 1758, Syst.
 Nat., ed. 10, p. 710 (*vide* Ellis and
 Messina, 1940 *et seq.*).
Ammonia beccarii (Linné). Frizzel
 and Keen, 1949, p. 106.
Ammonia inflata (Seguenza)
Rosalina inflata Seguenza, 1862, p.
 106, pl. 1, fig. 6.
Ammonia japonica (Hada)
Rotalia japonica Hada, 1931, p. 137,
 fig. 93.
Ammonia japonica (Hada). Ujiie,
 1963, p. 236, pl. 2, figs. 3, 4.
Ammonia ketienziensis (Ishizaki)
Pl. 21, fig. 1
Streblus ketienziensis Ishizaki, 1948, p.
 59, pl. 1, fig. 2.
Ammonia ketienziensis (Ishizaki).
 Huang, 1964, p. 53, pl. 1, fig. 13.
Ammonia okinawaensis (LeRoy)
Rotalidium okinawaensis LeRoy, 1964,
 p. 38, pl. 4, figs. 29-31.
Ammonia okinawaensis (LeRoy).
 Loeblich and Tappan, 1964, p. 607.
Ammonia takanabensis (Ishizaki)
Pl. 15, fig. 15
Streblus takanabensis Ishizaki, 1948, p.
 57, pl. 1, fig. 5.
Ammonia takanabensis (Ishizaki).
 Huang, 1964, p. 56, pl. 1, fig. 2.
Ammoscalaris pseudospiralis (William-
 son)
Proteonina pseudospiralis Williamson,
 1858, p. 1.
Ammoscalaris pseudospiralis (William-
 son). Parker, Phleger and Peirson,
 1953, p. 6, pl. 1, figs. 29, 35.
Amphicoryna pauciloculata (Cushman)
Nodosaria pauciloculata Cushman,
 1921, p. 205, pl. 36, figs. 10-12.
Amphicoryna pauciloculata (Cush-
 man). Hasegawa, 1979, p. 142.
Amphicoryna proxima (Silvestri)
Nodosaria proxima Silvestri, 1872, p.
 63, pl. 6, figs. 138-147. (*vide* Ellis
 and Messina, 1940 *et seq.*).
Amphicoryna proxima (Silvestri).
 (*vide* Barker, 1960).
Amphicoryna scalaris (Batsch)
Pl. 17, fig. 7
Nautilus scalaris Batsch, 1791, pl. 2,
 fig. 4. (*vide* Ellis and Messina, 1940
et seq.).
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Amphicoryna scalaris sagamiensis
 (Asano)
Lagenodosaria scalaris sagamiensis
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 (Asano). Matoba, 1967, p. 251, pl.
 25, fig. 4.
Amphistegina madagascariensis d'Or-
 bigny
Amphistegina madagascariensis
 d'Orbigny, 1826, p. 304.
Amphistegina radiata (Fichtel and Moll)
Nautilus radiatus Fichtel and Moll,
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 and Messina, 1940 *et seq.*).
Amphistegina radiata (Fichtel and

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- Amphistegina wanneriana* Fischer
Amphistegina wanneriana Fischer, 1927, p. 167, pl. 217 (6), fig. 131. (fide Ellis and Messina, 1940 *et seq.*)
- Anomalinoidea cavus* Belford
Anomalinoidea cavus Belford, 1966, p. 180, pl. 33, figs. 1-8.
- Anomalinoidea glabrata* (Cushman)
Anomalina glabrata Cushman, 1924, p. 39, pl. 12, figs. 5-7.
- Anomalinoidea glabulosa* (Chapman and Parr) Pl. 20, fig. 6
Anomalina glabulosa Chapman and Parr, 1937, p. 117.
- Aschemonella scabra* Brady
Aschemonella scabra Brady, 1879, p. 44, pl. 3, figs. 6, 7.
- Astaculus crepidulus* (Fichtel and Moll)
Nautilus crepidulus Fichtel and Moll, 1798, p. 107, pl. 19, figs. g, i. (fide Ellis and Messina, 1940 *et seq.*)
Astaculus crepidulus (Fichtel and Moll). Galloway and Wissler, 1927, p. 45.
- Astrononion italicum* Cushman and Edward
Astrononion italicum Cushman and Edwards, 1937, p. 35, pl. 3, figs. 19, 20.
- Astrononion stellatum* Cushman and Edwards Pl. 15, fig. 9
Astrononion stellatum Cushman and Edwards, 1937, p. 32, pl. 3, figs. 9-11.
- Astrononion stelligerum* (d'Orbigny) Pl. 15, fig. 8
Nonionina stelligera d'Orbigny, 1839b, p. 128, pl. 3, fig. 7.
Astrononion stelligerum (d'Orbigny). Cushman and Edwards, 1937, p. 31, pl. 3, fig. 7.
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cf. *Rotalia trispinosa* Thalman, 1933, p. 248, figs. 2, 3. (fide Ellis and Messina, 1940 *et seq.*)
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Pulvinulina philippinensis Cushman, 1921, p. 331, pl. 58, fig. 2.
Baggina philippinensis (Cushman). Cushman and Todd, 1944, p. 104, pl. 17, figs. 1-3.
- Baggina philippinensis pilulifera* Cushman and Todd
Baggina philippinensis var. *pilulifera* Cushman and Todd, 1944a, p. 105, pl. 17, fig. 4.
- Bathysiphon filiformis* M. Sars
Bathysiphon filiformis M. Sars, 1871, p. 251. (fide Ellis and Messina, 1940 *et seq.*)
- Bathysiphon rufus* de Folin
Bathysiphon rufum de Folin, 1887, p. 283, pl. 6, fig. 8. (fide Ellis and Messina, 1940 *et seq.*)
- Biloculinella globula* (Bornemann)
Biloculina globulus Bornemann, 1855, p. 349, pl. 19, fig. 3. (fide Ellis and Messina, 1940 *et seq.*)
Biloculinella globula (Bornemann). Asano, 1951a, p. 20, figs. 134, 135.
- Biloculinella* cf. *japonica* Asano
cf. *Biloculinella japonica* Asano, 1956, p. 79, pl. 9, fig. 24.
- Bolivina canvallaria* Millett
Bolivina canvallaria Millett, 1900, p. 544, pl. 4, fig. 6.
- Bolivina digitalis* (d'Orbigny)
Polymorphina digitalis d'Orbigny, 1846, p. 235, pl. 14, figs. 1-4.
Bolivina digitalis (d'Orbigny). Papp and Schmid, 1985, p. 83, pl. 76, figs. 1-6.
- Bolivina earlandi* Parr
Bolivina earlandi Parr, 1950, p. 339, pl. 12, fig. 16. (fide Barker, 1960).
- Bolivina mayori* Cushman
Bolivina mayori Cushman, 1922a, p. 27, pl. 3, figs. 5, 6.
- Bolivina pisciformis* Galloway and Morrey
Bolivina pisciformis Galloway and Morrey, 1929, p. 36, pl. 5, fig. 10.
- Bolivina robusta* Brady

-Pl. 16, fig. 1; Pl. 22, fig. 13
Bolivina robusta Brady, 1881, p. 27.
- Bolivina semicostata* Cushman
Bolivina semicostata Cushman, 1911,
 p. 43, text-fig. 70.
- Bolivina sinuosum* Cushman
Loxostoma sinuosum Cushman, 1936b,
 p. 60, pl. 8, fig. 16.
- Bolivina spinescens* Cushman
Bolivina spinescens Cushman, 1911, p.
 46, text-fig. 76.
- Bolivina subspinescens* Cushman
Bolivina subspinescens Cushman,
 1922b, p. 48, pl. 7, figs. 5.
- Bolivina subtenuis* Cushman
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- Bolivinita quadrilatera* (Schwager)
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Bolivinita quadrilatera (Schwager).
 Cushman, 1927a, p. 90.
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 p. 297, pl. 15, fig. 1. (*fide* Ellis and
 Messina, 1940 *et seq.*).
- Brizalina amygdalaeformis* (Brady)
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 1884, p. 426, pl. 53, figs. 28, 29.
- Brizalina bradyi* (Asano)..Pl. 15, fig. 11
Bolivina bradyi Asano, 1938b, p. 603,
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Brizalina bradyi (Asano). Ta-
 kayanagi and Hasegawa, 1987, p. 7.
- Brizalina karreriana* (Brady)
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Bolivina karreriana Brady, 1881, p. 28.
Brizalina karreriana (Brady). Bel-
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- Brizalina kiiensis* (Asano)
Bolivina kiiensis Asano, 1958, p. 19, pl.
 4, figs. 7, 8.
Brizalina kiiensis (Asano). Ta-
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- Brizalina pacifica* (Cushman and McCul-
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 1942, p. 185, pl. 21, figs. 2, 3.
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 McCulloch). Hasegawa, 1979, p.
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Bolivina plicatella Cushman, 1930, p.
 46, pl. 8, fig. 10. (*fide* Ellis and Mes-
 sina, 1940 *et seq.*).
- Brizalina plicatella* (Cushman). Bel-
 ford, 1966, p. 26, pl. 1, figs. 8, 9,
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- Brizalina pygmaea* (Brady)
Bolivina pygmaea Brady, 1881, p. 59.
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 1966, p. 31, pl. 1, figs. 19-22.
- Brizalina seminuda* (Cushman)
Bolivina seminuda Cushman, 1911, p.
 34, text-fig. 55.
Brizalina seminuda (Cushman).
 Hasegawa, 1979, p. 143.
- Brizalina subangularis ogasaensis*
 (Asano)
Bolivina subangularis ogasaensis
 Asano, 1936c, p. 142, pl. 18, figs. 17-
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Brizalina subangularis ogasaensis
 (Asano). Takayanagi and Haseg-
 awa, 1987, p. 8.
- Brizalina subreticulata* (Parr)
Bolivina subreticulata Parr, 1932, p.
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Brizalina subreticulata (Parr). Bel-
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- Brizalina viscistriata* Belford
Brizalina viscistriata Belford, 1966, p.
 34, pl. 2, figs. 11-12.
- Buccella makiyamai* Chiji
Buccella makiyamae Chiji, 1961, p.
 234, text-fig. 2, pl. 1, figs. 13, 14.
- Bueningia crecki* Finlay
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Bulimina pseudotorta Cushman,
 1926a, p. 55, pl. 7, fig. 3.
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 Moll, 1798, p. 108, pl. 20, figs. a-c.
 (fide Ellis and Messina, 1940 *et seq.*).
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 Moll, 1798, p. 110, pl. 20, figs. d-f.
 (fide Ellis and Messina, 1940 *et seq.*).
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Rotalia spiculotesta Carter, 1877, p.
 470. (fide Ellis and Messina, 1940 *et seq.*).
Carterina spiculotesta (Carter). Loeblich
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 Barker, 1960).
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Cassidulina ? perumbonata Keyzer,
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 Ellis and Messina, 1940 *et seq.*).
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- 156, pl. 8, figs. 13-15.
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- Cibicides lobatulus* (Warker and Jacob)
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 (sic) Cushman, 1931, p. 118, pl. 21,
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- Cibicides macneili* LeRoy
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- Cibicides margaritiferus* (Brady)
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- Cibicides praecinctus* (Karrer)
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 158, p. 189, pl. 5, fig. 7. (*fide* Ellis
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 Barker, 1960, p. 196, pl. 95, figs. 1-3.
- Cibicides refulgens* Montfort
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- Cibicides* cf. *refulgens* Montfort
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- Cibicides robertsonianus* (Brady)
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- Cibicides subdepressa* (Asano)
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- Cibicides tenuimargo* (Brady)
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- Cibicides wuellerstorfi* (Schwager)
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 Messina, 1940 *et seq.*).
Cribrorbulina serpens (Seguenza).
 Selli, 1941, p. 83, pl. 1, figs. 1, 2, 5-
 10, 12-14; p. 86, text-figs. 1, 2.
- Cruciloculina japonica* Asano
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- Cyclammia bradyi* Cushman
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 113, text-fig. 174.
- Cyclammia cancellata* Brady
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- Cyclogyra crassisepta* (Brady)
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 714. (*fide* Ellis and Messina, 1940 *et*
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- Cyclogyra involvens* (Reuss)
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- Cyclogyra planorbis* (Schlutze)
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Cyclogyra planorbis (Schlutze).
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- Cymbaloporetta bradyi* (Cushman)
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- Cymbaloporetta bradyi* (Cushman).
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- Cystammia galeata* (Brady)
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- Cystammia pauciloculata* (Brady)
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- Dentalina communis* d'Orbigny
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- Dentalina filiformis* (d'Orbigny)
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- Dentalina intorta bradyensis* (Dervieux)
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 Messina, 1940 *et seq.*).
- Dentalina intorta* var. *bradyensis* (Der-
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- Dentalina* cf. *setanaensis* Asano
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- Dentalina mucronata* Neugeboren
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- Discanomalina coronata* (Parker and
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 Loeblich and Tappan, 1964, p. 575.
- Discorbinella baconica* (Hantken)
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- Discorbinella convexa* (Takayanagi)
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- Valentine, 1930, p. 31, pl. 10, figs. 1, 2.
- Dyocibicides uniserialis* Thalmann
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- Eggerella bradyi* (Cushman)
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- Eggerella pusilla* (Goës). Cushman and McCulloch, 1939, p. 96, pl. 10, figs. 2, 3.
- Eggerella scabra* (Williamson)
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- Ehrenbergina bosoensis* Takayanagi
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- Ehrenbergina bradyi* Cushman
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- Ehrenbergina hystrix* Brady
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- Ehrenbergina pacifica* Cushman
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- Ehrenbergina trigona* Goës
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Polystomella umbilicatula var. *incerta* Williamson, 1858, p. 44, pl. 3, fig. 82.
Elphidium incertum (Williamson). Cushman, 1930, p. 18, pl. 7, figs. 4-9.
- Elphidium jenseni* (Cushman)
Polystomella jenseni Cushman, 1924, p. 49, pl. 16, figs. 4, 6.
Elphidium jenseni (Cushman). Cushman, 1933b, p. 48, pl. 11, figs. 6, 7.
- Elphidium poeyanum* (d'Orbigny)
Polystomella poeyana d'Orbigny, 1839a, p. 55, pl. 6, figs. 25, 26.
Elphidium poeyanum (d'Orbigny). Cushman, 1930, p. 25, pl. 10, figs. 4, 5.
- Elphidium simplex* Cushman
Elphidium simplex Cushman, 1933b, p. 52, pl. 12, figs. 8, 9.
- Elphidium translucens* Natland
Elphidium translucens Natland, 1938, p. 144, pl. 5, figs. 3, 4. (*vide* Ellis and Messina, 1940 *et seq.*).
- Epistominella pulchra* (Cushman)
Pulvinulinella pulchra Cushman, 1933c, p. 92, pl. 9, fig. 10.
Epistominella pulchra (Cushman). Todd, 1965, p. 31, pl. 10, figs. 3, 4.
- Eponides tumidulus* (Brady)
Truncatulina tumidula Brady, 1884, p. 666, pl. 95, fig. 8.
Eponides tumidulus (Brady). Cushman, 1931, p. 55.
- Evolvocassidulina belfordi* Nomura
Evolvocassidulina belfordi Nomura, 1983a, p. 79, pl. 2, fig. 6; pl. 20, figs. 8-12.
- Evolvocassidulina brevis* (Aoki)
.....Pl. 17, fig. 5
" *Cassidulina* " *brevis* Aoki, 1968, p.

- 261, pl. 27, fig. 4.
Evolvocassidulina brevis (Aoki).
 Nomura, 1983b, p. 49, pl. 4, figs. 4-7;
 pl. 20, fig. 11; pl. 21, figs. 1-5.
- Evolvocassidulina kuwanoi* (Matoba)
Cassidulinoides kuwanoi Matoba, 1967,
 p. 253, pl. 29, figs. 1, 2.
- Evolvocassidulina kuwanoi* (Matoba).
 Nomura, 1983b, p. 46, pl. 4, fig. 1;
 pl. 20, figs. 4-7.
- Favocassidulina favus* (Brady)
Pulvinulina favus Brady, 1877, p. 535.
 (fide Ellis and Messina, 1940 *et seq.*).
Favocassidulina favus (Brady). Loeblich
 and Tappan, 1957, p. 230, pl. 73,
 figs. 7-11.
- Fissurina alveolata* (Brady)
Lagena alveolata Brady, 1884, p. 487,
 pl. 60, figs. 30, 32.
Fissurina alveolata (Brady). Parr,
 1950, p. 307. (fide Barker, 1960).
- Fissurina circulocosta* Asano
Fissurina circulocosta Asano, 1938c, p.
 219, pl. 30, fig. 17.
- Fissurina clathrata* (Brady)
Lagena clathrata, Brady, 1884, p. 485,
 pl. 60, fig. 4.
Fissurina clathrata (Brady). Parr,
 1950, p. 310. (fide Barker, 1960).
- Fissurina crebra* (Matthes)
Lagena crebra Matthes, 1939, p. 72, pl.
 5, figs. 66-70.
Fissurina crebra (Matthes). Barker,
 1960, p. 122, pl. 59, fig. 6.
- Fissurina cucullata* Silvestri
Fissurina cucullata Silvestri, 1902, p.
 146, text-figs. 23-25. (fide Ellis and
 Messina, 1940 *et seq.*).
- Fissurina formosa* (Schwager)
Lagena formosa Schwager, 1866, p.
 206, pl. 4, fig. 19.
Fissurina formosa (Schwager).
 Srinivasan and Sharma, 1980, p. 41,
 pl. 3, figs. 15, 16.
- Fissurina kerguelenensis* Parr
Fissurina kerguelenensis Parr, 1950, p.
 305, pl. 8, fig. 7. (fide Barker, 1960).
- Fissurina lacunata* (Burrows and Hol-
 land)
- Lagena lacunata* Burrows and Hol-
 land, 1895, p. 205, pl. 7, fig. 12. (fide
 Ellis and Messina, 1940 *et seq.*).
- Fissurina lacunata* (Burrows and Hol-
 land). Parr, 1945, p. 203.
- Fissurina laevigata* Reuss
Fissurina laevigata Reuss, 1850, p. 366,
 pl. 46, fig. 1. (fide Ellis and Messina,
 1940 *et seq.*).
- Fissurina lucida* (Williamson)
Entosolenia marginata (Montagu) var.
lucida Williamson, 1848, p. 17, pl. 2,
 fig. 7.
Fissurina lucida (Williamson). Loeblich
 and Tappan, 1953, p. 76, pl. 14,
 fig. 4.
- Fissurina marginata* (Montagu)
Vermiculum marginatum Montagu,
 1803, p. 524. (fide Ellis and Messina,
 1940 *et seq.*).
- Fissurina marginata* (Montagu).
 Murray, 1971, p. 97, pl. 39, fig. 4-6.
- Fissurina orbignyana* Seguenza
Fissurina orbignyana Seguenza, 1862,
 p. 66, pl. 2, figs. 24, 26.
- Fissurina quadrata* (Williamson)
Entosolenia quadrata Williamson,
 1870, p. 28, pl. 10, fig. 2. (fide Ellis
 and Messina, 1940 *et seq.*).
- Fissurina quadrata* (Williamson).
 Barker, 1960, p. 122, pl. 59, fig. 3.
- Fissurina quadrata* (Williamson) var.
rizzae Seguenza
Fissurina quadrata (Williamson) var.
rizzae Seguenza, 1862, p. 72, pl. 2,
 fig. 46.
- Fissurina radiata* Seguenza
Fissurina radiata Seguenza, 1862, p.
 70, pl. 2, figs. 2, 3.
- Fissurina sequenziana* (Fornasini)
Lagena sequenziana Fornasini, 1887, p.
 350. (fide Ellis and Messina, 1940 *et
 seq.*).
- Fissurina sequenziana* (Fornasini).
 Earland, 1934, p. 160.
- Fissurina semimarginata* (Reuss)
Lagena marginata Williamson var.
semimarginata Reuss, 1870, p. 468.
 (fide Ellis and Messina, 1940 *et seq.*).

- Fissurina semimarginata* (Reuss).
Loeblich and Tappan, 1953, p. 78,
pl. 14, fig. 3.
- Frondicularia kiensis* Barker
Frondicularia kiensis Barker, 1960, p.
138, pl. 65, fig. 18.
- Gaudryina arenaria* Galloway and Wiss-
ler
Gaudryina arenaria Galloway and
Wissler, 1927, p. 68, pl. 11, fig. 5.
- Gaudryina atlantica* (Bailey)
Textuiaria atlantica Bailey, 1851, p.
12, figs. 38-43.
- Gaudryina atlantica* (Bailey). Cush-
man, 1922b, p. 70, pl. 13, figs. 1-3.
- Gaudryina atlantica* (Bailey) var.
pacifica Cushman and McCulloch
Gaudryina atlantica (Bailey) var.
pacifica Cushman and McCulloch,
1939, p. 94, pl. 9, figs. 1, 2.
- Gaudryina flintii* Cushman
Gaudryina flintii Cushman, 1911, p.
63, text-figs. 102.
- Gaudryina matusimai* Asano
Gaudryina matusimai Asano, 1937, p.
1234, text-figs. 1-3.
- Gaudryina robusta* Cushman
Gaudryina robusta Cushman, 1913b, p.
636, pl. 78, figs. 2.
- Gaudryina siphonifera* (Brady)
Textuiaria siphonifera Brady, 1881, p.
53.
- Gaudryina siphonifera* (Brady). Cush-
man, 1928, p. 109, pl. 16, figs. 1-5.
- Gaudryina subglabrata* Cushman and
McCulloch
Gaudryina subglabrata Cushman and
McCulloch, 1939, p. 92, pl. 8, figs. 5-
7.
- Glabratella australensis* (Heron-Allen
and Earland)
Discorbis australensis Heron-Allen and
Earland, 1932, p. 416.
- Glabratella australensis* (Heron-Allen
and Earland). Loeblich and
Tappan, 1964, p. 588.
- Glabratella nakamurai* (Asano)
Discorbis nakamurai Asano, 1951d, p.
2, figs. 8-12.
- Glabratella nakamurai* (Asano). Ta-
kayanagi and Hasegawa, 1987, p. 14.
- Glabratella opercularis* (d'Orbigny)
Rosalina opercularis d'Orbigny, 1839a,
p. 93, pl. 3, figs. 24, 25.
- Glabratella opercularis* (d'Orbigny).
Matoba, 1970, p. 54, pl. 5, fig. 4.
- Glabratella ozawai* (Asano)
Discorbis ozawai Asano, 1951d, p. 3,
figs. 14-16.
- Glabratella ozawai* (Asano). Ta-
kayanagi and Hasegawa, 1987, p. 14.
- Glabratella patelliformis* (Brady)
Discorbina patelliformis Brady, 1884,
p. 647, pl. 88, fig. 3; pl. 89, fig. 1.
- Glabratella patelliformis* (Brady).
Loeblich and Tappan, 1964, p. 588.
- Glabratella subopercularis* (Asano)
Discorbis subopercularis Asano, 1951d,
p. 3, figs. 14-16.
- Glabratella subopercularis* (Asano).
Matoba, 1970, p. 54, pl. 5, fig. 5.
- Glabratella tabernacularis* (Brady)
Discorbis tabernacularis Brady, 1881,
p. 65.
- Glandulina laevigata* (d'Orbigny)
Nodosaria laevigata d'Orbigny, 1826,
p. 252, pl. 10, figs. 1-3.
- Glandulina nipponica* Asano
Glandulina nipponica Asano, 1951e, p.
14, figs. 71, 72.
- Globobulimina auriculata* (Bailey)
Bulimina auriculata Bailey, 1851, p.
12, figs. 25-27.
- Globobulimina auriculata* (Bailey).
Asano, 1958, p. 9, pl. 2, figs. 1-3.
- Globobulimina hanzawai* Asano
Globobulimina hanzawai Asano, 1958,
p. 10, pl. 2, figs. 4-6.
- Globobulimina pacifica* Cushman
..... Pl. 16, fig. 9
- Globobulimina pacifica* Cushman,
1927b, p. 67, pl. 14, fig. 12.
- Globocassidulina bisecta* Nomura
Globocassidulina bisecta Nomura,
1983a, p. 73, pl. 2, figs. 2, 3; pl. 2,
fig. 3; pl. 14, figs. 8-12; pl. 15, figs.
1-5.
- Globocassidulina brocha* (Poag)

-Pl. 18, fig. 7
Cassidulina brocha Poag, 1966, p. 426, pl. 8, figs. 33-35.
Globocassidulina brocha (Poag). Nomura, 1983a, pl. 18, figs. 8-12; 1983b, p. 31, pl. 3, figs. 3-5.
- Globocassidulina crassa* (d'Orbigny)
Cassidulina crassa d'Orbigny, 1839a, p. 56, pl. 7, figs. 18-20.
Globocassidulina crassa (d'Orbigny). Nomura, 1983b, p. 37, pl. 3, figs. 9, 10.
- Globocassidulina decorata* (Sidebottom)
Cassidulina decorata Sidebottom, 1910, p. 107, pl. 4, fig. 2. (*vide* Ellis and Messina, 1940 *et seq.*).
Globocassidulina decorata (Sidebottom). Nomura, 1983b, p. 27, pl. 2, figs. 14-16.
- Globocassidulina depressa* (Asano and Nakamura)
Cassidulina subglobosa Brady *depressa* Asano and Nakamura, 1937, p. 148, pl. 13, fig. 8.
Globocassidulina depressa (Asano and Nakamura). Nomura, 1983a, pl. 15, figs. 11, 12, pl. 6, figs. 1-4; 1983b, p. 35, pl. 3, fig. 6.
- Globocassidulina elegans* (Sidebottom)
.....Pl. 18, fig. 5
Cassidulina elegans Sidebottom, 1910, p. 106, pl. 4, fig. 1. (*vide* Ellis and Messina, 1940 *et seq.*).
- Globocassidulina gemma* (Todd)
.....Pl. 18, fig. 3
Cassidulina gemma Todd, in Cushman, Todd and Post, 1954, p. 366, pl. 90, figs. 26, 27.
Globocassidulina gemma (Todd). Belford, 1966, p. 147, pl. 24, figs. 22-25, text-fig. 16, nos. 11, 12.
- Globocassidulina moluccensis* (Germeraad)
.....Pl. 18, fig. 6
Cassidulina moluccensis Germeraad, 1946, p. 72, pl. 2, figs. 29-32. (*vide* Ellis and Messina, 1940 *et seq.*).
- Globocassidulina moluccensis* (Germeraad). Nomura, 1983b, p. 19, pl. 2, fig. 6, 7; pl. 11, fig. 12; pl. 12, figs. 1, 2.
- Globocassidulina mucronata* Nomura
Globocassidulina mucronata Nomura, 1983a, p. 63, pl. 1, fig. 15; pl. 12, fig. 13.
- Globocassidulina neobrocha* Nomura
Globocassidulina neobrocha Nomura, 1983a, p. 67, pl. 1, fig. 16; pl. 19, figs. 3-6.
- Globocassidulina okinawaensis* (LeRoy)
Cassidulina okinawaensis LeRoy, 1964, p. F40, pl. 11, figs. 21, 22.
Globocassidulina okinawaensis (LeRoy). Nomura, 1983b, p. 29, pl. 2, fig. 20; pl. 18, fig. 2.
- Globocassidulina oriangulata* (Belford)
.....Pl. 18, fig. 2
Cassidulina oriangulata Belford, 1966, p. 148, pl. 25, figs. 1-5, text-figs. 16, nos. 13, 14.
Globocassidulina oriangulata (Belford). Nomura, 1983b, p. 43, pl. 3, figs. 16, 17; pl. 6, fig. 16; pl. 16, figs. 11, 12; pl. 17, figs. 1, 2.
- Globocassidulina paratortuosa* (Kuwano)
.....Pl. 18, fig. 8
Cassidulina paratortuosa Kuwano, 1954, p. 34, figs. 4-6.
Globocassidulina paratortuosa (Kuwano). Nomura, 1983b, p. 28, pl. 2, fig. 19.
- Globocassidulina parva* (Asano and Nakamura)
.....Pl. 18, fig. 4
Cassidulina subglobosa Brady *parva* Asano and Nakamura, 1937, p. 148, pl. 13, fig. 5.
Globocassidulina parva (Asano and Nakamura). Nomura, 1983a, pl. 15, figs. 6-10; 1983b, p. 41, pl. 3, fig. 13.
- Globocassidulina parviapertura* Nomura
.....Pl. 18, fig. 1
Globocassidulina parviapertura Nomura, 1983a, p. 56, pl. 1, fig. 6; pl. 12, figs. 5-8.
- Globocassidulina pseudoquadrata* Nomura
Globocassidulina pseudoquadrata Nomura, 1983a, p. 62, pl. 1, fig. 14; pl. 12, figs. 5, 6.
- Globocassidulina subglobosa* (Brady)

- Cassidulina subglobosa* Brady, 1881, p. 30.
- Globocassidulina subglobosa* (Brady). Belford, 1966, p. 149, pl. 25, figs. 11-16.
- Globocassidulina subparva* Nomura
Globocassidulina subparva Nomura, 1983a, p. 64, pl. 1, fig. 11.
- Globocassidulina tsuchidai* Nomura
Globocassidulina tsuchidai Nomura, 1983a, p. 70, pl. 2, fig. 4; pl. 17, figs. 3-7.
- Globocassidulina venustas* Nomura
Globocassidulina venustas Nomura, 1983a, p. 60, pl. 1, figs. 7, 8; pl. 14, figs. 4-7.
- Glomospira charoides* (Jones and Parker)Pl. 11, fig. 7; Pl. 13, fig. 6
- Trochammina squamata charoides* Jones and Parker, 1860, p. 304. (*fide* Ellis and Messina, 1940 *et seq.*).
- Glomospira charoides* (Jones and Parker). Cushman, 1925c, p. 25, pl. 2, fig. 12.
- Glomospira gordialis* (Jones and Parker)Pl. 11, fig. 8; Pl. 13, fig. 5
- Ammodiscus gordialis* Jones and Parker, 1880, pl. 5, fig. 22. (*fide* Ellis and Messina, 1940 *et seq.*).
- Glomospira gordialis* (Jones and Parker). Cushman and Jarvis, 1928, p. 87, pl. 12, figs. 7, 8.
- Gyroidina altiformis* R.E. and K.C. Stewart
Gyroidina soldanii var. *altiformis* R.E. and K.C. Stewart, 1930, p. 67, pl. 9, fig. 2.
- Gyroidina* cf. *broeckhiana* (Karrer)
 cf. *Rotalia broeckhiana* Karrer, 1878, p. 98, pl. 5, fig. 26. (*fide* Ellis and Messina, 1940 *et seq.*).
- cf. *Gyroidina broeckhiana* (Karrer). Hofker, 1951, p. 403, figs. 281, 282.
- Gyroidina cushmani* BoomgaardPl. 21, fig. 3
- Gyroidina cushmani* Boomgaard, 1949, p. 125, pl. 14, fig. 1.
- Gyroidina neosoldanii acuta* Boomgaard
Gyroidina neosoldanii acuta Boomgaard, 1949, p. 124, pl. 9, fig. 9.
- Gyroidina orbicularis* d'Orbigny
Gyroidina orbicularis d'Orbigny, 1826, p. 278, modèles, no. 13.
- Gyroidina soldanii* d'OrbignyPl. 24, fig. 5
- Gyroidina soldanii* d'Orbigny, 1826, p. 278, modèles, no. 36.
- Gyroidinoides nipponicus* (Ishizaki)Pl. 21, fig. 4
- Gyroidina nipponica* Ishizaki, 1944, p. 102, pl. 3, fig. 3. (*fide* Ellis and Messina, 1940 *et seq.*).
- Gyroidinoides nipponicus* (Ishizaki). Matoba, 1967, p. 255, pl. 29, fig. 13.
- Hanzawaia nipponica* AsanoPl. 20, fig. 4
- Hanzawaia nipponica* Asano, 1944, p. 99, pl. 4, fig. 1, 2.
- Haplophragmoides columbiensis* CushmanPl. 11, fig. 2; Pl. 13, fig. 8
- Haplophragmoides columbiensis* Cushman, 1925a, p. 39, pl. 6, fig. 2.
- Haplophragmoides columbiensis evolutum* Cushman and McCulloch
Haplophragmoides columbiensis evolutum Cushman and McCulloch, 1939, p. 73, pl. 5, figs. 11, 12; pl. 6, figs. 1, 2.
- Heronallenia lingulata* (Burrows and Holland)
Discorbina lingulata Burrows and Holland, 1896, p. 297, pl. 7, fig. 33. (*fide* Ellis and Messina, 1940 *et seq.*).
- Heronallenia lingulata* (Burrows and Holland). Chapman and Parr, 1931, p. 236, pl. 9, fig. 6.
- Heterolepa haidingerii* (d'Orbigny)
Rotalia haidingerii d'Orbigny, 1846, p. 154, pl. 8, figs. 7-9.
- Heterolepa subhaidingerii* (Parr)Pl. 23, fig. 3
- Cibicides subhaidingerii* Parr, 1950, p. 364. (*fide* Ellis and Messina, 1940 *et seq.*).
- Heterolepa subpraecincta* (Asano)
Eponides subpraecinctus Asano, 1951d, p. 12, figs. 88-90.
- Heterolepa subpraecincta* (Asano).

- Takayanagi and Hasegawa, 1987, p. 19.
- Hoeglundina elegans* (d'Orbigny)
.....Pl. 21, fig. 7; Pl. 24, fig. 6
- Rotalia elegans* d'Orbigny, 1826, p. 276.
- Hoeglundina elegans* (d'Orbigny).
Brotzen, 1948, p. 92.
- Hormosina globulifera* Brady
Hormosina globulifera Brady, 1879, p. 60, pl. 4, figs. 4, 5.
- Hyalinea balthica* (Schröter)
.....Pl. 20, fig. 5
- Nautilus balthicus* Schröter, 1783, p. 20, pl. 1, fig. 2. (*vide* Ellis and Messina, 1940 *et seq.*).
- Hyalinea balthica* (Schröter). Hofker, 1951, p. 508, figs. 346-348.
- Hyperammia friabilis* Brady
Hyperammia friabilis Brady, 1884, p. 258, pl. 23, figs. 1-3, 5, 6.
- Hyperammia laevigata* Wright
Hyperammia elongata Brady var. *laevigata* Wright, 1891, p. 466, pl. 20, fig. 1. (*vide* Ellis and Messina, 1940 *et seq.*).
- Hyperammia laevigata* Wright. Cushman, 1918, p. 77, pl. 29, figs. 5, 6.
- Islandiella kazusaensis* (Asano and Nakamura)
Cassidulina kazusaensis Asano and Nakamura, 1937, p. 146, pl. 14, fig. 2.
- Islandiella kazusaensis* (Asano and Nakamura). Nomura, 1983b, p. 6, pl. 1, fig. 5; pl. 11, figs. 10, 11.
- Jaculella acuta* Brady
Jaculella acuta Brady, 1879, p. 35, pl. 3, figs. 12-13.
- Karreriella apicularis* (Cushman)
.....Pl. 12, fig. 2
- Gaudryina apicularis* Cushman, 1911, p. 69, text-fig. 110.
- Karreriella* (*Karrerulina*) *apicularis* (Cushman). Barker, 1960, p. 94, pl. 46, figs. 17-19.
- Karreriella baccata* (Schwager)
Gaudryina baccata Schwager, 1866, p. 200, pl. 4, fig. 12.
- Karreriella baccata* (Schwager). Cushman, 1937a, p. 133, pl. 15, figs. 20-24.
- Karreriella bradyi* (Cushman)
Gaudryina bradyi Cushman, 1911, p. 67, fig. 107.
- Karreriella bradyi* (Cushman). Barker, 1960, p. 94, pl. 46, fig. 1-4.
- Karreriella subrotundata* (Schwager)
Gaudryina subrotundata Schwager, 1866, p. 198, pl. 4, fig. 9.
- Karreriella subrotundata* (Schwager). Srinivasan and Sharma, 1980, p. 17, pl. 2, figs. 11, 18.
- Lagena acuticosta* Reuss
Lagena acuticosta Reuss, 1861, p. 305, pl. 1, fig. 4. (*vide* Ellis and Messina, 1940 *et seq.*).
- Lagena distoma* Parker and Jones
Lagena distoma Parker and Jones, 1865, p. 467, pl. 48, fig. 6.
- Lagena elongata* (Ehrenberg)
Miliola elongata Ehrenberg, 1844, p. 274. (*vide* Ellis and Messina, 1940 *et seq.*).
- Lagena elongata* (Ehrenberg). Brady, 1884, p. 456, pl. 56, fig. 29.
- Lagena filicosta* Reuss
Lagena filicosta Reuss, 1863, p. 328, pl. 4, figs. 50, 51. (*vide* Ellis and Messina, 1940 *et seq.*).
- Lagena flexa* Cushman and Gray
Lagena flexa Cushman and Gray, 1946, p. 68, pl. 12, figs. 18-21.
- Lagena gracilis* Williamson
Lagena gracilis Williamson, 1848, p. 13, pl. 1, fig. 5.
- Lagena gracillima* (Seguenza)
Amphorina gracillima Seguenza, 1862, p. 51, pl. 1, fig. 37.
- Lagena gracillima* (Seguenza). Brady, 1884, p. 456, pl. 56, figs. 19-26, (not 27, 28).
- Lagena hispida* Reuss
Lagena hispida Reuss, 1858, p. 434. (*vide* Ellis and Messina, 1940 *et seq.*).
- Lagena laevis* (Montagu) ..Pl. 19, fig. 17
Vermiculum laevis Montagu, 1803, p. 524. (*vide* Ellis and Messina, 1940 *et seq.*).

- Lagena laevis* (Montagu). Cushman, 1913a, p. 5, pl. 1, fig. 3; pl. 38, fig. 5.
- Lagena parri* Loeblich and Tappan
Lagena parri Loeblich and Tappan, 1953, p. 64, pl. 11, figs. 11-13.
- Lagena perlucida* (Montagu)
Vermiculum perlucidum Montagu, 1803, p. 525, pl. 14, fig. 3. (*fide* Ellis and Messina, 1940 *et seq.*).
- Lagena perlucida* (Montagu). Brown, 1827, pl. 1, fig. 29. (*fide* Cushman and McCulloch, 1950).
- Lagena semistriata* Williamson
Lagena vulgaris var. *semistriata* Williamson, 1858, p. 6, pl. 1, fig. 9.
- Lagena setigera* Millett
Lagena clavata (d'Orbigny) var. *setigera* Millett, 1901, p. 491, pl. 8, fig. 9.
- Lagena setigera* Millett. Loeblich and Tappan, 1953, p. 66, pl. 11, figs. 23, 24.
- Lagena striata* (d'Orbigny)
Oolina striata d'Orbigny, 1839b, p. 21, pl. 5, fig. 12.
- Lagena striata* (d'Orbigny). Asano, 1938c, p. 217, pl. 27, fig. 26; pl. 29, fig. 28.
- Lagena striata toddi* LeRoy
Lagena striata (d'Orbigny) var. *toddii* LeRoy, 1964, p. 25, pl. 13, figs. 39.
- Lagena substriata* Williamson
Lagena substriata Williamson, 1848, p. 15, pl. 2, fig. 12.
- Lagena sulcata laevicostata* Cushman and Gray
Lagena sulcata (Walker and Jacob) var. *laevicostata* Cushman and Gray, 1946, p. 68, pl. 12, figs. 13, 14.
- Lagena sulcata peculiaris* Cushman and McCulloch
Lagena sulcata (Walker and Jacob) var. *peculiaris* Cushman and McCulloch, 1950, p. 361, pl. 48, figs. 11-13.
- Lagena sulcata spicata* Cushman and McCulloch
Lagena sulcata var. *spicata* Cushman and McCulloch, 1950, p. 360, pl. 48, figs. 3-7.
- Lamarckina scabra* (Brady)
Pulvinulina oblonga (Williamson) var. *scabra* Brady, 1884, p. 689, pl. 106, fig. 8.
- Lamarckina scabra* (Brady). Cushman, 1931, p. 35, pl. 7, fig. 6.
- Lamarckina ventricosa* (Brady)
Discorbis ventricosa Brady, 1884, p. 654, pl. 91, fig. 7.
- Lamarckina ventricosa* (Brady). Cushman, 1926b, p. 12.
- Lenticulina gibba* (d'Orbigny)
Cristellaria gibba d'Orbigny, 1826, p. 292, no. 17.
- Lenticulina gibba* (d'Orbigny). Thalmann, 1932, p. 305.
- Lenticulina kamakuraensis* Asano
Lenticulina kamakuraensis Asano, 1936b, p. 612, pl. 31, fig. 1.
- Lernella inflata* (LeRoy)
Cassidulina inflata LeRoy, 1944a, p. 37, pl. 4, figs. 30, 31.
- Lernella inflata* (LeRoy). Nomura, 1983b, p. 86, pl. 2, fig. 9.
- Lernella ogasawarai* Nomura
Lernella ogasawarai Nomura, 1983a, p. 88, pl. 2, fig. 8.
- Lernella seranensis* (Germeraad)
Cassidulina seranensis Germeraad, 1946, p. 72, pl. 5, figs. 5, 6. (*fide* Ellis and Messina, 1940 *et seq.*).
- Lernella seranensis* (Germeraad). Nomura, 1983a, p. 24, figs. 6, 7; 1983b, p. 56, pl. 4, figs. 15-17.
- Lituotuba lituiformis* (Brady)
Trochammia lituiformis Brady, 1879, p. 59, pl. 5, fig. 16.
- Lituotuba lituiformis* (Brady). Rhumbler, 1895, p. 83. (*fide* Barker, 1960).
- Marginulina glabra* d'Orbigny
Marginulina glabra d'Orbigny, 1826, p. 259, modèles, no. 55.
- Marginulina tenuis* Bornemann
Marginulina tenuis Bornemann, 1855, p. 326, pl. 13, figs. 14. (*fide* Ellis and Messina, 1940 *et seq.*).
- Marsipella cylindrica* Brady
Marsipella cylindrica Brady, 1882, p.

714.
Marsipella elongata Norman. . Pl. 12, fig. 9
Marsipella elongata Norman, 1878, p. 281, pl. 16, fig. 7. (*vide* Ellis and Messina, 1940 *et seq.*)
Martinottiella communis (d'Orbigny)
Clavulina communis d'Orbigny, 1826, p. 268.
Martinottiella communis (d'Orbigny). Cushman, 1933a, p. 37, pl. 4, figs. 6-8.
Melonis barleeanus (Williamson)
Nonionina barleeanus Williamson, 1858, p. 32, pl. 3, figs. 68, 69.
Melonis barleeanum (Williamson). Woodruff, 1985, p. 167.
Melonis pacificus (Cushman)
Nonionina umbilicatula (Montagu) var. *pacifica* Cushman, 1924, p. 48, pl. 16, fig. 3.
Melonis pacificus (Cushman). Hasegawa, 1979, p. 151.
Melonis parkerae (Uchio)
.....Pl. 17, fig. 2; Pl. 18, fig. 9
Nonion parkerae Uchio, 1960, p. 60, pl. 4, figs. 9, 10.
Melonis parkerae (Uchio). Matoba, 1967, p. 256, pl. 29, fig. 15.
Melonis sphaeroides Voloshinova
Melonis sphaeroides Voloshinova, 1958, p. 153, pl. 3, figs. 8, 9.
Miliolinella australis (Parr)
.....Pl. 15, fig. 14; Pl. 22, fig. 5
Quinqueloculina australis Parr, 1932, p. 7, pl. 1, fig. 8.
Miliolinella (?) *australis* (Parr). Barker, 1960, p. 10, pl. 5, figs. 10, 11.
Miliolinella circularis (Bornemann)
Pl. 15, fig. 12; Pl. 17, fig. 9; Pl. 22, fig. 6
Triloculina circularis Bornemann, 1855, p. 349, pl. 19, fig. 4. (*vide* Ellis and Messina, 1940 *et seq.*)
Miliolinella circularis (Bornemann). Asano, 1951a, p. 9, figs. 65-67.
Miliolinella inflata LeRoy
.....Pl. 15, fig. 13; Pl. 22, fig. 4
Miliolinella inflata LeRoy, 1964, p. 21, pl. 12, figs. 36, 37.
Miliolinella oblonga (Montagu)
Vermiculum oblonga Montagu, 1803, p. 522, pl. 14, figs. 9. (*vide* Ellis and Messina, 1940 *et seq.*)
Miliolinella oblonga (Montagu). Cushman, 1933b, p. 15, fig. 6.
Miliolinella subrotunda (Montagu)
Vermiculum subrotunda Montagu, 1803, p. 521. (*vide* Ellis and Messina, 1940 *et seq.*)
Miliolinella subrotunda (Montagu). Wiesner, 1931, p. 107. (*vide* Barker, 1960).
Mississippina concentrica (Parker and Jones)Pl. 24, fig. 9
Pulvinulina concentrica Parker and Jones, 1865, p. 393. (*vide* Ellis and Messina, 1940 *et seq.*)
Mississippina concentrica (Parker and Jones). Uchio, 1952a, p. 197, pl. 18, figs. 3-5.
Neoconorbina pacifica LeRoy
Neoconorbina pacifica LeRoy, 1964, p. 36, pl. 4, figs. 26-28.
Neoconorbina terquemi (Rzehak)
Discorbina terquemi Rzehak, 1888, p. 228. (*vide* Ellis and Messina, 1940 *et seq.*)
Neoconorbina terquemi (Rzehak). Hofker, 1951, p. 433.
Neoconorbina tuberculata (Chapman)
Discorbina tuberculata Chapman, 1900, p. 11, pl. 1, fig. 9.
Neoconorbina tuberculata (Chapman). Todd, 1965, p. 17, pl. 1, figs. 8, 9.
Neoconorbina procerus (Brady)
Pulvinulina procerus Brady, 1884, p. 698, pl. 105, fig. 7.
Neoconorbina procerus (Brady). Reiss, 1960, p. 17.
Nodellum membranaceum (Brady)
.....Pl. 12, fig. 8; Pl. 13, fig. 1
Reophax membranaceum Brady, 1879, p. 53, pl. 4, fig. 9.
Nodellum membranaceum (Brady). Rhumbler, 1913, p. 474. (*vide* Barker, 1960).

- Nodosaria aculeata* d'Orbigny
Nodosaria aculeata d'Orbigny, 1846, p. 35, pl. 1, figs. 26, 27.
- Nodosaria milletti* Cushman
Nodosaria milletti Cushman, 1917, p. 654.
- Nodosaria subraphana* Asano
Nodosaria subraphana Asano, 1938c, p. 212, pl. 29, fig. 19; pl. 30, figs. 33, 35.
- Nonion granosum* (d'Orbigny)
Nonionina granosum d'Orbigny, 1826, p. 294, no. 8.
Nonion granosum (d'Orbigny). Cushman, 1939, p. 11, pl. 2, figs. 17, 18.
- Nonion manpukuziensis* Otuka
Nonion manpukuziensis Otuka, 1932, p. 654, fig. 5.
- Nonion japonicum* Asano . . Pl. 14, fig. 6
Nonion japonicum Asano, 1938a, p. 593, pl. 15, figs. 1, 2.
- Nonion pauperatum* (Balkwell and Wright)
Nonionina pauperata Balkwell and Wright, 1885, p. 353, pl. 13, figs. 25, 26. (*fide* Ellis and Messina, 1940 *et seq.*).
Nonion pauperatum (Balkwell and Wright). Cushman, 1930, p. 13, pl. 5, figs. 4, 5, 7.
- Nonionella amplilabrata* Belford
Nonionella amplilabrata Belford, 1966, p. 159, pl. 31, figs. 16-20.
- Nonionella basiloba* Cushman and McCulloch
Nonionella basiloba Cushman and McCulloch, 1940, p. 162, pl. 18, fig. 3.
- Nonionella bradii* (Chapman)
Nonionina scapha var. *bradii* Chapman, 1917, p. 71, pl. 5, fig. 42.
Nonionella bradii (Chapman). Barker, 1960, p. 224, pl. 109, fig. 16.
- Nonionella miocenica* Cushman
Nonionella miocenica Cushman, 1926a, p. 64.
- Nonionellina labradorica* (Dawson)
. Pl. 14, fig. 13
Nonionina labradorica Dawson, 1860, p. 191, text-fig. 4. (*fide* Ellis and Messina, 1940 *et seq.*).
- Nonionellina labradorica* (Dawson). Cushman, 1939, p. 23, pl. 6, figs. 13-16.
- Nuttallides umbonifera* (Cushman)
. Pl. 14, fig. 8
Pulvinulinella umbonifera Cushman, 1933c, p. 90, pl. 9, fig. 9.
Nuttallides umbonifera (Cushman). Todd, 1965, p. 29, pl. 11, fig. 1.
- Oolina costata* (Williamson)
Entosolenia costata Williamson, 1858, p. 9, pl. 1, fig. 18.
Oolina costata (Williamson). Loeblich and Tappan, 1953, p. 68, pl. 13, figs. 4-6.
- Oolina globosa* (Montagu)
Vermiculum globosum Montagu, 1803, p. 523 (*fide* Ellis and Messina, 1940 *et seq.*).
Oolina globosa (Montagu). Parr, 1950, p. 302. (*fide* Barker, 1960).
- Oolina hexagona* (Williamson)
Entosolenia sequamosa (Montagu) var. *hexagona* Williamson, 1858, p. 13, pl. 1, fig. 32.
Oolina hexagona (Williamson). Parr, 1950, p. 304. (*fide* Barker, 1960).
- Oolina melo* d'Orbigny
Oolina melo d'Orbigny, 1839a, p. 20, pl. 5, fig. 9.
- Oolina ovum* (Ehrenberg)
Miliola ovum Ehrenberg, 1843, p. 166. (*fide* Ellis and Messina, 1940 *et seq.*).
Oolina ovum (Ehrenberg). Barker, 1960, p. 114, pl. 56, fig. 5.
- Operculina ammonoides* (Gronovius)
. Pl. 15, fig. 11
Nautilus ammonoides Gronovius, 1781, p. 282, pl. 19, figs. 5, 6. (*fide* Ellis and Messina, 1940 *et seq.*).
- Operculina ammonoides* (Gronovius). Carpenter, Parker and Jones, 1862, p. 310. (*fide* Barker, 1960).
- Operculina venosa* (Fichtel and Moll)
Nautilus venosa Fichtel and Moll, 1798, p. 59, pl. 8, figs. e-h. (*fide* Ellis and Messina, 1940 *et seq.*).
- Operculina venosa* (Fichtel and Moll).

- Cushman, 1921, p. 39, pl. 14, fig. 6.
Ophthalmidium acutimargo (Brady)
Spiroloculina acutimargo Brady, 1884,
 p. 154, pl. 10, figs. 12-15.
Ophthalmidium acutimargo (Brady).
 Loeblich and Tappan, 1964, p. 448,
 fig. 340, 2.
Oridorsalis pauciapertura Belford
Oridorsalis pauciapertura Belford,
 1966, p. 173, pl. 30, figs. 7-13, text-
 fig. 22, no. 3.
Oridorsalis tener (Brady) ..Pl. 21, fig. 5
Truncatulina tenera Brady, 1884, p.
 665, pl. 95, fig. 11.
Oridorsalis tener (Brady). Pflum and
 Frerichs, 1976, p. 106, pl. 6, figs. 2-4.
Oridorsalis umbonatus (Reuss)
Pl. 24, fig. 4
Rotalia umbonata Reuss, 1851, p. 75,
 pl. 5, fig. 35.
Oridorsalis umbonatus (Reuss). Par-
 ker, 1964, p. 626, pl. 99, figs. 4-6.
Osangularia culter (Parker and Jones)
Planorbulina culter Parker and Jones,
 1865, p. 421, pl. 19, fig. 1.
Osangularia culter (Parker and Jones).
 Todd, 1965, p. 25, pl. 15, fig. 1.
Osangularia rugosa (Phleger and Parker)
Pseudoparrella (?) *rugosa* Phleger and
 Parker, 1951, p. 28, pl. 15, figs. 8, 9.
Osangularia rugosa (Phleger and Par-
 ker). Pflum and Frerichs, 1976, p.
 122, pl. 7, figs. 2-4.
Paracassidulina miuraensis (Higuchi)
Pl. 15, fig. 4
Cassidulina miuraensis Higuchi, 1956,
 p. 58, text-figs. 58, 59.
Paracassidulina miuraensis (Higuchi).
 Nomura, 1983a, pl. 5, fig. 3; pl. 25,
 figs. 4-6; 1983b, p. 70, pl. 6, figs. 7-
 10.
Paracassidulina neocarinata (Thalmann)
Pl. 17, fig. 4
Cassidulina neocarianta Thalmann,
 1950, p. 44.
Paracassidulina neocarinata (Thal-
 mann). Nomura, 1983b, p. 63, pl. 5,
 fig. 11.
Paracassidulina nipponensis (Eade)
Pl. 19, fig. 2
Globocassidulina nipponensis Eade,
 1967, p. 65, pl. 13, figs. 1, 2, 4.
Paracassidulina nipponensis (Eade).
 Nomura, 1983a, p. 95, pl. 25, fig. 3;
 1983b, pl. 6, figs. 13, 14.
Paracassidulina oshimai (Aoki)
Cassidulina oshimai Aoki, 1967, p.
 381, pl. 1, figs. 10-15.
Paracassidulina oshimai (Aoki).
 Nomura, 1983b, p. 71, pl. 6, figs. 11,
 12.
Paracassidulina quasiearinata Nomura
Pl. 19, fig. 5
Paracassidulina quasiearinata Nomura,
 1983a, p. 100, pl. 2, fig. 19; pl. 25,
 figs. 9-11.
Paracassidulina sulcata (Belford)
Pl. 19, fig. 1
Cassidulina sulcata Belford, 1966, p.
 142, 144, pl. 24, figs. 11-14, text-fig.
 16, nos. 7, 8.
Paracassidulina sulcata (Belford).
 Nomura, 1983b, p. 69, pl. 6, figs. 4-6.
Paracassidulina tomiyensis (Uchio)
Pl. 19, fig. 3
Cassidulina tomiyensis Uchio, 1951, p.
 40, pl. 3, fig. 3.
Paracassidulina tomiyensis (Uchio).
 Nomura, 1983b, pl. 5, figs. 12, 13.
Parafissurina lateralis Cushman
Parafissurina lateralis Cushman,
 1913a, p. 9, pl. 1, fig. 1.
Pararotalia calcar (d'Orbigny)
Pl. 15, fig. 16
Calcarina calcar d'Orbigny, 1826, p.
 276, no. 1.
Pararotalia nipponica (Asano)
Rotalia nipponica Asano, 1936b, p.
 614, pl. 30, fig. 2.
Pararotalia nipponica (Asano). Ujiie,
 1966, p. 192, text-figs. 1-3; pl. 24,
 figs. 1-7; pl. 25, figs. 1-5.
Parrelloides bradyi (Trauth)
Pl. 20, fig. 2; Pl. 23, fig. 8
Truncatulina bradyi Trauth, 1918, p.
 235.
Parrelloides bradyi (Trauth).
 Belford, 1966, p. 100, pl. 11, figs. 10-

19.
Patellina corrugata Williamson
Patellina corrugata Williamson, 1858, p. 46, pl. 3, figs. 86-89.
Patellinella hanzawai Asano
Patellinella hanzawai Asano, 1936b, p. 613, pl. 30, fig. 3.
Patellinella jugosa (Brady)
Textularia jugosa Brady, 1884, p. 358, pl. 42, fig. 7.
Patellinella jugosa (Brady). Cushman, Todd and Post, 1954, p. 358, pl. 89, fig. 3.
Pelosina cylindrica Brady
Pelosina cylindrica Brady, 1884, p. 236, pl. 26, figs. 1-6.
Peneropsis pertusus (Forsk.)
Nautilus pertusus Forskal, 1775, p. 125, no. 65. (*vide* Ellis and Messina, 1940 *et seq.*).
Peneropsis pertusus (Forsk.). Cushman, 1921, p. 481.
Planispirinella exigua (Brady)
Hauerina exigua Brady, 1879, p. 267.
Planispirinella exigua (Brady). Wiesner, 1931, p. 69. (*vide* Barker, 1960).
Planularia australis Chapman
Planularia australis Chapman, 1915, p. 24. (*vide* Ellis and Messina, 1940 *et seq.*).
Planularia planulata (Galloway and Wissler)
Astacolus planulata Galloway and Wissler, 1927, p. 46, pl. 8, fig. 5.
Planularia planulata (Galloway and Wissler). Cushman and McCulloch, 1950, p. 303, pl. 40, figs. 1-5.
Planulina ariminensis d'Orbigny
Planulina ariminensis d'Orbigny, 1826, p. 280, pl. 5, figs. 1-3.
Planulina tricarlinella (Reuss)
Cristellaria tricarlinella Reuss, 1862, p. 68, pl. 7, fig. 9; pl. 12, figs. 2-4. (*vide* Ellis and Messina, 1940 *et seq.*).
Planulina tricarlinella (Reuss). Asano, 1938c, p. 207, pl. 30, figs. 2, 3.
Poroepionides cribrorepiandus Asano and Uchio Pl. 24, fig. 1
Poroepionides cribrorepiandus Asano and Uchio, in Asano, 1951d, p. 18, figs. 134, 135.
Praeglobobulimina ovata (d'Orbigny)
Bulimina ovata d'Orbigny, 1846, p. 185, pl. 11, figs. 13, 14.
Praeglobobulimina ovata (d'Orbigny). Haymes, 1954, p. 190, text-figs. 9-12, 17-19, pl. 35, figs. 2, 3.
Praeglobobulimina pupoides (d'Orbigny)
Bulimina pupoides d'Orbigny, 1846, p. 185, pl. 11, figs. 11, 12.
Praeglobobulimina pupoides (d'Orbigny). Loeblich and Tappan, 1964, p. 561, figs. 442, 14, 15.
Praeglobobulimina pyrula (d'Orbigny)
Bulimina pyrula d'Orbigny, 1846, p. 184, pl. 9, figs. 9, 10.
Praeglobobulimina spinescens (Brady)
Bulimina pyrula d'Orbigny var. *spinescens* Brady, 1884, p. 400, pl. 50, figs. 11, 12.
Praeglobobulimina spinescens (Brady). Loeblich and Tappan, 1964, p. 561, figs. 442, 12, 13.
Psammospaera fusca Schulze
Psammospaera fusca Schulze, 1875, p. 113, pl. 2, fig. 8. (*vide* Ellis and Messina, 1940 *et seq.*).
Psammospaera parva Flint. . Pl. 12, fig. 5
Psammospaera parva Flint, 1897, p. 268, pl. 9, fig. 1.
Pseudoepionides nakazatoensis (Kuwano)
Epistomaria (*Epistominella*) *nakazatoensis* Kuwano, 1950, p. 316, figs. 4, 11.
Pseudoepionides nakazatoensis (Kuwano). Asano, 1951d, p. 19, figs. 141-143.
Pseudononion auricula (Heron-Allen and Earland)
Nonionella auricula Heron-Allen and Earland, 1930, p. 192, pl. 5, figs. 68-70. (*vide* Ellis and Messina, 1940 *et seq.*).
Pseudononion decora (Cushman and McCulloch)
Nonionella decora Cushman and

- McCulloch, 1940, p. 160, pl. 17, figs. 11, 12.
- Pseudononion grateloupi* (d'Orbigny)
Nonionina grateloupi d'Orbigny, 1826, p. 294.
- Pseudononion grateloupi* (d'Orbigny). Hasegawa, 1979, p. 152.
- Pseudononion japonicum* Asano
Pseudononion japonicum Asano, 1936a, p. 347, text-figs. a-c.
- Pseudoparrella exigua* (Brady)
Pl. 20, fig. 7; Pl. 24, fig. 3
- Pulvinulina exigua* Brady, 1884, p. 696, pl. 103, figs. 13, 14.
- Pseudoparrella exigua* (Brady). Phleger and Parker, 1951, p. 28, pl. 15, figs. 6, 7.
- Pseudorotalia gaimardii* (d'Orbigny)
Pl. 21, fig. 2
- Rotalia gaimardii* d'Orbigny, 1826, p. 275, no. 46.
- Pseudorotalia gaimardii* (d'Orbigny). Matoba, 1967, p. 257, pl. 27, fig. 4.
- Pullenia bulloides* (d'Orbigny)
Pl. 17, figs. 11, 12; Pl. 22, fig. 15
- Nonionina bulloides* d'Orbigny, 1846, p. 107, pl. 5, figs. 9, 10.
- Pullenia bulloides* (d'Orbigny). Parker and Jones, 1862, 106 p. 184. (*vide* Loeblich and Tappan, 1964).
- Pullenia elegans* Cushman and Todd
Pullenia elegans Cushman and Todd, 1943, p. 23, pl. 4, fig. 11.
- Pullenia quinqueloba* (Reuss)
Pl. 17, fig. 13
- Nonionina quinqueloba* Reuss, 1851, p. 71, pl. 5, fig. 31. (*vide* Ellis and Messina, 1940 *et seq.*).
- Pullenia quinqueloba* (Reuss). Cushman and Todd, 1943, p. 11, pl. 2, fig. 8.
- Pullenia salisburyi* R.E. and R.C. StewartPl. 17, fig. 14
- Pullenia salisburyi* R.E. and R.C. Stewart, 1930, p. 72, pl. 8, fig. 2.
- Pyrgo depressa* (d'Orbigny)
Biloculina depressa d'Orbigny, 1826, p. 298, no. 7, modèles, no. 91.
- Pyrgo depressa* (d'Orbigny). Cushman, 1929, p. 71, pl. 19, figs. 4, 5.
- Pyrgo elongata* (d'Orbigny)
Biloculina elongata d'Orbigny, 1826, p. 298, no. 4.
- Pyrgo elongata* (d'Orbigny). Cushman, 1929, p. 70, pl. 19, figs. 2, 3.
- Pyrgo fornasinii* Chapman and Parr
Pyrgo fornasinii Chapman and Parr, 1935, p. 5.
- Pyrgo murrhina* (Schwager)
Biloculina murrhina Schwager, 1866, p. 203, pl. 4, fig. 15.
- Pyrgo murrhina* (Schwager). Cushman, 1929, p. 71, pl. 19, fig. 6, 7.
- Pyrgo pacifica* Asano
Pyrgo pacifica Asano, 1956, p. 78, pl. 9, fig. 3.
- Pyrgo serrata* (Bailey)
Biloculina serrata Bailey, 1862, p. 350, pl. 8, fig. E. (*vide* Ellis and Messina, 1940 *et seq.*).
- Pyrgo serrata* (Bailey). Cushman, 1929, p. 73.
- Pyrgo vespertilio* (Schlumberger)
Biloculina vespertilio Schlumberger, 1891, p. 174, pl. 10, figs. 74-76.
- Pyrgo vespertilio* (Schlumberger). Thalmann, 1932, p. 295.
- Pyrulina angusta* (Egger)
Polymorphina (Globulina) angusta Egger, 1857, p. 290, pl. 13, figs. 13-15.
- Pyrulina angusta* (Egger). Barker, 1960, p. 150, pl. 72, figs. 1-3.
- Pyrulina gutta* d'Orbigny
Pyrulina gutta d'Orbigny, 1826, p. 267, no. 28, modèles, no. 30.
- Quadrिमorphina laevigata* (Phleger and Parker)Pl. 24, fig. 7
- Valvulineria laevigata* Phleger and Parker, 1951, p. 25, pl. 13, figs. 11, 12.
- Quadrिमorphina laevigata* (Phleger and Parker). Belford, 1966, p. 155, pl. 37, figs. 21-25.
- Quinqueloculina akneriana* d'Orbigny
Quinqueloculina akneriana d'Orbigny, 1846, p. 290, pl. 18, figs. 16-20.
- Quinqueloculina bicostata* d'Orbigny

- Quinqueloculina bicostata* d'Orbigny, 1839a, p. 195, pl. 12, figs. 8-10.
- Quinqueloculina boueana* d'Orbigny
- Quinqueloculina boueana* d'Orbigny, 1846, p. 293, pl. 19, figs. 7-9.
- Quinqueloculina bradyana* Cushman
- Quinqueloculina bradyana* Cushman, 1917, p. 52, pl. 18, fig. 2.
- Quinqueloculina contorta* d'Orbigny
- Quinqueloculina contorta* d'Orbigny, 1846, p. 298, pl. 20, figs. 4-6.
- Quinqueloculina costata* d'Orbigny
- Quinqueloculina costata* d'Orbigny, 1826, p. 301, no. 3.
- Quinqueloculina ferussaci* d'Orbigny
- Quinqueloculina ferussaci* d'Orbigny, 1826, p. 301, no. 18.
- Quinqueloculina hashimotoi* Asano
- Quinqueloculina hashimotoi* Asano, 1938e, p. 92, pl. 10, fig. 5.
- Quinqueloculina lamarckiana* d'Orbigny
- Quinqueloculina lamarckiana* d'Orbigny, 1839a, p. 189, pl. 11, figs. 14, 15.
- Quinqueloculina monroei* Cushman and Todd
- Quinqueloculina monroei* Cushman and Todd, 1946, p. 79, pl. 14, figs. 1, 2.
- Quinqueloculina polygona* d'Orbigny
- Quinqueloculina polygona* d'Orbigny, 1839a, p. 198, pl. 12, figs. 21-23.
- Quinqueloculina reticulata* (d'Orbigny)
- Triloculina reticulata* d'Orbigny, 1826, p. 209.
- Quinqueloculina reticulata* (d'Orbigny). Cushman, 1917, p. 55, pl. 16, figs. 1-3.
- Quinqueloculina seminulum* (Linné)
- Serpula seminulum* Linné, 1767, p. 1267, No. 791. (fide Ellis and Messina, 1940 et seq.).
- Quinqueloculina seminula* (Linné). Cushman and Gray, 1946, p. 3.
- Quinqueloculina vulgaris* d'Orbigny
- Quinqueloculina vulgaris* d'Orbigny, 1826, p. 302.
- Quinqueloculina yabei* Asano
- Quinqueloculina yabei* Asano, 1936, p. 942, pl. 51, fig. 1.
- Rectobolivina bifrons* (Brady)
- Sagrina bifrons* Brady, 1881, p. 34.
- Rectobolivina bifrons* (Brady). Cushman, 1927b, p. 204, pl. 23, figs. 13, 14.
- Rectobolivina columellaris* (Brady)
- Sagrina columellaris* Brady, 1881, p. 64.
- Rectobolivina columellaris* (Brady). Barker, 1960, p. 156, pl. 75, figs. 15-17.
- Rectobolivina limbata* (Brady)
- Bolivina limbata* Brady, 1881, p. 27.
- Rectobolivina limbata* (Brady). Belford, 1966, p. 49, pl. 4, figs. 12-15.
- Rectobolivina limbata* (Brady) var. *costulata* (Cushman)
- Bolivina limbata* Brady var. *costulata* Cushman, 1922, p. 26, pl. 3, fig. 8.
- Rectobolivina raphanus* (Parker and Jones) Pl. 15, fig. 6; Pl. 16, fig. 4
- Uvigerina* (*Sagrina*) *raphanus* Parker and Jones, 1865, p. 364, pl. 18, figs. 16, 17.
- Rectobolivina raphana* (Parker and Jones). Loeblich and Tappan, 1964, p. C533, figs. 438 (9-11).
- Reophax bacillaris* Brady
- Reophax bacillaris* Brady, 1881, p. 49.
- Reophax dentaliniformis* Brady
- Reophax dentaliniformis* Brady, 1881, p. 49.
- Reophax difflugiformis* Brady
- Pl. 12, fig. 6
- Reophax difflugiformis* Brady, 1879, p. 51, pl. 4, fig. 3.
- Reophax distans* Brady
- Pl. 12, fig. 7; Pl. 13, fig. 2
- Reophax distans* Brady, 1881, p. 50.
- Reophax fusiformis* (Williamson)
- Proteonina fusiformis* Williamson, 1858, p. 1, pl. 1, fig. 1.
- Reophax fusiformis* (Williamson). Siddall, 1879, p. 4. (fide Brady, 1884).
- Reophax guttifer* Brady
- Reophax guttifer* Brady, 1881, p. 49.
- Reophax nodulosus* Brady

- Reophax nodulosus* Brady, 1849, p. 52, pl. 4, figs. 7, 8.
- Reophax pilulifer* Brady
- Reophax pilulifer* Brady, 1884, p. 292, pl. 30, figs. 18-20.
- Reophax scorpiurus* Montfort
- Reophax scorpiurus* Montfort, 1808, p. 331, fig. 83d. (*fide* Ellis and Messina, 1940 *et seq.*).
- Reophax spiculifera* Brady Pl. 13, fig. 3
- Reophax spiculifera* Brady, 1879, p. 54, pl. 4, figs. 10, 11.
- Reussella aculeata* Cushman
- Reussella aculeata* Cushman, 1945, p. 41, pl. 7, figs. 10, 11.
- Reussella aequa* Cushman and McCulloch
- Reussella aequa* Cushman and McCulloch, 1948, p. 251, pl. 31, fig. 7.
- Reussella insueta* Cushman
- Reussella insueta* Cushman, 1945, p. 40, pl. 7, fig. 9.
- Reussella pacifica* Cushman and McCulloch
- Reussella pacifica* Cushman and McCulloch, 1948, p. 251, pl. 31, fig. 6.
- Reussella simplex* (Cushman)
- Trimosina simplex* Cushman, 1929, p. 158, text-fig. 2.
- Reussella simplex* (Cushman). Cushman, 1945, p. 40, pl. 7, fig. 5.
- Reussella spinulosa* (Reuss) Pl. 15, fig. 1
- Verneuilina spinulosa* Reuss, 1850, p. 374, pl. 47, fig. 12. (*fide* Ellis and Messina, 1940 *et seq.*).
- Reussella spinulosa* (Reuss). Cushman, 1945, p. 33, pl. 6, figs. 8, 9.
- Rhabdammina abyssorum* M. Sars
-Pl. 12, fig. 10; Pl. 14, fig. 4
- Rhabdammina abyssorum* M. Sars, 1868, p. 248. (*fide* Ellis and Messina, 1940 *et seq.*).
- Rhizammima algaeformis* Brady
- Rhizammima algaeformis* Brady, 1879, p. 39, pl. 4, figs. 16, 17.
- Robulus abensis* Asano
- Robulus abensis* Asano, 1936c, p. 327, pl. 37, figs. 5, 10.
- Robulus calcar* (Linné)
-Pl. 15, fig. 10; Pl. 17, fig. 6
- Nautilus calcar* Linné, 1767, p. 1162. (*fide* Ellis and Messina, 1940 *et seq.*).
- Robulus calcar* (Linné). d'Orbigny, 1846, p. 99, pl. 4, figs. 18-20.
- Robulus chitanii* Yabe and Asano
- Robulus chitanii* Yabe and Asano, 1937, p. 100, pl. 18, fig. 14.
- Robulus cultratus* Montfort
- Robulus cultrata* Montfort, 1808, p. 215. (*fide* Ellis and Messina, 1940 *et seq.*).
- Robulus kotiensis* Asano
- Robulus kotiensis* Asano, 1938c, p. 203, pl. 29, fig. 35.
- Robulus lucida* (Cushman)
- Cristellaria lucida* Cushman, 1923, p. 111, pl. 30, fig. 2.
- Robulus lucida* (Cushman). Asano, 1951e, p. 5, figs. 21, 22.
- Robulus nikobarensis* (Schwager)
- Cristellaria nikobarensis* Schwager, 1866, p. 243, pl. 6, fig. 87.
- Robulus nikobarensis* (Schwager). Asano, 1938c, p. 204, pl. 28, figs. 5, 6; pl. 29, fig. 8.
- Robulus orbicularis* d'Orbigny
-Pl. 22, fig. 1
- Robulus orbicularis* d'Orbigny, 1826, p. 288, pl. 15, figs. 8, 9.
- Robulus obetusata* (Reuss)
- Cristellaria obetusata* Reuss, 1870, p. 479, No. 2. (*fide* Ellis and Messina, 1940 *et seq.*).
- Robulus obetusata* (Reuss). Thalmann, 1932, p. 304.
- Robulus pseudorotulatus* Asano
- Robulus pseudorotulatus* Asano, 1938c, p. 201, pl. 25, figs. 1, 3, 4; pl. 26, fig. 28; pl. 31, figs. 3, 4.
- Robulus sagamiensis* Asano
- Robulus sagamiensis* Asano, 1938c, p. 201, pl. 25, fig. 6; pl. 26, figs. 11-13, pl. 28, fig. 12; pl. 29, fig. 16.
- Robulus surugaensis* Asano
- Robulus surugaensis* Asano, 1936c, p. 328, pl. 37, fig. 4.
- Robulus tumidus* Asano
- Robulus tumidus* Asano, 1938c, p. 200,

- pl. 24, fig. 9.
- Robulus tangens* LeRoy
Robulus tangens LeRoy, 1939, p. 233, pl. 8, figs. 12, 13.
- Robulus vortex* (Fichtel and Moll)
Nautilus vortex Fichtel and Moll, 1798, p. 33, pl. 2, figs. d-e. (*fide* Ellis and Messina, 1940 *et seq.*).
Robulus vortex (Fichtel and Moll). Barker, 1960, p. 144, pl. 69, figs. 14-16.
- Rosalina australis* (Parr)
Discorbis australis Parr, 1932, p. 227.
Rosalina australis (Parr). Matoba, 1970, p. 60, pl. 4, fig. 7.
- Rosalina bradyi* (Cushman)
Discorbis globularis var. *bradyi* Cushman, 1915, p. 12, pl. 8, fig. 1.
Rosalina bradyi (Cushman). Matoba, 1970, p. 60, pl. 4, fig. 8.
- Rosalina floridana* (Cushman)
Discorbis floridana Cushman, 1922a, p. 39, pl. 5, figs. 11, 12.
Rosalina floridana (Cushman). Parker, 1954, p. 524, pl. 8, figs. 19, 20.
- Rosalina globularis* d'Orbigny
Rosalina globularis d'Orbigny, 1826, p. 217, pl. 13, figs. 1-4, modèles, no. 69.
- Rosalina micens* (Cushman)
Discorbis micens Cushman, 1933c, p. 89, pl. 9, fig. 5.
Rosalina micens (Cushman). Todd, 1965, p. 12, pl. 4, fig. 2.
- Rosalina ponticulus* Belford
Rosalina ponticulus Belford, 1966, p. 94, pl. 13, figs. 12-16.
- Rosalina vilardeboana* d'Orbigny
.....Pl. 22, fig. 16
Rosalina vilardeboana d'Orbigny, 1839a, p. 44, pl. 6, figs. 13-15.
- '*Rotalia*' *murrayi* Heron-Allen and Earland
Rotalia murrayi Heron-Allen and Earland, 1915, p. 720, pl. 53, figs. 27-34.
'*Rotalia*' *murrayi* Heron-Allen and Earland. Graham and Millitante, 1959, p. 100, pl. 15, fig. 5.
- Rutherfordoides mexicanus* (Cushman)
.....Pl. 19, fig. 9
- Virgulina mexicana* Cushman, 1922a, p. 120, pl. 23, fig. 8.
- Rutherfordoides mexicana* (Cushman). Kohl, 1985, p. 89, pl. 18, fig. 3.
- Rutherfordoides subtenuis* (Nomura)
Hastilina subtenuis Nomura, 1983a, p. 83, pl. 2, fig. 14; pl. 3, fig. 8; pl. 25, fig. 12.
Rutherfordoides subtenuis (Nomura). Takayanagi and Hasegawa, 1987, p. 26.
- Rutherfordoides virga* (Nomura)
Hastilina virga Nomura, 1983a, p. 84, pl. 2, fig. 15.
Rutherfordoides virga (Nomura). Takayanagi and Hasegawa, 1987, p. 26.
- Saccamina sphaerica* M. Sars
.....Pl. 12, fig. 1
Saccamina sphaerica M. Sars, 1872, p. 250 (*fide* Ellis and Messina, 1940 *et seq.*).
- Saccorhiza ramosa* Brady
.....Pl. 12, fig. 11; Pl. 14, fig. 1
Saccorhiza ramosa Brady, 1879, p. 33, pl. 3, figs. 14, 15.
- Saracenaria angularis* Natland
Saracenaria angularis Natland, 1938, p. 143, pl. 5, figs. 1, 2.
- Saracenaria latifrons* (Brady)
Cristellaria latifrons Brady, 1884, p. 544, pl. 68, fig. 19; pl. 113, fig. 11.
Saracenaria latifrons (Brady). Asano, 1951e, p. 13, figs. 67, 68.
- Schlumbergerina alveoliniformis* (Brady)
Miliolina alveoliniformis Brady, 1879, p. 268.
Schlumbergerina alveoliniformis (Brady). Cushman, 1929, p. 36.
- Seabrookia pellucida* Brady
Seabrookia pellucida Brady, 1890, p. 570. (*fide* Loeblich and Tappan, 1964).
- Sigmavirgulina atlantica* (Cushman)
Bolivina tortuosa Brady var. *atlantica* Cushman, 1936b, p. 57, pl. 8, fig. 8.
- Sigmavirgulina tortuosa* (Brady)
Bolivina tortuosa Brady, 1881, p. 57.
Sigmavirgulina tortuosa (Brady). Loeblich and Tappan, 1957, p. 227.

- text-fig. 30, pl. 73, figs. 1, 2.
- Sigmoidella kagaensis* Cushman and Ozawa
Sigmoidella kagaensis Cushman and Ozawa, 1928, pl. 2, fig. 14.
- Sigmoidella pacifica* Cushman and Ozawa
Sigmoidella pacifica Cushman and Ozawa, 1928, p. 19, pl. 2, fig. 13.
- Sigmoilopsis schlumbergeri* (Silvestri)
Pl. 17, fig. 8; Pl. 22, fig. 8
- Sigmoilina schlumbergeri* Silvestri, 1904, p. 267. (*fide* Ellis and Messina, 1940 *et seq.*).
- Sigmoilopsis schlumbergeri* (Silvestri). Finlay, 1947, p. 270.
- Sigmomorphina gallowayi* Cushman and Ozawa
Sigmomorphina gallowayi Cushman and Ozawa, 1930, p. 135, pl. 36, fig. 4.
- Sigmomorphina* cf. *ozawai* (Hada)
 cf. *Sigmomorpha ozawai* Hada, 1931, p. 115, text-figs. 73, 74.
 cf. *Sigmomorphina ozawai* (Hada). Asano, 1951b, p. 12, figs. 57, 58.
- Sigmomorphina semitecta* (Reuss)
Polymorphina semitecta Reuss, 1867, p. 91, pl. 3, fig. 10. (*fide* Ellis and Messina, 1940 *et seq.*).
- Sigmomorphina semitecta* (Reuss). Cushman and Ozawa, 1930, p. 129, pl. 33, figs. 6, 7.
- Sigmomorphina semitecta* (Reuss) var. *terquemiana* (Fornasini)
Polymorphina amygdaloides Reuss var. *terquemiana* Fornasini, 1902, p. 72, fig. 25. (*fide* Ellis and Messina, 1940 *et seq.*).
- Sigmomorphina semitecta* (Reuss) var. *terquemiana* (Fornasini). Cushman and Ozawa, 1930, p. 129, pl. 33, figs. 4, 5; pl. 34, figs. 2, 3; pl. 35, fig. 1.
- Siphonina tubulosa* Cushman
Siphonina tubulosa Cushman, 1924, p. 40, pl. 13, figs. 1, 2.
- Siphotextularia concava* (Karrer)
Plecanium concavum Karrer, 1868, p. 129, pl. 1, fig. 3. (*fide* Ellis and Messina, 1940 *et seq.*).
- Siphotextularia concava* (Karrer). Barker, 1960, p. 86, pl. 42, figs. 13, 14.
- Siphotextularia crispata* (Brady)
Textularia crispata Brady, 1884, p. 359, pl. 113, figs. 2.
- Siphotextularia crispata* (Brady). Cushman, Todd and Post, 1954, p. 330.
- Siphotextularia saulcyana* (d'Orbigny)
Textularia saulcyana d'Orbigny, 1839a, p. 146, pl. 1, fig. 21, 22.
Textularia (*Siphotextularia*) *saulcyana* d'Orbigny. Asano, 1950c, p. 7, figs. 33, 34.
- Sphaeroidina austriaca* d'Orbigny
Sphaeroidina austriaca d'Orbigny, 1846, p. 284, pl. 20, figs. 19-21.
- Sphaeroidina bulloides* d'Orbigny
Sphaeroidina bulloides d'Orbigny, 1826, p. 267.
- Sphaeroidina compacta* Cushman and Todd
Sphaeroidina compacta Cushman and Todd, 1949, p. 19, pl. 4, fig. 14.
- Spirillina limbata* Brady ..Pl. 23, fig. 9
Spirillina limbata Brady, 1879, p. 278, pl. 8, fig. 26.
- Spirolina arietina* (Batsch)
Nautilus (*Lituus*) *arietinus* Batsch, 1791, p. 4, pl. 6, fig. 15. (*fide* Ellis and Messina, 1940 *et seq.*).
- Spirolina arietina* (Batsch). Cushman, 1933b, p. 43, pl. 15, figs. 4, 5.
- Spiroloculina angulata* Cushman
Spiroloculina grata var. *angulata* Cushman, 1917, p. 36, pl. 7, fig. 5.
- Spiroloculina angulata* Cushman. Cushman and Todd, 1944a, p. 50. (*fide* Barker, 1960)
- Spiroloculina antillarum* d'Orbigny
Spiroloculina antillarum d'Orbigny, 1839, p. 166, pl. 9, figs. 3, 4.
- Spiroloculina canaliculata* d'Orbigny
Spiroloculina canaliculata d'Orbigny, 1846, p. 269, pl. 16, figs. 10-12.
- Spiroloculina communis* Cushman and Todd

- Spiroloculina communis* Cushman and Todd, 1944b, p. 63, pl. 9, figs. 4, 5, 7, 8.
- Spiroloculina communis incisa* Cushman
Spiroloculina grateloupi d'Orbigny *incisa* Cushman, 1921, p. 397, pl. 78, fig. 5.
- Spiroloculina communis incisa* Cushman. Asano, 1951a, p. 13, figs. 89, 90.
- Spiroloculina hadai* Thalmann
Spiroloculina hadai Thalmann, 1933, p. 354.
- Spiroloculina dentata* Cushman and Todd
Spiroloculina dentata Cushman and Todd, 1944b, p. 71, pl. 9, figs. 33, 34.
- Spiroplectammina higuchii* Takayanagi
.....Pl. 14, fig. 4
Spiroplectammina higuchii Takayanagi, 1953, p. 27, pl. 4, fig. 1.
- Stainforthia exilis* (Brady) var. *tenuata* (Cushman)
Buliminella subfusiformis Cushman var. *tenuata* Cushman, 1927, p. 149, pl. 2, fig. 9.
- Bulimina exilis* Brady var. *tenuata* (Cushman). Cushman and Parker, 1947, p. 124, pl. 28, fig. 29.
- Streptochilus globigerum* (Schwager)
Textularia globigera Schwager, 1866, p. 252, pl. 7, fig. 100.
- Streptochilus globigerum* (Schwager). Brönnimann and Resig, 1971, p. 1288, pl. 51, fig. 2. (*vide* Srinivasan and Sharma, 1980).
- Stilostomella ketienziensis* (Ishizaki)
Ellipsonodosaria ketienziensis Ishizaki, 1943, p. 684, text-figs. 1, 6, 11.
- Stilostomella ketienziensis* (Ishizaki). Takayanagi and Hasegawa, 1987, p. 15.
- Stilostomella lepidula* (Schwager)
Nodosaria lepidula Schwager, 1866, p. 210, pl. 5, figs. 27, 28.
- Stilostomella lepidula* (Schwager). LeRoy, 1964, p. F35.
- Textularia abbreviata* d'Orbigny
Textularia abbreviata d'Orbigny, 1846, p. 249, pl. 15, figs. 9-12.
- Textularia agglutinans* d'Orbigny
Textularia agglutinans d'Orbigny, 1839a, p. 136, pl. 1, figs. 17, 18, 32, 34.
- Textularia articulata* d'Orbigny
Textularia articulata d'Orbigny, 1846, p. 250, pl. 15, figs. 16-18.
- Textularia astutia* Lalicker and McCulloch
Textularia astutia Lalicker and McCulloch, 1940, p. 119, pl. 13, fig. 4.
- Textularia aura* Lalicker and McCulloch
Textularia aura Lalicker and McCulloch, 1940, p. 119, pl. 13, fig. 5.
- Textularia calva* Lalicker
Textularia calva Lalicker, 1935, p. 1, pl. 1, figs. 1, 2. (*vide* Ellis and Messina, 1940 *et seq.*).
- Textularia candeiana* d'Orbigny
Textularia candeiana d'Orbigny, 1839a, p. 143, pl. 1, figs. 25-27.
- Textularia catenata* Cushman
Textularia catenata Cushman, 1911, p. 23, text-figs. 39, 40.
- Textularia conica* d'Orbigny
Textularia conica d'Orbigny, 1839a, p. 143, pl. 1, figs. 19, 20.
- Textularia fistula* Cushman
Textularia agglutinans d'Orbigny var. *fistula* Cushman, 1911, p. 10, fig. 11.
- Textularia fistula* Cushman. Cushman and McCulloch, 1940, p. 127, pl. 14, fig. 10.
- Textularia rugosa* (Reuss)
Plecanium rugosa Reuss, 1869, p. 453, pl. 1, fig. 3. (*vide* Ellis and Messina, 1940 *et seq.*).
- Textularia rugosa* (Reuss). Brady, 1884, p. 362, pl. 42, figs. 23, 24.
- Textularia sagittula* Defrance
Textularia sagittula Defrance, 1824, p. 177. (*vide* Ellis and Messina, 1940 *et seq.*).
- Textularia sagittula* Defrance var. *atrata* Cushman
Textularia sagittula Defrance var. *atrata* Cushman, 1911, p. 7, figs. 2-5.
- Textularia sagittula* Defrance var.

- fistulosa* Brady
Textularia sagittula Defrance var.
fistulosa Brady, 1884, p. 362, pl. 42,
 figs. 19-22.
- Textularia schencki* Cushman and Valen-
 tine
Textularia schencki Cushman and Val-
 entine, 1930, p. 8, pl. 1, fig. 3.
- Textularia scripta* Lalicker and McCul-
 loch
Textularia scripta Lalicker and
 McCulloch, 1940, p. 141, pl. 16, fig.
 25.
- Textularia semialta* Cushman
Textularia semialta Cushman, 1913b,
 p. 634, pl. 80, figs. 6, 7.
- Textularia* cf. *trochus* d'Orbigny
 cf. *Textularia trochus* d'Orbigny, 1840,
 p. 45, pl. 4, figs. 25, 26.
- Textularia vola* Lalicker and McCulloch
Textularia vola Lalicker and McCul-
 loch, 1940, p. 142, pl. 16, fig. 27.
- Textulariella pacifica* Cushman
Textulariella pacifica Cushman, 1937,
 p. 67, pl. 7, figs. 11-13.
- Thalmannammina parkerae* (Uchio)
Recurvoidella parkerae Uchio, 1960, p.
 53, pl. 1, figs. 18, 19.
- Thalmannammina parkerae* (Uchio).
 Loeblich and Tappan, 1964, p. 226,
 fig. 136, 8.
- Tosaia hanzawai* Takayanagi
Pl. 22, fig. 10
- Tosaia hanzawai* Takayanagi, 1953, p.
 30, pl. 4, fig. 7.
- Trifarina bradyi* Cushman. .Pl. 15, fig. 5
Trifarina bradyi Cushman, 1923, p. 99,
 pl. 22, figs. 3-9.
- Trifarina carinata* (Cushman)
Angulogerina carinata Cushman,
 1927c, p. 159, pl. 4, fig. 3.
- Trifarina hughesi* (Galloway and Wiss-
 ler)
Uvigerina hughesi Galloway and Wiss-
 ler, 1927, p. 76, pl. 12, fig. 5.
- Trifarina occidentalis* (Cushman)
Uvigerina occidentalis Cushman, 1923,
 p. 169.
- Triloculina tricarinata* d'Orbigny
Pl. 22, fig. 7
- Triloculina tricarinata* d'Orbigny,
 1826, p. 299.
- Triloculina trigonula* (Lamarck)
Pl. 17, fig. 10
- Miliolites trigonula* Lamarck, 1804, p.
 351, No. 3. (*vide* Ellis and Messina,
 1940 *et seq.*).
- Triloculina trigonula* (Lamarck).
 d'Orbigny, 1826, p. 299, pl. 16, figs.
 5-9, modèles, no. 93.
- Trochammina charlottensis* Cushman
Trochammina charlottensis Cushman,
 1925, p. 39, pl. 6, fig. 4.
- Trochammina discorbis* Earland
Trochammina discorbis Earland, 1934,
 p. 104, pl. 3, figs. 28-31.
- Trochammina globigeriniformis* (Parker
 and Jones) Pl. 11, fig. 6; Pl. 13, fig. 9
- Lituola nautiloidea* var. *globigeriniformis*
 Parker and Jones, 1865, p. 407,
 pl. 15, figs. 46, 47.
- Trochammina globigeriniformis* (Par-
 ker and Jones). Poag, 1981, p. 85,
 pl. 13, fig. 1; pl. 14, fig. 1.
- Trochammina inflata* (Montagu)
Pl. 11, fig. 4
- Nautilus inflata* Montagu, 1808, p. 81,
 pl. 18, fig. 3. (*vide* Ellis and Messina,
 1940 *et seq.*).
- Trochammina inflata* (Montagu).
 Carpenter, Parker and Jones, 1862,
 p. 141, pl. 11, fig. 5. (*vide* Brady,
 1884).
- Trochammina nana* (Brady)
Haplophragmium nanum Brady, 1881,
 p. 50.
- Trochammina nana* (Brady). Barker,
 1960, p. 72, pl. 35, figs. 6-8.
- Trochammina nitida* Brady
Trochammina nitida Brady, 1881, p.
 52.
- Trochammina pacifica* Cushman
Pl. 11, fig. 5
- Trochammina pacifica* Cushman, 1925,
 p. 39, pl. 6, fig. 3.
- Trochammina squamata* Jones and Par-
 ker
Trochammina squamata Jones and

- Parker, 1860, p. 304. (*vide* Ellis and Messina, 1940 *et seq.*).
- Trochammina squamiformis* Cushman and McCulloch
Trochammina squamiformis Cushman and McCulloch, 1939, p. 108, pl. 12, fig. 4.
- Trochammina vesicularis* Goës
Trochammina vesicularis Goës, 1894, p. 31, pl. 6, figs. 235-237. (*vide* Ellis and Messina, 1940 *et seq.*).
- Tubulogenerina zanzibarica* (Cushman)
Brizalina zanzibarica Cushman, 1936b, p. 58, pl. 8, fig. 12.
Tubulogenerina zanzibarica (Cushman). Loeblich and Tappan, 1964, p. 654.
- Uvigerina asperula* Čížek
Uvigerina asperula Čížek, 1848, p. 146, pl. 13, figs. 14, 15. (*vide* Ellis and Messina, 1940 *et seq.*).
- Uvigerina flintii* Cushman
Uvigerina flintii Cushman, 1923, p. 165, pl. 42, fig. 13.
- Uvigerina hispida* Schwager
.....Pl. 16, fig. 11
Uvigerina hispida Schwager, 1866, p. 249, pl. 7, fig. 95.
- Uvigerina hispidocostata* Cushman and ToddPl. 16, fig. 10
Uvigerina hispido-costata Cushman and Todd, 1945, p. 51, pl. 7, figs. 27, 31.
- Uvigerina interrupta* Brady
Uvigerina interrupta Brady, 1879, p. 274, pl. 8, figs. 17, 18.
- Uvigerina nitidula* Schwager
Uvigerina nitidula Schwager, 1866, p. 248, pl. 7, fig. 93.
- Uvigerina peregrina* Cushman
Uvigerina peregrina Cushman, 1923, p. 166, pl. 42, figs. 7-10.
- Uvigerina peregrina pavula* Cushman
Uvigerina peregrina pavula Cushman, 1923, p. 168, pl. 42, fig. 11.
- Uvigerina peregrina* Cushman *shiwoensis* Asano
Uvigerina peregrina Cushman *shiwoensis* Asano, 1958, p. 35, pl. 6, figs. 5-8.
- Uvigerina porrecta* Brady ..Pl. 15, fig. 2
Uvigerina porrecta Brady, 1879, p. 274, pl. 8, figs. 15, 16.
- Uvigerina proboscidea* Schwager
.....Pl. 22, fig. 12
Uvigerina proboscidea Schwager, 1866, p. 250, pl. 7, fig. 96.
- Uvigerina proboscidea vadescens* CushmanPl. 16, fig. 12
Uvigerina proboscidea Schwager var. *vadescens* Cushman, 1933, p. 85, pl. 8, figs. 14, 15.
- Uvigerina pseudoampullacea* Asano
Uvigerina pseudoampullacea Asano, 1938d, p. 613, pl. 17, figs. 28, 29.
- Uvigerian schencki* Asano ..Pl. 16, fig. 13
Uvigerian schencki Asano, 1950b, p. 17, figs. 74, 75.
- Uvigerina schwageri* Brady
Uvigerina schwageri Brady, 1884, p. 575, pl. 74, figs. 8-10.
- Vaginulinopsis asanoi* Ishizaki
.....Pl. 22, fig. 3
Vaginulinopsis asanoi Ishizaki, 1942, p. 106, text-figs. 1-3.
- Vaginulinopsis pacifica* (Cushman and Hanzawa)
Polymorphinella pacifica Cushman and Hanzawa, 1936, p. 47.
Vaginulinopsis pacifica (Cushman and Hanzawa). Parr, 1950, p. 325. (*vide* Barker, 1960).
- Vaginulinopsis tasmanica* Parr
Vaginulinopsis tasmanica Parr, 1950, p. 324, pl. 11, figs. 13, 14.
- Valvobifarina mackinnoni* (Millett) var. *robusta* (Sidebottom)
Bifarina mackinnoni (Millett) var. *robusta* Sidebottom, 1818, p. 125, pl. 3, figs. 17, 18. (*vide* Ellis and Messina, 1940 *et seq.*).
- Valvulineria hamanaoensis* (Ishiwada)
Anomalina hamanaoensis Ishiwada, 1958, p. 18, text-fig. 3, pl. 1, figs. 24-27.
- Valvulineria hamanaoensis* (Ishiwada). Matoba, 1970, p. 63, pl. 4, figs. 12, 13.
- Valvulineria rugosa* (d'Orbigny)

- Rosalina rugosa* d'Orbigny, 1839c, p. 42, pl. 2, figs. 12-14.
Valvulineria rugosa (d'Orbigny). Barker, 1960, p. 180, pl. 87, fig. 3.
Valvulineria rugosa (d'Orbigny) var. *minuta* (Schubert)
Discorbina rugosa d'Orbigny var. *minuta* Schubert, 1904, p. 420. (*vide* Ellis and Messina, 1940 *et seq.*).
Valvulineria rugosa (d'Orbigny) var. *minuta* (Schubert). Barker, 1960, p. 188, pl. 91, fig. 4.
Vertebralina striata d'Orbigny
Vertebralina striata d'Orbigny, 1826, p. 283, no. 1.
Wiesnerella auriculata (Egger)
Planispirina auriculata Egger, 1893, p. 245, pl. 3, figs. 13-15. (*vide* Ellis and Messina, 1940 *et seq.*).
Wiesnerella auriculata (Egger). Cushman, 1933d, p. 33, pl. 3, figs. 7-9.

DESCRIPTION OF NEW SPECIES

Superfamily Discorbacea
 Ehrenberg, 1938
 Family Discorbidea Ehrenberg, 1938
 Genus *Valvulineria* Cushman, 1926
Valvulineria gunjii
 Akimoto, n. sp.

Pl. 21, fig. 6.

Description: Test medium, ovoid in outline, longer than broad, biconvex, trochospirally coiled, umbilical side more convex than spiral side, periphery rounded, not lobulate; composed of two whorls; chambers somewhat inflated, seven in last whorl, increasing gradually in size as added; suture distinct, not limbate, radial, gently curved, somewhat depressed on spiral side, umbilical side not limbate, radial, nearly straight,

slightly depressed: aperture interiomarginal, extraumbilical-umbilical; several continuous series of rounded opening along suture; umbilicus covered with rhomboidal valvular plates.

Dimension (in mm):

	length	breadth	thickness
Holotype	0.37	0.23	0.18
Paratypes	0.36	0.22	0.18
	0.36	0.23	0.18

Type and occurrence: Holotype, IGPS coll. cat. no. 100603, from sample KT86-11-P8, Nankai Trough.

Occurrence: Rare, only at type locality.

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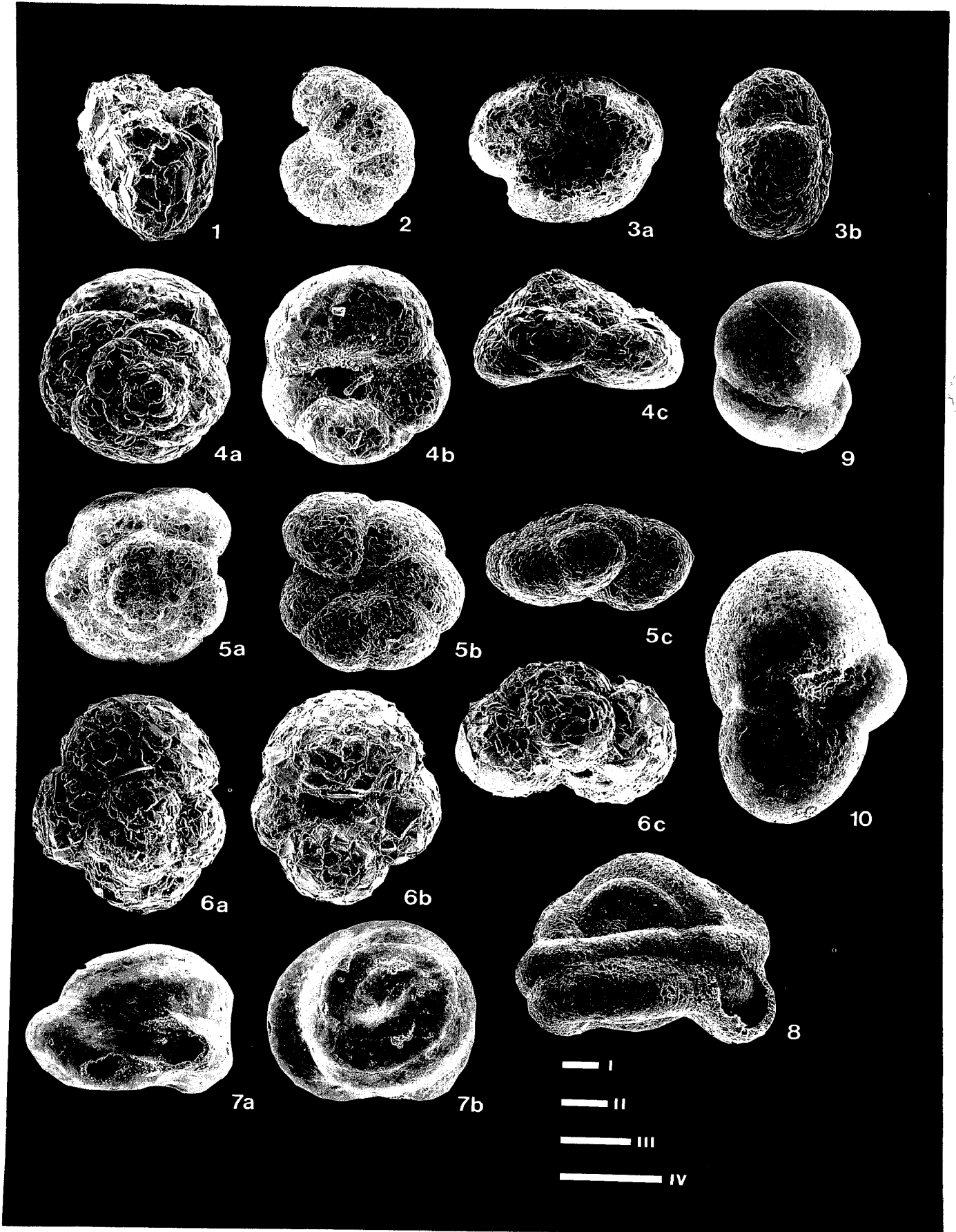
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Plate 11

- Fig. 1. *Adercotryma glomeratum* (Brady)
(IGPS 101001), Sample GH80-4-D372, Hachijojima area, $\times 200$.
- Fig. 2. *Haplophragmoides columbiensis* Cushman
(IGPS 101002), Sample GH80-4-D372, Hachijojima area, $\times 200$.
- Fig. 3. *Alveolophragmium subglobosa* (G.O. Sars)
(IGPS 101003), Sample GH80-4-D372, Hachijojima area, $\times 150$.
- Fig. 4. *Trochammina inflata* (Montagu)
(IGPS 101004), Sample GH80-4-D376, Hachijojima area, $\times 300$.
- Fig. 5. *Trochammina pacifica* Cushman
(IGPS 101005), Sample GH80-4-D372, Hachijojima area, $\times 200$.
- Fig. 6. *Trochammina globigeriniformis* (Parker and Jones)
(IGPS 101006), Sample GH80-4-D372, Hachijojima area, $\times 300$.
- Fig. 7. *Glomospira chaloides* (Jones and Parker)
(IGPS 101007), Sample GH80-4-D372, Hachijojima area, $\times 300$.
- Fig. 8. *Glomospira gordialis* (Jones and Parker)
(IGPS 101008), Sample GH80-4-D372, Hachijojima area, $\times 300$.
- Fig. 9. *Alveolophragmium ringens* (Brady)
(IGPS 101009), Sample GH80-4-D372, Hachijojima area, $\times 300$.
- Fig. 10. *Cystammina pauciloculata* (Brady)
(IGPS 101010), Sample GH80-4-D376, Hachijojima area, $\times 300$.

Scale bar : 0.1 mm ; I : 9 ; II : 3 ; III : 1, 2, 5 ; IV : 4, 6, 7, 8, 10.



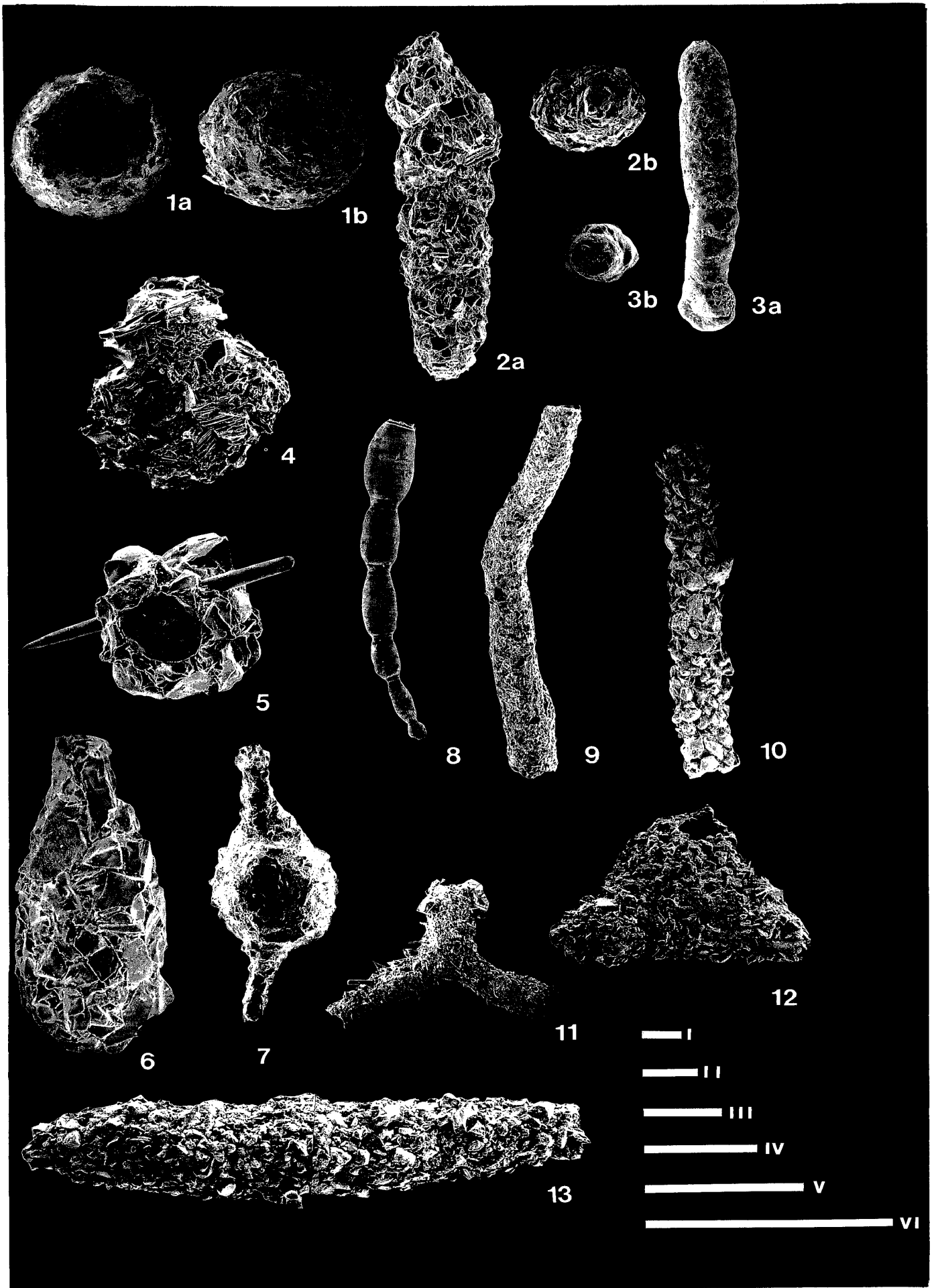


Plate 12

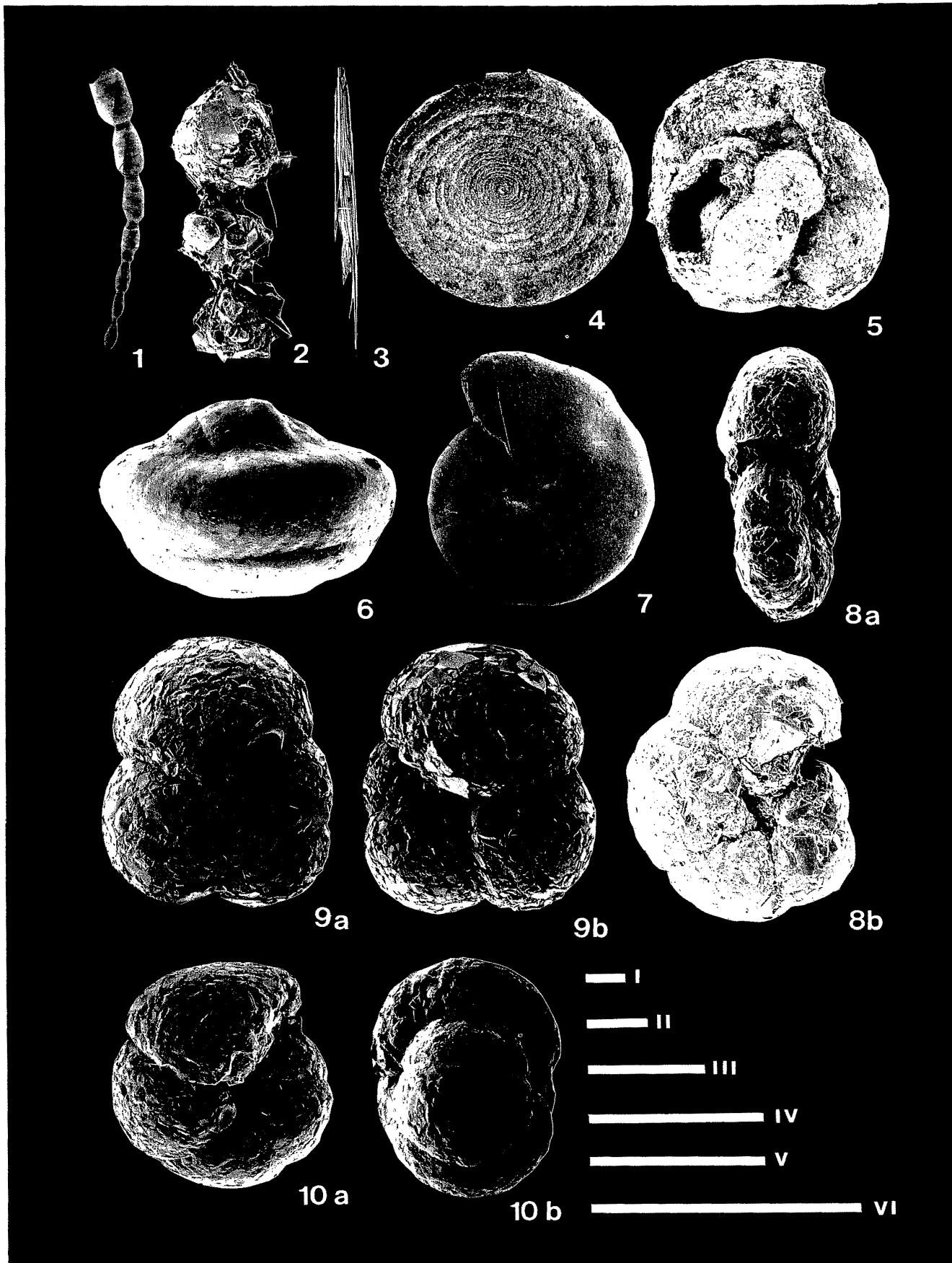
- Fig. 1. *Saccamina sphaerica* M. Sars
(IGPS 101011), Sample GH80-4-D372, Hachijojima area, $\times 100$.
- Fig. 2. *Karreriella apicularis* (Cushman)
(IGPS 101012), Sample GH80-4-D372, Hachijojima area, $\times 150$.
- Fig. 3. *Ammobaculites agglutinans filiformis* Earland
(IGPS 101013), Sample GH80-4-D372, Hachijojima area, $\times 200$.
- Fig. 4. *Ammobaculites agglutinans* (d'Orbigny)
(IGPS 101014), Sample GH80-4-D372, Hachijojima area, $\times 70$.
- Fig. 5. *Psammosphaera parva* Flint
(IGPS 101015), Sample GH80-4-D372, Hachijojima area, $\times 200$.
- Fig. 6. *Reophax difflugiformis* Brady
(IGPS 101016), Sample GH80-4-D372, Hachijojima area, $\times 300$.
- Fig. 7. *Reophax distans* Brady
(IGPS 101017), Sample GH80-4-D372, Hachijojima area, $\times 70$.
- Fig. 8. *Nodellum membranaceum* (Brady)
(IGPS 101018), Sample GH80-4-D372, Hachijojima area, $\times 150$.
- Fig. 9. *Marsipella elongata* Norman
(IGPS 101019), Sample GH80-4-D373, Hachijojima area, $\times 100$.
- Fig. 10. *Rhabdammina abyssorum* M. Sars
(IGPS 101020), Sample GH80-4-D375, Hachijojima area, $\times 70$.
- Fig. 11. *Saccorhiza ramosa* Brady
(IGPS 101021), Sample GH80-4-D373, Hachijojima area, $\times 45$.
- Fig. 12. *Rhabdammina abyssorum* M. Sars forma A
(IGPS 101022), Sample GH80-4-D375, Hachijojima area, $\times 45$.
- Fig. 13. *Rhabdammina abyssorum* M. Sars forma A
(IGPS 101023), Sample GH80-4-D375, Hachijojima area, $\times 45$.

Scale bar : 0.1 mm ; I : 1, 9 ; II : 2, 8 ; III : 3, 5. IV : 6 ; 1.0 mm ; V : 11, 12, 13 ; VI : 4, 7, 10.

Plate 13

- Fig. 1. *Nodellum membranaceum* (Brady)
(IGPS 101024), Sample GH84-3-3, Tanegashima area, $\times 100$.
- Fig. 2. *Reophax distans* Brady
(IGPS 101025), Sample GH84-3-3, Tanegashima area, $\times 100$.
- Fig. 3. *Reophax spiculifer* Brady
(IGPS 101026), Sample GH84-3-3, Tanegashima area, $\times 45$.
- Fig. 4. *Ammodiscus pacifica* Cushman and Valentine
(IGPS 101027), Sample GH84-3-3, Tanegashima area, $\times 45$.
- Fig. 5. *Glomospira gordialis* (Jones and Parker)
(IGPS 101028), Sample GH84-3-3, Tanegashima area, $\times 100$.
- Fig. 6. *Glomospira charoides* (Jones and Parker)
(IGPS 101029), Sample GH84-3-3, Tanegashima area, $\times 300$.
- Fig. 7. *Cyclammima bradyi* Cushman
(IGPS 101030), Sample GH84-3-3, Tanegashima area, $\times 150$.
- Fig. 8. *Haplophragmoides columbiensis* Cushman
(IGPS 101031), Sample GH84-3-52, Tanegashima area, $\times 200$.
- Fig. 9. *Trochammima globigeriniformis* (Parker and Jones)
(IGPS 101032), Sample GH84-3-3, Tanegashima area, $\times 150$.
- Fig. 10. *Alveolophragmium subglobosa* (G.O. Sars)
(IGPS 101033), Sample GH84-3-3, Tanegashima area, $\times 70$.

Scale bar : 0.1 mm ; I : 1, 2, 5 ; II : 7, 8, 9 ; III : 6. 1.0 mm ; IV : 3, 4 ; V : 10.



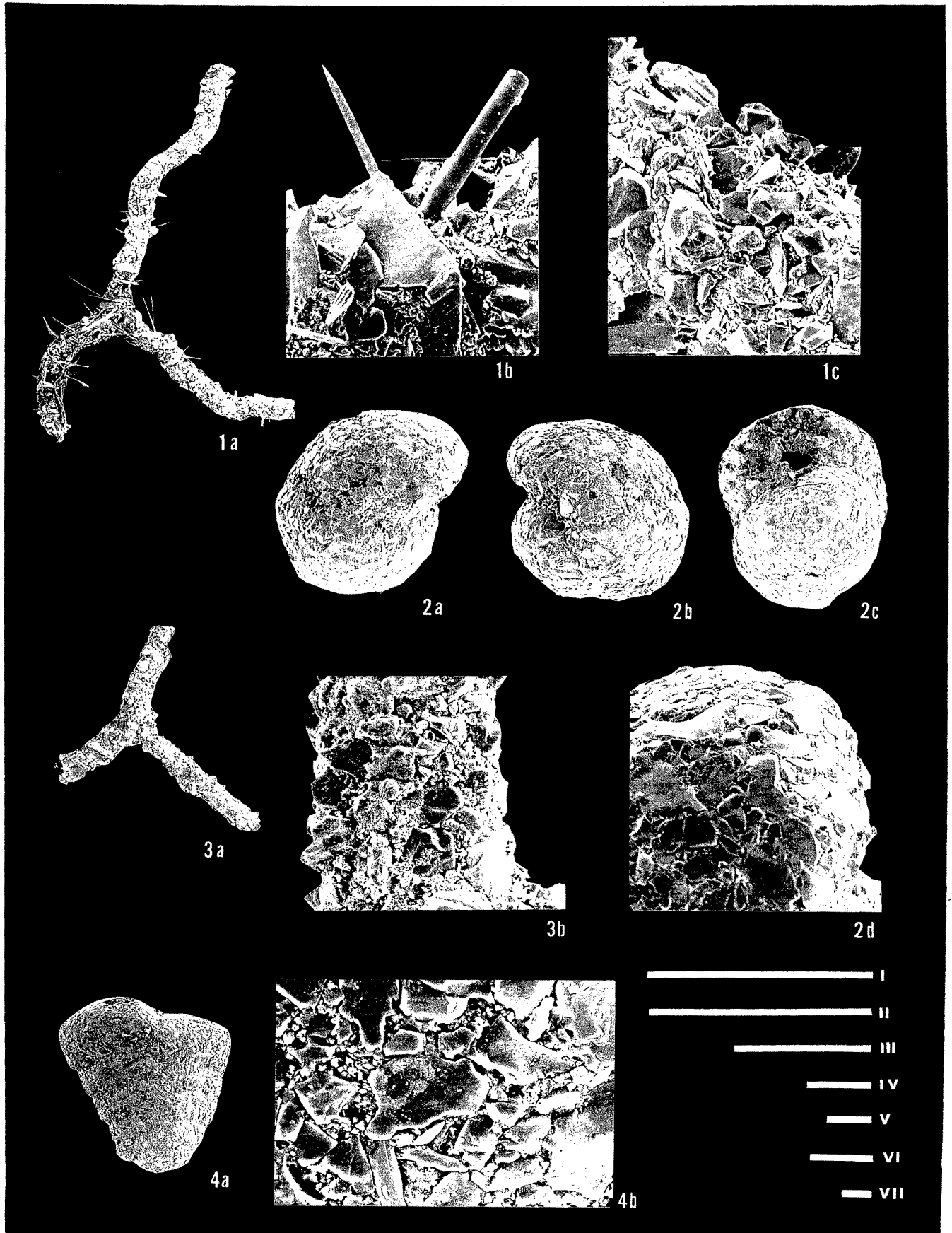


Plate 14

Fig. 1. *Saccorhiza ramosa* Brady

(IGPS 101034), Sample KT85-6-G7, Enshunada area, a, $\times 70$; b, c, $\times 700$.

Fig. 2. *Alveolpharagmium subglobosa* (G.O. Sars)

(IGPS 101035), Sample KT86-11-MP8, Nankai Trough, a, b, c, $\times 150$; d, $\times 450$.

Fig. 3. *Rhabdammina abyssorum* M. Sars

(IGPS 101036), Sample KT-85-6-G7, Enshunada area, a, $\times 150$; b, $\times 1,000$.

Fig. 4. *Spiroplectammia higuchii* Takayanagi

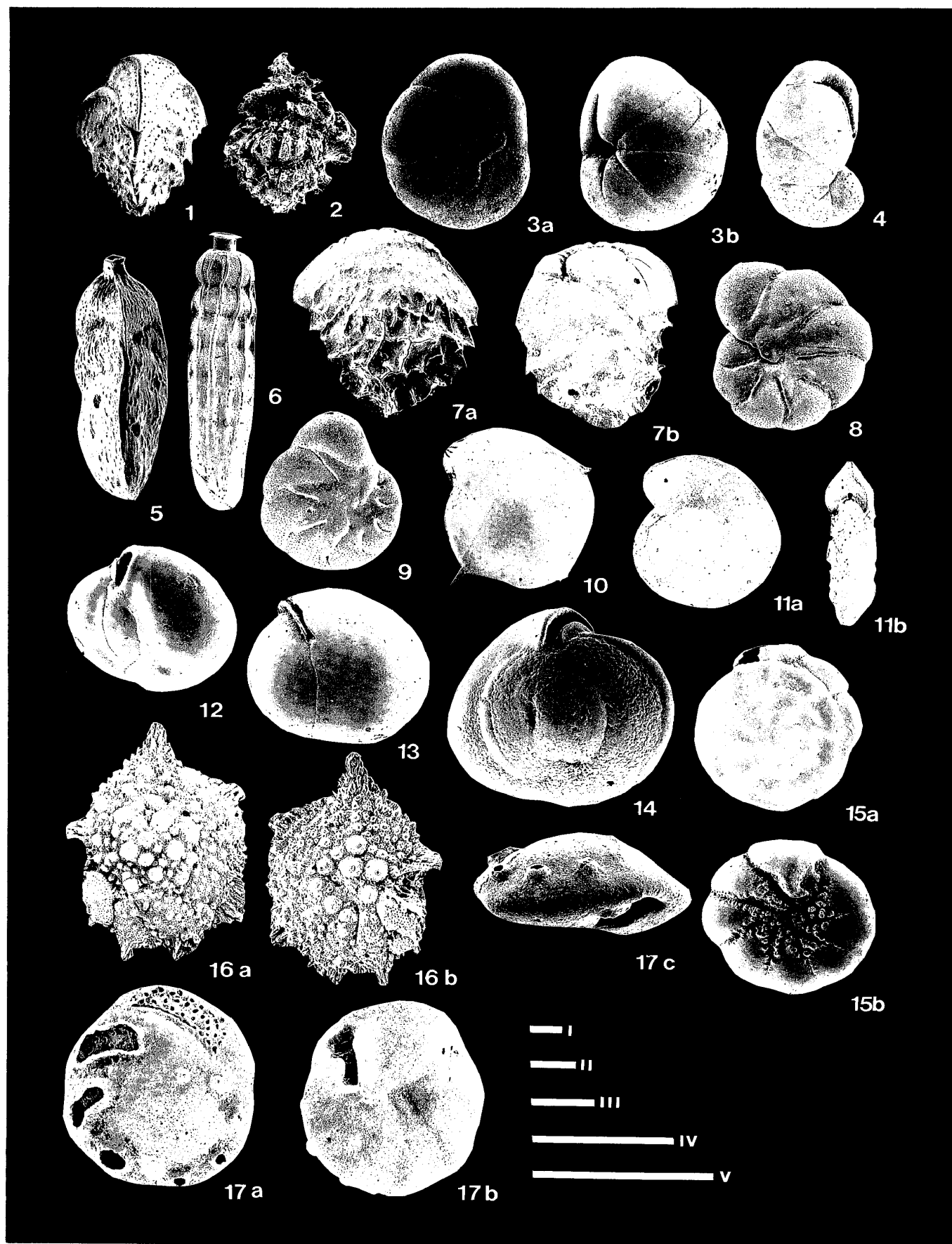
(IGPS 101037), Sample KT85-6-G5, Enshunada area, a, $\times 200$; b, $\times 2,000$.

Scale bar: 1.0 mm; I: 1a. 0.1 mm; II: 1b, c; III: 2d; IV: 4a; V: 2a-c, 3a. 0.01 mm; VI: 3b;
VII: 4b.

Plate 15

- Fig. 1. *Reussella spinulosa* (Reuss)
(IGPS 101038), Sample GH84-3-54, Tanegashima area, $\times 150$.
- Fig. 2. *Uvigerina porrecta* Brady
(IGPS 101039), Sample GH84-3-55, Tanegashima area, $\times 200$.
- Fig. 3. *Ceratobulimina pacifica* Cushman and Harris
(IGPS 101040), Sample GH84-3-3, Tanegashima area, $\times 100$.
- Fig. 4. *Paracassidulina miuraensis* (Higuchi)
(IGPS 101041), Sample GH84-3-52, Tanegashima area, $\times 200$.
- Fig. 5. *Trifarina bradyi* Cushman
(IGPS 101042), Sample GH84-3-52, Tanegashima area, $\times 200$.
- Fig. 6. *Rectobolivina raphanus* (Parker and Jones)
(IGPS 101043), Sample GH84-3-55, Tanegashima area, $\times 100$.
- Fig. 7. *Ehrenbergina hysteric* Brady
(IGPS 101044), Sample GH84-3-54, Tanegashima area, $\times 200$.
- Fig. 8. *Astrononion stelligerum* (d'Orbigny)
(IGPS 101045), Sample GH84-3-52, Tanegashima area, $\times 150$.
- Fig. 9. *Astrononion stellatum* Cushman and Edwards
(IGPS 101046), Sample GH84-3-52, Tanegashima area, $\times 200$.
- Fig. 10. *Robulus calcar* (Linné)
(IGPS 101047), Sample GH84-3-54, Tanegashima area, $\times 70$.
- Fig. 11. *Operculina ammonoides* (Gronovius)
(IGPS 101048), Sample GH84-3-101, Tanegashima area, $\times 70$.
- Fig. 12. *Miliolinella circularis* (Bornemann)
(IGPS 101049), Sample GH84-3-54, Tanegashima area, $\times 100$.
- Fig. 13. *Miliolinella inflata* LeRoy
(IGPS 101050), Sample GH84-3-55, Tanegashima area, $\times 150$.
- Fig. 14. *Miliolinella australis* (Parr)
(IGPS 101051), Sample GH84-3-54, Tanegashima area, $\times 200$.
- Fig. 15. *Ammonia takanabensis* (Ishizaki)
(IGPS 101052), Sample GH84-3-52, Tanegashima area, $\times 100$.
- Fig. 16. *Pararotalia calcar* (d'Orbigny)
(IGPS 101053), Sample GH84-3-101, Tanegashima area, $\times 100$.
- Fig. 17. *Alabamina tubulifera* (Heron-Allen and Earland)
(IGPS 101054), Sample GH84-3-52, Tanegashima area, $\times 200$.

Scale bar : 0.1 mm ; I : 3, 6, 12, 15, 16 ; II : 1, 8, 13 ; III : 2, 4, 5, 7, 9, 14, 17. 1.0 mm ; IV : 11 ; V : 10.



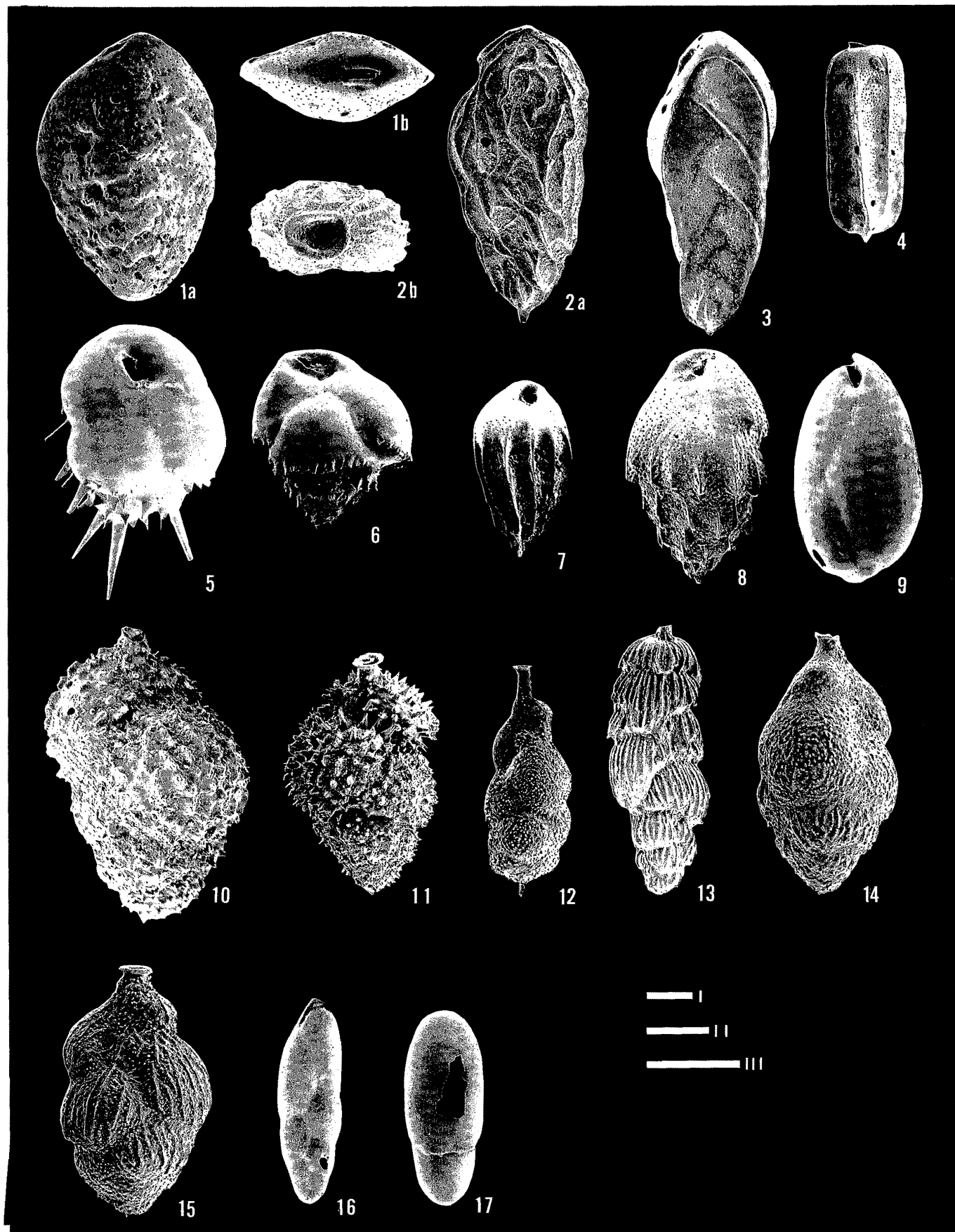


Plate 16

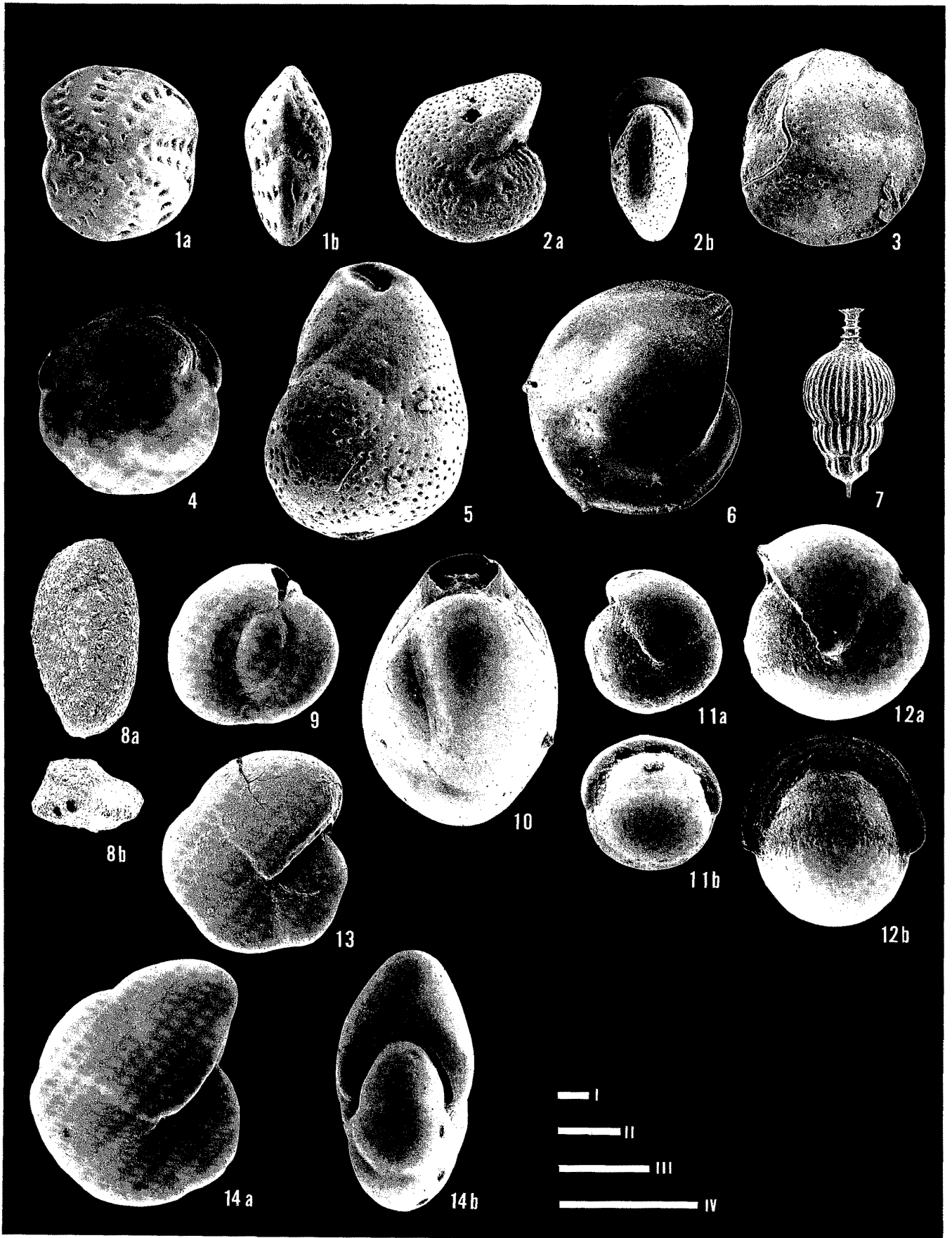
- Fig. 1. *Bolivina robusta* Brady
(IGPS 101055), Sample KT85-6-G2, Enshunada area, $\times 300$.
- Fig. 2. *Brizalina karreriana* (Brady)
(IGPS 101056), Sample KT85-6-G3, Enshunada area, $\times 300$.
- Fig. 3. *Bolivinita quadrilatera* (Schwager)
(IGPS 101057), Sample KT85-6-G2, Enshunada area, $\times 200$.
- Fig. 4. *Rectobolivina raphanus* (Parker and Jones)
(IGPS 101058), Sample KT85-6-G4, Enshunada area, $\times 200$.
- Fig. 5. *Bulimina aculeata* d'Orbigny
(IGPS 101059), Sample KT85-6-G1, Enshunada area, $\times 300$.
- Fig. 6. *Bulimina marginata* d'Orbigny
(IGPS 101060), Sample KT85-6-G4, Enshunada area, $\times 200$.
- Fig. 7. *Bulimina rostrata* Brady
(IGPS 101061), Sample KT85-6-G2, Enshunada area, $\times 200$.
- Fig. 8. *Bulimina striata* d'Orbigny
(IGPS 101062), Sample KT85-6-G2, Enshunada area, $\times 200$.
- Fig. 9. *Globobulimina pacifica* Cushman
(IGPS 101063), Sample KT86-11-MP8, Nankai Trough, $\times 150$.
- Fig. 10. *Uvigerina hispidocostata* Cushman and Todd
(IGPS 101064), Sample KT85-6-G1, Enshunada area, $\times 200$.
- Fig. 11. *Uvigerina hispida* Schwager
(IGPS 101065), Sample KT85-6-G1, Enshunada area, $\times 150$.
- Fig. 12. *Uvigerina proboscidea vadescens* Cushman
(IGPS 101066), Sample KT85-6-G1, Enshunada area, $\times 200$.
- Fig. 13. *Uvigerina schencki* Asano
(IGPS 101067), Sample KT85-6-G5, Enshunada area, $\times 150$.
- Figs. 14, 15. *Uvigerina* sp.
14, (IGPS 101068), Sample KT85-6-G4, Enshunada area, $\times 300$.
15, (IGPS 101069), Sample KT85-6-G4, Enshunada area, $\times 300$.
- Fig. 16. *Brizalina pacifica* (Cushman and McCulloch)
(IGPS 101070), Sample KT86-11-MP8, Nankai Trough, $\times 200$.
- Fig. 17. *Chilostomella oolina* Schwager
(IGPS 101071), Sample KT86-11-MP6, Nankai Trough, $\times 150$.

Scale bar : 0.1 mm ; I : 1, 2, 5, 14, 15 ; II : 3, 4, 6, 7, 8, 10, 12, 16 ; III : 9, 11, 13, 17.

Plate 17

- Fig. 1. *Elphidium advena* (Cushman)
(IGPS 101072), Sample KT85-6-G5, Enshunada area, $\times 200$.
- Fig. 2. *Melonis parkerae* (Uchio)
(IGPS 101073), Sample KT85-6-G1, Enshunada area, $\times 200$.
- Fig. 3. *Cassidulina carinata* Silvestri
(IGPS 101074), Sample KT85-6-G2, Enshunada area, $\times 300$.
- Fig. 4. *Paracassidulina neocarinata* (Thalmann)
(IGPS 101075), Sample KT85-6-G4, Enshunada area, $\times 300$.
- Fig. 5. *Evolocassidulina brevis* (Aoki)
(IGPS 101076), Sample KT85-6-G4, Enshunada area, $\times 450$.
- Fig. 6. *Robulus calcar* (Linne)
(IGPS 101077), Sample KT85-6-G4, Enshunada area, $\times 200$.
- Fig. 7. *Amphicoryna scalaris* (Batsch)
(IGPS 101078), Sample KT85-6-G4, Enshunada area, $\times 100$.
- Fig. 8. *Sigmoilopsis schlumbergeri* (Silvestri)
(IGPS 101079), Sample KT85-6-G2, Enshunada area, $\times 100$.
- Fig. 9. *Miliolinella circularis* (Bronemann)
(IGPS 101080), Sample KT85-6-G6, Enshunada area, $\times 200$.
- Fig. 10. *Triloculina trigonula* (Lamarck)
(IGPS 101081), Sample KT85-6-G6, Enshunada area, $\times 300$.
- Figs. 11, 12. *Pullenia bulloides* (d'Orbigny)
11, (IGPS 101082), Sample KT85-6-G1, Enshunada area, $\times 200$.
12, (IGPS 101083), Sample KT85-6-G1, Enshunada area, $\times 200$.
- Fig. 13. *Pullenia quinqueloba* (Reuss)
(IGPS 101084), Sample KT85-6-G1, Enshunada area, $\times 300$.
- Fig. 14. *Pullenia salisburyi* R.E. and R.C. Stewart
(IGPS 101085), Sample KT85-6-G1, Enshunada area, $\times 200$.

Scale bar : 0.1 mm ; I : 7, 8 ; II : 1, 2, 6, 9, 11, 12, 14 ; III : 3, 10, 13 ; IV : 4, 5.



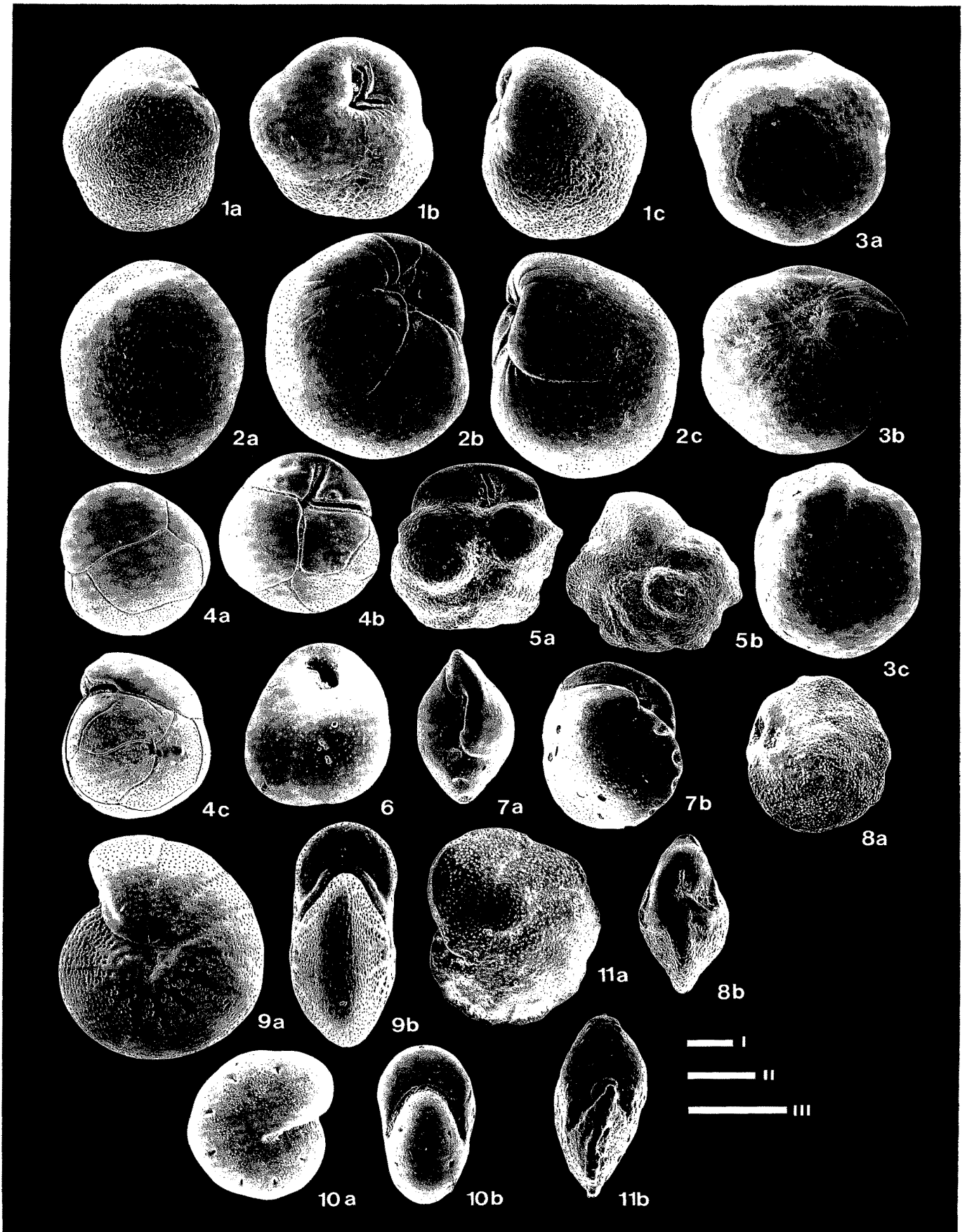


Plate 18

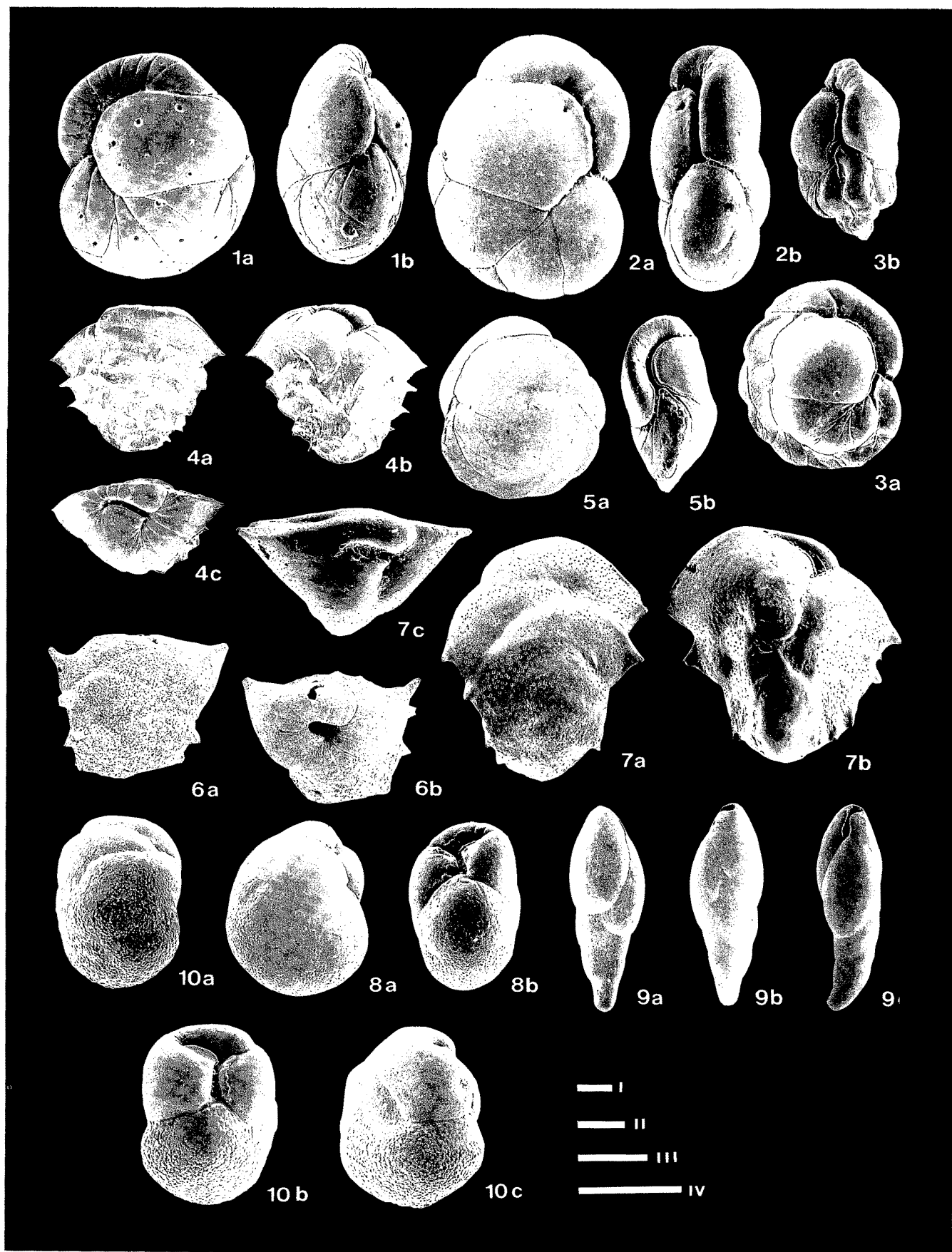
- Fig. 1. *Globocassidulina pseudoquadrata* Nomura
(IGPS 101086), Sample GH80-4-94, Hachijojima area, $\times 300$.
- Fig. 2. *Globocassidulina oriangulata* (Belford)
(IGPS 101087), Sample GH80-4-94, Hachijojima area, $\times 300$.
- Fig. 3. *Globocassidulina gemma* (Todd)
(IGPS 101088), Sample GH80-4-D370, Hachijojima area, $\times 150$.
- Fig. 4. *Globocassidulina parva* (Asano and Nakamura)
(IGPS 101089), Sample GH80-4-94, Hachijojima area, $\times 150$.
- Fig. 5. *Globocassidulina elegans* (Sidebottom)
(IGPS 101090), Sample GH80-4-D370, Hachijojima area, $\times 200$.
- Fig. 6. *Globocassidulina moluccensis* (Germeraad)
(IGPS 101091), Sample GH80-4-RC102, Hachijojima area, $\times 200$.
- Fig. 7. *Globocassidulina brocha* (Poag)
(IGPS 101092), Sample GH80-4-94, Hachijojima area, $\times 200$.
- Fig. 8. *Globocassidulina paratortuosa* (Kuwano)
(IGPS 101093), Sample GH80-4-D370, Hachijojima area, $\times 200$.
- Fig. 9. *Melonis parkerae* (Uchio)
(IGPS 101094), Sample GH80-4-RC102, Hachijojima area, $\times 150$.
- Fig. 10. *Astrononion* sp.
(IGPS 101095), Sample GH80-4-RC102, Hachijojima area, $\times 200$.
- Fig. 11. *Nonionellina* sp.
(IGPS 101096), Sample GH80-4-97, Hachijojima area, $\times 300$.

Scale bar : 0.1 mm ; I : 3, 4, 9 ; II : 5, 6, 7, 8, 10 ; III : 1, 2, 11.

Plate 19

- Fig. 1. *Paracassidulina sulcata* (Belford)
(IGPS 101097), Sample GH80-4-94, Hachijojima area, $\times 300$.
- Fig. 2. *Paracassidulina nipponensis* (Eade)
(IGPS 101098), Sample GH80-4-96, Hachijojima area, $\times 300$.
- Fig. 3. *Paracassidulina tomiyensis* (Uchio)
(IGPS 101099), Sample GH80-4-94, Hachijojima area, $\times 150$.
- Fig. 4. *Ehrenbergina hystrix* Brady
(IGPS 101100), Sample GH80-4-97, Hachijojima area, $\times 200$.
- Fig. 5. *Paracassidulina quasicarinata* Nomura
(IGPS 101101), Sample GH80-4-96, Hachijojima area, $\times 200$.
- Fig. 6. *Ehrenbergina* sp. A
(IGPS 101102), Sample GH80-4-D367, Hachijojima area, $\times 150$.
- Fig. 7. *Ehrenbergina pacifica* Cushman
(IGPS 101103), Sample GH80-4-D370, Hachijojima area, $\times 200$.
- Figs. 8, 10. *Globocassidulina pseudoquadrata* Nomura forma A
Fig. 8, (IGPS 101104), Sample GH80-4-86, Hachijojima area, $\times 300$.
Fig. 10, (IGPS 101106), Sample GH80-4-94, Hachijojima area, $\times 300$.
- Fig. 9. *Rutherfordoides mexicanus* (Cushman)
(IGPS 101105), Sample GH80-4-86, Hachijojima area, $\times 300$.

Scale bar : 0.1 mm ; I : 9 ; II : 3, 6 ; III : 4, 5, 7 ; IV : 1, 2, 8, 10.



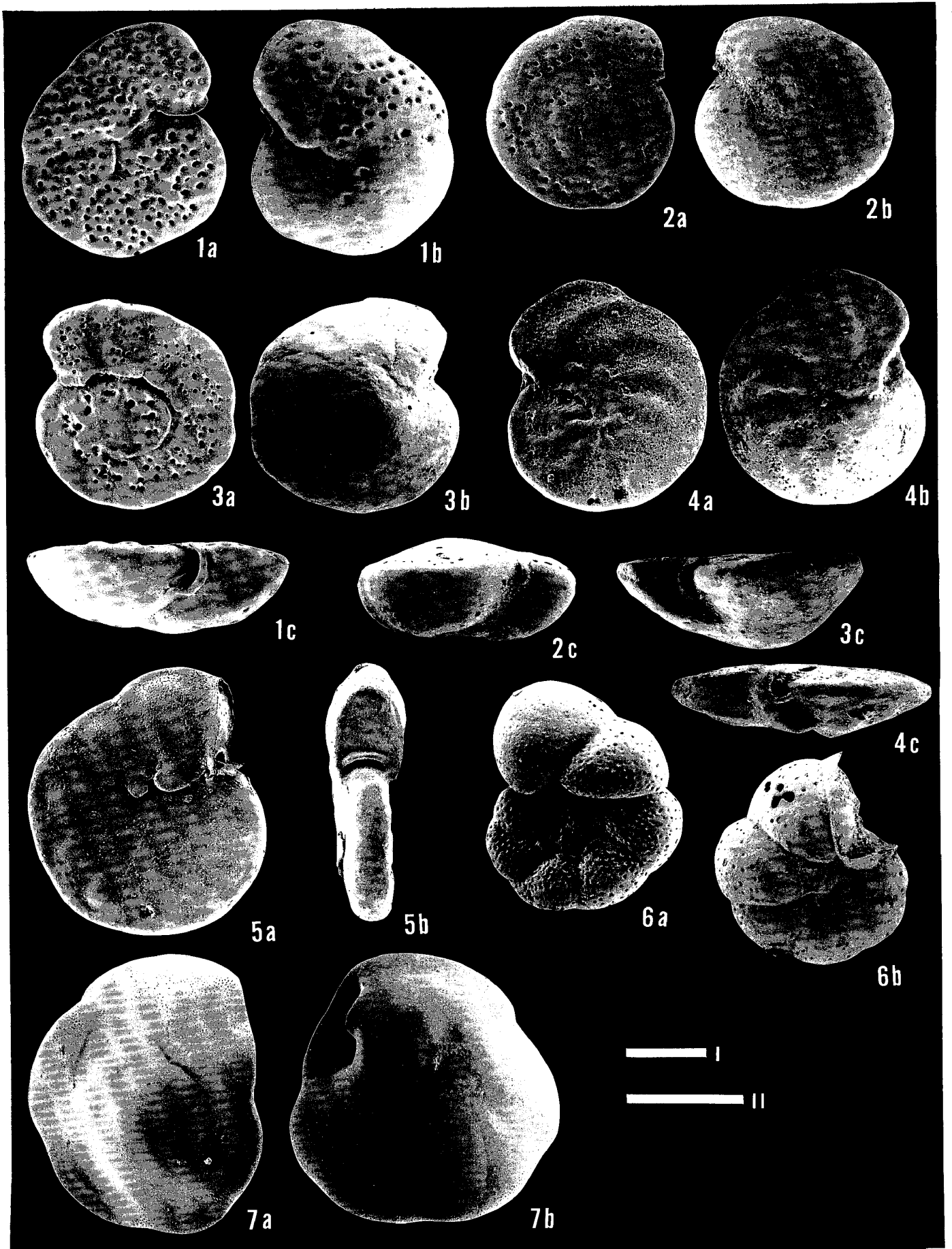


Plate 20

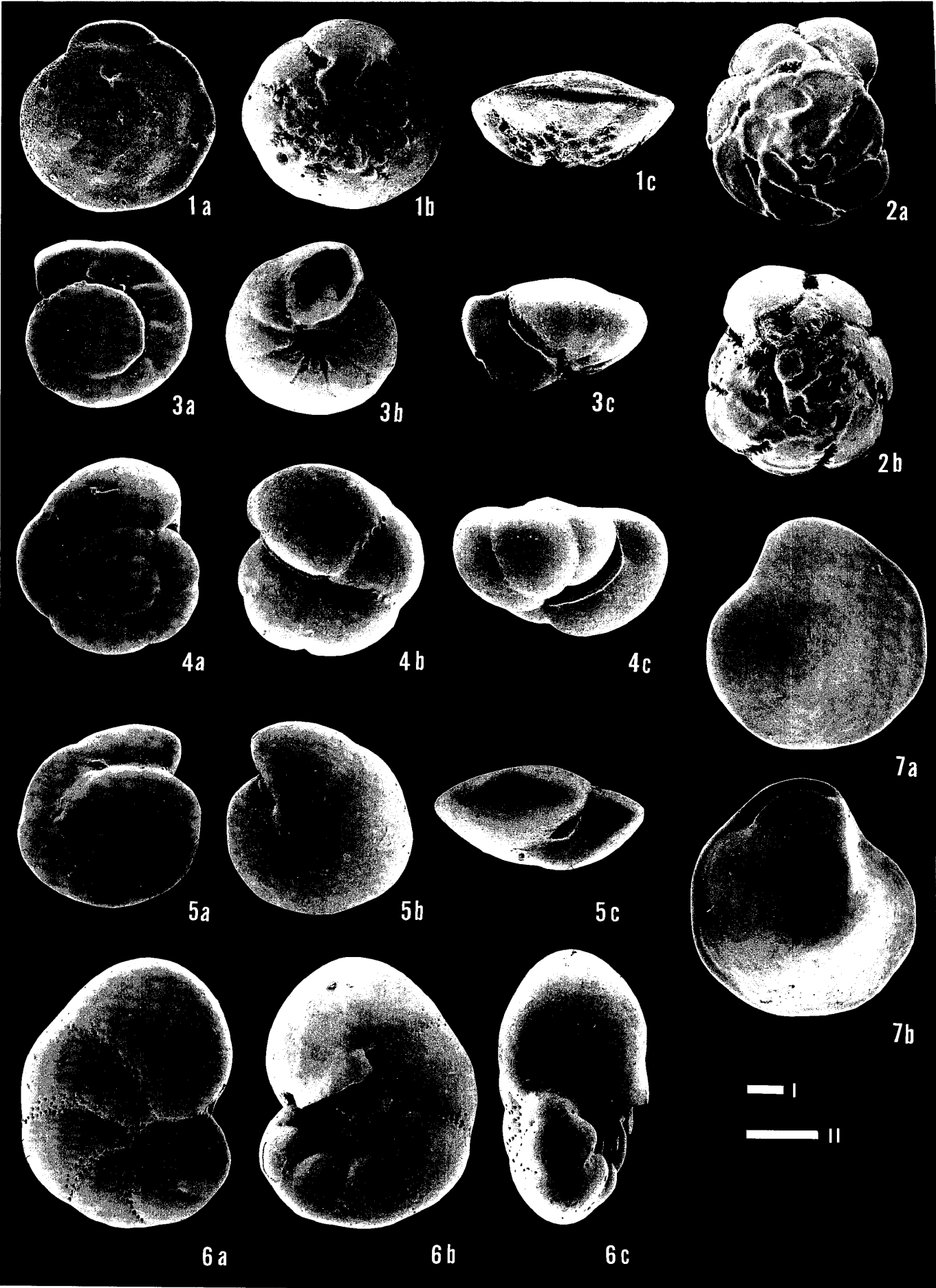
- Fig. 1. *Cibicides aknerianus* (d'Orbigny)
(IGPS 101107), Sample KT85-6-G4, Enshunada area, $\times 200$.
- Fig. 2. *Parrelloides bradyi* (Trauth)
(IGPS 101108), Sample KT85-6-G1, Enshunada area, $\times 200$.
- Fig. 3. *Cibicidoides mediocris* (Finlay)
(IGPS 101109), Sample KT-85-6-G6, Enshunada area, $\times 200$.
- Fig. 4. *Hanzawaia nipponica* Asano
(IGPS 101110), Sample KT85-6-G5, Enshunada area, $\times 200$.
- Fig. 5. *Hyalinea bathica* (Schröter)
(IGPS 101111), Sample KT85-6-G5, Enshunada area, $\times 300$.
- Fig. 6. *Anomalinoides globulosa* (Chapman and Parr)
(IGPS 101112), Sample KT85-6-G6, Enshunada area, $\times 200$.
- Fig. 7. *Pseudoparrella exigua* (Brady)
(IGPS 101113), Sample KT86-11-MP6, Nankai Trough, $\times 300$.

Scale bar : 0.1 mm ; I : 1, 2, 3, 4, 6 ; II : 5, 7.

Plate 21

- Fig. 1. *Ammonia ketienziensis* (Ishizaki)
(IGPS 101114), Sample KT85-6-G5, Enshunada area, $\times 200$.
- Fig. 2. *Pseudorotalia gaimardii* (d'Orbigny)
(IGPS 101115), Sample KT85-6-G6, Enshunada area, $\times 100$.
- Fig. 3. *Gyroidina cushmani* Boomgraat
(IGPS 101116), Sample KT-85-6-G5, Enshunada area, $\times 200$.
- Fig. 4. *Gyroidinoides nipponicus* (Ishizaki)
(IGPS 101117), Sample KT85-6-G4, Enshunada area, $\times 200$.
- Fig. 5. *Oridorsaris tener* (d'Orbigny)
(IGPS 101118), Sample KT85-6-G1, Enshunada area, $\times 200$.
- Fig. 6. *Valvulineria gunjii* Akimoto, n. sp.
Holotype (IGPS 101119), Sample KT86-11-MP8, Nankai Trough, $\times 200$.
- Fig. 7. *Hoeghundina elegans* (d'Orbigny)
(IGPS 101120), Sample KT86-11-MP6, Nanakai Trough, $\times 100$.

Scale bar : 0.1 mm ; I : 2, 7 ; II : 1, 3, 4, 5, 6.



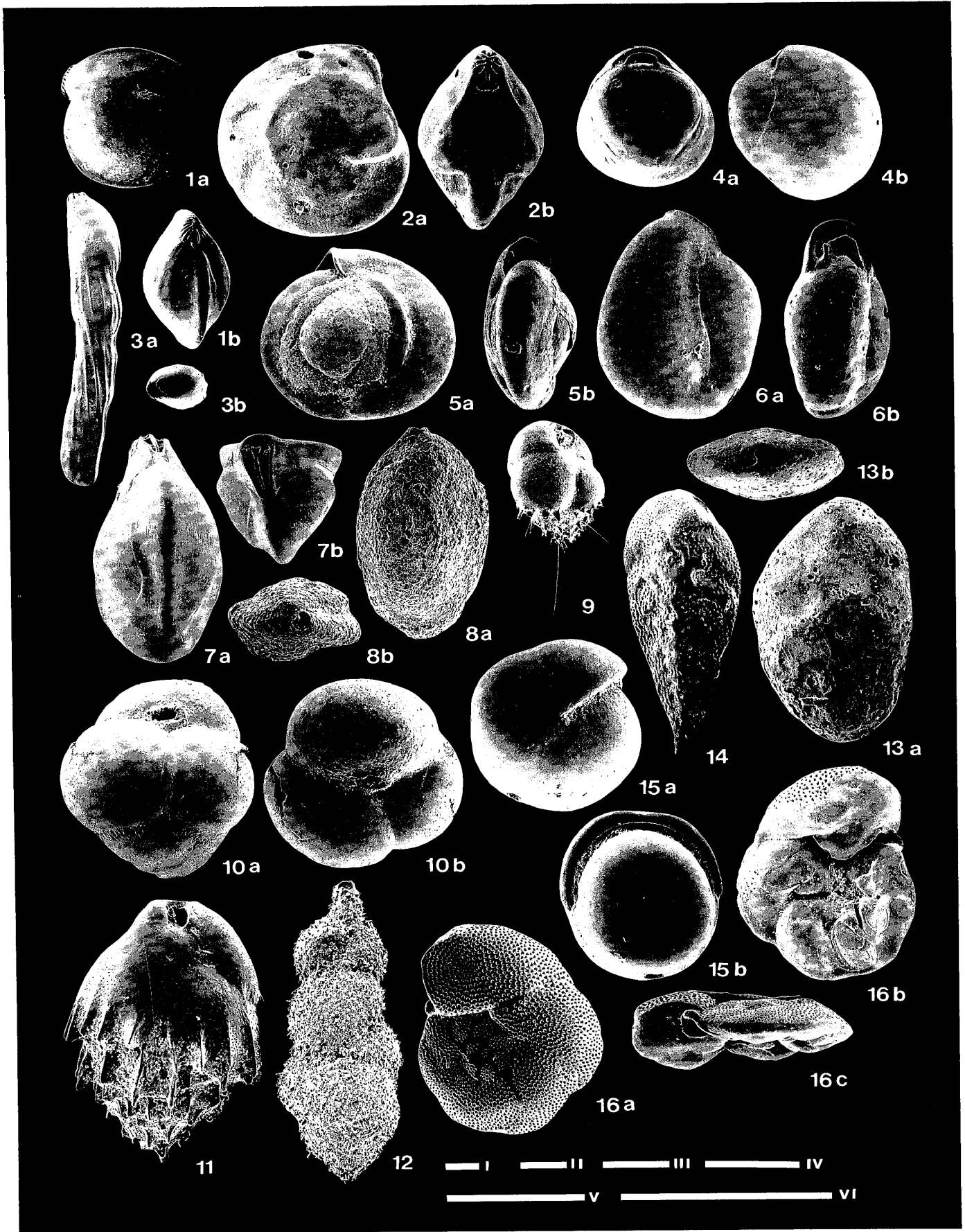


Plate 22

- Fig. 1. *Robulus orbicularis* d'Orbigny
(IGPS 101121), Sample GH80-4-85, Hachijojima area, $\times 70$.
- Fig. 2. *Cribrorbulina serpens* (Seguenza)
(IGPS 101122), Sample GH80-4-85, Hachijojima area, $\times 200$.
- Fig. 3. *Vaginulinopsis asanoi* Ishizaki
(IGPS 101123), Sample GH80-4-97, Hachijojima area, $\times 45$.
- Fig. 4. *Miliolinella inflata* LeRoy
(IGPS 101124), Sample GH80-4-85, Hachijojima area, $\times 150$.
- Fig. 5. *Miliolinella australis* (Parr)
(IGPS 101125), Sample GH80-4-85, Hachijojima area, $\times 200$.
- Fig. 6. *Miliolinella circularis* (Bornemann)
(IGPS 101126), Sample GH80-4-94, Hachijojima area, $\times 200$.
- Fig. 7. *Triloculina tricarinata* d'Orbigny
(IGPS 101127), Sample GH80-4-85, Hachijojima area, $\times 100$.
- Fig. 8. *Sigmoilopsis schlumbergeri* (Silvestri)
(IGPS 101128), Sample GH80-4-D370, Hachijojima area, $\times 100$.
- Fig. 9. *Bulimina aculeata* d'Orbigny
(IGPS 101129), Sample GH80-4-RC102, Hachijojima area, $\times 200$.
- Fig. 10. *Tosaia hanzawai* Takayanagi
(IGPS 101130), Sample GH80-4-D373, Hachijojima area, $\times 200$.
- Fig. 11. *Bulimina straiata* d'Orbigny
(IGPS 101131), Sample GH80-4-D370, Hachijojima area, $\times 200$.
- Fig. 12. *Uvigerina proboscidea* Schwager
(IGPS 101132), Sample GH80-4-D371, Hachijojima area, $\times 150$.
- Fig. 13. *Bolivina robusta* Brady
(IGPS 101133), Sample GH80-4-97, Hachijojima area, $\times 300$.
- Fig. 14. *Bolivina robusta* Brady forma A
(IGPS 101134), Sample GH80-4-D371, Hachijojima area, $\times 150$.
- Fig. 15. *Pullenia bulloides* (d'Orbigny)
(IGPS 101135), Sample GH80-4-D370, Hachijojima area, $\times 150$.
- Fig. 16. *Rosalina vilardeboana* d'Orbigny
(IGPS 101136), Sample GH80-4-85, Hachijojima area, $\times 100$.

Scale bar : 0.1 mm ; I : 7, 8, 16 ; II : 4, 12, 14, 15 ; III : 2, 5, 6, 9, 10, 11 ; IV : 13. 1.0 mm ; V : 3 ; VI : 1.

Plate 23

- Fig. 1. *Carpenteria monticularis* Carter
(IGPS 101137), Sample GH80-4-85, Hachijojima area, $\times 45$.
- Fig. 2. *Cibicidoides mediocris* (Finlay)
(IGPS 101138), Sample GH80-4-85, Hachijojima area, $\times 150$.
- Fig. 3. *Heterolepa subhaidingeri* (Parr)
(IGPS 101139), Sample GH80-4-85, Hachijojima area, $\times 100$.
- Fig. 4. *Cibicides praecinctus* (Karrer)
(IGPS 101140), Sample GH80-4-94, Hachijojima area, $\times 150$.
- Fig. 5. *Cibicides refulgens* Montfort
(IGPS 101141), Sample GH80-4-94, Hachijojima area, $\times 70$.
- Fig. 6. *Cibicides tenuimargo* (Brady)
(IGPS 101142), Sample GH80-4-94, Hachijojima area, $\times 45$.
- Fig. 7. *Cibicides wuellerstorfi* (Schwager)
(IGPS 101143), Sample GH80-4-D371, Hachijojima area, $\times 100$.
- Fig. 8. *Parrelloides bradyi* (Trauth)
(IGPS 101155), Sample GH80-4-D371, Hachijojima area, $\times 150$.
- Fig. 9. *Spirillina limbata* Brady
(IGPS 101144), Sample GH80-4-94, Hachijojima area, $\times 200$.
- Fig. 10. *Spirillina* sp.
(IGPS 101145), Sample GH80-4-97, Hachijojima area, $\times 150$.

Scale bar : 0.1 mm ; I : 3, 7 ; II : 2, 4, 8, 10 ; III : 9. 1.0 mm ; IV : 1, 6 : V : 5.



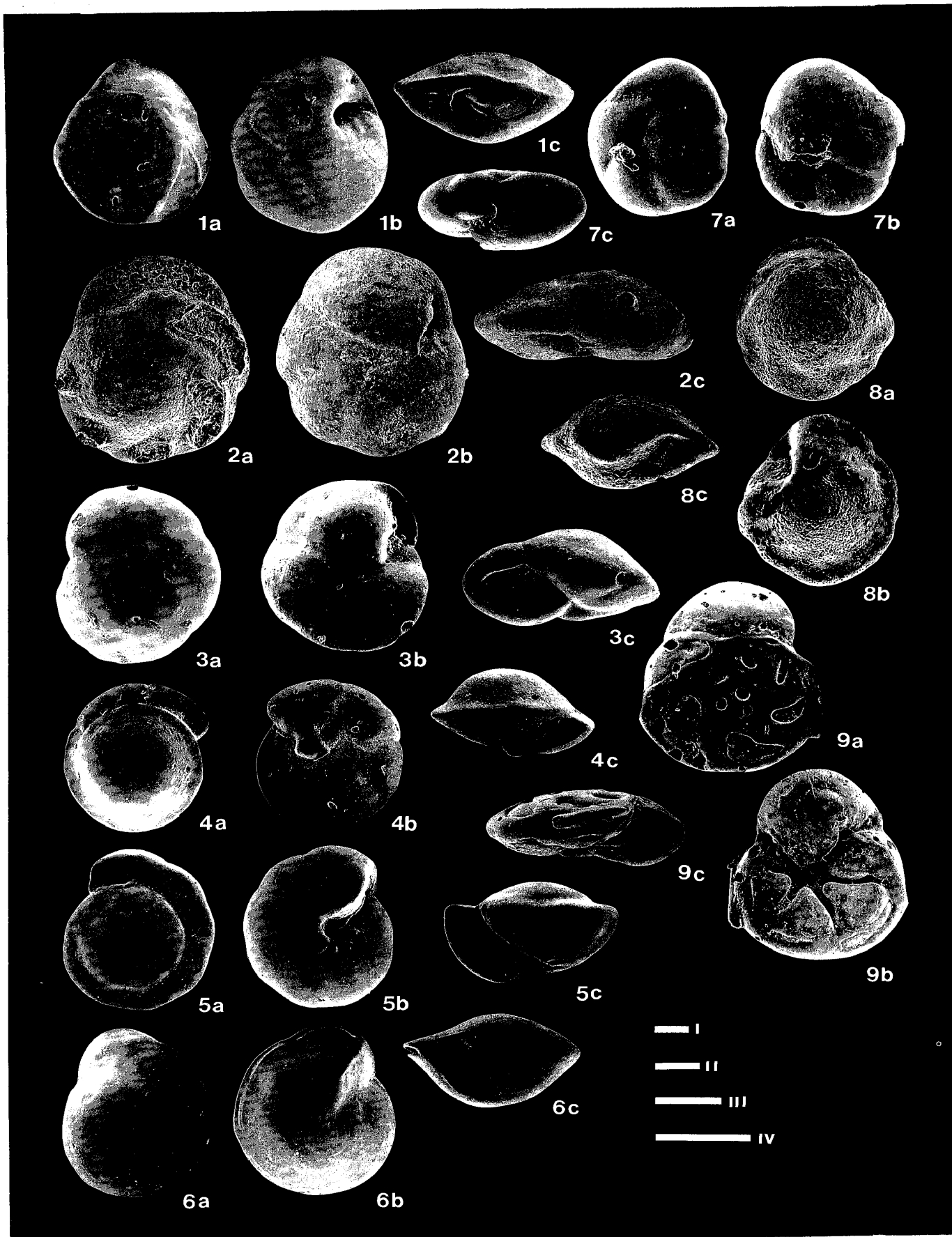


Plate 24

- Fig. 1. *Poroeponides cribrorephandus* Asano and Uchio
(IGPS 101146), Sample GH80-4-85, Hachijojima area, $\times 100$.
- Fig. 2. *Alabama tubulifera* (Heron-Allen and Earland)
(IGPS 101147), Sample GH80-4-85, Hachijojima area, $\times 150$.
- Fig. 3. *Pseudoparrella exigua* (Brady)
(IGPS 101148), Sample GH80-4-RC102, Hachijojima area, $\times 300$.
- Fig. 4. *Oridorsalis umbonatus* (Reuss)
(IGPS 101149), Sample GH80-4-D370, Hachijojima area, $\times 100$.
- Fig. 5. *Gyroidina soldanii* d'Orbigny
(IGPS 101150), Sample GH80-4-D370, Hachijojima area, $\times 100$.
- Fig. 6. *Hoeglundina elegans* (d'Orbigny)
(IGPS 101151), Sample GH80-4-96, Hachijojima area, $\times 150$.
- Fig. 7. *Quadriformina laevigata* (Phleger and Parker)
(IGPS 101152), Sample GH80-4-RC102, Hachijojima area, $\times 200$.
- Fig. 8. *Nuttallides umbonifera* (Cushman)
(IGPS 101153), Sample GH80-4-D376, Hachijojima area, $\times 200$.
- Fig. 9. *Mississippina concentrica* (Parker and Jones)
(IGPS 101154), Sample GH80-4-85, Hachijojima area, $\times 100$.

Scale bar: 0.1 mm; I: 1, 4, 5, 9; II: 2, 6; III: 7, 8; IV: 3.