

Sub Grouping of Lipase Enzymes According to Tween Substrate in Different Organs of Rat

Dr. Mahmoud Mishaal (M.B.Ch.B) Dr. Maan H. Al-Khalisy (M.B.Ch.B, M.Sc, Ph.D)
University of Baghdad, College of medicine, Department of Anatomy

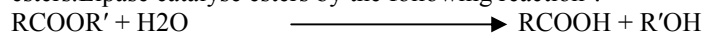
Abstract

Lipase is a general term involved types of lipase enzymes that hydrolyse long chain esters. Yet, no one knows which type of lipase is specific for certain organ or tissue. Therefore the aim of this study is to highlight which type of lipase enzymes subgroup is specific for each organ, using different tween substrates. Norway rats had been used. Different tissues had been taken from these animals. Detection of lipase enzyme, for each tissue, had been done by using Gomori method using different types of tween substrates. Different organs revealed different lipase enzymes activities according to the different substrates used in this experiment. From this study, different organs elicited their own lipase enzymes subgroups since different organs utilized different types of tweens as substrate and in different intensity.

Keywords: Lipase, Gomori method, tween substrates.

1. Introduction

Lipase is a general term that applied to group of enzymes that have the ability to hydrolyse long chain esters. Lipase catalyse esters by the following reaction¹:



In mammals, dietary lipid digestion is commonly assumed to be mediated by three main enzymes^{2,3}:

- Pre-duodenal (lingual or gastric) lipase.
- Pancreatic lipase.
- Post-duodenal lipase.

Types of Lipase

Cholesterol ester lipase: also called pancreatic ester lipase.

- a. Lysophospho-lipase: this enzyme could be isolated from from pancreas, liver, intestine, kidney, and muscles (including skeletal and cardiac).
- b. Choline-esterase: which are distinguished from the other esterases by their ability to hydrolyse esters of choline^{4,5}.
- c. Triacylglycerol lipase: this enzyme is generally referred to as lipase, functioning primarily in the hydrolysis of long-chain fatty acids. This hydrolyses triglycerides to form monoglycerides⁶. This enzyme is present in adipose tissue, liver, kidney, salivary glands, stomach, intestine, brain, heart and muscle⁵.
- d. Lipoprotein Lipase (LPL): also called body lipase. LPL hydrolyses triglycerides in plasma lipoproteins into fatty acids available for energy production, storage, or other metabolic reactions^{6,7}. LPL acts on triglycerides but has no effect on cholesterol⁸.

Lipase enzymes activities could be identified by many methods and as follows⁹:

1. Titrimetry.
2. Spectroscopy (photometry, fluorimetry, infra-red).
3. Chromatography.
4. Radio-activity.
5. Interfacial tensiometry.
6. Turbidimetry.
7. Conductimetry.
8. Immuno-chemistry.
9. Microscopical histo-chemistry.

Histo-chemical method for detection of lipase enzymes is done using Gomori method^{by} using tweens as substrate¹⁰.

There is no previous study to determine which of the tween substrates are suitable to elicited specific lipase enzyme in different mammalian tissues.

Aim of the study: is using different tissues to elicited lipase enzymes activities using different tween substances as a substrate in different tissue, putting in consideration sex and sexual maturity.

2. Material and method

Norway albino rats were used in this experiment. Both sexes including mature and immature rats were used. Forty animals had been used and divided into four groups and as follows:

Table1: number and criteria of rats in experimental groups

Maturity of animals	Age	Sex	Number
mature	7-10 weeks	Male	10
immature	3-4 weeks	Male	10
mature	7-10 weeks	Female	10
immature	3-4 weeks	Female	10

From each animals, different tissues had been obtained,including, skeletal muscle, pancreas, small intestine, testis, liver, kidney, and ovary.

To identify the activity of lipase enzyme, the following histo-chemical method had been used. This method is Gomori method (1952) using different tween substances as a substrates¹⁰.

Here, different types of tween substances had been used i.e. tween 20, 40, 60, and 80.

In this work, tween substances used as substrate. Lipase will hydrolyse tween substances to form fatty acid, which in turn combined with calcium ions (from already prepared calcium chloride solution). This combination will form relatively insoluble calcium soap. This later will be treated with lead ions, then with ammonium sulphide . This will form visible dark brown-black lead sulphide deposite at the site of lipase activity within the tissue¹¹.

The activity of lipoprotein lipase was measured using the method of (+) as shown in the following table⁹:

Table 2: scaling of LPL activity.

<u>Activity of LPL (intensity of reaction)</u>	<u>Presentation in (+)</u>
No LPL activity	-
Very weak LPL activity	+/-
Weak LPL reaction	+
Weak to moderate LPL reaction	+/++
Moderate LPL reaction	++
Moderate to strong LPL reaction	++/+++
Strong LPL reaction	+++

3. Results

The intensity of activity of lipase enzyme in skeletal muscle was elicited as shown in table (3).

Table 3: the intensity of lipase activity in skeletal muscles using different types of tweens.

MATURITY AND SEX	TWEENS			
	80	60	40	20
Mature male	+	+/-	-	-
Premature male	+/-	-	-	-
Mature female	+	-	-	-
Premature female	+/-	-	-	-

The intensity of activity of lipase enzyme in intestine was elicited in table (4).

Table 4: the intensity of lipase activity in intestine using different types of tweens.

MATURITY AND SEX	TWEENS			
	80	60	40	20
Mature male	+++	-	-	-
Premature male	++	-	-	-
Mature female	+++	-	-	-
Premature female	++	-	-	-

The intensity of activity of lipase enzyme in pancreas had been shown in table (5).

Table 5: The intensity of lipase activity in pancreas in different age and sex groups using different tweens.

MATURITY AND SEX	TWEENS			
	80	60	40	20
Mature male	+++	++/+++	+++	++
Premature male	++/+++	++	++	+/++
Mature female	++/+++	++/+++	++/+++	++/+++

The intensity of activity of lipase enzyme in the testis was shown in table (6).

Table 6: the intensity of lipase activity in testis using different types of tweens.

MATURITY AND SEX	TWEENS			
	80	60	40	20
Mature male	++	+	+	-/+
Premature male	-	-	-	-

The intensity of activity of lipase enzyme in ovary had been shown in table (7).

Table 7: the intensity of lipase activity in ovary using different types of tweens

MATURITY AND SEX	TWEENS			
	80	60	40	20
Mature female	+	+	+	-
Premature female	-	-	-	-

The intensity of activity of lipase enzyme in the liver had been shown in table (8).

Table 8: the intensity of lipase activity in the liver using different types of tweens.

MATURITY AND SEX	TWEENS			
	80	60	40	20
Mature male	-	+	+++	-
Premature male	-	-/+	++	-
Mature female	-	-/+	++	-
Premature female	-	-/+	++	-

4. Discussion

Almost all the previous studies involved in detection of lipase enzymes activities, using Gomori method, had been used tween substances as substrate¹¹. Yet, no one mentioned which tween substances were specific substrate for specific tissues. This raises a question that which tween (of the four tweens used as a substrate) is specific for each tissue lipase when Gomori method has been used.

Regarding activity of lipase enzyme in skeletal muscles, this study revealed no enzyme activity in all groups of animal tissues and in all tween substrates used except tween 80 which revealed weak reaction. This result agreed with that of the Oscai et al¹². And Levak –Frank et al¹³.

Lipase activity in intestine elicited strong lipase activity when used tween 80 only, as shown in table (4). Otherwise no reaction could be elicited using other tween substrates. This means that tween 80 is the only tween substrate that could showed activity of lipase enzyme in the intestine. This findings coincides with the findings of Montin¹⁴ and petridou and Mougios¹⁵.

Regarding pancreatic tissues, in both sexes, tween 80 showed strong reaction of lipase enzyme among all tissues lipase. Although among all tween substrates, tween 80 showed the strongest reaction. These results go in the same line with the finding of Winkler et al.¹⁶, Withers-Martinez et al.¹⁷ and Petridou et al.¹⁸. This strong activity is expected since pancreas is the most strongest gland responsible for lipid metabolism^{17,19}.

The gonads, both in male and female, showed mild to moderate activity when lipase enzymes activities had been detected using tween substrates. Yet, tween 80 showed the strongest reaction among other tween substances. However, ovaries revealed lesser lipase activity than testes. These results agreed with findings of Kuwajima et al.³, Soterion and Cryer²⁰, Holst et al.²¹, and Chung et al.²².

The liver had unexpected results. Lipase enzyme activity revealed strong reaction when tween 40 had been used, while tween 80 and 20 showed no reaction. Besides, the female rat's liver tissue showed lesser reaction when tween 40 had been used in comparison with that of male. This difference in activity between male and female could be attributed to the differences in metabolic activities between the two sexes. Besides sex hormones could play important role. However these findings coincide with the findings of Rug et al²³, Schulz et al²⁴, and Elliot and Elliot²⁵.

References

- Stoward P. J. and Pearse A. G. E., 1991a. Esterase. In: Histochemistry, theoretical and applied Enzyme histochemistry, 4th ed., Churchill livingstone, Edinburg, London, Melbourne, New York and Tokyo, Vol.3 : 219-241.
- Tsujita T., Sumiyoshi M., Takaku T., Momsen W.E., Lowe M.E., and Brockman H.L., 2003. Inhibition of lipases by ξ -polylysine. *J. Lipid Res.*, American Society for Biochemistry and Molecular Biology; DOI 10.1194/Jlr.M300151-JLR200.
- Kuwajima M, Foster DW, Mcgarry JD. 1988. Regulation of lipoprotein lipase in different rat tissues. *Metab Clin Exp.*; 37: 597-601.
- Olivecrona G. & T. olivecrona 1995. Triglyceride Lipases and atherosclerosis. *Curr. Opin.Lipidol*; 6:291-305
- Belfrage P., Fredrikson G., Stralfors P., and Tornquist H., 1984. In: Borgstrom B., Brockman H.L. (Eds), lipases. Elsevier, Amsterdam, : 365.
- Olivecrona T., Bengtsson-Olivecrona, G., T., 1987. Mouse preheparin plasma contains high levels of hepatic lipase with low affinity for heparin. *Biochem. Biophys. Acta*; 878: 65-70.
- Goldberg, I. J., 1996. Lipoprotein lipase and lipolysis: central roles in lipoprotein metabolism and atherogenesis. *J. Lipid Res.*; 37: 693-707.
- Montgomery R, Conway T.W., Spector D.A., and chappell D., 1996. Lipoproteins. In: *Biochemistry: A case – Oriented Approche*. 6th ed., Mosby, St. Louis.,: 356-383.
- Beisson F., Tiss A., Riviere C., Verger R., 2000. Methods for lipase detection and assay; a critical review. *Eur. J. Lipid Sci. Technol.*, WILEY-VCH Verlag GmbH, Weinheim; 133-153.
- Gomori G, 1952. Histochemistry of esterase. *International Review of cytology*; : 323.
- Bancroft J., 1982. Enzyme histochemistry. In: Bancroft J. and Stevens A. (Ed.s). *Theory and Practice of histochemical Techniques*. 2nd ed., Churchill livingstone, London, : 392-393.
- Oscai, L.B.; Gorski, J.; Omiler, W.C.; Balmece, W.K., 1988. Role of the alkaline TG lipase in regulating intramuscular TG content. *MEDICINE and SCEINCE in SPORTS and EXERCISE*, Lippincott Williams and Wilkins; 20(6): 539-544.
- Levak-Frank. S., Weinstock P.H., Hayek T., Verdery R., Hofmann W., Ramakrishnan R., Sattler W., Breslow J.L., and Zechner R., 1997. Induced Mutant Mice Expressing Lipoprotein Lipase Exclusively in Muscle Have Subnormal Triglycerides Yet Reduced High Density Lipoprotein Cholestrol Levels in Plasma. *Jbc, the American Society for Biochemistry and Molecular Biology, Inc.*; 272(27): 17182-17190.
- Martin B.F., 1956. The histochemical distribution of 'lipase' in the alimentary tract and associated glands of laboratory animals. *J Anat.*; 90(Pt 3): 440-454.
- Pertidou A. and Mougios V., 2002. Acute changes in triacylglycerol lipase activity of human adipose tissue during exercise. *J Lipid Res.*; 43(8): 133-134.
- Wikkler Fk. D'Arcy A, and W Hunziker, 1990. "Structure of human pancreatic lipase". *Nature* 343(6260): 771-774.
- Withers-Martinez C, Crriere F, verger R, Bourgeois D, and C Cambillau (1996). "Pancreatic lipase with

- phospholipase A1 activity: Crystal structure of achimeric pancreatic lipase-related protein 2 from guinea pig". *Structure* 4(11): 1363-1374.
- Petridou A. and Mougious V., 2002. Acute changes in triacyl-glycerol lipase activity of human adipose tissue during exercise. *J Lipid Res*; 43(8): 1331-1334.
- Shils ME, Shike M and Rose AC. eds, 2005. *Modern nutrition in health and disease*. 10th ed. Philadelphia, Lippincott Williams and Wilkins.
- Soterion A and crier A., 1992. "purification and characterization of lipoprotein lipase from adipose tissue, skeletal muscle, cardiac muscle, mammary gland, ovary and lung tissues of the rat". *Int. J. Biochem.*;25(10): 1483-1490.
- Holst L.S., Langin D., Mulder H., Laurell H., Grober J., Bergh A., Mohrenweiser H.W., Edgren G., and Holm C., 1996. Molecular Cloning, Genomic Organization, and Expression of a testicular Isoform of Hormone-Sensitive Lipase. *Science direct*. 35 (5): 441-447.
- Chung S, Shu Pei Wang, Linge Pan, Grant Mitchell, Jacquetta Trasler and Louis Hermo, 2001. Infertility and Testicular Defects in Hormone-Sensitive Lipase-Deficient Mice. *Endocrinolog*; 142 (10): 4272-4281.
- Ruge T., Bergo M., Huttin M, Olivercrona Q., and Olivercrona T., 2000. Nutritional regulation of binding sites for lipoprotein lipase in rat liver and heart. *AJP-Endocrinology and metabolism*; 278(2): 211-218.
- Schultz C.J., Mackie E.J., and Scow R.O., 2000. Adrenal and liver in normal and cld/cld mice synthesis and secrete hepatic lipase, but the lipase is in active in cld/cld mice. *J. of Lipid Research*; 41: 214-225.
- Elliot W.H., and Elliot D.C., 2003. Biochemical Mechanisms Involved in Food Transport, Storage and Metabolization. In: *Biochemistry and Molecular Biology*. 2nd ed.,: 125.

The IISTE is a pioneer in the Open-Access hosting service and academic event management. The aim of the firm is Accelerating Global Knowledge Sharing.

More information about the firm can be found on the homepage:

<http://www.iiste.org>

CALL FOR JOURNAL PAPERS

There are more than 30 peer-reviewed academic journals hosted under the hosting platform.

Prospective authors of journals can find the submission instruction on the following page: <http://www.iiste.org/journals/> All the journals articles are available online to the readers all over the world without financial, legal, or technical barriers other than those inseparable from gaining access to the internet itself. Paper version of the journals is also available upon request of readers and authors.

MORE RESOURCES

Book publication information: <http://www.iiste.org/book/>

Academic conference: <http://www.iiste.org/conference/upcoming-conferences-call-for-paper/>

IISTE Knowledge Sharing Partners

EBSCO, Index Copernicus, Ulrich's Periodicals Directory, JournalTOCS, PKP Open Archives Harvester, Bielefeld Academic Search Engine, Elektronische Zeitschriftenbibliothek EZB, Open J-Gate, OCLC WorldCat, Universe Digital Library, NewJour, Google Scholar

