Avondale College ResearchOnline@Avondale

Nursing and Health Papers and Journal Articles

School of Nursing

10-8-2018

Statins Do Not Impair Whole-Body Fat Oxidation During Moderate-Intensity Exercise in Dyslipidemic Adults

Maria A. Matuszek University of New South Wales

Ross Grant Australasian Research Institute, ross.grant@sah.org.au

Follow this and additional works at: https://research.avondale.edu.au/nh_papers

Part of the Nursing Commons

Recommended Citation

Matuszek, M. A., & Grant, R. (2018). Statins do not impair whole-body fat oxidation during moderateintensity exercise in dyslipidemic adults. *Exercise Medicine*, *2*:12. doi:10.26644/em.2018.012

This Article is brought to you for free and open access by the School of Nursing at ResearchOnline@Avondale. It has been accepted for inclusion in Nursing and Health Papers and Journal Articles by an authorized administrator of ResearchOnline@Avondale. For more information, please contact alicia.starr@avondale.edu.au.

Exercise Medicine

ONLINE MANUSCRIPT SUBMISSION

Title:	Statins do not impair fat metabolism during moderate-intensity exercise in dyslipidemic adults			
Type of Manuscript:	Research Article			
Running Title:	Statins, fat metabolism and exercise			

Abstract

Objective: Some lipid-lowering agents, for example, nicotinic acid and fibrates, decrease an individual's ability to metabolise fat during exercise. However, it is unclear whether statins affect fat metabolism during exercise in patients. This study investigated whether fatty acid oxidation is impaired in a dyslipidemic population, while walking at a moderate intensity. Methods: Patients (n=16), walked for 45 minutes on a treadmill at 50% of their estimated $\dot{V}O2max$, in the absence and presence of their prescribed statin. Fat metabolism was investigated by examining respiratory data, and circulating plasma glycerol and free fatty acids. Results: Analysis of respiratory data indicated a progressive increase in fat oxidation over time, along with a decrease in carbohydrate oxidation, for all patients during exercise, in both the absence and presence of a statin (P \leq 0.05). The increase in the percent of energy derived from fat was further supported by the observation of a significantly progressive increase in circulating glycerol and free fatty acids respiratory increase in circulating glycerol and free fatty acids during the exercise period. However no significant difference in the extent of change was observed when comparing the respiratory and biochemical response to physical activity in the absence and presence of the prescribed statin. Conclusions: There is no evidence of a negative impact of statins on the ability to metabolise fat as a fuel for moderate-intensity aerobic exercise. Given the importance of physical activity, this result encourages patients to participate in regular exercise without concern of premature fatigue.

Editorial members *Exercise Medicine* Editorial Office Department of Preventive Medicine, College of Medicine, Gachon University 155 Gaetbeol-ro, Yeonsu-gu, Incheon 21999, Korea TEL: +82-32-899-6433 FAX: +82-504-372-0664 E-mail : exercmed@gmail.com Website: http://submit.exercmed.org/

> Department of Preventive Medicine, College of Medicine, Gachon University 155 Gaetbeol-ro, Yeonsu-gu, Incheon 21999, Korea

> > Copyright© Sapientia Publishing Group. All rights reserved.

1	Statins do not impair	fat metabolism during	g moderate-intensity	v exercise in dy	slipidemic adults.

- Hercise Medicink Running title: Statins, fat metabolism and exercise.

2425 Abstract

26	Objective: Some lipid-lowering agents, for example, nicotinic acid and fibrates, decrease an individual's
27	ability to metabolise fat during exercise. However, it is unclear whether statins affect fat metabolism during
28	exercise in patients. This study investigated whether fatty acid oxidation is impaired in a dyslipidemic
29	population, while walking at a moderate intensity.
30	Methods: Patients (n=16), walked for 45 minutes on a treadmill at 50% of their estimated $\dot{V}O_{2max}$, in
31	the absence and presence of their prescribed statin. Fat metabolism was investigated by examining
32	respiratory data, and circulating plasma glycerol and free fatty acids.
33	Results: Analysis of respiratory data indicated a progressive increase in fat oxidation over time, along
34	with a decrease in carbohydrate oxidation, for all patients during exercise, in both the absence and
35	presence of a statin (P \leq 0.05). The increase in the percent of energy derived from fat was further
36	supported by the observation of a significantly progressive increase in circulating glycerol and free fatty
37	acids during the exercise period. However no significant difference in the extent of change was observed
38	when comparing the respiratory and biochemical response to physical activity in the absence and
39	presence of the prescribed statin.
40	Conclusions: There is no evidence of a negative impact of statins on the ability to metabolise fat as a
41	fuel for moderate-intensity aerobic exercise. Given the importance of physical activity, this result
42	encourages patients to participate in regular exercise without concern of premature fatigue.
10	

- 45 Key Words: adipose tissue, aerobic exercise, cholesterol-lowering drugs, fat oxidation, physical activity

Introduction 47

atorvastatin and 34 for rosuvastatin [2].

48 Lipid-lowering drugs are amongst the most commonly subsidized prescribed drugs world-wide, 49 including Europe and the United States (USA). In 2016, atorvastatin and simvastatin were ranked in 50 positions 1 and 7 in the USA [1]. In the same year, in Australia, atorvastatin and rosuvastatin were in 51 positions 1 and 3, with the number of Australians per thousand taking the drug daily being 53 for 52

53 Physiologically, these drugs inhibit liver enzyme HMG-CoA reductase, blocking cholesterol synthesis. They are recognized as effective in the secondary prevention of cardiovascular disease and 54 55 all-cause mortality [3]. However, it is now well established that a healthy diet and regular physical 56 activity are valuable contributors to maintaining healthy blood lipids, with exercise improving cardiovascular fitness; reducing all-cause mortality and the risk of coronary heart and cardiovascular 57 disease, stroke, type 2 diabetes mellitus (T2DM), and obesity. A recent meta-analysis highlighted that 58 59 physical activity alone is potentially as effective as many drug interventions on mortality outcomes in coronary heart disease, stroke, heart failure, and prediabetes [4]. Despite this, there exists a bias against 60 61 exercise intervention with a subsequent progression towards lowering the threshold for pharmaceutical 62 treatment, and an increased likelihood that individuals embarking on an exercise regime are already taking a cholesterol-lowering agent as a secondary or even primary prevention against cardiovascular 63 risk. To maximize the benefit of pharmaceutical intervention and exercise, it is important therefore, to 64 65 understand the effect of these drugs on the body's response to exercise.

66 Lipid stores, critical for fueling prolonged exercise, exist as triglycerides in adipose tissue and 67 muscle, and also as circulating triglycerides. Hormone-sensitive lipase (HSL), an intracellular enzyme in 68 adipose tissue, and lipoprotein lipase (LPL), an enzyme which hydrolyzes the triglyceride core of 69 circulating chylomicrons, both liberate free fatty acids (FFA) for use by exercising muscle. Any drugs 70 acting at these sites, could potentially alter FFA availability, resulting in increasing dependence on

muscle glycogen stores and earlier fatigue [5]. Studies in healthy volunteers undertaking exercise have
found that the lipid-lowering drugs, acipimox, and fibrates such as bezafibrate and gemfibrozil, have a
negative impact on fat oxidation [5,6]. This increases demand for carbohydrate (CHO), and is thought to
contribute to a reduction in exercise tolerance. However, no negative impact of simvastatin, fluvastatin
or atorvastatin has been observed in healthy exercising participants [5,6,7].

76 In contrast, the impact of stating on fat metabolism in patient cohorts is less clear. The available 77 research in patients undergoing exercise has produced conflicting results, with studies confounded by 78 the co-administration of a kaleidoscope of other cholesterol-lowering agents, or with β -blockers, which 79 inhibit the release of FFA from adipose tissue and complicate the interpretation of the data [8,9]. One study in patients has reported reduced fat oxidation with atorvastatin, during lower-intensity exercise 80 [10]. Patients have also self-reported increased fatigue with exertion after taking simvastatin for a period 81 82 of 6 months [11]. In contrast, other studies have reported no negative impact of statins on fat metabolism 83 [12,13].

Given the importance of exercise in these patients, and the limited and conflicting data on the interaction between cholesterol-lowering agents and physical activity, the current study investigated in a dyslipidemic population, whether statins impair fatty acid oxidation during exercise of moderateintensity. This study is unique as it appears to be the first longitudinal investigation of fat metabolism in both the absence and presence of a statin during exercise, within the same group of patients.

- 89
- 90
- 91
- 92

- 93 Method
- 94

95 **Participants**

96 The study engaged 16 Caucasian adults taking a statin for hypercholesterolemia. The study was 97 approved by the Human Research Ethics Committees from The University of New South Wales (UNSW) 98 (HREC05310) and the Sydney Adventist Hospital (HREC12/06) and all procedures were in accordance 99 with institutional guidelines. Participants were recruited by newspaper advertisements, and required 100 written permission from their medical practitioner to participate. Individuals also provided written 101 informed consent, and were aware of their right to withdraw at any time. Participants were considered 102 able to engage in physical activity comprising of a brisk walk, by satisfactorily completing a physical 103 activity readiness questionnaire (PAR-Q), as well as an additional questionnaire on family history, 104 medical history, present symptoms, and current pattern of physical activity [14,15]. Patients with T2DM 105 were excluded as this condition may influence fat metabolism. In addition, recruited participants were 106 not taking other medications which could knowingly interfere with fat metabolism.

107

108 **Experimental design**

Each participant made a total of three visits to the laboratory. All participants ceased lipid-lowering medication for a minimum of three weeks prior to Visit 1. The purpose of Visit 1 was to (a) prescreen the participant, (b) perform a submaximal exercise test on a treadmill to predict the participant's maximum oxygen uptake ($\dot{V}O_{2max}$), and (c) calculate the individual treadmill settings (speed and gradient) which would enable the participant to walk at a power output of 50% $\dot{V}O_{2max}$ in Visits 2 and 3. The latter were identical to each other with the notable exception that Visit 2 continued in the absence of the drug and Visit 3 in the presence of the drug. Visit 3 occurred after the participant had resumed their lipid-lowering

116 medication for a minimum of 3 weeks. Statin administration was verified verbally and by assessment of

117 plasma lipids measured at the end of the drug versus no drug phases, on the mornings of Visits 2 and 3.

118

119 Visit 1: Prescreening and submaximal exercise test

Resting heart rate (HR), blood pressure, anthropometric assessment (height, weight, body mass index
(BMI), waist and hip circumference), and percent body fat by bioelectrical impedance (Tanita BWB-800,
Tanita Corporation, Japan) and by skinfolds analysis (biceps, tripceps, subscapular, suprailiac and
abdominal) was measured [16].

Participants performed a submaximal exercise test on a treadmill to predict $\dot{V}O_{2max}$ and to 124 125 calculate the individual treadmill setting for each participant to exercise at 50% of their maximal capability in Visits 2 and 3. Ventilation (VE, 1.min⁻¹), oxygen uptake (VO₂, 1.min⁻¹), expired carbon 126 dioxide ($\dot{V}CO_2$, 1.min⁻¹) and respiratory exchange ratio (RER) ($\dot{V}CO_2/\dot{V}O_2$) were measured using a 127 metabolic cart (ParvoMedics TrueMax 2400). HR was monitored via an electrocardiogram (ECG) and a 128 129 HR monitor. Participants chose a comfortable walking speed at 0% gradient. With the speed constant, exercise intensity was increased a further two times, by increasing the gradient of the treadmill belt 130 every 3-5 min after a stable exercise HR was recorded. The predicted maximum oxygen consumption 131 132 was calculated by plotting exercise HR against oxygen consumption, with a line of best fit extrapolated 133 to the point of each participant's maximal predicted HR [15]. The relative perceived exertion (RPE) [15] 134 and finger prick blood lactate, confirmed participants exercised at a submaximal level.

135

136 Visits 2 and 3: Exercise at 50% $\dot{V}O_{2max}$ in the absence and presence of a statin

137 The exercise trials occurred at Visit 2 (-statin) and Visit 3 (+statin). Visit 3 occurred after participants

138 had resumed medication for a minimum of three weeks. The two visits were otherwise identical.

139 Participants arrived at the laboratory after an overnight fast (water permitted). A forearm vein was

140 cannulated using a 20 gauge indwelling line (BD Insyte Autoguard, Becton Dickinson, UK). As the act

141 of venepuncture can increase some hormone levels by more than 50% [17], a stabilization period of 30 142 min was allowed after the cannula was inserted into the forearm vein, and before collection of a fasting 143 blood sample (6ml) into vacutainer tubes (Becton Dickinson, UK) containing ethylenediaminetetra-144 acetic acid (EDTA). Fasting blood lipids, lactate and glucose were measured immediately. Participants 145 then consumed a low fat 1MJ CHO meal, consisting of Special K cereal and low fat dairy or lactose free 146 milk, followed by rest in a semi-inclined position. Participants commenced exercise 90 min after their 147 meal. They walked on the treadmill at an intensity of 50% of their calculated $\dot{V}O_{2max}$ (determined in 148 Visit 1), corresponding to a 'brisk walk' for 45 minutes. The treadmill speed for each participant was 149 that of Visit 1, and the treadmill gradient was determined using the formula: $\dot{VO}_2 = (0.1 \text{ x S}) + (1.8 \text{ x S x})$ G)+3.5; where $\dot{V}O_2$ is gross oxygen consumption mL.kg⁻¹.min⁻¹ at 50% of estimated $\dot{V}O_{2max}$, speed (S) 150 is in metres.min⁻¹ and G is percent grade expressed as a fraction [15]. The predetermined treadmill 151 152 settings (speed and gradient) for each individual participant remained constant throughout the exercise session. VE, VO₂, VCO₂, and RER were measured to determine the proportion of CHO and fat 153 154 metabolism during exercise. Calculation of carbohydrate and fat oxidation was by the classical 155 stoichiometric equations of indirect calorimetry [18]; with carbohydrate oxidation (mg/min) 156 =4.58 $\dot{V}CO_2$ -3.2 $\dot{V}O_2$; and fat oxidation (mg/min) =1.7 $\dot{V}O_2$ -1.7 $\dot{V}CO_2$. HR and RPE were monitored. Additional blood (each 6ml) was collected into EDTA-containing tubes immediately before exercise, 157 158 and at time 15, 30 and 45 minutes of exercise, for the immediate measurement of glucose and lactate. 159 Remaining blood samples were centrifuged (4 °C, 1560g, 10min, Heraeus Megafuge 1.0R, Hanau, 160 Germany) and plasma was stored at -86 °C prior to the analysis of glycerol and FFA. 161

Biochemistry 162

163 Lipids (total cholesterol (TC), triglycerides (TG), low density lipoprotein-cholesterol (LDL-C) and high 164 density lipoprotein-cholesterol (HDL-C)) (Cholestech LDX, CA, USA), glucose (HemoCue Glucose

- 165 201, Sweden), and lactate (Lactate Pro, ARKRAY, Inc., Japan) were measured in whole blood by
- 166 reflectance photometry. Glycerol was measured in plasma using two colorimetric assays for comparison
- 167 (Sigma, USA; and Cayman Chemical, USA) and the concentration determined at 550nm using an Expert
- 168 Plus Microplate reader (Asys Hitech, Austria) and a Versa Max Microplate reader (Molecular Devices,
- 169 USA). FFA were measured in plasma by colorimetric assay (Cell Biolabs, USA) at 570nm using the
- 170 Versa Max Microplate reader.
- 171

172 Food diary

- 173 Participants completed a 3-day food diary, to calculate total caloric intake, and percentages of
- 174 macronutrient and micronutrient intake during two non-consecutive weekdays and one weekend day.
- 175 Data were entered into a nutritional database (Serve Nutrition Systems, Australia).
- 176

177 Statistical Analysis

178 Power calculations [19], based on a previous study [5] where participants walked at 50% \dot{VO}_{2max} in the 179 presence or absence of a lipid-lowering agent, acipimox, determined that 11 participants would be 180 sufficient to detect a significant decrease in plasma glycerol, assuming a 0.05 significance level and 80% power. The current study recruited 16 participants. All data are expressed as mean ±standard error of the 181 182 mean (SEM). The Student's paired *t*-test compared fasting blood lipids ±statin. A two-way repeated 183 measures ANOVA examined differences between the two within-subject factors (treatment and time) for 184 the physiological variables (HR, RPE, fat and CHO oxidation, % energy derived from fat and CHO, RER, $\dot{V}O_2$, glucose, lactate, glycerol, and FFA), measured during exercise. Where a significant two-way 185 186 interaction was found (FFA), simple main effects examining differences between trials at each level of 187 time and treatment were determined using the General Linear Model (GLM) Repeated Measures 188 procedure. For cases without a significant two-way interaction, the main effect for treatment and time

- 189 were determined. Variables which did not meet Mauchly's test of sphericity were interpreted using a
- 190 Greenhouse-Geisser correction. For all data comparisons, unless indicated, significance is at P ≤ 0.05 . In
- 191 using the GLM Repeated measures procedure for the FFA data, a Bonferroni adjustment was applied for
- 192 the testing of multiple main effects, with a significance of $p \le .05/3 = p \le .016$. Analysis was conducted
- 193 using SPSS (SPSS Inc.,USA).
- 194
- 195

exercise Medicin

196 **Results**

197 Patient characteristics

198 The study engaged 13 males and 3 females, aged 39-82 years, taking a statin for hypercholesterolemia. 199 In addition to hyperlipidemia, some patients had cardiovascular conditions (hypertension, coronary 200 artery disease, heart murmur, aortic valve replacement, mild myocardial regurgitation, varicose veins), 201 and arthritis. Two participants, with the permission of their medical practitioner, ceased taking 202 metoprolol for the entire duration of the study as it interferes with fat metabolism [8]. 203 Patients were non-smokers or previous smokers who had ceased smoking 15-35 years earlier. 204 Body composition data revealed that participants were on average overweight, had an increased disease 205 risk relative to weight and waist circumference, and a moderate disease risk based on waist-hip ratio [15]. Regarding physical activity, participants reported an average occupational activity intensity of 206 207 1.5±0.2 (1=sedentary; 5=heavy labour), sport and leisure physical activity intensity of 6±0.4 (1=very 208 light, 10=intense breathless), and exercise duration of 8 ± 1 hours per week. Activities included cycling, swimming, board paddling, running, walking, dance, stair climbing, gym (circuit, weight training, 209 210 aerobic classes), tennis, rollerblading, and martial arts. Aerobic fitness, indicated by a predicted VO_{2max}, was categorized as 'average' for males (35±3 ml.kg⁻¹.min⁻¹) and 'fair' for females (23±0.5 ml.kg⁻¹.min⁻¹) 211 212 [15]. The average percent daily consumption of protein and CHO was slightly higher, and lower, than 213 the recommended 10-15% and 50-60%, respectively. The average percent daily consumption of <30%214 total fat was as recommended [20]. (Table 2)

- 215
- 216

217 Lipid-lowering agents and blood lipoproteins

218 Of the 16 patients, 11 were taking atorvastatin, with a daily dose range of 10-80mg (1=10mg, 6=20mg,

219 3=40mg, 1=80mg). Rosuvastatin, at 10 mg/day was prescribed for 1 patient, and the remaining 4

patients were administering simvastatin at a dose range of 10-40mg/day (1=10mg, 1= 20mg, 2=40mg). Patients had been taking their drug for 3.5 ± 0.6 years (range 0-10 years). There was one newly diagnosed individual who had not yet commenced his statin medication. All patients ceased taking their statin for 3.4 ± 0.2 weeks prior to their blood lipids analysis in Visit 2. Following this visit, patients recommenced taking their medication for 26 ± 0.7 days. A significant decrease was observed in TC, LDL-C and TC/HDL-C when taking a statin, compared with in the absence of the drug (Table 3). Though not significant, a decrease in TG was also observed (*p*=.073).

227

228 Energy expenditure and metabolism during exercise

All participants completed the exercise tests. Exercise performed at 50% of $\dot{V}O_{2max}$ for 45 minutes required a total energy expenditure of 268±20 kcal or 282±21 kcal in the absence or presence of a statin, respectively.

232 No two-way interaction (between treatment and time) was found for fat or CHO oxidation, % energy derived from fat or CHO, RER, VO₂, glycerol, glucose, lactate, HR, and RPE (Table 4). There 233 234 was a two-way interaction between treatment and time for FFA (F(1.171, 12.879)=5.366, p=.033, 235 epsilon=.58) at 45 minutes (p=.024). However, this effect was disregarded following correction for the testing of multiple main effects, with a Bonferroni adjustment, (p < .016), resulting in non-significance. 236 237 As there was no 2x3 interaction, the main effect for treatment (irrespective of time point) and the 238 main effect for time (irrespective of treatment) were examined. Regarding treatment, a difference 239 between the conditions (\pm statin) was found for HR and \dot{VO}_2 ($p \le 0.05$), with these variables lower in the 240 presence of the statin. The main effect of time was significant for all variables ($p \le 0.05$).

241

242 **Discussion**

243 The current study has shown no evidence of an adverse effect of statins in dyslipidemic patients on the 244 ability to use fat as a metabolic fuel during a moderate-intensity 45-minute walk. The results in this 245 patient group are in agreement with previous research in healthy volunteers [5], where simvastatin 246 treatment had no impact on fat oxidation or circulating plasma free fatty acids, glycerol and glucose 247 during walking. This result is encouraging for patients, given that the importance of regular physical 248 activity is recognized as a positive contributor to improving and maintaining health [4]. Indeed, this 249 study supports positive messaging to patients taking statins; that they may engage in regular exercise. 250 This does to some extent counteract the recently suggested association of stating with an increased risk 251 of T2DM [21]. Furthermore, implementation of a healthy diet and regular aerobic exercise, while on 252 statin medication, may for some patients, enhance the possibility of a medically managed reduction in 253 their statin dose or even complete cessation of this medication. The current study provides reassurance 254 for both patients and their advising physicians, that it is possible to engage effectively in moderateintensity exercise without the prescribed statin compromising the ability to use fat stores as a fuel to 255 256 carry out the exercise session.

257 The lack of an adverse effect on fatty acid mobilization by statins may be due to the fact that 258 their action is thought to be largely confined to the liver [22], rather than adipose tissue. Preliminary 259 research by our group on ezetimibe (Ezetrol), a non-statin whose action is confined to inhibiting 260 cholesterol absorption from the intestine [23], has also shown no effect of this drug on fat metabolism 261 during 45 minutes of moderate-intensity walking (data not published). In contrast, acipimox and fibrates, 262 significantly inhibit HSL in adipose tissue, which results in inhibiting the release of FFA during exercise 263 from this site [5,6]. Fibrates may also reduce availability of FFA to skeletal muscle, by increasing the 264 uptake of FFA into liver with the induction of a fatty acid transporter protein [24].

The findings from this study, therefore make a valuable contribution to the somewhat confusing information on statins and metabolism. It has been suggested that statins, by lowering cholesterol, may affect mouse membrane calveolae, important in fatty acid transport [25]. Statins have been implicated in impairing mitochondrial function, evidenced by increased lactate production, elevated intramuscular lipid stores, decreased mitochondrial activity (measured by a decrease in cytochrome C activity) and lower CoQ10 levels [26,27]. However, despite the above, it has been found that a decrease in circulating CoQ10 does not result in a reduction in muscle CoQ10 [28]. Furthermore, reviews to date, have not concluded that CoO10 is required when taking statins [29,30].

273 Other conflicting results include reports in humans of increased resting RER with statins. 274 indicating a possible shift from fat to CHO metabolism [10,31]. A 12-week aerobic training study 275 measured a greater increase in \dot{VO}_{2max} in a -statin, compared with +statin group. The authors reported a 276 decline in muscle citrate enzyme activity, suggesting that statins may affect mitochondria [32]. However, in contrast to the above, there are reports of simvastatin therapy in patients having no effect on RER or 277 278 aerobic fitness ($\dot{V}O_{2max}$) suggesting no effect on fat metabolism and aerobic capacity [30]. Additionally, 279 a 30% increase in $\dot{V}O_{2max}$, indicating an increase in aerobic fitness, has been reported in patients on 280 rosuvastatin undergoing exercise training for 10 weeks [13].

281 The conflicting results in the literature, are due to many variables, including that of studying 282 different patient groups in either the presence or absence of the drug. The current study eliminated this 283 variable, adopting a longitudinal design, with all participants required to undertake exercise with and 284 without a statin. Based on previous research in healthy volunteers [5], the current study adopted a 3-285 week treatment period verified by assessment of plasma lipids at the end of the drug versus no drug 286 phases. The treatment period significantly lowered TC and LDL-C, confirming that it was of sufficient 287 length to obtain the desired effect of lowering circulating lipids, enabling patients to serve as suitable 288 controls.

To minimize disruption to patients, the current study did not adopt a cross-over design, with all patients experiencing the prescreening session and the first 45-minute walk in the absence of the drug,

and the second 45-minute walk in the presence of the drug. Additionally, researchers did not interfere with each individual patient's statin type and therapeutic dosage, to keep with their real-life situation, controlled by their physician. This, along with the age range of the patients, was not considered a limitation, as the strength of this study is in having each patient serve as their own effective control.

This study also concentrated on the effect of statins without the confounding variable of decreased blood glucose during exercise. The mean energy expenditure during exercise was 1.1MJ (282 kcal), well below the CHO energy reserve of 8-11MJ for a well-nourished adult [5,33]. Additionally, participants consumed a low fat 1MJ CHO meal 90 minutes prior to exercise. Blood glucose levels remained stable during both trials. The decreasing contribution of CHO and increasing contribution of fat as a fuel during the walk was similar in the ±statin exercise sessions. At 45 minutes of walking, fat oxidation peaked at an average of 0.28g/min.

The chosen modality and intensity of exercise, with a duration of 45 minutes, was adopted as it has been reported as manageable for patients and ideal for maximizing cardiovascular health benefits, significantly reducing morbidity and mortality [5,34]. Though it appears that statins do not impact fat metabolism when exercising under these conditions, future studies examining different exercise modalities, intensities and duration, are recommended.

There are many people world-wide on statins. Amongst this group are individuals who are considered capable of adopting a healthier lifestyle incorporating regular exercise. This study demonstrates that statins do not compromise fat metabolism during moderate-intensity aerobic exercise, and therefore do not contribute to premature fatigue and an inability of the patient to complete the exercise. More emphasis may therefore be placed on encouraging regular physical activity in patients to maximize the benefits of concomitant therapies such as pharmaceutical intervention.

313

314 Acknowledgements

315	The authors thank Associate Professor Stephen Boutcher from UNSW, for use of the metabolic cart, and
316	for his advice and encouragement.
317	
318	Funding
319	This study was funded by a grant (051101) from the Australasian Research Institute (ARI), at the
320	Sydney Adventist Hospital; and by a Career Advancement Fund awarded to Maria Matuszek from the
321	University of New South Wales.
322	
323	Conflict of Interest
324	The authors declare no conflict of interest exists with respect to research, authorship and publication.
325	
326	Author contribution
327	MM and RG contributed to the conception of the study. MM designed the study, acquired, analysed and
328	interpreted the data, and drafted the manuscript. MM and RG critically revised the manuscript. Both
329	gave final approval and agree to be accountable for all aspects of work ensuring integrity and accuracy.
330	
331	
332	
333	

334 **References**

335 336 1. LowestMed 2016. Top 50 Prescription drugs filled in the US in Q1 of 2016. [Updated 16 June, 2016]. 337 https://lowestmed.com/top-50-prescription-drugs-filled. (Accessed 15 February, 2017) 338 339 2. NPS MedicineWise (2016) Top 10 Drugs 2015-16. Aust Prescr 2016;39:2205. 340 341 3. Naci H, Brugts JJ, Fleurence R, Tsoi B, Toor H, Ades AE. Comparative benefits of statins in the 342 primary and secondary prevention of major coronary events and all-cause mortality: a network meta-343 analysis of placebo-controlled and active-comparator trials. Eur J Prev Cardiol 2013;20:641-57. 344 345 4. Naci H, Ioannidis JPA. Comparative effectiveness of exercise and drug interventions on mortality 346 outcomes: metaepidemiological study. BMJ 2013;347:f5577. 347 348 5. Head A, Jakeman PM, Kendall MJ, Cramb R, Maxwell S. The impact of a short course of three lipid 349 lowering drugs on fat oxidation during exercise in healthy volunteers. Postgrad Med J 1993;69:197-203. 350 351 6. Eagles CJ, Kendall MJ, Maxwell S. A comparison of the effects of fluvastatin and bezafibrate on 352 exercise metabolism: a placebo-controlled study in healthy normolipidaemic subjects. Br J Clin 353 Pharmacol 1996;41:381-87. 354 355 7. Chung J, Brass EP, Ulrich RG, Hiatt WR. Effect of Atorvastatin on energy expenditure and skeletal 356 muscle oxidative metabolism at rest and during exercise. Clin Pharmacol Ther 2008;83:243-50. 357 8. Eagles CJ, Kendall MJ. The effects of combined treatment with β_1 -selective receptor antagonists and 358 359 lipid-lowering drugs on fat metabolism and measures of fatigue during moderate intensity exercise: a 360 placebo-controlled study in healthy subjects. Br J Clin Pharmacol 1997;43:291-300. 361 9. Fisher NM, Meksawan K, Limprasertkul A, Isackson PJ, Pendergast DR, Vladutiu GD. Statin therapy 362 363 depresses total body fat oxidation in the absence of genetic limitations to fat oxidation. J Inherit Metab 364 Dis 2007;30:388-99. 365 10. Limprasertkul A, Fisher NM, Awad AB, Pendergast DR. Statin therapy depresses fat metabolism in 366 367 older individuals. J Am Coll Nutr 2012;31:32-8. 368 369 11. Golomb BA, Evans MA, Dimsdale JE, White HL. Effects of statins on energy and fatigue with 370 exertion: results from a randomized controlled trial. Arch Intern Med 2012;172:1180-82. 371 372 12. Parker BA, Thompson PD. Effect of statins on skeletal muscle: Exercise, myopathy, and muscle 373 outcomes. Exerc Sport Sci Rev 2012;40:188-94. 374 375 13. Coen PM, Flynn MG, Markofski MM, Pence BD, Hannemann RE. Adding exercise training to 376 rosuvastatin treatment: influence on serum lipids and biomarkers of muscle and liver damage. Metab 377 Clin Exp 2009;58:1030-38. 378 379 14. British Columbia Ministry of Health. Department of National Health and Welfare, Canada. Physical 380 Activity Readiness Questionnaire – PAR-Q. Revised 2002, Canadian Society for Exercise Physiology. 381 Available from: http://www.csep.ca/forms

383 15. Dwyer GB, Davis SE, editors. ACSM's Health-Related Physical Fitness Assessment Manual. 384 Phildelphia: Lippincott Williams and Wilkins. 2005, p 43-171. 385 386 16. Durnin JVGA, Womersley J. Body fat assessment from total body density and its estimation from skinfold thickness: measurements on 481 men and women aged from 16-72 years. Br J Nutr 387 388 1974;32:77-97. 389 390 17. Dimsdale JE, Ziegler MG. What do plasma and urinary measures of catecholamines tell us about 391 human response to stressors? *Circulation* 1991;83(Suppl. II):II-36-II-42. 392 393 18. Frayn KN. Calculation of substrate oxidation rates in vivo from gaseous exchange. J Appl Physiol 394 1983;55:628-34. 395 396 19. Faul F, Erdfelder E, Lang A-G, Buchner A. G*Power 3. A flexible statistical power analysis 397 program for the social, behavioral, and biomedical sciences. Behav Res Methods 2007;39:175-91. 398 20. Smolin LA, Grosvenor MB. Nutrition. Science and Applications. 3rd ed. Orlando: Harcourt College 399 400 Publishing. 2000, p 92-197. 401 402 21. Crandall JP, Mather K, Rajpathak SN, Golberg RB, Watson K, Foo S, et al. Satin use and risk of 403 developing diabetes:results from the Diabetes Prevention Program. BMJ Open Diab Res Care 404 2017;5:e000438doi:10.1136/bmjdrc-2017-000438 405 22. Reents S. Antilipemic agents. In: Sport and Exercise Pharmacology. Champaign, IL: Human 406 407 Kinetics. 2000, p 217-34. 408 23. Kreisberg RA, Oberman A. Medical management of Hyperlipidemia/Dyslipidemia. J Clin 409 410 Endocrinol Metab 2003;88:2445-61. 411 412 24. Staels B, Dallongeville J, Auwerx J, Schoonjans K, Leitersdorf E, Fruchart J-C. Mechanism of 413 action of fibrates on lipid and lipoprotein metabolism. Circulation 1998;98:2088-93. 414 415 25. Khan T, Hamilton MP, Mundy DI, Chua SC, Scherer PE. Impact of Simvastatin on adipose tissue: 416 pleiotropic effects in vivo. Endocrinology 2009;150:5262-72. 417 418 26. Apostolopoulou M, Corsini A, Roden M. The role of mitochondria in statin-induced myopathy. Eur 419 J Clin Invest 2015;45:745-54. 420 421 27. Golomb BA, Evans MA. Statin Adverse Effects: A review of the literature and evidence for a 422 mitochondrial mechanism. Am J Cardiovasc Drugs 2008;8:373-418. 423 424 28. Laaksonen R, Jokelainen K, Laakso J, Sahi J, Härkönen M, Tikkanen MJ, et al. The effect of 425 simvastatin treatment on natural antioxidants in low-density lipoproteins and high-energy phosphates 426 and ubiquinone in skeletal muscle. Am J Cardiol 1996;77:851-54. 427 428 29. Marcoff L, Thompson PD. The role of coenzyme Q10 in statin-associated myopathy: a systematic 429 review. J Am Coll Cardiol 2007;49:2231-37.

- 30. Banach M, Serban C, Sahebkar A, Ursoniu S, Rysz, J, Muntner P, et al. Effects of coenzyme Q10 on statin-induced myopathy: a meta-analysis of randomized controlled trials. Lipid and Blood Pressure meta-analysis collaboration group. Mayo Clin Proc 2015;90:24-34.
- 31. Paolisso G, Barbagallo M, Petrella G, Ragno E, Barbieri M, Giordano M, et al. Effects of simvastatin and atorvastatin administration on insulin resistance and respiratory quotient in aged dyslipidemic non-insulin dependent diabetic patients. Atherosclerosis 2000;150:121-27.

- 32. Mikus CR, Boyle LJ, Borengasser SJ, Oberlin DJ, Naples SP, Fletcher J, et al. Simvastatin impairs exercise training adaptations. J Am Coll Cardiol 2013;62:709-14.
- 33. Martin WH, Klein S. Use of endogenous carbohydrate and fat as fuels during exercise. *Proc Nutr* Soc 1998;57:49-54.
- 34. O'Keefe JH, Schnohr P, Lavie CJ. The dose of running that best confers longevity. Heart tercise 2013;99:588-90.

Table 1. Patient characteristics.

Variable		-
Ν	16	
Gender	13M, 3F	
Age (years)	57±3	
RHR (mmHg)	70±3	
SBP (mmHg)	125±2	
DBP (mmHg)	80±2	
MAP (mmHg)	94 <u>+</u> 2	
Height (cm)	173±2	
Weight (kg)	84±3	
BMI (kg/m^2)	28 ± 1	
Waist (cm)	95±3	
Hip (cm)	103±2	
Waist-hip ratio	0.91±0.02	
%BF (skinfolds)	33±2	
%BF (Tanita)	27 <u>+2</u>	
\dot{VO}_{2max} (ml.kg ⁻¹ .min ⁻¹)	33±3	

ctercis

458 N:number of participants; M:male; F:female; RHR:resting heart rate; SBP:systolic blood pressure;

DBP:diastolic blood pressure; MAP:mean arterial pressure; BMI:body mass index; %BF:percent body
 fat; VO_{2max}:maximal oxygen uptake. Values are mean ±SEM.

Table 2. Dietary intake.

	Energy or
	% Macronutrient intake
Total energy (kJ/day) Carbohydrate (%) Protein (%) Fat (%)	$10,111 \pm 927 \\ 48 \pm 2 \\ 17 \pm 1 \\ 28 \pm 2$
Monounsaturated fat (%)	11±0.8
Polyunsaturated fat (%)	5±0.5
Saturated fat (%) Alcohol (%)	9±0.5 7±2
	, <u> </u>
Values are mean±SEM.	

520 Table 3. Fasting blood lipids (mmol/L) in the absence(-) and presence(+) of a statin.

521

	-statin	+statin	<i>p</i> -value	
T- (-1 -1 -1 - (1	576.021	4 11 .0 14	000	
Total cholesterol	5.76±0.31	4.11±0.14	.000	
Triglyceride	1.69±0.41	1.14±0.15	.073	
HDL-C	1.31±0.10	1.28±0.10	.908	
LDL-C	3.77±0.21	2.32±0.10	.000	
TC/HDL	4.54±0.40	3.39±0.20	.001	
TC/HDL	4.54±0.40	3.39±0.20		

522

523 HDL-C:high density lipoprotein cholesterol; LDL-C:low density lipoprotein cholesterol; TC/HDL:total

524 cholesterol/high density lipoprotein ratio. Values are mean \pm SEM. $p \le 0.05$ significantly different between 525 \pm statin (Student's paired t-test).

> Hedicik Exercise

		-statin			+statin		
Variable	15 min	30 min	45 min	15 min	30 min	45 min	<i>p</i> -value
Fat oxidation	0.16±0.02	0.23 ± 0.01	0.29 ± 0.02	0.13±0.02	0.19±0.02	0.27 ± 0.02	.622
(g/min)							
CHO oxidation	1.8±0.15	1.5±0.13	1.3±0.13	1.7±0.15	1.5±0.13	1.3±0.11	.312
(g/min)							
%Fat	19±2	31±2	38±2	19±3	28±2	38±2	.567
%CHO	82±2	70±2	63±2	82±3	74±3	63±2	.390
RER	$.93 \pm .007$	$.90 \pm 005$	$.88 \pm .006$	$.93 \pm .009$	$.91 \pm .009$	$.88 \pm .007$.393
Glycerol (µmol/l)	175±24	202±20	232±22	159±15	167 ± 15	180±13	.080
(Cayman)							
Glycerol (µmol/l)	102±8	125 ± 11	150±15	99±15	105 ± 14	125±13	.178
(Sigma)							
FFA (µmol/L)	47±8	82±12	138±22	53±13	70±16	104±20	.033
\dot{VO}_2 (ml.kg ⁻¹ .min ⁻¹)	16±1.2	15±1.3	15±1.3	15±1.2	15±1.3	15±1.3	.452
Glucose (mmol/l)	4.2±0.2	4.0±0.2	4.2±0.1	4.3±0.3	4.2±0.2	4.3±0.1	.883
Lactate (mmol/l)	1.3 ± 09	$1.0 \pm .06$	$1.0 \pm .09$	$1.2 \pm .08$	1.0 ± 06	0.9 ± 03	.426
HR (bpm)	109±3	110±3	111±3	101±3	103±3	104±3	.756
RPE	11±0.4	11±0.4	12±0.4	11±0.4	11±0.4	11±0.4	.596

Table 4. Physiological variable measured in participants during a walk at time = 15, 30 and 45
minutes in either the absence(-) or presence(+) of a statin.

529

530 CHO:carbohydrate; %Fat:percentage kcal derived from fat; %CHO:percentage kcal derived from

531 carbohydrate; RER:respiratory exchange ratio; FFA:free fatty acid; VO₂:oxygen uptake; HR(bpm):heart

rate (beats per minute); RPE:rating of perceived exertion. Data are mean \pm SEM. Significance is P \leq 0.05,

533 repeated measures analysis of variance (ANOVA). No significance achieved for 'treatment and time' for

any variables (exception FFA). For FFA, significance is $p \le 0.016$ (Bonferroni adjustment) with the

535 simple main effect also not statistically significant.