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Hedgepeth, M. Y., Doyle, R. T., & Stehlik, L. (1979) A Laboratory Analysis of Kepone Depuration by Spot, Leiostomus xanthurus. Special Reports in Applied Marine Science and Ocean Engineering (SRAMSOE) No. 229. Virginia Institute of Marine Science, College of William and Mary. https://doi.org/10.21220/ V5KJ05

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A LABORATORY ANALYSIS OF KEPONE DEPURATION BY SPOT, LEIOSTOMUS XANTHURUS

by Marion Y. Hedgepeth, Robert T. Doyle, and Linda L. Stehlik



Special Report In Applied Marine Science and Ocean Engineering #229

A Laboratory Analysis of Kepone Depuration by Spot, Leiostomus xanthurus

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ACKNOWLEDGEMENTS

This research was conducted at the Virginia Institute of Marine Science with financial support received through the Commonwealth of Virginia from funds endowed to the State by Allied Chemical, Inc. for the purposes of Kepone research.

We wish to thank Drs. J. V. Merriner, H. M. Austin and M. E. Bender for receiving this report. Also, we acknowledge the assistance of Barbara Taylor and Nancy Peters in typing the manuscript.

INTRODUCTION

Dectectable residue levels of the pesticide Kepone have been found in resident and migratory finfishes from the James River, Virginia (Bender et al., 1977). As a result, the James River was closed to commercial finfishing in early 1976 (with the exceptions of channel catfish and American shad for a short period of time). In addition, the United States Food and Drug Administration established an action level of 0.3 ppm of Kepone in finfishes utilized for human consumption.

Residue levels of Kepone in fishes such as spot, Atlantic croaker, bluefish, striped bass and American shad were investigated to determine if these migratory species present a health hazard to the public in areas beyond the James River system. Bender et al. (1977) found that residue levels in finfishes were dependent upon the species of fish and the length of residence in the James River. Also, they maintained that residue levels in finfishes declined as distance from the James was increased.

In 1977, additional Kepone studies were begun at the Virginia Institute of Marine Science (VIMS) to determine the rates of Kepone depuration in contaminated fishes from the James River. In a laboratory analysis of Kepone depuration by Atlantic croaker, <u>Micropogonias undulatus</u>, Doyle et al. (In Press) observed a significant drop in Kepone concentration in the 24th week sample. Furthermore, it was noted that this significant change in mean residue levels coincided with a rise in the ambient water temperature to above 15°C; however, additional studies were needed to confirm this relationship. In our study we chose to observe the effect of temperature on the rate of Kepone depuration by contaminated spot, Leiostomus xanthurus, from the James River.

Bender et al. (1977) reported a mean Kepone level of 0.81 ppm in spot from the James River and the lower Chesapeake Bay. This was attributed to the biomagnification of Kepone through the food chain and/or direct uptake from the water (Schimmel and Wilson, 1977). Bahner et al. (1977) confirmed this belief in a study in which spot were fed live mysids which had grazed on Kepone laden brine shrimp. Consequently, the spot accumulated concentrations of Kepone near that in their diet. Spot which had been exposed to Kepone in water were able to reduce Kepone residues in their tissues to 30-50 percent within 24-28 days in Kepone free water.

MATERIALS AND METHODS

On November 11, 1977, approximately 550 spot were obtained from the lower James River with a 30 foot semi-balloon trawl. They ranged in size from 86 mm in fork length and 8 grams in total weight to 233 mm in fork length and 165 grams in total weight. They were transported to VIMS and distributed randomly to four circular four-foot tanks (approximately 200 gallons each). All tanks were supplied with Kepone-free York River water in a flowthrough system and strong aeration. Fish were fed chopped Keponefree squid daily (8-12 percent body weight).

After one month of acclimation at ambient river temperature, three of the tanks were heated with water from a large header

tank equipped with two 220-volt heaters. Heated and unheated water were combined in mixing boxes, and flow rates were adjusted so that temperatures were maintained at approximately 22°, 17° and 12°, respectively, in the three experimental tanks. The fourth tank remained at ambient temperature except for a period between January and March in which a small heater was added to keep the water above 5°C. All tanks were insulated with cotton padding and aluminium foil. At times, temperatures in the heated tanks fluctuated as a result of sand clogging the pipes and disrupting the established flow rates. In the spring, river water temperature rose until, in June, all tanks were above 22°C. Throughout the experiment, salinity and dissolved oxygen were measured weekly, while temperatures were taken daily. In addition, water samples were analyzed periodically for Kepone.

During the acclimation period, two samples of twenty fish (five per tank) were sacrificed on Day 0 and Day 31 and analyzed for Kepone. Thereafter, biweekly samples of ten spot per tank were taken for several weeks. Since, it appeared that the spot were depurating slowly, the time interval was increased later to four weeks. Kepone concentrations (whole body, micrograms/gram, μ g/g; and parts per million, ppm) were determined by electron capture gas chromatography. Mass spectrometry was utilized when concentrations were high. For the exact methodology of the chemical analysis see Appendix A.

RESULTS

Contaminated spot depurated considerable amounts of Kepone within a period of two hundred days (Fig. 1, dotted line). A mean Kepone concentration of 1.63 ppm (N = 20) was found for spot sacrificed on the day of collection (t = 0); whereas, a mean Kepone concentration of 0.45 ppm (N = 30) was found for spot sacrificed two hundred days later (t = 200). In a statistical analysis utilizing mean concentration values for the periods t = 0 and t = 31, spot eliminated approximately 53 percent of the Kepone residues in their tissues; however, 95 percent confidence intervals were broad during this period of acclimation (Fig. 1). Bahner et al. (1977) reported residue declines of 30-50 percent in spot after 24-28 days in Kepone-free water.

Further demonstration of Kepone depuration in the spot was provided by Pearson correlation coefficients (r) of -0.7252 (p = .001) for the variables Kepone concentration with total number of days in tank (t) and -0.6231 (p = .001) for the variables Kepone concentration with total number of days in tank squared (t²). A multiple regression analysis of mean Kepone concentrations by t of each tank produced the following regression equation: Kepone concentration + 1.48183-0.145133 t + $4.5612 \times 10^{-5} t^2$, $r^2 = 0.6948$, p = .001 and the regression curve (solid line) of figure 1.

The levels of Kepone concentrations in spot varied by period (t) and by tank (Appendix B). The appearance of a rise in concentrations after day 31, when heat was applied to tanks 1, 2 and 3, was attributed to no net loss of Kepone while spot were reacclimating to the rise in temperatures (Appendix C) and to possible random samples of highly contaminated fish. Thus, the actual acclimation period for the spot might be considered as the first sixty or seventy days. Once the tanks had achieved their respective temperatures (between days 59 and 73) mean Kepone concentrations in the spot samples began to change. Spot in the warmer tanks demonstrated lower mean Kepone concentrations. In fact, spot in Tank 1 (22°C) generally exhibited lower concentrations (Fig. 1). Unfortunately, Tank 1 was discontinued after a short period of excessively high temperatures which caused a high mortality among the spot.

No significant relationships were found between the level of Kepone residues (ppm) in spot and the length, weight, or sex of the fish. Values of micrograms of Kepone per gram fish (Appendix C) produced comparable results in statistical analyses. Furthermore, no substantial growth was observed in the spot during the study period. Thus, dilution of Kepone residue in the tissues due to growth was not a factor in the rate of depuration.

Although spot and Atlantic croaker are closely related species, Kepone concentrations in spot were generally higher than those Doyle et al. (1978) found in Atlantic croaker. Both species were collected from the James River at approximately the same time of year (October-November) although in different years (1976-1977). Initially, spot depurated Kepone at a faster rate than Atlantic croaker (Fig. 2). In fact, there was no significant decrease in Kepone levels of Atlantic croaker until after a period of fiftysix days. On the other hand, Atlantic croaker that were sacrificed after a period of one hundred and fourteen days, depurated at a slightly faster rate than spot from Tank 4 (ambient temperature).

CONCLUSIONS

Spot, like other fishes, depurated Kepone at a slower rate than some invertebrate species (see: Bahner et al. 1977). A mean loss in Kepone residues of 72 percent occurred between the initial spot sample (t = 0) and the eight spot sample (t = 200). A plot of the variables, mean Kepone concentration by period (t) (Figure 1) demonstrated the fact that a negative relationship existed between Kepone concentration in spot and the amount of time a spot was allowed to depurate in Kepone free water. Nonetheless, only 30 percent of the spot (N = 309) utilized in the test were below the established action level for human consumption (0.3 Therefore, it appears that it would be impractical to re-. (mag move spot from a contaminated area and to maintain them in a holding facility for the purpose of depuration and later commercial sale. Whether wild spot from the James River and the lower Chesapeake Bay can or cannot eliminate Kepone from their bodies while in the overwintering grounds of Virginia and North Carolina is still another question. To answer this question and other management questions, we would have to establish the Kepone levels in fish from offshore and returning populations which would be very difficult and costly.

Temperature was an important factor in the rate of Kepone depuration in spot. Spot held in warmer water exhibited lower mean Kepone concentrations; however, we were unable to observe the effect of the lower temperature extremities for any length of time in the cooler tanks because of the rise in temperature during the later spring months. In response to the warmer temperatures, Kepone concentrations in spot indicated that the rate of elimination of Kepone from body tissues is probably a function of the rate of an individual's metabolism. Thus, an increase in the matabolic rate as a result of an increase in body temperature may cause an acceleration in the depuration rate; however, it may not be apparent until after a period of acclimation.

It is regretable that the cost of Kepone body burden analysis is so high that sample sizes must remain small. In the future, we should take a closer look at the processes of uptake and accumulation of Kepone in eggs, larvae, juvenile and adult life stages. Also, we must have a better understanding of how Kepone concentrations in fish are related to uptake, accumulation and the lipid composition of fish.

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Figure 1. Depuration of Kepone from Spot, Leiostomus xanthurus.





Appendix A

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Chemical Analysis for the Pesticide Kepone

Appendix A

Chemical Analysis for the Pesticide Kepone

Whole fish were ground in a meat grinder into hamburger consistency. A mixture of anhydrous sodium sulfate and Quso^R G-30 (precipitated silica, Philadelphia Quartz Co.) was added for desiccation. The proportions of sample to the desiccants 30 g fish - 54 g Na, SO₄ - 6 g Quso. Then samples were were: frozen at -5°C for 24 hours to rupture the cells. After thawing the desiccated samples were ground with a blender to a powdery consistency and transferred to pre-extracted paper thimbles for Soxhlet extraction. Extraction was carried out using 1:1 ethyl ether-petroleum ether for 16 hrs. Extracts were then concentrated by evaporation and cleaned by activated fluorisil column chromatography (EPA, 1975). The Kepone containing elutriate was analyzed by electron capture gas chromatography utilizing packed columns with one or more of the following liquid phases: 4% SE-30 + 6% OV 210; 1.5% OV-17 + 1.95% QF-1 + 3% CV-1. On occasion, when concentrations were sufficiently high, Kepone presence was confirmed by mass spectrometry.

Appendix B

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Descriptive Statistics of Kepone Concentrations Broken Down by Tank and Sampling Period

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REGRESSICN ANALYSIS FOR SPOT DEPURATION STUDY					11/27/70	B PAGE	3		
FILE CSPCT	CREATICN	CATE = 11	Mean micro	Appendix B5. grams of Kepone per gram fi	ish (µg/g) values	for spot samples.	-		
						\$			
CRITERICA VARIA	BLE NER					5			
ERCKEN COWN	EV TEAY By Tankni	101 G INI	TAL NUMBER OF DAYS IN Cicates one of four (: TANK Gatrcl Temperatur					
						,		• •	
VARIABLE		CCLE	VALLE LABEL	SUM	MEAN	STD DEV	VARIANCE		N
FOR ENTIRE PGPU	LATION			7539.5694	24.3212	38-5215	1483-5021	C	310)
TEAN		0.	EDESANDI E	A681 A6A	74. 30 67	79-9743	6395-8877	4	26)
TANKAC		0.	TT COPHICE	686-1836	34.3092	79.9743	6395.8877	i	20)
TEAY		31.	IST SAMPLE	516.9828	28.8491	39-3826	1550-9912	٤.	20)
TANKNC		С.		576.9828	28.8491	39.3826	1550.9512	(201
TEAY		45.	SECONE SAMPLE	1178.6245	29.4656	37.4720	1404-1498		40)
TANKAC		1.		247.6199	24.7620	27.4161	751.7521		10)
TANKNC		ź.		178.1039	17-8164	19.4443	378.0817	(10)
TANKNC		3.		331-1999	33.12CC	56.6346	2563.8608	Ę.	10)
TANKAC		4.		421-7668	42.1701	44.9584	2021+2588	1	101
TCAY		55.	THIRD SAMPLE	1628_8555	40.7225	53.5146	2663.8099	ť	401
TANKNC		1.		464.3159	46.4320	67.9159	4612.5746	(10)
TANKNC		2.		431.5599	43 . 156C	53.9748	2913-2750	(10)
TANKNC		3.		237.5659	33.7576	46.7663	2378.1477	(10)
TANKNC		4.		355.4498	39.5450	49.0672	2407.5916	(10)
TEAY		73.	FEURTH SAMPLE	830.1347	20.7534	27.4251	752.1352	(40)
TANKAC		1.		128.2000	12.8200	£.C695	65.1168	(16)
TANKAC		2.		211.1755	21-1180	36.1400	908.4175	(10)
TANKNC		3.		185.8059	18.5810	15.2741	371.4914	(16)
TANKNC		4.		304.5449	30.4545	41.6350	1733-4711	C	10)
τιαν		122.	FIFTH SAMPLE	717.5548	17.9389	21.5632	464.9730	ť	40)
TANKNC		1.		71.1500	7.115C	6.7327	45.3292	(10)
TANKNC		ž.		167.65CC	16.7850	19.6316	385.3599	(10)
1ANK NC		3.		309.4649	36.9465	30.0739	904.4377	(16)
TANKNC		4.		165.6859	16.9090	18.5443	358.8867	t	10)
TCAY		152.	SIXTE SAPPLE	651.4358	16.2860	22.4033	501.5059	(4Ú)
TANKNE		1.		156.5966	15.0590	17-1141	292.8 920	(10)
TANKNE		ź.		242.6799	24.2680	36.3176	1468.2410	(10)
14N#NC		3.		65-19CC	6.5150	5.4601	29.8123	1	10)
TANKNC		4.		152.5755	15-2980	13.9801	195.4421	(10)
TEAN		178-	SEVENTE SAPPLE	736.5857	16.4147	24.4548	559.9951	(40)
TANENC	•	1.		85.3500	8,9350	11.0222	121.4899	(10)
TANKNE		2.		147.5300	14.7530	11.2755	127.1375	{	10)
TANKAC		3.		375-0198	37.5020	41.4401	1717 2045	,	1.01
TANFNC		4.		124.6900	12.4690	8.6738	75.2349	Ċ	101
1649		200	FINAL SANDIE	£33 1660	17 3336				
TANKAC		200.	I INPL SPPPLE	233-1079	1/.//20	31.6315	1000.5523	(30)
TANKAC		2.		136.4266 262 8600	13.0420	10.0274	256-8764	(10)
TANKAC		4_		1 CH _ Q40C	27.3000	71+1165 11 2007	2012.2200	4	10)
				100.7400	1000340	11.5667	121.1675	i	10)

NEUNEDZIUP FIMLIDID	IUN JELI ULPLKA	ITTON STOUT	Appendix B6.	11/21/10		•	
FILE ESFCT (CRE	EATICN CATE = 11	(27/78) Mean length in	millimeters (mm) of sp	ot samples.			
		CESCRIPTIC	CF SUEPO	PULATIONS			-
CRITERICN VARIABLE	LENGTE TOT	AL LENGTH IN MILLIMET	RS	• -	•		
ERCKEN COWN BY	TCAY TCI	AL NUMBER OF DAYS IN	IANK	•			
EY		ICATES ONE OF FOUR CO	VTRCL TEMPERATUR				
VARIABLE	COCE	VALLE LABEL	SUP	MEAN	STD DEV	VARIANCE	
			40105 0000	126 2710	25 23/8	641 6516	
FLK ENTINE PUPULATIO			40103-0000	123-3710	2303360		
TEAY	٤.	FRESAMPLE	1945.0000	97.2500	11.1255	123-7763	
1 ANKAC	C -		1945.0000	97.2500	11.1255	123.7763	
TCAY	31.	IST SAMFLE	2599.0000	129.9500	33.3868	1114-6816	
TANENC	C.		2595.0000	129-9500	33.3868	1114-6816	
TCAY	45.	SECCNE SAFFLE	4505-0000	122.7250	16.4986	272.2045	
TANKNC	1.		1154-0000	115.4000	10.5325	110.9333	
TANKNC	٤.		1143.0000	114.3000	12.7545	162-6778	
TANKNC	3.		1279.0000	127.5000	18-5379	343-6556	
1 <i>4</i> N KN C	4.		1293.0000	129.3000	19.8329	393-3444	
TCAY	59.	THIRE SAMPLE	5221.COCO	136-5256	25.1630	633.1788	
TANKNC	1.		1273.000	127.3000	20.1332	405.3444	
TANKNC	2.		1279.0000	127.9000	26.7268	714.3222	
JAVKAC	3.		1272.0000	127.2000	21.7552	473.2889	
TANKNC	4.		1397.0000	139.7000	32.0275	1025.7869	
TEAY	73.	FCLRTH SAMPLE	5441.COGC	136.0250	26.2668	686.7942	
TANKNC	1-		1314-CCCC	131-4000	21.5555	482.0444	
TANKAC	ź.		1398.0000	139.8000	26.8816	122.0222	
TANKNO	3.		1358-0000	135.8000	25.415/	042.5220	
TANKNE	4.		1371-000	137.1000	32.5324	1084. 3444	
TEAY	122.	FIFTH SAMPLE	5314.0000	132.8500	25.0983	625.5256	
TANKAC	1.	•	1265.0000	126.9000	21.0895	444.1661	
TANKNC	2.		1361.0000	136.1000	30-2120	912.7667	
TANKAC	3.		1414.0000	141.4000	20.1302	114.9333	
148880	4.		1276.6666	127.000	22.0000	400.0003	
TDAY	152.	SIXTE SAMPLE	5365.0000	134.1250	27.2333	741.6506	
TANKNC	1.		1295.000	129-5000	28.2013	66234600	
TANKNC	2.		1448.000	144-8606	36.1603 26.6301	968.4660	
TANKAC	3.		1255.0000	125.9000	26.4801	417.4333	
TANKNC	4-		1363.0000	130-3000	27.2204	037.3444	
TDFY	179-	SEVENTH SAPPLE	5340.0000	133.5000	21.5585	466.5128	
TANKNC	1.	-	1266 6060	135-6000	21.6920	467.6111	
TANKNC	2.		1:52.000	13340000	21.0220	40763111	
TANKNE	3.		1477.000	147.7000	24.9491	627-4554	
TANFNC	. 4-		1298.0000	129-8000	12.1454	147.5111	
TEAY	200-	FINAL SAMPLE	3571.0000	132.3667	22.2734	456-1023	
TANENC	2.		1332-0000	133.2000	23.4132	548.1778	
TANFNE	3.		1301.0000	130.1000	21.4565	462,1000	

TCTAL CASES = 310

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REGRESSION ANALYSIS FOF SPOT DEPURATION STUDY					11/27/78 FAGE 11				
FILE DSPCT (CREA	TICN CATE = 11	127/76) <u>Mean weight i</u>	n grams (g) of spot samp	les.			·		
CRITERICA VAKIABLE BRCKEN COMN BY	NEIGHI ICI TCAY ICI	C E S C R I P T I G Ial beight in graps Ial nupber of cays in	N GF SUBPO TANK	PULATION	· S	· 			
8Y	TANKNC IN	ICATES CNE GF FCUR CG	NTROL TEPPERATUR						
VAFIABLE	CCCE	VALUE LAREL	SUM	MEAN	STD CEV	VARIANCE		N	
FCF ENTIPE FUPULATION			16501.8995	25.1674	25.8903	670.3069	ť	310	
						• .			
Τ Γ ΑΥ ΤΔΝΚΝΟ	C. 0-	PRESAMPLE	320.60C0 320.60C0	16-030C 16-0300	15.2458 15.2498	370.5548 370.5548	t t	201 201	
		÷		•••••					
TDAY TANKNC	31. C.	IST SAMFLE	678.0C0C 678.CCCC	33.9000 33.9000	32.7316 32.7316	1071.3575 1071.3579	(t	20) 20)	
TEAN	45	SECCAR SAMPLE	1126-3000	26.1575	13-9141	193-6020	· ·	401	
TANKIC	1.	3000110 201122	235.0000	23.9000	7.8521	61.6556	i	10	
TANENC	2.		215.70CG	21.9700	9.0620	82.1201	L	10)	
TANKAC	.ن		326.7000	32.0700	14.5194	210.8135	(101	
TANKNC	4.		346.9000	34.6500	16-9011	357.2495	(10	
TCAY	55.	THIRC SAMPLE	1415.0000	35.3750	27.0135	725.1276	(4C)	
TANKAC	1.		285.0000	26.5000	15.4722	235.3685	t	10	
TANKNC	ź.		331.0000	33.100C	23.6287	530.3222	(101	
TANKNE	3.		333.0000	33.3000	23.1855	537.5667	(101	
TANINC	4.		466.000	46.6000	46.6426	1651.8222	C	101	
TEAY	73.	FCURTH SAMPLE	1556.0000	38.9000	25.8601	666.7462	(40)	
TANKNC	1.		328.0000	32.8COC	16.6986	£70.8444	(101	
TANKAC	2.		416.0000	41.6GOG	22.2221	453.8222	(101	
TANKNC	3-		360.5000	36.0500	25.7601	663.5206	ç	101	
TANKNC	4.		431.5000	43.1500	37.3668	1351-3301	ſ	101	
1041	122.	FIFTH SAMPLE	1443.0000	36.0750	26.0713	675.7122	£	471	
TANKNC	1.		262.0000	28.2000	21.2697	452.4000	(101	
TANKNC	2.		428.0000	42.8000	37.5227	1407.4556	(101	
TANKNC	3.		430.0000	43.0000	23.4852	551.5556	(101	
ΤΑΝΚΝΟ	4.		363.0000	36.3000	16.0003	324.0111	•	101	
TCAY	152.	SIXTH SAMPLE	1597.0000	35.9250	29.8641	891.8660	ſ	4 C (
TANKAC	1.		375.0000	37.5000	31.6763	1003.3885	(10)	
TANKNC	ž.		496.000	45.0000	25.0593	844.4444	(101	
TANKNE	3.		280.000	28.0000	15.2607	232 8385	(131	
TANKNC	4.		452.0000	45.2006	35.6515	1457.6667	C	101	
TCAY	176.	SEVENTH SANPLE	1535.0000	38-3750	23.5260	553.4712	ť	46	
TANKIC	1.		262.0000	26.2000	14.74ė3	217.5111	ſ	101	
TANKNC	ž.		403-0000	46.3000	30.1835	953.7889	(1 C)	
TANKAC	3.		517.GCCC	51.7000	26.8537	721.1222	· (:03	
TANKNC	4.		353.0000	35.3000	11.3730	129.3444	C	ìC	
TC#Y	200.	FINAL SAMPLE	1231_0000	41.0333	27.8524	775.7575	t	30	
TANKNC	2.		437.0000	43.7000	32.4004	1645 7889	i	10	
TANKNC	3.		351.000	35.1000	18.5270	335.87 10	i	101	
TANKNE	4•		443.GOCC	44.3000	32.4826	1055.1222	(161	

1CTAL CASES = 310

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Appendix C

Water Quality Analysis During the Experimental Period







Appendix C2. Dissolved oxygen concentrations for all four tanks.



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Appendix C3. Average salinity for all four tanks.