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# Microsatellite genotyping of old Slovenian grapevine varieties (*Vitis vinifera* L.) of the Primorje (coastal) winegrowing region

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### Summary

Thirty-three grapevine varieties from Slovenia were genotyped at 21 microsatellite loci in order accurately to identify varieties and to evaluate their synonyms and homonyms, including varieties cultivated in neighbouring countries. Among Slovenian varieties some previously assumed synonyms were confirmed and some new ones were discovered: 'Poljšakica Drnovk' = 'Istrska Malvazija', 'Pikolit Italy' = 'Pikolit Vienna', 'Vitovska grganja' = 'Racuk' and 'Prosecco' = 'Glera' = 'Števerjana'. Types of Zelen ('Zelen Pokov', 'Zelen 66' and 'Zelen 2.4') were distinct at several microsatellite loci so can only be considered to be homonyms. Two 'Picolit' types were considered to be 'true-to-type' on the basis of comparison with 'Picolit' clones from Italy. Synonymy between 'Heunisch' and 'Belina' was not confirmed in our study, since 'Belina Pleterje' differentiated from 'Heunisch weiss' at 13 out of 19 loci. Comparison of 'Vitovska grganja' from Slovenia with 'Vitouska' from Italy also showed dissimilarities at the majority of the analysed loci.

K e y words: *Vitis vinifera*, microsatellite, SSR, genotyping.

## Introduction

Varied climatic and geological conditions have contributed to the great diversity of Vitis vinifera L. varieties in Slovenia. Around 50 well-known and rare varieties exist in Slovenia today, the majority of them being listed in the International List of Vine Varieties and Their Synonyms (OIV 2005). Although morphological and agronomical descriptions of the majority of autochthonous varieties exist, this is mostly unexplored plant material, and some of these varieties survived only in less productive vineyards or in germplasm collections. In 1980 we started systematically to collect our old and/or neglected varieties and set up four grapevine collections which include old varieties and clonal candidates. One of the problems in the management of these germplasm collections is the use of synonymic and homonymic designations, which are not completely reliable mainly because of inadequate documentation and poor preservation of historical facts related to grape growing and trade. The identification of plant material by ampelographic methods often results in misinterpretations (DETT- WEILLER 1993). In contrast, DNA-based markers are independent of environmental factors and are therefore more reliable for variety identification (Botta *et al.* 1995). In the last decade, more than 60 SSR (Short Sequence Repeat) primers from the genomic libraries of *Vitis vinifera* L. have been developed and used for identification purposes (Thomas and Scott 1993, Bowers *et al.* 1996, 1999, Sefc *et al.* 1999, Lefort *et al.* 2001).

The aim of our work was to genotype old Slovenian grapevine varieties using 21 SSR markers in order to examine their synonyms and homonyms, also in comparison with varieties cultivated in neighbouring countries. Additionally, genetic variability was assessed among analysed varieties.

#### **Material and Methods**

Plant material: The study was carried out on 32 old Slovenian grapevine varieties plus 'Chardonnay', 'Pinot noir', 'Cabernet-Sauvignon', 'Sultanine', 'Touriga Nacional' and 'Barbera' as reference varieties and *Vitis rupestris* L. as an out-group variety (Table). The vines are grown in grapevine germplasm collections in the Primorje winegrowing region (collection vineyards: Nova Gorica, Vipava and Goriška Brda).

D N A isolation: DNA was extracted from young leaves of shoot tips of individual vines using the modified CTAB method described by Kump and Javornik (1996).

Microsatellite analysis: The old Slovenian grapevines, the reference variety 'Chardonnay' and Vitis rupestris L. were genotyped at 21 microsatellite loci: VVS2 (THOMAS and SCOTT 1993); VVMD5, VVMD6, VVMD7, VVMD8 (Bowers et al. 1996); VVMD17, VVMD24, VVMD25, VVMD27, VVMD31, VVMD32, VVMD36 (Bowers et al. 1999); ssrVrZAG21, ssrVr-ZAG47, ssrVrZAG62, ssrVrZAG67, ssrVrZAG79, ssrVr-ZAG83 and ssrVrZAG112 (Sefc et al. 1999); ssrVvUch11 and ssrVvUch29 (Lefort et al. 2001). Five other reference varieties 'Pinot noir', 'Cabernet-Sauvignon', 'Sultanine', 'Touriga Nacional' and 'Barbera' were genotyped at 8 SSR loci (Table). The choice of these loci made the results comparable to the grapevine genotyping results of neighbouring countries and allows the integration of the results into the European grapevine database. The high number of SSR loci was useful in comparing allelic profiles of synonymic varieties. The data for comparison were obtained from dif-

T a b l e Grapevine genotypes at twenty-one SSR loci represented by allele sizes in base pairs (bp)

Вагьега		200	244 260		241 257 186 190	251 271	223 223 249 253	131
IsnoiseM aginuoT		194	246 2		251 2 257 2 182 1	239 2		141 1
Sultanine		188	248 2		241 2 251 2 182 1	249 2		143
Cabernet-Sauvignon		188 1	248 248 2		241 2 251 2 176 1	239 2		137
Pinot toniq		188 1	240 2246 2		241 2 251 2 186 1 190 1	239 2		135 1
Chardonnay	6 6 6 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8			08 5 5 6 8 6 5 6 5			232 2 236 2 236 2 208 2 239 2 243 2 146	
istrska Malvazija							220 238 238 238 208 206 239 239 239 239 239 239 239 239 239 239	
Prosecco	' ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' '						224 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2	
Rdeča žlahtnina							226 234 2 234 2 206 2 206 2 247 2 247 2 142 1 14	
Vitis rupestris							234 2 234 2 204 1 214 2 257 2 257 2 261 2 170 1	
Pikolit Italy							230 230 236 2236 2208 2208 2247 2247 2247 2247 2247 2247 2247 224	
Pikolit Vienna							230 2 236 2 198 1 198 2 208 2 247 2 247 2 140 1 166 1	
Klarnea	' ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' '						224 2 226 2 206 1 206 2 239 2 249 2 249 2 249 2 2 249 2 2 249 2 2 2 2	
Števerjana							224 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2	
Rečigia							226 232 232 232 232 240 249 249 249 249 249 249 249 249 249 249	
Zunek							224 22 238 238 238 2206 206 206 207 2247 2247 2247 2247 2247 2247 2247	
Cividin							236 2 236 2 206 1 208 2 247 2 247 2 261 2 142 1	
Касик							224 238 228 229 239 2239 2255 225 225 225 225 225 225 225 225 2	
Volovnik							224 22 238 228 208 19 208 19 224 224 224 224 224 224 224 224 224 22	
Dolga Petija							232 232 232 232 232 208 208 208 239 2249 2249 2249 2249 2249 2249 2249	
Delge Betlie							224 232 233 233 233 233 239 239 239 239 247 247 247 247 247 247 247 247 247 247	
Vrtovka							224 22 232 232 232 2206 18 207 2247 2249 2449 244 142 142 142 144 144 144 144 144 144	
Danijela							226 22 236 236 236 2008 2008 2008 2009 2009 2009 2009 2009	
Pergolin							224 23 224 23 206 20 208 20 239 24 243 243 243 243 243 243 243 243 243 243	
kinsgaD.sakvotiV							224 2238 2238 2238 2239 2339 2339 2339 2339	
Bratana Grannia Grannia								
							234 234 234 234 208 208 208 208 247 247 247 247 247 247 247 247 247 247	
Briška Glera	74 242 72 262 90 190 96 200 8 158 8 172						224 244 244 244 26 26 26 26 27 27 27 27 27 27 27 27 27 27 27 27 27	
Bela Glera	4 234 2 262 0 200 6 206 8 158 8 158						224 8 244 8 188 8 188 206 9 239 2 239 2 142 1 154	
Planika							0 224 8 238 8 208 6 208 6 209 9 239 9 253 2 142	
Poljšakica Drnovk	4 234 4 240 4 194 4 202 8 158 0 158			8 248 8 248 1 215 9 297 2 220 2 220 6 208				3 141
Poljšakica Lože	4 234 2 234 6 204 6 214 6 214 0 158 0 160						8 238 8 188 8 188 8 208 9 239 9 247 0 154	
Zelenika Zelenika	4 234 2 242 4 206 6 206 8 160 0 160			248 27 248 39 297 2 220 8 208				1 131
4.2 nələX	2 234 2 242 0 204 6 206 2 158 2 160			25 262 262 297 309 297 0 212 0 220 2 208			232 88 208 80 208 66 208 7 2 247 7 2 249 140	
Zelen Pokov	0 242 8 262 0 190 6 206 8 162 2 172		6 250 8 252 8 190 8 200 7 242			2 216 2 216 3 239 1 271 2 250 2 252		
99 nələZ	, , , , , , , , ,			2 246 2 246 3 211 9 211 2 220 0 220 8 208				
sləniq							5 232 8 208 8 208 6 208 7 247 7 249 7 249 6 142 6 142	
gnilziA idžka	0 230 2 234 0 190 6 206 8 164 0 168						224 236 88 198 8 206 5 247 7 257 0 142 0 146	
Belina Pleterje							5 232 6 244 8 188 6 208 6 208 9 247 0 140 2 140	
Kebula	240 240 206 206 206 164 164	188 200 128 160	252 252 188 188	248 285 220 220 212	214 245 251 186 186	203 204 271 242 252	226 226 198 206 243 249 140	145
Name	ssrvrZAG112 ssrvrZAG112 ssrvrZAG21 ssrvrZAG21 ssrvrZAG47	ZAG62 ZAG62 SSIVIZAG67 SSIVIZAG67	ZAG79 ZAG79 SSrVrZAG83 SSrVrZAG83	ssrvvUch 11 ssrvvUch 29 ssrvvUch 29 vVMD17 vVMD17	VVMD24 VVMD25 VVMD27 VVMD27	VVMD31 VVMD31 VVMD32 VVMD32 VVMD36 VVMD36	VVMDS VVMDS VVMD6 VVMD7 VVMD8 VVMD8 VVMD8	VVS2 VVS2

ferent publications, in which reference varieties were not always used, but normalization and comparison was possible on the basis of difference between two alleles.

PCR conditions: In a total volume of 10  $\mu$ l the PCR reaction mixture contained 20 ng of genomic DNA, 1 x PCR buffer (Fermentas), 0.2 mM of each dNTP's (Sigma), 2 mM MgCl<sub>2</sub> (Fermentas), 0.5  $\mu$ M of each primer and 0.25 U of *Taq* DNA polymerase (Fermentas). One of the primers for each locus was labelled with fluorescent Cy-5 dye for fluorescent detection (IDT lnc., BioSciences). The amplification of microsatellite loci was performed in a Whatman Biometra T-Gradient thermocycler with the following steps: hot start for 5 minutes at 95 °C; 26-40 cycles of denaturation at 94 °C for 30-45 s, annealing at 50-58 °C for 30-45 s and an extension step at 72 °C for 90 s. Reactions were completed by incubating at 72°C for 8 min.

After PCR optimization and amplification of individual loci, the amplification products were separated on 6 % polyacrylamide, 7 M urea and 1 x TBE gels, running in 0.5 x TBE buffer on an ALFexpress DNA automated sequencer (GE Healthcare). The allele sizes were analyzed with AlleleLocator 1.03 software (GE Healthcare). Alleles were precisely sized against an ALFexpress sizer 50-500 (GE Healthcare) and by internal DNA standards of different sizes amplified from plasmid.

Statistical analysis: Expected heterozygosity (He; Nei 1978), observed heterozygosity (Ho) and probability of identity (PI; PAETKAU *et al.* 1995) were calculated using Identity 1.0 software (Wagner and Sefc 1999). This program was also used to detect identical genotypes. Genetic distances between varieties were calculated in Microsat (Minch 1997) as 1- proportion of shared alleles.

# **Results and Discussion**

Allele sizes for each accession are reported in the table. A total of 169 alleles were detected, with an average of 8 alleles per locus. Thirty-seven alleles from 18 loci were variety specific, but 55 % of these alleles were specific for *Vitis rupestris* L. The average expected heterozygosity (0.779) was slightly higher than observed heterozygosity (0.767). This may be due to the ancient common origin of most accessions, which had not been subjected to a high degree of human selection, which acts against homozygosity (Sefc *et al.* 2000). The product of the PI values across all loci was 2.33x10<sup>-18</sup>, thus demonstrating the high distinguishing capacity of these markers. The average similarity of all varieties is 34 % of shared alleles, which is close to the average similarity observed for mid-European cultivars (40 %, Sefc *et al.* 1998).

Groups of cultivars with similar names or identical allelic profiles were investigated in order to assess their relationship. The variety 'Picolit' is represented in Slovenia by two distinct, morphologically different types 'Picolit Italia' and 'Picolit Wienna'. Microsatellite markers revealed no differences at 21 loci. Both our samples of 'Picolit' variety were further compared with 'Picolit' clones from Italy (Zulin et al. 2005), revealing the same allelic profiles at

all 7 SSR loci and they can therefore be considered to be 'true-to-type' 'Picolit'.

The synonymy found between 'Vitovska grganja' and 'Racuk' could not yet be confirmed, despite obtaining identical allelic patterns, because an accurate morphological characterization of 'Racuk' is still lacking. 'Vitovska grganja' is an old grape variety cultivated in Slovenia in winegrowing districts of the Vipava valley and the Karst and also in the north-east part of Italy, where it was an important variety in the past. The allelic profiles of our 'Vitovska grganja' were further compared with the Italian variety 'Vitouska', recently published by Crespan *et al.* (2007). Dissimilarity was revealed at 14 out of 16 loci, but the two varieties share at least one allele per locus at 15 loci, suggesting that they might be in a parent/progeny relationship.

Varieties 'Prosecco, 'Briška Glera' and 'Števerjana' form another group of synonyms. 'Prosecco' and 'Glera' had already been shown to be synonyms on the basis of morphological descriptors and isoenzyme analysis (Rose 1995), while 'Števerjana' has not previously been considered to be their synonym. The comparison of 16 SSR loci of our 'Prosecco' with Italian 'Prosecco tondo', which was recently analysed by Crespan *et al.* (2007), revealed no differences. On the basis of this comparison, it can be said that 'Prosecco' = 'Prosecco tondo' = 'Glera' but, according to Crespan *et al.* (2007), the synonym 'Glera' is mainly applied to 'Prosecco lungo' and less frequently to 'Prosecco tondo'.

Comparison between the two Gleras ('Briška Glera' and 'Bela Glera') included in our analysis revealed differences at 16 out of 21 loci, so they are considered homonyms. They also differ in the shape and compactness of the grape cluster, as well as in the white hairiness of the lower part of the leaf, which is expressed only in 'Bela Glera' ('White Glera').

Two varieties, 'Poljšakica Drnovk' and 'Poljšakica Lože', which were expected to have the same genetic profile, were different at many analysed microsatellite loci. Moreover, 'Poljšakica Drnovk' resulted in the same allelic pattern as 'Istrska Malvazija'. It therefore seems that the wrong name has been assigned to this variety in the Goriška Brda collection vineyard, where it is cultivated.

The synonymy of 'Heunisch' = 'Ranfol' = 'Belina', which was first mentioned by Goethe (1887), was also analysed. Comparing 19 SSR loci of 'Belina Pleterje' with 'Heunisch weiss' (REGNER et al. 2000), discrepancies were found at 13 loci, but the two varieties share one common allele at all 13 loci. Goethe (1887) described a range of varieties named Belina, differing in the pilose and whiteness of the underside of the leaf. Furthermore, the variety 'Belina Pleterje' was compared to the synonymic variety 'Ranfol Bijeli' from Croatia using microsatellite data of MALETIĆ et al. (1999). Comparison of 8 SSR loci resulted in the same allelic profiles, except at VVMD7, where a triallelic profile was observed in 'Belina Pleterje' (234, 246, 248) instead of the usual diallelic profile observed in the Croatian 'Ranfol Bijeli' (236, 246). These triallelic genotypes may be periclinal chimeras, in which two cell layers,

L1 and L2, of the plant meristem are genetically different (Hocquigny *et al.* 2004). Mutations in relationship to locus VVMD7 have also been previously reported (IBANEZ *et al.* 2000; Hocquigny *et al.* 2004).

The SSR profiles of different types of Zelen ('Zelen Pokov', 'Zelen 66' and 'Zelen 2.4') were compared. They showed differences at several microsatellite loci and it can thus be concluded that Zelen varieties from the Primorje winegrowing region are a heterogeneous group consisting of several genotypes.

Among 32 Slovenian grapevine accessions genotyped at the 21 microsatellite markers, 23 displayed different allelic profiles. In pairwise comparison, the highest genetic dissimilarity (86 %) was found between 'Rebula' – 'Volovnik', 'Dolga Petlja' – 'Guštana' and 'Klarnca' – 'Pikolit'. Other studied varieties also showed great differences on the basis of the microsatellie markers used, so it can be concluded that Slovenia has a diverse gene pool of *Vitis vinifera* L. varieties, which was also confirmed by their morphological descriptions (Tomažič and Korošec-Koruza, 1997).

In conclusion, microsatellites are very powerful means of identifying synonyms in germplasm collections, thereby allowing the removal of duplicates. The 32 varieties investigated in this study are only a small selection of the varieties grown in Slovenia. However, within this limited selection, we have begun the evaluation of rare and neglected grapevine varieties. The genetic characterization of a large number of varieties will contribute to improved organization of grapevine collections and the possibility of identifying and exchanging true-to-type material. This work will allow the integration of the obtained allele data into the European grapevine genetic resource database defined by the European project GrapeGen06.

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