



PREPARATION OF MAGNETIC ANTIBACTERIAL COMPOSITE BEADS Fe₃O₄/ALGINATE/Ag

Le Thi Thu Ha^{1,2}, Vu Xuan Minh¹, Le Thi My Hanh¹, Le Trong Lu^{1,2},
Pham Thi Lan¹, Nguyen Tuan Dung^{1,2,*}

¹*Institute for Tropical technology, VAST, 18 Hoang Quoc Viet, Cau Giay, Ha Noi*

²*Graduate University of Science and Technology, VAST, 18 Hoang Quoc Viet, Cau Giay, Ha Noi*

*Email: ndung@itt.vast.vn

Received: 17 July 2018; Accepted for publication: 9 September 2018

ABSTRACT

In recent years, the synthesis of magnetic polymer beads has much attractive attention from many technological fields such as biomedical and environmental. In addition of stabilizing effect for magnetic particles core, the polymer shells can also be further functionalized, thus enlarging the spectrum of possible applications of magnetite nanoparticles. In this study, the novel magnetic composite beads of Fe₃O₄/alginate/Ag were synthesized and characterized for point-of-use water disinfection. Magnetite nanoparticles were prepared from waste spent pickling liquors and then were encapsulated by natural alginate to form the spherical beads of the size about 1-2 mm. Silver nanoparticles were loaded on the beads with the large content (24.62 wt.%). The as-prepared beads have a high saturation magnetization value, $M_s = 59$ emu/g, and very good antibacterial activity against both gram (-) and gram (+) bacteria. With the concentration 35 $\mu\text{g mL}^{-1}$ the magnetic beads can kill 100 % *Staphylococcus aureus*, *Escherichia coli* and *Pseudomonas aeruginosa*, whereas this value found 70 $\mu\text{g mL}^{-1}$ for *Lactobacillus fermentum*, 105 $\mu\text{g mL}^{-1}$ for *Salmonella enterica*, and 140 $\mu\text{g mL}^{-1}$ for *Bacillus subtilis*.

Keywords: magnetic antibacterial beads, sodium alginate, silver nanoparticles, water disinfection.

1. INTRODUCTION

Magnetic nanoparticles have received recently a considerable amount of attention due to their wide applications in various fields, which ranges from biomedical to industrial. Especially, in the field of water and wastewater treatment where the challenges associated with separation and recovery of the small size of adsorbent materials from contaminated water are becoming increasingly important [1]. However, the practical use of magnetic nanoparticles raises a series of problems concerning their intrinsic instability [2, 3]. Hence, they should be chemically stabilized by grafting or coating with organic or inorganic substances. Synthesis of magnetic polymer beads constitutes a new topic of research rapidly developing last decade. In addition of

stabilizing effect, the polymer shells can also be further functionalized, thus enlarging the spectrum of possible applications of magnetite nanoparticles [4, 5].

Alginate is a natural polysaccharide extracted from brown seaweed, it has many advantages such as low cost and availability, biocompatibility, biodegradability, and ease of gelation [6-8]. It has found numerous applications in diverse fields such as pharmaceutical, biomedical, agriculture, and environmental. There are some studies reported recently on the encapsulation of magnetic nanoparticles in the alginate matrix for different applications: drug delivery [6], heavy metal adsorption [7] or dye removal [8].

In the field of water purification, chlorination is the most widely used method for disinfecting water because of their high efficiency and low cost. But chlorine may react with natural organic matter to form the toxic by-products, some of which have been identified as carcinogenic [9]. In recent years, silver nanoparticles have been a well-known antibacterial agent due to its higher antimicrobial activities against gram-positive and gram-negative bacteria. Ag nanoparticles were also studied recently to immobilized onto various inorganic or organic substrates, to improve their safe and effective use in practice [10,11].

In this study, we synthesize the magnetic antibacterial composite beads Fe₃O₄/alginate/Ag for point-of-use water disinfection. The waste spent pickling liquors was used for magnetic nanoparticles synthesis, they have been encapsulated by the natural alginate and further immobilized with Ag nanoparticles. The as-prepared beads were characterized by FE-SEM, EDX methods, the magnetic and antibacterial properties were also examined.

2. EXPERIMENTAL

2.1. Chemicals

The Fe₃O₄ nanoparticles were synthesized from the spent pickling liquors of Hoa Phat Steel Co. according to our recent work [12]. Sodium alginate is an extracted product from natural brown algae of Nha Trang Institute of Technology Research and Application. Ca(NO₃)₂ of reagent grade was supplied from Xilong Scientific Co. (China). AgNO₃, NaBH₄ of analytical grade were purchased by Merck Co. (Germany).

2.2. Preparations and characterization of Fe₃O₄/alginate/Ag beads

The alginate bead formation was investigated at first, using Ca²⁺ solution: an aqueous solution of sodium alginate with various concentrations, from 0.2 to 1.5 wt%, was slowly injected into 0.3 M Ca(NO₃)₂ solution. After 24 hours for aging, the resulting beads were washed with distilled water three times. The shape of obtained beads was observed by the ZEISS optical microscope.

For the composite beads preparation, Fe₃O₄ nanoparticles were dispersed in the sodium alginate solution, the mass ratio of Fe₃O₄:alginate was 2:1. Injecting this solution to 0.3 M Ca(NO₃)₂ solution for composite granulation. After aging for 24 hours, the Fe₃O₄/alginate beads were collected by a magnet, rinsed three times with distilled water. The resulting Fe₃O₄/alginate beads were further dispersed in the AgNO₃ solution, with Ag⁺ concentration varied from 5 to 20 mM, stirring gently for 12 hours, then adding NaBH₄ to reduce Ag⁺ to Ag⁰, molar ratio NaBH₄:AgNO₃ was 1:1. Collect the product with a magnet, rinse it several times with distilled water.

The obtained composite beads were characterized by scanning electron microscope (S-4800, HITACHI, Japan), and energy dispersive X-ray spectroscopy (JSM-6510LV, Jeol, Japan). Magnetic properties were determined on the vibration sampling system (VSM).

2.3. Antibacterial test

Antibacterial activity of magnetic composite beads was tested by using of standard microdilution method which enables to determine the MIC (minimum inhibitor concentration) or MBC (minimum bactericidal concentration). Firstly, a serial dilution of composite is added in a 96-wells plate from high concentration, for example $128 \mu\text{g mL}^{-1}$, to zero, as blank sample. Then a bacterial suspension with 5×10^5 CFU/mL is added in every well. After 24 h at 37°C incubation the wells are visually evaluated for turbidity. For determining the MBC, 100 μL last three transparent suspensions are chosen to be spread on new agar plates. After another 24 h incubation at 37°C colony formation shows up and the lowest concentration with biocidal activity is taken as MBC. Different bacteria of both gram (+) and gram (-) were used for the tests: *Staphylococcus aureus* (Sa), *Bacillus subtilis* (Bs), *Lactobacillus fermentum* (Lf), *Salmonella enterica* (Se), *Escherichia coli* (E.coli), *Pseudomonas aeruginosa* (Pa).

3. RESULTS AND DISCUSSION

3.1. Formation of Fe_3O_4 /alginate bead and magnetic property

The bead formation is occurred immediately upon the contact of the alginate and Ca^{2+} , the mechanism of this gelation process involves guluronic residues with the specific chelation of Ca^{2+} forming the so-called “egg-box” structure [13]. The size and shape of the beads are highly dependent on the concentration of sodium alginate salt. Figure 1 presents the images of alginate beads observed on the optical microscope, the beads were prepared with different alginate concentrations. It is clear that with the low concentrations (less than 1 wt%), no uniform and round grain was produced. In the case of higher concentrations, from 1-1.5 %, the uniform spherical beads of diameter 1-2 mm were well formed.

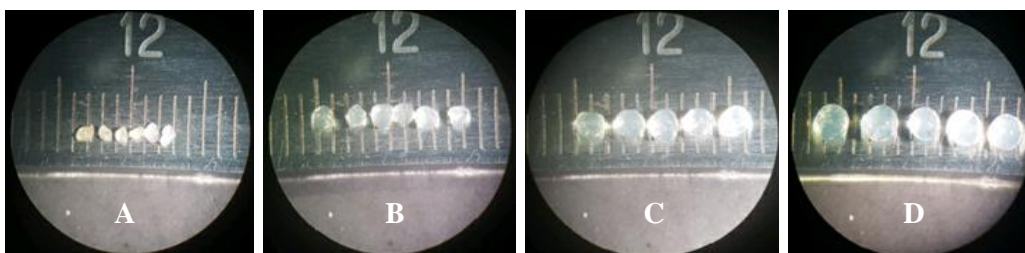


Figure 1. Optical images of alginate bead at different concentrations:

(A) 0.2 %; (B) 0.5 %; (C) 1 %; (D) 1.5 %.

The Fe_3O_4 /alginate composite beads were prepared by dispersion of Fe_3O_4 nanoparticles in 1 % sodium alginate solution, then the mixture was injected in Ca^{2+} solution. The prepared magnetic composite beads have a similar shape to a sole alginate beads (Figure 2A) and they can be collected by a magnet (Figure 2B).

The mass ratio of Fe_3O_4 :alginate was chosen 2:1. In principle the magnetic property of composite beads will be increased with increasing the Fe_3O_4 content, but it is difficult to

granulate the beads in the case of higher Fe_3O_4 content. Figure 2B presents the magnetization hysteresis curves of sole Fe_3O_4 nanoparticles and $\text{Fe}_3\text{O}_4/\text{alginate}$ beads, measured by the VSM magnetometry, under a magnetic field ranging from -10 to 10 kOe at room temperature. The results showed the high saturation magnetization for Fe_3O_4 nanoparticles, $M_s = 79 \text{ emu/g}$. This value decreased to 59 emu/g for the $\text{Fe}_3\text{O}_4/\text{alginate}$ composite beads.

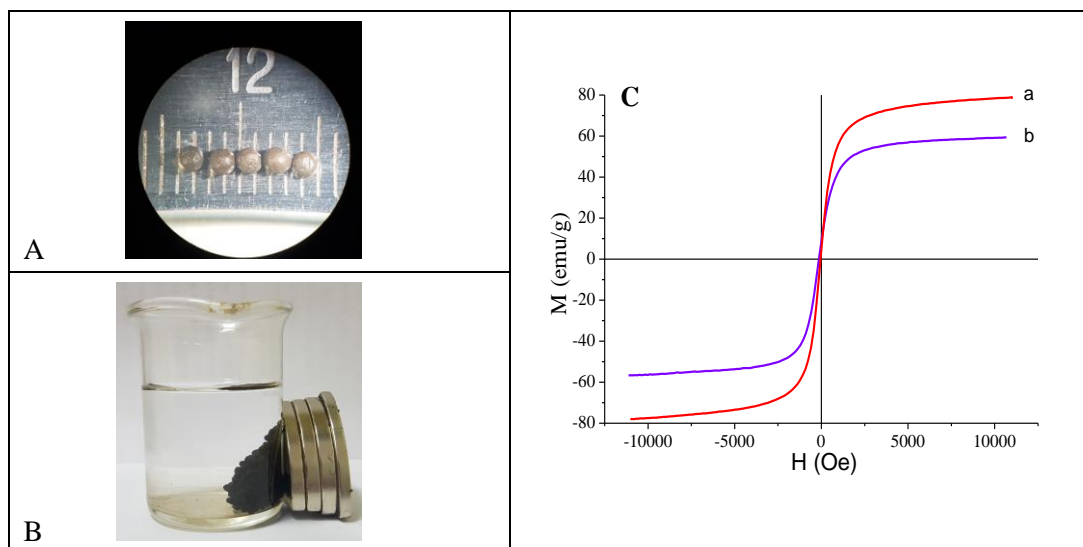


Figure 2. Optical image (A) and photo of $\text{Fe}_3\text{O}_4/\text{alginate}$ beads collection with a magnet (B); C: magnetization curve of Fe_3O_4 (a) and $\text{Fe}_3\text{O}_4/\text{alginate}$ (b).

3.2. Synthesis of $\text{Fe}_3\text{O}_4/\text{alginate}/\text{Ag}$ composite beads

$\text{Fe}_3\text{O}_4/\text{alginate}/\text{Ag}$ composite beads were synthesized by loading Ag^+ ion from AgNO_3 solution on the surface of $\text{Fe}_3\text{O}_4/\text{alginate}$ beads, then Ag^+ ions were reduced by NaBH_4 to metallic silver. The presence of silver in the composite beads can be examined by EDX analysis.

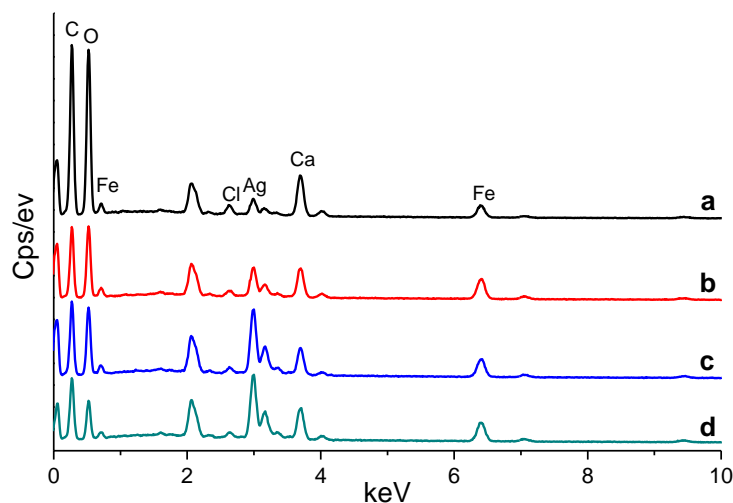


Figure 3. EDX spectra of $\text{Fe}_3\text{O}_4/\text{alginate}/\text{Ag}$ composite beads synthesized with different AgNO_3 concentrations: 5 mM (a), 10 mM (b), 15 mM (c), 20 mM (d).

The obtained EDX results demonstrated in Fig. 3 indicate clearly the chemical composition of the as-prepared magnetic beads that includes Fe, C, O, Ca, Cl and Ag. Intensity of the Ag peak at 3 keV was increased with increasing the concentration of AgNO₃ solution from 5 to 20 mM.

The chemical composition of Fe₃O₄/alginate/Ag beads has been also quantitatively determined and presented in Table 1. The Ag amount in the composite beads increased obviously when AgNO₃ concentration increased from 5 to 15 mM, in the case of 20 mM, the silver content increased a little, so the value 15 mM was selected for the synthesis of Fe₃O₄/alginate/Ag composite beads, with the incorporated Ag amount 23.8 wt% .

Table 1. Elements of Fe₃O₄/alginate/Ag nanocomposite beads synthesized with different concentrations of AgNO₃ solution.

Element	AgNO ₃ concentration							
	5 mM		10 mM		15 mM		20 mM	
	Weight (%)	Atomic (%)	Weight (%)	Atomic (%)	Weight (%)	Atomic (%)	Weight (%)	Atomic (%)
Fe	11.71	3.49	10.67	3.33	10.87	4.05	11.96	4.56
O	44.54	46.31	45.76	49.79	37.79	49.08	34.13	45.43
C	34.05	47.16	29.94	43.39	23.11	39.98	23.82	42.23
Ca	5.34	2.22	4.16	1.81	4.07	2.11	5.10	2.71
Cl	0.47	0.22	0.48	0.23	0.33	0.20	0.36	0.22
Ag	3.89	0.60	8.98	1.45	23.81	4.59	24.62	4.86
Total	100.00		100.00		100.00		100.00	

The structural morphology of Fe₃O₄/alginate/Ag composite beads (synthesized with 15 mM AgNO₃ solution) was characterized by field emission scanning electron microscope (FE-SEM), Fe₃O₄ nanoparticles and Fe₃O₄/alginate beads were also analyzed for comparison, the obtained results are shown in Figure 4.

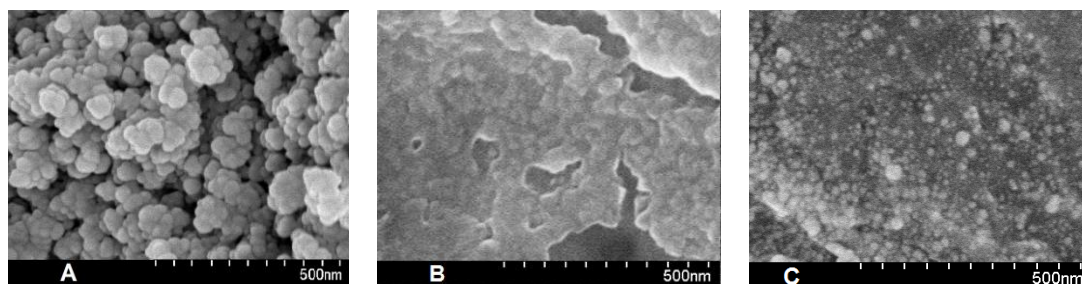


Figure 4. FE-SEM images of Fe₃O₄ particles (A), Fe₃O₄/alginate (B) and Fe₃O₄/alginate/Ag (C).

FE-SEM image of Fe₃O₄ showed the spherical particles with an average size of about 20 nm (Fig. 4A). Figure 4B shown the FE-SEM image of Fe₃O₄/alginate beads, the magnetic

nanoparticles homogenously dispersed within the polymer matrix. After the Ag incorporation onto magnetic beads, by Ag⁺ adsorption (in AgNO₃ solution) and then reduction by NaBH₄, the silver nanoparticles of about several nm were appeared in the bead surface as seen in Figure 4C.

3.4. Antibacterial tests

The antimicrobial activity of Fe₃O₄/alginate/Ag composite beads against different bacteria, *Staphylococcus aureus* (*Sa*), *Bacillus subtilis* (*Bs*), *Lactobacillus fermentum* (*Lf*); and Gram (-): *Salmonella enterica* (*Se*), *Escherichia coli* (*E.coli*), *Pseudomonas aeruginosa* (*Pa*), was evaluated, by using standard microdilution method which enables to determine the MBC (minimum bactericidal concentration) value. The obtained results were shown in Table 2.

Table 2. Antibacterial activity of Fe₃O₄/alginate/Ag composite beads.

Fe ₃ O ₄ /alginate/Ag content (µg/ml)	Percentage of inhibition of micro-organism (%)					
	Gram (+) bacteria			Gram (-) bacteria		
	<i>Sa</i>	<i>Bs</i>	<i>Lf</i>	<i>Se</i>	<i>E.coli</i>	<i>Pa</i>
140	100	100	100	100	100	100
105	100	0	100	100	100	100
70	100	0	100	20	100	100
35	100	0	40	0	100	100

As can be seen in Table 2, the magnetic beads Fe₃O₄/alginate/Ag present an excellent antibacterial activity for both gram (+) and gram (-) microorganisms. They can strongly inhibit *Sa*, *E.coli* and *Pa*, with low value MBC = 35 µg mL⁻¹, this value was 70 µg mL⁻¹ for *Lf*, 105 µg mL⁻¹ for *Se*, and 140 µg mL⁻¹ for *Bs*. Sole Fe₃O₄ and Fe₃O₄/alginate beads were also tested under the similar conditions for a comparison, they both exhibited no antibacterial activity, even with concentration of 140 µg mL⁻¹. Furthermore, Fe₃O₄/alginate/Ag composite beads are easily separated and recycled due to their good magnetic property. These results are very promising and we believe that the magnetic Fe₃O₄/alginate/Ag beads could be developed into the novel effective disinfection material.

4. CONCLUSION

In this work, we synthesized and characterized the magnetic composite beads Fe₃O₄/alginate/Ag of the size about 1-2 µm, with the large Ag content (24.62 wt.%). The as-prepared material has a high saturation magnetization value, M_s = 59 emu/g, and very good antibacterial activity against both gram (-) and gram (+) bacteria. With the concentration 35 µg mL⁻¹ the magnetic beads can kill 100 % *Staphylococcus aureus*, *Escherichia coli* and *Pseudomonas aeruginosa*, whereas this value found 70 µg mL⁻¹ for *Lactobacillus fermentum*, 105 µg mL⁻¹ for *Salmonella enterica*, and 140 µg mL⁻¹ for *Bacillus subtilis*. The results are well promising and we believe that this novel material has potential to be used in water disinfectant applications.

Acknowledgment. This research was financially supported by the project from Vietnam Academy of Science and Technology (code: VAST07.04/17-18). The authors also would like to thank Mr. Pham Hong Nam, Institute of Materials Science – VAST, for VSM measurements.

REFERENCES

1. Ambashta R. D., Sillanpää M. - Water purification using magnetic assistance: a review, *J. Hazard. Mater.* **180** (1-3) (2010) 38-49.
2. Tran V. H., Tran D. L., Nguyen N. T. - Preparation of chitosan/magnetite composite beads and their application for removal of Pb (II) and Ni (II) from aqueous solution, *Materials Science and Engineering: C*. **30** (2) (2010) 304-310.
3. Lu A. H., Salabas E. L., Schuth F. - Magnetic nanoparticles: synthesis, protection, functionalization, and application, *Angew. Chem. Int. Ed.* **46** (2007) 1222–1244.
4. Kalia S., Kango S., Kumar A., Haldorai Y., Kumari B., Kumar R. - Magnetic polymer nanocomposites for environmental and biomedical applications, *Colloid Polym. Sci.* **292** (2014) 2025–2052.
5. Ngoan N. T., Lam T. D., Cuong N. D., Loc N. T., Cham T. B., Binh N. H., Duong T. B., Nam P. H., Dzung N. T., Thai Hoa T., Dien P. G. – Facile synthesis of multifunctional Ag/Fe₃O₄-CS nanocomposites for antibacterial and hyperthermic applications, *Curr. Appl. Phys.* **15** (2015) 1482-1487.
6. Kondaveetia S., Cornejo D.R., Freitas D., Petria S. - Alginate/magnetite hybrid beads for magnetically stimulated release of dopamine, *Colloids and Surfaces B: Biointerfaces*, **138** (2016) 94–101.
7. Lakouraj M. M., Mojerlou F., Zare E. N. - Nanogel and superparamagnetic nanocomposite based on sodium alginate for sorption of heavy metal ions, *Carbohydr. Polym.* **106** (2014) 34–41.
8. Rocher V., Bee A., Siaugue J. M., Cabuil V. - Dye removal from aqueous solution by magnetic alginate beads crosslinked with epichlorohydrin, *J. Hazard. Mater.* **178** (2010) 434–439.
9. Feliziani E., Lichter A., Smilanick J. L., Ippolito A. - Disinfecting agents for controlling fruit and vegetable diseases after harvest, *Postharvest Biol. Technol.* **122** (2016) 53–69.
10. Oyanedel-Craver V. A., Smith J. A. - Sustainable colloidal-silver impregnated ceramic filter for point-of-use water treatment, *Environ. Sci. Technol.* **42** (3) (2008) 927-933.
11. Mthombeni N. H., Mpenyana-Monyatsi L., Onyango M. S., Momba Maggie N. B. - Breakthrough analysis for water disinfection using silver nanoparticles coated resin beads in fixed-bed column, *J. of Hazard. Mater.* **217-218** (2012) 133-140.
12. Nguyen T. D., Pham T. L., Le T. T. H., Vu X. M., Le T. M. H., Le T. M. H., Pham H. N., Le T. L. - Synthesis and antibacterial properties of a novel magnetic nanocomposite prepared from spent pickling liquors and polyguanidine, *RSC Adv.* **8** (2018) 19707–19712.
13. Grant G. T., Morris E. R., Rees D. A., Smith P. J. C., Thom D. - Biological interactions between polysaccharides and divalent cations: the egg-box model, *FEBS Letters* **32** (1) (1973) 195–198.