



http://www.arjournals.org/index.php/ijpm/index

International Automational Physiological Phy

**Original Research Article** 

## Phytochemical investigation and antibacterial activity of *Gymnema sylvestre* and *Andrographis paniculata* from western ghats Sukesh K<sup>1</sup>, Shafi Thompson T<sup>2</sup>, Densingh J<sup>1</sup>

#### \*Corresponding author:

#### T. Shafi thompson

Assistant Professor, Department of Biotechnology, Mar Ivanios College, Thiruvananthapuram Dist-695 015, Kerala, India. Tel: +919847793663, shafithompson@gmail.com

### Abstract

In this study the antibacterial activity of Gymnema sylvestre (Retz.) and Andrographis paniculata collected from Western Ghats, Kanyakumari district, South India was made against common bacterial pathogens by filter paper disc-agar diffusion procedure. Both the plants showed considerable levels of antibacterial activity. Hexane extract of Gymnema sylvestre showed maximum inhibition against Serratia marcescens MTCC 86 and the Andrographis paniculata was effective against penicillin resistant Staphylococcus aureus MTCC 87. Phytochemical study revealed the presence of steroids/ terpenoids and coumarins in the extracts of both the plants. The bio-active compounds in the medicinal plants were extracted with Hexane and Chloroform (99 %) and further resolved by thin layer chromatography. Hexane extract of Gymnema sylvestre (Retz.) have 15 compounds and in its chloroform extract 25 compounds whereas 19 compounds in Chloroform extract and five compounds with Hexane extract was found in Andrographis paniculata. Further research is needed to characterize the compounds obtained and their mechanism of action on the bacterial pathogens.

### Introduction

Historically, the medicinal value of plants was tested by trial and error. Modern approaches to determining the medicinal properties of plants involve collaborative efforts of ethno-botanists, anthropologists, pharmaceutical chemists, and physicians. Many modern medicines had their origin in medicinal plants. Examples include aspirin from willow bark (Salix spp.), digitalis from foxglove (Digitalis purpurea), and vinblastine from Madagascar periwinkle (Vinca rosea). More than 13,000 plants have been studied during the last 5 years period [Dahanukar et al., 2000]. The Indian Systems of medicine can be classified into traditional and classical systems. The traditional system is the local folk stream, which is prevalent in rural and tribal villages in India. The system like Ayurveda, Sidha, Unani, Yoga, and Naturopathy are

expressions of classical systems. Thus the term Indian Systems of Medicine covers the system which originated in India or which originated outside but got adapted in the course of time [Sharma, 1995]. Plants have an almost limitless ability to synthesize aromatic substances, most of which are phenols or their oxygen-substituted derivatives [Geissman, 1963]. Most are secondary metabolites, of which only 12,000 have been isolated, which are estimated to be less than 10% of the total [Schultes, 1978]. Of these, only small percentage has been investigated phyto-chemically and the fraction submitted to biological or Pharmacological screening is even lower. Since plants may contain hundreds or even thousands of metabolites, there is currently a great interest in the medicinal plant research as a possible source of new lead compounds for introduction therapeutical screening into

programmes. The new branch of Science, ethnobotany (or ethno-pharmacology), whose goal is to utilize the impressive array of knowledge assembled by indigenous peoples about the plant and animal products they have used to maintain health [Borris, 1996]. The investigation was to analyze the bioactive potential and phytochemical constituents of two medicinal plants Gymnema sylvestre and Andrographis paniculata collected from the Western Ghats, part in Kanya kumari District, Tami Nadu, India. The motivation behind the study was detection of a phytoremedy for microbial infections as an alternative to chemotherapeutics and to have constructive exploitation of bioresources of a region thereby developing special protection over rare species.

# Materials and Methods

Medicinal Plant Material: Leaves of *Gymnema* sylvestre and *Andrographis paniculata* were collected from the lower region of Western Ghats [Lat. 8° 19'N; Long. 77° 10' S] and authenticated taxonomy by standard procedures. *Gymnema* sylvestre [Retz.] R.Br. ex Roem. & Schult is medicinally important in antidiabetic activity. It has shown experimental or clinical anti-diabetic activity and it boosts the insulin level. Andrographis paniculata, a native medicinal plant commonly known as 'Kalmegh' (Hindi) 'Kiriath' (Malayalam); Nilavempui (Tamil).

Preparation of Medicinal Plant Extracts: The hexane and chloroform extracts of *Gymnema* sylvestre and Andrographis paniculata were used for the study. 500 gram leaves of both medicinal plant samples were shade dried and pulverized. From this 15 gram was mixed with 50 ml of hexane and was placed in a rotary shaker. The residual powder after hexane extraction was dried and was mixed with 50 ml of chloroform and placed in a rotary shaker followed by evaporation. The percentage yield of *Gymnema* sylvestre and Andrographis paniculata extracts were calculated.

Bacterial Pathogens: Eight different pathogenic bacteria viz., Staphylococcus aureus [PRSA] MTCC 87, Salmonella typhi MTCC 531, Pseudomonas aeruginosa MTCC 424, Clostridium perfringens MTCC 450, Serratia marcescens MTCC 86, Bacillus subtilis MTCC 1427, Enterobacter aerogenes MTCC 111 and Shigella flexneri MTCC 1457 obtained from Microbial Type Culture Collection (MTCC, Chandigarh, India] were used in the present study.

Antibacterial Activity Study: Filter paper discagar diffusion procedure (Kirby-Bauer method) was used to study the antimicrobial activity [Bauer et al., 1966; Dey and Harborne, 1991]. Filter paper discs (Whatman No.41) of uniform size (5 mm diameter), impregnated with specified concentrations of the different extracts (100µg/disc) and control were placed on the surface of the agar plate, that has been pre-seeded with the bacterial pathogens on Muller Hinton Agar (Himedia, India) plates with a pH of 7.4. The plates were incubated at 37°C for 17 hours. The antimicrobial activity was determined by measuring the diameter of the zone of inhibition in millimeter. Chloroform (95% v/v) and Hexane (95% v/v) were used as negative control and the antibiotic tetracycline was used as positive control for Salmonella sp and penicillin for Staphylococcus sp. All the experiments were done in triplicates. The average of triplicate values was calculated.

Phytochemical Investigation: The extracts were subjected to following phytochemical investigation such as Liebermann-Burchard test for steroids/ terpenoids, Shinoda's test for flavanoids, Molisch's test for carbohydrates, Wiefferering field test for iridoids, test for coumarins, Test for saponins, Dragendorff's test for alkaloids [Sofowora, 1984 and Kepm, 1986].

Thin Layer Chromatography: The crude extracts were subjected to TLC with developing solvent systems using different ratios of Hexane, Chloroform and ethanol [Sadasivam and Manickam, 1996]. The TLC plates were exposed to iodine vapour in a glass chamber for locating unsaturated compounds and Rf values were calculated.

#### Results

The extractive percentage was higher with hexane extract of *Gymnema sylvestre* 5 %

whereas chloroform extracts of *Andrographis* paniculata showed 2% as its maximum (Fig 1).

The results of antibacterial activity of hexane and chloroform extracts of *Gymnema sylvestre* and *Andrographis paniculata* are shown in Table 1.

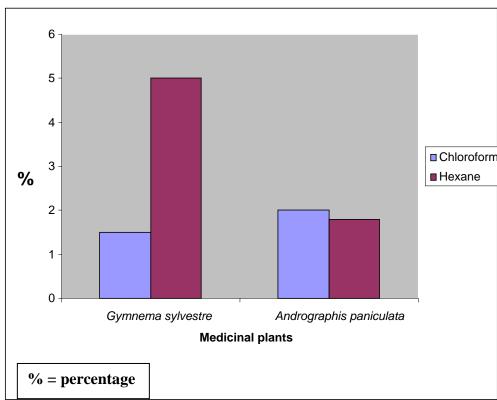


Figure 1. Extractive Percentage of G. sylvestre and A.paniculat.

Postorial Pathogona		hibition by y <i>lvestre</i> (mm)	Zone of inhibition by A. paniculata (mm)	
Bacterial Pathogens	Hexane extract	Chloroform extract	Hexane extract	Chloroform extract
Staphylococcus aureus [PRSA] MTCC 87	12	7	17	19
Salmonella typhi MTCC 531	8	Т	9	11
Pseudomonas aeruginosa MTCC 424	13	15	8	14
Clostridium perfringens MTCC 450	10	10	Т	8
Serratia marcescens MTCC 86	18	Т	17	14
Bacillus subtilis MTCC 1427	-	Т	12	9
Enterobacter aerogenes MTCC 111	12	Т	15	17
Shigella flexneri MTCC 1457	12	Т	8	11

 Table 1. Antibacterial Effect of Gymnema sylvestre and Andrographis paniculata

'-'= No activity; T= Trace <7.0; MTCC= Microbial Type Culture Collection. (Values are the average of Triplicates)

Zone of inhibition of hexane extract of *Gymnema* sylvestre ranges from 8.0 mm (*Pseudomonas* 

aeruginosa MTCC 424) to 18.0 mm for Serratia marcescens MTCC 86 whereas the activity by its

chloroform extract was maximum for *Pseudomonas aeruginosa* MTCC 424 (15.0 mm) followed by *Clostridium perfringens* MTCC 450 ((10.0 mm) and *Staphylococcus aureus* [PRSA] MTCC 87 (7.0 mm) but other pathogens showed trace response i.e., less than 7.0 mm.

Staphylococcus aureus [PRSA] MTCC 87 (19.0 mm), Enterobacter aerogenes MTCC 111 (17.0 mm), Pseudomonas aeruginosa MTCC 424 and Serratia marcescens MTCC 86 (14.0 mm each), Salmonella typhi MTCC 531 and Shigella flexneri MTCC 1457 (11.0 mm each) were the diameter of zone of inhibition by the chloroform extracts of Andrographis paniculata. Its hexane extract activity was maximum towards Staphylococcus aureus [PRSA] MTCC 87 and

Serratia marcescens MTCC 86 (17.0 mm) followed by Enterobacter aerogenes MTCC 111 (15.0 mm), Bacillus subtilis MTCC 1427 (12.0 mm), Salmonella typhi MTCC 531 (9.0 mm) and Pseudomonas aeruginosa MTCC 424 and Shigella flexneri MTCC 1457 (8.0 mm each) but Clostridium perfringens MTCC 450 showed less sensitivity.

The phytochemical screening of *Gymnema* sylvestre showed the presence of steroids or terpenoids and coumarins in the chloroform and steroids or terpenoids alone was found in its hexane extract. Incase of *Andrographis* paniculata both the extracts showed the presence steroids or terpenoids and coumarins (Table 2).

Table 2. I	Phytochemical	screening of G.	sylvestre and A.	paniculata.

Experiments	Phytochemical analysis of G. sylvestre		Phytochemical analysis of A. paniculata	
	Hexane extract	Chloroform extract	Hexane extract	Chloroform extract
Liebermann-Burchard test for steroids or terpenoids	+	+	+	+
Shinoda's test for flavanoids	-	-	_	_
Molisch's test for carbohydrates	-	-	_	_
Wiefferering field test for iridoids	-	-	_	_
Test for coumarins	-	+	+	+
Test for saponins	-	-	_	_

- = Absent; + = Present

 Table 3. TLC of Gymnema sylvestre (Retz. R.Br.ex Roem. & Schult.

	Solvent system	No of spots obtained	<b>Rf values</b>
	Chloroform (100%)	5	0.137, ,0.168, 0.475, 0.893, 0.906
	Chloroform-	7	0.438, 0.451, 0.380, 0.651, 0.677,
Chlorofor	Ethanol(9.5:0.5)		0.845, 0.974
m Extract	Chloroform-Ethanol(9:1)	3	0.024, 0.325, 0.487
	Chloroform-Ethanol (7:3)	5	0.069, 0.333, 0.837, 0.852, 0.883
	Chloroform- Ethanol(8.5:1.5)	5	0.096, 0.103, 0.482, 0.848, 0.896
Hexane extract	Hexane-Chloroform	6	0.192, 0.144, 0.192,
	(9.5:0.5)	0	0.269, 0.576, 0.932
	Hexane-Chloroform (7:3)	9	0.047, 0.085, 0.114, 0.171, 0.228, 0.571, 0.619, 0.809, 0.980

Chlorofor m Extract	Solvent system	No of spots obtained	Rf value
	Chloroform (100%)	3	0.264, 0.052, 0.523
	Chloroform-ethanol (9:1)	5	0.160, 0.400, 0.120, 0.560, 0.600
	Chloroform-ethanol (9.5:0.5)	5	0.021, 0.425, 0.702, 0.709, 0.758
	Chloroform-Ethanol (9:1)	3	0.780, 0.950, 0.957
	Chloroform-Ethanol (7:3)	6	0.240, 0.263, 0.829 0.845, 0.914, 0.902
Hexane	Hexane-Chloroform (9.5:0.5)	3	0.144. 0.192,0.576
extract	Hexane-Chloroform (3:7)	2	0.056, 0.099

Sukesh *et al.* International Journal of Phytomedicine 3 (2011) 254-260 Table 4 TLC of *Andrographis paniculata* (Burm f) Wall, ex Nees

The result of thin layer chromatography of hexane and chloroform extract of *Gymnema* sylvestre Retz. R.Br.ex Roem. & Schult is shown in Table 3. The chloroform-ethanol (9:1) gave three fractions and the second fraction showed the highest Rf value (0.950). Totally Hexane extract of *Gymnema sylvestre* [Retz.] revealed to have 15 compounds and 25 compounds by its chloroform extract. The Hexane extract of *Andrographis paniculata* showed the presence of five compounds whereas 22 compounds were noticed in its Chloroform extracts (Table 4).

### Discussion

Plants are rich in a wide variety of secondary metabolites such as tannins, alkaloids, flavanoids which have been found in vitro to have antimicrobial properties. Among these, only around ten percentages of all the plants had been investigated in detail for bioactive agents [Sanberg and Bruhn, 1979]. So it is imperative to characterize plants with bioactive properties to develop consumer friendly therapeutics with challenging effectiveness. The major advantage of this type studies may form a solution for emergence of multiple drug resistantence or even the complete cure or an alternative to diverse diseases. Various plant extracts are reported to be beneficial in the control of Hepatitis, and have antiviral, antibacterial, antimalarial and immune modulating factors and are with highly potent antibacterial and antimalarial ingredients [Mcrea and Towers, 1984]. The bioactive substances in plants though produced as secondary metabolites not only serves as developmental stage specific, but also organ and tissue specific [Daroker et al., 1988]. Selection of plant samples of this study considered all these remarkable properties of medicinal plants. The plant samples were collected from the Western Ghats is considered as one of the global biodiversity hot-spots. It comprises of over 5000 vascular plants, of which about 30% are endemic to the Ghats and this area has become a hotspot of discovery of new species in the last decade.

All the eight pathogenic bacteria viz., Staphylococcus aureus [PRSA] MTCC 87, Salmonella typhi MTCC 531, Pseudomonas aeruginosa MTCC 424, Clostridium perfringens MTCC 450, Serratia marcescens MTCC 86, Bacillus subtilis MTCC 1427, Enterobacter aerogenes MTCC 111 and Shigella flexneri MTCC 1457 obtained from Microbial Type Culture Collection (MTCC, Chandigarh, India] were sensitive to any of the extracts used (Table 1).

The Hexane extract of *Gymnema sylvestre* inhibited the growth of all the bacterial isolates analysed except Salmonella typhi MTCC 531 and Bacillus subtilis MTCC 1427. Its Chloroform extract was active against Pseudomonas aeruginosa MTCC 424, Clostridium perfringens MTCC 450, and Staphylococcus aureus MTCC 87 whereas Serratia marcescens MTCC 86, Bacillus subtilis MTCC 1427, Enterobacter aerogenes MTCC 111 and Shigella flexneri MTCC 1457 and Salmonella typhi MTCC 531 showed less sensitivity. The chloroform and hexane extracts of *Andrographis paniculata* was inhibitory to all the bacterial pathogens. [Mishra et al., 2009] reported the antimicrobial activity of Cinnamonum zeylanicum extracts against human and plant pathogenic dermatiaceous moulds. The antibacterial effect of Azadirachta indica extracts against food borne pathogens and spoilage bacteria was reported by [Mahfuzul Hoque et al., 2007].

The highest inhibition towards Methicillin resistant S. aureus by both Chloroform and Hexane extracts of Andrographis paniculata is the remarkable finding of the study. The high antimicrobial properties of Andrographis paniculata (Burm.f.) are due to high phenolic composition [Tripathi et al., 2000; Patra et al., Similar result was reported 2004]. by [Dharmaratne et al., 1999] in which the sensitivity of MRSA strain with xanthones of Calophyllum sp. has been revealed. Further he suggested that the variations in activity among the organic solvents may be due to the polarity which determines the type of the reaction and solubility of the compounds. Antimicrobial activity of Syzygium aromaticum extracts against multiple drug resistant bacterial strains and fungus of clinical origin was reported by [Khan et al., 2009].

Presence of steroids/ terpenoids and coumarins (Table 2). The detection of 15 compounds in hexane and 25 compounds in chloroform extract of *Gymnema sylvestre* (Table 3) and five compounds of hexane and 22compounds in Chloroform extracts of *Andrographis paniculata* (Table 4) indicates that both plants studied are with higher numbers of compounds with potential antibacterial activity. Plants, rich in the secondary metabolites such as alkaloids, glycosides, steroids and relative active metabolites are used as drugs in pharmaceutical industry [Vlietinck et al., 1995]. Further research is needed to characterize the compounds obtained. More over, the presence of different active fractions in the TLC studies further reiterates the antibacterial properties of the selected plants. The antimicrobial activity of the extracts and their potency can be quantitatively assessed by determining the minimum inhibitory concentration (MIC) and Minimum Bactericidal Concentration (MBC).

The results suggest that the hexane and chloroform extracts of Gymnema sylvestre and Andrographis paniculata posses potent antibacterial activity. Further works are proceeding in Malankara catholic college laboratory to identify and characterize the compounds of these plants and also to check its antimycotic activity and response towards other than the bacteria studied.

# Acknowledgement

The authors are thankful to Rev Fr. Prem Kumar, MSW, The Correspondent and Secretary of Malankara Catholic College, for providing facilities and moral support.

## References

- 1. Dahanukar SA, Kulkarni RA and Rege NN. Pharmacology of medicinal plants and natural products. Indian J. Pharmacol. 2000;32:81-118.
- 2. Sharma PV. Glimpses of Indian ethnopharmacology. Proceedings of the First National Conference on Ethnopharmacology, May 24-26, TBGRI Publication. 1995;pp: 233-242.
- 3. Geissman TA. Flavonoid Compounds, Tannins, Lignins and Related Compounds. In: Pyrrole Pigments, Isoprenoid Compounds and Phenolic Plant Constituents, Stotz, E.H. (Ed.) Elsevier, New York. 1963;Vol. 9:pp: 265.
- Schultes RE. The Kingdom of Plants. In: Medicines from the Earth, Thompson, W.A.R. (Ed.), McGraw-Hill Book Co., New York. 1978;pp:208.
- 5. Borris RP. Natural product research: Perspective from a major pharmaceutical

company. J. Ethnopharmacol. 1996;51:29-38.

- 6. Bauer AW, Kirby WMM, Sherris JC and Truck M. Antibiotic susceptibility testing by standard single disk diffusion method. Am. J. Clin. Path. 1966;45:493-496.
- 7. Dey PM and Harborne JB. Methods in Plant Biochemistry. Vol.1., Academic Press, London. 1991;pp: 47-58.
- Sofowora A. Medicinal Plants and Traditional Medicine in Africa. John Wiley and Sons Ltd, New York. 1984;pp: 256.
- Kepm W. Qualitative Organic Analysis: Spectrochemical Techniques. 2<sup>nd</sup> Edn., McGraw- Hill, London and New York, ISBN 0070841586. 1986;pp: 197.
- 10. Sadasivam S and Manickam A. Biochemical Methods. 2nd Edn., New Age International Publishers, New Delhi. 1996;pp: 1-255.
- Sanberg F and Bruhn JG. Sreeninig of Plants for Bioactive Substances in Africa. O.A.U. Press, Nigeria. 1979;pp: 119.
- 12. Mcrea WD and Towers GHN. Biological activities of lignans. Phytochemistry. 1984;23:1207-1220.
- 13. Daroker MP, Mathur A, Dwivedi S, Bhalla R, Khanuja SPS and Kumar S. Detection of antibacterial activity in the floral petals of some higher plants. Curr. Sci. 1988;75:187–189.
- 14. http://203.190.147.121/bitstream/123456789 /83/1/J\_Current%20Science\_75\_187.pdf.
- 15. Dharmaratne HRW, Wijesinghe WMN and Thevanasem V 1999. Antimicrobial activity xanthones from of <I>Calophyllum species</I>, against methicillin-resistant <I>Staphylococcus aureus</I> (MRSA). J. Ethnopharmacol., 339-342. 10.1016/S0378-66: doi: 8741(98)00239-6.

- 16. Khan R, Islam B, Akram M, Shakil S and Ahmad A <I>et al</I>. Antimicrobial activity of five herbal extracts against Multi Drug Resistant (MDR) strains of bacteria and fungus of clinical origin. Molecules. 2009;14:586-597.
- 17. Mishra AK, Mishra A, Kehri HK, Sharma B and Pandey AK. Inhibitory activity of Indian spice plant <I>Cinnamomum zeylanicum</I> extracts against human and plant pathogenic dematiaceous moulds. Ann. Clin. Microbiol. Antimicrob. 2009;8:9-9.doi:10.1186/1476-0711-8-9.
- Mahfuzul Hoque MD, Bari ML, Inatsu Y, Juneja VK and Kawamoto S. Antibacterial activity of guava (<I>Psidium guajava</I> L.) and Neem (<I>Azadirachta indica</i> A. Juss.) extracts against foodborne pathogens and spoilage bacteria. Foodborne Pathog. Dis. 2007;4:481-488.

doi:10.1089/fpd.2007.0040.

- 19. Tripathi AK, Verma N, Prajapati V, Khanuja SPS, Shasany AK, Satpathy S and Kumar S. Plant extracts and compounds active against the crop insectpest <I>Helicoverpa armigera</I>. J. Med. Arom. Plant Sci. 2000;22/23:125-145.
- 20. Patra DD, Chattopadhyay A, Mishra HO, Alam M and Khanuja SPS <I>et al</I>.. Agrotechnology of kalmegh (<I>Andrographis paniculata</I>). J. Med. Arom. Plant Sci. 2004;26:534–537.
- 21. Vlietinck AJ, van Hoof L, Totte J, Lasure A, Vanden Berghe D, Rwangabo PC and Mvukiyumwami J. Screening of hundred Rwandese medicinal plants for antimicrobial and antiviral properties. J. Ethnopharmacol. 1995;46:31-47.