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Coral Growth and Skeletal Density Relationships in Some Branching Corals of the Red Sea, Egypt

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Abstract

The coral reefs at the exposed zones are exhibited to strong wave action, renewable water masses, clear seawater and small temperature variations, however, in the sheltered zones they exposed to high turbidity, high temperature variations and protected from the surge waves and currents. In situ measurements of seasonal and annual growth rates and the branch thicknesses using Varner Caliber and the laboratory measurements of skeletal densities using Archimedes's Principle were done in four branching coral species growing in the exposed and sheltered zones of Hurghada and Hamrawin at the northern Red Sea. Acropora humilis recorded the highest seasonal and annual growth rates at all zones; 0.68±0.02, 0.76±0.03, 0.66±0.03 and 0.69±0.02mm/month and 7.25±0.20, 7.96±0.33, 7.10±0.11 and 7.34±0.14mm/yr respectively. Pocillopora damicornis recorded the highest averages of skeletal density at the different zones of Hurghada and Hamrawin; 2.04±0.35, 1.64±0.26, 2.64±0.66 and 1.96±0.18gm/cm³ respectively and the highest averages of the branch thicknesses at the exposed and sheltered zones of Hurghada (1.66±0.42cm², 1.51±0.30cm²) while, A. humilis recorded the highest average of the branch thicknesses at the exposed and sheltered zones of Hamrawin (1.49±0.16cm², 1.14±0.08cm²). A. humilis was the fastest growing species in the worm season at the exposing and sheltered zones of Hurghada and Hamrawin, while P. damicornis was the slowest growing species because of it tends to form thick and dense branches. The oceanographic and local conditions as; temperature variations, aragonite saturation, turbidity, effects of surge waves and light intensity are responsible about the differences in the skeletal parameters of the studied species.

Keywords: Branching corals – Growth rates - Skeletal density – Branch thicknesses – exposed and sheltered zones

INTRODUCTION

Scleractinian corals have a complex porous skeleton containing more or less dense structure lattice. Branching corals grow rapidly at their branch tips (Hughes, 1987; Tunnicliffe, 1983), but they are often easily broken during storms and trampling (Woodly *et al.*, 1983; Hawkins and Roberts, 1994) or bleached by the interaction of the physical conditions and/or biological effects (Mohammed and Mohammed, 2005). Skeletal density of corals is a growth mode that has been studied by many authors as a function of increasing in height and volume of the newly grown skeletons (Oliver, 1984; Hughes, 1987; Ammar *et al.*, 2005). Kotb (2002) pointed out that the linear extensions and skeletal densities of corals are independent growth modes. The growth and skeletal deposition mechanisms are making the effect of each factor investigation separately very difficult (Loya, 1985). Coral growth or skeletogenesis is driven by calcification, the process whereby calcium (Ca⁺²) and carbonate (CO₃⁻²) ions obtained from seawater precipitate beneath the calcioblastic ectoderm (CE) of the coral polyp to form crystals of the calcium carbonate (CaCO₃) mineral polymorph, aragonite (Wooldridge, 2013). Growth rates are inherently variable among different corals species, partly in accordance with differences in gross morphology, skeletal structure and polyp size (Hall and Hughes 1996). Spatial, temporal and taxonomic differences in coral growth play an important role in the ecology and dynamics of coral reef ecosystems, affecting reef productivity, heterogeneity, and growth (Anderson *et al.*, 2012).

The annual growth rates and the skeletal densities can be affected by different physical factors such as temperature (Klein *et al.*, 1993), light and depth (Davies, 1991), reproduction (Tarrant *et al.*, 2004), salinity and sedimentation rates (Charuchinda and Hylleberg, 1984), water movement and wave actions (Bottjer, 1980; Brown and Howard., 1985; Rodgers *et al.*, 2003) in addition to the interference of a complex set of biotic or abiotic factors (Mohammed *et al.*, 2007). The changes in temperature, salinity, and sedimentation can influence not only growth but also diversity and abundance of corals (Lirman *et al.*, 2003), often, high energy and high sedimentation together can reduce growth (Cruz-Pinon *et al.*, 2003).

The present work aims to study the effects of oceanographic and local conditions on the growth rates, skeletal densities and branch thicknesses of the branching corals species at the exposed and sheltered zones of the Red Sea.

MATERIALS AND METHODS

1- Geomorphic and environmental settings of the studied localities

Two localities with different environmental settings and anthropogenic effects at Hurghada and Hamrawin in the northern Red Sea (Fig., 1) were selected to study the coral growths, skeletal densities and branches thicknesses



of four branching coral species; *Acropora humilis, Acropora hyacinths, Stylophora pistillata* and *Pocillopora damicornis*. These coral species inhabit the exposed and sheltered zones of the localities of investigation. The exposed zones were characterized by strong surge waves and current, renewable water masses, clear seawater and small temperature variations as well as these corals didn't suffer from exposing during low tide time. The corals in the sheltered zones are protected from the surge waves and currents but they are intensively suffering from high turbidity rates, high temperature variations as well as the solar radiations during the low tide time.

The sheltered zone at Hurghada is in front of NIOF (National Institute of Oceanography and Fisheries) is sheltered from the surge waves, strong currents and direct wind effects but slightly affected by overfishing and touristic activities (diving and snorkeling). It is also suffered from the neap tide events, long time of areal exposure during the semidiurnal tidal regime (Mohammed and Mohammed, 2005), direct sunlight exposure, retain high temperature in summer and low temperature in winter inside their basins, high sedimentation rates, landfilling, fishing and tourist activities (Dar *et al.*, 2012). The second zone at Hurghada is in the lee side of El Fanadir Rocky Islands directly exposed to intensive surge waves, wind induced waves and marine currents. Also it was intensively used for diving and snorkeling activities (Fig., 1b).

Hamrawin is a phosphate harbour and mining occupation located about 120km to the south of Hurghada along the Red Sea coast. It lies directly on small embayment to the south (Dar, 2005) at the downstream of Wadi Hamrawin that was used as navigation basin for shipping phosphate raw materials. Throughout the shipment processes, the transferring raw materials are exposing in most days to intensive winds, subsequently, the finest particles are smothering then fall down to sea. In the reverse wind days, the amount of volatized dusts geminates reaching to the embayment and the southern tidal area is about four or five folders due to the generated eddy winds. The sheltered zone of Hamrawin was inside the southern shipping embayment while the exposed zone included in the back and fore reef areas (Fig., 1c).

Materials and Methods

Sampling and *in situ* measurements were carried out in the shallow reefs using SCUBA diving at the exposed and sheltered zones of Hurghada and Hamrawin. Five colonies of each species of; *Acropora humilis*, *A. hyacinthus*, *Stylophora pistillata* and *Pocillopora damicornis* were chosen and marked at depth between 3 to 5m of each zone to study the growth rates. Five branches from each colony were tagged by plastic string at about 1.5-2.0 cm apart from the tip. The growth rates of the tagged branches were investigated by measuring the increasing in length (linear extension) bi-seasonally in winter and summer, from the plastic string to the tip of the branch (Mohammed, 2003) then the annual growth rates were calculated. After one year of the experiment, five fragment specimens of each species at each zone were collected and cleaned from the adhered materials then dried to study the skeletal densities and the branches thickness. The skeletal densities were measured using Archimedes's principle by weighting them then suspended briefly from an analytical balance into water (Graus and Macintyre, 1982; Ammar *et al.* 2005) while, the branch thicknesses were measured using vernier caliper of 0.01 mm accuracy (Mohammed *et al.* 2007; Dar and Mohammed, 2009). Some of the physico-chemical parameters were measured *in situ* at each location using Hydrolab (Hanna Model).

Results and Discussion

1- Oceanographic parameters of the studied sites

Seawater temperature at Hurghada was varied between 19.56° C in winter and 29.38° C in summer and at Hamrawin, it was changed between 18.89° C in winter and 29.03° C in summer (Table 1). Salinity was fluctuated between 39.75% and 42.43% at Hurghada and from 40.18% to 42.15% a Hamrawin. The pH Values shows the great variation at Hurghada in summer (7.83 - 8.61) and at Hamrawin in winter (7.7 - 8.05). The relatively high values of salinity and pH at Hurghada were due to shallowness the sheltered zone, the exposing to high temperatures for long time and dismissing the continuous water mixing.

2- Annual and Seasonal Growth rates

The annual linear growth rates of *Acropora humilis* shows an average of 7.25±0.20mm/yr at exposed zone and the average of 7.96±0.33mm/yr at the sheltered zone of Hurghada. At Hamrawin, it recorded the average of 7.10±0.11mm/yr at the exposed zone 7.34±0.14mm/yr at the sheltered zone (Table 2). *A. hyacinthus* recorded an average of 6.60±0.24mm/yr and an average of 6.88±0.23mm/yr at the exposed and sheltered zones of Hurghada. At Hamrawin, it was recorded the averages of 5.20±0.20mm/yr and 6.04±0.21mm/yr at the exposed and sheltered zones. According to Attalla *et al.*, (2011), the linear growth rates of *A. humilis* varied significantly according to the sheltering condition, with an overall mean of 8.89 and 6.98mm/year in exposed and sheltered sites of Hurghada, Red Sea. They added, the highest linear growth rate of *A. humilis* was estimated at the exposed site of Gotta Abu Ramada, being 9.17mm/year, while the lowest rate of 6.61mm/year was recorded at the sheltered site of El-Fanadir Reef at Hurghada. Mohammed *et al.*, (2007) founded that *Acropora humilis* have the higher growth rates (7.49 mm/y) than *Stylophora pistillata* (6.87mm/y). Aamer (2004) recorded linear growth rates of *A. humilis* at Sharm El-Shiekh, northern Red Sea between 6.17mm/year and 9.80mm/year, while



Davies (1989) reported that *A. humilis* in Caribbean Sea varied between 2-13.3mm/year at 2m depth. Yap *et al.*, (1992) documented that the *Acropora hyacinthus* consistently exhibited the lowest growth rates among the studied species in a northern Philippine.

The annual linear growth rate of *Stylophora pistillata* at the exposed zone of Hurghada recorded the average of 6.17±0.24mm/yr and at the sheltered zone recorded an average of 6.27±0.31mm/yr. At Hamrawin, *S. pistillata* showed the average of 6.02±0.19mm/yr at the exposed zone and 6.48±0.02mm/yr at the sheltered zone. *P. damicornis* showed annual growth rate averages of 6.37±0.15mm/yr and 7.13±0.21mm/yr at the exposed and sheltered zones of Hurghada. At Hamrawin *P. damicornis* linear growth rate averages were 5.97±0.13mm/yr at the exposed zone and 6.36±0.11mm/yr at the sheltered zone. Kotb (2001) recorded that the estimated annual rates of linear growth for the three corals considered at the different depths (5m, 15m, and 30m) were 9.24, 7.48, and 6.51mm/y for *S. pistillata*; 6.34, 9.24, and 5.90mm/y for *Acropora granulosa*; and 7.40 and 6.6mm/y for *P. damicornis*, respectively; *P. damicornis* was not recorded at 30m depth at Na'ama Bay, south of Sinai, northern Red Sea. Vago *et al.*, (1997) documented that the relative growth rate of *S. pistillata* does not remain constant throughout its lifetime, meaning that in this species, growth accelerates with colony size and age, probably until a certain size and age of incipient senescence are reached.

Seasonally, *Acropora humilis* shows the average of linear growth rate reaching 0.68 ± 0.02 mm/month through summer season and 0.53 ± 0.02 mm/month through winter season at the exposed zone of Hurghada. At the sheltered zone, it showed the averages of 0.76 ± 0.03 mm/month in summer and 0.56 ± 0.04 mm/month in winter. At Hamrawin, *A. humilis* recorded the growth rate averages 0.66 ± 0.03 mm/month and 0.52 ± 0.02 mm/month in summer and winter seasons respectively at the exposed zone, and at the sheltered zone the averages were 0.69 ± 0.02 mm/month in summer and 0.54 ± 0.01 mm/month in winter. The linear growth rate of *A. hyacinthus* showed the averages of 0.58 ± 0.03 mm/month and 0.52 ± 0.03 mm/month in summer and winter seasons at the exposed zone and it was averaged 0.64 ± 0.04 mm/month and 0.51 ± 0.01 mm/month in summer and winter seasons at Hurghada. At Hamrawin, the linear growth of *A. hyacinthus* recorded the averages of 0.50 ± 0.02 mm/month and 0.37 ± 0.02 mm/month through summer and winter seasons at the exposed zone and the averages of 0.55 ± 0.0 mm/month and 0.46 ± 0.04 mm/month through the same seasons at the sheltered zone respectively (Table 2)

S. pistillata linear growth averaged of 0.56±0.01 and 0.47±0.04 mm/month in summer and winter at the exposed zone and 0.58±0.03 and 0.47±0.04 mm/month in summer and winter at the sheltered zone of Hurghada. At Hamrawin, S. pistillata recorded the averages of 0.55±0.02, 0.46±0.02, 0.56±0.00 and 0.52±0.00mm/month in summer and winter seasons at the exposed and sheltered zones respectively. P. damicornis showed the averages of 0.58±0.02mm/month in summer and 0.48±0.01mm/month in winter at the exposed zone and 0.61±0.02mm/month in summer and 0.57±0.03mm/month in winter at the sheltered zone of Hurghada. At Hamrawin, the averages of linear growths were; 0.51±0.01, 0.48±0.01, 0.56±0.00 and 0.50±0.01mm/month in summer and winter seasons at the exposed and sheltered zones respectively. Gillette (2012) recorded that P. damicornis showed the highest skeletal extension rates under optimal (26°C) temperatures. Jokiel and Coles (1977) found that the skeletal growth in P. damicornis in Hawaii is greatest at about 26–27°C. Piniak and Brown (2008) documented that, P. damicornis growth rates in Ma'alaea Harbor were highest in the summer, when both light and temperature were at their maximum. Anderson et al., (2012) found that, P. damicornis was the slowest growing corals (0.54-1.20 mm/month) at Lord Howe Island and the linear extension for P. damicornis were 29-68% lower when compared to the summer growth rates at the Solitary Islands (Harriott 1999) because of P. damicornis is allocating most of its energy for reproduction in the summer accounting for the reduced summer growth. Inversely, Al-Sofyani and Floos (2013) found that the highest skeletal growth rate of P. damicornis was during winter whilst the lowest value was during summer but the difference was not significant.

Changes in environmental conditions may have both positive and negative effects on coral growth. Most corals are adapted to local environmental temperature (Hughes *et al.* 2003), and typically bleach and die if the local temperature exceeds the normal summer maxima by >1°C for 3 to 4 weeks (Hoegh-Guldberg, 1999). Branching corals are generally considered to be much more susceptible to extreme temperature (Marshall and Baird 2000), compared to massive coral species (Anderson *et al.*, 2012). Small changes in temperature can cause significant (up to 50%) changes in growth rates (James *et al.*, 2005). In general, the studied species were develop well in the worm season at the sheltered zones than those in the exposed zones whereas the temperature degree of the seawater around the average of 28°C. Anderson *et al.*, (2012) reported that the coral growth is currently limited by the cool winter temperatures and climate related increases in ocean temperature may extend the summer growing period. Marshall and Baird (2000) pointed out that the elevated temperatures have been shown to have a greater affect on branching species, such as *Acropora* and *Pocillopora*. Crossland (1981) documented that the greater skeletal elongation with increasing temperature is observed seasonally in the subtropics and the rising global temperatures could support greater subtropical coral growth rates. At some subtropical reefs, coral growth is currently limited by cool winter temperatures (Harriott 1999; Crossland 1981), whereby coral growth is negligible during winter months (Anderson *et al.*, 2012). Anomalously low (below 18 °C) and high (much



above 30 °C) seawater temperatures can impair coral physiology (Tambutté *et al.*, 2011). Gladfelter (1984) Correlated skeletal growth of *Acropora cervicornis* and four environmental parameters; temperature, daylight hours, sun hours, plankton abundance. She found that linear extension does not change during the year with the possible exception of April. It is suggested that temperatures outside an optimal range (ca. 26°–29°C for staghorn *Acroporas*) might cause a decrease in linear extension.

A. humilis recorded the highest increasing percentage in the seasonal growth among the studied coral species at the exposed and sheltered zones of Hurghada (29.13% and 35.85% respectively) and sheltered zone of Hamrawin (28.48%), A. hyacinthus recorded the highest growth percentage at the exposed zone of Hamrawin (34.54%) and the lowest seasonal growth percentage at the exposed zone of Hurghada (9.82%). S. pistillata recorded the lowest seasonal growth percentage at the sheltered zone of Hamrawin (7.30%), while P. damicornis recorded the lowest growth percentage (7.52% and 6.66%) at the sheltered zone of Hurghada and the exposed zone of Hamrawin respectively (Fig., 2). A. humilis was the fastest growing species in the worm season at the exposing and sheltered zones of Hurghada and Hamrawin, while P. damicornis was the slowest species in growth rate under the different conditions because of P. damicornis tends to form thick and dense branches relative to the other species especially under the sheltered conditions. Veron and Pichon (1976) reported that the growth forms typically show a gradual transition from a compact shape, under exposed (high energy fore-reef) conditions, to a thinner branching shape under sheltered (low energy back reef or lagoonal reefs) conditions. Such morphological variations are reflected in the annual extension and density pattern of the growth forms, where linear extension rates are lowest and bulk densities highest at the exposed reef sites (Scoffin et al., 1992; Logan and Tomascik 1994). Harriott (1998) suggested that the high growth rate and trend towards thinner branches for the lagoonal sites is consistent with the appearance of colonies in such habitats. She added, the lagoonal basins are frequently dominated by monogeneric stands of staghorn corals, which are tall with finebranching structure. Such habitats are protected from prevailing weather conditions with little water movement. Muko et al., (2013) attributed the decline in growth rates of Acropora in the exposed sites to the physical disturbances and in the sheltered sites to the thermal stresses. Guzman and Cortes (1989) suggested that the seasonal growth, which is greater during the dry season, may be affected by variations in available light, cloud cover, turbidity, salinity and reproductive time rather than temperature changes. Anderson et al., (2012) documented that the growth rates of Acropora corals at Lord Howe Island recorded (4.15-3.40mm/month) which are much greater than the recorded in the present study and the growth rates recorded by Harriott (1999) at the Solitary Islands; 0.46 mm/month for A. cytherea and 0.80 mm/month for A. valida. Lough (2008) reported that the average linear extension and calcification rates in the massive coral Porites are significantly linearly related to average sea surface temperature (SST). Brachert et al., (2013) concluded that the predicted SST rise over the next decades is likely hazardous to coral health because precipitation of the calcareous skeleton depends primarily on SST.

3- Skeletal densities

The different coral species recorded the highest averages of the skeletal densities and branch thicknesses at the exposed zones of Hurghada and Hamrawin. P. damicornis recorded the highest averages of skeletal density at the different zones of Hurghada and Hamrawin (Fig., 3); 2.04, 1.64, 2.64 and 1.96gm/cm³ respectively, A. hyacinthus recorded the lowest averages of skeletal density at exposed and sheltered zones of Hurghada (1.13, 1.09 and 1.26gm/cm³) and the sheltered zone of Hamrawin, while S. pistillata recorded the lowest skeletal density average (1.62gm/cm³) at the exposed zone of Hamrawin (Table 3). Skeletal densities at exposed zone were higher than the sheltered zone of Hurghada with percentages between 1.6% in S. pistillata and 32.20% in A. humilis. At Hamrawin, the exposed zone species were higher in the skeletal densities than the sheltered zone species by a percentage varied between 26.46% in S. pistillata and 34.52% in P. damicorins. Also, the species of exposed zone of Hamrawin were denser than those in the exposed zone of Hurghada with a percentage fluctuated between 26.79% in A. humilis and 66.54% in S. pistillata and the species of the sheltered zone of Hamrawin were denser than the species in the sheltered zone of Hurghada by a percentage varied between 16.91% in A. hyacinthus and 38.35% in S. pistillata (Table 3). Tambutté et al., (2011) found up to a 4.5% increasing in calcification with a 1°C elevation in seawater temperature. Harriott (1998) demonstrated that coral calcification rate was positively and significantly correlated (R=0.8) to seawater temperature. According to Lough and Barnes, (2000), a 1°C increase in mean annual temperature increased mean annual coral calcification rate by about 3.1%. Brown and Howard (1985) reported that the corals in exposed areas can have denser skeletons and hence may be more resistant to breakage than those in more sheltered areas. The high-extension, low density and skeletal growth are the common sclera-chronological signal of combined sedimentation and eutrophication effects on corals (Edinger et al., 2000; Carricart-Ganivet and Merino, 2001). Harriott (1997) examined variability of skeletal density in Acropora formosa between sites within the Houtman Abrolhos - Australia, and between the Abrolhos and a more tropical site. She found that corals from more protected sites at the Abrolhos were less heavily calcified than corals from more exposed sites, and that corals from the tropical site were less heavily calcified than those from Houtman Abrolhos. A detailed study of bulk density of Acropora formosa showed that



porosity varies significantly with degree of exposure at Houtman Abrolhos, with lagoonal corals being more lightly calcified than those from more exposed sites.

4- Branch thicknesses

P. damicorins showed the highest averages of the branch thicknesses at the exposed and sheltered zones of Hurghada (1.66cm², 1.51cm²), A. humilis recorded the highest average of the branch thicknesses at the exposed and sheltered zones of Hamrawin (1.49cm², 1.14cm²), A. hyacinthus recorded the lowest branch thicknesses at the exposed and sheltered zones of Hurghada (0.87cm², 0.70cm²) and S. pistillata showed the lowest averages of branch thicknesses (0.82cm² and 0.67cm²) at the exposed and sheltered zones of Hamrawin respectively (Fig., 4). S. pistillata recorded the highest variation percentage in branch thickness (53.33%), while P. damicornis showed the lowest variation percentage (13.09%) between the exposed and sheltered zones of Hurghada. At Hamrawin, P. damicornis recorded the highest variation percentage (40.29%) in branch thickness between the exposed and sheltered zones and A. hyacinthus recorded the lowest variation percentage (20.29%). Dar and Mohammed (2009) found that, Acropora humilis and Stylophora pistillata have thicker branches at the exposed sites than those at the sheltered sites, indicating to that the coral species are developing well under the effects of sea currents and surge waves. Harriott, (1998) pointed out, in protected habitats, linear extension is faster, but branches are more lightly calcified, so that net accretion of calcium carbonate does not vary greatly between habitats. Where water movement is high, thicker, shorter, more heavily calcified branches are produced which would be more resistant to damage from wave action.

The relatively higher growth rates, skeletal densities and branch thicknesses of the studied species in the exposed and sheltered zones of Hamrawin than those at Hurghada may also be affected by the phosphate particulates in the water column that are scavenged and introduced to the skeletal frameworks of the different species. This process may reduce the required energy to buildup the framework of the corals from the ionic forms of calcium and carbonates. Godinot et al., (2012) found that skeletal growth rate increased by 31% between un-enriched and P-enriched corals, whereas, phosphate-enriched corals incorporated 1.7 times more phosphorus into their skeleton than did un-enriched corals. They added that phosphate enrichment mainly affected the coral symbionts, by increasing their carbon, nitrogen, and phosphorus contents and their specific growth rate. Phosphate enrichment also affected the skeletal compartment, by increasing the skeletal growth and the P/Ca ratio. Dunn et al., (2012) found linear extension and tissue growth increasing for Acropora muricata exposed to phosphate levels of 0.09, 0.20, and 0.50 mg L⁻¹ for four months and the growth rates were highest at a phosphate concentration of 0.50 mg L⁻¹. They suggested that the phosphate enhanced growth was due to increased zooxanthellar populations and photosynthetic production within the coral. Skeletal density reduction may be due to phosphate binding at the calcifying surface and the creation of a porous and structurally weaker calcium carbonate/calcium phosphate skeleton. Increased phosphate concentrations, often characteristic of eutrophic conditions, caused increased coral growth but also amore brittle skeleton.

The high sedimentation levels limit light availability and consequently reduce the coral growth (Barnes and Lough, 1999; James et al., 2005). On the other hand, these sediment particles in water column may increase the aragonite saturation of seawater that may also help in skeleton formation. Aragonite saturation declines with increasing latitude and climate induced ocean acidification may further reduce the capacity for growth of calcifying organisms at the latitudinal limits of reef growth (Anderson et al., 2012). Ohde and Hossain (2004) found direct relationship between aragonite saturation in the seawater and the coral calcification. They reported, such coral calcification plays an important role in constructing reefs in warm shallow-water in which many organisms grow. Since a decrease in coral calcification leads to a decrease in reef-building capacity of corals, it is important to elucidate factors controlling coral calcification. The calcium carbonate budget, and in turn skeletal density, varies over space and time as a function of location in water current (Le-Campion-Alsumard et al., 1993), light intensity (Davies, 1991) or temperature (Tunnicliffe, I 983). Ohde and van Woesik (1999) suggested that coral calcification depends on the saturation state of surface seawater with respect to aragonite. The increasing in temperature during summer season increases aragonite saturation in the seawater, consequently the rate of calcification and coral growth increasing too. Silverman et al., (2007) indicated that summer (April-October) and winter (November-March) in the northern Red Sea have average calcification rates of corals varied between 60±20 and 30±20 mmol.m⁻².d⁻¹, respectively. They added, in general, calcification increased with temperature and aragonite saturation state of reef water with an apparent effect of nutrients, which is in agreement with most laboratory studies and in situ measurements of single coral growth rates. Calcium carbonate accretion is most strongly correlated with number of sun hours (Gladfelter, 1984). Harriott (1998) pointed out that carbonates of Acropora formosa were deposited in similar amounts, but either as porous, rapidly ex-tending branches, or as denser branches which extended more slowly. Lough and Barnes (1997) observed an increase in calcification rate of 3.5% for every 1°C rise in SST for the massive coral Porites lutea between 1906 and 1982 on the Great Barrier Reef, Positive relationships between temperature and calcification were also noted in studies that examined coral growth rates over latitudinal scales (Lough and Barnes, 2000; Carricart-Ganivet, 2004). Cooper et al., (2007) defined an optimum temperature of 26.7°C for extension and calcification of



massive *Porites* on the Great Barrier Reef, with extension and calcification rates decreasing by 15% per 1°C either side of this temperature. They also observed a decrease in calcification (~21%), linear extension (~16%) and bulk density (~6%) of colonies of the massive coral *Porites* on the northern Great Barrier Reef over a 16-year period (1988–2003) that also coincided with an increase in SST (0.24°C per decade) over the 16 years examined. Tanzil *et al.*, (2009) suggested that the warmer sea temperature regime at Phuket is a factor associated with a more precipitous decline or whether the corals in each location are thermally acclimatized to their respective local regimes. They also found a decrease in only calcification and linear extension rates while bulk density remained unchanged. Gladfelter (1984) recorded the monthly rates of linear extension and calcification with environmental variables in *Acropora cervicornis* by, suggested that temperatures outside the optimal range might cause a decrease in linear extension while calcification was more influenced by light availability.

5- Growth rates, skeletal densities and branch thicknesses relationships

From the data, there inverse relationships were observed between the coral growth in a side with coral density and branch thickness in the other side in the studied coral species and positive relationships were observed between coral density and branch thickness with different significances. These relationships were clearly observed in A. humillis (r = -0.53, -0.50 and 0.54 respectively (Fig. 5 2a,b,c) but they were decreased in their significances with the other species depending up on the continuous variability in the surrounding oceanographic conditions at the studied zones (Figs, 6a,b and 7). Also, the inverse relationships of the linear growth with coral density and branch thickness indicated that the more dense species and the thick branches at the exposed zones of Hurghada and Hamrawin are less porous than those in sheltered zones. The inverse relationships of skeletal density and extension rates are affected by light and temperature in the shallower areas (Lough and Barnes, 2000; smith et al., 2007; Tanzil, et al., 2009). Lough, (2008) found inverse relationship of the average skeletal density versus the linear extension rate and calcification rate whereas the linear extension is the main source of variability in calcification rate. He added, any effects of declining calcification on linear extension may also be offset by reductions in skeletal density. While the exposed areas are suffering from the high and surge waves and the observed high water currents, so they consumed some of their energies in the calcification process rather than the growth. So that, the branch thickness increases as a result of calcification and consequently their skeletal density increases. Rodgers et al., (2003) pointed out that, the skeletal strength was correlated with the wave energy present in the environment in the region they inhabit; suggesting that structural strength of corals is an adaptive response to hydraulic stress. The great uncertainties observed in the different illustration figures attributed to the effect of the different oceanographic variables at the studied zone. Temperature, light, sedimentation rate, dissolved nutrients and wave energy were the most effective oceanographic variables that control coral growths, densities and branch thicknesses.

Conclusions

- Linear growth rates, coral densities and branch thicknesses were studied in four branch coral species; *Acropora humilis*, *A. hyacinthus*, *Stylophora pistillata* and *Pocillopora damicornis* at two localities under the exposed and sheltered conditions.
- The growth rates of the selected coral species were noticed biseasonally (winter and summer) throughout a year. After a year of the experiment, coral density and branch thicknesses were measured.
- Acropora humilis recorded the linear highest annual and seasonal growth rates the two localities under the different conditions, while Stylophora pistillata showed the lowest annual and seasonal linear growth rates Hurghada and Acropora hyacinthus at Hamrawin.
- A. humilis was the fastest growing species in the worm season at the exposing and sheltered zones of Hurghada and Hamrawin, while P. damicornis was the slowest species in growth rate under the different conditions because of P. damicornis tends to form thick and dense branches relative to the other species especially under the sheltered conditions.
- *P. damicornis* recorded the highest averages skeletal density at the different zones of Hurghada and Hamrawin, *A. hyacinthus* recorded the lowest averages of skeletal density at exposed and sheltered zones of Hurghada and the sheltered zone of Hamrawin, while *S. pistillata* recorded the lowest skeletal density average at the exposed zone of Hamrawin.
- P. damicorins showed the highest averages of the branch thicknesses at the exposed and sheltered zones of Hurghada, A. humilis recorded the highest average of the branch thicknesses at the exposed and sheltered zones of Hamrawin, A. hyacinthus recorded the lowest branch thicknesses at the exposed and sheltered zones of Hurghada and S. pistillata showed the lowest averages of branch thicknesses at the exposed and sheltered zones of Hamrawin.
- Inverse relationships were observed between the coral growth in a side with coral density and branch thickness in the other side in the studied coral species and positive relationships were observed between coral density and branch thickness with different significances. These relationships were clearly



- observed in A. humillis but these relationships were decreased in their significances with the other species depending up on the continuous variability in the surrounding oceanographic conditions at the studied zones
- Temperature rates, aragonite saturation, phosphate particulates, sedimentation rates, effects of surge waves and light intensity are the main variables that controlling the coral growths, densities and branch thicknesses at the different zones.

REFERENCES

- Aamer, M.A.A., (2004). The role of zooplankton and water quality on some biological and ecological aspects of corals along the Egyptian Red Sea coast. A Ph.D. Thesis, Suez Canal University, Ismailia, Egypt; 325 pp.
- Al-Sofyani, A.A. and Floos, Y.A.M. (2013). Effect of temperature on two reef-building corals *Pocillopora* damicornis and *P. verrucosa* in the Red Sea. **OCEANOLOGIA**, 55 (4): 917–935.
- Ammar, M. S. A.; Mohammed, T. A. A.; Mahmoud, M. A. (2005): Skeletal density (strength) of some corals in an actively flooding and a non flooding site, south Marsa Alam, Red Sea, Egypt. *J. Egypt. German Soci.* Zool. 1-15.
- Anderson, K.; Pratchett, M. and Baird, A. (2012). Summer growth rates of corals at Lord Howe Island, Australia. 4C Coral reef response to multiple stresses: organisms to ecosystems. Proceedings of the 12th International Coral Reef Symposium, Cairns, Australia, 9-13 July.
- Attalla, T.M.; Hanafy, M.H. and Aamer, M.A. (2011). Growth rates of the two reef-building species, *Acropora humilis* and *Millepora platyphylla* at Hurghada, Red Sea, Egypt. Egypt J. Aquat. Biol. & Fish., 15(2): 1-15.
- Barnes, D.J. and Lough J.M. (1999) Porites growth characteristics in a changed environment: Misima Island, Papua New Guinea. Coral Reefs 18:213-218.
- Bottjer, D. J. (1980). Branching morphology of the reef coral Acropora cervicornis in different hydraulic regimes J. Paleobiol., 54: I 102.-1107.
- Brachert, T. C.; Reuter, M.; Kruger, S.; Bocker, A.; Lohmann, H.; Mertz-Kraus, R. and Fassoulas, C. (2013). Density banding in corals: barcodes of past and current climate change. Coral Reefs, 32:1013–1023.
- Brown, B.E. and L.S. Howard, (1985). Assessing the effects of "stress" on reef corals. *Adv. Mar. Biol., Vol. 22*, pp. 1-63.
- Carricart-Ganivet JP, Vásquez-Bedoya, LF, Cabanillas-Terán N, Blanchon P (2013). Gender-related differences in the apparent timing of skeletal density bands in the reef-building coral *Siderastrea siderea*. Coral Reefs. Volume 32, Issue 3, pp.769-777
- Carricart-Ganivet, J. P. and Merino, M. (2001). Growth responses of the reef-building coral *Montastraea* annularis along a gradient of continental influence in the southern Gulf of Mexico, Bull. Mar. Sci., 68, 133–146
- Carricart-Ganivet, J.P. (2004). Sea surface temperature and the growth of the West Atlantic reef-building coral Montastraea annularis. J. Exp. Mar. Biol. Ecol. 302, 249–260.
- Charuchinda, M. and Hylleberg, J. (1984) Skeleton extension of Acropora formasa at a fringing reef in the Andaman Sea. Coral Reefs 3: 215-219.
- Cooper, T.F.; De'ath, G.; Fabricius, K.E. and Lough, J.M. (2007). Declining coral calcification in massive Porites in two nearshore regions of the northern Great Barrier Reef. Global Change Biol 14:529–538.
- Crossland, C.J. (1981). Seasonal growth of *Acropora cf. formosa* and *Pocillopora damicornis* on a high latitude reef (Houtman Abrolhos, Western Australia). Proc 4th Int Coral Reef Sym 1:663-667
- Cruz-Pinon G, Carricart-Ganivert JP, Espinoza-Avalos J (2003). Monthly skeletal extension rates of the hermatypic corals *Montastrea annularis* and *Montastrea faveolata*: biological and environmental controls. Mar. Biol. 143:491–500.
- Dar, M.A. (2005). Coastal habitats degradation due to chronic and recent landfilling along the Red Sea. First Ain Shams University International Conference on Environmental Engineering, Faculty of Engineering, Ain Shams Univ., 9-11 April, Cairo, Egypt (773-786).
- Dar, M. A. and Mohammed, T. A. A. (2009): Seasonal variations in the skeletogensis process in some branching corals of the red sea. International Journal of Marine Sciences (Thalassas), 25 (1): 31-44
- Dar, M. A.; Mohammad, T. A. and El-Saman M. I. (2012): Geomorphic and Geographic Distributions of Reef Building and Soft Coral Communities in the Inshore Zone of the Red Sea. *Egyptian Journal of Aquatic Research*, 38: 105–117.
- Davies, P. S. (1989). Short-term growth measurements of corals using an accurate buoyant weighing technique. Mar. Biol., 101, 389-395.
- Davis, P S. (1991). Effect of daylight variations on the energy budgets of shallow-water corals. Mar. Biol., 108,137: 144
- Dunn, J.G.; Sammarco, P.W. and LaFleur Jr., G. (2012). Effects of phosphate on growth and skeletal density in the scleractinian coral *Acropora muricata*: A controlled experimental approach. Journal of Experimental



- Marine Biology and Ecology 411: 34-44.
- Edinger, E. N., Limmon, G. V., Jompa, J., Widjatmoko, W., Heikoop, J., and Risk, M. (2000). Normal coral growth rates on dying reefs: are coral growth rates good indicators of reef health?, Mar. Pollut. Bull., 40, 404–425.
- Gillette, P. (2012). Intraspecific genetic variability in temperature tolerance in the coral *Pocillopora damicornis*: Effects on growth, photosynthesis and survival, Ph.D. Thesis, University of Miami, USA.
- Gladfelter, E.H. (1984). Skeletal development in Acropora cervicornis: III A comparison of monthly rates of linear extension and calcium carbonate accretion measured over a year. Coral Reefs, 3:51–57.
- Godinot C.; Ferrier-Pagès C.; Montagna P. and Grover, R. (2011). Tissue and skeletal changes in the scleractinian coral Stylophora pistillata Esper 1797 under phosphate enrichment. Journal of Experimental Marine Biology and Ecology. 200-207.
- Graus, R. R. and MacIntyre, I. G. (1982). Variation in growth forms of the reef coral M. annularis (Ellis & Solander): a quantitative evaluation of growth response to light distribution using computer simulation. In: Rützler, K., Macintyre, I. G. (ed.) The Atlantic Barrier Reef ecosystem at Carrie Bow Cay, Belize. I. Structure and communities. Smithson. Contr. mar. Sci. 12: 441-464.
- Guzman, H.M. and Cortes, J. (1989). Growth rates of eight species of scleractinian corals in the Eastern Pacific (Costa Rica). BULLETIN OF MARINE SCIENCE, 44(3): 1186-1194.
- Hall, V.R. and Hughes, T.P. (1996). Reproductive strategies of modular organisms: comparative studies of reefbuilding corals. Ecology 77: 950-963.
- Harriott, V.J. (1997). Skeletal bulk density of the scleractinian coral *Acropora formosa* (Dana 1846) in tropical and subtropical Western Australia. In: Wells FE (ed) The ora and fauna of the Houtman Abrolhos Islands, Western Australia. (Proc 7th int. mar. boil. Wkshop) Vol. 1. Western Australian Museum, Perth, pp 75-82.
- Harriott, V. J. (1998). Growth of the staghorn coral Acropora Formosa at Houtman Abrolhos, Western Australia. Marine Biology, 132: 319-325.
- Harriott, V.J. (1999). Coral growth in subtropical eastern Australia. Coral Reefs, 18: 281-291.
- Hawkins, J.P. and Roberts, C.M. (1994). The growth of coastal tourism in the Red Sea: Present and future effects on coral reefs. Ambio 23, 503–508.
- Hoegh-Guldberg, O. (1999). Climate change, coral bleaching and the future of the world's coral reefs. Mar Freshw Res 50: 839-866
- Hughes, T.P.; Baird, A.H.; Bellwood, D.R.; Card. M.; Connolly, S.R.; Folke, C.; Grosberg, R.; Hoegh-Guldberg, O.; Jackson, J.B.C.; Kleypas, J.; Lough, J.M.; Marshall, P.; Nyström, M.; Palumbi, S.R.; Pandolfi, J.M.; Rosen, B. and Roughgarden, J. (2003). Climate change, human impacts, and the resilience of coral reefs. Science 301: 929-933.
- Hughes, T.P. (1987). Skeletal density and growth form of corals. Mar. Ecol. Prog. Ser., 35: 259-266.
- James, M.; Crabbe, C. and Smith, D.J. (2005). Sediment impacts on growth rates of *Acropora* and *Porites* corals from fringing reefs of Sulawesi, Indonesia. Coral Reefs, 24: 437–441.
- Jokiel, P. L. and Coles. S. L. (1977). Effects of temperature on the mortality and growth of Hawaiian reef corals. Mar. Biol. (Berl.). 43:201–208.
- Klein, R., Patzold, J., Wefer, G. and Loya, Y. (1993) Depth related timing of density band formation in *Porites* spp. corals form the Red Sea inferred from X ray chronology and stable isotope composition. *Marine Ecology Progress Series*, **97**, 99 104.
- Kotb, M.M.A. (2002). Skeletal density of three reef-building coral reef species in the Northern Red Sea, Egypt. J. Egypt. German Soci. Zool. Vol.39D: 1-16.
- Kotb, M.M.A. (2001). Growth rates of three reef-building coral species in the northern Red Sea, Egypt. Egypt J. Aquat. Biol. And Fisheries, 5(4):165-185.
- Le-Campion-Alsumard. T.; Romano, J. C; Peyrot. Clausade. M; LE. Campion, J. and Paul, R. (1993) Influence of some coral communities on calcium carbonate budget of Tiahura Reef (Moorea, French Polynesia). Mar. Biol. I 15 (4): 685-693.
- Lirman, D.; Orlando, B.; Macia, S.; Maqnzello, D.; Kaufman, L.; Biber, P. and Jones, T. (2003). Coral communities of Biscayne Bay, Florida and adjacent offshore areas; diversity abundance, distribution and environmental correlates. Aqu. Conserv. Mar. Freshwater Ecosyst. 13:121–135.
- Logan, A. and Tomascik, T. (1994). Linear extension growth rates in two species of *Diploria* from high-latitude reefs of Bermuda, Coral Reefs, 13, 225–230.
- Lough, J.M. (2008). Coral calcification from skeletal records revisited. Mar Eco Prog Ser 373: 257-264
- Lough, J.M. and Barnes, D.J. (2000) Environmental controls on growth of the massive coral Porites. Journal of Experimental Marine Biology and Ecology, 245, 225 243.
- Lough, J.M. and Barnes, D.J., (1997). Several centuries of variation in skeletal extension, density and calcification in massive Porites colonies from the Great Barrier Reef: a proxy for seawater temperature and a background of variability against which to identify unnatural change. J. Exp. Mar. Biol. Ecol. 211 (1), 29–



67

- Loya, Y. (1985). Seasonal changes in growth rate of a Red Sea coral population. Fifth International Coral Reef Congress, Tahiti, 6:187-191
- Marshall, P.A. and Baird, A.H. (2000). Bleaching of corals on the Great Barrier Reef: differential susceptibilities among taxa. Coral Reefs 19: 155-163.
- Mohammed, T. A. A. (2003): Study of growth and reproduction of some corals at Hurghada region with reference to the effect of some pollutants in the area. Ph. D. Thesis. Zool. Dept. F. Science. Suez Canal Uni. 204 P.
- Mohammed, T. A. A. Mohammed, M. A. (2005): Some ecological factors affecting coral reefs assemblages off Hurghada, Red Sea, Egypt. Egypt. J. aquat. Res. Vol. 31 (1): 133 145.
- Mohammed, T. A-A.; Kotb, M. M. A.; Ghobashy, A-F A.; El-Deek, M. S. (2007): Reproduction and growth rate of two scleractinian coral species in the northern Red Sea, Egypt. Journal of Aquatic Research. Vol. 33 (2): 70-86
- Muko, S.; Arakaki, S.; Nagao, M and Sakai, K. (2013). Growth form-dependent response to physical disturbance and thermal stress in Acropora corals. Coral Reefs, 32:269–280
- Ohde, S. and Hossain, M.M.M. (2004). Effect of CaCO₃ (aragonite) saturation state of seawater on calcification of *Porites* coral. Geochemical Journal, 38: 613-621.
- Ohde, S. and van Woesik, R. (1999). Carbon dioxide flux and metabolic processes of a coral reef, Okinawa. *Bull. Mar. Sci.* 65, 559–576.
- Oliver, J. K. (1984). Intra-colony variation in the growth of *Acropora formosa*: extension rates and skeletal structure of white (zooxanthellae free) and brown-tipped branches. Coral Reefs 3:139–147.
- Piniak, G.A. and Brown, E.K. (2008). Growth and mortality of coral transplants (*Pocillopora damicornis*) along a range of sediment influence in Maui, Hawaii. Pacific Science, 62(1):39–55.
- Rodgers, K.; Cox, E. and Newtson, C. (2003). Effects of mechanical fracturing and experimental trampling on Hawaiian corals. Environ. Manag., 31(3): 377-384.
- Scoffin, T. P., Tudhope, A. W., Brown, B. E., Chansang, H., and Cheeney, R. F. (1992). Patterns and possible environmental controls of skeletogenesis of *Porites lutea*, South Thailand, Coral Reefs 11:1-11.
- Silverman, J.; Lazar, B. and Erez, J. (2007). Effect of aragonite saturation, temperature, and nutrients on the community calcification rate of a coral reef, J. Geophys. Res., 112:1-14.
- Smith, L.W.; Barshis, C. and Birkeland, C. (2007). Phenotypic plasticity for skeletal growth, density and calcification of Porites lobata in response to habitat type. Coral Reefs, 26:559-567.
- Tambutté, S.; Holcomb, M.; Ferrier-Pagès, C.; Reynaud, S.; Tambutté, E.; Zoccola, D. and Allemand, D. (2011). Coral biomineralization: From the gene to the environment. Journal of Experimental Marine Biology and Ecology, 408:58-78.
- Tanzil, J.T.I.; Brown, B.E.; Tudhope, A.W. and Dunne, R.P. (2009). Decline in skeletal growth of the coral Porites lutea from the Andaman Sea, South Thailand between 1984 and 2005. Coral Reefs, 28(2): 519-528.
- Tarrant, A. M.; Atkinson, M. J. and Atkinson, S. (2004). Effects of steroidal estrogens on coral growth and reproduction. Mar. Ecol. Prog. Ser., 269: 121-129.
- Tunnicliffe, V. (1983). Caribbean staghorn coral populations: pre-hurricane Men conditions in Discovery Bay, Jamaica. Bull. mar. Sci. 33: 132-151,
- Vago, R.; Dubinsky, Z.; Genin, A. and Kizner, Z. (1997). Growth rates of three symbiotic corals in the Red Sea. Limnol. Oceanogr. By the American Society of Limnology Oceanography, Inc 42(S), 1997, 1814-1819.
- Veron, J. E. N. and Pichon, M. (1976). Scleractinia of eastern Australia Part 1, In: Australian Institute of Marine Science Monograph Series Vol. 1., Australian Government Publishing Service, Canberra,.
- Woodly, D. T., C. N. Rao, J. R. Hassell, L. A. Liotta, G. R. Martin, and H. K. Kleinman. 1983. Interactions of basement membrane components. Bioehim. Biophys. Acta. 761:278-283.
- Wooldridge, S. (2013). A new conceptual model of coral biomineralisation: hypoxia as the physiological driver of skeletal extension. Biogeosciences, 10, 2867–2884.
- Yap, H.T.; Alino, P. M. and Gomez, E. D. (1992). Trends in growth and mortality of three coral species (Anthozoa: Scleractinia), including effects of transplantation. Mar. Ecol. Prog. Ser., 83: 91-101.



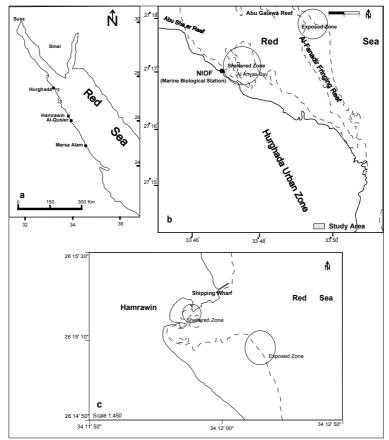


Fig. (1) Location map shows the exposed and sheltered zones at each locality.

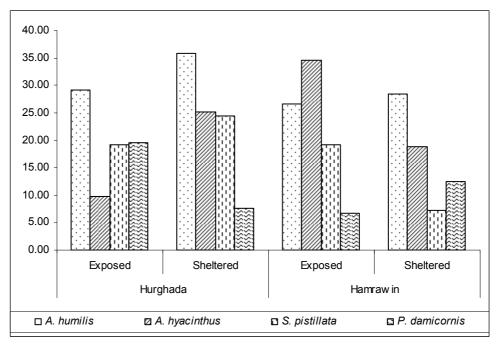


Fig., (2) Growth rate variations between summer and winter seasons of the different species at the different zones.



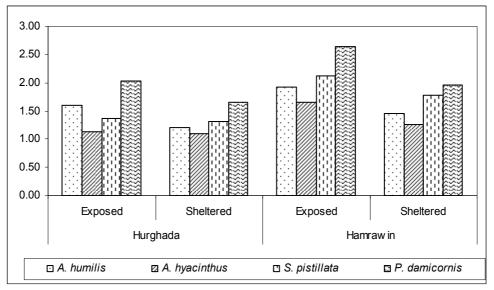


Fig., (3) the differences of skeletal density of the studied species at the different zones.

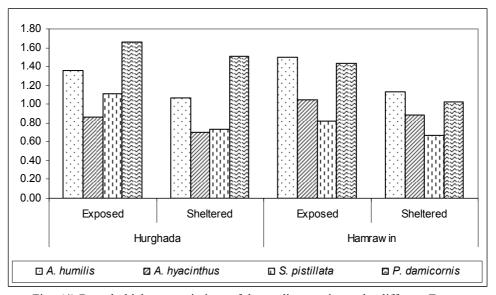


Fig., (4) Branch thickness variations of the studies species at the different Zones.



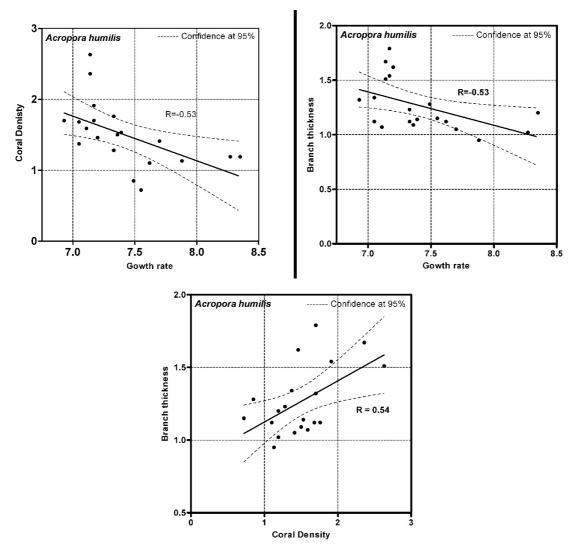


Fig., (5). Linear regression relationships of Growth rate, coral density and branch thickness in A. humilis.

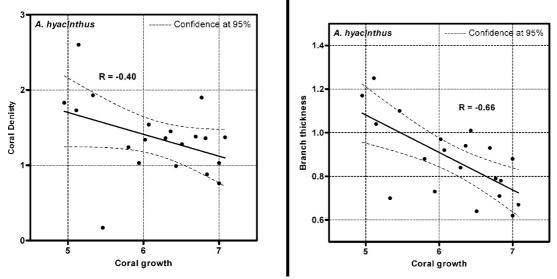


Fig., (6). Linear regression relationships of Growth rate against coral density and branch thickness in A. hyacinthus.



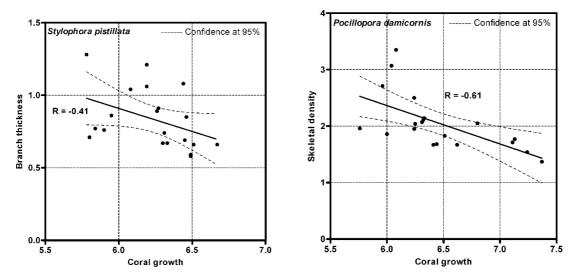


Fig., (7). The linear regression relationships of Growth rate against branch thickness and coral density in *S. pistillata* and *P. damicornis*.

Table (1): The means of some oceanographic parameters in summer and winter at the studied locations:

Locations	Season	Tempe	erature (°C)	p]	Н	Salinity (‰)		
		Max.	Min.	Max.	Min.	Max.	Min.	
Hurghada	Winter	21.94	19.56	8.6	7.98	41.18	39.75	
	Summer	29.38	26.91	8.61	7.83	42.43	41.61	
Hmrawin	Winter	23.57	18.89	8.05	7.7	40.92	40.18	
	Summer	29.03	26.94	8.01	7.88	42.15	41.79	

Table (2). The recorded seasonal and annual growth rates of the studied species at the studied zones of Hurghada and Hamrawin:

	Hurghada						Hamrawin						
	Exposed				Sheltered			Exposed			Sheltered		
		Summer	Winter	annual									
A. humilis	1	0.66	0.53	7.17	0.75	0.53	7.70	0.66	0.53	7.17	0.69	0.53	7.33
	2	0.66	0.53	7.14	0.80	0.59	8.35	0.66	0.53	7.14	0.69	0.54	7.36
	3	0.69	0.54	7.33	0.75	0.52	7.62	0.69	0.52	7.20	0.66	0.53	7.11
	4	0.68	0.50	7.05	0.78	0.60	8.27	0.68	0.50	7.05	0.70	0.53	7.39
	5	0.71	0.55	7.55	0.73	0.58	7.88	0.61	0.54	6.93	0.70	0.55	7.49
	Av.	0.68 ± 0.02	0.53 ± 0.02	7.25±0.02	0.76 ± 0.03	0.56 ± 0.04	7.96 ± 0.33	0.66 ± 0.03	0.52 ± 0.02	7.10 ± 0.11	0.69 ± 0.02	0.54 ± 0.01	7.34±0.14
A. hyacinthus	1	0.60	0.53	6.77	0.66	0.52	7.08	0.50	0.39	5.33	0.53	0.48	6.07
	2	0.61	0.53	6.84	0.62	0.52	6.82	0.50	0.35	5.14	0.54	0.46	6.02
	3	0.54	0.54	6.43	0.66	0.51	7.00	0.52	0.39	5.46	0.54	0.52	6.36
	4	0.57	0.48	6.29	0.58	0.50	6.51	0.50	0.35	5.11	0.55	0.42	5.80
	5	0.57	0.55	6.69	0.67	0.49	7.00	0.46	0.36	4.95	0.55	0.44	5.94
	Av.	0.58 ± 0.03	0.52 ± 0.03	6.60 ± 0.24	0.64 ± 0.04	0.51 ± 0.01	6.88 ± 0.23	0.50 ± 0.02	0.37 ± 0.02	5.20±0.02	0.55 ± 0.01	0.46 ± 0.04	6.04±0.21
	1	0.58	0.46	6.19	0.60	0.45	6.31	0.53	0.47	5.95	0.56	0.53	6.51
	2	0.57	0.50	6.44	0.61	0.50	6.67	0.57	0.49	6.33	0.56	0.52	6.49
S. pistillata	3	0.54	0.50	6.27	0.54	0.51	6.30	0.53	0.44	5.84	0.55	0.52	6.45
S. pisuuaia	4	0.56	0.47	6.19	0.58	0.47	6.26	0.55	0.43	5.90	0.57	0.52	6.49
	5	0.55	0.42	5.78	0.57	0.40	5.80	0.55	0.46	6.08	0.56	0.52	6.46
	Av.	0.56 ± 0.01	0.47 ± 0.04	6.17±0.24	0.58 ± 0.03	0.47 ± 0.04	6.27±0.31	0.55 ± 0.02	0.46 ± 0.02	6.02±0.19	0.56 ± 0.00	0.52 ± 0.00	6.48±0.02
P. damicornis	1	0.59	0.48	6.41	0.61	0.58	7.13	0.52	0.49	6.04	0.56	0.49	6.31
	2	0.57	0.47	6.25	0.62	0.59	7.24	0.53	0.48	6.08	0.56	0.48	6.24
	3	0.56	0.48	6.24	0.61	0.52	6.80	0.51	0.49	6.00	0.57	0.51	6.44
	4	0.56	0.50	6.33	0.59	0.60	7.11	0.51	0.49	5.96	0.57	0.52	6.51
	5	0.61	0.49	6.62	0.65	0.58	7.37	0.49	0.46	5.76	0.56	0.49	6.32
	Av.	0.58 ± 0.02	0.48 ± 0.01	6.37±0.15	0.61 ± 0.02	0.57 ± 0.03	7.13 ± 0.21	0.51 ± 0.01	0.48 ± 0.01	5.97±0.13	0.56 ± 0.00	0.50 ± 0.01	6.36 ± 0.11



Table (2). The recorded coral densities and branch thicknesses of the studied species at the studied zones of Hurghada and Hamrawin:

			Coral	Density		branch thickness					
		Hurghada		Hamrawin		Hurg	ghada	Hamrawin			
		Exposed	Sheltered	Exposed	Sheltered	Exposed	Sheltered	Exposed	Sheltered		
A. humilis	1	1.70	1.41	1.91	1.76	1.79	1.05	1.54	1.12		
	2	2.63	1.19	2.36	1.50	1.51	1.20	1.67	1.09		
	3	1.28	1.10	1.46	1.59	1.23	1.12	1.62	1.07		
	4	1.68	1.19	1.37	1.53	1.12	1.02	1.34	1.14		
	5	0.72	1.13	1.70	0.85	1.15	0.95	1.32	1.28		
	Av.	1.60 ± 0.70	1.20 ± 0.12	1.91±0.40	1.45±0.35	1.36 ± 0.29	1.07±0.10	1.49 ± 0.16	1.14 ± 0.08		
A. hyacinthus	1	1.90	1.37	1.93	1.54	0.79	0.67	0.70	0.92		
	2	0.88	1.36	2.60	1.34	0.78	0.71	1.04	0.97		
	3	0.99	1.03	0.17	1.45	1.01	0.62	1.10	0.94		
	4	1.36	1.28	1.73	1.24	0.84	0.64	1.25	0.88		
	5	1.38	0.76	1.83	1.03	0.93	0.88	1.17	0.73		
	Av.	1.13 ± 0.25	1.09 ± 0.26	1.66 ± 0.90	1.26 ± 0.19	0.87 ± 0.10	0.70 ± 0.10	1.05 ± 0.21	0.89 ± 0.10		
	1	1.96	1.46	2.11	2.20	1.21	0.74	0.86	0.66		
	2	1.33	1.81	2.77	1.52	1.08	0.66	0.67	0.59		
C mintillata	3	1.08	0.87	2.13	2.04	0.91	0.67	0.77	0.69		
S. pistillata	4	1.36	1.20	2.16	1.41	1.06	0.89	0.76	0.58		
	5	1.01	1.58	1.62	1.68	1.28	0.71	1.04	0.85		
	Av.	1.37±0.38	1.31 ± 0.36	2.11 ± 0.41	1.79 ± 0.34	1.11±0.14	0.73 ± 0.09	0.82 ± 0.14	0.67 ± 0.11		
P. damicornis	1	1.67	1.77	3.07	2.07	2.20	1.71	1.43	1.09		
	2	2.04	1.54	3.35	1.95	1.75	1.75	1.33	0.86		
	3	2.50	2.05	1.86	1.68	1.15	1.73	1.77	1.28		
	4	2.14	1.71	2.71	1.83	1.33	1.11	1.05	0.84		
	5	1.67	1.37	1.96	2.11	1.88	1.25	1.63	1.08		
	Av.	2.04 ± 0.35	1.64 ± 0.26	2.64 ± 0.66	1.96 ± 0.18	1.66 ± 0.42	1.51 ± 0.30	1.44 ± 0.28	1.03 ± 0.18		