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# Load-dependent NMR low-field profiling and relaxation dispersion study of osteoarthritic articular cartilage



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#### ABSTRACT

At low magnetic fields,  $T_1$  variation within cartilage represents a robust parameter that is employed to quantify the layered structure in the tissue and is sensitive to factors such as enzymatic degradation, external load, and degeneration such as osteoarthritis. Variable-field relaxometry, on the other hand, provides access to the quadrupolar dips, i.e. enhanced relaxation rates of <sup>1</sup>H particularly at field strengths between 50 and 70 mT, that probe proton-nitrogen interaction and thus the content and local order of macromolecular constituents, namely glycosaminoglycans and collagen. At the same time, an strong overall dispersion of  $T_1$  is observed over the whole accessible range of magnetic fields upward from 0.25 mT.

In this study on 20 human cartilage samples, low-field and variable-field techniques were combined for the first time to correlate corresponding NMR parameters and the response to load with the severity of osteoarthritis. The magnitude of the quadrupolar dips, as well as cartilage thickness obtained from profile measurements, is found to correlate with the severity of osteoarthritis. At the same time, a significant correlation was identified for relaxation time variation before and after uniaxial compression at 0.6 MPa, a typical value for forces appearing in the human knee and hip joint. This finding is of importance since the spatial resolution of 50 µm obtained with the single-sided scanner is about one order of magnitude better than the one in clinical high-field or low-field scanners, thus allowing a much more detailed investigation and yet providing constraints for the interpretation of averaged values obtained with whole-body scanners.

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#### 1. Introduction

Clinical investigations of joint tissue has become of increasing importance during the last decade and have been developed into a standard and routine procedure for a number of health issues, osteoarthritis being one of the most important ones. However, clinical studies usually rely on spatial information, such as measuring the distance between bone surfaces, and frequently do not attempt to quantify properties such as NMR relaxation times. As far as transverse relaxation is concerned, cartilage possesses some of the shortest values in the body apart from its solid components, which limits the application of imaging sequences if one assumes conventional, commercially available gradient systems. This particularly restricts also the spatial resolution of the cartilage tissue with its typical thickness of 2–3 mm to a few pixels at most. While T<sub>2</sub> relaxation contains significant information and is strongly affected by the layered structure of the collagen network with distinctly different order parameters [1–3] which suggest correlation with OA [4], T<sub>1</sub> remains – without the addition of contrast agents – less affected by the degeneration, but also not prone to the magic angle effect.

The observations above are made at typical magnetic field strengths found in clinical systems. At lower fields, however,  $T_1$  varies in a similar fashion as  $T_2$ , possibly with an even larger range

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of values [5]. A further variable that introduces potential changes in the relaxation characteristics of cartilage is mechanical load: if the joint is subjected to "realistic" conditions, i.e. by applying the body weight onto the knee joint, the water distribution as well as arrangement of macromolecules and, consequentially, its relaxation properties are affected. While normal, healthy tissue would react in a well-defined way, by water being expelled from the cartilage tissue during loading and re-entering within several seconds after un-loading, the osteoarthritic tissue has much less capacity in resorbing water due to reduced swelling pressure, a fact which is confirmed by the reduced mechanical modulus found in diseased tissue [6-8]. This has led to the desire to compare MRI images with and without loading; apart from mechanical devices that apply pressure on the extended leg, the alternate approach is by putting the patient into a natural, upright position and acquiring images in two positions, with and without load. To this end, lowfield scanners have become available that are able to rotate the magnet system with the patient fixed inside [9].

The existence of such low-field scanners, typically operating at field strengths well below 0.5 T, has highlighted the shortage of available NMR data on tissue in this respective field range, where it has been shown that in particular T<sub>1</sub> is strongly field-dependent. One further aspect is the occurrence of so-called quadrupolar dips in the relaxation profiles below about 80 mT [5]. These dips correspond to cross-relaxation phenomena of the observed <sup>1</sup>H with <sup>14</sup>N nuclei of nitrogen-containing compounds such as proteins and collagen. The dips are correlated to the concentration of these substances as well as their mobility, in the sense that freely tumbling molecules would not give rise to the extra relaxation mechanism due to motional averaging. Since osteoarthritis is correlated with a decrease of glycosaminoglycan (GAG) concentration and with an increase of water in the tissue [10], a decrease in the strength of these quadrupolar dips can be expected and has actually been proven in a pilot study [11]. In an investigation of enzymatically degraded tissue and its constituents, it was shown that both GAG and collagen contribute to the presence of quadrupolar dips [12]. However, the identification of these features requires an instrument capable of measuring T<sub>1</sub> at different field strengths; so far, only prototypes have been developed with imaging capability [13], while field-cycling relaxometers without imaging units are commercially available.

The purpose of this study is to combine different methods available at low magnetic field strengths, and to suggest suitable parameters that can be employed for a possible identification of osteoarthritis. To this end, we have combined, for the first time, a detailed study of field-cycling relaxometry with investigations at high spatial resolution in the profiling dimension, which can be provided by a single-sided NMR scanner, the NMR-MOUSE working at a field comparable to commercial MRI systems (0.27 T). The latter allows measurement of the samples under mechanical load and is thus suitable to quantify the structural changes occurring during tissue compression. To this end, a total of 20 human knee cartilage samples have been analyzed, first, for spatially resolved  $T_1$  with an NMR-MOUSE scanner, and subsequently studied with a variable-field relaxometer providing volume-averaged relaxation times.

#### 2. Experimental

A total of 20 osteochondral plug samples of 6 mm diameter were extracted from human tibial plateaus from patients undergoing total knee arthroplasty, and were stored frozen at -20 °C in tubes filled with phosphate buffer solution [14]. The experiments were

approved by the Ethical Committee of the Northern Ostrobothnia Hospital District, Oulu, Finland (191/2000).

Each sample was allowed to equilibrate for 24 h at  $+6 \degree C$  before being exposed to room temperature, and was then placed in a tightly fitting cylindrical container that allowed the application of mechanical load in the vertical direction via a hydrostatic pressure cell, with small holes drilled in order to release excess water. This cell was mounted on top of an NMR-MOUSE single-sided scanner (Magritek, Aachen, Germany) operating at a <sup>1</sup>H Larmor frequency of 11.7 MHz, and the relaxation times T<sub>2</sub> and T<sub>1</sub> of the tissue were determined with a one-dimensional resolution of 50 µm, averaging over the cylinder diameter of 6 mm. The same experiment was repeated under constant pressure of 0.6 MPa immediately afterwards. Consequently, samples were taken out of the cell and the cartilage was separated from the bone and calcified tissue. The cartilage samples were subsequently measured in a SpinMaster 2000 Fast Field Cycling relaxometer (Stelar, Mede, Italy), and T<sub>1</sub> dispersion was measured in the frequency range 10 kHz ... 20 MHz with particular emphasis on the region of quadrupolar dips between 1.5 and 4 MHz where sampling was performed with higher density. All signals were acquired at a detection field corresponding to the Larmor frequency of 16.7 MHz, followed by a single 90° pulse and integrating the FID. The signal decays were fitted by an exponential function; no significant deviation from monoexponential behavior was observed within the accuracy of these experiments.

Following the NMR protocol, samples were stored in a 10% formalin solution, then cut, stained and categorized according to the Mankin grading system by three individuals, the results of which were averaged. Mankin grading assigns numbers to certain properties of the tissue guided by visual inspection, covering the range between 0 (unaffected) and 14 (highest degree of OA) [15]. Pearson and Spearman (rank) correlation coefficients of all determined parameters with the averaged grade and among themselves, respectively, were computed; both values were mostly identical. Note that for a sample size of 20, all correlation coefficients magnitudes larger than 0.5 were considered significant, with a corresponding p < 0.05 value.

## 3. Results and discussion

T<sub>1</sub> and T<sub>2</sub> relaxation time profiles showed significant variation with increasing cartilage degeneration (Figs. 1 and 2). Furthermore, it becomes apparent that the general shape of the T<sub>1</sub> distribution is only weakly affected by load in the healthy tissue, whereas the reduction of particularly T<sub>1</sub> and the volume change is much more evident in the diseased tissue. Also, after compression the dynamic range of T<sub>1</sub> is reduced by about a factor of 3 compared to the situation before application of pressure. The pattern is similar for  $T_2$ , however, here the error bars are significantly larger, particularly for the diseased tissue, a fact that can be attributed to strongly multiexponential relaxation due to tissue heterogeneity within the sensitive region, i.e. a plane of 50 µm thickness. However, due to the limited SNR, a multiexponential analysis was not attempted at this stage. The common interpretation of both relaxation times is a reduction of the free water component with its long  $T_1$  ( $T_2$ ), due to water being expelled by the external pressure. The fraction of free water, due to pressure, is reduced while some of the residual water in the tissue remains in contact with the proteins and the collagen fibers, which together give rise to shorter relaxation times much as the surface relaxivity of an interface does in a porous medium. Additionally, the shortening of T<sub>2</sub> is also attributed to the realignment of the collagen network that leads to



**Fig. 1.**  $T_1$  across (a) healthy (Mankin grade 1) and (b) osteoarthritic (Mankin grade 11) cartilage plug measured at a <sup>1</sup>H Larmor frequency of 11.7 MHz. Filled symbols represent unloaded tissue, open symbols represent loading at 0.6 MPa. Depth is counted from the tissue surface.

altered magic angle conditions and associated dipolar relaxation effects. Assuming fast exchange conditions, one might approximate the measured relaxation times with the residual water content [10,16]. Note that the relation of different tissue density with corresponding variation of water diffusivity [17] and the restricted diffusion in the matrix [18] have been described before, as well as the connection with relaxation properties of the water protons themselves [19,20]. Due to the complex and unresolved microstructure of the tissue, an attempt towards an analysis with respect to a surface-to-volume weighted interaction is not made in this work.

Both healthy and degenerated cartilage samples showed significant  $T_1$  relaxation time dispersion (Fig. 3). At first sight, no obvious trend is observable, apart from a possibly more pronounced dispersion of the diseased tissue particularly at high fields. In order to separate the essential features, the exponent of apparent power-laws  $T_1 \sim \omega^{\alpha}$  was separately fitted for the regions below and above 1 MHz in Larmor frequency, and the area under the quadrupolar peaks, i.e. their shape when plotted as a function of inverse relaxation time  $R_1 = T_1^{-1}$ , being an additive parameter for relaxation contributions, was obtained phenomenologically by subtracting a polynomial fit to the background dispersion, which was shown to provide the smallest residuals (see Ref. [12]). Note that this is only an approximation of the actual contribution since the range of <sup>1</sup>H-<sup>14</sup>N interaction is not limited to the immediately visible dips, but it is sufficient for a quantitative comparison among datasets of similar properties.



**Fig. 2.** As in Fig. 1, but showing T<sub>2</sub>.

In order to identify correlations between observed parameters with (i) each other and (ii) the state of disease, quantities derived from the experimental data were statistically analyzed. From the  $T_1$  dispersion curves, these correlations were the mentioned power-law exponents above and below 1 MHz (a single, averaged value was considered insufficient to fit the experimental data), as well as the area of the quadrupolar dips. From the MOUSE measurements, these were, among others, the average and maximum values of  $T_1$  and  $T_2$ , the position of their relative maximum measured from the tissue surface, the position of the maximum in signal intensity, the tissue thickness, and the same parameters under load as well as the relative changes when comparing measurements with and without load. Only a selection of the correlations will be discussed here.

### 3.1. Correlation between osteoarthritis stage and quadrupolar dips

The correlation between Mankin grade and the area of the quadrupolar peaks in R<sub>1</sub> (see above) is shown in Fig. 4. A negative correlation coefficient of -0.67 was observed, i.e. more pronounced tissue degeneration corresponds to a weaker effect of  $^{1}H^{-14}N$  relaxation. This correlation has indeed been found in an earlier study [11], likewise for human cartilage samples. The finding is, in fact, expected, since the depletion of GAG as a consequence of OA is well known and accessible by Gd-based tracer addition [21], just as the increase of the water content [22]. Both effects point in the same direction, as was demonstrated by comparing the individual components GAG and collagen as a function of hydration [12]. While a decrease of available  $^{14}N$  leads to a corresponding decrease



**Fig. 3.** Relaxation time as a function of Larmor frequency,  $T_1(\nu)$ , for the cartilage samples from Fig. 1 and 2 (open symbols: Mankin grade 1; filled symbols: Mankin grade 11). Straight lines represent the fitted slopes for the two regions above and below 1 MHz, respectively. The insert shows a magnification of the region of the quadrupolar dips.



**Fig. 4.** Pearson correlation coefficients between Mankin Grade of cartilage tissues and the area under the quadrupolar relaxation rate peaks (description see text).

of the quadrupole interaction, a surplus of water will further lengthen  $T_1$  proportionally and may also lead to a higher mobility of the <sup>14</sup>N-containing species, thus reducing interaction even more. Previously a high correlation between  $T_1$  and cartilage water content has been demonstrated for bovine tissue [10]. Note, however, that in the derivation of correlations such as the mentioned one, the Mankin grade is necessarily considered as a linearly variable quantity, which must remain a gross approximation since it is an artificial, partially subjective measure of disease. However, the observed connection is confirmed by corresponding t-tests where the data set has been divided into "healthy" and "diseased" tissue (this discussion data will be presented in a more detailed manuscript).

#### 3.2. Maximum value of $T_1$

Both the actual value of the  $T_1$  maximum across the tissue, as well as its position with respect to the surface, are parameters that show correlation with the state of the tissue. As is demonstrated in Fig. 5a for the case without load, the position of  $T_{1,max}$  correlates with the Mankin grade, and is moving closer to the surface, a fact



**Fig. 5.** (a) Pearson correlation coefficients between Mankin Grade of cartilage tissues and the position of the maximum  $T_1$  as obtained at 11.7 MHz Larmor frequency without mechanical load. (b) Pearson correlation coefficients between Mankin Grade of cartilage tissues and the value of the maximum  $T_1$  without (filled symbols) and with mechanical load (open symbols) as obtained at 11.7 MHz Larmor frequency.

that is not surprising considered the effect of disease on the overall thickness of the tissue (see below). If one takes this into account by normalizing the relative position within the tissue, the correlation remains, but becomes statistically insignificant (r = -0.36). In contrast, no correlation is found for the position of  $T_{2,max}$ .

The actual value of  $T_{1,max}$  itself is found to correlate only in the compressed tissue (r = -0.49, see Fig. 5b).  $T_{2,max}$ , on the other hand, appears to correlate both with and without load, but the effect is generally more pronounced in the compressed cartilage (data not shown). Longer relaxation times in general point to water in an environment of lower concentration of GAG and/or collagen, thus experiencing less relaxation contributions; this water is preferentially removed under compression so that, in a qualitative way, it can be understood that the structural changes of OA tissue can be reflected in the relaxation times values. The fact that the observation is different for longitudinal and transverse relaxation, however, suggests that the explanation of this empirically found correlation is indeed less straightforward.

Note that these findings cannot be generalized for the average value of T<sub>1</sub>, which means that an "easy" assignment of tissue properties cannot be done in a low-resolution imaging device operating at low fields. On the other hand, correlations were indeed identified in a study carried out at 9.4 T [14]. It is, however, possible, at least in principle, to determine the spread of the relaxation time distribution, and thus the maximum and minimum values, by a



**Fig. 6.** Pearson correlation coefficients between Mankin Grade of cartilage tissues and relative thickness change without (filled symbols) and with mechanical load (open symbols).

thorough analysis of the signal decay. Whether this can be achieved reliably for the typical width of  $T_1$  in cartilage at 0.27 T, i.e. about a factor of 3 between shortest and longest components for healthy cartilage and considerably less for diseased cartilage, is the topic of ongoing research.

#### 3.3. Tissue thickness

During severe OA, the cartilage is known to become thinner; early stage OA may differ from this trend. Furthermore, degeneration is associated with cartilage softening. In this study, a negative correlation was found between Mankin grade and thickness in the unloaded (r = -0.58) and loaded tissue (r = -0.63), while the relative percentile thickness change under constant load correlates positively (r = 0.61, see Fig. 6). In a simplified view, the tissue becomes "softer" and will be compressed more, thereby losing its ability to maintain resistance against load during normal human activity. A stronger compressibility, corresponding to s smaller compression modulus or Young's modulus, has indeed been reported in several high-field studies [6-8]. In a loading/unloading experiment with sufficient spatial resolution, this relation can thus be exploited for diagnostic purposes. If such resolution is not available, different parameters need to be employed - for instance, a strong positive correlation between change in thickness and change in  $T_{1,max}$  was demonstrated (r = 0.66), while the latter itself did not correlate significantly with Mankin grade. This observation may provide a hint towards choosing an advanced processing of parameters that avoids the problematic assumption of Mankin grading as a linear function.

#### 3.4. Further observations

No compression experiments were carried out on the variable field relaxometer, so that only unloaded tissue samples could be analyzed. Apart from the area of the quadrupolar dips, the overall shape of the dispersion curve  $T_1(\omega)$  is of relevance, and it was successfully approximated by power-laws  $T_1 \sim \omega^{\alpha}$  with two distinct exponents in the regions below and above a limiting Larmor frequency of 1 MHz. At this stage, such an analysis must remain purely phenomenological, considering the complexity of the system. Nevertheless, a certain body of literature exists that interprets the behaviour of water interacting with proteins or surfaces, so that the

assumption of power-laws is not unrealistic [23]. In the present study, the exponent for frequencies below 1 MHz was found to be essentially constant ( $\alpha \approx 0.27 \pm 0.01$ ), but the value above 1 MHz was strongly varying between about 0.5 and 0.9, and a correlation was found, for instance, with thickness change and signal intensity change with/without load at 0.27 T, the latter corresponding to water content. Correlation with the Mankin grade itself is positive but below the level of statistical significance (r = 0.40).

# 4. Conclusions

Spatially resolved NMR parameter measurements of ex vivo human articular cartilage at constant, low magnetic field were combined with unresolved relaxation measurements at variable field, and were compared to a parameter quantifying the severity of osteoarthritis in the tissue. A number of significant correlations were identified, most notably the magnitude of the quadrupolar dips resulting from <sup>1</sup>H-<sup>14</sup>N cross relaxation, and the position of the maxima of T<sub>1</sub> within the tissue. However, more pronounced correlations could be found when subjecting tissue to physiologically meaningful lateral pressure (0.6 MPa) where the structural changes derived from OA resulted in significant effects on relaxation times, thickness and water distribution. This study serves as a first step towards establishing a strategy for in vivo low-field investigations of human joints, to be carried out at commercial low-field scanners or field cycling MRI equipment. These first results highlight the necessity of improved analysis tools for multi-exponential signal decays as well as of an alternative approach to grading of OA, which is more suitable for establishing correlations with the new methods presented in this study.

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# References

- [1] Y. Xia, T. Farquhar, N. Burton-Wurster, E. Ray, L.W. Jelinski, Magn. Reson. Med. 31 (1994) 273.
- [2] Y. Xia, J.B. Moody, H. Alhadlaq, Orientational dependence of T2 relaxation in articular cartilage: a microscopic MRI (μMRI) study, Magn. Reson. Med. 48 (2002) 460–469.
- [3] M.T. Nieminen, J. Rieppo, J. Toyras, J.M. Hakumaki, J. Silvennoinen, M.M. Hyttinen, H.J. Helminen, J.S. Jurvelin, Magn. Reson. Med. 46 (2001) 487–493.
- [4] E. David-Vaudey, S. Ghosh, M. Ries, S. Majumdar, Magn. Reson. Med. 22 (2004) 673–682.
- [5] E. Rössler, C. Mattea, A. Mollova, S. Stapf, J. Magn. Reson 213 (2011) 112–118.
- [6] L.A. Setton, D.M. Elliott, V.C. Mow, Osteoarthr. Cartil. 7 (1999) 2–14.
- [7] H.A. Alhadlaq, Y. Xia, J.B. Moody, J.R. Matyas, Ann. Rheum. Dis. 63 (2004) 709-717.
- [8] S. Knecht, B. Vanwanseele, E. Stüssi, Clin. Biomech. 21 (2006) 999–1012.
- [9] I.J. Dahabreh, N. Hadar, M. Chung, Ann. Intern. Med. 155 (2011) 616–624.
- [10] J.E. Berberat, M.J. Nissi, J.S. Jurmelin, M.T. Nieminen, Magn. Reson. Imaging 27 (2009) 727–732.
- [11] L. Broche, G.P. Ashcroft, D.J. Lurie, Magn. Reson. Med. 68 (2012) 358-362.
- [12] E. Rössler, C. Mattea, S. Stapf, Magn. Reson. Med. 73 (2015) 2005–2014.
   [13] D.J. Lurie, S. Aime, S. Baroni, N.A. Booth, L.M. Broche, C.H. Choi, G.R. Davies,
- S. Ismail, D. O'Hogain, K.J. Pine, Compt. Rend. Phys. 11 (2010) 136–148. [14] J. Rautiainen, M.J. Nissi, E.-N. Salo, V. Tiitu, M.A.J. Finnilä, O.-M. Aho,
- S. Saarakkala, P. Lehenkari, J. Ellermann, M.T. Nieminen, Magn. Reson. Med. 74 (2015) 249–259.
- [15] H.J. Mankin, H. Dorfman, L. Lippiell, A. Zarins, J. Bone Jt. Surg. A 53 (1971) 523–537.
- [16] S.Q. Zheng, Y. Xia, J. Magn. Reson 198 (2009) 188–196.

- [17] D. Burstein, M.L. Gray, A.L. Hartman, R. Gipe, B.D. Foy, J. Orthop. Res. 11 (1993) 465-478.
- [18] R. Knauss, J. Schiller, G. Fleischer, J. Kärger, K. Arnold, Magn. Reson. Med. 41 (1999) 285–292.
- (1933) 200–232.
   Y. Xia, T. Farquhar, N. Burton-Wurster, E. Ray, L.W. Jelinski, Magn. Reson. Med. 31 (1994) 273–282.
- [20] P. Fantazzini, F. Galassi, V. Bortolotti, R.J.S. Brown, F. Vittur, New J. Phys. 13 (2011) 065007.
- [21] A. Bashir, M.L. Gray, J. Hartke, D. Burstein, Magn. Reson. Med. 41 (1999) 857-865.
- [22] M. Venn, A. Maroudas, Ann. Rheumat. Dis. 36 (1977) 121–129.
  [23] G. Diakova, J.-P. Korb, R.G. Bryant, Magn. Reson. Med. 68 (2012) 272–277.