ISSN: Print - 2277 - 0755 Online - 2315 - 7453 © FUNAAB 2017 Journal of Agricultural Science and Environment

# EFFECT OF THE BOTANICAL PELLETINGS AND STORAGE PERIODS ON THE GERMINATION OF AFRICAN STAR APPLE (*CHRYSOPHYLLUM ALBIDUM* G. DON) SEEDS

# D. O. ADELANI, O.S. AMOS AND S. MAIKANO

Federal College of Forestry Mechanization, P.M.B 2273, Afaka, Kaduna, Nigeria **\*Corresponding Author**: adelani.olusegun@yahoo.com **Tel**: 2347038953146

#### ABSTRACT

There is paucity of quantified information on the simple, safe and affordable methods of invigorating seeds and storage of recalcitrant seeds. In this light, investigation was conducted in the screen house of the Federal College of Forestry Mechanization, Afaka, Kaduna on the effect of the botanical pelletings and storage periods of C. albidum seeds. A seed pellet is a coating, usually of clay mixed with other inerts, which streamlines the size, shape, and uniformity of a small, non-round seed. A split-plot experimental design with four (4) replications was laid down to assess the effect of botanical pelletings with the leaves of six agro-forestry tree species (Gliricidia sepium, Leucaena leucocephala, Albizia lebbeck, Acacia auriculiformis, Parkia biglobosa and Dacryodus edulis) and storage periods (0, 1, 2, 3 and 4 weeks) on the germination of C. albidum seeds stored in the refrigerator. The data were subjected to analysis of variance (ANOVA) using SAS (2003) software. The percentage germination of seeds pelletized with G. sepium, L leucocephala, A. lebbeck, A. auriculiformis, P. biglobosa and D. edulis ranged between 18.50%-42.00%, 8.3%-13.60%, 5.2%-18.80%, 3.6% -16.30%, 5.0%-14.30%, and 2.5%-11.10%, for 2-8 weeks respectively. The percentage germination of seeds stored for 0, 1, 2, 3 and 4 weeks ranged between 8.50%-21%, 4.70%-24.58%, 9.42%-22.75%, 8.67%-19.58% and 3.33%-8.83% respectively. The percentage germination of seeds pelletized with the leaves of G. sepium, L. leucocephala, A. lebbeck, A. auriculiformis, P. biglobosa and D. edulis and stored for storage periods ranged between 1-65%, 0-30%, 0-30.5%, 1-25%, 0-20% and 0-30%. Botanical pelletings and storage periods significantly (P<0.05) enhanced the germination of C. albidum seeds. Highest germination percentage of 65% was recorded for seeds pelletized with G. sepium leaves and stored for 3 weeks. The research therefore recommends seed pelleting with G. sepium leaves and storage for 3 weeks for mass production of C. albidum seedlings for agro-forestry systems.

Keywords: Chrysophyllum albidum, Agro-forestry, Seed storage periods, seed germination,

# INTRODUCTION

*Chrysophyllum albidum* known as African star apple, belongs to the family Sapotaceae which has up to 800 species (Ehiagbonare *et al.*, 2008). It is a tropical forest tree species that has been incorporated into traditional agro-forestry systems. It is indigenous multipurpose tree species with enormous potentials for agro-forestry (Ureigho and Ekeke, 2010). The species is called "Udala", Agbalumo and Agbaluma in Igbo, Yoruba, and Hausa languages, respectively. Its natural occurrences have been reported in diverse ecozones in Nigeria, Uganda, Niger Republic, Cameroon and Coted'Ivoire (Bada, 1997).

J. Agric. Sci. & Env. 2017, 17(2):37-49

In recent times, C. albidum has become a crop with commercial value in Nigeria (Oboh et al., 2009) due to awareness of its nutritional and medicinal value (Onyekwelu and Stimm, 2011). ICRAF (2007) noted that the fleshy and juicy fruits of the species, which are popularly eaten, are potential source of soft drink. The fleshy pulp of the fruits has been found to have higher contents of ascorbic acid than oranges and guava (Amusa et al., 2003). The fruits are also suitable for the production of fruit jams and jellies (Ureigho and Ekeke, 2010). Also, C. albidum fruit has been also reported to be an excellent source of vitamins, irons, and flavours to diets (Adisa, 2000). The seeds are used for local games or discarded (Bada, 1997). The seeds of the species are sources of oil, which are used for diverse purposes (Ugbogu and Akukwe, 2009). The fruits contain 90% anacardic acid which is used industrially in protecting wood and as source of resin. Chrysophyllum albidum leaves are used as emollients and for the treatment of skin eruptions, diarrhoea and stomach ache, etc (Adisa, 2000).

In spite of the enormous benefits of C. al*bidum*, it has been neglected with the respect to its domestication. To avert total extinction of some of the important forest fruit species, they must be cultivated more intensely through improved propagation either by seed or stem cuttings(Dolor, 2013). Domestication through propagation serves as ex-situ conservation. Ex-situ conservation efforts are necessary to minimize the loss of genetic diversity, reduce the risk of extinction (Godefroid et al., 2010), and produce propagation material for the expansion or re-introduction of populations (Godefroid et al., 2010). This approach is preferred in situations where the populations are in real danger of physical destruction or genetic

deterioration due to excessive pressures in their natural habitat (Nguyen et al., 2014). Seeds are still important starting materials for propagation of many vital tree species (Mng'ombaet al.,2007). Furthermore, the use of seeds as propagules has been considered the easiest and cheapest, and the most common means for many agro-forestry and timber tree species (Akinnifesi et al. 2007). Generally, this has been attributed to the fact that seeds are often easy to produce and handle (Black, 1989). Comparatively, vegetative propagation methods such as grafting, budding and air layering require some skills and knowledge. Seeds are important in mass multiplication of tree planting materials as well as tree improvement through selection (Mng'omba et al., 2007). According to Bewley and Black (1983), seed conservation has been the most reliable and widely used method for ex situ storage. Propagation from seed is inexpensive and usually effective and is therefore a variable method for their exsitu conservation (Abirami et al., 2010).

Often, seed ripening and collection do not correspond with the time of seedling production, while some species do not produce seeds all year round (Dolor, 2013). Even if fruiting is regular and abundant every year, it may be more cost-efficient to collect surplus seeds to cover the needs of few consecutive years (Schmidt, 1988). Hence, efficient storage of seeds is indispensable to ensure continuous and cost-effective supply of tree seedlings, which is a prerequisite for the success of any mass afforestation programme (Umarani *et al.*, 2015).

Moreover, the intelligent manipulation and appropriation of storage life of seeds gives 95% germinability and enhances seedling vigour in the nursery (Adelani *et al.*, 2014a). Normally, *C. albidum* seeds should not be kept longer than 5<sup>th</sup> day of extraction before sowing or else it will lose viability (Adelani *et al.*, 2016). Denton (1997) reported that *C. albidum* seeds are recalcitrant and should therefore be sown almost immediately after maturing to ensure adequate germination. Recalcitrant seeds, maintain relatively high moisture content (typically 40-50%), are damaged by desiccation and are sensitive to chilling. *Chrysophyllum albidum* seeds have very high viability (> 80% at maturity), which may decline rapidly with storage, because it is recalcitrant (Aduradola *et al.*, 2005; Adegoke *et al.*, 2014; Adelani *et al.*, 2016).

Seed management techniques are employed to invigorate the seed by modifying the physiological stamina of the seed (Srimathi *et al.*, 2013). Pelleting is one such physical treatment given to seeds to invigorate its vigour. A seed pellet is a coating, usually of clay mixed with other inerts, which streamlines the size, shape, and uniformity of a small, non-round seed such as those of lettuce, carrots, onions, and many herbs and flowers. Pelleting results in easier, safer, and more accurate mechanical seeding, thus reducing gaps in the field and the need for labour-intensive thinning (Gatch, 2016).

Some of the common benefits of pelleting are uniformity in seed size, precision planting; uniform stands with reduced seed rate, more insect and disease resistance, better performance under stress conditions and additional nourishment to the seedlings (Halmer, 2003). Pelleting was employed to overcome the problem of poor germination and seedling growth as well as survival of finger and brown top millets and vegetables (jute, amaranth, cat's whiskers) (Patrick *et al.*, 2016). Pelleting increased seed size by about 33-150% and a significant improve-

ment in germination was observed for pelleted seed (Patrick *et al.*, 2016).

The studies on pelleting in agriculture especially in field crops are well exploited but in case of tree species it is very scanty (Srimathi *et al.*, 2013). There is dearth of information on the pelleting and storage of *C. albidum* seeds. Inadequate germinability and storability lead to paucity of seedlings for afforestation programme in agro-forestry systems. In an attempt to increase the germinability and storability of *C. albidum* seeds, the study investigates its pelleting and storage periods.

# MATERIALS AND METHOD Experimental Site

The pot experiment was conducted during the dry season (that is December) of 2015 at the screen house of Federal College of Forestry Mechanization, Afaka, Kaduna state, Nigeria. The college is located within the Northern Guinea Savanna ecological zone and situated in Igabi Local Government Area of Kaduna State, Nigeria. The college lies between latitude 10° 35' and 10° 34'N and longitude 7° 21' and 7° 20'E (Adelani, 2015). The mean annual rainfall is approximately 1000 mm, with the lowest mean monthly relative humidity of about 29%. The soil is classified as sandy loam. The vegetation is open woodland with tall broad leave trees (Otegbeye *et al.*,2001).

# Fruit Collection and Preparation

The fruits were collected from the mother tree at village near Federal University of Agriculture, Abeokuta, Ogun State. The seeds were extracted from the fruits, washed and air dried for thirty minutes. The river sand was collected from the floor of the College dam, passed through (2mm) sieve and sterilized at 160°C for 24 hours (Adelani and Joseph, 2014). The polypots of 20 x 25 x 25

cm<sup>3</sup> capacity filled with sterilized river sand in the nursery was used.

#### Experiment

The effect of the botanical pelletings and storage periods on the germination of *C. albidum* seeds was laid down using a split plot experimental design. Pelletings with leaves of six agro-forestry tree species (*Gliricidia sepium, Leucaena leucocephala, Albizia lebbeck, Acacia auriculiformis, Parkia biglobosa and Dacryodus edulis*) and storage periods (0, 1, 2, 3, and 4 weeks) constituted the main and subplot treatments, respectively. The effects of pelletings with leaves of each of the agro-forestry tree species and the period of storage on the germination of *C. albidum* seeds were assessed for 8 weeks beginning

from the first day of germination. Ten (10) C. albidum seeds represented a replicate. Forty (40) seeds were coated with the mixture of clay (1.5g/seed), fungicides (0.5ml/seed), and leaves of agro forestry trees species (2.4g /seed) and 1ml of water. Pelleted seeds were stored on 0, 1, 2, 3, and 4weeks. Forty (40) seeds per week of storage in refrigerator were sown in the acid washed river sand at 4cm depth and watered twice daily at 200ml/seed. Seeds that were not coated and not stored served as control. Seeds were considered to have germinated after the emergence of plumule above the ground. Germination percentages were calculated as total number of seed germinated per total number of seed planted multiplied by 100.

 $=\frac{Totalseedgerminated}{Totalseedsown} \times 100$ 

Germination Percentage

Each sample of pulverized leaves of agroforestry tree species was analysed chemically for nitrogen, phosphorus and potassium (NPK) content at the Ahmadu Bello University laboratory. Determination of total nitrogen and available phosphorus were done by MacroKjeldahi method. Extracts from the digestion of the leaves of the agro-forestry species were used to determine potassium by flame photometry. The dry weights of the leaves of agroforestry tree species were determined, by the use of Mettle Top Loading Weighing Balance (Model-Mettler PM 11-K), after oven dried at 70°C for 72hours (Umar and Gwaram, 2006).

#### Data analysis

The data were collected on the potentials of botanical pelletings and storage periods on the germination of *C. albidum* seeds. The data were subjected to analysis of variance

(ANOVA)using SAS (2003)software. Separation of significant means was accomplished using Fisher's Least Significant Difference (LSD) at 5% level of significance.

# **RESULTS AND DISCUSSION** *Percentage NPK composition of the leaves of Agro-forestry tree species*

The result of the analysis of NPK composition of the leaves of agro-forestry tree species is presented in Table 1. *G. sepium* leaves had highest percentage composition of 30.89% and 19.6% for nitrogen and phosphorus respectively. *D. edulis* leaves had the highest percentage composition value of 2.71% for potassium. The significant performance of *G. sepium* leaves is traceable to its highest composition of nitrogen and phosphorus. Similar observation was reported by Adelani *et al.* (2014b) who stated that the highest growth parameters recorded for maize planted in acid washed sand amended

J. Agric. Sci. & Env. 2017, 17(2):37-49

#### EFFECT OF THE BOTANICAL PELLETINGS AND STORAGE...

with the leaves of *Gliricidia sepium* was adduced to its ability to release its highest nitrogen composition among other agroforestry trees species investigated.

Nitrogen which is component of *G. sepium* has ability to improve water absorption and protein synthesis and equally promote cell division and elongation of seeds (Crawford

*et al.*, 2000). Nitrogen in *G. sepium* regulated developmental process as germination (Alboresi *et al.*, 2005). Phosphorus is an important component of *G. sepium* (Adelani *et al.*, 2014b) which help in the germination of seeds (Mengel and Kirkby 2001; Smith, 2014).

Agro-forestry tree spp	N%	P%	K%
Gliricidia sepium	30.89	19.6	2.07
Leucaena leucocephala	27.68	3.1	2.05
Albizia lebbeck	21.05	3.4	1.91
Acacia auriculiformis	26.96	0.1	2.17
Parkia biglobosa	27.44	10.6	0.84
Dacryodus edulis	27.64	1.1	2.71

# Table 1: Percentage NPK composition of the leaves of Agroforestry tree species

# Effect of the botanical pelletings and storage periods on the germination of *C. albidum* seeds

The result of the effect of the botanical pelletings and storage periods on the germination of C. albidum seeds is represented in Table 2. The percentage germination of seed pelletized with the leaves of G. sepium, L. leucocephala, A. lebbeck, A. auriculiformis, P. biglobosa and D. edulis ranged between 8.3%-13.60%, 18.50%-42.00% 5.2%-18.80%, 3.6%-16.30%, 5.0%-14.30%, and 2.5%-11.10%, for 2-8 weeks respectively. Highest germination of 42% was recorded in seed pelletized with G. sepium leaves. The excellent performance of G. sepium leaves could be adduced to the fact that it enhanced the fertilization of C. albidum seeds. This is in consonance with the re-

ports of Vijaya and Balamurugan (2011) who stated that seed pelleting added needy substance to the individual seed so that the seeds get invigorative effect on absorption of such materials, at initial watering by enriching the rhizosphere region of each and every seed as nutritive without physiological modification of seed but by simple physical alterations of the seed.

Similar reports have been made by Scott, 1989; Mani *et al.*, 1999; Gurunathan *et al.*, 2006; Govinden and Levantard (2008) and Srimathi *et al.*, (2013). This is also corroborated by the reports of Srimathi *et al.* (2013) on pelleting of *Jatropha curcasand Pongamia pinnata*with the leaf powder of *Azadirachta indica, Pongamia pinnata*and *Adhatoda vasica.* Srimathi *et al.* (2013) attributed the excellent

J. Agric. Sci. & Env. 2017, 17(2):37-49

performance of *Pongamia pinnata* to its ability to correct soil moisture, contains gibberellins like substance which was synergistically activated to form indole acetic acid (IAA) and ability of its leaf component to act as chelating agent and activated the growth and development.

All pelletized C. albidum seeds performed better than that of the unpelletized seeds. This in consonance with the report of Woods (2014) who stated that pelleted seeds registered significantly higher germination and seedling vigour in red cedar and paper compared to non-pelleted red (control) under all soil types. Similar observation has been made on pelletings of Jatropha curcas and Pongamia pinnata (Srimathi et al., 2013). This is also corroborated by Patrick et al. (2016) on dicotyledon and grass seeds. Contrary to earlier reporters, Arshadi and Asgharipour (2011) stated that pelleting with usage of extra covering material for sugar beet seed pelleting decreased viability and uniformity of seed germination.

# Effect of storage periods on the germination of C. albidum seeds

The result of the effect of storage periods on the germination of *C.albidum* seeds is presented Table 2. The germination percentage values of seeds stored for 0, 1, 2, 3 and 4 ranged between 8.50%-21%, 4.70%-24.58%, 9.42%-22.75%, 8.67%-19.58% and 3.33%-8.83%, respectively. Highest germination percentage of 24.58% was recorded in seeds stored for 1 week. Based on this result, it can be inferred that *C. albidum* seeds can be stored for one week, for viability not to fall because it is recalcitrant. This is corroborated by the reports of Adu-

radola et al. (2005) and Adelani et al. (2014a).

The increasing periods of storage decrease the germination percentage of C. albidum seeds. Similar observation has made by Temel *et al.* (2011) who reported significant reductions in germination rate (from 79.93%) to 30.68%) and germination percentage (from 95.99% to 58.41%) of *Pinus nigra* after 10-year storage. Also, Missanjo and Kapira (2015) reported that germination percentage and germination energy of *Pinus kesiya* seeds decreased with an increase of storage period. This is in consonance with the reports of Hilli et al. (2003) who stated that the germination of 10-year stored Pinus sylvestris seeds reduced significantly for both cold and frozen storage conditions. Contrary to earlier reporters, Kamotho et al. (2013) reported a significant increase in germination percentage of *Cleome gynandra* after 6 months of storage.

The ability of the *C.albidum* seeds to have decrease germination percentage with the increasing period of storage shows it is recalcitrant. Recalcitrant seeds are shed at relatively high moisture contents and are sensitive to desiccation (Anandalakshmi et al., 2005). Similar observation has been made by lakovoglou (2005) on recalcitrant seeds of the Leucobalanus subgenus of Quercus poses. Recalcitrant seeds germinate rapidly when sown fresh, but are sensitive to desiccation and freezing (Berjak and Pammenter, 2004; McDonald, 2004). This is in line with the reports of Liang and Sun (2000). Some recalcitrant seeds are shed when seed moisture is high and usually start germinating before and after shedding (Rajeeswari and Kaveriappa, 2000).

2       4       6       8       2       4       6       8         Gliricidia. spium       18.50cd       21.5c       31.0b       42.0a       -	Agoforestry tree species	Weeks	of	Germination		Period of Storage	Weeks	of	Germination	
18.50cd       21.5c       31.0b       42.0a       -       -       -       -       -       -         hala       8.3b       9.5b       13.6ab       18.30a       0       8.50c       13.42b       17.67ab         mis       5.20b       7.3b       14.40ab       18.8a       1       9.0d       17.75c       24.5b         mis       3.6c       8.40b       15.10a       16.30a       2       9.42b       14.42ab       17.08ab         8.67ab       10.92ab       15.80a       19.5a       3       8.67ab       10.92ab       15.80a         7.60a       8.00b       11.1a       11.1a       4       3.33ab       5.83a       7.92a         3.15       3.15       3.15       3.15       3.15       3.15       5       2.88       2.88       2.88		2	4	9	8		2	4	6	8
hala $8.3b$ $9.5b$ $13.6ab$ $18.30a$ $0$ $8.50c$ $13.42b$ $17.67ab$ $5.20b$ $7.3b$ $14.40ab$ $18.8a$ $1$ $9.0d$ $17.75c$ $24.5b$ $3.6c$ $8.40b$ $15.10a$ $16.30a$ $2$ $9.42b$ $14.42ab$ $17.08ab$ $8.67ab$ $10.92ab$ $15.80a$ $19.5a$ $3.67ab$ $10.92ab$ $15.80a$ $7.60a$ $8.00b$ $11.1a$ $11.1a$ $4$ $3.33ab$ $5.83a$ $7.92a$ $3.15$ $3.15$ $3.15$ $3.15$ $3.15$ $3.15$ $5E$ $2.88$ $2.88$ $2.88$	Gliricidia. sepium	18.50cd	21.5c	31.0b	42.0a	ı	ı	ı		,
5.20b       7.3b       14.40ab       18.8a       1       9.0d       17.75c       24.5b         3.6c       8.40b       15.10a       16.30a       2       9.42b       14.42ab       17.08ab         8.67ab       10.92ab       15.80a       19.5a       3       8.67ab       10.92ab       15.80a         7.60a       8.00b       11.1a       11.1a       4       3.33ab       5.83a       7.92a         3.15       3.15       3.15       3.15       3.15       5.8       2.88       2.88       2.88	Leucaena leucocephala	8.3b	9.5b	13.6ab	18.30a	0	8.50c	13.42b	17.67ab	21.0a
mis       3.6c       8.40b       15.10a       16.30a       2       9.42b       14.42ab       17.08ab         8.67ab       10.92ab       15.80a       19.5a       3       8.67ab       10.92ab       15.80a         7.60a       8.00b       11.1a       1       3.33ab       5.83a       7.92a         3.15       3.15       3.15       3.15       3.15       5.88       2.88       2.88	Albizia lebbeck	5.20b	7.3b	14.40ab	18.8a	-	9.0d	17.75c	24.5b	47.0a
8.67ab 10.92ab 15.80a 19.5a 3 8.67ab 10.92ab 15.80a 7.60a 8.00b 11.1a 11.1a 4 3.33ab 5.83a 7.92a 3.15 3.15 3.15 $3.15 {}_{SE} \pm 2.88 2.88 2.88$	Acacia auriculiformis	3.6c	8.40b	15.10a	16.30a	2	9.42b	14.42ab	17.08ab	22.75a
ryodus edulis 7.60a 8.00b 11.1a 11.1a 4 3.33ab 5.83a 7.92a $\pm$ 3.15 3.15 3.15 3.15 $5_{\text{E}} \pm$ 2.88 2.88 2.88 2.88	Parkia biglobosa	8.67ab	10.92ab	15.80a	19.5a	S	8.67ab	10.92ab	15.80a	19.5a
$\pm$ 3.15 3.15 3.15 3.15 3.15 $= 3.15$ $= 3.15$ $= 3.15$ $= 2.88$ 2.88 2.88	Dacryodus edulis	7.60a	8.00b	11.1a	11.1a	4	3.33ab	5.83a	7.92a	8.83a
	SE ±	3.15	3.15	3.15	3.15		2.88	2.88	2.88	2.88

EFFECT OF THE BOTANICAL PELLETINGS AND STORAGE...

J. Agric. Sci. & Env. 2017, 17(2):37-49

43

#### Interactive effect of the botanical pelletings and storage periods on the germination of C. albidum seeds

The result of interactive effect of the botanical pelletings and storage periods on the germination of *C. albidum* seeds is presented in Table 3. The percentage germination of seeds pelletized with the leaves of *G. sepium*, *L.leucocephala*, *A. lebbeck*, *A. auriculiformis*, *P. biglobosa and D. edulis* and stored for 0, 1, 2, 3 and 4weeks ranged between 1-65%, 0-30%, 0-30.50%, 1-25%, 0-20% and 0-30% respectively. Highest germination percentage of 65% was recorded in seeds pelletized with *G. sepium* leaves and stored for 3 weeks. This is traceable to the fact that *G. sepium* leaves enhanced the storability of *C. albidum* seeds. Similar observation was recorded by Srimathi *et al.* (2013) who stated that seed pelleted with *Pongamia pinnata* leaf powder could be stored for up to six months with 48% germination.

 Table 3: Interactive effect of pelleting and storage on the germination of *C. albidum* seeds

			Weeks of Germination		
Agro-forestry tree Species	Period of storage	2	4	6	8
Gliricidia sepium	0	1.00 <sup>c</sup>	5.00 <sup>c</sup>	10.5 <sup>b</sup>	20.0ª
Gliricidia sepium	1	1.50 <sup>d</sup>	10.50 <sup>c</sup>	25.00b	35.0 <sup>a</sup>
Gliricidia sepium	2	15.50 <sup>d</sup>	21.00 <sup>c</sup>	45.0 <sup>b</sup>	50.0 <sup>a</sup>
Gliricidia sepium	3	25.00 <sup>c</sup>	40.00 <sup>c</sup>	60.00 <sup>b</sup>	65.00ª
Gliricidia sepium	4	20.00c	35.00 <sup>b</sup>	40.00 <sup>a</sup>	40.00 <sup>a</sup>
Leucaena leucocephala	0	15.50 <sup>bc</sup>	20.00b	25.00b	25.00ª
Leucaena leucocephala	1	15.50 <sup>bc</sup>	15.50 <sup>bc</sup>	20.00b	25.00ª
Leucaena leucocephala	2	6.00 <sup>c</sup>	15.00 <sup>b</sup>	15.50 <sup>b</sup>	30.0
Leucaena leucocephala	3	0.00 <sup>b</sup>	1.50 <sup>b</sup>	1.50 <sup>b</sup>	30.00 <sup>a</sup>
Leucaena leucocephala	4	0.00 <sup>b</sup>	0.00b	0.50 <sup>b</sup>	11.00ª
Albizia lebbeck	0	10.50 <sup>b</sup>	11.00 <sup>b</sup>	15.00 <sup>b</sup>	25.50ª
Albizia lebbeck	1	5.50 <sup>c</sup>	15.50 <sup>c</sup>	25.50 <sup>ab</sup>	30.00 <sup>a</sup>
Albizia lebbeck	2	0.00 <sup>c</sup>	5.00 <sup>c</sup>	15.50 <sup>b</sup>	30.50ª
Albizia lebbeck	3	0.00 <sup>c</sup>	5.00 <sup>c</sup>	10.50 <sup>b</sup>	20.50ª
Albizia lebbeck	4	0.00 <sup>c</sup>	0.0 <sup>c</sup>	1.50 <sup>ab</sup>	2.00 <sup>a</sup>
Acacia auriculiformis	0	6.50 <sup>c</sup>	20.00 <sup>b</sup>	25.00 <sup>a</sup>	25.00ª
Acacia auriculiformis	1	1.00 <sup>c</sup>	10.50 <sup>b</sup>	20.50 <sup>ab</sup>	25.00ª
Acacia auriculiformis	2	5.00 <sup>b</sup>	6.00 <sup>b</sup>	15.50ª	15.50ª
Acacia auriculiformis	3	5.50 <sup>b</sup>	5.50 <sup>b</sup>	15.50 <sup>ab</sup>	20.00ª
Acacia auriculiformis	4	0.00 <sup>b</sup>	0.00b	0.00b	5.00 <sup>a</sup>
Parkia biglobosa	0	11.00 <sup>ab</sup>	15.00 <sup>a</sup>	15.50ª	15.50ª
Parkia biglobosa	1	10.00 <sup>b</sup>	15.00 <sup>a</sup>	15.50ª	16.00ª
Parkia biglobosa	2	0.00 <sup>b</sup>	0.50 <sup>b</sup>	20.00 <sup>a</sup>	20.00ª
Parkia biglobosa	3	0.00c	0.00c	15.00 <sup>b</sup>	20.00ª
Parkia biglobosa	4	0.00a	0.00a	0.00a	0.00a
Dacryodus edulis	0	6.00 <sup>ab</sup>	10.00 <sup>b</sup>	15.00ª	15.00ª
Dacryodus edulis	1	6.50 <sup>c</sup>	20.00 <sup>b</sup>	25.50 <sup>ab</sup>	30.00 <sup>a</sup>
Dacryodus edulis	2	0.00 <sup>b</sup>	0.00 <sup>b</sup>	0.50 <sup>b</sup>	5.00ª
Dacryodus edulis	3	0.00 <sup>b</sup>	0.00 <sup>b</sup>	2.00b	10.00 <sup>a</sup>
Dacryodus edulis	4	0.00a	0.00ª	0.00ª	0.00 <sup>a</sup>
se ±		7.04	7.04	7.04	7.04

\* Means on the same row on each treatment with the same alphabet are not significantly difference at (P<0.05)

# CONCLUSION

Botanical pelleting was employed to overcome the problem of poor germination and storage of *C. albidum* seeds. The result revealed that pelleting and storage of *C. albidum* seeds enhanced germination. Seeds pelletized with *G. sepium* leaves gave highest percentage germination. Seeds of *C. albidum* need to be stored for 1 week to have highest germination percentage. *C. albidum* seeds pelletized with leaves of *G. sepium* and stored for 3 weeks had highest germination percentage. Botanical pelletings is used to invigorate the seeds by modifying its physiological stamina.

# REFERENCES

Abirami, K., Rema, J., Mathew, P. A., Srinivasan, V., Hamza, S. 2010. Effect of different propagation media on seed germination, seedling growth and vigour of nutmeg (*Myristica fragrans* Houtt). Journal of Medicinal Plants Research 4(19): 2054-2058.

Adegoke, F.F., Ogunwande, O.A., Olaitan, A.O., Agbeje, M.A. 2014. Rooting Potentials of the cuttings of Chrysophyllum albidum as influenced by different rooting hormones at different levels of concentration. Forests and Forest Products: Key to Sustainable Livelihood. In: Adedire. M.O: Onyekwelu, J.C; Oke, D.O; Adekunle, Jayeola, O.A and Oladoye, A.O. V.A.J. (Eds). Proceedings of the 4th Biennial National Conference of the Forests and Forest Product Socie*ty* pp54-56.

Adelani, D.O., Joseph, A. 2014. Storability of Japanese Acacia (*Acacia auriculiformis* A.cunn. Ex Benth) in Northern Guinea Savannah Ecological Zone of Nigeria. *Forest and Forest Product Journal* 7:1-10.

**Ia**, **A.M.**, **Anamyi**, **S.E.** 2014a. Storability of tropical forest seeds for sustainable development in the world: A review. *Biological and Environmental Sciences for the Tropics* 11 (1): 38-42

Adelani, D.O., Suleiman, R.A., Aduradola, M.A., Akesode, H.A. 2014b. Assessment of leaf litters of some tree species on the growth of *Zea* mays (L) in the Northern Guinea Savanna ecology. *Journal of Organic Agriculture and Environment* 2:117-130.

**Adelani**, **D.O.** 2015. Effect of hydropriming and potassium nitrate priming on the germination of *Balanites aegyptiaca*. *Applied Tropical Agriculture 20* (2): 17-23.

Adelani, D.O., Aduradola, M.A., Maisamari, I.J. 2016. Storability and presowing treatments of *Chrysophyllum albidum* seeds: A step towards Biodiversity Conservation in Borokini, I.T and Babalola, F.D (eds). MDGsto SDGs: Towards Sustainable, Biodiversity Conservation in Nigeria. Proceedings of Joint Biodiversity Conservation, Conference of Nigeria Tropical Biology Association (NTBA) and Nigeria Chapter of Society for Conservation Biology (NSCB) PP80-86.

Adisa, S.A. 2000. Vitamin C, protein and mineral contents of African star apple (*Chrysophyllum albidum*) in Proceedings of the 18<sup>th</sup> Annual Conference of NI ST (eds) Garba, S.A, IJA-gbone, I.F, Iyagbo, A.O., Iyamu, A.O., Kilani, A.S., Ufaruna Pp.141-146.

Aduradola, A.M., Adeola, A.O., Adedire, M.O. 2005. Enhancing germination in seeds of *Chrysophyllum albidum* (G.Don). *Journal of Food, Agriculture and Environment* 3(2): 292-294.

Adelani, D.O., Adedire, M.O., Adurado- Akinnifesi, F. K., Sileshi, G., Mkonda, A.,

Ajayi, O.C., Mhango, J., Chilanga, T. 2007. Germplasm supply, propagation and nursery management of Miombofruit trees. In: Akinnifesi, F.K., Sileshi, G., Ajaji, O., Tchoundjeu, Z., and Matakala, P(Eds). *Indigenous Fruit Trees in the Tropics: Domestication, Utilization and Commercialization,* CABI publishing, UK, chapter 19, pp 341.

Alboresi, A., Gestin, C., Leydecker, M. T., Bedu, M., Meyer, C., Truong, H.N. 2005. Nitrate, a signal relieving seed dormancy in *Arabidopsis. Plant Cell and Environment* 28: 500-512.

Anandalakshmi, R., V. Sivakumar, R. R. Warrier, R. Parimalam, S. N. Vijayachandran., B. G. Singh. 2005. Seed storage studies in *Syzigium cuminii. Journal of Tropical Forest Science* 17(4): 566-573.

Amusa, N.A., Ashaye, O.A., Oladapo, M.O. 2003. Biodeterioration of African starapple (*Chrysophyllum albidum*) in storage and the effect on its food value. *African Journal of Biotechnology* 2:56-59.

**Arshadi, J., Asgharipour, M.R.** 2011. The Effects of seed size on germination and early seedling growth of pelleted seeds of sugar beet. *Journal of Applied Sciences Research, 7*(8): 1257-1260.

**Bada, S.O.** 1997. Preliminary information on the ecology of *Chrysophyllum albidum* (G. Don) in West and Central Africa in proceedings of National Workshop on the Potentials of the African star apple in Nigeria (eds) Denton DA, Ladipo D.O, Adetoro M.A. and Sarumi M.B. Pp 16-25.

Berjak, P., Pammenter, N.W. 2004. *Recalcitrant Seeds.* In: Benech-Arnold R.L, Sánchez, R.A (Eds) Handbook of Seed Physiology, Application to Agriculture, The Haworth Press, Inc, NY, USA, pp 305-345

**Bewley, J. D., Black, M.** 1983. Physiological and Biochemistry of Seeds in Relation to Germination. (Vol 1) Development, Germination, and Growth, Sprin-ger-Verlag, Berlin, Germany, 306 pp.

**Black, M.**1989. Seed research – past, present and future. *In*: Taylorson, R.B (Ed) *Recent Advances in the Development and Germination of Seeds.* Series A, Life Sciences Vol.187, Plenum Press, NY, 295 pp.

Crawford, N. M., Khan, M. L., Leustek, T., Long, S. R. 2000. Nitrogen and sulphur. *In: Biochemistry and Molecular biology of plants*, (eds.) Buchanan, B., Gruissem, W. and Jones, R. (Publ.) *American Society of Plant Physiologist* pp. 786-849.

**Denton, L.** 1997. Germplasm, collection and strategy for *Chrysophyllum albidum*. *In: Proceedings of a National Workshop on the Potentials of the Chrysophyllum albidum in Nigeria*. Gbile, Z. (eds). Pp. 130

Dolor, D. E. 2013. Propagation of *Treculia africana* as influenced by seed storage and propagation media. *Agricultura Tropica et Sub-tropica*, 46/2: 52-57.

Ehiagbonare, J.E., Onyibe, H.I., Okoegwale, E.E. 2008. Studies on the Isolation of normal and abnormal seedlings of *Chrysophyllum albidum*. A step towards sustainable management of the taxon in the 21st century. *Science Research Essay*, *3*(12): 567-570.

**Gatch**, **E**. 2016. Organic seed treatments and coatings. *Organic Seed Resource Guide* (1):1 -6.

**Godefroid, S., Vanderborght, T.** 2010. Seed banking of endangered plants: are we conserving the right species to address climate change? *Biodiversity and Conservation* 19 (11):3049-3058.

**Govinden, S. J., Levantard, M.** 2008. Comparative studies of seed priming and Pelleting on percentage and mean time to germination of seeds of tomato (*Lycopersicon esculentum* Mill.). *African Journal of Agricultural Research* 3(10):725-731.

Gurunathan, N., Srimathi, P., Paramathma, M., Kumaran, K., Parthiban, K.T. 2006. Seed pelleting for production of quality seeding in *Jatropha curcas*. Abstr. XII National Seed Seminar, 24-26, Feburary 2006, ANGRAU, Hyderbad. Pp 12.

Halmer, P. 2003. Enhancing seed performance for better yield and quality. *Asian Seed Planting Materials* 10(2):4-6.

**Hilli, A., Tillman-Sutela, E., Kauppi, A.** 2003. Germination of pre-treated scotspine seeds after long term storage. *Canadian Journal of Forest Research* 33: 47-53.

**ICRAF**. 2007. Chrysophyllum albidum International Center for Research in Agroforestry, Kenya. Pp 3

Kamotho, G. N., Mathenge, P.W., Muasya, R. M., Dulloo, M. E. 2013. Effects of packaging and storage conditions on quality of spider plant (*Cleome gynandra* L.) seeds. *African Journal of Food, Agriculture, Nutrition and Development,* 13(5): 8368-8387.

**lakovoglou**, **V.** 2005. "Desiccation and nitrous oxide storage effects on the recalcitrant seeds of *Quercus alba* and *Quercus macrocarpa*". *Retrospective Theses and Dissertations.* 

J. Agric. Sci. & Env. 2017, 17(2):37-49

**Liang, Y., W.Q. Sun.** 2000. Desiccation tolerance of recalcitrant *Theobroma cacao* embryonic axes: the optimal drying rate and its physiological basis. *Journal of Experimental Botany* 51:1911-1919.

**McDonald, M. B.** 2004. Orthodox seed deterioration, its repair. In: Benech-Arnold R. L, Sánchez RA (Eds). Handbook of Seed Physiology, Application to Agriculture, Haworth Press, Inc, NY, USA, pp 273-298

Mani, G., Ponnuswamy, A.S and Vanangamudi', K. 1999. Performance of seed pelletization in *Acacia leucophloea* (Roxb.) under different soil types. "*Tropical J\\$ticuUumt Research anrf ?xtension.* 2(1): 30-32.

**Mengel, K., Kirkby. E. A.** 2001. Principles of Plant Nutrition. Kluwer Academic Publishers, Dordrecht, The Netherlands. p. 833.

**Missanjo, E., Kapira, D.** 2015. Storage conditions and period effects on quality of *Pinuskesiya* seeds from Malawi. *Scholars Academic Journal of Biosciences (SAJB) 3*(3):315-319

Mng'omba ,S. A.,du Toit, E.S., Akinnifesi, F. K. 2007. Germination characteristics of tree seeds: spotlight on Southern African tree species. *Tree and Forestry Science and Biotechnology 1*(1): 1-8.

Nguyen, T.Q., Hamilton, P., Ziegler, T. 2014. *Shinisaurus crocodilurus*. The IUCN Red List of Threatened Species. Version 2014. 2. <www.iucnredlist.org>. Downloaded on 30 October 2014.pp5.

**Oboh, L.O., Aluyor, E.O., Audu, T.O.K.** 2009. Use of *Chrysophyllumalbidum* for the removal of metal ions from aqueous solution.

Scientific Research and Easy4 (6): 632-635.

**Onyekwelu, J.C., Stimm, B.** 2011. *Chrysophyllum albidum*. In:Roloff, A; Weisger ber, HLang, U; Stimm, B.(Eds): Enzyklopadieder Holzgewachse, Willey, VCH, Weinheim, 59. Erg.Lfg.10/11,12pp

**Otegbeye G.O., Owonubi, J.J., Oviasuyi** P.K. 2001. Interspecipic variation growth *Eucalyptus* growing in northern Nigeria in Popoola. L. Abu. J.E and Oni, P.I (eds). *Proceedings of 27th Annual Conference of the Forestry Association of Nigeria* Pp 12-16.

**Patrick, T., Thomas, A., Rutto, L. K.** 2016. Effect of seed pelleting on germination and survival of select small seeded species. *Virginia Journal of Science* 67 (1):4-5.

**Rajeeswari, D. B., Kaveriappa, K. M.** 2000. Effect of desiccation and temperature on germination and vigour of the seeds, *Hopeaparviflora* Beddome and *Hopea ponga* (Dennst.) Mabb. *Seed ScienceTechnology*28: 497 –506.

**SAS.** 2003. Statistical Analysis System. SAS release 9.1 for windows, SAS Institute Inc.cary,NC, USA.

Schmidt, W. C. 1988. Lodge pole pine: an ecological opportunist. In Proceedings, Symposium: *The Management of Lodgepole Pine to Minimize the Losses to the Mountain Pine Beetle* (ed. Amman, G.D.), Kalispell, MT, USDA Forest Service GTR-INT-1988, 12–14, July, pp. 14–20.

**Scott, J. M** 1989. Seed coatings and treatments and their effects on plant establishment. *Advances in Agronomy*, 42: 43-83.

Smith, R.A. 2014. Plant Nutrients. 1(1): 1-4

J. Agric. Sci. & Env. 2017, 17(2):37-49

http//www/plant nutrient.htm. Accessed on 10/03/2015.

Srimathi, P., Mariappan, N., Sundaramoorthy, L., Paramathma, M. 2013. Effect of organic seed pelleting on seed storability and quality seedling production in biofuel tree species. *Journal of Horticulture and Forestry 5*(5): 68-73.

**Temel, F., Gülcü, S., Ölmez, Z., Göktürk, A.** 2011. Germination of Anatolian black pine (*Pinus nigra* subsp. *pallasiana*) seeds from the Lakes Region of Turkey: Geographic variation and effect of storage. *Notulae Botanicae Horti Agro botanici Cluj-Napoca; 39*(1): 267-274.

**Ugbogu O. C., Akukwe, A. R.** 2009. The antimicrobial effect of oils from *Pentacle thra macrophylla* (Bent), *Chrysophyllum albidum* (G. Don) and Persea gratissima (Gaerth F) on some local clinical bacterial isolate. African Journal of Biotechnology 8 (2): 285-287.

**Umar, T. Gwaram, A. B.** 2006. Foliar nutrient contents of four indigenous trees of the sudan savanna. In: Popoola, L. (Eds). *Proceedings of 31st Annual Conference of Association of Nigeria* 131-139.

Umarani, R., Aadhavan, E. K., Faisal, M. M. 2015. Understanding poor storage potential of recalcitrant seeds. *Current science 108* (11): 2023-2034

**Ureigho, U.N., Ekeke, B.A.** 2010. Nutrient value of *Chrysophyllum albidum* African star apple as a domestic income plantation species. *International multi-disciplinary Journal, Ethiopia* 4(2): 50-56.

**Vijaya Geetha, V., Balamurugan, P.** 2011. Organic Seed Pelleting in Mustard. *Research Journal of Seed Science, 4: 174-180.*  Woods, J .2014. Report on the Quality Assurance (QA) Results with pelleted seeds of Western redcedar, red alder and paper birch

(Manuscript received:29th June, 2017; accepted: 12th November, 2018).