

W&M ScholarWorks

Reports

1965

Key to Some of the Marine Diatom Genera in Virginia Waters

Richard A. Mulford Virginia Institute of Marine Science

Morris H. Roberts Jr. Virginia Institute of Marine Science

Follow this and additional works at: https://scholarworks.wm.edu/reports

Part of the Marine Biology Commons, and the Zoology Commons

Recommended Citation

Mulford, R. A., & Roberts, M. H. (1965) Key to Some of the Marine Diatom Genera in Virginia Waters. Educational series; no. 12.. Virginia Institute of Marine Science, College of William and Mary. https://doi.org/10.21220/V5PK6Q

This Report is brought to you for free and open access by W&M ScholarWorks. It has been accepted for inclusion in Reports by an authorized administrator of W&M ScholarWorks. For more information, please contact scholarworks@wm.edu.

VIMS OH 91.15 E38 Jub, 12 c, 2

Key to Some of the Marine Diatom Genera

in Virginia Waters

by

R. A. Mulford and M. H. Roberts, Jr.

VIRGINIA INSTITUTE OF MARINE SCIENCE Gloucester Point, Virginia 1965

Educational Series No. 12

LIBRARY of the VIRGIMIA INSTITUTE of MARINE SCIENCE

CONTENTS	Page
Introduction	1
Classification of Bacillariophyceae	1
Key to Common Genera of Lower Chesapeake Bay	4
Description of Genera	7
Glossary	16
Bibliography	18

INTRODUCTION

The information compiled herein is intended to assist students and scientific staff in making preliminary generic identifications of diatoms they encounter when definite speciation is not necessary. Other more extensive keys (e.g. Cupp, 1950 and Hendey, 1964) are available and should be consulted when verification is needed or when identification to species is desired. Generic descriptions and drawings have been included to supplement the key.

Acknowledgements

The generic descriptions have been adapted from Boyer (1916),

Cupp (1943) and Van Heurck (1880-1885). Some of the figures have been copied, with modification, from Griffith (1961).

CLASSIFICATION OF BACILLARIOPHYCEAE

The following classification, revised by R. A. Mulford from Hendey (1937), includes only the more common genera.

Class Bacillariophyceae Order Bacillariales Suborder Discineae Family Coscinodiscaceae Subfamily Melosiroideae

Melosira Agardh (8)

Subfamily Skeletonemoideae

Skeletonema Greville (11) Stephanopyxis Ehrenberg (11)

Subfamily Thalassiosiroideae

<u>Coscinosira</u> Gran (7) <u>Lauderia</u> Cleve (9) <u>Schroderella</u> Pavillard (10) <u>Thalassiosira</u> Cleve (6, 7)

Subfamily Coscinodiscoideae

<u>Actinocyclus</u> Ehrenberg (13) <u>Coscinodiscus</u> Ehrenberg (14) <u>Planktoniella</u> Schutt (13)

Family Actinodiscaceae Subfamily Actinoptychoideae Actinoptychus Ehrenberg (14) Suborder Biddulphineae Family Biddulphiaceae Subfamily Biddulphioideae Bellerochea Van Heurck (28) Biddulphia Gray (23) Cerataulina H. Peragallo ex Schutt (9) Subfamily Triceratioideae Ditylum Bailey (26) Lithodesmium Ehrenberg (28) Triceratium Ehrenberg (27) Subfamily Hemiauloideae Hemiaulus Ehrenberg (23) Subfamily Eucampioideae Climacodium Grunow (25) Eucampia Ehrenberg (24) Streptotheca Shrubsole (25) Family Chaetoceraceae Subfamily Chaetoceroideae Chaetoceros Ehrenberg (21) Suborder Soleniineae Family Bacteriastraceae Subfamily Bacteriastroideae Bacteriastrum Shadbolt (17) Family Rhizosoleniaceae Subfamily Rhizosolenioideae Guinardia H. Peragallo (19) Rhizosolenia (Ehrenberg) ex Brightwell (15) Family Leptocylindraceae Subfamily Leptocylindroideae Dactyliosolen Castracane (18) Leptocylindrus Cleve (19) Family Corethronaceae Subfamily Corethronoideae Corethron Castracane (17) - 2 -

Suborder Araphidineae Family Fragilariaceae Subfamily Fragilarioideae

Asterionella Hassal ex Wm. Smith (42) Fragilaria Lyngbye (41) Thalassionema Grunow ex Hustedt (41) Thalassiothrix Cleve and Grunow (42)

Subfamily Tabellarioideae

<u>Grammatophora</u> Ehrenberg (39) <u>Licmophora</u> Agardh (38) <u>Rhabdonema</u> Kutzing (39)

Suborder Monoraphidineae Family Achnanthaceae Subfamily Achnanthoideae

Achnanthes Bory (36)

Subfamily Cocconeioideae

Cocconeis Ehrenberg (36)

Suborder Biraphidineae Family Naviculaceae Subfamily Naviculoideae

> <u>Gyrosigma</u> Hassall (33) <u>Navicula</u> Bory (31) <u>Pleurosigma</u> Wm. Smith (33)

Subfamily Amphiproroideae

<u>Amphiprora</u> Ehrenberg (34) <u>Tropidoneis</u> Cleve (34)

Family Cymbellaceae Subfamily Cymbelloideae

Amphora Ehrenberg (32)

Family Bacillariaceae Subfamily Nitzschioideae

Nitzschia Hassall (30)

KEY TO COMMON GENERA OF LOWER CHESAPEAKE BAY

	1	 Raphe or pseudoraphe present 29 Raphe or pseudoraphe absent 2
· (1) 2	. Cells circular or oval in valve view, narrowly rectangular
	2	. Cells in all views angular or gonoid, not naviculoid, girdle view not narrowly rectangular, or with projections at angles 20
(3	2) 3 3	 Pervalvar axis about twice diameter or less 4 Pervalvar axis 4-20 times diameter 15
(3	3) 4 4	. Colonial 5 . Solitary 12
(4	 5 5 5 	 Valves connected by mucous thread(s) 6 Valves connected directly by faces 8 Valves connected by marginal spines 10
(5) 6.	Cells united as chains 7 Cells united as aggregates of no definite shape <u>Thalassiosira</u>
(6) 7. 7.	Mucous thread singleMucous thread fasciculate <u>Coscinosira</u>
(5) 8. 8.	Valves punctate, radially furrowed <u>Melosira</u> Valves hyaline 9
(8)) 9. 9.	Marginal process single, reduced to an apicule <u>Lauderia</u> Marginal processes two, each bearing a small mucro <u>Cerataulina</u>
(5)	10. 10.	Spines minute, one spine centrally located <u>Schroderella</u> Spines greater than one-half pervalvar axis 11
(10)	11. 11.	Valve surface coarsely areolateStephanopyxis Valve surface hyalineSkeletonema
(4)	12. 12.	Valves with extensions or ocelli 13 Valves without extensions or ocelli 14
(12)	13. 13.	Valves with loculate extensionsPlanktoniella Valves with marginal ocelli <u>Actinocyclus</u>
(12)	14. 14.	Valves with coarse areolation, not undulatingCoscinodiscus Valves with radial markings forming undulating seg- ments <u>Actinoptychus</u>
(3)	15. 15.	Valve face conical Rhizosolenia Valve face flat or weakly convex 16
(15)	16. 16.	Valves with marginal corona of setae 17 Valves without marginal corona of setae 18
(16)	17. 17.	Setae bifurcateBacteriastrum Setae not bifurcate Corethron

(16)	18. 18.	Intercalary bands spirally arranged <u>Dactyliosolen</u> Intercalary bands not spirally arranged
(18)	19. 19.	Valves flat, with marginal spur <u>Guinardia</u> Valves convex, lacking marginal spur <u>Leptocylindrus</u>
(2)	20. 20.	Cells with two angles in valve view 21 Cells with more than two angles in valve view 26
(20)	21. 21.	Each angle of cell bearing one long seta <u>Chaetoceros</u> Each angle of cell bearing more than one process 22
(21)	22. 22.	Processes cornutate 23 Processes simple, boss-like projections 24
(22)	23. 23.	Processes terminated with spines <u>Hemiaulus</u> Processes not terminated with spines <u>Biddulphia</u>
(22)	24. 24.	Cells in curved, ribbon-like chains <u>Eucampia</u> Cells in chains with a slight spiral twist25
(24)	25. 25.	Intercellular spaces large and elliptical <u>Climacodium</u> Intercellular spaces lacking, cells weakly siliceous Streptotheca
(20)	26. 26.	Valves with long central spine <u>Ditylum</u> Valves lacking central spine 27
(26)	27. 27.	Valve surfaces hexagonally areolate, areolation transverse, angular processes cornutate <u>Triceratium</u> Valves not as above 28
(27)	28. 28.	Valve surfaces contiguousBellerochea Valve surfaces not contiguous <u>Lithodesmium</u>
(1)) 29. 29.	Valves both with true raphe 30 One or both valves with pseudoraphe 35
(29) 30. 30.	Raphe located medially or eccentrically but not on keel 31 Raphe located eccentrically on keel <u>Nitzschia</u>
(30) 31.	Valves linear to oval-lanceolate, punctate, striation reaching to margin of valve
(31) 32. 32. 32.	Cells sigmoid 33 Cells spirally twisted 34 Cells sublunate <u>Amphora</u>
(32) 33.	Striations transverse and oblique <u>Pleurosigma</u> Striations transverse and longitudinal <u>Gyrosigma</u>
(32	2) 34. 34	 Raphe medial and straight <u>Tropidoneis</u> Raphe not medial and sigmoid; with alate projections <u>Amphiprora</u>

(29)	35.	Cells with raphe on one valve and pseudoraphe
	35.	on other 36 Cells lacking true raphe on either valve 37
(35)	36. 36.	Valves symmetrical about apical and transapical axes- <u>Cocconeis</u> Valves asymmetrical about apical axis <u>Achnanthes</u>
(35)	37. 37.	Internal septa in valvar plane, often perforated 38 No internal septa 40
(37)	38. 38.	Valves broadly clavate, girdle cuneate <u>Licmophora</u> Valves not clavate 39
(38)	39. 39.	Cells in zig-zag chains, septa 4-12, straight or nearly so <u>Rhabdonema</u> Cells solitary or colonial, usually 2 septa at either end, wavy <u>Grammatophora</u>
(37)	40. 40.	Bilaterally symmetrical about transapical axis 41 Asymmetrical about transapical axis 42
(40)	41. 41.	Cells in ribbon-like chains, valves usually linear <u>Fragilaria</u> Cells solitary with small terminal spines <u>Thalassionema</u>
(40)	42. 42.	Cells in stellate clusters, valve clavate Asterionella Cells solitary, spirally twisted, often in dense masses, one end of cell only slightly large than other <u>Thalassiothrix</u>

DESCRIPTION OF GENERA

Achnanthes Bory - Plate II, Fig. 13

Frustule stipitate, solitary or in short chains, flexed. Valves elliptical or lanceolate, naviculoid, dissimilar, the lower with a raphe and median and terminal nodules, the upper with a pseudoraphe.

Actinocyclus Ehrenberg - Plate I, Fig. 3

Single cells with slightly convex valves having flat centers; valves circular to elliptical. Central space circular to irregular, sometimes inconspicuous. Markings in radiating rows of unequal length, leaving radiating hyaline spaces; rows sometimes fasciculate. Margin distinct or indistinct, with clear round spot (ocellus); small marginal or submarginal spines sometimes present.

Actinoptychus Ehrenberg - Plate I, Fig. 2

Frustule cylindrical, length less than diameter, in girdle view undulated. Valve divided into six or more sectors alternately raised and depressed, areolate and punctate alternately. Marginal processes three or more. Umbilicus circular or angular, hyaline.

Amphiprora Ehrenberg - Plate II, Fig. 17

Frustules twisted in the longitudinal axis, constricted in middle. Valve lanceolate, acute. Raphe confined within a sigmoid keel, central and terminal nodules indistinct. Striae transverse, punctate, with coarser striae at junction of keel and lower part of valve.

Amphora Ehrenberg - Plate II, Fig. 16

Valves asymmetrical along the longitudinal axis, with the plane passing through the dorsal and ventral sides of one valve at an angle with that of the other.

- 7 -

<u>Asterionella</u> Hassall - Plate II, Fig. 15

Cells clavate, united by larger ends into star-shaped or spirally curved colonies, sometimes almost straight bands. Intercalary bands and septa absent. In girdle view linear, usually widened at the ends, sometimes pervalvarly constricted. Valve with delicate transapical striae and narrow median pseudoraphe. Chromatophores a small plate or small granules.

Bacteriastrum Shadbolt - Plate I, Fig. 10

Cells cylindrical, bound into loose chains by fusion of bristles regularly arranged around margins of cells, Terminal setae different from others, often curved; not fused and therefore not bifurcate. Intercalary bands absent, apertures between cells of varying widths. Cell wall delicate and hyaline, Chromatophores numerous, small, roundish, more or less lobed. Resting cells near middle of cell with small spines on one valve. All species marine pelagic.

Bellerochea Van Heurck - Plate I, Fig. 9

Cells in narrow chains with corners and centers of neighboring valves touching; creating narrow, elliptical apertures; weakly siliceous. Valve 3 or 4 sided, with undulating margins; raised marginal parts connecting with valve of next cell. Cell usually broader than long.

Biddulphia Gray - Plate I, Fig. 18

Frustule prismatic or subcylindrical. Cells in filamentous or zig-zag chains or free. Valve triangular, polygonal, elliptical or subcircular, convex, more or less elevated at the angles into processes or horns.

Cerataulina Peragallo - Plate I, Fig. 13

Cells cylindrical, usually in chains. Valves slightly arched, with two blunt projections or processes near margin, attached to adjacent cell by means

- 8 -

of a fine, small, curved, hair-like process which fits into the valve of the adjacent cell. Intercalary bands numerous, annular. Chromatophores numerous, small. Cell wall soft and weakly siliceous, sculpturing very delicate.

Chaetoceros Ehrenberg - Plate I, Fig. 19

Cells oval to almost completely circular in valve view; in broad girdle view quadrangular with straight sides and concave, flat, or slightly convex ends. Valve with more or less flat surface and mantle, bound together without a suture. A long thick or thin seta, bristle, or awn, at each end of valve on corners. Opposite setae of neighboring cells touching one another near their origin, usually directly or sometimes by a bridge, and fusing firmly at a point near their bases to hold the cells in chains. Usually with large or small apertures between cells. Basal portion of setae parallel to the pervalvar axis or directed diagonally outward with the outer portion frequently perpendicular to the axis of the chain. Most species with special end cells bearing terminal setae usually shorter, thicker and more nearly parallel to the chain axis than are the other setae. A few species with cells solitary. Chromatophores varying greatly in size, number, form, and position in different species but constant for a given species and consequently indispensible for species determination.

Climacodium Grunow - Plate II, Fig. 5

Cells straight but usually forming somewhat twisted chains. Apertures oval or squarish-oblong as a result of the hammer-like ends of cells. Cell wall weakly siliceous, without visible sculpturing. Intercalary bands absent. Pervalvar axis of cell usually short. Chromatophores numerous, small.

Cocconeis Ehrenberg - Plate II, Fig. 11

Valves elliptical, dissimilar, upper valve with a pseudoraphe, lower valve with a true raphe and nodules; usually with an annulus. Epiphytic.

- 9 -

Corethron Castracane - Plate I, Fig. 6

Cells solitary; cylindrical, with rounded valves having a marginal crown of long thin spines or setae directed outward at an angle. Intercalary bands numerous, scale-like, often distinct. Cell wall delicate, weakly siliceous. Chromatophores numerous, small.

Coscinodiscus Ehrenberg - Plate I, Fig. 1

Frustules solitary, cylindrical, compressed; valve circular or elliptical; surface flat or sometimes convex near border; markings more or less angular, radiating, sometimes fasciculate. Central space, if present, hyaline.

Coscinosira Gran - Plate II, Fig. 9

Cells drum-shaped to short cylindrical with flat or convex valves. United in loose chains by several long gelatinous threads. Intercalary bands distinct. Valves with large areolae. Margins with spinulae, rarely with spines. Chromatophores numerous, small, round plates.

Dactyliosolen Castracane - Plate I, Fig. 21

Cells cylindrical, solitary or in long, stiff, closed chains, attached by the flat valve surfaces. Valves circular, without noticeable spines or processes. Intercalary bands numerous, mantle surface with fine or coarse areolae. Girdle band frequently unilaterally displaced and considerably closer to younger valve.

Ditylum Bailey - Plate I, Fig. 11

Cells elongate, prismatic to box-shaped; solitary except immediately after division. Valves with 3 to 4 corners, seldom bipolar, with a strong central siliceous hollow spine and a marginal ridge strengthened by ribs. Intercalary bands more or less numerous. Valve surface more or less wavy, with usually poorly developed humps on the corners. The 3-cornered elevated region often with a circle of short pervalvar-directed spines. Outer valve margin wavy, giving the appearance of lines running from valve to valve. Cell wall weakly siliceous, valve

- 10 -

membrane delicately areolated. Chromatophores numerous, small.

Eucampia Ehrenberg - Plate II, Fig. 1

Valves elliptical in surface view, with two blunt processes, without spines or setae. Numerous intercalary bands difficult to see in water mounts. Chains spirally curved. Large apertures between cells.

Fragilaria Lyngbye - Plate II, Fig 13

Frustules rectangular, valve lanceolate, oblong or elliptical in general outline, with convex or sinuate margins; lacking costae; pseudoraphe narrow or indistinct; striae transverse.

Grammatophora Ehrenberg - Plate II, Fig. 7

Frustules quadrangular, adnate, in zigzag chains, united by an isthmus or free. Valve linear or oblong, sometimes with sinuate sides, and with a pseudoraphe and transverse punctate lines.

Guinardia H. Peragello - Plate I, Fig. 8

Cells cylindrical, longer than broad, with a straight or slightly curved pervalvar axis, solitary or in straight to twisted, close-set chains. Intercalary bands numerous, collar-like or with wedge-shaped ends. Valves circular, surface flat, with an asymmetrical lateral rudimentary tooth at valve margin. Chromatophores numerous, roundish, more or less lobed.

Gyrosigma Hassall - Plate II, Fig. 6a

1

Valve lanceolate, sigmoid, axial area very narrow, central area small; transverse and longitudinal striae punctate.

Hemiaulus Ehrenberg - Plate I, Fig. 22

Cells solitary or in chains. Valves elliptical, with two narrow, pointed, rather long processes at ends of apical axis and parallel to pervalvar axis. One or more hyaline claws on the end of the processes. Intercalary bands indistinct or absent, without septa. Valve strongly or weakly siliceous, finely or coarsely areolate or punctate. Chromatophores numerous.

- 11 -

Lauderia Cleve - Plate II, Fig. 8

Cells cylindrical. Valves rounded. Cells united in straight chains by very fine gelatinous threads; cells touching or separated. An unpaired, oblique outwardly-directed apicule on each valve; numerous very small spinulae or slime canals at margin and over most of surface. Center of valve slightly concave. Intercallary bands numerous; chromatophores numerous small plates. Valve surface radially striated, mantle surface of intercalary bands delicately areolate.

Leptocylindrus Cleve - Plate I, Fig. 14

Cells long, cylindrical, united into chains by whole valve surface. Valves flat, without spines or processes. Intercalary bands present but seen with difficulty. Cells thin-walled, hyaline, without visible sculpturing. Chromatophores one or many roundish plates or granules.

Lichmophora Agardh - Plate II, Fig. 12

Frustules wedge-shaped, joined together into fan-shaped stipitate fascicles. Valve cuneate, rounded at both ends, septate. An epiphyte, attached to substrate by gelatinous threads, but often found in plankton.

Lithodesmium Ehrenberg - Plate I, Fig. 7

Cells united in usually long, straight chains with concealed apertures. Valves triangular, with marginal pervalvar-directed membrane by which adjacent cells are joined. Long, thin, hollow spine in center of valve. Intercalary bands present, collar-like. Chromatophores numerous, small.

Melosira Agardh - Plate I, Fig. 20

Frustules globose, ellipsoidal or cylindrical, concatenate, closely joined together. Valve either simply punctate or areolate. Cell wall constricted, forming a furrow between edge of valve and girdle.

Navicula Bory - Plate II, Fig. 13

Valve linear to elliptical; ends acute, rounded, rostrate, capitate or truncate; axial area usually distinct, rounded, or rarely extended into a

- 12 -

transverse fascia; striae transverse or radiate, punctate.

Nitzschia Hassall - Plate II, Fig. 14

Frustules usually free, sometimes enclosed in tubes or united into a filament. Valves keeled, keels of two valves diametrically opposed; keel punctae short or prolonged.

Planktoniella Schutt - Plate I, Fig. 4

Cells solitary, disk-shaped, with a hyaline alate expansion on circumference consisting of extra-cellular chambers strengthened by radial rays. Alate expansion weakly siliceous. Valves areolate; chromatophores numerous small plates lying along valve surface.

Pleurosigma Wm. Smith - Plate II, Fig. 6b

Valve lanceolate, sigmoid; axial area very narrow, central area small; transverse and oblique striae punctate.

Rhabdonema Kützing - Plate I, Fig. 16

Frustules quadrangular, concatenate, composed of numerous septate partitions with transverse costae or rows of punctae. Valves elliptical, with a pseudoraphe and transverse apparent costae and punctate lines, partitions with one or several foramina.

Rhizosolenia (Ehrenberg) Brightwell - Plate I, Fig. 17

Cells cylindrical with greatly elongated pervalvar axis; solitary or in chains. Cells usually straight or more rarely curved, forming spirally twisted chains. Crosssection elliptical or circular. Intercalary bands usually very numerous, obscure in some species. Valves may be almost flat or symmetrically conical but are usually eccentric, sharp, or hood-shaped; valves may bear processes of various kinds or may be drawn out to form a process. Chromatophores usually small, numerous, distributed on entire cell wall, but especially massed in girdle zone about the nucleus.

Schroderella Pavillard - Plate II, Fig. 3

Cells cylindrical with slightly convex valves, somewhat concave in center; form

- 13 -

straight chains but sometimes solitary. Distinct spine in central depression of valve is joined by spine belonging to adjacent cell. Margin surrounded by a row of small spinulae from each of which arise two gelatinous threads which diverge and join the corresponding thread of the next cell, thus forming a characteristic zigzag. Threads sometimes straight or appearing so. Intercalary bands numerous, forming incomplete hoops, with minute punctae. Chromatophores small, strongly slit plates, usually with four straight ends.

Skeletonema Greville - Plate II, Fig. 4

Cells circular, lens-shaped, oblong, or cylindrical. Valves circular, somewhat arched, lacking distinct structures, with a row of fine spines at edge of valve parallel to longitudinal, pervalvar axis. Spines interlocking midway between adjacent cells and uniting into chains. Chromatophores one or two per cell.

Stephanopyxis Ehrenberg - Plate I, Fig. 5

Cells oblong, oval, or nearly circular, with hexagonal areolations. Usually in short chains. Margins rounded, with a crown of stout spines or hollow needles, nearly parallel with pervalvar axis. Intercalary bands absent. Chromatophores numerous, small rounded.

Streptotheca Shrubsole - Plate II, Fig 2.

Cells extremely flat. outline square, sometimes triangular; united by valve faces, leaving very little, if any aperture. Cell wall weakly siliceous. Chains strongly twisted. Valves narrowly elliptical, with a rudimentary central knob. Chromatophores numerous small granules.

Thalassionema Grunow - Plate II, Fig. 10

Cells forming zigzag bands or star-shaped colonies, adjacent cells united to each other by small gelatinous cushions on one cell end. In girdle view, linear. Intercalary bands and septae absent. Valve linear to narrow-lanceolate. Valve with numerous tiny marginal spines placed at regular intervals. Cell wall otherwise structureless.

Thalassiosira Cleve - Plate I, Fig. 12

Cells usually drum- or disk-shaped, united in flexible chains by a cytoplasmic or gelatinous thread, living in formless gelatinous masses, or sometimes solitary. Valve with one or more intercalary bands and with areolate or delicate radial rows of punctae. Marginal spinulae or little spines present.

Thalassiothrix Cleve and Grunow - Plate II, Fig. 10

Cells living singly or forming star-shaped colonies, zigzag bands, or bunches, united to one another by a gelatinous cushion on the end of the cell. In girdle view linear or slightly lanceolate, ends unlike. Valve borders often with small spines. Valve surface with short marginal striae or structureless.

Triceratium Ehrenberg - Plate I, Fig. 15

Cells typically box-shaped, pervalvar axis shorter or longer due to connecting bands. Valvar plane with 2-4 corners, in some variation bipolar, rarely circular. Valve edges with humps, lacking special terminal claws. Cell wall strongly siliceous. Delicate punctae next to very large areolae, pore canals passing through costae next to fine pores in shallow cavities. Structure of hump generally differs from that of remainder of valve. Chromatophores numerous, small.

Tropidoneis Cleve - Plate II, Fig. 18

Frustule oblong, constricted medially; keel not sigmoid. Axial area not evident. Striae very fine. punctate, in longitudinal lines.

- 15 -

GLOSSARY

<u>Alate</u> - wing-like

Annulus - a ring-like part

Apical axis - longitudinal axis of the valve

Apicule - single large marginal nodule penetrated by a pore channel

Areolate - having polygonal or rounded areas or cavities

Bifurcate - forked, having two branches

Bilaterally symmetrical - capable of being halved in one, and only one, plane in such a way that the two halves are mirror images of each other

Boss-like - short, squat

Capitate - enlarged at the tip

Chromatophore - a cytoplasmic inclusion containing chlorophyll and other pigments; may appear yellow, golden brown, or greenish brown

Clavate - club-shaped

Concatenate - linked together or united as in chains

Concentric - having a common center, as circles and spheres

Contiguous - adjoining or nearly so

Cornutate - horned

Corona - crown-like structure

Costa - conspicuous rib on the valve

Cuneate - wedge-shaped

Fascia - a band or flat strip

Foramen (pl. foramina) - a small opening

Frustule - individual diatom cell

Girdle - connection point or overlap of two valves

Gonoid - round, not disced

Hyaline - transparent, glassy, or translucent

- 16 -

<u>Intercalary</u> <u>band</u> - a ring-like or scale-like intermediate band between valve and connecting band

<u>Keel</u> - plate-like projection of valve face, usually lateral

Laceolate - narrow and tapering, like the head of a lance

Loculus - small cavity, cell or chamber

Mucro - an abrupt point, tip or process

<u>Nodule</u> - small internal thickening of a rounded or conical shape, generally containing a cavity

Ocellus - a small rounded thickening or "little eye"

<u>Peripheral</u> - outer or external

Pervalvar axis - cell axis through the center point of adjoining valves

<u>Pseudoraphe</u> - a narrow hyaline axial area without a central nodule or striae

Puncta - a small cavity situated within ridges of walls

Raphe - longitudinal slit in valve, varying in form and structure

Rostrate - having a beak-like process

<u>Septa</u> - ingrowths of the intercalary bands, more or less perforated; and nearly parallel to valve face

Seta - a fine bristle or hair

Sigmoid - S-shaped

ł

Siliceous - composed of silica

Stellate - star-shaped, coming out like rays from the center

Stipitate - growing on a stalk

Striae - prolonged furrows closed to the outside and open on the inside

Sublunate - curved, tending toward, but not, cresent-shaped

Transapical axis - transverse axis of a valve

Umbilicus - a navel-like depression

Valve - one of the two largest segments of the frustule

- 17 -

1. <u>Coscinodiscus</u> Ehrenberg

- 2. Actinoptychus Ehrenberg
- 3. Actinocyclus Ehrenberg
- 4. <u>Planktoniella</u> Schutt
- 5. Stephanopyxis Ehrenberg
- 6. <u>Corethron</u> Castracane
- 7. Lithodesmium Ehrenberg
- 8. Guinardia H. Peratello
- 9. <u>Bellerochea</u> Van Heurck
- 10. Bacteriastrum Shadbolt
- 11. <u>Ditylum</u> Bailey
- 12. <u>Thalasiosira</u> Cleve
- 13. Cerataulina Peragallo
- 14. Leptocylindrus Cleve
- 15. Triceratium Ehrenberg
- 16. <u>Rhabdonema</u> Kützing
- 17. Rhizosolenia (Ehrenberg) Brightwell
- 18. <u>Biddulphia</u> Gray
- 19. Chaetoceros Ehrenberg
- 20. <u>Melosira</u> Agardh
- 21. <u>Dactyliosolen</u> Castracane
- 22. <u>Hemiaulus</u> Ehrenberg



ł















M





m

uuu



















Plate II

1. Eucampia Ehrenberg

ŗ

- 2. <u>Streptotheca</u> Schrubsole
- 3. <u>Schroderella</u> Pavillard
- 4. <u>Skeletonema</u> Greville
- 5. <u>Climacodium</u> Grunow
- 6a. <u>Gyrosigma</u> Hassall
- b. Pleurosigma Wm. Smith
- 7. Grammatophora Ehrenberg
- 8. Lauderia Cleve
- 9. Coscinosira Gran
- 10. Thalassionema Grunow

Thalassiothrix Cleve and Grunow

- 11. Cocconeis Ehrenberg
- 12. Licmophora Agardh
- 13. Navicula Bory

Achnanthes Bory

Fragilaria Lyngbye

- 14. Nitzschia Hassall
- 15. Asterionella Hassall
- 16. Amphora Ehrenberg
- 17. Amphiprora Ehrenberg
- 18. Tropidoneis Cleve



· . .

Boyer, C.C. 1916. The Diatomaceae of Philadelphia and vicinity. Lippincott, Philadelphia, 143 pp.

Boyer, C.S. 1926-27. Synopsis of North Americal Diatomaceae. Phil. Nat. Acad. Sci. Proc. 78(Suppl.): 1.228; 79 (Suppl.); 229-583.

Cleve-Euler, A. 1951-55. Die Diatomen von Schweden und Finland, Vol. I-V. K. Sv. Vet. Akad. Handl., Ser. 5, Bd. 2, No. 1;, Bd. 3, No. 3; Bd. 4, No. 1, 5; Bd. 5, No. 4.

Cupp, E.E. 1943. Marine plankton diatoms of the west coast of North America. Scripps Inst. Oceanogr. Bull., Tech. Ser., 5: 1-237.

Gran, H. H. 1908. Diatomaceae, XIX pp. 1-146 In: Brandt and Apstein, Nordisches Plankton, I, Kiel and Leipzig.

Griffith, R. E. 1961. Phytoplankton of Chesapeake Bay. Ches. Biol. Lab. Contr. No. 172, iv. 79 pp.

Helmcke, J. G. & W. Krieger. 1960-64. Diatomeenschalen im Electronenmikroskopischen. Bild I-V.

Hendey, N. I. 1937. The plankton diatoms of the southern seas. Discovery Repts. 16: 151-364.

Hendey, N. I. 1964. An introductory account of the smaller algae of British coastal waters. Part V. Bacillariophyceae. Gt. Brit. Min. Fish. Investi. Ser. 4, xxii, 317 pp.

Heurck, H. F. van 1880-1885. Synopsis des Diatomees do Belgique. Anvers. Edite par l'auteur, 4 Vol., plates.

Heurck, H. F. van 1896 (1964). A treatise on the Diatomaceae. Translated by W. E. Baxter. London, William Wesley and Son. 558 pp., 35 pls., 291 text figs.

Hustedt, F. 1930. Die Kieselalgen Vol. VII in: L. Rabenhorsts, Kryptogramen-Flora, Leipzig.

Hustedt, F. 1955. Marine littoral diatoms of Beaufort, N.C. Duke Univ. Mar. Sta. Bull. 6, 67 pp.

LeBour, M. V. 1930. The plankton diatoms of northern seas, Ray Soc. ix, 244 pp. 4 pls.

Werff, H. H. van der 1958-61. Diatomeenflora van Nederland. afl. 1-6, (unbound sheets).