

Fluorinated analogues of lipidic dialkynylcarbinol pharmacophores: synthesis and cytotoxicity in HCT116 cancer cells

Pauline Rullière^a, François Lizeaux^a, Etienne Joly^b, Stéphanie Ballereau^a, Hafida Gaspard^a, Valérie Maraval^c, Remi Chauvin^{c*}, Yves Génisson^{a*}

^a UMR CNRS 5068, LSPCMIB, Université de Toulouse, Université Paul Sabatier, 118 Route de Narbonne, 31062 Toulouse cedex 9, France.

^b UMR CNRS 5089, IPBS (Institut de Pharmacologie et de Biologie Structurale), 205 Route de Narbonne, 31077 Toulouse Cedex, France.

^c CNRS, LCC (Laboratoire de Chimie de Coordination), 205 route de Narbonne, BP 44099, 31077 Toulouse Cedex 4, France.

chauvin@lcc-toulouse.fr; genisson@chimie.ups-tlse.fr

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Lipidic alkynylcarbinols (LACs) have been identified as potential antitumor compounds, and a thorough understanding of their pharmacophoric environment is now required to elucidate their biological mode of action. In the dialkynylcarbinol (DAC) series, a specific study of the pharmacophore potential has been undertaken by focusing on the synthesis of three fluorinated derivatives followed by their biological evaluation. This work highlights the requirement of an electron-rich secondary carbinol center as a key structure for cytotoxicity in HCT116 cells.

Introduction

Originally identified as natural bioactive compounds extracted from marine sponges or constitute a class of potential antitumoral compounds [2]. Over the past few years, efforts have been devoted to the delineation of both optimal structural features [3] and cellular targets [4] of LAC derivatives exhibiting *in vitro* anti-proliferative activity against the HCT116 colon

tumor cell line. Noteworthy is a significant and systematic chirality effect, *i.e.* a dependence of the cytotoxicity on the absolute configuration of racemic LAC sample is thus expected to be roughly two times higher than that of the corresponding eutomer: results in the racemic series are however relevant prior to optimization

The IC₅₀ value of a

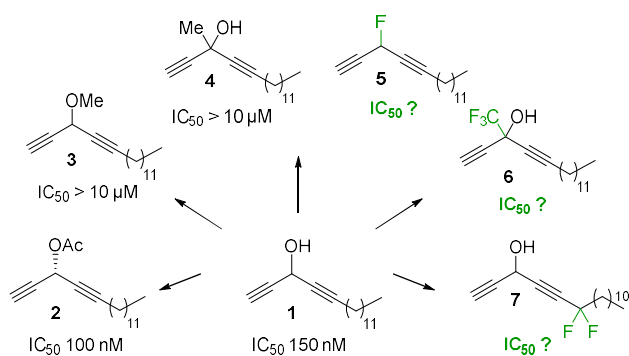


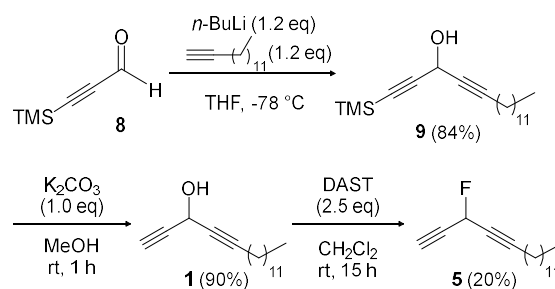
Figure 1. Insight into structure-activity relationships for the cytotoxicity of DACs through systematic pharmacophore modulation.

in the eutomeric series. In the search for the highest potency among various LAC pharmacophore models [3b], the dialkynylcarbinol (DAC) **1** was selected as a reference for its high bioactivity ($IC_{50} = 150$ nM), structural simplicity and ready synthetic accessibility (**Figure 1**) [5]. The impact on cytotoxicity of further alterations of the DAC warhead and lipidic environment of **1** was then explored (**Figure 1**). We reported that the acetylated analogue (*S*)-**2** displayed a cytotoxicity in HCT116 similar to that of (*S*)-**1**, probably due to *in cellulo* hydrolysis of the acetate moiety, releasing the free DAC [5]. In contrast, the *O*-methylated derivative **3** proved to be inactive ($IC_{50} > 10$ μ M) [6]. This dramatic effect of a slight modification (H \rightarrow Me) could be attributed to the stability of the methyl ether group in biological medium, persistently masking the alcohol moiety. Furthermore, the *C*-methylated tertiary alcohol **4** displayed the same lack of activity. Overall, the results for **3** and **4** may thus reflect the carbinol inability to be oxidized, which may be important for its

cytotoxicity. Further investigations were thus envisaged through the impoverishment of the DAC warhead by fluorination, the targeted fluorinated analogues being expected to provide a new panel to the systematic study of the DAC pharmacophore.

Results and discussion

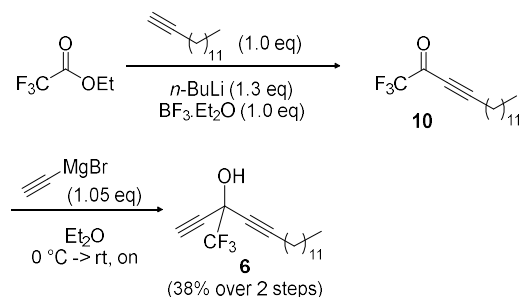
The racemic bis-propargylic fluoride **5**, deoxyfluorinated analogue of **1**, was readily obtained by direct treatment of **1** with diethylaminosulfur trifluoride (DAST, **Scheme 1**) [7].



Scheme 1. Synthesis of the deoxyfluorinated analogue **5** of the reference DAC **1**.

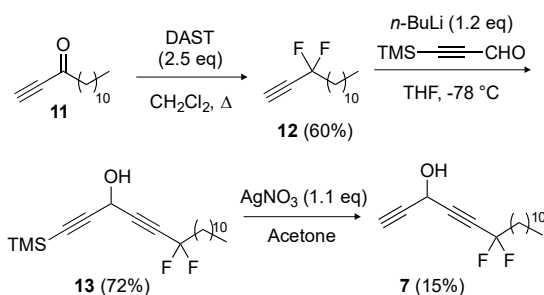
While full conversion of the alcohol was observed by TLC monitoring, **5** was isolated in low yield. This outcome can be attributed to the particularly high volatility of the hydrocarbon fluoride **5**, impeding efficient purification steps and drying procedures. The trifluoro analogue **6** of the tertiary alcohol **4** was secured by addition of ethynylmagnesium bromide onto the trifluoromethylated ynone **10** (**Scheme 2**) [8], itself obtained by reaction of a single equivalent of tetradecynyllithium on commercial ethyl trifluoroacetate in the presence of a

stoichiometric amount of boron trifluoride etherate [9].



Scheme 2. Synthesis of trifluoromethylated tertiary dialkynylcarbinol **6**.

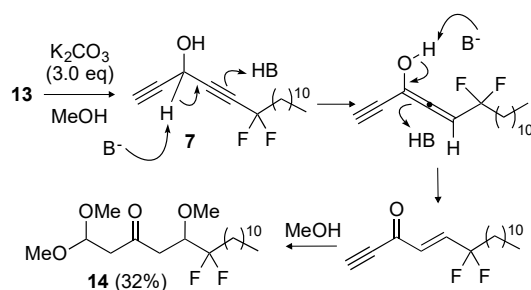
The difluorinated analogue **7** with a CF₂ group in place of the propargylic methylene group of **1** was obtained through a concise three-step sequence (**Scheme 3**). 3,3-Difluorotetradec-1-yne **12** was prepared by DAST-mediated deoxofluorination of the corresponding ynone **11** [10], and its lithium salt was in turn added to TMS-propynal to give **13** with 72% yield (**Scheme 3**). Desilylation was not observed even under fluorination conditions with an over-stoichiometric amount of DAST.



Scheme 3. Synthesis of the difluorinated dialkynylcarbinol **7**.

Removal of the TMS group, first attempted under standard basic conditions (K₂CO₃/MeOH), led to the formation of an unexpected compound. The latter was identified as the product resulting from the conjugate addition of three molecules of

MeOH onto the Michael acceptor formed by a base-induced isomerization of the difluorinated alkynylcarbinol **7** [11]. This structural assignment was notably based on several key data from mass spectrometry (*m/z* = 380), ¹H NMR (3 OCH₃ signals at 3.49, 3.40 and 3.39 ppm, absence of the characteristic terminal acetylenic proton) and ¹³C NMR (C=O signal at 205 ppm). Even though several isomers of this structure may be formed, under basic conditions, the structure **14** appears the most likely (**Scheme 4**).



Scheme 4. Base-induced isomerization of **7** and subsequent transformation into the triple Michael adduct **14** upon desilylation of **13**.

The final desilylation was smoothly accomplished using AgNO₃ (**Scheme 3**). The highly sensitive difluoro-DAC **7** was isolated with 15% yield after column chromatography. It gave characterization data in full agreement with the expected structure.

The antitumoral potential of the fluorinated DAC analogues **5-7** was then assessed by *in vitro* cell viability assays in cancer cells (**Table 1**). The HCT116 cell line was selected as a model. This representative human colon carcinoma cell line is frequently used in the biological evaluation of potential antitumor compounds. In addition,

cytotoxicity data against HCT116 allow direct comparison with our previous work [2], [3], [4].

Table 1. Cytotoxic activity of DAC analogues in HCT 116 cancer cells.

DAC derivatives	IC ₅₀ [μM]	DAC derivatives	IC ₅₀ [μM]
<i>rac-1</i>	0.15 ^{a,2a}	<i>rac-5</i>	>10 ^a
(<i>S</i>)- <i>2</i>	0.10 ^{a,5}	<i>rac-6</i>	>10 ^a
<i>rac-3</i>	>10 ^{a,6}	<i>rac-7</i>	>10 ^a
<i>rac-4</i>	>50 ^{b,6}		

^aCells were seeded in 96-well plates and were treated with concentrations ranging from 10 nM to 10 μM; after 48 or 72 h, the number of live cells was evaluated by a standard MTT test. ^bCells were seeded in 96-well plates and were treated with concentrations ranging from 10 nM to 50 μM; after 96 h, the number of live cells was evaluated by a WST test.

None of the fluorinated electron-impoverished DAC analogues **5**, **6** and **7** displayed any detectable activity at concentrations up to 10 μM (Table 1). These results indicate a correlation between the oxidability of DACs or analogues and their cytotoxicity (see Introduction and Figure 1): deoxyfluorination or proximal fluorination of the carbinol center of **1** indeed induces an increase of the oxidation potential in **5-7**. These results point to the requirement of an electron-rich carbinol center for inducing cytotoxicity against HCT116 cells.

Conclusions

The disclosed results highlight key features for cytotoxicity of DAC pharmacophores, *i.e.* the actual presence, secondary character and electron-richness of the carbinol center. Although the corresponding dialkynylketone was described as non-cytotoxic [6], the essential need for a readily oxidizable

carbinol pharmacophore provides a guideline for the design of DACs with improved cytotoxicity. While a prominent criterion for cytotoxic drug candidates is their selectivity towards tumor cells vs. normal cells, the very first criterion is their intrinsic activity. The disclosed fluorinated LACs being basically not cytotoxic, they serve as "negative guidelines" for the design of non-fluorinated candidates, for which this concern will be addressed in due course after optimization.

Experimental part

General.

All reagents were obtained from commercial suppliers and used without any further purification. Dichloromethane and tetrahydrofuran (THF) were obtained by filtration through a drying column on a filtration system. Thin-layer chromatography (TLC) analyses were performed on silica gel precoated plates (Merck 60 F254). Visualization of the developed chromatogram was performed by UV light (254 nm) and using 10 % phosphomolybdic acid in EtOH, or aqueous potassium permanganate (KMnO₄) stain. Flash column chromatography purifications were performed using flash silica gel (SDS 35-70 mm). Nuclear magnetic resonance spectra were recorded in CDCl₃ on a Bruker Advance 300 MHz spectrometer. Chemical shifts for ¹H NMR spectra are recorded in parts per million (ppm) with the solvent resonance as the reference

CDCl₃ (δ = 7.26 ppm). Data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet and br = broad), coupling constant in Hz and integration. Chemical shifts for ¹³C NMR spectra are recorded in parts per million using the central peak of CDCl₃ (δ = 77.16 ppm) as the reference. All ¹³C NMR spectra were obtained with complete proton decoupling. Chemical shifts for ¹⁹F NMR spectra are recorded in parts per million (ppm) with an internal standard resonance as the reference fluorobenzene (δ = -113.15 ppm). IR analysis were run on a Thermo-Nicolet Diamond ATR (4 cm⁻¹ of resolution, 16 scans) equipped with a DTGS detector. High-resolution mass spectrometry (HRMS) was performed on a Thermo-Finnigan MAT 95 XL instrument.

Experimental procedures and characterizations.

3-fluorohexadeca-1,4-diyne (5). DAST (0.1 mL, 0.8 mmol, 2.0 equiv.) was added to a solution of previously described [5] heptadeca-1,4-diyne-3-ol (100 mg, 0.4 mmol) in CH₂Cl₂ (27 mL) at -10 °C. The reaction mixture was warmed up to rt and was stirred overnight. Pentane and water (1:1) were added. The aqueous phase was extracted with pentane and the organic layer was separated. The combined organic layers were dried over MgSO₄ and concentrated under reduced pressure. The crude product was submitted to flash chromatography over silica gel (up to pentane/Et₂O, 9 : 1) to give compound **5** (20 mg, 20%) as a colorless oil: *R*_f = 0.61 (100% pentane). ¹H NMR (300 MHz, CDCl₃) δ 5.68

(dq, *J* = 46.9, 2.1 Hz, 1H), 2.74 (dd, *J* = 5.3, 2.3 Hz, 1H), 2.27 (qd, *J* = 7.0, 2.1 Hz, 2H), 1.6-1.45 (m, 2H), 1.42-1.22 (m, 18H), 0.95-0.80 (m, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 86.3, 83.3 (d, *J* = 252.0 Hz), 77.1, 72.3, 68.2, 52.3, 29.7, 29.6, 29.2, 29.0, 28.9, 28.6, 28.4, 18.8, 18.5. ¹⁹F NMR (282 MHz, CDCl₃) δ -160.4 (m). MS (DCI-NH₃ neg.): *m/z* 249 (M-1). FTIR (cm⁻¹) (neat): 3300, 2922, 2853, 2319, 2242, 2135, 1467, 1313, 1154, 999, 934.

3-(trifluoromethyl)heptadeca-1,4-diyne-3-ol

(6). An ethynylmagnesium bromide solution (1.88 mL, 1.05 equiv., 0.5 M in THF) was added to a solution of 1,1,1-trifluorohexadec-3-yn-2-one **10** (260 mg, 0.90 mmol) in THF (5 mL) under N₂ atmosphere at -78 °C. The mixture was stirred at rt overnight. A saturated aqueous NH₄Cl solution was added, the aqueous layer was extracted with Et₂O and the organic layer was separated. The combined organic layers were dried over MgSO₄ and concentrated under reduced pressure. The crude product was submitted to flash chromatography over silica gel (up to pentane/Et₂O, 8 : 2) to give trifluoromethylated dialkynylcarbinol **6** (108 mg, 38% over two steps) as a brown oil: *R*_f = 0.5 (pentane/Et₂O, 8 : 2). ¹H NMR (300 MHz, CDCl₃) δ 2.94 (s, 1H), 2.68 (s, 1H), 2.26 (t, *J* = 7.1 Hz, 2H), 1.63-1.53 (m, 2H), 1.43-1.27 (m, 18H), 0.92 (t, *J* = 6.7 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 122.0 (q, *J* = 285.0 Hz), 88.9, 74.4, 72.8, 68.1, 32.1, 29.8, 29.8, 29.7, 29.6, 29.5, 29.1, 28.9, 27.9, 22.8, 18.7. HRMS (DCI-

CH₄): calcd for C₁₈H₂₈F₃O [M+H]⁺: 317.2092 *m/z*, found: 317.2102 *m/z*. **FTIR** (cm⁻¹) (neat): 3408 (br), 3307, 2924, 2854, 2246, 2129, 1266, 1193, 1173, 1073, 998.

6,6-difluoroheptadeca-1,4-diyn-3-ol (7). 6,6-Difluoro-1-(trimethylsilyl)heptadeca-1,4-diyn-3-ol **13** (320 mg, 0.9 mmol) was stirred overnight with AgNO₃ (168 mg, 0.99 mmol, 1.1 equiv.) in wet acetone (5 mL). The reaction was quenched with water and Et₂O was added. The aqueous layer was extracted with Et₂O (3 x 4 mL) and the organic layer was separated. The combined organic layers were dried over MgSO₄ and concentrated under reduced pressure. The crude product was submitted to flash chromatography over SiO₂ (with a gradient elution up to pentane/Et₂O, 9 : 1) to give the desired product **7** (38 mg, 15%) as a brown oil: *R_f* = 0.5 (pentane/Et₂O 9:1). **¹H NMR** (300 MHz, CDCl₃) δ 5.22 – 5.16 (m, 1H), 2.62 (d, *J* = 2.4 Hz, 1H), 2.31 (d, *J* = 8.0 Hz, 1H), 2.14 – 1.91 (m, 2H), 1.62 – 1.45 (m, 2H), 1.28 (d, *J* = 12.2 Hz, 16H), 0.88 (t, *J* = 6.7 Hz, 3H). **¹³C NMR** (75 MHz, CDCl₃) δ 114.8 (t, *J* = 233.4 Hz), 83.1 (t, *J* = 6.9 Hz), 79.4 (t, *J* = 2.3 Hz), 77.8, 74.2, 51.9, 39.1 (t, *J* = 25.5 Hz), 32.1, 29.7, 29.6, 29.5, 29.4, 29.1, 22.8, 22.7 (t, *J* = 3.5 Hz), 14.3. **¹⁹F NMR** (282 MHz, CDCl₃) δ -83.99 (td, *J* = 15.1, 3.8 Hz). **HRMS** (DCI-CH₄): calcd for C₁₇H₂₇F₂O [M+H]⁺: 285.2030 *m/z*, found: 285.2027 *m/z*. **FTIR** (cm⁻¹) (neat): 3364 (br), 3310, 2925, 2855, 2262, 2126, 1031.

1,1,1-trifluorohexadec-3-yn-2-one (10). According to previously reported reaction conditions [9], to a stirred solution of 1-tetradecyne (1.73 mL, 7.04 mmol) in THF (11 mL) was added *n*-butyllithium (3.7 mL, 9.15 mmol, 2.5 M in hexanes) dropwise at -78 °C. The resulting solution was stirred at the same temperature for 30 min, at which time a solution of ethyl trifluoroacetate (0.84 mL, 7.04 mmol) and boron trifluoride diethyl etherate (0.87 mL, 7.04 mmol) in THF (2 mL) was introduced dropwise. After reaction completion (ca. 1 h), a saturated aqueous NH₄Cl solution was added. The aqueous layer was separated and extracted with Et₂O. The combined organic extracts were washed with brine, dried over MgSO₄, filtered and concentrated under reduced pressure to give the crude product, which was purified by flash chromatography on silica gel (up to pentane/Et₂O, 8 : 2) to afford desired trifluoromethylketone **10** (260 mg, 13%). Spectral data were comparable with those previously reported in the literature [9b] and are not described in detail.

Tetradec-1-yn-3-one (11). Dess-Martin periodinane reagent (202 mg, 0.48 mmol, 1 equiv.) was added to a solution of tetradec-1-yn-3-ol (100 mg, 0.48 mmol) in CH₂Cl₂ (2 mL) under N₂ atmosphere. The mixture was stirred at rt overnight. A saturated aqueous NaHCO₃ solution was added, the aqueous layer was extracted with CH₂Cl₂ and the organic layer was separated. The combined organic layers were

dried over MgSO₄ and concentrated under reduced pressure. The crude product was submitted to flash chromatography over silica gel (up to pentane/Et₂O, 8 : 2) to give ketone **11** (86 mg, 87%) as a yellow oil: *R_f* = 0.57 (pentane/Et₂O, 8 : 2). ¹H NMR (300 MHz, CDCl₃) δ 3.19 (s, 1H), 2.58 (t, *J* = 7.4 Hz, 1H), 1.67 (t, *J* = 7.2 Hz, 2H), 1.57 (s, 1H), 1.33-1.22 (m, 16H), 0.95 (t, *J* = 6.0 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 187.8, 81.6, 78.4, 45.6, 32.0, 29.7, 29.5, 29.4, 29.0, 23.9, 22.8, 14.3. FTIR (cm⁻¹) (neat): 3299, 3255, 2853, 2924, 2093, 1683, 1466.

3,3-difluorotetradec-1-yne (12). Tetradec-1-yn-3-one **11** (1.5 g, 7.2 mmol) and DAST (2.23 mL, 18 mmol, 2.5 equiv.) were heated neat at 56 °C and stirred under nitrogen atmosphere over 5 h. The crude product was submitted to flash chromatography over silica gel (up to pentane/Et₂O, 95 : 5) to give the difluorinated product **12** (1.0 g, 60%) as a yellow oil: *R_f* = 0.57 (pentane/Et₂O, 8 : 2). ¹H NMR (300 MHz, CDCl₃) δ 2.74 (t, *J* = 4.9 Hz, 1H), 2.10-1.94 (m, 2H), 1.60-1.50 (m, 2H), 1.39-1.22 (m, 16H), 0.93 (t, *J* = 5.4 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 114.6 (t, *J* = 232.0 Hz), 75.1 (t, *J* = 6.8 Hz), 39.2 (t, *J* = 25.4 Hz), 32.1, 29.9, 29.7, 29.6, 29.5, 29.1, 23.0, 14.3. ¹⁹F NMR (282 MHz, CDCl₃) δ -83.9 (td, *J* = 15.1 Hz, 5.0 Hz). FTIR (cm⁻¹) (neat): 3305, 2923, 2854, 2134, 2095, 1685, 1466.

6,6-difluoro-1-(trimethylsilyl)heptadeca-1,4-diyn-3-ol (13). *n*-Butyllithium solution (2.6 mL, 0.65 mmol, 1.2 equiv., 2.5 M in hexanes) was added to a solution of 3,3-difluorotetradec-1-yne

12 (125 mg, 0.54 mmol) in THF (15 mL) under N₂ atmosphere at -78 °C. After 45 min of stirring, 3-(trimethylsilyl) propionaldehyde (0.1 mL, 3.17 mmol, 5.8 equiv.) was added at -78 °C. Temperature and stirring were maintained during 30 min. A saturated aqueous NH₄Cl solution was added, the aqueous layer was extracted with Et₂O and the organic layer was separated. The combined organic layers were dried over MgSO₄ and concentrated under reduced pressure. The crude product was submitted to flash chromatography over silica gel (up to pentane/Et₂O, 9 : 1) to give dialkynylcarbinol **13** (140 mg, 72%) as a brown oil: *R_f* = 0.47 (pentane/Et₂O, 9 : 1). ¹H NMR (300 MHz, CDCl₃) δ 5.17 (dt, *J* = 7.5, 3.7 Hz, 1 H), 2.26 (d, *J* = 7.3 Hz, 1H), 2.12-1.96 (m, 2H), 1.60-1.49 (m, 2H), 1.37-1.22 (m, 16H), 0.94 (t, *J* = 13.4 Hz, 3H), 0.20 (s, 9H). ¹³C NMR (75 MHz, CDCl₃) δ 127.2, 117.8, 114.7, 111.6, 100.0, 91.2, 83.5 (t, *J* = 7.0 Hz), 77.3, 52.3, 39.1 (t, *J* = 25.5 Hz), 31.9, 29.6, 29.5, 29.4, 29.0, 23.3, 14.1, 0.0. ¹⁹F NMR (282 MHz, CDCl₃) δ -83.79 (td, *J* = 15.0 Hz, 3.6 Hz). FTIR (cm⁻¹) (neat): 3390 (br), 2926, 2856, 1467, 1252, 1043, 845, 761.

6,6-difluoro-1,1,5-trimethoxyheptadecan-3-one (14): 6,6-Difluoro-1-(trimethylsilyl)heptadeca-1,4-diyn-3-ol **13** (140 mg, 409 μmol) was stirred overnight with K₂CO₃ (169 mg, 1.2 mmol, 3 equiv.) and MeOH (2 mL). A saturated aqueous NH₄Cl solution was added, the aqueous layer was extracted with Et₂O (3 x 5 mL) and the organic layer was separated. The

combined organic layers were dried over MgSO₄ and concentrated under reduced pressure. The crude product was submitted to flash chromatography over SiO₂ (up to pentane/Et₂O, 9 : 1) to give the triple Michael adduct **14** (50 mg, 33%) as a brown oil: *R*_f = 0.5 (pentane/Et₂O 9:1). ¹H NMR (300 MHz, CDCl₃) δ 4.84 (t, *J* = 5.6 Hz, 1H), 4.05 – 3.92 (m, 1H), 3.49 (s, 3H), 3.40 (s, 3H), 3.39 (s, 3H), 2.81 (dd, *J* = 5.6, 2.0 Hz, 2H), 2.76 (d, *J* = 5.8 Hz, 2H), 2.07 – 1.72 (m, 2H), 1.59 – 1.47 (m, 2H), 1.33-1.19 (m, 16H), 0.97 (t, *J* = 6.7 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 204.5 (C=O), 125.7 (t, *J* = 245.9 Hz, CF₂), 101.4, 77.4 (dd, *J* = 30.4, 27.8 Hz), 60.0 (OMe), 54.2 (OMe), 53.8 (OMe), 47.4, 43.7 (dd, *J* = 3.8, 1.9 Hz), 32.7 (t, *J* = 24.1 Hz), 32.0, 29.7, 29.6, 29.6, 29.5, 29.5, 22.8, 21.4 (t, *J* = 4.3 Hz), 14.3. MS (DCI-NH₃ neg.): *m/z* 280 (M-1).

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References

[1] (a) Kuklev DV, Dembitsky VM. Epoxy acetylenic lipids: Their analogues and derivatives. *Prog Lipid Res.* 2014;56:67-91. (b) Kuklev DV, Domb AJ, Dembitsky VM. Bioactive acetylenic metabolites. *Phytomedicine.* 2013;20(13):1145-59. (c) Siddiq A, Dembitsky V. Acetylenic anticancer agents. *Anticancer Agents Med Chem.* 2008;8(2):132-170. (d) Shun ALKS, Tykwinski RR. Synthesis of Naturally Occurring Polyynes. *Angew Chem Int Ed.* 2006;45(7):1034-57. (e) Dembitsky VM.

Anticancer activity of natural and synthetic acetylenic lipids. *Lipids.* 2006;41(10):883-924.

[2] El Arfaoui D, Listunov D, Fabing I, Oukessou M, Frongia C, Lobjois V, Samson A, Ausseil F, Ben-Tama A, El Hadrami EM, Chauvin R, Génisson Y. Identification of Chiral Alkenyl- and Alkynylcarbinols as Pharmacophores for Potent Cytotoxicity. *ChemMedChem.* 2013;8(11):1779-86. (b) Hallock YF, Cardellina JH, Balaschak MS, Alexander MR, Prather TR, Shoemaker RH, Boyd MR. Antitumor Activity and Stereochemistry of Acetylenic Alcohols from the Sponge *Cribrorchalina vasculum*. *J Nat Prod.* 1995;58(12):1801-7. (c) Aiello A, Fattorusso E, Menna M, Pansini M. Further Bioactive Acetylenic Compounds from the Caribbean Sponge *Cribrorchalina vasculum*. *J Nat Prod.* 1992;55(9):1275-80. (d) Gunasekera SP, Faircloth GT. New acetylenic alcohols from the sponge *Cribrorchalina vasculum*. *J Org Chem.* 1990;55(25):6223-5.

[3] (a) Listunov D, Maraval V, Chauvin R, Génisson Y. Chiral alkynylcarbinols from marine sponges: asymmetric synthesis and biological relevance. *Nat Prod Rep.* 2015;32(1):49-75. (b) Listunov D, Joly E, Rullière P, Gaspard H, Bernardes-Génisson V, Génisson Y, Maraval V, Chauvin R. From natural to artificial antitumor lipidic alkynylcarbinols: asymmetric synthesis, enzymatic resolution and refined SARs. *Synthesis.* 2018;50(16):3114-30.

[4] Listunov D, Mazères S, Volovenko Y, Joly E, Génisson Y, Maraval V, Chauvin R. Fluorophore-tagged pharmacophores for antitumor cytotoxicity: Modified chiral lipidic dialkynylcarbinols for cell imaging. *Bioorg Med Chem Lett.* 2015;25(20):4652-6.

[5] Bourkhis M, Gaspard H, Rullière P, de Almeida DKC, Listunov D, Joly E, Abderrahim R, de Mattos MC, de Oliveira MCF, Maraval V, Chauvin R, Génisson Y. Skeletal Optimization of Cytotoxic Lipidic Dialkynylcarbinols. *ChemMedChem.* 2018;13(11):1124-30.

[6] Listunov D, Billot C, Joly E, Fabing I, Volovenko Y, Génisson Y, Maraval V, Chauvin R. Extended structural

modulation of bio-inspired chiral lipidic alkynylcarbinols as antitumor pharmacophores. *Tetrahedron*. 2015;71(41):7920-30.

[7] (a) Madiot V, Lesot P, Grée D, Courtieu J, Grée R. Highly enantioselective propargylic monofluorination established by carbon-13 and fluorine-19 NMR in chiral liquid crystals. *Chem Commun*. 2000;(2):169-70. (b) Prakesch M, Kerouredan E, Grée D, Grée R, DeChancie J, Houk KN. The propargylic route as efficient entry to monofluoro and gem-difluoro compounds: mechanistic considerations. *J Fluor Chem*. 2004;125(4):537-41. (c) Fauq AH, Singh RP, Meshri DT. Diethylaminosulfurtrifluoride. *e-EROS Encyclopedia of Reagents for Organic Synthesis*. 2004;1 :1-14.

[8] (a) Rives A, Baglai I, Barthes C, Maraval V, Saffon-Merceron N, Saquet A, Voitenko Z, Volovenko Y, Chauvin R. Carbo-cyclohexadienes vs. carbo-benzenes: structure and conjugative properties. *Chem Sci*. 2015;6(2):1139-49. (b) Maraval V, Leroyer L, Harano A, Barthes C, Saquet A, Duhayon C, Shinmyozu T, Chauvin R. 1,4-Dialkynylbutatrienes: Synthesis, Stability, and Perspectives in the Chemistry of carbo-Benzenes. *Chem Eur J*. 2011;17(18):5086-100.

[9] (a) Hsieh M-T, Kuo S-C, Lin H-C. Solvent- and Transition Metal Catalyst-Dependent Regioselectivity in the [3+2] Cyclocondensation of Trifluoromethyl- α,β -ynones with Hydrazines: Switchable Access to 3- and 5-Trifluoromethylpyrazoles. *Adv Synth Catal*. 2015;357(4):683-9. (b) Linderman RJ, Lonikar MS. Addition of organocuprates to acetylenic di- and trifluoromethyl ketones. Regiospecific synthesis of β,β -disubstituted unsaturated fluoro ketones. *J Org Chem*. 1988;53(26):6013-22.

[10] Grée D, Grée R. Effect of fluorine or oxygen atom(s) in propargylic position on the reactivity in click chemistry. *Tetrahedron Lett*. 2010;51(17):2218-21.

[11] El Dine A N, Khalaf A, Grée D, Tasseau O, Fares F, Jaber N, Lesot P, Hachem A, Grée R. Synthesis of enones, pyrazolines and pyrrolines with gem-difluoroalkyl side chains. *Beilstein J Org Chem*. 2013;9:1943-8.