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## Aspects of systematics, morphology, life history and feeding of western Atlantic sciaenidae (pisces: perciformes)

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LIFE HISTORY AND FEEDING OF WESTERN  
ATLANTIC SCIAENIDAE (PISCES:PERCIFORMES).

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ASPECTS OF SYSTEMATICS, MORPHOLOGY,  
LIFE HISTORY AND FEEDING OF WESTERN  
ATLANTIC SCIAENIDAE (PISCES:PERCIFORMES)

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A Dissertation

Presented to

The Faculty of the School of Marine Science  
The College of William and Mary in Virginia

In Partial Fulfillment

Of the Requirements for the Degree of  
Doctor of Philosophy

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by

Labbish Ning Chao

1976

APPROVAL SHEET

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the requirements for the degree of

Doctor of Philosophy

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## ABSTRACT

### Part I. A Basis for Classifying Western Atlantic Sciaenidae (Pisces; Perciformes)

The sciaenids of the western Atlantic consist of 21 genera and 56 species placed in 11 supra-generic groups. They are the *Cynoscion*, *Larimus*, *Lonchiurus*, *Menticirrhus*, *Micropogonias*, *Nebris*, *Pogonias*, *Sciaena*, *Sciaenops*, *Stellifer* and *Umbrina* groups. The phylogenetic relationships of all western Atlantic genera are assessed on the basis of swimbladder, otolith (sagitta and lapillus) and external morphology. The *Stellifer* group differs from all other western Atlantic sciaenids by having a two-chambered swimbladder and an enlarged lapillus. Phylogenetic and ontogenetic trends of the swimbladder is proposed as from simple carrot-shaped to a more complicated structure with anterior diverticula and horns, to a very complicated lateral diverticula system. The sagitta usually is oval or elongated in shape. The thickness and the impression of the sulcus on the inner surface of the inner surface of the sagitta are diagnostic among genera. External morphology, especially related to the feeding habits, habitat, or both are adaptive, but a trend is evident that the closely related genera often have similar body shape, mouth position and other external features. The species of the genus *Stellifer* are unique by having many diverse mouth positions or feeding habits. The synopsis section of the paper includes a diagnosis, a primary synonymy and types of nominal species for each taxonomic category. Four genera and 23 nominal species of New World freshwater sciaenids are also included. A tested field key to species and genera of all western Atlantic sciaenids is included. The range of distribution and some meristic counts are listed under each species. This paper serves as a basis for further revision of western Atlantic sciaenids.

## ABSTRACT

Part II. Life history, feeding habits and functional morphology of juvenile sciaenid fishes in the York River estuary, Virginia.

Four abundant sciaenid fishes, *Cynoscion regalis*, *Bairdiella chrysoura*, *Micropogonias undulatus* and *Leiostomus xanthurus* use the York River, Virginia as a nursery ground and as an adult seasonal feeding ground. In addition, six species of sciaenids, *Menticirrhus saxatilis*, *M. americanus*, *Sciaenops ocellata*, *Cynoscion nebulosus*, *Pogonias cromis* and *Larimus fasciatus* are present in the estuary occasionally. Yearling *C. regalis* were first caught in April and young of the year in July or August. Yearling *B. chrysoura* were first caught in March or April and young of the year in July or August. Juvenile *M. undulatus* and *L. xanthurus* may be present in the York River all year round. Young of the year *L. xanthurus* were first caught in April and *M. undulatus* were first caught in August. Small *M. undulatus* (15 to 20 mm TL) were caught from August to June, which may indicate a prolonged spawning season or a late spawning stock. Emigration to the ocean was found in all the four species during late fall or early winter. The relative abundance (catch per unit effort) of these four species indicated that water temperature and dissolved oxygen seemed to be the most important factors in the spatial and temporal distributions of these four species in the York River.

Six sciaenid species, *Larimus fasciatus*, *Cynoscion regalis*, *Bairdiella chrysoura*, *Micropogonias undulatus*, *Menticirrhus saxatilis* and *Leiostomus xanthurus* were examined for their food habits and functional morphology of feeding apparatus. The mouth position, dentition, gill rakers, digestive tract, pores and barbels, nares and body shape were found to be important for these sciaenids in locating and ingesting prey in the water column. Stomach contents indicated that the food partitioning of these six species was closely correlated with the habitat where each species was adapted to feed in. *Leiostomus fasciatus*, *Cynoscion regalis*, and *Bairdiella chrysoura* fed mainly above the bottom, whereas *M. undulatus*, *M. saxatilis* and *L. xanthurus* fed on epifauna, infauna or both.

Juvenile sciaenids are able to coexist in the same area because of differences in spatial and temporal distribution, relative abundance and food habits.

PART I  
A BASIS FOR CLASSIFYING WESTERN  
ATLANTIC SCIAENIDAE (PISCES:PERCIFORMES)

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## INTRODUCTION

Sciaenid fishes are characterized by their large otoliths and, with few exceptions, by the large vacuous lateral line pores on the snout and lower jaw, and by the extension of the lateral line to the tip of the caudal fin. They are also characterized by a large and often complex swimbladder and by the presence of well-developed drumming muscles. This paper treats 21 genera and 56 species of Sciaenidae in the western Atlantic Ocean. There are also four genera and 23 nominal species of sciaenid fishes present in freshwater river systems of the New World.

Current concepts of sciaenid classification are largely based on the morphology of swimbladder, otoliths (sagitta and lapillus), snout (rostral) and mental (mandibular) pores and/or barbels (Chu, Lo and Wu, 1963; Trewavas 1962, 1964; Robins and Tabb, 1965; Gilbert, 1966; Mohan, 1969; and Chao and Miller, 1975). Other characters, such as the position of the mouth, dentition, body form, size of the second anal spine, marginal serration of the preopercle, arrangement of lateral line scales, color of peritoneal lining, and numbers of gill rakers, fin ray counts and vertebrae are important in distinguishing species, and have been used by many authors (see section "A brief history of the study of western Atlantic Sciaenidae") to assign species to genera or to assess the generic relationships of western Atlantic Sciaenidae. However, the generic boundaries have not been well-defined and many nomenclatural problems prevail.

The purpose of this paper is to clarify the generic boundaries, to define supra-generic groups, and to describe the evolutionary trends of western Atlantic Sciaenidae primarily on the basis of the morphology of the swimbladder, otolith and snout and mental pore and/or barbel systems.

To classify the taxa above the generic level, the term supra-generic group is used here. Different attempts have been made to group sciaenid genera. Trewavas (1962) grouped eastern Atlantic sciaenids into tribes. Chu, Lo and Wu (1963) grouped sciaenids of the Chinese coasts into subfamilies. Mohan (1969) grouped Indian sciaenids into both subfamilies and tribes. The supra-generic groups used here are comparable to the tribes of Trewavas (1962) and the subfamilies of Chu, Lo and Wu (1963), and avoid the taxonomic problems inherent in utilizing formal taxonomic categories. Like those studies, the present assessment is also a regional study. Variations in swimbladder, otolith, and external morphology are found at different taxonomic levels. Therefore, a comparative study of sciaenids from several geographic regions, especially the eastern Pacific area, is necessary before allocating the western Atlantic sciaenid genera to formal taxonomic categories (tribes or subfamilies).

## MATERIALS

Type material and other preserved specimens were examined from the following institutions:

- AMNH - American Museum of Natural History, New York, New York.
- ANSP - Academy of Natural Sciences, Philadelphia, Pennsylvania.
- BMNH - British Museum (Natural History), London, England.
- CAS - California Academy of Sciences, San Francisco, California.
- FMNH - Field Museum of Natural History, Chicago, Illinois.
- LACM - Los Angeles County Museum of Natural History, Los Angeles, California.
- MCZ - Museum of Comparative Zoology, Harvard University, Cambridge, Massachusetts.
- MNHN - Museum National d'Histoire Naturelle, Paris, France.
- NHMV - Naturhistorisches Museum, Vienna, Austria.
- NMFS - National Marine Fisheries Service, NOAA, U.S.A.
- RMNH - Rijksmuseum van Natuurlijke Historie, Leiden, Netherland.
- USNM - United States National Museum, Washington, D. C.
- VIMS - Virginia Institute of Marine Science, Gloucester Point, Virginia.
- ZMA - Institute voor Taxonomische Zoologie, Universiteit van Amsterdam, Amsterdam, Netherland.
- ZMK - Zoological Museum, Copenhagen, Denmark.

Freshly frozen and formalin-preserved specimens were also obtained from various biological surveys, along the Atlantic Seaboard

of the U. S. These surveys were conducted by the Virginia Institute of Marine Science on various research vessels, by NMFS, Woods Hole, on R/V *Albatross IV* and Sandy Hook, on R/V *Atlantic Twin*, and by National Science Foundation's R/V *Eastward*, Beaufort. Specimens examined from Gulf of Mexico and Atlantic coast of South America were collected mainly by NMFS research vessels, the M/V *Oregon I* and *Oregon II* from 1962 to 1975. Type-specimens examined are indicated under the synonymy of each species. Numerous additional specimens have been studied, most of which are deposited at VIMS and USNM.

## METHODS

The comparative morphology of swimbladder, otolith and pore systems were examined. Other characters important in the recognition of species and genera were the arrangement and size of lateral line scales, serrations of the preopercular margin, size of the second anal fin spines, color of the branchial chamber and peritoneal linings, meristics and morphometrics. The standard methods of Hubbs and Lagler (1958) were used for all counts and measurements except for some modifications described by Chao and Miller (1975). In addition, scales perforated by lateral line tubes or pores were counted as lateral line scales from the upper end of the gill slit to the end of the hypural plate. On those species lacking well-defined lateral line tubules or pores, counts were made on the scale series immediately above the lateral line. Vertebral counts were determined from radiographs and from cleared and stained specimens (Taylor, 1967). The first caudal vertebra was identified by the absence of pleural ribs on the haemal process and usually by a short haemal spine just behind the elongated proximal pterygiophore of the anal fin.

Otoliths of most species were extracted from freshly caught specimens but some came from preserved specimens. The inner surface of the right sagitta and lapillus (in the *Stellifer* group) were illustrated. Ontogenetic changes of otoliths and swimbladder were studied in some species.

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All available type-specimens were examined to assure the true identities of species and genera of sciaenids studied in this paper. A primary synonymy is listed for each species to clarify nomenclatural problems.

Morphological terminology used here follows Chu, Lo and Wu (1963), Trewavas (1962, 1964) and Chao and Miller (1975). A modification of the terms used for the snout pores was suggested by Dr. E. Trewavas of BMNH, and for otoliths by Mr. J. Fitch of the California Department of Fish and Game. Snout pores along the margin of the rostral fold are termed marginal (snout) pores and pores in front or above them are termed upper (snout) pores (Fig. 1). The surface of the sagitta, with a "tadpole-shaped" sulcus is the inner surface and the obverse side is the outer surface (Fig. 2) because of the *in situ* position of the sagitta (Fig. 3). Also, the lateral margins of the sagitta are termed dorsal and ventral margins (Frizzell and Dante, 1965). The "head section" of the sulcus is called the "ostium" and the "tail section" is called the "cauda" (Stinton, 1975 and J. Fitch, personal communication).



## A BRIEF HISTORY OF THE STUDY OF WESTERN ATLANTIC SCIAENIDAE

Prior to the binominal system of Linnaeus, Catesby in his 1743 edition of "The natural history of Carolina, Florida and the Bahama Islands", illustrated "Perca marina" (= *Micropogonias undulatus*), the croaker (plate 3), from Chesapeake Bay and "Alburnus Americanus" (= *Menticirrhus americanus*), the whiting (plate 12), from Charleston, South Carolina. Edwards (1751) in the appendix of "A Natural history of birds" illustrated (plate 210) a "Ribband Fish" (= *Eques lanceolatus*) from the Caribbean Islands. Linnaeus (1758) in his tenth edition of "Systema naturae", Pisces Thoracici, recorded the genus *Sciaena* with five species, none of them from the western Atlantic. However, two species of western Atlantic Sciaenidae were named as *Chaetodon lanceolata* (= *Equetus lanceolatus*) based on Edwards' plate and *Cyprinus americanus* (= *Menticirrhus americanus*) from "the whiting" of Catesby. In the 12th edition of "Systema naturae", 1766, Linnaeus added *Labrus chromis* (= *Pogonias cromis*), *Perca alburnus* (= *Menticirrhus americanus*), *Perca punctata* (= *Bairdiella chrysoura*), *Perca ocellata* (= *Sciaenops ocellata*) and *Perca undulata* (= *Micropogonias undulatus*), although he did not place them with the Sciaenidae of Europe.

Bloch and Schneider (1801) in their "Systema ichthyologiae" reported six genera and nine species of western Atlantic Sciaenidae. They were *Johnius regalis* (= *Cynoscion regalis*), *J. saxatilis* (= *Menticirrhus saxatilis*), *Sciaena chromis* (= *Pogonias cromis*),

(= *Pogonias cromis*), *Lonchurus depressus* (= *Lonchiurus lanceolatus*), *Lonchurus ancyloдон* (= *Macrodon ancyloдон*), *Eques americanus* (= *Equetus lanceolatus*), *Eques punctatus*, *Grammistes accuminatus* (= *Pareques acuminatus*) and *Bodianus stellifer* (= *Stellifer stellifer*). A year later (1802) Lacépède in volume three of "Histoire Naturelle des Poissons" recorded *Dipterodon chrysourus* (= *Bairdiella chryoura*), *Pogonias fasciatus* (= *P. cromis*) and *Cheilodipterus acoupa* (= *Cynoscion acoupa*) and in volume four (1803) he added *Lutjanus cayennensis* (= *Cynoscion acoupa*), *Lutjanus triangulum* (= *Sciaenops ocellata*), *Centropomus alburnus* (= *Menticirrhus americanus*), *Sciaena croaker* (= *Micropogonias undulatus*) and *Leiostomus xanthurus* of the western Atlantic and also a fresh water sciaenid *Perca furcroid* (= *Pachypops fourcroid*) from Surinam.

The boundaries of the species, genera and family Sciaenidae were not well-defined, until Cuvier. In the first edition of "Règne Animal" (1817) Cuvier defined limits to the application of some generic names (e.g. *Umbrina* and *Sciaena*). Also four new genera of western Atlantic Sciaenidae were described: scientific names *Stellifer*, *Umbrina*, *Otolithes* and *Ancyloдон* were derived from Cuvier's vernacular names by Oken (Isis, 1817). In the second edition, Cuvier (1829) added a new generic name *Corvina*. Cuvier and Valenciennes (1830) in volume five of "Histoire Naturelle des Poissons", described seven genera and 21 species of western Atlantic sciaenids. There were four new genera, i.e., *Larimus*, *Nebriis*, *Lepipterus* and *Micropogon* and five freshwater nominal species, *Corvina oscula* (= *Aplodinotus grunniens*), *C. furcroea* (= *Pachypops fourcroid*) and *Lepipterus francisci* (= *Pachyrus francisci*) were included. They also described and illustrated the swimbladders of

six western Atlantic sciaenid fishes. Günther (1960) recorded 12 genera and 45 species of Sciaenidae from the western Atlantic including three genera and nine species from freshwater. He used the shape of the swimbladder as one of the diagnostic characters in some of his generic descriptions. Jordan and Eigenmann (1889) listed 22 genera and 60 species, including four genera and 11 species of freshwater sciaenids. Jordan and Everman (1895) included only 21 genera and 50 species of Sciaenidae from the Atlantic in the "Fishes of North and Middle America" including three genera and five species from freshwater. The studies of Desmarest (1823), Agassiz (1829), Castlenau (1855), Steindachner (1863), Bleeker (1865 & 1873), Poey (1881), Berg (1895) and more recently of Miranda Ribeiro (1915), Meek and Hildebrand (1925), Ginsburg (1929), Fowler (1941 & 1954), Schultz (1945 & 1949), Travassos and Paiva (1957), Cervigon (1966a & 1973), Vazzoler (1969), Travassos and Rego-Barros (1971), Roux (1973) and Jardim (1973) have contributed to the knowledge of taxonomy and distribution of western Atlantic sciaenids in different geographic regions. At present, 21 genera and 56 species of sciaenids are recognized from the western Atlantic. In addition, there is one monotypic genus (*Aplodinotus*) recognized from the freshwaters of North America, and three genera (*Pachypops*, *Placioscion* and *Pachyurus*) from the freshwaters of South America.

Few studies have been made on the relationships of genera, and higher divisions of western Atlantic Sciaenidae. Gill (1861 a,b,c & d) revised the genera and named six subfamilies for North American sciaenids, i.e. Corvinae, Haplidonotinae, Lariminae, Liostominae,

Otolithinae and Sciaeninae. Later, only five subfamilies of Sciaenidae were listed, the Corvininae was not mentioned (Gill, 1863a & 1873). Genera and subfamilies were defined by external characters and the boundaries between them were not clear. Bleeker (1876) in part two of "Systema Percarum Revisum" names two phalanxes (= tribes), the Sciaenini and Hemisciaenini for the familiar "Sciaenoidei". He also described three subphalanxes (= subtribes) in the Sciaenini, i.e. Equili, Aplodinoti and Johnii, and two subphalanxes in the Hemisciaenini, i.e. Pseudosciaeni and Otolithi. The western Atlantic sciaenid genera were described under each subphalanx. The diagnostic characters for each taxon were determined from the external morphology, e.g. mouth position, dentition, scales and fin rays.

Jordan and Eigenmann (1889) and Jordan and Evermann (1895) divided the Sciaenidae into two subfamilies, the Otolithinae and Sciaeninae, based on the number of precaudal and caudal vertebrae, lower jaw position, dentition, and size of the second anal spines. In the present study, I did not find these characters useful above the generic level. The first comprehensive studies on the species and generic relationships of sciaenids were published by Trewavas (1962) for tropical west African sciaenids and Chu, Lo and Wu (1963) for Chinese sciaenids. Trewavas (1962) grouped the nine west African sciaenid species into five tribes, based mainly on swimbladder structure; and the diagnostic characters of each tribe were clearly defined. Chu, Lo and Wu (1963) grouped 37 species of Chinese sciaenid fishes into seven subfamilies based on both swimbladder and otolith structure. Relationships among genera and subfamilies were also

clearly defined. Because the authors lacked access to type material, the true identity of a few species may be in doubt. Trewavas (1964) discussed the relationships of sciaenid fishes with a single mental barbel. She used the number of barbels and the structure of the swimbladder to assess the relationships of some barbeled sciaenid genera of the New World and suggested possible links of certain genera with Old World sciaenids. Similarly, Mohan (1969) used morphology of the swimbladders and otoliths as diagnostic characters to define each of the 14 genera of Indian Sciaenidae. Although Mohan (1969) placed 14 genera in five subfamilies and recognized two tribes in each of two subfamilies, the restrictions of some subfamilies and tribes remained unclear.

## DESCRIPTION AND RELATIONSHIPS

The morphology of the swimbladders, otoliths, snout and mental pores and barbels are diagnostic in the classification of sciaenid fishes. Within each of the characters, there are several structural patterns which may reflect phyletic relationships among groups of genera. The variations within each pattern are important in understanding the boundaries of different taxa. At the species level, the ontogenetic development of certain characters may suggest possible trends in phylogenetic relationships in higher taxa of the family. The presence of drumming muscles in males or in both males and females of certain taxa is also useful in assessing the relationships of sciaenids. Other characters essential for the study of relationships at the genus and species level but not at the family level are described in the key (see section "Field key to the genera and species of western Atlantic Sciaenidae").

Based on swimbladder, otoliths and external morphology of all western Atlantic sciaenid fishes, phyletic relationships are proposed here for 21 marine and two freshwater genera (Fig. 4). The relationships among the supra-generic groups were determined primarily by swimbladder morphology and secondarily by the morphology of the otoliths. External morphology was of some use as long as the adaptive and potentially convergent nature of these characters was kept in mind. By following the tactics used by Bolin (1947) for

the marine Cottidae of California, the actual or "convenient" classification level for tribes may be drawn as between dotted lines C and D on Figure 4. The subfamily level may be drawn between lines A and B.

#### I. Relationships Based on the Morphology of the Swimbladders.

Sciaenid fishes have a physoclistous swimbladder with a single or two inter-connected chambers. The usually well-developed swimbladder is attached very firmly to the ventral surface of the third to sixth vertebrae, under the post cardinal vein. The associated swimbladder (drumming) muscles usually present in males, are extrinsic except in *Pogonias*, in which they are intrinsic. The drumming muscles are composed of red striated fibres with abundant cytoplasm and conspicuous nuclei (Tower, 1908; Jones and Marshall, 1953). Tower (1908) also concluded that contractions of the drumming muscle set up vibrations in the bladder walls, thus producing sounds. Sound production in sciaenid fishes may function as recognition signals associated with spawning, feeding and aggressive behavior (Dijkgraaf, 1947; Knudsen, et. al. 1948; Johnson, 1948, Jones and Marshall, 1953; Chu, Lo and Wu, 1963). The sounds produced by different species of sciaenids also have different ranges of frequencies (Knudsen, et. al. 1948; Fish and Mowbray, 1970). This may result from the different morphology of swimbladders in different sciaenids. Comparison of sciaenid swimbladders in the present study is confined to external morphology and *in situ* position within the fish.

## 1. The morphological patterns of swimbladders.

All western Atlantic sciaenids have well-developed swimbladders except the genera *Menticirrhus* and *Lonchiurus*. The swimbladders in *Menticirrhus* atrophy during the juvenile stage. Only a vestige remains in adults. In adult *Lonchiurus*, the relative size of the swimbladder is much reduced. Based on swimbladders, western Atlantic sciaenids can be divided into two groups. One group has a single chambered swimbladder (Figs. 5 to 14) and the other has a two chambered swimbladder (Figs. 15 & 16). A total of eight distinct structural patterns of swimbladders are recognized here.

### A. The group with a single chambered swimbladder.

#### 1) *Sciaena* pattern:

A simple carrot-shaped swimbladder without well-developed anterior horns or diverticula (Fig. 5), its anterior end slightly in front of septum transversum and never reaching the skull. Swimbladders of nine genera have this pattern: *Aplodinotus*, *Ctenosciaena*, *Equetus*, *Larimus*, *Leiostomus*, *Menticirrhus*, *Pareques*, *Sciaena* and *Umbrina*. Although the swimbladder of *Larimus breviceps* has an additional pair of anterior projections (Fig. 5,F), *L. fasciatus* (Fig. 5,E) lacks the projections. Thus, *Larimus* should be included in this category. *Pachyurus schomburgkii* has a simple swimbladder and differs from *Pachyurus bonariensis* (Fig. 6). The generic status of *Pachyurus* is discussed in the synopsis section.

The swimbladder of *Menticirrhus* degenerates or atrophies in the adult. *M. saxatilis* has a well-developed simple bottle-shaped swimbladder in young specimens up to 90-100 mm SL (Fig. 7,A & B), but



only a trace of the swimbladder remains in the adult (Fig. 7,C & D). Bearden (1963) reported the length and condition of the swimbladders of 420 *M. americanus*. His results showed a gradual atrophy of the swimbladder from specimens of 56 to 60 mm SL (5.6% atrophied) to specimens of 125 to 130 mm SL (100% atrophied) in *M. americanus*. During this study, 28 specimens of *M. littoralis* were examined from 50 to 108 mm SL. Five specimens 71.1 to 108 mm SL had completely atrophied swimbladders and the rest had only a trace of the swimbladder less than 7 mm in length. *M. americanus* and *M. littoralis* have their atrophied swimbladder even in juvenile specimen. The juvenile swimbladder of the genus *Menticirrhus* may also be included in the *Sciaena* pattern.

2) *Nebris* pattern:

The swimbladder has a pair of broad diverticula which originate on the antero-lateral margin of the main chamber, extend posterior and loop anteriorly near the posterior end of the main chamber (Fig. 8). The diverticula are tapered and terminate anteriorly at the septum transversum. *Nebris* is the only genus with this pattern of swimbladder.

3) *Micropogonias* pattern:

The swimbladder of *Micropogonias* has a pair of tube-like diverticula which originate from the posterior half of the main chamber laterally (Fig. 9). The diverticula extend anteriorly to the septum transversum, but never reach the skull, then curve posteriorly to the anterior border of the swimbladder medially. *Micropogonias* is the only genus with this pattern of swimbladder.

4) *Pogonias* pattern:

A very complex lateral system of diverticula (Fig. 10) characterize the swimbladder of *Pogonias*, the only genus possessing this pattern. A study of the ontogenetic development of this swimbladder pattern (Fig. 10) may reveal the developmental pathway of other swimbladder patterns.

5) *Sciaenops* pattern:

The swimbladder of *Sciaenops* has a pair of tube-like anterior diverticula (Fig. 11,A). In large specimens, the complicated lateral diverticula and a pair of sac-like projections are found dorso-laterally at the anterior portion (Fig. 11,B & C). These diverticula are constructed of numerous small labyrinthine chambers (Fig. 11,c). Sometimes the lateral diverticula are completely covered with fat tissue (Fig. 11,B). A pair of holes is also present dorso-laterally on the body wall, between the third and fourth pleural ribs, to receive the dorsal projections of the swimbladder. *Sciaenops* is the only genus with this pattern of swimbladder.

6) *Cynoscion* pattern:

A pair of prominent horns develops anteriorly from the simple carrot-shaped swimbladder without lateral diverticula (Fig. 12); the well-developed anterior horns sometimes reaching to the back of the skull. Four genera have this pattern: *Cynoscion*, *Isopisthus*, *Macrodon*, and *Plagioscion*. Variations among genera and species (Figs. 12 & 13) with this pattern of swimbladder are limited to the

extension of the horns, whether they are broad or narrow, straight or curved. These genera, except the freshwater *Plagioscion*, have similar external features, such as an oblique terminal mouth, canine-like dentition, and similar habitats. They form a distinct group among western Atlantic sciaenids.

7) *Lonchiurus* pattern:

The swimbladder has a pair of forked anterior horns, each with a long posteriorly directed branch and a short branch directed anteriorly (Fig. 14). *Lonchiurus*, *Paralonchurus* and *Pachypops fourcroy* have this pattern. The anterior horns of swimbladders in *Lonchiurus* and *Paralonchurus* are rather stout and short (Fig. 14,A to C). They are thinner and longer in *Pachypops fourcroy* (Fig. 14,D), pierce the septum transversum and curve under the base of the skull. The relative size of the swimbladder in *Lonchiurus* is much reduced (Fig. 14,A, A' & A").

B. The group with a two chambered swimbladder.

8) *Stellifer* pattern:

The swimbladder is divided into two chambers by an anterior constriction. The anterior chamber reaches to the skull, is yoke-shaped and the posterior chamber is carrot-shaped (Fig. 15). *Bairdiella*, *Odontoscion*, *Ophioscion*, and *Stellifer* have this pattern. A pair of diverticula is present on the anterior chamber posterolaterally in *Stellifer* (Figs. 15,B & 16). These genera usually have two pairs of enlarged otoliths (Fig. 2,A & B, also see otolith section), a rather strong second anal spine and are relatively small in size.

## 2, Phylogenetic relationships based on swimbladder patterns.

Western Atlantic sciaenids can be separated into two distinctive groups (Fig. 17), based on the number of the chambers of the swimbladder. The *Stellifer* group differs from all other Sciaenidae by having two chambers (Fig. 15). Single chambered swimbladders are generally carrot-shaped with or without lateral diverticula except for the *Cynoscion* and *Lonchiurus* patterns (Figs. 5 to 14). In the *Cynoscion* pattern, the well-developed anterior horns are unique (Fig. 12). The *Lonchiurus* pattern also develops a pair of posterior diverticula from the base of the anterior horns (Fig. 14).

The genus *Menticirrhus* has only a vestige of the swimbladder in adults (Fig. 7). This trend of swimbladder reduction is also found in fishes with the *Lonchiurus* pattern swimbladders. There is a gradual reduction in size of the swimbladder from *Paralonchurus brasiliensis*, to *P. elegans*, then to *Lonchiurus lanceolatus* (Fig. 14), the latter apparently lacking a functional swimbladder. The gradual ontogenetic reduction in the relative size of swimbladders in the species of *Menticirrhus* and in *L. lanceolatus* may indicate close relationships among these fishes.

Further divisions of this character (Fig. 17) rely on the complexity of the swimbladders. *Sciaena* pattern is a simple carrot-shaped swimbladder without well-developed diverticula (Fig. 5). Although *Larimus breviceps* has a pair of small anterior projections (Fig. 5,F), this appears to be a variation within the *Sciaena* pattern. The swimbladder pattern with well-developed diverticula can also be divided into two groups (or more) by the complexity of their lateral diverticula (Fig. 17). *Nebriis* and *Micropogonias* have only a pair of

tube-like diverticula. The diverticula in *Nebris* originate from the antero-lateral margin of the main chamber (Fig. 8). In *Micropogonias*, the paired tube-like diverticula originate from the posterior half of the main chamber (Fig. 9). Another pattern of a rather simple diverticulated swimbladder is found in *Pachyurus bonariensis* (Fig. 6,A). It has a pair of anterior tube-like diverticula directed posteriorly.

*Sciaenops* and *Pogonias* have much more complicated lateral diverticula in adults. *Sciaenops* has a simple short pair of diverticula in young specimens (Fig. 11,A) and the "sac-like" projections located dorso-laterally develop in larger specimens (Fig. 11,C). The complicated diverticula on the swimbladder of *Pogonias* also develop ontogenetically (Fig. 10). The ontogenetic development of the swimbladder in *Pogonias* indicates that the diverticula were developed from the simple swimbladder from anterior to posterior portions of the main chamber. This evidence may also reflect the phylogenetic development of the swimbladder in the Sciaenidae.

#### Specific characters.

There are variations of swimbladder morphology within the general patterns. In the *Stellifer* pattern, the species of *Stellifer* have a pair of "bulb-like" or "tube-like" diverticula at the posterior-lateral margins of the anterior chamber (Figs. 15 & 16). Slight variations are also found in species such as *S. griseus* and *S. microps* (Fig. 16,B & C). *Bairdiella*, *Odontoscion*, and *Ophioscion* lack diverticula on the anterior chamber of the swimbladder (Fig. 15,A). The anterior horns in the *Stellifer* pattern terminate near the articulation between the opercular and hyomandibular bones (J.

Wintersteen, personal communication). In *Bairdiella* and *Odontoscion*, these horns curve forward abruptly and their tips end just behind the *adductor operculi* muscle and medially to the *dialator* and *levator operculi*. In *Ophioscion* and *Stellifer*, the anterior horns of the swimbladder diverge more laterally and their tips end subcutaneously, and are frequently visible on superficial inspection.

In the *Cynoscion* pattern, variations of the anterior horn morphology was found among different genera (Fig. 12), species and also ontogenetically within species (Fig. 13). There is little variation among the swimbladder morphology of the *Sciaena* pattern (Fig. 5). The anterior projections are present in *Larimus breviceps* and absent in *L. fasciatus*, indicating the possible extent of variation of swimbladder morphology within a genus and/or a pattern.

Although swimbladder morphology has been viewed as basic to the scheme of sciaenid classification (Trewavas, 1962; Chu, Lo and Wu, 1963; Mohan, 1969), features of some defined swimbladder patterns may vary widely. In setting generic limits, ontogenetic variations and intraspecific variation in certain species (not with growth) are important. Assessing the phyletic relationships of western Atlantic Sciaenidae is limited when complex swimbladders are involved. The criterion of simplicity or complexity is difficult among some taxa, but the evidence does indicate the trend of swimbladder development, i.e., from simple to complex with the diverticula probably evolving from the anterior portion towards the posterior portion of the main swimbladder chambers.

The drumming muscle is also useful in assessing relationships within sciaenid fishes. The development of the drumming muscles seems

to be correlated with the size of swimbladders and the sound producing function in some western Atlantic sciaenids. Generally, the drumming muscles are present in male sciaenid fishes with a well-developed swimbladder. In fishes with the *Lonchiurus* pattern swimbladder (Fig. 14), the drumming muscles are well-developed in males of *Paralonchurus* but absent in *Lonchiurus*, which is correlated with the size (or function) of the swimbladder. *Menticirrhus* has only a vestigial swimbladder and lacks drumming muscles. Drumming muscles are present in both male and female *Micropogonias*. *Pogonias* has a unique drumming muscle among western Atlantic sciaenids, located intrinsically on the complicated swimbladder in both sexes (Fig. 10). The drumming muscles of male *Cynoscion regalis* are only well-developed during the spawning season, and regress after spawning (J. Merriner, personal communication).

Possible relationships of western Atlantic sciaenids to sciaenids of other geographic areas may be revealed by the swimbladder shapes. Trewavas (1964) suggested a possible link of *Pachypops* and *Polyclemus* (= *Paralonchurus*), the *Lonchiurus* pattern swimbladder, with the tropical West African tribe Pseudotolithini. And the swimbladder of *Atractoscion aequidens* (Cynoscionini of Trewavas) agrees with the *Cynoscion* pattern of the present study. The diverticula on the swimbladder of *Pachyurus bonariensis* (Fig. 6,A) is also similar to that of *Bahaba flavolabiata* of the Indo-West Pacific (Chu, Lo and Wu, 1963). The complex swimbladder of the *Pogonias* and *Sciaenops* patterns resemble those of many Indo-Pacific (Chu, Lo and Wu, 1963; Mohan, 1969) and some East Pacific sciaenids (e.g. *Roncardor stearnsii*). Their relationships are not known at present.

## II. Relationships Based on the Morphology of the Otoliths.

The inner ear of sciaenids consists of three semicircular canals and a large sacculus, wherein the sagitta is contained (Fig. 3). The utriculus is an expanded area of the anterior semicircular canal containing the lapillus near the junction with the horizontal semicircular canal. This section is greatly expanded in fishes with a large lapillus (Fig. 3, B & B'). Posteriorly, the lagena is a small sac, located postero-dorsally to the sacculus, where the asteriscus is contained (Parker, 1908; and Chu, Lo and Wu, 1963). The relative size of the three otoliths, particularly the sagitta in all and lapillus in some sciaenids, are larger than in most Perciformes. The discussion here is based on the general morphology of the sagitta and the lapillus. The sagitta lies obliquely in the sacculus with the smooth surface inside (medially) and is oriented ventro-laterally *in situ* (Fig. 3). The lapillus is enclosed in the utriculus obliquely, with the smooth surface inside and is oriented laterally (Fig. 3, B & B').

### 1. The morphological patterns of otoliths.

The sagitta of Sciaenidae is characterized by having a distinct "tadpole-shaped" sulcus always on its inner surface (Fig. 2). A marginal groove is usually present between the dorsal margin and the sulcus of the sagitta. The anterior portion of the sulcus usually is expanded into a pear or oval shape and is termed the "ostium". The elongated narrow posterior portion is usually bent obliquely and is termed the "cauda" of the sulcus (Frizzell and Dante, 1965;



Stinton, 1975). The sagitta distinguishes sciaenids from other families in that the ostium is merely outlined and never channeled as in other families, whereas the cauda is always channeled (or scooped out) in the Sciaenidae. The outer surface usually has crest-like or blotch-like projections or granulations (Fig. 2, B' & C'). Western Atlantic sciaenids can readily be divided into two groups based on their otoliths: a group with both the lapillus and the sagitta enlarged and a group with only the sagitta enlarged (Fig. 2). Based on the shape, thickness and size of the sagitta and lapillus, and the morphology of the ostium and the cauda of the sulcus, 11 distinct patterns of otoliths are recognized here.

A. The group with only the sagitta enlarged.

1) *Sciaena* pattern:

The sagitta is more or less oval, usually with a smoothly convex ventral margin and a straight or crenulated dorsal margin (Fig. 18). Laterally the outer surface of the sagitta is usually much thicker in the middle. The ostium of the sulcus is usually broad, pear-shaped and reaches the anterior margin of the sagitta. The cauda is J-shaped with a relatively long and narrow distal end. A marginal groove is usually present between the sulcus and dorsal margin of the sagitta. *Ctenosciaena*, *Equetus*, *Leiostomus*, *Pachyurus*, *Pareques*, *Plagioscion*, *Sciaena* and *Umbrina* have this pattern (Fig. 18). Variations in this sagitta pattern are mainly in the position of the ostium and the length and the curvature of the cauda, both of which vary among genera and species. In most species, the ostium is straight or bent ventrally (to the left of Fig. 18). *Leiostomus xanthurus* (Fig. 18,D) is unique

in having the ostium bent to the dorsal margin and the sagitta slightly elongated and much thinner. *Plagioscion surinamensis* (Fig. 18,L) has a very long and curved distal end of the cauda. They are included in the *Sciaena* pattern for the convenience of the present study.

2) *Pogonias* pattern:

The sagitta is more or less semi-circular in shape with an evenly curved ventral margin and a straight but finely crenulate dorsal margin (Fig. 19). The ostium of the sulcus is broad and does not reach the anterior margin of the sagitta. The cauda is J-shaped with a pointed distal end and does not reach to the margin. The marginal groove is present between the sulcus and the dorsal margin of the sagitta. *Pogonias* and *Aplodinotus* have this pattern. The relative size of the "tadpole-shaped" sulcus is larger in *Pogonias* than in *Aplodinotus*. The sagitta is more curved laterally in *Pogonias* than *Aplodinotus*, but the latter has more granulations on the outer surface (Fig. 19,B' & b).

3) *Larimus* pattern:

The sagitta is slightly elongate and ovoid with rather straight and slightly crenulate dorsal margin, and convex ventral margin (Fig. 20). The ostium of the sulcus is large, pear-shaped and does not reach the anterior margin of the sagitta. The J-shaped cauda is bent acutely with a pointed distal end. The marginal groove is not distinct. *Larimus* and *Sciaenops* have this pattern. The sagitta of *Larimus* is broader anteriorly than posteriorly (Fig. 20, A & B), both ends are about equal in *Sciaenops* (Fig. 20, C & D). The posterior

portion of the sagitta is laterally very thick in *Larimus* (Fig. 20, a & b). *Sciaenops* has a rather thin sagitta (Fig. 20, c) and the shape of the sulcus is different from *Larimus*, therefore, a distinct otolith pattern may also be recognized.

4) *Nebris* pattern:

The sagitta in *Nebris* is very thick and oval with a notch at the posterior margin (Fig. 21). The ostium of the sulcus is large and ovoid and does not reach the anterior margin of the sagitta. The cauda is sharply bent with a very deeply grooved and enlarged distal portion. The marginal groove is absent. The outer surface of the sagitta is extremely elevated and very thick laterally (Fig. 21, a).

5) *Cynoscion* pattern:

The sagitta is elliptical and thin (Figs. 22, A & 23), the inner margin usually smoothly convex and the ventral margin slightly concave and finely crenulate. Laterally, the posterior portion of the sagitta is thicker than the anterior portion. The sulcus is usually elongate, the ostium ovoid or pear-shaped (Figs. 22,A & 24). The anterior end of the ostium may or may not reach the anterior margin of the sagitta. This varies among species and also within species ontogenetically (Fig. 23). The cauda is long and bent with a short distal end (Figs. 22,A & 23). The outer surface usually has a granular appearance, especially in large adult specimens (Fig. 23,a, b & c). *Cynoscion* species have this pattern. Sometimes a notch is present at the middle of the dorsal margin of the sagitta (Fig. 23,C & C') in species of *Cynoscion*.

6) *Menticirrhus* pattern:

The sagitta is elliptical and thin (Fig. 22,B, B' & b), the dorsal margin usually straight and finely crenulate, and the ventral margin slightly convex. Laterally, the middle portion is the thickest. The sulcus is elongate, the ostium is pear-shaped (Fig. 22,B) and does not reach the anterior margin of the sagitta. The cauda is long and J-shaped. The outer surface usually has crests and granulations. *Menticirrhus* species have this pattern.

7) *Lonchiurus* pattern:

The sagitta is usually thin and elongate (Fig. 24), the ventral margin usually smoothly convex and the dorsal margin slightly concave or crenulate. Laterally, the posterior portion of the sagitta is thicker than the anterior portion. The sulcus on the inner surface is elongate (Fig. 24). The ostium is pear-shaped and bent toward the ventral margin of the sagitta anteriorly. The cauda is J-shaped with a rather expanded distal end, close or reaching to the ventral posterior margin of the sagitta. The outer surface has granulations (Fig. 24,a, b & c). *Lonchiurus* and *Paralonchurus* have this pattern. There are two distinctive forms of sulcus on sagittae of this pattern. *Lonchiurus lanceolatus* and *Paralonchurus elegans* have a rather narrow ostium of the sulcus (Fig. 24,A & B). *P. brasiliensis* has a relatively thicker sagitta and the ostium of the sulcus is broader (Fig. 24,C & c).

8) *Isopisthus* pattern:

The sagitta is slightly elongate, rather broad and thick (Fig. 25,A, A' & a). The sulcus is elongate, the ostium is round and

reaches to the anterior margin of the sagitta. The cauda is long and slightly bent at the distal end. The outer surface has crest-like elevations, especially thickened at the posterior half (Fig. 25,a).

*Isopisthus* has this pattern.

9) *Macrodon* pattern:

The sagitta is elongate and thin (Fig. 25,B, B' & b). The dorsal margin has a deep notch on the posterior half and the ventral margin has a projection in the middle. Laterally, the anterior portion of the sagitta is slightly thicker than the posterior portion. The sulcus is elongate, the ostium is elongate with a broader anterior portion and reaches to the anterior margin of the sagitta. The cauda is rather straight and short with a disc-like distal end (Fig. 25,B). *Macrodon* has this pattern.

10) *Micropogonias* pattern:

The sagitta in *Micropogonias* is very thick and shield-shaped, often with a shelf or flange on the outer surface or on the dorsal margin (Fig. 26). The ostium of the sulcus is large and does not reach the anterior margin of the sagitta. The cauda is oblique and bent only slightly towards the ventral margin with a round disc-like distal end. The sagittal shelf on the outer surface of *M. furneri* and the lateral flange of *M. undulatus* vary ontogenetically (Fig. 26).

Sagittae of *Sciaena*, *Larimus* and *Nebriis* patterns are usually oval in shape and very thick in the middle. *Pogonias* pattern is usually semicircular in shape and thin. Sagittae of the *Cynoscion*, *Menticirrhus*, *Lonchirus* and *Macrodon* patterns are usually more

elongate and thin. *Isopisthus* pattern is elongate and thick in the posterior portion. Sagitta of the *Micropogonias* pattern is irregular in shape and very thick.

B. The group with both sagitta and lapillus enlarged:

11) *Stellifer* pattern:

The size of the sagitta is obviously reduced and an enlarged lapillus more than two thirds the size of the sagitta is present (Fig. 27). The ostium of the sulcus lacks its anterior portion and the cauda is well bent in a J-shape reaching almost to the ventral margin of the sagitta. *Bairdiella*, *Odontoscion*, *Ophioscion* and *Stellifer* have this pattern. There are considerable morphological variations within this otolith pattern (Fig. 27).

*Stellifer* and *Ophioscion* have a parallelogram-shaped sagitta and a sub-oval lapillus; both otoliths are about the same size (Fig. 27, A, A', B & B'). The ostium of the sulcus is short and reaches to the anterior margin of the sagitta. The cauda is bent obliquely towards the ventral margin of the sagitta. A marginal groove is evident along the dorsal margin of the cauda. The whole sagitta appears to be truncated at the middle of the ostium. The lapillus is large, sometimes larger than the sagitta (e.g., *Stellifer*, Fig. 28), a deep groove is present at the antero-ventral end of its inner surface with an open end to the dorsal margin (Fig. 28, A', B', C' & D').

The sagitta of *Bairdiella* has an inverse triangular shape with a projection at the antero-dorsal corner dorsally (Fig. 27, C). The ostium of the sulcus is further reduced. The cauda is J-shaped and broadened at the middle. The marginal groove is well-defined. The

lapillus is smaller than the sagitta and is irregular in shape (Fig. 27,C'). The groove along the antero-ventral end of the lapillus opens to the ventral margin.

The sagitta of *Odontoscion* is sub-triangular in shape. The ostium of the sulcus is nearly absent except for a small piece at the anterior margin (Fig. 27,D). The cauda is wide and curves vertically towards the ventral margin. The marginal groove is absent from the sagitta. The lapillus is ovoid and its antero-ventral groove is indistinct (Fig. 27,D').

A total of 11 patterns of otoliths are described here for western Atlantic Sciaenidae. Among them, the *Stellifer* pattern is most different and present only on the Atlantic and Pacific coasts of the American continents. The *Sciaena* pattern is the most generalized and resembles the "Sciaena-form" of Indo-West Pacific Sciaenidae (Chu, Lo and Wu, 1963; Mohan, 1969). The *Macrodon* pattern is similar to the "Johnius-form" and the *Micropogonias* pattern shares the disc-like "tail" end (cauda) with the "Pseudosciaena-form" of Chu, Lo and Wu, (1963). Whether these should be interpreted as convergences or as possible evidence of relationships is not known at present. Many sagitta patterns, such as *Nebris*, *Micropogonias*, *Cynoscion*, *Menticirrhus*, *Isopisthus* and *Macrodon* are monotypic. These basic patterns are defined here for the convenience of assessing the relationships of western Atlantic Sciaenidae. Further division or grouping will be essential for comparisons with sciaenids of other geographic regions.

2) Phylogenetic relationship based on otolith patterns.

Based on the morphology of otoliths, western Atlantic sciaenids can readily be divided into two groups (Fig. 29 and Chao & Miller, 1975). The *Stellifer* pattern differs from all other sciaenids by having a large lapillus more than one half the size of the sagitta (Fig. 27). The other patterns of the otoliths can be further divided into two groups (Fig. 29), one having a more or less ovoid or oval elongate sagitta and the other having a shield-like sagitta as found in *Micropogonias* (Fig. 26). Within the former group, the *Cynoscion*, *Menticirrhus*, *Lonchiurus*, *Isopisthus* and *Macrodon* patterns usually have thin, elongate sagittae (Figs. 22 to 25) and the rest have a broader and thicker sagitta. In the general outline of the sagitta, the *Cynoscion* and *Menticirrhus* patterns are most similar to each other (Fig. 29) and have the posterior portion of their sagittae slightly thicker than the anterior portion (Fig. 22). The *Lonchiurus* and *Isopisthus* patterns have the posterior portion of their sagittae further thickened (Figs. 24, a, b, c & 26, a), which may be variations among the generally elongated and thin sagittae. The sagitta of *Macrodon* is unique in having the anterior portion of the sagitta thicker than the posterior portion (Fig. 26, b). Among the remaining otolith patterns, the *Nebris* pattern (Fig. 21) is unique in the general outline of its sagitta and the morphology of the sulcus (Fig. 29). The *Larimus* pattern has a rectangular sagitta (Fig. 20), which differs from the more generalized oval-shaped sagitta of the *Sciaena* and *Pogonias* patterns (Figs. 18, 19 & 29).



### Specific Characters.

The sizes of the lapillus and sagitta in the *Stellifer* pattern vary among the genera (Fig. 27). The trend of reducing the size of the sagitta and increasing the size of the lapillus is evident from *Stellifer* to *Odontoscion*, then to *Bairdiella* (Fig. 27). *Cynoscion* has the most number of species of western Atlantic sciaenid genera, a total of 12. Therefore, it is not surprising that morphological variation in their sagittae is great (Fig. 23). *Menticirrhus* has a sagitta similar to *Cynoscion* (Fig. 22), but the relative size of the sagitta is smaller. Smith (1905) found the otoliths in *Menticirrhus* are relatively smaller than in other genera of sciaenids he examined. *Micropogonias* has the most distinctive sagitta, with the sagitta further ossified and a lateral flange developed along the dorsal margin (Fig. 26). *Nebris* is also unique in having an oval, thick sagitta (Fig. 21) with a notch on the posterior margin which does not resemble any other western Atlantic sciaenid. Although the *Pogonias* pattern (Fig. 19) has a semicircular sagitta, the sulcus is similar to that of the *Sciaena* pattern (Fig. 18). Variations among the genera of the *Sciaena* pattern are mainly in the position and shape of the sulci. The sagittae of *Larimus* and *Sciaenops* are more or less rectangular (Fig. 20), but the shapes of the sulci differ. *Leiostomus xanthurus* differs from other *Sciaena* pattern by having its sagitta (Fig. 18,D & d) slightly elongated and thin. It is probably an intermediate form between the elongated thin and ovoid thick forms of sagittae.

The morphology of the otoliths appears more variable than that of the swimbladder within a given taxon. Chu, Lo & Wu (1963) relied

heavily on the relative position of the "head" (ostium) and "tail" (cauda) sections of the "tadpole-shaped" sulcus to diagnose the subfamilies of Chinese Sciaenidae. In some species of western Atlantic sciaenids, the relative position of the ostium and cauda shift slightly as the fish grows (Figs. 20, 23 & 26). This feature should be viewed cautiously when used to group taxa above the generic level.

Fossil materials of sciaenid otoliths are important in assessing the phyletic relationships of modern sciaenids. John Fitch of the California Department of Fish and Game is currently studying this aspect of sciaenid relationships.

### III. Relationships Based on External Morphology.

Prior to Trewavas (1962) and Chu, Lo & Wu (1963), most authors used external morphology to classify sciaenid fishes (also see the section "A brief history of the study of western Atlantic Sciaenidae"). The characters used were body shape, presence or absence of the mental barbel and/or pores, size of the second anal fin spine and position of the mouth. Unfortunately these characters are extremely adaptive, evolutionarily plastic and convergent. They are more indicative of feeding adaptations and habitats rather than phylogenetic relationships (Fig. 30). Even so, these characters do show that the external morphology is also diagnostic at the generic and species levels. The following discussion assesses the limits and usefulness of these characters.

#### 1. Patterns of pore and barbel systems.

The pores at the tip of the snout and lower jaw (mandible) of sciaenids are the openings of the well-developed cavernous lateral-line canals on the head (Fig. 1). The snout (rostral) pores can be divided into upper pores, those present at the tip of snout and marginal pores, those present along the edge of the rostral fold (Fig. 1). Usually, there are three or five distinct upper pores, although some genera lack upper pores and some may have more than five as adults. Typically there are five marginal pores, one median and two pairs of lateral pores (Fig. 31). Some genera have only two marginal pores. The mental (mandibular) pores are most often five (four to six) one median and two pair of lateral pores (Fig. 32), whereas some genera

completely lack mental pores. One or more mental barbels may be present in many western Atlantic sciaenid fishes. The number of pores and barbels are correlated with the mouth position and feeding niches of the species (see part II of whole study). The variations are broad among the genera (Fig. 33, 34 & 35) and species (Fig. 36).

A. Structural patterns of snout pores:

There are five snout pore patterns recognized in western Atlantic Sciaenidae (Fig. 31).

1) Two (or no) marginal pores:

This pattern has only two marginal pores and no upper pores (Fig. 31,A). The rostral fold is thin and complete without notches. *Cynoscion*, *Macrodon*, *Nebris* and *Plagioscion* have this pattern. All species have a very oblique large mouth, the lower jaw projecting in front of the upper jaw, and sometimes no pores on the snout, except *Plagioscion*, which has a slightly oblique and terminal mouth. Fishes with this snout pore pattern and mouth position are mid- to upper water column feeders.

2) Five marginal pores:

This pattern has five marginal pores with no upper pores. The rostral fold has notches at the openings of the lateral pores (Fig. 31, B). *Aplodinotus* and *Larimus* have this pattern. *Aplodinotus* has a moderate sized mouth, terminal or slightly inferior, a typical lower water column to bottom feeder. *Larimus* has a large mouth, very oblique, and feeds in mid- to the upper water column.

3) Five marginal pores and two upper pores:

This pattern has a slightly indented rostral fold (Fig. 31,C). *Odontoscion* is the only genus of western Atlantic sciaenid with this pattern. *Odontoscion* has a moderate sized terminal mouth, and is typically a mid- and lower water column feeder.

4) Five marginal and three upper pores:

The rostral fold of this pattern is indented at the openings of the marginal pores (Fig. 31,D). *Bairdiella*, *Ctenosciaena*, *Menticirrhus*, *Ophioscion*, *Paralonchurus*, *Sciaena* and *Stellifer* have this pattern. The position and size of the mouths in this group of fishes suggests that some feed in midwater and some on the bottom. In bottom feeders such as *Menticirrhus* and *Paralonchurus*, the rostral folds (upper lips) are very deeply indented below the marginal pores (Fig. 33, C & D). For the mid- or lower mid-water feeders, *Stellifer lanceolatus* and *Ophioscion punctatissimus*, the rostral folds are only slightly indented (Fig. 33,A & B). *O. punctatissimus* also has two minute pairs of upper pores present dorso-laterally to the outer lateral marginal pores (Fig. 33,B).

5) Five (or more) marginal and five (or more) rostral pores:

This pattern usually has five distinct rostral and five marginal pores (Fig. 31,E). Sometimes an extra pair or two pairs of minute pores are developed lateral to the upper and marginal pores (Fig. 31,F). The rostral fold is slightly notched or smooth below the openings of the marginal pores. *Equetus*, *Leiostomus*, *Lonchiurus*, *Micropogonias*

*Pachyurus*, *Pareques*, *Pogonias*, *Sciaenops* and *Umbrina* have this pattern and have a horizontal or inferior mouth, and are typically bottom feeders. Variation within this snout pore pattern is mainly in the presence of additional minute pores (Fig. 33, E to H), which may also vary ontogenetically.

B. Structural patterns of mental pores and barbels:

There are seven mental pore and barbel patterns recognized in western Atlantic Sciaenidae (Fig. 32). Variations are rather common within the genera especially in speciose genera, such as *Stellifer* and *Bairdiella*.

1) No pores:

This pattern has neither pores nor barbels on the lower jaw (Fig. 32,A) and is found in *Cynoscion*, *Isopisthus*, *Macrodon* and *Plagioscion*. All fishes with this pattern have a large oblique mouth and are upper or mid-water feeders, although *Plagioscion* has a nearly terminal mouth.

2) Four pores:

This pattern has four pores at the anterior part of the lower jaw (Fig. 32,B) which may vary in arrangement among different genera (Fig. 34,A to D). *Nebris*, *Odontoscion*, *Larimus* and *Stellifer lanceolatus* have this pattern and all have a terminal or oblique mouth, and feed in the mid- to upper water column.

## 3) Five pores:

This pattern has five pores without a barbel, one median pore located at the center of the lower jaw symphysis and two lateral pairs (Fig. 32,C). The arrangement of the five pores vary among genera (Fig. 34,E to H). *Aplodinotus*, *Equetus*, *Leiostomus*, *Ophioscion*, *Pachyurus*, *Pareques*, *Sciaena*, *Sciaenops* and some species of *Bairdiella* (*B. ronchus* and *B. sanctaeluciae*) and *Stellifer* (*S. microps*, *S. stellifer* and *S. venezuelae*) have this pattern.

## 4) Six pores:

This pattern has six pores and no barbel, the two median pores usually close together, and two pairs of lateral pores (Fig. 32,D). *Bairdiella chrysoura*, *Stellifer colonensis*, *S. griseus* and *S. rastrifer* have this pattern, which is a derivative of the five-pored pattern. Fishes with this pattern of mental pores have a terminal to slightly inferior mouth and feed from lower mid-water to the bottom.

## 5) Four pores and one barbel:

This pattern has two pairs of lateral pores and a barbel at the anterior tip of lower jaw (Fig. 32,E). *Ctenosciaena*, *Menticirrhus* and *Umbrina* have this pattern. The morphology of the barbel varies among these three genera (Fig. 35). *Ctenosciaena* has a thin barbel tapering at the end. *Menticirrhus* and *Umbrina* have a short rigid barbel and an apical pore which is also present at the tip of the barbel in *Umbrina*. They all have inferior mouths and are bottom feeders.

6) Four pores and two barbels:

This pattern has two pairs of lateral pores and a pair of thin barbels originating from inside the posterior pair of mental pores (Fig. 32,F). *Lonchiurus* is the only genus with this pattern and has an inferior mouth, typical of bottom feeders.

7) Five pores and many barbels:

This pattern has five pores, one median and four lateral, together with three to 19 pairs of minute barbels at the symphysis of the mandibles and along the rami of the lower jaws (Fig. 31,G & H). *Micropogonias*, *Paralanchurus* and *Pogonias* have this pattern. The arrangement of the barbels are different among the genera, especially in *Paralanchurus*. Three pairs of minute barbels form a tuft situated lateral to the median pore and 12 to 16 pairs of small barbels are distributed along the rami of the lower jaw (Fig. 32,H). *Micropogonias* and *Pogonias* lack the tufts of barbels at the symphysis of the lower jaw (Fig. 32,G). Fishes with this pore and barbel pattern have an inferior mouth, and are typically bottom feeders.

When comparing the pore and barbel systems of western Atlantic Sciaenidae with the Indo-Pacific (Chu, Lo & Wu, 1963) and East Atlantic and East Pacific Sciaenids, the five marginal and three to five upper pores on the snout seem to be generalized in most sciaenids. Also the five pores on the lower jaw is typical in sciaenids. Sciaenids with a single mental barbel are found in the Indo-West Pacific, on both sides of the Atlantic and in the eastern Pacific (Trewavas, 1964). However, sciaenids with more than one barbel are found only along the Atlantic and Pacific coasts of America.



2) Phylogenetic relationships based on external morphology.

Western Atlantic sciaenids can be readily separated into two groups by the presence or absence of the mental barbels (Fig. 30). The group without barbel can be further divided by their mouth positions and body shape (Fig. 30). *Cynoscion*, *Isopisthus* and *Macrodon* differ from other barbelless sciaenids by having an elongate body, an oblique mouth with a pair of enlarged canines at the tip of the upper jaw and a protruding lower jaw (Fig. 37,A). Those species that lack canines can be divided into groups by their mouth positions and body shapes (Fig. 30). *Nebris* and *Larimus* have a very large and oblique mouth (Fig. 37,B). *Bairdiella* and *Odontoscion* both have a terminal mouth (Fig. 37,C). The genus *Stellifer* consists of species with inferior (*S. microps*), terminal (*S. rastrifer*) and oblique (*S. lanceolatus*) mouth (Fig. 38). *Sciaena*, *Leiostomus*, *Pareques*, *Equetus* and *Sciaenops* have inferior mouths. *Sciaenops* differs from these fishes by having an elongate body. *Sciaena* is characterized by a terminal and horizontal mouth.

Sciaenids with a mental barbel usually have an inferior mouth and can be divided into different categories by the number and position of barbels (Fig. 30). *Ctenosciaena* has a thin tapered barbel and a terminal mouth (Fig. 35,A), unique among barbeled western Atlantic sciaenids. Both *Umbrina* and *Menticirrhus* have an apical pore at the tip of the single rigid barbel (Fig. 35,B & C). *Lonchurus* has a pair of long slender barbels at the symphysis of the lower jaw (Fig. 32,F). *Paralonchurus* has three pairs of minute barbels in a tuft at the symphysis of the lower jaw lateral to the median mental pore and 12 to 16 pairs of small barbels along the rami of the lower jaw

(Fig. 32,H). *Micropogonias* has three to four pairs of small mental barbels and *Pogonias* has 12 to 13 pairs that never form a tuft at the symphysis of the lower jaw (Fig. 32,G).

The combination of morphological characters described above reflects habitat or similarities in feeding habits. The genera listed at the top of Figure 30 generally feed in the upper to mid-water column and the genera listed near the bottom feed in the lower water column to the bottom. Within the different snout pore, mental pore and barbel patterns of western Atlantic Sciaenidae, three different feeding modes may be recognized (Fig. 30). The mid- to upper water column feeders have only marginal pores (two or five) on the snout, and have zero to four mental pores without barbels on the lower jaw. The mid- to lower water column feeders have five marginal pores and two to three upper pores on the snout, and four to five mental pores without barbel(s) on the lower jaw. The third group, bottom feeders, have five marginal and three to five rostral pores on the snout, and have four to five mental pores and one to many barbels on the lower jaw. The number of pores and barbels seem to increase in feeding niches that are closer to the bottom.

#### Specific characters.

Western Atlantic sciaenids with two marginal snout pores and with neither upper pores (Fig. 31,A) nor mental pores (Fig. 31,A) are upper water column feeders, such as *Cynoscion*. Although *Nebris* has a very oblique mouth (Fig. 37,B) and snout pore arrangement similar to *Cynoscion*, it also has four mental pores (Fig. 34,A) and a rounded body with a flat ventral surface. This suggests that *Nebris* is a

bottom dweller feeding from the bottom upward. Sciaenid fishes with five marginal pores and no upper pore on the snout are mid- to upper midwater column feeders, such as *Larimus*, which has four mental pores similar to *Nebris*, (Fig. 34,B). But *Larimus* has a compressed body which may not be adaptive to a bottom habitat as in *Nebris*.

*Odontoscion* has five marginal pores and two upper pores on the snout (Fig. 31,C). It also has four mental pores and a terminal mouth (Fig. 37,C) which suggests that it is a mid- to lower midwater column feeder.

Sciaenid fishes with a structural pattern of five marginal pores and three upper pores (Fig. 33,A to D) are rather common among different genera, such as *Bairdiella*, *Ophioscion*, *Stellifer* and *Menticirrhus*. Species of *Menticirrhus* all have one barbel and four pores on the lower jaw, and are bottom feeders. But among the genera and species of *Stellifer*, the number of mental pores varies from four to six (Fig. 36). Their feeding niches are mainly correlated with the positions of the mouth. Although the number of mental pores among the species of *Bairdiella* varies from five to six, they all have a terminal mouth and are the mid- to lower mid-water feeders. Species of *Stellifer* not only have a variable number of mental pores, from four to six, but also have different mouth positions, from inferior to slightly oblique (Fig. 38). The species with a slightly oblique mouth, such as *S. lanceolatus*, *S. stellifer* and *S. rastrifer*, are apparently mid-water column feeders. But each species has a different number of mental pores (Fig. 36): *S. lanceolatus* four, *S. stellifer* five and *S. rastrifer* six. In addition, other genera of western Atlantic sciaenids, *Ctenosciaena*, *Paralónchurus* and *Sciaena*, also

have a similar snout pore pattern. Among them, *Ctenosciaena* and *Sciaena* both have terminal mouths and probably feed in mid-water down to the bottom. *Ctenosciaena* also has one barbel and four pores on the lower jaw (Fig. 35,A). *Sciaena* has five pores and no barbel on the lower jaw. Based on these variations, the three upper and five marginal snout pore pattern is probably the more generalized among western Atlantic sciaenids with different feeding niches.

Sciaenid fishes with five or more upper and marginal pores (Fig. 31,E & F) are all bottom feeders with an inferior mouth. They usually have five pores on the lower jaw with or without mental barbels. *Umbrina* has four pores and a short perforated barbel with an apical pore on the lower jaw (Fig. 35,C). *Lonchiurus* has four pores and two long tapered barbels on the lower jaw (Fig. 32,F).

The external characters described above are sometimes variable within a genus and yet similarities are found among different genera. Neither the differences nor the similarities necessarily reflect phylogenetic relationships. These external morphological characters and those mentioned in the key should be treated as "key characters" only. Their usefulness in assessing relationships of sciaenids is usually not above the species level.

A SYNOPSIS OF THE SUPRA-GENERIC GROUPS,  
GENERA, AND SPECIES OF WESTERN ATLANTIC SCIAENIDAE

I. Diagnoses and Primary Synonymy of the Supra-generic Groups,  
Genera and Species

Based on the structural patterns of swimbladder, otolith, and pore and barbel systems, western Atlantic sciaenids can be grouped into 11 supra-generic groups. They are the *Sciaena*, *Umbrina*, *Larimus*, *Sciaenops*, *Cynoscion*, *Micropogonias*, *Lonchiurus*, *Menticirrhus*, *Nebrius*, *Pogonias*, and *Stellifer* groups. The freshwater sciaenid genera, *Pachypops* and *Pachyurus* of South America are not included in these groups, due to lack of comparative material. The taxonomic position of these generic groups is between genus and family. Neither tribes nor subfamilies are utilized for western Atlantic Sciaenidae, until comparisons can be made with the sciaenids of other regions.

The following descriptions of western Atlantic sciaenid groups are arranged in order from simple to more complex swimbladder patterns. In each group description, the diagnosis of the supra-generic groups and genera are given and the species are listed after each genus. A primary synonymy is given for each genus and species for nomenclatural purposes. All available type-specimens are listed in the synonymy for each species.

*Sciaena* Group

Diagnosis: Swimbladder in a simple carrot-shape, without diverticulum (*Sciaena* pattern, Fig. 5); sagitta more or less oval-shaped and thick (Fig. 18), lapillus not enlarged, snout with five marginal pores and three to seven rostral pores (Fig. 31,D to F); lower jaw with five pores and no barbels (Fig. 32,C). There are three genera in this group; *Equetus*, *Pareques* and *Sciaena*. *Leiostomus* thinner and longer (Fig. 18,D). But, *Leiostomus* is probably closely related to the *Sciaena* group than other western Atlantic sciaenid groups.

Genus *Equetus* Rafinesque

Diagnosis: Body oblong, compressed, back much elevated, rapidly tapering to a narrow caudal peduncle; sides with broad oblique bands; spinous dorsal fin very long and filamentous. Mouth small and inferior, teeth in villiform bands; gill rakers few, short and slender. Vertebrae 10+15=25; no free interneurals anterior to first dorsal fin (McPhail, 1961). Swimbladder simple, carrot or bottle-shaped (Fig. 5,C). Sagitta oval-shaped, the ostium of the sulcus reaching to anterior margin (Fig. 18,B & C); snout with five to seven upper pores; lower jaw without barbel. Tropical western Atlantic endemic. Two species: *E. lanceolatus* and *E. punctatus*. Mainly inhabits coral reefs, sometimes shallow coastal waters.

*Eques* Bloch, 1793, pt. 7:90 (type-species: *Eques americanus* Bloch, by monotypy = *Chaetodon lanceolatus*, Linnaeus).

*Equetus* Rafinesque 181:86 (emendation of *Eques* Bloch, 1793 and substitute name for *Eques*, therefore taking the same type-species: *Eques americanus* Bloch; preoccupied by *Eques* Linnaeus, 1758:459, a genus of lepidopteran insect).

*Equetus lanceolatus* (Linnaeus)

"Ribband Fish" Edwards, 1751, pl. 210, middle figure, Caribbean Islands (non-binomial).

*Sciaena edwardi* Gronow, 1754 (ed. by Gray 1854):53, (West) Indian Sea (after Edwards).

*Chaetodon lanceolatus* Linnaeus, 1758:277, Caribbean Islands (after Edwards, pl. 210).

"Serranas" Parra, 1787, pl. 2, upper figure, Cuba (non-binomials).

*Eques americanus* Bloch, 1793: 91. 347, fig. 1, West Indies.

*Eques balteatus* Cuvier, 1829:175 (after Edwards and Bloch).

*Equetus punctatus* (Bloch & Schneider)

"Serranas" Parra, 1787, pl. 2, lower figure, Cuba, (non-binomials).

*Eques punctatus* Bloch & Schneider, 1801:105, pl. 3, fig. 2 (based on Parra).

*Pareques* Gill

Diagnosis: Body oblong, compressed, anterior profile steep, back tapering rapidly to a narrow caudal peduncle; sides with longitudinal stripes. Mouth small, inferior; teeth in villiform bands; gill rakers short and blunt. Vertebrae 10+15=25; three free interneurals anterior to first dorsal fin. Swimbladder simple, carrot-shaped. Sagitta

oval, the ostium of the sulcus reaching to anterior margin (Fig. 18,E & F), the cauda long, J-shaped. Snout with five to seven upper pores (Fig. 33,H); lower jaw without barbel. Tropical eastern Pacific and western Atlantic, about five species. Two western Atlantic species: *P. acuminatus* and *P. umbrosus*. Inhabits sandy and muddy bottoms of high salinity waters and coral reefs.

*Pareques* Gill in Goode 1876:50 (type-species: *Grammistes acuminatus* Bloch and Schneider 1801, by original designation).

*Pareques acuminatus* (Bloch & Schneider)

"Chaetodon, lineis fuscis" Seba, 1761:72, pl. 26, fig. 33, no locality (non-binomial).

*Grammistes acuminatus* Bloch & Schneider, 1801:184 (after Seba).

*Eques lineatus* Cuvier, 1830:169, Brazil (syntypes: MNHN 7475, one specimen, 135 mm SL; MNHN 43, not located).

*Eques pulcher* Steindachner, 1867:349, Barbados (type: not located in NHMV, Vienna).

*Pareques umbrosus* (Jordan & Eigenmann)

*Eques acuminatus umbrosus* Jordan & Eigenmann, 1889:440, Charleston, South Carolina (holotype: USNM 25981, 165 mm SL).

Remarks: *P. acuminatus* and *P. umbrosus* were thought to be the same species until Jordan & Eigenmann (1889) proposed the subspecies name *E. acuminatus umbrosus*, USNM 25981 was the specimen described by Jordan and Eigenmann (1889) as *Eques acuminatus* (Bloch & Schneider), and collected by G. C. Leslie. These two species can be separated



by their coloration and counts (also see "Key to the species of *Pareques*"). The sagittae of these two species differ from each other by the features of the ostium of the sulcus (Fig. 18,E & F). *P. umbrosus* is a more northerly species distributed from Chesapeake Bay to Florida and also recorded from Texas over sandy mud bottom of inshore and offshore waters. *P. acuminatus* is a more southerly species from Florida to Brazil in coral reef and rocky bottom coastal waters. L. Woods (FMNH) and G. Miller (NMFS, Miami) have new species of *Pareques* from western Atlantic in preparation.

#### *Sciaena* Linnaeus

**Diagnosis:** Body elongate, compressed, dorsal profile slightly elevated or evenly arched; spinous dorsal not elongated. Mouth horizontal to inferior; teeth in bands; gill rakers short and slender, widely spaced. Vertebrae 25 (11+14 in *S. bathytatos* and 10+15 in *S. trewavasae*). Swimbladder simple, carrot-shaped (Fig. 5,A). Sagittae oval, the ostium of the sulcus just reaching to the anterior margin (Fig. 18,G & H). Snout with three upper pores (Fig. 31,D); lower jaw without barbel. Two species in the western Atlantic; *S. bathytatos* and *S. trewavasae*, inhabit deeper subtropical waters and one species in East Atlantic, and Mediterranean, *S. umbra*.

*Sciaena* Linnaeus, 1758:228 (type-species: *Sciaena umbra* Linnaeus, by subsequent designation of International Commission for Zoological Nomenclature (1972), Opinion 988.

**Remarks:** The nomenclature of the genus *Sciaena* was not stabilized until *S. umbra* Linnaeus, 1758, was designated as the

type-species of the genus under the Plenary Powers of the International Commission for Zoological Nomenclature (1972, Opinion 988) and interpreted by the neotype designated by Trewavas (1966). The history of the case and synonymies are not repeated here. This genus is the type-genus of the tribe Sciaenini (Trewavas, 1962). The genus *Sciaena* is still not well-defined and it may contain more species than currently recognized (Chao and Miller, 1975).

*Sciaena bathytatos* Chao & Miller

*Sciaena bathytatos* Chao & Miller, 1975:267, fig. 9, Caribbean Sea off Colombia (holotype: USNM 211514, 208 mm SL).

*Sciaena trewavasae* Chao & Miller

*Sciaena trewavasae* Chao & Miller, 1975:262, fig. 8, Caribbean Sea off Colombia (holotype: USNM 211513, 155 mm SL).

*Leiostomus* Lacépède

Diagnosis: Body oblong, compressed, back elevated, sides with oblique stripes and a dark humeral spot behind upper end of gill slit; spinous dorsal fin not elongated. Mouth small and inferior; teeth in villiform band; gill rakers short and slender. Vertebrae 10+15=25. Swimbladder in simple carrot-shape (Fig. 5,B). Sagitta oval and thin, the ostium of the sulcus reaching the anterior margin and bent to the ventral margin (Fig. 18,D). Snout with five rostral pores; lower jaw without barbel. Northwestern Atlantic. Monotypic: *L. xanthurus*. Inhabits sandy and mud bottom in estuarine rivers and inshore waters.

*Leiostomus* Lacépède, 1803, vol. 4:439 (type-species: *Leiostomus xanthurus* Lacépède, by monotypy).

*Liostomus* Gill, 1861c:93 (invalid emendation of *Leiostomus* Lacépède 1803, therefore taking the same type-species: *Leiostomus xanthurus* Lacépède).

*Leiostomus xanthurus* Lacépède

*Leiostomus xanthurus* Lacépède 1803, vol. 4:439, pl. 10, fig. 1,

Carolina (syntypes: MNHN 7599, two specimens, 148 & 163 mm SL).

*Mugil obliquus* Mitchill, 1814 (ed. by Gill, 1896):16, New York.

*Sciaena multifasciata* LeSueur, 1822:255, east Florida (no length of the type-specimen reported).

*Leiostomus humeralis* Cuvier, 1830:141, pl. 110, New York (syntypes: MNHN 9733, one dried stuffed specimen, 146 mm SL; another syntype of Cuvier's description not located).

*Umbrina* Group

Diagnosis: Swimbladder simple carrot-shaped, without diverticula (*Sciaena* pattern, Fig. 5); sagitta oval, (*Sciaena* pattern, Fig. 18); snout with five marginal pores and three to seven upper pores; lower jaw with four pores and one barbel, an apical pore present. Two genera in this group: *Ctenosciaena* and *Umbrina*.

*Ctenosciaena*

Diagnosis: Body oblong, moderately compressed, head moderate and blunt with well-developed cavernous canals. Mouth terminal to slightly inferior, snout protruding; lower jaw with a barbel tapering to a fine

point without an apical pore at the tip (Fig. 35,A). Vertebrae 10+15=25. Sagitta oval, the ostium of the tadpole-shaped sulcus slightly in contact with the anterior margin. Snout with three upper pores. Tropical western Atlantic. Monotypic, *C. gracilicirrhus* inhabits offshore waters over hard sandy mud bottom.

*Sciaena* (*Ctenosciaena*) Fowler & Bean, 1924:15 (type-species: *Sciaena dubia* Fowler & Bean, by monotypy).

*Ctenosciaena gracilicirrhus*

*Umbrina gracilicirrhus* Metzelaar, 1919:72, fig. 24, Venezuela

(holotype: ZMA 113.103, 105 mm SL).

*Sciaena* (*Ctenosciaena*) *dubia* Fowler & Bean, 1924:16, Wilks Exploring Expedition, no locality (holotype: USNM 83309, 111 mm SL).

*Umbrina* Cuvier

Diagnosis: Body moderately elongate, back slightly arched. Head oblong, snout thick and protuberant. Mouth small, horizontal to inferior; teeth in villiform bands; gill rakers short. Vertebrae 11+14=25. Swimbladder simple carrot-shaped, without diverticula. Sagitta oval, the ostium of the sulcus reaching to anterior margin (Fig. 18,I & J). Snout with five to seven upper pores (Fig. 31,E & F); lower jaw with four pores and a short, thick barbel, an apical pore present at the tip of the anterior surface. Eastern Pacific and both sides of the Atlantic, about eight species. Four species in western Atlantic; *U. broussonetii*, *U. canosai*, *U. coroides* and *U. milliae*. Inhabits inshore and offshore waters with sandy mud bottoms and around coral reefs.

*Sciaena* Linnaeus, 1758:288 (of Artedi, 1738; in part; includes *S. cirrosa*; type-species: *Sciaena umbra*, by subsequent designation, Cuvier, 1817:297).

*Umbrina* Cuvier, 1817:297 (type-species: *Sciaena cirrosa* Linnaeus, by original designation).

*Attilus* Gistel, 1848:109 (type-species, *Sciaena cirrosa* Linnaeus, by original designation).

Remarks: The name *Chromis* in the footnote of Lacépède, 1803, volume 3, p. 546 was from the sketch of Le Pere Plumier, a missionary in Martinique, for the "grygry" or "grogro", which is *Umbrina coroides* Cuvier 1830. The name *Chromis* is older than *Umbrina* but later than *Cromis* Browne (after Jordan 1917). Also see Gilbert (1966) for more detailed synonymy.

*Umbrina broussonetii*

*Umbrina broussonetii* Cuvier, 1830:187, Jamaica (holotype: MNHN 7471, 151 mm SL).

*Umbrina canosai*

*Umbrina canosai* Berg, 1895:56, Montevideo, Uruguay to Mar del Plata Argentina (no type-specimens nor specific localities mentioned in the original description).

*Umbrina coroides*

*Umbrina coroides* Cuvier, 1830:187, pl. 117, Brazil (holotype: MNHN 5343, 174 mm SL).

*Umbrina milliae*

*Umbrina milliae* Miller, 1971:303, fig. 1, Colombia (holotype: USNM 204932, 193 mm SL).

*Larimus* Group

Diagnosis: Swimbladder carrot-shaped with or without a pair of small tube-like diverticula at the anterior end (*Sciaena* pattern, Fig. 5, E & F). Sagitta with broader anterior portion and narrower posterior portion, the sulcus not reaching to the margins (*Larimus* pattern, Fig. 20, A & B). Snout with five marginal pores, no upper pores (Fig. 31, B), lower jaw with four pores, no barbel (Fig. 34, B). *Larimus* is the only genus in this group.

*Larimus* Cuvier

Diagnosis: Body short and robust, compressed dorsal profile slightly elevated, ventral convex. Head with prominent cavernous canal; snout short. Mouth large and oblique, teeth conical in a single row on both jaws. Gill rakers, long and slender. Vertebrae 11+14=25. Other diagnostic characters as in the *Larimus* group. Tropical eastern Pacific and western Atlantic, about four species. Two western Atlantic species. *L. breviceps* and *L. fasciatus*. Inhabits estuarine and inshore open waters with sandy and muddy bottoms.

*Larimus* Cuvier, 1830:145 (type-species: *Larimus breviceps* Cuvier, by monotypy).

*Amblyscion* Gill, 1863b:165 (type-species: *Amblyscion argenteus* Gill by monotypy).

*Monosira* Poey, 1881:326 (type-species: *Monosira stahli* Poey, by monotypy).

*Larimus breviceps* Cuvier

*Larimus breviceps* Cuvier, 1830:146, pl. 140, Saint Dominique & Brazil, (syntypes: MNHN 7578, 1, 139 mm SL, St. Dominique; MNHN 7636, 1, 156 mm SL, Brazil).

*Monosira stahli* Poey, 1881:326, pl. 6, Puerto Rico (type: 190 mm long, not located in MCZ nor in USNM).

*Larimus fasciatus* Holbrook

*Larimus fasciatus* Holbrook, 1855:153, pl. 22, fig. 1, Charleston (type: not located).

*Nebris* Group

Diagnosis: Swimbladder with carrot-shaped main chamber and a pair of diverticula, which almost extend to the tapering end of the main chamber then loop back (Fig. 8) reaching the septum transversum anteriorly (*Nebris* pattern). Sagitta oval-shaped, with a notched posterior margin, very thick (Fig. 21); the ostium of the sulcus large, ovoid; the cauda J-shaped, with broad and deeply grooved distal end; snout with only two marginal pores, without upper pores; lower jaw with four minute pores and no barbel. One genus in this group: *Nebris*

*Nebris* Cuvier

Diagnosis: Body elongate, tapering posteriorly. Head extremely cavernous, interorbital space very broad, eye very small. Mouth very large, oblique, nearly vertical; teeth in narrow conical bands. Gill rakers long and slender. Vertebrae 12+13=25. Other diagnostic characters as in the group diagnosis. Tropical eastern Pacific and western Atlantic, two species. One species in the western Atlantic, *N. microps*. Inhabits sandy to muddy bottoms in coastal and estuarine waters.

*Nebris* Cuvier, 1830:149 (type-species: *Nebris microps* Cuvier, by monotypy).

*Nebris microps* Cuvier

*Nebris microps* Cuvier, 1830:149, pl. 112, Surinam (type: in Berlin, not examined).

*Micropogonias* Group

Diagnosis: Swimbladder carrot-shaped with a pair of tube-like diverticula extending anteriorly from the middle of lateral sides to septum transversum. (*Micropogonias* pattern, Fig. 9). Sagitta very heavy, irregular in shape (*Micropogonias* pattern, Fig. 26), the sulcus not reaching to margins, the ostium broad, the cauda oblique with an expanded distal end. Snout with five marginal pores and five upper pores (Fig. 31,E); lower jaw with five pores and six to eight pairs of minute barbels (Fig. 32,G). One genus in this group, *Micropogonias*.



*Micropogonias* Bonaparte

Diagnosis: Body elongate, dorsal profile slightly elevated, ventral profile nearly straight. Head conical, preopercular margin with 10 to 14 spines, two to five strong spines at the angle. Mouth inferior, teeth in villiform bands. Gill rakers short. Vertebrae 10+15=25. Other diagnostic characters as in the *Micropogonias* group. Tropical eastern Pacific and western Atlantic, about five species. Two species in western Atlantic; *M. furnieri* and *M. undulatus*. Inhabits estuarine rivers and coastal waters with sandy-muddy bottom.

*Micropogon* Cuvier, 1830:213 (type-species: *Micropogon lineatus*

Cuvier = *Umbrina furnieri* Desmarest, see remarks).

*Micropogonias* Bonaparte, 1831, 52:170 (substitute name for *Micropogon*, therefore taking the same type-species: *M. lineatus* Cuvier; preoccupied by *Micropogon* Boie, 1826:977, Aves).

*Micropogonias furnieri* (Desmarest)

*Umbrina furnieri* Desmarest 1823:182, pl. 17, fig. 3, Havana, Cuba  
(type: see remarks).

*Micropogon lineatus* Cuvier, 1830:215, pl. 119 & pl. 138 (swimbladder)  
(lectotype: MNHN 4968, 109 mm SL, also see remarks).

*Micropogon argenteus* Cuvier, 1830:218, Surinam (type: not located in MNHN).

*Ophioscion woodwardi* Fowler 1937:311, figure on p. 312, Haiti  
(holotype: ANSP 68257, 108 mm SL).

*Micropogonias undulatus* (Linnaeus)

"*Perca marina*" Catesby, 1743, "Croker", pl. 3, upper figure,  
Chesapeake Bay (non-binomial).

*Perca undulata* Linnaeus, 1766:483, South Carolina (syntypes:  
Linnaean Society, London no. 112, right side of a dried skin,  
212 mm SL and no. 113, left side of a dried skin, 241 mm SL).

*Sciaena croker* Lacépède, 1802, vol. 4:309, Carolina (type: not  
located in MNHN).

*Bodianus costatus* Mitchill, 1815:417, New York.

*Sciaena opercularis* Quoy & Gaimard, 1824:347, Rio de Janeiro.

*Micropogon lineatus* Cuvier, 1830:215 (syntypes: MNHN 7457, two  
specimens, 62.8 & 138 mm SL, New York; MNHN 7459, 273 mm SL,  
Montevideo).

Remarks: Cuvier's (1830) description of *M. lineatus* fits both  
*M. furnieri* and *M. undulatus*. The type-specimens obtained from New  
York (MNHN 7457) by Milbert are apparently *M. undulatus* (there is  
also a specimen of *Leiostomus xanthurus* 87.2 mm SL, in the same jar).  
The type-specimen from Cuba (MNHN 4968) was labeled as "type de *Umbrina*  
*fournieri* Desmarest", and the specimen illustrated on plate 119 and a  
swimbladder illustrated on plate 138 appeared to be *M. furnieri*.  
The diverticula on the swimbladder of *M. furnieri* usually originates  
further back than that of *M. undulatus* (Fig. 9, C & D). MNHN 7456  
from Montevideo contains two syntypes in the jar, the smaller specimen,  
159 mm SL is *M. furnieri* and the larger one, 273 mm SL is *M.*  
*undulatus*. In this case, the MNHN 4968, 109 mm SL of Cuba is

selected as the lectotype. Then the type-species of *Micropogonias* is designated as *M. furnieri* (Desmarest). Jordan (1917) designated *M. lineatus* Cuvier = *Perca undulatus* Linnaeus as type-species of *Micropogonias*. His reason was that Cuvier's (1830) description on p. 215, mentioned the New York Specimens (*M. undulatus*) first.

Some morphometrics and meristics of the lectotype MNHN 4968 are listed as follows:

Morphometrics (in mm)	
TL	110
SL	109
Snout to anal fin origin	72.6
Snout to dorsal fin origin	40.5
Snout to pectoral fin origin	34.7
Snout to pelvic fin origin	36.0
Maximum depth	28.2
Head length	33.6
Snout length	9.7
Eye diameter	8.5
Interorbital width	8.1
Pectoral fin lengths	19.5 (left) - 22.5 (right)
Pelvic fin lengths	16.0 (left) - 19.2 (right)
Length of spinous dorsal fin base	20.2
Length of soft dorsal fin base	42.7
Length of anal fin base	12.5
Length of second anal fin spine	13.4
Depth of caudal peduncle	9.2

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Meristics

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D. X+I, 27

A. II, 8

Pectoral rays, 18-17

Lateral gill rakers on the first right gill arch;

8 (upper arm) + 15 (lower arm)

Medial (or inner) gill rakers on the first right gill arch;

3 (upper arm) + 10 (lower arm)

Lateral line scales (a row above the lateral line);

Left side: 52 scales

Right side: 48 scales

Transverse lateral line scales;

from soft dorsal fin origin to lateral line

vertical: 9

oblique: 12

from lateral line to anal fin origin

vertical: 12

oblique: 15

Other nominal species of this genus were discussed by Roux (1973). The true identity of *M. argenteus* Cuvier and *Sciaena opercularis* Quoy & Gaimard are not certain at present. The distinguishing characters of the two species of *Micropogonias* (see also "Key to the species of *Micropogonias*") used here are only valid for adult specimens. The sagitta morphology of *M. furnieri* usually occurs south of the Antilles and off South America and *M. undulatus* have been reported from the Gulf of Maine to Argentina.

*Sciaenops* Group

Diagnosis: Swimbladder carrot-shaped with a pair of short tube-like diverticula at the anterior lateral corners of the swimbladder (*Sciaenops* pattern, Fig. 11). In large specimens, a pair of "sac-like" projections which consist of numerous labyrinth chambers (Fig. 11,C) present on the dorsal surface of the swimbladder; a hole present between the third and fourth lateral ribs on both sides of the body wall to receive the paired "sac-like" projections. Sagitta rectangular, sulcus not reaching to the margins (*Larimus* pattern). Snout with five marginal and five upper pores (Fig. 33,F); lower jaw with five pores and no barbel (Fig. 34,F). *Sciaenops* is the only genus in this group.

*Sciaenops*

Diagnosis: Body elongate, robust, back slightly elevated, ventral surface flat. Head long and low; snout long. Mouth inferior; teeth villiform in bands. Gill rakers short. Vertebrae 10+15=25. Other diagnostic characters as in *Sciaenops* group. Subtropical northwestern Atlantic. Monotypic: *S. ocellata*. Inhabits estuarine and inshore waters with sandy bottoms.

*Sciaenops* Gill, 1963a:30 (type-species: *Perca ocellata* Linnaeus by monotypy).

*Sciaenops ocellata* Linnaeus

*Perca ocellata* Linnaeus, 1766:483, South Carolina (syntypes: Linnaean Society, London, no. 106, one dried right side skin, 394 mm TL and no. 107, one dried right side skin, 399 mm TL).

*Lutjanus triangulum* Lacépède, 1803, vol. 4:181, figure in vol. 3, pl. 24, fig. 3, Sumatra (type: not located in MNHN).

*Sciaena imberbis* Mitchill, 1815:411, New York.

*Pogonias* Group

Diagnosis: swimbladder with very complicated diverticula along the side of main chamber (Fig. 10). Sagitta semi-circular in shape, the "tadpole-shaped" sulcus not reaching to the margins, (Fig. 19,A). Snout with five rostral pores and five marginal pores; (Fig. 31,E); lower jaw with five to seven pores and 12 to 13 pairs of small barbels (Fig. 32,G). Monotypic: *Pogonias*.

*Pogonias* Lacépède

Diagnosis: Body oblong and deep, dorsal profile elevated, ventral nearly straight. Head large, snout conical not projecting in front of upper jaw. Mouth moderate, horizontal and slightly inferior; teeth in villiform band. Gill rakers very short. Vertebrae 10+14=24. Complexity of swimbladder increases with size (Fig. 10); other diagnostic characters as in group diagnosis. Tropical and temperate western Atlantic. Monotypic: *Pogonias cromis*. Inhabits coastal waters especially in areas with large river runoffs over sandy and sandy-muddy bottoms.

*Pogonias* Lacépède, 1802:137 (type-species: *Pogonias fasciatus*

Lacépède = *Labrus cromis* Linnaeus, by monotypy).

*Pogonathus* Lacépède, 1803, vol. 5:120 (type-species: *Pogonathus*

*courbina* Lacépède = *Labrus cromis* Linnaeus, by subsequent designation of Jordan, 1917:65).

*Pogonias cromis* (Linnaeus)

*Labrus cromis* Linnaeus, 1766:479, Carolina.

*Pogonias fasciatus* Lacépède, 1802, vol. 3:137, figure in vol. 2, pl.

16, fig. 2, Brazil (syntypes: MNHN 7461, one dried stuffed specimen, 103 mm TL; MNHN 7460, one dried stuffed specimen, 181 mm SL).

*Pogonathus courbina* Lacépède, 1803, vol. 5:120, Rio de La Plata (from the manuscript of Commerson).

*Mugil grunniens* Mitchill, 1814:15, New York.

*Mugil gigas* Mitchill, 1814:16, New York.

*Sciaena fusca* Mitchill, 1815:409, New York.

*Cynoscion* Group

Diagnosis: Swimbladder with a pair of well-developed horns (Fig. 12) from anterior-lateral corners of main chamber, tapering forward to the back of skull. Sagitta elongate oval with smooth or notched margins. Snout with only two (or no) marginal pores and without upper pores (Fig. 31,A); lower jaw without pores or barbels (Fig. 32,A). Four genera in this group: *Cynoscion*, *Isopisthus*, *Macrodon* and *Plagioscion*.

*Cynoscion* Gill

Diagnosis: Body elongate, compressed, predorsal outline nearly straight, ventral evenly arched. Head conical, snout pointed. Mouth large, oblique, lower jaw projecting; teeth sharp in narrow ridges, tip of upper jaw with two large canines (no canine teeth in *C. steindachneri*), tapering from base to tip. Preopercular margin membranous or ciliated. Gill rakers short to long slender. Vertebrae usually  $13(12)+12(13)=25$ , except  $15+12=27$  in *C. nothus*. Swimbladder with a pair of anterior horns, some straight and some curved (Fig. 13). Sagitta oval, the ostium of the sulcus usually not reaching to the anterior margin in most species (Fig. 23); the cauda usually short and expanded. Eastern Pacific and western Atlantic, about 20 species. This genus including four northwestern Atlantic species: *C. arenarius*, *C. nebulosus*, *C. nothus* and *C. regalis*; and eight Caribbean and southwestern Atlantic species: *C. acoupa*, *C. jamaicensis*, *C. leiarchus*, *C. microlepidotus*, *C. similis*, *C. steindachneri*, *C. striatus* and *C. virescens*. Inhabits estuarine and inshore waters.

*Cestreus* Gronow, 1754 (ed. by Gray, 1854):49 (type-species: *Cestreus carolinensis*, by monotypy, preoccupied by *Cestreus* Klein, 1749, type-species: see remarks and not of *Cestreus* McClelland, 1842, type-species: *Cestreus minimus* = *Atherina donius* Hamilton).

*Cynoscion* Gill, 1861b:81 (type-species: *Otolithus regalis* Cuvier, = *Johnius regalis* Bloch & Schneider, by original designation).

*Apseudobranchus* Gill, 1862:18 (type-species: *Otolithus toe-roe* Cuvier, by original designation).



*Archoscion* Gill, 1862:18 (type-species: *Otolithus analis* Jenyns, 1842, by original designation).

*Atractoscion* Gill, 1862:18 (type-species: *Otolithus aequidens* Cuvier by original designation).

*Buccone* Jordan & Everman, 1895:394 (type-species: *Cestreus praedatorium* Jordan & Gilbert, by original designation).

*Symphysoglyphus* Miranda Ribeiro, 1915, Sciaenidae: 43 (type-species: *Otolithus bairdi* Steindachner, by monotypy).

*Eriscion* Jordan & Evermann, 1927:506 (type-species: *Cynoscion nebulosus* Cuvier, by monotypy).

Remarks: In Klein (1749), page 23 and 24, species of "Mugil" were described and "Sciaena" was described in a footnote. Type-species of *Sciaena*: *Mugil cephalus* Linnaeus = "Cestreus dorso repando" Klein, was subsequently designated by Walbaum (1793).

*Cynoscion acoupa* (Lacépède)

*Cheilodipterus acoupa* Lacépède, 1800, vol. 2:540; Cayenne (holotype: MNHN 5502, 262 mm SL, Surinam).

*Lutjanus cayennensis* Lacépède, 1803, vol. 4:196 (syntypes: MNHN 5502, one specimen, 262 mm SL, Surinam; MNHN A.5617, two specimens, 522 & 567 mm SL, Martinique; MNHN A.4562, two specimens, 278 & 288 mm SL, Brazil).

*Otolithus toe-roe* Cuvier, 1830:72, pl. 103 (syntypes: MNHN 5500, one specimen, 175 mm SL, Surinam; MNHN 4616, one specimen, 347 mm SL, Cayenne; MNHN A.4518, two specimens, 493 & 485 SL, Cayenne).

*Cynoscion maracaiboensis* Schultz, 1949:160, fig. 20, Maracaibo,  
Venezuela (holotype: USNM 12742, 251 mm SL).

*Cynoscion arenarius* Ginsburg

*Cynoscion arenarius* Ginsburg 1929:83, Texas (holotype, USNM 89385,  
245 mm SL).

*Cynoscion jamaicensis* (Vaillant & Bocourt)

*Otolithus jamaicensis* Vaillant & Bocourt, 1915:156, pl. 6, fig. 1,  
Jamaica (holotype: MNHN A.557, 205 mm SL).

*Archoscion petranus* Miranda Ribeiro, 1915, Sciaenidae: 42, Brazil  
(type: not listed in Miranda Ribeiro, 1953).

*Cynoscion leiarchus* (Cuvier)

*Otolithus leiarchus* Cuvier, 1830:78, Cayenne, Brazil (syntypes: all  
from Brazil, MNHN 5503, 2 specimens 152 & 234 mm SL; MNHN A.2690,  
1 specimen, 187 mm SL; MNHN A.5422, one dried stuffed specimen,  
112 mm SL).

*Cynoscion microlepidotus* (Cuvier)

*Otolithus microlepidotus* Cuvier, 1830:79, Surinam (type: in Berlin,  
not examined).

*Otolithus bairdi* Steindachner, 1879:40, pl. 1, fig. 2, Santos  
(NHMV 51130, 152 mm SL, donated by Steindachner, is  
probably the holotype).

*Cynoscion nebulosus* (Cuvier)

*Labrus squetaeque* var. *maculatus* Mitchill, 1815:396, New York  
(not *Labrus maculatus* Bloch).

*Otolithus nebulosus* Cuvier, 1830:79, no locality (holotype: MNHN  
7527, 233 mm SL).

*Otolithus carolinensis* Valenciennes, 1833:475, South Carolina  
(holotype: MNHN 7507, 335 mm SL).

*Otolithus drummondi* Richardson, 1836:70, New Orleans.

*Cynoscion maculatum* Jordan & Gilbert, 1882:285, Pensacola,  
Galveston (after Mitchill).

*Cynoscion nothus* (Holbrook)

*Otolithus nothus* Holbrook, 1855:134, pl. 19, fig. 1, South  
Carolina.

*Cynoscion regalis* (Bloch & Schneider)

*Cestreus carolinensis* Gronow, 1754 (ed. by Gray, 1854):49 (holotype:  
BMNH 1853, 11, 12, 42, a dried skin, 344 mm TL, not of  
*O. carolinensis* Valenciennes).

*Johnius regalis* Bloch & Schneider, 1801:75, New York.

*Roccus comes* Mitchill, 1814:26, New York.

*Labrus squeteague* Mitchill, 1815:396, pl. 2, fig. 1, New York.

*Otolithus thalassinus* Holbrook, 1855:132, pl. 18, fig. 2, Charleston  
South Carolina.

*Cynoscion similis* Randall & Cervigon

*Cynoscion similis* Randall & Cervigon, 1968:170, fig. 2, Isla de  
Margarita, Venezuela (holotype: USNM 201382, 284 mm SL).

*Cynoscion steindachneri* (Jordan)

*Cestrus steindachneri* Jordan in Jordan & Eigenmann, 1889:372, Curuca  
(holotype: MCZ 10922, 318 mm SL).

*Cynoscion striatus* (Cuvier)

"Quatucupa" Marcgrave, 1648 (translated by Taunay, 1914):177  
(non-binomial).

*Otolithus striatus* Cuvier, 1839:173 (after Marcgrave).

*Otolithus quatuccupa* Cuvier, 1830:75, Montevideo (syntypes: MNHN  
7517, two specimens, 366 & 358 mm SL).

*Cynoscion virescens* (Cuvier)-

*Otolithus virescens* Cuvier, 1830:72, Surinam (type: in Berlin, not examined).

*Otolithus microps* Steindachner, 1879:38 (type: not located in NHMV, Vienna).

*Isopisthus* Gill

Diagnosis: Body elongate, compressed, dorsal profile nearly straight, ventral evenly arched. Head pointed, rather compressed. Mouth large, oblique, lower jaw projecting; teeth in narrow band, tip of upper jaw with two enlarged canine-like teeth. Preopercular margin membranous with fine ciliae. Spinous and soft portions of dorsal fin well-separated. Gill rakers moderate. Vertebrae 11+14=25. Swimbladder with a pair of tube-like anterior horns (Fig. 12,B). Sagitta nearly oval, the ostium of the sulcus reaching to anterior margin (Fig. 25,A); the cauda oblique, slightly bent at the distal end. Tropical eastern Pacific and western Atlantic, two species. One western Atlantic species: *I. parvipinnis*. Inhabits inshore waters with sandy to muddy bottoms.

*Isopisthus* Gill, 1862:18 (type-species: *Ancylodon parvipinnis* Cuvier by monotypy).

*Paraplesichthys* Bleeker, 1876 (from Kaup's manuscript, 1862): 335

(type-species: *Isopisthus parvipinnis* Gill = *Ancylodon parvipinnis* Cuvier, by monotypy).

*Isopisthus parvipinnis* (Cuvier)

*Ancylodon parvipinnis* Cuvier, 1830:84, pl. 105, Cayenne (syntypes:

MNHN 745, three specimens, 100-140 mm SL).

*Isopisthus affinis* Steindachner, 1879:43, pl. 2, fig. 2, Port Alegre,

Brazil (syntypes: NHMV 15190, two specimens, 168 & 172 mm SL).

*Isopisthus harroweri* Fowler, 1916:402, fig. 3, Colon, Panama (holotype:

ANSP 45236, 145 mm SL).

*Macrodon* Schinz

Diagnosis: Body elongate, moderately compressed; dorsal profile slightly arched, ventral evenly arched. Head pointed, compressed. Mouth large, oblique; lower jaw projecting in front of upper jaw; teeth in narrow ridge, upper jaw with a pair of much enlarged canine (Fig. 37,A), lance-shaped; lower jaw with several enlarged canines at the tip, exposed externally when mouth closed. Preopercular margin membranous. Gill rakers slender. Vertebrae 13+12=25. Swimbladder with a pair of anterior fork-like horns (Fig. 12,C). Sagitta oval, lateral margins concave or notched (Fig. 25,B); the ostium of the sulcus reaching to the anterior margin; the cauda nearly straight with an expanded distal end (*Macrodon* pattern). Tropical eastern Pacific and western Atlantic, two species. One western Atlantic species: *M. ancylodon*. Inhabits inshore waters.

*Ancylodon* Bosc, 1816:497 (type-species: "Lonchures" Schneider = *Lonchurus ancylodon* Bloch & Schneider, by original designation, not of *Ancylodon* Illiger).

"Ancylodons" Cuvier, 1817:299, vernacular name Latinized as *Ancylodon* by Oken, 1817:1182 (type-species: *Lonchurus ancylodon* Bloch & Schneider, by original designation, not of *Ancylodon* Illiger).

*Macrodon* Schinz, 1822:482 (substitute name for *Ancylodon*, therefore taking the same type-species: *L. ancylodon* Bloch & Schneider; preoccupied by *Ancylodon* Illiger 1811, a genus of mammals).

*Nomalus* Gistel, 1848, p. VIII (substitute name for *Ancylodon*, therefore taking the same type-species: *L. ancylodon* Bloch & Schneider).

*Sagenichthys* Berg, 1895:52 (substitute name for *Ancylodon*, therefore taking the same type-species: *L. ancylodon* Bloch & Schneider).

*Macrodon ancylodon* (Bloch & Schneider)

*Lonchurus ancylodon* Bloch & Schneider 1801:102, pl. 25, Surinam.

*Ancylodon jaculidens* Cuvier, 1830:81, Cayenne (syntypes: MNHN 7451, one specimen, 212 mm SL: MNHN 7454, one specimen, 144 mm SL).

*Ancylodon atricauda* Gunther, 1880:12, Rio de La Plata (type: not located in BMNH).

*Lonchiurus* Group

Diagnosis: Swimbladder carrot-shaped with a pair of anterior horns, each horn with a long posterior branch (*Lonchiurus* pattern, Fig. 14); sometimes the swimbladder is reduced in size. Sagitta oval elongate to rectangular in shape, the "tadpole-shaped" sulcus elongate;

the ostium reaching to the margin and the cauda long, J-shaped (*Lonchiurus* pattern, Fig. 24). Snout with five marginal pores and three to five upper pores (Fig. 31, D & E); lower jaw with two to many pairs of barbels (Fig. 32, F & H). Two genera of western Atlantic Sciaenidae in this group; *Lonchiurus* and *Paralonchurus*.

#### *Lonchiurus* Bloch

Diagnosis: Body long and rounded; dorsal profile evenly arched, ventral nearly flat. Pectoral fin very long, head conical, low and broader than body; snout projecting. Mouth inferior; teeth conical in bands. Gill rakers short. Vertebrae 11+18=29. Swimbladder much reduced in size, but retaining typical shape with anterior horns more prominent, posterior end of the main chamber tapering into a fine point (Fig. 14,A). Snout with five upper pores; lower jaw with four pores and two barbels (Fig. 31,F), longer than eye diameter.

Distribute in tropical west Atlantic. Monotypic: *L. lanceolatus*.

Inhabits sandy and muddy bottom along the coast and in estuarine waters.

*Lonchiurus* Bloch, 1793:143 (type-species: *Lonchiurus barbatus*

Bloch, by monotypy).

*Lonchurus* Bloch & Schneider, 1801:102 (invalid emendation of *Lonchiurus*).

#### *Lonchiurus lanceolatus* (Bloch)

*Perca lanceolata* Bloch, 1788:383, fig. 3, India (= West Indies).

*Lonchiurus barbatus* Bloch, 1793:144, pl. 360, fig. 1, Surinam.

*Lonchurus depressus* Bloch & Schneider, 1801:102, Surinam.



*Paralonchurus* Bocourt

Diagnosis: Body elongate, slightly rounded; dorsal profile slightly elevated, ventral nearly flat. Pectoral fin long or short. Head broad, nape slightly convex; snout projecting. Mouth small and inferior, teeth in conical bands. Gill rakers short. Total vertebrae 25 or 29 (see "Key to the species of *Paralonchurus*"). Swimbladder well-developed (Fig. 14,B & C). Sagitta elongate, the ostium of the sulcus reaching to anterior margin; the cauda, J-shaped (Fig. 24,B & C). Snout with three upper pores; lower jaw with 12 to 16 pairs of minute barbels, anterior three pairs form a tuft at the symphysis of lower jaw and other barbels along the rami of lower jaw (Fig. 32,H). Tropical eastern Pacific and western Atlantic, about five species. Two species from the western Atlantic in this genus: *P. brasiliensis* and *P. elegans*. Inhabits estuarine and inshore waters with sandy to muddy bottoms.

*Paralonchurus* Bocourt, 1869:21 (type-species: *Paralonchurus petersi* Bocourt, by monotypy).

*Polycirrhus* Bocourt, 1869:22 (type-species: *Polycirrhus dumerili* Bocourt by monotypy, not *Polycirrhus* Grube).

*Polyclemus* Berg, 1895:54 (substitute name for *Polycirrhus*, therefore taking the same type-species: *P. dumerili*; preoccupied by *Polycirrhus* Grube, 1850, a genus of worms, quote from Neave, 1940).

*Paralonchurus* (*Zonoscion*) Jordan & Evermann, 1895:401 (type-species: *Polycirrhus rathbuni* Jordan & Bollman, 1889, by monotypy).

*Paralonchurus* (*Zaclemus*) Gilbert in Jordan & Evermann, 1895: 402 (type-species: *Paralonchurus goodei* Gilbert, manuscript name, by monotypy).

*Paralonchurus brasiliensis* (Steindachner)

*Genyanemus brasiliensis* Steindachner; 1875a:476, Para, Santos. (type: not located in NHMV).

*Micropogon ornatus* Günther, 1880:13, pl. 7, fig. A, Rio de La Plata (holotype: BMNH 1879.5. 14. 289, 175 mm SL).

*Paralonchurus rathbuni* Puyo, 1949:215, fig. 114 (not Jordan & Bollman, 1889:162).

*Paralonchurus elegans* Boeseman

*Paralonchurus elegans* Boeseman, 1948:3, Surinam (holotype: RMNH 390, 200 mm SL).

Remarks: *P. elegans* and *L. lanceolatus* have very similar sagittae (Fig. 24) and both have a long black pectoral fin. *P. brasiliensis* has a total vertebrae count of 29 as in *L. lanceolatus*. *P. brasiliensis* and *P. elegans* both have well-developed swimbladders and many minute barbels on the chin. The interrelationship of these three species could not be determined without comparative material of the eastern Pacific species of this group. The reduced size of the swimbladder in *L. lanceolatus* (Fig. 14,A) has been viewed as an important diagnostic character (Günther, 1860), and this may also link this group with *Menticirrhus*. *Paralonchurus brasiliensis* may be considered to belong in a separate genus, *Polyclemus* as in Berg (1895). The type-species of *Paralonchurus* (*P. petersi*) and *Polyclemus* (*P. dumerili*) are eastern Pacific species. Study of these species will be necessary to clarify the generic status of these fishes.

*Menticirrhus* Group

Diagnosis: Swimbladders atrophy as the fish grow, only a vestige in adult. Sagitta oval, elongate, the sulcus not reaching to the margin (*Menticirrhus* pattern, Fig. 22,B). Snout with three upper pores and five marginal pores, rostral fold deeply indented (Fig. 33,D); lower jaw with a short, rigid barbel, knob-like and four pores (Fig. 35,B). Anal fin with only one weak spine. One genus in this group; *Menticirrhus*.

*Menticirrhus* Gill

Diagnosis: Body elongate, rounded, dorsal profile slightly arched, ventral nearly flat. Head conical, low and broad, snout projecting. Mouth small and inferior; teeth in villiform bands. Gill rakers short and tubercle-like. Vertebrae 10+15=25. Other diagnostic characters as in the group diagnosis. Tropical and temperate eastern Pacific and western Atlantic, nine species. Three species from the western Atlantic: *M. americanus*, *M. littoralis* and *M. saxatilis*. Inhabits shallow coastal waters with sandy bottoms, juveniles often found in estuaries.

*Menticirrhus* Gill, 1861b:86 (type-species: *Perca alburnus* Linnaeus = *Cyprinus americanus* Linnaeus by original designation).

*Cirrimens* Gill, 1862:17 (type-species: *Umbrina ophiocephalus* Jenyns, 1842:45, by original designation).

*Menticirrhus* (*Umbrula*) Jordan and Eigenmann, 1889:424 (type-species: *Umbrina littoralis* Holbrook, by subsequent designation of Jordan and Everman, 1898:1469).

*M. americanus* (Linnaeus)

"Alburnus Americanus" Catesby, 1743, "Carolina whiting", pl. 12, lower figure, Charleston (non-binomial, also see remarks).

*Sciaena alburnus* Gronow, 1754 (ed. by Gray, 1854):108 (non *Perca alburnus* Linnaeus, holotype BMNH 1853. 11. 12. 75, a dried skin, 228 mm TL).

*Cyprinus americanus* Linnaeus, 1758:321 (based on Catesby).

*Perca alburnus* Linnaeus, 1766:482, South Carolina (holotype: Linnaean Society, London, no. 111, a dried left side of skin, 282 mm TL).

*Umbrina arenata* Cuvier, 1830:190, Brazil (syntype: MNHN 7500, one specimen, 76.4 mm SL; MNHN 7472, one specimen, dried and stuffed, in ethanol, 341 mm TL).

*Umbrina martinicensis* Cuvier, 1830:186, Martinique (holotype: MNHN 7498, 193 mm SL).

*Umbrina gracilis* Cuvier, 1830:189, Brazil (syntypes: MNHN 9037, one specimen, 97 mm SL; and MNHN 44 one specimen, 156 mm SL).

*Umbrina phalaena* Girard, 1859:167, Brazos Santiago, Texas (syntypes: USNM 154721, 88.3 mm SL; other syntypes not located).

*Umbrina januarina* Steindachner, 1877:170, Rio Janeiro (type-specimens, not located).

Remarks: The description of "Alburnus americanus" by Catesby (1743) stated that "under the lip having five or six fleshy barbels, resembling teeth hanging to it on the outside. It had one small fin on the middle of the back". The lower figure of plate 12, "Alburnus" agrees with this description. Except for these features, the general appearance of the figure is a species of *Menticirrhus* but it is

impossible to identify it with any of the *Menticirrhus* species. The name *Cyprinus americanus* Linnaeus, 1758 is based on Catesby. The holotype of *Perca alburnus* Linnaeus, 1766, is designated here as neotype of *Cyprinus americanus* Linnaeus, 1758, to restrict the application of this name to the species to which it is usually applied.

A specimen, MNHN 9037, was catalogued as a type-specimen of *Umbrina gracilis* Cuvier 1830, in MNHN, Paris and was identified by Irwin (1970) as *M. littoralis* (Holbrook). It is not a type of *U. gracilis*, because the locality and the donor of the specimen does not agree with the original description of Cuvier (Dr. M. Bauchot, MNHN, Paris, personal communications).

*Menticirrhus littoralis* (Holbrook)

*Umbrina littoralis* Holbrook, 1855:142, South Carolina (holotype: ANSP 11567, 198 mm SL).

*Menticirrhus saxatilis* (Bloch & Schneider)

*Johnius saxatilis* Bloch & Schneider, 1801:75, New York. (holotype: HU 8792, Humboldt-Universität, Berlin, a stuffed skin, dried in poor condition, from Irwin, 1970).

*Sciaena nebulosa* Mitchill, 1815:408, pl. 3, New York.

*Menticirrhus focaliger* Ginsburg, 1952:97, St. Joseph Bay, Florida (holotype: USNM 144101, 104 mm SL).

*Menticirrhus atlanticus* McFarland, 1963:98, Mustang Island, Texas (Nomen nudum).

*Stellifer* Group

Diagnosis: Swimbladder separated into two chambers by a constriction; the anterior one yoke-shaped and the posterior one simple carrot-shaped (*Stellifer* pattern, Fig. 15). Sometimes a pair of diverticula present on the anterior chamber posterior-laterally. Otolith with both sagitta and lapillus enlarged (*Stellifer* pattern, Fig. 27). Snout with five marginal pores and two to five upper pores (Fig. 31,C to F); lower jaw with four to six pores and without barbels (Fig. 32,B to D). Four genera of western Atlantic sciaenids in this group; *Bairdiella*, *Odontoscion*, *Ophioscion* and *Stellifer*.

Genus *Bairdiella*

Diagnosis: Body oblong, compressed; dorsal profile slightly elevated, ventral nearly straight. Head usually with strong spines at the angle of preopercular margin. Mouth terminal, slightly oblique; upper jaw teeth in conical band, lower jaw teeth in a narrow ridge. Gill rakers long and slender. Vertebrae 12+13=25. Swimbladder in two chambers, without diverticula on anterior chamber (Fig. 15,A). Sagitta triangular; the ostium of the sulcus incomplete; lapillus more than 1/2 the size of sagitta (Fig. 27,C). Snout with five marginal and three upper pores; lower jaw with five or six pores the median pair of pores in six pored form very closely set (Fig. 32,C & D). Tropical eastern Pacific and western Atlantic, about six species. Four species of western Atlantic Sciaenidae in this genus: *B. batabana*, *B. chryoura*, *B. ronchus* and *B. sanctaeluciae*.

*Bairdiella* Gill, 1861b:83 (type-species: *Bodianus argyroleucus* Mitchill, by original designation).

*Bairdiella* (*Nector*) Jordan & Evermann, 1898:1432 (type-species: *Bairdiella chrysoleuca* Günther (by original designation)).

*Corvula* Jordan & Eigenmann, 1889:377 (type-species: *Johnius batabanus* Poey, by original designation).

*Bairdiella batabana* (Poey)

*Johnius batabanus* Poey, 1860, 2:184, Batabano, Cuba (holotype: MCZ 21957, 189 mm SL).

*Corvula sialis* Jordan & Eigenmann, 1889:379, Key West, Florida (holotype: USNM 26575, 132 mm SL).

*Bairdiella chrysoura* (Lacépède)

*Perca punctata* Linnaeus, 1766:482, South Carolina (a junior homonym of *Perca punctatus* of Linnaeus, 1758:291, = *Bodianus fulvus punctatus*; syntypes?: Linnaean Society of London, no. 108, a left side dried skin, 144 mm TL and no. 109, a left side dried skin, 153 mm TL).

*Dipterodon chrysourus* Lacépède, 1803, vol. 4:166, South Carolina (type: not located in MNHN).

*Bodianus argyro-leucus* Mitchill, 1815:417, pl. 6, fig. 9, New York.

*Bodianus exiguus* Mitchill, 1815:419, New York.

*Bodianus pallidus* Mitchill, 1815:420, New York.

*Homoprion xanthurus* Holbrook, 1855:170, pl. 24, fig. 2, South Carolina (not *Leiostomus xanthurus*, Lacépède).

*Bairdiella ronchus* (Cuvier)

*Corvina ronchus* Cuvier, 1830:107 (tyntypes: MNHN 95, two specimens, 123 & 149 mm SL, St. Dominique; MNHN 5345, two specimens, 144 & 156 mm SL, Surinam, MNHN 5345, one dried stuffed specimen, 247 mm SL, Maracaibo?).

*Corvina subaequalis* Poey, 1875:58, Cuba (holotype: 245 mm TL, not located in MCZ nor in USNM).

*Sciaena bedoti* Regan, 1905:391, pl. 6, fig. 1, Cuba (syntypes; BMNH 1905. 3.182, one specimen, 155 mm SL; Museum d'Histoire Naturelle, Geneve, no. 678.1, not examined).

*Bairdiella verae-crucis* Jordan & Dickerson, 1908:17, fig. 1, Vera Cruz (holotype; USNM 61676, 194 mm SL).

*Bairdiella sanctae-luciae* (Jordan)

*Corvula sanctae-luciae* Jordan, 1889:649, Port Castries, St. Lucia (holotype: USNM 4173, 105 mm SL).

*Odontoscion* Gill

Diagnosis: Body oblong, compressed; profile evenly arched. Head conical; snout short, blunt; eye large. Mouth large, terminal, slightly oblique; enlarged conical teeth in a single row on both jaws, two teeth at the tip of lower jaw large canine-like (Fig. 37,C). Gill rakers long and stiff. Vertebrae 12+13=25. Swimbladder with two chambers, anterior one yoke-shaped and posterior one carrot-shaped, without diverticula (Fig. 15,A). Sagitta and lapillus both enlarged, only a trace left of the ostium of the sulcus (Fig. 27,D). Snout



with five marginal pores and two upper pores (Fig. 31,C); lower jaw with four pores and no barbel (Fig. 34,C). Tropical eastern Pacific and western Atlantic, two species. One species in western Atlantic: *O. dentex*. Inhabits coral reefs and offshore waters over sandy bottoms.

*Odontoscion* Gill, 1862:18 (type-species: *Corvina dentex* Cuvier, by original designation).

*Odontoscion dentex* (Cuvier)

*Corvina dentex* Cuvier, 1830:139, pl. 109, St. Dominica (holotype: MNHN 144, 114 mm SL).

*Ophioscion* Gill

Diagnosis: Body robust, dorsal profile elevated, ventral slightly convex. Head broad, conical top with prominent cavernous canals (Fig. 39,A); bony interorbital width 3.5 or more in head. Mouth small, inferior, teeth conical in bands. Gill rakers short. Vertebrae 11+14=25. Swimbladder with two chambers, anterior one yoke-shaped and posterior one carrot-shaped, both without diverticula (Fig. 15,A). Both sagitta and lapillus enlarged; the ostium of the "tadpole-shaped" sulcus incomplete (Fig. 27,A). Snout with three large and two to four minute upper pores, and five marginal pores, rostral fold deeply indented (Fig. 32,B). Tropical eastern Pacific and western Atlantic, about 5 species. Three nominal species of this genus from the western Atlantic recognized here: *O. adustus*, *O. panamensis* and *O. punctatissimus*. Inhabits inshore waters over sandy to muddy bottoms.

*Ophioscion* Gill, 1863b:164 (type-species: *Ophioscion typicus* Gill,  
by monotypy).

*Ophioscion* (*Sigmurus*) Gilbert, in Jordan & Evermann, 1898:1452  
(type-species: *Corvina vermicularis* Günther by monotypy).

*Ophioscion adustus* (Agassiz)

*Corvina* (*Sciaena*) *adusta* Agassiz, 1829:126, pl. 70, Montevideo (type:  
not located in MCZ).

Remarks: Schultz (1945) indicated that the description of *Corvina adusta* by Agassiz (1829) did not agree with the fish figured by Spix (Agassiz, 1829). He compared the descriptions of *Ophioscion adustus* of several authors, then suggested it probably was a senior synonym of *Ophioscion woodwardi* Fowler, 1937 (= *Micropogonias furnieri*). The true identity of this species is doubtful.

*Ophioscion panamensis* (Schultz)

*Ophioscion panamensis* Schultz, 1945:134, fig. 8, Fox Bay, Colon,  
Panama (holotype: USNM 122612, 52 mm SL).

*Ophioscion punctatissimus* Meek & Hilderbrand

*Ophioscion punctatissimus* Meek & Hilderbrand, 1925:644, pl. 68,  
Panama (holotype: USNM 81766, 131 mm SL, Canal Zone).

Remarks: The type-specimens of *O. panamensis* are all juveniles (11, 23.3-52 mm SL) and their true identity will be in doubt until the morphology of the swimbladder can be examined. No specimens have been reported except the types.

*Stellifer* Oken

Diagnosis: Body robust, back elevated. Head broad, rather flat on top with apparent cavernous canals (Fig. 39,A); interorbital wide, bony interorbital width 3.5 or less in head. Mouth moderate to large, inferior to oblique (Fig. 38); teeth in conical or villiform bands. Gill rakers usually long and slender, some moderate to short. Vertebrae 11+14=25. Swimbladder in two chambers, the anterior chamber yoke-shaped with a pair of diverticula, tube-like or bulb-like (Fig. 16), variation also found within the species and the posterior chamber in simple carrot-shape. Both sagitta and lapillus enlarged, the ostium of sulcus on sagitta incomplete (Fig. 28). Snout with five marginal pores, the median one rounded, separate from rostral fold and arranged rhomboidally with three upper pores at the tip of snout (Fig. 1,B); lower jaw with four to six pores, no barbel (Fig. 26). Tropical eastern Pacific and western Atlantic, about 15 species. Nine species of western Atlantic sciaenids belong to this genus: *S. brasiliensis*, *S. colonensis*, *S. griseus*, *S. lanceolatus*, *S. microps*, *S. naso*, *S. rastrifer*, *S. stellifer* and *S. venezuelae*. Inhabits inshore waters of sandy bottoms, also in estuaries and around coral reefs.

"Stelliféres" Cuvier, 1817:283, vernacular name Latinized as *Stellifer* by Oken, 1817:1182 (type-species: *Bodianus stellifer* Bloch, by original designation).

*Stelliferus* Stark, 1828:459 (type-species: *Stellifer capensis* Stark = *Bodianus stellifer* Bloch, by monotypy).

*Homoprion* Holbrook, 1855:168 (type-species: *Homoprion lanceolatus* Holbrook, by subsequent designation of Gill, 1861b:83).

*Stellifer* (*Zestis*) Gilbert in Jordan & Evermann, 1898:1439 (type-species *Stellifer oscitans* Jordan & Gilbert, by original designation).

*Stellifer* (*Zestidium*) Gilbert in Jordan & Everman, 1898:1439 (type-species: *Stellifer illecebrosus* Gilbert, by original designation).

*Stellifer* (*Stellicarens*) Gilbert in Jordan & Evermann, 1898:1439 (type-species: *Stellifer zestocarus* Gilbert, by original designation).

*Stellifer brasiliensis* (Schultz)

*Ophioscion brasiliensis* Schultz, 1945:128, fig. 6, Santos, Brazil (holotype: USNM 87742, 77 mm SL).

*Stellifer colonensis* Meek & Hilderbrand

*Stellifer colonensis* Meek & Hilderbrand, 1925:623, pl. 46, fig. 1, Mindi, Panama (holotype: USNM 81729, 99 mm SL).

*Stellifer griseus* Cervigon

*Stellifer* sp. Cervigon, 1966a:509, fig. 209, Morro de Puer to Santos, Venezuela.

*Stellifer griseus* Cervigon, 1966b:1, fig. 1, North Peninsula de Araya (holotype in Museo de Historia Natural La Salle, Venezuela, MHNLS 1.875, 120 mm SL, not examined; paratypes: USNM 200782, two specimens, 108 & 123 mm SL).

*Stellifer lanceolatus* (Holbrook)

*Homoprion lanceolatus* Holbrook, 1855:168, pl. 23, Beaufort, South Carolina.

*Stellifer microps* (Steindachner)

*Corvina microps* Steindachner, 1864:205, pl. 2, fig. 2, Guiana (type: not located in NHMV).

*Stellifer naso* (Jordan)

*Stelliferus naso* Jordan in Jordan & Eigenmann, 1889:395, Cachira, Brazil (syntypes: USNM 130630, 5 specimens, 68-75 mm SL).

*Stellifer rastrifer* (Jordan)

*Stelliferus rastrifer* Jordan in Jordan & Eigenmann, 1889:393, Brazil (holotype: MCZ 10815, 128 mm SL, Santos).

*Stellifer stellifer* (Bloch)

*Bodianus stellifer* Bloch, 1790:55, pl. 231, fig. 1, Cape of Good Hope.

*Corvina trispinosa* Cuvier, 1830:109, Cayenne, Brazil.

*Stellifer mindii* Meek & Hilderbrand, 1925:626, pl. 66, fig. 2, Mindi Reef, Panama (holotype: USNM 81730, 93.4 mm SL).

*Stellifer venezuelae* (Schultz)

*Ophioscion venezuelae* Schultz, 1945:131, fig. 7, Venezuela (holotype: USNM 121749, 140 mm SL).

## Freshwater Sciaenidae of America

Four genera, *Aplodinotus*, *Pachypops*, *Pachyurus* and *Plagioscion*, and 23 nominal species of sciaenids are recognized here from the river systems of the Atlantic drainage in America. *Aplodinotus* is the only North America freshwater inhabitant, the other three genera are endemic to continental South America. Because of lack of sufficient material for the South American genera, the diagnosis of these genera are based mainly on the literature and type-specimens. The exact limits of these genera are not clear at present.

Genus *Aplodinotus* Rafinesque

Diagnosis: Body oblong, snout blunt, back elevated and compressed; mouth small, horizontal to inferior; lower pharyngeals very large, completely united. Swimbladder simple carrot-shaped; sagitta more or less semi-circular, the sulcus not reaching the margin (Fig. 19,B); snout without upper pores (Fig. 31,B); lower jaw without mental barbels. Vertebrae 10+14=24. Freshwater North and Central America. Monotypic: *A. grunniens*.

*Aplodinotus* Rafinesque, 1819:418 (type-species: *Aplodinotus grunniens* Rafinesque, by monotypy).

*Haploidonotus* Gill, 1861d:102 (invalid emendation of *Aplodinotus*, Rafinesque, 1819, therefore taking the same type-species: *Aplodinotus grunniens* Rafinesque).

*Eutyachelithus* Jordan, 1876:242 (type-species: *Corvina richardsoni* Cuvier, by monotypy = *Aplodinotus grunniens* Rafinesque).

Remarks: The original description of *Amblodon* Rafinesque (1819: 421) included two species of "buffalo-fish" (= Catostomidae), *A. bubalus* and *A. niger*. The unique lower pharyngeal teeth of *Aplodinotus grunniens* were wrongly attributed to these fishes. Rafinesque's (1820: 24) subsequent replacement of *Aplodinotus* with *Amblodon* does not make *Amblodon* an available name in the Sciaenidae.

*Aplodinotus grunniens* Rafinesque

*Aplodinotus grunniens* Rafinesque, 1819:419, Ohio River (no type mentioned in the description).

*Sciaena oscula* LeSueur, 1822:252, pl. 13, Lake Erie (holotype: MNHN A.5696, 308 mm SL).

*Sciaena grisea* LeSueur, 1822:254, Ohio River (type: "18 to 24 inches in total length", not located in MNHN).

*Corvina richardsoni* Cuvier, 1830:100, Lake Huron (type: not located in MNHN, Paris).

*Amblodon concinnus* Agassiz, 1854:307, Tennessee River (no type mentioned in the description).

*Amblodon lineatus* Agassiz, 1854:307, Osage River (no type mentioned in the description).

*Amblodon neglectus* Girard, 1859:167, Rio Grande del Norte, Rio Bravo (holotype: USNM 639, 84.3 mm SL).

Remarks: Two catalogue numbers, MNHN A.5696 and MNHN 7536 are indicated to be syntypes of *Corvina oscula* LeSueur, MNHN A.5696, a dried stuffed specimen, 308 mm SL, collected by LeSueur from Lake Erie was examined. MNHN 7536 from New Orleans was not located.

*Pachypops* Gill

Diagnosis: Body moderately elongate, dorsal profile elevated and descending nearly straight under soft portion of dorsal fin, ventral profile nearly straight. Head oblong, snout convex and projecting; suborbital region much swollen and translucent. Mouth small, inferior, teeth in villiform bands; supramaxillary bones entirely concealed under the suborbitals. Preopercular margin slightly serrated. Snout with five upper pores and five marginal pores; lower jaw with five to six pores and three to many barbels. From Cuvier (1930), pl. 138, and Trewavas' (1964) description, the swimbladders of *P. furcroid* and *P. trifilis* have a pair of anterior appendages each dividing into a longer posterior branch and a shorter anterior one (*Lonchiurus* pattern, Fig. 14,D). Sagitta of a *Pachypops furcroid* (153 mm SL) examined, which has a similar appearance of the sagitta of *Pachyurus schomburgkii* (Fig. 18, K). Four nominal species are recognized here; *P. adspersus*, *P. camposi*, *P. furcroid* and *P. trifilis*.

*Pachypops* Gill, 1861b:87 (type-species: *Micropogon trifilis* Müller & Troschel, by original designation).

*Pachypops adspersus* (Steindachner)

*Pachyurus adspersus* Steindachner, 1879:123, Tio San Antonio, Brazil  
(type: not located in MNHV).

*Pachypops camposi* Fowler

*Pachypops camposi* Fowler, 1954, 9:252, Rupununi River, British Guiana  
(holotype: ANSP 39773, 117 mm SL, labeled as *P. steindachneri*).



*Pachypops fourcroyi* (Lacépède)

*Perca fourcroyi* Lacépède, 1803, vol. 4:398, no locality (holotype: MNHN 7539, 135 mm SL).

*Pachypops trifilis*

*Micropogon trifilis* Müller & Troschel, 1848:622, British Guiana (type: not located).

*Pachyurus* Agassiz

Diagnosis: Body moderately elongate, dorsal profile slightly elevated, ventral nearly straight or slightly arched. Head conical; snout bluntly, swollen and translucent. Eye moderate to large. Mouth horizontal, inferior or terminal; teeth in villiform bands. Snout with five marginal pores and usually without upper pores, some with three to five upper pores; lower jaw with five pores and no barbel. Swimbladder simple, carrot-shaped or with a pair of tube-like diverticula extending from anterior corners of swimbladder, tapering back to the end of main chamber (Fig. 6). Sagitta moderate to large. Second spine of anal fin moderate to strong. This genus may be divisible into two genera based on the above characters. Nine nominal species are here recognized: *P. biloba*, *P. bonariensis*, *P. francisci*, *P. grunniens*, *P. lundii*, *P. natteri*, *P. paranensis*, *P. schomburgkii* and *P. squamipinnis*.

*Pachyurus* Agassiz, 1829:125 & 127 (type-species: *Pachyurus squamipinnis* Agassiz, by monotypy).

*Lepipterus* Cuvier, 1830:152 (type-species: *Lepipterus francisci* Cuvier, by monotypy).

*Pachyurus biloba* Cuvier

*Corvina biloba* Cuvier, 1830:112, no locality (holotype: MNHN 7683, 75.6 mm SL).

*Pachyurus bonariensis* Steindachner

*Pachyurus (Lepipterus) bonariensis* Steindachner 1870:80:125, La Plata, Argentine (NHMV 15181, may be one of the syntypes, 188 mm SL, La Plata, Argentine; "cotype": CAS, Indiana University No. 11353, 152 mm SL, Buenos Aires, this specimen probably not a type-specimen).

*Pachyurus francisci* (Cuvier)

*Lepipterus francisci* Cuvier, 1830:152, pl. 113, Riviere de Saint-Francois, Brazil (type: not located).

*Pachyurus corvina* Reinhardt in Lütken, 1875:284, Rio das Velhas, Brazil (syntypes: ZMUC 345, three specimens; 217, 243 & 233 mm SL).

*Pachyurus grunniens* (Schomburgki)

*Corvina grunniens* Schomburgki, 1843:136, pl. 2, Comacca Island, Essequibo, British Guiana.

*Pachyurus lundii* Reinhardt

*Pachyurus lundii* Reinhardt in Lütken, 1875:248, pl. 20, Rio das Velhas, Brazil (syntypes: ZMUC, 16-3, 1871, two specimens; 274 & 366 mm SL).

*Pachyurus natteri* (Steindachner)

*Pachyurus natteri* Steindachner, 1863:10, pl. 3 (type: not located in MNHV).

*Pachyurus paranensis* Dameri

*Pachyurus paranensis* Dameri, 1956:6, fot. 1 (original description not seen, after Travassos & Rego-Barros, 1971:60).

*Pachyurus schomburgkii* Günther

*Pachyurus schomburgkii* Günther, 1860:282, rivers of Brazil (holotype: BMNH 49.11.8.22, 197 mm SL).

*Pachyurus squamipinnis* Agassiz

*Pachyurus squamipinnis* Agassiz 1839:128, pl. 71, Brazil.

*Plagioscion* Gill

Diagnosis: A freshwater genus of Sciaenidae from South American rivers, some species occasionally in estuaries, rarely marine. Body elongate, dorsal profile evenly arched, ventral nearly straight. Head slightly conical, snout protuberant. Mouth moderate, terminal, slightly oblique, no enlarged canine-like teeth at the tip of upper jaw. Scales along lateral line of some species, concealed by many smaller scales, appeared as one enlarged scale (Fig. 40). Gill rakers rather long and slender. Swimbladder with a pair of horn-like diverticula, originating from anterior 1/4 of main chamber and hooked at the distal ends (Fig. 12,D). Sagitta sub-oval in shape, the ostium

of the "tadpole-shaped" sulcus reaching to anterior margin and the cauda deeply curved in a J-shape (Fig. 18,L). Nine nominal species in this genus: *P. auratus*, *P. heterolepis*, *P. macdonaghi*, *P. microps*, *P. monacantha*, *P. pauciradiatus*, *P. squamosissimus*, *P. surinamensis* and *P. ternetzi*.

*Plagioscion* Gill, 1861b:82 (generic description, no species or type indicated, compared with *Corvina* of Cuvier; (type-species: *Sciaena squamosissima* Heckel, by subsequent designation of Jordan & Eigenmann, 1889:380).

*Diplolepis* Steindachner, 1863:2 (type-species: *Diplolepis squamosissimus* Steindachner = *Sciaena squamosissima* Heckel, by monotypy, spelled *Diplolepis* on p. 3; preoccupied by *Diplolepis* Geoffroy, 1762, cynipid insect and Fabricius, 1805, calcid insect, quote from Neave, 1940).

*Plagioscion auratus* (Castelnau)

*Johnius auratus* Castelnau, 1855:12, pl. 4, fig. 2; Ucayala, Brazil (holotype: MNHN 7622, 203 mm SL).

*plagioscion heterolepis* (Bleeker)

*Johnius heterolepis* Bleeker, 1873:456, Surinam (syntypes: RMNH 6042, two specimens, 123 & 129 mm SL, these type-specimens are similar to the *Stellifer* group).

*Plagioscion macdonaghi* Dameri

*Plagioscion macdonaghi* Dameri, 1954:179, fig. 1, Rio de La Plata  
(holotype in Coleccion Nacional del Instituto Nacional de  
Investigación de las Ciencias Naturales, Brazil, no. 4197, 147  
mm TL, not examined).

*Plagioscion microps* Steindachner

*Plagioscion microps* Steindachner, 1917:657, pl. 1, fig. 1, Amazon  
(one specimen of NHMV 15180, 168 mm SL and 206 mm TL donated by  
Steindachner, collected from Surinam, may be one of the syntypes:  
208 & 214 mm TL of original description).

*Plagioscion monacantha* (Cope)

*Corvina monacantha* Cope, 1867:402, near Paramaribo, Surinam (holotype:  
ANSP 11519, 167 mm SL).

*Sciaena magdalena* Steindachner, 1878:22, pl. 1, fig. 1, Magdalenen  
River (types: 270 to 540 mm TL, not located in NHMV).

*Plagioscion pauciradiatus* Steindachner

*Plagioscion pauciradiatus* Steindachner, 1917:660, figure on p. 661,  
Paramaribo (type: not located in NHMV).

*Plagioscion squamosissimus* (Heckel)

*Sciaena squamosissima* Heckel, 1840:438, Amazon.

*Sciaena rubella* Schomburgk, 1843:133, river of British Guiana.

*Johnius courvina* Castelnau, 1855:11, pl. 5, fig. 1, Rio Crixas, Rio Arognay, Brazil (holotype: MNHN 7503, 366 mm TL).

*Johnius amazonicus* Castelnau, 1855:12, pl. 4, fig. 1, Amazon (syntypes: MNHN 7504, two specimens, 96.3 & 145 mm SL).

*Plagioscion surinamensis* (Bleeker)

*Pseudosciaena surinamensis* Bleeker, 1873:458, Surinam (holotype: RMNH 5995, 88.1 mm SL).

*Plagioscion ternetzi* Boulenger

*Plagioscion ternetzi* Boulenger, 1895:523, Paraguay (syntypes: BMNH 1895.5.17.1-2, two specimens, 88.4 & 263 mm SL).

## II. Phylogenetic Relationships of the Supra-generic Groups and Genera.

The morphological patterns of swimbladders, otoliths and external features of western Atlantic sciaenid fishes are considered together here (Table 1) to assess phylogenetic relationships (Fig. 4). In the following discussion, relationships will be discussed among the genera, then among supra-generic groups.

### 1. Relationships of the genera within supra-generic groups.

#### *Sciaena* group:

The members of this group all have a simple swimbladder (*Sciaena* pattern, Fig. 5) and a thick sagitta (*Sciaena* pattern, Fig. 18). The shape and position of the sulci vary within the genera. *Equetus* and *Paraquet* are very similar in having an elevated dorsal profile and dark stripes on the sides. They are both coral reef dwellers. *Sciaena* lives in deeper water (Chao & Miller, 1975) than other sciaenids. Members of *Sciaena* group have enclosed inferior mouths except for *Sciaena* with a terminal and slightly inferior mouth. Habitat diversification of this group is greatest among all western Atlantic supra-generic groups of sciaenids.

#### *Umbrina* group

This group is most similar to the *Sciaena* group, except these fishes have a single barbel on the lower jaw. The swimbladder is simple, carrot-shaped (*Sciaena* pattern, Fig. 5). The sagitta of *Ctenosciaena gracilicirrhus* is most similar to *Umbrina millae* and *Sciaena trewavasae* (Fig. 18). This suggests a relationship between the

*Sciaena* and *Umbrina* groups. Although *Ctenosciaena* has a mental barbel, it lacks an apical pore (Fig. 35,A). It also has a terminal and slightly inferior mouth, and inhabits waters slightly offshore and deeper than *Umbrina* species. The deeper water habitat of *Ctenosciaena* and corresponding morphological adaptations are similar to those of *Sciaena*. This may suggest a convergence between these respective genera.

*Lonchiurus* group:

*Paralonchurus* and *Lonchiurus* show a sequence of reduction in swimbladder size (Fig. 14). This is significant in assessing the relationships of the *Menticirrhus* and *Lonchiurus* groups. *Lonchiurus lanceolatus* has the most reduced swimbladder (Fig. 14,A) and lacks drumming muscles in both sexes. It is a coastal and estuarine dweller. Its habitat is similar to that of *Menticirrhus* which has only a vestige of the swimbladder remaining in adults. The reduced swimbladder of *L. lanceolatus* also shows a basic two-horned structure, which resembles that of the *Cynoscion* group (Fig. 12). *Paralonchurus* has a well-developed swimbladder, drumming muscles are present in males and it inhabits deeper inshore waters than *Lonchiurus*. In comparing the sagitta (Fig. 24) and external morphology of *Lonchiurus* and *Paralonchurus*. The similarities among *L. lanceolatus*, *P. brasiliensis* and *P. elegans* indicate that they might all be validly treated as one genus. The South American freshwater sciaenid genus *Pachypops* may also belong to this group, based on the swimbladder of *P. furcroi* (Fig. 14,D). Few species examined from this genus all have inferior mouths and three to many pairs of barbels.



*Cynoscion* group:

Although genera of this group have similar swimbladder shapes, (Fig. 12) the sagittae (Figs. 23 & 25) and external morphology are rather variable. *Macrodon ancylodon* has a thin elongate sagitta (Fig. 25,B) similar to *Cynoscion* (Fig. 23), but the general outline and the sulcus differ. The external morphology is also similar, except that *Macrodon* has a pair of large lanceolate canines at the tip of the upper jaw (Fig. 37,A). *Isopisthus* differs from all other sciaenids by having a space between the spinous and soft dorsal fins, but its general body shape resembles *Cynoscion*. The South American freshwater genus *Plagioscion* has a terminal horizontal mouth, and lacks enlarged canines. Its body shape is deeper and is not as fusiform as other genera of this group. This may be an adaptation to the freshwater habitat, whereas, the fast swimming and fusiform *Cynoscion* may be adapted to an open water habitat. The sagittae of the genera in the *Cynoscion* group differ from each other. In *Plagioscion surinamensis* (Fig. 18,L) and *Isopisthus parvipinnis* (Fig. 25,A), the sagittae have a similar thickened posterior half. Sciaenidae usually have a thick sagitta, therefore, this similarity is not indicative of generic relationships.

*Stellifer* group:

Genera of this group all have a two chambered swimbladder (Fig. 15) and an enlarged lapillus (Fig. 27). The genus *Stellifer* has a pair of diverticula postero-laterally on the anterior chamber of the swimbladder (Fig. 16). The different morphology of these diverticula indicate the range of variation. *Ophioscion* has a sagitta and lapillus

shaped similarly to *Stellifer* (Fig. 27,A & B) and the body shape, cavernous head (Fig. 29,A) and swimbladder position are similar. Although *Ophioscion* lacks diverticula on the anterior chamber of the swimbladder, it could be considered as a member of *Stellifer* (Fig. 4). *Bairdiella* and *Odontoscion* (Fig. 37,C) have a terminal mouth, and lack apparent cavernous canals on the head (Fig. 39,B). *Odontoscion* inhabits clear water reefs and has large eyes and large canine-like teeth (Fig. 37,C). Other genera of this group are mainly estuarine inhabitants.

Genus, *Leiostomus*, is similar to *Sciaena* group, but it has a much thinner sagitta (Fig. 18,D). In addition, the ostium of its sulcus is bent toward the dorsal margin (to the right of Fig. 18). The phylogenetic position of this genus is tentatively put with the *Sciaena* and *Umbrina* groups. The North American freshwater *Aplodinotus* has a semi-circular sagitta (*Pogonias* pattern, Fig. 19). Although the swimbladder of *Aplodinotus* is a simple carrot-shaped (*Sciaena* pattern, Fig. 5), it may still be more closely related to *Pogonias* than any other western Atlantic sciaenid genus (Fig. 4). Osteological studies of Topp and Cole (1968) on *Sciaenops* and Mohsin (1973) on four *Cynoscion* species of the Gulf of Mexico are also useful in assessing relationship within the genera. Other monotypic supra-generic groups of western Atlantic sciaenids may contain more genera from other geographic regions, especially the eastern Pacific.

## 2. Relationships of the supra-generic groups:

The *Sciaena* and *Umbrina* groups are similar in swimbladder (*Sciaena* pattern, Fig. 5) and otolith (*Sciaena* pattern, Fig. 18)

morphology. Their external morphology is similar except that the *Umbrina* group has a mental barbel (Fig. 35,A & C). Members of both groups are mainly bottom feeders with inferior mouths. The genus *Sciaena* of the *Sciaena* group and the genus *Ctenosciaena* of the *Umbrina* group have entered deeper waters and both have a relatively larger and more terminal mouth to feed in mid-water. The cluster of *Sciaena* and *Umbrina* groups are in turn most closely related to the *Larimus* group (Fig. 4). The species of *Larimus* have a basic simple swimbladder (Fig. 5,E & F). A pair of anterior projections found on the swimbladder of *L. breviceps* appear to be a modification of the *Sciaena* swimbladder pattern. The sagitta of *Larimus* species (Fig. 20,A & B) has a unique outline and the ostium of the sulcus is much larger and does not reach to the anterior margin of the sagitta. The lack of upper pores on the snout (Fig. 31,B) and four minute mental pores (Fig. 34,B) in *Larimus* are adaptive characters correlated with its large oblique mouth and upper water column feeding habits (Fig. 30 and Table 1).

Among the supra-generic groups, both *Pogonias* and *Sciaenops* have a complicated swimbladder as adults (Figs. 10 & 11), but the structural patterns are different and their sagittae also differ (Figs. 19 & 20). Although *Pogonias* and *Sciaenops* have a similar inferior mouth and inhabit inshore coastal and estuarine waters, the body shape of *Sciaenops* is more elongate and less compressed than *Pogonias*. This may be an adaptation of *Sciaenops* to the shallow water surf zone habitat.

The *Menticirrhus* group is closely related to the *Lonchiurus* group on the basis of their reduced swimbladders (Figs. 7 & 14). In

comparison to other diagnostic characters, the *Menticirrhus* and *Lonchiurus* groups, whether this trend of swimbladder reduction may be interpreted as indicating phylogenetic relationships or ecological convergence is unknown. The sagittae of the *Menticirrhus* and *Lonchiurus* groups (Figs. 22,B & 24) are both elongate and thin but the shapes of the sulci are different. The body shape is elongate and rounded in both groups. The flat ventral side of the body is an adaptation for inhabiting the bottom or specifically the surf zone. Both genera have inferior mouths and barbels. Although the *Menticirrhus* group has a pores single barbel (Fig. 35,B) similar to that of *Umbrina* (Fig. 35,C), this probably should be viewed as ecologically convergent for bottom feeding rather than as phylogenetically important.

Other generic groups of western Atlantic sciaenids may not be as closely related as the groups already discussed. Their relationships appear to be above the tribal level (between line C and line D of Figure 4) in the present study. The next higher level of clustering (between line B and line C of Figure 4) indicates that the *Pogonias* and *Sciaenops* are most closely related to the *Larimus*, *sciaena* and *Umbrina* groups. The generic groups included in this cluster (Z on Figure 4) show a basic carrot-shaped swimbladder with diverticula present or absent. Their sagittae are basically sub-oval shaped and most members are bottom feeders. This cluster (Z) of supra-generic groups is probably the main scheme of the phylogenetic development of western Atlantic Sciaenidae. The trend of swimbladder development in western Atlantic sciaenids is clearly demonstrated from the simple *Sciaena* pattern with extrinsic drumming muscles in the male to the complicated

*Pogonias* pattern with intrinsic drumming muscles in both sexes.

Furthermore, the ontogenetic development of the swimbladder in *Pogonias cromis* (Fig. 10) and *Sciaenops ocellata* (Fig. 11) may also reflect the phylogenetic development of the swimbladder, from simple to complicated with diverticula developing on the swimbladder from the anterior to the posterior end.

Further clustering of the taxa (between line A and line B of Figure 4) is more difficult due to the limitations of a regional study. There are gaps, especially in assessing the relationships of the *Micropogonias* and *Nebris* groups. *Nebris microps* has a pair of well-developed anterior diverticula on its swimbladder (Fig. 8) and a very thick oval-shaped sagitta (Fig. 21). *Micropogonias* species have a pair of posteriorly originating tube-like diverticula on their swimbladders (Fig. 9) and a rather thick and shield-like irregular shaped sagitta (Fig. 26). At present, I suggest that the *Nebris* is more closely related to the Z cluster on Figure 4 than does the *Micropogonias*. This clustering is based mainly on the position of the diverticula. The swimbladder diverticula of *Micropogonias* are developed more posteriorly than that in the *Nebris* group. Both of them have a sagitta different from the members of Z cluster (Fig. 4). *Nebris* group has also evolved a different way of feeding, by having an elongate round body and a very large and oblique mouth (Fig. 37,B). This form of mouth structure and body shape enables them to feed from the bottom upward. The *Micropogonias* has an inferior mouth and four to six pairs of mental barbels (Fig. 37,D). typical structures of bottom feeding sciaenids.

The grouping of the *Cynoscion* group with the *Menticirrhus* and *Lonchiurus* groups (Y on Fig. 4) is based on the basic shape of the main chamber of the swimbladder and their elongate thin sagittae. Although the members of the *Lonchiurus* group have a pair of posteriorly directed diverticula on their swimbladders (Fig. 14), the main chambers have two anterior horns resembling those of the *Cynoscion* group (Fig. 12). Especially in the reduced form of *Lonchiurus lanceolatus* (Fig. 14,A), the anterior horns remain unchanged.

The X and Y clusters on Figure 4 are more closely related to each other phyletically than to the *Stellifer* group, because they all have a single chambered swimbladder and only the sagitta enlarged. The swimbladder of the *Stellifer* group has an additional chamber (Fig. 15) in front of the main carrot-shaped chamber. This yoke-shaped anterior chamber is located in front of the septum transversum and its anterior ends reach the skull. The sagitta of the *Stellifer* group is reduced in size and the lapillus is enlarged (Fig. 27). The sequences of sagitta reduction and lapillus enlargement are evident among the modern genera of the *Stellifer* group. The unique swimbladder and otolith characters of the *Stellifer* group suggest that they are a distinct group from all other western Atlantic sciaenids. The two chambered swimbladder has not been reported in sciaenid fishes of other geographic regions other than the New World. Therefore, the *Stellifer* group could be treated as a subfamily of the Sciaenidae.

In conclusion, western Atlantic Sciaenidae can be readily divided into two groups. One group is characterized by a swimbladder with two chambers and with two pairs of enlarged otoliths (lapillus and

sagitta). The other group is characterized by a single chambered swimbladder and only one pair of enlarged otoliths (sagitta). Further divisions are based mainly on swimbladder structure, secondly on the morphology of the sagittae and thirdly on external morphology.

Swimbladder structure may be graded from simple to complex. Such a gradation is also reflected in the ontogenetic changes of several species such as *Pogonias cromis* (Fig. 10) and *Sciaenops ocellata* (Fig. 11). The primitive condition of the swimbladder (simple *Sciaena* pattern) exists in juveniles and adults of many species. Further development of the swimbladder is determined by the development of the diverticula from the anterior to the posterior end of the main chamber. Whether the reduction in size or loss of the swimbladder in adults of *Menticirrhus* and *Lonchiurus* groups (Figs. 7 & 14) is attributed to phylogenetic relationships or ecological convergence is undetermined at present.

The general morphology of the sagitta and its sulcus were used to determine relationships of different taxa especially at the generic level. In the primitive condition of this character, the sagitta is thick and the sulcus opens to the anterior margin of the sagitta, i.e. the ostium of the sulcus reaches the anterior margin (*Sciaena* pattern, Fig. 18). This is the most generalized pattern in many groups of western Atlantic sciaenids. The external morphology of the western Atlantic Sciaenidae, especially the mouth positions and body shapes are more diverse than any other perciform family in the region. These characters are more or less adaptive and plastic, but may be used to augment other characters in assessing the relationships of the western

Atlantic sciaenids. Biochemical characters of parvalbumins have been studied by Sullivan, et. al. (1975) and Rao, et. al. (1976) to assess the systematic relationships of some sciaenid species from the Atlantic American coast. Their findings in the future would contribute much more to the current knowledge of phylogenetic relationships of western Atlantic Sciaenidae. There are four genera of Sciaenidae; *Aplodinotus*, *Pachyurus*, *Pachypops* and *Plagioscion* which live in freshwater for their entire life history. The freshwater sciaenids are endemic to the Atlantic drainages of the New World. Knowledge of their taxonomy and biology is rather sparse. Further studies on these freshwater sciaenids and on eastern Pacific sciaenids should fill many gaps in the present study.



FIELD KEY TO THE GENERA  
AND SPECIES OF WESTERN ATLANTIC SCIAENIDAE

- 1a Lower jaw with one or more barbels (Fig. 32, E to H)
  - 2a Only one barbel at the tip of lower jaw
    - 3a Anal fin with one spine; body elongate fusiform; swimbladder absent or degenerate in adult. *Menticirrhus*
    - 3b Anal fin with two spines; body compressed, not fusiform; swimbladder present in adult
    - 4a Mental barbel short, rigid and perforate, with an apical pore (Fig. 35,C); two pairs of large pores at the base of barbel; mouth inferior; body with longitudinal stripes and vertical bars. *Umbrina*
    - 4b Mental barbel short, flexible and tapered at end, without an apical pore (Fig. 35,A); lateral pores minute; mouth slightly inferior, subterminal; body uniform in color, a black spot at pectoral fin origin. [D.X, I+21-24; A.II, 7-8; gill rakers short, (7-9)+(13-17)=21-25]. *Ctenosciaena gracilicirrus*  
(South Caribbean to Brazil)
  - 2b Tuft of barbels at tip and/or small barbels along rami of lower jaw
    - 5a Second anal fin spine strong and long, more than two-thirds length of first anal ray; eye moderate, five times or less in head

- 6a Barbels mostly in tuft at tip of chin (Fig. 32, H); eye diameter large (about three to four times in head); freshwater . . . . . *Pachypops*  
(Freshwater South America)
- 6b Row of small barbels along rami of lower jaw, not in tuft at tip of chin (Fig. 32, G); eye diameter medium (about five times in head); marine and estuarine
- 7a Preopercular margin strongly serrate; body covered with relatively small scales, 64 to 72 in the row above lateral line; swimbladder with a pair of tube-like diverticula (Fig. 9); body silvery with pinkish cast, with many oblique stripes along the scale rows . . . . .  
. . . . . *Micropogonias*
- 7b Preopercular margin smooth; body covered with relatively large scales, 41 to 45 in the row above lateral line; swimbladder very complex with many interconnected diverticula (Fig. 10); body gray to dark, juveniles with 4 to 5 broad vertical bars. [D. X+I, 19-21; A. II, 5-6; gill rakers short, (4-6)+(12-16)=16-21]. . . . . *Pogonias cromis*  
(Gulf of Maine to Brazil)
- 5b Second anal fin spine weak and short less than two-thirds length of first anal ray; eye small about nine times in head

- 8a Lower jaw with a tuft of three pairs minute barbels at the tip and more than 10 pairs barbels along rami of lower jaw (Fig. 32, H); pectoral fin large or small; swimbladder well-developed (Fig. 14, B & C). . . . . *Paralonchurus*
- 8b Lower jaw with two barbels only (Fig. 32, F), barbels longer than eye diameter; pectoral fins very large, jet black; swimbladder atrophied in adult (Fig. 14, A). [D. X-XI+I, 37-39, A. II, 7-9; gill rakers short, (4-6)+(11-13)=15-18]. . . . .  
. . . . . *Lonchiurus lanceolatus*  
(Venezuela to Brazil)
- 1b Lower jaw without barbel (Fig. 32, A to D)
- 9a Body fusiform; mouth distinctively oblique; a pair of enlarged canine-like teeth usually at tip of upper jaw; preopercular margin entire never serrate; swimbladder with a pair of horns at anterior end (Fig. 12)
- 10a Spinous and soft portions of dorsal fin well separated; anal fin base about equal to the length of soft dorsal fin base; anal fin rays 16-20. [D. VII+I, 18-20; gill rakers moderate, (2-3)+(7-9)=9-12]. *Isopisthus parvipinnis*  
(Guiana to southern Brazil)
- 10b Spinous and soft portion of dorsal fin, with a deep notch in between, but not separated; soft dorsal fin base much longer than anal fin base; anal fin rays seven to 12

- 11a Enlarged canine teeth lance-shaped (Fig. 37, A);  
lower jaw with canine-like teeth exposed externally  
when mouth closed. [D. X+I, 27-29; A. II, 8-9;  
gill rakers slender, (2-3)+(7-9)=9-12]. . . . .  
. . . . . *Macrodon ancylodon*  
(Guiana to southern Brazil)
- 11b Enlarged canine teeth not lance-shaped but tapering  
gradually from base to tip; lower jaw teeth never  
exposed externally when mouth closed. *Cynoscion*
- 9b Body fusiform or deeply compressed, mouth inferior, terminal,  
horizontal or oblique; no enlarged canine-like teeth at tip  
of upper jaw; preopercular margin serrate or not; swimbladder  
with or without anterior horns
- 12a Preopercular margin serrate, sometimes strong with one  
or more spines at the angle
- 13a Scales along lateral line considerably enlarged but  
almost entirely concealed by small scales (Fig. 40);  
swimbladder with one chamber and a pair of anterior  
horns present (Fig. 12, D); lapillus (small  
earstone) small, less than one-tenth the size of  
sagitta (large earstone); freshwater and  
estuarine. . . . . *Plagioscion*  
(freshwater and estuarine South America)
- 13b Scales along lateral line about same size as  
adjacent rows; swimbladder with two chambers  
anterior one yoke-shaped and separated by a  
constriction from main posterior chamber (Fig. 15);

lapillus (small earstone) large, more than half the size of sagitta (larger earstone) (Fig. 27);  
marine and estuarine

14a Anterior horn of swimbladder terminating subcutaneously, frequently visible on superficial inspection in adult and juvenile under the upper end of gill slit. Mouth inferior to oblique; snout usually projecting; dorsal view of head blunt, cavernous (Fig. 39, A); interorbital width usually more than 1.2 times of eye diameter

15a Anterior chamber of swimbladder with a pair of variably developed posterior diverticula (Fig. 16); bony interorbital width 3.5 or less in head. . *Stellifer*

15b Anterior chamber of swimbladder without posterior diverticula; bony interorbital width 3.5 or more in head. . *Ophioscion*

14b Anterior horn of swimbladder only visible in juvenile externally under the upper end of gill slit. Mouth horizontal to oblique; snout not projecting; dorsal view of head tapered; not cavernous (Fig. 39, B)  
interorbital width usually less than 1.2 times of eye diameter. . . . . *Bairdiella*

12b Preopercular margin without strong serrations, sometime ciliated but never with spines at the angle

16a Mouth large, oblique or nearly vertical; snout not projecting; lower jaw prominent

17a Mouth slightly oblique, terminal; lower jaw with a row of enlarged canine-like teeth, not projecting beyond upper jaw (Fig. 37, C); swimbladder with two chambers, anterior one yoke-shaped, posterior one in carrot-shape; lapillus (small earstone) large more than two-thirds the size of sagitta (larger earstone); a distinct black blotch at pectoral fin base. /D. XI-XII+I, 23-26; A. II, 8-9; gill rakers long and stiff, (5-9)+(14-17)=19-25/. . . . . *Odontoscion dentex* (Florida to Brazil)

17b Mouth very oblique or vertical; lower jaw without canine-like teeth, projecting in front of upper jaw (Fig. 37, B); swimbladder in one chamber, lapillus (small earstone) small, less than one-tenth the size of sagitta (large earstone), without a distinct black blotch at pectoral fin base

18a Eye small, about eight to 10 times in head; body elongate; scales small, cycloid, about 90 in the row above lateral line; caudal fin pointed in adult. /D. VIII+I, 31-32; A. II, 9-10; gill rakers long and slender, (5-9)+(14-

15)=20-24]. . . . . *Nebriis microps*

(Costa Rica to Brazil)

18b Eye large, about three to four times in head; body short; scales large, ctenoid, about 50 in the row above lateral line; caudal fin biconcave in adult. *Larimus*

16b Mouth small, inferior; snout projecting in front of upper jaw; lower jaw included

19a Body short, deep; dorsal profile elevated anteriorly

20a Body silvery with faint oblique stripes along the oblique scale rows dorsally; a dark spot above upper angle of gill slit; gill rakers more than 30 on first gill arch. [D. IX+I, 29-35; A. II, 12-13; gill rakers (8-12)+(20-23)=30-36].  
 . . . . . *Leiostomus xanthurus*

(U.S. Atlantic and Gulf of Mexico coasts)

20b Body pale to dark brown; with dark longitudinal stripes and/or oblique bars on sides, without a dark spot above gill slit; gill rakers less than 20 on first gill arch

21a Height of spinous dorsal longer than head, usually with long filament; body brownish pale with three oblique bars on sides; third bar running

- from nape to caudal fin; dorsal fin soft rays more than 45. *Equetus*
- 21b Height of spinous dorsal shorter than head; body pale brown to dark brown with longitudinal stripes on sides; dorsal fin soft rays less than 40. . . . . *Pareques*
- 19b Body elongate, dorsal profile not elevated
- 22a Snout with peculiar conical appearance, from the preorbital region being swollen and enlarged; maxillary entirely concealed under snout. . . *Pachyurus*  
(freshwater South America)
- 22b Snout rounded, not swollen, maxillary exposed under snout
- 23a Body elongate; one or more large dark spots (larger than eye) on the base of caudal peduncle and/or side of body. [D. X+I, 23-25; A. II, 8-9; gill rakers (4-5)+(7-9)=12-14]. . . . .  
. . . . . *Sciaenops ocellata*  
(U.S. Atlantic and Gulf of Mexico coasts)
- 23b Body robust; color uniform, no black spot on caudal base, no dark spots on sides of body. [D. IX-X+I, 27-33; A. II, 7; gill rakers 6+(10-12)=



16-18]. . . *Aplodinotus grunniens*  
(freshwater, Canada to Guatamala)

Key to the species of *Bairdiella*

- 1a Preopercular margin with distinct strong spines at the angle
- 2a Second anal fin spine very strong, 21-26% of standard length; preopercular margin with strong spine-like serration. D. X+I, 21-26 (usually 23-25); A. II, 7-9 (usually 8); gill rakers slender,  $(6-10)+(15-18)=21-27$  (usually 24-25). *B. ronchus* (Caribbean islands to Brazil)
- 2b Second anal fin spine not as strong, 13-20% of standard length; preopercular margin with distinct strong spines. D. X, I, 19-23; A. II, 8-10 (usually 9); gill rakers slender,  $(7-8)+(14-16)=22-24$ . . . . . *B. chrysoura* (U.S. Atlantic and Gulf of Mexico coasts)
- 1b Preopercular margin with only weak serrations, no spines at the angle
- 3a Preopercular margin with strong lateral ridge and weak serrations; second anal fin spine about  $2/3$  length of first ray; D. X+I, 25-29; A. II, 7-8; gill rakers short,  $(5-6)+(13-16)=18-22$ . . . . . *B. batabana* (Florida to Caribbean islands)
- 3b Preopercular margin membranous without distinct serrations; second anal fin spine less than  $2/3$  length of first ray; D. X-IX, I+22-24; A. II, 9 (rarely 8); gill rakers long,  $(7-8)+(16-18)=23-26$ . . . . . *B. sanctaeluciae* (Florida to Guiana)

Key to the Species of *Cynoscion*

## 1a Scales cycloid on body

2a Soft portion of dorsal fin base covered with small scales;  
with less than 25 rays

3a Inner row teeth of lower jaw much larger than external  
ones, some central ones canine-like; small scales cover  
2/3 of soft dorsal fin; D. X+I, 22-25 (usually 23-24);  
A. II, 10-12; gill rakers (2-3)+(6-9)=8-11; 150-160  
scales along the row above lateral line; anterior horns  
of swimbladder long and straight (Fig. 13, A). . . . .  
. . . . . *C. microlepidotus*  
(Venezuela to Brazil)

3b Inner row teeth of lower jaw slightly enlarged, no  
canine-like teeth; small scales cover less than 1/3 of  
soft dorsal fin; D. X+I, 20-24 (usually 21-23); A. II,  
8-10; gill rakers (2-3)+(5-8)=7-11; 115-125 scales  
along the row above lateral line; anterior horns of  
swimbladder curved (Fig. 13, D & E). *C. leiarchus*  
(Venezuela to Brazil)

2b Soft portion of dorsal fin naked, with 23-31 rays; A. II,  
8-9; gill rakers (1-3)+(6-8)=7-11; 120-130 scales along the  
row above lateral line; anterior horn of swimbladder slightly  
curved; sagitta with a notch on ventral margin (Fig. 23, C). .  
. . . . . *C. virescens*  
(Panama to Brazil)

## 1b Scales ctenoid on body

- 4a More than 20 gill rakers. [D. X+I, 18-21; A. II, 8-9; gill rakers long,  $(7-9)+(14-17)=21-26$ ]. . . . *C. striatus*  
(Brazil to Argentina)
- 4b Less than 20 gill rakers
- 5a Dorsal fin usually with fewer than 21 rays; anal with eight to nine soft rays; gill rakers long,  $(2-6)+(8-10)=10-16$ . . . . . *C. acoupa*  
(Panama to Brazil)
- 5b Dorsal fin usually with more than 21 rays; anal fin with eight to 11 soft rays; gill rakers long or short.
- 6a Pectoral fin short, about two times or more in head
- 7a A pair of canine-like teeth at the tip of upper jaw; D. IX-X+I, 25-29; A. II, 10-12; gill rakers  $(3-4)+(9-10)=12-14$ ; about 60 (58-62) scales in the row above lateral line . . . . .  
. . . . . *C. arenarius*  
(U.S. Gulf of Mexico coast)
- 7b Canine-like teeth reduced; D. X+I, 21-24; A. II, 10-12; gill rakers  $(3-5)+(8-10)=11-15$ ; about 70 (67-72) scales in the row above lateral line . . . . *C. steindachneri*  
(Guiana to Brazil)
- 6b Pectoral fin long, much less than two times in head
- 8a Soft portion of dorsal fin not covered by small scales

- 9a Gill rakers long,  $(3-4)+(6-9)=9-13$ ;  
 D. X+I, 24-29; A. II, 8-10; body color  
 brownish to pale without dark spots on  
 the side. . . . . *C. similis*  
 (Venezuela to Surinam)
- 9b Gill rakers short,  $(2-3)+(7-9)=9-12$ ;  
 D. IX-X, 25-28, A. II, 10-11; body with  
 large black spots on the back and sides. .  
 . . . . . *C. nebulosus*  
 (U.S. Atlantic and Gulf of Mexico coasts)
- 8b Soft portion of dorsal fin usually densely  
 covered with small scales
- 10a Anal fin with 11-13 soft rays; body  
 usually with small dark spots forming  
 oblique lines on the side. [D. X+I,  
 26-29; gill rakers  $(4-5)+(10-12)=14-17$ ]. .  
 . . . . . *C. regalis*  
 (Gulf of Maine to Florida)
- 10b Anal fin with 8-10 soft rays; side of  
 body without distinct spots
- 11a Soft portion of dorsal fin covered  
 with small scales to about three-  
 fourths of soft dorsal fin height;  
 D. X+I, 23-27; gill rakers  $(2-3)+$   
 $(7-10)=9-13$ ; total vertebrae 25. . .  
 . . . . . *C. jamaicensis*  
 (Panama to Argentina)

- 11b Soft portion of dorsal fin covered with small scales less than two-thirds of soft dorsal fin height; D. X+I, 26-30; gill rakers (3-4)+(8-10)=11-14; total vertebrae 27. . . . .*C. nothus*  
(Chesapeake Bay to Texas)

Key to the Species of *Equetus*

- 1a Body with two narrow longitudinal stripes above and below the third bar; dorsal, anal and caudal fin dark brown with white spots; pectoral dark brown. [D. XI-XII+I, 45-47; A. II, 6-8; gill rakers short, 5+(10-13)=15-18]. . . . .*E. punctatus*  
(Florida, Antilles, Panama to Brazil)
- 1b Body without narrow longitudinal stripes; broad oblique bars with distinct white margins; pectoral, dorsal, and anal fins pale. [D. XII-XIII+I, 47-55; A. II, 6; gill rakers short, (5-6)+(10-13)=14-18]. . . . .*E. lanceolatus*  
(Florida, Antilles and Venezuela)

Key to the Species of *Larimus*

- 1a Body dark gray above and silvery below without vertical bars on the sides; [D. X+I, 26-28; gill rakers slender, longer than eye diameter; (9-11)+(19-22)=28-33]. . . . .*L. breviceps*  
(Caribbean Islands to Brazil)

- 1b Body dark with 7 to 9 vertical bars on the sides; [D. X+I, 24-27; gill rakers slender, about equal to eye diameter, (11-13)+(22-25)=34-36]. . . . . *L. fasciatus*  
(U.S. Atlantic and Gulf of Mexico coasts)

Key to the Species of *Menticirrhus*

- 1a Breast scales not uniform in size; those towards head notably smaller than scales along lateral line; molar teeth present on pharyngeal plates; pectoral fins short, not reaching beyond tip of pelvic fins; usually three or more gill rakers on lower limb of first branchial arch; color plain silvery gray; juveniles (less than 100 mm SL) with only a vestige of swimbladder. Lateral line scales 72-74. [D. X=XI+19-26; gill rakers short, 3-5+0-8=3-12]. . . . . *M. littoralis*  
(Chesapeake Bay to Brazil)
- 1b Breast scales uniform in size; about as large as those along lateral line; no molar teeth on pharyngeal plates; pectoral fin reaching to or beyond tip of pelvic fin; gill rakers tuberculate or absent on the lower limb of first branchial arch; side with dark oblique bars

- 2a Anal fin rays usually 7 (6-8); depressed spinous portion of dorsal fin seldom extends past origin of soft portion of dorsal fin; longest dorsal spine 16.2-24.1% of SL; juveniles (less than 100 mm SL) with only a vestige of swimbladder. [D. X, I+20-26; gill rakers short, (2-3)+(0-7)=2-10]. . . . . *M. americanus*  
(Chesapeake Bay to Argentina)
- 2b Anal fin ray usually 8 (7-9); depressed spinous portion of dorsal fin extends past origin of soft portion of dorsal fin; longest dorsal spine 24.6-38.9% of SL; juveniles (less than 100 mm SL) with well-developed swimbladder. [D. X, I+22-27; gill rakers short, (3-5)+(0-7)=3-12]. . . . . *M. saxatilis*  
(Gulf of Maine to Mexico)

Key to the Species of *Micropogonias*

- 1a Dark spots on scales above lateral line forming continuous streaks nearly as wide as interspaces; 6 to 7 scales between dorsal fin origin and lateral line in vertical series; pelvic fin longer, less than 1.6 times in head; D. X+I, 26-30 (usually 26-28); A. II, 7-8; gill rakers short,  $(7-9)+(12-15)=21-25$ . . . . *M. furnieri* (Venezuela to Brazil)
- 1b Dark spots on scales above lateral line not forming continuous streaks; 8 to 9 scales between dorsal fin origin and lateral line in vertical series; pelvic fin shorter, more than 1.6 times in head; D. X+I, 27-30 (usually 28-29); A. II, 8-9; gill rakers short,  $(8-10)+(14-18)=22-29$ . . . . *M. undulatus* (Chesapeake Bay to Brazil)

Key to the Species of *Ophioscion*

- 1a Dorsal fin with more than 25 rays; anal fin with 9 rays; D. X+I, 28-29; gill rakers  $9+17=26$ ; lateral line scales 50-57; body with oblique streaks above lateral line (also see remarks in Synopsis). . . . *O. adustus* (Caribbean Islands to Brazil)
- 1b Dorsal fin with less than 25 rays; anal fin with 7 rays
- 2a Lateral line pores 49-52; D. X+I, 20-21; gill rakers  $(7-9)+14=21-23$ . . . . *O. panamensis*
- 2b Lateral line pores 54-57; D. X+I, 23-24; gill rakers  $(7-8)+(13-16)=20-24$ . . . . *O. punctatissimus* (Panama to Brazil)



Key to the Species of *Paralonchurus*

- 1a Pectoral fin short and pale, not extending beyond the tip of pelvic fin; eye diameter moderate (about 5 times in head); soft dorsal with 28-31 rays; body with vertical dark stripe and dark humeral spot above the pectoral fin origin; vertebrae 11+18=29; gill rakers short,  $(3-5)+(6-9)=10-14$ . . . . .*P. brasiliensis*  
(Venezuela to Brazil)
- 1b Pectoral fin very long and jet black, reaching to anal fin origin; eye diameter small (about 9 times in head); soft dorsal 31-33 rays; body uniform brown; vertebrae 10+15=25; gill rakers short,  $3+4=7$ . . . . .*P. elegans*  
(Surinam to Brazil)

Key to the Species of *Pareques*

- 1a Side with 7-10 narrow longitudinal stripes narrower than pupil, pectoral fin pale, [D. XI-X+I, 38-40, A.II, 7; gill rakers short,  $(4-6)+(10-12)=15-18$ ]. . . . .*P. umbrosus*
- 1b Side with 3 to 5 broad longitudinal stripes wider than pupil; and narrow stripes in between them, pectoral fin dark brown. [D. VIII-IX+I, 37-41, A, II, 7-8, gill rakers short,  $(5-6)+(9-14)=14-20$ ]. . . . .*P. accuminatus*

Key to the Species of *Stellifer*

- 1a Preopercular margin with two or three strongly developed spines at the angle

- 2a Preopercular margin with three strong spines (occasionally four), mouth oblique (Fig. 38, B); gill rakers (12-14)+(20-25)=32-38; [D. XI+I, 18-20; A. II, 8; longer than 2/3 of eye; pectoral fin about 28.5 to 31% of SL (72-106 mm SL specimens)]. . . . . *S. stellifer*  
(Venezuela to Brazil)
- 2b Preopercular margin with two strong spines; mouth horizontal; gill rakers on first arch 40 or more; [D. XI-XII+I, 21-24; A. II, 8-9]
- 3a Gill rakers 16-20+(24-31)=40-51; pectoral fin length about 26.8 to 28.8% of SL (90-100 mm SL specimens); one to several median predorsal rows of ctenoid scales on nape. . . . . *S. rastrifer*  
(Venezuela to Brazil)
- 3b Gill raker 19-22+(32-35)=51-57; pectoral fin length about 32.5 to 34% of SL (90-100 mm SL specimens); no predorsal ctenoid scales on nape. . . . *S. griseus*  
(Venezuela)
- 1b Preopercular margin with four or more strongly developed spines at the angle
- 4a Gill rakers 28 or fewer; upper jaw 2.8 or more in head, mouth more or less ventral in position; when mouth closed, snout projecting in front of premaxillae (upper jaw)
- 5a Scales on top of head mostly cycloid; tip of pectoral fin extends to about level of tip of pelvic fin; swimbladder with tube-like diverticula

- 6a Gill rakers  $(8-9)+(13-14)=21-23$ ; D.  $X=XI+I$ , 19-21; A. II, 8-10 (usually 9); diverticula on swimbladder short, digit-like (Fig. 16, C). . . .*S. microps*  
(Columbia to Brazil)
- 6b Gill rakers  $8+(14-16)=22-24$ ; D.  $X+I$ , 22; A. II, 9; diverticula on swimbladder very long, U-shaped (Fig. 16, D). . . . .*S. brasiliensis*  
(Brazil)
- 5b Scales on top of head ctenoid; tip of pectoral fin extends beyond tip of pelvic fin; swimbladder with bulb or bean-shaped diverticula (Fig. 16, A)
- 7a Tip of pelvic fin ends slightly anterior to the tip of pectoral fin; gill rakers  $(8-9)+(15-16)=23-25$ ; [D.  $XI+I$ , 20-22; A. II, 8]. . .*S. naso*  
(Venezuela to Brazil)
- 7b Tip of pelvic fin end far before the tip of pectoral fin; gill rakers  $(9-10)+(16-19)=26-28$ ; [D.  $XI=XII+I$ , 21-22; A. II, 8 (rarely 9)]. . . . .  
. . . . .*S. venezuelae*  
(Venezuela)
- 4b Gill rakers 29 or more; upper jaw 2.6 or less in head, mouth more or less oblique in position; when mouth closed, snout equal to or projecting slightly in front of premaxillae (upper jaw)
- 8a Tip of upper lip on horizontal with or above ventral margin of eye; snout not projecting beyond upper lip; D.  $XI-XII+I$ , 20-24; gill rakers  $(10-13)+(22-23)=32-36$ ;

eye 4.7-5.5 times in head . . . . .*S. lanceolatus*  
(Chesapeake Bay to Gulf of Mexico coasts)

- 8b Tip of upper lip on horizontal line well below ventral margin of eye; snout projecting beyond upper lip;  
D. XI+I, 23-24; gill rakers (10-12)+(19-22)=29-33; eye  
5.5-6.2 times in head . . . . .*S. colonensis*  
(Caribbean islands and Central America)

Key to the species of *Umbrina*

- 1a Anal fin rays 7 to 8; mental (chin) barbel short; total gill rakers 19-20
- 2a Eye larger, about 9.8 to 10.7% of SL; caudal peduncle circumferential scales 22; mental barbel with an apical pore; D. X, I+24-25; A, II, 7; gill rakers (8-9)+(12-13)=20-22. . . . .*U. canosai*  
(southern Brazil to Argentina)
- 2b Eye smaller, about 5.9 to 6.2% of SL; caudal peduncle circumferential scales 18 to 19; mental barbel with a pore on the middle anterior surface; D. X., I+22-23; A. II, 8; gill rakers (7-8)+(11-13)=19-20. . . . .*U. milliae*  
(offshore Columbia)
- 1b Anal fin rays 6; mental (chin) barbel relatively long and slender; total gill rakers 13 to 15
- 3a Scale rows beneath spinous dorsal fin more or less parallel to lateral line; scales in diagonal series from dorsal fin origin to lateral line 5-6; stripes on body less distinct;

D. X+I, 23-26; [gill rakers (5-7)+(7-10)=13-15] . . . . .

. . . . . *U. broussonetii*

(Caribbean Sea)

3b Scale rows beneath spinous dorsal fin situated at an angle of about 30° to lateral line; scales in diagonal series of dorsal fin origin to lateral line usually 8, sometimes 7; stripes on body distinct; D. X+I, 26-31. (except for specimens from western Gulf of Mexico, which have slightly lower counts, 24-26); [gill rakers (5-7)+(7-10)=13-15] . . . . .

. . . . . *U. coroides*

(Florida to Brazil)

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Table 1. Summary of morphological characters of the supra-generic groups of western Atlantic Sciaenidae

SUPRA-GENERIC		MORPHOLOGICAL CHARACTERS			
GROUPS	SWIMBLADDERS	OTOLITHS	SNOUT PORES	MENTAL PORES & BARBELS	
<i>Micropogonias</i>	with a pair tube-like diverticula	sagitta shield-like, very thick	5 upper 5 marginal	5 pores many barbels	
<i>Nebris</i>		olive-shaped, cauda of sulcus very deep	no upper 2 marginal	4 pores no barbel	
<i>Pogonias</i>	with complicate diverticula in adults	sagitta oval or sub-oval, moderately thin to very thick	5 upper 5 marginal	5 pores many barbels	
<i>Sciaenops</i>				5 pores no barbel	
<i>Larimus</i>	simple without complicate diverticula, sometime with anterior projection		no upper 5 marginal	4 pores no barbel	
<i>Sciaena</i>			3 to 7 upper 5 to 7 marginal	5 pores no barbel	
<i>Umbrina</i>				4 pores 1 barbel	
<i>Menticirrhus</i>	atrophied in adults	thin and elongate, some partially thickened			
<i>Lonchirus</i>	with a pair of anterior horns, some also with a pair of tube-like diverticula, some reducing size in adult			5 pores 2 or many barbels	
<i>Cynoscion</i>			no upper 2 marginal	no pores no barbel	
<i>Stellifer</i>	with two chambers, anterior one yoke-shaped, posterior one carrot-shaped	both sagitta and lapillus enlarged	3 to 7 upper 5 marginal	4 to 6 pores no barbel	

Fig. 1

Terminology of pore system on head of western Atlantic sciaenids. Snout; upper pores (above the dotted line) and marginal pores (below the dotted line). Lower jaw; mental pores and barbel(s). A. ventral view of the mouth, *Menticirrhus saxatilis*. B. tip of the snout, *Stellifer lanceolatus*.

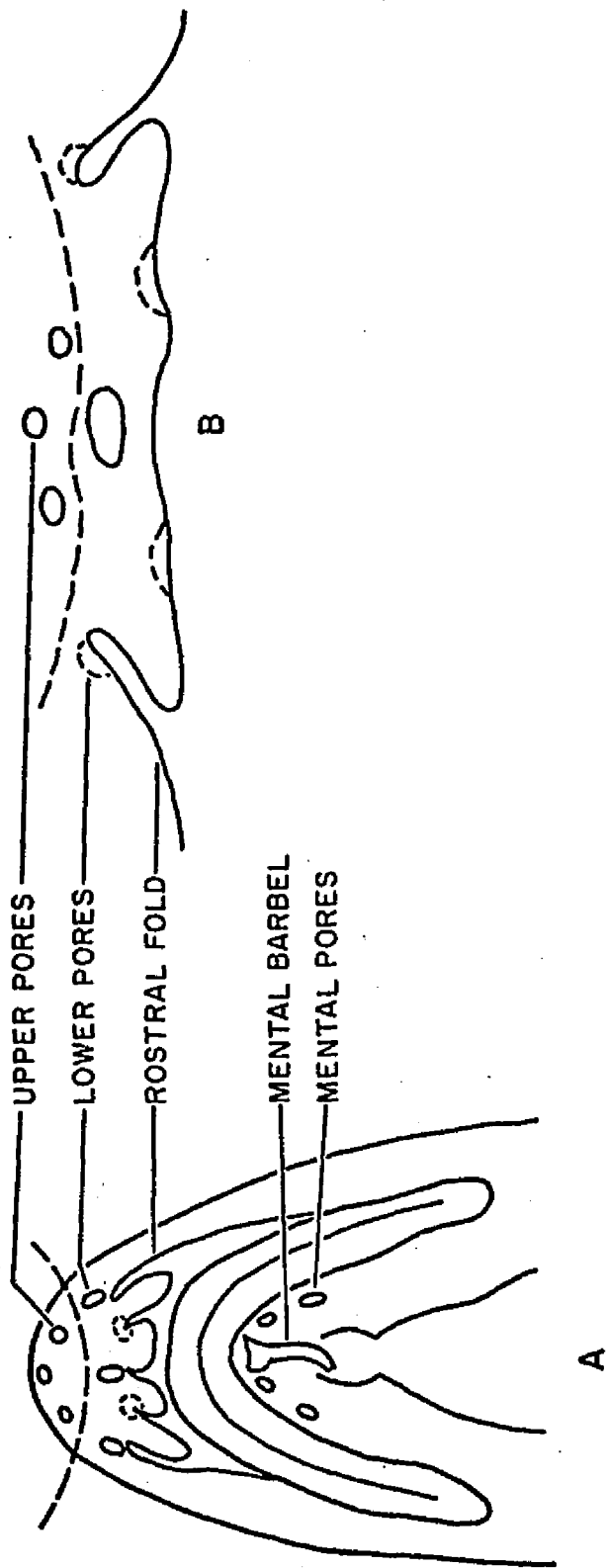
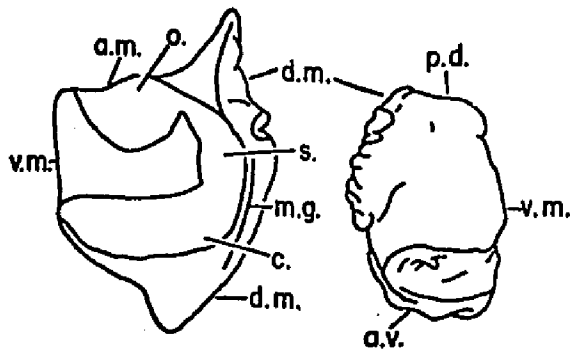


Fig. 2

Terminology of otoliths. A, A' & a, sagitta and B, B' & b, lapillus of *Bairdiella chrysoura*; C, C' & c, sagitta of *Leiostomus xanthurus*. A, B & C, inner surface; A', B' & C' outer surface; a, b & c, lateral view with inner surface down and anterior end to the left. a.m. anterior margin; a.v. anterior ventral margin; d.m. dorsal margin; c. cauda section of sulcus; m.g. marginal groove; o. ostium section of sulcus; p.d. posterior dorsal margin; s. "tadpole-shaped" sulcus; v.m. ventral margin. All the figures of otoliths (sagittae and lapilli) are oriented the same as this figure.

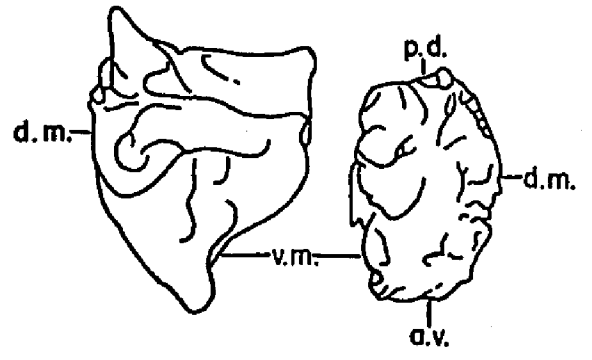
INNER SURFACE



A

B

DORSAL SURFACE



A'

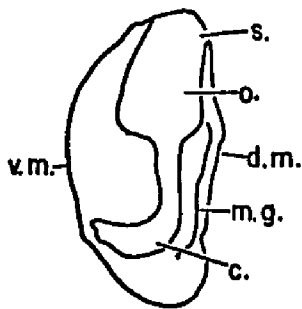
B'



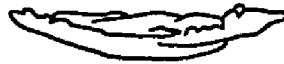
a



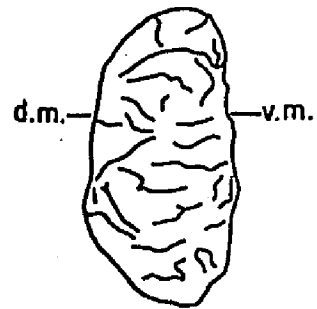
b



C



c

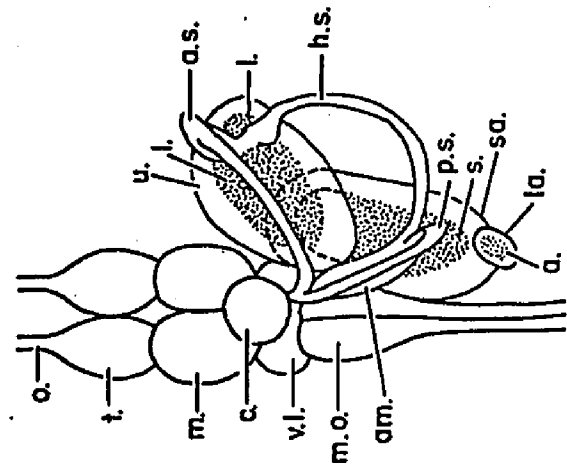


C'

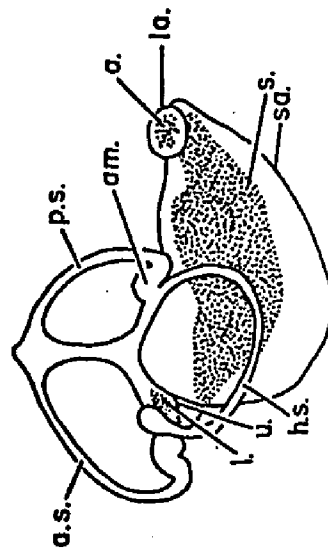
Fig. 3

The inner ears of sciaenids. A & A', with only sagitta enlarged, *Pseudosciaena crocea* (Richardson), after Chu, Lo and Wu (1963; fig. 8, A & B). B & B', with both sagitta and lapillus enlarged, *Stellifer lanceolatus* (Holbrook). A & B, dorsal view of right inner ear. A' & B', lateral view of left inner ear. a. asteriscus; am. ampulla; a.s. anterior semicircular canal; c. cerebellum, h.s. horizontal semicircular canal; l. lapillus; la. lagena; m. mesencephalon; m.o. medulla oblongata; o. olfactory nerve; p.s. posterior semicircular canal; s. sagitta; sa. sacculus; t. telencephalon; u. utriculus; v. vergus nerve; v.l. vegal lobe.

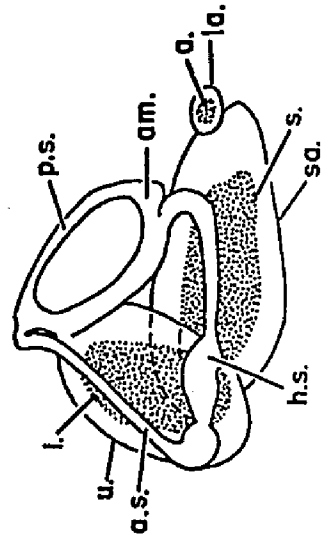




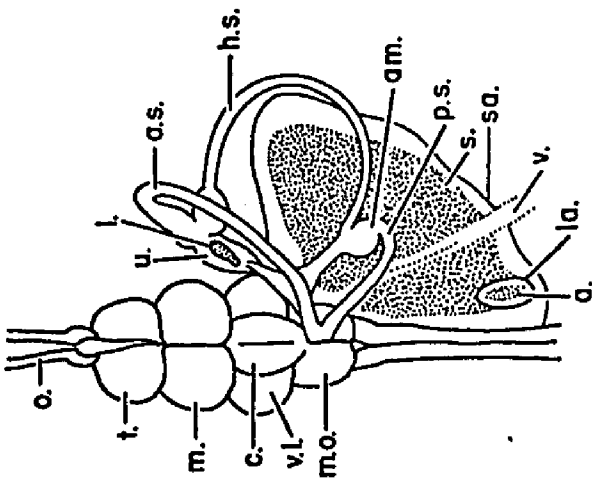
A



A'



B'



B

Fig. 4

Phylogenetic relationships of genera and supra-generic groups of western Atlantic Sciaenidae. Subfamily level lies between lines A and B; tribal level lies between lines C and D; X, Y, & Z represent clusters discussed in the text.

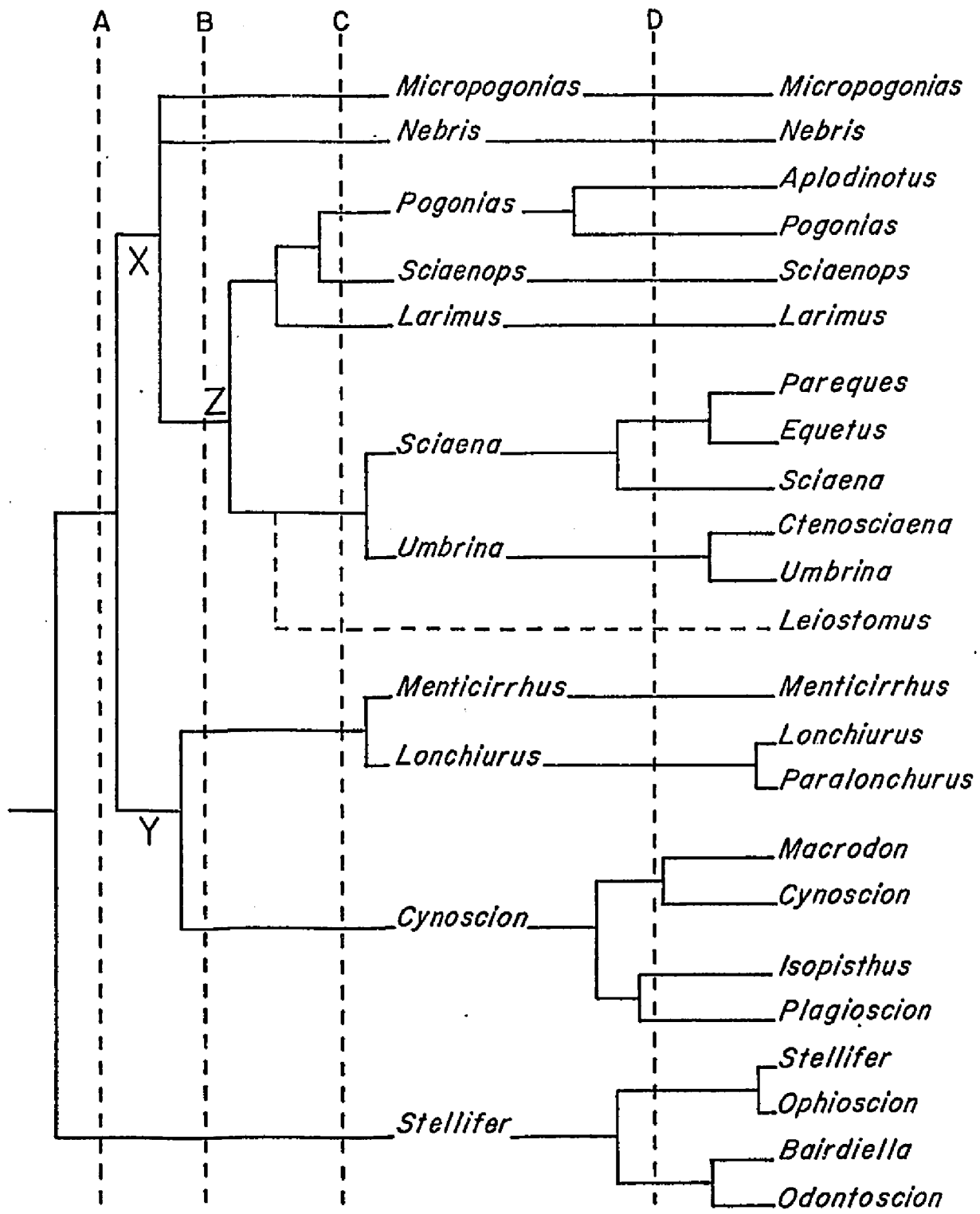
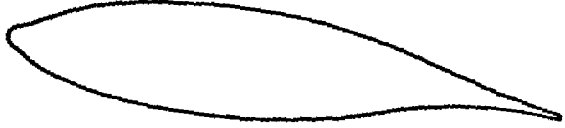


Fig. 5

Swimbladders of the *Sciaena* pattern. A. *Sciaena trewavasae*; B. *Leiostomus xanthurus*; C. *Equetus punctatus*; D. *Aplodinotus gruniens*; E. *Larimus fasciatus*; F. *Larimus breviceps*.



A



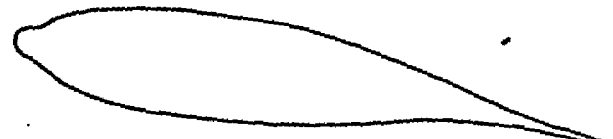
B



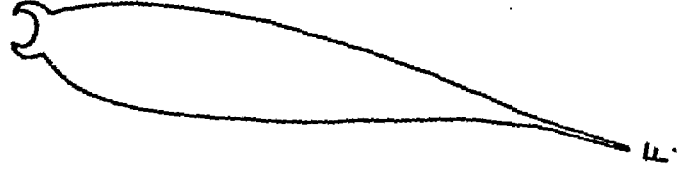
C



D



E



F

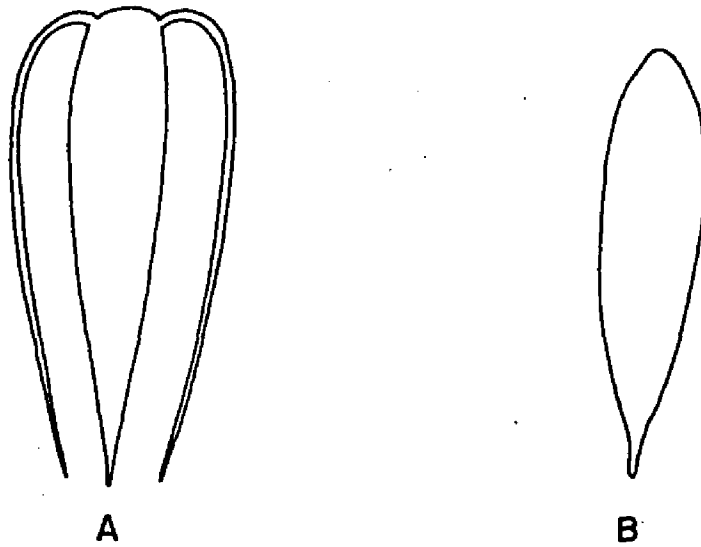
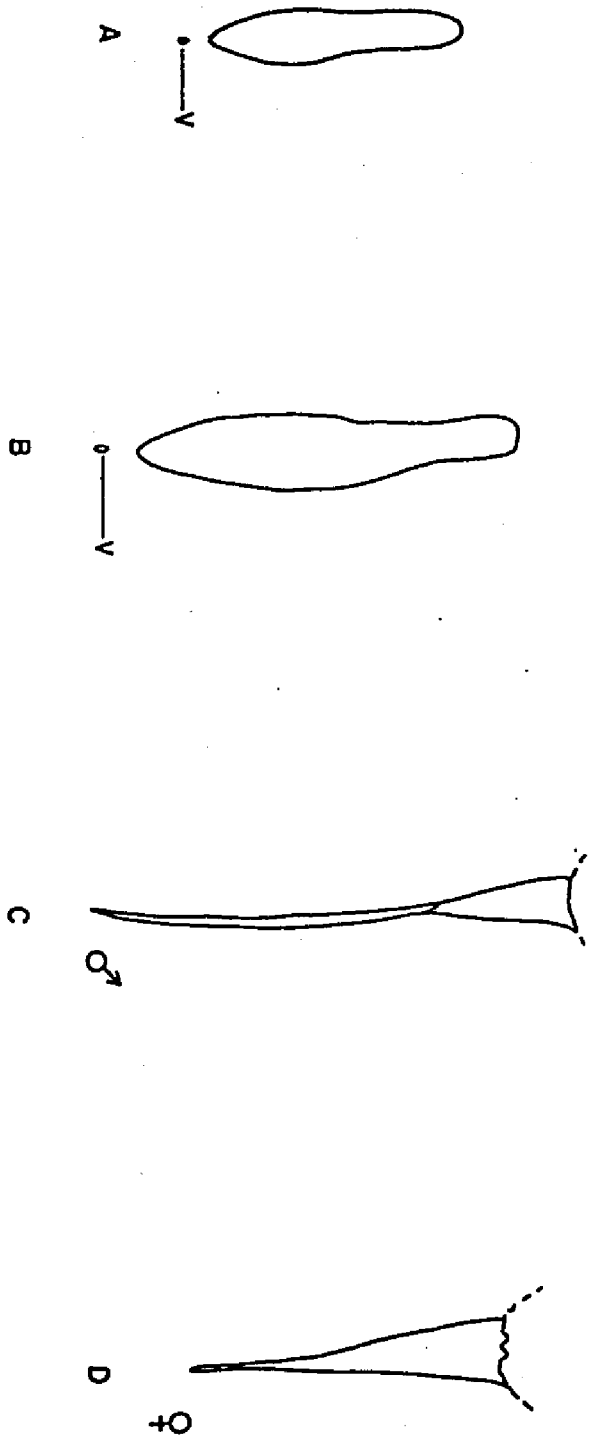
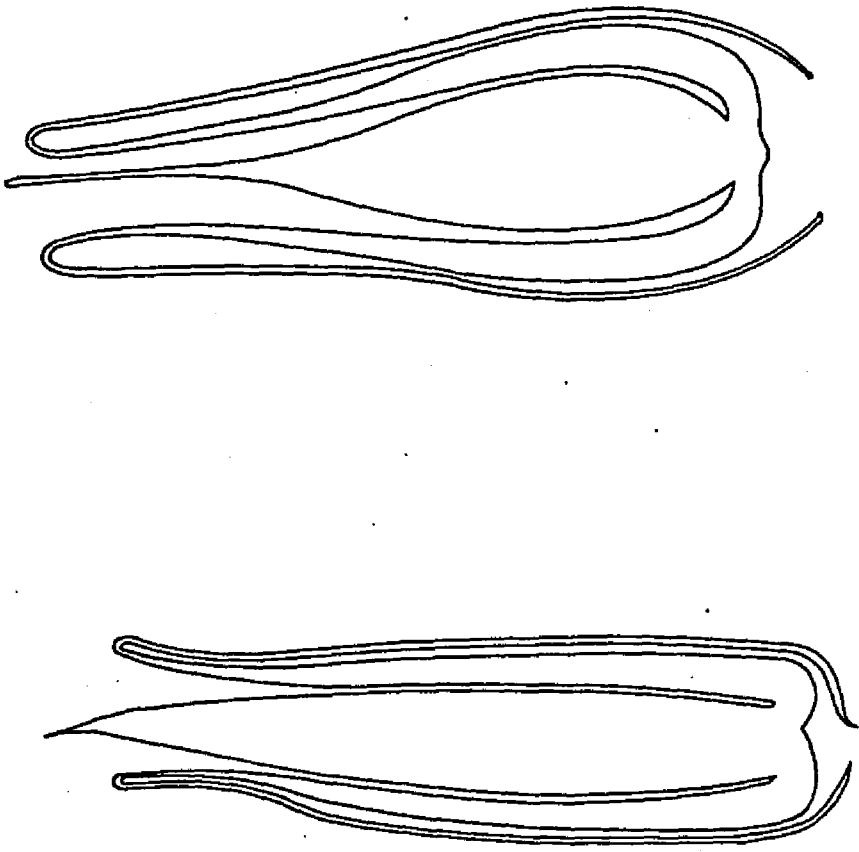


Fig. 6. Swimbladders of two species of *Pachyurus*. A. *P. bonariensis*;  
B. *P. schomburgkii*.

Fig. 7. Swimbladders of *Menticirrhus saxatilis* (x2). A. 58 mm SL; B. 75.5 mm SL; C. 185 mm SL; D. 182 mm SL.





A

B

Fig. 8. Development of swimbladder in *Nebris microps*. A. 125 mm SL (x2); B. 273 mm SL (x1).



Fig. 9

Variation and development of swimbladders of the  
*Micropogonias* (xl). *M. undulatus*, A. 67.1 mm SL;  
A'. 180 mm SL and *M. furnier*, B. 160 mm SL; B'.  
162 mm SL. v. vent.

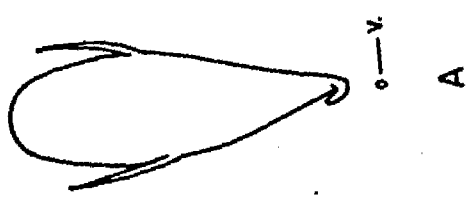
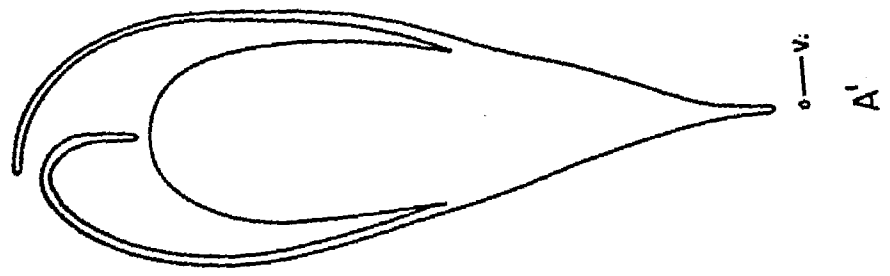
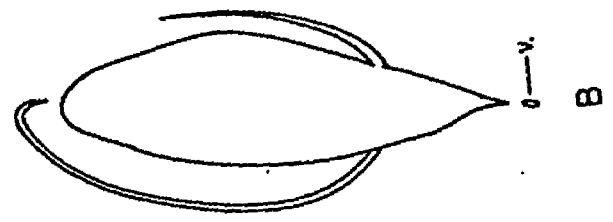
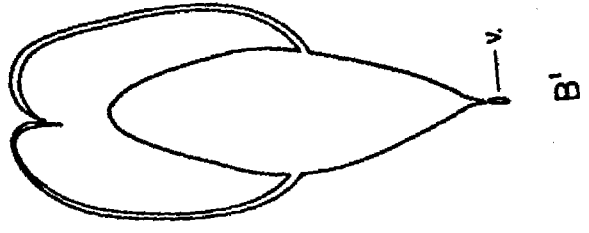


Fig. 10

Ontogenetic development of swimbladders in  
*Pogonias cromis*. A. 24 mm TL; B. 57.4 mm TL;  
C. 150 mm TL; D. 236 mm TL; E. 1090 mm TL.  
C, D & E on the left sides of dotted line  
represent dorsal view of the swimbladders  
(diverticula omitted); the shaded areas represent  
intrinsic drumming muscles.

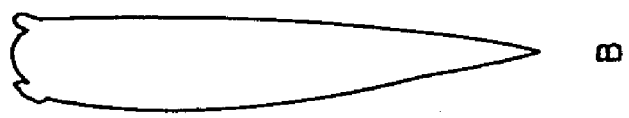
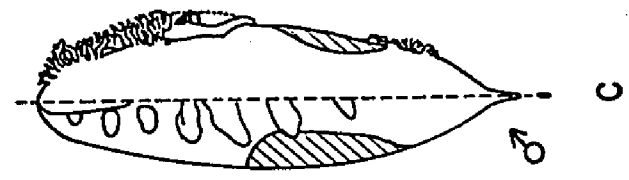
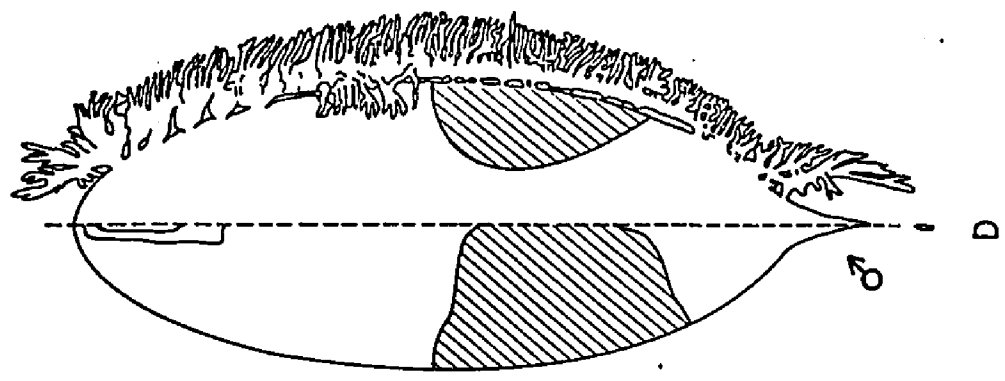
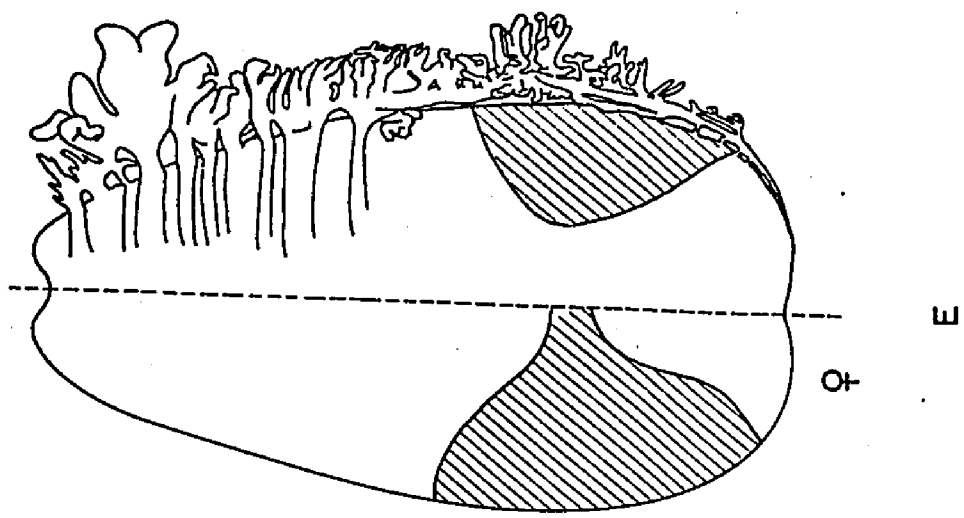


Fig. 11

Ontogenetic development of swimbladder in *Sciaenops ocellata*. A. 197 mm SL; B. 980 mm SL, shaded area represents fat tissue; C. 1082 mm SL, dorsal view of a portion of the swimbladder with the left side of the "sac-like" projection; c. a cross section of the dorsal projection to show labyrinthine chambers.

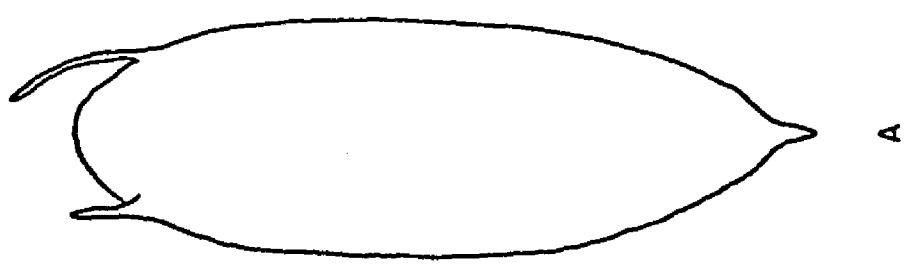
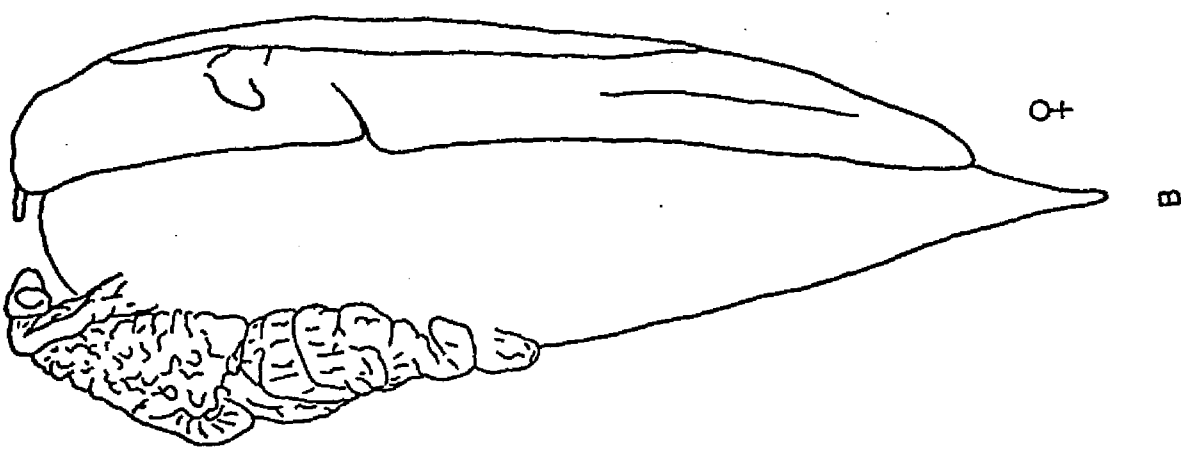
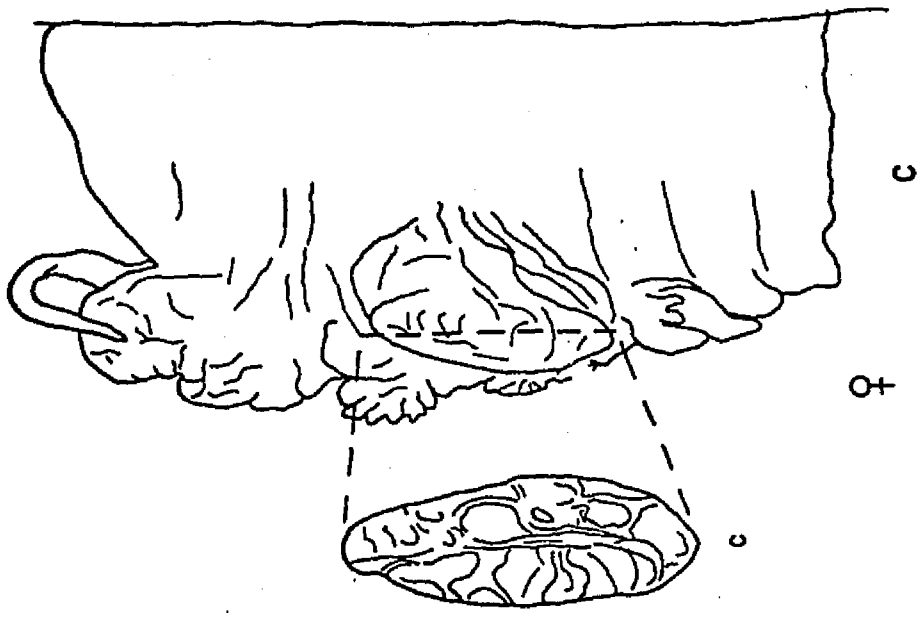
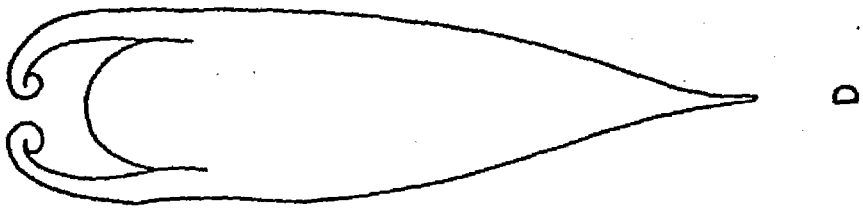
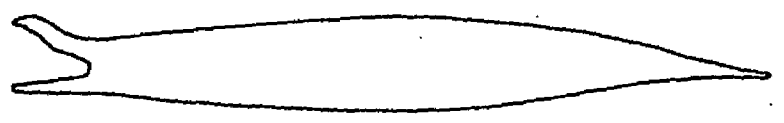


Fig. 12

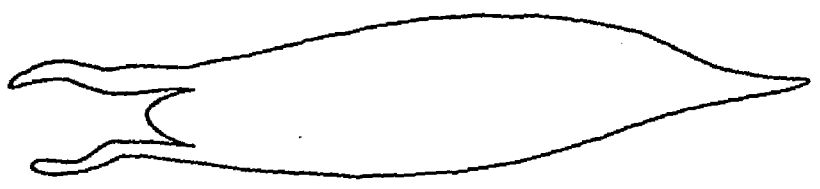
Swimbladders of the *Cynoscion* pattern. A.  
*Cynoscion regalis*; B. *Isopisthus parvipinnis*;  
C. *Macrodon ancylodon*; D. *Plagioscion*  
*surinamensis*.



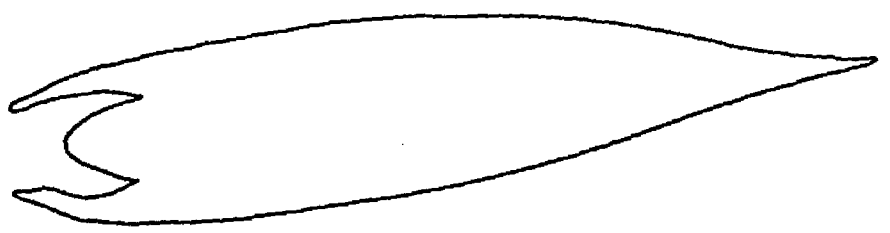
D



C



B



A

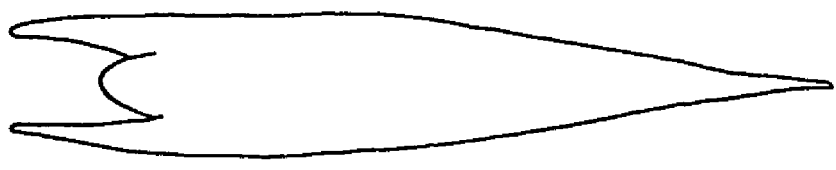


Fig. 13

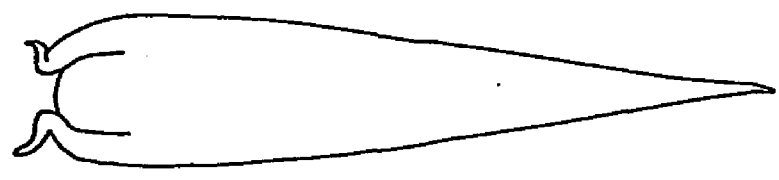
Variation of the swimbladders in the genus  
*Cynoscion*. A. *C. microlepidtus*; B. *C. nothus*;  
C. *C. virencens*; D. *C. leiarchus* (395 mm SL)  
D'. *C. leiarchus* (191 mm SL).



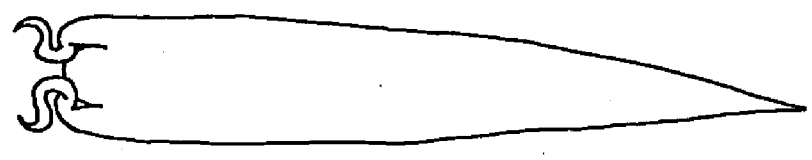
A



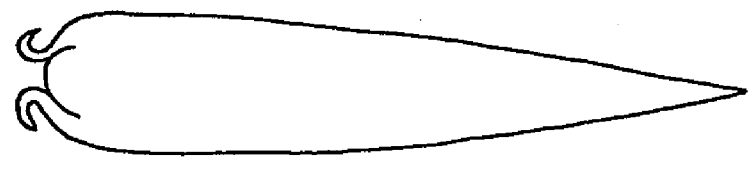
B



C



D



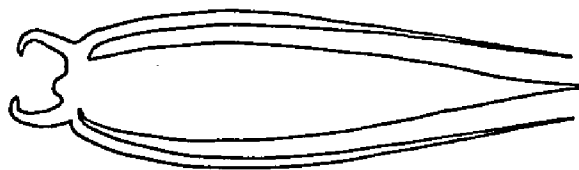
D'

Fig. 14

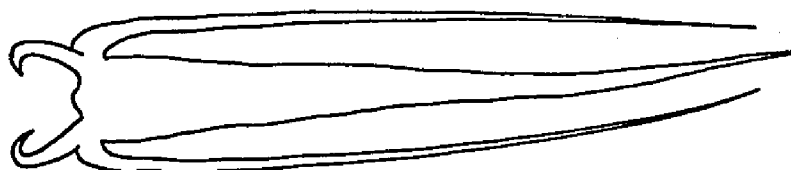
Relative size and development of swimbladders in the *Lonchiurus* pattern (x2). A. *Lonchiurus lanceolatus* (87.7 mm SL); A'. *L. lanceolatus* (103 mm SL); A". *L. lanceolatus* (114 mm SL); B. *Paralonchurus elegans* (180 mm SL); C. *P. brasiliensis* (157 mm SL); D. *Pachypops furcroides*, after Cuvier and Valenciennes (1830; pl. 138, "Corb Fourcroy", x1), original size unknown.  
v. vent.



D

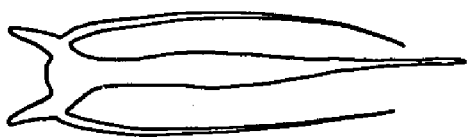


C



o-v.

B



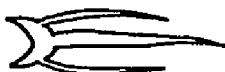
o-v.

A''



o-v.

A'



o-v.

A

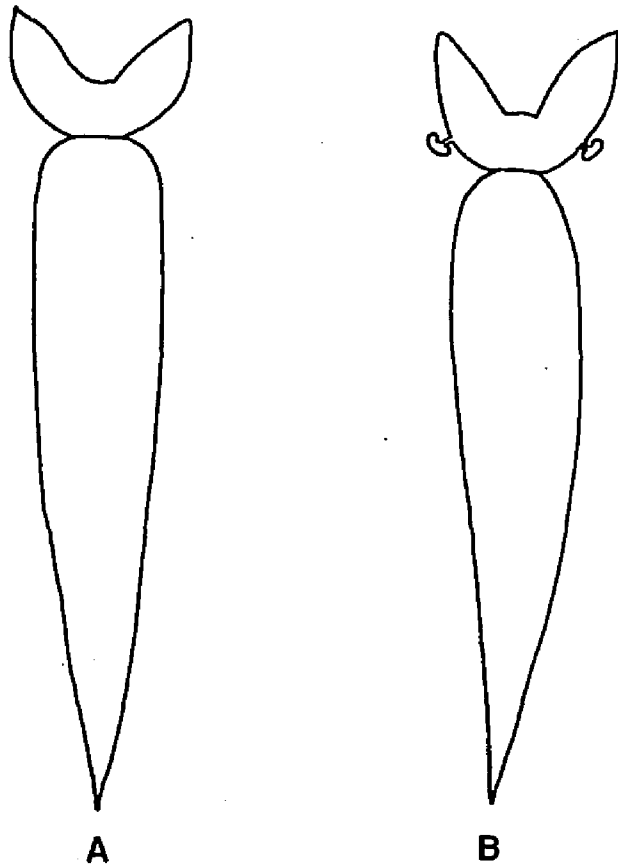
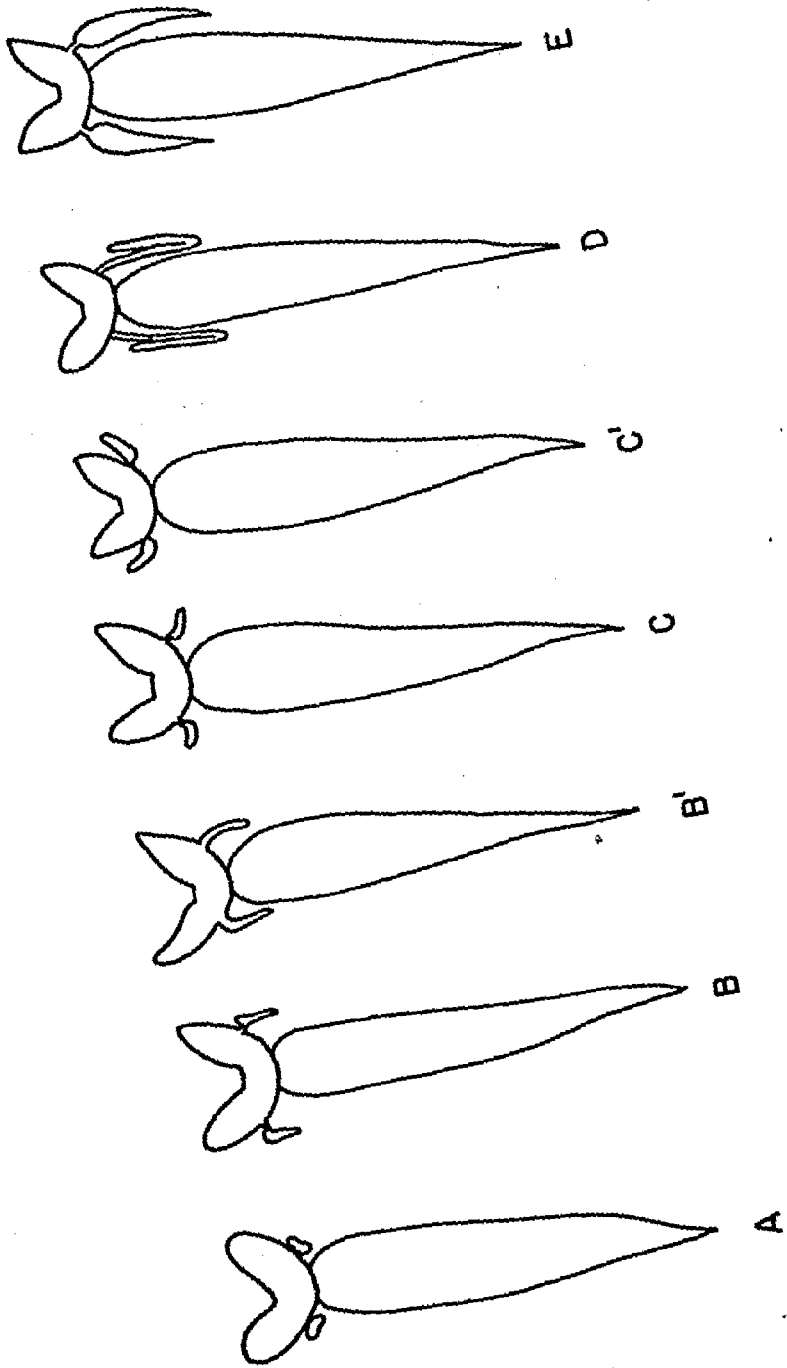


Fig. 15. Swimbladders of the *Stellifer* pattern. A. *Bairdiella*,  
*Odontoscion* and *Ophioscion*; B. *Stellifer lanceolatus*.

Fig. 16

Variation of the swimbladder diverticula in the genus *Stellifer*. A. *S. naso*; B. *S. griseus* (84.2 mm SL); B'. *S. griseus* (108 mm SL); C. *S. microps* (129 mm SL); C'. *S. microps* (105 mm SL); D. *S. brasiliensis*; E. *S. rastrifer*.



SWIMBLADDER PATTERNS

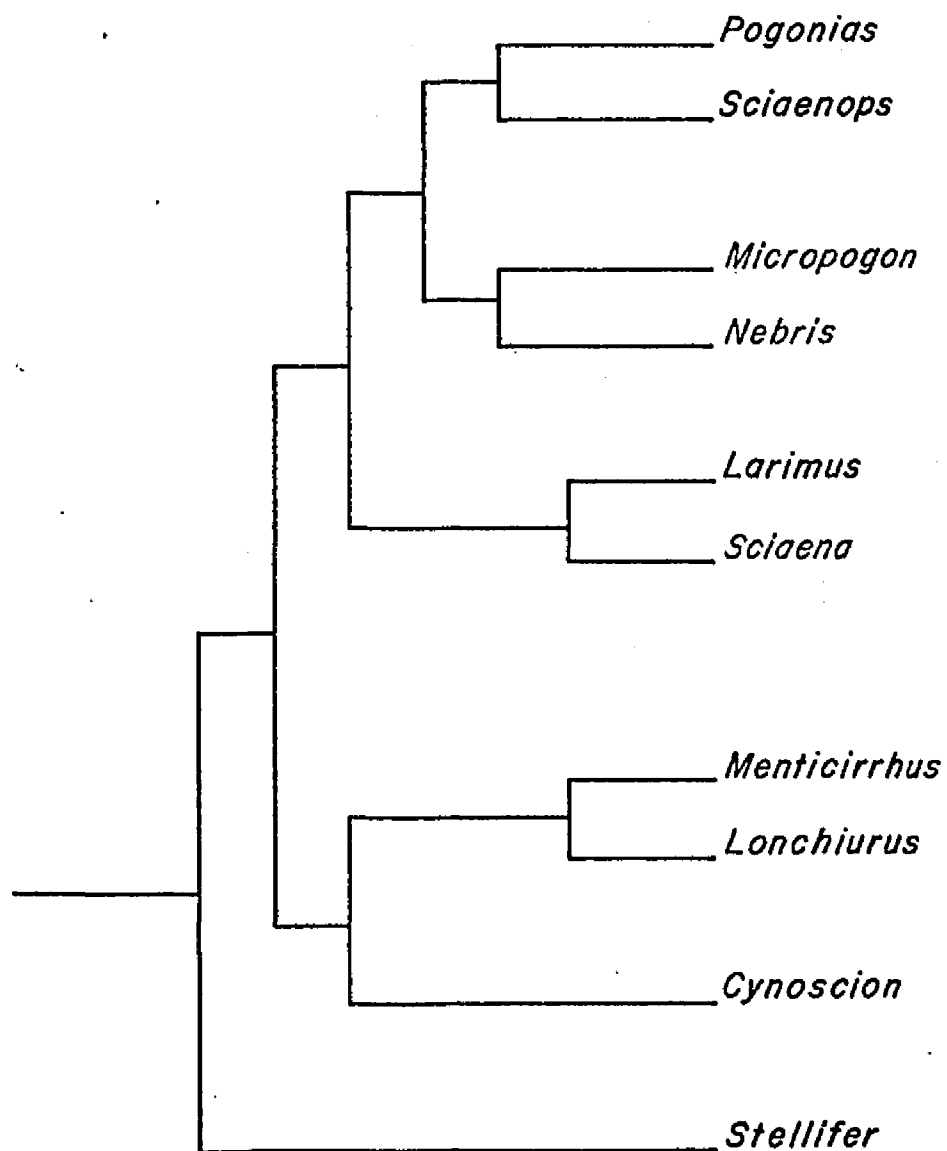


Fig. 17. Phylogenetic relationships of western Atlantic Sciaenidae as shown by swimbladder patterns.



Fig. 18

Inner surface (capital letters) and lateral view (lower case letters) of sagittae of the *Sciaena* pattern. A & a, *Ctenosciaena gracillicirrus*; B & b, *Equetus lanceolatus*; C & c, *Equetus punctatus*; D & d, *Leiostomus xanthurus*; E & e, *Paraques acuminatus*; F & f, *Paraques umbrosus*; G & g, *Sciaena trewavasae*; H & h, *Sciaena bathytatos*; I & i, *Umbrina coroides*; J & j, *Umbrina millae*; K & k, *Pachyurus schomburgkii*; L & l, *Plagioscion surinamensis*.

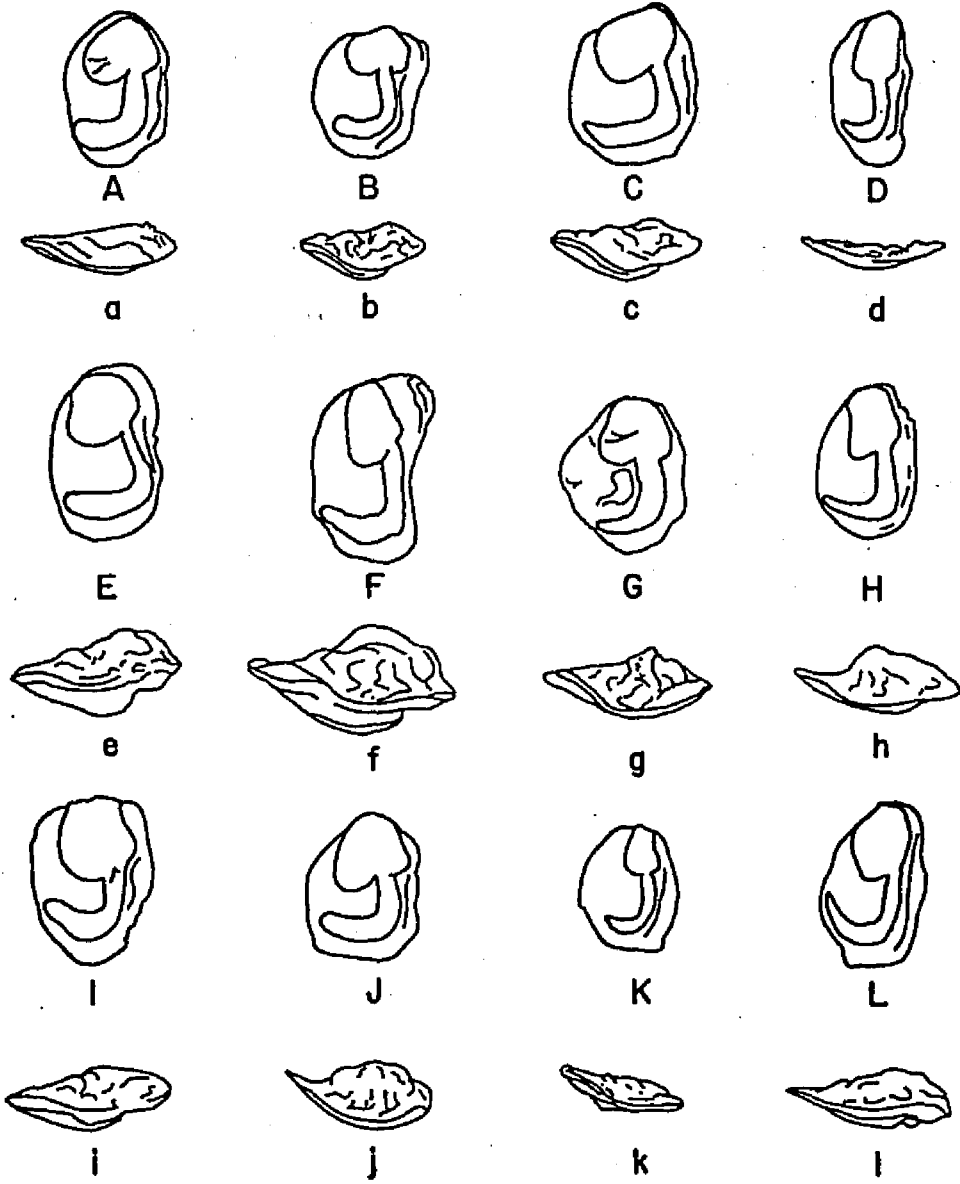
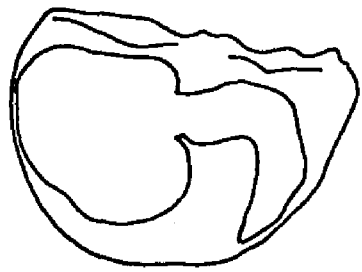


Fig. 19

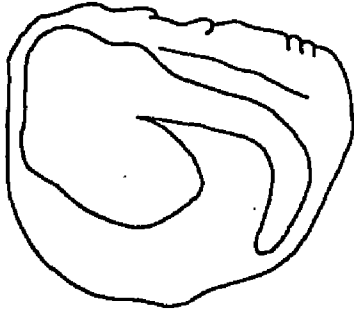
Sagittae of the *Pogonias* pattern. A, A' & a, *Pogonias cromis*; B, B' & b, *Aplodinotus grunniens*. A & B, inner surface; A' & B', outer surface; a & b, lateral view.



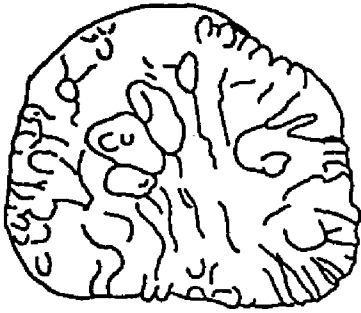
A



A'



B



B'



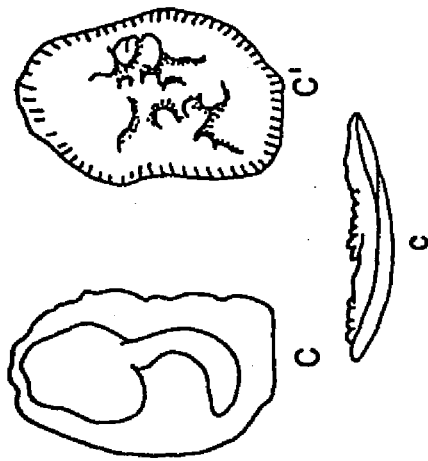
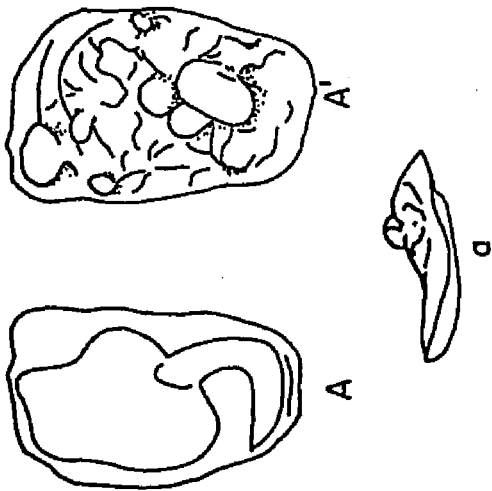
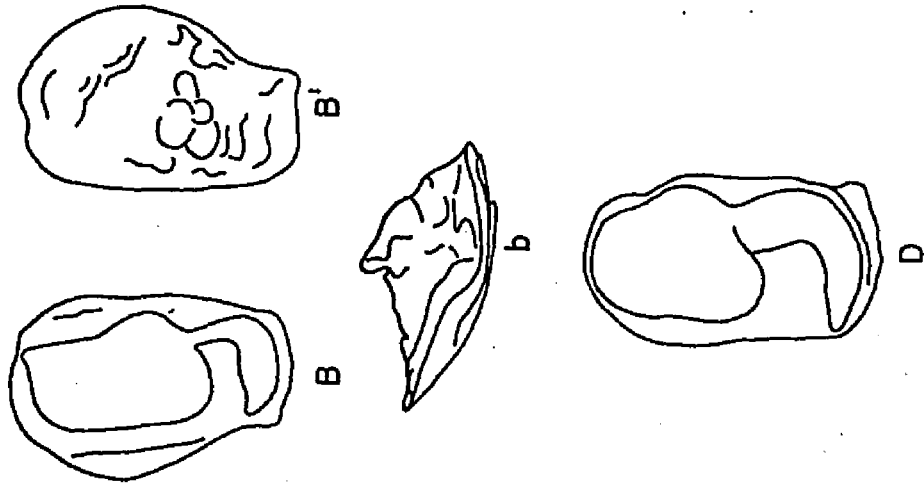
a



b

Fig. 20

Sagittae of the *Larimus* pattern. A, A' & a, *Larimus fasciatus*; B, B' & b, *Larimus breviceps*; C, C' & c, *Sciaenops ocellata*, 357 mm TL; D. *S. ocellata*, 1100 mm TL. A, B, C & D, inner surface; A', B' & C', outer surface; a, b & c, lateral view.



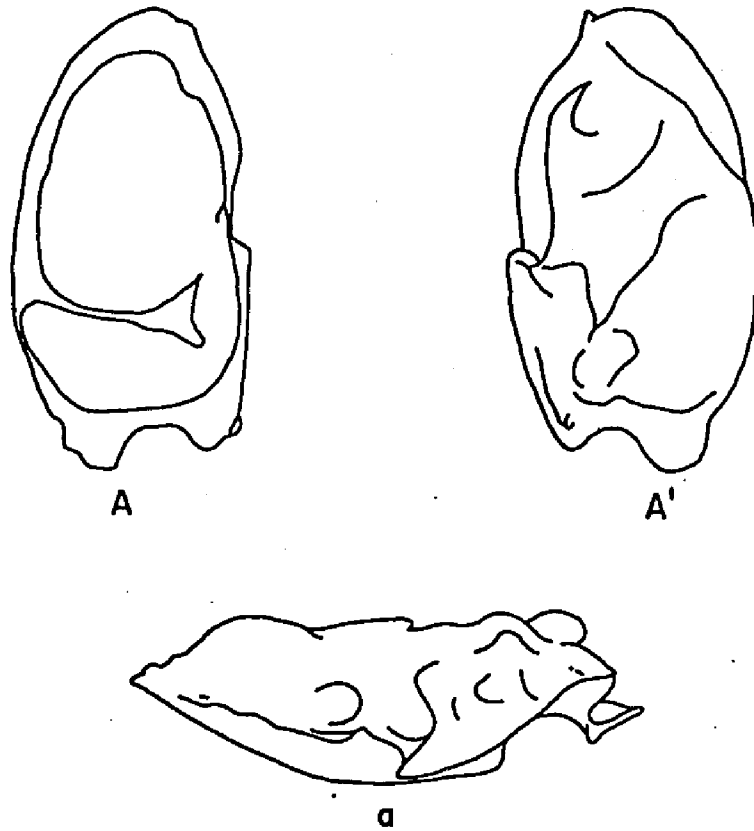


Fig. 21. Sagitta in *Nebris microps*. A. inner surface; A', outer surface; a, lateral view.

Fig. 22

Sagittae of the *Cynoscion* and the *Menticirrhus* patterns. A, A' & a, *Cynoscion nebulosus*; B, B' & b, *Menticirrhus saxatilis*. A & B, inner surface; A' & B', outer surface; a & b, lateral view.



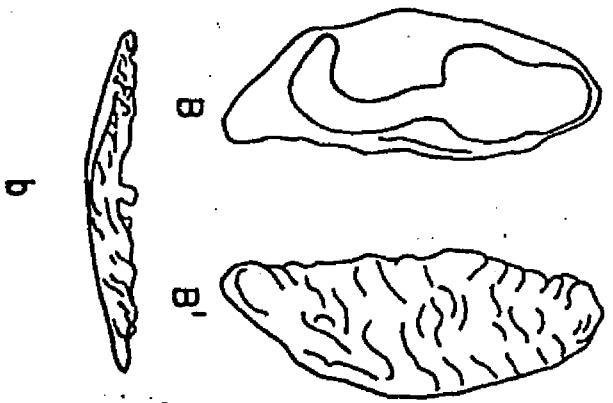
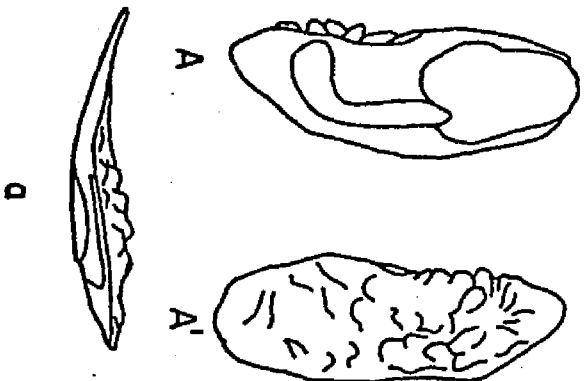
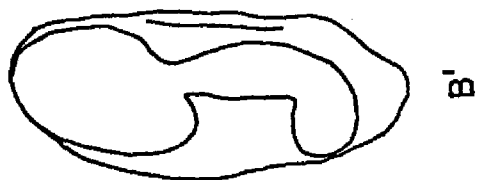


Fig. 23

Specific and ontogenetic variations of sagittae  
in species of *Cynoscion*. *C. regalis*, A & a, 370  
mm TL; A', 516 mm TL. *C. similis*, B & b, 166  
mm TL; B', 350 mm TL. *C. virens*, C & c,  
260 mm TL; C', 350 mm TL. A, A', B, B', C, &  
C', inner surface; a, b & c, lateral view.



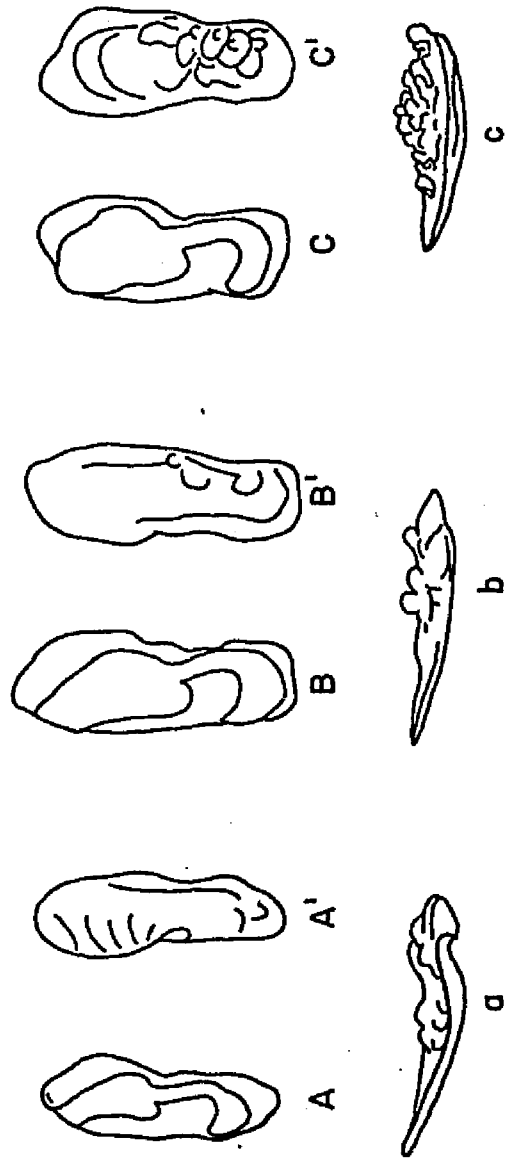


Fig. 24. Sagittae of the *Lonchurus* pattern. A, A' & a, *Lonchurus lanceolatus*; B, B' & b, *Paralonchurus elegans*; C, C' & c, *Paralonchurus brasiliensis*. A, B & C, inner surface; A', B' & C', outer surface; a, b & c, lateral view.

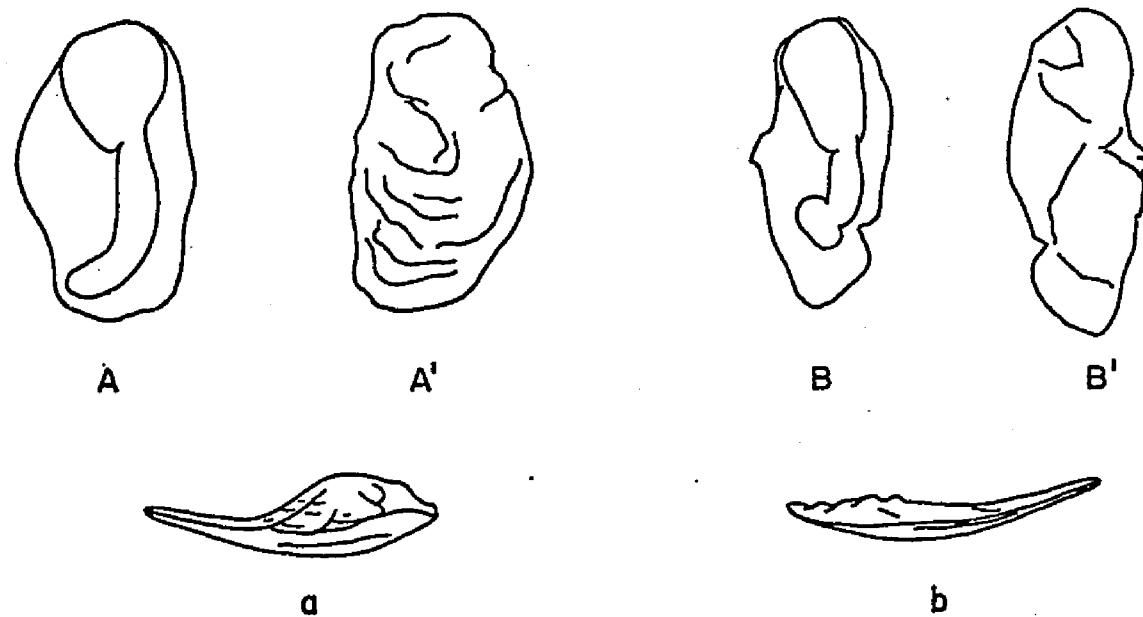


Fig. 25. Sagittae of the *Isopisthus* and the *Macrodon* pattern. A, A' & a, *Isopisthus parvipinnis*; B, B' & b, *Macrodon ancylodon*; A & B, inner surface; A' & B', outer surface; a & b, lateral view.

Fig. 26

Ontogenetic variations of the sagittae of the  
*Micropogonias*. *M. furnieri*, A, A' & a, 235 mm  
TL; B, B' & b, 295 mm TL; C, C' & c, 330 mm TL.  
*M. undulatus*, D, D' & d, 141 mm TL; E, E' & e,  
250 mm TL; F, F' & f, 350 mm TL. A, B, C, D,  
E & F, inner surface; A', B', C', D', E' & F',  
outer surface; a, b, c, d, e & f, lateral view.

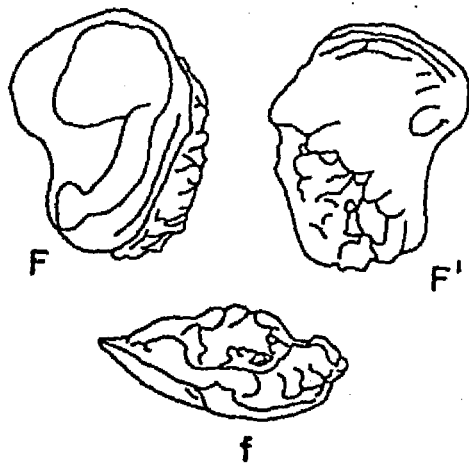
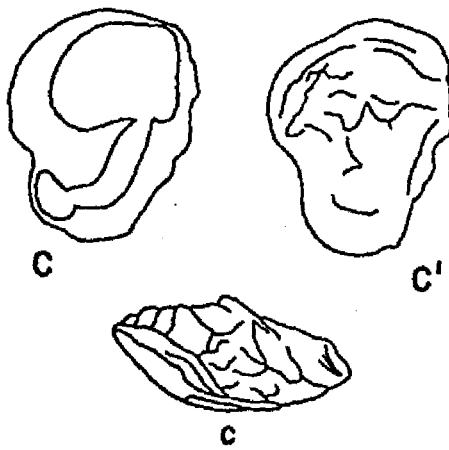
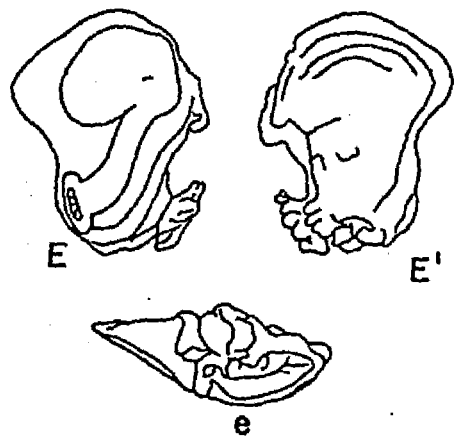
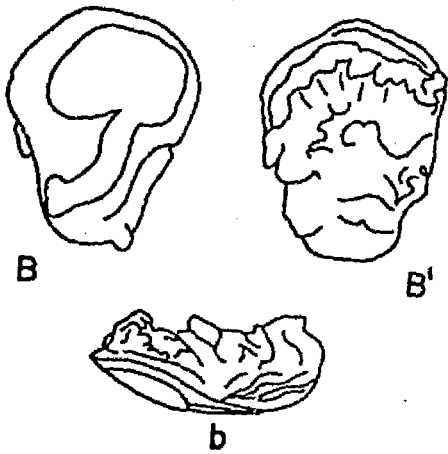
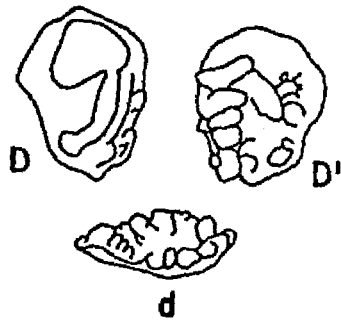
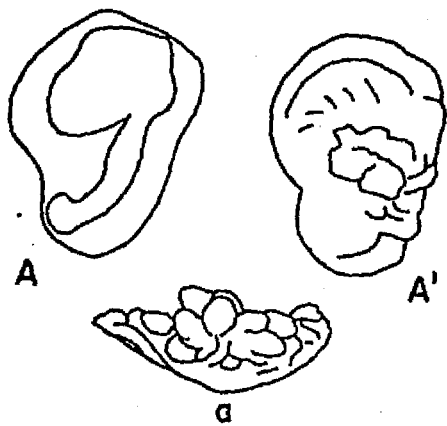
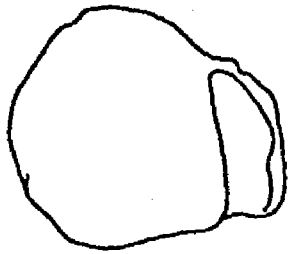


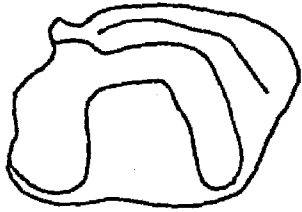
Fig. 27

Inner surface of right sagittae and lapilli of the *Stellifer* pattern. A & A', *Ophioscion punctatissimus*; B & B', *Stellifer lanceolatus*; C & C', *Bairdiella chrysoura*; D & D', *Odontoscion dentex*. A, B, C & D, sagittae; A', B', C' & D', lapilli.





B'



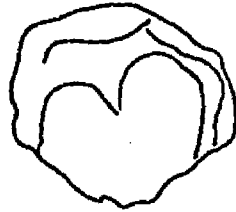
B



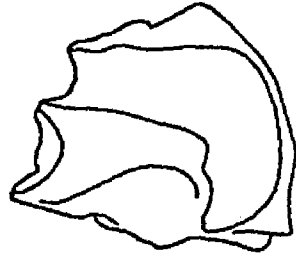
A'



A



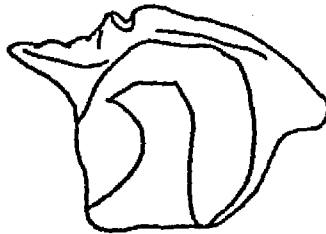
D'



D



C'



C

Fig. 28

Inner surface of right sagittae and lapilli in species of *Stellifer*. A & A', *S. colonensis*; B & B', *S. griseus*; C & C', *S. microps*; D & D', *S. rastrifer*. A, B, C & D, sagittae; A', B', C' & D', lapilli.



B'



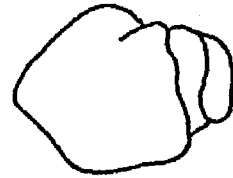
B



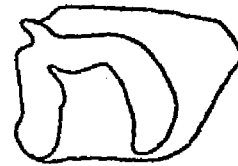
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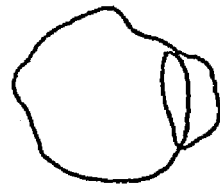
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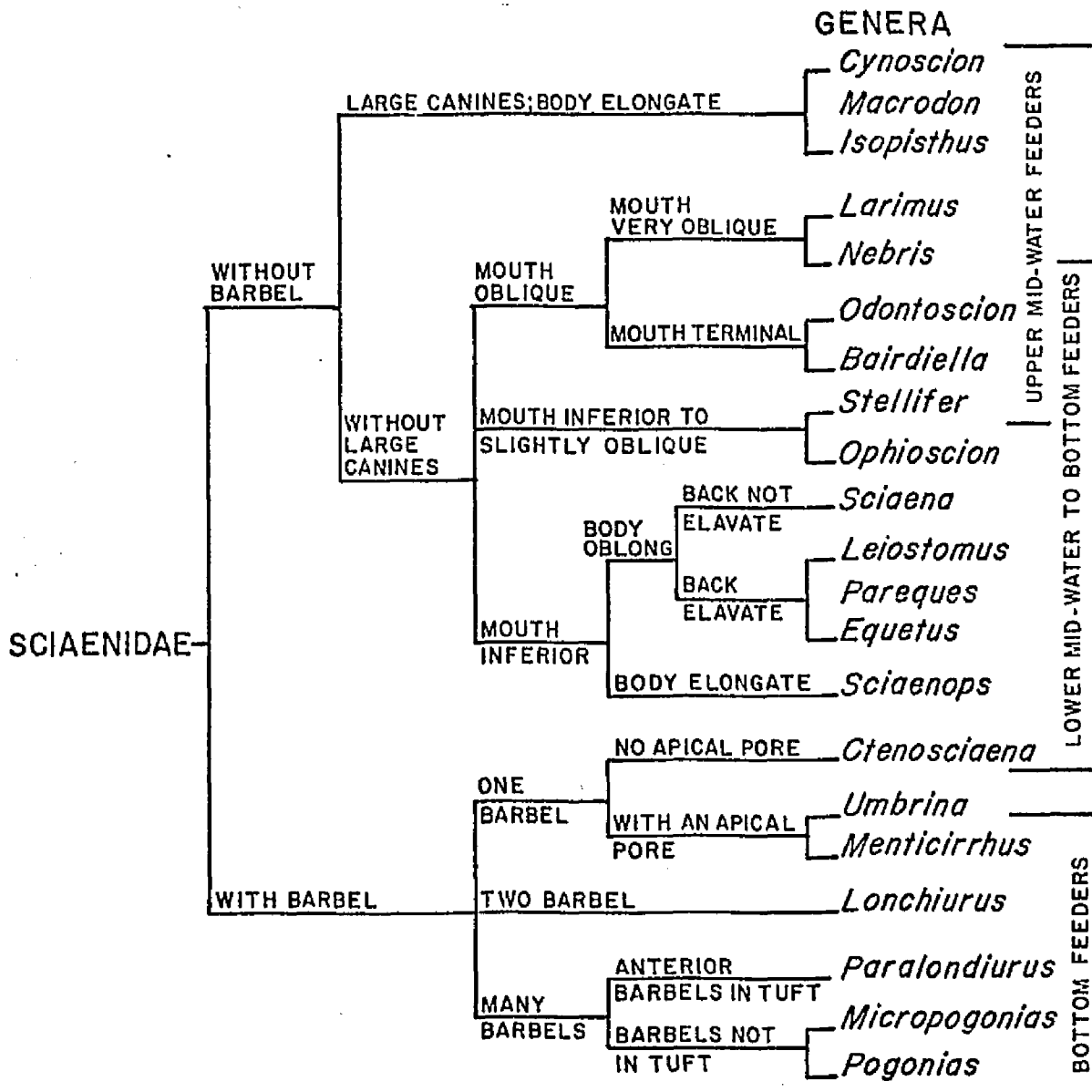


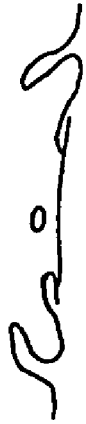
Fig. 30. Phylogenetic relationships of western Atlantic Sciaenidae as shown by external morphology.

Fig. 31

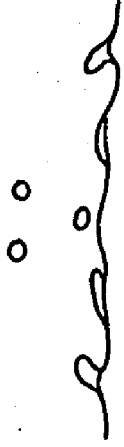
Snout pore patterns (diagramatic) of western Atlantic Sciaenidae. A. no upper and two marginal pores; B. no upper and five marginal pores; C. two upper and five marginal pores; D. three upper and five marginal pores; E. five upper and five marginal pores; F. five or more upper and marginal pores.



A



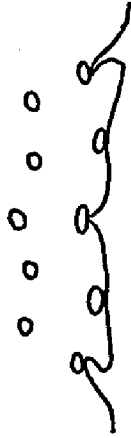
B



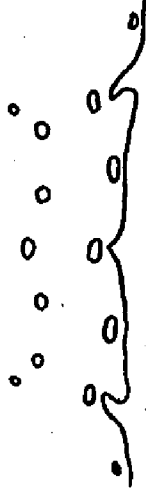
C



D



E

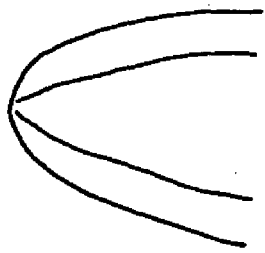


F

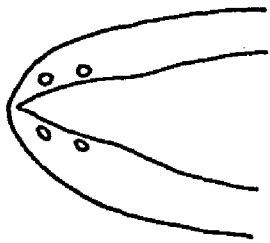
Fig. 32

Mental pore and barbel patterns (diagramatic)  
on the lower jaw of western Atlantic Sciaenidae.  
A. no pore and no barbel; B. four pores and no  
barbel; C. five pores and no barbel; D. six pores  
and no barbel; E. four pores and one barbel; F.  
four pores and two barbels; G. five pores and  
many barbels, not in tuft; H. five pores and many  
barbels, anterior three pairs of barbels in tuft.

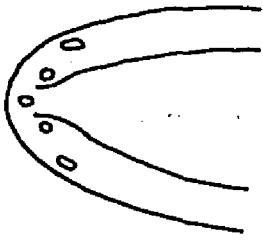




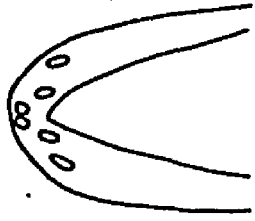
A



B

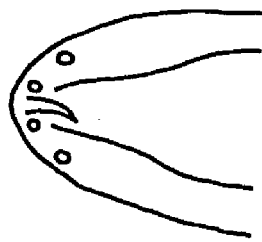


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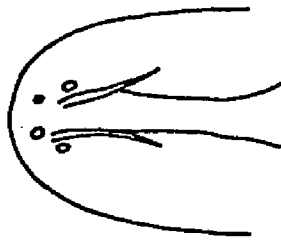


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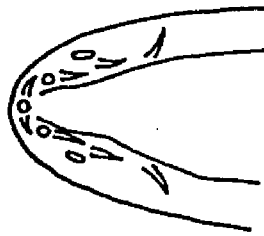
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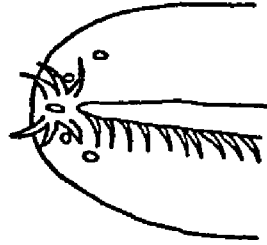
E



F



G



H

Fig. 33

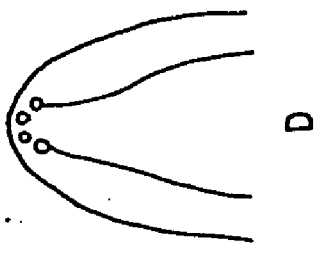
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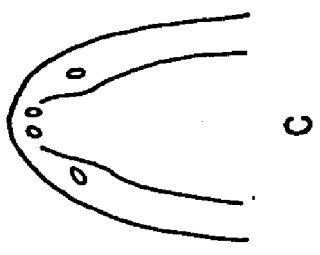
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Fig. 34

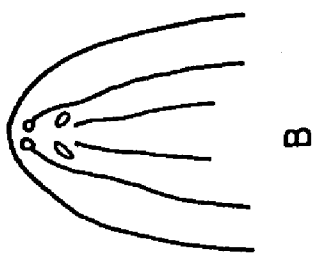
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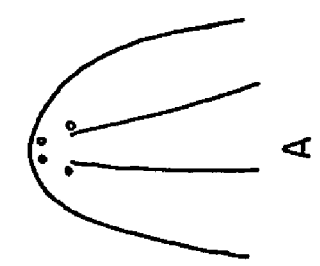
A



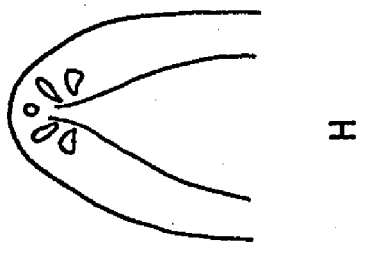
B



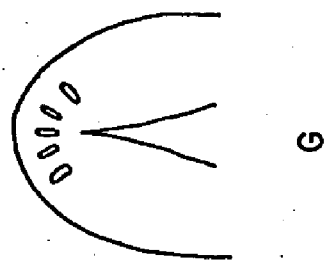
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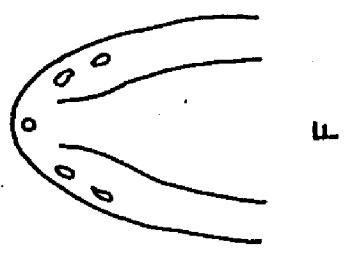
D



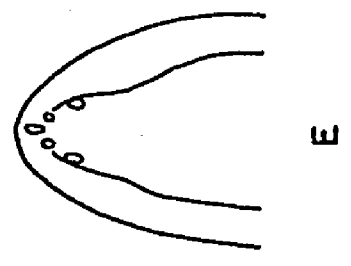
E



F



G



H

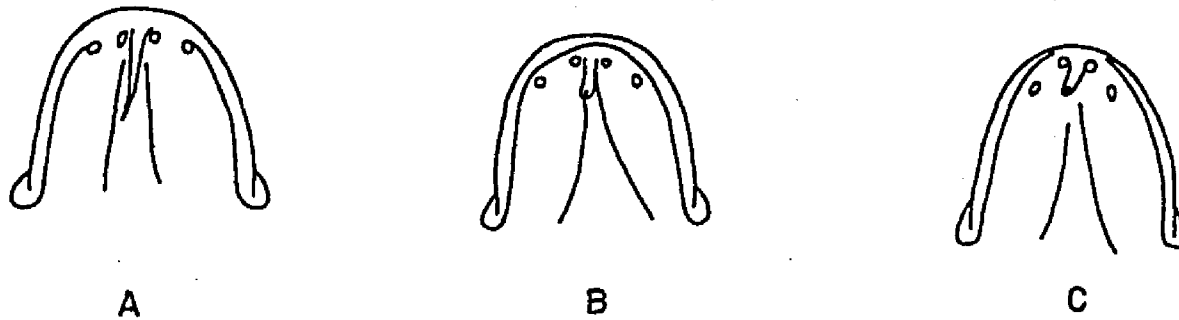


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Fig. 36

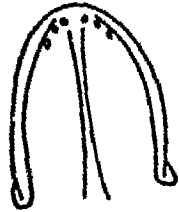
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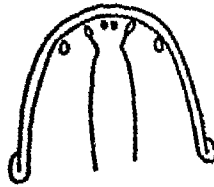
A



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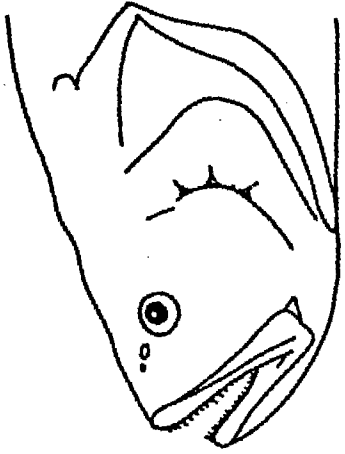


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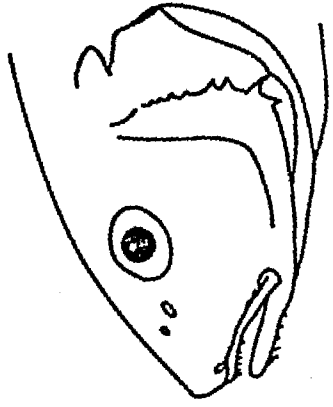


Fig. 37

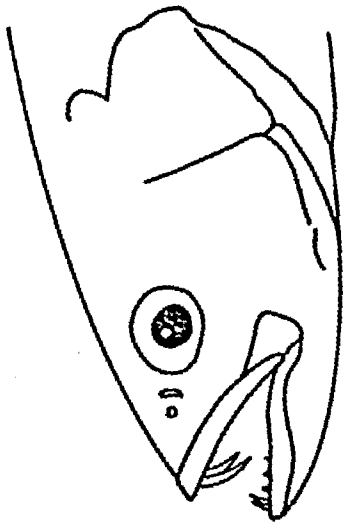
Four typical mouth positions and dentitions of western Atlantic Sciaenidae. A. *Macrodon ancylodon*; B. *Nebris microps*; C. *Odontoscion dentex*; D. *Micropogonias undulatus*.



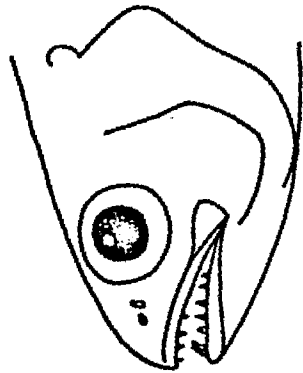
B



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C

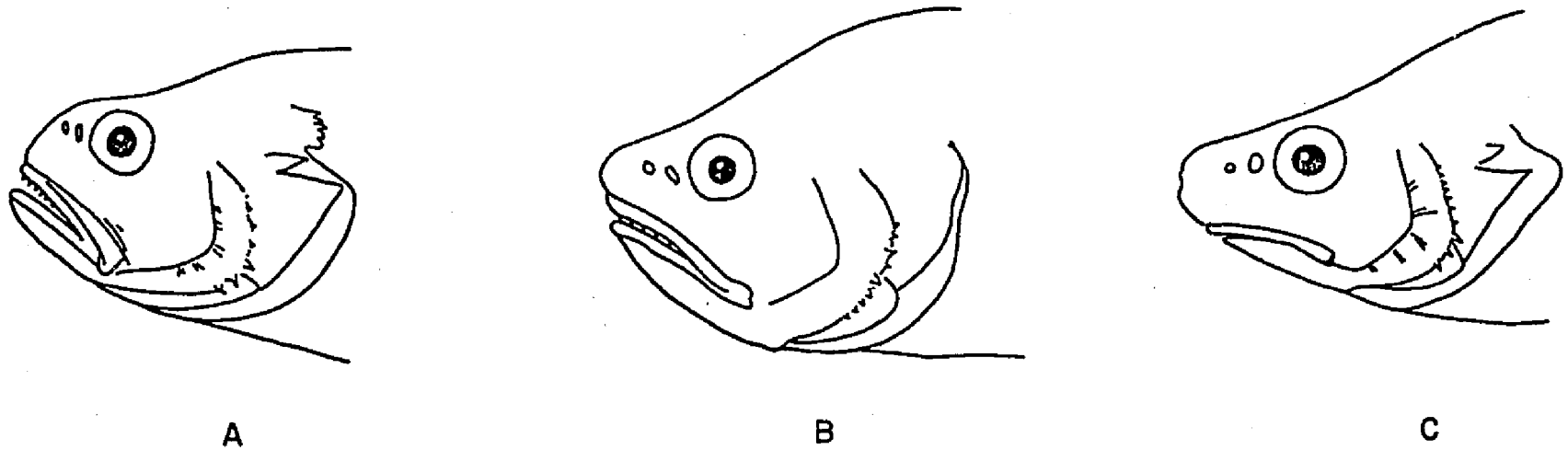


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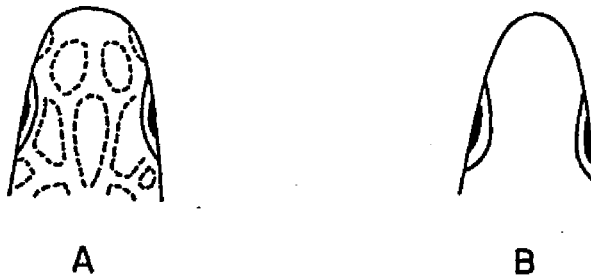


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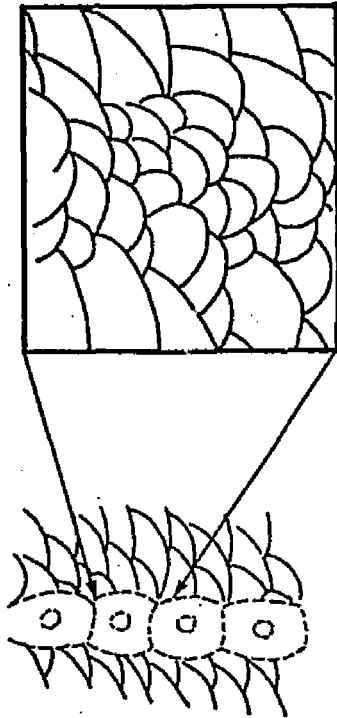


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PART II  
LIFE HISTORY, FEEDING HABITS AND FUNCTIONAL  
MORPHOLOGY OF JUVENILE SCIAENID FISHES  
IN THE YORK RIVER ESTUARY, VIRGINIA

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## INTRODUCTION

Sciaenid fishes are among the most important inshore bottom fishery resources of the Atlantic and Gulf of Mexico coasts of the United States (Roithmayr, 1965; Joseph, 1972; Gutherz, et. al., 1975). Sciaenid fishes usually use the estuary as a nursery ground, and as a seasonal feeding ground for part of their lives. Among the 14 species of sciaenids recorded from Chesapeake Bay proper (Musick, 1972), young-of-the-year of ten species of sciaenid fishes enter the York River (Fig. 1, a major Chesapeake estuary) and its tributaries in the course of a year. Among them *Leiostomus xanthurus*, *Micropogonias undulatus*, *Bairdiella chrysoura* and *Cynoscion regalis* are the most abundant species. *Menticirrhus saxatilis*, *M. americanus*, *Sciaenops ocellata*, *Cynoscion nebulosus*, *Pogonias cromis* and *Larimus fasciatus* are caught only occasionally.

Juvenile sciaenids, except croakers (*M. undulatus*), usually enter the York River in late spring and leave in late fall. During this period, young-of-the-year sciaenids dominate bottom trawl catches in the York River (Colvocoresses, 1975; Markle, 1976). The purpose of this study is to determine how these juvenile species coexist in the York River system, Virginia. Several ecological parameters are examined, including relative abundance, temporal and spatial distribution, length frequency, movement, and feeding habits. Morphological structures related to feeding habits and habitats are also examined. Specimens of the banded drum, *Larimus fasciatus* and the northern kingfish,

*Menticirrhus saxatilis*, are included in the morphological structure and feeding habit studies to show the spectra of variation in juvenile sciaenids. The distribution and food habit studies are based on York River bottom trawl surveys conducted by the Virginia Institute of Marine Science from January, 1972 to December, 1974. A community study has been partially reported by Colvocoresses, et. al. (1975). Colvocoresses is currently preparing complete results of these surveys. Therefore, description of the sampling program, study area, hydrographic data and relative abundance are greatly simplified for the purposes of the present study.

The study area includes the York River and its major tributaries, the Pamunkey and Mattaponi rivers, an estuarine system which is relatively well known biologically, and is relatively undisturbed (Boesch, 1971). The general trend of geomorphology, hydrography (Salinity, dissolved oxygen and temperature), ecology and alteration by man of the area were described by McHugh (1967), Brehmer (1970) and Boesch (1971).

## MATERIALS AND METHODS

### 1. Survey programs:

All fishes examined were collected from the York River system, Virginia. Seven longitudinal strata (A, B, C, D, E, F and G) and three cross-sectional strata (north shoal, channel and south shoal) within each stratum were sampled monthly (Fig. 1). Randomly numbered square grids (540 m = 600 yard on a side) were assigned as trawl stations. In the lower 16 km (10 miles) of the York River strata A, B, C and D were sampled from March 1972 to December 1974. The upper part of the York River was sampled from January 1972 to March 1974, but the random method was not used until June 1972 and the strata (E, F and G) were not designated until January 1973. One to 12 stations were randomly selected monthly after the random sampling methods were used. Before then, fixed channel station samples were set at 8 km (5 miles) intervals from the mouth of the York River (mile zero) up to 45 km (mile 28). Data from fixed station samples (January to May 1972) were combined within the strata for analyses. Lower portions of the Mattaponi and Pamunkey rivers have been sampled since January 1973. Three strata (1, 2 and 3) were set at 8 km (5 miles) intervals for the lower 24 km (15 miles) upstream from their confluence with the York River (about 45 km from the York River mouth). Four to six gridded stations (540 m on a side) were randomly sampled monthly from each stratum.

## 2. Gear:

Bottom trawl tows were against the current of five minutes duration with a 4.9 m (16 ft) semi-balloon otter trawl (7 m rope, 1.9 cm bar mesh, 0.63 cm bar mesh cod end liner), 7 m bridle, and 0.6 m weighted otter doors at a speed of approximately 90 m per minute. Nine stations were sampled monthly with beach seines along the shores of lower parts (strata A to D) of the York River (Fig. 1) and three replicate hauls were made with a 15.25 m (50 ft) bag seine (1.8 m deep with a square bag, 0.64 cm bar mesh in the wing and 0.48 cm bar mesh in the bag). Thirteen beach seine stations were selected along the shores of the upper part of the York River (strata E to G, Fig. 1). These stations were only sampled from July to October in 1972 and 1973 with a 30.5 m (100 ft) bag seine. Beach seine data are used only for length frequency analysis in the present study. Hydrographic (salinity, temperature and dissolved oxygen) data were collected from both surface and bottom water. Results are summarized in Figure 2 and the Appendix.

## 3. Sampling procedure:

All fishes were identified, counted and weighed in the field or laboratory. Total length, measured from snout of the fish to the middle of the caudal fin, was taken to the nearest millimeter. All individuals of each species were measured from each trawl haul, except for very large catches. Then at least 25 individuals were subsampled. Specimens for stomach analyses were randomly selected and preserved in 10% formalin, the stomachs were dissected and transferred to 40% isopropanol or 70% ethanol. Stomach contents were identified to the lowest practical taxon and the frequencies of occurrences were recorded.

The standard methods of Hubbs and Labler (1958) were used for all counts and measurements if applicable. Digestive tracts were removed from the fish and intestines were straightened with slight tension before measuring lengths from the junction of the intestine with the stomach to the anus. Osteological observations were made on cleared and stained specimens, prepared by Taylor's (1967) method.

## RESULTS AND DISCUSSIONS

### I. Hydrographic Description.

Data on the depth, temperature, salinity and dissolved oxygen samples during this study are given in Appendix. The benthic environment was of principal importance to the present study. The means of bottom temperature, salinity and dissolved oxygen of each stratum from May, 1972 to August, 1973 are summarized in Figure 2, to show seasonal patterns in the York River estuary.

#### Temperature:

The bottom water temperature of the York River (Fig. 2) was lowest in January and highest in July (1973) or August (1972). The gradual increase of temperature in April to June and the decrease in October to December are most important to the migratory fishes (Markle, 1976). In winter months (December to February), the bottom temperature of the upper portion of the York River (Fig. 2) was lower than that of the lower portion. No apparent differences in temperature were found among the shoal and the channel stations. In spring months (March to May), bottom temperatures increased rapidly, and the upper portion of the York River had slightly higher temperatures than the lower portion (Fig. 2). The shoal stations also showed a slightly higher mean bottom temperature than the channel stations. In summer months (June to August), the bottom temperature of the upper portion of the river was



higher than the lower portion (Fig. 2). The shoal stations also showed a higher mean bottom temperature than the channel stations. In fall months (September to November), the bottom temperature decreased rapidly and the upper portion of the river had slightly higher temperatures than the lower portion in the early fall (September). In late fall (November), the bottom water temperature was slightly higher in the lower portion of the river (Fig. 2). No apparent differences were found among the shoal and channel stations.

#### Salinity:

The salinity decreases toward the upper portion of the York River (Fig. 2). Lower salinities usually are found in the spring (March to May) and the winter (December to February). The extremely low salinities of June to August, 1972, were caused by the hurricane Agnes (Fig. 2). The salinity at channel stations is usually higher than at shoal stations, especially in the lower portion of the river (Fig. 2) from March to June.

#### Dissolved oxygen:

The dissolved oxygen (D.O.) in the York River (Fig. 2) is generally lower in the warmer months (May to October) and higher in the colder months (November to April). In the warmer months, the D.O. was lowest at the deeper channel stations (Fig. 2). There was no apparent difference between the upper and lower portions of the York River. In the colder months the D.O. was slightly higher in the upper portion of the river (Fig. 2) and no apparent difference was found among shoal and channel stations.

## II. Temporal and Spatial Distributions.

Young sciaenids are among the most abundant migratory finfishes in the York River (Massman, 1962; Colvocoresses, 1975; Markle, 1976). Temporal and spatial distributions of juveniles of the four most abundant sciaenids, *Cynoscion regalis*, *Bairdiella chrysoura*, *Micropogonias undulatus* and *Leiostomus xanthurus* are compared here (Figs. 3 to 5) to determine ecological partitioning during their estuarine life. The relative abundance of each species is expressed by the geometric mean,  $\log_{10} (\bar{x}+1)$ , of the individual catches per tow within the substrata. Four months (July, October, January and April) were selected to represent the seasonal abundances from different parts of the York River (Fig. 3). Monthly mean catches per tow by river distance (strata) and depths (substrata) were compared (Figs. 3 to 5). Fishes caught from Mattaponi and Pamunkey rivers are compared only by river distance (Fig. 4).

In July (1972 and 1973), all four species of juvenile sciaenids were present in all parts of the estuary except the upper part (Figs. 3 to 5). The relative abundance (mean number of individuals per tow) varied among species (Fig. 3). *B. chrysoura* were more abundant in the lower and middle part of the river, while *C. regalis* and *M. undulatus* were more abundant in the upper part of the river. *L. xanthurus* were abundant in most parts of the river. *M. undulatus* showed a trend of gradual decline in abundance upstream in both the Mattaponi and Pamunkey rivers (Figs. 4 and 5). *L. xanthurus* catches were quite variable in the Pamunkey River. This may be caused by the contagious distribution of this species. Comparing depth distribution, sciaenids

were more abundant in shoal stations (Fig. 3) than channel stations, especially in July 1972. Colvocoresses (1975) and Markle (1976) noted a general decline in the mean number of species and individuals of fishes per month in the summer from channel stations. This may be attributed to a reduction in the dissolved oxygen concentration at the bottom of the channel (Brehmer, 1970 and Markle, 1976) and was apparently the case in the present study (Fig. 2). Catches of *C. regalis* did not decline in channel stations, but this species is the best adapted for pelagic life of the four species studied (see sections on "Correlation of feeding structures and food habits").

In October (1972 and 1973) juveniles of all four species of sciaenids were present in all parts of the estuary (Fig. 3) and all reached their highest total abundance (Markle, 1976). *C. regalis* were more abundant in the lower parts of the York River. *B. chrysoura* and *L. xanthurus* were more abundant in the middle part of the river. *M. undulatus* were more abundant in the upper part of the river, and especially in the Mattaponi and Pamunkey rivers (Figs. 4 and 5). Mean catch per tow increased up estuary. Depth distribution of these four species of sciaenids indicated that they were more abundant in the channel stations (Fig. 3). The relative abundance at south shoal stations was higher than at north shoal stations. The area was larger and the sampling depth was greater in the south shoal than the north shoal area (see Appendix). Also, the average size of young sciaenids, especially the young-of-the-year groups, were larger in the channel than in the shoal stations (see section on "Length frequencies and distributions"). Larger sized juvenile sciaenids might use deeper areas to seek food and shelter.

In January (1972 to 1974), the numbers of individual sciaenid fishes were considerably reduced except for *M. undulatus* (Figs. 3 and 5). *C. regalis*, *B. chrysoura* and *L. xanthurus* were caught only sporadically. During the winter months, the resident fish species were more abundant than transients, especially in the upper tributaries of the York River (Markle, 1976). *M. undulatus* was the most abundant sciaenid fish in the middle part of the York River (Fig. 5). Depth distribution in January, 1973 (Fig. 3), indicated that most fish were caught in the channel. Bottom temperatures of the shoal stations were lower than channel stations (Fig. 2), which might be the major factor causing the concentration of young sciaenids in the channel.

In April (1972 to 1974), *C. regalis*, *M. undulatus* and *L. xanthurus* were caught (Figs. 3 and 5). *C. regalis* was caught sporadically in 1972 and 1974. *M. undulatus* were more abundant in the upper part of the river and *L. xanthurus* were more abundant in the lower reaches (Figs. 4 and 5), apparently because the young-of-the-year *L. xanthurus* had just entered the estuary (see section on "Length frequencies and distributions"). Depth distribution of these two species (Fig. 3) showed that they were more abundant in shoal areas, especially *M. undulatus*. *B. chrysoura* was completely absent.

The relative abundance of all major dominant fishes (including *C. regalis*, *M. undulatus* and *L. xanthurus*) are highly variable in the York River from year to year (Colvocoresses 1975 and Markle 1976). The comparisons here are not suitable for assessing the year class strengths of these sciaenid fishes.

### III. Length Frequencies and Distributions.

Length frequency distributions (Figs. 6-19) indicate that juvenile *Leiostomus xanthurus*, *Bairdiella chrysoura*, *Cynoscion regalis* and *Micropogonias undulatus* enter the York River consecutively from April on, and all but *M. undulatus* leave the York River by December. Seasonal size distributions of these four species in the York River will be discussed individually and compared with studies from other areas.

#### *Leiostomus xanthurus* Lacépède

##### 1. Early life history in the York River:

Young-of-the-year spot, *L. xanthurus*, entered the trawl and beach seine catches in early April and most left by December (Fig. 6, mode I). A few smaller fish stayed in the estuary over winter. Yearling spot usually entered the study area from March to May and left the area in September (Fig. 6, mode II). The intermediate mode (between mode I and II) on Figure 6, April and May 1972 was not found in 1973 and 1974 samples. This may indicate a late spawning in the previous year (1971). The length frequencies of young spot from May to July during 1972 to 1974 were pooled and grouped by river strata (Fig. 7). Young-of-the-year spot move up to the confluence of the Pamunkey and Mattaponi rivers (Fig. 1). Most yearling spot stayed in the lower parts of the York River. During the same periods, no differences were found between the length frequency distributions in both shoal and channel stations (Fig. 8) of either young-of-the-year or yearling spot.

Spot caught by beach seine (Fig. 8) were obviously smaller than those taken by trawls. Spot was the most abundant sciaenid in the beach seine zone (depth less than 1.5 m) for collections with the 15.25 m and 30.5 m seine. The length frequency distribution of spot caught by beach seine was typically unimodal; mostly young-of-the-year (Fig. 9). Some smaller yearlings were taken occasionally (Fig. 9, 1974, mode II) and individuals larger than 135 mm TL were captured only with the 30.5 m seine (Fig. 9, August and September, 1972).

In summary, young-of-the-year spot entered the York River in April and used the estuary as a nursery ground. In December, most spot left though some smaller fish stayed in the estuary through the winter, joining the yearlings as they returned to the river in the next spring. The yearlings left the estuary after an extended feeding period from March to October.

## 2. Other studies:

Selected length frequency data for spot along the Atlantic and Gulf of Mexico coasts of the U.S. are summarized (Table 1) for comparison with the present study. Hildebrand and Schroeder (1928) and Pacheco (1957 and 1962a) reported length frequency of spot from the present study area (York River and Chesapeake Bay). Across all areas (Table 1), young-of-the-year spot (Group 0 on Table 1) enter the estuarine nursery grounds during the first half of the year. Spot may enter estuaries as early as January (Table 1; Hildebrand & Cable, 1930; Springer and Woodburn 1960 and Sundararaj, 1960). Spot first enter the estuary in February on the Atlantic coast (Georgia, Music, 1974) and

Gulf of Mexico coast (Florida, Townsend, 1956; Louisiana, Dunham, 1972; Texas, Parker, 1971). In South Carolina (Dawson 1958), North Carolina (Shealy, et. al., 1974) and the lower Chesapeake Bay (Hildebrand and Schroeder, 1928 and present study) young-of-the-year spot first entered the estuary in April (Table 1). In upper Chesapeake Bay (Young, 1953) and Delaware River (Thomas, 1971), young-of-the-year spot probably do not appear until May (Table 1). The smallest young-of-the-year spot from trawl catches are about 15 to 20 mm TL in all areas. Which indicates that the young-of-the-year spot in northern areas enter the estuary later than southern ones. When spot first enter estuaries, gear selectivity (Table 1) affects the size ranges of spot captured; beach seines usually catch only the small specimens (Young, 1954 and Fig. 9), but pound nets (Pacheco, 1957) and large otter trawls (Music, 1974) usually catch larger fishes. Offshore movements of spot during the winter season are evident in all areas studied, because spot completely absent or in low abundance in inshore catches (Table 1). Yearling or older spot (Group I of Table 1) usually leave the estuary after September and do not return until spring of the next year (Table 1). Some young-of-the-year spot over-winter in the estuary (Fig. 6 and Table 1). Tagged spot (Pacheco, 1962b) have moved from the Chesapeake Bay to an area west of Diamond Shoals, North Carolina. Similarly, a spot tagged and released from Delaware Bay in October 1930 was recovered from Ocracoke Inlet, North Carolina in December, 1930 (Pearson, 1932). Thus, spot from these areas may have a common coastal feeding or spawning ground during the winter, although Struhsaker (1969) reported a winter offshore movement of spot into deeper water (lower-shelf habitat off South Carolina). These offshore

spots may be a mixture of northern and southern populations or just southern residents. The late fall or early winter spawning time of spot appears to be the same in both Atlantic and Gulf waters (Welsh and Breder, 1923). Later spawning by a northern component is evidence in the length ranges of postlarvae and juvenile spot (Table 1). Young-of-the-year spot begin to enter the estuarine nursery grounds of Delaware, Maryland, Virginia, North and South Carolina during April (Table 1).

*Cynoscion regalis* (Bloch and Schneider)

1. Early life history in the York River:

Weakfish, *C. regalis*, showed distinct seasonal fluctuations. Young-of-the-year weakfish first entered trawl catches in July or August and virtually all left the estuary in the winter (Fig. 10, mode I). Yearling weakfish returned to the river in April or May and left in September or October (Fig. 10, mode II). Larger weakfish (two years or older) were caught only sporadically during this study because of gear avoidance. The length mode of small weakfish in August showed a rapid increase (Fig. 10). This increase may be due to the recruitment of yearlings or an earlier spawned group of young-of-the-year. Length frequencies for weakfish (under 250 mm TL) caught from August to October, 1972-1974, were pooled to compare distribution by size in the York River and its tributaries (Fig. 11). Smaller fishes were more abundant in the Pamunkey and Mattaponi rivers, than in the York River proper. Yearling weakfish also showed a movement up river (Fig. 11). This suggests that young weakfish entered the low salinity nursery



ground (upper portion of the York River) and then moved down river as they grew. Pooled length frequency distributions revealed an apparent difference between shoal and channel areas of the York River (Fig. 12). Yearling weakfish (or larger ones) are proportionally more abundant in the channel. The 15.25 m beach seine catches contained no weakfish, but occasionally the 30.5 m seine caught some young-of-the-year weakfish in the summer.

## 2. Other studies:

Major populations of weakfish are confined to the Atlantic coast of the U.S. from the Gulf of Maine to Florida. Existing data indicate young-of-the-year weakfish enter estuarine or coastal catches from May to July (Table 2). The smallest sizes of the weakfish in the catches differ with area and may be due to gear and/or time of sampling. Small fishes with less size variation (about 5 mm) were taken over a longer period of time in southern areas than northern areas (Table 2). Young-of-the-year weakfish do not occur in catches during winter months in northern coastal areas or estuaries (Perlmutter, 1956; Massman et. al., 1958; Thomas, 1971; Markle, 1976). Year round catches of weakfishes from Beaufort, North Carolina (Hildebrand & Cable, 1934) and Georgia (Mahood, 1974) were from sounds and short coastal rivers. Most of the studies suggest the age Group 0 on Table 2 were a combination of young-of-the-year and yearlings. No distinct mode could be identified for young-of-the-year from these studies. This may be due to the multiple spawning of the females (Merriner, 1973) and/or the recruitment of the young-of-the-year from different spawning populations.

The reproductive biology weakfish is better known than other sciaenid fishes studied here. Welsh and Breder (1923) described the eggs and development of weakfish and they also indicated the Delaware Bay as a spawning ground of weakfish. Merriner (1973) indicated that weakfish have an extended spawning season in North Carolina (March to August) and are characterized by high fecundity and possible multiple spawning by some females. Pearson (1941) took plankton tows of lower Chesapeake Bay from May to August in 1929 and 1930 and reported greater densities of weakfish (1.5 to 17 mm TL) in subsurface tows (average 67 per tow) than in surface tows (average 13 per tow). The density of planktonic weakfish decreased at those stations within Chesapeake Bay, compared with sites near the bay mouth. Harmic (1958) reported that newly hatched larval weakfishes averaged 1.8 mm TL. Soon after hatching, the larvae became demersal and were dispersed into the nursery areas of Delaware Bay by means of the "salt wedge". The smallest weakfishes taken in the bottom trawl were 6-10 mm TL (Hildebrand and Cable, 1934). The young-of-the-year weakfish in York River are probably progeny from adults spawning near the mouth of Chesapeake Bay. Weakfish tagged and released in lower Chesapeake Bay (Nesbitt, 1954) were later recovered to the north in New York and New Jersey, and southward in North Carolina. Nesbitt (1954), Perlmutter, et. al. (1956) and Harmic (1958) cited the presence of a northern spawning population in New York and northern New Jersey waters and a southern spawning population from New Jersey to North Carolina. Seguin (1960) found that morphometric and meristic variation of weakfish exists along the middle Atlantic coast and suggested that three possible population segments may exist; a New

York group, a Delaware and lower Chesapeake group and a North Carolina group. Joseph (1972) questioned the division of weakfish into northern and southern stocks and did not consider the decline of weakfish in Chesapeake Bay to be a result of the trawl fisheries in the shallow coastal waters and bays of North Carolina. Joseph (1972) also cited Chesapeake Bay as a major spawning area and nursery ground, but also cited failure to obtain one weakfish larva per tow in extensive VIMS ichthyoplankton studies during 1959-1963. As a result of the present study, I am not convinced that Chesapeake Bay is a major spawning area of weakfish even though weakfish eggs and larvae were reported by Hildebrand and Schroeder (1928), and Pearson (1941). I agree with Massmann's (1963) implication, that Chesapeake Bay weakfish are from southern spawning populations or stocks. No ripe female weakfish were caught from the lower Chesapeake Bay in present study. Pearson (1932) described the winter trawl fishery off North Carolina and cited higher total catches of weakfish from area B (southwest of Cape Hatteras) than from area A (northeast of Cape Hatteras) in deeper waters. I assume that most young-of-the-year and larger weakfish left the York River and moved southward to their wintering ground, off Cape Hatteras. In spring, weakfishes disperse from the wintering ground; some fish move north and spawning may occur from late spring to summer along the coast from North Carolina to New York.

*Bairdiella chrysoura* (Lacépède)

1. Early life history in the York River:

Silver perch, *B. chrysoura*, were present from April to December

and were most abundant from August to October (Fig. 13). Total catches were reduced in 1973 and 1974. Young-of-the-year silver perch first entered the catches in July and most silver perch left the river in November. Yearlings may enter the river as early as April (Fig. 13, 1974) and most leave the river in October. There were no silver perch taken from January to March during the present study (1972-1974).

Pooled length frequencies from August to October, 1972 to 1974, indicated that silver perch were most concentrated in the lower part of the York River (Fig. 14) and larger specimens tended to stay in the channel (Fig. 15). The 30.5 m beach seine caught young-of-the-year occasionally but the 15.25 m seine rarely caught any silver perch.

## 2. Other studies:

Silver perch occur along the U.S. coast from New York to Texas. The seasonal distribution pattern is similar in all Atlantic coastal states (Table 3). Young-of-the-year silver perch were first caught in bottom trawls during June or July. Size of the smallest young-of-the-year silver perch during a given month decreases as latitude of the nursery ground increases on the Atlantic coast and west coast of Florida (Table 3). Silver perch are present almost all year round south of Chesapeake Bay (Table 3), which may be due to the higher salinity or temperature of these study areas. The embryonic development of silver perch from Beaufort, North Carolina was described by Kuntz (1913). Welsh and Breder (1923) made further observations from material obtained at Atlantic City, New Jersey. Jannke (1971) described larval silver perch from the Everglades National Park, Florida and showed that the larvae of 2 to 3 mm "notochord" length were present all year

round. Hildebrand and Schroeder (1928) reported ripe fish of both sexes in Chesapeake Bay (24 m deep, off Crisfield, Maryland). This indicated that silver perch spawn in the deeper waters of lower Chesapeake Bay in late spring and early summer but, because of its relatively small size, commercial catches of silver perch were relatively small. Silver perch might move oceanward or to the deeper part of the Chesapeake Bay in winter (Hildebrand and Schroeder, 1928).

*Micropogonias undulatus* (Linnaeus)

1. Early life history in the York River:

Young-of-the-year croaker, *M. undulatus*, first entered the trawl and beach seine catches in August and stayed in the York River throughout the winter (Fig. 16). They left the estuary between August and September of the following year as yearlings (Fig. 16, mode III). Large croaker (more than 1.5 years old) were caught only sporadically in this study due to gear avoidance, but they were present from February to September. There were apparently two to three length groups (modes) of young-of-the-year croaker in September (1972 to 1974). The mode II was different from mode I and mode III of 1972 and 1974 (Fig. 16). The former group did not stay in the York River over winter, they entered the estuary as early as May (Fig. 16, mode II) and most left in November.

Size may be a determining factor for migration of young croaker from the York River. From 1972 to 1974, the length frequencies (Fig. 16) indicate that very few young-of-the-year croakers larger than 130 mm TL stayed in the York River during the winter months. Young-of-the-year croakers were present in the York River in large numbers all

year round except during the summer months (June to August). Young croakers showed slower growth rate over winter (Fig. 16). Those entering the estuary between September and November were the main strength of the year class (mode I of Fig. 16). Whether they represent progeny from a different spawning population compared to the earlier group (mode II of Fig. 16) is unknown at present.

Length frequencies of croakers taken between September and November, 1972-1974, were pooled to compare distribution by size in the York River (Fig. 17). The size composition indicated that smaller fish were caught in the upper part of the York River and saline portions of the Mattaponi and Pamunkey rivers. Larger fish were proportionally more abundant in the lower part of the river. Larger fish also constituted a larger portion of the croaker catch in the channel than in the shoal area (Fig. 18). Beach seines (Fig. 19) caught exclusively yearlings in the 30.5 m beach seine. The 15.25 m seine caught almost no croakers.

In summary, young-of-the-year croaker enter the estuary in May and from August on. The earlier group entered in May and left the estuary in November, as do older year classes. The later group (August-November) stay in the estuary until the summer months of the following year. Young croaker moved to the upper part of the York River and the saline portions of major tributaries after first entry, then move down the York River into more saline waters as they grew. Smaller fishes (less than 130 mm TL) stayed in the river throughout the winter.

## 2. Other studies:

Croaker occur from the Gulf of Maine to Argentina, along the coasts of the Atlantic and Gulf of Mexico. Length frequency

distributions exist for different areas of the U.S., including Wallace (1940) and Haven (1957) for the lower Chesapeake Bay and York River (Table 4). Studies usually show that small croakers (10-20 mm TL) were present in the estuary during all except the summer months (June to August). Croakers seemingly have a long spawning season since small individuals (<20 mm TL) were present from October to May in different estuarine areas (Table 4). Some croaker were very small (< 20 mm TL) in spring because of slow growth of fish spawned late in winter, or because they were spawned in spring. Such groups found in this study (mode II, Fig. 16) were not found in previous Chesapeake Bay studies. Tagging of adult croakers from Maryland and Virginia by Haven (1959) showed spring time movement of croaker up the estuaries and up the Chesapeake Bay, as well as an oceanward and southerly migration in fall (some recoveries were from off the North Carolina coast). Pearson (1932) reported a high percentage of croaker in the catches of the commercial trawl fishery during November (88%) and December (76%) from the fishing grounds off the North Carolina coast. Croaker may migrate out from the estuary to a southern coastal or offshore wintering and spawning ground. Hildebrand and Cable (1930) believed croaker spawning probably began in August in Chesapeake Bay and northward, in September at Beaufort (North Carolina), and in October in Texas. Croaker from all areas may have the same spawning period, probably from July to January (Table 4). A later group apparently spawning from March to May (mode II, Fig. 16) was found in the present study. White and Chittenden (1976) indicated that some croaker in the Gulf of Mexico may lack the first (overwinter) ring on the scales. This suggests that some croaker may spawn in the spring in the Gulf of Mexico.

Massmann and Pacheco (1960) reported the disappearance of young croakers from the York River but their conclusion may have been in error because of selectivity of their fishing gear. Haven's (1957) length frequencies for croaker during 1952 and 1953 differs from those presented by Massmann and Pacheco (1960) for the same years. No fish less than 100 mm TL were reported by Massmann and Pacheco (1960) and their gear was a net with three quarter inch mesh, whereas Haven (1957) used quarter inch mesh. Joseph (1972) attributed the decline of croaker in the commercial catches of the middle Atlantic coast to climatic trends. The apparent increase of juvenile croakers in 1973 and 1974 was probably due to the warmer winter months in those years. The main year class strength of croaker in the York River was dependent on the success of the late young-of-the-year group (Fig. 16, mode I). Because they stayed in the estuary through the winter. Milder winters probably were the most important factor determining the success of a given year class. VIMS York River trawl data showed mass mortalities of young-of-the-year croaker during cold winters (VIMS, Ichthyology Department, unpublished).



#### IV. Feeding Mechanisms.

Sciaenidae have the widest spectrum of feeding niches of any fish family in the Chesapeake Bay. The four most abundant species of sciaenids are *Cynoscion regalis*, *Bairdiella chrysoura*, *Micropogonias undulatus* and *Leiostomus xanthurus*. These species are most abundant in the estuary from late spring to fall, especially young-of-the-year and yearlings (Figs. 6 to 19). Under these conditions possible food resources may be limiting and division of feeding niches may have evolved in order to reduce competitive exclusion among the dominant species. Fishes that are closely related and show feeding niche segregation also often show morphological differentiation in the feeding apparatus in *Larimus fasciatus*, *Cynoscion regalis*, *Bairdiella chrysoura*, *Micropogonias undulatus*, *Menticirrhus saxatilis* and *Leiostomus xanthurus* to test the hypothesis that feeding niche division has evolved among those six species.

The feeding structures examined were mouth position and size, dentition, number of gill rakers and intestine length, which directly effect the size and kind of food ingested and digested. Other accessory structures examined, such as the pore and barbel system on the snout and/or lower jaw, the nares and body shape were more important in locating food. Morphological differentiation and adaptation of these structures was found to be correlated with feeding partitioning of these species.

##### Mouth Position:

Mouth position and size of the opening limit the size of prey and

habitats in which the predator can effectively capture the prey. *Larimus fasciatus* have the most oblique mouth (Fig. 20,A) with the lower jaw projecting strongly in front of the nonprotrusible upper jaw. The maxilla (Fig. 21,A) is under the lateral margin of the rostral fold and its anterior end is firmly attached to the premaxilla and skull (dermethmoid). As the mouth opens, the distal ends of the premaxilla and maxillae push forward as the lower jaw is lowered (Fig. 20,A'). *Cynoscion regalis* have a large oblique mouth with the tip of the lower jaw projecting in front of the nonprotrusible upper jaw (Fig. 20,B). The anterior end of the maxilla is firmly attached to the premaxilla and articulates with the dermethmoid (Fig. 21,B). As the mouth opened, the posterior end of the premaxilla and the lower jaw move forward (Figs. 20,B'). *Bairdiella chrysoura* has a similar mechanism of jaw movement (Fig. 20,C'), but the mouth is only slightly oblique with the lower jaw about equal in length to the upper jaw (Figs. 20,C and 21,C). *Micropogonias undulatus* has an inferior mouth with the tip of the lower jaw enclosed by the protrusible upper jaw (Fig. 20,D). The anterior end of the maxilla is loosely attached to the premaxilla (Fig. 21,D). As the mouth is opened, the entire premaxilla and the lower jaw move antero-ventrally (Fig. 20,D'). *Menticirrhus saxatilis* and *Leiostomus xanthurus* have a similar mechanism of jaw movement but their upper jaws seem more protrusible (Figs. 20,E' & F' and 21,E & F). In *M. saxatilis*, the mouth is inferior and the lower jaw is enclosed by the upper jaw (Fig. 20,E). *L. xanthurus* also has a small inferior mouth (Fig. 20,F).

The mouth position indicates that *L. fasciatus*, *C. regalis* and *B. chrysoura* are the pelagic feeders (Fig. 20,A, B & C) and that *M. undulatus*, *M. saxatilis* and *L. xanthurus* feed on the bottom (Fig. 20,D, E & F). The relative length of the premaxillary and dentary bones decreases and the height of the anterior dorsal process of the premaxilla increased from fishes adapted to feed in mid-water to those adapted to feed on the bottom (Fig. 2). This trend is also evident in the relative mouth size (Fig. 21,a, b, c, d, e & f, Table 5). The index number, the length of the upper jaw multiplied by the length of the lower jaw then divided by head length, decreases as the species trends toward a bottom feeding habit.

In bottom feeders, *Micropogonias undulatus*, *Leiostomus xanthurus* and *Menticirrhus saxatilis*, have protrusible premaxillae (Figs. 20,D', E' & F' and 21,D, E & F). This can be advantageous in getting the mouth opening close to food that is to be sucked in from the bottom (Alexander, 1967). The mid-water feeders, *Larimus fasciatus*, *Cynoscion regalis* and *Bairdiella chrysoura*, have lost the protrusibility of premaxillae (Figs. 20,A', B' & C' and 21,A, B & C). *C. regalis* and *B. chrysoura* may compensate for this with faster swimming speed. Gero (1952) and Nyberg (1971) have discussed this aspect in detail. *Larimus fasciatus* differs from other sciaenids studied here. It may swim around with its mouth open and using its gill rakers as a filter similar to that of *Engraulis* (Gunther, 1962).

#### Dentition:

Teeth on the premaxilla and dentary are important in capturing prey whereas the pharyngeal teeth are used for grinding and/or transporting

food to the esophagus. In sciaenids, members of the genus *Cynoscion* usually have a pair of enlarged canine teeth at the tip of the upper jaw (Fig. 21,B). Other teeth are conical and present on narrow ridges of the premaxilla and dentary. The tips of the upper and lower jaws are broad and have several rows of teeth which decrease in number to a single prominent row in the narrower posterior portion of the jaws. Small teeth also develop inside the larger functional row of upper jaw teeth and outside the lower jaw teeth. *L. fasciatus* and *B. chrysoura* have a band of teeth similar to *C. regalis* but lack large canine teeth at the tip of the upper jaw (Fig. 21,A & C). *L. fasciatus* teeth are finer than those of *B. chrysoura*. *M. undulatus*, *L. xanthurus* and *M. saxatilis* have broad villiform bands of teeth on the premaxillae and dentaries and also lack canine teeth.

Pharyngeal teeth are generally conical in sciaenids (Fig. 22). The lower pharyngeal teeth form a pair of separate narrow tooth patches and are situated on the most medial pairs of ceratobranchial bones. The upper pharyngeal teeth are mainly formed by two pairs of patches on the two most medial pairs of epibranchial bones. The pharyngeal plates are relatively small and narrow in *Larimus fasciatus* and *Cynoscion regalis* compared to the other sciaenids examined (Fig. 22,A & B). The pharyngeal teeth of *L. fasciatus* and *C. regalis* are sharp, conical and directed backward but in *Bairdiella chrysoura* the pharyngeal teeth are blunt and the median ones are enlarged (Fig. 22,C). *Micropogonias undulatus* has much stronger and more enlarged pharyngeal teeth along the median rows (Fig. 22,D). *Leiostomus xanthurus* develops molariform teeth medially on the pharyngeal plates (Fig. 22,F). *Menticirrhus saxatilis* has fine and sharp pharyngeal teeth (Fig. 22,E).

The sequential morphological changes in pharyngeal teeth reflect the feeding niches differentiation from mid-water to benthic.

Gill rakers:

Gill rakers on the branchial arches of fishes are important in protecting the delicate gill filaments from abrasion by ingested materials and may also be adapted to particular food and feeding habits (Nikolsky, 1963). In sciaenids, the gill rakers reflect feeding niches by their numbers, size and shape. They are found on the dorsolateral surface of the branchial arch (Fig. 22) and along the inner surface of the branchial arch. In sciaenids, the lateral gill rakers are well-developed only on the first gill arch and the inner (or medial) gill rakers occur only as tubercles on all five gill arches. Only the rakers on the first gill arch are discussed here.

*Menticirrhus saxatilis* and *Cynoscion regalis* have the fewest gill rakers (Table 6). *B. chrysoura* and *Micropogonias undulatus* have an intermediate number and *Leiostomus xanthurus* and *Larimus fasciatus* have the most gill rakers. Numbers of inner gill rakers (Table 6) follow a similar sequence. The relative size of the gill rakers and their morphology differ among species (Fig. 23). *L. fasciatus* has the longest and the most closely spaced gill rakers (Fig. 23,A). Each raker has many minute spicules scattered on it (Fig. 23,a). *C. regalis* and *B. chrysoura* have long gill rakers compared with the length of the gill filaments (Fig. 23,B & C). Numerous minute spicules are also present on each raker, especially on the basal portion of the raker (Fig. 23,b & c). *M. undulatus* has relative shorter gill rakers (Fig. 23,D) with seemingly strong serrations limited to the basal half

of the raker (Fig. 23,d). The relative lengths of the lateral gill rakers in *M. saxatilis* and *L. xanthurus* are the shortest (Fig. 23, E & F) and lack strong spicules (Fig. 23,e & f). *L. xanthurus* has only slightly denticulated gill rakers and *M. saxatilis* has smooth gill rakers.

The inner gill rakers are knob-like, sometimes with spicules or teeth on their distal ends (Fig. 23,a', b', c', d', e' & f'). *C. regalis*, *M. undulatus* and *M. saxatilis* have broad, short inner gill rakers, with the height not longer than the width of the base. *C. regalis* and *M. undulatus* have prominent spicules at the distal ends of their inner gill rakers (Fig. 23,b' & d'). *M. saxatilis* lacks spicules on its inner gill rakers (Fig. 23,e'). *L. fasciatus*, *B. chrysourea*, and *L. xanthurus* have long inner gill rakers, with the height longer than the width of the base. *L. fasciatus* and *B. chrysourea* have prominent spicules at the distal ends of their inner gill rakers (Fig. 23,b' & c'). *L. xanthurus* has minute spicules on its inner gill rakers (Fig. 23,f'). Furthermore, in *L. fasciatus* a small inner gill raker is often present inbetween the larger inner gill rakers (Fig. 23,a'). This is rather common among western Atlantic sciaenids.

The lateral and inner gill rakers on the second to fifth gill arches are similar in size and structure to the inner gill rakers on the first gill arch. The gill arches of these six species also differ in the relative lengths of the epibranchial (upper) arm and ceratobranchial (lower) arm (Fig. 23). *L. xanthurus* has the shortest upper arm and *M. saxatilis* has the shortest lower arm. The numbers and size of the gill rakers indicate that mid-water feeders have lateral rakers longer than those of bottom feeders. The relative lengths of

inner rakers are longer in fishes with higher numbers of lateral rakers, e.g. *L. fasciatus* and *L. xanthurus* (Fig. 23,a' & f' and Table 6). Although *M. undulatus* has the strongest spicules on the lateral gill rakers (Fig. 23,d), the mid-water feeders usually have better developed spicules on the lateral rakers than the bottom feeders (Fig. 23). The morphological differentiation of gill rakers seems to relate to the filter feeding habits in the water column. Higher numbers of rakers (both inner and lateral) suggest that the filtering mechanism is more important in these fishes.

#### Digestive tract:

The digestive tract of sciaenids includes four parts: esophagus, stomach, pyloric caeca and intestine. The intestine usually has two loops (Fig. 24) except that of *Cynoscion regalis*, which is a straight tube from stomach to anus (Fig. 24,B). The relative position and size of the stomach and intestine vary with the amount of food present. The numbers of pyloric caeca and the relative length of the intestine may be correlated with the feeding habits (Suyehiro, 1942). The relative length of the intestine of these six species of sciaenid fishes (Table 7), may be grouped into three general categories. *C. regalis* has the shortest intestine. *Bairdiella chrysoura* has an intermediate intestine length. *Micropogonias undulatus*, *Leiostomus xanthurus*, *Larimus fasciatus*, and *Menticirrhus saxatilis* have a long intestine. The numbers of pyloric caeca (Table 8) in these six sciaenid fishes also show a similar trend. *C. regalis* has the least number of pyloric caeca, four to five. *B. chrysoura* and *M. saxatilis* have six to eight pyloric caeca, *M. undulatus* and *L. xanthurus* have

seven to 10 pyloric caeca. *L. fasciatus* has 10 to 11 pyloric caeca. *L. fasciatus* and *L. xanthurus* have both a longer intestine and more pyloric caeca, but *L. fasciatus* is a mid-water feeder and *L. xanthurus* is a bottom feeder. They both have large amounts of small crustaceans in their diet. *C. regalis* has the shortest intestine and the fewest pyloric caeca, its diet is mainly composed of large crustaceans and fishes (see section "Food specialization"). Thus, the relative lengths of the intestine and the numbers of pyloric caeca in these sciaenids may be correlated with the size of the food rather than the feeding position in the water column, or their phylogenetic position (see part I of the whole study).

#### Pores and Barbels:

The mucus pores on the snout and the tip of the lower jaw, and mental barbels in fishes are sense organs probably involved in touch, taste, or both. The number and arrangement of the pores and barbels in sciaenid fishes are closely related to their feeding habitats (see Part I of the whole study). These six species of sciaenid fishes show a gradual increase in the number of pores from upper water column feeders to lower water column and bottom feeders (Fig. 25). *Larimus fasciatus* has five marginal pores on the snout and four pores at the tip of the lower jaw (Fig. 25,A & a). *Cynoscion regalis* has only two marginal pores on the snout and no pores or barbels on the lower jaw (Fig. 25,B & b). *Bairdiella chrysoura* has five marginal and five upper pores on the snout, and six mental pores at the tip of lower jaw (Fig. 25,C & c). *Leiostomus xanthurus* has five marginal and five upper pores on the snout, and five mental pores at the tip of the lower jaw (Fig. 25,F & f).



*Micropogonias undulatus* also has five marginal and five upper pores on the snout, and five mental and six minute barbels at the tip of the lower jaw (Fig. 25,D & d). *Menticirrhus saxatilis* has five marginal pores and three upper pores on the snout, and four mental pores and a short, rigid barbel at the tip of lower jaw (Fig. 25,E & e). An apical pore is also present on the barbel of *M. saxatilis*. The anterior margin of the snout (rostral fold) in *L. fasciatus* and *C. regalis* are complete without notches (Fig. 25,A & B). *B. chrysoura* and *L. xanthurus* have a similar indented rostral fold, although the former has a terminal mouth and the latter has an inferior mouth (Fig. 25,C & F). Both *M. undulatus* and *M. saxatilis* have deeply notched rostral folds (Fig. 25,D & E), correlated to their inferior mouth positions. The mental pores of *L. fasciatus* (Fig. 25,a) are the smallest of these sciaenids. The barbels of *M. undulatus* and *M. saxatilis* may differ in function as well as to number, because the single barbel of *M. saxatilis* has a pore at the tip, whereas barbels of *M. undulatus* do not (Fig. 25,d & e). The numbers of pores increase as the feeding niche of the fish tends toward the bottom and barbels are present only in the bottom feeders.

#### Nares:

Sciaenid fishes have two pairs of closely set nostrils. The anterior one is usually round; the posterior one is oval and elongate (Fig. 26). A flap of skin is sometimes also present along the posterior margin of the anterior nostril in bottom feeding species. The nasal cavity is generally oval shaped with a cluster of olfactory laminae forming a nasal rosette anteriorly (Fig. 26). *Larimus fasciatus* has

the shortest nasal cavity (Fig. 26,A) and *Leiostomus xanthurus* has the longest (Fig. 26,F). The shape of the nasal rosettes and olfactory laminae are similar in these six species of sciaenid fishes. The mean number of laminae (averaging both sides per specimen and rounding upward) differs among these species (Table 9) and is variable within a species. The numbers of laminae are 13 to 14 in *L. fasciatus*; 14 to 22 in *Cynoscion regalis*; 16 to 31 in *L. xanthurus* and 18 to 19 in *Menticirrhus saxatilis*. *L. fasciatus*, *C. regalis*, and *B. chrysoura* average fewer laminae than *M. undulatus*, *L. xanthurus* and *M. saxatilis* (Table 9). Within a species, the number of nasal laminae seems higher in larger specimens. The number of nasal laminae is greater in bottom feeding fishes.

#### Other Morphological Characters:

Differences in body shape, mouth structure, food specialization and habitat preferences of fishes may act to restrict interspecific competition within a fauna (Keast and Webb, 1966). The six species of sciaenid fishes discussed here show a correlation between body shape and feeding habitat (Fig. 27). Young *Larimus fasciatus* are oblong, and relatively deep and have a compressed body and a pointed tail (Fig. 27,A). These features, in combination with a strong oblique mouth and large eyes (Fig. 20,A & A' and Table 5) indicate that *L. fasciatus* is a moderate swimmer that feeds in the upper water column by sight. Young *Cynoscion regalis* have a fusiform and compressed body, and a long pointed tail, (Fig. 27,B). These features, in combination with a large oblique mouth and relatively large eyes (Fig. 20,B & B' and Table 5) indicate that *C. regalis* is a fast swimmer that feeds in the

upper water column by sight. Young *Bairdiella chrysoura* have an oblong and compressed body, and a broad and slightly rounded to truncate tail (Fig. 27,C). These features, together with its terminal mouth and relatively large eyes (Fig. 20,C & C' and Table 5) indicate that *B. chrysoura* is a moderate swimmer that feeds in the middle water column. Young *Micropogonias undulatus* have an elongate and less compressed body and a long pointed tail (Fig. 27,D). These features, combined with an inferior mouth with barbels and relatively smaller eyes (Fig. 20,D & D' and Table 5), indicate that *M. undulatus* is a moderate swimmer that feeds in the lower water column by sight, olfaction and touch. The young of *Leiostomus xanthurus* have a rather short and deep body, and a broad and truncate tail (Fig. 27,F). These features, combined with an inferior mouth and large eyes (Fig. 20,F & F' and Table 5), indicate that *L. xanthurus* is a slow swimmer that feeds in the lower water column by sight and olfaction. Young *Menticirrhus saxatilis* have an elongate, round and narrow body, and a relatively pointed tail (Fig. 27,E). These features, combined with an inferior mouth with a pored-barbel (Fig. 25,e) and relative smaller eyes (Table 5) indicate that *M. saxatilis* is a slow swimmer that feeds in the lower water column by sight, olfaction, and touch.

The cross sections of these young sciaenid fishes (Fig. 27) also indicate their habitat. *L. fasciatus*, *C. regalis*, and *B. chrysoura* are compressed and have relatively narrow ventral surfaces (Fig. 27,A, B & C) in comparison to *M. undulatus*, *L. xanthurus* and *M. saxatilis* (Fig. 27,D, E & F). Some of these morphological characters such as the shape of the tails and the size of the eyes vary ontogenetically. Generally, most juvenile sciaenids have pointed tails and relatively larger eyes than adults.

## V. Food Specialization:

The food habits of young sciaenids have been studied by numerous authors and the information reported by them is scattered and presented in different ways. Some of this work has been summarized for comparison with the present study (Tables 10 to 14). Only those studies having some sort of quantitative analysis were chosen for the comparison. Different authors have used different taxonomic categories to analyze their information. The classification of the food items in the present study has been modified from Darnell (1961) and Qasim (1972). Six major food groups were employed more or less according to their vertical occurrence in the water column, from the upper water column to the bottom. They were fishes, macrozooplankton, microzooplankton, epibenthos, infauna and other organic matters. Within each food group, several items were listed and the trivial and generic names of the primary prey species in the study area were indicated. The boundaries for these six food groups are not definite, because some prey taxa move vertically in the water column and some taxa may also include both pelagic and benthic species in parts of the water columns. Generalized terms used by many authors such as shrimps, annelids, mollusks, crabs, et. al. were placed under respective food groups for the convenience of comparison. The food habits of each species were compared with previous studies from different geographic areas and seasons. Food items were listed in different categories for each species. Under each listed item, there were cases where more than a single food taxon was listed by the original authors. Then the one

that had the highest frequency (by occurrence, volume, or weight) was chosen to represent that item.

All fish specimens used for stomach analyses in this study were randomly selected from specimens collected in June to November (1972 to 1974). During this period, these sciaenids reach their maximum abundance and degree of sympatry. All specimens were young-of-the-year or yearlings.

*Larimus fasciatus:*

In this study, stomachs of 12 specimens of *L. fasciatus* (14 to 125 mm TL) were examined. All stomachs contained crustaceans, exclusively: *Neomysis americana* in seven stomachs, Cumacea in five, Amphipoda (mostly *Gammarus*) in four and calanoid copepoda (mostly *Acartia tonsa*) in two. Most of these prey species were of small size.

Published information on food habits of *L. fasciatus* was scarce. Welsh and Breder (1923) reported on food of four *L. fasciatus* (50-110 mm SL) from Mississippi and Texas. Only two stomachs had food, one with a post-larval clupeoid and the other with "schizopodous forms" (crustacean remains).

*Cynoscion regalis:*

In this study, stomachs of 36 specimens (67-183 mm TL) of *C. regalis* were examined (Table 10). They fed only on *Anchoa mitchilli* and *Neomysis americana*. *A. mitchilli* was very abundant in the same area as *C. regalis* in the same months (Colvocoresses, 1975; Markle, 1976). Fishes and planktonic crustaceans were the major food items of *C. regalis* (Table 10). A shift of food habits with growth was noted by Thomas (1971), Merriner

(1975), and Stickney et. al. (1975). The smaller weakfish fed more on mysid shrimp and the larger weakfish fed more on fishes.

*Bairdiella chrysoura*:

In this study, stomachs of 68 specimens (57-190 mm TL) of *B. chrysoura* were examined (Table 11). They fed mainly on *Neomysis americanus* and *Anchoa mitchilli*. Juvenile *B. chrysoura* fed mainly on crustaceans and fishes (Table 11). Smaller specimens (less than 40 mm SL) fed mostly on copepods but as they grew they fed more on *Neomysis americanus*, amphipods and other larger crustaceans. Fishes became more important food items for specimens over 70 mm SL (Thomas, 1971; Carr and Adams, 1973; Stickney, et. al., 1975).

*Micropogonias undulatus*:

In this study, stomachs of 69 specimens (65-199 mm TL) of *M. undulatus* were examined (Table 12). They showed as wide a variety of prey items as have previous studies from other geographic areas (Table 12). Polychaetes and crustaceans were the main food items of the juvenile *M. undulatus* in the study area. Juvenile *M. undulatus* fed on a large variety of invertebrates and sometimes fishes (Table 12). Stickney, et. al. (1975) indicated that smaller specimens (less than 100 mm SL) depend extensively on hapacticoid copepods, which were mainly bottom dwellers. As the fish grew, they became more generalized feeders (Parker, 1971). Geographic variation in food habits of juvenile *M. undulatus* (Table 12) probably is attributable to availability of food species in the area.

*Menticirrhus saxatilis*:

In this study, stomachs of 20 specimens (36.5-118 mm TL) of *M. saxatilis* were examined. They all had crustaceans in the stomach and polychaetes were also important in their diet. The occurrence of organic detritus was also frequent. This suggests that *M. saxatilis* is a bottom feeder. Juvenile *M. saxatilis* also feed mainly on crustaceans and polychaetes (Table 13). Welsh & Breder (1923) indicated that *M. saxatilis* fed mainly on relatively large crustaceans.

*Leiostomus xanthurus*:

In this study, stomachs of 77 specimens (73-205 mm TL) of *L. xanthurus* were examined. Although they showed a wide variety of food species, the major part of the food was benthic. *Pectinaria gouldii*, a burrowing polychaete, was a major food item in the diet of *L. xanthurus* in the study area. This indicated that *L. xanthurus* not only feeds on the bottom but also on infauna. Juvenile *L. xanthurus* fed mostly on benthic invertebrates (Table 14). Stickney, et. al. (1975) chosed that harpacticoid copepods were the main food for juvenile *L. xanthurus* and that seasonal variations in diet were slight. Organic detritus and unidentified remains were also common in stomachs (Table 14).

To summarize the feeding habits of the juveniles of the six sciaenid species, a chart (Fig. 28) had been prepared for six food groups defined previously. The main food group of *Larimus fasciatus* was mostly planktonic (Fig. 28) and the primary food species was *Neomysis americana*. *Cynoscion regalis* and *Bairdiella chrysoura* fed

mainly on fishes and macrozooplankton (Fig. 28), the primary food species were *Anchoa mitchilli* and *N. americana*, respectively. *Micropogonias undulatus* fed on a wide variety of food including all six food groups (Fig. 28), with the dominant food organisms *N. americana* and *Nereis succinea*. *Menticirrhus saxatilis* fed mainly on macrozooplankton and epibenthos (Fig. 28), with the primary food organisms being *N. americana* and polychaetes. *Leiostomus xanthurus* also fed on a wide variety of food including five food groups (Fig. 28). The dominant food organisms were *N. americana* and *Pectinaria gouldii*. Food specialization or selectivity of the fishes *per se* does not seem sufficient to indicate food partitioning among them.

*Neomysis americana* was a very abundant and available food item to all species of sciaenids in the study area. This shrimp migrates vertically in response to change in ambient light (Herman, 1963). *N. americana* is negatively phototactic. In shallow turbid water (as in the study area) during daylight it might concentrate near the bottom in the darkest sector of the vertical light gradient (Stickney, et. al. 1975). Because of the abundance and availability of *N. americana*, the other prey items should provide a better indication of feeding specialization. As has been repeatedly shown (Tables 10 to 14), most fishes were sufficiently opportunistic in their food habits to take advantage of extremely abundant prey species. All the fishes in the present study were sampled by bottom trawl during the daytime. Therefore, both the prey and predators might have been dwelling close to the bottom.

Polychaetes were also the major food resources for the bottom feeders (Tables 12 to 14); *Micropogonias undulatus*, *Leiostomus*



*xanthurus* and *Menticirrhus saxatilis*. But, *M. undulatus* fed more on the "crawling" species of worms (Table 12) such as *Nereis* and *Nephtys* (Barnes, 1969) and *L. xanthurus* fed more on "tubicolous" or "burrowing" species of worms (Table 14) such as *Pectinaria* and *Amphitrite*. This is contradictory to the findings of Roelofs (1954) and Stickney et. al. (1975). Observations of the feeding behavior of these two species in the aquarium agreed with Roelofs (1954). But, *L. xanthurus* seemed to "dive" into the bottom sand much more often than *M. undulatus*, and the depth of the "dives" by *L. xanthurus* was not shallower than *M. undulatus* as stated by Roelofs (1954).

## VI. Correlation of Feeding Structures and Food Habits.

*Larimus fasciatus* and *Cynoscion regalis* have oblique mouths (Fig. 20,A & B) and their upper jaws are slightly or not protrusible (Fig. 21,A & B). These features allow them to feed anteriorly and dorsally to the longitudinal axis of their bodies along their swimming course. Their mouth opens as the lower jaw drops antero-ventrally and the distal ends of the premaxillae move forward (Fig. 20,A' & B'). The mouth openings of *L. fasciatus* and *C. regalis* are relatively larger than in the other species studied (Table 5). The anterior views of their mouth openings (Fig. 21,a, a', b & b') show that the upper jaws (premaxillae) are larger or equal to the lower jaws (dentaries). Although both of them feed "anterio-dorsally and pelagically" they did show differences in diet (Fig. 28). The following morphological characters are correlated with the dietary differences. The premaxillary and dentary teeth of both species are sharp and ridge-like. *C. regalis* has much larger teeth than *L. fasciatus* (Fig. 21,A & B) especially a pair of large canines at the tip of upper jaw in *C. regalis*. These large sharp teeth are adaptations for grasping larger swimming prey. Both species have small sharp pharyngeal teeth (Fig. 23,A & B). The arrangement and size of the gill rakers (Fig. 23,A & B) in *L. fasciatus* are much denser and longer than those of *C. regalis*. These differences reflect the food contents in the stomachs of *L. fasciatus*, which consisted of small crustaceans collected by the filtering function of the gill rakers. The stomach contents of *C. regalis* consisted of large crustaceans and fishes (Table 10). *L. fasciatus* has a much longer two-looped intestine than the straight intestine of *C. regalis* (4 to 5).

These morphological differences are probably correlated with the size of food ingested. The pore systems of *C. regalis* and *L. fasciatus* are not well-developed. *C. regalis* has only two marginal pores on the snout (Fig. 25,B) whereas *L. fasciatus* has five marginal pores on the snout and four pores on the tip of the lower jaw (Fig. 25,A). In addition, the fusiform *C. regalis* (Fig. 27,B) is adapted for fast swimming and active predation. The robust, and presumably slower moving *L. fasciatus* (Fig. 27,A) is probably better adapted for a plankton picking type of feeding (Davis & Birdsong, 1973).

*Bairdiella chrysoura* has a slightly oblique terminal mouth (Fig. 20,C) and a slightly protrusible upper jaw (Fig. 20,C'). These features allow the fish to feed directly in front of its body axis along its swimming course. Its mouth opens as the lower jaw drops antero-ventrally and the premaxillae move forward (Fig. 20,C'). The relative size of the mouth opening in *B. chrysoura* (Table 5) is similar to *L. fasciatus* and *C. regalis*. The anterior view of its mouth opening shows equal upper and lower jaws (Fig. 20,c'). Although *B. chrysoura* feeds "anteriorly", a "median" pelagic feeder, its stomach contents are similar to those of *C. regalis* (Fig. 28) except for a smaller proportion of fishes. The difference in the morphology of feeding structures may also give a clue to food partitioning between *B. chrysoura* and *C. regalis*. The jaw teeth of *B. chrysoura* are strong, conical and arranged in narrow ridges, but lacks a pair of canines at the tip of the premaxilla (Fig. 21,C). Its pharyngeal teeth are relatively stronger and blunter than in *C. regalis* (Fig. 22,C), especially along the median rows. Gill rakers of *B. chrysoura* are intermediate between *L. fasciatus* and *C. regalis* in number (Table 6)

and length (Fig. 23,C). The intestine of *B. chrysoura* has two loops and its relative length and number of pyloric caeca (6 to 8) are also intermediate between *L. fasciatus* and *C. regalis* (Fig. 24,C and Tables 7 & 8). These intermediate features reflect the intermediate feeding habits of *B. chrysoura* (Fig. 28). In addition, the body shape of *B. chrysoura* is oblong (Fig. 27,C) and not fusiform as in *C. regalis*, thus resulting in slower swimming and less efficiency in capturing fishes, as reflected in the diet. The relatively well-developed pore systems of *B. chrysoura* (Fig. 25,C), three upper and five marginal pores on the snout and six mental pores on the tip of the lower jaw, also may indicate that *B. chrysoura* depends more on "taste" and "touch" than "sight" feeding lower in the water column than *L. fasciatus* and *C. regalis*.

*Micropogonias undulatus*, *Leiostomus xanthurus* and *Menticirrhus saxatilis* have inferior mouths (Fig. 20,D, E & F) and rather protrusible premaxillae (Fig. 20,D', E' & F'). These features enable them to feed "anteriorly and ventrally" to their body axis along their swimming courses. Their mouths open as the lower jaw drops ventrally backward and premaxillae protrudes antero-ventrally (Fig. 20,D', E' & F'). Their mouth openings are relatively smaller than the pelagic feeders described previously (Table 5). The anterior views of their mouth openings (Fig. 20,d, e & f) show that the upper jaws (premaxillae) are smaller than the lower jaws (dentaries). Although they all feed antero-ventrally, and benthically, there are differences in their feeding habits (Fig. 28). These differences are reflected in the structural differences in the feeding apparatus and feeding behavior among them. The jaw teeth of *M. undulatus*, *L. xanthurus* and *M. saxatilis* are all

in bands and the outer row of teeth in the upper jaw and the inner row of teeth in the lower jaws are slightly enlarged. The pharyngeal teeth of *M. undulatus* and *M. saxatilis* are conical (Fig. 22,D & E) and the median rows are larger and blunt. *L. xanthurus* has smaller pharyngeal teeth and the median ones are molariform (Fig. 22,F). The gill rakers of these three bottom feeding sciaenids differ in number (Table 6) and size (Fig. 23,D, E & F). *M. saxatilis* has the fewest and shortest gill rakers among them. *M. undulatus* has fewer but longer gill rakers than *L. xanthurus*. The inner gill rakers of *L. xanthurus* (Fig. 23,f') are best developed, and most numerous (Table 6). This is reflected in the larger amounts of small crustaceans (e.g. copepods), ingested by *L. xanthurus* (Table 14). The relative length of intestines (Table 7) and *in situ* position (Fig. 24,D, E & F) are similar among benthic feeders. The average relative intestinal length of *M. undulatus* and *M. saxatilis* is slightly shorter than in *L. xanthurus* (Table 7). The intestine of *L. xanthurus* is much thinner than in *M. undulatus* and *M. saxatilis*, and the food is visible through the intestinal wall. *L. xanthurus* is the only sciaenid fish in the study area with a completely black peritoneal lining. The numbers of pyloric caeca of these bottom feeders are similar (Table 8). The pore and barbel system differ among *M. undulatus*, *L. xanthurus* and *M. saxatilis*. They all have five upper and five marginal pores on the tip of snout (Fig. 25,D, E & F). *M. undulatus* and *M. saxatilis* also have a deeply notched rostral fold. Ventrally, *M. undulatus* has five pores and six miniature barbels (Fig. 25,d); *M. saxatilis* has four pores and a short rigid barbel with an apical pore (Fig. 25,e); *L. xanthurus* has five pores and no barbel (Fig. 25,f). *M. saxatilis* also has the most pronounced snout and most

elongate and rounded body form (Fig. 27,E). *L. xanthurus* has the least pronounced snout and shortest and deepest body form (Fig. 27,F). *M. undulatus* is intermediate in snout and body form between *M. saxatilis* and *L. xanthurus*.

The length of snout and body form reflect the feeding habits of these three species. Food habits (Fig. 28) indicate that *M. saxatilis* and *M. undulatus* feed on the substrate, on the epifauna, more than they feed "into" the substrate on the infauna. *L. xanthurus* feeds more on the infauna. The long projecting snout seems to be an obstacle for fishes with an inferior mouth to forage into the bottom for food. Roelofs' (1945) observations on feeding behavior of *M. undulatus* and *L. xanthurus* in aquaria with sandy bottoms were repeated during the present study. Juveniles of both species foraged into the bottom sand often, especially when the substrate was freshly dug from the beach. Foraging tapered off gradually, especially in *M. undulatus*, apparently as the food in the substrate decreased. Brine shrimp (*Artemia*) were fed to these two species in the aquarium. Both *M. undulatus* and *L. xanthurus* were able to feed on brine shrimp just below the water surface. *M. undulatus* fed on the surface shrimp in an oblique to vertical position. To feed on brine shrimp close to the surface, *L. xanthurus* occasionally maneuvered in an oblique upside down position, with the dorsal fin pointing toward the bottom to overcome its inferior mouth position.

Other accessory organs of feeding, such as the nares and eyes also reflect the feeding habits of young sciaenid fishes. The numbers of nasal laminae of the six species (Table 9) overlap due to ontogenetic changes. The absolute number of the nasal laminae increase as the

fishes grow larger. The upper ends of ranges in number (Table 9) are most important, as they indicate the highest number of nasal laminae each species may develop. Generally, the bottom feeders, *M. undulatus* and *L. xanthurus*, have more nasal laminae than *L. fasciatus*, *C. regalis* and *B. chrysoura* (Table 9). *M. saxatilis* has relatively fewer nasal laminae than other benthic feeders, but it has a pored mental barbel on the lower jaw. This may suggest that *M. saxatilis* depends more on "touch" sense for foraging than other benthic feeders. The relative eye size of *M. saxatilis* is smaller than in other sciaenid fishes studied here (Table 5). Larger eye sizes are found among the pelagic feeders, *L. fasciatus*, *C. regalis* and *B. chrysoura* (Table 5).

Allometrically, the relative eye size of all these sciaenid fishes is larger in young specimens and smaller in adults. For benthic feeders, decrease in relative eye size with growth is faster than for the pelagic feeders. The relative roles of olfaction, touch and vision in feeding in young sciaenids studied may be hypothesized as follows. The benthic feeders, *M. undulatus*, *L. xanthurus* and *M. saxatilis* depend more on their senses of "smell" or "touch" or both to locate their prey. The pelagic feeders, *L. fasciatus*, *C. regalis* and *B. chrysoura* depend more on their senses of "sight" to catch their prey, especially in *C. regalis* and *B. chrysoura* which prey on *Anchoa mitchilli*, an active small anchovy.

Morphological differences in the feeding apparatus, especially the mouth position, size and protrusibility, the form of teeth and the gill raker structure are limiting factors for the habitat and the size of the prey species which can be eaten by the fish. Evidently, the off-bottom feeders, *L. fasciatus*, *C. regalis* and *B. chrysoura* almost

completely lack any sedentary benthos in their diets (Fig. 28). Even among the bottom feeders, *M. undulatus* feeds more on epibenthic polychaete species (Table 13) and *L. xanthurus* feeds more on the burrowing polychaete species (Table 14).

Morphological differences of the digestive tract, the number of pyloric caeca and the length of intestine may be an adaptation to more efficient use of food already ingested. As is evident in *L. fasciatus* and *L. xanthurus*, size of the food items is important, *L. fasciatus* fed exclusively on small crustaceans (small Mysidacea and Amphipoda); *L. xanthurus* fed mainly on copepods (Table 14). *L. fasciatus* is mainly a pelagic feeder and *L. xanthurus* is mainly a benthic feeder. Both species have longer intestines (Table 7) and more pyloric caeca (Table 8) than other species in their respective groups (pelagic and benthic).

The so-called "selective feeding habits" of these young sciaenids reported by many previous authors (see citation of Tables 10-14) are not evident in the present study. Partitioning of food among these young sciaenids depends more on the habitats of the prey species than on the "selective preferences" of the fishes. These juvenile sciaenids feed opportunistically in a limited depth range in the water column, probably within or close to two meters above the bottom. Within this layer of the water column, *Larimus fasciatus*, *Cynoscion regalis*, and *Bairdiella chrysoura* feed in the upper portion of the water column and *Micropogonias undulatus*, *Leiostomus xanthurus* and *Menticirrhus saxatilis* feed in the lower portion of the water column to the bottom. The difference in habitat preference of juvenile sciaenids from York River was also demonstrated by Cooke (1974).



Feeding niche division and resulting dietary differences among these species of sciaenids in the Chesapeake estuary area are attributable to differences in feeding behavior imposed upon these species by adaptive morphological limitations rather than to selective feeding *per se*.

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Table 1. Growth of Spot, Leiostomus xanthurus, from different estuarine areas along U.S. Atlantic and Gulf of Mexico coasts.

Author	Thomas, 1971	Young, 1953	Hildebrand & Schroeder, 1928
Locality	Delaware River, Delaware	Chesapeake Bay, Maryland	Chesapeake Bay
Period	1969 (June 1968 to Sept. 1970)	May to October 1951	Prior to 1928
Gear*	16 ft T & S	75 x 4 ft Haul S.	?
Source	Table 68	Tables 4 & 5	Table on p. 273
Length (mm)	Total length	Total length	Total length
Age groups*	O I	O I	O I
January			75-149
February			
March			15-19
April			20-24
May			15-74
June	19.2	16-60	20-99
July	30-80	26-80	40-124
August	45-100	(26-130)	65-149
September		86-90	94-170
October		86-106	100-184
November		(71-155)	190-209
December			190-299
			75-184
			75-119

\* Gear: G, gill net; P, pound net; Pl, plankton net; Pu, push net; R, rotenone; S, seine; T, trawl; Tr, trammel net.

Age groups: O represents smallest group of young-of-the-year first taken from January on, other fishes (including overwintering young-of-the-year) are included in age group I. Parentheses indicate that the boundary of age 0 and I groups is indistinguishable.



Table 1. (continued, growth of Leiostomus xanthurus)

Author	Chao, 1976	Facheco, 1957	Hildebrand & Cable, 1930
Locality	York River, Virginia	Lower Chesapeake Bay & York River, Virginia	Beaufort, North Carolina
Period	Jan. 1972 to Dec. 1974	May 1955 to Feb. 1956	Prior to 1930
Gear*	16 ft T & S	P & 30 ft T	P1 & T
Source	Fig. 6 (present study)	Table 3	Tables 7 & 8
Length (mm)	Total length	Total length	Total length
Age groups*	0 O I	0 O I	0 O I
January		155-255	82-195
February		150-275	91-200
March			93-200
April			84-214
May	80-105	130-225	97-215
June	115-(140)	(145)-210	122-198
July	115-(150)	(155)-230	130-228
August	125-(180)	(200)-245	140-219
September	135-(185)	(190)-260	155-234
October	135-(235)		175-269
November	165-(205)		190-264
December	155-185	220-240	84-188
			1.5-9.2

Table 1. (continued, growth of Leiostomus xanthurus)

Author	Shealy, et. al., 1974	Music, 1974	Townsend, 1956
Locality	South Carolina	Georgia	Alligator Harbor, Florida
Period	Feb. 1973 to Jan. 1974	Oct. 1970 to Sept. 1973	March 1955 to May 1956
Gear*	20 ft T	40 ft T, 12 ft S, 300 ft G	150 & 600 ft S
Source	Table 27	Fig. 10	Table I
Length (mm)	Total length	Total length	Total length
Age groups*	O I	O I	O I
January			
February			
March			
April			
May			
June			
July			
August			
September			
October			
November			
December			

Table 1. (continued, growth of Leiostomus xanthurus)

Author	Springer & Woodburn, 1960		Sundararaj, 1960		Dunham, 1972	
Locality	Tampa Bay, Florida		Lake Ponchartrain, Louisiana		Coast, Louisiana	
Period	Jan. to Dec. 1958		July 1953 to May 1955		July 1969 to June 1972	
Gear*	T, 80 ft S & Pu		T, Tr & S,R		16 ft T	
Source	Table 13		Fig. 17		Fig. 21	
Length (mm)	Standard length		Total length		Total length	
Age groups*	O	I	O	I	O	I
January	13-31		15-25	90-165		100-170
February	13-49		15-40	115-165	10-80	110-170
March	10-73		10-75	140-230	20-100	110-170
April	19-79		30-100	140-255	40-110	
May	25-85		45-120	(120)-240	50-125	
June	31-103		55-145	150-255	50-155	
July	48-118		40-160	165-250	20-155	
August	49-103		85-(180)	(180)-215	70-160	
September	52-82		95-(150)	(150)-210	90-170	210
October	52-97		90-150	170-190	120-160	
November	67-91		110-(170)	(170)-205	100-180	
December	76-109		135-165		70-180	

Table 1. (continued, growth of Leiostomus xanthurus)

Author	Nelson, 1969		Parker, 1971		Pearse, 1928	
	Locality	Period	Locality	Period	Locality	Period
	Mobile Bay, Alabama	May 1963 to April 1964	Galveston Bay, Texas	Jan. 1963 to Dec. 1965	Subline River to Rio Grande, Texas	March 1926 to May 1927
Gear*	16 ft T		4.0 m. T		Tr, T, S & G	
Source	Table 9		Table 2		Table 31	
<u>Length (mm)</u>	<u>Total length</u>	<u>Total length</u>	<u>Total length</u>	<u>Total length</u>	<u>Total length</u>	<u>Total length</u>
Age groups*	O	I	O	I	O	I
January		75-160		60-170		
February		90-125	30	70-180		
March		90-180	10-30	60-190		
April		90-165	10-70	90-160		
May	45-70	(130)-171	30-100	110-190	10-90	120-250
June	45-(125)	170-180	30-110	140-190	40-120	130-250
July	50-140	200	30-140	170	70-(150)	(150)-230
August	55-145		30-150	170-180	80-140	150-230
September	80-135		30-160		110-(220)	230-270
October	95-(190)		50-160		110-(240)	250-260
November	95-165		60-150		110-(170)	(170)-260
December	90-175	200	70-180		130-190	200-250
					130-190	200-250

Table 2. Growth of Weakfish, Cynoscion regalis from different estuarine areas along U. S. Atlantic coast.

Author	Thomas, 1971	Pearson, 1941	Chao, 1976
Locality	Delaware River, Delaware	Lower Chesapeake Bay	York River, Virginia
Period	1969	1929-1930	Jan. 1972 to Dec. 1974
Gear*	T & S	Pl & P	16 ft T
Source	Table 4	Fig. 23	Figs. 10 to 12 (present study)
Length (mm)	Total length	Total length	Total length
Age groups*	0	0	0
January			60-200
February			315
March			65-175
April		130-250	155-330
May			140-385
June	5-70		105-305
July	15-125		100-370
August	15-(185)		20-55
September	70-(185)		10-(95)
October	40-(175)	(130)-180	70-(110)
November			35-(135)
December			65-(140)
			95-(170)

\*Gear: P, pound net; Pl, plankton net; S, seine; T, trawl.

Age group: 0 represents smallest groups of young-of-the-year taken from January on, other fishes (including overwintering young-of-the-year) are included in age group I. Parentheses indicate that the boundary of age 0 and I groups is indistinguishable.

Table 2. (continued, growth of Cynoscion regalis)

Author	Hildebrand & Cable, 1934	Shealy et. al., 1974	Mahood, 1974
Locality	Beaufort, North Carolina	South Carolina coast	Georgia coast
Period	?	Feb. 1973 to Jan. 1974	Oct. 1970 to Sept. 1973
Gear*	Fl, P & T	20 ft T	40 ft T.
Source	Table 4	Table 32	Table 7
Length (mm)	Total length	Total length	Total length
Age groups*	O I	O I	O I
January	75-204		68-438
February	105-274	138-327	68-398
March	90-230	155	83-358
April	80-284	118-188	78-408
May	125-224		48-358
June	95-279	23-47	(133)-328
July	40-379	23-(52)	(178)-363
August	65-369	(73)-182	(208)-323
September	80-314	(68)-208	(218)-388
October	100-329	(73)-228	(228)-313
November	(100)-329	58-72	(238)-348
December	(95)-299	88-92	(238)-348

Table 3. Growth of silver perch, Bairdiella chrysoura, from different estuarine areas along U.S. Atlantic and Gulf of Mexico coasts.

Author	Thomas, 1971	Chao, 1976	Hildebrand & Cable, 1930
Locality	Delaware River, Delaware	York River, Virginia	Beaufort, North Carolina
Period	1969	Jan. 1972 to Dec. 1974	Spring 1926 to Summer 1927
Gear*	16 ft T	16 ft T	Pl & T
Source	Table 28	Figs. 13 to 15 (present study)	Tables 5 & 6
Length (mm)	Total length	Total length	Total length
Age groups*	0 I I I	0 I I	0 I I
January			74-204
February			90-209
March			98-204
April			93-195
May			85-204
June	5-20	85-200	1-6
July	5-65	145-185	1-38
August	45-100	120-190	9-76
September	70-120	100-205	20-92
October	65-130	160-210	45-122
November		160-220	73-115
December		210	68-143
		73-110	78-124

\*Gear: Pl, plankton net; Pu, puchnet; S, seine; T, trawl.

Age groups: 0 represents smallest group of young-of-the-year first taken from January on, other fishes (including overwintering young-of-the-year) are included in age group I. Parentheses indicate that the boundary of age 0 and I groups is indistinguishable.

Table 3. (continued, growth of Bairdiella chrysoura)

Author	Springer & Woodburn, 1960	Reid, 1954
Locality	Tampa Bay, Florida	Cedar Key, Florida
Period	Oct. 1957 to Dec. 1958	June 1950 to May 1951
Gear*	T, S & Pu	15 ft T, S & Pu
Source	Fig. 12	Fig. 10
Length (mm)	Standard length	Standard length
Age groups*	O I	O I
January		55-60
February	18-(72)	67
March	93-182	52-76
April	88-137	67-73
May	98-172	
June	73-182	5-40
July	113-152	15-50
August	123-132	20-70
September	128-192	5-80
October	143-172	10-82
November	138-177	40-95
December	33-87	50-70
	58-107	
	73-132	
	78-(187)	
	98-(172)	
	98-(182)	



Table 4. Growth of Croaker, Microponias undulatus, from different estuarine areas along U.S. Atlantic and Gulf of Mexico coasts.

Author	Thomas, 1971		Haven, 1957		Chao (1976)	
	Locality	Period	Locality	Period	Locality	Period
	Delaware River, Delaware	June 1968 to Dec. 1970	York River, Virginia	Oct. 1952 to July 1953	York River, Virginia	Jan. 1972 to Dec. 1974
Gear*	16 ft T		30 ft T		16 ft T & S	
Source	Table 70		Fig. 7		Figs. 16 to 19 (present study)	
Length (mm)	Total length		Total length		Total length	
Age groups*	O	I	O	I	O	I
January			15-(85)	(95)-105	20-120	(20)-240
February			10-60	(45)-100	20-155	(60)-245
March			10-(70)	(70)-120	20-175	(80)-250
April			10-(65)	70-100	25-120	(70)-240
May			25-(90)	70-140	20-30	(60)-245
June			40-(120)	(120)-155	20-70	(70)-240
July			75-145	(135)-175	30-(110)	(70)-240
August			N.S.	N.S.	30-(90)	(70)-195
September			N.S.	N.S.	10-(100)	(100)-250
October	20	135-140	10-(40)	(40)-85	10-(110)	(60)-250
November	25		15-(60)	(60)-115	15-100	(60)-250
December	20-50		10-(60)	(60)-120	20-110	165-175

\*Gear: Pl, plankton net; S, seine; T, trawl.

Age group: 0 represents smallest group of young-of-the-year first taken from January on, other fishes (including overwintering young-of-the-year) are included in age group I. Parentheses indicate that the boundary of age 0 and age I groups is indistinguishable. N.S.; no sample.



Table 4. (continued, growth of Microgogonias undulatus)

Author	Hansen, 1969	Suttkus, 1955	Parker, 1971
Locality	Pensacola, Florida	Lake Pontchartrain, & Louisiana Coast	Galveston Bay, Texas
Period	Aug. 1963 to Dec. 1965	July 1953 to Oct. 1954	Jan. 1963 to Dec. 1965
Gear*	5 m T	T & S	4.9 m T
Source	Fig. 2	Table 1	Fig. 21
Length (mm)	Total length	Total length	Total length
Age groups*	O I	O I	O I
January	15-20 45-95	10-79 120-189	10-(80) 90-200
February	20-25 40-95	10-89 130-179	10-(90) (90)-250
March	15-35 75-85	20-119 120-259	10-(90) (100)-250
April	N.S.	20-129 130-339	10-(120) (130)-250
May	20-(75) (60)-135	30-139 140-319	10-(130) (130)-240
June	30-(95) (90)-150	30-139 140-329	40-(140) (156)-250
July	35-(90) (90)-145	50-159 160-380	30-(150) (160)-230
August	35-(110) (100)-150	80-169 170-319	60-160 170-250
September	40-(90) (90)-150	80-169 170-319	60-(170) (170)-190
October	45-(110) (110)-150	90-189 (170)-349	10-40 60-220
November	50-105	20-59 130-309	10-(60) 60-210
December	10-95	10-79 120-299	10-(70) 70-230

Table 5. Relative size of mouth and eye diameter in six species of juvenile sciaenids from York River.

Species	SL (mm)		Index of mouth size		S.D.		N.		SL (mm)		Eye diameter in % of SL	
	Range	$\bar{x}$	Range	$\bar{x}$	S.D.	N.	Range	$\bar{x}$	S.D.	N.		
<u>Larimus fasciatus</u>	55.3-107	4.634	3.17-5.90	4.634	0.957	20	55.3-107	7.38-9.84	8.602	0.672	21	
<u>Cynoscion regalis</u>	35.2-75.3	2.827	1.929-3.54	2.827	0.518	22	35.2-82.4	8.20-11.45	9.55	0.782	26	
<u>Bairdiella chrysoura</u>	38.4-76.2	2.494	1.76-3.08	2.494	0.431	17	30.0-75.3	8.27-10.82	9.407	0.677	20	
<u>Micropogonias undulatus</u>	35.5-116	1.686	1.20-2.41	1.686	0.325	30	35.5-116	6.45-9.46	7.850	0.837	30	
<u>Menticirrhus saxatilis</u>	29.2-99.6	0.957	0.50-1.37	0.957	0.264	30	29.2-99.6	6.03-8.56	7.043	0.644	30	
<u>Leiostomus xanthurus</u>	47.4-146	1.472	0.77-2.64	1.472	0.477	30	47.4-163	7.05-11.11	9.139	0.889	46	

\* Index of mouth size = (upper jaw length x lower jaw length) / head length.

Table 6. Total number of lateral and medial gill rakers in six species of juvenile sciaenids from York River.

Species	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	N	$\bar{x}$				
<u>Menticirrhus saxatilis</u>						6	11	9	3	1											30	12.04				
(29.2-99.6 mm SL)	[5	20	4	1]																	[ 30	6.73]				
<u>Cynoscion regalis</u>			[1		4	10	8	2	2]	1	8	13	13	2							37	17.19				
(35.2-75.3 mm SL)																					[ 27	11.40]				
<u>Micropogonias undulatus</u>														1	3	16	15	7			42	22.55				
(35.5-116 mm SL)									[2	8	16	4]									[ 30	15.73]				
<u>Bairdiella chrysoura</u>														2	3	13	14	1			33	24.27				
(38.4-75.3 mm SL)										[1	2	5	6	6]							[ 20	15.70]				
<u>Leiostomus xanthurus</u>	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	N	$\bar{x}$		
(47.4-148 mm SL)													4	7	6	12	9	13	3	1			55	32.29		
<u>Larimus fasciatus</u>																							[ 44	27.18]		
(55.3-107 mm SL)																				5	9	5	2	1	22	38.00
																							[ 21	21.04]		

[ ] medial gill rakers

Table 7. Relative length of intestine in six species of juvenile sciaenids from York River.

Species	SL (mm)	Intestine length in % of SL		
		Range	$\bar{x}$	S.D. N.
<u>Cynoscion regalis</u>	35.2-152	35.5-49.6	40.24	3.07 36
<u>Bairdiella chrysoura</u>	30.0-151	46.1-64.1	55.34	5.92 30
<u>Micropogonias undulatus</u>	35.5-145	52.3-88.6	65.57	6.56 39
<u>Menticirrhus saxatilis</u>	29.2- 91.2	56.6-88.2	76.06	6.67 26
<u>Larimus fasciatus</u>	35.3- 99.8	73.1-97.7	83.87	9.08 14
<u>Leiostomus xanthurus</u>	47.4-166	73.6-97.8	84.69	6.95 30

Table 8. Number of pyloric caeca in six species of juvenile sciaenids from York River.

Species	4	5	6	7	8	9	10	11	N	$\bar{x}$
<u>Cynoscion regalis</u> (35.2-82.4 mm SL)	20	14							34	4.41
<u>Bairdiella chrysoura</u> (30.0-75.3 mm SL)			8	20	1				29	6.76
<u>Menticirrhus saxatilis</u> (29.2-99.6 mm SL)			11	19					30	6.63
<u>Microponias undulatus</u> (35.5-116 mm SL)			1	25	11				37	8.27
<u>Leiostomus xanthurus</u> (47.4-148 mm SL)			6	13	8	1			28	8.14
<u>Larimus fasciatus</u> (55.3-107 mm SL)						9	6	15		10.4

Table 9. Number of laminae in olfactory rosettes in six species of juvenile sciaenids from York River.

Species	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	N	$\bar{x}$	
<u>Larimus fasciatus</u> (55.3-107 mm SL)	3	5	2	5																			15	12.6	
<u>Cynoscion regalis</u> (35.2-86.4 mm SL)	1	6	6	4	7	5	1	2	1	1	2												36	15.89	
<u>Bairdiella chrysoura</u> (30.0-75.3 mm SL)	2	2	2	8	4	6	2	6	3	1	1												37	16.81	
<u>Microgogonias undulatus</u> (35.5-116 mm SL)	1	1		3	3	4	3	3	2	1	2	1	2	3	2	2	1	1	1	1	1	1	1	34	19.53
<u>Menticirrhus saxatilis</u> (29.2-99.6 mm SL)	1			1	6	6	3	3	6	3	2	1											32	17.31	
<u>Leiostomus xanthurus</u> (47.4-148 mm SL)							1	2	4	4	3	2	7	6	1	5	1	1	1	1	1	1	37	21.70	



Table 10. Stomach contents of weakfish, Cynoscion regalis from different estuarine areas along U.S. Atlantic coast.

Author	Chao, present study		Welsh & Breder, 1923	
Locality	York River, Virginia	Acushnet River, Massachusetts	Cape Charles, Virginia	Winyah Bay, South Carolina
Period	June to Aug. 1973	Sept. 1882	Sept. 1916	July 1915
Source	Original	p. 159	p. 160	p. 161
Number of specimens	36	28	45	34
Empty stomachs	2	5	0	5
Length of specimens	70-183 mm TL	7-11 cm SL	43-11.5 cm SL	2.8-6.2 cm SL
Quantitative Method	% of occurrence	% of volume	% of volume	% of volume
<b>Fishes</b>				
<i>Anchoa mitchilli</i>	72.2	48.0	2.0	9
Others & remains	8.3			18
<b>Macrozooplankton</b>				
Nysid sp.	2.8			
<i>Neomysis americana</i>	63.9			6
Isopoda		0.5	0.5	
Delapoda (shrimps)		47.0	91.0	83
Others & remains				46
<b>Microzooplankton</b>				
Copepoda			3.5	2
Calanoid sp.				
Others & remains				
<b>Epibenthos</b>				
Polychaeta		0.5		
<i>Nereis succinea</i>			3.0	
Amphipoda				
<i>Gammarus</i> sp.				
Others & remains				
Unidentified remains	5.6	4.0		18

Table 10. (continued, stomach contents of Cynoscion regalis)

Author	Thomas, 1971			
Locality	Delaware River, Delaware			
Period	June 1969	July 1969	Aug. 1969	Sept. 1969
Source	Table 20	Table 20	Table 20	Table 20
Number of specimens	71	94	94	66
Empty stomachs	10	11	10	12
Length of specimens	11-76 mm TL	5-123 mm TL	15-180 mm TL	20-180 mm TL
Quantitative method	% of occurrence	% of occurrence	% of occurrence	% of occurrence
Fishes	7.0*	14.9*	16.0*	34.8*
<u>Anchoa mitchilli</u>	1.4	2.1	1.1	4.5
Others & remains	2.8	7.4	13.8	30.3
Macrozooplankton	74.6	59.6	65.8	66.7
<u>Nyctid sp.</u>				
<u>Neomysis americana</u>				
<u>Isopoda</u>		4.3	2.1	1.7
<u>Deiapoda (shrimps)</u>		2.1	3.2	6.7
Others & remains				10.6
Microzooplankton	19.7	4.3	2.1	3.3
<u>Copepoda</u>				
<u>Calanoid sp.</u>				
Others & remains	9.9	4.3	1.1	0.8
Epibenthos				
<u>Polychaeta</u>				
<u>Neris succinea</u>				
<u>Amphipoda</u>				
<u>Gammarus sp.</u>	9.9	58.5	58.5	28.3
Others & remains				
Unidentified remains				28.8

\* All fishes combined

Table 10. (continued, stomach contents of Cynoscion regalis)

Author	Merriner, 1975	Stickney, et. al., 1975
Locality	Pamlico sound & Morehead City, North Carolina	Savannah River & Oseabow sound, Georgia
Period	June 1967 to Jan. 1970	May 1972 to July 1973
Source	Table 1	Table 1
Number of specimens	2159	120
Empty stomachs	1342	35
Length of specimens	135-481 mm SL	30-169 mm SL
Quantitative method	% of occurrence	% of volume
Fishes		% of occurrence
<u>Anchoa mitchilli</u>	58.1	2.5
Others & remains	15.7	31.7
Macrozooplankton		
Nysid sp.		0.8
<u>Neomysis americana</u>	31.0	55.0
Isopoda		2.5
Deiapoda (shrimps)	0.1	2.5
Others & remains	1.5	
Microzooplankton		
Copepoda		5.0
Calanoid sp.		2.5
Others & remains		
Epibenthos		
Polychaeta	0.5	15.0
<u>Nereis succinea</u>		2.5
Amphipoda		1.7
<u>Gammarus</u> sp.		9.2
Others & remains	1.5	2.5
Unidentified remains	96.8	2.5

Table 11. Stomach contents of silver perch, Bairdiella chrysoura from different estuarine areas along U.S. Atlantic and Gulf of Mexico coasts.

Author	Chao, present study	Thomas, 1971	Welsh & Ertter, 1923
Locality	York River, Virginia	Delaware River, Delaware	Cape Charles, Virginia
Period	June to Aug. 1973	Aug. to Oct. 1969	Sept. 1916
Source	Original	Table 46	P. 174-175
Number of specimens	68	99	21
Empty stomachs	10	9	0
Length of specimens	57-153 mm TL	5-130 mm TL	6-8.2 cm TL
Quantitative method	% of occurrence	% of occurrence	% of occurrence
Fishes		12.1*	
Anchoa mitchilli	5.1	3.0	1
Others & remains	20.7	8.1	
Macrozooplankton		89.9	
Nysid sp.	1.7		
Nemysis americana	74.1	1.0	5
Isopoda	1.7	17.2	87
Decapoda (shrimp)	10.3		
Others & remains	1.7		
Microzooplankton			
Copepoda			
Others & remains			
Epibenthos			
Annelida (polychaete)	3.5		2
Nereis succinea	3.5		
Cnidaria	1.7		
Amphipoda	1.7	15.2	5
Gammarus sp.	3.5	62.6	
Crabs		3.0	
Others & remains	1.7		
Infauera (bivalve & nemaloda)	3.5		
Unidentified Remains	6.9	2.0	

\* All fishes combined

Table 11. (continued, contents of Bairdiella chrysoura)

Author	Stickney, et. al., 1975	Reid, 1954	Carr & Adams, 1973
Locality	Savannah River & Ossabow sound, Georgia	Cedar key, Florida	Crystal River, Florida
Period	May 1972 to July 1973	June 1950 to May 1951	Oct. 1970 to Aug. 1971
Source	Table 1	Table 5	Estimate from Fig. 9
Number of specimens	161	45	195
Empty stomachs	48	0	43
Length of specimens	30-149 mm TL	25-99 mm SL	5-130 mm TL
Quantitative method	% of occurrence	% of occurrence	% of occurrence
<b>Fishes</b>			
<u>Anchoa mitchilli</u>	2.7		
Others & remains	6.6	4.4	31.2
<b>Macrozooplankton</b>			
Nysid sp.	0.6		
<u>Neomysis americana</u>	25.1		
Isopoda	1.1		
Decapoda (shrimp)	5.5	73.3	51.6
Others & remains	8.2	4.4	
<b>Microzooplankton</b>			
Copepoda	3.9		
Others & remains	2.2	4.0	
<b>Epibenthos</b>			
Annelida (polychaete)	0.6	2.2	7.3
<u>Neris succinea</u>	8.2		
Cumacea			
Amphipoda	2.2	33.3	9.2
<u>Gammarus</u> sp.	6.0		
Crabs	8.2		
Others & remains	0.6	16.6	16.6
<b>Infaua (bivalve &amp; nemaloda)</b>			
Unidentified Remains		6.6	8

Table 12. Stomach contents of croaker, Microponogonias undulatus from different estuarine areas along U.S. Atlantic and Gulf of Mexico coasts.

Author	Chao	Thomas, 1971	Røelofs, 1954	Welsh & Breder, 1923
Locality	York River, Virginia	Delaware River, Delaware	Coast, North Carolina	Winyah Bay Cape Canaveral South Carolina Florida
Period	June to Aug. 1973	Nov. to Dec. 1970	All seasons 1950	July 1915 Dec. 1919
Source	Original	Table 71	Table 1	P. 184 P. 183
Number of specimens	69	25	159	37
Empty stomachs	5	3	?	0
Length of specimens	56-199 mm TL	23-50 mm TL	60-140 mm TL	4.2-6.2 cm SL
Quantitative method	% of occurrence	% of occurrence	% of occurrence	% of volume
Fish & Remains	20.3		3.1	0.7
Macrozooplankton				
Mysidace	3.1		5.7	
Neomysis americana	35.9	64.0		
Isopoda	1.6	4.0		
Decapoda (or shrimp)	9.4		3.1	7.0
Insecta	1.6			
Others & Remains	3.1			7.0
Microzooplankton				
Copepoda	3.1			
Calanoid	10.9	76.0	25.2	
Harpacticoid				
Ostracoda				
Others & Remains	6.3		4.4	2.0

Table 12. (continued, stomach contents of Microgogonias undulatus)

Author	Chao	Thomas, 1971	Roslofs, 1954	Welsh & Breder, 1923 Winyah Bay Cape Canaveral
Epibenthos				
Annelids (or polychaets)	67.2		89.9	29.0
Neris succinea	18.8			3.0
Glycinde solitaria	9.4			
Pyliodocid	1.6			
Spionid	6.3			
Cumacea	4.7			2.0
Amphipoda	21.9		5.7	
Gammarus sp.	7.9			1.8
Crabs	3.1			1.0
Others & Remains	6.3			2.5
Infauna				
<u>Pectinaria gouldii</u>	15.6			
<u>Ampharetid</u>	1.6			
Gastropoda	1.6			
Pelecypoda			11.3	
Nematoda			0.6	
Others & Remains				
Unidentified Remains & Organic Matters	23.5		4.4	22.0
				48.0

Table 12. (continued, stomach contents of Microgogonias undulatus)

Author	Stickney, et. al., 1975	Hansen, 1969	Lake Pontchartrain, Louisiana	Parker, 1971 Clear Lake, Texas
Epibenthos				
Annelids (or polychaets)	12.4		17.0	6.0
Neris succinea	3.0	61.5		6.5
<u>Glycynde solitaria</u>				
Phyllococid				
Spionid	1.0			
Cumacea	6.0			
Amphipoda	8.0		4.0	4.5
Gammarus sp.	9.5			5.0
Crabs	4.5		17.0	10.0
Others & Remains	7.0		17.0	18.5
				16.0
				22.0
				10.5
				18.0
Infauuna				
<u>Pectinaria gouldii</u>				
Ampharetid				
Gastropoda	2.5		9.0#	3.0#
Pelecypoda		3.5#		18.0#
Nematoda		0.3		
Others & Remains	1.5			
Unidentified Remains & Organic Matters	35.6	2.5	17.0	12.0
* Arthropods				
# Mollusks				
				9.0



Table 12. (continued, stomach contents of Microponogonias undulatus)

Author	Stickney, et. al., 1975	Hansen, 1969	Parker, 1971
Locality	Savannah River & Ossabow Sound, Georgia	Pensacola, Florida	Lake Pontchartrain, Louisiana Clear Lake, Texas
Period	May, 1972 to July, 1973	Aug. 1963 to Dec: 1965	Jan. 1963 to Dec. 1965
Source	Table 1	Table 5	Table 28
Number of specimens	196	2520	63
Empty stomachs	15	?	2
Length of specimens	39-180 mm SL	76-173 mm TL	10-74 mm TL 75-124 mm TL 10-96 mm TL 70-110 mm TL
Quantitative Method	% of occurrence	% of volume	% of occurrence
Fish & Remains	7.5	15.9	11
Macrozooplankton		14.3*	4
Mysidace	1.0		2
Neomysis americana	16.9		4.5
Isopoda	1.0		8.0
Decapoda (shrimp)	3.0		17.0
Insecta	0.5		3
Others			18.5
Microzooplankton			13.0
Copepoda	7.0		2.0
Calanoid	1.0		6.5
Harpacticoid	10.0		1.0
Ostracoda	0.5		8.0
Others & Remains			12.0
			9.0
			3.0
			18.0
			14.5
			20.0

Table 13. Stomach contents of northern kingfish, Menticirrhus saxatilis, from different estuarine areas along U.S. Atlantic coast.

Author	Chao		Welsh & Breder, 1945	
Locality	York River, Virginia	March 1972 to Dec. 1974	Cape May, New Jersey	Falmouth, Massachusetts
Period			August, 1916	August, 1892
Source	Original		p. 194	p. 194
Number of specimens	20		21	17
Empty Stomachs	0		0	4
Length of Specimens	37-118 mm TL		1.9-7.2 cm SL	2.4-7.4 cm SL
Quantitative Methods	% of Occurrence		% of Volume	% of Volume
<b>Macrozooplankton</b>				
Neomysis americana	35.0		5.0	
Isopoda			9.0	42.0
Decapoda (shrimp)				
<u>Crangon septemspinosus</u>	5.0			
<u>Palaeomonetes</u>	10.0			
Insecta	5.0			
Others & remains	70.0		9.0	42.0
<b>Microzooplankton</b>				
Copepoda	5.0			
Calanoid	5.0			
Others & remains				
<b>Epibenthos</b>				
Polychaete	70.0		19.0	
<u>Glycinidae solitaria</u>	10.0			
<u>Spionids sp.</u>	15.0			
Amphipoda	35.0		30.0	
<u>Gammarus sp.</u>	15.0			
<u>Others &amp; remains</u>	40.0			
Unidentified Remains & Organic Matters	50.0		26.0	16.0

Table 14. Stomach contents of spot, Leiostomus xanthurus from different estuarine areas along U.S. Atlantic and Gulf of Mexico coasts.

Author	Chao	Roelofs, 1954	Stickney, et. al. 1975
Locality	York River, Virginia	North Carolina	Savannah River & Ossabow Sound, Georgia
Period	June to Aug. 1973	All Season, 1950?	May 1972 to July 1973
Source	Original	Table 1	Table 1
Number of specimens	77	73	126
Empty stomachs	4	0	7
Length of specimens	73-202 mm TL	60 to 140 mm TL	50 to 149 mm SL
Quantitative method	% of occurrence	% of occurrence	% of occurrence
Fish & Remains	8.2	6.8	5.0
Macrozooplankton			
Mysidace	8.2	4.1	
<i>Neomysis americana</i>	27.4	7.4	
Isopoda	2.7		
Decapoda (or shrimp)	1.4	5.5	0.8
Insecta	2.7	1.4	1.7
Others & Remains	1.4		0.8
Microzooplankton			
Copepoda	21.9	100	
Cyclopoid	19.2		33.1
Calanoid	13.7		88.4
Harpacticoid	20.5		5.8
Ostracoda		2.7	
Others & Remains	1.4	6.8	7.4

Table 14. (continued, stomach contents of Leiostomus xanthurus)

Author	Chao	Roelofs, 1954	Stickney, et. al. 1975
Epibenthos			
Annelids (or polychaete)	56.6	32.9	11.6
<u>Neris succinea</u>	27.4		9.1
<u>Glycinde solitaria</u>	37.0		
<u>Nephtys</u> sp.	11.0		
<u>Phyllodoce</u>	6.8		
Spionid	6.8		
Oligochaete	4.1		1.7
Cumacea	21.9		13.2
Amphipoda	24.7		
<u>Gammarus</u> sp.	12.3		11.6
Crabs	1.4		
Cnidaria	9.6		
Others & Remains	5.5	13.7	2.5
Infauna			
<u>Pectinaria gouldii</u>	53.4		
<u>Ampharetid</u>	19.2		
Gastropoda	20.5		0.8
Pelecypoda	27.4	11.0	
Nematoda	34.2	71.2	
Others & Remains	5.5		5.0
Unidentified Remains & Organic Matters	42.5	23.3	35.6

Table 14. (continued, stomach contents of Leiostomus xanthurus)

Author	Welsh & Breder, 1945	Townsend, 1956	Parker, 1971
Locality	St. Vincent Sound, Florida	Alligator Harbor, Florida	Lake Pontcha Train, Clear Lake, Texas Louisiana
Period	April 1915	June 1955 to May 1956	July 1959 to March 1961
Source	P. 179	Table 3	Table 13
Number of specimens	50	45	22
Empty stomachs	0	9	4
Length of specimens	2.1-3.5 cm SL	16-163 mm SL	40-99 mm TL 18-99 mm TL
Quantitative method	% of volume	% of occurrence	% of occurrence
Fish & Remains		11.1	19
Macrozooplankton			14
Mysidace			8.5
Neomysis americana			16.0
Isopoda		5.5	15.0
Decapoda (or shrimp)			19.0
Insecta			14.0
Others & Remains			
Microzooplankton			
Copepoda	8.0	66.7	3.0
Cyclopoid			1.0
Calanoid			
Harpacticoid			
Ostracoda	72.0	2.8	5.0
Others & Remains	1.0		14.5
			13.0

Table 14. (continued, stomach contents of Leiostomus xanthurus)

Author	Welsh & Breder, 1945	Townsend, 1956	Louisiana	Parker, 1971	Texas
Epibenthos					
Annelids (or polychaete)	1.0		14.5	13.0	
Neris succinea					
<u>Glycinde solitaria</u>					
<u>Nephtys</u> sp.					
Phyllococid					
Spionid					
Oligochaete					
Cumacea	2.0		7.0	8.5	
Amphipoda		16.7			
<u>Gammarus</u> sp.					
Crabs					
Cnidaria	0.5	16.6	19.0	12.0	
Others & Remains					
Infauna					
<u>Pectinaria gouldii</u>					
Ampharetid			4.0	17.0	
Gastropoda			1.5	10.0	
Pelecypoda					
Nematoda		30.6			
Others & Remains					
Unidentified Remains & Organic Matters	14.0	36.1	14.5	7.0	

Fig. 1

The trawl strata, substrata, and beach seine stations (·) in the York River estuary, Virginia. Strata: A, B, C, D, E, F, G, M and P. Substrata: N. north shoal; C. channel; and S. south shoal. Substrata in Mattaponi River expressed as M-1, M-2 and M-3, in Pamunkey River as P-1, P-2 and P-3. River distances from the mouth of York River (0 km) are indicated in kilometers.

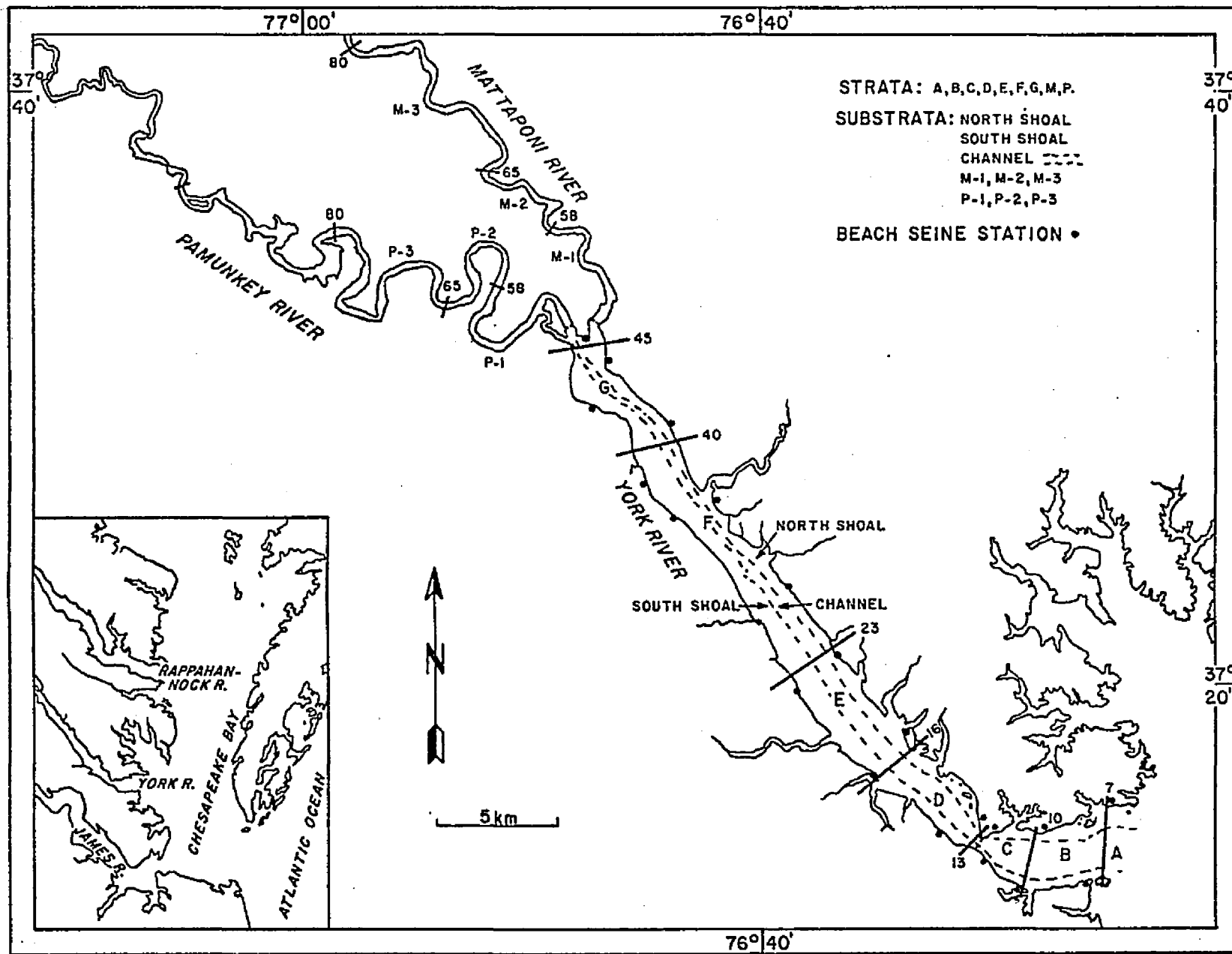
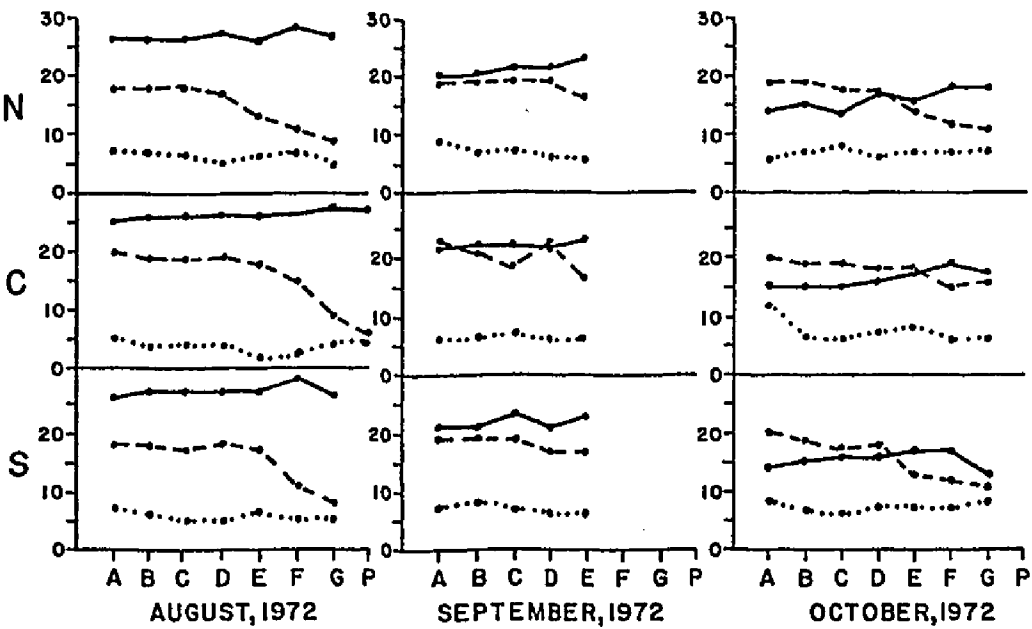
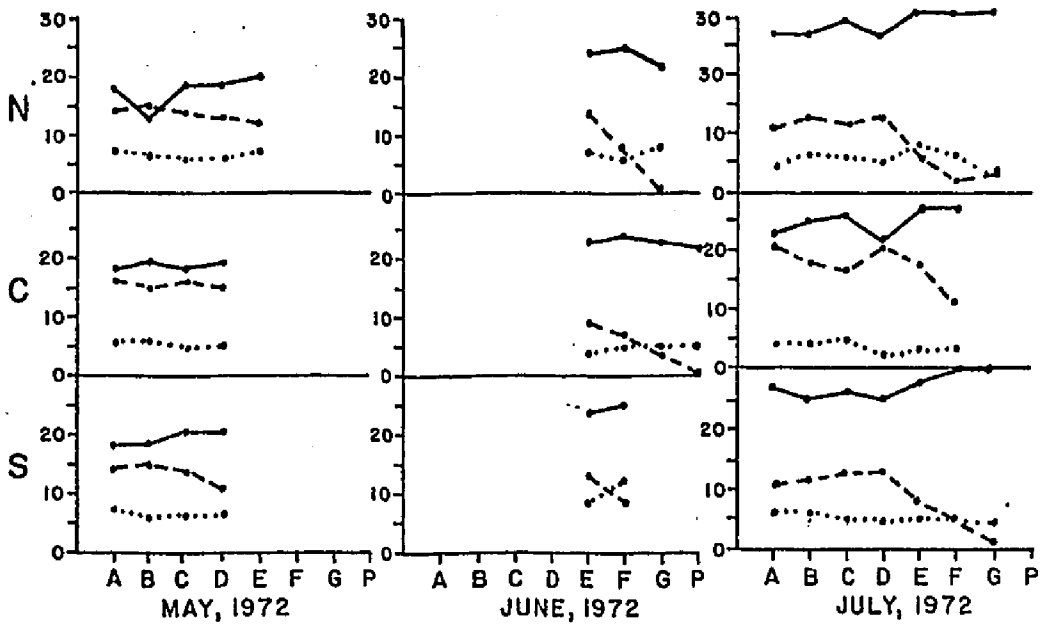


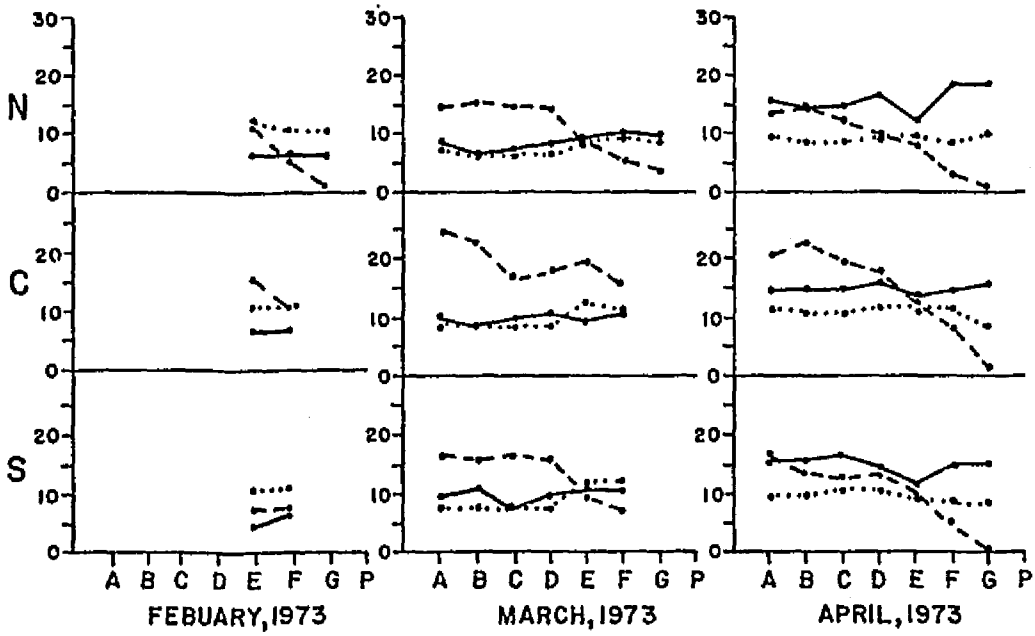
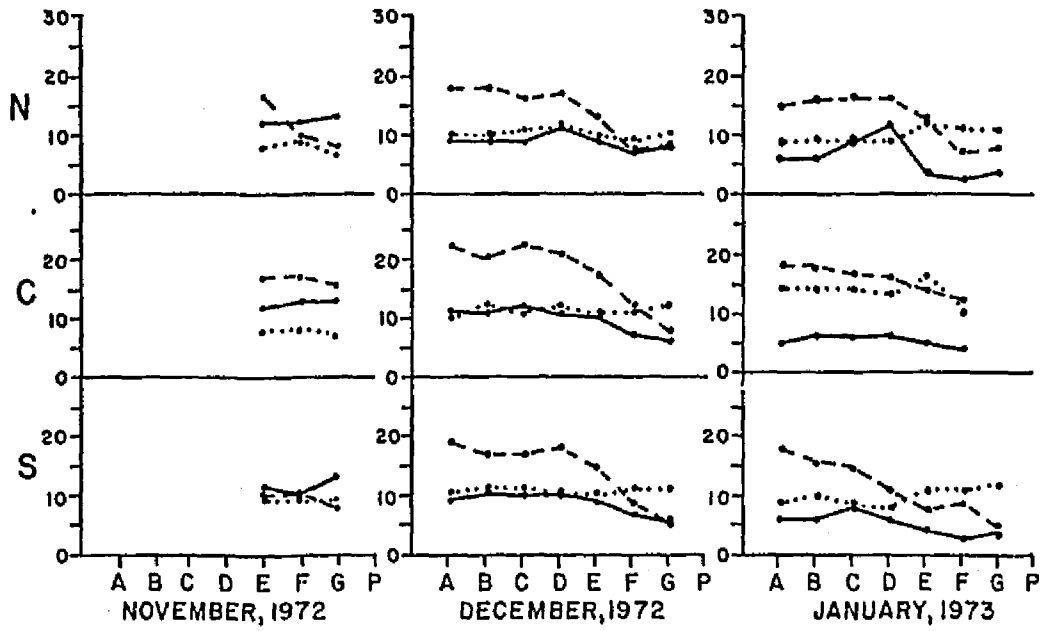


Fig. 2

Monthly means of the bottom temperature, salinity and dissolved oxygen in the York River estuary from May, 1972 to August, 1973. Strata: A to G in York River and P in Pamunkey River. Substrata: N. north shoal, C. channel and S. south shoal.



DISSOLVED OXYGEN (mg./l) .....  
 SALINITY (‰) ----  
 TEMPERATURE (°C) ———



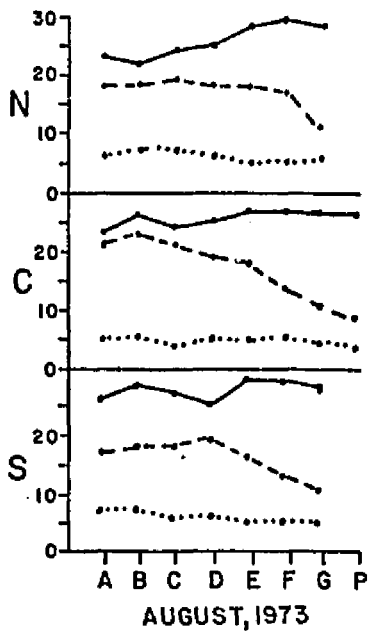
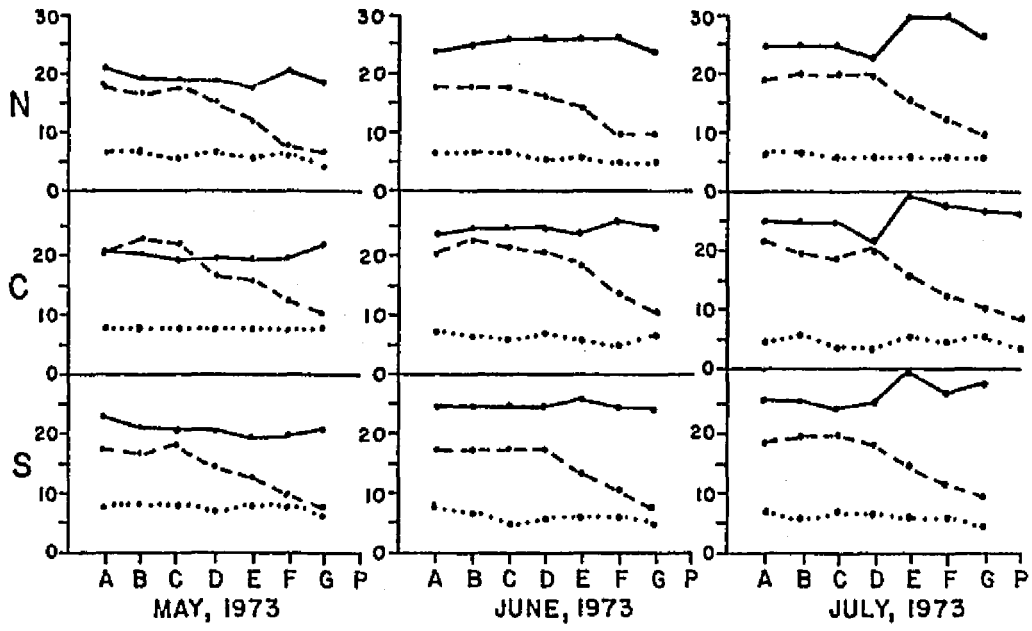
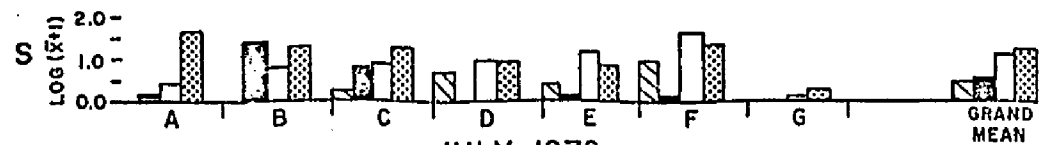
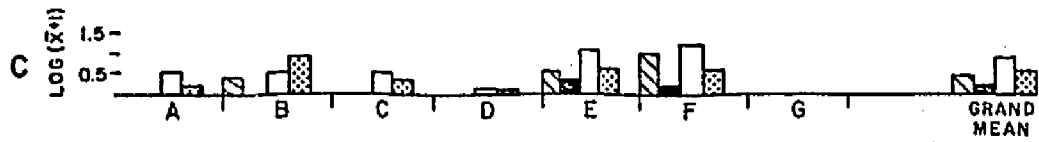
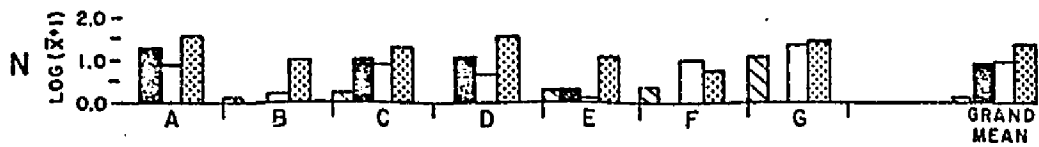


Fig. 3

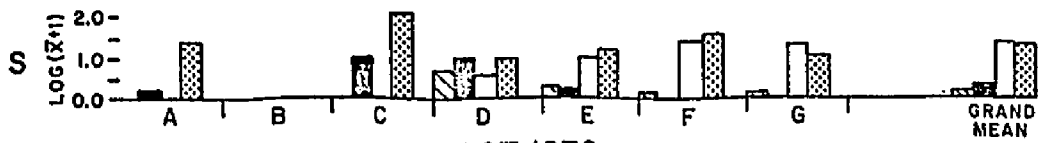
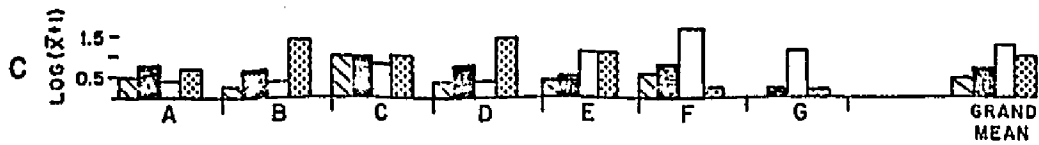
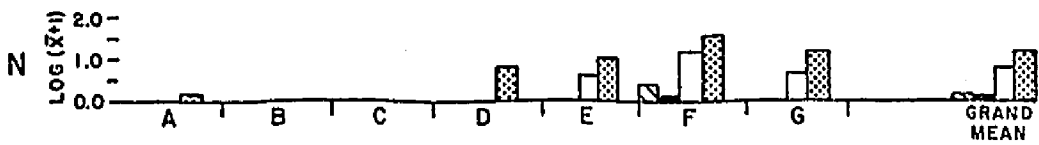
Seasonal abundance of four juvenile sciaenids with  
depth and distance upstream in the York River.

Mean numerical catch per tow expressed as  $\log(\bar{x}+1)$ .

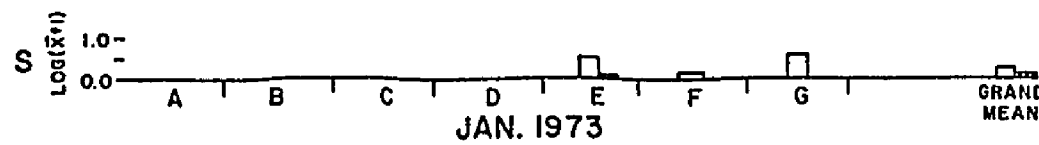
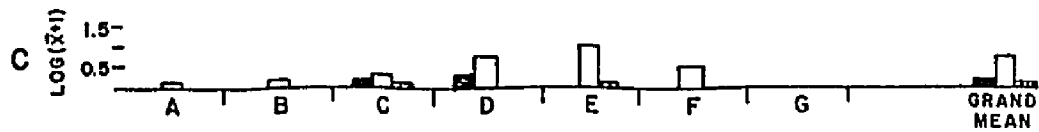
Strata: A to G; substrata: N. north shoal, C.  
channel and S. south shoal.



JULY 1972



OCT. 1972



JAN. 1973

*Cynoscion regalis*

*Micropogonias undulatus*

*Bairdiella chysoura*

*Leiostomus xanthurus*

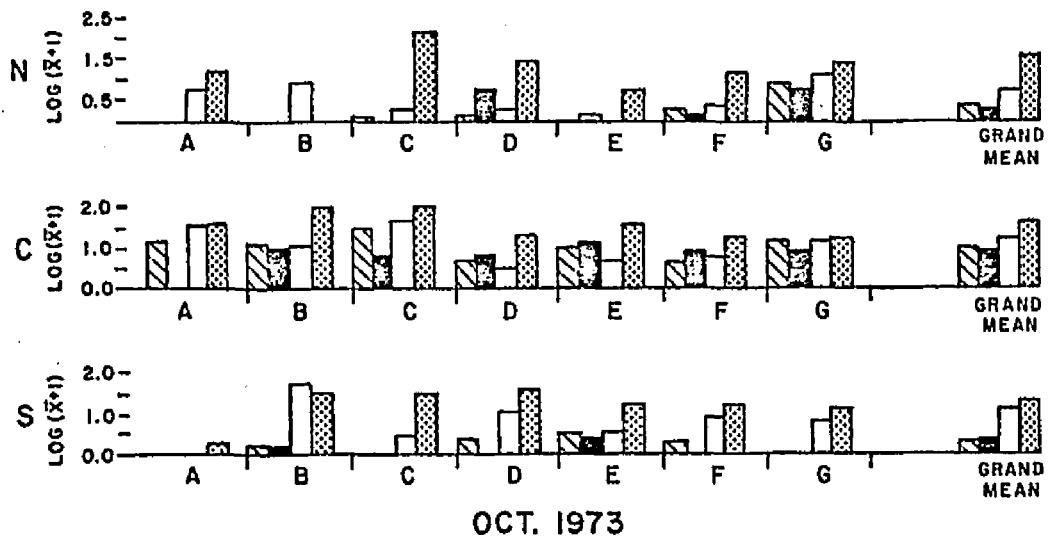
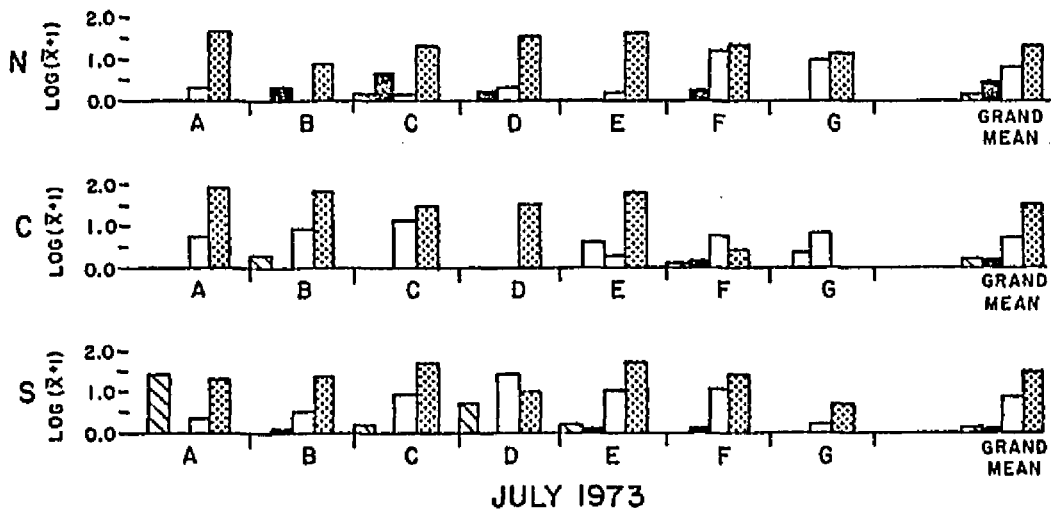
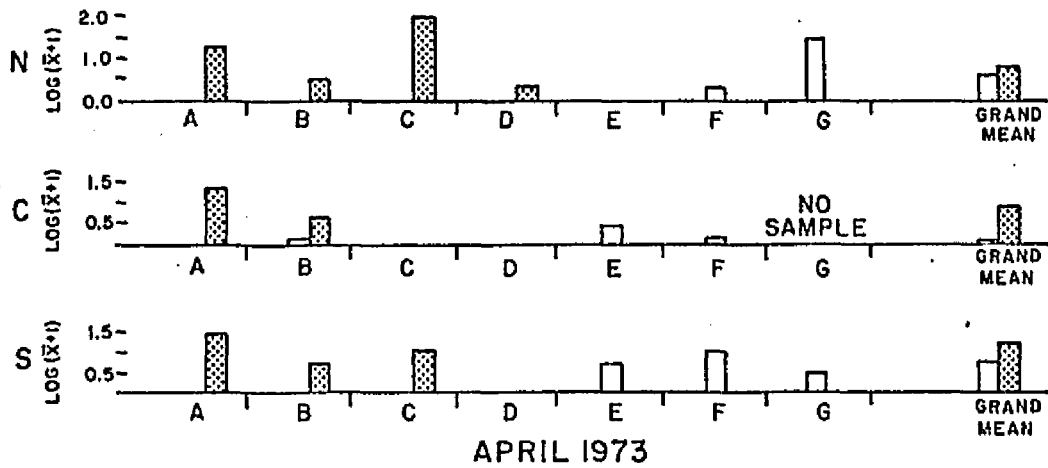


Fig. 4

Seasonal abundance of four juvenile sciaenids in  
Mattaponi and Pamunkey rivers. Mean numerical  
catch per tow expressed as  $\log(\bar{x}+1)$ . Strata: M.  
Mattaponi River, P. Pamunkey River. Substrata: 1,  
2 and 3 designated by river distance upstream.



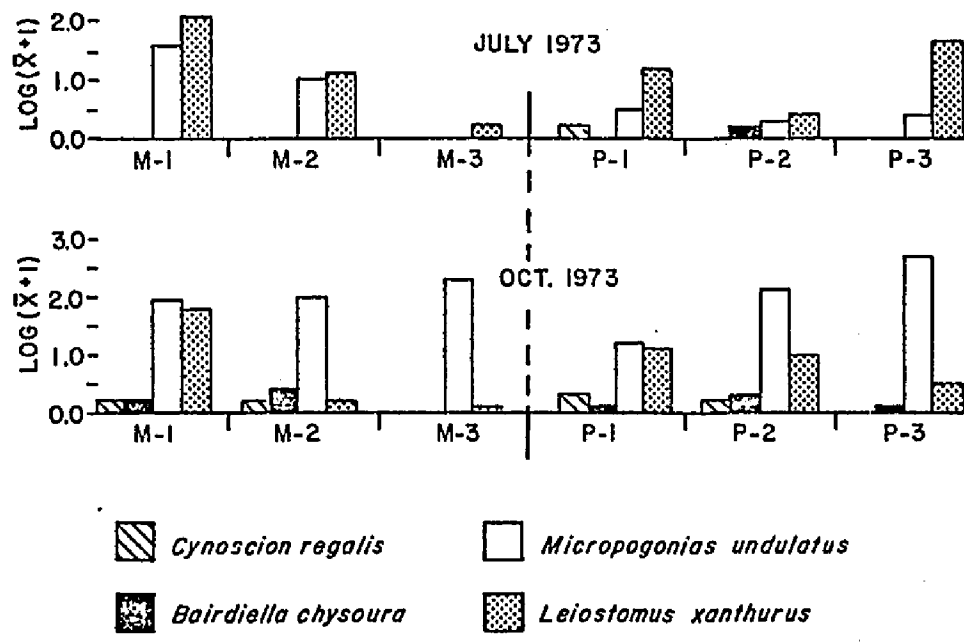
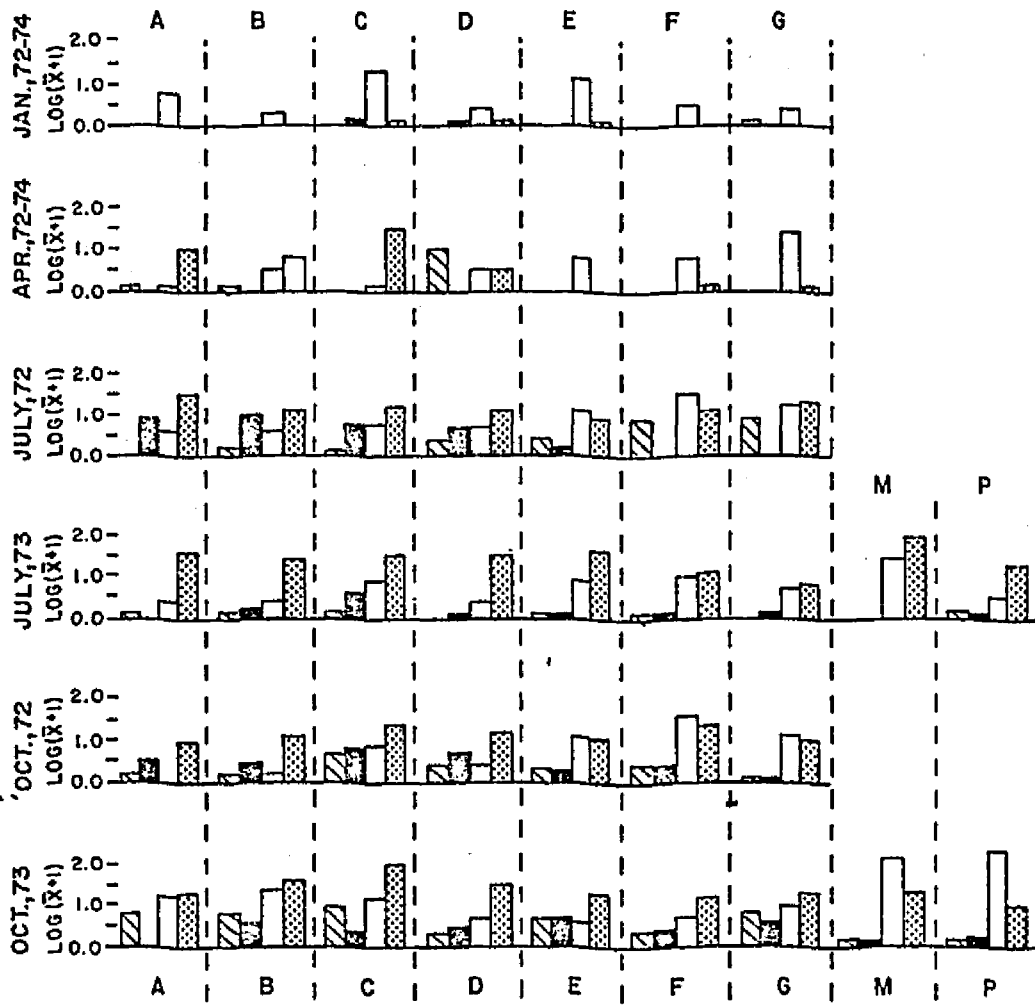


Fig. 5

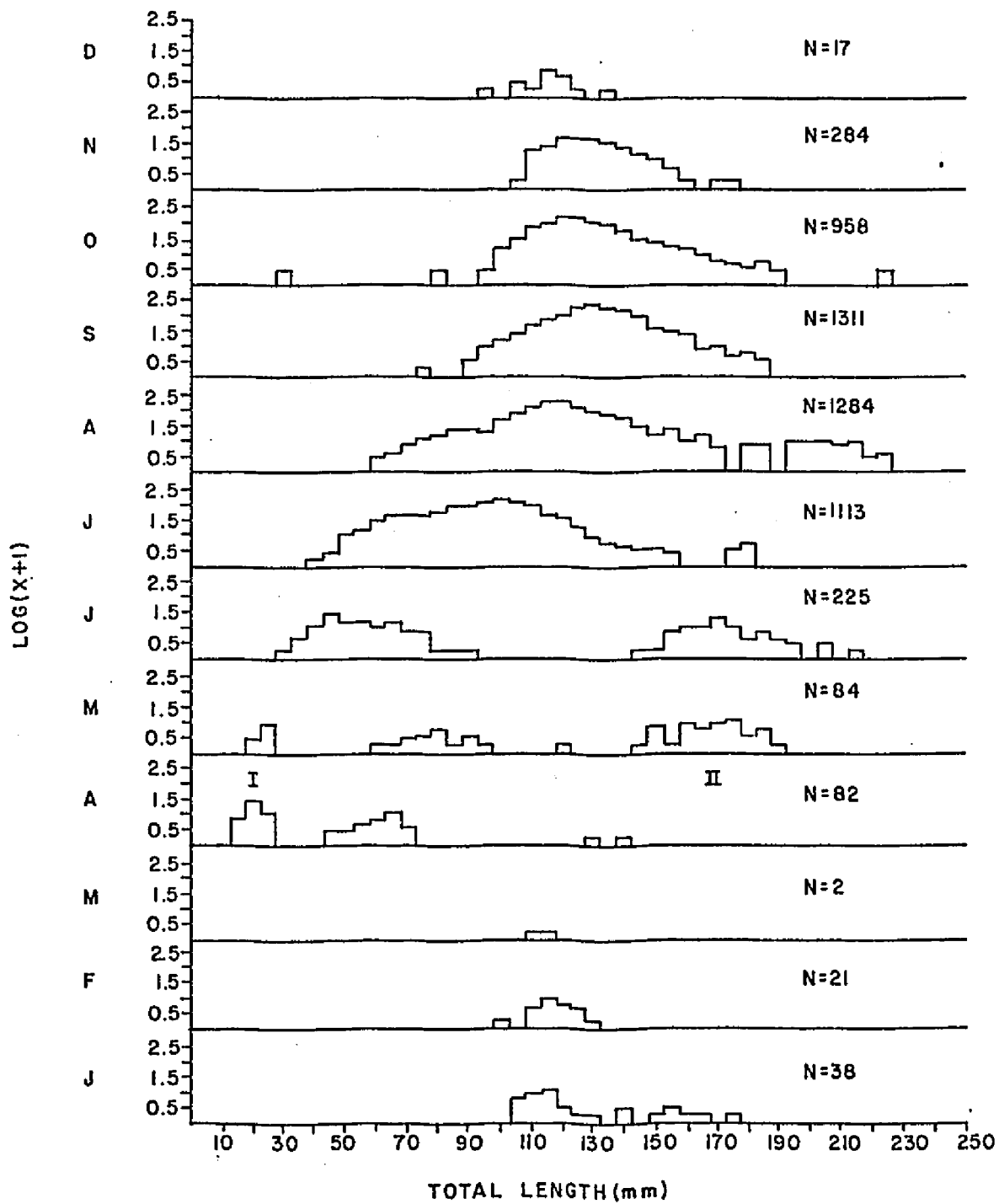
Seasonal mean abundance of four juvenile sciaenids along the salinity gradient (strata) in the York River estuary. Grand mean numerical catch of channel and shoals per tow expressed as  $\log(\bar{x}+1)$ . Strata: A to G in York River, M. Mattaponi River, P. Pamunkey River.



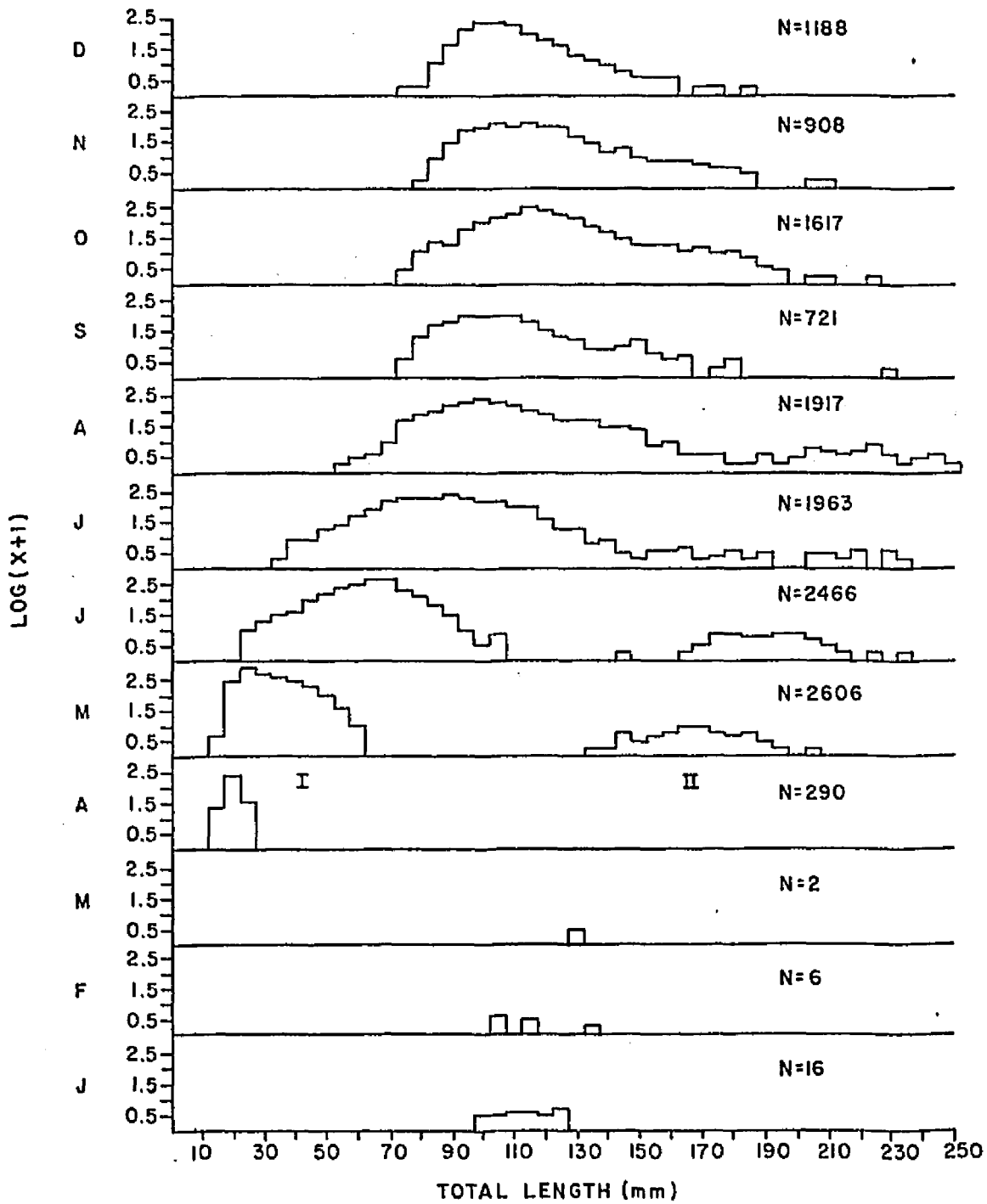
 *Cynoscion regalis*       *Micropogonias undulatus*  
 *Bairdiella chysoura*       *Leiostomus xanthurus*

Fig. 6

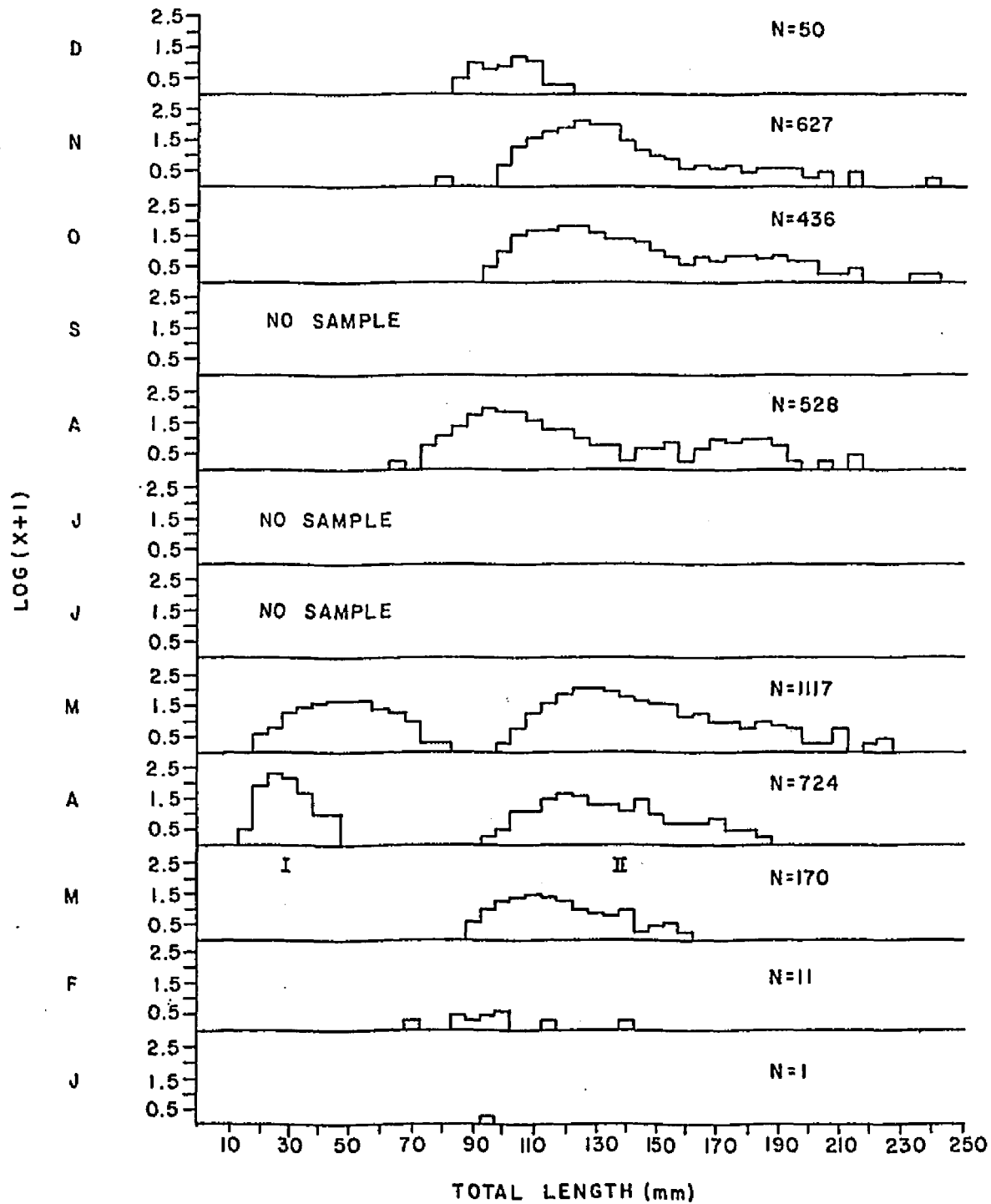
Monthly length frequency distributions of juvenile spot, *Leiostomus xanthurus*, from York River, 1972-74. Mode I, young-of-the-year; mode II, yearlings. Frequencies expressed as  $\log(x+1)$  at 5 mm increments. Only the lower portion of river (strata A to D) represented in 1974.



*Leiosomus xanthurus*, 1972 Total



*Leiostomus xanthurus*, 1973 Total

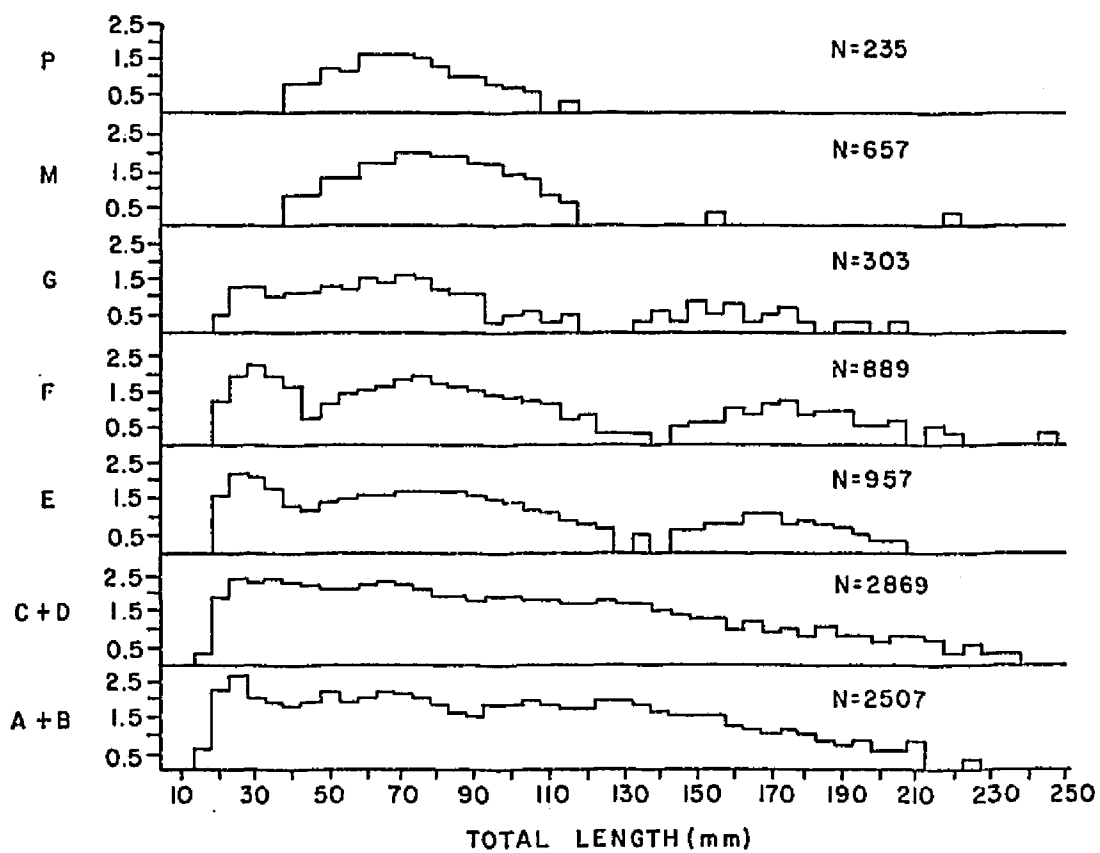


*Leioostomus xanthurus*, 1974 Total.

Fig. 7

Length frequency distributions of spot, *Leiostomus xanthurus*, by river distance (strata) upstream in the York River estuary. Pooled total, May to July 1972-74. Strata: A to G in York River, M. Mattaponi River, P. Pamunkey River. Frequencies expressed as  $\log (x+1)$  at 5 mm increments.

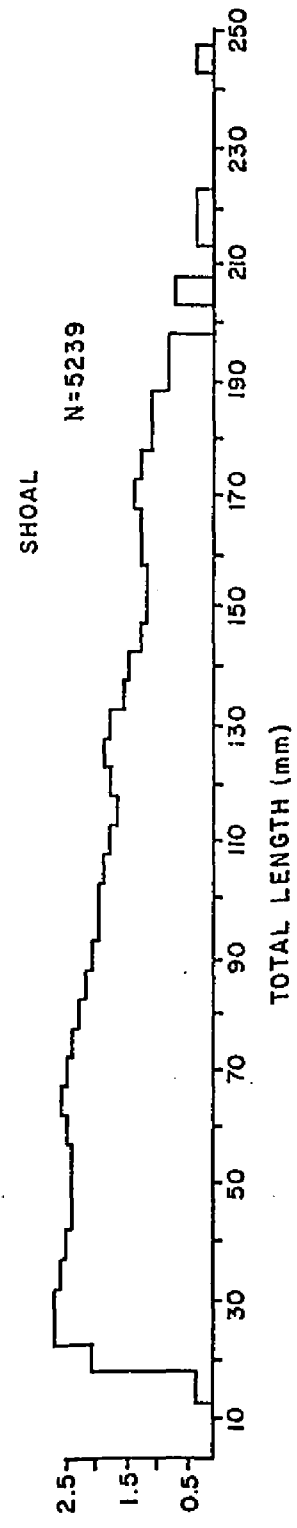
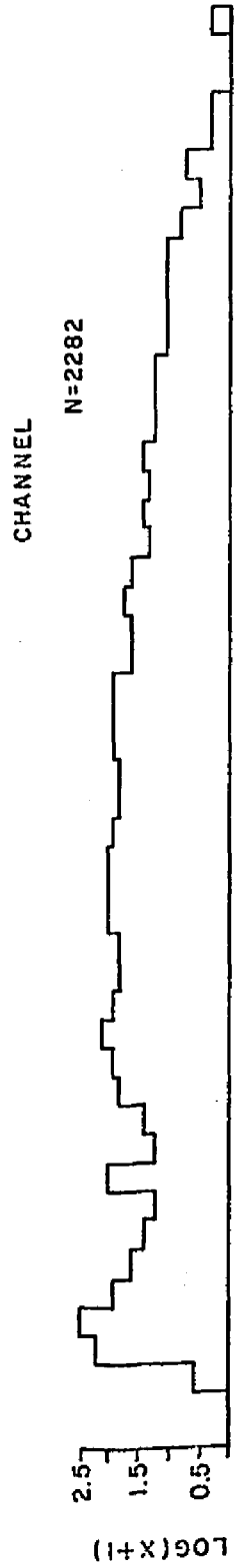
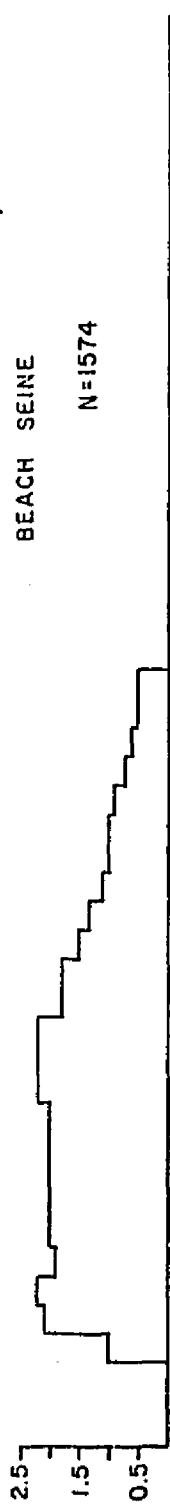




*Leiosomus xanthurus* May to July, 1972-1974

Fig. 8

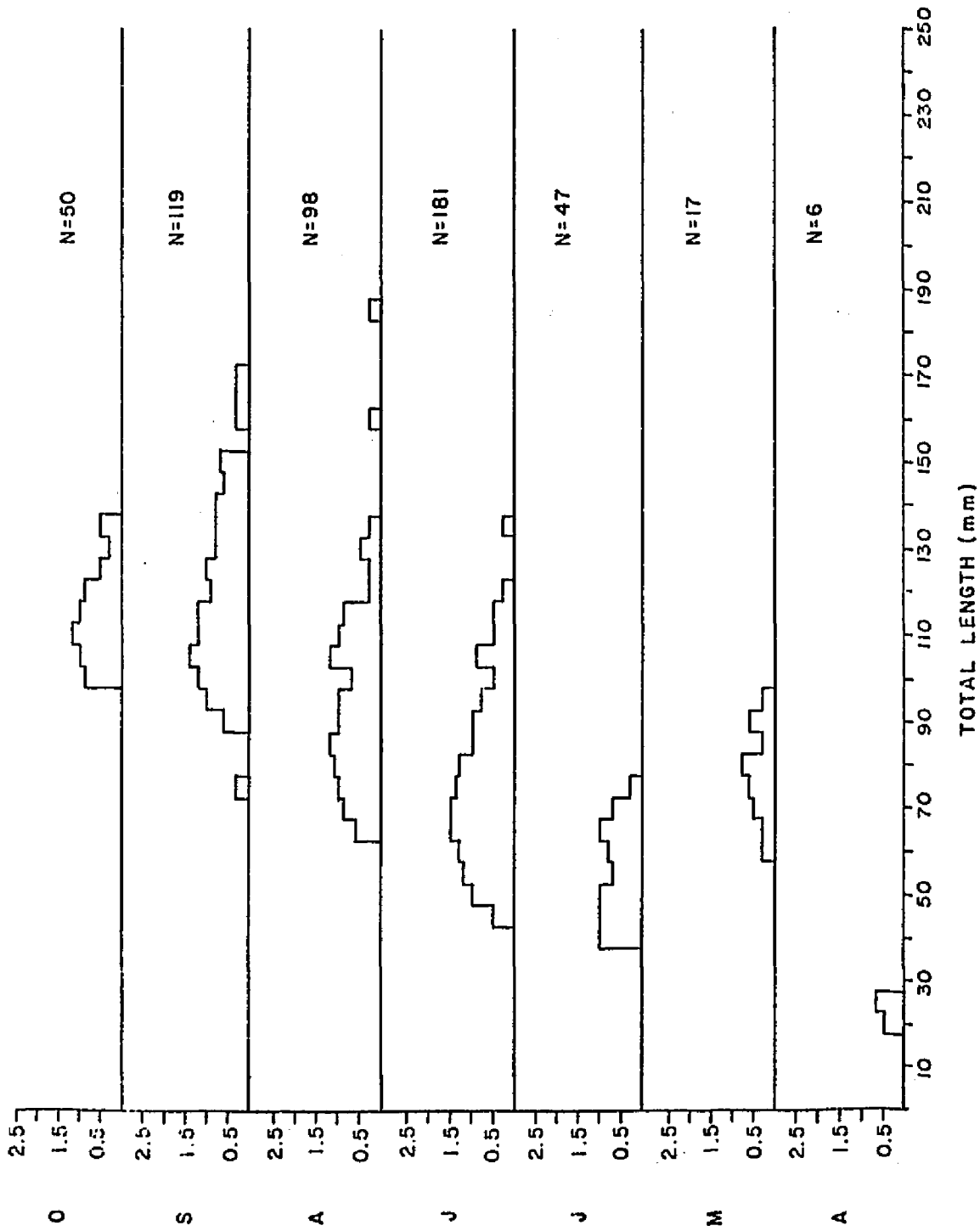
Length frequency distributions of spot, *Leiostomus xanthurus*, by depth of York River. Pooled total, May to July 1972-74. Frequencies expressed as  $\log(x+1)$  at 5 mm increments.



*Leiostomus xanthurus* May to July, 1972-1974

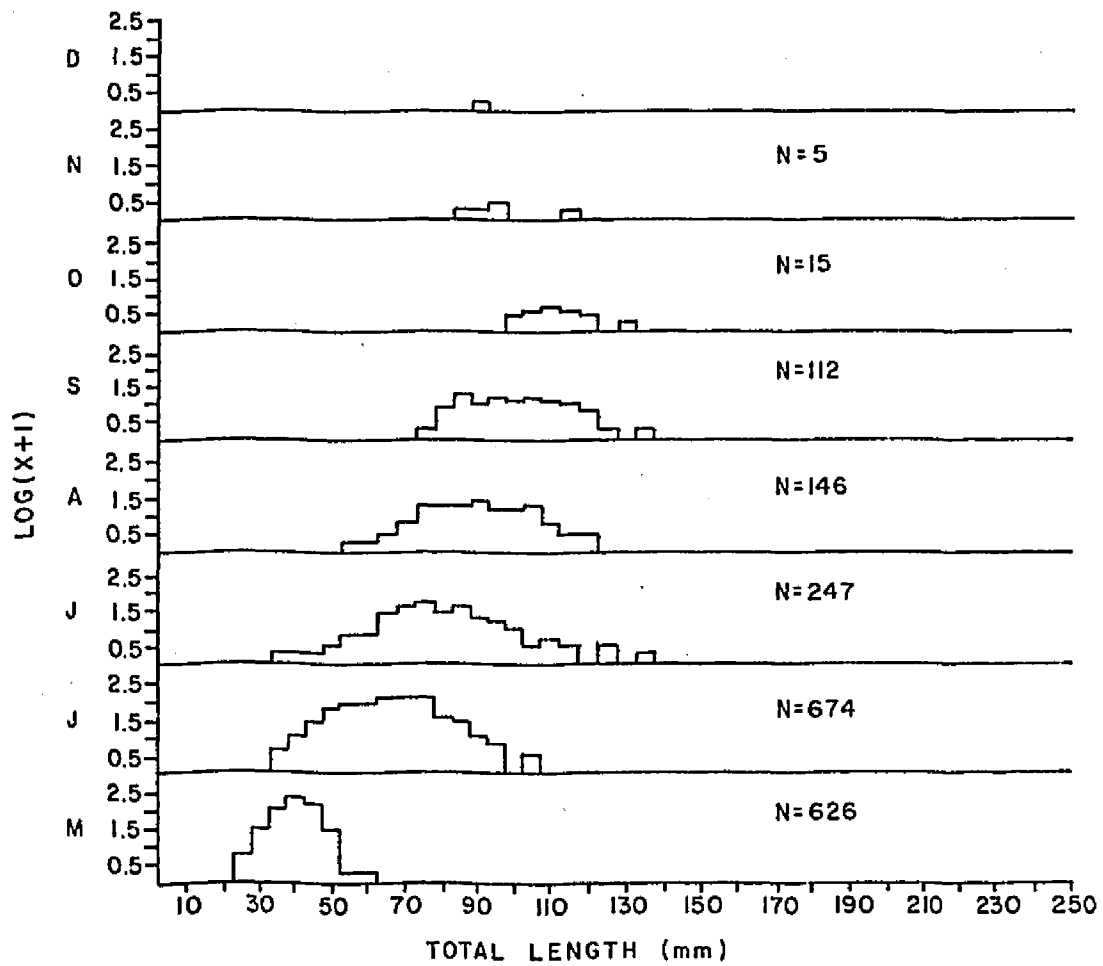
Fig. 9

Monthly length frequency distributions of spot, *Leiostomus xanthurus*, from the beach seine catches of York River, 1972-74. Frequencies expressed as  $\log (x+1)$  at 5 mm increments.

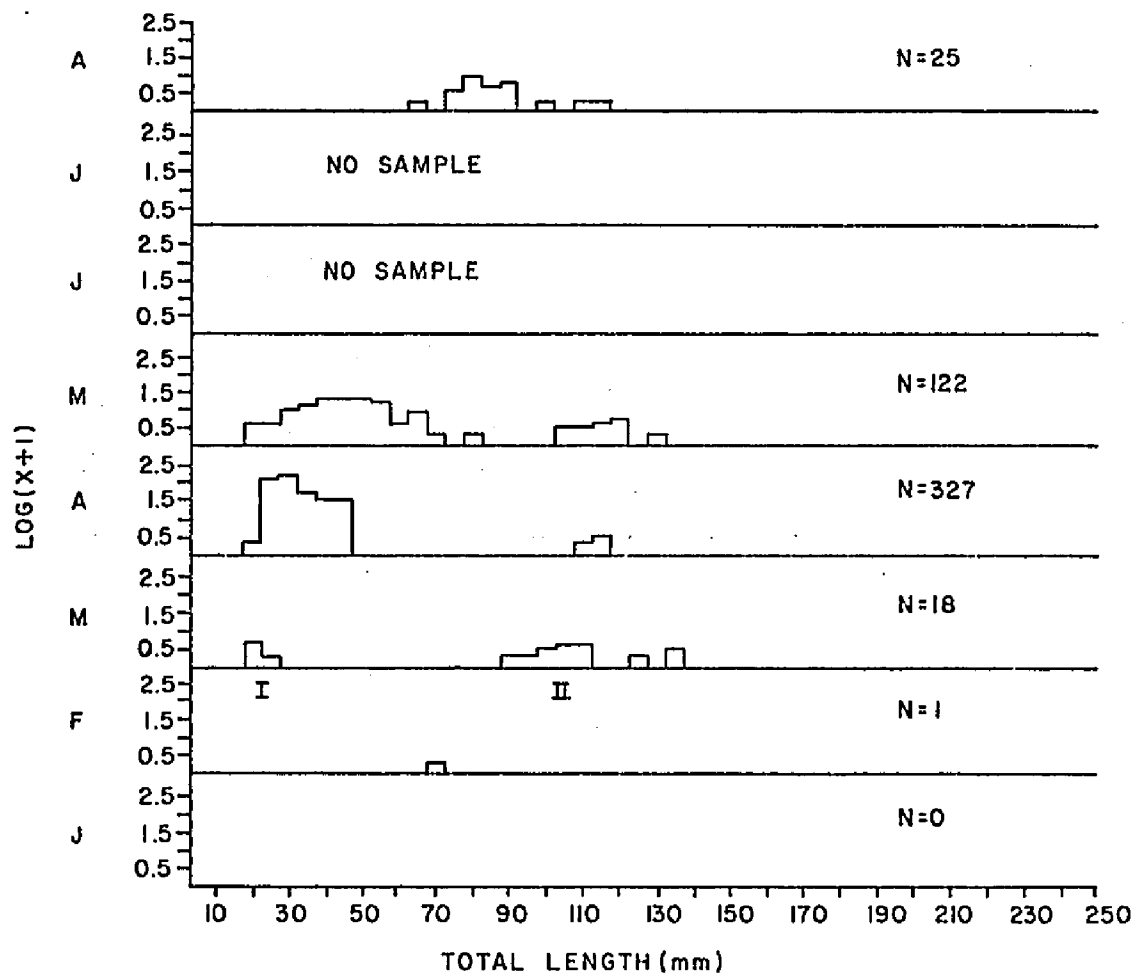


(1+X) 901

*Leiostomus xanthurus* 1972 Beach Seine



*Leiostomus xanthurus* 1973 Beach Seine

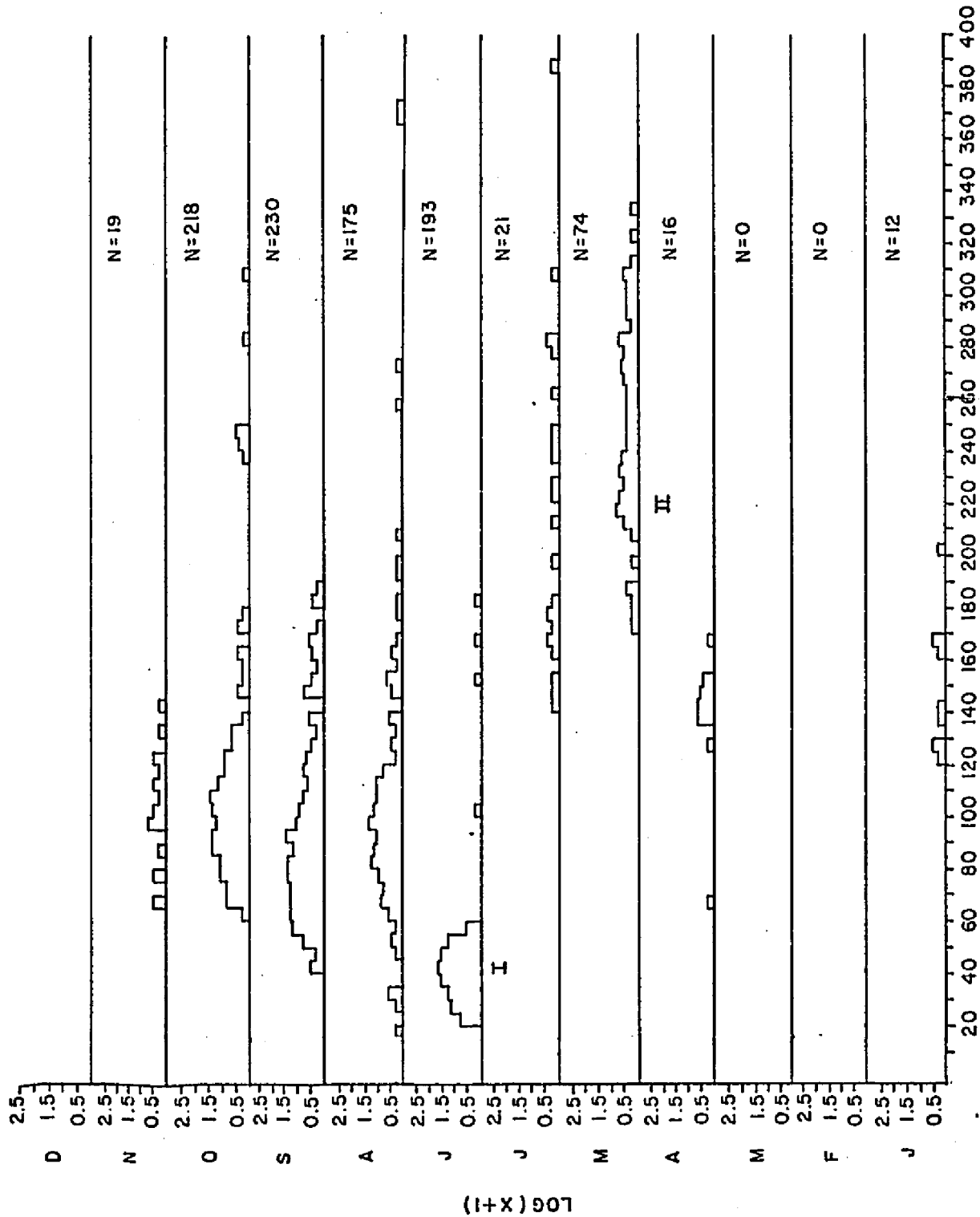


*Leioostomus xanthurus* 1974 Beach Seine

Fig. 10

Monthly length frequency distributions of weakfish, *Cynoscion regalis*, from York River, 1972-74. Mode I, young-of-the-year; mode II, yearlings. Frequencies expressed as  $\log (x+1)$  at 5 mm increments. Only the lower portion of river (strata A to D) represented in 1974.

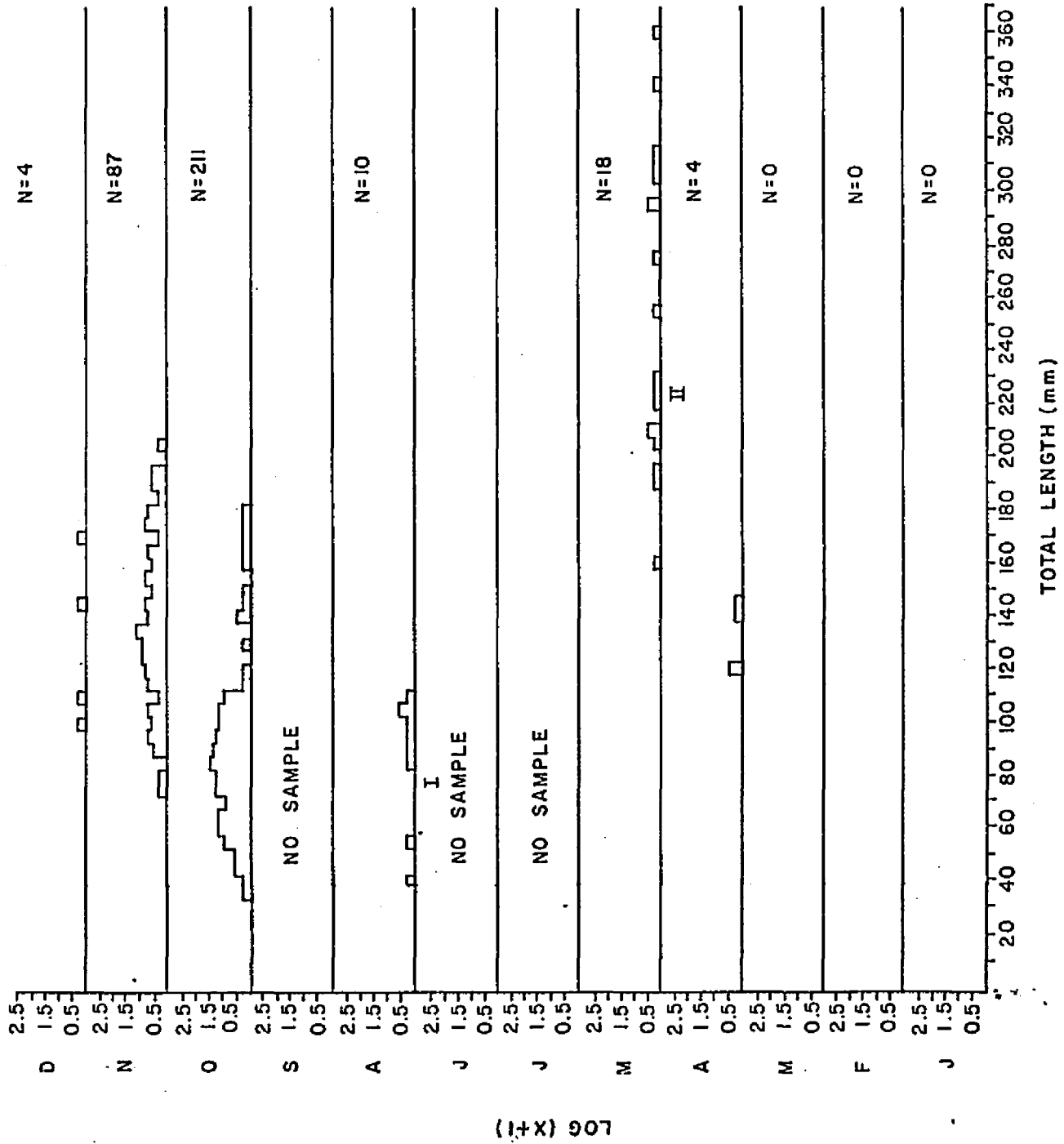




TOTAL LENGTH (mm)

*Cynoscion regalis* 1972 Total

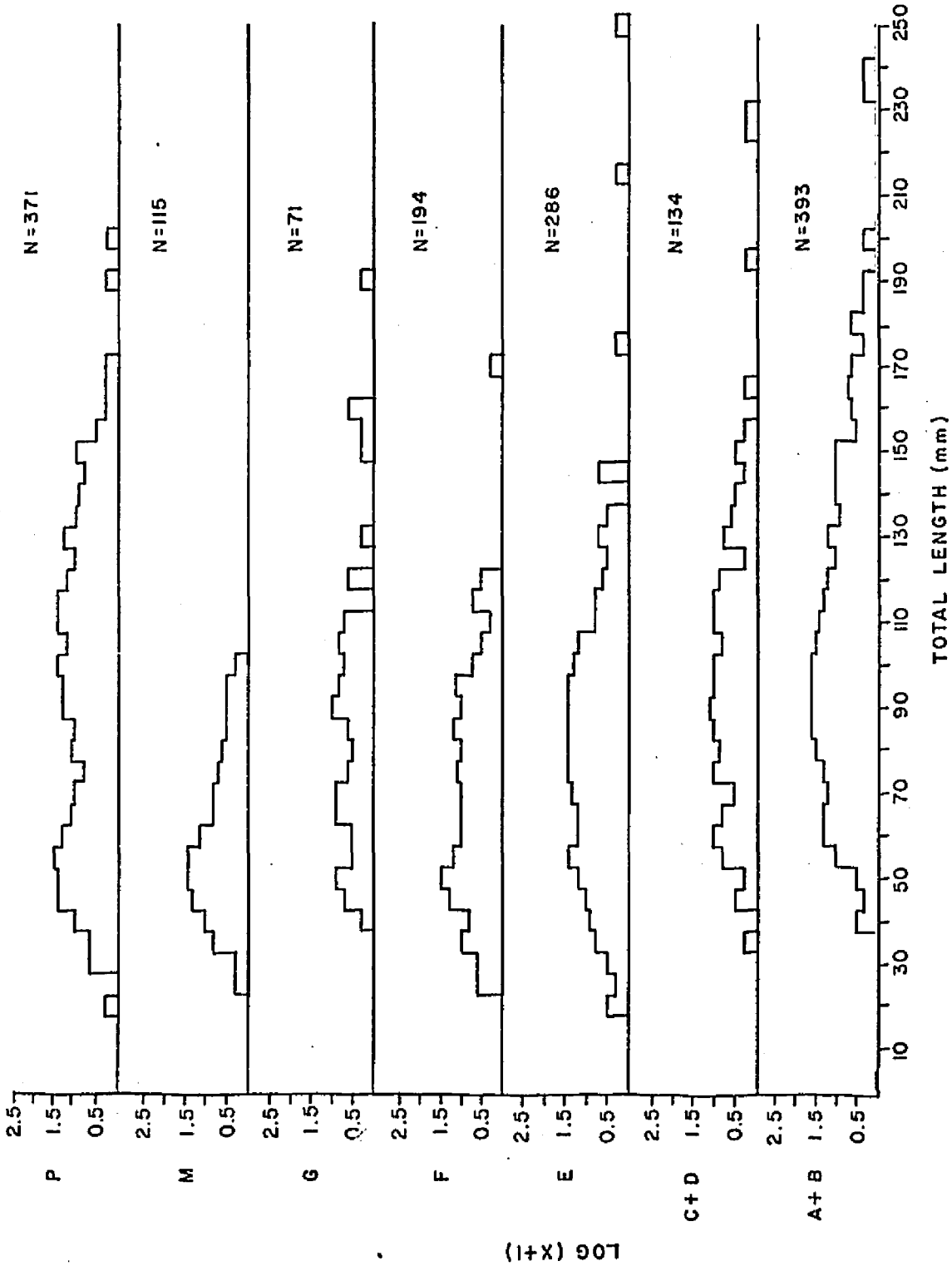




*Cynoscion regalis* 1974 Total

Fig. 11

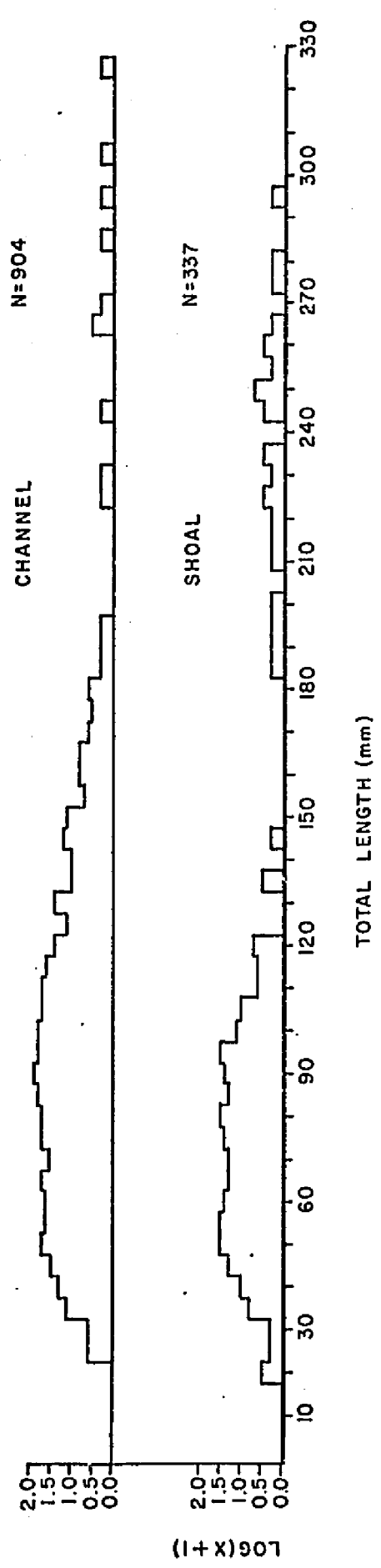
Length frequency distributions of weakfish, *Cynoscion regalis*, by river distance (strata) upstream of the York River estuary. Pooled total, August to October 1972-74. Strata: A to G in York River, M. Mattaponi River, P. Pamunkey River. Frequencies expressed as  $\log(x+1)$  at 5 mm increments.



*Cynoscion regalis* August to October, 1972-1974

Fig. 12

Length frequency distributions of weakfish, *Cynoscion regalis*, by depth of York River. Pooled total, August to October, 1972-74. Frequencies expressed as  $\log (x+1)$  at 5 mm increments.

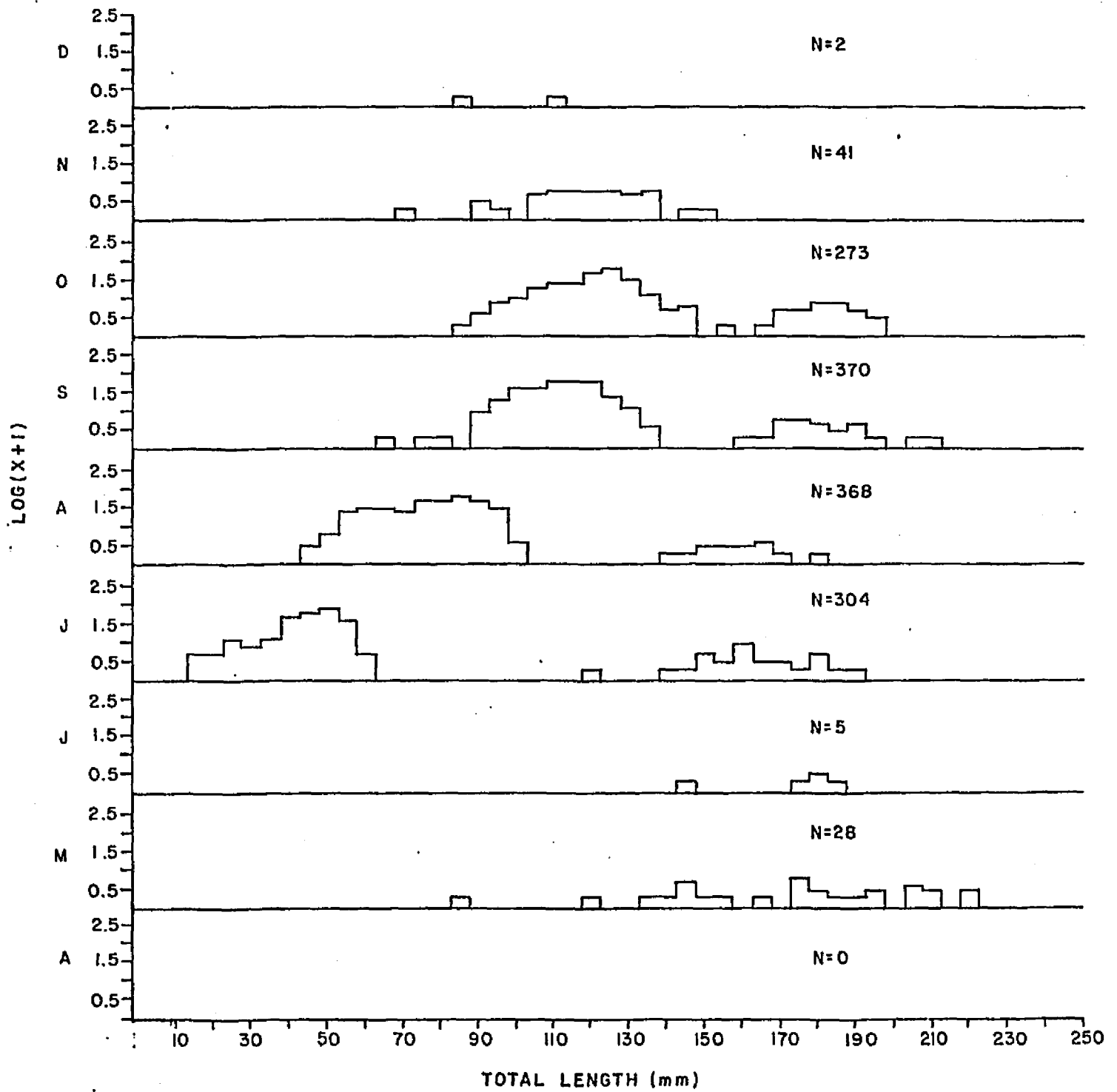


*Cynoscion regalis* August to October, 1972-1974

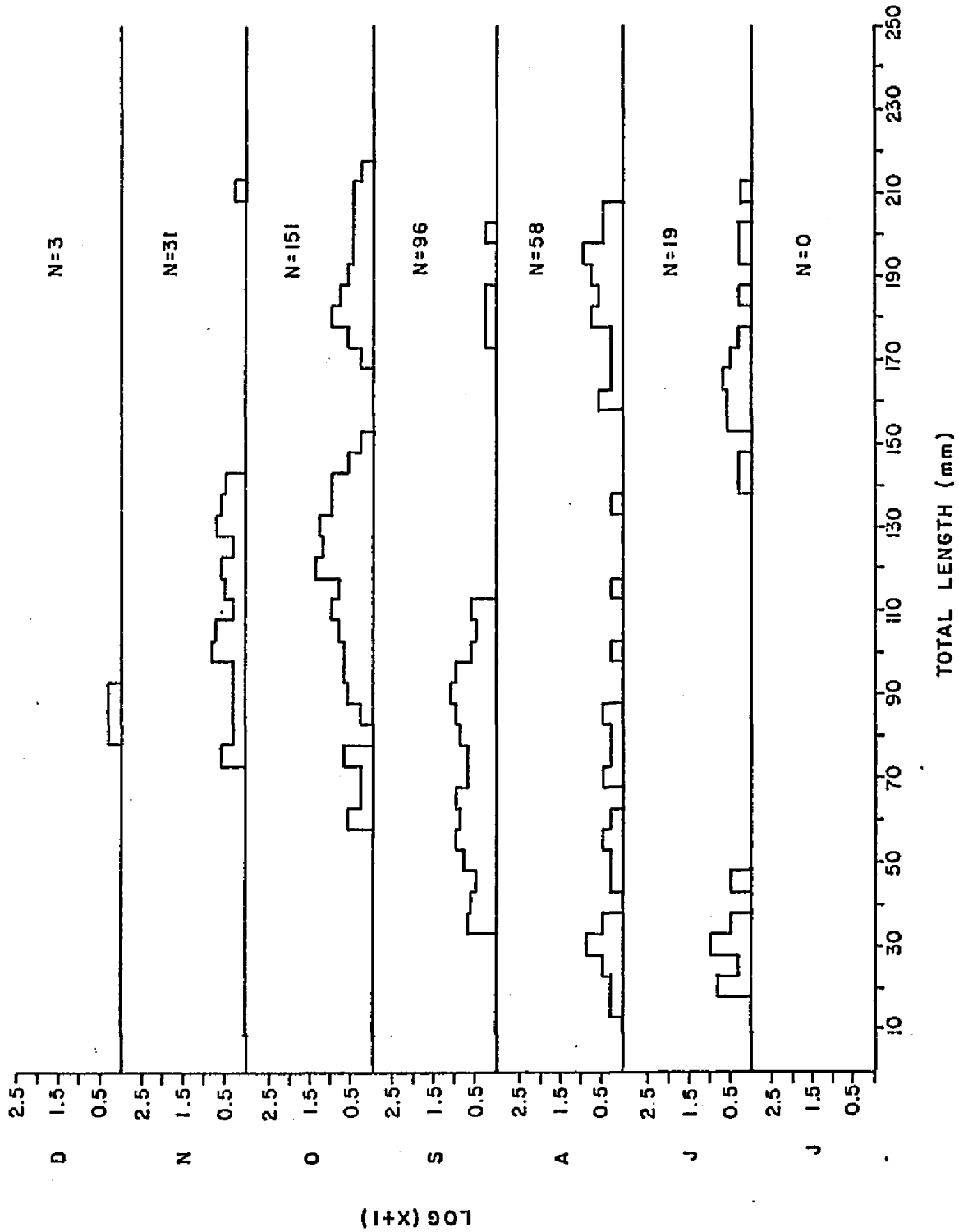
Fig. 13

Monthly length frequency distributions of silver perch, *Bairdiella chrysoura* from York River, 1972-74. Frequencies expressed as  $\log(x+1)$  at 5 mm increments. Only the lower portion of river (strata A to D) represented in 1974.

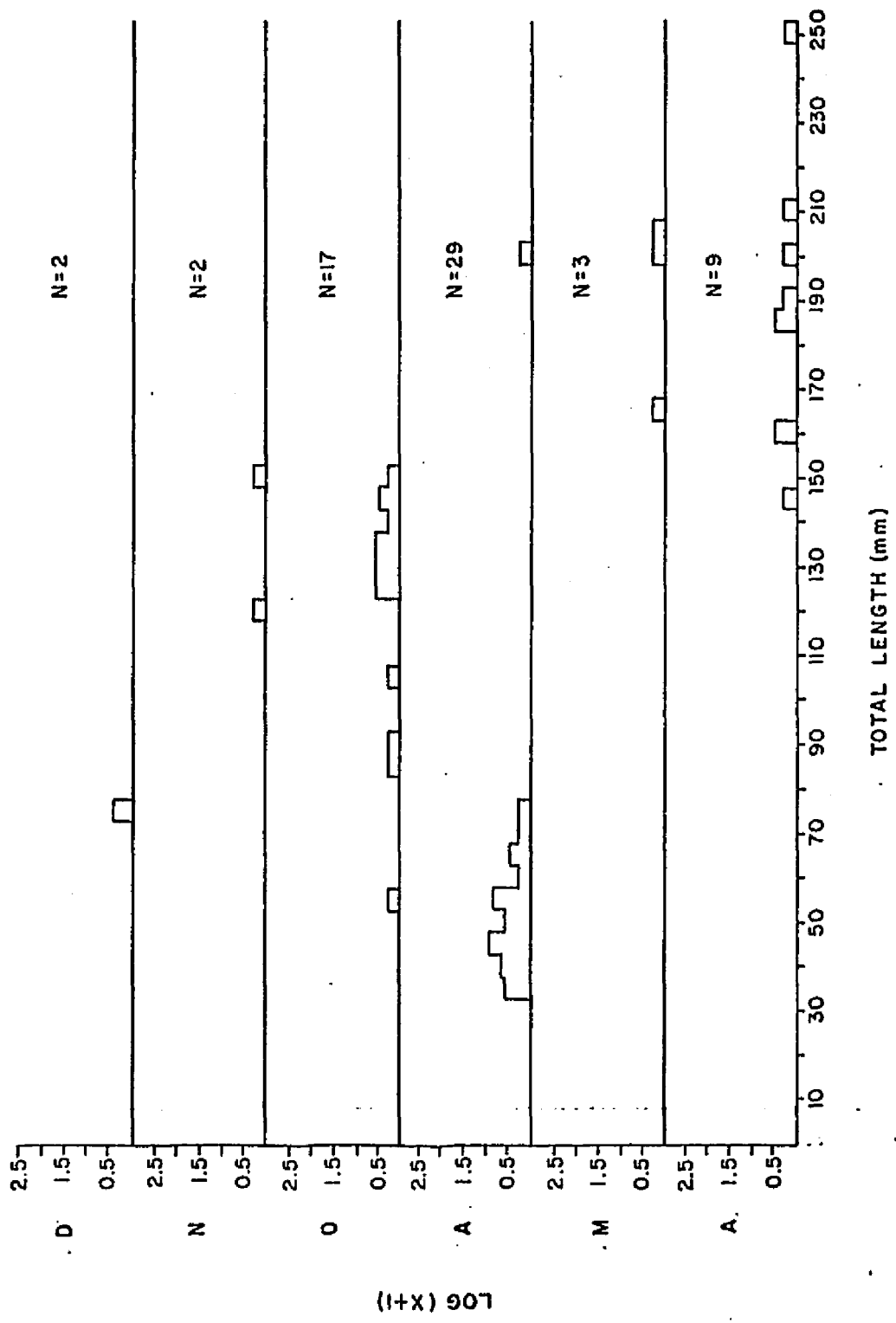




*Bairdiella chrysoura* 1972 Total



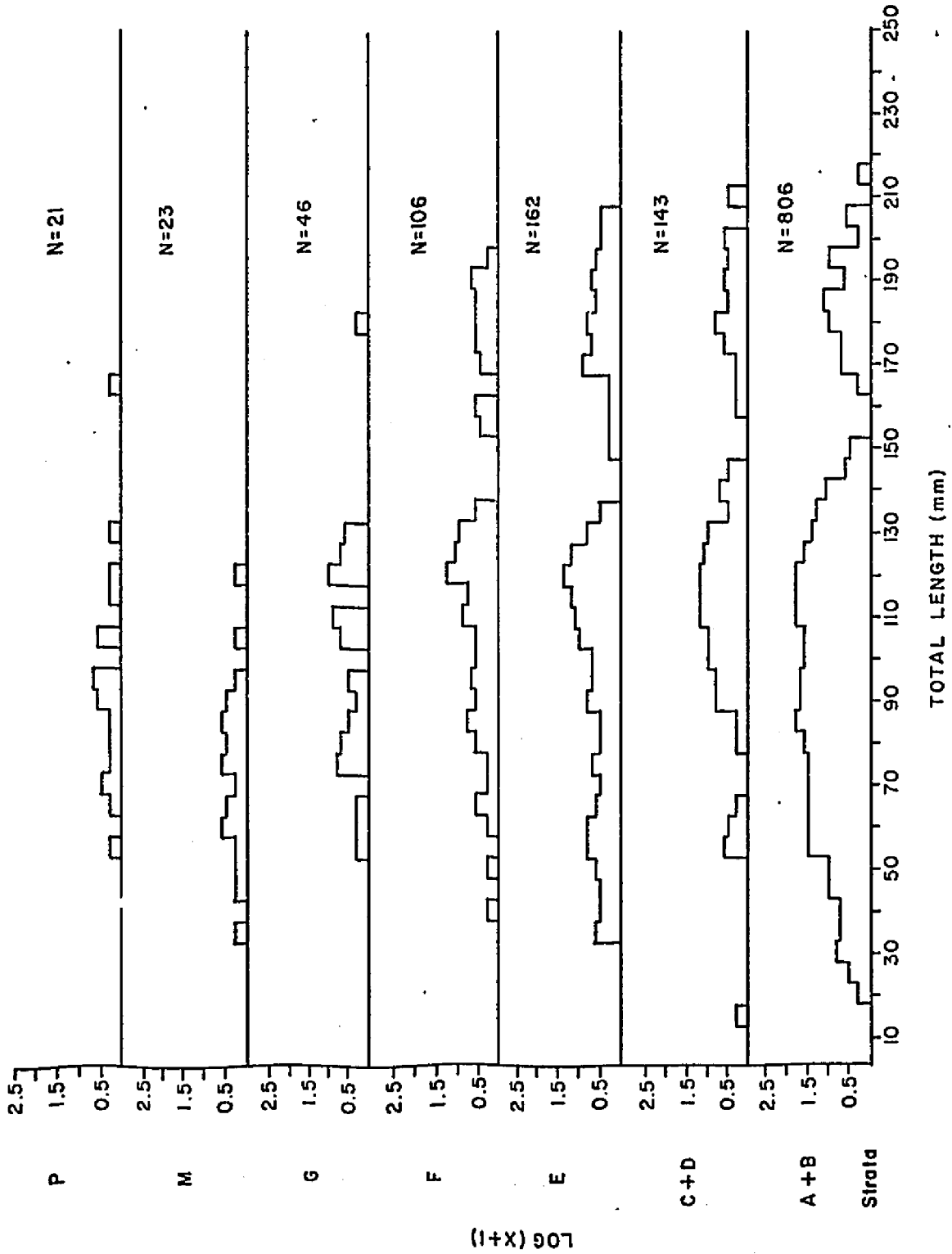
*Bairdiella chrysoura* 1973 Total



*Bairdiella chrysourea* 1974 Total

Fig. 14

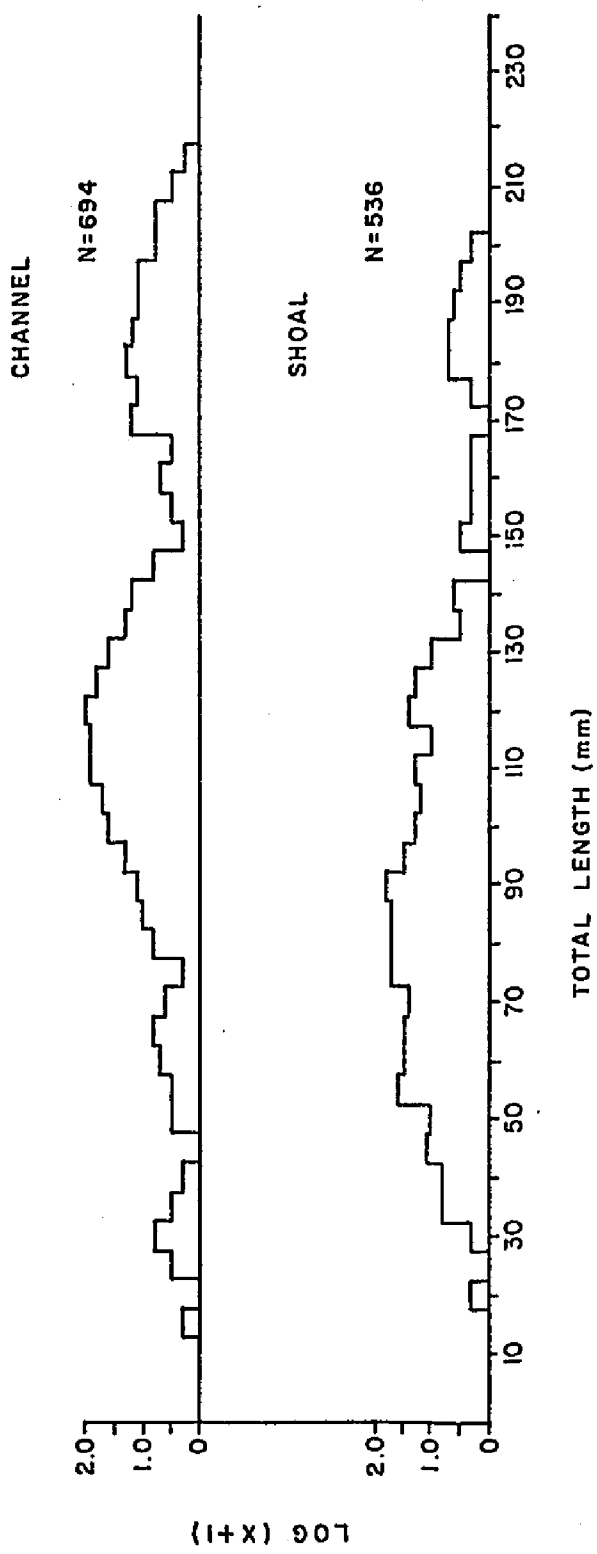
Length frequency distributions of silver perch, *Bairdiella chrysoura* by river distance (strata) upstream of the York River estuary. Pooled total, August to October, 1972-74. Strata: A to G in York River, M. Mattaponi River, P. Pamunkey River. Frequencies expressed as  $\log(x+1)$  at 5 mm increments.



*Bairdiella chrysoura* August to October, 1972-1974

Fig. 15

Length frequency distributions of silver perch,  
*Bairdiella chrysoura* by depth of York River.  
Pooled total, August to October, 1972-74.  
Frequencies expressed as  $\log (x+1)$  at 5 mm  
increments.

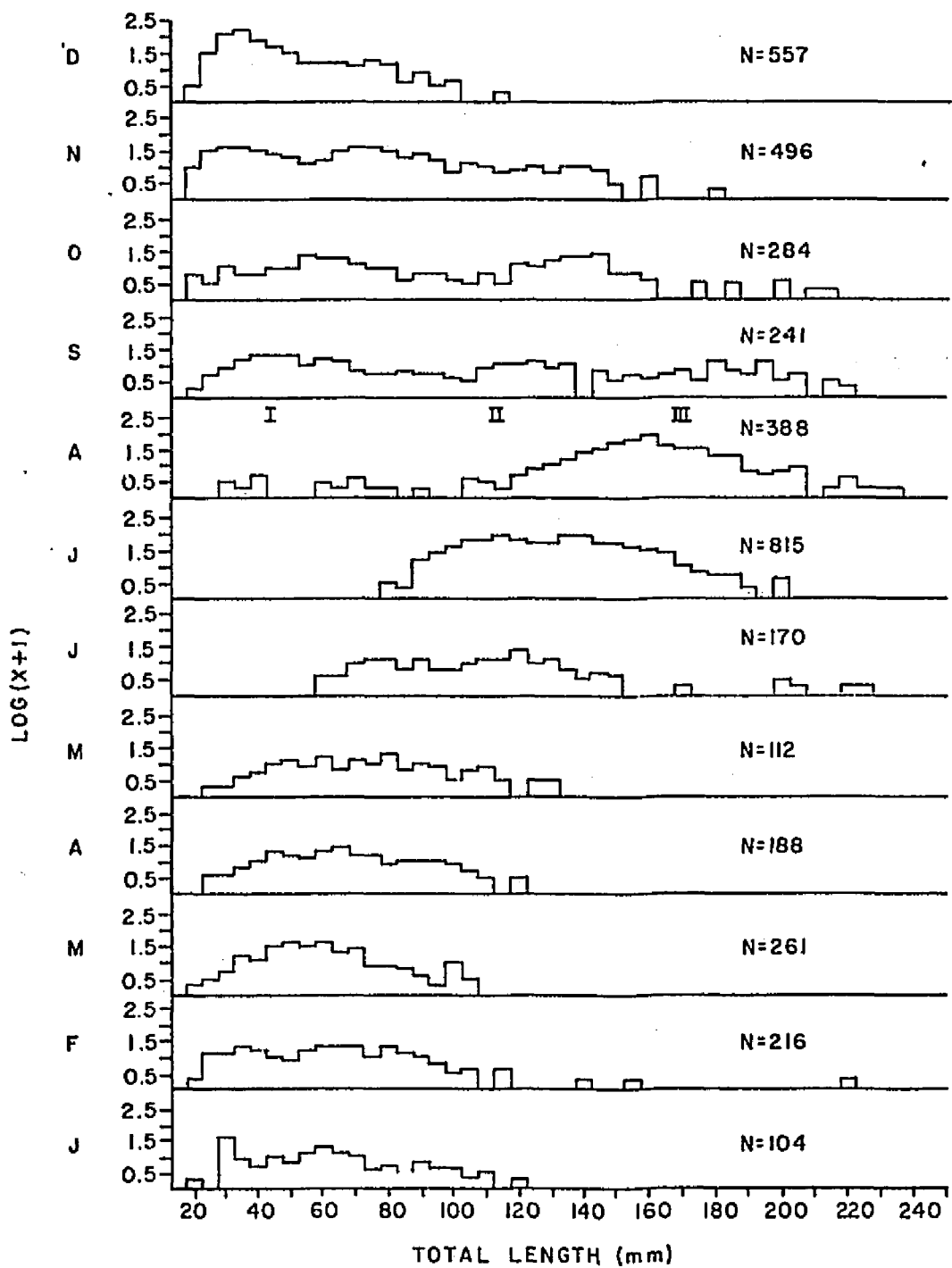


*Bairdiella chrysoura* August to October, 1972-1974

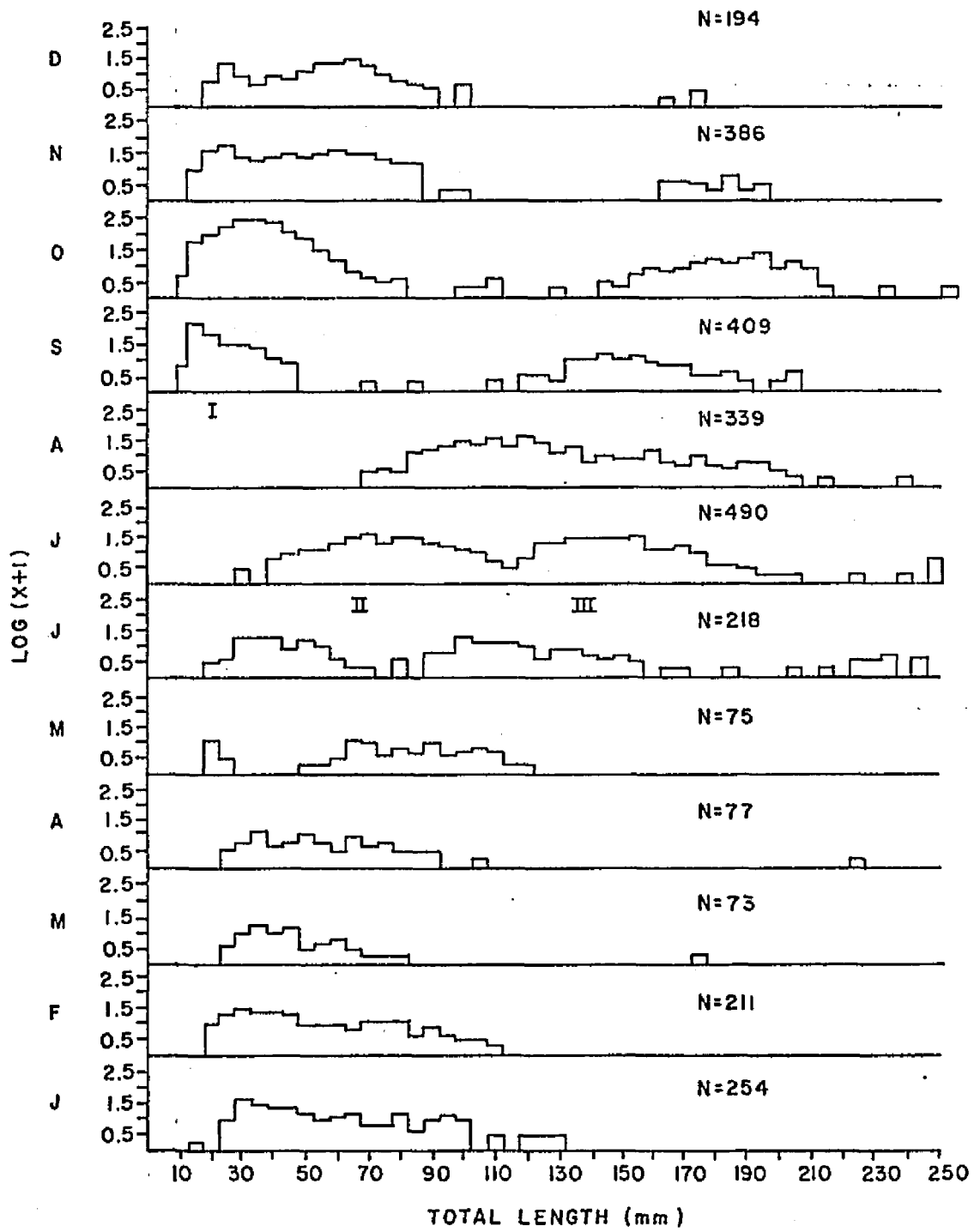
Fig. 16

Monthly length frequency distributions of croaker,  
*Micropogonias undulatus* from York River, 1972-74.  
Modes I & II, young-of-the-year; mode III, yearling.  
Frequencies expressed as  $\log(x+1)$  at 5 mm  
increments. Only the lower portion of river (strata  
A to D) represented in 1974.

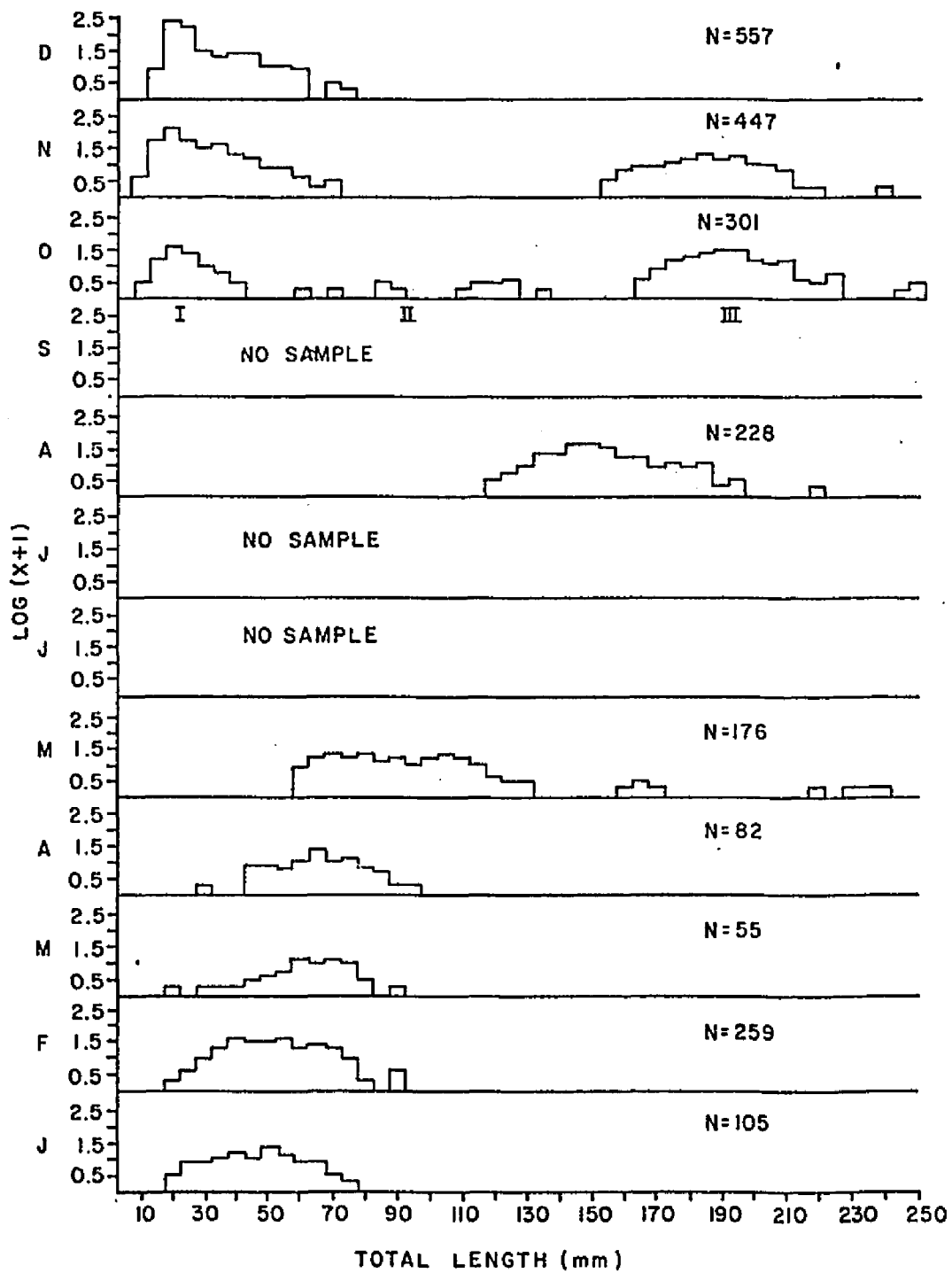




*Micropogonias undulatus* 1972 Total



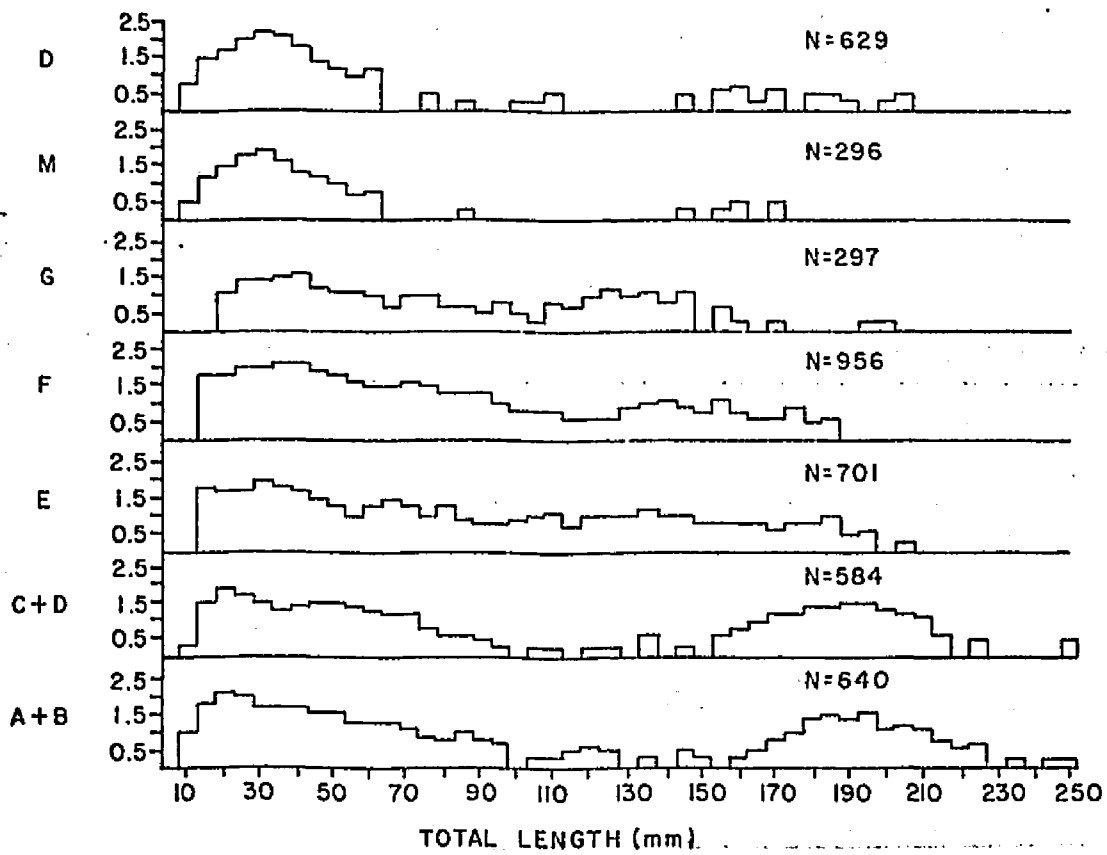
*Micropogonias undulatus* 1973 Total



*Micropogonias undulatus* 1974 Total

Fig. 17

Length frequency distributions of croaker, *Micropogonias undulatus* by river distance (strata) upstream of the York River estuary. Pooled total, September to November, 1972-74. Strata: A to G in York River, M. Mattaponi River, P. Pamunkey River. Frequencies expressed as  $\log(x+1)$  at 5 mm increments.



*Micropogonias undulatus* September to November, 1972-1974

Fig. 18

Length frequency distributions of croaker,  
*Micropogonias undulatus*, by depth of York River.  
Pooled total, September to November, 1972-74.  
Frequencies expressed as  $\log (x+1)$  at 5 mm  
increments.

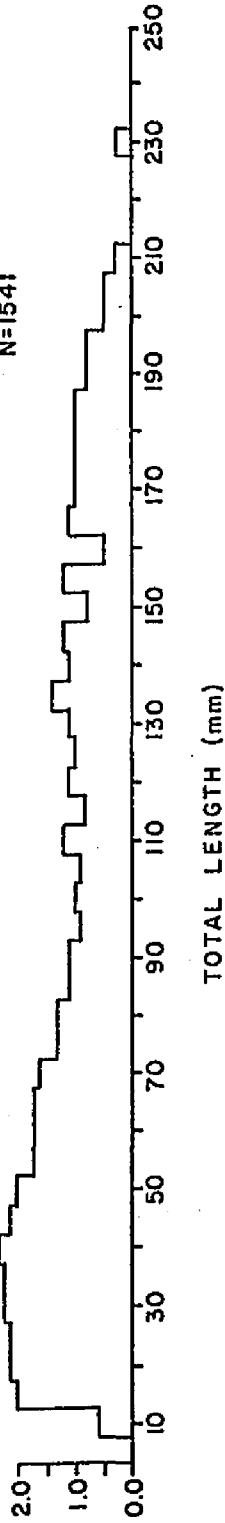
CHANNEL

N=1899



SHOAL

N=1541

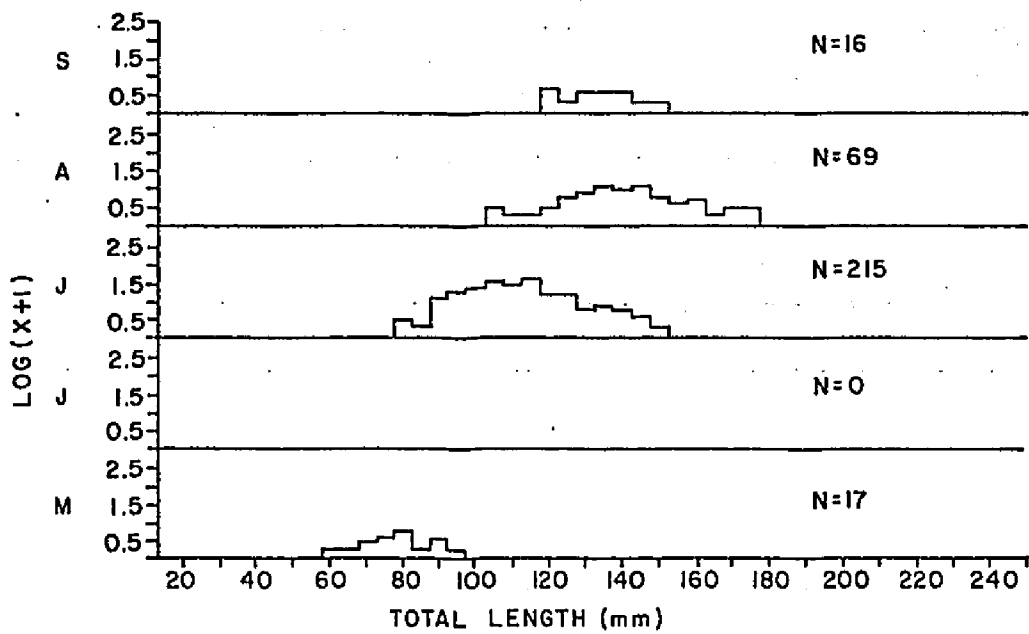


*Micropogonias undulatus* September to November, 1972-1974

Fig. 19

Length frequency distributions of croaker,  
*Micropogonias undulatus*, from beach seine catches  
of York River, May to September, 1972. Frequencies  
expressed as  $\log (x+1)$  at 5 mm increments.

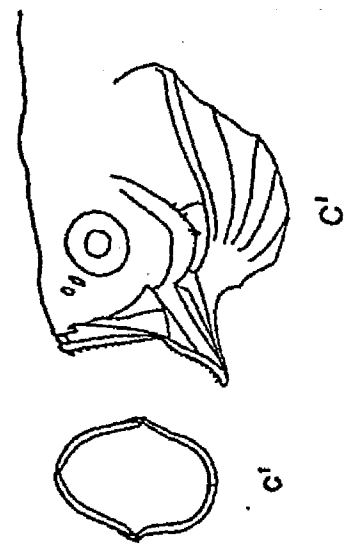
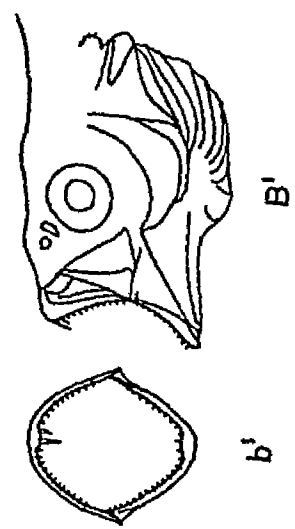
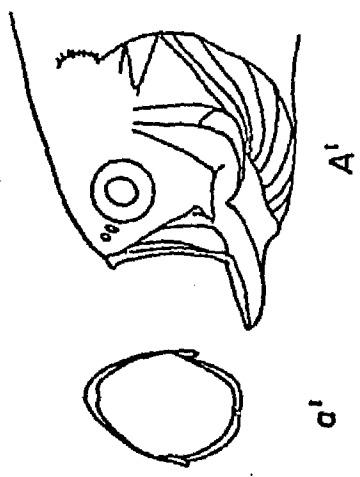
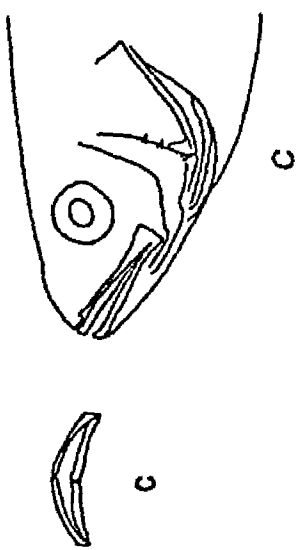
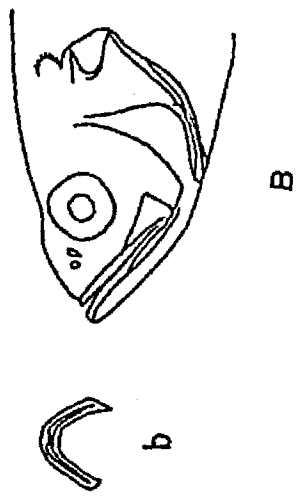
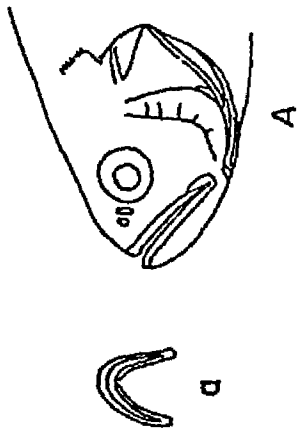




*Micropogonias undulatus* May to September 1972, Beach Seine

Fig. 20

Mouth position and opening in six species of juvenile sciaenids from York River. A, A', a & a', *Larimus fasciatus*; B, B', b & b', *Cynoscion regalis*; C, C', c & c', *Bairdiella chrysoura*; D, D', d & d', *Micropogonias undulatus*; E, E', e & e', *Menticirrhus saxatilis*; F, F', f & f', *Leiostomus xanthurus*. A, B, C, D, E & F, mouth closed. A', B', C', D', E' & F' mouth wide open. Front view of mouth openings (lower case letters) in corresponding positions.



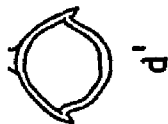
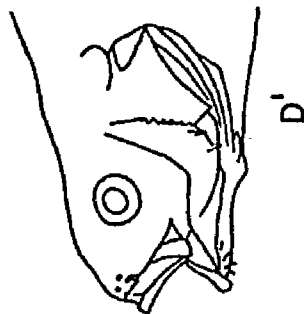
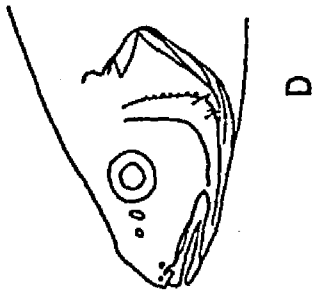
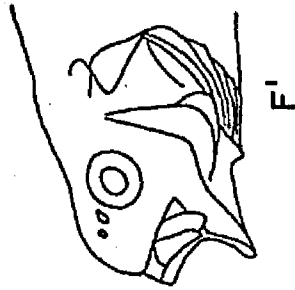
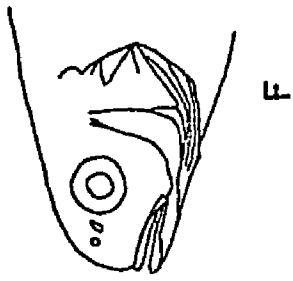
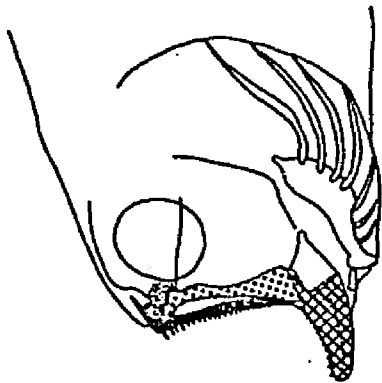


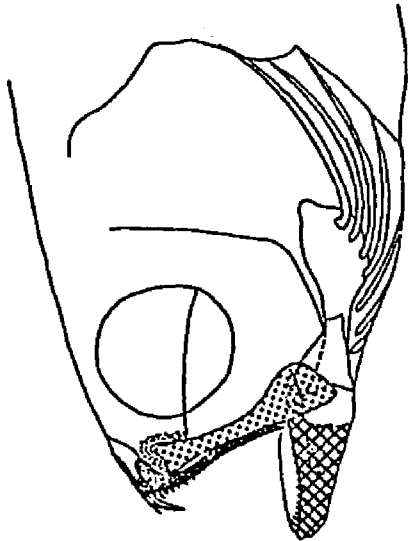
Fig. 21

Osteology of mouth opening in six species of juvenile sciaenids from York River. A. *Larimus fasciatus*; B. *Cynoscion regalis*; C. *Bairdiella chrysoura*; D. *Micropogonias undulatus*; E. *Menticirrhus saxatilis*; F. *Leiostomus xanthurus*.

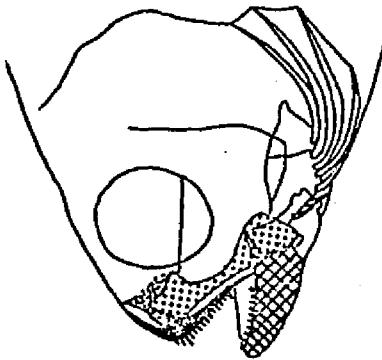
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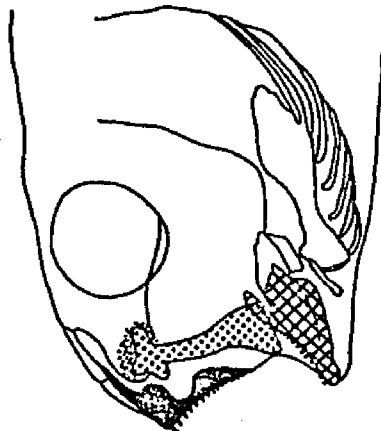
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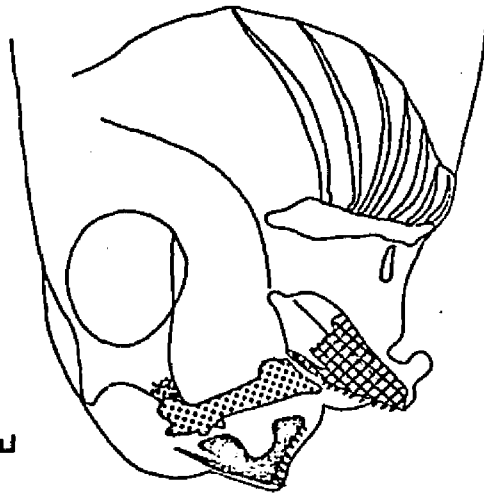
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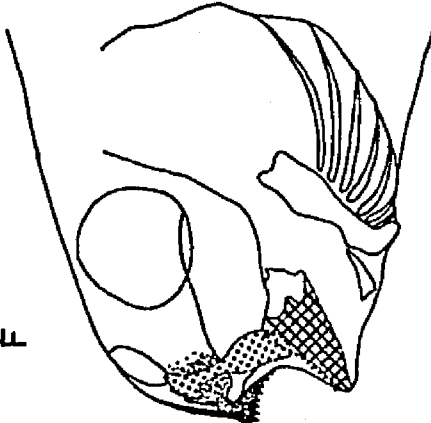
D



E



F



ETHMOID REGION



DENTARY



PREMAXILLA



MAXILLA

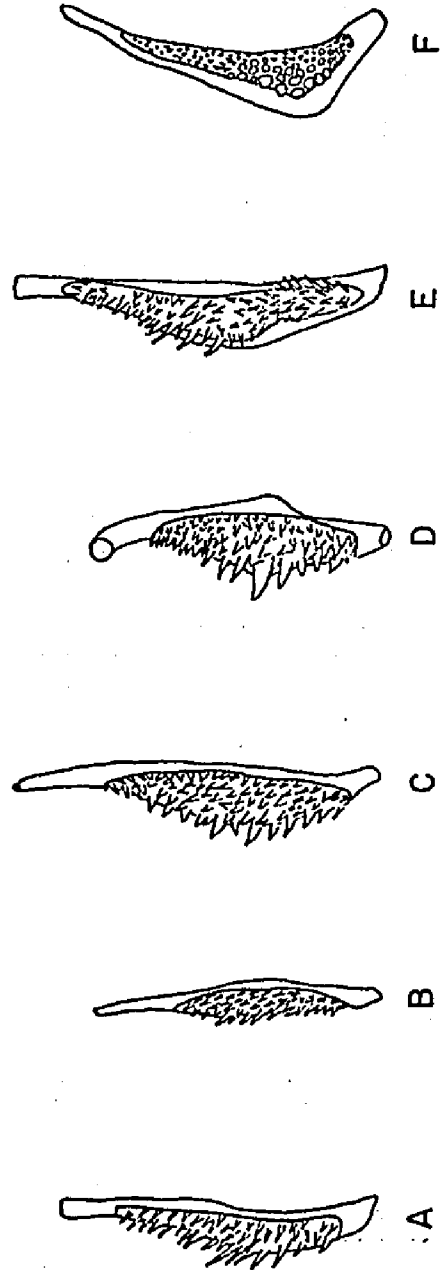


Fig. 22

Portions of right pharyngeal teeth in six species of juvenile sciaenids from York River. A. *Larimus fasciatus*; B. *Cynoscion regalis*; C. *Bairdiella chrysoura*; D. *Micropogonias undulatus*; E. *Menticirrhus saxatilis*; F. *Leiostomus xanthurus*. Anterior end toward the middle of the page.



UPPER PHARYNGEAL TEETH

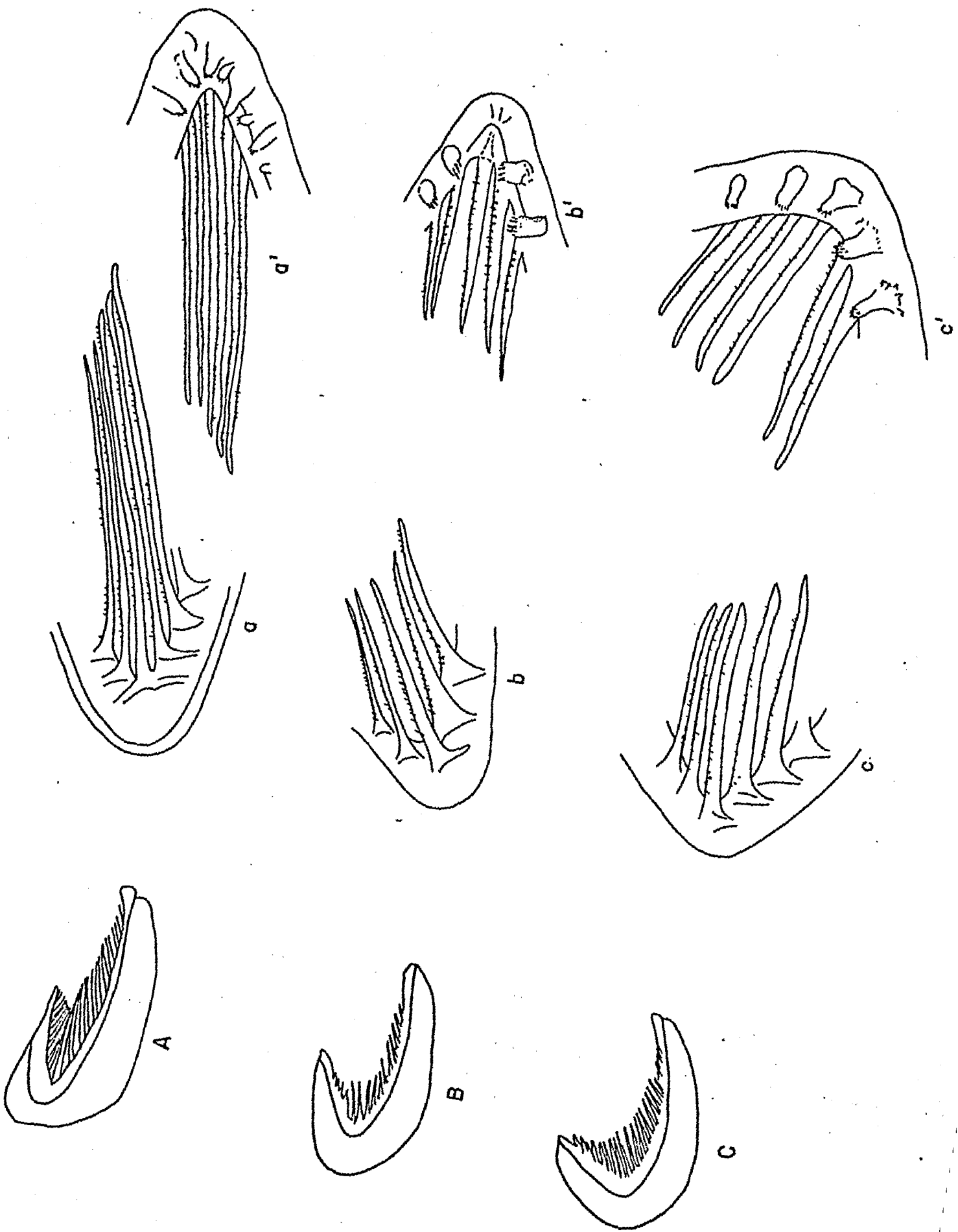


LOWER PHARYNGEAL TEETH



Fig. 23

First right gill arch in six species of juvenile sciaenids from York River. A, a & a', *Larimus fasciatus*; B, b & b', *Cynoscion regalis*; C, c & c', *Bairdiella chrysoura*; D, d & d', *Micropogonias undulatus*; E, e & e', *Menticirrhus saxatilis*; F, f & f', *Leiostomus xanthurus*. a, b, c, d, e & f, lateral view at the corner; a', b', c', d', e', & f', medial view at the corner.



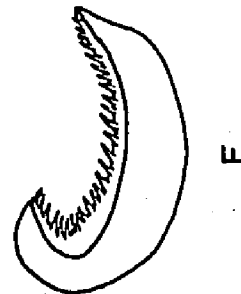
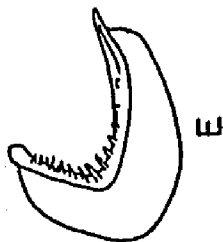
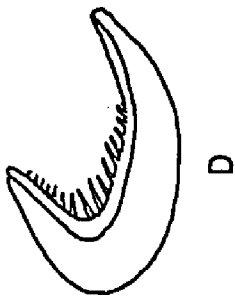
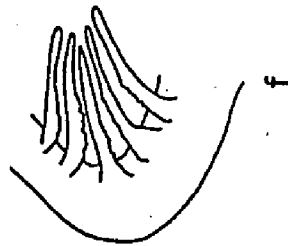
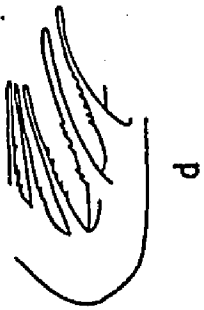
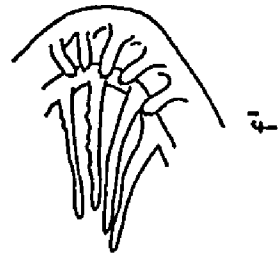
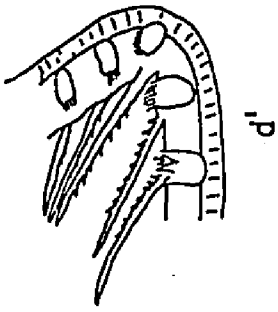
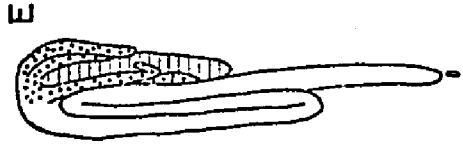
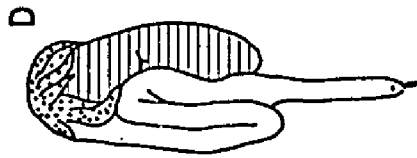
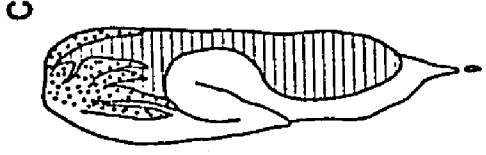
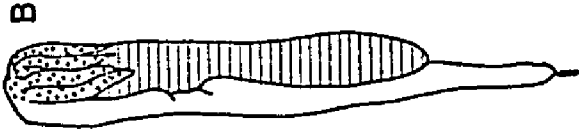
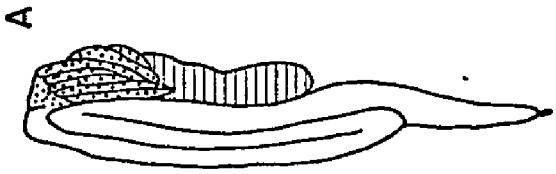


Fig. 24

Ventral view of the digestive tract of six species of juvenile sciaenids from York River. A. *Larimus fasciatus*; B. *Cynoscion regalis*; C. *Bairdiella chrysoura*; D. *Micropogonias undulatus*; E. *Menticirrhus saxatilis*; F. *Leiostomus xanthurus*.



PYLORIC  
CAECA



STOMACH



INTESTINE



Fig. 25

Snout pores (capital letters) and mental pores and barbels (lower case letters) in six species of juvenile sciaenids from York River. A & a, *Larimus fasciatus*; B & b, *Cynoscion regalis*; C & c, *Bairdiella chrysoura*; D & d, *Micropogonias undulatus*; E & e, *Menticirrhus saxatilis*; F & f, *Leiostomus xanthurus*.

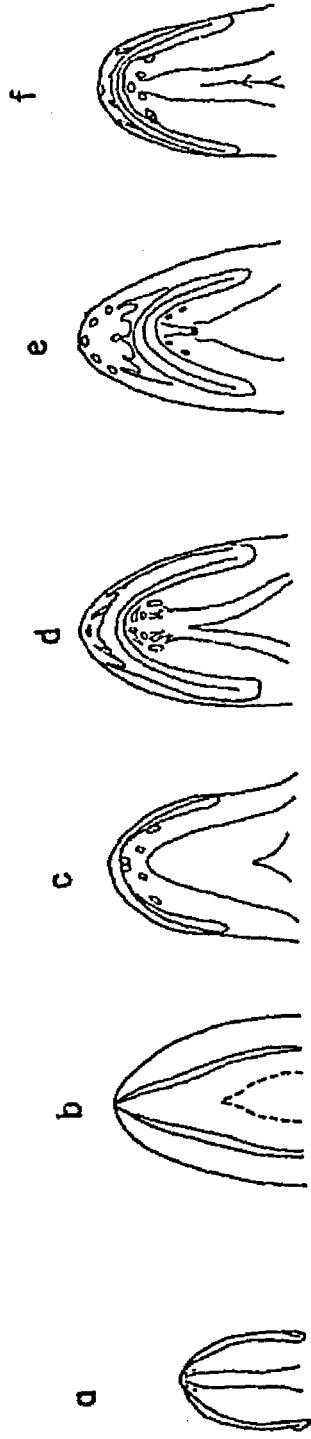
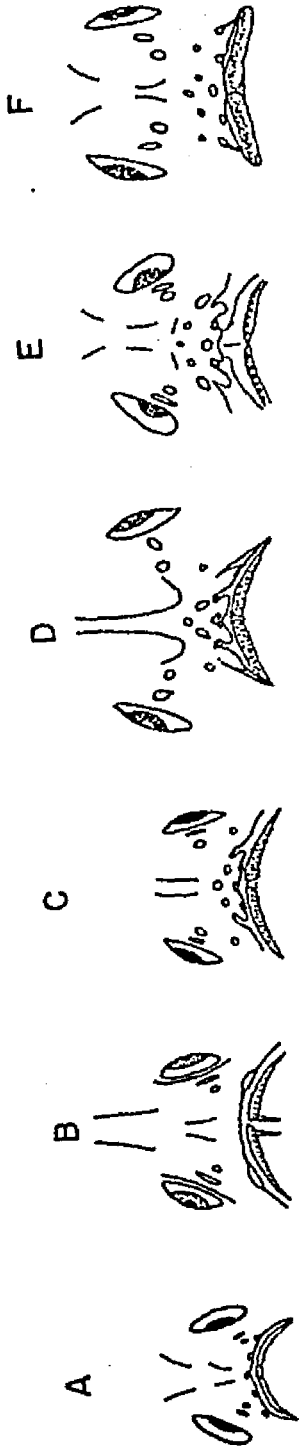
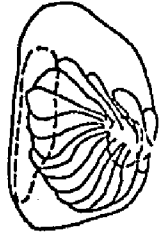


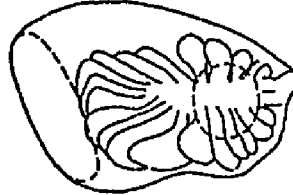
Fig. 26

Right olfactory rosette and nasal cavity in six species of juvenile sciaenids from York River. A. *Larimus fasciatus*; B. *Cynoscion regalis*; C. *Bairdiella chrysoura*; D. *Micropogonias undulatus*; E. *Menticirrhus saxatilis*; F. *Leiostomus xanthurus*. Dotted circles represent nostrils, the anterior nostril to the right.

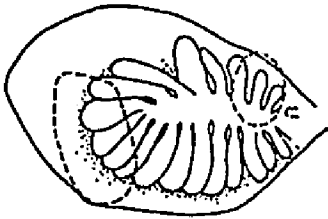




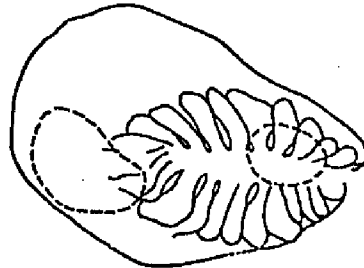
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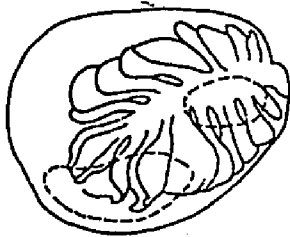
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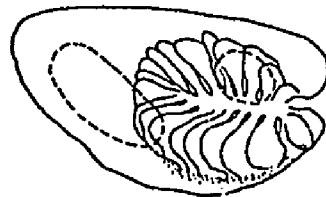
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D



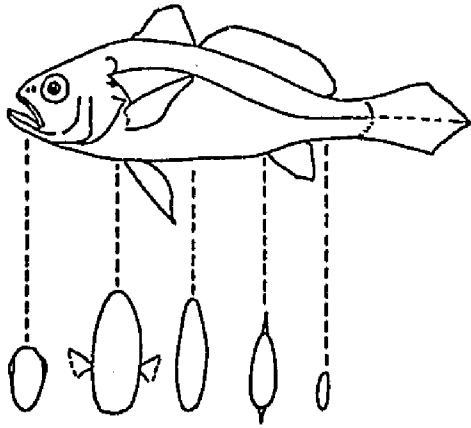
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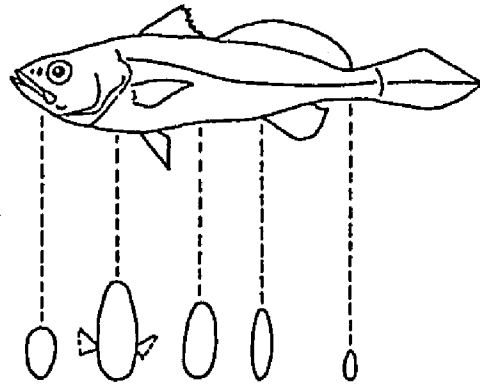
F

Fig. 27

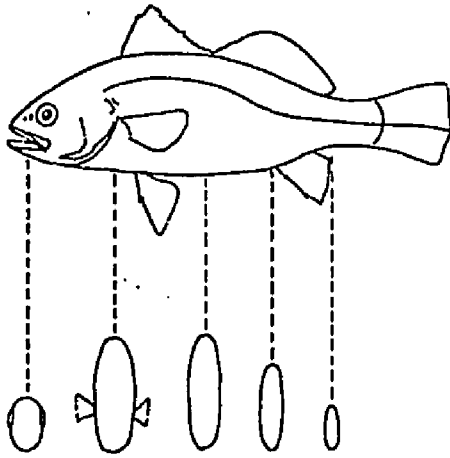
Body shape and cross sections of six species of juvenile sciaenids from York River. A. *Larimus fasciatus*; B. *Cynoscion regalis*; C. *Bairdiella chrysoura*; D. *Micropogonias undulatus*; E. *Menticirrhus saxatilis*; F. *Leiostomus xanthurus*.



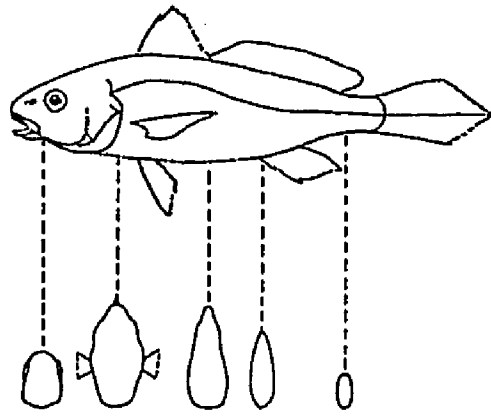
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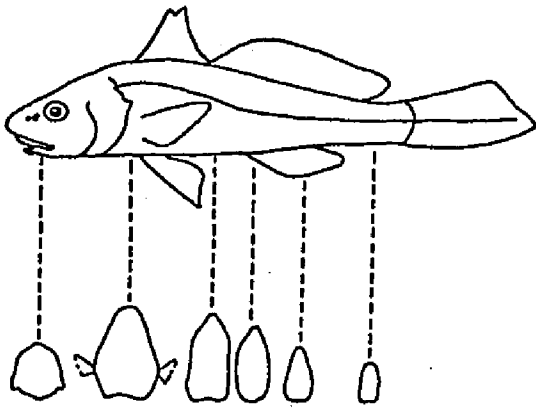
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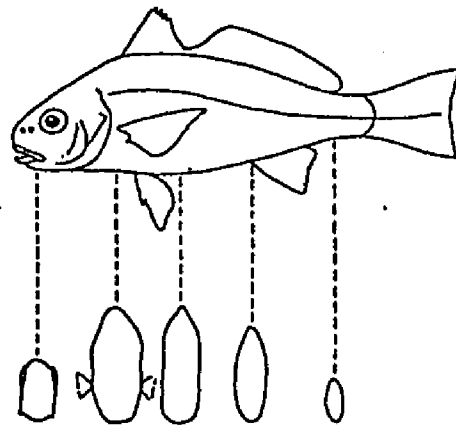
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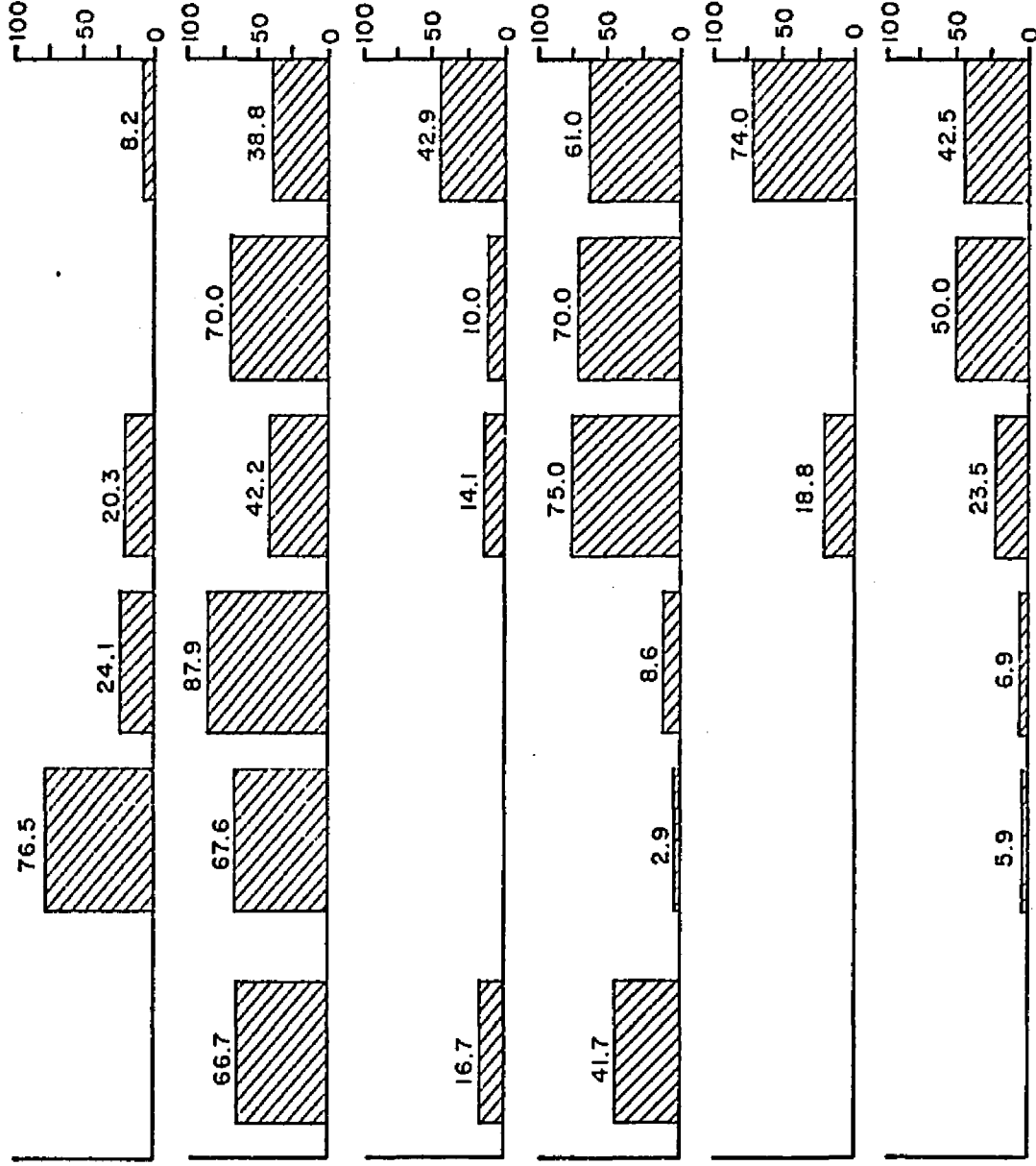
E



F

Fig. 28

Frequencies of occurrence of various categories  
of food groups in stomachs of six species of  
juvenile sciaenids from York River.



TOTAL LENGTH (mm)  
NUMBER OF STOMACHS

*Larimus fasciatus*    *Cynoscion regalis*    *Bairdiella chrysaora*    *Microgogonias undulatus*    *Menticirrhus saxatilis*    *Leiostomus xanthurus*

## APPENDIX

Mean surface and bottom water temperature (TEMP), salinity (SALN) and dissolved oxygen (D.O.) of the York River estuary from March 1972 to March 1974. Strata are expressed as A, B, C, D, E, F & G for the York River and M for the Mattaponi and P for Pamunkey rivers. Substrata are expressed as N (north shoal), C (channel) and S (south shoal) for each stratum of the York River.

Quantitative units:

Depth; in meters.

TEMP; in °C.

SALN; in parts per ten thousand for strata A to D, others in .

D.O.; in one tenth of mg/l for strata A to D, others in mg/l.

YEAR = 72 MONTH = 03

SURFACE

	TEMP			SALN			O.D.			NO		
	MEAN	MAX	MIN	S	D	NO	MEAN	MAX	MIN	S	D	NO
A-N	10.4	11.5	10.0	0.67		11	169	173	163	3.96		11
A-C	10.0	10.0	10.0	0.00		8	163	172	117	7.52		8
A-S	10.0	10.0	10.0	0.00		6	155	172	122	3.57		6
B-N	9.7	11.0	9.0	0.86		11	164	170	159	3.75		11
B-C	9.3	10.0	9.0	0.47		6	163	170	159	4.76		6
B-S	9.1	9.5	9.0	0.19		11	152	161	146	6.04		11
C-N	9.3	10.0	9.0	0.39		11	163	166	160	3.42		11
C-C	9.0	9.0	9.0	0.00		8	153	166	144	8.47		8
C-S	10.3	14.5	9.0	2.23		14	156	164	145	8.62		14
D-N	9.5	10.0	9.0	0.40		14	133	161	123	5.58		14
D-C	9.1	9.5	9.0	0.20		10	134	140	129	3.53		10
D-S	9.6	10.0	9.0	0.43		11	134	155	119	3.43		11

DEPTH

	DEPTH			TEMP			SALN			O.D.			NO			
	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	S	D	NO	MEAN	MAX	MIN	S	D	NO
A-N	3	5	1	8	9.8	10.0	9.0	0.43		8	172	173	171	1.00		8
A-C	13	16	8	8	9.1	9.5	9.0	0.22		8	193	220	170	5.31		8
A-S	3	6	4	6	9.7	10.0	9.5	0.24		6	172	172	171	0.47		6
B-N	3	5	1	11	8.6	9.5	8.0	0.54		8	165	169	162	2.59		8
B-C	15	23	5	6	8.3	9.0	8.0	0.47		6	190	207	170	0.60		6
B-S	2	5	1	11	8.6	9.5	8.0	0.55		8	151	160	146	5.26		8
C-N	2	3	1	11	8.4	9.5	8.0	0.65		8	163	166	156	3.04		8
C-C	10	14	6	8	8.4	9.0	8.0	0.41		8	163	177	145	1.61		8
C-S	2	4	1	14	9.0	9.0	9.0	0.00		8	150	154	147	2.69		8
D-N	2	2	1	14	9.5	10.0	9.0	0.55		8	132	143	127	0.50		8
D-C	14	17	9	10	9.0	9.0	9.0	0.00		10	171	199	155	4.47		10
D-S	3	5	1	8	9.4	10.0	9.0	0.41		8	130	148	130	6.12		8

YEAR = 72 MONTH = 04

	TEMP				SALN				SURFACE				D.O.							
	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO
A-N	15.4	16.5	14.5	0.77	15	160	167	155	4.38	15	88	99	49	5.51	15	87	99	58	1.70	12
A-C	15.1	16.0	14.5	0.58	10	162	164	160	1.79	10	76	81	73	2.79	10	63	82	47	2.31	10
A-S	15.5	16.0	14.0	0.77	10	162	165	159	1.95	10	84	92	69	8.38	10	80	89	64	0.01	10
B-N	15.7	17.0	15.0	0.94	9	164	171	158	5.11	9	84	92	69	8.80	9	68	76	54	8.62	8
B-C	14.8	15.0	14.0	0.43	8	160	165	156	3.34	8	63	90	71	7.70	8	83	87	79	3.54	8
B-S	17.0	21.0	14.5	2.25	14	164	173	154	8.07	14	108	129	84	8.47	14	72	74	69	2.50	4
C-N	15.3	16.0	15.0	0.45	11	160	167	157	4.56	11	103	129	84	6.95	11	64	81	50	1.45	6
C-C	14.8	15.0	14.5	0.25	4	155	158	151	3.50	4	81	82	80	1.00	4	83	87	79	3.54	8
C-S	17.2	20.5	14.5	2.50	17	160	171	142	9.71	17	86	99	58	4.82	17	64	78	50	1.45	6
D-N	16.4	16.0	15.0	1.20	17	159	165	146	7.20	17	86	99	50	4.82	17	52	60	47	5.56	6
D-C	15.2	15.5	15.0	0.24	6	133	141	134	3.09	6	61	82	47	3.23	6	65	88	46	6.76	6
D-S	15.9	17.0	14.5	1.04	11	144	160	130	4.30	11	80	89	64	9.55	11					

BOTTOM

	DEPTH				TEMP				SALN				D.O.						
	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO
A-N	2	5	1	15	15.1	16.0	14.0	0.71	12	159	152	156	2.61	12	87	99	58	1.70	12
A-C	12	15	7	10	12.8	13.0	12.0	0.40	10	215	233	160	7.98	10	63	82	47	2.31	10
A-S	4	5	2	10	15.1	16.0	13.0	1.11	10	162	164	160	1.36	10	80	89	64	0.01	10
B-N	3	5	1	9	14.7	15.0	14.5	0.24	6	162	163	162	0.47	6	79	89	64	9.59	6
B-C	14	18	9	8	13.1	14.0	12.0	0.74	8	167	229	160	6.28	8	68	76	54	8.62	8
B-S	1	3	1	14	14.5	15.5	13.5	1.22	8	157	160	154	3.00	8	83	87	79	3.54	8
C-N	3	5	1	11	14.5	15.0	14.0	0.35	6	160	163	157	2.38	6	83	87	79	3.54	8
C-C	11	12	10	4	13.8	14.0	13.5	0.25	4	155	162	147	7.50	4	72	74	69	2.50	4
C-S	1	3	1	17	14.3	15.0	14.0	0.47	6	150	154	144	4.52	6	64	78	50	1.45	6
D-N	2	3	1	17	14.3	14.5	14.0	0.24	6	159	160	140	5.25	6	64	78	50	1.45	6
D-C	12	14	9	6	13.8	14.0	13.5	0.24	6	147	167	136	4.38	6	52	60	47	5.56	6
D-S	3	5	2	6	14.5	14.5	14.5	0.00	6	137	139	133	2.62	6	65	88	46	6.76	6



YEAR = 72 MONTH = 05

	TEMP				SALN				D.U.C.						
	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO
A-N	21.1	24.0	18.0	2.92	12	150	159	134	0.19	12	73	81	62	6.14	12
A-C	18.5	18.8	18.3	0.21	6	144	149	137	5.25	6	60	62	55	4.08	6
A-S	18.2	18.4	18.1	0.12	6	138	142	131	5.19	6	68	70	66	1.70	6
B-N	20.9	24.0	18.5	2.69	14	132	161	61	7.27	14	70	90	57	0.94	14
B-C	18.8	19.0	18.5	0.24	6	147	151	143	5.30	6	62	67	63	1.70	6
B-S	19.4	20.5	18.0	1.11	12	156	163	148	6.29	12	61	65	57	2.54	12
C-N	19.1	19.5	18.5	0.30	14	146	156	136	9.02	14	64	80	57	7.11	14
C-C	19.1	19.2	19.0	0.09	6	151	141	123	7.48	6	62	62	64	0.47	6
C-S	21.1	24.0	19.0	1.96	21	146	155	132	6.87	21	71	88	49	0.81	21
D-N	20.4	23.5	19.0	2.03	20	142	148	129	7.16	20	71	80	49	0.99	20
D-C	19.8	20.5	19.0	0.62	6	115	133	102	4.38	6	64	76	56	7.95	6
D-S	20.9	22.0	19.5	1.13	12	122	138	105	6.13	12	69	83	56	8.75	12

DEPTH

	DEPTH				TEMP				SALN				D.U.C.			
	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO	
A-N	3	5	1	6	18.2	18.5	18.0	0.24	6	141	150	134	0.80	6	69	
A-C	16	18	13	6	18.0	18.0	18.0	0.00	6	172	190	150	0.66	6	64	
A-S	3	5	2	6	18.2	18.3	18.0	0.12	6	139	142	134	5.56	6	66	
B-N	2	3	2	8	13.5	18.5	0.1	8.19	11	147	148	147	0.43	8	65	
B-C	10	12	9	6	18.7	19.0	18.5	0.24	6	151	153	145	3.68	6	61	
B-S	2	2	1	6	18.3	18.5	18.0	0.44	6	149	151	148	1.22	6	59	
C-N	2	5	1	14	18.5	19.5	18.5	0.41	8	141	144	138	2.29	8	59	
C-C	12	24	6	6	18.5	19.0	17.5	0.68	6	164	212	151	4.87	6	52	
C-S	2	4	1	15	19.8	21.5	19.0	1.13	6	139	140	139	0.47	6	60	
D-N	1	2	1	14	19.0	19.0	19.0	0.00	8	135	140	129	4.03	8	60	
D-C	12	14	10	6	19.3	19.5	19.0	0.21	6	153	171	132	5.06	6	57	
D-S	2	4	1	12	19.8	20.5	19.3	0.52	6	112	120	105	6.13	6	61	

YEAR = 72 MONTH = 06

	TEMP			SALIN			D.O.					
	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	NO
A-N	23.5	25.5	23.5	6	156	156	156	6	80	80	80	6
B-N	27.0	27.0	27.0	6	150	150	150	6	92	92	92	6
B-S	25.0	25.0	25.0	6	159	159	159	6	85	85	77	6
C-N	24.0	24.0	24.0	6	156	156	156	6	81	85	77	6
C-S	23.2	31.0	24.0	10	159	140	140	10	74	80	62	10
D-N	24.0	24.0	24.0	5	142	142	142	5	60	80	80	5
D-S	27.0	27.0	27.0	6	151	151	151	6	106	106	106	6

WILSON

	DEPTH			TEMP			SALIN			D.O.		
	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	NO
B-N	1	1	1	6								
B-S	1	1	1	6								
C-N	1	1	1	6								
C-S	1	1	1	10								
D-S	1	1	1	6								

YEAR = 72 MONTH = 06

SURFACE

	TEMP			SAL			D.O.		
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN
E-N	24.0	24.6	24.0	13.97	9.33	2.21	6.80	7.90	5.30
E-C	24.30	24.6	24.0	5.77	9.33	2.21	6.60	7.90	5.30
E-S	24.0	24.6	24.0	12.71	9.33	2.21	8.70	7.90	5.30
F-N	25.25	25.5	25.0	7.46	8.16	6.76	6.60	6.60	6.60
F-C	24.25	25.5	24.0	3.46	7.63	0.27	6.33	8.20	4.90
F-S	25.50	26.0	25.0	8.46	10.61	6.16	7.43	7.50	7.30
G-N									
G-S									
P	22.85	23.2	22.5	0.28	0.48	0.11	5.00	4.00	6.00

S D			S D			S D		
1	1	1	2	2	1	2	2	1
2	2	2	1	1	1	2	2	1
6	6	6	3	3	3	6	6	3
3	3	3	2	2	2	3	3	2
2	2	2	2	2	2	2	2	2
2	2	2	2	2	2	2	2	2

BOTTOM

	DEPTH			TEMP			SALN			D.O.		
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN
E-N	2.0	6.0	9.0	24.00	22.00	23.50	13.60	14.85	3.53	6.60	4.60	4.30
E-C	7.50	6.0	9.0	22.75	22.00	23.50	9.19	14.85	3.53	5.660	4.60	4.30
E-S	3.0	3.0	2.0	24.00	25.0	25.0	12.86	7.82	6.50	0.660	6.30	6.10
F-N	2.50	3.0	2.0	25.00	24.0	23.0	7.16	12.31	0.49	3.550	7.40	3.19
F-C	8.17	10.0	6.0	23.52	25.5	25.0	6.64	11.28	6.12	2.107	8.10	6.70
F-S	2.67	3.0	2.0	25.33	25.5	25.0	8.73	7.60	8.10	0.638	8.10	6.70
G-N	6.0			22.00			6.18	8.20				
G-C												
G-S												
P	6.66	9.0	5.0	22.40	22.6	22.0	0.18	0.43	0.05	0.216	8.20	2.20

SURFACE

	TEMP			SALIN			D.O.			NO			
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN				
A-N	27.4	28.0	27.0	0.49	111	117	100	7.08	73	57	7.53	14	
A-C	27.1	27.5	27.0	0.20	105	109	100	3.74	70	67	4.12	10	
A-S	27.0	27.0	27.0	0.00	105	107	100	2.77	66	59	4.49	6	
B-N	27.4	28.0	27.0	0.49	117	120	110	3.15	68	53	9.47	14	
B-C	28.0	28.0	28.0	0.00	109	115	105	3.50	73	66	5.61	6	
B-S	28.0	31.0	12.9	4.36	114	129	105	3.72	69	49	0.02	15	
C-N	28.1	30.0	27.0	1.07	117	120	112	3.29	70	49	0.57	13	
C-C	28.0	29.0	27.0	0.82	116	120	112	3.50	53	34	3.74	6	
C-S	30.5	33.0	27.0	2.03	111	131	73	1.24	42	37	4.68	17	
D-N	27.1	27.5	27.0	0.19	103	131	77	6.15	42	37	5.23	12	
D-C	27.5	28.5	27.0	0.71	121	126	117	3.05	50	68	14	0.79	6
D-S	30.0	33.0	27.0	3.00	105	125	78	5.34	74	55	8.38	10	

EDDIUM

	DEPTH			TEMP			SALIN			D.O.			NO
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	
A-N	2	5	1	14	27.1	26.0	25.5	0.54	108	120	102	7.07	6
A-C	11	12	9	10	22.8	27.0	20.0	2.71	210	260	106	2.38	10
A-S	3	4	2	8	26.9	27.0	26.5	0.22	107	142	101	4.15	8
B-N	2	4	1	14	27.0	27.0	27.0	0.00	126	141	111	1.02	8
B-C	10	11	9	6	24.8	26.5	22.0	2.01	173	250	132	1.73	6
B-S	2	3	1	9	25.6	27.5	13.0	4.46	119	150	109	9.00	9
C-N	3	5	1	13	27.9	29.0	27.0	0.89	118	116	117	0.50	4
C-C	13	23	7	6	25.5	29.0	21.5	3.06	160	243	114	3.60	6
C-S	2	3	1	11	26.7	27.0	26.5	0.24	150	151	128	1.41	6
D-N	2	3	1	12	26.5	26.5	26.5	0.00	130	131	129	0.94	6
D-C	15	20	10	6	22.2	23.5	21.0	1.02	214	237	201	6.51	6
D-S	3	4	2	6	26.0	26.5	25.0	0.71	134	145	126	6.16	6

YEAR = 72 MONTH = 07

SURFACE

	TEMP			SAL			D.O.			S D			NO
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	
E-N	30.07	30.5	29.5	5.33	5.77	5.08	9.13	9.50	8.50	0.450	0.450	0.450	3
E-C	29.78	30.5	29.0	5.61	9.69	3.64	8.63	11.10	6.20	1.595	1.595	1.595	6
E-S	29.79	31.0	29.0	4.82	5.53	3.67	8.31	11.50	5.60	1.577	1.577	1.577	8
F-N	30.88	32.0	30.0	1.92	3.31	0.65	6.45	7.20	5.80	0.610	0.610	0.610	4
F-C	30.13	30.5	29.5	1.76	2.32	1.23	4.17	6.50	3.00	1.429	1.429	1.429	4
F-S	30.30	31.0	30.0	3.34	4.28	2.23	5.40	6.30	3.30	1.092	1.092	1.092	5
G-N	30.93	31.5	30.3	2.59	6.88	0.37	4.60	5.50	3.80	0.698	0.698	0.698	3
G-C													
G-S	29.97	30.9	29.5	1.14	1.16	1.13	4.90	6.10	4.30	0.849	0.849	0.849	3

BOTTOM

	DEPTH			TEMP			SALIN			D.O.			NO
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	
E-N	2.00	2.0	2.0	29.67	30.5	28.5	8.01	8.01	5.26	1.250	1.250	1.250	3
E-C	9.33	11.0	7.0	26.50	30.0	25.0	13.67	13.67	9.97	1.244	1.244	1.244	6
E-S	2.50	3.0	2.0	27.94	31.5	25.5	12.84	12.84	5.01	3.077	3.077	3.077	8
F-N	2.00	2.0	2.0	30.20	30.5	30.0	3.31	3.31	1.34	0.711	0.711	0.711	4
F-C	6.75	9.0	5.0	27.13	27.5	26.5	10.98	10.98	9.91	0.457	0.457	0.457	4
F-S	2.20	3.0	1.0	29.5	31.0	27.0	9.30	9.30	2.91	2.336	2.336	2.336	5
G-N	1.67	2.0	1.0	29.57	30.5	28.7	4.12	4.12	0.71	1.405	1.405	1.405	3
G-C													
G-S	2.00	2.0	2.0	29.67	30.0	29.0	2.21	2.21	1.06	0.542	0.542	0.542	3

YEAR = 72 MONTH = 08

SURFACE

	TEMP				SALN				D.O.						
	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D			
A-N	27.9	31.0	25.3	2.21	18	164	161	149	2.71	16	90	120	48	3.00	18
A-C	26.9	27.0	26.6	0.16	8	161	165	176	2.59	8	81	93	71	9.50	8
A-S	26.0	27.0	20.1	2.00	10	130	161	176	1.20	10	86	97	60	6.94	8
B-N	27.3	30.5	25.4	2.38	14	166	169	150	3.78	14	100	120	60	7.74	14
B-C	26.0	26.1	25.3	0.12	6	176	177	173	1.89	6	53	56	52	1.89	6
B-S	26.3	27.7	25.5	0.90	13	165	177	152	0.93	13	72	97	53	4.79	13
C-N	27.1	28.0	25.7	0.93	14	160	176	140	7.43	14	74	97	53	5.63	14
C-C	26.1	26.5	25.7	0.33	6	180	183	177	2.49	6	60	64	52	5.44	6
C-S	26.9	28.0	26.0	0.85	16	150	171	146	0.06	16	73	87	38	4.79	16
D-N	27.9	29.0	26.8	0.95	14	153	163	133	2.71	14	72	87	38	5.30	14
D-C	26.7	26.8	26.5	0.14	6	169	172	164	3.40	6	34	54	50	1.55	6
D-S	27.4	27.8	26.9	0.28	12	147	169	127	0.06	12	74	87	48	4.55	12

BOTTOM

	DEPTH				TEMP				SALN				D.O.				
	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	
A-N	4	6	2	12	26.1	25.8	25.3	0.49	12	176	192	162	1.09	12	87	38	5.65
A-C	12	13	12	8	25.4	25.5	25.3	0.07	8	196	199	197	0.87	8	64	30	2.52
A-S	4	6	3	10	26.1	26.6	25.7	0.33	10	185	197	175	6.47	10	71	48	4.56
B-N	3	5	1	14	25.8	26.4	25.2	0.43	8	178	179	176	1.30	8	67	48	3.63
B-C	14	21	10	6	25.5	25.9	25.0	0.37	6	186	200	176	0.08	6	50	28	1.87
B-S	2	5	1	13	26.6	27.6	25.6	0.72	8	176	176	175	0.43	8	64	57	4.27
C-N	2	4	1	14	26.2	27.0	25.4	0.73	6	178	182	173	3.19	8	64	57	4.27
C-C	7	8	7	6	25.9	25.9	25.0	0.37	6	186	190	183	2.94	6	58	29	3.44
C-S	2	3	1	16	26.7	26.9	26.3	0.21	6	170	171	169	0.82	6	54	52	2.62
D-N	2	2	1	14	27.0	27.3	26.5	0.31	6	169	177	163	5.12	8	53	41	5.15
D-C	12	13	11	6	25.6	25.8	25.7	0.05	6	185	188	181	2.67	6	34	28	4.19
D-S	3	5	2	6	26.5	27.2	26.0	0.50	6	176	186	167	7.79	6	47	31	2.28

YEAR = 72 MONTH = 08

SURFACE

	TEMP			SAL			D.O.			S D			NO					
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN
E-N	26.26	26.50	26.2	12.15	12.56	10.94	13.46	16.03	11.88	7.62	7.90	7.10	13.46	16.03	11.88	5.68	6.10	4.60
E-C	26.96	27.10	26.4	11.69	12.45	10.79	17.83	20.21	16.61	8.16	9.30	7.20	17.83	20.21	16.61	1.59	2.40	0.80
E-S	27.19	27.90	26.3	11.01	12.10	10.19	11.92	15.65	10.74	7.51	9.00	5.80	11.92	15.65	10.74	5.56	6.70	3.40
F-N	27.12	27.20	26.90	10.79	11.61	9.80	10.73	11.44	9.79	7.25	7.80	6.50	10.73	11.44	9.79	6.67	7.70	5.50
F-C	26.93	28.1	26.0	10.95	11.8	9.59	14.70	17.24	12.20	7.43	10.3	6.30	14.70	17.24	12.20	2.47	4.00	1.50
F-S	27.08	29.0	26.4	10.68	11.59	9.89	11.18	12.94	9.83	6.67	9.00	5.50	11.18	12.94	9.83	5.43	7.20	3.50
G-N	27.20	28.0	26.7	7.78	9.19	5.75	8.52	9.87	6.32	7.33	7.70	7.00	8.52	9.87	6.32	5.03	5.80	4.30
G-C	26.70	27.2	26.2	7.21	7.53	6.90	8.46	8.72	8.20	6.65	7.80	5.50	8.46	8.72	8.20	4.25	4.50	4.00
G-S	25.90	25.9	25.9	7.02	8.05	5.99	7.66	8.60	6.72	5.75	6.10	5.40	7.66	8.60	6.72	4.85	5.60	4.10
P	28.3			4.82			6.55	7.10	6.01	8.8			6.55	7.10	6.01	5.30	6.00	4.60

BOTTOM

	DEPTH			TEMP			SALIN			D.O.			NO					
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN
E-N	2.00	2.0	2.0	26.16	26.2	26.0	13.46	16.03	11.88	5.68	6.10	4.60	5.68	6.10	4.60	5.68	6.10	4.60
E-C	8.71	11.0	6.0	25.84	26.8	24.8	17.83	20.21	16.61	1.59	2.40	0.80	1.59	2.40	0.80	1.59	2.40	0.80
E-S	2.38	4.0	1.0	27.07	28.0	26.7	11.92	15.65	10.74	5.56	6.70	3.40	5.56	6.70	3.40	5.56	6.70	3.40
F-N	1.50	2.0	1.0	28.02	28.30	27.90	10.73	11.44	9.79	6.67	7.70	5.50	6.67	7.70	5.50	6.67	7.70	5.50
F-C	8.57	11.0	5.0	26.51	28.00	24.90	14.70	17.24	12.20	2.47	4.00	1.50	2.47	4.00	1.50	2.47	4.00	1.50
F-S	2.09	4.0	1.0	27.68	28.90	26.70	11.18	12.94	9.83	5.43	7.20	3.50	5.43	7.20	3.50	5.43	7.20	3.50
G-N	3.00	4.0	2.0	27.40	27.60	27.20	8.52	9.87	6.32	5.03	5.80	4.30	5.03	5.80	4.30	5.03	5.80	4.30
G-C	4.50	5.0	4.0	27.25	27.50	27.00	8.46	8.72	8.20	4.25	4.50	4.00	4.25	4.50	4.00	4.25	4.50	4.00
G-S	2.00	2.0	2.0	26.05	26.10	26.00	7.66	8.60	6.72	4.85	5.60	4.10	4.85	5.60	4.10	4.85	5.60	4.10
P	3.5	2.0	5.0	27.20	27.40	27.00	6.55	7.10	6.01	5.30	6.00	4.60	5.30	6.00	4.60	5.30	6.00	4.60

YEAR = 72 MONTH = 09

	TEMP				SALIN				SURFACE				D.U.							
	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO
A-N	20.7	21.5	19.0	0.97	16	18.7	18.7	17.6	4.95	16	78	89	65	0.37	16	86	92	74	6.83	10
A-C	21.2	21.5	21.0	0.24	10	188	190	184	2.40	10	54	96	79	5.99	10	56	70	41	9.65	10
A-S	20.3	21.5	20.0	0.56	8	190	192	186	1.79	8	74	87	58	0.36	8	73	89	47	6.87	8
B-N	21.1	22.0	20.0	0.63	14	183	188	177	5.55	14	75	87	58	0.07	14	62	106	40	0.88	6
B-C	21.0	21.0	21.0	0.00	6	191	192	191	0.47	6	64	75	56	3.18	6	72	86	58	0.40	8
B-S	21.1	22.0	20.0	0.78	13	183	191	175	6.43	13	72	89	49	2.09	13	65	71	59	4.92	6
C-N	21.5	22.6	20.0	0.82	14	182	195	173	8.36	14	71	89	49	1.75	14	64	86	52	1.51	6
C-C	21.7	22.5	21.0	0.62	6	193	205	181	7.09	6	75	82	71	4.78	6	61	72	52	1.51	6
C-S	22.9	26.0	21.0	1.63	17	178	187	173	6.33	17	79	92	57	1.71	17	62	86	52	1.53	14
D-N	21.2	22.0	19.5	0.84	14	172	185	163	8.90	14	60	92	57	1.53	14	62	70	49	7.34	6
D-C	21.3	21.5	21.0	0.24	6	177	184	170	5.73	6	62	70	49	7.34	6	61	76	45	2.26	6
D-S	21.3	22.0	20.5	0.69	12	183	174	156	7.27	12	68	89	47	0.46	12	61	76	45	2.26	6

ADDITION

	DEPTH				TEMP				SALIN				D.U.						
	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO
A-N	4	6	2	10	20.0	21.0	19.0	0.71	10	187	197	176	0.93	10	86	92	74	6.83	10
A-C	12	14	11	10	21.7	22.0	21.0	0.40	10	230	238	222	6.58	10	56	70	41	9.65	10
A-S	4	6	2	8	20.8	21.0	20.0	0.43	8	191	198	189	3.96	8	73	89	47	6.87	8
B-N	2	5	1	14	20.0	20.5	19.5	0.55	8	188	188	187	0.50	8	73	89	47	6.87	8
B-C	12	15	8	6	21.7	22.0	21.5	0.24	6	206	210	198	5.44	6	62	106	40	0.88	6
B-S	2	3	1	13	20.6	21.0	20.0	0.41	8	188	191	184	2.49	8	72	86	58	0.40	8
C-N	2	3	1	14	20.8	22.0	19.5	0.90	8	188	190	186	1.58	8	72	86	58	0.40	8
C-C	9	12	7	6	21.8	22.0	21.5	0.24	6	192	200	185	5.44	6	65	71	59	4.92	6
C-S	2	5	1	17	22.5	24.0	21.5	1.06	6	187	187	187	0.00	6	65	86	52	1.51	6
D-N	2	4	1	14	20.8	21.0	20.0	0.43	8	186	191	181	3.57	8	64	86	52	9.98	8
D-C	11	12	11	6	22.2	22.5	22.0	0.24	6	225	230	218	5.10	6	54	57	52	2.16	6
D-S	6	62	1	12	20.5	20.5	20.5	0.00	6	171	174	168	2.45	6	61	76	45	2.26	6



YEAR = 72 MONTH = 09

SURFACE

	TEMP			SAL			D.O.					
	MEAN	MAX	MIN	S	D	NO	MEAN	MAX	MIN	S	D	NO
E-N	23.45	23.6	23.3	0.150	0.150	4	16.85	17.55	16.28	0.548	0.548	4
E-C	23.33	23.4	23.3	0.045	0.045	7	16.81	16.99	16.45	0.162	0.162	7
E-S	22.70	23.4	22.20	0.576	0.576	4	16.52	16.83	16.22	0.262	0.262	4
F-N												
F-C												
F-S												
G-N												
G-C												
G-S												

BOTTOM

	DEPTH			TEMP			SALN			D.O.						
	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	S	D	NO	MEAN	MAX	MIN	S	D	NO
E-N	1.50	2.0	1.0	4	23.07	23.3	23.0	0.130	0.130	4	17.07	17.58	16.47	0.462	0.462	4
E-C	7.71	10.0	5.0	7	23.14	23.2	23.1	0.049	0.049	7	17.22	17.30	17.03	0.088	0.088	7
E-S	3.00	4.0	2.0	4	22.87	23.1	22.8	0.130	0.130	4	16.71	17.13	16.32	0.379	0.379	4
F-N																
F-C																
F-S																
G-N																
G-C																
G-S																

YEAR = 72 MONTH = 10

	TEMP			SALT			SURFACE			D.U.		
	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	NO
A-N	15.5	17.0	13.9	12	182	188	170	6	69	102	82	12
A-C	14.7	15.1	14.0	6	187	189	155	6	34	97	77	6
A-S	14.7	14.9	14.0	4	186	183	187	4	91	108	74	4
B-W	16.0	17.0	14.6	12	184	190	179	12	87	108	75	12
B-C	15.1	15.2	15.0	8	188	194	182	8	65	83	65	8
B-S	16.0	18.0	18.0	6	170	170	170	6	67	87	87	6
C-N	16.6	18.0	14.5	10	181	182	180	10	65	87	81	10
C-C	15.9	16.4	14.9	6	179	180	177	6	77	91	87	6
C-S	20.3	23.0	15.8	14	169	177	166	14	64	84	44	14
D-N	17.4	18.0	16.1	18	157	175	146	18	87	90	44	18
D-C	16.0	16.4	15.3	8	177	179	174	8	63	90	53	8
D-S	22.2	25.4	18.2	12	158	177	159	12	60	88	60	12

BULLION

	DEPTH			TEMP			SALT			D.O.		
	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	NO
A-N	3	4	2	6	14.4	14.6	14.3	6	200	200	189	6
A-C	10	12	6	6	14.7	14.9	14.5	6	204	204	195	6
A-S	5	3	2	4	14.4	14.9	14.0	4	204	204	194	4
B-N	2	4	1	12	14.7	14.9	14.4	6	187	187	185	6
B-C	11	14	8	8	14.8	15.0	14.6	8	198	198	182	8
B-S	1	1	1	6	13.0	14.4	12.8	4	181	181	161	4
C-N	1	2	1	10	15.4	15.8	14.7	6	193	193	189	6
C-C	15	23	11	6	15.8	15.8	15.0	6	170	170	170	6
C-S	2	7	1	14	16.6	16.8	16.3	6	175	175	172	6
D-N	1	2	1	18	16.1	16.3	15.9	8	181	181	178	8
D-C	13	18	9	8	16.1	16.2	16.0	8	179	179	179	8
D-S	3	5	1	12	16.1	16.2	16.0	6	179	179	179	6

YEAR = 72 MONTH = 10

SURFACE

	TEMP			SAL			D.O.			MEAN			NO		
	MAX	MIN	NO	MAX	MIN	NO	MAX	MIN	NO	MAX	MIN	NO	MAX	MIN	NO
E-N	16.00	16.8	3	16.53	9.22	3	7.80	8.50	3	7.80	8.50	3	7.80	8.50	3
E-C	16.02	16.8	6	15.30	9.27	6	7.58	7.70	6	7.58	7.70	6	7.58	7.70	6
E-S	17.29	20.0	12	15.45	8.99	12	7.69	8.30	12	7.69	8.30	12	7.69	8.30	12
F-N	18.11	21.1	9	14.56	9.63	9	7.23	7.50	9	7.23	7.50	9	7.23	7.50	9
F-C	18.11	21.1	7	14.76	9.06	7	6.89	7.60	7	6.89	7.60	7	6.89	7.60	7
F-S	17.04	20.6	15	14.25	8.98	15	7.18	7.60	15	7.18	7.60	15	7.18	7.60	15
G-N	18.04	21.1	8	11.45	8.34	8	6.85	8.00	8	6.85	8.00	8	6.85	8.00	8
G-C	18.37	21.7	3	8.78	7.69	3	7.20	8.30	3	7.20	8.30	3	7.20	8.30	3
G-S	17.70	18.9	2	10.99	7.71	2	7.90	8.20	2	7.90	8.20	2	7.90	8.20	2

BOTTOM

	DEPTH			TEMP			SALIN			D.O.			MEAN			NO		
	MAX	MIN	NO	MAX	MIN	NO	MAX	MIN	NO	MAX	MIN	NO	MAX	MIN	NO	MAX	MIN	NO
E-N	2.00	2.0	3	15.6	0.424	3	16.38	8.86	3	13.85	16.38	3	7.40	7.60	3	7.40	7.60	3
E-C	9.17	10.0	6	15.6	1.354	6	19.24	17.72	6	18.26	19.24	6	7.58	7.90	6	7.58	7.90	6
E-S	2.54	4.0	12	14.2	1.443	12	15.46	9.68	12	12.69	15.46	12	7.36	8.50	12	7.36	8.50	12
F-N	1.89	5.0	9	14.8	2.502	9	14.66	9.50	9	12.20	14.66	9	6.83	7.40	9	6.83	7.40	9
F-C	1.86	10.0	7	16.2	2.039	7	17.33	13.60	7	15.80	17.33	7	6.43	7.50	7	6.43	7.50	7
F-S	2.11	4.0	19	14.7	1.972	15	15.48	8.40	15	11.68	15.48	15	7.25	8.10	15	7.25	8.10	15
G-N	2.88	5.0	8	16.4	1.687	8	12.37	9.33	8	11.37	12.37	8	6.92	8.10	8	6.92	8.10	8
G-C	4.33	5.0	3	17.0	1.910	3	12.55	9.12	3	11.01	12.55	3	6.60	8.40	3	6.60	8.40	3
G-S	2.00	2.0	2	16.8	1.300	2	11.84	10.96	2	11.40	11.84	2	7.90	7.90	2	7.90	7.90	2

YEAR = 72 MONTH = 11

	TEMP				SALN				SURFACE				J.G.								
	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO	
B-S	8.2	8.2	8.2	0.00	6	152	155	152	0.00	6	106	106	106	0.00	6						
C-S	11.1	14.2	8.1	3.05	12	132	137	126	5.50	12	105	109	52	7.36	12						
D-N	12.5	12.5	12.5	0.00	6	124	124	124	0.00	6	107	109	109	0.00	6						
D-S	7.2	7.2	7.2	0.00	6	104	104	104	0.00	6	110	110	110	0.00	6						

BOTTOM

	DEPTH				TEMP				SALN				D.U.						
	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D

YEAR = 72 MONTH = 11

SURFACE

	TEMP			SAL			D.O.			S D			NO		
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	S	D	NO	S	D	NO
E-N	11.53	12.1	11.0	15.98	17.16	13.72	8.27	8.60	8.00	0.249	3	8.47	8.70	8.30	3
E-C	11.20	11.4	11.0	14.65	15.38	14.15	8.75	9.70	8.20	0.594	4	8.07	10.40	6.60	4
E-S	10.98	11.4	10.5	13.97	15.33	12.80	8.89	9.50	8.30	0.351	9	8.67	9.90	7.30	9
F-N	12.70	13.4	12.2	9.07	10.28	8.13	8.65	9.20	7.90	0.431	6	9.07	12.0	7.80	6
F-C	12.05	13.2	11.4	9.89	12.57	8.39	8.48	9.00	8.00	0.319	5	7.78	8.90	6.60	6
F-S	10.51	11.3	9.7	9.40	12.60	8.37	8.55	9.80	6.80	0.866	11	8.57	10.00	6.60	10
G-N	12.60	12.9	12.3	7.93	8.93	6.93	7.95	8.00	7.90	0.050	2	7.40	7.50	7.30	2
G-C	11.55	12.60	10.50	9.38	9.52	9.24	7.80	8.3	7.3	0.500	2	6.85	7.10	6.60	2
G-S	12.9			7.53			9.90				1	8.60			1

BOTTOM

	DEPTH			TEMP			SALIN			D.O.			S D			NO		
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	S	D	NO	S	D	NO	S	D	NO
E-N	2.00	2.0	2.0	11.9	11.5	0.170	16.54	16.99	15.64	0.634	3	8.47	8.70	8.30	0.170	3		
E-C	9.0	11.0	6.0	12.8	11.5	0.524	17.21	19.67	15.23	1.799	4	8.07	10.40	6.60	1.561	4		
E-S	2.67	4.0	1.0	12.2	10.8	0.490	15.00	18.90	13.17	2.061	9	8.67	9.90	7.30	0.830	9		
F-N	2.00	2.0	2.0	13.5	11.7	0.593	10.01	14.60	8.12	2.271	6	9.07	12.0	7.80	1.376	6		
F-C	7.17	10.0	6.0	13.4	12.8	0.189	17.03	19.41	15.64	1.322	6	7.78	8.90	6.60	0.901	6		
F-S	1.91	2.0	1.0	11.70	9.10	0.826	10.07	12.96	8.79	1.061	11	8.57	10.00	6.60	0.878	10		
G-N	2.5	2.0	3.0	12.5	12.5	0.000	7.70	7.77	7.64	0.650	2	7.40	7.50	7.30	0.100	2		
G-C	6.5	7.0	6.0	13.0	13.0	0.000	16.15	16.21	16.09	0.060	2	6.85	7.10	6.60	0.250	2		
G-S	2.0			12.60			8.01				1	8.60				1		

MURKALE

	TEMP				SALN				D.O.					
	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	NO	
A-N	7.0	10.0	2.0	3.56	18	174	199	2.76	18	194	114	99	9.16	18
A-C	9.1	9.2	9.0	0.08	8	165	193	3.12	8	169	113	104	3.27	8
A-S	9.2	9.7	8.9	0.36	6	166	192	6.18	6	107	111	104	2.07	6
B-N	8.6	12.5	4.0	3.51	18	170	190	9.85	18	107	111	104	1.89	18
B-C	9.2	9.8	8.8	0.38	10	165	190	9.69	10	107	110	102	3.07	10
B-S	8.4	10.0	7.0	1.40	12	156	193	3.25	12	90	114	84	9.03	12
C-N	8.3	9.0	7.0	0.94	18	152	169	1.61	18	94	114	64	6.63	18
C-C	9.5	9.8	9.2	0.28	8	160	171	3.39	8	96	101	90	5.92	8
C-S	9.2	11.0	7.0	1.67	18	160	170	3.94	18	91	130	69	1.62	18
D-N	7.9	11.0	3.3	3.09	24	134	192	6.94	24	94	111	69	1.76	24
D-C	10.0	10.0	9.0	0.08	10	159	172	1.48	10	103	106	95	3.79	10
D-S	6.5	10.0	3.0	3.47	12	151	170	1.70	10	99	109	86	9.23	12

ADLION

	DEPTH				TEMP				SALN				D.O.					
	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	NO	
A-N	4	6	2	6	9.4	10.0	9.0	0.43	6	181	189	170	8.18	6	104	101	3.09	6
A-C	14	20	10	8	9.3	9.9	9.2	0.25	8	211	224	209	7.58	8	100	95	3.98	8
A-S	4	5	2	6	9.4	9.5	9.2	0.14	6	195	192	174	7.72	6	104	100	2.95	6
B-N	3	3	2	6	9.4	9.5	9.2	0.17	6	175	189	169	7.12	6	104	100	2.05	6
B-C	13	19	9	10	9.7	10.2	9.3	0.36	10	193	204	162	5.51	10	107	104	2.48	10
B-S	2	4	1	12	10.1	10.6	9.2	0.66	6	166	180	151	2.38	6	105	105	2.49	6
C-N	2	3	1	12	9.4	9.5	9.3	0.09	6	164	165	163	0.62	6	106	105	2.49	6
C-C	16	23	11	8	10.7	12.0	10.1	0.75	6	205	215	157	8.54	8	98	92	6.34	8
C-S	2	5	1	12	10.3	10.5	10.2	0.12	6	166	168	163	2.50	4	110	106	1.70	6
D-N	2	3	1	12	10.8	11.0	10.3	0.33	6	165	179	153	9.67	6	110	108	1.70	6
D-C	13	17	8	10	10.4	11.0	9.8	0.59	10	200	204	198	1.87	10	105	97	5.85	10
D-S	3	3	3	6	10.3	10.5	10.2	0.12	6	181	189	170	8.18	6	98	95	2.50	4

YEAR = 72 MONTH = 12

SURFACE

	TEMP			SAL			D.O.			NO
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	
E-N	9.2			13.29			10.20			1
E-C	8.95	9.1	8.8	12.80	13.20	12.47	10.20	10.40	10.00	6
E-S	8.95	9.1	8.8	12.55	14.28	11.37	10.22	10.90	9.70	8
F-N	7.47	8.8	4.9	8.07	13.61	4.80	9.23	12.00	7.50	6
F-C	6.67	8.7	5.5	7.60	8.72	5.90	9.75	10.60	8.90	2
F-S	7.11	8.9	4.8	8.07	11.01	4.65	10.53	11.8	9.50	9
G-N	8.37	8.6	8.2	4.46	6.43	2.82	9.55	9.90	9.00	4
G-C	5.20	5.2	5.2	4.65	4.89	4.41	11.25	11.40	11.10	2
G-S	5.27	5.4	5.1	4.28	4.99	3.40	11.50	11.70	11.20	3

BOTTOM

	DEPTH			TEMP			SALN			D.O.			NO
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	
E-N	2.0			13.34			18.65	16.20	0.899	1			
E-C	8.67	11.0	5.0	17.70	18.59	11.90	18.59	11.90	2.025	6			
E-S	2.75	4.0	2.0	15.01	18.48	6.77	18.48	6.77	0.532	8			
F-N	2.0	2.0	2.0	7.46	18.50	10.20	11.88	14.94	2.165	6			
F-C	6.67	9.0	5.0	11.88	14.94	10.20	7.50	7.81	6.23	3			
F-S	2.22	4.0	1.0	7.50	7.81	6.23	6.96	8.22	5.70	9			
G-N	2.00	2.0	2.0	6.96	8.22	5.70	4.78	5.17	4.43	4			
G-C	5.50	7.0	4.0	4.78	5.17	4.43	0.303	0.303	0.303	2			
G-S	2.0	2.0	2.0	0.050	0.141		11.47	11.90	11.90	3			

YEAR = 73 MONTH = 01

	TEMP				SALN				J.U.				
	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	NU
A-N	5.4	6.5	4.0	1.07	11	14.1	1.42	11	194	111	87	6.98	11
A-C	4.9	6.0	4.0	0.85	16	16.1	1.36	16	69	100	69	9.15	16
A-S	4.4	5.0	4.1	0.42	6	130	3.57	6	64	100	38	3.52	6
B-N	7.1	8.5	5.2	1.44	12	15.5	1.49	12	85	107	58	7.31	12
B-C	5.9	6.3	5.0	0.56	10	150	1.42	10	92	101	77	6.78	10
B-S	4.8	6.5	4.3	0.79	12	136	1.13	12	100	106	88	7.48	12
C-N	8.1	26.3	6.1	5.26	13	146	1.93	13	100	106	88	7.37	13
C-C	6.4	6.3	6.0	0.24	10	142	1.50	10	94	104	86	5.43	10
C-S	5.5	11.0	3.8	2.33	18	137	1.50	12	100	106	67	6.79	18
D-N	5.5	6.5	5.9	1.19	18	136	1.62	18	100	106	67	6.79	18
D-C	5.5	6.2	5.1	0.41	10	157	1.70	10	83	96	69	7.88	10
D-S	3.3	5.5	2.0	1.62	10	126	1.62	10	93	104	69	4.65	10

319

	DEPTH				TEMP				SALN				D.O.				
	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	NU
A-N	2	3	2	6	5.5	5.8	5.2	0.24	6	154	162	146	9.55	6	67	1.72	6
A-C	12	15	10	16	5.4	6.8	4.8	0.84	16	164	211	169	4.76	16	97	0.44	16
A-S	3	5	2	6	5.5	5.9	5.0	0.37	6	178	198	165	4.52	6	104	4.86	6
B-N	4	7	2	6	6.3	7.3	6.0	0.47	6	159	161	158	1.41	6	104	4.86	6
B-C	14	21	8	10	5.9	7.0	5.2	0.59	10	184	205	162	7.81	10	90	3.50	10
B-S	2	3	2	6	6.0	7.2	5.2	0.83	6	160	170	145	0.60	6	98	2.49	6
C-N	3	4	2	7	9.2	25.0	6.3	6.85	7	165	197	157	3.89	7	98	2.63	7
C-C	10	12	6	10	6.1	6.5	6.0	0.20	10	163	191	147	7.91	8	99	3.93	10
C-S	2	3	2	6	7.8	9.0	7.2	0.85	6	143	154	144	4.32	6	100	6.78	6
D-N	2	3	2	6	7.2	9.0	6.3	0.70	6	160	162	157	2.05	6	100	6.78	6
D-C	12	15	8	10	6.3	6.5	6.0	0.20	10	163	166	153	4.88	10	83	4.03	10
D-S	3	3	3	4	5.8	5.9	5.8	0.05	4	163	163	162	0.50	4	78	3.50	4



YEAR = 73 MONTH = 01

SURFACE

	TEMP			SAL			D.O.			S D			NO					
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	S	D	NO	MEAN	MAX	MIN	S	D	NO
E-N	3.70	4.6	2.5	11.41	14.00	7.35	2.470	7	11.06	11.70	10.60	0.444	7	11.06	11.70	10.60	0.444	7
E-C	3.91	4.9	3.3	10.67	14.08	8.34	1.737	9	11.32	11.50	10.80	0.210	9	11.32	11.50	10.80	0.210	9
E-S	3.80	4.2	2.9	10.99	13.32	9.05	1.035	10	11.38	11.70	10.80	0.244	10	11.38	11.70	10.80	0.244	10
F-N	3.55	3.6	3.5	10.66	14.12	7.21	3.455	2	11.45	11.5	11.4	0.050	2	11.45	11.5	11.4	0.050	2
F-C	3.70	3.7	3.7	6.90	9.04	5.59	1.485	3	11.47	11.50	11.40	0.047	3	11.47	11.50	11.40	0.047	3
F-S	3.24	3.8	2.2	7.56	9.13	5.95	1.288	9	11.36	11.50	11.00	0.171	9	11.36	11.50	11.00	0.171	9
G-N	4.15	4.3	4.0	5.58	5.75	5.42	0.165	2	11.35	11.50	11.20	0.150	2	11.35	11.50	11.20	0.150	2
G-C																		
G-S	3.80	3.8	3.8	4.04	4.04	4.04	0.000	2	11.40	11.40	11.40	0.000	2	11.40	11.40	11.40	0.000	2

BOTTOM

	DEPTH			TEMP			SALIN			D.O.			S D			NO						
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	S	D	NO	MEAN	MAX	MIN	S	D	NO	
E-N	1.29	2.0	1.0	3.90	2.60	0.417	7	12.82	15.44	8.65	2.037	7	11.06	11.70	10.70	0.374	7	11.06	11.70	10.70	0.374	7
E-C	8.00	11.0	5.0	5.00	4.20	0.260	9	13.78	16.14	11.97	1.370	9	10.63	11.70	5.10	1.966	9	10.63	11.70	5.10	1.966	9
E-S	2.40	4.0	1.0	4.90	3.40	0.415	10	12.67	16.55	9.80	1.714	10	11.40	11.70	10.80	0.224	10	11.40	11.70	10.80	0.224	10
F-N	1.00	1.0	1.0	2.80	2.70	0.050	2	7.25	7.35	7.16	0.095	2	11.30	11.40	11.20	0.100	2	11.30	11.40	11.20	0.100	2
F-C	7.33	10.0	5.0	4.70	3.90	0.330	3	12.47	14.22	10.98	1.335	3	11.40	11.50	11.30	0.082	3	11.40	11.50	11.30	0.082	3
F-S	1.89	4.0	1.0	4.40	2.10	0.803	8	9.41	13.11	6.80	2.231	9	11.40	11.60	11.20	0.149	9	11.40	11.60	11.20	0.149	9
G-N	2.00	3.0	1.0	2.00	2.10	0.803	1	7.85	9.41	6.30	1.555	2	11.20	11.30	11.10	0.100	2	11.20	11.30	11.10	0.100	2
G-C																						
G-S	1.00	1.0	1.0	4.0	4.0	0.00	2	4.81	4.81	4.81	0.00	2	11.60	11.60	11.60	0.000	2	11.60	11.60	11.60	0.000	2



YEAR - 73 MONTH - 02

SURFACE

	TEMP			SAL			D.O.			S D			NO		
	MAX	MIN	NO	MAX	MIN	NO	MAX	MIN	NO	MAX	MIN	NO	MAX	MIN	NO
E-N	6.90	6.7	4	8.88	6.27	4	11.47	12.70	4	11.47	12.70	4	0.715	10.90	4
E-C	6.80	6.6	7	11.43	7.33	7	11.03	12.00	7	11.03	12.00	7	0.423	10.70	8
E-S	5.65	5.0	11	6.99	4.85	11	10.97	11.30	11	10.97	11.30	11	0.287	10.50	11
F-N	6.62	6.5	5	4.55	2.87	5	10.98	11.10	5	10.98	11.10	5	0.147	10.70	5
F-C	6.70	6.6	4	4.68	2.90	4	10.77	10.90	4	10.77	10.90	4	0.083	10.70	4
F-S	6.77	6.5	8	5.35	3.69	8	11.31	12.50	8	11.31	12.50	8	0.518	10.60	8
G-N	6.6		1	1.52		1	11.20		1	11.20		1			1

G-C  
G-S

BOTTOM

	DEPTH			TEMP			SALN			D.O.			S D			NO		
	MAX	MIN	NO	MAX	MIN	NO	MAX	MIN	NO	MAX	MIN	NO	MAX	MIN	NO	MAX	MIN	NO
E-N	2.00	1.0	4	7.10	6.50	4	11.58	15.36	4	7.01	3.001	4	11.07	11.50	4	0.383	10.60	4
E-C	7.43	5.0	7	6.70	6.40	7	15.65	16.76	7	14.24	0.777	7	10.96	11.10	7	0.090	10.80	7
E-S	1.82	1.0	11	6.90	4.10	11	7.76	13.45	11	4.67	3.167	11	10.90	11.30	11	0.226	10.70	11
F-N	1.00	1.0	5	6.70	6.10	5	6.11	12.02	5	3.28	3.192	5	10.92	11.50	5	0.306	10.60	5
F-C	6.50	5.0	4	6.60	6.50	4	10.87	12.27	4	9.91	0.887	4	10.77	11.00	4	0.179	10.00	4
F-S	1.63	1.0	8	6.80	6.60	8	7.88	12.55	8	4.33	2.958	8	11.30	11.90	8	0.384	10.80	8
G-N	1.0		1	6.60	0.070	1	1.46		1			1	10.90		1			1

G-C  
G-S



YEAR = 73 MONTH = 03

SURFACE

	TEMP			SAL			D.O.		
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN
E-N	12.20			10.11			13.10		
E-C	11.50			10.53			12.40		
E-S	11.15	11.2	11.1	8.92	9.25	8.60	13.20	13.60	12.80
F-N	11.50			4.24			11.00		
F-C	11.90	12.5	11.3	5.77	7.26	4.28	11.10	11.50	10.70
F-S	11.05	11.2	10.9	7.04	7.49	6.60	11.60	12.00	11.20
G-N	11.2			4.84			10.50		
G-C									
G-S									

BOTTOM

	DEPTH			TEMP			SALN			D.O.		
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN
E-N	2.00			10.70			10.34			11.40		
E-C	8.00			8.9			18.73			11.80		
E-S	1.00	2.0	1.0	11.15	11.4	10.9	10.43	10.94	9.92	12.45	12.60	12.30
F-N	1.00			11.60			6.57			10.50		
F-C	10.0	10.0	10.0	9.70	10.00	9.40	14.71	16.40	13.03	10.75	11.00	10.50
F-S	1.00	1.0	1.0	11.25	11.50	11.00	8.10	8.43	7.78	11.50	11.70	11.30
G-N	1.0			11.00			5.24			10.30		
G-C												
G-S												

YEAR = 73 MONTH = 04

DUKFALE

	TEMP			SALN			D.O.		
	MEAN	MAX	NO	MEAN	MAX	NO	MEAN	MAX	NO
A-N	15.9	17.1	15.0	135	140	3.04	94	112	75
A-C	16.0	17.5	16.0	157	167	3.26	107	112	105
A-S	16.3	16.5	16.0	152	151	1.70	107	112	103
B-N	16.1	17.6	15.2	149	137	3.24	95	112	69
B-C	16.4	17.2	16.0	148	155	4.54	110	113	104
B-S	16.4	16.3	15.9	164	128	1.59	109	112	105
C-N	15.5	16.8	14.5	144	135	4.63	109	112	105
C-C	16.6	17.5	15.5	149	115	3.60	107	109	105
C-S	16.7	20.0	15.5	140	126	4.79	98	112	69
D-N	16.4	15.0	15.0	151	100	1.24	99	112	69
D-C	18.0	16.0	17.9	113	104	3.50	110	111	109
D-S	17.4	19.0	16.5	123	95	0.65	99	110	89

GRILIN

	DEPTH			TEMP			SALN			D.O.		
	MEAN	MAX	NO	MEAN	MAX	NO	MEAN	MAX	NO	MEAN	MAX	NO
A-N	4	4	3	10.7	17.0	16.1	154	156	122	106	112	6
A-C	11	18	6	14.4	17.1	13.0	202	251	155	99	111	16
A-S	3	4	2	15.9	16.5	14.9	150	170	151	89	110	6
B-N	3	4	2	10.4	17.3	15.6	156	171	146	69	110	6
B-C	13	18	9	13.7	15.1	13.0	219	233	187	98	107	10
B-S	4	6	2	15.5	16.5	14.5	144	154	152	100	107	6
C-N	3	4	2	15.6	16.0	15.5	142	143	149	103	107	6
C-C	10	17	5	14.2	15.0	13.0	159	211	164	104	104	10
C-S	3	6	2	16.9	18.5	16.0	131	135	129	108	108	6
D-N	2	2	2	17.7	18.0	17.0	113	115	109	108	108	7
D-C	13	14	12	14.7	15.6	14.1	172	163	164	107	110	8
D-S	2	2	2	15.2	15.2	15.1	136	137	114	105	106	6

YEAR = 73 MONTH = 04

SURFACE

	TEMP			SAL			D.O.			S D			NO		
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	S	D	NO	S	D	NO
E-N	13.70	14.5	12.9	9.87	12.23	7.08	2.013	5	10.42	11.20	9.90	0.534	5		
E-C	13.65	14.4	12.9	9.04	12.17	6.55	2.196	6	11.08	12.60	10.00	0.857	6		
E-S	12.42	17.2	11.5	10.23	12.79	5.87	1.871	12	9.82	11.50	8.50	0.755	12		
F-N	15.03	16.2	14.0	5.43	6.19	4.91	0.548	3	10.07	10.80	8.80	0.899	3		
F-C	14.77	15.9	14.2	5.59	6.36	4.92	0.608	4	10.35	10.90	9.00	0.789	4		
F-S	15.35	17.1	12.1	5.36	8.18	3.31	1.615	10	9.40	10.60	8.60	0.711	10		
G-N	14.80	14.9	14.7	1.73	2.08	1.38	0.350	2	10.70	10.70	10.70	0.000	2		
G-C	15.00	15.2	14.8	0.61	1.01	0.21	0.400	2	8.75	8.80	8.70	0.050	2		
G-S	15.60			1.32			0.400	1	9.00				1		

BOTTOM

	DEPTH			TEMP			SALN			D.O.			S D			NO		
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	S	D	NO	S	D	NO
E-N	1.80	4.0	1.0	14.20	15.30	13.30	0.718	5	12.14	7.08	1.929	5	10.54	11.30	10.00	0.492	5	
E-C	8.50	10.0	7.0	12.55	12.90	12.20	0.350	6	13.58	10.36	1.278	6	10.62	11.60	9.40	0.699	6	
E-S	2.33	4.0	1.0	12.11	17.90	10.70	2.099	12	13.32	6.48	1.963	12	9.96	12.60	8.60	0.919	12	
F-N	1.33	2.0	1.0	15.20	16.00	14.50	0.616	3	5.83	4.78	0.439	3	9.73	10.40	8.50	0.873	3	
F-C	8.00	10.0	5.0	13.80	14.20	13.30	0.324	4	10.99	5.90	2.045	4	10.12	10.80	8.80	0.798	4	
F-S	2.30	4.0	1.0	14.55	16.50	11.50	1.512	10	9.10	3.87	1.652	10	9.22	10.40	8.40	0.741	10	
G-N	1.00	1.0	1.0	15.30	15.60	15.00	0.300	2	2.07	1.39	0.340	2	10.65	10.80	10.50	0.150	2	
G-C	5.50	7.0	4.0	14.80	14.90	14.70	0.100	2	2.12	0.87	0.625	2	8.35	8.46	8.30	0.050	2	
G-S	1.0			15.40			0.100	1				1	8.80					

YEAR = 75 MONTH = 05

SURFACE

	TEMP			SALN			D.U.			NO					
	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D						
A-N	20.1	21.0	19.5	0.97	12	165	175	154	0.32	12	80	90	50	0.49	12
A-C	19.2	20.2	14.2	1.34	16	170	190	161	1.09	10	62	60	63	1.43	16
A-S	19.8	20.0	19.5	0.21	6	164	171	154	0.35	6	60	63	63	1.07	6
B-N	20.2	21.5	18.5	1.34	12	150	163	154	0.40	12	73	97	63	3.76	12
B-C	19.6	20.0	19.0	0.36	10	160	162	150	1.02	10	67	72	59	4.71	10
B-S	18.7	20.2	18.0	0.92	10	153	159	151	0.12	10	82	99	68	0.31	10
C-N	19.3	20.5	19.0	0.69	12	159	163	140	4.93	12	81	99	62	4.77	12
C-C	19.3	20.3	18.7	0.54	10	155	162	147	0.13	10	66	73	62	3.97	10
C-S	19.1	19.3	18.0	0.75	16	155	199	140	7.71	13	63	93	63	3.05	16
D-N	19.2	19.8	18.2	0.72	16	137	147	135	0.27	16	63	99	63	3.65	13
D-C	19.6	19.8	19.2	0.24	3	127	135	121	0.73	3	58	70	66	1.48	6
D-S	18.5	19.3	17.5	1.05	12	136	146	119	0.32	12	73	94	61	2.25	12

DEPTH

	TEMP			SALN			D.U.			NO					
	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D						
A-N	2	2	1	6	20.7	21.5	19.3	0.77	6	175	175	174	0.47	6	69
A-C	11	13	6	16	19.3	20.0	18.6	0.42	16	189	216	159	0.11	16	64
A-S	2	2	2	6	21.5	21.6	21.3	0.12	6	165	172	163	0.25	6	66
B-N	3	4	2	6	19.1	19.5	18.8	0.29	6	174	182	170	0.44	6	66
B-C	12	15	3	10	18.6	19.3	18.0	0.50	10	200	215	201	0.46	10	61
B-S	1	2	1	10	19.9	20.5	19.3	0.60	4	161	162	159	1.50	4	67
C-N	4	7	2	5	18.5	19.2	18.0	0.50	5	160	196	162	4.70	6	64
C-C	11	15	6	10	18.4	19.3	17.5	0.57	10	200	203	195	4.43	10	61
C-S	2	2	1	12	19.5	20.0	18.5	0.45	6	166	169	152	6.39	6	66
D-N	2	2	2	6	19.4	19.6	19.2	0.17	6	145	150	136	0.10	6	60
D-C	10	12	6	8	18.5	19.5	18.0	0.20	6	153	194	119	7.34	6	56
D-S	2	2	1	6	19.5	19.8	19.3	0.22	6	130	145	129	0.80	6	63



YEAR = 73 MONTH = 05

SURFACE

	TEMP			SAL			D.O.			S D			
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	NO	S	D	NO
E-N	18.58	18.7	18.5	12.41	13.51	10.38	6.86	7.10	6.10	5	0.388	0.388	5
E-C	13.17	18.8	2.0	11.27	13.11	10.12	6.73	7.30	6.20	3	0.450	0.450	3
E-S	19.02	19.7	18.7	11.11	12.44	10.01	6.80	7.30	5.90	8	0.406	0.406	8
F-N	19.72	20.50	19.00	7.60	8.76	5.87	7.58	8.10	7.20	6	0.339	0.339	6
F-C	20.10	21.2	19.1	8.12	10.04	3.97	7.16	8.10	6.70	12	0.444	0.444	12
F-S	19.51	20.2	19.2	8.41	10.49	6.01	7.38	8.00	6.70	10	0.389	0.389	10
G-N	20.5			5.47			6.00			1			1
G-C	22.0			3.72			6.60			1			1
G-S	21.9			4.30			8.20			1			1

BOTTOM

	DEPTH			TEMP			SALIN			D.O.			S D			
	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	NO
E-N	1.40	2.0	1.0	5	18.3	18.5	17.9	5	13.03	13.79	10.23	5	6.10	6.70	5.80	5
E-C	7.33	9.0	5.0	3	17.97	18.5	17.4	3	14.46	14.76	14.22	3	6.20	6.60	5.00	3
E-S	2.88	4.0	1.0	8	18.51	19.8	17.5	8	12.48	13.79	10.71	8	6.69	7.80	6.10	8
F-N	1.17	2.0	1.0	6	19.57	20.6	19.0	6	7.65	9.02	5.86	6	7.30	7.80	6.90	6
F-C	7.75	10.0	5.0	12	17.61	20.0	1.90	12	10.73	13.84	6.40	12	6.42	7.30	5.90	12
F-S	1.70	4.0	1.0	10	19.23	19.8	18.7	10	8.81	10.80	6.65	10	7.01	7.70	6.10	10
G-N	1.0			1	19.2			1	6.81			1	5.20			1
G-C	5.0			1	20.1			1	8.79			1	6.20			1
G-S	4.0			1	19.8			1	6.74			1	6.10			1

YEAR = 73 MONTH = 06

SURFACE

	TEMP				SALG				D.U.				NU			
	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D		MEAN	MAX	MIN
A-N	25.5	27.5	24.0	1.61	16	17.5	18.1	17.0	4.39	16	30	36	71	4.97	16	16
A-C	24.9	25.6	24.0	0.65	10	17.6	18.1	17.1	5.83	10	75	77	72	1.72	10	10
A-S	25.3	26.8	24.2	0.95	10	17.8	18.3	17.1	5.19	10	79	102	74	8.05	10	10
B-N	26.3	28.0	24.7	1.43	14	17.9	18.5	17.6	2.55	14	73	102	74	9.91	14	14
B-C	25.1	25.2	25.0	0.09	6	18.1	18.5	18.0	1.25	6	75	76	74	0.94	6	6
B-S	26.5	28.5	24.3	1.73	14	18.2	18.4	18.1	1.29	14	74	77	70	3.04	14	14
C-N	23.7	26.4	20.5	2.74	14	17.4	18.4	16.5	6.38	14	74	77	70	3.04	14	14
C-C	25.0	25.3	24.7	0.25	6	18.3	18.4	18.1	1.25	6	72	77	63	3.06	6	6
C-S	28.2	32.1	25.0	2.86	13	17.9	18.1	17.7	1.76	13	69	90	55	8.94	18	18
D-N	26.3	27.5	25.2	0.92	20	17.0	17.5	16.3	4.27	20	69	90	55	8.65	20	20
D-C	25.6	26.0	25.1	0.37	6	16.7	17.4	16.2	4.99	6	68	70	64	2.19	6	6
D-S	27.0	28.8	24.9	1.82	12	17.5	17.6	16.7	3.55	12	118	670	36	6.72	12	12

BOTTOM

	DEPTH				TEMP				SALN				D.U.				NO				
	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D		MEAN	MAX	MIN	S D
A-N	4	10	1	10	24.3	24.7	23.9	0.31	10	183	190	173	5.91	10	71	55	9.10	10	10	10	10
A-C	10	12	6	10	23.3	25.4	23.3	0.64	10	402	222	183	2.54	10	68	70	61	2.93	10	10	10
A-S	3	4	2	10	24.7	25.0	24.3	0.25	10	182	187	175	4.28	10	152	670	66	9.44	10	10	10
B-N	3	4	2	8	24.6	24.8	24.2	0.24	8	184	185	185	0.83	8	147	670	68	7.66	8	8	8
B-C	13	17	10	6	23.0	24.4	23.0	0.59	6	217	242	203	7.72	6	58	68	45	9.53	6	6	6
B-S	3	4	2	8	25.1	25.8	24.5	0.47	8	184	186	183	1.09	8	72	76	64	3.48	8	8	8
C-N	3	3	2	8	25.6	27.0	24.6	0.37	8	144	187	182	1.79	8	72	76	64	3.48	8	8	8
C-C	10	14	8	6	23.5	24.0	22.4	0.75	6	207	229	190	6.31	6	50	67	39	2.19	6	6	6
C-S	2	4	1	12	25.0	25.2	24.6	0.28	6	184	190	181	4.03	6	54	64	41	9.63	6	6	6
D-N	1	2	1	8	25.0	26.0	25.5	0.21	8	160	184	183	2.80	8	50	64	41	8.75	8	8	8
D-C	13	14	12	6	24.4	24.8	24.0	0.31	6	196	202	190	4.92	6	56	57	54	1.25	6	6	6
D-S	3	4	2	6	24.7	25.2	24.5	0.33	6	176	183	167	6.50	6	60	61	59	0.82	6	6	6

YEAR = 73 MONTH = 06

SURFACE

	TEMP			SAL			D.O.			D.O.			
	MEAN	MAX	MIN	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO
E-N	25.70	25.8	25.6	14.04	15.12	12.71	0.897	4	6.65	6.90	6.40	0.180	4
E-C	25.85	26.1	25.6	13.59	14.53	11.11	1.129	4	6.40	6.80	5.70	0.342	4
E-S	26.37	27.3	25.4	13.24	14.72	11.22	1.223	9	6.63	7.10	6.30	0.221	9
F-N	26.47	27.6	24.5	8.90	9.81	6.83	1.002	6	6.03	6.30	5.80	0.170	6
F-C	26.80	26.8	26.8	9.21	9.21	9.21	0.000	2	6.10	6.10	6.10	0.000	2
F-S	25.50	26.0	25.0	10.57	11.79	8.91	0.910	8	6.00	6.20	5.70	0.166	8
G-N	25.0			7.03				1	5.80				1
G-C	25.33	25.5	25.0	7.57	9.22	6.20	1.249	3	6.33	6.40	6.30	0.047	3
G-S	25.63	26.0	25.0	6.32	7.26	5.98	0.543	4	6.42	6.60	6.30	0.109	4

BOTTOM

	DEPTH			TEMP			SALN			D.O.			
	MEAN	MAX	MIN	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO
E-N	1.25	2.0	1.0	25.60	25.8	25.4	0.200	4	16.53	12.69	1.592	0.141	4
E-C	8.00	11.0	6.0	23.40	23.7	23.1	0.300	4	18.70	16.67	0.775	0.345	4
E-S	1.78	4.0	1.0	25.80	26.6	24.7	0.636	9	15.57	12.08	0.887	0.250	9
F-N	2.00	4.0	1.0	26.12	26.9	24.5	0.843	6	12.31	6.98	1.746	0.351	6
F-C	9.0			25.3				1	13.11				1
F-S	1.63	4.0	1.0	25.20	25.4	25.0	0.200	2	14.03	8.73	1.633	0.332	8
G-N	2.0			24.0				1	11.48				1
G-C	5.00	7.0	3.0	23.67	24.0	23.0	0.471	3	11.97	8.56	1.387	1.580	3
G-S	2.25	4.0	1.0	24.75	25.5	24.0	0.559	4	9.09	5.89	1.229	0.480	4

YEAR = 73 MONTH = 07

SURFACE

	TEMP			SALN			D.O.					
	MEAN	MAX	MIN	S	D	NO	MEAN	MAX	MIN	S	D	NO
A-N	26.6	27.6	26.0	0.74		16	190	200	180	6.21		16
A-C	26.1	26.2	26.0	0.06		10	189	199	180	5.04		10
A-S	26.1	26.4	26.0	0.15		10	191	195	187	3.16		10
B-N	26.4	27.2	25.4	0.72		14	196	200	185	2.15		14
B-C	25.7	25.8	25.7	0.05		4	189	199	192	0.00		4
B-S	26.2	31.2	25.7	2.60		14	201	203	197	2.31		14
C-N	26.2	25.5	25.5	0.43		10	200	201	200	0.40		10
C-C	25.8	25.9	25.8	0.05		4	197	197	197	0.00		4
C-S	29.5	33.0	25.6	2.96		18	199	203	193	3.94		18
D-N	27.5	32.3	23.0	3.42		20	196	200	191	3.16		20
D-C	22.8	25.0	21.3	1.60		6	191	205	180	0.33		6
D-S	26.7	29.3	21.5	2.91		12	194	196	190	2.61		12

BOTTOM

	DEPTH			TEMP			SALN			D.O.						
	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	S	D	NO	MEAN	MAX	MIN	S	D	NO
A-N	3	4	2	10	25.4	25.7	25.2	0.19		10	189	200	184	6.05		10
A-C	15	18	9	10	24.5	25.0	23.5	0.52		10	220	232	200	1.37		10
A-S	4	6	2	10	26.0	26.6	25.4	0.46		10	194	198	190	2.61		10
B-N	3	3	2	8	25.4	25.8	25.0	0.30		6	159	201	195	2.45		6
B-C	10	10	10	4	25.1	25.4	24.8	0.30		4	202	205	201	1.00		4
B-S	4	6	1	8	25.6	26.0	25.1	0.32		8	200	202	199	1.30		8
C-N	2	5	1	10	24.8	25.2	24.5	0.35		4	202	202	201	0.50		4
C-C	11	11	10	4	24.5	24.9	24.2	0.35		4	194	207	180	3.50		4
C-S	4	5	2	6	25.2	25.5	24.6	0.31		6	202	209	193	4.97		6
D-N	2	3	1	14	23.3	25.2	22.5	1.12		8	198	201	194	2.74		8
D-C	12	14	8	6	21.6	25.0	19.7	2.43		6	211	222	191	4.36		6
D-S	3	5	2	6	25.8	26.0	25.5	0.24		6	192	197	186	3.86		6

YEAR = 73 MONTH = 07

SURFACE

	TEMP			SAL			D.O.			NO				
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	S D	NO
E-N	30.05	31.5	28.3	15.81	16.19	14.97	0.489	4	5.62	6.30	4.10	0.893	4	4
E-C	29.60			13.24				1	6.40				1	1
E-S	29.73	31.2	29.3	14.09	15.00	13.70	0.435	10	5.46	6.70	1.40	1.937	10	10
F-N	28.93	30.10	27.0	12.15	13.20	10.86	0.900	6	5.93	6.60	5.50	0.427	6	6
F-C	29.23	30.3	27.5	11.95	13.23	9.79	1.301	9	5.61	6.80	3.30	1.135	9	9
F-S	26.99	27.4	26.3	12.64	16.66	11.18	1.669	8	6.42	7.00	6.00	0.295	8	8
G-N	27.2			9.46				1	5.10				1	1
G-C	27.5			10.66				1	6.00				1	1
G-S	28.60	30.2	27.0	9.28	9.93	8.64	0.645	2	5.10	5.90	4.30	0.800	2	2
P	28.05	28.4	27.8	7.46	7.84	6.97	0.424	4	3.53	3.9	3.0	0.450	4	4

BOTTOM

	DEPTH			TEMP			SALN			D.O.			NO					
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	S D	NO	
E-N	1.25	2.0	1.0	29.1	29.9	28.0	0.791	4	15.79	16.29	14.68	0.649	4	6.05	6.50	5.50	0.364	4
E-C	9.00			29.70				1	15.97				1	6.00				1
E-S	1.80	4.0	1.0	20.62	31.2	28.7	0.685	10	14.50	14.90	13.99	0.288	10	5.86	6.40	3.70	0.739	10
F-N	1.50	4.0	1.0	28.82	30.4	27.0	1.268	6	12.53	13.31	10.91	0.802	6	6.27	7.10	5.70	0.556	6
F-C	7.33	10.0	4.0	28.52	30.0	26.9	0.998	9	12.68	14.00	10.67	1.228	9	5.30	6.40	3.50	0.970	9
F-S	1.13	2.0	1.0	26.76	27.6	26.1	0.534	8	12.25	13.20	11.33	0.661	8	6.27	6.80	5.50	0.380	8
G-N	1.0			26.50				1	9.80				1	5.90				1
G-C	4.0			26.90				1	10.91				1	5.80				1
G-S	1.00	1.0	1.0	28.35	29.9	26.8	1.550	2	9.84	10.52	9.17	0.675	2	4.95	5.50	4.40	0.550	2
P	4.75	6.0	3.0	26.9	27.3	26.3	0.489	4	8.57	9.57	7.87	0.769	4	3.8	4.00	3.60	0.230	4

YEAR = 73 MONTH = 03

SURFALL

	TEMP				SALN				D.C.			
	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D
A-N	27.1	32.0	23.5	3.80	185	187	151	2.96	77	97	64	5.22
A-C	25.5	26.8	24.8	0.77	183	188	179	3.52	69	77	62	6.50
A-S	26.8	27.3	26.0	0.47	180	192	179	1.10	74	79	66	3.42
B-N	28.2	31.8	22.4	3.28	184	189	179	3.96	74	79	66	3.79
B-C	27.6	28.0	27.3	0.29	186	189	181	3.74	64	90	75	8.05
B-S	27.6	28.2	27.0	0.50	186	189	181	3.45	73	82	62	7.82
C-N	26.7	28.3	24.8	1.31	192	201	182	6.06	71	82	58	7.90
C-C	25.3	26.0	24.8	0.52	189	190	189	0.47	65	73	56	6.54
C-S	28.1	28.4	27.0	0.49	190	191	189	0.74	79	105	55	8.42
D-N	29.7	33.5	26.4	2.89	190	194	186	2.84	82	125	55	8.51
D-C	26.5	27.0	26.2	0.34	196	199	195	2.49	74	92	41	3.16
D-S	26.5	26.9	25.5	0.52	190	195	186	4.22	78	97	68	8.65

	NO	MEAN	MAX	MIN	S D	NO
A-N	10	185	187	151	2.96	16
A-C	10	183	188	179	3.52	10
A-S	10	180	192	179	1.10	10
B-N	14	184	189	179	3.96	14
B-C	6	186	189	181	3.74	6
B-S	14	186	189	181	3.45	14
C-N	17	192	201	182	6.06	17
C-C	6	189	190	189	0.47	6
C-S	12	190	191	189	0.74	18
D-N	20	190	194	186	2.84	20
D-C	6	196	199	195	2.49	6
D-S	12	190	195	186	4.22	12

BOLIDS

	DEPTH				TEMP				SALN				D.U.			
	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D
A-N	2	4	2	10	24.1	24.5	23.2	0.46	185	187	182	4.52	69	92	59	3.52
A-C	12	18	6	10	23.5	26.0	20.8	1.76	222	260	187	4.53	84	92	32	3.14
A-S	3	5	2	10	27.3	28.0	26.3	0.60	182	188	179	3.26	78	97	68	9.27
B-N	3	5	2	8	23.2	26.0	21.2	2.11	186	188	183	1.79	76	90	68	7.43
B-C	11	12	9	6	26.6	27.0	26.1	0.39	192	193	192	0.47	59	60	56	0.94
B-S	2	4	1	14	27.5	27.8	27.1	0.27	185	189	182	3.08	75	80	76	2.00
C-N	2	6	1	17	24.7	26.1	21.6	1.46	196	201	182	6.72	76	80	66	4.89
C-C	11	15	6	6	24.7	25.5	23.7	0.74	216	236	193	7.68	52	69	36	3.52
C-S	2	3	1	12	27.3	27.3	27.0	0.33	191	191	190	0.47	59	76	66	4.71
D-N	2	5	1	14	28.3	28.6	28.0	0.23	193	197	191	2.28	69	76	66	4.35
D-C	12	15	9	6	28.4	28.7	28.3	0.14	204	206	202	1.70	63	71	54	6.97
D-S	4	5	2	6	26.0	26.2	25.9	0.12	199	202	194	3.50	66	72	63	4.03

YEAR = 73 MONTH = 08

SURFACE

	TEMP			SAL			D.O.			NO			
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	NO
E-N	28.50	28.8	28.2	18.12	18.14	18.12	6.27	6.60	6.00	0.009	0.009	0.277	4
E-C	28.68	29.2	28.2	15.86	17.19	15.08	6.62	6.80	6.40	0.932	0.932	0.133	5
E-S	29.79	30.4	28.5	16.18	17.43	15.38	6.31	6.80	5.90	0.647	0.647	0.284	11
F-N	29.50	29.8	28.9	12.65	13.27	12.20	6.45	7.00	5.50	0.396	0.396	0.568	4
F-C	28.87	29.3	28.5	13.33	14.27	11.58	6.51	6.90	5.60	0.820	0.820	0.419	7
F-S	29.20	29.6	28.9	15.86	15.86	12.65	6.32	6.60	5.80	1.137	1.137	0.286	9
G-N	29.03	30.1	28.4	11.36	11.85	10.71	6.00	6.50	5.40	0.478	0.478	0.455	3
G-C	28.8			10.3			6.10						1
G-S	28.96	29.0	28.9	9.37	10.97	8.57	5.27	5.30	5.20	1.385	1.385	0.057	3
P													

BOTTOM

	DEPTH			TEMP			SALN			D.O.			NO		
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN
E-N	1.25	2.0	1.0	27.82	28.0	27.3	19.14	19.14	18.40	0.311	0.311	5.62	5.90	4.80	4
E-C	9.20	10.0	8.0	28.00	28.3	27.7	19.51	19.51	18.08	0.614	0.614	5.60	6.00	5.10	5
E-S	2.55	4.0	1.0	28.86	30.5	27.7	18.78	18.78	16.15	0.760	0.760	6.44	7.40	5.60	11
F-N	2.00	5.0	1.0	29.30	29.8	28.9	13.19	13.19	12.09	0.389	0.389	6.45	7.00	6.10	4
F-C	6.43	8.0	6.0	28.10	28.6	27.8	17.22	17.22	11.27	2.362	2.362	5.64	5.90	5.50	9
F-S	1.11	2.0	1.0	29.13	29.5	28.8	16.21	16.21	12.95	1.112	1.112	6.33	6.80	5.60	7
G-N	2.33	4.0	1.0	28.47	29.0	27.9	13.13	13.13	11.56	0.641	0.641	5.67	6.30	4.80	3
G-C	1.0			28.4			11.95					5.70			1
G-S	4.50	5.0	4.0	28.40	28.6	28.0	11.55	11.55	9.57	1.114	1.114	4.766	4.80	4.70	3
P															

YEAR = 73 MONTH = 09

SURFACE

	TEMP			SALN			D.O.			NO			
	MEAN	MAX	MIN	S	D	NO	MEAN	MAX	MIN		S	D	NO
A-N	25.0	23.0	25.0	2.36	1.32	16	195	177	194	112	76	7.24	16
A-C	24.1	24.5	23.5	0.38	2.06	8	193	199	194	79	64	5.41	8
A-S	24.0	24.5	23.5	0.33	0.00	10	199	199	199	78	64	5.55	10
B-N	25.7	27.8	25.3	1.86	1.48	14	196	197	194	78	64	4.75	14
B-C	23.7	23.8	23.0	0.09	0.00	6	197	197	197	72	69	1.25	6
B-S	26.8	27.5	25.2	0.75	2.53	14	193	196	190	83	65	9.18	14
C-N	23.9	24.0	23.0	0.47	2.65	14	194	198	191	89	65	9.16	14
C-C	24.1	25.5	23.2	1.02	0.47	6	194	194	193	71	72	0.82	6
C-S	27.3	31.0	24.9	2.39	4.03	18	193	193	194	103	60	2.12	18
D-N	24.9	27.8	23.0	1.90	0.52	20	186	196	179	103	60	2.56	20
D-C	23.4	23.9	23.0	0.39	2.05	6	192	194	189	73	67	2.19	6
D-S	26.1	26.9	23.0	2.79	1.40	12	147	192	106	68	49	6.04	12

BOTTOM

	DEPTH			TEMP			SALN			D.O.					
	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	S	D	NO	MEAN	MAX	MIN	S	D
A-N	2	3	2	10	23.6	24.0	23.0	0.57	1.0	196	197	195	60	5.96	10
A-C	10	15	6	8	24.5	25.0	23.2	0.75	8	198	199	196	64	2.68	8
A-S	3	4	2	10	24.4	25.0	24.0	0.42	10	199	200	199	49	6.48	10
B-N	3	3	2	8	24.0	24.5	23.8	0.29	6	197	197	197	49	7.14	6
B-C	15	18	13	6	24.0	24.0	24.0	0.00	6	196	196	197	58	0.94	6
B-S	3	6	2	8	25.1	26.0	24.5	0.65	8	197	198	195	66	2.12	8
C-N	2	3	1	14	23.2	23.8	23.0	0.33	8	196	197	194	65	2.12	8
C-C	11	13	7	6	23.5	24.5	23.0	0.71	6	196	197	195	68	0.82	6
C-S	4	4	4	6	25.2	25.5	25.0	0.24	6	195	195	194	60	3.11	6
D-N	2	2	2	8	23.2	23.4	23.0	0.14	8	195	197	192	65	5.35	8
D-C	11	12	10	6	23.5	23.8	23.0	0.36	6	193	195	192	67	0.47	6
D-S	2	3	2	6	23.1	23.2	23.0	0.09	6	192	193	191	69	1.89	6



YEAR = 73 MONTH = 10

SUREALL

	TEMP			SALIN			D.O.			NO					
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN						
A-N	15.6	18.0	11.5	3.15	16	195	200	197	1.00	16	79	85	75	4.59	16
A-C	18.2	18.5	18.0	0.24	10	199	205	194	3.76	10	74	77	70	2.50	10
A-S	18.7	19.0	18.0	0.40	10	199	201	195	2.53	10	78	87	73	3.99	10
B-N	15.5	18.0	13.0	2.19	14	197	194	195	1.50	14	78	87	75	3.60	14
B-C	17.9	18.0	17.8	0.09	6	197	198	196	0.94	6	77	78	76	0.94	6
B-S	17.0	19.0	15.0	1.75	14	197	200	194	2.80	14	75	77	73	1.67	14
C-N	16.6	18.2	15.5	0.99	14	196	199	189	3.33	14	75	77	73	1.67	14
C-C	18.2	18.7	18.0	0.33	6	188	193	180	3.91	6	73	75	71	1.63	6
C-S	18.1	19.5	16.5	1.26	18	197	201	190	4.08	18	77	82	82	3.20	18
D-N	15.1	18.5	12.5	2.84	20	191	198	178	6.32	20	77	82	85	4.07	20
D-C	16.0	18.0	16.0	0.00	6	189	194	182	5.10	6	74	77	72	2.05	6
D-S	15.4	18.8	14.5	0.92	12	191	194	189	1.83	12	75	79	70	2.14	12

MULION

	DEPTH			TEMP			SALIN			D.O.			NO		
	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN	MEAN	MAX	MIN			
A-N	3	5	1	10	17.9	18.0	17.5	0.20	10	202	206	197	3.14	10	76
A-C	10	14	8	10	18.2	19.0	17.8	0.42	10	205	208	202	2.84	10	74
A-S	2	3	1	10	18.7	19.0	18.0	0.38	10	200	205	195	3.16	10	75
B-N	2	3	1	8	17.0	17.7	16.0	0.60	8	196	198	195	1.09	8	75
B-C	14	15	12	6	17.5	17.5	17.5	0.00	6	205	206	204	0.82	6	70
B-S	2	5	1	14	18.1	19.0	17.5	0.65	8	195	197	195	1.58	8	74
C-N	2	2	1	14	17.4	18.5	17.0	0.62	6	196	198	192	2.29	8	74
C-C	7	9	4	6	18.5	19.0	18.0	0.41	6	198	200	192	3.27	6	72
C-S	2	5	1	18	18.5	19.0	18.0	0.41	6	192	193	190	1.25	6	77
D-N	2	3	1	9	16.6	18.0	15.0	1.19	8	188	195	178	7.46	8	76
D-C	11	14	8	6	15.8	17.0	16.5	0.24	6	233	247	218	1.90	6	68
D-S	2	5	1	12	16.2	16.5	16.0	0.24	6	193	197	187	4.50	6	72

YEAR = 73 MONTH = 11

	TEMP				SALIN				D.U.				
	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	NO
A-N	13.9	13.9	13.8	0.05	210	218	214	1.70	90	92	76	1.70	12
A-C	13.7	14.5	13.3	0.34	217	220	219	1.58	93	89	75	3.16	16
A-S	14.4	15.0	14.1	0.40	220	221	219	0.94	81	82	78	1.89	6
B-N	14.1	14.4	13.9	0.25	214	215	213	1.00	84	93	78	4.71	12
B-C	13.7	14.1	13.5	0.27	222	246	215	2.12	89	96	86	3.50	10
B-S	13.9	15.0	12.8	1.07	213	215	213	0.75	87	88	82	1.66	12
C-N	13.3	13.9	13.0	0.38	213	215	212	1.15	87	88	82	1.66	12
C-C	13.1	13.6	12.9	0.25	215	220	211	3.01	86	95	80	5.69	10
C-S	14.6	15.7	12.9	1.53	212	214	210	1.75	89	99	78	7.63	18
D-N	12.2	14.4	9.4	2.03	206	220	191	0.88	89	99	78	7.03	18
D-C	12.9	13.8	12.0	0.81	155	213	175	7.01	82	87	75	4.32	6
D-S	13.3	14.1	12.2	0.59	204	214	182	0.17	83	93	77	4.26	12

BOTTOM

	DEPTH				TEMP				SALIN				D.U.				
	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	NO
A-N	3	5	2	6	14.6	14.9	14.4	0.24	217	217	216	0.47	90	99	80	9.17	6
A-C	11	17	7	16	14.2	14.5	13.8	0.27	253	242	224	1.72	80	87	63	5.50	16
A-S	4	5	3	6	15.0	15.5	14.2	0.51	217	222	210	5.10	83	93	77	5.80	6
B-N	2	3	2	6	15.1	15.3	14.9	0.17	215	216	214	0.94	83	93	77	5.80	6
B-C	10	15	3	10	14.0	14.8	13.4	0.47	229	240	216	7.83	84	93	77	6.11	10
B-S	2	3	2	6	13.4	15.0	12.2	1.16	213	214	213	0.47	85	88	80	2.69	6
C-N	2	3	2	6	14.3	14.8	13.9	0.29	214	215	214	0.47	85	88	80	2.69	6
C-C	8	10	7	10	13.5	13.8	12.6	0.47	218	227	214	4.71	82	84	80	1.50	10
C-S	2	3	1	6	13.5	15.4	12.2	1.39	213	214	212	0.94	85	94	77	6.03	6
D-N	2	3	1	6	14.0	15.4	11.7	1.84	203	209	192	7.59	85	94	77	6.03	6
D-C	12	16	9	8	13.0	13.9	11.9	0.93	222	223	219	1.84	83	88	79	3.27	8
D-S	3	5	2	6	13.9	15.3	11.4	1.75	201	214	189	4.82	85	87	83	1.83	6

YEAR = 73 MONTH = 12

SURFACE

	TEMP				NO	SALN				NO	D.O.				
	MEAN	MAX	MIN	S D		MEAN	MAX	MIN	S D		MEAN	MAX	MIN	S D	NO
A-N	5.5	7.8	5.0	2.35	12	217	219	215	1.21	12	104	108	98	4.02	12
A-C	8.6	9.0	8.2	0.35	14	219	220	217	1.12	14	96	106	95	4.25	14
A-S	8.2	8.3	8.2	0.05	6	216	220	210	4.50	6	101	105	97	2.54	6
B-N	5.7	5.7	5.7	0.00	6	217	217	217	0.00	6	101	105	97	2.54	6
B-S	7.9	7.9	7.9	0.00	6	212	212	212	0.00	6	100	100	99	0.50	6
C-N	9.6	9.6	9.6	0.00	3	211	211	211	0.00	3	99	99	99	0.00	3
D-N	4.1	4.1	4.1	0.00	6	197	197	197	0.00	6	102	106	97	2.69	6
D-S	5.5	5.5	5.5	0.00	6	193	193	193	0.00	6	99	100	96	1.60	6

BULLOCK

	DEPTH				NO	TEMP				NO	SALN				NO	D.O.			
	MEAN	MAX	MIN	NJ		MEAN	MAX	MIN	S D		MEAN	MAX	MIN	S D		MEAN	MAX	MIN	S D
A-N	2	3	1	6	7.9	8.0	7.4	0.26	6	216	216	218	0.00	6	102	106	97	2.69	6
A-C	13	16	11	14	8.3	8.7	7.8	0.26	14	220	222	219	1.05	14	96	100	95	4.40	14
A-S	3	3	2	6	8.0	8.2	7.8	0.16	6	217	220	210	4.71	6	99	100	96	1.60	6
C-N	1	1	1	3															

YEAR = 74 MONTH = 01

	TEMP				SALN				SURFACE				D.O.							
	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO
A-N	8.2	9.9	7.2	1.05	12	135	157	111	2.02	12	131	159	105	4.16	12	131	159	105	4.16	12
A-C	9.2	10.0	8.7	0.58	14	157	159	154	1.77	14	107	113	103	3.90	14	107	113	103	3.90	14
A-S	9.0	10.0	4.9	1.71	7	161	176	156	6.48	7	193	207	105	5.96	7	193	207	105	5.96	7
B-N	9.1	10.4	8.7	0.50	12	154	159	111	3.03	12	162	207	94	5.49	12	162	207	94	5.49	12
B-C	9.4	10.2	9.0	0.43	10	159	160	157	1.10	10	102	106	91	5.95	10	102	106	91	5.95	10
B-S	9.5	12.0	7.9	1.80	12	158	191	117	0.87	12	126	149	96	2.53	12	126	149	96	2.53	12
C-N	8.9	10.0	8.0	0.88	12	140	159	120	3.51	12	126	149	96	2.53	12	126	149	96	2.53	12
C-C	9.4	10.0	9.0	0.42	10	158	167	151	5.60	10	100	114	89	6.00	10	100	114	89	6.00	10
C-S	10.2	12.8	7.4	2.23	18	124	161	98	4.75	16	120	146	81	2.06	16	120	146	81	2.06	16
D-N	8.7	10.0	7.9	0.85	18	121	144	107	5.50	18	120	146	81	2.06	18	120	146	81	2.06	18
D-C	10.1	10.2	10.0	0.09	8	139	145	126	7.76	8	99	102	90	4.97	8	99	102	90	4.97	8
D-S	8.9	10.2	7.8	1.12	12	126	144	114	2.70	12	103	139	97	4.59	12	103	139	97	4.59	12

	DEPTH				TEMP				SALN				D.O.						
	MEAN	MAX	MIN	NO	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO	MEAN	MAX	MIN	S D	NO
A-N	4	5	3	6	6.8	9.0	6.6	0.17	6	158	159	157	0.54	6	102	113	81	9.93	6
A-C	12	25	6	14	8.4	10.3	7.3	0.98	14	168	216	156	1.91	14	97	104	85	6.00	14
A-S	4	6	2	7	8.7	9.5	5.0	1.55	7	165	176	156	7.20	7	102	108	97	3.38	7
B-N	3	4	1	6	9.8	10.0	9.4	0.26	6	158	159	157	0.82	6	101	105	97	2.52	6
B-C	12	14	11	10	8.4	9.5	8.0	0.56	10	169	214	159	0.45	10	85	95	78	6.28	10
B-S	2	3	1	12	10.9	11.5	9.9	0.87	6	161	161	160	0.47	6	98	105	83	7.37	6
C-N	3	6	1	6	9.8	10.3	8.8	0.88	6	164	177	158	0.56	6	98	105	83	7.37	6
C-C	12	23	7	10	9.0	9.5	8.5	0.33	10	181	214	153	2.79	10	88	95	81	5.46	10
C-S	2	4	1	12	10.5	11.1	9.5	0.71	6	160	161	159	1.41	6	87	99	72	1.22	6
D-N	2	4	1	12	10.1	10.2	10.0	0.00	6	146	149	144	2.36	6	87	99	72	1.22	6
D-C	13	14	10	8	9.2	9.9	8.3	0.40	8	171	183	149	3.97	8	91	99	87	4.92	8
D-S	3	5	2	6	10.1	10.2	10.0	0.00	6	146	155	130	1.15	6	99	100	97	1.25	6

YEAR = 74 MONTH = 02

SURFACE

	TEMP			SALN			D.U.					
	MEAN	MAX	MIN	S	D	NO	MEAN	MAX	MIN	S	D	NO
A-N	7.7	9.7	5.4	1.99		12	167	171	158	4.73		12
A-C	5.4	6.0	5.0	0.37		16	164	166	161	1.60		16
A-S	4.7	5.1	4.3	0.37		5	175	176	169	3.12		5
B-N	7.3	8.7	5.8	1.42		12	162	164	158	2.43		12
B-C	5.7	6.1	5.2	0.32		10	162	164	160	1.53		10
B-S	8.0	10.2	5.0	2.24		12	169	170	159	7.76		12
C-N	6.1	6.4	5.8	0.19		12	150	162	153	2.73		12
C-C	5.8	6.8	5.0	0.61		12	152	160	146	4.22		12
C-S	8.0	12.8	5.2	3.40		13	161	173	150	9.04		18
D-N	7.3	9.2	6.2	1.13		18	160	178	154	9.24		18
D-C	6.4	6.5	6.3	0.08		6	167	171	162	3.68		6
D-S	8.0	9.6	6.2	1.60		12	159	163	143	7.21		12

AUXILIARY

	DEPTH			TEMP			SALN			D.U.			
	MEAN	MAX	MIN	NO	S	D	NO	MEAN	MAX	MIN	S	D	NO
A-N	4	5	3	6	5.7	0.26	6	162	165	157	3.56		6
A-C	12	18	3	16	4.8	0.36	18	188	224	168	7.12		18
A-S	3	4	2	5	4.7	0.11	5	172	176	168	3.61		5
B-N	3	4	2	6	5.8	0.41	6	174	207	158	3.10		6
B-C	12	13	10	10	4.7	0.54	10	193	212	174	3.53		10
B-S	1	2	1	12	5.0	0.57	6	161	164	159	2.36		6
C-N	3	3	2	6	6.1	0.22	6	166	173	157	0.60		6
C-C	13	24	6	12	2.6	0.52	12	182	206	161	7.50		12
C-S	3	5	1	12	5.0	0.19	6	153	154	121	1.25		6
D-N	2	3	2	6	5.7	0.14	6	170	179	165	3.58		6
D-C	12	16	7	6	6.0	0.09	6	177	178	170	0.94		6
D-S	2	3	1	12	6.2	0.05	6	158	168	143	0.80		6

YEAR = 74 MONTH = 03

SURFACE

	TEMP				SALIN				D.U.						
	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	NO		
A-N	11.0	14.5	7.3	3.54	12	179	181	177	1.88	12	100	102	98	1.73	12
A-C	8.7	9.5	8.3	0.44	16	134	135	132	0.93	16	98	104	81	0.82	16
A-S	8.7	8.9	8.3	0.16	6	155	137	133	1.63	6	98	99	90	4.07	6
B-N	11.9	14.8	8.9	2.85	12	163	184	162	0.90	12	99	103	90	0.03	12
B-C	9.3	10.0	9.2	0.30	10	151	132	129	1.26	10	91	98	81	0.49	10
B-S	11.2	12.6	9.2	1.45	12	178	179	175	1.41	12	100	111	83	3.28	12
C-N	11.2	13.6	8.0	2.46	12	179	182	177	2.15	12	100	111	83	3.28	12
C-C	9.2	9.5	9.0	0.13	10	175	151	177	1.90	10	94	97	90	2.94	10
C-S	13.0	17.2	9.4	3.34	18	174	173	159	3.54	18	103	110	84	7.14	18
D-N	12.0	14.5	9.5	1.97	18	171	175	167	2.27	18	103	110	84	7.14	18
D-C	9.3	9.5	9.2	0.11	8	160	152	159	1.30	8	97	103	90	6.50	8
D-S	10.3	11.5	9.2	1.05	12	161	156	154	2.37	12	103	109	99	3.72	12

DEPTH

	TEMP				SALIN				D.U.					
	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	MEAN	MAX	MIN	S D	NO	
A-N	3	5	1	12	7.4	7.8	7.2	0.28	6	181	184	180	1.89	6
A-C	13	15	11	16	8.9	9.8	8.7	0.54	16	202	209	183	7.94	16
A-S	3	3	3	6	8.9	9.0	8.8	0.09	6	185	185	184	0.47	6
B-N	3	3	2	6	8.6	9.2	7.6	0.73	6	184	184	183	0.47	6
B-C	18	24	11	10	8.8	9.2	8.1	0.43	10	205	217	192	0.97	10
B-S	2	3	2	6	9.1	9.4	9.0	0.19	6	177	180	176	1.59	6
C-N	2	3	2	6	8.3	8.9	7.6	0.45	6	183	184	182	0.54	6
C-C	11	20	6	10	9.0	9.2	8.8	0.18	10	187	192	181	4.22	10
C-S	2	5	1	12	10.0	12.2	8.8	1.58	6	175	177	175	0.82	6
D-N	2	2	1	6	9.3	9.7	9.0	0.14	6	170	175	166	3.06	6
D-C	11	12	9	8	9.1	9.2	9.0	0.03	6	185	194	183	4.87	8
D-S	2	3	1	6	9.5	9.6	9.0	0.35	6	158	162	153	4.24	6

VITA

Chao, Labbish Ning

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