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Review

Natural sources as potential anti-cancer agents: A review Abhishek Bhanot, Rohini Sharma, Malleshappa N. Noolvi*

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Abstract

Natural products remain an important source of new drugs, new drug leads and new chemical entities. The plant based drug discovery resulted mainly in the development of anticancer agents including plants (vincristine, vinblastine, etoposide, paclitaxel, camptothecin, topotecan and irinotecan), marine organisms (citarabine, aplidine and dolastatin 10) and micro-organisms (dactinomycin, bleomycin and doxorubicin). Beside this there is numerous agents identified from fruits and vegetables can used in anticancer therapy. The agents include curcumin (turmeric), resveratrol (red grapes, peanuts and berries), genistein (soybean), diallyl sulfide (allium), S-allyl cysteine (allium), allicin (garlic), lycopene (tomato), capsaicin (red chilli), diosgenin (fenugreek), 6-gingerol (ginger), ellagic (pomegranate), ursolic acid (apple, pears, prunes), silymarin (milk thistle), anethol (anise, camphor, and fennel), catechins (green tea), eugenol (cloves), indole-3-carbinol (cruciferous vegetables), limonene (citrus fruits), beta carotene (carrots), and dietary fiber. In this review active principle derived from natural products are offering a great opportunity to evaluate not only totally new chemical classes of anticancer agents, but also novel lead compound and potentially relevant mechanisms of action.

Keywords: Cancer, vincristin, vinblastin, fruit, vegetables.

Introduction

Cancer continues to be one of the major causes of death worldwide and only modest progress has been made in reducing the morbidity and mortality of this disease [1]. Cancers may be caused in one of three ways, namely incorrect diet, genetic predisposition, and via the environment. As many as 95% of all cancers are caused by life style and may take as long as 20–30 years to develop. Current estimates from the American Cancer Society and from the International Union Against Cancer indicate that 12 million cases of cancer were diagnosed last year, with 7 million deaths worldwide; these numbers are expected to double by 2030 (27 million cases with 17 million deaths) [2].

According to a report of World Health Organization, more than 80% of world's populations depend on traditional medicine for their primary health care needs [3,4]. Plants have a long history of use in the treatment of cancer and it is significant that over 60% of currently used anti-cancer agents are come from natural sources [5]. Naturally occurring drugs that are part of the war against cancer include (vincristine, vinca alkaloids vinblastine, vindesine, vinorelbine), taxanes (paclitaxel, docetaxel), podophyllotoxin and its derivative (etoposide, teniposide), camptothecin and its derivatives (topothecan, irinothecan), anthracyclines (doxorubicin, daunorubicin. epirubicin, idarubicin) and others. In fact, half

of all anti-cancer drugs approved internationally were either natural products or their derivatives and were developed on the basis of knowledge gained from small molecules or macromolecules that exist in nature [6,7]. In between 2001 and 2005, 23 new drugs derived from natural products were introduced for the treatment of disorders such as bacterial and fungal infections, cancer, diabetes, dyslipidemia, atopic dermatitis, Alzheimer's disease and genetic diseases such as tyrosinaemia and Gaucher disease out of these 4 drugs have been approved as anti cancer agents. The approved anti cancer agents in 2002 doxorubicin, in 2002 estradiol, in 2004 cholorophyll and l- aspartic acid and taxol nanoparticles in 2005 [8]. Three new drugs also introduced in 2007 originate from microbial sources for the treatment of cancer is marine alkaloid trabectedin, epothilone derivative ixabepilone and temsirolimus [9].

Nature is an attractive source of new therapeutic candidate compounds as a tremendous chemical diversity is found in millions of species of animals, marine organisms plants. microorganisms as potential anti-cancer agent [10,11]. In this present study the potential antiagent cancer from plants. microorganisms and dietary (fruits, vegetables, spices) sources with some recent advancement in the field of cancer research were discussed.

Plants as source of anti-cancer agents:

The history of plant as source of anti-cancer agents started in earnest in the 1950s with the discovery and development of the vinca alkaloids (vinblastine and vincristine) and the isolation of the cytotoxic podophyllotoxins. Vinca alkaloid was responsible for an increase in the cure rates for Hodgkin's disease and some forms of leukemia [12]. Vincristine inhibits microtubule assembly, inducing tubulin self-association into coiled spiral aggregates [13]. Etoposide is a epipodophyllotoxin, derived from the mandrake plant Podophyllum peltatum and

the wild chervil Podophyllum emodi [14]. It has also significant activity against small-cell lung carcinoma [15]. Etoposide is a topoisomerase II inhibitor, stabilizing enzyme-DNA cleavable complexes leading to DNA breaks [16]. The taxanes paclitaxel and docetaxel has been show antitumor activity against breast, ovarian and other tumor types in the clinic trial. Paclitaxel stabilizes microtubules and leading to mitotic arrest [17]. In addition, the camptothecin derivatives irinotecan and topotecan, have shown significant antitumor activity against colorectal and ovarian cancer respectively [18,19]. These compounds were initially obtained from the bark and wood of Nyssacea Camptotheca accuminata and act by inhibiting topoisomerase I [20]. The taxanes and the camptothecins are presently approved for human use in various countries (Table 1).

Table 1: Plant based anticancer agents in clinical practice.

practi	се.		
S.No.	Compound	Uses	Status
1.	Vincristine	Leukemia, lymphoma,	Phase
		breast, lung, pediatric	III/IV
		solid cancers and	
		others	
2.	Vinblastine	Breast, lymphoma,	Phase
		germ-cell and renal	III/IV
		cancer	
3.	Paclitaxel	Ovary, breast, lung,	Phase
		bladder and head and	III/IV
		neck cancer	
4.	Docetaxel	Breast and lung cancer	Phase III
5.	Topotecan	Ovarian, lung and	Phase
		pediatric cancer	II/III
6.	Irinotecan	Colorectal and lung	Phase
		cancer	II/III

Rohitukine the plant alkaloid, isolated from the leaves and stems of Dysoxylum binectariferum (Maliaceae) [21,22]. Synthetic flavone derived from rohitukine, Flavopiridol representing the first cyclin-dependent kinase inhibitor to enter the clinical trial [23]. The mechanism of action involves interfering with the phosphorylation of cyclin-dependent kinases and arrest cell-cycle progression at growth phase G1 or G2 [24,25].

Homoharringtonine an alkaloid isolated from the Chinese tree Cephalotaxus harringtonia (Cephalotaxacea) [26]. The mechanism of action is the inhibition of protein synthesis and blocking cell-cycle progression [27]. It has shown efficacy against various leukemias [28]. A lung-cancer-specific antineoplastic agent 4-Ipomeanol is isolated from the sweet potato Ipomoea batata (Convolvulaceae) [29]. The mechanism of action is converted into DNA-binding metabolites upon metabolic activation by cytochrome P450 enzymes that are present in

cells of the lung [30]. DNA topoisomerase I inhibitor β -lapachone, that induces cell-cycle delay at G1 or S (synthesis) phase before inducing either apoptotic or necrotic cell death in a variety of human carcinoma cells, including ovary, colon, lung, prostate and breast [31].

Beside this there are so many plants which are used in cancer; following enlist the plant which prevent and target for future studies as potential anticancer agent (Table 2):

Table 2: Plants used as anti-cancer.

	Plants used as anti-cancer.			
S.No	Plant Species	Family	Plant Part	References
1.	Salvia officinalis	Labiatae	Leaves	[32]
2.	Viscum album	Loranthaceae	Leaves	[33]
3.	Combretum caffrum	Combretaceae	Bark	[34]
4.	Melaleuca alternifolia	Myrtaceae	Leaves	[35]
5.	Lavandula angustifolia	Labiatae	Leaves	[35]
6.	Aglaia foveolata	Meliaceae	Fruit	[36]
7.	Maytenus serrata	Celastraceae	Seed	[37]
8.	Tabebuia impetiginosa	Bignoniaceae	Stem bark and trunk wood	[38,39]
9.	Tabebuia rosea	Bignoniaceae	Stem bark and trunk wood	[38,39]
10.	Tabebuia serratifolia	Bignoniaceae	Stem bark and trunk wood	[38,39]
11.	Dipteryx odorata	Fabaceae	Seed	[40]
12.	Thapsia garganica	Apiaceae	Fruit	[41]
13.	Indigofera tinctoria	Leguminosae	Aerial part	[42]
14.	Matricaria chamomilla	Asteraceae	Flower	[43]
15.	Erythroxylum pervillei	Erythroxylaceae	Root	[44]
16.	Broussonetia papyrifera	Urticaceae	Entire	[45]
17.	Cyclopia intermedia	Fabaceae	Leaves	[46]
18.	Scutellariae radix, Scutellariae indica	Labiatae	Root	[47]
19.	Physalis philadelphica	Solanaceae	Seed	[48]
20.	Dysoxylum binectariferum	Meliaceae	Stem bark	[49]
21.	Aristotelia chilensis	Elaeocarpaceae	Leaf and Stem	[50]
22.	Cyathostemma argentium	Annonaceae	Root	[51]
23.	Epimedium hunanense	Berberidaceae	Aerial parts	[52]
24.	Croton urucurama	Euphorbiacaeae	Bark	[53]
25.	Epilobium hirsutum	Onagraceae	Entire	[54]
26.	Pleione bulbocodioides	Orchidaceae	Tuber	[55]
27.	Cassia quinquangulata	Caesalpiniaceae	Root	[56]
28.	Begonia glabra	Begoniaceae	Entire	[57]
29.	Celastrus orbiculatus	Celastraceae	Entire	[57]
30.	Croton draco	Euphorbiacaeae	Aerial parts	[57]
31.	Smilax sieboldii	Liliaceae	Entire	[58]
32.	Ximenia Americana	Olacaceae	Root	[58]

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33.	Maytenus emarginata	Celastraceae	Entire	[59]
34.	Sarcandra glabra	Choranthaceae	Entire	[60]
35.	Salvia plebeian	Labiatae	Aerial	[61]
36.	Scutellaria barbata	Labiatae	Entire	[62]
37.	Ocotea caparrapi	Lauraceae	Essential oil	[63]
38.	Caragana cuneata	Leguminosae	Leaf	[64]
39.	Croton flavens	Euphorbiacaeae	Leaf	[65]
40.	Euphorbia heterophylla	Euphorbiacaeae	Stem	[65]
41.	Echites vucatanensis	Apocynaceae	Latex	[65]
42.	Thevetia ahouia	Apocynaceae	Leaf and Stem	[65]
43	Thevetia gaumeri	Apocynaceae	Leaf and Stem	[65]
44.	Thevetia peruciana	Apocynaceae	Leaf and Stem	[65]
45.	Euphorbia ebracteolata	Euphorbiacaeae	Aerial parts	[66]
46.	Dioscorea collettii	Dioscoreaceae	Rhizome	[67]
47.	Juglans mandshurica	Juglandaceae	Root	[68]
48.	Maackia tenuifolia	Leguminosae	Root	[69]
49	Juncus acutus	Juncaceae	Leaf	[70]
50.	Hedyotis chrysotricha	Rubiaceae	Entire	[71]
51.	Arisaema erubescens	Araceae	Root	[72]
52.	Leptadenia hastate	Asclepiadaceae	Bark	[73]
53.	Viscum calcaratum	Loranthaceae	Entire	[74]
54	Aphanamixis polystachya	Meliaceae	Stembark	[75]
55.	Pratia nummularia	Campanulaceae	Entire	[76]
56.	Aeonium arboretum	Crassulaceae	Leaf	[77]
57.	Ocotea foetens	Lauraceae	Branchlets	[77]
58.	Maytenus canariensis	Celastraceae	Fruit juice	[78]
59.	Sedum alboroseum	Crassulaceae	Entire	[79]
60.	Euphorbia micractina	Euphorbiacaeae	Entire	[80]
61.	Euphorbia prolifera	Euphorbiacaeae	Latex	[81]
62.	Scirpus holoschoenus	Cyperaceae	Inflorescence	[82]
63.	Dillenia suffruticosa	Dilleniaceae	Fruit	[83]
64	Hypoxis rooperii	Hypoxiaceae	Tuber	[84]
65.	Inula linariaefolia	Compositae	Flowers	[85]
66.	Ziziphus mauritiana	Rhamnaceae	Stem bark and Fruit	[86]
67.	Adiantum macrophyllum	Pteridaceae	Entire	[87]
68.	Thalictrum fabri	Ranunculaceae	Root	[88]
69.	Scutellaria indica	Labiatae	Root	[89]
70.	Hypericum japonicum	Guttiferae	Entire	[90]
71.	Cyathea fauriei	Cyatheaceae	Shoot	[91]
72.	Fissistigma oldhamii	Annonaceae	Stem	[92]
73.	Monnina obtusifolia	Polygalaceae	Aerial parts	[93]
74.	Coriolus versicolor	Polyporaceae	Fruitbody	[94]
75.	Melastoma malabathricum	Melatomataceae	Flower	[95]
76.	Carapa guianensis	Meliaceae	Seed oil	[96]
70. 77.	Swietenia humilis	Meliaceae	Seed	[97]
77. 78.	Ficus pretoiae	Moraceae	Sap	[98]
70.	i icus premiae	ivioraccac	Jup	[٥٥]

79.	Croton lechleri	Euphorbiacaeae	Latex	[99]
80.	Aster amellus	Compositae	Entire	[100]
81.	Crassocephalum bojeri	Compositae	Entire	[101]
82.	Echinops grijisii	Compositae	Root	[101]
83.	Adenium obesum	Apocynaceae	Leaf	[102]
84.	Ipomea batata	Convolvulaceae	Rhizome	[103]
85.	Uncaria tomentosa	Rubiaceae	Bark	[104]
86.	Plantago asiatica	Plantaginaceae	Leaf	[105]
87.	Phymatosorus diversifolium	Polydiaceae	Root	[105]
88.	Rabdosia rubescens	Labiatae	Leaf	[106]
89.	Salvia chinensis	Labiatae	Entire	[107]
90.	Ganoderma lucidum	Ganodermataceae	Fruitbody	[108]
91.	Euphorbia kansui	Euphorbiacaeae	Root	[109]
92.	Echinops latifolius	Compositae	Root	[110]
93.	Euphorbia marginata	Euphorbiacaeae	Entire	[111]
94.	Ligustrum lucidum	Oleaceae	Seed	[112]
95.	Phytolacca esculenta	Phytolaccaceae	Root	[113]
96.	Pinus parviflora	Pinaceae	Strobilus	[114]
97.	Dysosma pleiantha	Berberidaceae	Root	[115]
98.	Alnus japonica	Betulaceae	Wood	[116]
99.	Ruellia tuberose	Acanthaceae	Bark	[117]
100.	Acacia xanthophloea	Leguminosae	Fruit	[118]
101.	Lannea stuhlmannii	Anacardiaceae	Root	[118]
102.	Maytenus obscura	Celastraceae	Leaf	[118]
103.	Plicosepalus sagittifolius	Loranthaceae	Branches	[118]
104.	Piper latifolium	Piperaceae	Leaf	[119]
105.	Morinda citrifolia	Rubiaceae	Root	[119]
106.	Knema tenuinervia	Myristicaceae	Stembark	[120]
107.	Deeringia amaranthoides	Amaranthaceae	Fruit	[121]
108.	Cynanchum hancoekianum	Asclepiadaceae	Entire	[122]
109.	Azadirachta indica	Meliaceae	Leaf	[123]
110.	Virola bicuhyba	Myristicaceae	Seed	[124]
111.	Sempervivum armenum	Crassulaceae	Leaf	[125]
112.	Sempervivum arvense	Crassulaceae	Leaf	[125]
113.	Hippophae salicifolia	Elaeagnaceae	Fruit	[126]
114.	Hypoxis nyasica	Hypoxiaceae	Rhizome	[127]
115.	Astragalus membranaceus	Leguminosae	Root	[128]
116.	Maytenus macrocarpa	Celastraceae	Stembark	[129]
117.	Cephalotaxus Harrington	Cephlotaxaceae	Entire	[130]

Dietary source of anti cancer agents:

Natural dietary agents including fruits, vegetables, and spices have drawn a great deal of attention from both the scientific community and the general public owing to their demonstrated ability to suppress cancers. Recent

studies suggest that the consumption of food rich in fruits, vegetables and spices have a lower incidence of cancers (stomach, esophagus, lung, oral cavity and pharynx, endometrium, pancreas and colon) [131-133].

Dietary agents consist of a wide variety of active components that are biologically responsible for the anti-cancer effects like curcumin, genistein, resveratrol, diallyl sulfide, S-allyl cysteine, allicin, lycopene, capsaicin, diosgenin, gingerol, ellagic acid, ursolic acid, silymarin, anethol, catechins, eugenol, isothiocyanates, isoeugenol, dithiolthiones, indole-3-carbinol, isoflavones, saponins, phytosterols, inositol hexaphosphate, Vitamin C, D-limonene, lutein, folic acid, beta carotene,

selenium, Vitamin E and flavonoids (Table 3). Many of which have been used in traditional medicines for thousands of years. These dietary agents are believed to suppress inflammatory that processes lead to transformation, hyperproliferation, and initiation of carcinogenesis. Their inhibitory influences may ultimately suppress the final steps of carcinogenesis i.e angiogenesis and metastasis [134].

Table 3: Dietary sources as anticancer agent.

S. No.	Botanical Name	Source	Compound	Reference
1	Carica papaya, Family- Caricaceae	Berries	β-Cryptoxanthin	[135]
2	Glycyrrhiza glabra; Glycyrrhiza radix;	Licorice root	Glycyrrhizin	[136]
	Glycyrrhiza uralensis, Family- Leguminosae			
3	Cannabis sativa, Family- Cannabiaceae	Hemp	Cannabinol	[137]
4	Rosmarinus officinalis, Family- Lamiaceae	Rosemary	Carnosol	[138]
5	Pueraria lobata radix, Family- Fabaceae		Genistein	[139]
6	Glycine max, Family- Fabaceae	Soybeans	Genistein	[139]
7	Prunus armeniaca, Family- Rosaceae	Apricots	Carotenoids	[140]
8	Zingiber officinale, Family- Zingiberaceae	Tuber	Gingerol	[141]
9	Lycopersicon esculentum, Family- Solanaceae	Tomato	Lycopene, Lutein,	[141]
			Kaempferol	
10	Piper nigrum; Piper longum, Family- Piperaceae	Black pepper	Purpurogallin;	[142]
			Piperine	
11	Ocimum sanctum, Family-Lamiaceae	Basil	Ursolic acid	[143]
12	Betula alba, Family- Betulaceae	Birch tree	Betulinic acid	[144]
13	Crocus sativus, Family- Iridaceae	Saffron	Carotenoids	[146]
14	Silymarin marianum, Family- Asteraceae	Milk thistle	Silymarin	[147]
15	Capsaicum annum; Capsaicum frutens, Family-	Red chilli	Capsaicinoids,	[148]
	Solanaceae		Capsaicin	
16	Camellia sinensis, Family- Theaceae	Green and	Catechin and	[149]
		black teas	theaflavins	
17	Vitis vinifera, Family- Vitaceae	Grapes	Resveratrol	[150]
18	Daucus carota sativus, Family-	Carrot	β-Carotene	[151]
	Apiaceae/umbelliferae			
19	Tabebuia avellanedae, Family- Bignoniaceae	Lapacha tree	Lapachone	[31]
20	Citrus aurantium, Family- Rutaceae	Orange	Hesperidin	[152]
21	Prunus dulcis, Family- Rosaceae	Almond	Morin	[153,154]
22	Aloe arborescens, Family- Asphodelaceae	Aloe vera	Emodin	[155]
23	Opium poppy, Family- Paparveraceae	Poppy	Morphine and its analogues	[157]
24	Curcurbita moschata, Family-Cucurbitaceae	Pumpkin	β-Carotene	[158]
25	Azadirachata indica, Family- Meliaceae	Neem	Polyphenolics	[159]

Marines as source of anti-cancer agents:

Marine organisms are a rich source for natural products [160]. In recent time, advancement in deep-sea collection and aqua culture technology gives significant number of compounds derived from marine organisms entering preclinical and early clinical evaluation as potential anticancer agent [161,162]. Overall, more than 3000 new substances have been identified from marine organisms that demonstrate the great potential as a source of novel chemical classes [163]. Marine belongs to very diverse structural classes including polyketides, terpenes, steroids and peptides. The organisms yielding these bioactive marine compounds include invertebrate animals, algae, fungi and bacteria [164].

The first anticancer product didemnin B, a cyclic depsipeptide isolated from the tunicate Trididemnum solidum from marine source enter in clinical trials. Preliminary results showed a against non-Hodgkin's partial activity lymphoma [165]. It can inhibit protein synthesis and arrest G1 phase of cell-cycle. Another depsipeptide Aplidine appear to be more active as comparison with didemninB in preclinical trial and does not produce life-threatening neuromuscular toxicity. Preclinical data indicate that aplidine is active against several tumors through blockade of cell-cycle progression at phase [166]. There are number of G1 ecteinsscidins have been isolated from the marine source tunicate Ecteinascidia turbinata. One of these ecteinascidins (ET-743) was selected for clinical trials and antitumor effects have been observed in phase I studies [167]. ET-743 is a tetrahydroisoguinilone alkaloid and they acts by selective alkylation of guanine residues in the DNA minor groove [168] and also interacts with nuclear proteins [169]. In Europe and the United States ET-743 is currently in phase II clinical trials [167]. The dolastatins are a class of peptides obtained from the Indian Ocean, Dolabella auricularia. These pentides have cytotoxic activity and now a day.

dolastatin 10 and dolastatin 15 of this class have received the greatest clinical interest Dolastatin10 has entered in Phase I and Phase II clinical trials. after showing significant antitumor activity in preclinical models [170]. Its mechanism of action involves inhibition of microtubule assembly ultimately result in cellcycle arrest in metaphase [171,172]. The bryostatins, 20 macrocyclic lactones isolated from Bugula neritina and other marine bryozoa. These macrocyclic compounds have shown activity lymphocytic significant against leukemia cell line [173]. Bryostatin1 has recently entered phase II clinical trials for the of melanoma, non-Hodgkin's treatment lymphoma, renal cancer and colorectal cancer [174-176] and continues to be evaluated in phase I clinical trials. Bryostatin1 has been found to promote the normal growth of bone marrow progenitor cells, to provide in vivo protection against normally lethal doses of ionizing radiation and to serve as an immune stimulant, enhancing the normal production of interleukin2 and interferons [177].

Beside this there are the number of compounds isolated from marine as potential anti-cancer agents included in Table 4 [178,179].

Microorganisms as source of anticancer agents:

Antitumor antibiotics are among the most important cancer chemotherapeutic agents, and members of the anthracycline, include mitomycin bleomycin, actinomycin, aureolic acid families [6]. Clinically useful agents from these above families are the daunomycin and related agents like doxorubicin, idarubicin and epirubicin; the peptolides (exemplified by dactinomycin), the mitosanes (such as mitomycin C) and the glycosylated anthracenone mithramycin. The anthracyclines are among the most used antitumor antibiotics in the clinic and exert antitumor activity mainly by inhibiting topoisomerase II [180,181].

Table 4: Marine derived potential anticancer agent.

S.No.	Compound	Organism	Chemistry	Mechanism of action
1.	Aaptamine	Sponge	Alkaloid	Induction of p21 and G2/M cell cycle
				arrest
2.	Cortistatin A	Sponge	Alkaloid	Selective inhibiton of angiogensis
3.	Aplidine	Ascidian	Depsipeptide	Oxidation and inactivation of low
				molecular weight-protein tyrosine
				phosphatase activity
4.	Bastadine 6	Sponge	Alkaloid	Inhibition of angiogenesis in vitro and
				in vivo involves apoptosis
5.	Fucoxanthinol	Ascidian	Carotenoid	Induction of apoptosis
6.	Lamellarin D	Mollusk	Alkaloid	ErbB3 protein and PI3K- Akt pathway
				involved in necrosis induction
7.	Clavulone II	Soft coral	Prostanoid	G1 cell cycle arrest and apoptosis
8.	Geodiamolides	Sponge	Peptide	Disorganization of actin filaments
9.	Ircinin-1	Sponge	Sesterterpene	G1 phase inhibition and apoptosis
				induction
10.	Laxaphycins A and	Bacterium	Cyclic peptides	Increased polyploidy by putative
	В			topoisomerase II alterations
11.	Leptosins C and F	Fungus	Alkaloid	DNA topoisomerase I and II inhibition
				and apoptosis induction
12.	Onnamide A	Sponge	Polyketide	Protein synthesis inhibition
13.	Philinopside A	Sea cucumber	Saponin	Inhibition of angiogenesis and
				receptor tyrosine kinases
14.	Variolin B	Sponge	Alkaloid	Inhibition of cyclin-dependent kinases
				and apoptosis induction
15.	Aplidine	Ascidian	Depsipeptide	Induction of apoptosis with
				concomitant G1 arrest and G2
				blockage
16.	Ascididemin	Ascidian	Alkaloid	Direct iminoquinone reduction and
				reactive oxygen species generation
17.	Cammbrescidin	Sponge	Alkaloid	Induction of eythroid differentiation
	800			and cell cycle arrest
18.	Dideoxypetrosynol	Sponge	Fatty acid	Induction of apoptosis via
	A			mitochondrial signaling pathway
19.	Dolastatin 10	Mollusc	Peptide	Binds to amino-terminal peptide of β-
				tubulin containing cysteine
20.	Girolline	Sponge	Alkaloid	Induction of G2/M cell cycle arrest
				and p53 proteasome recruitment
21.	Halichondrin B	Sponge	Macrolide	Induction of mitotic blockage and
	analogues		derivative	apoptosis
	Lissoclinolide	Ascidian	Fatty acid	G2/M cell cycle arrest
22.	Lissociiionac			
22. 23.	Neoamphimedine	Sponge	Alkaloid	Induction of topoisomerase II α-

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24.	Psammaplin A	Sponge	Alkaloid	Inhibition of aminopeptidase N and suppression of angiogenesis in vitro
25.	Alkylpyridinium	Sponge	Alkaloid	Induction of apoptosis and reduced cell adhesion
26.	Aeroplysinin	Sponge	Alkaloid	Induction of apoptosis on proliferating endothelial cells
27.	Bryostatin-1	Bryozoan	Macrolide	Potentiation of ara-C induced apoptosis by PKC-dependent release of TNF-α
28.	Cephaiostatin	Worm	Steroid	Apoptosis and increased mitochondrial matrix density
29.	Chondropsin A	Sponge	Macrolide	In Vitro inhibition of V-ATPase enzyme
30.	Dehydrothrysiferol	Alga	Triterpene	Enhanced apoptosis induction in estrogen receptor negative breast cancer cells
31.	Diazonamide-A	Ascidian	Peptide	Disruption of mitosis and cellular microtubules with inhibition of GTP hydrolysis
32.	Dictyostatin	Sponge	Polyketide	Induction of tubulin polymerization
33.	Dolastatin 11	Mollusc	Peptide	F-actin stabilization by connection between two long-pitch strands
34.	Ecteinascidin- 743	Ascidian	Isoquinoline alkaloid	Telomere dysfunction increases susceptibility to ET-743
35.	GA3 polysaccharide	Alga	Polysaccharide	Inhibition of topoisomerase I and II
36.	Hemiasterlin analogue	Sponge	Tripeptide	Induction of microtubule depolymerisation
37.	Kahalalide F	Mollusc	Depsipeptide	Potent cytotoxicity and induction of necrosis
38.	Lamellarin D	Mollusc	Alkaloid	Potent inhibition of topoisomerase I
39.	omega-3 fatty acids	Fish	Fatty acid	

Many pharmaceutical agents have been discovered by screening natural products from a wide range of microorganisms. Rapamycin and its analogs are products of Streptomyces hygroscopicus have potent immunosuppressive activity. They inhibit signaling pathways required for T-cell activation and proliferation.

Rapamycin blocks progression of the cell cycle at middle-to-late G1 phase in T cells and B cells, and osteosarcoma and rhabdomyosarcoma cell lines, among others [182]. Geldanamycin is a benzoquinone ansamycin natural fermentation product and inhibits heat-shock protein HSP 90 [183].

Table 5: Microorganism derived anti-cancer agents.

S.No.	Compound	Microorganism	Used in Cancer
1.	Actinomycin	Streptomyces spp.	Sarcoma and germ-cell tumors
2.	Bleomycin	Streptomyces verticillus	Germ-cell, cervix and head and neck
			cancer
3.	Daunomycin	Streptomyces coeruleorubidus	Leukemia
4.	Doxorubicin	Streptomyces Pneuceticus	Lymphoma, breast, ovary, lung and
			sarcomas
5.	Epirubicin	Streptomyces pneuceticus	Breast cancer
6.	Idarubicin	Streptomyces Pneuceticus	Breast cancer and leukemia
7.	Mitomycin C	Streptomyces caespitosus	Gastric, colorectal, anal and lung
			cancer
8.	Geldanamycin	Streptomyces Hygroscopicus	Experimental
9.	Rapamicin	Streptomyces hygroscopicus	Experimental
10.	Wortamannin	Talaromyces wortmanni	Experimental

Wortmannin is a product of the fungus Talaromyces wortmanni and inhibits signal transduction pathways by forming a covalent complex with an active-site residue of phosphoinositide 3 kinase (PI3K), inhibiting PI3K activity [184] (Table 5). Thus, toxins that evolved originally to kill competing micoorganisms can have a variety of physiological effects in animals. In many cases, the targets of these compounds are components of signal transduction cascades that are conserved in many species, and that have been considered novel targets for anticancer drug discovery [185].

Conclusion:

Natural products have been a prime source for the treatment of many forms of cancer, many of which are consumed daily with the diet. They provide significant protection against various cancers and many other diseases. The antioxidant medicinal plants and their products prevent from the cancer and other diseases by protecting cells from damage. Thus, consuming a diet rich in antioxidant fruits, vegetables, herbs etc. will provide health-protective effects. Microbes and marine organisms also have been offering the great role in the prevention and treatment of cancer. All the natural products discussed in this review exhibit anticancer

activities. Natural products offer a great opportunity to evaluate not only totally new chemical classes of anticancer agents, but also novel and potentially relevant mechanisms of action

References

- 1. Hail Jr N. Mitochondria: a novel target for the chemoprevention of cancer. Apoptosis. 2005;10:687–705.
- 2. Aggarwal BB, Danda D, Gupta S, Gehlot P. Models for prevention and treatment of cancer: Problems vs promises. Biochem Pharmacol. 2009;78:1083–1094.
- 3. Farnsworth NR. Ethnobotany and the Search for New Drugs, Wiley Willett WC. Diet and health: what should we eat. Science. 1994;264(5158):532–7.
- 4. Duraipandiyan V, Ayyanar M, Ignacimuthu S. Antimicrobial activity of some ethnomedicinal plants used by Paliyar tribe from Tamil Nadu, India. BMC Comp Alter Med. 2006;6:35-41.
- 5. Cragg GM, Kingston DGI, Newman DJ. Anticancer Agents from Natural Products. Taylor & Francis Group, Boca Raton, FL. Brunner-Routledge Psychology Press; 2005.
- 6. Cragg GM, Newman D J, Weiss RB. Coral reefs, forests, and thermal vents: the worldwide exploration of nature for novel antitumor agents. Semin Oncol. 1997;24:156-163.

- 7. Newman DJ, Cragg GM. Natural products as sources of new drugs over the last 25 years. J Nat Prod. 2007;70:461–477.
- 8. Butler MS. Natural products to drugs: natural product derived compounds in clinical trials. Nat Prod Rep. 2005;22:162–195.
- 9. Bailly C. Ready for a comeback of natural products in oncology. Biochem Pharmacol. 2009;77:1447–1457.
- 10. Newman DJ, Cragg GM, Snader KM. Natural products as sources of new drugs over the period 1981–2002. J Nat Prod. 2003;66: 1022–1037.
- 11. Butler MS. The role of natural product chemistry in drug discovery. J Nat Prod. 2004;67:2141–2153.
- 12. DeVita VT Jr, Serpick AA, Carbone PO. Combination chemotherapy in the treatment of advanced Hodgkin's disease. Ann Intern Med. 1970;73:881-895.
- 13. Noble RL. The discovery of the vinca alkaloids chemotherapeutic agents against cancer. Biochem Cell Biol. 1990;68:1344-1351.
- 14. Stähelin H. Actvity of a new glycosidic lignan derivative (VP-16-213) related to podophyllotoxin in experimental tumors. Eur J Cancer. 1973;9:215-221.
- 15. Harvey AL. Medicines from nature: are natural products still relevant to drug discovery. Trends Pharmacol Sci. 1999;20:196-198.
- 16. Liu LF. DNA topoisomerase poisons as antitumor drugs. Annu Rev Biochem. 1989;58:351-375.
- 17. Wani MC, Taylor HL, Wall ME, et al. Plant antitumor agents. VI. The isolation and structure of taxol, a novel antileukemic and antitumor agent from Taxus brevifolia. J Am Chem Soc. 1971;93:2325-2327.
- 18. Creemers GJ, Bolis G, Gore M, et al. Topotecan, an active drug in the second-line treatment of epithelial ovarian cancer. J Clin Oncol. 1996;14:3056-3061.
- 19. Bertino JR. Irinotecan for colorectal cancer. Semin Oncol. 1997;24:S18-S23.
- 20. Liu LF, Desai SD, Li TK, et al. Mechanism of action of camptothecin. Ann New York Acad Sci. 2000;922:1-10.
- 21. Harmon AD, Weiss U, Silverton JV. The structure of rohutukine, the main alkaloid of

- Amoora rohituka (syn. Aphanamixis polystachya) (Maliaceae). Tetrahydron. 1979;20:721-724.
- 22. Cragg G, Suffness M. Metabolism of plant-derived anticancer agents. Pharmacol Ther. 1988;37:425-432.
- 23. Losiewicz MD, Carlson BA, Kaur G, et al. Potent inhibition of cdc2 kinase activity by the flavonoid L86-8275. Biochem Biophys Res Commun. 1994;201:589-595.
- 24. Worland PJ, Kaur G, Stetler-Stevenson M, et al. Alteration of the phosphorylation state of p32cdc2 kinase by the flavone L86-8275 in breast carcinoma cells. Biochem Pharmacol. 1993;46:1831-1836.
- 25. Kelland LR. Flavopiridol, the first cyclindependent kinase inhibitor to enter the clinic: current status. Ex Opin Inv Drugs. 2000;9:2903-2911.
- 26. Powell RG, Weisleder D, Smith CR Jr, et al. Structures of harringtonine, isoharringtonine, and homoharringtonine. Tetrahydron Lett. 1970;11:815-818.
- 27. Zhou DC, Zittoun R, Marie JP. Homoharringtonine: an effective new natural product in cancer chemotherapy. Bull Cancer. 1995;82:987-995.
- 28. Kantarjian HM, O'Brien S, Anderlini P, et al. Treatment of myelogenous leukemia: current status and investigational options. Blood. 1996;87:3069-3081.
- 29. Rowinsky EK, Noe DA, Ettinger DS, et al. PhaseI and pharmacological study of the pulmonary cytotoxin 4-ipomeanol on a single dose schedule in lung cancer patients: hepatotoxicity is dose limiting in humans. Cancer Res. 1993;53:1794-1801.
- Rehm S, Devor DE. Acute effects of 4ipomeanol on experimental lung tumors with bronchiolar or alveolar cell features in Syrian hamsters or C3H/HeNCr mice. J Cancer Res Clin Oncol. 1993;120:41-50.
- Li YZ, Li CJ, Pinto AV, et al. Release of mitochondrial cytochrome c in both apoptosis and necrosis induced by βlapachone in human carcinoma cells. Mol Med. 1999;4:232-239.
- 32. Vukovic-Gacic B, Nikcevic S, Beric-Bjedov T, et al. Antimutagenic effect of essential oil of sage (Salvia officinalis L.) and its monoterpenes against UV-induced mutations

- in Escherichia coli and Saccharomyces cerevisiae. Food Chem Toxicol. 2006;44:1730–1738.
- 33. ÖnayUçar E, Karagöz A, Arda N. Antioxidant activity of Viscum album sp. Fitoterapia. 2006;77: 556–560.
- 34. Pinney KG, Jelinek C, Edvardsen K, et al. The discovery and development of the combretastatins. In: Cragg GM, Kingston DGI, Newman DJ. Anticancer Agents from Natural Products. Taylor & Francis Group, Boca Raton, FL Brunner-Routledge Psychology Press; 2005:23–46 (Chapter 3).
- 35. Evandri MG, Battinelli L, Daniele C, et al. The antimutagenic activity of Lavandula angustifolia (lavender) essential oil in the bacterial reverse mutation assay. Food Chem Toxicol. 2005;43:1381–1387.
- 36. Hwang BY, Su BN, Chai H, et al. Silvestrol and episilvestrol, potential anticancer rocaglate derivatives from Aglaia silvestris. J Org Chem. 2004;69(10):3350-3358.
- 37. Cassady JM, Chan KK, Floss HG, et al. Recent developments in the Maytansanoid antitumor agents. Chem Pharma Bulletin. 2004;52:1–26.
- 38. Suffness M, Douros J. Miscellaneous natural products with antitumor activity. In: Cassady JM, Douros JD. Anticancer Agents Based on Natural Product Models. New York, Academic Press; 1980:474 (Chapter 14).
- 39. Ravelo AG, Estevez-Braun A, Chavez-Orellana H, et al. Recent studies on natural products as anticancer agents. Curr Topics Med Chem. 2004;4:241–265.
- 40. Jang DS, Park EJ, Hawthorne ME, et al. Potential cancer chemopreventive constituents of the seeds of Dipteryx odorata (tonka bean). J Nat Prod. 2003;66(5):583–587.
- 41. Denmeade SR, Jakobsen CM, Janssen S, et al. Prostate-specific antigen-activated thapsigargin prodrug as targeted therapy for prostate cancer. J Natl Cancer Inst. 2003;95:990–1000.
- 42. Newman DJ, Cragg GM, Holbeck S, et al. Natural products as leads to cell cycle pathway targets in cancer chemotherapy. Curr Cancer Drug Targets. 2002;2:279–308.
- 43. Hernandez-Ceruelos A, Madrigal-Bujaidar E, de la Cruz C. Inhibitory effect of chamomile

- essential oil on the sister chromatid exchanges induced by daunorubicin and methyl methanesulfonate in mouse bone marrow. Toxicol Lett. 2002;135:103–110.
- 44. Silva GL, Cui B, Chavez D, et al. Modulation of the multidrug-resistance phenotype by new tropane alkaloid aromatic esters from Erythroxylum pervillei. J Nat Prod. 2001;64(12):1514–1520.
- 45. Lee D, Bhat KPL, Fong HHS, et al. Aromatase inhibitors from Broussonetia papyrifera. J Nat Prod. 2001;64(10):1286–1293
- 46. Marnewick JL, Gelderblom WCA, Joubert E. An investigation on the antimutagenic properties of South African herbal teas. Mutat Res. 2000;471:157–166.
- 47. Abdulla M, Gruber P. Role of diet modification in cancer prevention. Biofactors. 2000;12:45–51.
- 48. Dinkova-Kostova AT, Talalay P. Persuasive evidence that quinine reductase type 1 (DT diaphorase) protects cells against the toxicity of electrophiles and reactive forms of oxygen. Free Radic Biol Med. 2000;29(3–4): 231–240.
- 49. Sausville EA, Zaharevitz D, Gussio Z, et al. Cyclin-dependent kinases: initial approaches to exploit a novel therapeutic target. Pharmacol Thera. 1999;82:285.
- 50. He K, Valcic S, Timmerman BN, et al. Indole alkaloids from Aristotelia chilensis (Mol.) Stuntz. Int J Pharmacog. 1997;35:215–217.
- 51. Khamis SB, Brown JE, Bibby MC, et al. Evaluation of two plants species used in Malay traditional medicine for the treatment of breast cancer. J Pharma Pharmacol. 1997;49:113.
- 52. Liang HR, Vuorela P, Vuorela H, et al. Isolation and immunomodulatory effect of flavonol glycosides from Epimedium hunanense. Planta Medica. 1997;63:316 319.
- 53. Peres MTLP, Monache FD, Cruz AB, et al. Chemical composition and antimicrobial activity of Croton urucurana Baillon (Euphorbiaceae). J Ethnopharmacol. 1997;56:223 226.

- 54. Barakat HH, Hussein SAM, Marzour MS, et al. Polyphenolic metabolites of Epilobium hersutum. Phytochemistry. 1997;46:935–941.
- 55. Bai L, Yamaki M, Takagi S. Lignans and a bichroman from Pleione bulbocodioides. Phytochemistry. 1997;44:341–343.
- 56. Jang M, Cai L, Udeani GO, et al. Cancer chemopreventive activity of resveratrol, a natural product derived from grapes. Science. 1997;275(5297):218–220.
- 57. Gupta MP, Monge A, Karikas GA, et al. Screening of Panamanian medicinal plants for brine shrimp toxicity, crown gall tumor inhibition, cytotoxicity and DNA intercalation. Int J Pharmacog. 1996;34:19 27
- 58. Suh HW, Song DK, Son KH, et al. Antinociceptive effect of Smilaxin B administered intracerebroventricularly in the mouse. Planta Med. 1996;62:141 145.
- Shirota O, Tamemura T, Morita H, et al. Triterpenes from Brazilian medicinal plant "Chuchuhuasi" (Maytenus krukobii). J Nat Prod. 1996;59:1072–1075.
- 60. Tsui WY, Brown GD. Cycloeudesmanolides from Sarcandra glabra. Phytochemistry. 1996;43:819–821.
- 61. Um SH, Lee KR, Zee OP, Pyo S. The effect of Salvia plebeia on murine macrophage-mediated cytotoxicity. Nat Prod Sci. 1996;2: 43 47.
- 62. Ducki S, Hadfield JA, Lawrence CY, et al. Isolation of E-1-(4'-hydroxyphenyl)-But-1-en-3-one from Scutellaria barbata. Planta Med. 1996;62:185–186.
- 63. Palomino E, Maldonado C, Kempff MB, et al. Caparratriene, an active sesquiterpene hydrocarbon from Ocotea caparrapi. J Nat Prod. 1996;59:77 79.
- 64. Singh VK, Ali ZA, Zaidi STH, et al. Ethnomedicinal uses of plants of Gonda district forests of Uttar Pradesh, India. Fitoterapia. 1996;67:129 139.
- 65. Flores JS, Ricalde RV. The secretions and exudates of plants used in mayan traditional medicine. J Herbs Spices Med Plants. 1996;4:53–59.
- 66. Ahn BT, Oh KJ, Ro JS, Lee KS. A new flavonoid from Euphorbia ebracteolata. Planta Med. 1996;62:383–384.

- 67. Hu K, Dong A, Yao XS, et al. Antineoplastic agents I. Three spirostanol glycosides from rhizomes of Dioscorea collettii var. hypoglauca. Planta Med. 1996;62:573 575.
- 68. Joe YK., Son JK, Park SH, et al. New naphthalenyl glucosides from the roots of Juglans mandshurica. J Nat Prod. 1996;59:159 160.
- 69. Zeng, JF, Li GL, Zhu DY. Two isoprenoid-substituted isoflavans from roots of Maackia tenuifolia. Phytochemistry. 1996;43:893 896.
- 70. Martinez-Lirola MJ, Gonzalez-Tejero MR, Molero-Mesa J. Ethnobotanical resources in the province of Almeria, Spain, Campos De Nijar. Econ Bot. 1996;50:40 56.
- 71. Peng JN, Feng XZ, Liang XT. A new Iridoid from Hedyotis crysotrichae. Chin Chem Lett. 1995;6:965–966.
- 72. Ducki S, Hadfield JA, Lawrence NJ, et al. Isolation of paeonol from Arisaema eubescens. Planta Med. 1995;61:586–587.
- 73. Aquino R, Pizza C, De Tommasi N, et al. New polyoxypregnane ester derivatives from Leptadenia hastata. J Nat Prod. 1995;58:67.
- 74. Tiwari VJ. Ethnobotanical survey of Halbi tribe of Chandrapur and Gadchiroli districts of Maharashtra State, India. Fitoterapia. 1995;66:346 350.
- 75. Rabi T, Gupta RC. Antitumor and cytotoxic investigation of Amoora rohituka. Int J Pharmacog. 1995;33:359–361.
- 76. Ho LK, Ou JC, Sun ML, Sun CM. Two rare alkaloids from Pratia nummularia. Planta Med. 1995;61:567–568.
- 77. Rivera D, Obon C. The Ethnopharmacology of Madeira and Porto Santo islands, a review. J Ethnopharmacol. 1995;46:73–93.
- 78. Shirota O, Morita H, Takeya K, Itokawa H. Structures of xuxuarines, stereoisomeric triterpene dimmers from Maytenus chuchuhuasca. Tetrahedron. 1995;51:1107–1120.
- 79. Fujita T, Sezik E, Tabata M, et al. Traditional medicine in Turkey VII. Folk medicine in middle and west Black sea regions. Econ Bot. 1995;49:406–422.
- 80. Shi JG, Jia ZJ, Cui YX. Novel Tricyclic Diterpenoids from Euphorbia micractina. J Nat Prod. 1995;58:51–56.

- 81. Wu DG, Sorg B, Hecker E. New myrsinolrelated polyfunctional pentacyclic diterpene esters from roots of Euphorbia prolifera. J Nat Prod. 1995;58:408–413.
- 82. Gonzalez-Tejero MR, Molero-Mesa J, Casares-Porcel M, et al. New Contributions to the ethnopharmacology of Spain. J Ethnopharmacol. 1995;45:157–163.
- 83. Ahmad FB, Holdsworth DK. Traditional medicinal plants of Sabah, Malaysia part III. The Rungus people of Kudat. Int J Pharmacog. 1995;33:262–264.
- 84. Amusan OOG, Msonthi JD, Makhubu LP. Molluscicidal activity of Spathodea campanulata, Andrachneovalis, Phytolacca dodecandra and Hypoxis rooperi. Fitoterapia. 1995;66:113–116.
- 85. Zani CL, Chaves PPG, Queiroz R, et al. Brine shrimp lethality assay as a prescreening system for anti-trypanosoma cruzi activity. Phytomedicine. 1995;2:47–50.
- 86. Pisha E, Chai H, Lee IS, et al. Discovery of betulinic acid as a selective inhibitor of human melanoma that functions by induction of apoptosis. Nat Med. 1995;1(10):1046–1051.
- 87. Laferriere JE. Medicinal plants of the lowland Inga people of Colombia. Int J Pharmacog. 1994;32:90–4.
- 88. Lin LZ, Hu SF, Zaw K, et al. Thalifaberidine, a cytotoxic aporphine-benzylisoquinoline alkaloid from Thalictrum faberi. J Nat Prod. 1994;57:1430–1436.
- 89. Bae K H, Min BS, Park KI, Ahn BZ. Cytotoxic flavonoids from Scutellaria indica. Planta Med. 1994;60:280–281.
- 90. Ishiguro K, Nagata S, Fukumoto H, et al. A 2-pyrone derivative from Hypericum japonicum. Phytochemistry. 1994;37:283–284.
- 91. Chiang HC, Juilo Y, Lu FJ. Xanthine oxidase inhibitors from the leaves of Alsophila spinulosa (Hook) Tryon. J Enzyme Inhib. 1994;8:61–71.
- 92. Wu JB, Cheng YD, Kuo SC, et al. Fissoldhimine, a novel skeleton alkaloid from Fissistigma oldhamii. Chem Pharm Bulletin. 1994;42:2202–2204.
- 93. Pinto DCG, Fuzzati N, Pazmino XC, Hostettmann K. Xanthone and antifungal

- constituents from Monnina obtusifolia. Phytochemistry. 1994;37:875–878.
- 94. Liu WK, Ng TG, Sze SF, Tsui KW. Activation of peritoneal macrophages by polysaccharopeptide from the mushroom, Coriolus 6ersicolor. Immunopharmacology. 1993;26:139–146.
- 95. Mohandoss S, Ravindran P. Flavonoids from Melastoma melabathricum. Fitoterapia. 1993;64:277–278.
- 96. Hammer M L A, Johns EA. Tapping an Amazonian plethora, four medicinal plants of Marajo Island, Para (Brazil). J Ethnopharmacol. 1993;40:53-75.
- 97. Segura-Correa R, Mata R, Anaya AL, et al. New tetranortriterpenoids from Swietenia humilis. J Nat Prod. 1993;56:1567–1574.
- 98. Ghazanfar SA, Al-Sabahi MA. Medicinal plants of Northern and Central Oman (Arabia). Econ Bot. 1993;47:89–98.
- 99. Cai Y, Chen JP, Phllipson J. Diterpenes from Croton lechleri. Phytochemistry. 1993;32:755–760.
- 100. Hussain SJ, Alvi AB, Jahan M. A study on Unani medicinal plants, Asteratiqus. J Res Educ Indian Med. 1993;2:35–39.
- 101. Lin CC, Lin CH. Pharmacological and pathological studies on Taiwan folk medicine. IX. The hepatoprotective effect of the methanolic extract from Echinops grijisii. Am J Chin Med. 1993;21:33–44.
- 102. Omino EA, Kokwaro JO. Ethnobotany of Apocynaceae species in Kenya. J Ethnopharmacol. 1993;40:167–180.
- 103. Rehm S, Devor DE. Acute effects of 4-ipomeanol on experimental lung tumors with bronchiolar or alveolar cell features in Syrian hamsters or C3H/HeNCr mice. J. Cancer Res Clin Oncol. 1993;120:41-50.
- 104. Stuppner H, Sturm S, Geisen G, et al. A differential sensitivity of oxindole alkaloids to normal and leukemic cell lines. Planta Med. 1993;supplement A:583.
- 105. Holdsworth DK. A preliminary study of medicinal plants of Easter Island, South Pacific. Int J Pharmacog. 1992;30:27–32.
- 106. Sun HD, Zhou QZ, Fujita T, et al. Rubescensin D, a diterpenoid from Rabdosia rubescens. Phytochemistry. 1992;31:1418–1419.

- 107. Qian TX, Li LN. Isosalvianolic acid C, a depside possessing a dibenzooxepin skeleton. Phytochemistry. 1992;31:1068–1070.
- 108. Furusawa E, Chou SC, Furusawa S, et al. Antitumour activity of Ganoderma lucidum, an edible mushroom, on intraperitoneally implanted lewis lung carcinoma in synergenic mice. Phytother Res. 1992;6:300 304.
- 109. Ding YL, Jia ZA. Two phenolic derivatives from Euphorbia kansui. Phytochemistry. 1992;31:1435 1436.
- 110. Lin CC, Lin CH, Chiu HF, et al. The Pharmacological and pathological studies on Taiwan folk medicine (VII). The anti-inflammatory effect of Echinops grjiisii. Am J Chin Med. 1992;22:127 134.
- 111. De Feo V, Aquino R, Menghini A, et al. Traditional phytotherapy in the peninsula Sorrentina, Campania, Southern Italy. J Ethnopharmacol. 1992;36:113 125.
- 112. Wong BYY, Lau BHS, Teel RW. Chinese medicinal herbs modulated mutagenesis, DNA binding and metabolism of Benzo[A]Pyrene. Phytother Res. 1992;6:10 14.
- 113. Hua YY. Two new saponins from the roots of Phytolacca esculenta. Planta Med. 1992;58:99 101.
- 114. Sakagami H, Konno K, Kawazoe Y, et al. Multiple immunological functions of extract from the cone of Japanese white pine, Pinus par6iflora Sieb. et Zucc. Adv Exp Med Biol. 1992;319:331 335.
- 115. Kao WJ, Hung DZ, Tsai WJ, et al. Podophyllotoxin intoxication. Toxic effect of Bajiaolian in herbal therapeutics. Hum Exp Toxicol. 1992;11:480 487.
- 116. Lee IR, Song JY, Lee YS. Cytotoxicity of folkloric medicines in murine and human cancer cells. Kor J Pharmacog. 1992;23:132 – 136.
- 117. Reddy MB, Reddy KR, Reddy MN. Ethnobotany of Cuddapah district, Andhra Pradesh, India. Int J Pharmacog. 1991;29:273 280.
- 118. Chhabra SC, Mahunnah RLA, Mshiu MN. Plants used in traditional medicine in Eastern Tanzania. V. Angiosperms (Passifloraceae to Sapindaceae). J Ethnopharmacol. 1991;33:143 157.

- 119. Holdsworth DK. Traditional medicinal plants of Rarotonga, Cook Islands. Part II. Int J Pharmacog. 1991;29:71–79.
- 120. Kijjoa A, Gonzalez MJTG, Pinto MMM, et al. Constituents of Knema laurina and Knema tenuiner6ia subsp. setosa. Planta Med. 1991;57:575–577.
- 121. Sati OP, Bahuguna S, Uniyal S, et al. A new saponin from Deeringia amaranthoides. J Nat Prod. 1990;53:466–469.
- 122. Konda Y, Iguchi M, Harigaya Y, et al. Hancokinol, a novel triterpene, from Cynanchum hancokianum. Tetrahedron Lett. 1990;31:5315 5318.
- 123. Bhat RB, Eterjere EO, Oladipo VT. Ethnobotanical studies from Central Nigeria. Econ Bot. 1990;44:382 390.
- 124. Plotkin MJ, Schultes RE. Virola, a promising genus for ethnopharmacological investigation. J Psychoactive Drugs. 1990;22:357–361.
- 125. Novaretti R, Lemordant D. Plants in the traditional medicine of the Ubaye valley. J Ethnopharmacol. 1990;30:1–34.
- 126. Uniyal MR. Utility of hitherto unknown medicinal plants traditionally used in Ladakh. J Res Educ Indian Med. 1990;9:89 95
- 127. Vinesi P, Serafini M, Nicoletti M, et al. Plant regeneration and hypoxoside content in Hypoxis obtusa. J Nat Prod. 1990;53:196 199.
- 128. Chu DT, Wong WL, Mavlight GM. Immunotherapy with Chinese medicinal herbs: immune restoration of local xenogenic graft-versus-host reaction in cancer patients by fractionated Astragalus membranacueus in vitro. J Clin Lab Immunol. 1988;25(3):19-23.
- 129. Kokwaro JO. Medicinal plants of east Africa. East African Literature Bureau, Nairobi;1976.
- 130. Powell RG, Weisleder D, Smith CRJr, et al. Structures of harringtonine, isoharringtonine, and homoharringtonine. Tetrahydron Lett. 1970;11: 815-818.
- 131. Block G, Patterson B, Subar A. Fruit, vegetables, and cancer prevention: a review of the epidemiological evidence. Nutr Cancer. 1992;18(1):1–29.

- 132. Steinmetz KA, Potter JD. Vegetables, fruit, and cancer prevention: a review. J Am Diet Assoc. 1996;96(10):1027–39.
- 133. Reddy L, Odhav B, Bhoola KD. Natural products for cancer prevention: a global perspective. Pharmacol Ther. 2003;99(1):1–13.
- 134. Aggarwal BB, Shishodia S. Molecular targets of dietary agents for prevention and therapy of cancer. Biochem Pharmacol. 2006;71:1397–1421.
- 135. Hoyoku N, Michiaki M, Harukuni T, et al. Cancer prevention by carotenoids. Arch Biochem Biophys. 2009;483:(2):165-168.
- 136. Wan X, Luo M, Li X, et al. Hepatoprotective and anti-hepatocarcinogenic effects of glycyrrhizin and matrine. Chem Biol Interact. 2009;181:(1)15-19.
- 137. Caihua L, Michael DMc, Carmen M, et al. A Population-Based Case-Control Study of Marijuana Use and Head and Neck Squamous Cell Carcinoma. Cancer Prev Res. 2009;2(8):759-768.
- 138. Shiu-Chen H, Chi-Tang H, Shoei-Yn L, et al. Carnosol inhibits the invasion of B16/F10 mouse melanoma cells by suppressing metalloproteinase-9 through down-regulating nuclear factor-kappa B and c-Jun. Biochem Pharmacol. 2005;69(2):221-232.
- 139. Li M, Zhang Z, Hill DL, et al. Genistein, a dietary isoflavone, down-regulates the MDM2 oncogene at both transcriptional and posttranslational levels. Cancer Res. 2005;65(18):8200–8.
- 140. Ruiz D, Egea J, Tomas-Barberan FA, et al. Carotenoids from new apricot (Prunus armeniaca L.) varieties and their relationship with flesh and skin color. J Agric Food Chem. 2005;53:6368-74.
- 141. Kim GY, Kim JH, Ahn SC, et al. Lycopene suppresses the lipopolysaccharide-induced phenotypic and functional maturation of murine dendritic cells through inhibition of mitogen-activated protein kinases and nuclear factor-kappaB. Immunology. 2004;113(2):203–11.
- 142. Sunila ES, Kuttan G. Immunomodulatory and antitumor activity of Piper longum Linn. and piperine. J Ethnopharmacol. 2004;90:(2-3):339-346.

- 143. Shishodia S, Majumdar S, Banerjee S, et al. Ursolic acid inhibits nuclear factor-kappaB activation induced by carcinogenic agents through suppression of IkappaBalpha kinase and p65 phosphorylation: correlation with downregulation of cyclooxygenase 2, matrix metalloproteinase 9, and cyclin D1. Cancer Res. 2003;63(15):4375–83.
- 144. Takada Y, Aggarwal BB. Betulinic acid suppresses carcinogen-induced NF-kappa B activation through inhibition of I kappa B alpha kinase and p65 phosphorylation: abrogation of cyclooxygenase-2 and matrix metalloprotease-9. J Immunol. 2003;171(6):3278–86.
- 145. Cheng YH., Shen TF, Pang VF, et al. Effects of aflatoxin and carotenoids on growth performance and immune response in mule ducklings. Comp Biochem Physiol Toxicol Pharmacol. 1999;128:19–26.
- 146. Abdullaev FI. Cancer chemopreventive and tumoricidal properties of saffron (Crocus sativus L.). Exp Biol Med. 2002;227:20-5.
- 147. Kohno H, Marda M, Honjo S. Prevention of colonic preneoplastic lesions by the beta-cryptoxanthine and hesperidine rich powder prepared from Citrus Unshiu Marc. Juice in male F344 rats. J Toxicol Pathol. 1999;12:209-215.
- 148. Han SS, Keum YS, Seo HJ, et al. Capsaicin suppresses phorbol ester-induced activation of NF-kappaB/Rel and AP-1 transcription factors in mouse epidermis. Cancer Lett. 2001;164(2):119–26.
- 149. Yang F, Oz HS, Barve S, et al. The green tea polyphenol (-)-epigallocatechin-3-gallate blocks nuclear factor-kappa B activation by inhibiting I kappa B kinase activity in the intestinal epithelial cell line IEC-6. Mol Pharmacol. 2001;60(3):528–33.
- 150. Manna SK, Mukhopadhyay A, Aggarwal BB. Resveratrol suppresses TNF-induced activation of nuclear transcription factors NF-kappa B, activator protein-1, and apoptosis: potential role of reactive oxygen intermediates and lipid peroxidation. J Immunol. 2000;164(12):6509–19.
- 151. Cheng YH, Shen TF, Pang VF, et al. Effects of aflatoxin and carotenoids on growth performance and immune response in mule

- ducklings. Comp Biochem Physiol Toxicol Pharmacol. 1999;128:19–26.
- 152. Berkarda B, Koyuncu H, Soybir GT, et al. Inhibitory effect of hesperidin on tumor initiation and promotion in mouse skin. Res Exp Med. 1998;198:93-99.
- 153. Laughton MJ, Evans PJ, Moroney MA, et al. Inhibition of mammalian 5-lipoxygenase and cyclooxygenase by flavonoids and phenolic dietary additives. Relationship to antioxidant activity and to iron ionreducing ability. Biochem Pharmacol. 1991;42(9):1673–81.
- 154. Soliman KF, Mazzio EA. In vitro attenuation of nitric oxide production in C6 astrocyte cell culture by various dietary compounds. Proc Soc Exp Biol Med. 1998;218(4):390–7.
- 155. Kumar A, Dhawan S, Aggarwal BB. Emodin (3-methyl-1,6,8-trihydroxyanthraquinone) inhibits TNF-induced NFkappaB activation, IkappaB degradation, and expression of cell surface adhesion proteins in human vascular endothelial cells. Oncogene. 1998;17(7):913–8.
- 156. Geng Z, Rong Y, Lau BH. S-Allyl cysteine inhibits activation of nuclear factor kappa B in human T cells. Free Radic Biol Med. 1997;23(2):345–50.
- 157. Sueoka N, Sueoka E, Okabe S, et al. Anticancer effects of morphine through inhibition of tumour necrosis factor-a release and mRNA expression. Carcinogenesis. 1996;17(11):2337-2341.
- 158. Kune GA, Bannerman S, Field B, et al. Diet, smoking, serum beta-carotene and vitamin A in male nonmelanocytic skin cancer patients and controls. Nutr Cancer. 1992;18(3):237.
- 159. Gogate SS. Cytotoxicity of neem leaf extract: an antitumor. Natl Med J India. 1991;9:297.
- 160. Pomponi AS. The bioprocess-technological potential of the sea. J Biotechnol. 1999;70:5-13.
- 161. Schwartsmann G. Marine organisms and other novel natural sources of new anticancer drugs. Ann Oncol. 2000;11:235-243.
- 162. Schwartsmann G, Rocha AB, Berlinck R, et al. Marine organisms as a source of new anticancer agents. Lancet Oncol. 2001;2:221-225.
- 163. Schweitzer J, Handley FG, Edwards J, et al. Summary of the workshop on drug

- development, biological diversity, and economic growth. J Natl Cancer Inst. 1991;83:1294-1298.
- 164. Rinehart KL. Antitumor compounds from tunicates. Med Res Rev. 2000;20:1-27.
- 165. Chun HG, Davies B, Hoth D, et al. Suffness. Didemnin B The first marine compound entering clinical trials as an antineoplastic agent. Invest New Drugs. 1986;4:279-284.
- 166. Geldof AA, Mastbergen SC, Henrar REC, et al. Cytotoxicity and neurocytoxicity of new marine anticancer agents evaluated using invitro assays. Cancer Chemother Pharmacol. 1999;44:312-318.
- 167. Demetri G, Garcia-Carbonero R, Harmon D, et al. Ecteinascidin-743 (ET-743) induces objective responses and disease control in patients with advanced non-osseous sarcomas: results from phaseII trials. Ann Oncol. 2000;11(Suppl 4):126.
- 168. Erba E, Bergamaschi D, Bassano L, et al. Ecteinascidin-743 (ET-743), a natural marine compound, with a unique mechanism of action. Eur J Cancer. 2001;37:97-105.
- 169. Damia G, Silvestri S, Carrassa L, et al. Unique pattern of ET-743 activity in different cellular systems with defined deficiencies in DNA-repair pathways. Int J Cancer. 2001;92:583-588.
- 170. Poncet J. The dolastatins, a family of promising antineoplastic agents. Curr Pharm Des. 1999;5:139-162.
- 171. Bai R, Pettit GR, Hamel E. Dolastatin10, a powerful cytostatic peptide derived from a marine animal. Inhibition of tubulin polymerization mediated through the vinca alkaloid binding domain. Biochem Pharmacol. 1990;39:1941-1949.
- 172. Pathak S, Multani AS, Ozen M, et al. Dolastatin10 induces polyploidy, telomeric associations and apoptosis in a murine melanoma cell line. Oncol Res. 1998;5:373-376.
- 173. Pettit GR. The bryostatins. Fortschr Chem Org Naturst. 1991;57:153-195.
- 174. Pagliaro L, Daliani D, Amato R, et al. Phase II trial of bryostatin-1 for patients with metastatic renal cell carcinoma. Cancer. 2000;89:615-618.
- 175. Varterasian ML, Mohammad RM, Shurafa MS, et al. PhaseII trial of bryostatin1 in

- patients with relapsed low-grade non-Hodgkin's lymphoma and chronic lymphocytic leukemia. Clin Cancer Res. 2000;6:825-828.
- 176. Zonder JA, Shields AF, Zalupski M, et al. A phase II trial of bryostatin 1 in the treatment of metastatic colorectal cancer. Clin Cancer Res. 2001;7:38-42.
- 177. Ahmad I, Al-Katib AM, Beck FW, et al. Sequential treatment of a resistant chronic lymphocytic leukemia patient with bryostatin1 followed by 2-chlorodeoxyadenoside: case report. Clin Cancer Res. 2000;6:1328-1332.
- 178. Mayer A and Gustafson KR. Marine pharmacology in 2002–2004: Anti-tumour and cytotoxic compounds. Eur J Cancer. 2006;42:2241–2270.
- 179. Mayer A and Gustafson K R. Marine pharmacology in 2005–2006: Anti-tumour and cytotoxic compounds. Eur J Cancer. 2008;44: 2257–2287.
- 180. Binaschi M, Farinosi R, Borgnetto ME, et al. In vivo site specificity and human isoenzyme selectivity of two topoisomerase II poisoning anthracyclines. Cancer Res. 2000;60:3770-3776.

- 181. Patrick Y. Major microbial diversity initiative recommended. Am Soc Microbiol News. 1997;63:417-421.
- 182. Alberts MW, Williams RT, Brown EJ, et al. KBP-Rapamycin inhibits a cyclin-dependent kinase activity and a cyclin D1-Cdk association in early Gl of an osteosarcoma cell line. J Biol Chem. 1993;268:22825-22829.
- 183. Schulte TW, Neckers LM. The benzoquinone ansamycin 17-allylamino-17 demethoxygeldanamycin binds to HSP90 and shares important biologic activities with geldanamycin. Cancer Chemother Harmacol. 1998;42:273-279.
- 184. Cadenas ME, Sandfrison A, Cutler NS, et al. Signal transduction cascades as targets for therapeutic intervention by natural products. Trends Biotechnol. 1998;16:427-433.
- 185. Adjei AA. Signal transduction pathway targets for anticancer drug discovery. Curr Pharmaceut Design. 2000;6:361-378.