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Multifunctional Nanocomposites based on Bacterial Cellulose

by

Qisheng Jiang

A dissertation presented to
The Graduate School
of Washington University in
partial fulfillment of the
requirements for the degree
of Doctor of Philosophy

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List of Abbreviations

4-NP	4-Nitrophenol
AFM	Atomic force microscopy
ATCC	American type culture collection
AuNPs	Gold nanoparticles
AuNRs	Gold nanorods
BET	Brunauer-Emmett-Teller
BNC	Bacterial nanocellulose
CV	Capacitance-voltage
DLS	Dynamic light scattering
<i>E. coli</i>	<i>Escherichia coli</i>
EDLC	Electric double-layer capacitor
FTIR	Fourier-transform infrared spectroscopy
GCD	Galvanostatic charge/discharge
GO	Graphene oxide
HPA	Hypophosphorous acid
MB	Methylene blue
MO	Methyl orange
NIR	Near infrared
PBS	Phosphate-buffered saline
PDA	Polydopamine
PDMS	Polydimethylsiloxane

PdNPs	Palladium nanoparticles
PEDOT:PSS	Poly(3,4-ethylenedioxythiophene) polystyrene sulfonate
PET	Polyethylene terephthalate
pMBA	4- Mercaptobenzoic acid
Psi	Pound per square
PVA	Polyvinyl alcohol
R6G	Rhodamine 6G
RGO	Reduced graphene oxide
SCs	Supercapacitors
SEM	Scanning electron microscopy
SERS	Surface enhanced Raman scattering
TGA	Thermogravimetric analysis
UV-vis	Ultraviolet-visible
XPS	X-ray photoelectron spectroscopy
XRD	X-ray diffraction

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Qisheng Jiang

Washington University in St. Louis

December 2018

Dedicated to my parents, who brought me into this world and raised me to be a rebel against our
gene master.

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Abstract of the Dissertation

Multifunctional Nanocomposites based on Bacterial Cellulose

by

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Cellulose is biodegradable, renewable, and abundant in nature thus cellulose (or paper)-based products can be inexpensively produced and recycled. Among cellulosic materials, bacterial nanocellulose (BNC) draws a special research attention due to the inherent three-dimensional nanofibrous structure, excellent mechanical flexibility, high purity and well-defined surface chemistry, and cost-efficient, scalable and environment-friendly synthesis. BNC can be biosynthesized by *Gluconacetobacter xylinus*, which is the most characterized BNC producer among various microorganisms. BNC is composed of highly pure cellulose nanofibrils, produced from well-defined dextrose through biochemical steps and subsequent self-assembling of the secreted cellulose fibrils which has the dimension ranges from 25 to 100 nm in diameter from bacteria in the culture medium. During the biosynthesis of BNC, shape-controlled hydrogels with well-defined network structure pore diameters below 10 μm can be easily achieved. For all the above-mentioned reasons, BNC is a highly promising platform material for the fabrication of functional composites through *in situ* growth or adsorption of pre-synthesized nanostructures on the nanoscale cellulose fibers.

In this work, we have designed and demonstrated novel strategies to realize BNC-based functional nanocomposites with applications in sensing, water purification and energy storage. We have

demonstrated a BNC film-based surface enhanced Raman scattering (SERS) substrate which has 3D porous structure and ultrafine fibers with uniform and dense adsorption of plasmonic nanostructures, resulting large SERS enhancement and excellent uniformity of SERS activity.

For the first time, we have demonstrated a novel, highly scalable, cost-effective and green strategy to realize functional BNC-based foams/membranes. Functional materials such as graphene oxide (GO), polydopamine (PDA) can be efficiently incorporated within BNC matrix during its growth. Owing to the intercalation of functional materials within the layered BNC matrix, the functional composites showed excellent mechanical robustness and flexibility, which is crucial for efficient, large-scale applications, either as a foam or as a membrane.

Specifically, we have designed and developed a bilayered hybrid biofoam comprised of BNC and RGO and a completely biodegradable bilayered foam based on BNC and PDA for highly efficient solar steam generation, which can be a sustainable solution to alleviate global water crisis. An innovative water filtration membrane based on BNC and RGO which harvests sunlight to kill microorganisms has been developed to provide a novel anti-biofouling approach. We have also demonstrated a robust filtration membrane based on BNC loaded with GO and PdNPs, which exhibited excellent dye degradation performance for highly efficient wastewater treatment. Furthermore, the *in situ* fabrication approach has been extended to polymeric materials such as poly(3,4-ethylenedioxythiophene) polystyrene sulfonate (PEDOT:PSS) to realize hybrid flexible supercapacitor electrodes based on RGO, BNC and PEDOT:PSS.

The fabrication strategies and materials design demonstrated in this work can be easily extended to realize various BNC-based nanocomposites with applications in water purification, energy harvesting, sensing, catalysis, and life sciences.

Chapter 1: Introduction and Motivation

1.1 Bacterial Nanocellulose

Cellulose is biodegradable, renewable, and abundant in nature thus cellulose (or paper) based products can be inexpensively produced and recycled.¹ Among cellulosic materials, bacterial nanocellulose (BNC) draws a special research attention due to the inherent three-dimensional nanofibrous structure, excellent mechanical flexibility, high purity and well-defined surface chemistry, and cost-efficient, scalable and environment-friendly synthesis.²⁻⁶ BNC can be biosynthesized by *Gluconacetobacter xylinus* under static culture conditions, which is the most characterized BNC producer among various microorganisms.⁷ BNC is composed of highly pure cellulose nanofibrils, produced from well-defined dextrose through biochemical steps and subsequent self-assembly of the secreted cellulose fibrils from bacteria in the culture medium.^{6, 8-}
⁹ In contrast to paper substrates obtained from plant cellulose, which comprise of carbonyl and carboxyl functional groups along with hydroxyl, BNC possesses only hydroxyl functional groups.¹⁰ The dimensions of BNC nanofibrils range from 25 to 100 nm in diameter and several micrometers in length. BNC as a biomaterial offers the special benefit of *in situ* control of cellulose formation, such as the shape and supramolecular structure of the products.³ During the biosynthesis of BNC, shape-controlled hydrogels with well-defined network structure pore diameters below 10 μm can be easily achieved. The as synthesized BNC is harvested and processed to obtain nanocellulose hydrogel with extremely high-water content (~99% by weight). For all the above-mentioned reasons, BNC is a highly promising platform material for the fabrication of functional composites through *in situ* growth or adsorption of pre-synthesized nanostructures on the nanoscale cellulose fibers.¹¹⁻²⁰

1.2 Surface Enhanced Raman Scattering (SERS) and Flexible SERS swab

Surface enhanced Raman scattering (SERS) involves the large enhancement (under ideal conditions up to 10^{11} times) of Raman scattering signal from molecules adsorbed on or in close proximity to a nanostructured metal surface.²¹⁻²⁷ SERS is a highly promising transduction platform for the detection of trace levels of known and unknown chemical compounds with applications spanning from forensics to homeland security.²⁸⁻³³ The advantages of SERS as an analytical tool include (i) rich and unique molecular fingerprint corresponding to the target analytes; (ii) ultrahigh sensitivity (single molecule sensitivity under ideal conditions); (iii) absence of interference from water (making it tolerant to humidity variations); and (iv) ease of sample preparation. Extensive efforts have focused on the design and fabrication of SERS substrates with high sensitivity and large-scale uniformity. For many real-world applications, the ease and efficiency of the sample collection is very important. For example, collection and detection of trace amounts of hazardous and pathogenic chemical and biological analytes present on solid surfaces is a powerful screening method. Among various sample collection methods, physical swabbing has been recognized as a highly efficient and practical method to maximize the sample collection on a solid surface using a flexible and functional swab.^{34-35,36-37} Paper-based SERS swabs developed by our group have been successfully demonstrated to collect chemical analytes from solid surfaces.³⁸ However, past efforts have been limited to the use of off-the-shelf filter paper substrates, providing limited control over the micro/nanoscale roughness, which determines the contact between the SERS swabs and the surface of interest. Thus, novel methods and materials that offer better control over the micro/nanoscale structure of these flexible SERS swabs are of great interest.

1.3 Novel Materials and Processes for Energy-efficient Water Treatment

Water scarcity is recognized to be one of the most critical global issues of the 21st century as the demand for clean and safe water, critical for human survival, food production and manufacturing, continues to rise with increasing world's population.³⁹⁻⁴¹ Water scarcity is tightly intertwined with two other global challenges, namely, energy crisis and climate change. Innovative methods for producing fresh water from sea water (or unpurified water) with significantly reduced energy- and economic burden are paramount.

1.31 Solar steam generation and processes for energy-efficient water treatment

Solar energy is the most abundant and the cleanest source of energy available on earth that can be tapped into for addressing the global fresh water shortage. Steam generation through efficient harvesting of solar energy and thermal concentration is a highly promising technique and has attracted tremendous attention in the past few years.⁴²⁻⁴³ Various solar steam generation devices (also called as interfacial evaporators) have been demonstrated within the past four years.

From the device design standpoint, most of these interfacial thermal evaporators share a few common features: (i) the top surface is comprised of materials that exhibit broad light absorption over visible and near infrared wavelengths combined with an efficient photothermal conversion; (ii) support materials with low thermal conductivity to minimize the heat transfer from the evaporative surface to the bulk of the water; and (iii) open porous structure and hydrophilic nature of supporting materials that facilitate a rapid transport of water from bulk to the evaporative surface. It is highly desirable to utilize materials and processes that are sustainable and energy efficient in the fabrication of these interfacial thermal evaporators. For example, biodegradability of the evaporators is an important consideration since the disposal of these devices can quickly pose a

significant threat to the environment and marine ecosystems, considering that these evaporators will be most commonly utilized in aqueous environments.⁴⁴ Thus, evaporators based on renewable, biodegradable and scalable materials holds a great potential for ultimate solar steam generation applications.

1.32 Graphene oxide (GO)-based membranes for water purification

Graphene oxide (GO) has emerged as an excellent membrane material owing to the excellent mechanical strength, facile synthesis and the ability to be readily dispersed in water and restacked into membrane using various methods, including vacuum filtration, spin-coating and drop-casting.⁴⁵⁻⁴⁸ In contrast to graphene flakes, which have a strong tendency to stack and aggregate in aqueous solutions, GO flakes exhibits excellent water dispersibility and thus aqueous processability because of the presence of oxygen-containing functional groups (carboxyl, epoxy, hydroxyl, and carbonyl groups).⁴⁹

Owing to the excellent mechanical strength, facile synthesis and the ability to be readily dispersed in water and restacked into membrane using various methods, including vacuum filtration, spin-coating and drop-casting, GO has emerged as an excellent membrane material.⁴⁵⁻⁴⁸ However, the stability of current GO-based membranes is compromised under vigorous agitation, pH, and ionic strength variations that are within the range of feed water.⁵⁰ Thus, there is a need to develop new scalable approaches to fabricate stable GO-based membranes.

1.33 Photothermal effect as a unique solution for tackling membrane biofouling

In response to the water scarcity, various membrane technologies have emerged and are being actively investigated for water purification and reclamation.⁵¹⁻⁵³ However, fouling and consequent degradation in performance remain to be a ubiquitous problem in most of these membranes.⁵⁴

Specifically, mineral scaling, organic fouling, and biofouling are three major fouling mechanisms, which lead to either temporary or permanent decline in the water flux.⁵⁵ Among them, biofouling, which is ascribed to more than 45% of all membrane fouling, is the “Achilles heel” of the membrane technologies due to the difficulty associated with the complete removal of microorganisms.⁵⁶⁻⁵⁸

Various strategies, such as adjusting pH, adding disinfectants and biocides, introducing quorum sensing molecules to inhibit the formation of biofilm, have been suggested for controlling biofouling.⁵⁹⁻⁶² However, most of these strategies introduce considerable operational costs and/or potential hazardous contaminants. Researchers have also investigated the incorporation of nanomaterials (e.g., silver nanoparticles, titanium dioxide nanoparticles and graphene oxide nanosheets), polymers (e.g., polyethylene glycol, polyvinyl alcohol, polyvinylpyrrolidone and zwitterionic polymers), and other materials (e.g., small organic molecules and biomacromolecules) to engineer fouling-resistant membrane surfaces that can reduce biofilm growth and inactivate bacteria.^{55, 63-68} However, most of these methods involve additional thermal or chemical treatment steps. Furthermore, most of the above techniques are effective only for a short period of time because biofilm has the ability to gradually adapt to imposed harsh environments.⁵⁶ Even if 99.9% of biofilm is removed once, the residual cells are sufficient to grow back and form a new biofilm. Therefore, there is an immediate need for highly efficient and cost-effective methods that overcome biofouling on water purification membranes over a long period of time.

Photothermal effect, which involves the conversion of incident light into heat, can offer a unique solution, obviating the need for harsh chemical treatments to achieve bacterial lysis. In our previous study, gold nanostars grown on graphene oxide (GO) flakes modified on commercial membrane were utilized as nanoheaters. Owing to the photothermal properties of gold nanostars,

adjacent *Escherichia coli* were efficiently killed with laser irradiation to prevent the formation of biofilm.⁶⁸ While this study provided a promising example of utilizing photothermal effect to minimize biofouling on membranes, it would be more beneficial if the membrane itself is comprised of photothermal materials and even more promising, if renewable and sustainable solar illumination can be harnessed as a light source.

1.34 Metal and transition metal nanoparticles as a novel and efficient catalyst for dye-contaminated wastewater treatment

Water contamination caused by chemical and biological species released from industrial and agricultural practices is a formidable environmental challenge.^{51, 69-70} Among all the contaminants such as cleaning agents, agricultural chemicals, and noxious microorganisms, organic dyes, which are widely employed in textile, leather, cosmetic, pharmaceutical industries, pose significant threats to aquatic life and humans due to their carcinogenic and mutagenic nature.⁷¹⁻⁷³ Due to the high resistance of organic dyes to traditional biological, physical, and chemical treatment methods, there is a critical need for the development of highly efficient and cost-effective methods for dye degradation and removal from contaminated water.⁷⁴⁻⁷⁶

Noble and transition metal nanoparticles, which have been extensively employed in catalysis, organic synthesis, hydrogen storage, and water treatment, have shown great promise for organic dye removal.⁷⁷⁻⁸⁰ In particular, palladium nanoparticles (PdNPs) have attracted special attention for dye degradation owing to their excellent catalytic properties and low environmental impacts.⁸¹⁻⁸⁵ To achieve maximum catalytic activity and prevent aggregation of PdNPs, immobilizing PdNPs on various substrates such as carbon (e.g., carbon microspheres, carbon nanotubes, and graphene oxide), silica, and metal oxides (e.g., titanium dioxide) has been utilized.⁸⁶⁻⁹⁰ GO is a particular promising substrate material because of its high mechanical, chemical, thermal stability, and high

surface area.⁹¹⁻⁹⁶ Various self-assembled GO-based composites have been demonstrated for wastewater treatment.⁹⁷⁻¹⁰² To the best of our knowledge, a GO-based filtration membrane with transition metal nanoparticles as catalyst has not been reported since previous developed GO-based composites have limited aqueous stability and mechanical strength under pressure.

As mentioned in Chapter 1.1, bacterial nanocellulose (BNC) can serve as an ideal material for realizing functional composites by incorporating functional nanomaterials within BNC matrix because of its desirable properties such as excellent mechanical strength and toughness, rich chemical functionality, and high specific surface area in aerogel state. It is highly desirable to realize a composite based on BNC and GO/PdNPs, which synergizes high catalytic activity and mechanical robustness.

1.4 Flexible supercapacitors and paper-based supercapacitors

1.41 Hybrid materials for high performance supercapacitor (SCs)

With an explosive development of portable and flexible electronics such as bendable mobile phones, flexible displays and wearable devices, there is an urgent need for high performance, flexible energy-storage devices.¹⁰³⁻¹⁰⁹ Supercapacitors (SCs) have emerged as an important class of energy storage devices due to their numerous advantages such as high power density, long cycle-lifetime and low environmental impact.¹¹⁰⁻¹¹² Owing to various advantages compared to other energy storage devices such as flexibility, lightweight, reliability, and safe operation over a wide temperature range, extensive efforts have been devoted to developing flexible all solid-state SCs, which hold great potential for next generation of flexible energy storage devices.¹¹³⁻¹¹⁹

Generally, active materials used in supercapacitors can be classified into two groups: i) Electrochemical double layer capacitor (EDLC) materials, which physically store charges via

reversible ion adsorption at the electrode-electrolyte interface; ii) Pseudocapacitors materials, which chemically store charges via redox reaction at the vicinity of the surface. Typically, EDLC SCs rely on carbon-based materials (including activated carbon, carbon nanotubes, graphene, reduced graphene oxide (RGO) etc.), while pseudocapacitors typically involve transition metal oxides (*e.g.* RuO₂, MnO₂, CoO_x, NiO, Fe₂O₃, etc.) and conducting polymers (*e.g.* polypyrrole, polyaniline, poly(3,4-ethylenedioxythiophene), etc.). EDLC material core and pseudocapacitive material shell constitutes an attractive combination of core–shell design due to the following merits: i) EDLC materials as core have better cycle stability due to their electrostatic charge and discharge storage mechanism (*i.e.* no phase changes), which makes them robust backbones even for large number of cycles; ii) EDLC core material also have better electrical conductivity compared to pseudocapacitive materials, which facilitates efficient transport of electrons; iii) pseudocapacitive materials as shell can effectively offset the low capacitance contribution from EDLC materials core as they can introduce Faradaic charge transfer due to reversible redox reactions upon charging and discharging, which leads to higher capacitance and energy density.¹²⁰⁻¹⁴⁰

1.42 Flexible paper-based SCs and bacterial nanocellulose-based SCs

Soft and compliant elastomers or plastics such as polydimethylsiloxane (PDMS) and polyethylene terephthalate (PET) are often used as substrates or packaging materials for the fabrication of flexible SCs. However these plastics introduce considerable cost and environmental impact after disposal.¹⁴¹ Cellulose-based paper substrates, on the other hand, are a low cost, highly flexible, biodegradable option, which possess a fibrous structure, which facilitates interactions with active materials and also provides pathways for ion transport, making them ideal substrates for flexible supercapacitors and energy storage devices in general.¹⁴²⁻¹⁴⁶

BNC shows great promise for the fabrication of functional composites through *in situ* growth or adsorption of pre-synthesized nanostructures on the nanoscale cellulose fibers due to its excellent mechanical properties, tunable porosity, chemical functionalizability, ease of synthesis, high scalability.^{11, 13, 19, 147-148} Due to these advantages, BNC has been adapted for realizing paper based flexible supercapacitors.¹⁴⁹⁻¹⁵³ However, in order to make high performance BNC-based supercapacitors, the fabrication techniques typically involve vacuum filtration^{152, 154}, pyrolysis activation^{150, 155-156}, which offer poor prospects in terms of scalability and/or compromise the mechanical properties of BNC. These considerations highlight the need for a radically different approach to integrate active materials with BNC that not only preserves the favorable nanofibrous structure and mechanical properties of BNC but also synergistically enhances the energy storage performance of the active materials.

1.5 Research Goals and Objectives

The ultimate goal of this research effort is to design and demonstrate novel bacterial nanocellulose (BNC)-based functional composites with tailored properties for diverse emerging applications, such as chemical sensing, water treatment and energy storage (Figure 1.1). Towards this goal, we have pursued the following specific objectives:

Objective 1: Design and demonstrate a flexible surface enhanced Raman scattering (SERS) swab based on bacterial nanocellulose.

Objective 2: Design and demonstrate novel materials and processes for energy-efficient water treatment using bacterial nanocellulose-based composites.

Objective 2A: Design and develop a strategy of realizing scalable and efficient reduced graphene oxide and bacterial nanocellulose interfacial photothermal evaporator for highly efficient solar steam generation.

Objective 2B: Design and demonstrate a completely biodegradable interfacial photothermal evaporator using polydopamine microparticles and bacterial nanocellulose.

Objective 2C: Design a biofouling-resistant ultrafiltration membrane by harnessing sunlight.

Objective 2D: Design and demonstrate a novel membrane for organic dye-contaminated wastewater treatment.

Objective 3: Design and demonstrate a novel bacterial nanocellulose-based flexible supercapacitor electrode materials.

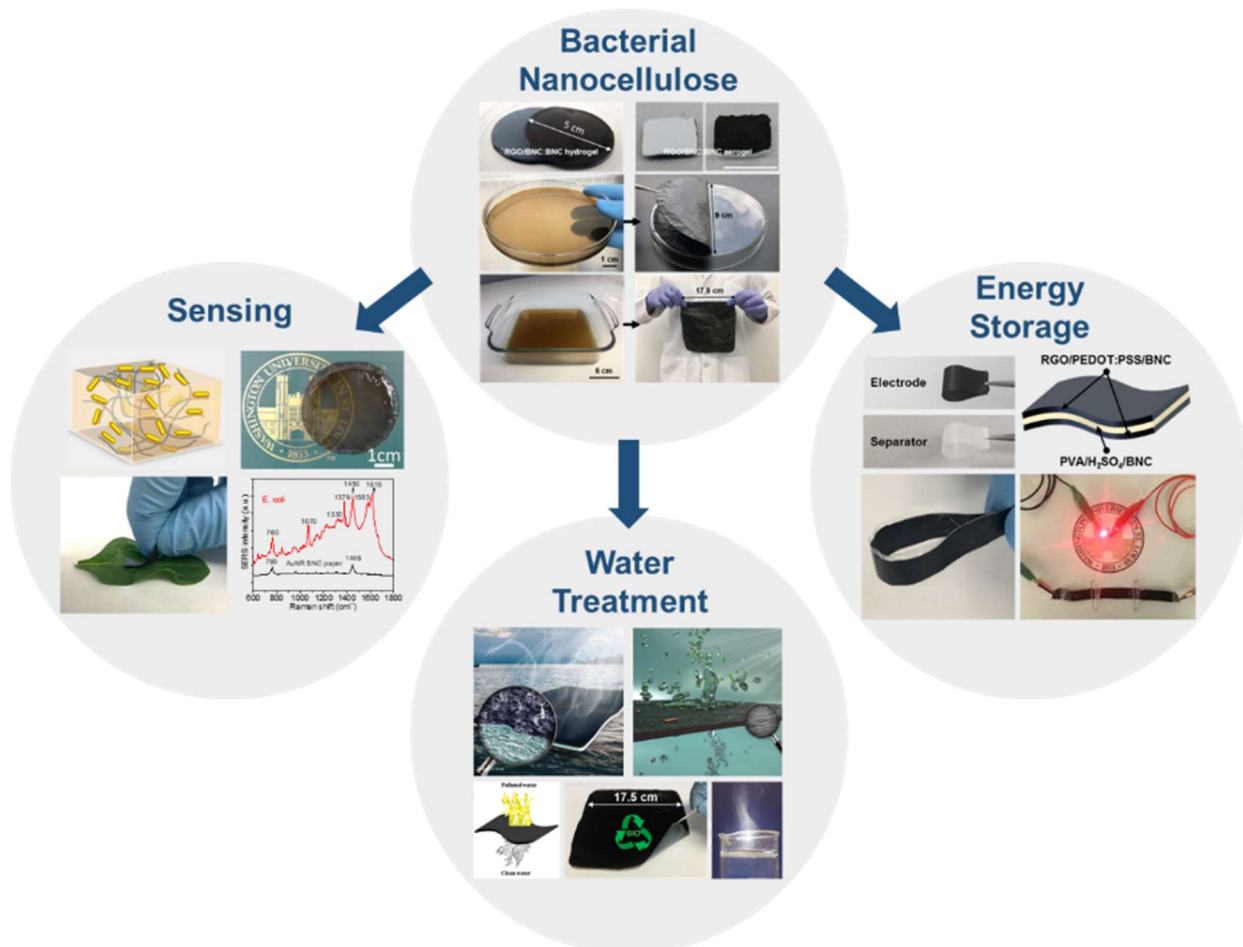


Figure 1.1. Illustration outlining the objectives of the overall research focus.

1.6 Overview of the Dissertation

The current chapter (Chapter 1) provides an overview of the research efforts and provides a brief background of BNC and functional composites based on BNC for application in sensors, water and energy.

Chapter 2 describes a BNC film-based SERS substrate fabricated by gravity-assisted filtration method. Due to the 3D porous structure of BNC and its ultrafine fibers network, the uniform and dense adsorption of plasmonic nanostructures on the surface and sub-surface regions lead to large SERS enhancement and excellent uniformity of SERS activity over the entire substrate. The ultra-smooth BNC-based SERS substrate serves as an ideal platform for the collection, detection and recognition of microorganisms.

In Chapter 3, we introduce an approach for the fabrication of photothermally-active biofoam that involves the *in situ* incorporation of GO flakes into BNC during its growth. We also demonstrate the versatility of the approach by making a bilayer foam simply achieved by sequential addition of BNC growth medium. The bilayered GO/BNC composite foam is tailored for high optical absorption, photothermal conversion, heat localization and water transport, resulting in a highly efficient solar steam generation.

Chapter 4 describes a complete biodegradable composites foam made of BNC and PDA as a interfacial photothermal evaporator for solar steam generation. In addition to 2D materials such as GO, we demonstrate that 0D nanomaterials such as PDA particles can also be incorporated into BNC during its growth, leading to a complete biodegradable foam with comparable solar steam generation performance.

In Chapter 5, a novel anti-biofouling ultrafiltration membrane based on BNC and GO is described, which involves *in situ* incorporation of GO flakes into BNC during its growth and air-drying. In

stark contrast to previously reported GO-based membranes for water treatment, the RGO/BNC membrane exhibited excellent aqueous stability under environmentally relevant pH conditions, vigorous mechanical agitation/sonication and even high pressure. Importantly, the membrane exhibited bactericidal capability enabled by light, due to its excellent photothermal properties, which lead to efficient bacterial lysis, obviating the need for any treatment of the feed water or external energy.

Chapter 6 introduces a simple and scalable approach for the fabrication of Pd/GO/BNC membrane for highly efficient dye degradation. The fabrication of Pd/GO/BNC membrane involves the incorporation of GO flakes into the BNC matrix during its growth followed by *in situ* growth of PdNPs on GO flakes. The layered structure of Pd/GO/BNC, which forms nanocapillaries throughout the membrane, maximizes the contact between organic dye contaminants and PdNPs anchored on the GO flakes.

In Chapter 7, we demonstrate that BNC can serve as an ideal layered matrix for incorporation of active electrochemical active materials to realize flexible and scalable supercapacitor electrode materials. GO flakes and PEDOT:PSS were incorporated into bacterial nanocellulose (BNC) matrix during its growth for the fabrication of flexible and light-weight electrodes for high performance supercapacitors. The fabricated flexible super-capacitor electrode exhibited a high specific capacitance and stability over 1000 testing cycles. Solid-state supercapacitor formed by sandwiching a BNC-based separator between RGO/PEDOT:PSS/BNC electrodes exhibited excellent energy storage performance and mechanical flexibility.

Chapter 2: Bacterial Nanocellulose-based Flexible Surface Enhanced Raman Scattering Substrate

2.1 Abstract

Owing to high purity, simple surface chemistry and three-dimensional (3D) nanofibrous structure, biosynthesized bacterial nanocellulose (BNC) is a highly attractive biomaterial for a wide range of applications. Previously, we demonstrated that conventional cellulose-based laboratory filter paper, adsorbed with plasmonic nanostructures can be employed as a flexible surface enhanced Raman scattering (SERS) substrate. In this work, we report a BNC film-based SERS substrate fabricated by gravity-assisted filtration method. The 3D porous structure of BNC facilitates uniform and dense adsorption of plasmonic nanostructures on the surface and in sub-surface regions, which results in large SERS enhancement. Furthermore, significantly lower surface roughness of BNC compared to conventional filter paper resulted in an excellent uniformity of SERS activity across the entire substrate. Harnessing the smooth surface of BNC, we show that BNC-based SERS substrate serves as an ideal platform for collection, detection and recognition of bacteria. The 3D plasmonic BNC composites demonstrated here are highly attractive for a broad range of applications including sensing, catalysis, and energy harvesting.

2.2 Introduction

Due to the numerous advantages, such as significant reduction in cost, high specific surface area, excellent wicking properties, and compatibility with conventional printing approaches (enabling multiplex detection and easy disposability), paper is gaining increased attention as a potential

substrate in chemical/biological sensing, tissue engineering, energy harvesting storage and implementing engineered gene circuits.¹⁵⁷⁻¹⁶²

Previously, we demonstrated that conventional laboratory filter paper decorated with plasmonic nanostructures could be employed as a flexible SERS substrate.^{34, 163-165} Very recently, we demonstrated 3D plasmonic foam, composed of bacterial nanocellulose aerogel with a high density of plasmonic nanostructures adsorbed on the cellulose nanofibrils.¹⁶⁶ The plasmonic biofoam provided extraordinarily large plasmonically-active sensing volume (under the laser footprint) compared to 2D substrates (*e.g.* filter paper). However, the thickness of plasmonic foams (1-5 mm) fabricated by adsorbing nanostructures on 3D cellulose network using immersion method is much higher than the light penetration depth (few tens of microns). Thus, there is a need to develop new fabrication technique to maximally utilized the plasmonic nanostructures.

In this work, to maximally utilize the plasmonic nanostructures, we employ a filtration method to localize the nanostructures to the top portion of the hydrogel while the bottom portion serves as a support layer. Furthermore, filtration method makes the processing much faster (30 min) compared to the immersion approach (typically more than 12 hours). We demonstrate that naturally-dried plasmonic BNC film with an ultrasmooth surface can serve as a mechanically-stable and flexible SERS substrate for efficient collection, recognition and detection of pathogenic bacteria.

2.3 Experimental Section

Synthesis of gold nanorods (AuNRs): Gold nanorods were synthesized using a seed mediated approach.¹⁶⁷⁻¹⁶⁸ Seed solution was prepared by adding 0.6 ml of an ice-cold sodium borohydride (NaBH_4) aqueous solution (10 mM) into a mixture of 9.75 ml of cetyltrimethylammonium bromide (CTAB, 0.1 M) and 0.25 ml of chloroauric acid (HAuCl_4 , 10 mM) solution under vigorous stirring

at room temperature (1000 rpm). The color of the seed solution immediately changed from yellow to brown after NaBH₄ addition. Growth solution was prepared by mixing 95 ml of CTAB (0.1 M), 1.0 ml of silver nitrate (10 mM), 4.75 ml of HAuCl₄ (10 mM), and 0.55 ml of ascorbic acid (0.1 M) in the same order, followed by gentle shaking to homogenize the solution. To the resulting colorless solution, 0.12 ml of freshly prepared seed solution was added to the growth solution. After 24 h of aging, AuNR solution was centrifuged at 10,000 rpm for 15 min to remove excess reactants and dispersed in nanopure water (18.2 MΩ-cm). The centrifugation was repeated once more to further remove excess CTAB for AuNRs/BNC film and AuNRs filter paper preparation.

Bacterial nanocellulose (BNC) and AuNRs/BNC substrate preparation: *Gluconacetobacter hansenii* (ATCC[®]53582) was grown in test tubes containing 6 ml of #1765 broth at 30°C under shaking at 250 rpm. The #1765 broth is composed of 2% (w/v) glucose, 0.5% (w/v) yeast extract, 0.5% (w/v) peptone, 0.27% (w/v) disodium phosphate, and 0.5% (w/v) citric acid. After 3 days of growth, 1.0 ml of the above solution was inoculated to 6ml of broth in a 6 cm petri dish, followed by gentle mixing and kept covered at room temperature without disturbance. After 3 days, a thin film of bacterial nanocellulose (~1.0 mm) formed on the air-liquid interface. To purify the film, the grown film was washed in a 500 ml of 0.1 M NaOH aqueous solution under boiling for 1 h, followed by dialysis for 2 days in nanopure water. To prepare AuNRs/BNC substrate, 3 ml of concentrated AuNRs solution with extinction intensity ~20 was filtered through a hydrated BNC film, which was placed on a glass filter. Once the infiltration is complete within 30 min, AuNRs/BNC hydrogel film was stored in hydrated condition for further use. The AuNRs density in the final SERS substrate can be tuned by varying the concentration of AuNRs in the feed solution.

Spectroscopy measurements: SERS substrates were exposed to R6G or pMBA solutions of known concentrations for 1 hour followed by thorough rinsing to remove unadsorbed or weakly bound molecules. Subsequently, SERS substrates were left to naturally dry before measurements. Raman spectra were obtained using a Renishaw inVia confocal Raman spectrometer mounted on a Leica microscope equipped with 20X objective (NA = 0.4) and 785 nm wavelength diode laser (0.7 mW). The spectra were obtained in the range of 600–1800 cm^{-1} with 10 s exposure time. Five spectra were collected from different spots across each sample. Shimadzu UV-1800 spectrophotometer was employed for collecting UV-vis extinction spectra from solution and the AuNRs/BNC substrate.

***E.coli* swab test:** *E.coli* (ATCC[®]35218) was cultured in nutrient broth medium for 18 hours at 30°C under shaking at 250 rpm until they reached their stationary growth stage. This corresponded to a bacteria concentration in the range of 10^7 - 10^8 CFU/ml.¹⁶⁹ After washing twice with phosphate buffer saline (PBS) buffer by centrifugation, 10 μl of the *E.coli* suspension was spread on a spinach leaf surface with surface area of $2 \times 2 \text{ cm}^2$. Following natural drying of *E.coli* on the spinach leaf, plasmonic BNC paper of $1 \times 2 \text{ cm}^2$, wetted with PBS buffer, was swabbed on the leaf surface with pre-spread *E. coli*. After collecting *E.coli* with plasmonic BNC paper, the samples were characterized using optical microscopy and Raman spectroscopy.

Microscopy Characterization: Scanning electron microscope (SEM) images were obtained using a FEI Nova 2300 Field Emission SEM at accelerating voltage of 10 kV. Plasmonic paper was gold sputtered for 60 sec prior to SEM imaging. Atomic force microscopy (AFM) images were obtained using Dimension 3000 (Digital instruments) AFM in light tapping mode.

2.4 Results and Discussion

2.41. Comparison of BNC film and Common Filter Paper

Bacterial nanocellulose (BNC) was biosynthesized under static culture conditions using *Gluconacetobacter xylinus*, which is the most extensively characterized nanocellulose producing bacterium among various microorganisms.⁷ BNC film is composed of highly pure cellulose nanofibrils, produced from dextrose through a series of biochemical steps followed by the self-assembly of the secreted cellulose fibrils from bacteria in the culture medium.^{6, 8-9} Similar to plant-derived cellulosic materials, BNC possesses surface hydroxyl groups, but does not contain carbonyl or carboxyl functional groups.^{6, 10} Owing to pure surface chemistry, nanofibrous and microporous structure, the properties of BNC are significantly different from those of conventional filter paper.¹⁷⁰⁻¹⁷² After growing the BNC film to a desired thickness (~1 mm), it is harvested and washed using sodium hydroxide and water to remove bacteria and residual growth medium (see Experimental Section for details). The resulting hydrogel is comprised of a highly swollen network of nanoscale cellulose fibers with up to 99 wt% water.

BNC exhibits strikingly different 3D structure and surface morphology compared conventional filter paper. Freeze-dried BNC aerogel exhibits large surface area, composed of nanofibrils of 20-100 nm in diameter (see appendix 1, Figure S1.1).¹⁶⁶ Considering that bacteria form the cellulose fibers at the air/liquid interface, the cellulose nanofibrils are preferentially oriented parallel to the surface (i.e. normal to the thickness) of the aerogel structure, which results in denser physical entanglements of the cellulose nanofibrils parallel to the surface compared to that along the thickness.^{3, 6} A representative cross-section SEM image of BNC aerogel reveals dense layers of cellulose fibers separated by a layer of sparse “tie” fibers resulting in a layered structure (Figure S1). BNC film of ~20 μm thickness, obtained by naturally drying ~1 mm thick BNC hydrogel,

exhibited a smooth surface morphology over a large scale due to the irreversible collapse of cellulose nanofibril network during dehydration (Figure 2.1A). AFM images revealed the RMS surface roughness of dry BNC film to be 25.6 nm over $3 \times 3 \mu\text{m}^2$ area (Figure 2.1B). In contrast, conventional filter paper exhibited an extremely rough, hierarchical fibrous morphology (Figure 2.1C). The filter paper largely consisted of micro scale (up to $20 \mu\text{m}$) cellulose fibrous strands interwoven together. Smaller microfibers made part of the large fibrous structure with nanofibers braided in between. In a relatively smooth area, the RMS surface roughness of the paper was measured to be 59.6 nm over $3 \times 3 \mu\text{m}^2$, which is still much higher compared to the naturally-dried BNC film.

2.42. Preparation and Characterization of Plasmonic BNC

We employed gold nanorods (AuNRs) as plasmonic nanostructures to fabricate a 3D SERS substrate based on BNC. Owing to their tunable localized surface plasmon resonance (LSPR) and strong electromagnetic field at the edges, AuNRs have been widely used for sensing applications.¹⁷³⁻¹⁷⁵ We employed simple filtration method to realize 3D plasmonic hydrogel, which enables the uniform adsorption and maximal utilization of plasmonic nanostructures on porous cellulose nanofibrils (Figure 2.2). Simple filtration not only facilitates the transport and adsorption of AuNRs into porous BNC hydrogel, but also makes the processing much faster (30 min) than immersion approach (typically more than 12 hours).

One of the distinct advantages of nanocellulose-based materials compared to conventional filter paper is the optical transparency. Transparent cellulose-based materials have gained significant interest in recent years as substrates for various electronic and optoelectronic devices.^{162, 176} The optical transmittance of BNC hydrogel was measured to be $\sim 80\%$ at 600 nm, which is significantly higher compared to that of the opaque filter paper (Figure 2.3A). The AuNRs employed in

plasmonic hydrogel were 51 ± 3 nm long and 14 ± 1 nm in diameter, making the aspect ratio approximately 3.6 (see appendix 1, Figure S1.2). UV-Vis extinction spectra of the AuNRs solution showed the two characteristic peaks at ~ 513 nm and 768 nm corresponding to the transverse and longitudinal plasmon resonances of AuNRs, respectively (Figure 2.3B).¹⁷⁷ After filtration of the AuNR solution through the BNC hydrogel, the AuNRs were either adsorbed or trapped within the first few microns of the BNC hydrogel. The colorless filtrate contained no AuNRs as confirmed by the absence of characteristic LSPR bands of AuNR in the UV-vis extinction spectra. Extinction spectrum obtained from plasmonic hydrogel fabricated with filtration method showed a red shift in longitudinal plasmon resonance wavelength with a slight broadening of the LSPR band, compared to the gold nanorod solution (Figure 2.3B). The observed red shift and LSPR band broadening can be attributed to plasmonic coupling between closely packed AuNRs and an effective increase in the refractive index of the surrounding medium (from water to water+cellulose). Due to the higher refractive index sensitivity of the longitudinal plasmon resonance compared to the transverse band, the longitudinal plasmon band exhibited a larger red shift (~ 25 nm) compared to the transverse band (< 5 nm).¹⁷⁸

We further examined the distribution of AuNRs incorporated in the plasmonic hydrogel. SEM images revealed uniform and dense adsorption of the AuNRs on the cellulose nanofibrils surface without any signs of large scale aggregation (Figure 2.3C). Furthermore, the presence of AuNRs in the subsurface regions (embedded under the cellulose fibrils at the surface) of the porous hydrogel is revealed by SEM images (inset of Figure 2.3C and Figure S1.3 in appendix 1). From a number of SEM images, the number density of AuNR monolayer on the BNC film surface was found to be $\sim 100/\mu\text{m}^2$ (Figure 2.3C). Based on the amount of AuNRs in the feed solution and the surface coverage of AuNR on the BNC, we estimate that approximately first 10 layers of

cellulose fibers in the BNC hydrogel are decorated with AuNRs. Gravity-assisted infiltration of AuNRs into the highly swollen BNC hydrogel enables the incorporation of AuNRs into the subsurface layers of BNC. Following the infiltration, the BNC hydrogel is naturally dried, thus collapsing the fiber network and trapping the nanostructures within the network. The overall distribution of AuNRs in the plasmonic hydrogel can be tuned by varying the concentration of AuNRs and the volume of the feed solution. Following the incorporation of AuNR into the BNC, even vigorous rinsing with water or alcohol did not result in noticeable desorption of the AuNRs from BNC, suggesting the stability of the BNC-based SERS substrate for deployment in liquid environments. Cellulose has a large number of hydroxyl groups, which facilitates the adsorption of positively charged species.¹⁷⁹⁻¹⁸⁰ The uniform, irreversible, and high density adsorption of the AuNRs is possibly due to the electrostatic and van der Waals interactions between the positively charged AuNRs and the hydroxyl-rich cellulose surface.¹⁸¹ In case of filter paper, AuNRs were found to form a monolayer on the surface of the microfibers of the paper surface. The number of AuNR bound on the surface of the filter paper substrate was found to be in agreement with the amount of AuNRs absent in the solution after incubation of the paper substrate, which confirms the absence of AuNR in subsurface regions (Figure 2.3D). Owing to ability to load AuNRs in the subsurface regions, the amount of AuNRs loaded in the BNC-based plasmonic hydrogel was found to be 10X higher than that adsorbed on the filter paper. Such higher loading of plasmonic nanostructures is expected to result in higher SERS activity of BNC compared to plasmonic paper.

2.43. SERS Performance of Plasmonic BNC

Now we turn our attention to the SERS performance of plasmonic hydrogel in comparison to the 2D plasmonic paper we developed previously. We exposed plasmonic hydrogel and plasmonic paper with similar surface coverage of AuNRs to the same concentration of rhodamine (R6G), a

model analyte with a strong affinity to gold. SERS spectrum of R6G adsorbed on AuNRs exhibited strong Raman bands at 612, 1311, and 1363/1509 cm^{-1} , which correspond to C-C-C ring in-plane vibration, C-O-C stretching, and aromatic C-C stretching vibration, respectively (Figure 2.4A).¹⁸²⁻

¹⁸³ The intensity of the SERS spectra obtained from plasmonic hydrogel was found to be nearly 10X higher than that from plasmonic paper.¹⁸¹ The significantly superior SERS performance of plasmonic hydrogel can be ascribed to the large plasmonically-active surface area of 3D BNC with higher number of AuNRs within the excitation laser footprint. While the porous nature of BNC hydrogel facilitates the diffusion and adsorption of the analytes on subsurface AuNR, the optical transparency of BNC ensures the excitation of the subsurface AuNR. Figure 2.4B shows the SERS spectra obtained from plasmonic hydrogel after exposure to different concentrations of R6G. The SERS intensity of the characteristic Raman bands of R6G increased with increasing concentration of the model analyte. The most intense Raman band at 1363 cm^{-1} was used to analyze the trace detection ability of plasmonic hydrogel. The plot of SERS intensity at Raman band 1363 cm^{-1} vs. concentration of R6G obtained from plasmonic hydrogel shows a monotonic increase of the SERS intensity with increasing concentrations of R6G (Figure 2.4C). From the higher resolution spectra, the Raman bands corresponding to R6G are clearly distinguishable (signal to noise ratio (SNR) > 3) down to a concentration of 1 nM (Figure 2.4D).

Apart from high SERS intensity, uniformity of SERS signals across the substrate is extremely important for quantitative detection of the analytes of interest.¹⁸⁴ The homogeneity of SERS signals determines the noise floor of the spectral intensity, which in turn determines the limit of detection. We set out to investigate the SERS uniformity of plasmonic hydrogel compared to plasmonic paper. To demonstrate that the plasmonic hydrogel can be used in other common solvents (i.e. not limited to water), we chose p-mercaptobenzoic acid (pMBA) as a model molecule

to investigate SERS uniformity. The SERS measurement was performed after exposing the plasmonic hydrogel to 1 mM of pMBA in ethanol, followed by thorough rinsing with ethanol. Bright-field optical images revealed the smooth surface morphology of BNC after natural drying (Figure 2.5A). This smooth surface morphology of BNC film, which in agreement with the SEM images described above, results from the irreversible collapse of BNC nanofibrous structure. A SERS map was collected over a $26 \times 34 \mu\text{m}^2$ area (indicated in the bright-field optical image) of plasmonic hydrogel after exposure to pMBA and naturally drying (Figure 2.5B). The SERS intensity map corresponding to the Raman band at 1590 cm^{-1} (aromatic ring mode of pMBA) exhibited remarkable uniformity with a relative standard deviation of $\sim 4\%$, calculated from 221 pixels in the map. Three representative spectra from different regions of the SERS map (indicated in the SERS map) further confirm the small variation in the SERS activity over the mapped region (Figure 2.5C). In contrast to BNC-based SERS substrate, the optical micrograph of plasmonic paper reveals the microfibrillar structure and rough surface morphology as evidenced by the focus variations within the field-of-view (Figure 2.5D). The SERS map obtained from plasmonic paper over a $30 \times 30 \mu\text{m}^2$ area revealed much larger variation in the intensity compared to that from the BNC substrate (Figure 2.5E). The spatial variation in the SERS intensity largely stems from the rough surface morphology of the paper, which results in focus variations within the mapped region. Representative SERS spectra from different regions of the SERS map confirm the larger variations in the intensity of SERS signals compared to that from BNC-based SERS substrate (Figure 2.5F). The relative standard deviation of the SERS intensity from the plasmonic paper substrates was found to be $\sim 25\%$, calculated from 225 pixels in the map, which is significantly higher compared to the BNC substrate.

2.44. BNC-based SERS Swab for Bacteria Collection and Detection

Owing to the extremely high sensitivity and strain-specific spectroscopic signature, SERS is considered to be a powerful tool to detect pathogens for food safety and healthcare applications.¹⁸⁵⁻

¹⁸⁸ Typical SERS substrates are based on rigid substrates, such as silicon and glass, which are not suitable for sample collection through physical swabbing. BNC-based SERS substrate developed here can be employed as a flexible SERS swab to collect pathogens present on various surfaces.

More importantly, the smooth surface morphology of BNC facilitates the easy recognition of bacteria for obtaining strain-specific fingerprint. As a proof-of-concept, we chose *Escherichia coli* (*E. coli*) as a model pathogen to demonstrate the efficient collection, recognition and detection of pathogens using a BNC-based SERS swab. To demonstrate the unique sample collection ability,

we swabbed a slightly wet (in buffer) plasmonic BNC substrate on the surface of a spinach leaf with $\sim 10^5$ *E. coli* spread over 2×2 cm² area (Figure 2.6A). As expected, owing to the low surface roughness of BNC substrate, the collected bacteria could be clearly visualized using optical microscope under both bright field and dark field illumination for further SERS spectroscopic analysis (Figure 2.6B, 2.6C). SERS spectra obtained from individual *E. coli* collected on the SERS

swab showed the characteristic Raman bands of *E. coli*, which can be easily analyzed considering the minimal interference from the BNC-based SERS substrate (Figure 2.6D). Four prominent Raman bands attributed to *E. coli* were observed: 1070 cm⁻¹ corresponding to O=P-O⁻ stretching in DNA, 1379 cm⁻¹ corresponding to COO⁻ stretching, 1583 cm⁻¹ corresponding to guanine and adenine of DNA, and 1618 cm⁻¹ corresponding to tyrosine, phenylalanine and amine I of *E. coli*.^{187,}

¹⁸⁹ Using additional data processing methods such as principal component analysis, it has been demonstrated that SERS can be employed to distinguish between different species and to classify different bacteria strains.¹⁹⁰ Considering the highly efficient sample collection, excellent

sensitivity and ease of fabrication, the flexible SERS substrate demonstrated here can serve as a promising platform for rapid collection and identification of microorganisms.

2.5 Conclusions

In conclusion, we have demonstrated a flexible SERS substrate based on BNC uniformly adsorbed with AuNRs through simple filtration method. Apart from the highly dense adsorption on the surface, the AuNRs are trapped within the subsurface layers (about 10 layers) of BNC, which result in nearly an order of magnitude higher SERS intensity compared to the SERS substrate based on filter paper with similar surface coverage of plasmonic nanostructures. The smooth surface morphology of plasmonic BNC film, stemming from the irreversible collapse of nanoscale fiber network, renders optical transparency and excellent uniformity in SERS activity compared to plasmonic paper. The plasmonic BNC film with large and uniform SERS activity combined with low surface roughness is employed as a flexible swab to demonstrate the collection, identification and detection of model pathogens on real-world surfaces. The approach demonstrated here can be easily extended to other functional nanostructures to realize biotic/abiotic composites with tailored optical, electrical, magnetic and catalytic properties for a broad range of applications.

2.6 Supporting information

Supporting Information for chapter 2 is provided in appendix 1.

2.7 Figures

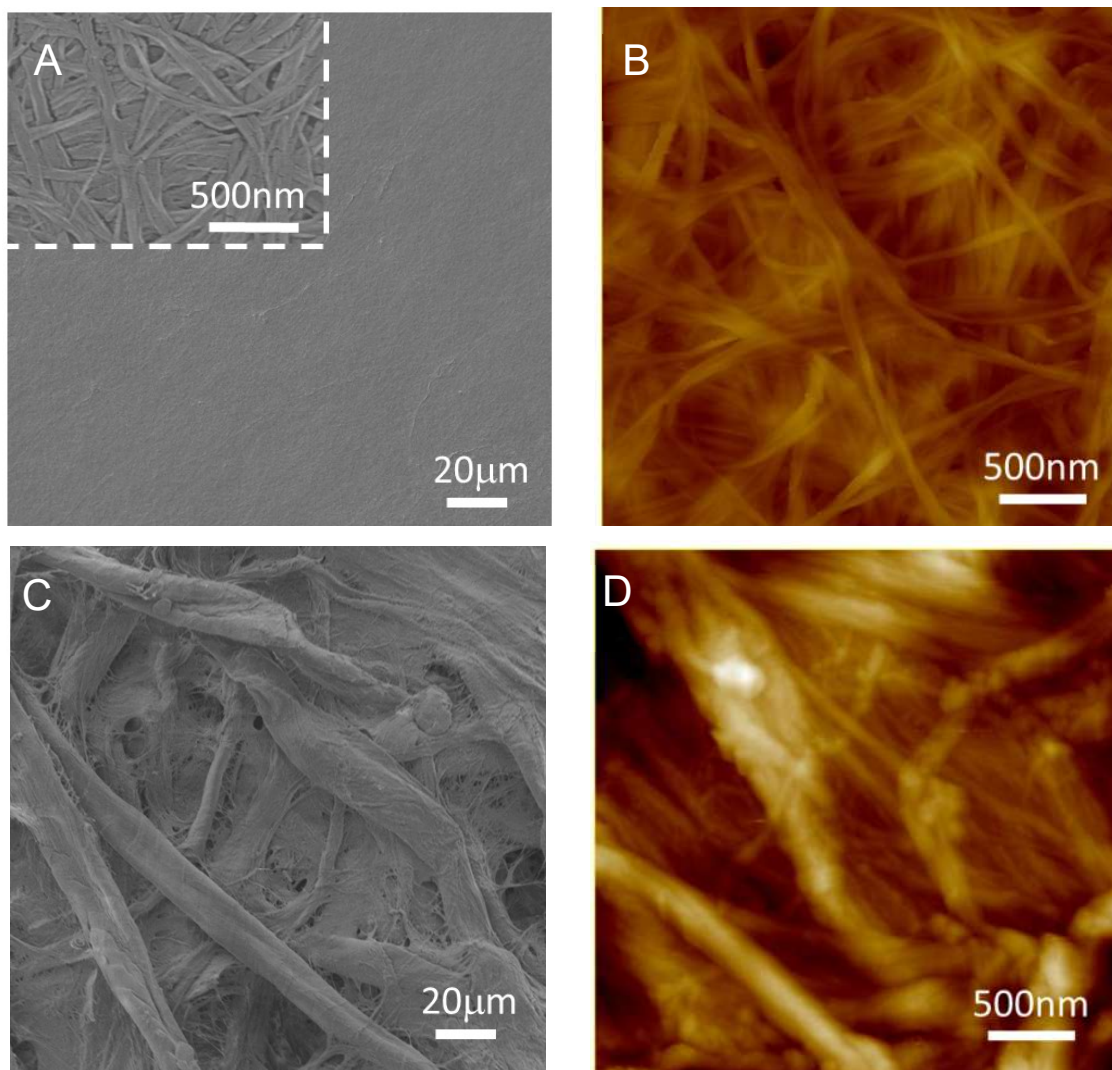


Figure 2.1. Surface morphology comparison of BNC and common filter paper. (A) SEM image of naturally dried pristine BNC paper (inset: higher magnification image showing the nanofibers of BNC paper). (B) AFM image of pristine BNC paper (Height scale: 250 nm). (C) SEM image of pristine filter paper (Whatman™ #1) composed of microfibers with a wide range of diameters. (D) AFM image showing the significantly rougher surface of filter paper compared to BNC paper (Height scale: 450 nm).

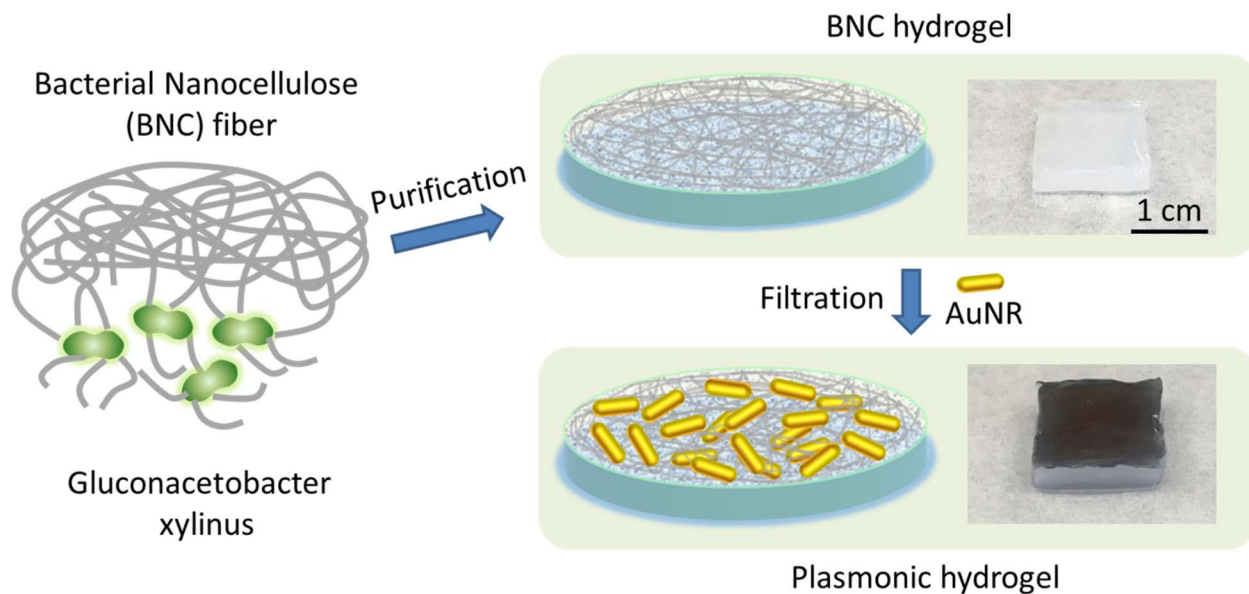


Figure 2.2. Schematic illustration showing the steps involved in the preparation of plasmonic hydrogel from bacterial nanocellulose and photographs showing BNC hydrogel and plasmonic BNC hydrogel.

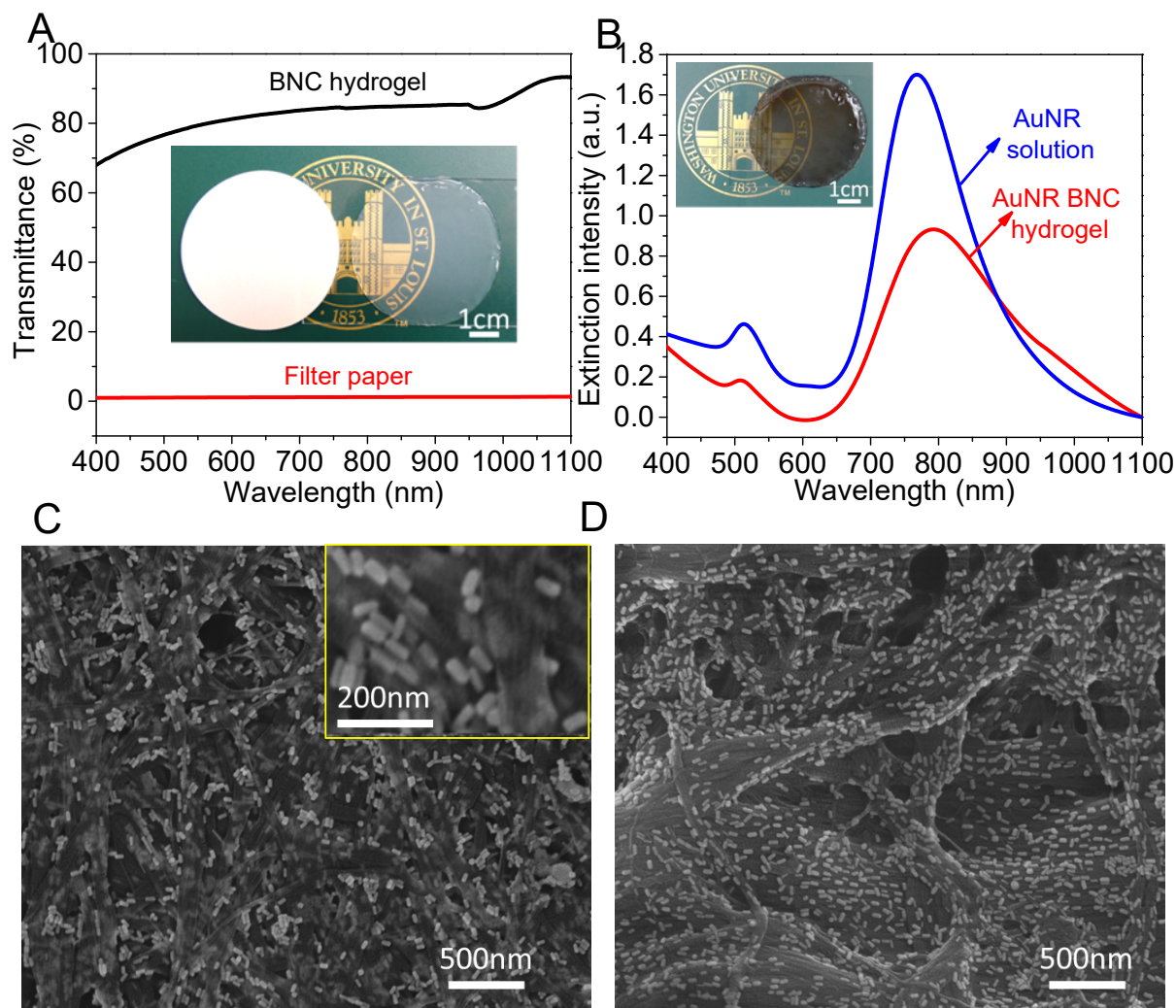


Figure 2.3. Optical properties and surface morphology of plasmonic BNC film and filter paper. (A) Transmission spectra showing the optical transparency of BNC hydrogel in comparison to the opaque filter paper (Inset photograph shows the filter paper and BNC hydrogel placed on Washington University logo to illustrate the contrast). (B) Extinction spectra of AuNRs solution and BNC hydrogel embedded with AuNRs (Inset photograph shows plasmonic hydrogel placed on Washington University logo). SEM images of (C) naturally dried BNC hydrogel embedded with AuNRs (Inset high resolution SEM image showing AuNRs penetrated into the subsurface layers of cellulose fibers) and (D) filter paper adsorbed with AuNRs.

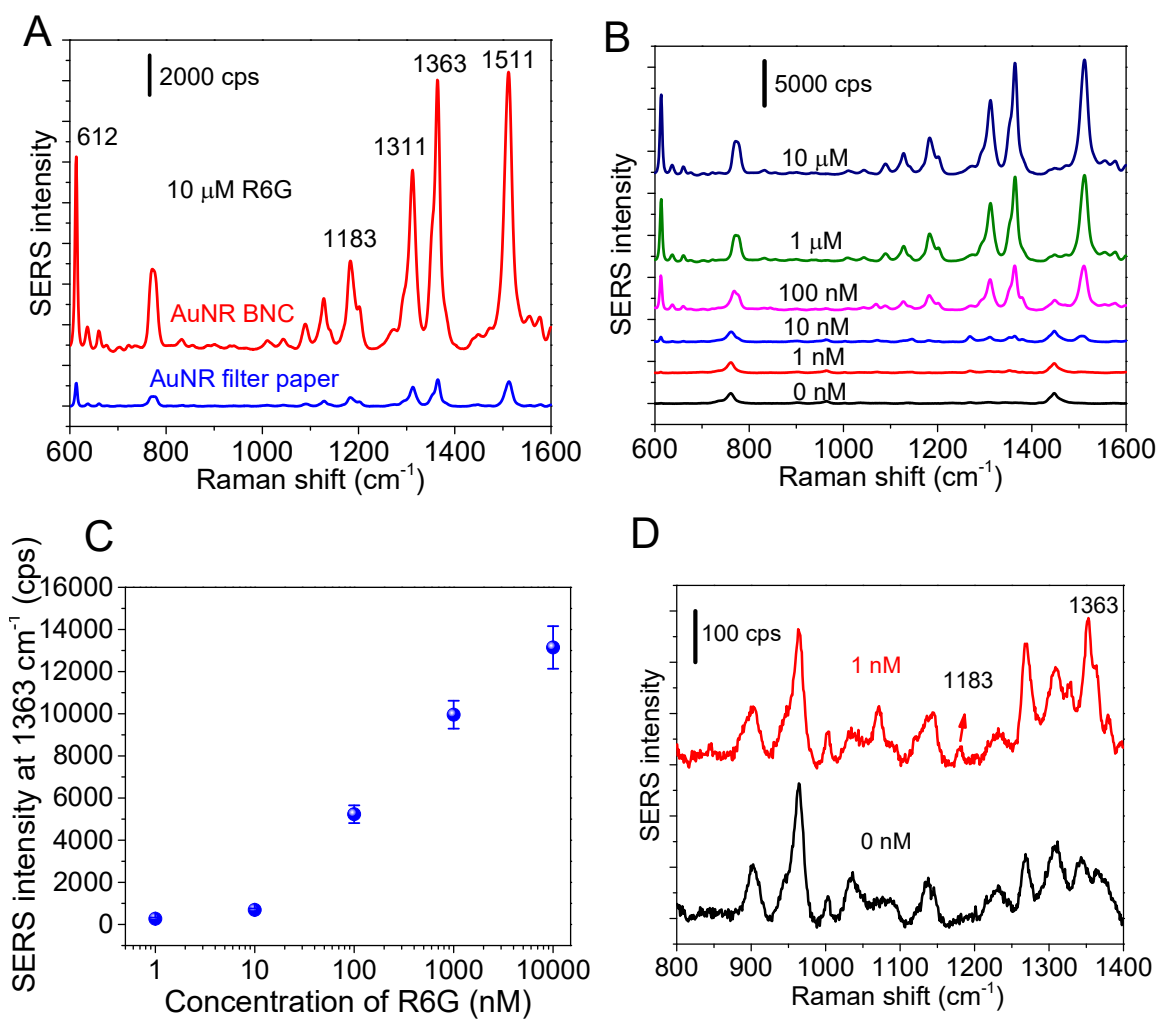


Figure 2.4. SERS performance of BNC-based plasmonic hydrogel. (A) SERS spectra obtained from BNC-based plasmonic hydrogel and filter paper after exposure to 10 μM of R6G aqueous solution. (B) SERS spectra obtained from AuNRs adsorbed on BNC after exposure to different concentrations of R6G. (C) Plot of SERS intensity as a function of R6G concentration obtained from AuNRs/BNC substrate. (D) Higher resolution SERS spectra showing the detection limit of R6G compared with AuNRs/BNC control sample.

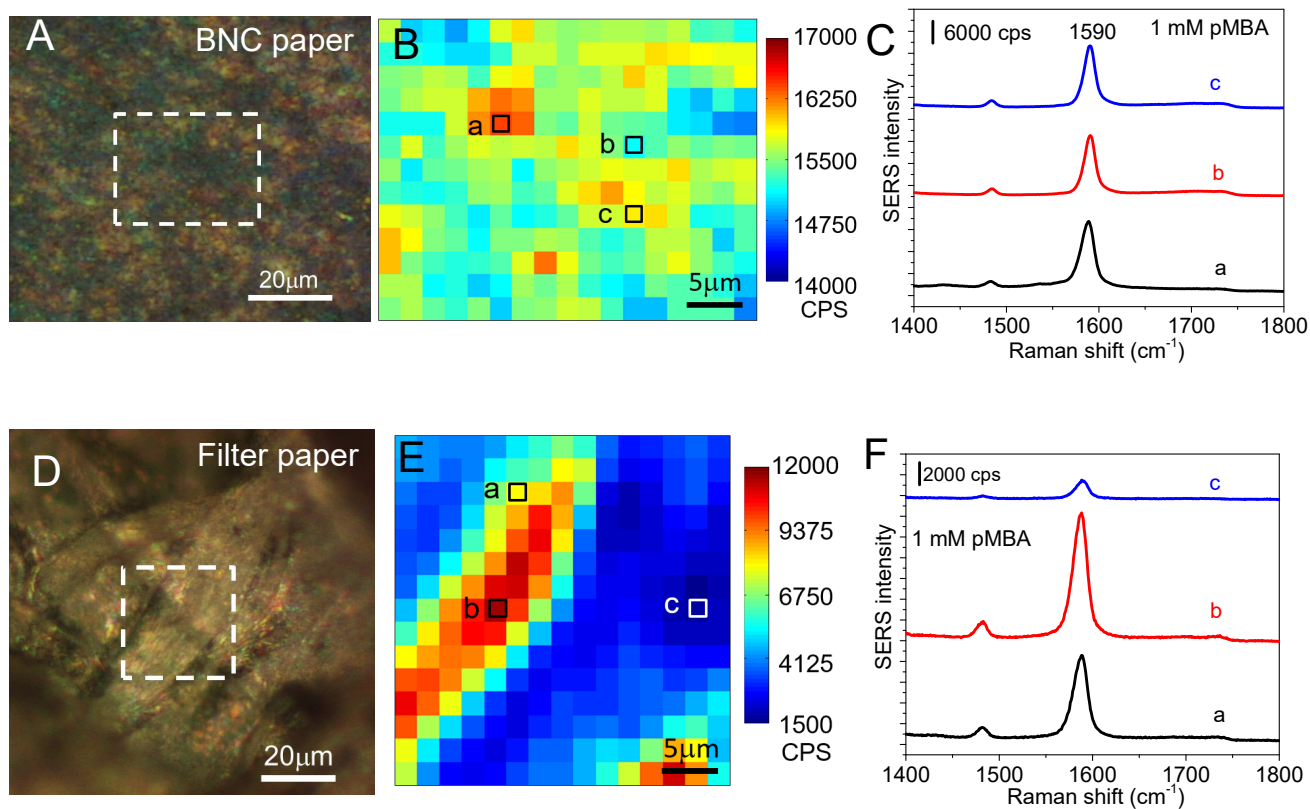


Figure 2.5. Uniformity of SERS intensity from BNC-based SERS substrate and plasmonic paper. Optical micrographs of (A) AuNRs/BNC and (D) AuNRs/filter paper substrates, and (B, E) corresponding SERS intensity maps of 1590 cm^{-1} band from the area indicated with dashed lines in the optical images A and D, respectively. Representative SERS spectra of pMBA collected from (C) BNC-based SERS substrate and (F) plasmonic paper substrates from the pixels marked with squares in B and E, respectively.

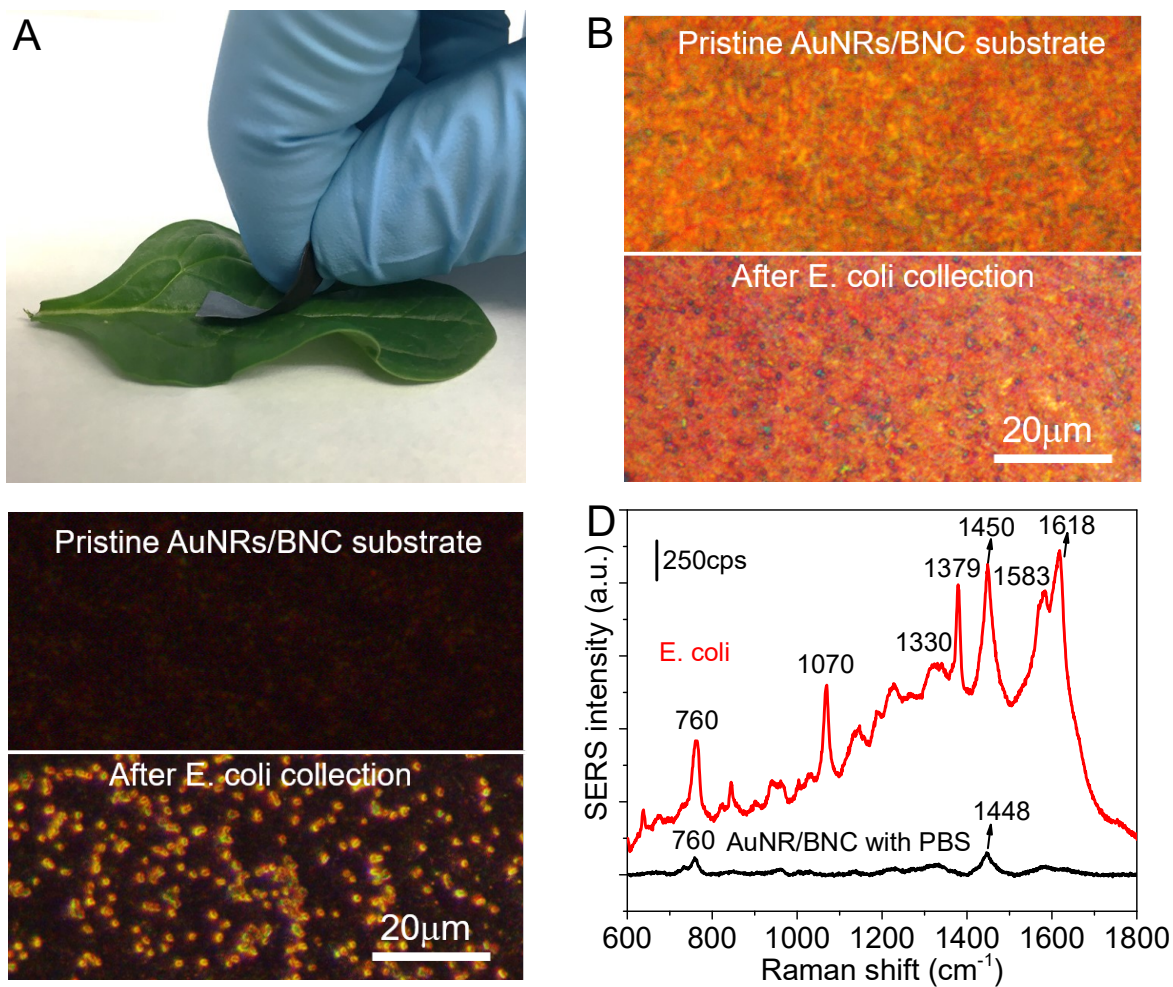


Figure 2.6. Flexible BNC-based SERS swab for bacteria collection and detection. (A) Photograph showing a plasmonic BNC SERS substrate being swabbed on a spinach leaf surface intentionally contaminated with *E.coli*. (B) Bright field and (C) dark field optical micrographs of pristine AuNRs/BNC paper and *E. coli* adsorbed on AuNRs/BNC paper collected by swabbing. (D) Representative SERS spectra collected from *E.coli* adsorbed on the AuNRs/BNC swab showing characteristic Raman bands of *E. coli*.

Chapter 3: Bilayered Biofoam for Highly Efficient Solar Steam Generation

3.1 Abstract

Solar steam generation by heat localization is highly promising for large-scale desalination and water purification, can alleviate water shortage and their consequences, especially in resource-limited regions. However, most of the interfacial evaporators demonstrated so far involve either expensive materials such as noble metal nanostructures or complex fabrication methods with poor prospects in terms of scalability. Here, a novel bilayered biofoam is realized using bacteria-mediated fabrication which brings bacterial nanocellulose and reduced graphene oxide together for highly efficient solar steam generation. The biofoam exhibits a solar thermal efficiency of ~83% under simulated solar illumination (10 kW/m²). The facile fabrication of this novel bilayered biofoam is highly scalable and cost-efficient.

3.2 Introduction

Steam generation (distillation) through efficient harvesting of solar energy is highly promising for large scale power generation, desalination and water purification. Most of the existing solar steam generation systems, which involve heating water using optical concentrators, suffer from poor efficiency due to losses associated with heating the bulk water. Confining heat to air/water interface (*i.e.* the evaporating surface) is a highly promising approach to improve the efficiency of evaporation and steam generation process. Over the past three years, various heat localization layers have been demonstrated to significantly improve the steam generation efficiency compared to conventional bulk heating methods. However, most of these designs involve either expensive

materials such as noble metal nanostructures or complex fabrication methods with poor prospects in terms of scalability. Thus, there is an immediate need for cost-effective novel materials and processes for highly-efficient solar distillation.

Graphene oxide (GO) exhibits a broad optical absorption over visible and near infrared (NIR) parts of the electromagnetic spectrum and excellent photothermal transduction.¹⁹¹⁻¹⁹⁵ Unlike graphene flakes, which tend to stack and aggregate in aqueous solutions, GO exhibits excellent water solubility. GO is essentially sheets of graphene with carboxylic functional groups at the edges and phenol hydroxyl and epoxide groups on the basal planes.⁴⁹ So, the hydrophilicity of GO is attributed to the hydroxyl and epoxide groups on the basal planes and carboxyl functionalities at the edges. Crumpled graphene oxide and crumpled reduced graphene oxide produced by aerosol synthesis methods also exhibit excellent stability and hydrophilicity.¹⁹⁶⁻¹⁹⁷ Graphene-based materials have been extensively investigated for various optoelectronic applications including transparent electrodes, photodetectors, and as electron and hole transport layers in photovoltaic devices.¹⁹⁸⁻²⁰⁰ While being electrically conductive, a single layer of graphene exhibits only small optical absorbance (~3%), making it an excellent candidate for transparent electrodes. However, the cumulative optical absorbance of a few graphene monolayers in tandem can quickly add up, making graphene materials an excellent choice for applications demanding high optical absorption. We hypothesize that a novel composite material that can stabilize the tandem configuration of RGO layers at the surface of the water and facilitate the facile transport of water to the photothermally-active interstitial space (between RGO layers) will be an excellent candidate for a solar steam generation.

In this work, we introduce a bilayered hybrid biofoam comprised of bacterial nanocellulose (BNC) and RGO for solar steam generation through heat localization at the evaporation surface. BNC is

composed of highly pure cellulose nanofibrils, produced from dextrose through a series of biochemical steps followed by the self-assembly of the secreted cellulose fibrils from bacteria in the culture medium.^{3,201} BNC is a highly attractive material for the fabrication of functional foams due to its large specific surface area, open microporous structure, excellent mechanical properties, and facile and scalable synthesis.¹¹ So far, most of the functional foams based on BNC rely on either in situ growth or adsorption of functional nanostructures or infiltration of polymers in the porous cellulose network after harvesting the BNC from the culture.¹³⁻¹⁹ Here, we demonstrate a novel approach for the fabrication of photothermally-active biofoam that involves the in situ incorporation of GO flakes into BNC during its growth. The bilayer structure of the functional foam is tailored for high optical absorption, photothermal conversion, heat localization, and water transport to the evaporation surface resulting in a highly efficient solar steam generation. The bilayer structure exhibited excellent stability even under vigorous mechanical agitation and harsh chemical conditions, which is quite remarkable considering the simplicity of the two-step fabrication approach demonstrated here.

3.3 Experimental Section

Preparation of RGO/BNC aerogel: *Gluconacetobacter hansenii* (ATCC[®]53582) was cultured in test tubes containing 16 ml of #1765 medium at 30°C under shaking at 250 rpm. The #1765 medium is composed of 2% (w/v) glucose, 0.5% (w/v) yeast extract, 0.5% (w/v) peptone, 0.27% (w/v) disodium phosphate, and 0.5% (w/v) citric acid. Graphene oxide was synthesized using an improved method reported by Tour and coworkers.²⁰² Graphene oxide solution (28 mL of 0.1 wt%) was centrifuged and redispersed in #1765 medium and then centrifuged again to leave a wet mixture of GO and medium after decanting supernatant. Bacterial culture solution (incubated 3 days) was added to the GO/medium wet mixture to make it to a total 7 ml (with GO concentration

of 0.4 wt %). The solution was subsequently transferred to petridish (diameter: 6 cm) and incubated at room temperature without disturbance. After 5 days, a thin film of GO/BNC was formed at the liquid/air interface. Subsequently, 7ml of bacterial growth solution was added on top of the GO/BNC film. After another 5 days, a bilayer of BNC and GO/BNC film was formed. For purification, the film was harvested from the petri dish and washed in a 500ml of 0.1 M NaOH aqueous solution under boiling condition for 2 h. The obtained RGO/BNC:BNC hydrogel was then dialyzed in nanopure water for 2 days. The purified RGO/BNC:BNC hydrogel was then cut into desired dimensions, typically 1 cm x 1 cm with a thickness of 21 mm, and then freeze-dried for 12 hours.

Microstructure characterization and properties measurement: Scanning electron microscope (SEM) images were obtained using a FEI Nova 2300 Field Emission SEM at accelerating voltage of 10kV. Atomic force microscopy (AFM) images were obtained using Dimension 3000 (Bruker Inc.) in light tapping mode. Shimadzu UV-1800 spectrophotometer was employed for collecting UV-vis extinction spectra in transmission mode. Raman spectra were obtained using a Renisha inVia confocal Raman spectrometer mounted on a Leica microscope with 20x objective and 514nm wavelength diode laser as an illumination source. Fourier transform infrared spectrometer (FTIR) spectra were recorded using a Nicolette Nexus 470 spectrometer. XPS analysis was performed using Physical Electronics® 5000 VersaProbe II Scanning ESCA (XPS) Microprobe. The specific surface areas of GO/BNC bilayer aerogel were measured by the Brunauer-Emmett-Teller (BET) method using an Autosorb-1C (AX1C-MP-LP) at 298 K.

Thermo- gravimetric analysis (TGA) to measure RGO loading in RGO/BNC:BNC: TGA was used to measure the mass loading of RGO in RGO/BNC:BNC. Both BNC film and air- dried RGO/BNC:BNC bilayer film are tested (see appendix 2, Figure S2.1). BNC film showed an initial

mass loss which has been attributed to absorbed water, a mass loss (~70 %) at ~280 °C is due to the degradation of cellulose and a mass loss (~25 %) at ~390 °C is due to the decomposition of cellulose residual which will generate CO₂ and H₂O.²⁰³ As for RGO/BNC:BNC, the first mass loss (~2 %) of at ~100 °C is still due to the loss of absorbed water. The second mass loss (~3 %) at ~200 °C is due to the decomposition of functional groups of GO. The third mass loss (~52 %) of at 280 °C is due to the degradation of cellulose. The fourth mass loss (~40 %) at 390 °C is due to the decomposition of cellulose residual and sublimation or burning of the damaged graphitic regions.²⁰⁴ Based on TGA results, the mass loading of RGO in RGO/BNC:BNC is calculated to be 27 wt%.

Stability of RGO/BNC:BNC: As the RGO/BNC:BNC hydrogel was base washed during cleaning, it proved its stability in strongly basic environments. For the stability in an acidic environment, we put one RGO/BNC:BNC hydrogel in a petridish filled with pH 1.5 solution and sonicated for one hour and the hydrogel was still intact (see appendix 2, Figure S2.3).

Thermal conductivity measurement of wet/dry RGO/BNC aerogel and bare BNC: The thermal conductivity of wet/dry RGO/BNC aerogel and BNC aerogel is measured by sandwiching the material between two glass microscope slides. The sandwich is placed between a hot plate and a glass slide with ice on top. The temperature distribution along the thickness was monitored by an IR camera (ICI 7320 USB camera). We assume the emissivity coefficient of glass slide and sample to be 0.9 to obtain the temperature distribution. Emissivity Coefficients of some common Materials, (February, 2016) Retrieved from http://www.engineeringtoolbox.com/emissivity-coefficients-d_447.html Fourier equation was used to calculate the thermal conductivity of each sample.²⁰⁵

$$q' = K \frac{\Delta T}{\Delta X}$$

Since the thermal conductivity (K) is known for glass slides ($1.05 \text{ W m}^{-1} \text{ K}^{-1}$ was used), the heat flux q' per unit area, was calculated. Assuming the samples and the glass slides were experiencing same heat flux, the thermal conductivity of the samples was calculated.

Solar steam generation experiment: A 1 cm x 1 cm bilayer RGO/BNC:BNC aerogel with a thickness of around 21 mm was floating on water in a plastic cuvette with dimensions of 12.5 mm (W) \times 12.5 mm (D) \times 49 mm (H). The solar beam from a solar simulator (Newport AM1.5) was concentrated using a magnifying lens and illuminated onto the floating aerogel. The power density of the solar beam at the sample surface was controlled to be 10 kW/m^2 . Each sample was illuminated for 15 min and the weight loss over the entire duration was recorded. The temperature was measured using an IR camera and the weight change from evaporation was measured using an electronic mass balance with an accuracy of 0.1 mg. We assume that the steam is generated at 100°C under 10 kW/m^2 illumination. The evaporation efficiency (η) is given by ²⁰⁵:

$$\eta = \frac{\dot{m}h_{LV}}{I}$$

Where η is evaporation efficiency, \dot{m} is the evaporation rate, h_{LV} is the total enthalpy of sensible heat (294 J/g, from 30°C to 100°C with specific heat 4.2 J/gK) and phase change of liquid to water (2256 J/g), and I is the incident laser power density.

3.4 Results and Discussion

3.41 The Fabrication of the Bilayered Biofoam

The fabrication of the bilayer structure involves growing *Gluconacetobacter hansenii* bacteria in the presence of GO flakes under aerobic and static growth conditions (Figure 3.1A). GO flakes were obtained using a method recently reported by Tour and co-workers.²⁰² It is known that the thickness of a monolayer of GO is $\sim 0.72 \text{ nm}$ and the higher thickness of GO compared to a

monolayer of graphene (0.34 nm) is ascribed to the presence of epoxy and hydroxyl groups on the basal plane.²⁰⁶ Atomic force microscope (AFM) images revealed the average thickness of GO flakes deposited on a silicon substrate to be $\sim 1 \pm 0.2$ nm, which corresponds to a bilayer of GO. Raman spectrum of graphene oxide showed characteristic graphitic band (G-band) at (1580-1600 cm^{-1}) assigned to the E_{2g} phonons at the Brillouin zone center and a defect band (D-band) at (1330-1340 cm^{-1}) associated with the defect-activated breathing modes of six-membered carbon rings (Figure 3.1D).²⁰⁷ GO flakes were dispersed in the broth solution with bacteria at a predetermined concentration to achieve a desired BNC growth rate (see experimental section for details). The mixture is homogenized and left under the static condition to obtain a BNC layer with desired thickness and uniformly embedded with GO flakes. Subsequently, additional broth with bacteria was added on the surface of the GO filled BNC hydrogel to form an additional layer of pristine BNC. The GO filled BNC/BNC bilayer film (called GO/BNC:BNC henceforth) was washed using NaOH solution (0.1 M) under high temperature to remove bacteria and residual growth medium. GO was found to be partially reduced during the washing process, which results in a color change of the hydrogel from brown to black (discussed in detail below). The as cleaned RGO/BNC:BNC hydrogel was freeze-dried to obtain highly porous bilayered aerogel (Figure 3.1A,B). The RGO/BNC:BNC has ~ 27 wt% of RGO loading based on TGA study (see appendix 2, Figure S2.1 and experimental section).

3.42 Probing the Structure of the Bilayered Biofoam

Pristine BNC aerogel obtained by freeze drying BNC hydrogel exhibited $\sim 98\%$ porosity, ultralow density ($\sim 20 \text{ kg m}^{-3}$) and extremely large specific surface area ($\sim 75 \text{ m}^2 \text{ g}^{-1}$) (Figure 3.2A). BNC aerogel is comprised of a highly open microporous non-woven 3D network of cellulose nanofibrils (diameter of 20-100 nm). The cellulose fibers are first formed by the bacteria at the air/medium

interface and fibers entangle to form a “layer” at the surface.^{3, 201} As the oxygen diffuses deeper into the medium, the first layer guides the formation of the subsequent BNC layers, which stack together to form a 3D BNC network. Considering that bacteria form the cellulose fibers at the air/liquid interface, the cellulose nanofibrils are preferentially oriented parallel to the surface (*i.e.* normal to the thickness) of the aerogel structure, which results in denser physical entanglements of the cellulose nanofibrils parallel to the surface compared to that along the thickness. A representative cross-section SEM image of BNC aerogel reveals dense layers of cellulose fibers separated by a layer of sparse “tie” fibers resulting in a layered structure (Figure 3.2A). Addition of the GO to the bacterial broth and base wash led to the formation of BNC hydrogel loaded with RGO (Figure 3.2B, b1). The “layered” formation of BNC starting from the liquid/air interface facilitated the layered arrangement of RGO flakes within the BNC (Figure 3.2B, b2 and b3). The layered incorporation of the RGO into the layered structure of BNC was clearly evident in the cross-sectional SEM images of the naturally dried RGO/BNC film (see appendix 2, Figure S2.2). SEM images of the aerogel also reveal the uniform incorporation of the RGO flakes into the cellulose fiber network. It is worth noting that most of the RGO flakes are arranged parallel to BNC layers (see SEM image of the cross-section in Figure 3.2B, b2).

3.43 The Stability of the Bilayered Biofoam

As briefly mentioned above, bilayered biofoam comprised of RGO/BNC and BNC layers was obtained by adding fresh broth containing bacteria following the formation of GO/BNC layer. The bilayered hydrogel subjected to rigorous mechanical agitation and strong basic solution during cleaning procedure to remove the bacterial and culture medium residue did not exhibit any signs of delamination. To further test the stability of the bilayer structure, we subjected the bilayer to a strong acidic environment (pH 1.5) and ultrasonication for 1 hr. The bilayered hydrogel subjected

to these extreme conditions remained mechanically stable and the RGO flakes remained embedded within the nanocellulose layers of BNC (see appendix 2, Figure S2.3). These findings suggest that the RGO/BNC:BNC has excellent mechanical and chemical stability even under extreme environments. Such strong interfacial interaction between the two layers formed in a sequential manner indicates that the nanoscale fibrils generated by the newly added bacteria entangled seamlessly with the existing BNC network. The bilayer exhibited a sharp interface in terms of RGO content between the RGO doped BNC and pristine BNC (Figure 3.2C; c1 and c2). The structures of the pristine and RGO filled regions of the bilayer were found to be identical to the corresponding monolayer films described above (Figure 3.2C; c3).

3.44 Probing the Properties of the Bilayered Biofoam

The RGO/BNC:BNC aerogel (with thickness of 2.1 mm) exhibited extremely small optical transmittance ($\sim 1.5\%$) and reflectance ($\sim 2.5\%$) in the visible and near infrared regions, indicating the large optical extinction ($\sim 96\%$) of the bilayer (Figure 3.3A). The large extinction of the bilayer owes to the optical absorption of the RGO flakes and the light scattering from the nanoscale cellulose fibers that increase the optical path length within the bilayer. Such large optical extinction of the bilayer structures combined with the excellent photothermal activity of RGO makes RGO/BNC:BNC an excellent candidate for a solar steam generation. Natural drying of RGO/BNC hydrogel (as opposed to freeze drying described above) resulted in the collapse of the 3D BNC structure into a flexible thin film (inset of Figure 3.3B). This bilayer thin film appeared light grey in color and exhibited a metallic luster, indicating the partial reduction of GO.²⁰⁸ We measured the electrical conductivity of the thin film on both sides (*i.e.* RGO/BNC and pristine BNC) under different bending angles (Figure 3.3B). As expected, the pristine BNC exhibited extremely small electrical conductivity ($\sim 2.4 \times 10^{-6} \text{ S} \cdot \text{m}^{-1}$). On the other hand, RGO/BNC exhibited

significantly higher electrical conductivity ($\sim 14.5 \text{ S}\cdot\text{m}^{-1}$), which was found to be insensitive to the bending angle of the flexible film (inset of Figure 3.3B). The electrical conductivity of RGO/BNC film without any special reduction procedure was higher compared to BNC/silk/MWCNT composite membrane ($0.2 \text{ S}\cdot\text{m}^{-1}$)²⁰⁹ and BNC/PANI nanocomposite membrane ($5 \text{ S}\cdot\text{m}^{-1}$)²¹⁰ and comparable to previous BNC/RGO nanocomposite ($23.8 \text{ S}\cdot\text{m}^{-1}$)²¹¹.

The high electrical conductivity and metallic luster of RGO/BNC film suggests the possible chemical reduction of GO during the BNC cleaning process. We investigated the effect of the high-temperature base wash (conditions employed in BNC cleaning process) on the GO flakes to further understand the chemical reduction of GO. When subjected to high-temperature base wash, the initially clear brown GO suspension turned into a black aggregate, which indicates the partial reduction and restacking of the RGO. We employed X-ray photoelectron spectroscopy (XPS) to confirm the chemical modification of GO upon base wash (Figure 3.3C, 3.3D). The 1s spectra of carbon can be deconvoluted into three peaks corresponding to sp^2 domains (C=C with a binding energy of 284 eV) and oxidized sp^3 domains (C–O with a binding energy of 286 eV and C=O with a binding energy of 288 eV).²¹² For as synthesized GO, the C/O ratio (1.74) obtained from the ratio of the area under the peaks suggests $\sim 57\%$ of the surface of GO is oxidized (Figure 3.3C). After extensive base wash, the C/O ratio increased to 4.95 indicating that only $\sim 20\%$ of the surface of GO is oxidized (Figure 3.3D). FTIR absorption spectrum of GO/BNC before base wash showed bands at 1743 cm^{-1} , 1620 cm^{-1} corresponding to C=O and COOH groups, respectively.²¹³ On the other hand, in the case of RGO/BNC, a dramatic decrease in the intensities of these peaks was observed, indicating the partial reduction of GO flakes (see appendix 2, Figure S2.4). These findings confirm that the base wash can remove the oxidative debris from the GO flakes.^{204, 214} While such large degree of reduction of GO can compromise their stability in aqueous solutions,

the RGO flakes embedded within the swollen network of nanocellulose fibers of BNC are locked in their tandem and layered configuration.

We then investigated the thermal conductivity of individual layers of RGO/BNC:BNC according to different conditions during steam generation. Since BNC layer is in a hydrated state during solar steam generation, it is important to understand the thermal conductivity of this layer in the wet state. We measured the thermal conductivity of BNC layer in the wet state using an infrared camera (see experimental section for details). The IR images of the BNC layer sandwiched between two glass slides maintained at different temperatures reveal a gradient in the temperature along the thickness of the BNC layer (insets of Figure 3.3E). The thermal conductivity of BNC layer was found to be $0.46 \text{ W m}^{-1} \text{ K}^{-1}$, which is lower than that of water ($0.6 \text{ W m}^{-1} \text{ K}^{-1}$ at room temperature).²¹⁵ The thermal conductivity of wet BNC layer is comparable to the heat insulating layers employed in recently demonstrated gold nanoparticle film and double layer carbon foam for solar steam generation in wet state.^{205, 216} As for RGO/BNC layer, it is important to measure its thermal conductivity in both dry and wet state because this layer is semi-wet during the steam generation. Despite the high electrical conductivity of the air-dried RGO/BNC film, the thermal conductivity of wet RGO/BNC ($0.816 \text{ W m}^{-1} \text{ K}^{-1}$) was found to be only slightly higher than water (see Figure 3.3F). It is worth noting that the thermal conductivity of wet RGO/BNC is lower than that of exfoliated graphite in the wet state ($0.959 \text{ W m}^{-1} \text{ K}^{-1}$)²⁰⁵, which has been employed as light-to-heat conversion layer for a solar steam generation. The lower thermal conductivity of wet RGO/BNC is due to the presence of the nanocellulose layers separating the RGO layers, which serve as thermal insulators. As for dry RGO/BNC, the thermal conductivity ($0.069 \text{ W m}^{-1} \text{ K}^{-1}$) is just slightly higher than air (see appendix 2, Figure S2.5). Due to high porosity, most of the dry RGO/BNC layer is comprised of air pockets with a thermal conductivity of $0.024 \text{ W m}^{-1} \text{ K}^{-1}$ (at

room temperature). Thus, the semi-wet RGO/BNC layer can greatly reduce the heat transferred from the evaporative surface to the following wet BNC layer. Overall, the low thermal conductivity of RGO/BNC:BNC results in the confinement of heat to the evaporative surface by minimizing the heat transfer (loss) to bulk water and improves the overall efficiency of the solar steam generation.

3.45 Photothermal and Solar Steam Generation Performance

Now we turn our attention to the photothermal and solar steam generation ability of RGO/BNC:BNC. We investigated the steam generation ability of RGO/BNC: BNC under near-infrared (NIR) laser (808 nm) and simulated solar illumination (Figure 3.4 and appendix 2, Figure S2.6). RGO exhibits large and broad optical absorption over visible and NIR regions and has been employed as a photothermal contrast agent for locoregional photothermal therapy.¹⁹³⁻¹⁹⁵ IR imaging was employed to monitor the temperature rise in the RGO/BNC:BNC floated at air/water interface upon irradiation with a simulated solar beam (power density of 10 KW/m²) or NIR laser (power density of 5 KW/m²). Upon irradiation, the temperature of the RGO/BNC rapidly increased from room temperature (30°C) to ~78°C in the case of the solar beam and to ~60°C in the case of 808 nm laser beam (Figure 3.4B, C and appendix 2, Figure S2.6). The temperature rapidly increased within tens of seconds after the irradiation and remained constant over the entire duration (900 sec) of the irradiation. On the other hand, the temperature rise of water in the absence of RGO/BNC layer was found to be significantly smaller (2-4°C). As briefly discussed above, the large rise in temperature owes to the high optical absorption of RGO combined with the multiple scattering of incident light from the nanoscale cellulose fibers resulting in an effective increase of the optical path length and maximizing the light absorption within the hydrated aerogel. The large

rise in temperature of RGO/BNC film illumination resulted in the appearance of steam above the cuvette, evidencing the rapid evaporation of water (Figure 3.4D).

The steam generation efficiency of the RGO/BNC:BNC was quantified by measuring the weight loss (due to water evaporation) as a function of irradiation time. The cumulative weight loss was found to increase linearly with irradiation time (Figure 3.4E). Under solar beam irradiation (10 kW/m^2), the weight loss over the duration of 900 sec was found to be 0.4 g over 1 cm^2 . The steady state evaporation rate, achieved after ~ 300 sec of irradiation, was calculated to be $11.8 \text{ kg/m}^2/\text{hr}$. On other hand, in the absence of the RGO/BNC:BNC, the evaporation rate of water under identical irradiation condition was found to be $4.8 \text{ kg/m}^2/\text{hr}$, which is nearly 2.5 times lower compared to that with the bilayer structure. Similarly, the steady state evaporation rate under NIR laser irradiation of RGO/BNC bilayer was found to be $6.1 \text{ kg/m}^2/\text{hr}$, which is nearly 13 times higher compared to the evaporation rate in the absence of RGO/BNC layer under identical irradiation condition ($0.46 \text{ kg/m}^2/\text{hr}$). The RGO/BNC:BNC were highly stable and could be reused multiple times without any noticeable degradation of the structure or steam generation ability. Over 8 cycles of reuse, the steady state evaporation rate and cumulative weight loss for 900 sec irradiation were found to exhibit less than 3% variation (Figure 3.4F).

Without considering the losses involved in the test facility, such as reflection and surface radiation, the evaporation efficiency of RGO/BNC:BNC aerogel was calculated to be 83% at a power density of 10 kW/m^2 (see experimental section for details). Replacing the RGO/BNC:BNC with RGO/BNC layer alone led to a drop in the evaporation efficiency by about 30%. This observation highlights the importance of bilayer structure in which the RGO/BNC layer at the top acts as a light-to-heat conversion layer and the pristine BNC layer at the bottom minimizes the heat loss to the bulk of by serving as a heat insulation layer. It is important to note that the BNC layer exhibited

lower thermal conductivity ($0.46 \text{ W m}^{-1} \text{ K}^{-1}$) compared to that of water ($0.6 \text{ W m}^{-1} \text{ K}^{-1}$). We suggest that the excellent steam generation efficiency of RGO/BNC:BNC aerogel owes its origin to the following four factors: (i) broad light absorption RGO flakes over visible and near infrared wavelengths combined with excellent photothermal conversion efficiency; (ii) high density of RGO flakes in the aerogel combined with the multiple light scattering enabled by nanocellulose fibers, which greatly increase the optical path length and light absorption; (iii) low thermal conductivity of RGO/BNC layer due to BNC matrix as well as low thermal conductivity of wet BNC supporting layer, which minimize the heat transfer from the evaporation surface to the bulk water; and (iv) open porous structure and hydrophilic nature of BNC aerogel enabling the rapid transport of water from bulk to the evaporative surface.

3.5 Conclusions

In summary, we have demonstrated the facile fabrication of a bilayered hybrid biofoam comprised of BNC and RGO for highly efficient solar steam generation. In stark contrast to conventional approaches, the functional biofoam is fabricated by incorporating GO flakes within the nanocellulose fiber layers during the bacteria-mediated growth of the BNC hydrogel. The bilayer structure exhibited excellent stability even under vigorous mechanical agitation and harsh chemical conditions. Owing to the large light absorption, excellent photothermal conversion, heat localization and efficient transport of water from bulk to evaporative surface, the novel bilayered structure exhibited a remarkably high solar thermal efficiency. The fabrication approach demonstrated here can be extended to other functional nanomaterials to realize biofoams with applications in energy harvesting, sensing, catalysis and life sciences.

3.6 Supporting Information

Supporting Information for chapter 3 is provided in appendix 2.

3.7 Figures

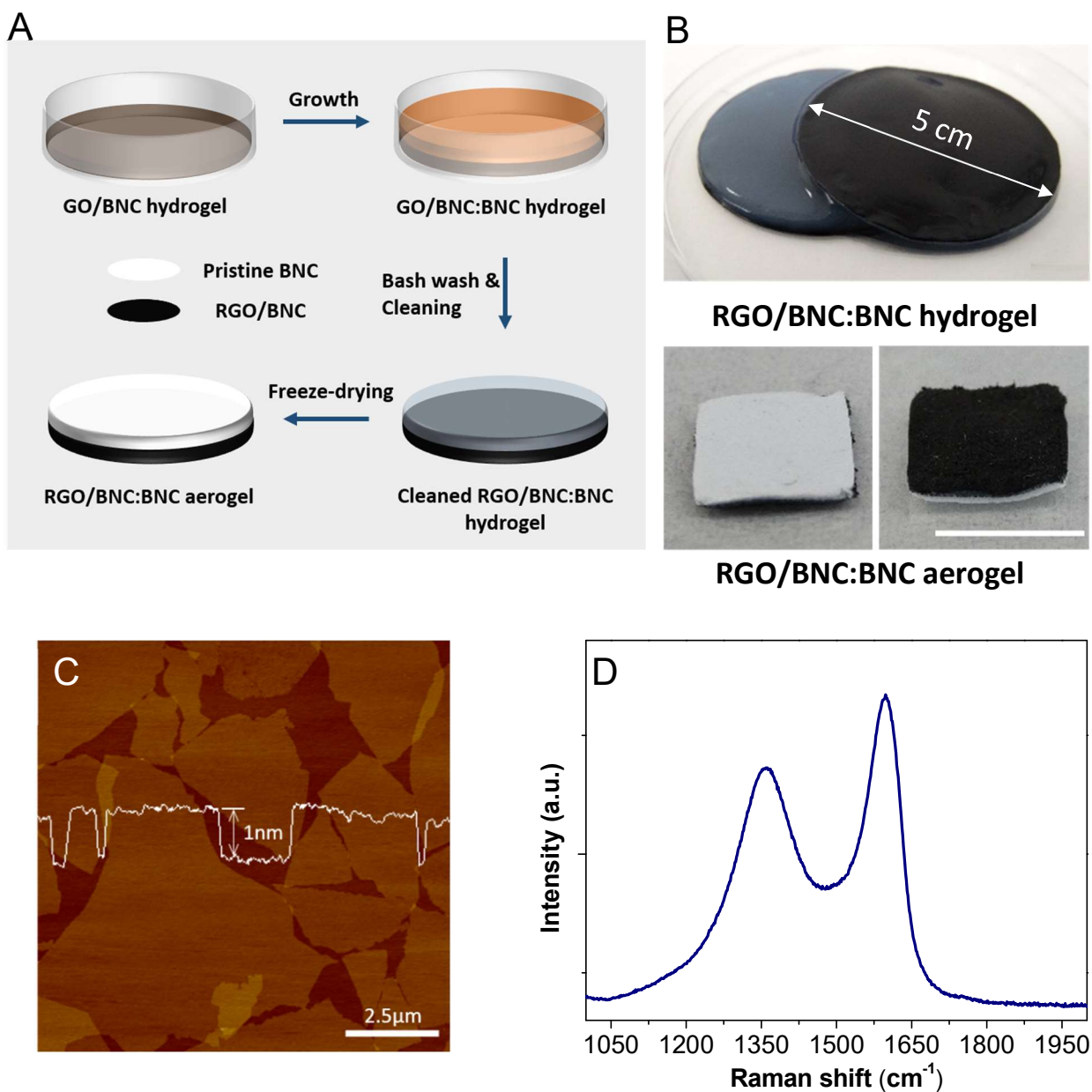


Figure 3.1. (A) Schematic illustration showing the fabrication of RGO/BNC:BNC aerogel. (B) Photographs of the *as cleaned* RGO/BNC:BNC hydrogel and RGO/BNC:BNC aerogel. The scale bars represent 1 cm.

(C) AFM image of GO flakes deposited on a silicon substrate. (D) Raman spectrum of GO flakes showing the characteristic G and D bands.

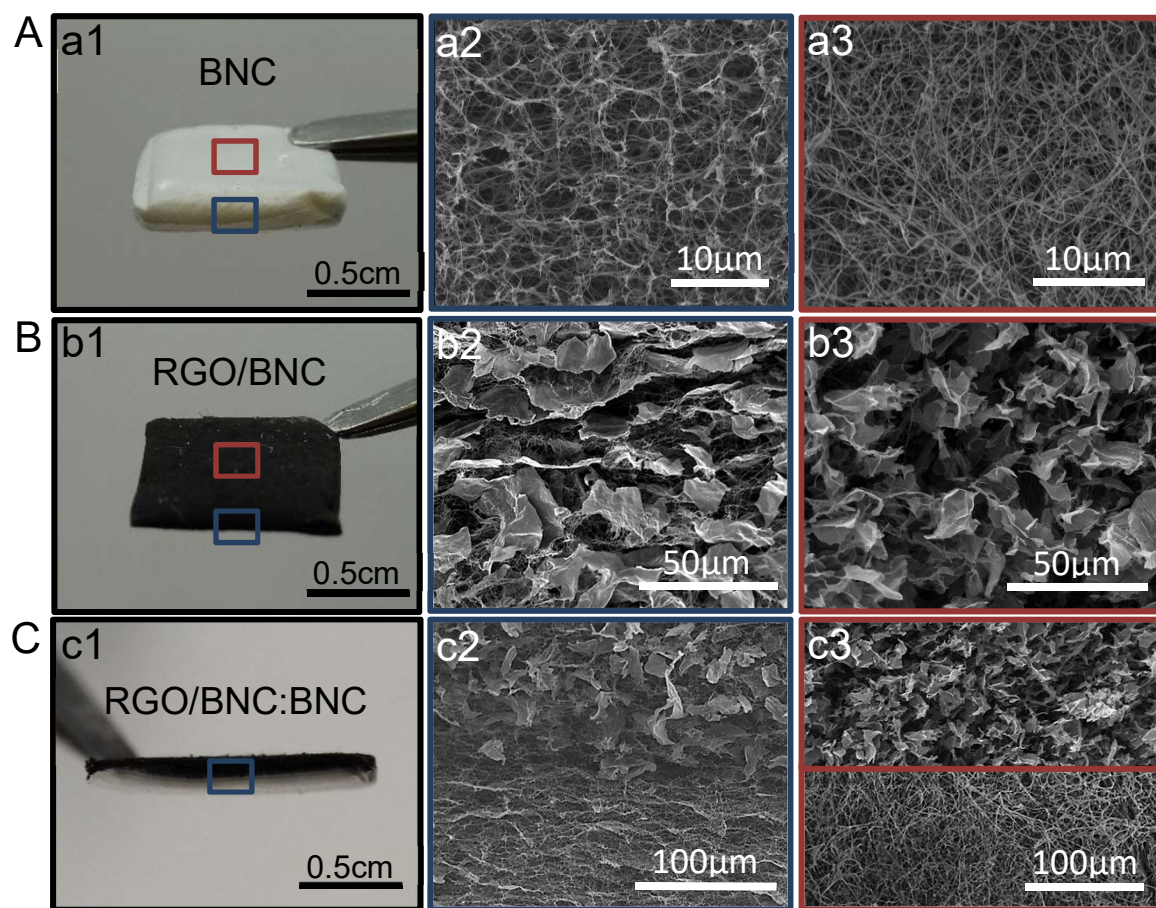


Figure 3.2. Structure of BNC and RGO/BNC and RGO/BNC:BNC aerogels. (A) Optical image (a1) and SEM images of cross-section (a2) and top surface (a3) of BNC aerogel. (B) Optical image (b1) and SEM images of cross-section (b2) and top surface (b3) of RGO/BNC aerogel. (C) Optical image (c1) and SEM images of cross-section at the interface (c2) and the two surfaces (c3) of RGO/BNC:BNC aerogel.

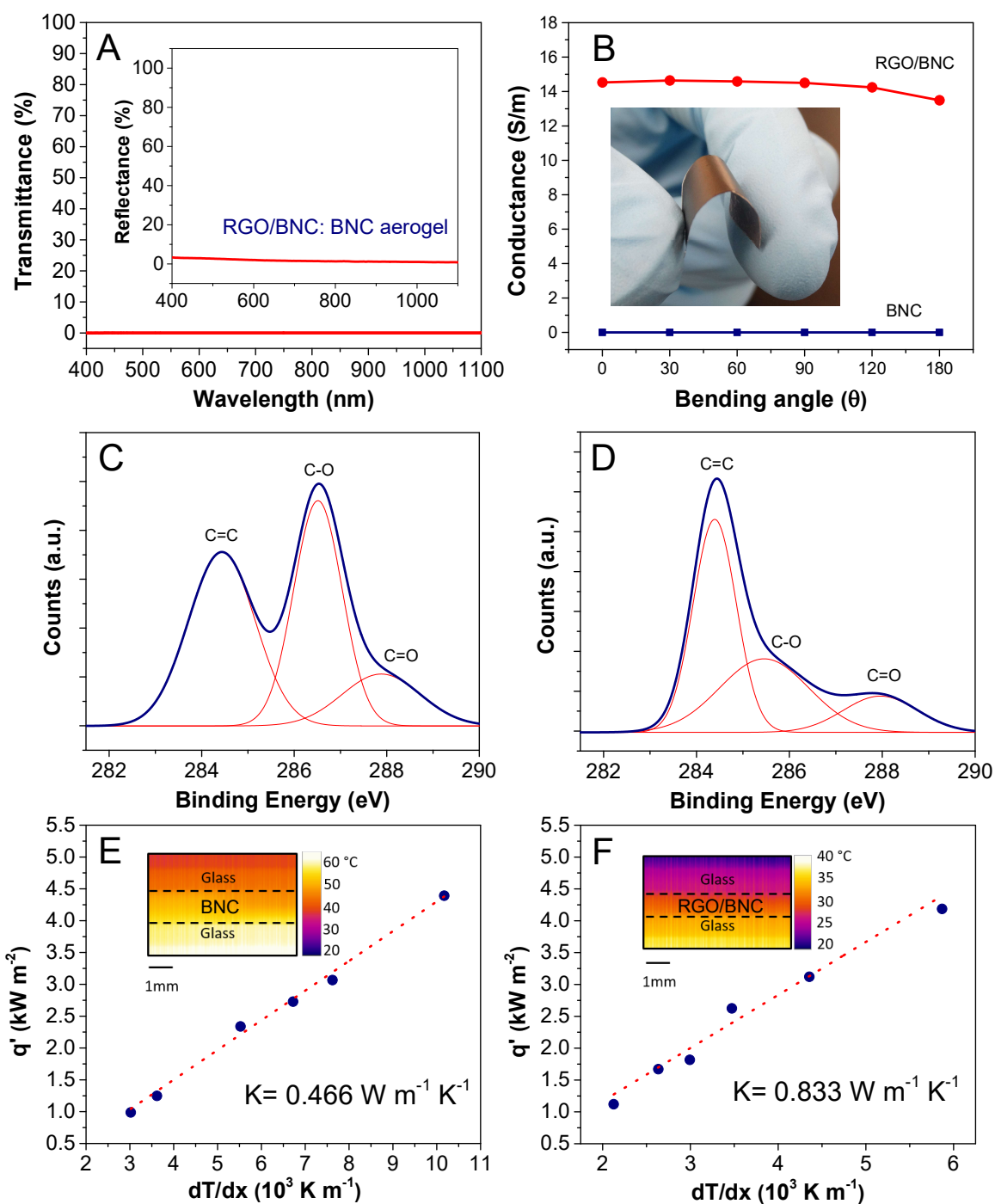


Figure 3.3. (A) Transmittance and reflectance spectra of RGO/BNC:BNC aerogel. (B) Conductance of air-dried RGO/BNC:BNC film under various degrees of bending. XPS spectra of (C) pristine GO and (D) base-washed RGO. Thermal conductivity of (E) wet BNC aerogel and (F) wet RGO/BNC aerogel. Insets: Representative IR images showing the temperature gradient along the thickness of hydrated BNC and RGO/BNC layers.

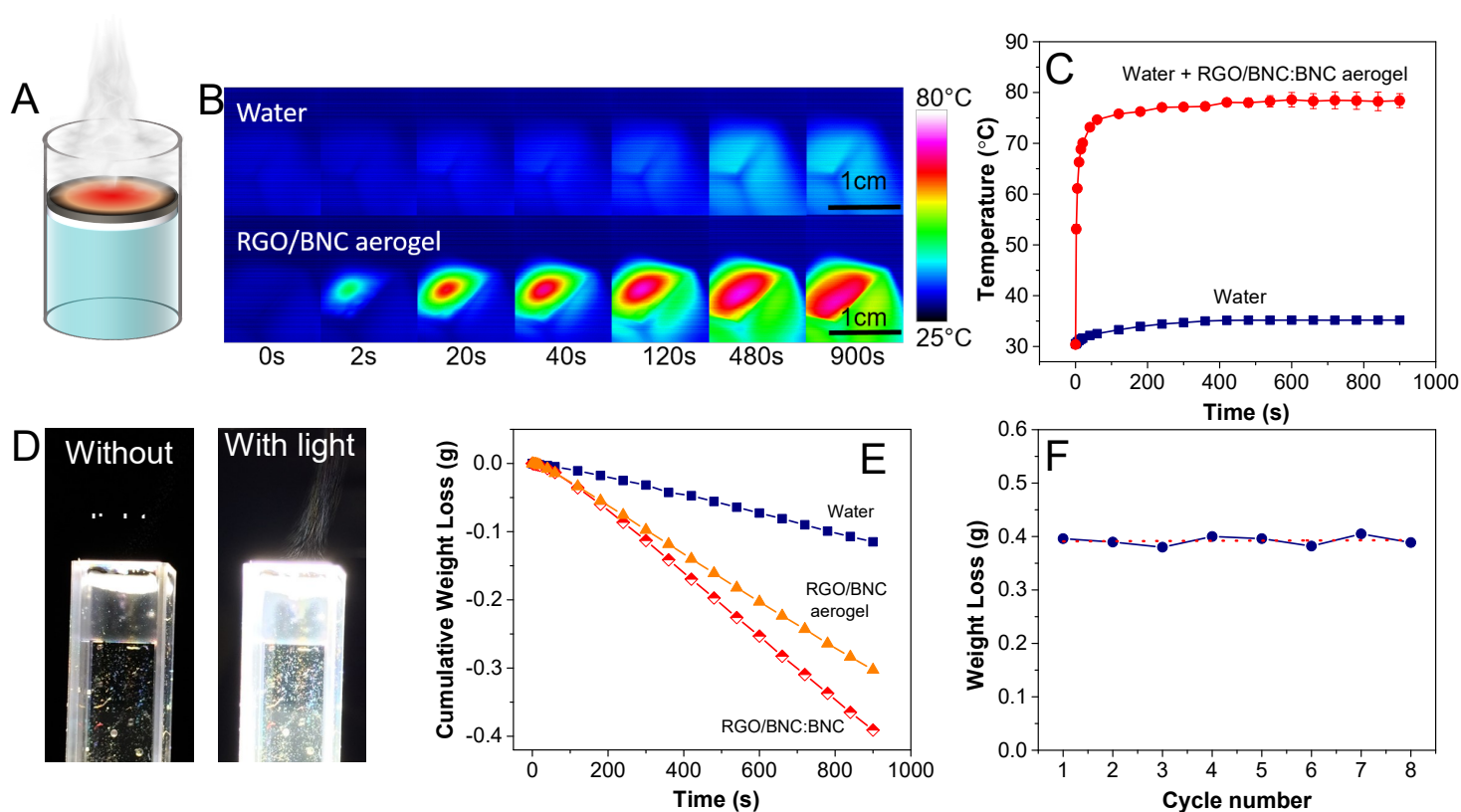


Figure 3.4. Photothermal and solar steam generation efficiency of RGO/BNC:BNC biofoam. (A) Schematic illustration of steam generation with the RGO/BNC:BNC biofoam floated at air/water interface. (B) IR images showing the temperature of water and RGO/BNC:BNC aerogel floated at air/water interface under solar illumination at various time points. (C) Plot showing the surface temperature of water and RGO/BNC:BNC aerogel at air/water interface under solar illumination of 10 kW/m² as function of irradiation time. (D) Photograph showing RGO/BNC:BNC aerogel floated at the air/water interface and steam generation under simulated solar illumination (also see video S1 in Supporting Information). (E) Plot showing the cumulative weight loss through water evaporation under solar illumination as a function of irradiation time. (F) Weight loss through water evaporation after 900 sec irradiation of RGO/BNC layer over 8 cycles showing the reusability of RGO/BNC:BNC.

Chapter 4: Polydopamine-filled Bacterial Nanocellulose as Biodegradable Interfacial Photothermal Evaporator

4.1 Abstract

Solar steam generation by heat localization is considered to be a highly efficient, sustainable way to alleviate water shortage in resource-limited regions. However, most of the interfacial photothermal evaporators demonstrated so far involve non-biodegradable nanoscale materials, which can quickly pose a significant threat to environment and ecosystems, especially marine ecosystems, after their disposal. For the first time, a flexible, scalable and, more importantly, completely biodegradable photothermal evaporator for highly efficient solar steam generation is introduced. The bilayered evaporator is comprised of bacterial nanocellulose densely loaded with polydopamine (PDA) particles during its growth. The biodegradable foam introduced here exhibits large light absorption and photothermal conversion, heat localization, and efficient water transportation, leading to excellent solar steam generation performance under one sun (efficiency of ~78%). The novel material and scalable process introduced here can be a sustainable solution to alleviate global water crisis.

4.2 Introduction

Water scarcity is one of the most critical global issues of the 21st century.³⁹⁻⁴¹ Increasing world population, unprecedented socioeconomic development, and significant depletion of traditional water sources call for more sustainable ways to secure clean and safe water. Steam generation

(distillation) through efficient harvesting of solar energy is highly promising for large-scale desalination and water purification, can alleviate water shortage and their consequences, especially in resource-limited regions.²¹⁷⁻²²³ In stark contrast to conventional solar steam generation systems which suffer from poor efficiencies due to losses associated with heating the entire water (bulk heating), recently introduced interfacial evaporation systems can convert incident sunlight to localized heat at the air-water interface, greatly improving the efficiency of steam generation process.^{205, 224} These evaporation systems are comprised of materials that have excellent photothermal, heat insulating, and water transport properties.

Over the last several years, various interfacial evaporator materials with tailored design and excellent performance have been demonstrated.^{216, 225-237} However, most of them involve non-biodegradable photothermal materials (graphite, reduced graphene oxide, carbon nanotubes, gold, aluminum, titanium oxide, polypyrrole, etc.) or support materials (aluminum, polystyrene thermal foam, stainless steel mesh, polyethyleneimine, etc.) leading to negative or unknown impacts on the environment. It should also be noted that most of these materials have a finite lifetime owing to pore clogging, degradation of the photothermal properties, and alteration of the surface properties of the water transport layer. Disposal of these materials can quickly pose a significant threat to the environment and ecosystems. In particular, degradation and leaching of nanoscale photothermal materials into marine ecosystems, where these materials are most likely deployed, can have lasting negative consequences.²³⁸⁻²⁴² These considerations clearly highlight the need for a scalable material platform that is environmentally benign and easy to produce, utilize, and dispose.

Polydopamine (PDA), formed by the oxidation of dopamine, is an important eumelanin-like biopolymer known for its versatile adhesion properties and universal surface modification.²⁴³ Due

to the structural similarity to eumelanin, PDA possesses many striking properties of naturally occurring eumelanin. For example, it can absorb 99% incident photon energy over a broad solar spectrum and rapidly convert it into heat within tens of picoseconds, thus offering the protection to living organisms against ultraviolet injury.²⁴⁴⁻²⁴⁶ Owing to its superb photothermal ability and biodegradability, extensive efforts have been dedicated to utilizing PDA nanostructures as contrast agents for photothermal therapy.²⁴⁷⁻²⁴⁹

Bacterial nanocellulose (BNC) is a highly pure form of cellulose that is produced from dextrose through a series of biochemical steps followed by the self-assembly of secreted cellulose fibrils from bacteria in the culture medium.³ BNC exhibits outstanding mechanical properties, high porosity, chemical functionalizability, ease of synthesis, high scalability, and more importantly, low environmental impact.²⁵⁰ Hence BNC is a highly versatile material platform for fabricating functional composites through *in situ* growth or adsorption of pre-synthesized nanostructures on the nanoscale cellulose fibers.^{11-20, 147} In addition, the hydrophilicity and low thermal conductivity of BNC provide effective water transport and heat management, making it a highly promising supporting material for interfacial solar steam generation.¹⁴⁷

In this work, we introduce a completely biodegradable, scalable, and flexible bilayered interfacial solar steam generator comprised of BNC and PDA impregnated BNC layers. The bilayered interfacial evaporator has outstanding optical absorption, photothermal conversion, heat insulation, and stability, leading to highly efficient solar generation under one sun condition (i.e., 1 kW m⁻²). The highly scalable and cost-effective materials and processing combined with the highly biocompatible and biodegradable nature of the materials makes this interfacial evaporator a great candidate for solar steam generation, water distillation in resource-limited regions around the world.

4.3 Experimental Section

Preparation of PDA/BNC: *Gluconacetobacter hansenii* (ATCC[®]53582) was cultured in test tubes containing 16 ml of #1765 medium at 30 °C under shaking at 250 rpm. The #1765 medium is composed of 2% (w/v) glucose, 0.5% (w/v) yeast extract, 0.5% (w/v) peptone, 0.27% (w/v) disodium phosphate, and 0.5% (w/v) citric acid. Polydopamine (PDA) particles were prepared using a method reported by Lu and co-workers.²⁵¹ To synthesize PDA particles with the size of 1 μm , ammonia solution (NH_4OH , 0.14 ml, 28-30%) was mixed with 31.5 ml of nanopure water (18 $\text{M}\Omega\cdot\text{cm}$) and 14 ml of ethanol and the above mixture was shaken for 30 minutes. Dopamine hydrochloride solution (3.5 ml, 0.05 g ml^{-1}) was added into the above solution and then transferred to a petridish. After 30 hours of mild shaking at room temperature, the PDA particles were collected by centrifugation (7000 rpm, 20 min) and washed with water for three times and dispersed in nanopure water (40 ml). Bacterial culture solution (3 ml, incubated 3 days) was added to #1765 medium (15 ml) to make a total 18 ml bacterial growth solution. The solution was subsequently transferred to a petridish (diameter: 6 cm) and incubated at room temperature without disturbance. After 5 days, a thick BNC hydrogel ($\sim 4\text{ mm}$) was obtained. PDA particle solution described above (40 ml) was centrifuged and dispersed in bacterial growth medium (7 ml) and was then added on top of the thick BNC hydrogel. After 12h, PDA particles precipitated on the BNC hydrogel and excess medium was removed. After another 12 h, a thin layer of PDA/BNC ($\sim 100\ \mu\text{m}$) was formed on top of the prior thick BNC hydrogel. The bilayered hydrogel was then harvested and washed in boiling water for 2 hours, then dialyzed in nanopure water for one day. The purified PDA/BNC bilayer was then freeze-dried overnight. For PDA/BNC with bigger size, above procedure were simply scaled up and performed in bigger containers.

Microstructure Characterization Methods: Scanning electron microscope (SEM) images were obtained using a FEI Nova 2300 Field Emission SEM. Transmission electron microscope (TEM) images were obtained using a JEOL JEM-2100F field emission microscopy. Dynamic light scattering (DLS) measurements were performed using Malvern Zetasizer (Nano ZS). Shimadzu UV-1800 spectrophotometer was employed for obtaining UV-vis extinction spectra and transmittance spectra. Reflectance spectra were obtained using a CRAIC micro spectrophotometer (QDI 302) coupled to a Leica optical microscope (DM 4000M) with 20× objective in the range of 450–800 nm with 10 accumulations and 100 ms exposure time in reflection mode. Raman spectra were obtained using a Renishaw inVia confocal Raman spectrometer mounted on a Leica microscope with 20 x objective and 785 nm wavelength diode laser as an illumination source. Thermogravimetric analysis (TGA) was performed using TA Instruments Q5000 IR Thermogravimetric Analyzer in air (at rate of 5 °C min⁻¹).

Thermal conductivity measurements of wet/dry PDA/BNC: The thermal conductivities of wet/dry PDA/BNC were measured by sandwiching the material between two glass microscope slides. The sandwich was placed between a hot plate and a glass slide with ice on top. The temperature distribution across the thickness was monitored by an IR camera (ICI 7320 USB camera). The emissivity coefficient of a glass slide and a sample was assumed to be 0.9 to obtain the temperature distribution. Fourier equation was used to calculate the thermal conductivity of each sample:¹⁴⁷

$$q' = K \frac{\Delta T}{\Delta X}$$

The heat flux (q') was calculated by assuming the thermal conductivity (K) of 1.05 W m⁻¹ K⁻¹ for glass slides. Because the glass slide and samples experience the same heat flux, the heat flux value obtained for glass slide was used to measure the thermal conductivity for PDA/BNC samples.

Solar steam generation experiment: In a typical test, a circular bilayer of PDA/BNC with 3 cm diameter and 2.1 mm thickness was floated on water in a 100-ml beaker. The solar beam from a solar simulator (Newport 66921 Arc Lamp) was directly or concentrated using a magnifying lens illuminated onto the PDA/BNC. The power density of the solar beam on the sample surface was controlled to be 1 and 3 kW m⁻². Each sample was illuminated for 45 min and the weight loss over the entire duration was recorded. For the cycling experiments, a 1 cm x 1 cm sample with 4 mm thickness floating on water in a plastic cuvette with dimensions of 12.5 mm (W) × 12.5 mm (D) × 49 mm (H) was used. The power density of the solar beam at the sample surface for cycling was controlled to be 7 kW m⁻² (7 sun) for 15 min illumination duration. The temperature was measured using an IR camera and the weight change from evaporation was measured using an electronic mass balance with an accuracy of 0.1 mg. It is assumed that the steam was generated at 100 °C. The evaporation efficiency (η) is given by:²⁰⁵

$$\eta = \frac{\dot{m}h_{LV}}{I}$$

Where η is evaporation efficiency, \dot{m} is the evaporation rate, h_{LV} is the total enthalpy of sensible heat (294 J g⁻¹, from 25 °C to 100 °C with specific heat 4.2 J g⁻¹ K⁻¹) and phase change of liquid to water (2256 J g⁻¹), and I is the incident illumination power density.

4.4 Results and Discussion

4.41 The fabrication of PDA/BNC and Characterizations of PDA particles and PDA/BNC

The fabrication of interfacial solar steam generator (called PDA/BNC henceforth) with desired size and shape involves a two-step BNC growth under aerobic and static growth conditions (Figure 4.1 and see experimental details). PDA particles were obtained using a method reported by Lu and co-workers through oxidation and self-polymerization of dopamine monomers in a mixture of

water, ethanol and ammonia at room temperature.²⁵¹ The size of PDA particles, which alters the optical properties, can be easily tuned by varying the ratio of ammonia to dopamine monomers. The size of the PDA particles was optimized to efficiently trap PDA particles within the BNC fiber matrix and to ensure significant overlap between the optical absorption of the PDA particles and the solar spectrum in the visible and near infrared region (Figure 4.2A). Transmission electron microscopy (TEM) and scanning electron microscopy (SEM) images revealed that the PDA particles were spherical in shape with a diameter of $\sim 1 \mu\text{m}$ (Figure 4.2B and appendix 3, Figure S3.1A). Dynamic light scattering (DLS) also revealed the hydrodynamic size of the PDA particles to be $\sim 1 \mu\text{m}$ (see appendix 3, Figure S3.1B). Raman spectrum of dopamine monomers showed characteristic bands of C-C stretching ($724, 948, 1324$ and 1422 cm^{-1}), C-O stretching (1290 cm^{-1}), and C-N stretching (795 cm^{-1}). The Raman spectrum of PDA particles exhibited two broad bands (1371 and 1578 cm^{-1}), suggesting catechol deformation. This provides an additional confirmation of successful PDA particles synthesis (Figure 4.2C).²⁵²⁻²⁵³

Pristine BNC aerogel exhibited $\sim 98\%$ porosity, ultralow density ($\sim 20 \text{ kg m}^{-3}$) and extremely large specific surface area (Figure 4.2D).^{13, 147} Furthermore, BNC aerogel forms a highly open microporous non-woven 3D network of cellulose nanofibrils (diameter of $20\text{-}100 \text{ nm}$) with highly abundant hydroxyl groups, promoting high hydrophilicity (Figure 4.2E, F). Thus, the highly porous and hydrophilic nature of the BNC aerogel facilitates the transport of water to the evaporative surface, making it as an ideal supporting material for an interfacial solar steam generator. The bilayered structure of PDA/BNC was achieved by growing a thin BNC layer ($\sim 100 \mu\text{m}$) with PDA particles (as a photothermally active layer) on top of a thick pristine BNC hydrogel ($\sim 4 \text{ mm}$, as a heat insulation and water transport layer) (Figure 4.2G). As the bacteria produce a dense entangled layer of cellulose fiber network at the air/bacteria-growth medium

interface, the high density of PDA particles can be loaded within the top layer, which is evident in the surface and cross-sectional SEM images (Figure 4.2H, I). A cross-sectional SEM image also shows the sharp interface between the PDA-loaded and pristine BNC layers. Thermogravimetric analysis (TGA) determined that the loading of PDA particles to be around 43 % (appendix 3, Figure S3.2).

4.42 Optical and Thermal Properties of PDA/BNC

To evaluate the light absorption properties of PDA/BNC, we measured the optical transmittance and reflectance of BNC and PDA/BNC. Pristine BNC hydrogel (~ 4 mm) showed high transmittance (~ 80%) and reflectance (~ 4.8%) in the visible region, which translates to relatively small light extinction (~ 15.2%) (Figure 4.3A). On the other hand, the PDA/BNC hydrogel (with thickness of ~ 4.1 mm) exhibited extremely small optical transmittance (~ 1.2%) and reflectance (~ 0.96%) in the visible region, indicating a large optical extinction (~ 98%) of the bilayer (Figure 4.3B). The large light extinction of the PDA/BNC results from the light absorption corresponding to the densely loaded PDA particles and the light scattering from the nanoscale cellulose fibers, which increases the optical path length within the bilayer. Such large optical extinction of the bilayer structures combined with the excellent photothermal activity of PDA particles makes PDA/BNC a highly promising candidate for the interfacial solar steam generation.

As mentioned above, one of the most intriguing properties of PDA is its strong adhesion to a broad range of materials with widely different surface chemistries.²⁴³ Hence, it is natural to imagine that PDA coating on BNC via self-polymerization under oxidative condition is a facile strategy for achieving PDA/BNC interfacial solar evaporator. However, even after five polymerization cycles (each for over 12 hours), the PDA/BNC hydrogel exhibited significantly lower light extinction (~ 87 %) than PDA-filled BNC (~ 98%) described above (appendix 3, Figure S3.3). The coating

process via polymerization results in only a thin layer of PDA on the nanofibers of the highly porous BNC. The significantly lower amount of PDA in the PDA-coated BNC compared to that in PDA particle-filled BNC results in the significantly lower light extinction of the former compared to the latter. This observation further underscores the importance of loading PDA particles with optimal optical properties (absorption matching solar spectrum) into the BNC matrix via an *in situ* growth strategy.

As the thermal conductivity of interfacial solar evaporator plays a key role in confining the heat to the evaporative surface, we investigated the thermal conductivity of the PDA/BNC under both dry and wet conditions considering that the PDA/BNC is in a semi-wet state during solar steam generation. PDA/BNC samples were sandwiched between two glass slides maintained at different temperatures at both ends. The gradient in the temperature along the thickness of samples was observed using an IR camera (insets of Figure 4.3C, D). The thermal conductivity of dry PDA/BNC ($0.037 \text{ W m}^{-1} \text{ K}^{-1}$) is just slightly higher than air (Figure 4.3C). The low thermal conductivity of dry PDA/BNC owes to the large porosity (*i.e.* filled with air pockets with a thermal conductivity of $0.024 \text{ W m}^{-1} \text{ K}^{-1}$ at room temperature) of the structure. On the other hand, wet PDA/BNC exhibited a thermal conductivity ($0.442 \text{ W m}^{-1} \text{ K}^{-1}$) lower than water ($0.600 \text{ W m}^{-1} \text{ K}^{-1}$ at room temperature) (Figure 4.3D). The low thermal conductivity of wet PDA/BNC results from the low thermal conductivity of bulk BNC layer and nanocellulose fibers surrounding PDA particles, which serve as thermal insulators. Thus, the semi-wet PDA/BNC can significantly reduce the heat transferred from the evaporative surface to bulk water. Therefore, the low thermal conductivity of PDA/BNC confines the photothermal heat close to the evaporative surface by minimizing the heat transfer (loss) to bulk water and improves the overall efficiency of the solar steam generation.

4.43 Solar Steam Generation Performance of PDA/BNC

Now we turn our attention to the solar steam generation performance of the PDA/BNC under 1 kW m⁻² (~ one sun) and 3 kW m⁻² (~ three sun). IR imaging was employed to monitor the temperature rise in the PDA/BNC floated at air/water interface upon irradiation with a simulated solar beam (power density of 1 kW m⁻² or 3 kW m⁻²). Upon irradiation (3 kW m⁻²), the temperature of the PDA/BNC rapidly increased from room temperature (25 °C) to ~ 43 °C in the case of 1 kW m⁻² irradiation and to ~ 72 °C (Figure 4.4A, B). On the other hand, the temperature rise of water in the absence of PDA/BNC layer was found to be significantly smaller (2-4 °C). The large rise in temperature of PDA/BNC film under 3 kW m⁻² illumination resulted in the appearance of steam above the 100-mL beaker, evidencing the rapid evaporation of water (Figure 4.4A).

The efficiency of the PDA/BNC interfacial evaporator was quantified by measuring the weight loss (due to water evaporation) as a function of irradiation time (all tests were performed for 45 min). The cumulative weight loss was found to increase linearly with irradiation time (Figure 4.4C). Under one sun irradiation, the evaporation rate was found to be 1.13 kg m⁻² h⁻¹. On the other hand, in the absence of the PDA/BNC steam generator, the evaporation rate of water under identical irradiation was only 0.46 kg m⁻² h⁻¹, which is around 2.45 times lower than that of the PDA/BNC. Similarly, the steady state evaporation rate under 3 kW m⁻² with PDA/BNC is 3.47 kg m⁻² h⁻¹, 2.4 times higher than unaided water. Without considering the losses involved in the test setup, such as reflection and surface radiation, the evaporation efficiency of PDA/BNC was calculated to be 78% under one sun, which is comparable to other reported evaporators that rely on potentially toxic carbon nanomaterials.^{232, 254-255} The evaporation efficiency of unaided water is only 32% due to the poor photothermal conversion and the large energy loss associated with bulk water heating. With increasing solar power density, the evaporation efficiency also increased,

reaching 82% under 3 kW m^{-2} (Figure 4.4D). We suggest that the excellent steam generation efficiency of PDA/BNC owes to the following four factors: (i) tailored absorption of PDA particles, which matches with solar spectrum combined with outstanding photothermal conversion efficiency; (ii) high density of PDA particles loaded in the top layer of PDA/BNC combined with the efficient light scattering enabled by nanocellulose fibers, which greatly increase the optical path length and light absorption; (iii) low thermal conductivity of PDA/BNC layer due to BNC matrix, which minimizes the heat transfer from the evaporation surface to the bulk water; and (iv) open porous structure and hydrophilic nature of BNC aerogel enabling the rapid transport of water from bulk to the evaporative surface.

4.44 Robustness of PDA/BNC

We then evaluated the robustness of the PDA/BNC foam. The PDA/BNC is subjected to rigorous mechanical agitation and boiling for 2 hours during cleaning procedure to remove the bacteria and culture medium residue. Despite the strong mechanical agitation, PDA/BNC did not exhibit any signs of disintegration or loss of PDA particles. To further test its stability, we subjected PDA/BNC to ultrasonic agitation (483 W) for 5 hours and vigorous shaking for 30 days. Even after the vigorous mechanical agitation, the membrane did not exhibit any signs of disintegration or loss of PDA particles (Figure 4.5A, left part). For comparison, we have prepared a PDA/BNC composite by depositing PDA particles onto BNC via vacuum filtration method. After only 15 min of ultrasonic agitation or shaking, most of the PDA particles detached from BNC, which indicates poor stability, highlighting the advantage of using *in situ* growth strategy to load PDA particles (Figure 4.5A, right part). The robustness of PDA/BNC solar steam generator enables us to reuse it multiple times without any noticeable degradation of the structure or steam generation ability. Over 20 cycles of reuse, the steady-state evaporation rate and cumulative weight loss over

15 min irradiations (even under a higher power density, 7 kW m^{-2}) was found to exhibit less than 6% variation (Figure 4.5B). The structure of PDA/BNC evaporator remained unaltered after cycling tests (involving around 5 h high temperature solar exposure), which is evident from SEM images of the PDA/BNC surface before and after cycling (Figure 4.5C). TGA results also suggested the excellent thermal stability of BNC (up to $280 \text{ }^{\circ}\text{C}$) and PDA (with 58% left up to $800 \text{ }^{\circ}\text{C}$) (appendix 3, Figure S3.2).

4.45 Biodegradability of PDA

Finally, we turn our attention to the biodegradability of the solar steam generator. As mentioned above, both BNC and PDA are well-known biocompatible and biodegradable materials. Numerous reports in the past have demonstrated the enzymatic degradation of BNC.²⁵⁶⁻²⁶² However, very limited amount of work has addressed the enzymatic degradation of PDA and PDA microparticles.²⁴⁴ It has been shown that natural occurring melanin can be degraded by microbial species due to the presence of melanin-degrading enzymes, such as manganese peroxidase, lignin peroxidase and laccase.²⁶³⁻²⁶⁶ Considering the structural similarity between eumelanin and PDA, we employed laccase to demonstrate the enzymatic degradation of PDA particles. After subjecting PDA particles to an extremely high enzymatic concentration (50 mg/mL , 0.5 U/mg) for 48 hours (accelerated testing), the colour of the PDA solution faded, indicating the enzymatic degradation of PDA (Figure 4.6A). UV-vis spectra of pristine PDA particles showed a strong absorbance band around 700 nm . On the other hand, laccase-treated PDA solution exhibited a broad and significantly weaker absorbance band around 550 nm (Figure 4.6B). The weak absorbance band corresponds to the residue PDA particles with significantly smaller size. The biodegradation of the PDA particles is also evident from SEM images obtained before and after exposing them to the enzyme (Figure 4.6C, D). It should be noted that the biodegradation tests demonstrated above

are highly accelerated by using high enzyme concentration. In reality, degradation timescales are expected to be much longer than the life-time of the photothermal evaporator. Furthermore, the high temperature at the surface of the solar steam generator can kill microorganisms, which will prolong the biodegradation time.

4.5 Conclusions

In summary, we have demonstrated a flexible, scalable and, more importantly, completely biodegradable photothermal evaporator for highly efficient solar steam generation. The PDA/BNC bilayered evaporator was achieved by high density loading PDA particles within a BNC hydrogel during its growth. The size of the PDA particles could be independently tailored to achieve light absorption properties matching the solar spectrum. Owing to its large light absorption, outstanding photothermal conversion, heat localization, and efficient water transportation, the PDA/BNC bilayer foams exhibited excellent solar steam generation performance under one sun (efficiency of ~78%). The material platform introduced here is completely biodegradable and highly environmentally-friendly, while not compromising the solar steam generation efficiency. Ultimately, the novel material and process introduced here can be a sustainable solution to alleviate global water crisis.

4.6 Supporting Information

Supporting Information for chapter 4 is provided in appendix 3.

4.7 Figures

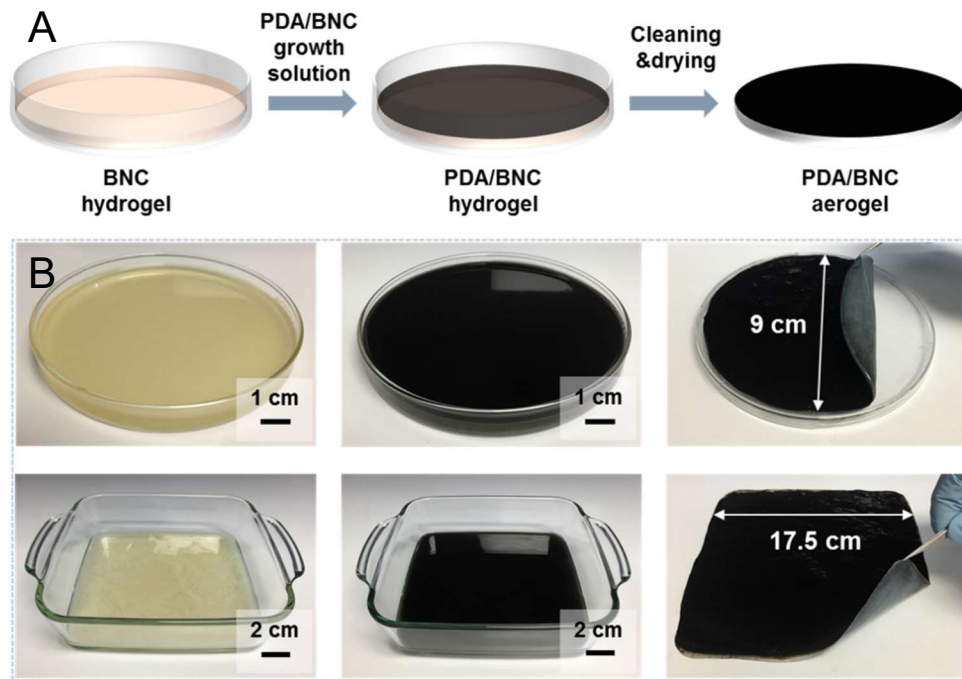


Figure 4.1. (A) Schematic illustration depicting the fabrication of bilayered PDA/BNC. (B) Photographs showing PDA/BNC hydrogel with tunable sizes and shapes.

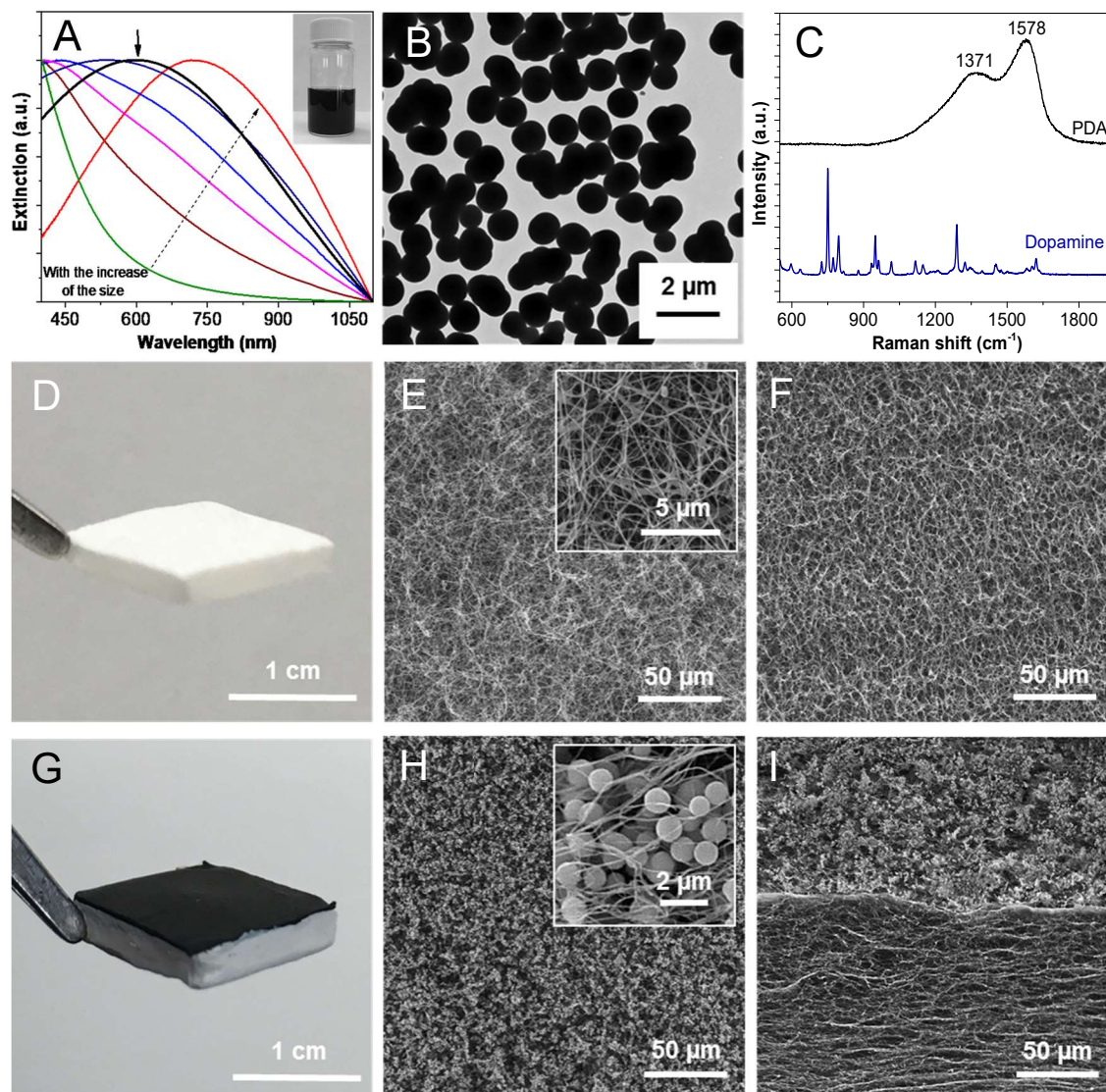


Figure 4.2. Characterizations of PDA particles and PDA/BNC. (A) Vis-NIR extinction spectra of PDA particles with varying sizes (inset is the photograph of PDA particles solution). (B) TEM images of the PDA particles. (C) Raman spectra of Dopamine and PDA. Optical image (D) and SEM images of (E) surface (inset is the image of higher resolution) and cross-section (F) of a pristine BNC foam. Optical image (G) and SEM images of (H) surface (inset is the image of higher magnification) and cross-section (I) of a PDA/BNC foam.

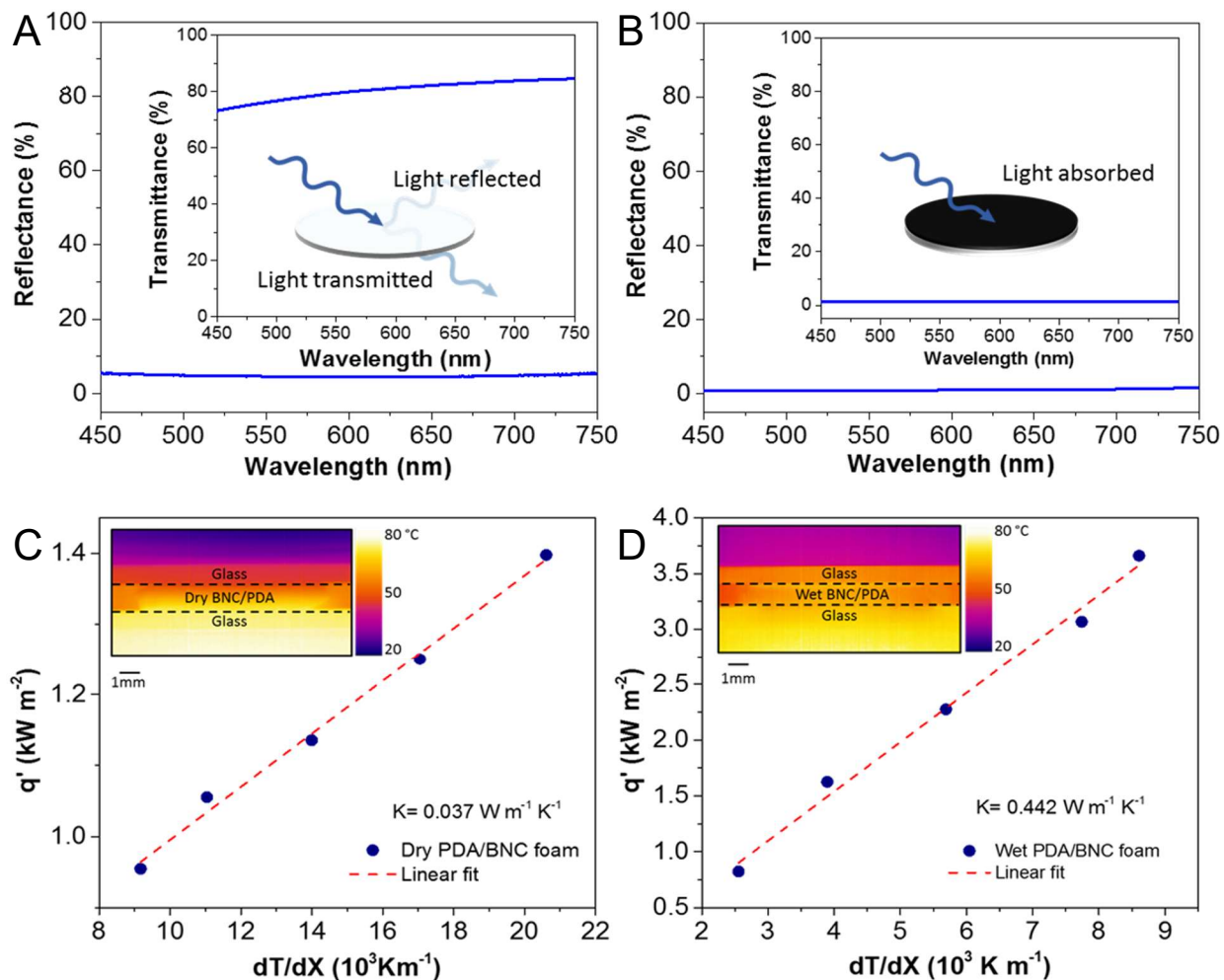


Figure 4.3. Optical and thermal properties of PDA/BNC. Transmittance and reflectance spectra of (A) BNC hydrogel and (B) PDA/BNC hydrogel. Thermal conductivities of (C) dry BNC/PDA foam and (D) wet PDA/BNC hydrogel. Insets: Representative IR images showing the temperature gradient along the thickness of the samples.

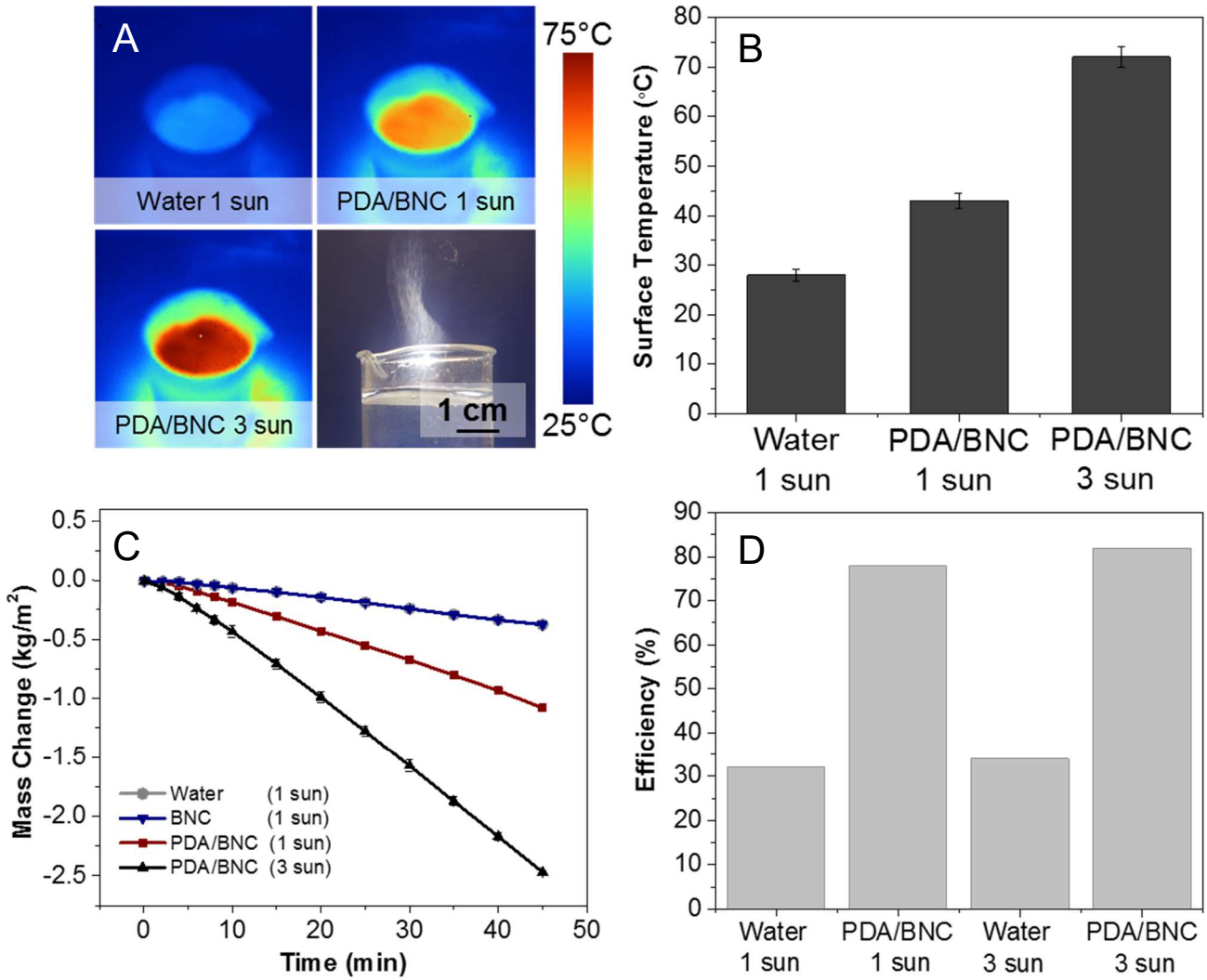


Figure 4.4. Solar steam generation performance of PDA/BNC. (A) IR images of water under 1 kW m^{-2} solar irradiation, PDA/BNC under 1 kW m^{-2} and 3 kW m^{-2} solar irradiation and optical image, showing visible steam generation under 3 kW m^{-2} . (B) Surface temperatures of water and PDA/BNC foam under 1, 3 kW m^{-2} irradianations. (C) Plot showing the cumulative weight losses through water evaporation of water and PDA/BNC foam under different solar irradianations. (D) Steam generation efficiencies of water and PDA/BNC foam under different solar irradianations.

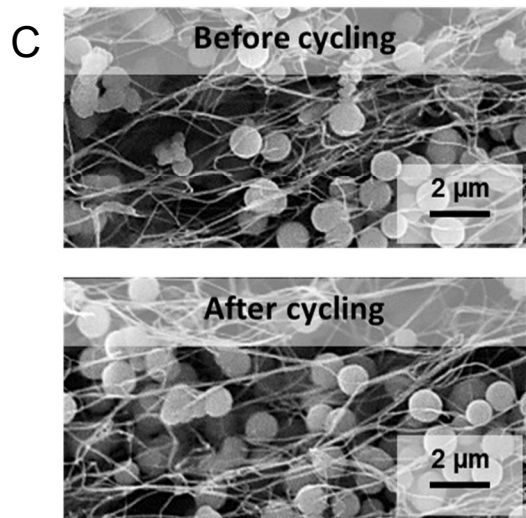
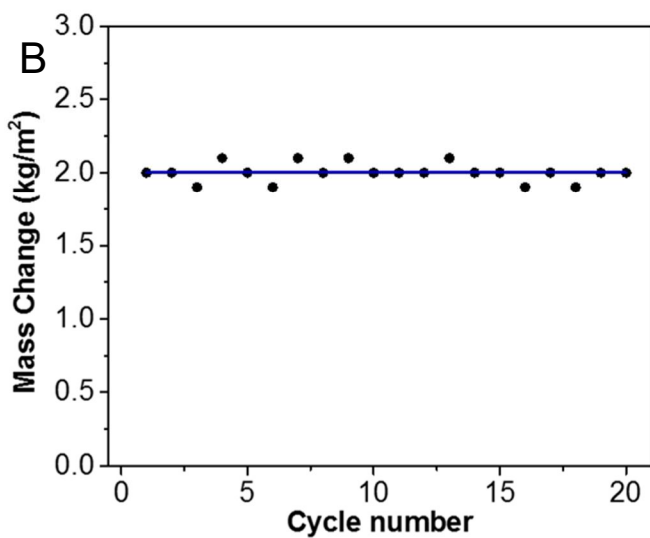
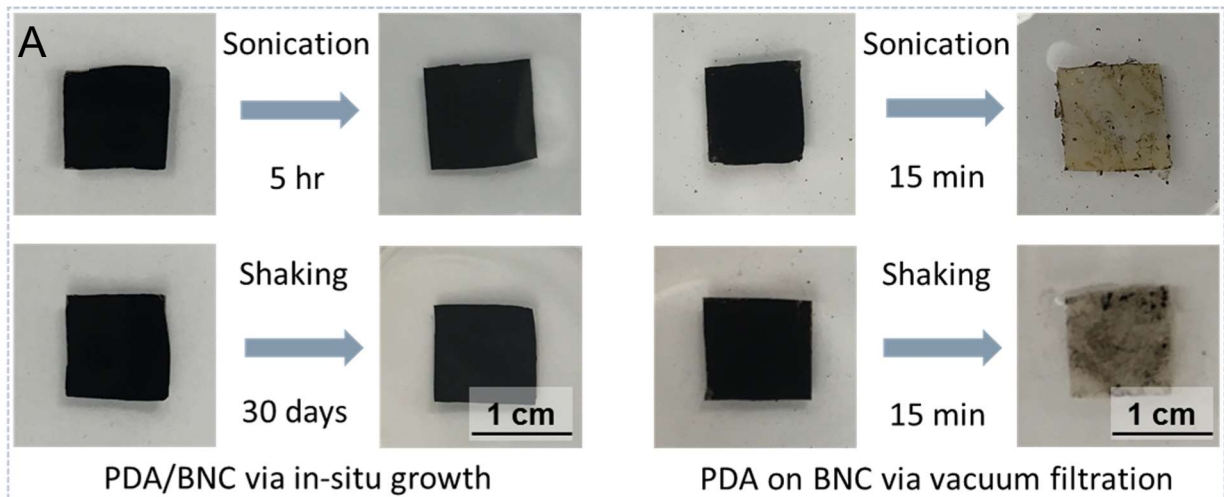


Figure 4.5. Stability of PDA/BNC. (A) Optical images of PDA/BNC foam achieved via *in situ* growth and vacuum filtration that have been subjected to sonication and shaking for extended duration. (B) Cycling solar steam generation tests under 7 kW m^{-2} solar irradiation for 15 min over 20 cycles. (C) High-resolution SEM images of the PDA/BNC surface before and after 20 cycles of solar steam generation depicting the intact structure of the PDA/BNC foam.

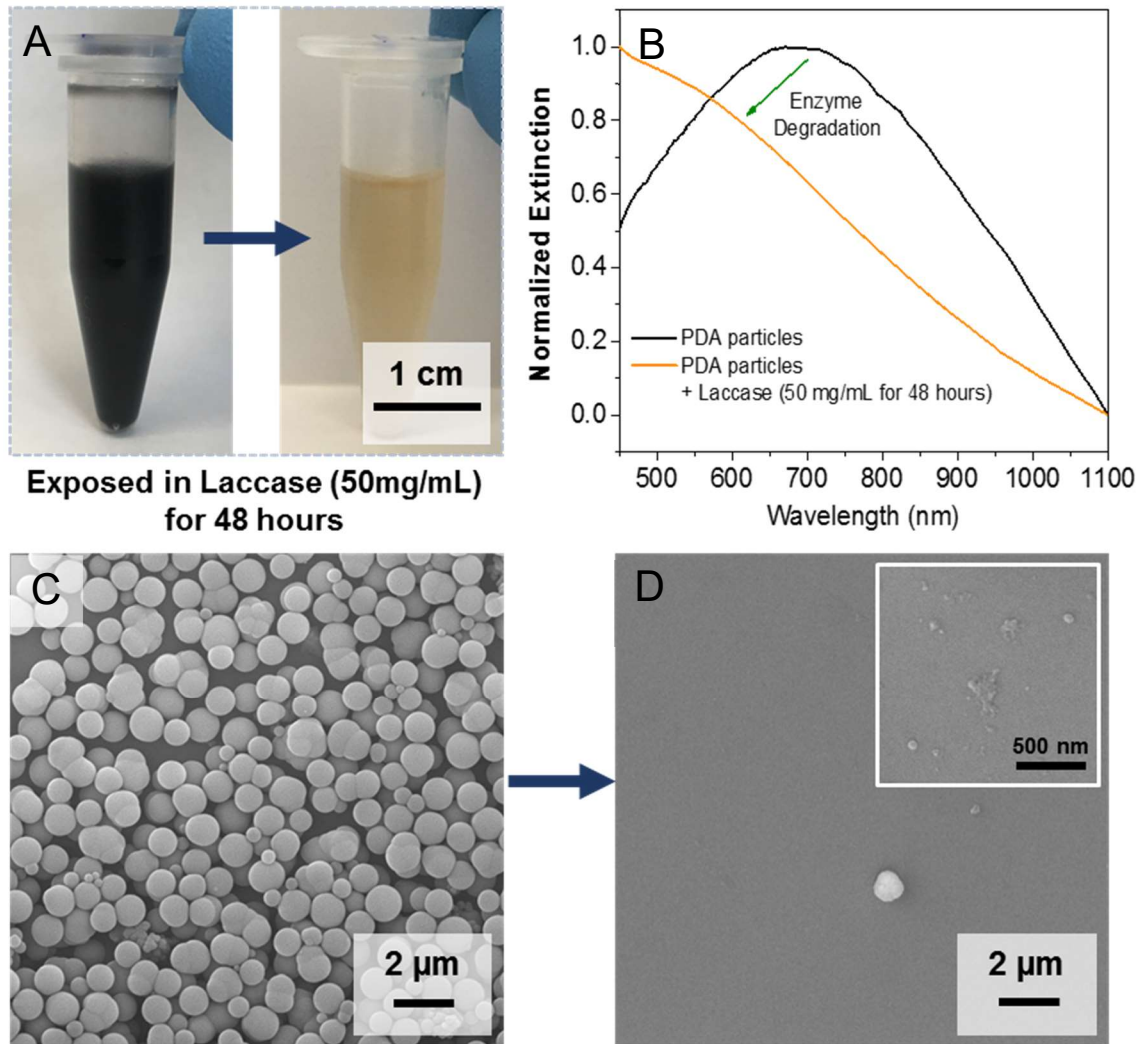


Figure 4.6. Biodegradability of PDA. (A) Optical images showing the pristine PDA and laccase-treated PDA solution. (B) Vis-NIR extinction spectra of pristine, laccase-treated PDA solution. SEM images of (C) pristine PDA and (D) laccase-treated PDA solutions.

Chapter 5: Biofouling-resistant, photothermally-active Ultrafiltration Membrane

5.1 Abstract

By severely decreasing water flux and driving up operational costs, biofouling poses one of the most serious challenges to membrane technologies. Here, we introduce a novel anti-biofouling ultrafiltration membrane based on reduced graphene oxide (RGO) and bacterial nanocellulose (BNC), which incorporates GO flakes into BNC in situ during its growth. In contrast to previously reported GO-based membranes for water treatment, the RGO/BNC membrane exhibited excellent aqueous stability under environmentally relevant pH conditions, vigorous mechanical agitation/sonication, and even high pressure. Importantly, due to its excellent photothermal properties, under light illumination, the membrane exhibited effective bactericidal activity, obviating the need for any treatment of the feed water or external energy. The novel design and in situ incorporation of the membranes developed in this study present a proof-of-concept for realizing new highly efficient and environmental-friendly anti-biofouling membranes for water purification.

5.2 Introduction

Water scarcity is recognized as one of the most critical global challenges in the 21st century.³⁹⁻⁴¹ In response to this dire need, various membrane technologies are being actively investigated for water purification and reclamation.⁵¹⁻⁵³ However, fouling and consequent degradation of membranes during performance still remain a ubiquitous problem.⁵⁴ The three major fouling mechanisms, mineral scaling, organic fouling, and biofouling, all lead to a decline in water flux.⁵⁵

Among them, biofouling, accounting for more than 45% of all membrane fouling, is the Achilles heel of the technologies, due to the difficulty of completely removing microorganisms.⁵⁶⁻⁵⁸

To inhibit the formation of biofilm, various biofouling controlling strategies, such as adjusting pH, adding disinfectants and biocides, and introducing quorum sensing molecules, have been suggested.⁵⁹⁻⁶² However, most of these strategies introduce considerable operational costs and/or potential hazardous contaminants.⁵⁹ Researchers have also investigated the incorporation of nanomaterials (e.g., silver nanoparticles, TiO₂ nanoparticles, and graphene oxide nanosheets), polymers (e.g., polyethylene glycol, polyvinyl alcohol, polyvinylpyrrolidone, and zwitterionic polymers), and other materials (e.g., small organic molecules and biomacromolecules) to engineer fouling-resistant membrane surfaces that can reduce biofilm growth and inactivate bacteria.^{55, 63-68} However, most of these methods involve additional thermal or chemical treatment steps. Furthermore, most of the above techniques are effective for only a short period of time, because biofilm gradually adapts to imposed harsh environments.⁵⁶ Even if 99.9% of biofilm is removed, the residual cells are sufficient to grow back and form a new biofilm. Highly efficient and cost-effective methods that overcome biofouling on water purification membranes over a long period of time would immediately help meet the grand challenge of providing access to clean water.

The photothermal effect of materials can offer a unique solution to biofouling, obviating the need for harsh chemical treatments to achieve bacterial lysis. Photothermally-active materials can effectively absorb light and then convert it into heat. In our recent study, gold nanostars grown on graphene oxide (GO) flakes coating a commercial membrane were utilized as nanoheaters.⁶⁸ In the study, with laser irradiation, the photothermal properties of the gold nanostars and GO were utilized to quickly kill adjacent *Escherichia coli* bacteria, inhibiting the formation of biofilm on reverse osmosis membranes. While this study provided a promising example of utilizing the

photothermal effect to minimize biofouling on membranes, it would be even more beneficial if the membrane itself were comprised of photothermal materials.

GO has been recognized as efficient photothermal material because the closely spaced energy level from loosely bonded π electrons absorb the broad electromagnetic spectrum.²⁶⁷⁻²⁶⁹ The absorbed light energy excites electrons, and its electron relaxes to ground state through non-radiative decay, releasing the energy by heat.²⁶⁷⁻²⁶⁹ In addition to having an intriguing photothermal property, GO has frequently been employed as a membrane component owing to its excellent mechanical strength, facile synthesis, and ease of attachment by vacuum filtration, spin-coating, and drop-casting.⁴⁵⁻⁴⁸ In contrast to graphene flakes, which have a strong tendency to stack and aggregate in aqueous solutions, GO flakes are easily dispersed, making the membrane preparation process easier in aqueous media.²⁷⁰⁻²⁷¹ This dispersibility comes from rich oxygen-containing functional groups (carboxyl, epoxy, hydroxyl, and carbonyl groups).⁴⁹ However, the stability of current GO-based membranes is compromised by vigorous agitation, and pH and ionic strength variations that are within the typical range of feed waters.⁵⁰ Thus, there is a need to develop new scalable approaches to fabricate stable GO-based membranes.

Bacterial nanocellulose (BNC) is a highly pure cellulose produced by bacteria with low-molecular weight sugar as a food source. Through a series of biochemical steps, the bacteria form exterior cellulose nanofibers in aqueous cultures, and these fibers become entangled to form a 3D network hydrogel.³ Similar to other cellulose nanofibers and cellulose nanocrystals (CNC), BNC is highly attractive for membrane technologies in view of its excellent mechanical properties, tunable porosity, chemical functionalizability, easy synthesis, high scalability, and most important, low environmental impact.²⁵⁰ Therefore, BNC is a promising material for fabricating functional

composites through *in situ* growth or by adsorption of pre-synthesized nanostructures on the nanoscale cellulose fibers.¹¹⁻²⁰

Here, we demonstrate a novel and facile approach for fabricating an anti-biofouling ultrafiltration membrane, involving *in situ* incorporation of GO flakes into BNC during its growth. The reduced graphene oxide (RGO) incorporated BNC membrane not only exhibited outstanding mechanical and chemical stability under environmentally-relevant pH conditions and vigorous mechanical agitation/sonication, but also showed stable water flux under high pressure. Particularly, owing to its photothermal properties, the membrane exhibited light-enabled bactericidal activity, avoiding the need for any treatment of the feed water or any external energy. The novel design and preparation method introduced here are suggested the first steps toward realizing highly efficient, environmental-friendly, and biofouling-resistant membranes for water purification.

5.3 Experimental Section

Fabrication of RGO/BNC membranes. *Gluconacetobacter hansenii* (ATCC®53582) was employed to synthesize the cellulose nanofibers. To produce a dense bacterial suspension, the bacteria were cultured in test tubes containing 16 ml of #1765 medium at 30 °C for 3 days under shaking at 250 rpm. The #1765 growth medium is composed of 2% (w/v) glucose, 0.5% (w/v) yeast extract, 0.5% (w/v) peptone, 0.27% (w/v) disodium phosphate, and 0.5% (w/v) citric acid. To synthesize GO, we employed an efficient oxidation process, reported by Tour and co-workers.²⁰² For *in situ* incorporation of GO sheets in a BNC membrane, GO solution (150 mL of 0.0725wt%) was sonicated (2 hours), centrifuged, re-dispersed in #1765 medium, and then centrifuged again to concentrate a wet mixture of GO and medium after supernatant was decanted. Then the densely cultured *Gluconacetobacter hansenii* suspension was added to the GO/medium wet mixture, making a total of 150 ml with 0.0725 wt% GO contents. The solution was

subsequently transferred to a Pyrex bakeware dish (18 cm × 18 cm) and incubated at room temperature without disturbance. After 2 days, a thin hydrogel of GO/BNC had formed, and it was harvested from the bakeware for cleaning. To remove the residual bacteria and growth medium, the hydrogel was boiled in 2.5 L of 0.1 M NaOH aqueous solution for 2 h. The obtained RGO/BNC hydrogel was then dialyzed in de-ionized water for 1 day. The purified RGO/BNC hydrogel was dried to obtain an RGO/BNC membrane.

Microstructure characterization and property measurements. Scanning electron microscopy (SEM, FEI Nova 2300 Field Emission SEM at an accelerating voltage of 10 kV) provided micron-scale images of RGO/BNC and pristine BNC. Atomic force microscopy (AFM) images were obtained for determining the thickness of GO flakes, using a Dimension 3000 (Bruker Inc.) instrument in light tapping mode. To investigate the relative oxygen and carbon ratio of GO and RGO flakes, carbon 1s peak was analyzed by X-ray photoelectron spectroscopy (XPS, a Physical Electronics® 5000 VersaProbe II Scanning ESCA Microprobe). The pore size distribution of RGO/BNC membranes was measured by the Brunauer-Emmett-Teller (BET) method using an Autosorb-1C (AX1C-MP-LP) at 298 K.

Stability tests of RGO/BNC membrane. To study the stability of the RGO/BNC membrane, we placed it in Petri dishes filled with solutions at pH 4, 7, and 9 and sonicated them for 5 hours. The pH values were chosen because they occur in many natural and engineering aqueous systems.²⁷² Subsequently, the release of RGO from the membrane was quantified from the UV-Vis absorbance spectra (Shimadzu UV-1800 spectrophotometer, 400 nm to 1000 nm) of the solutions. SEM was used to monitor the surface morphologies of the RGO/BNC film after sonication. To study the mass change of RGO/BNC membrane before and after sonication, thermo-gravimetric analysis

(TGA) was performed using a TA Instruments Q5000 IR Thermogravimetric Analyzer in air (at rate of 5 °C/min).

Because the GO membranes were frequently prepared by vacuum filtration, we made two types of control samples for comparison.⁴⁶⁻⁴⁸ First, a similar amount of base-washed RGO flakes was deposited on top of a BNC hydrogel, using the vacuum-assisted method, and dried to obtain a dry film. Second, BNC dispersions were pre-mixed with GO solutions and were then filtered to make membranes. Both control membranes were subjected to the same aqueous stability tests as the RGO/BNC membrane (appendix 4, Figure S4.1).

Evaluation of mass transport performance of RGO/BNC membranes. The mass transport performance of RGO/BNC membranes was evaluated by using a side-by-side diffusion cell system. An RGO/BNC membrane was first mounted between the two cells (Adams & Chittenden Scientific Glass, 5 mL volume). Then, ethanol and water were used to rinse the membrane several times to avoid subsequent air bubble formation. To test diffusion-driven transport through an RGO/BNC membrane, 0.5 mM of rhodamine 6G (R6G, ~ 1 nm, 479 Da) and lysozyme (3.8–4 nm, 14300 Da) were used. R6G is cationic dye, thus it is positively charged.²⁷³ The isoelectric points for lysozyme are 10.5, and its pH value at 0.5 mM concentration is 3.53, indicating that lysozyme is positively charged under our experimental condition.²⁷⁴ The solute was introduced on the feed side, while the dialysate side was just DI water. Solutions in both cells were subjected to vigorous stirring to minimize concentration polarization effects close to the membrane. The diffusing concentration of the solute was monitored in the wavelength range of 300–700 nm, using a Shimadzu UV-1800 spectrometer. For replicates, we utilized the RGO/BNC membranes grown in independent batches, and diffusion tests were conducted with three replicates.

Ultrafiltration efficiency and flux tests. The water fluxes of the RGO/BNC membrane and commercial ultrafiltration membrane (YMGESP3001, GE) were tested using a benchtop cross-flow system, and then compared those permeate flux. The commercial ultrafiltration membrane, used for comparison, was designed for pre-treatment, dye reduction, and purification with 1,000 Dalton cut-off sizes. The benchtop cross-flow system included a crossflow membrane cell (CF042D, Sterlitech Corp.) and a Hydracell pump (M03S, Wanner Engineering, Inc). During the measurement of the permeate flux, we set the pressure and the feed flux at 100 psi and 0.66 L/min, with 25 °C water. For replicates, we utilized the RGO/BNC membranes grown in independent batches and measured the water fluxes with three replicates. Furthermore, gold nanoparticles (AuNPs) with a diameter around 5 nm were synthesized using the seed-mediated growth method,²⁷⁵ and their size distribution was determined from TEM images. AuNP solutions were then filtered by RGO/BNC membranes, using above cross-flow system under 100 psi. Before and after filtration, AuNP concentrations in filtration/permeate solutions were measured by UV-Vis spectrometry (400 nm to 1000 nm). The rejection rate (RR) was calculated using the equation below:

$$RR = \frac{E_f}{E_p} \times 100,$$

where E_f is the optical extinction of the feed solution, and E_p is the optical extinction of the permeate solution.

Photothermal and bactericidal performance of RGO/BNC membranes under illumination.

The Photothermal performance of the membranes was tested using a solar simulator (Newport 66921, Arc Lamp). Both the RGO/BNC membrane and BNC membrane were illuminated at a power density of 2.9 kW/m² for 180 sec. The temperature map of the surface of both membranes under water was monitored by an IR camera. To test bactericidal activity, MG 1655 *E. coli* was

grown in Luria-Bertani liquid medium at 37 °C. All cultures were in 125 mL baffled shake flasks (25 mL working volume, shaking at 225 rpm). Cells in log phase ($>10^8$ live cells/ mL) were harvested after 12 h of incubation and then used for bactericidal tests. A layer of MG 1655 *E. coli* biofilm was grown on the RGO/BNC and BNC membrane surfaces, and then exposed to simulated sunlight for 180 secs at 2.9 kW/m². Before/after light illumination, the biofilms were exposed to fluorescent dyes (Molecular Probes Live/Dead Bacterial cell viability kit) for 30 min, and then imaged under a Leica microscope to identify live (blue fluorescent filter, 340-380 nm) and dead (green fluorescent filter, 450-490 nm) cells.²⁷⁶

5.4 Results and Discussion

5.4.1 The fabrication of RGO/BNC membrane

The RGO/BNC membranes were fabricated by *in situ* incorporation of GO flakes within the BNC network via bacteria-mediated growth under aerobic and static conditions (Figure 5.1). To determine the average thickness of synthesized GO flakes, they were deposited on a silicon substrate and measured with AFM. The thicknesses were $\sim 1.0 \pm 0.2$ nm, corresponding to a bilayer of GO (the thickness of a monolayer is ~ 0.7 nm) (Figure 5.2A).²⁰⁶ After washing, GO flakes were dispersed in broth solution with bacteria at an optimized concentration to achieve a desired BNC growth rate (Figure 5.1A). The GO dispersed solution was left undisturbed under ambient conditions to obtain GO/BNC hydrogels. To remove bacteria and residual broth solution from *in situ* grown GO/BNC hydrogel, it was immersed in NaOH solution (0.1 M) at boiling temperature, which partially reduced the GO flakes (discussed in detail below). The cleaned RGO/BNC hydrogel was dried to obtain a large, robust RGO/BNC membrane (Figure 5.1B).

5.42 Chemical Composition and Ultrastructure of RGO/BNC Membrane

To understand the chemical reduction of GO during cleaning, GO flakes were added to a boiling temperature 0.1 M NaOH solution, which turned uniformly distributed GO particles into black aggregated particles, indicating the partial reduction and restacking of the RGO. To confirm the reduction of GO after exposure to the basic solution, X-ray photoelectron spectroscopy (XPS) was utilized (Figure 5.2B, 2C). The high resolution 1s spectra of carbon were obtained and were deconvoluted into three peaks, corresponding one sp^2 domain (C=C with a binding energy of 284.6 eV) and two oxidized sp^3 domains (C–O with a binding energy of 286.6 eV, and C=O with a binding energy of 288.2 eV).²¹² The relative carbon and oxygen ratio was calculated based on the peak area, and this ratio was utilized to estimate the reduction extent of GO. For synthesized GO, the C/O ratio was 1.7, suggesting that ~ 58% of the GO surface was oxidized (Figure 5.2B). After an extensive base wash, the C/O ratio increased to 4.6, indicating that ~37% of the oxygen functional groups were reduced (Figure 5.2C). This result confirms that base washing to kill residual bacteria also reduces GO flakes in the BNC matrix.

In the absence of GO flakes, bacteria-mediated growth results in a white, translucent BNC membrane with outstanding flexibility and mechanical strength (Figure 5.2D).²⁰¹ Because bacteria-mediated synthesis of nanocellulose requires oxygen, a dense network of nanocellulose fibers (20 –100 nm in diameter) forms near the air/liquid interface, where abundant oxygen is available (Figure 5.2E). As the oxygen diffuses deeper into the medium, the first dense layer sinks, and makes way for the formation of subsequent BNC layers, which, upon sinking, stack together to form a 3D BNC network.^{3, 20} Due to this “layer-by-layer” formation, the cellulose nanofibrils are preferentially oriented parallel to the surface (i.e., normal to the thickness) of the membrane, which results in denser physical entanglements of the cellulose nanofibers parallel to the surface

(Figure 5.2F). To form RGO/BNC membranes, GO flakes were added during *in situ* growth, and a subsequent base washing process was conducted (Figure 5.2G). Compared to pristine BNC membrane, the RGO/BNC membrane is smoother and has fewer fibrils protruding from the surface, due to the presence of 2D RGO sheets (Figure 5.2H). Cross-sectional SEM images showed that the membrane was $\sim 8 \mu\text{m}$ thick (Figure 5.2I). The images also show the embedded RGO flakes between BNC layers due to the “layered” formation of BNC, starting from the liquid/air interface as described above (Figure 5.2I).

5.43 Stability of RGO/BNC Membrane

To investigate the mechanical and chemical stability of the membrane, we exposed it to ultrasonic agitation (483 W, 8892, Cole-Parmer) for 5 hours in solutions at pH 4, 7, and 9 (appendix 4, Figure S4.1). Even after this vigorous mechanical agitation at environmentally-relevant pH conditions, the membrane did not exhibit any signs of disintegration or loss of RGO flakes (Figure 5.3A and its inset). This result was further supported by SEM images of the membrane surface, which did not show a discernable change in the morphology after sonication (Figure 5.3B). Then, to evaluate the thermal stability of the RGO/BNC membrane, thermogravimetric analysis was conducted. The membrane showed a first mass loss of $\sim 2 \text{ wt.}\%$ at $\sim 100 \text{ }^\circ\text{C}$, due to the loss of absorbed water. A second mass loss of $\sim 3 \text{ wt.}\%$ at $\sim 200 \text{ }^\circ\text{C}$ was attributed to the decomposition of functional groups of GO.²⁰⁴ A third mass loss of $\sim 46 \text{ wt.}\%$ began at 280°C , and was due to the degradation of cellulose. A fourth mass loss ($\sim 49 \text{ wt.}\%$) at $390 \text{ }^\circ\text{C}$ was attributable to the continued decomposition of cellulose residual and sublimation or burning of damaged graphitic regions.²⁰³ Based on the mass loss profiles of the RGO/BNC membrane, RGO, and pristine BNC membrane, the mass loading of RGO in RGO/BNC was calculated to be $\sim 45 \text{ wt.}\%$, and it suggested excellent thermal stability of the RGO/BNC membrane up to $200 \text{ }^\circ\text{C}$. The RGO/BNC membrane after ultrasonic

treatment also showed an identical mass loss profile, implying that the embedded RGO flakes within the BNC matrix remained intact (Figure 5.3C).

As mentioned above, GO-based membranes have been extensively investigated in the past several years. Most of these membranes were fabricated by vacuum filtration of GO flakes onto a supporting membrane, but this coating method always leaves doubt about its long-term aqueous stability.⁵⁰ Here, we compared the mechanical stability of *in situ* embedded RGO/BNC with that of an RGO/BNC membrane prepared by depositing RGO particles (base-washed) on top of a BNC membrane using vacuum filtration. After 5 h of ultrasonic agitation in solutions at pH 4, 7, and 9, the RGO particles had disintegrated completely, and the solution exhibited broad absorbance, a feature of RGO flakes in solution (Figure 5.3D). This disintegration was further confirmed by surface SEM images and quantified by TGA. After sonication, the membrane prepared through vacuum filtration showed an initial mass loss (~5%) at 100 °C, attributed to absorbed water, and the decomposition of cellulose at ~280 °C (~63%) and at ~340 °C (~32%). All of these findings indicated the absence of RGO flakes (Figure 5.3F).

Conventional GO-based membranes rely on hydrogen bonding or metal ion incorporation for mechanical stability. However, mechanical agitation during water filtration or cleaning procedures, as well as pH variations in the feed water, can compromise the mechanical stability of these membranes (appendix 4, Figure S4.1). In contrast, *in situ* RGO/BNC membranes are robust because during the BNC growth, plate-like GO particles are physically locked into the layered BNC matrices, which provides its excellent mechanical and chemical stability. Moreover, intensive vortexing has been employed to wash used *in situ* RGO/BNC membrane without damaging it (appendix 4, Figure S4.1D). This finding further suggests the durability of the *in situ* RGO/BNC membrane for practical use.

5.44 Mass Transport Performance and Water Flux Tests

To probe the diffusive transport capability of small molecules across the RGO/BNC membranes, we employed a two-cell setup (Figure 5.4A) and tested it with 0.5 mM of two model solute systems, having different sizes and molecular weights: rhodamine 6G (R6G, ~ 1 nm, 479 Da) and lysozyme (3.8–4 nm, 14300 Da).²⁷⁷ Because ultrafilters have pore sizes between 1 nm to 100 nm, and thus remove contaminants via a size exclusion mechanism, these two different solutes helped to determine the filtration capability of RGO/BNC membranes.²⁷⁸ A UV-vis spectrometer was used to monitor the concentration of model solutes from the feed side to the permeate side (see experimental section for details). For pristine BNC membrane, all two solutes rapidly diffused through because the BNC fiber network is composed of micro-scale pores even if the nanofibers are densely packed (Figure 5.4B). By contrast, the addition of graphene oxide flakes within the matrix completely blocked the diffusion of lysozyme (3.8–4 nm), although R6G (~1 nm) passed through (Figure 5.4B). These results are in accordance with the BET measurements which indicate the pore size of the RGO/BNC membrane is around 2.2 nm (radius), which falls into the range of ultrafiltration membranes (appendix 4, Figure S4.3).²⁷⁹

Moreover, the unique mass transport properties of GO-based membranes originate from the nanocapillary network formed by lamellar stacking of GO, and the mass-transport behavior can be adjusted by tuning functional groups or inserting external species with desired dimensions.^{45,48} In this work, the presence of bacterial cellulose nanofibers between RGO flakes will lead to an overall tortuous network of pores in the membrane, even though no visible pores can be seen in a SEM image of the surface of RGO/BNC membranes.

To further demonstrate the potential of the novel RGO/BNC membrane for an ultrafiltration system, we performed flux tests and particle rejection tests using size-controlled gold nanoparticles

(AuNPs) via a benchtop cross-flow system (setup diagram in Figure 5.4C). For a particle rejection test, spherical gold nanoparticles with diameters of 5.15 ± 0.4 nm were synthesized using the seed-mediated method.²⁷⁵ The prepared AuNPs showed a very narrow size distribution (RSD < 8 %), which made the particle rejection study accurate (appendix 4, Figure S4.2B). The particle rejection rates were calculated by measuring the UV-vis extinction spectra of solutions before/after filtration through the RGO/BNC membranes (Figure 5.4D). This result suggested that AuNP of 5 nm diameter were $\sim 100\%$ rejected (inset of Figure 5.4D). In the same way, the rejection test was also performed for a commercial ultrafiltration membrane (pore size of around 1.66 nm), which also showed $\sim 100\%$ rejection for 5 nm gold nanoparticles (appendix 4, Figure S4.2C).

Under 100 psi, the RGO/BNC membranes showed higher water fluxes than a commercial ultrafiltration (UF) membrane. Because the pore size for both membrane types belongs to the range between UF and nanofiltration (NF), we intentionally tested water flux at a higher operating pressure than the usual operating pressure for UF (7.3 – 73 psi). Over a five-hour-long flux test after stabilization, the water flux of the RGO/BNC membrane was found to be 52.6 ± 2.5 L/m² h, and that of the commercial ultrafiltration membrane was 21.6 ± 0.8 L/m² h (Figure 5.4E). Importantly, the RGO/BNC membrane withstood an operating pressure as high as 100 psi without any supporting membrane. This performance emphasized the remarkable mechanical strength of RGO/BNC, considering that most of the GO-based membranes reported in the literature require a support membrane or a carefully designed apparatus due to their limited mechanical strength.²⁸⁰⁻

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5.45 Photothermal and Bactericidal Performance under Light

Next, we examined the photothermal and bactericidal ability of the RGO/BNC membrane using a Newport 66921 Arc Lamp with a power density of 2.9 kW/m². We used IR imaging to monitor

the temperature profile of the RGO/BNC membrane in an aqueous environment during illumination (Figure 5.5B). Upon illumination, the temperatures of the RGO/BNC membrane rapidly increased from room temperature (26 °C) to ~60 °C (Figure 5.5C). Specifically, the temperature rapidly increased during the first 20 seconds after the onset of irradiation and remained constant over the entire duration (120 sec). In comparison, the pristine BNC membranes showed only a small temperature increase (3 °C). The large rise in temperature was caused by the broad optical absorption of many RGO flakes arranged within the BNC matrix. Once light was absorbed by the RGO flakes, they immediately generated heat, which then dissipated to the surrounding water and BNC.²⁰ Due to the decrease of thermal radiation along the distance between IR camera and surface (~30 cm), the temperature profile obtained from the IR camera underestimated the actual temperature at the surface of the RGO/BNC membrane. Therefore, biological species were exposed to temperature higher than ~60 °C. To demonstrate that the heat generated by light exposure will not damage the mass transport and water flux performance of the RGO/BNC membrane, we performed BET and water flux tests before and after long duration of light exposure (2.9 kW/m²). Both the pore size distribution and the water flux performance of RGO/BNC membrane remained the same after light exposure (appendix 4, Figure S4.3, S4.4).

To test the bactericidal ability, the RGO/BNC membrane was covered with stained *E. coli* bacteria from a live/dead cell viability assay, and then light was shined on the membrane surface. Before irradiation, both RGO/BNC and pristine BNC membranes showed substantial and well distributed green fluorescence, corresponding to live bacteria, and no sign of red fluorescence, indicating the absence of dead bacteria (Figure 5.5D). After irradiation (2.9 kW/m²) for 180 secs, the bacteria on the RGO/BNC membrane exhibited predominantly red fluorescence (dead bacteria) and a complete visible absence of green fluorescence. However, the *E.coli*-covered pristine BNC

membrane exhibited green fluorescence corresponding to live bacteria, even after irradiation (Figure 5.5E). The SEM images showed morphology changes and leakage of bacteria, indicating that the high temperature at the RGO/BNC membrane had disrupted the cell walls and cell membranes (Figure 5.5F, G). Before light irradiation, live *E. coli* bacteria on an RGO/BNC membrane showed a rod-like structure, while after irradiation they were significantly shrunken and wrinkled. Because the cell walls of *E. coli* are known to deteriorate near 70 °C,²⁸⁵ this change implied that upon illumination, the surface of the RGO/BNC membrane had rapidly heated to above 70 °C and killed bacteria within a short time (180 sec). Live *E. coli* bacteria on an RGO/BNC membrane without illumination showed no dead bacteria cells even after 1 hour exposure, indicating the bactericidal activity of RGO/BNC membrane only occur upon light illumination (appendix 4, Figure S4.5). Here, we demonstrated the excellent bactericidal performance of the RGO/BNC membrane in harvesting light, and this capability makes the RGO/BNC membrane highly attractive for energy-saving and environmentally-friendly water purification applications.

5.5 Conclusions

In this study, we presented an innovative approach that uses the photothermal effect of RGO by embedding it in BNC structures. This new type of membrane can enhance the stability and durability of a membrane and inhibit or delay microorganism growth on its surface. While the most contemporary approaches to resisting biofouling rely on temporary chemical treatments, combining the photothermal effect with a noble membrane design shows that anti-biofouling can be achieved with sustainable and abundant sunlight.

We note that fully utilizing the photothermal property of RGO/BNC membranes can be challenging. A possible implementation involves modifying a spiral-wound module system, as

shown in Figure 5A. The inner and outer surfaces of the membrane modules or feed channel spacers can be equipped with low-energy LEDs for illumination, and can potentially be powered by renewable energy sources, such as low-cost photovoltaic devices or triboelectric nanogenerators (TENGs).²⁸⁶ In TENGs, mechanical energy generated through fluid flow in the UF process can be harnessed to produce light and to heat the membrane surface, reducing the overall operational expense. In addition to spiral-wound module system, this suggested system can also be applied to plate sheet membrane module by adding LED-equipped plates between the membrane modules. This would be an exciting future research direction for realizing photothermal membrane water filtration.

The novel fabrication method of incorporating RGO during ‘layer-by-layer’ growth of BNC yields a well stacked the structure, with a pore size in the UF membrane range. In addition, BNC production is considered eco-friendly because it needs only low-molecular weight sugar and oxygen as food sources.

Until now, many researchers prepared GO membranes through vacuum filtration or spin coating without a polymer matrix⁴⁵⁻⁴⁸, but these fabrication methods inevitably raised the mechanical stability concerns. However, the RGO/BNC membrane, reported here, exhibits stable water flux under 100 psi loading and maintains chemical stability at solution pH varying from 4 to 9. The water flux is higher than that of commercial UF membranes under identical pressure. In summary, RGO/BNC membrane demonstrated here not only provides a novel anti-biofouling approach powered by solar energy, but also suggests a scalable, eco-friendly, and cost-effective way to fabricate UF membranes for water purification.

5.6 Supporting Information

Supporting Information for chapter 5 is provided in appendix 4.

5.7 Figures

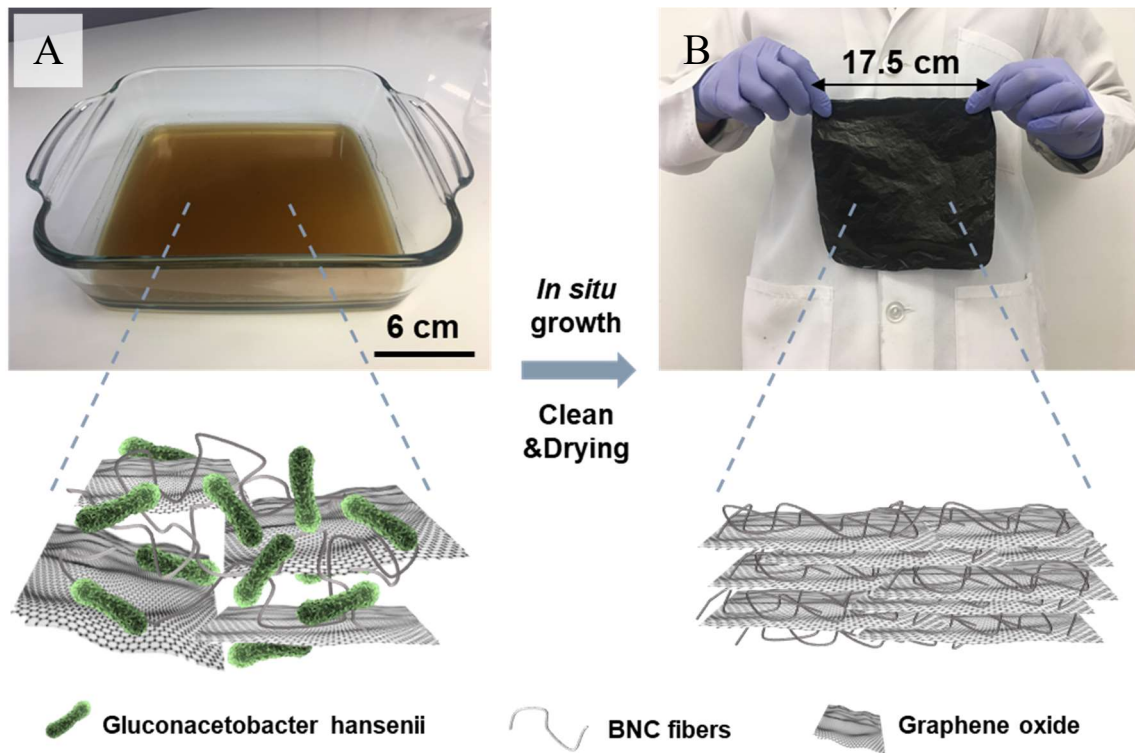


Figure 5.1. Fabrication of RGO/BNC membrane. Optical images showing (A) GO in bacterial medium and (B) *in situ* grown RGO/BNC membrane after cleaning and drying.

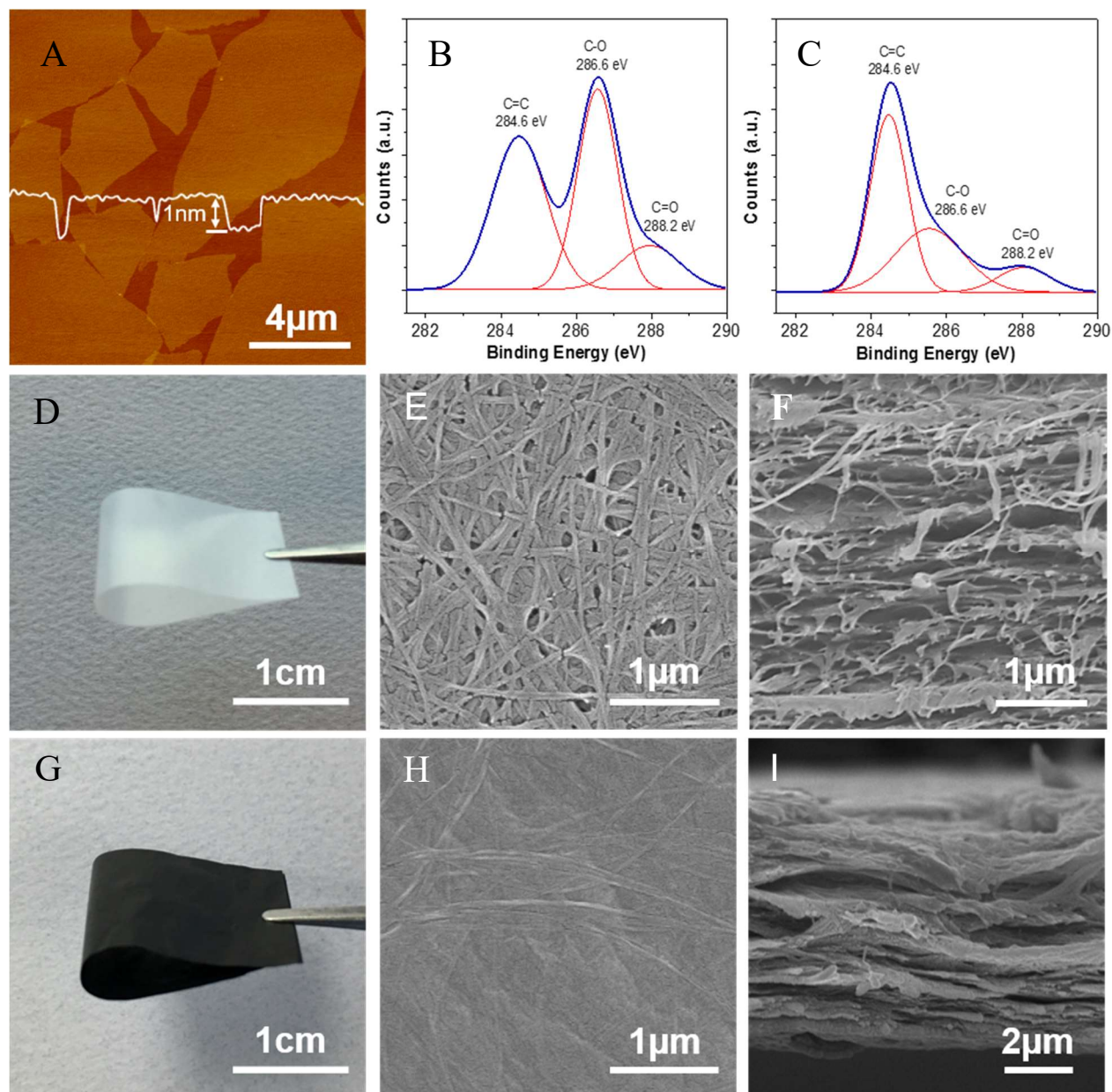


Figure 5.2. Chemical composition and ultrastructure of RGO/BNC membrane. (A) AFM image of GO flakes deposited on a silicon substrate. X-ray photoelectron spectra of (B) pristine and (C) base-washed GO. Optical image (D) and SEM images of (E) surface and (F) cross-section of a pristine BNC membrane. Optical image (G) and SEM images of (H) surface and (I) cross-section of a RGO/BNC membrane.

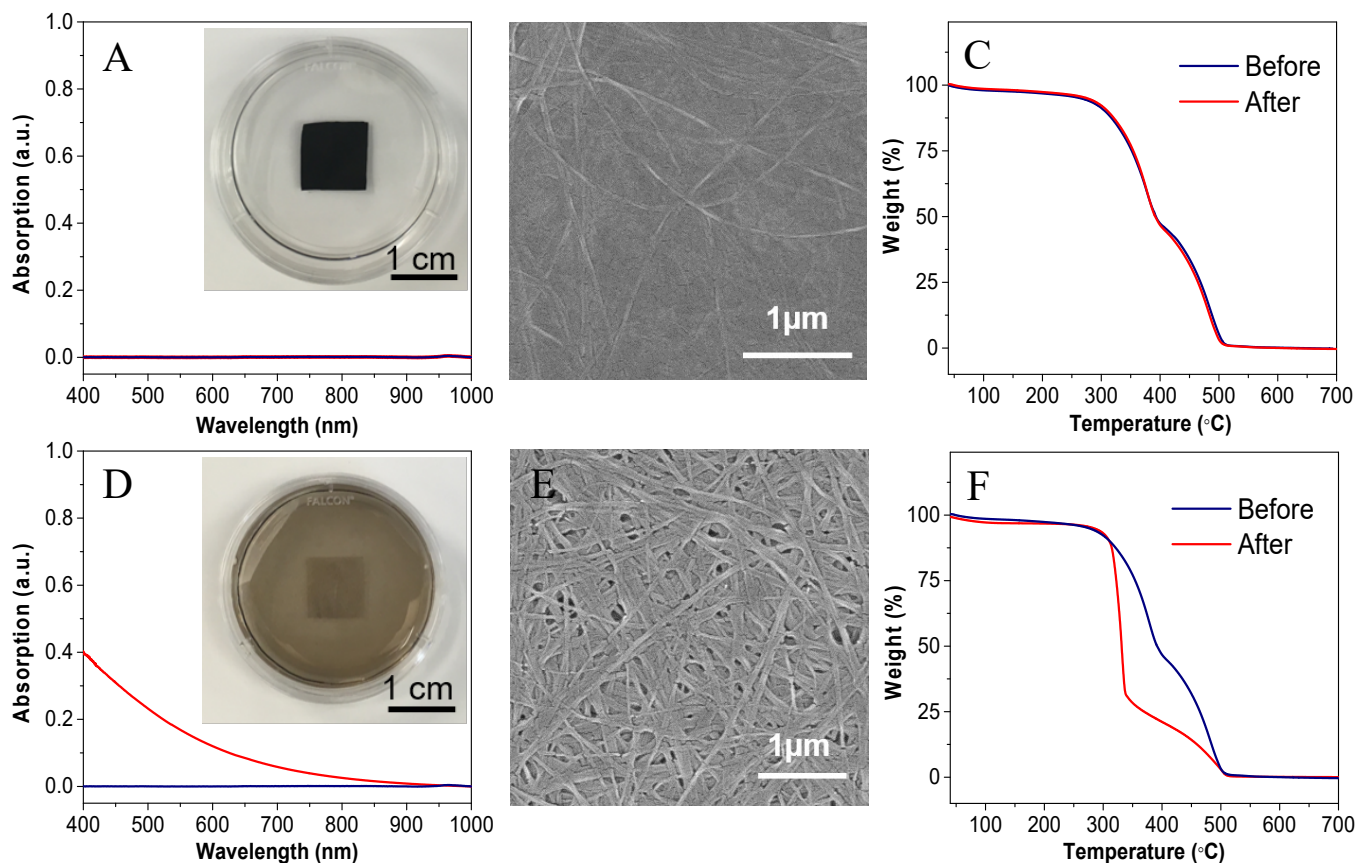


Figure 5.3. Stability of RGO/BNC membrane. UV-vis absorption spectra of (A) RGO/BNC and (D) RGO-coated BNC immersed solutions (at pH 7) before and after ultrasonic agitation for 5 h. The insets show the optical images of a RGO/BNC membrane and a RGO-coated BNC layer after sonication. SEM images of (B) a RGO/BNC and (E) a RGO-coated BNC layer before and after ultrasonic agitation. TGA analyses of (C) a RGO/BNC, (F) a RGO-coated BNC before and after ultrasonic agitation.

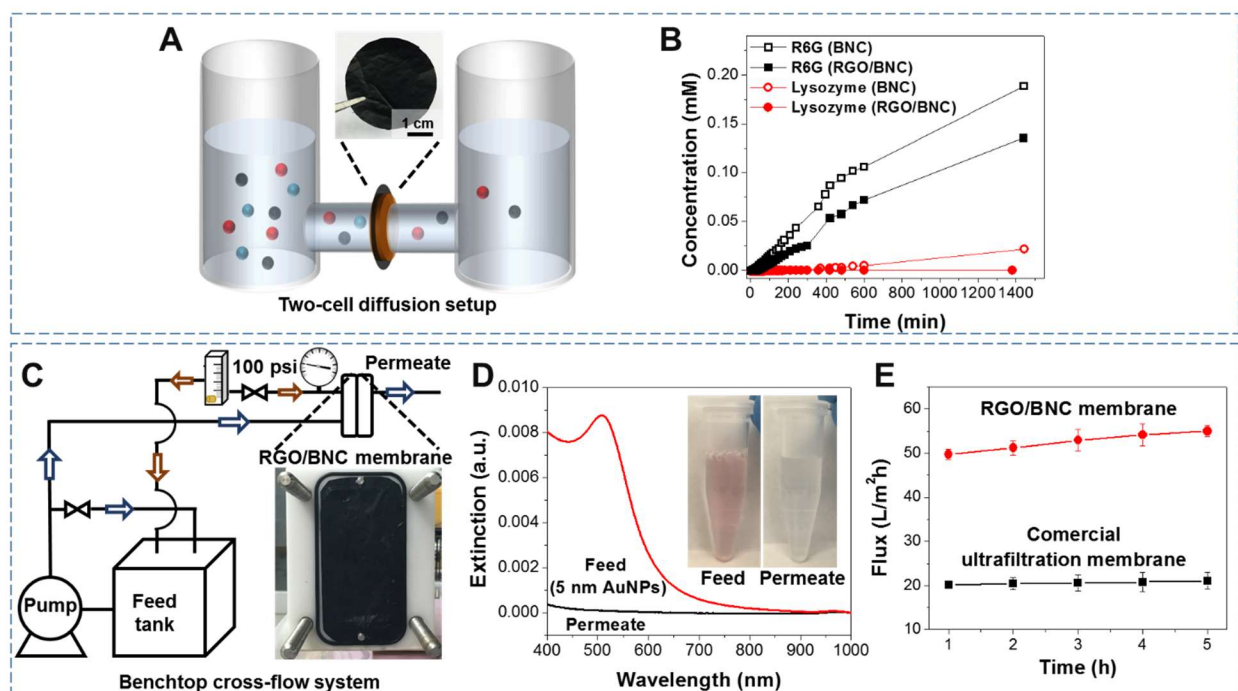


Figure 5.4. Mass transport performance and water flux tests. (A) Schematic diagram of a two-cells diffusion setup. (B) Diffusion of model solutes through pristine BNC membrane and RGO/BNC membrane. (C) Schematic diagram of cross-flow flux test setup. RGO/BNC membranes are placed in between the cross-flow cell and tightly sealed. (D) UV-vis extinction spectra indicating the rejection of AuNPs with 5 nm in diameter filtered through RGO/BNC membranes (inset is the picture showing feed and permeate solutions). (E) Water flux of RGO/BNC membranes ($\sim 8 \mu\text{m}$ thick) and commercial ultrafiltration membranes. 100 psi was applied for the flux tests.

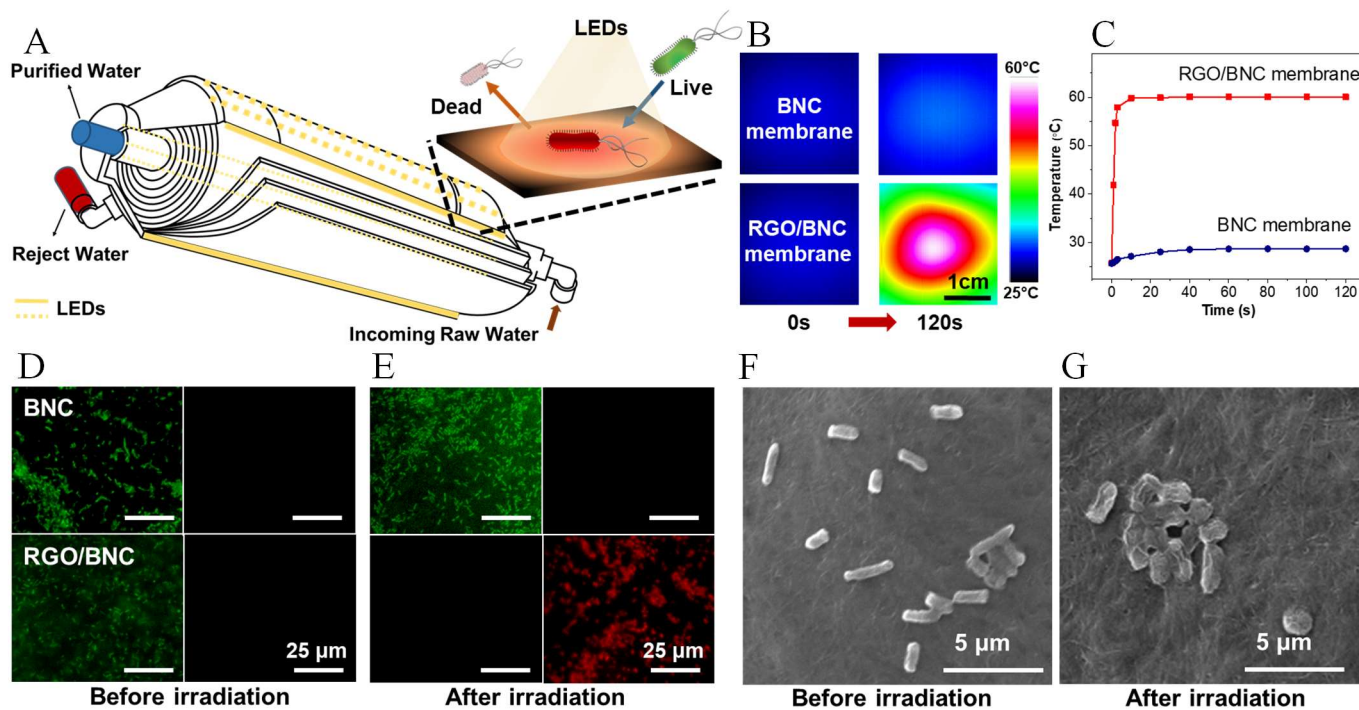


Figure 5.5. Photothermal and bactericidal performance under light. (A) Schematic showing antifouling mechanism of RGO/BNC membrane and a possible configuration of spiral wound UF module coupled with LEDs. (B) IR images showing the temperature of the pristine BNC and the RGO/BNC membrane in water under illumination at various time points. (C) Plot showing the temperature of pristine BNC and the RGO/BNC membrane in water under light of 2.9 kW/m² as a function of irradiation time. Fluorescence images of *E. coli* on BNC and RGO/BNC membranes (D) before and (E) after irradiation. SEM images of *E. coli* on RGO/BNC membranes (F) before and (G) after irradiation.

Chapter 6: Catalytically-active Bacterial Nanocellulose-based Ultrafiltration Membrane

6.1 Abstract

Large quantities of highly toxic organic dyes in industrial wastewater is a persistent challenge in wastewater treatment processes. Here, a novel membrane based on bacterial nanocellulose (BNC) loaded with graphene oxide (GO) and palladium nanoparticles (PdNPs) for highly efficient wastewater treatment is demonstrated. This Pd/GO/BNC membrane is realized through the *in situ* incorporation of GO flakes into BNC matrix during its growth followed by the *in situ* formation of palladium nanoparticles. The Pd/GO/BNC membrane exhibits highly efficient methylene orange (MO) degradation during filtration (up to 99.3% over a wide range of MO concentrations, pH and multiple cycles of reuse). Multiple contaminants (a cocktail of 4-nitrophenol, methylene blue and rhodamine 6G) can also be effectively treated by Pd/GO/BNC membrane simultaneously during filtration. Furthermore, the Pd/GO/BNC membrane demonstrates stable flux (33.1 L/m²·h) under 58 psi over long duration. The novel and robust membrane demonstrated here is highly scalable and holds a great promise for wastewater treatment.

6.2 Introduction

Water contamination caused by chemical and biological species released from industrial and agricultural practices is a formidable environmental challenge.^{51, 69-70} Among all the contaminants such as cleaning agents, agricultural chemicals, and noxious microorganisms, organic dyes, which are widely employed in textile, leather, cosmetic, pharmaceutical industries, pose significant threats to aquatic life and humans due to their carcinogenic and mutagenic nature.⁷¹⁻⁷³ Due to the

high resistance of organic dyes to traditional biological, physical, and chemical treatment methods, there is a critical need for the development of highly efficient and cost-effective methods for dye degradation and removal from contaminated water.⁷⁴⁻⁷⁶ Membrane technologies are being extensively investigated in wastewater treatments owing to its high efficiency, chemical stability, and low environmental impact.²⁸⁷⁻²⁸⁹ To achieve nearly complete removal of organic compounds from wastewater, a promising approach involves the utilization of membranes with specifically tailored micro- and nano-scale structures and permeation characteristics. These membranes can be further integrated with emerging technologies such as photocatalytic degradation, chemical oxidation, physical adsorption.²⁹⁰⁻²⁹² However, realizing scalable membranes with desired permeation properties and highly efficient organic dye removal performance remains to be a great challenge.

Noble and transition metal nanoparticles, which have been extensively employed in catalysis, organic synthesis, hydrogen storage, and water treatment, have shown great promise for organic dye removal.⁷⁷⁻⁸⁰ In particular, palladium nanoparticles (PdNPs) have attracted special attention for dye degradation owing to their excellent catalytic properties and low environmental impacts.⁸¹⁻⁸⁵ To achieve maximum catalytic activity and prevent aggregation of PdNPs, immobilizing PdNPs on various substrates such as carbon (e.g., carbon microspheres, carbon nanotubes, and graphene oxide), silica, and metal oxides (e.g., titanium dioxide) has been utilized.⁸⁶⁻⁹⁰ Recently, Hu and coworkers reported a novel mesoporous, three-dimensional (3D) wood which can be used as a robust substrate for PdNPs to achieve high-performance filtration membrane for efficient waste water treatment.²⁹³

Graphene oxide (GO) is a particular promising substrate material because of its high mechanical, chemical, thermal stability, and high surface area.⁹¹⁻⁹⁶ Various self-assembled GO-based

composites have been demonstrated for wastewater treatment.⁹⁷⁻¹⁰² To the best of our knowledge, a GO-based filtration membrane with transition metal nanoparticles as catalyst has not been reported since previous developed GO-based composites have limited aqueous stability and mechanical strength under pressure .

Owing to its desirable properties such as excellent mechanical strength and toughness, rich chemical functionality, and high specific surface area in aerogel state, bacterial nanocellulose (BNC) serves as an ideal material for realizing functional composites by incorporating functional nanomaterials within BNC matrix.^{11-19, 44, 250} Recently, we have demonstrated a novel, highly scalable, cost-effective and green strategy to realize functional GO/BNC-based membranes.²⁹⁴⁻²⁹⁵ GO sheets can be efficiently incorporated within BNC matrix during its growth, which can be applied to other two-dimensional (2D) materials. Owing to the intercalation of GO flakes within the layered BNC matrix, the membrane showed mechanical robustness, which is crucial for efficient, large scale water treatment.

Here, for the first time, we introduce a simple and scalable approach for the fabrication of Pd/GO/BNC membrane for highly efficient dye degradation. The fabrication of Pd/GO/BNC membrane involves the incorporation of GO flakes into the BNC matrix during its growth followed by *in situ* growth of PdNPs on GO flakes. The layered structure of Pd/GO/BNC, which forms nanocapillaries through out the membrane, maximizes the contact between organic dye contaminants and PdNPs anchored on the GO flakes. We demonstrate the novel membrane to effectively remove organic dyes, showing a great promise for wastewater treatment and separation applications.

6.3 Experimental Section

Preparation of GO/BNC aerogel: *Gluconacetobacter hansenii* (ATCC®53582) was cultured in test tubes containing 16 ml of #1765 medium at 30 °C under shaking at 250 rpm. The #1765 medium is composed of 2% (w/v) glucose, 0.5% (w/v) yeast extract, 0.5% (w/v) peptone, 0.27% (w/v) disodium phosphate, and 0.5% (w/v) citric acid. GO was synthesized using an approach reported by Tour and co-workers.²⁰² The as synthesized GO (150 mL of 0.0725 wt%) was centrifuged and redispersed in #1765 medium and then centrifuged again to obtain a wet mixture of GO and medium after decanting the supernatant. Subsequently, *Gluconacetobacter hansenii* culture solution and fresh medium were added into the above mixture to make a solution of total 150 mL (with GO concentration of 0.0725 wt%). The above solution was then transferred to a Pyrex bakeware (18 × 18 cm) and left without disturbance at room temperature. After 3 days, a hydrogel was formed, which is washed in boiling NaOH (0.1 M) aqueous solution for 2 h and then dialyzed in nanopure water for 12 h. The cleaned GO/BNC hydrogel was then cut into desired dimensions, typically 3×3 cm², and then freeze-dried for 12 h to obtain GO/ BNC aerogel.

Preparation of Pd/GO/BNC membrane: GO/BNC aerogel (3 × 3 cm²) was immersed in the PdCl₂ solution (2.5 wt% in 5 wt% of HCl) for 8h. The GO/BNC aerogel was transferred into a petridish and placed in an oven at 60 °C for 1h. The dried GO/BNC membrane was then immersed into NaBH₄ solution (80 mM) for 10 min to achieve *in situ* formation of Pd nanoparticles. The obtained Pd/GO/BNC membrane was dried in an oven at 60 °C for 1h.

Microstructure and chemical composition characterization: Scanning electron microscope (SEM) images were obtained using a FEI Nova 2300 Field Emission SEM at an acceleration voltage of 10 kV. Transmission electron microscopy (TEM) images were obtained using JEM-2100F (JEOL) field emission instrument. Raman spectra were obtained using a Renishaw inVia

confocal Raman spectrometer mounted on a Leica microscope with 50x objective and 785 nm wavelength diode laser as an illumination source. X-ray photoelectron spectroscopy (XPS) analysis was performed using Physical Electronics® 5000 VersaProbe II Scanning ESCA (XPS) Microprobe. The X-ray diffraction (XRD) measurements of the samples were obtained using a Bruker D8-Advance X-ray powder diffractometer using Cu K α radiation ($\lambda = 1.5406 \text{ \AA}$).

Thermalgravimetric analysis (TGA) to measure Pd loading in Pd/GO/BNC:

Thermogravimetric analysis (TGA) was performed in nitrogen using a TA Instruments Q5000 IR Thermogravimetric Analyzer at a heating rate of $5 \text{ }^\circ\text{C min}^{-1}$. GO/BNC, Pd/GO/BNC (with same loading of GO as GO/BNC membrane), Pd/BNC and pristine BNC were tested using above condition (appendix 3, Figure S3.2). For Pd/GO/BNC, the first mass loss ($\sim 10 \%$) can be attributed to absorbed water. The second mass loss ($\sim 13 \%$) at $\sim 160 \text{ }^\circ\text{C}$ is due to the decomposition of functional groups of GO. The third mass loss ($\sim 26 \%$) at $\sim 280 \text{ }^\circ\text{C}$ is due to the degradation of cellulose and a mass loss ($\sim 12.8 \%$) at $\sim 390 \text{ }^\circ\text{C}$ is due to the decomposition of cellulose residue, which generates CO_2 and H_2O and the sublimation or burning of the damaged graphitic regions.²⁰³⁻²⁰⁴ The remaining mass ($\sim 38 \%$) is composed of PdNPs and carbon residues. The mass of carbon residues for GO/BNC before the growth of Pd/GO/BNC is around 25.7% . Based on the above results, the mass loading of PdNPs in Pd/GO/BNC is calculated to be $12.3 \text{ wt}\%$.

Evaluation of catalytic activity of Pd/GO/BNC membrane: To investigate the catalytic activity of the Pd/GO/BNC membrane, the catalytic degradation of MO in the presence of NaBH_4 was employed as a model reaction. $40 \text{ }\mu\text{L}$ NaBH_4 solution (0.02 mg/L) was rapidly added into MO solution ($25.7 \text{ mg}\cdot\text{L}^{-1}$, 1.5 mL) under constant stirring. Subsequently, Pd/GO/BNC membrane ($1\times 0.5 \text{ cm}$) was placed in the above mixture. The concentration of reactant, MO (absorbance at 465 nm), was monitored by acquiring UV–vis absorption spectra using a Shimadzu UV-1800 UV–vis spectrometer. Pseudo-first-order kinetics was employed to evaluate the rate constants.

Thus, the rate of the degradation reaction of MO using the Pd/GO/BNC as catalyst can be calculated the following equation:²⁹⁶

$$\ln\left(\frac{C_0}{C_t}\right) = kt$$

Where, C is the concentration of the reactant, k is the reaction rate constant, t is the reaction time. In this reaction, the ratio of C_t to C_0 is calculated from the relative absorbance intensity of A_t/A_0 .

Evaluation of the performance of Pd/GO/BNC filtration membrane for wastewater treatment: The dye degradation performance was evaluated by filtering dye-contaminated water through the membrane via a benchtop filtration setup. Feed solutions containing different concentrations of MO (25.7 to 80 mg·L⁻¹, 10 mL) in the presence of NaBH₄ (140 mg·L⁻¹) at different pH (3, 5, 7, 9) were filtered through a bench-top filtration set-up under 0.8 bar (11.6 psi). The optical absorption of the solutions before and after the filtration was monitored using a Shimadzu UV-1800 spectrometer in the wavelength range of 300–700 nm. The dye-contaminated water treatment efficiency was calculated using the following equation:

$$\text{MO degradation efficiency (\%)} = \frac{C_0 - C_t}{C_0} \times 100 = \frac{A_0 - A_t}{A_0} \times 100$$

Where A_0 is the initial MO optical absorption and A_t is the MO optical absorption after filtration. A cocktail of contaminants was prepared by mixing 4-nitrophenol, rhodamine 6G, and methylene blue (50 mL, each with 10 mg·L⁻¹ in 140 mg·L⁻¹ NaBH₄). The cocktail was then filtered through the Pd/GO/BNC membrane. The optical absorption of the solutions before and after the filtration was monitored using a Shimadzu UV-1800 spectrometer in the wavelength range of 300–800 nm.

Flux and particle rejection tests: Water flux of Pd/GO/BNC membrane, commercial ultrafiltration membrane (YMGESP3001, GE, pore size ~ 1000 Da) and commercial nanofiltration membrane (YMDKSP3001, GE, pore size ~ 150-200 Da) was measured using a benchtop cross-

flow system, which includes a crossflow membrane cell (CF042D, Sterlitech Corp.) and a Hydracell pump (M03S, Wanner Engineering, Inc). Gold nanoparticles (AuNPs) with a diameter around 5 nm were synthesized using seed-mediated growth method.²⁷⁵ AuNPs solutions were then filtered by Pd/GO/BNC membranes using above cross-flow system under 58 psi. Before and after filtration, the optical extinction of filtration/permeate solutions were measured by UV-vis spectrometer (400 nm to 1000 nm). The rejection rate (RR) is calculated using equation below:

$$RR = \frac{E_f}{E_p} \times 100$$

Where E_f is the optical extinction of filtration solutions and E_p is the optical extinction of permeate solutions.

Stability tests: Freestanding GO membranes were prepared by vacuum-filtrating a GO suspension (0.4 wt%, 10 mL) onto a smooth PVDF membrane. After completely dried, the membranes were carefully peeled off. GO membranes and Pd/GO/BNC membranes were subjected to sonication, mechanical agitation and direct negative pressure without any support.

6.4 Results and Discussion

6.4.1 The fabrication of Pd/GO/BNC

The fabrication of the Pd/GO/BNC membranes involves the culture of *Gluconacetobacter hansenii* under static growth conditions with graphene oxide (GO) flakes incorporated into the growth medium. GO flakes are dispersed in bacteria growth medium at a desired concentration to achieve a GO/BNC hydrogel with a thickness ~2 mm (Figure 6.1). The presence of epoxy and hydroxyl groups on the basal plane of GO facilitates the uniform dispersion of GO flakes in the growth solution without aggregation or restacking into multilayer structures (Figure 6.1A). The uniform dispersion in turn enables the incorporation and intercalation of GO flakes within BNC matrix in

a “layer-by-layer” manner without any covalent crosslinking. The as grown hydrogel was cleaned in boiling alkaline water (pH 13) and freeze-dried to obtain GO/BNC aerogel. The resulting GO/BNC aerogels exhibit an open porous structure and large specific surface area (see appendix 5, Figure S5.1A). Subsequently, PdNPs were *in situ* grown within the matrix to obtain Pd/GO/BNC membrane (see supporting information for details). After washing and air-drying, a Pd/GO/BNC membrane was obtained (Figure 6.1B).

6.42 Micro and Nanostructure and Chemical Composition of Pd/GO/BNC Membrane

Pristine BNC film, prepared without GO flakes in the growth solution, is white and translucent as shown in Figure 6.2A1. Scanning electron microscopy (SEM) image of the top surface of pristine BNC membrane reveals the entangled network of nanocellulose fibers of 20–100 nm in diameter (Figure 6.2A2). Considering that bacteria form the densest cellulose fiber network at the air/liquid interface, the cellulose nanofibrils are preferentially oriented parallel to the surface (*i.e.* normal to the thickness) of the film, which results in denser physical entanglements of the cellulose nanofibrils parallel to the surface compared to that along the thickness. Cross-section image of the BNC film reveals such layered structure of nanocellulose fibers (Figure 6.2A3). Addition of GO flakes to the growth solution results in the formation of a GO/BNC composite hydrogel (see appendix 5, Figure S5.1A). GO/BNC membrane formed by air-drying GO/BNC hydrogel has smoother surface (less fibrillar) structure compared to pristine BNC membrane due to the presence of 2D GO sheets (see appendix 5, Figure S5.1B). Pd/GO/BNC membrane is dark and opaque due to the presence of GO flakes and PdNPs (Figure 6.2B1). As mentioned above, PdNPs were *in situ* grown in the GO/BNC composite by exposing the GO/BNC aerogel to Pd precursor solution. The highly porous nature of GO/BNC allows large uptake of palladium precursor over the entire matrix, resulting in a large loading of PdNPs in the final membrane (12.3 % from thermogravimetric

analysis, see experimental section and appendix 5, Figure S5.2 for details). Similar to GO/BNC, the surface of the Pd/GO/BNC membrane shows smoother structure compared to the bare BNC film because of the presence of GO flakes and dense, uniform distribution of PdNPs (Figure 6.2B2). The cross-sectional SEM image of the Pd/GO/BNC film reveals a laminated structure with a thickness of $\sim 7 \mu\text{m}$, which stems from the packing of GO flakes within the BNC fibers (Figure 6.2B3).

Transmission electron microscopy (TEM) image of Pd/GO/BNC membrane reveals that the *in situ* growth method employed here results in a uniform distribution of the Pd nanoparticles on the GO sheets (Figure 6.2C). The *in situ* grown PdNPs in GO/BNC matrix appears to be coalesced instead of individual nanoparticles owing to the absence of a surfactant or capping agent during the formation of PdNPs. Furthermore, a high density of nucleation sites on GO surface can also result in the formation of a branched network of PdNPs.²⁹⁷ The size of the PdNPs (estimated from the width of the PdNPs chains) was found to be $7.3 \pm 2.5 \text{ nm}$. High resolution TEM images of the PdNPs on GO/BNC show lattice fringes with a lattice spacing of 0.23 nm , corresponding to (111) planes of Pd (Figure 6.2D).²⁹⁸ Energy-dispersive X-ray spectroscopy (EDS) analysis in TEM further confirm the presence of Pd in the membrane (Figure 6.2E). The strong signal corresponding to Cu originates from the TEM grid made of copper. The presence of Pd nanoparticles in the membrane was also verified by XRD measurements. XRD pattern exhibited three peaks at 40.1 , 46.5 , and 68.1° corresponding to the (111), (200), and (220) planes of Pd, respectively (see appendix 5, Figure S5.3A).²⁹⁹

To further understand the chemical composition of Pd/GO/BNC membrane, we have performed XPS measurements. The binding energy of Pd $3d_{5/2}$ and Pd $3d_{3/2}$ obtained from the Pd/GO/BNC at 335.1 and 340.4 eV can be assigned to Pd(0), indicating the metallic form of Pd (PdNPs). The

peaks at 336.8 and 342.1 eV correspond to the oxidized form of Pd (Pd^{2+}).³⁰⁰⁻³⁰² It is calculated that the Pd(0): Pd(II) ratio is around 9:5, indicating the major species present on GO are Pd(0).³⁰³ High resolution spectrum of C1s region was deconvoluted into three peaks corresponding to sp^2 domains (C=C with a binding energy of 284.6 eV) and oxidized sp^3 domains (C–O with a binding energy of 286.6 eV and C=O with a binding energy of 288.2 eV) (see appendix 5, Figure S5.4A, B).²¹² The C/O ratio obtained from the area under the peaks can be used to evaluate the degree of reduction of GO. For GO/BNC, the C/O ratio was found to be 0.23 and it was found to increase to 0.98 after the growth of PdNPs, indicating the partial reduction of GO by NaBH_4 during the growth of PdNPs on the GO/BNC. The Raman scattering spectrum of both the GO/BNC membrane and the Pd/GO/BNC membrane exhibited Raman bands at 1590-1600 cm^{-1} and 1320-1330 cm^{-1} , which correspond to the G band and D band of GO, respectively (see appendix 5, Figure S5.4C).³⁰⁴ The ratio of the intensity of D/G bands was found to increase from 0.39 to 0.45, also indicating the reduction of GO.³⁰⁵

6.43 Dye Degradation Activity of Pd//GO BNC Membrane

To evaluate the catalytic activity of Pd/GO/BNC membrane, we selected the degradation of MO in the presence of NaBH_4 as a model catalytic reaction of organic dye molecules. UV-vis spectroscopy was employed to monitor the reaction kinetics in 20 sec intervals. A strong absorption peak at 465 nm can be ascribed to the characteristic absorbance of MO and no visible color change can be observed even after the addition of NaBH_4 solution, suggesting that the degradation reaction of the MO requires the presence of a catalyst. Once the Pd/GO/BNC membrane was immersed into the solution, the intensity of the absorption peak at 465 nm decreased rapidly and disappeared completely after 5 min, suggesting the complete degradation of MO (Figure 6.3A). The degradation is also evident from the color change of the solution, which

turned from orange to colorless (inset of Figure 6.3A). For comparison, BNC and GO/BNC membranes without PdNPs were immersed into the MO solution in the presence of NaBH₄. The intensity of absorption peak at 465 nm remained virtually unchanged within 5 min, suggesting that the PdNPs are primarily responsible for the catalytic activity of Pd/GO/BNC and MO degradation in the presence of NaBH₄ (Figure 6.3B).

To demonstrate the advantage of utilizing GO as a template for the *in situ* growth of PdNPs, a Pd/BNC membrane was prepared via *in situ* formation of PdNPs on BNC fibers and its catalytic activity was investigated along with Pd/GO/BNC under identical conditions. The rate of MO degradation with Pd/BNC membrane was found to be lower compared to Pd/GO/BNC (Figure 6.3B). Langmuir–Hinshelwood apparent first-order kinetics was employed to determine the reaction rate constants of MO degradation.³⁰⁶ From the first-order kinetics results, the rate constant with Pd/GO/BNC membrane is found to be 0.0152 s⁻¹, which is higher than that with Pd/BNC membrane (0.0111 s⁻¹) (Figure 6.3C).

The recovery of catalysts is critical for recycling the catalyst, which can be an important consideration for dye removal technologies in industrial settings. Both Pd/GO/BNC and Pd/BNC could be easily withdrawn from the reaction mixture and washed with distilled water/ethanol for the next reaction cycle. The Pd/GO/BNC retained 94.7% of its catalytic activity even after ten cycles, which indicates the excellent catalytic stability of the Pd/GO/BNC membrane. In stark contrast, the MO degradation efficiency of Pd/BNC catalyst dropped to ~67.7% after ten cycles, due possibly to desorption and leaking of PdNPs from the BNC matrix during multiple cycles of extensive washing (Figure 6.3D).

Based on the results, Pd/GO/BNC membrane exhibits superior catalytic efficiency and stability compared to Pd/BNC membrane. We believe that remarkable properties of Pd/GO/BNC largely

stem from the following factors: (i) the presence of GO increases the surface area for efficient *in situ* growth and anchoring of PdNPs, leading to higher loading of PdNPs; (ii) the distribution of PdNPs on GO flakes is more uniform compared to that on pristine BNC fibers, which exhibited severe aggregation (see appendix 5, Figure S5.5, S5.6); and (iii) strong interactions between *in situ* formed PdNPs and GO due to abundant chemical functionality on GO flakes and tight packing of GO flakes, reducing the loss of PdNPs during multiple reaction cycles (see appendix 5, Figure S5.7).

6.44 Pd/GO/BNC as a Filtration Membrane for Organic-contaminated Water Treatment

Next, we set out to investigate the applicability of Pd/GO/BNC as a filtration membrane for a dye-contaminated water treatment. The performance was evaluated by filtering dye-contaminated water through the membrane via a benchtop filtration setup (Figure 6.4B). It can be seen that the orange-colored MO solution became completely colorless after passing through the membrane and the filtration only took a few minutes. Noticeably, the Pd/GO/BNC membrane exhibited outstanding MO degradation performance over a wide range of MO concentrations (Figure 6.4C). The fact that the Pd/GO/BNC membrane resulted in 99.3% dye degradation efficiency for all MO concentrations up to $60 \text{ mg}\cdot\text{L}^{-1}$ (which is close to the lethal dose to animals, $60 \text{ mg}\cdot\text{L}^{-1}$) is remarkable.³⁰⁷ Even at a concentration of $85 \text{ mg}\cdot\text{L}^{-1}$, the degradation efficiency dropped only slightly (to 90.5%) indicating the excellent catalytic efficiency of the membrane. The dye degradation of Pd/GO/BNC was found to be insensitive to pH variation of the feed water (Figure 6.4D). The degradation efficiency remained very high (99.3%) across a broad range of environmentally-relevant pH (pH 3 to 9), which occur in many natural and engineered aqueous systems.²⁷² The Pd/GO/BNC membrane also exhibited a good cycling stability and its performance decreased only slightly (to 99.2%) after six cycles of filtration (Figure 6.4E). In

contrast, GO/BNC and bare BNC membrane without PdNPs exhibited negligible dye degradation efficiency, confirming that the dye degradation of Pd/GO/BNC stems from the catalytic efficiency of Pd rather than the adsorption or rejection of MO (see appendix 5, Figure S5.8).

Besides MO, an anionic dye, we have also evaluated the performance of Pd/GO/BNC membrane for the treatment of cationic dye and organic contaminant. A cocktail (50 mL) of contaminants (4-nitrophenol, a toxic organic compound; rhodamine 6G and methylene blue, cationic dyes with non-overlapping characteristic absorbance band, each with a concentration of $10 \text{ mg}\cdot\text{L}^{-1}$) was filtered through the Pd/GO/BNC membrane (Figure 6.4F). It can be seen that all the contaminants can be effectively treated (4-nitrophenol can be reduced to 4-aminophenol, leading to a new absorbance peak around 300 nm) simultaneously (Figure 6.4G).

6.45 Particle Rejection and Water Flux Tests

To further demonstrate the potential of Pd/GO/BNC membrane for large-scale water treatment, we tested the water flux and particle rejection capability of the Pd/GO/BNC membrane using a benchtop cross-flow system (schematic setup diagram in Figure 6.5A). We examined the particle rejection capability using gold nanoparticles (AuNPs) with diameters of $5.15 \pm 0.4 \text{ nm}$, which were synthesized by a seed-mediated method.²⁷⁵ The as-synthesized AuNPs exhibited very narrow size distribution, which is critical for accurate study of particle rejection (Figure 6.5B). The particle rejection rates were determined by measuring the UV-vis extinction spectra of feed/permeate solutions containing AuNPs (Figure 6.5D). The Pd/GO/BNC membrane showed $\sim 100\%$ rejection for particles with a diameter of 5 nm, indicating that the pore size of the Pd/GO/BNC is below 5 nm (inset of Figure 6.5D). Considering that Pd/GO/BNC membrane showed no rejection of MO (with molecular size of $\sim 1.2 \text{ nm}$) in the absence of NaBH_4 during filtration tests, the Pd/GO/BNC membrane falls into the range of ultrafiltration membrane (see appendix 5, Figure S5.9).^{279,308} The

nanochannels within GO-based membrane, which determine the mass transport properties can be altered by introducing exogenous spacers of desired dimensions.^{45, 48} In this work, the presence of BNC fibers and PdNPs between GO flakes lead to an overall tortuous network of pores throughout the membrane even though no visible pores can be observed on the surface of the membrane. The Pd/GO/BNC membrane showed stable water flux (33.1 L/m²h) over a 6-hour flux test after stabilization under positive pressure of 58 psi (4 bar), which is 2.3 and 2.8 times higher than the flux performances of commercial ultrafiltration color reduction membrane (YMGESP3001, pore size ~ 1000 Da) with MO rejection rate of 95.2% (14.5 L/m²h under 58 psi) commercial nanofiltration membrane (YMDKSP3001, pore size ~ 150-200 Da) with MO rejection rate of 100% (11.8 L/m²h under 100 psi) (Figure 6.5E and Figure S5.10). The above results demonstrate that Pd/GO/BNC membrane holds potential for large-scale ultrafiltration process.

It is remarkable that the Pd/GO/BNC membrane can withstand the operation pressure as high as 58 psi without any supporting membrane while most of the GO-based membranes require a porous support membrane and/or carefully designed setups because of their limited aqueous stability and poor mechanical stability under pressure.²⁸⁰⁻²⁸³ We performed additional stability tests to demonstrate the advantage of the *in situ* fabricated Pd/GO/BNC membrane against pure GO membrane (see details in supporting information). Pd/GO/BNC and GO membranes have been subjected to sonication, mechanical agitation and direct negative pressure without any support. Pd/GO/BNC membrane exhibited superior robustness under above testing conditions while GO membrane disintegrated or broke inevitably (see appendix 5, Figure S5.11).

6.5 Conclusions

In conclusion, we showed a novel membrane based on BNC loaded with GO and PdNPs for highly efficient wastewater treatment. Three main factors can enable the Pd/GO/BNC membrane to be

effective in dye-degradation: i) the excellent catalytic ability of PdNPs with ultrafine size; ii) uniform and high loading of PdNPs within the entire membrane; and iii) the lamellar structure of the membrane allowing mass transport through the nanocapillaries between GO flakes, greatly increasing the path length of organic contaminants, thus maximizing the interaction with the catalytic PdNPs and the probability of degradation. The bacteria-mediated growth of the BNC membrane, *in situ* incorporation of the GO flakes and *in situ* synthesis of the PdNPs makes the whole process simple and highly scalable. The excellent dye degradation performance (degradation of methylene orange (MO) up to 99.3% even at a concentration of 60 mg/L) as well as the ability for the treatment of multiple contaminants simultaneously, stable flux (33.1 L/m²h under 58 psi), and the scalable fabrication of the membrane collectively make the approach suggested here highly attractive for wastewater treatment even at industrial scales. The novel membrane design suggested here can be easily adapted to realize other GO/BNC-based functional membranes with potential applications in catalysis, separations, energy storage, and environmental remediation.

6.6 Supporting Information

Supporting Information for chapter 6 is provided in appendix 5.

6.7 Figures

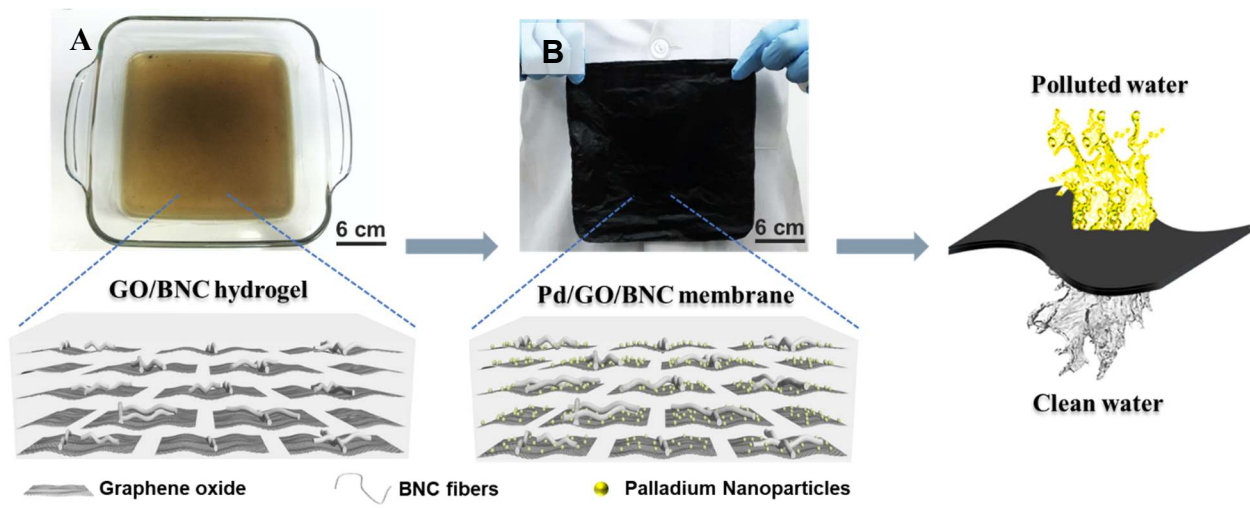


Figure 6.1. Schematic illustration and photographs showing the various steps including (A) GO/BNC hydrogel (B) Pd/GO/BNC membrane involved in the fabrication of Pd/GO/BNC membrane.

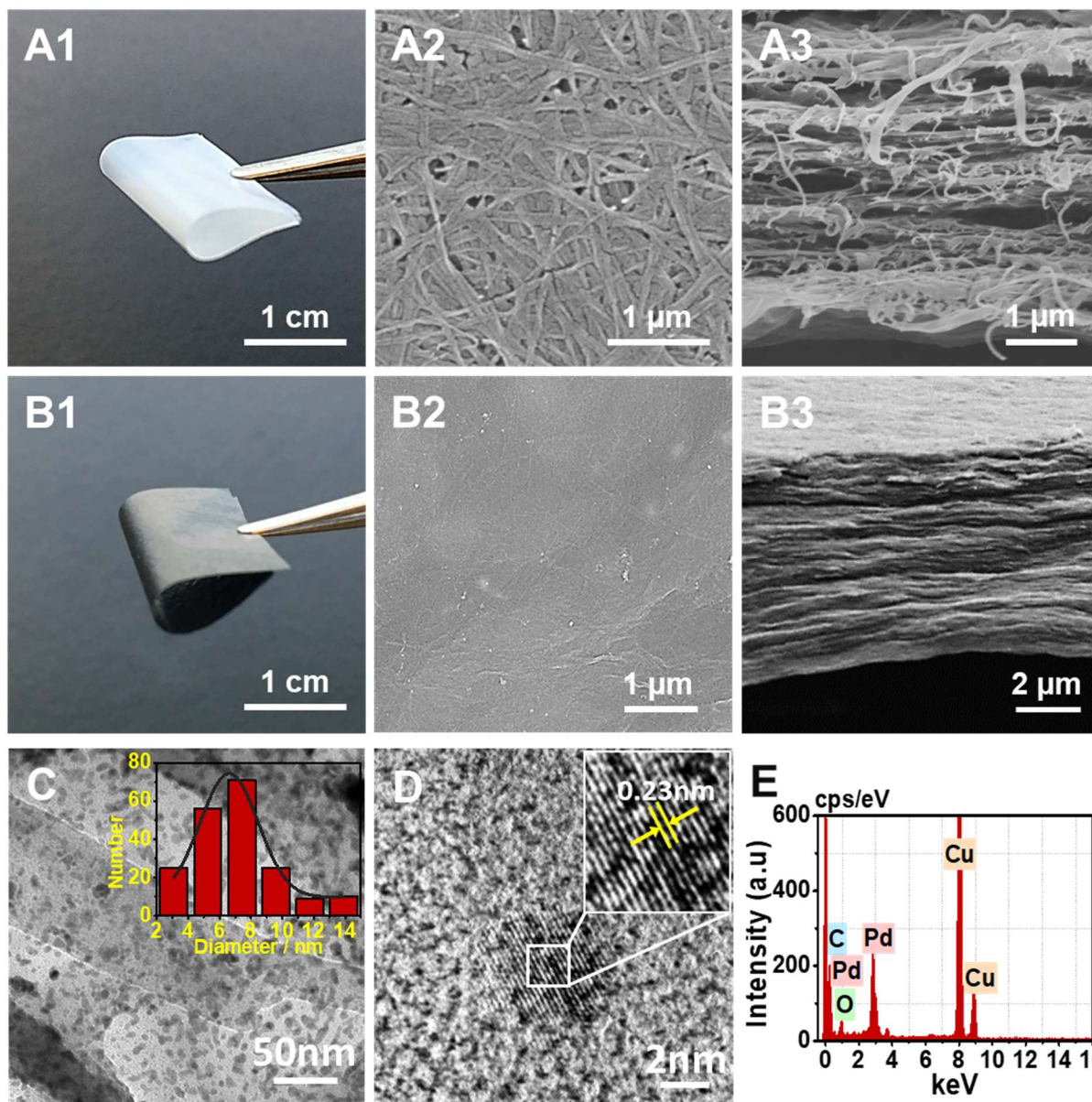


Figure 6.2. Micro and nanostructure and chemical composition of Pd/GO/BNC membrane. (A) Pristine BNC membrane: Photograph (A1) and SEM images of top surface (A2) and cross-section (A3). (B) Pd/GO/BNC membrane: Photograph (B1), SEM image of top surface (B2), and SEM images of cross-section (B3). (C) TEM image of Pd/GO/BNC membrane and the inset show the size distribution of Pd NPs. (D) HRTEM image of a single Pd NP on the GO/BNC membrane. (E) EDS of the Pd/GO/BNC membrane (strong Cu peaks correspond to the TEM grid made of Cu).

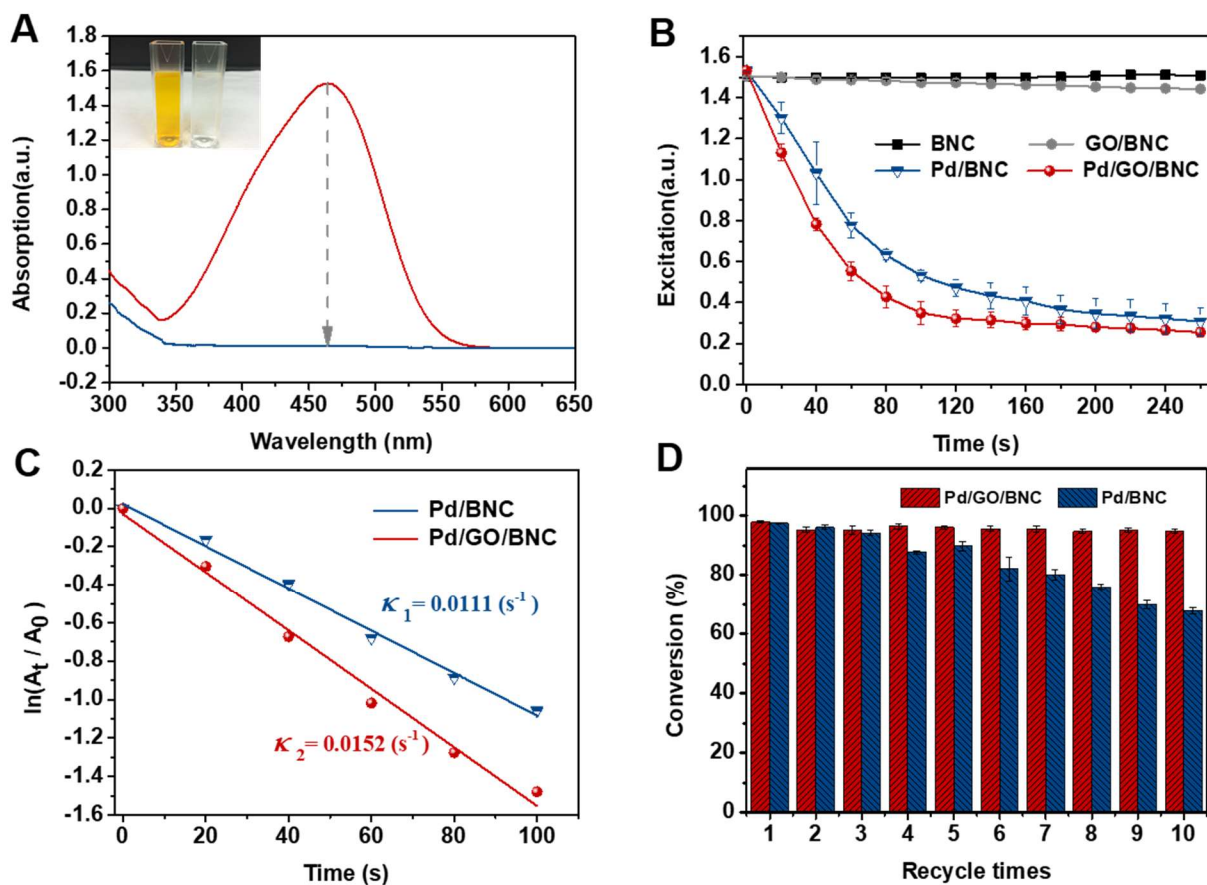


Figure 6.3. Dye degradation activity of Pd/GO/BNC membrane. (A) UV-vis spectra showing the degradation of MO in the presence of NaBH₄ and Pd/GO/BNC membrane. (B) Plot showing the degradation of MO in the presence of NaBH₄ over time. (C) Langmuir-Hinshelwood apparent rate constant for MO degradation by Pd/BNC and Pd/GO/BNC as catalysts. (D) The degradation performance of MO solution (25.7 mg L⁻¹) with NaBH₄ over 10 cycles for the Pd/GO/BNC membrane and Pd/BNC membrane catalysts.

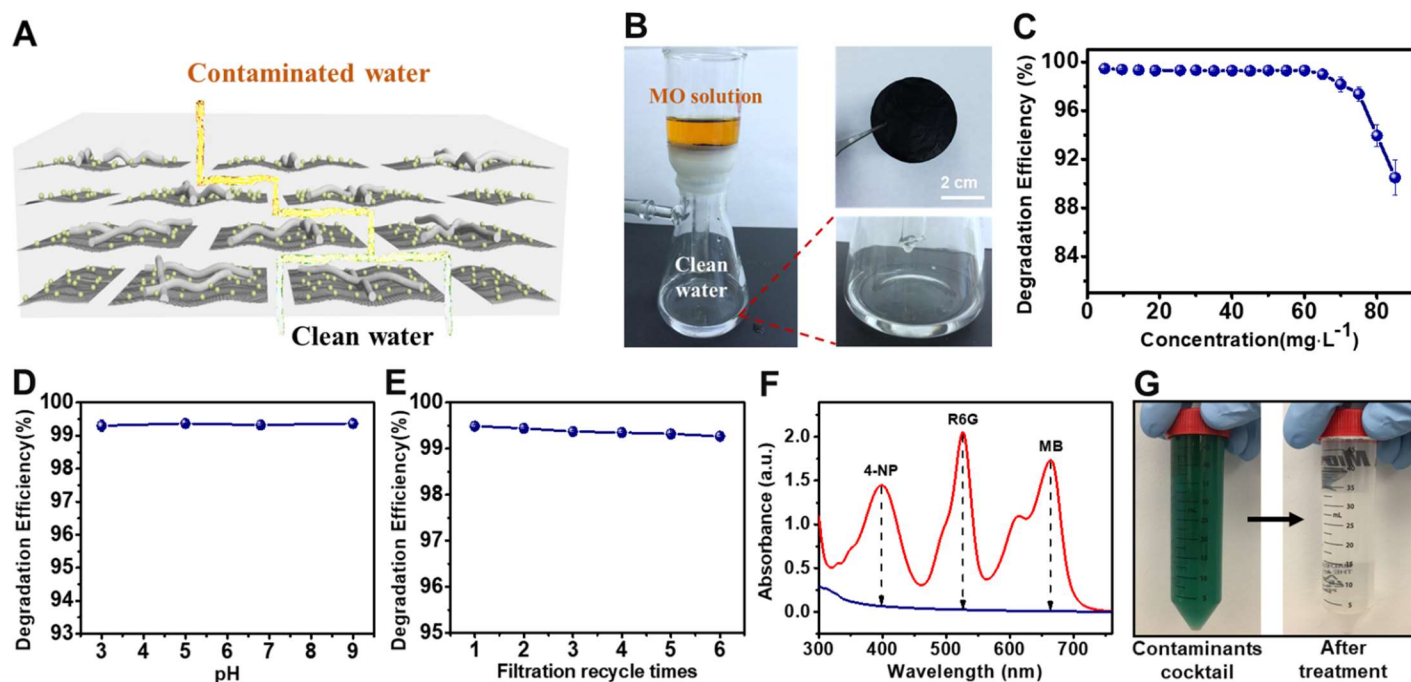


Figure 6.4. Pd/GO/BNC as a filtration membrane for organic-contaminated water treatment. (A) Schematic view to illustrating the filtration mechanism of the Pd/GO/BNC film. (B) Photograph of the bench-top filtration setup using Pd/GO/BNC film as a membrane. The yellow colored feed solution contains MO and NaBH₄. Dye degradation efficiency of Pd/GO/BNC film at different (C) MO concentrations and (D) different pH values. (E) Dye degradation efficiency of Pd/GO/BNC film over multiple cycles of reuse showing recyclability. (F) UV-vis spectra showing the degradation of a cocktail (50 mL) of organic contaminants: 4-nitrophenol (4-NP), rhodamine 6G (R6G) and Methylene blue (MB) (each with a concentration of 10 mg·L⁻¹) in the presence of NaBH₄ (140 mg·L⁻¹). (G) Images showing the contaminants cocktail before and after filtration treatment.

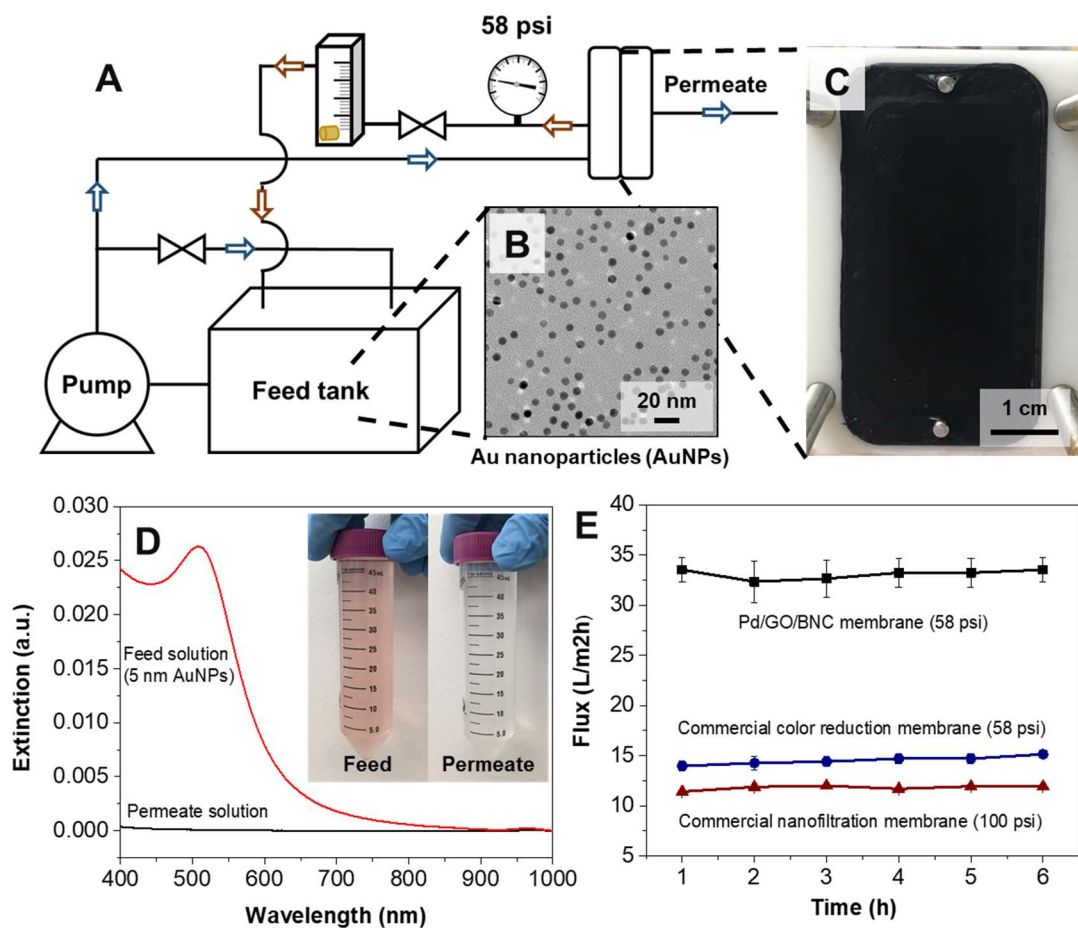


Figure 6.5. Particle rejection and water flux tests. (A) Schematic diagram of cross-flow flux test setup. (B) TEM images of gold nanoparticles with a diameter of 5 nm. (C) Image showing Pd/GO/BNC membrane placed in the cross-flow cell. (D) UV-vis extinction spectra indicating the rejection of AuNPs filtered through Pd/RGO/BNC membranes (inset is the picture showing feed and permeate solutions.) (E) Water flux of Pd/GO/BNC membrane, commercial color reduction membrane (58 psi was applied for the flux tests) and commercial nanofiltration membrane (100 psi was applied).

Chapter 7: An *In situ* Grown Bacterial Nanocellulose/Graphene Oxide Composite for Flexible Supercapacitor

7.1 Abstract

Recently, the development of flexible supercapacitors has received significant attention due to their application in flexible electronics such as bendable mobile phones, flexible displays and wearable devices. Owing to numerous advantages such as excellent mechanical strength, low cost, high porosity and natural abundance, bacterial nanocellulose (BNC) is considered to be highly attractive for the fabrication of flexible supercapacitors. This work demonstrates that BNC can serve as an ideal layered matrix for incorporation of active two-dimensional (2D) materials. A novel strategy for the incorporation of graphene oxide (GO) sheets into layered BNC during its growth is presented. GO flakes can be interlocked within nanocellulose network during BNC growth, enabling facile chemical reduction of GO sheets, which prevents their restacking and loss of active area, and leads to excellent energy storage performance as well as mechanical flexibility. Significantly, the fabrication approach demonstrated here can be extended to other 2D nanomaterials to realize flexible BNC-based energy storage devices.

7.2 Introduction

With the rapid development of portable and flexible electronics such as bendable mobile phones, flexible displays and wearable devices, there is a great need for high performance, flexible energy-storage devices.¹⁰³⁻¹⁰⁹ Supercapacitors (SCs) have emerged as an important class of energy storage

devices due to their high power density, long cycle-lifetime, fast charge-discharge characteristics and low environmental impact.¹¹⁰⁻¹¹² Owing to various advantages such as flexibility, light-weight, reliability, and safe operation over a wide temperature range, extensive efforts have been devoted to developing flexible all solid-state SCs, which hold great potential for next generation of flexible energy storage devices.^{113-119, 309-311}

Carbon based materials, such as activated carbon, carbon nanotubes, graphene and its derivative reduced graphene oxide, are widely used as the active electrode material for flexible supercapacitors due to their high electrical conductivity, large surface area, chemical stability, which leads to long cycle life-time, and natural abundance.¹²⁰⁻¹³¹ Carbon materials are commonly combined with pseudocapacitive materials including transitional metal oxides (*e.g.*, MnO₂, CoO, NiO) and conducting polymers, such as polyaniline (PANI), poly(3,4-ethylenedioxythiophene) (PEDOT), polypyrrole (PPy), for preparing flexible supercapacitors, which store charges through fast and reversible redox (Faradaic) reactions, leading to higher capacitance and energy density.^{132-140, 312-313}

In order to obtain flexibility while maintaining mechanical strength, soft and bendable plastics such as polydimethylsiloxane (PDMS) and polyethylene terephthalate (PET) are often used as substrates or packaging materials, however these plastics introduce considerable cost and environmental burden after disposal.¹⁴¹ Cellulose-based substrates, on the other hand, are a low cost, highly flexible, renewable, biodegradable option. Paper substrates possess fibrous structure that facilitates interactions with active materials and provides pathways for ion transport, which make them ideal for use as substrates for flexible supercapacitors.¹⁴²⁻¹⁴⁶ Recently, Hu and co-workers have reported an all-wood supercapacitor with excellent performance and almost all components coming from the renewable and biodegradable materials, further signifying the

advantages of cellulose-based supercapacitors.³¹⁴ Among all cellulose materials, bacterial nanocellulose (BNC), which is a highly pure form of cellulose produced from dextrose through a series of biochemical steps followed by the self-assembly of secreted cellulose fibrils from bacteria in the culture medium, shows great promise for the fabrication of functional composites through *in situ* growth or adsorption of pre-synthesized nanostructures on the nanoscale cellulose fibers due to its excellent mechanical properties, tunable porosity, chemical functionalizability, ease of synthesis, high scalability.^{11, 13, 19, 147-148} Due to these advantages, BNC has been adapted for fabricating BNC-based flexible supercapacitors.¹⁴⁹⁻¹⁵³ However, in order to make high performance BNC-based supercapacitors, the fabrication techniques usually involve vacuum filtration^{152, 154}, pyrolysis activation^{150, 155-156}, which either have prospects in terms of scalability or compromise the mechanical properties of BNC. Thus, there is an immediate need for facile and scalable methods for the fabrication of BNC-based electrode with high energy storage performance. In this work, we introduce a novel strategy that involves the *in situ* incorporation of graphene oxide (GO) flakes and poly(3,4-ethylenedioxythiophene)-poly(styrenesulfonate) (PEDOT:PSS) into BNC matrix during its growth for the fabrication of flexible, light-weight BNC-based electrodes, which exhibit excellent electrochemical performance (373 F/g at 1 A/g) and cycling stability (~85% capacitance retention over 1000 cycles). Using a BNC-based separator, RGO/PEDOT:PSS/BNC electrodes are assembled to obtain all solid-state supercapacitors, which exhibited excellent energy storage performance, flexibility and robustness. The facile and scalable fabrication method combined with the low cost of the materials, makes the approach suggested here highly attractive for all solid-state supercapacitors.

7.3 Experimental Section

Materials: Graphite flakes, PEDOT:PSS solution (Orgacon™ HIL-1005, 1 wt% in water, high conductivity grade), hypophosphorous acid (HPA) and all chemicals for bacterial medium are purchased from Sigma-Aldrich. *Gluconacetobacter hansenii* was purchased from ATCC (ATCC® 53582).

Preparation of RGO/PEDOT:PSS/BNC electrodes: *Gluconacetobacter hansenii* (ATCC®53582) was cultured in test tubes containing 16 ml of #1765 medium at 30°C under shaking at 250 rpm. The #1765 medium is composed of 2% (w/v) glucose, 0.5% (w/v) yeast extract, 0.5% (w/v) peptone, 0.27% (w/v) disodium phosphate, and 0.5% (w/v) citric acid. Graphene oxide was synthesized using an improved method reported by Tour and coworkers. Graphene oxide solution (28 mL of 0.1wt%) was centrifuged and redispersed in #1765 medium and then centrifuged again to leave a wet mixture of GO and medium after decanting supernatant. Bacterial culture solution (incubated 3 days) and 2 ml of 1 wt% PEDOT:PSS (Sigma Aldrich) was added to the GO/medium wet mixture to make it to a total 8ml (with GO concentration of 0.35 wt%, PEDOT:PSS concentration of 0.25 wt%). The solution was subsequently transferred to petridish (diameter: 6 cm) and incubated at room temperature without disturbance for 5 days to obtain a GO/PEDOT:PSS/BNC semi-dry film. For purification, the film was harvested from the petri dish and washed in a 500 ml of 0.1 M NaOH aqueous solution under boiling condition for 2 h. The GO/PEDOT:PSS/BNC hydrogel was then dialyzed in nanopure water for 2 days. The as cleaned hydrogel was then immersed in 5 wt% HPA and then heated to 60 °C for 24 hours, dialyzed in nanopure water and dried under ambient conditions. To show the scalability of the fabrication technique, a large electrode was prepared in a Pyrex bakeware (18×18 cm) in the same manner mentioned above.

Preparation of flexible all-solid state supercapacitors: A PVA/H₂SO₄ gel electrolyte was prepared by mixing 3 g of PVA powder, 3 g of H₂SO₄, and 30 mL nanopure water together, and then heated to 85 °C under vigorous stirring until the solution became clear. After the solution cooled down to room temperature, two pieces of RGO/PEDOT:PSS/BNC electrodes and a BNC paper as separator were immersed into the PVA/H₂SO₄ gel for 10 mins and they were assembled into a flexible all- solid state device. The device was dried overnight in fume hood to remove excess water.

Microstructure characterization and properties measurement: Scanning electron microscope (SEM) images were obtained using a FEI Nova 2300 Field Emission SEM at accelerating voltage of 10kV. Atomic force microscopy (AFM) images were obtained using Dimension 3000 (Bruker Inc.) in light tapping mode. Raman spectra were obtained using a Renisha inVia confocal Raman spectrometer mounted on a Leica microscope with 50x objective and 785 nm wavelength diode laser as an illumination source. XPS analysis was performed using Physical Electronics® 5000 VersaProbe II Scanning ESCA (XPS) Microprobe.

Electrochemical characterizations: Cyclic voltammetry (CV) and galvanostatic charge/discharge studies analysis were carried out using CHI 760 bipotentiostat (CH Instruments, Austin, USA). Chronopotentiometry technique used to run the charge/discharge measurements in the potential range from 0 to 0.8 V. Electrochemical studies were carried out using three electrode and two electrode configurations (solid-state device). In three electrode setup, RGO/PEDOT:PSS/BNC composite served as working electrode, Ag/AgCl as reference, and platinum wire as counter electrode. The electrolyte solution used here is 1 M H₂SO₄.

Preparation of PEDOT:PSS/BNC, GO/BNC and rGO/BNC films: PEDOT:PSS/BNC film was prepared by adding 2ml of 1 wt% PSS:PEDOT solution to 6ml BNC culture solution (overall

8ml, the concentration of PSS:PEDOT in solution is 0.25 wt%. After 5 days, the hydrogel was harvested and cleaned in the manner discussed above. GO/BNC film was prepared by adding a mixture of BNC culture solution and graphene oxide solution (overall 8ml), the concentration of GO in solution is 0.3 wt%. After 5 days, the hydrogel was harvested and cleaned in the manner discussed above. RGO/BNC film was prepared by adding a mixture of BNC culture solution and graphene oxide solution (overall 8ml, the concentration of GO in solution is 0.3 wt%. After 5 days, the hydrogel was harvested, cleaned, dried and reduced in the manner mentioned above.

Calculation details: The mass specific capacitances ($C_{s, CV}$) of the electrodes and device are all calculated from CV profile by the equation below:

$$C_{s, CV} = \frac{I_a - I_c}{m * (dV/dt)}$$

where I_a , I_c , m , and dv/dt are the anodic current, cathodic current, electrode mass, and scan rate, respectfully.

The mass specific capacitances ($C_{s, C-D}$) of the electrodes and device are all calculated from GCD profiles by the equation below:

$$C_{s, C-D} = \frac{I \Delta t}{m \Delta V}$$

Where I , Δt , m , ΔV are the applied current, the elapsed time, the mass of the electrode, and the voltage range, respectfully.

7.4 Results and Discussion

7.41 The Fabrication of RGO/PEDOT:PSS/BNC Electrode

The fabrication of the RGO/PEDOT:PSS/BNC electrode is achieved by culturing *Gluconacetobacter hansenii* bacteria in the presence of GO flakes and PEDOT:PSS under aerobic

and static growth conditions (Figure 7.1A). GO flakes were synthesized using a method reported by Tour and co-workers.²⁰² It is known that the thickness of a monolayer of GO is ~ 0.72 nm and the higher thickness of GO compared to a monolayer of graphene (0.34 nm) is ascribed to the presence of epoxy and hydroxyl groups on the basal plane.²⁰⁶ Atomic force microscope (AFM) images revealed the average thickness of GO flakes deposited on a silicon substrate to be $\sim 1 \pm 0.2$ nm, which corresponds to a bilayer of GO (appendix 6, Figure S6.1B). GO flakes were washed and dispersed in the broth solution with PEDOT:PSS and bacteria at a predetermined concentration to achieve a desired BNC growth rate (Figure 7.1B and see experimental section for details). The mixture is homogenized and set aside under static condition for 5 days to obtain a GO/PEDOT:PSS/BNC semi-dry film with desired thickness and uniformly embedded with GO flakes and PEDOT:PSS. The GO/PEDOT:PSS/BNC semi-dry film was then washed using boiling NaOH solution (0.1 M) with mechanical agitation and then nanopure water to remove bacteria and residual growth medium. The as cleaned film was then treated by 6% hypophosphorous acid (HPA) at 85 °C overnight to reduce GO flakes and washed in nanopure water for 2 days and then air-dried to obtain a RGO/PEDOT:PSS/BNC film with thickness of ~ 8 μm and active materials loading of 0.54 mg/cm^2 and overall density of 0.64 mg/cm^2 (Figure 7.1C). Note that during the rigorous aqueous processing mentioned above, the film remained intact and the mechanical robustness can be attributed to the mechanical interlocking of the loaded RGO flakes and PEDOT:PSS within the nanofiber network during the BNC growth. The entangled network of highly crystalline BNC nanofibers and imbedded active materials resulted in the excellent tensile strength of the electrode ($\sigma_s = \sim 240$ MPa, see appendix 6, Figure S6.6)

7.42 Characterizations and Optimization of The RGO/PEDOT:PSS/BNC Electrode

Pristine BNC film (produced in the absence of GO flakes and PEDOT:PSS in the broth) is white and translucent and shows excellent flexibility and mechanical strength (Figure 7.2A).²⁰¹ The bacteria first form an entangled layer of cellulose fibers (20-100 nm in diameter) at the air/medium interface (Figure 7.2A1). As the oxygen diffuses deeper into the medium, the subsequent BNC layers are formed below the surface layer, which stack together to form a 3D BNC network.^{3, 20} This “layer-by-layer” formation leads to network of cellulose nanofibrils that are preferentially oriented parallel to the surface (*i.e.* normal to the thickness) of the film. The cellulose nanofibril network exhibits denser physical entanglements parallel to the surface compared to that along the thickness (Figure 7.2A2). Addition of GO flakes to the bacterial broth followed by base wash and HPA treatment led to the formation of RGO/BNC film which appeared light grey in color and exhibited a metallic luster, indicating the reduction of GO (Figure 7.2B).²⁰⁸ The surface of the RGO/BNC film is smoother and less fibrillar compared to the pristine BNC film due to the incorporation of GO flakes (Figure 7.2B1). The “layered” formation of BNC starting from the liquid/air interface facilitated the layered arrangement of RGO flakes between the BNC layers as evidenced by the cross-sectional SEM images of the RGO/BNC film (Figure 7.2B2). Addition of PEDOT:PSS in GO/BNC broth and subsequent treatment resulted in a dark blue RGO/PEDOT:PSS/BNC film (Figure 7.2C). Compared to RGO/BNC film, the surface of RGO/PEDOT:PSS/BNC film exhibited granular surface morphology, possibly due to the formation of polymer aggregates.³¹⁵ The PEDOT:PSS fills the interstices within the RGO/BNC layered structure as evidenced by the cross-sectional SEM images of the RGO/PEDOT:PSS/BNC film (Figure 7.2C2). Raman scattering spectrum of the pristine GO showed the characteristic graphitic band (G-band) at 1580–1600 cm^{-1} and defect band (D-band) at 1330–1340 cm^{-1} .²⁰⁷

Pristine PEDOT:PSS exhibited characteristic bands of symmetric stretching mode of the aromatic C=C (1425 cm^{-1}), antisymmetric C-C and C-C stretching deformations (1530 cm^{-1} , 1374 cm^{-1} and 1261 cm^{-1}) and also oxyethylene ring deformation (989 cm^{-1} , 577 cm^{-1}).³¹⁶ The Raman spectrum of the RGO/PEDOT:PSS/BNC film also exhibited these characteristic bands, although with a weaker intensity (Figure 7.2D).

To understand the reduction state of GO flakes during the fabrication, we subjected as-synthesized GO flakes to high-temperature base wash and/or HPA treatment. X-ray photoelectron spectroscopy (XPS) was employed to probe the degree of reduction of GO upon base wash and/or HPA treatment. The 1s spectra of carbon can be deconvoluted into three peaks corresponding to sp^2 domains (C=C with a binding energy of 284 eV) and oxidized sp^3 domains (C–O with a binding energy of 286 eV and C=O with a binding energy of 288 eV).²¹² For as synthesized GO, the C/O ratio (1.7) obtained from the ratio of the area under the peaks indicates ~58% of the surface of GO is oxidized. After extensive base wash, the C/O ratio increased to 4.6 indicating that only ~21% of the surface of GO is oxidized. After further HPA treatment, the C/O ratio increased to 7.8, which indicates that only ~11% of the surface GO is oxidized (Figure 7.2E and appendix 6, Figure S6.2). The chemical reduction of GO flakes in the BNC matrix lead to an increased electrical conductivity. With the increase of GO content in the film up to 78 wt%, (corresponding to a concentration of 0.35 wt% in the broth), the conductivity of the RGO/BNC film increased to $15.6\text{ S}\cdot\text{cm}^{-1}$ (appendix 6, Figure S6.3). With further increase of GO concentration in the broth (*i.e.* above ~0.4 wt%), the viscosity of broth becomes too high leading to insufficient oxygen transport and unsuccessful film growth. The highest GO loading content achieved using the *in situ* growth and HPA reduction method is not sufficient to allow fast surface charge transfer, thus leading to poor capacitance performance (Figure 7.3A). With the addition of PEDOT:PSS into the BNC

matrix, the electrical conductivity and capacitance of the hybrid electrode can be greatly enhanced (Figure 7.2F and Figure 7.3A). However, the amount of PEDOT:PSS added to the bacterial broth needs to be carefully optimized. The PEDOT:PSS solution is highly acidic (pH 1.2 for 1 % aqueous solution) and addition of excess PEDOT:PSS solution causes a drop in the pH of the broth solution below pH 3 (optimal pH for BNC film growth), and also dilutes the nutrients in the broth, both of which compromise the growth of BNC film. Hence the concentration of PEDOT:PSS in the broth is a tradeoff between the film growth and conductivity of the composite film. Film with highest conductivity (274 S cm^{-1}) and highest capacitance was achieved at PEDOT:PSS and GO concentrations of 0.25 wt% and 0.35 wt% in the broth solution, respectively (which correspond to 22 wt% PEDOT:PSS and 63 wt% RGO in the dried film) (Figure 7.2F). For subsequent studies, BNC-based electrodes were prepared using broth solution corresponding to the highest conductivity and capacitance mentioned above.

7.43 Electrochemical Behavior of the RGO/PEDOT:PSS/BNC electrodes

We then investigated the electrochemical behavior of the RGO/PEDOT:PSS/BNC electrodes using cyclic voltammetry (CV) and galvanostatic charge-discharge (GCD) in a three-electrode configuration in H_2SO_4 (1 M) aqueous solution. The advantage of combining RGO and PEDOT:PSS for electrode preparation is evident from the CV curve of the RGO/PEDOT:PSS/BNC electrode, which has a larger integral area and rectangular shape than that of both RGO/BNC and PEDOT:PSS/BNC with same RGO or PEDOT:PSS loading, indicating a higher capacitance and faster charge transfer rate (Figure 7.3A). With an increase in the scan rate from 5 to 100 mV/s, the peak current density gradually increased; however, the shape of the CV curves deviates from the rectangular shape, which represents an ideal capacitance, to a spindle shape due to charge transfer resistance from the BNC matrix (Figure 7.3B). Specific capacitance

($C_{s,CV}$) of the electrodes calculated from the CV curves exhibited a decrease from 376 F/g to 55 F/g as the scan rates increased from 5 mV/s to 100 mV/s (Figure 7.3C).

GCD was performed at various current densities to study the cycling performance and capacitance retention (Figure 7.3D). It can be seen that the charge-discharge curves retain approximately symmetrical shape at different current densities from 0.5 to 5 A/g, indicating that the electrodes can be operated over a wide current range with reversible redox reaction and fast charge-discharge rate. From the charge-discharge curves, the specific capacitance ($C_{s,C-D}$) of RGO/PEDOT:PSS/BNC electrode was found to decrease from 470 F/g to 194 F/g (corresponds to 41% retained specific capacitance) with an increase in current density from 0.5 to 5 A/g. The $C_{s,C-D}$ (470 F/g) at 0.5 A/g and $C_{s,CV}$ (376 F/g) at 5 mV/s are higher compared to previous RGO/PEDOT based electrodes at the same current density and scan rate despite the internal resistance of non-conductive BNC matrix due possibly to the high conductivity PEDOT:PSS ($382 \pm 7.2 \text{ S cm}^{-1}$) employed here. More comparison between the presented electrode and other paper-based, carbon-based supercapacitors can be found in Table S6.1 in appendix 6.

The long-term stability of the electrode was examined using a galvanostatic charge-discharge method for 1000 cycles in aqueous solution. The capacitance retention of the electrode was calculated for each cycle based on the charge-discharge curves at a current density of 5 A/g (Figure 7.3F). It can be seen that although the capacity drops slightly in the first 200 cycles, the electrode retains more than 85% of its capacitance with no discernable changes in the capacitance over the last 800 cycles. These results indicate RGO/PEDOT:PSS/BNC electrode has excellent cycling stability, which is also evident from almost identical charge-discharge curves for 20 consecutive cycles randomly chosen (inset of Figure 7.3F). The high degree of cycling stability makes the RGO/PEDOT:PSS/BNC electrode promising for use in solid-state flexible supercapacitor devices.

7.44 The Performance of All Solid-state Flexible Supercapacitors devices

Now we turn our attention to exploring the advantages of using RGO/PEDOT:PSS/BNC electrodes and BNC papers for preparing all solid-state flexible supercapacitor devices. The devices were prepared by sandwiching a H₂SO₄/PVA/BNC paper as separator between two RGO/PEDOT:PSS/BNC electrodes without using any additional flexible substrates such as polyethylene terephthalate (PET) (see experimental section and Figure 7.4A). CV curves of the solid-state device exhibited rectangular shape up to scan rate of 50 mV/s, indicating good performance of the device at higher scan rate (Figure 7.4B). GCD was performed at different current densities ranging from 0.5 A/g to 8.0 A/g and these curves closely match the ideal symmetrical triangular curve. Even with a discharge at a current density of 4 A/g, the discharge time was found to be 48 s with no major sign of self-discharging (Figure 7.4C). One of the advantages of incorporating RGO and PEDOT:PSS in BNC matrix during the growth of the latter is that the electrode and device can inherit the excellent mechanical properties of BNC, obviating the need for flexible substrates such as PET. The fabricated devices exhibited excellent mechanical flexibility and could be bent to almost 180° multiple times without signs of fracture or delamination of the layers (Figure 7.4F). CV curves (at a scan rate of 50 mV/s) were also found to be insensitive to the bending angle between 0 to 180°, indicating excellent stability and flexibility of the device (Figure 7.4D). Subsequently, the cyclic stability of the devices was evaluated over 4,500 charge-discharge cycles (Figure 7.4E). The capacitance of the electrodes stabilized after the first 1000 cycles and remained essentially constant for the following 3500 cycles. To further demonstrate the potential application of the RGO/PEDOT:PSS/BNC electrodes-based flexible all solid-state supercapacitors, three devices were connected in tandem to power a red LED for a total of 3 minutes (Figure 7.4G).

7.5 Conclusions

In conclusion, we have demonstrated a novel and facile fabrication of RGO/PEDOT:PSS/BNC flexible electrode achieved via incorporation of GO flakes and conducting polymer PEDOT:PSS within the nanocellulose matrix during bacteria-mediated growth of the BNC hydrogel followed by aqueous processing for reduction of GO. The obtained electrodes exhibited excellent electrochemical performance (373 F/g at 1 A/g) and cycling stability (~85% capacitance retention over 1000 cycles) and they can be easily assembled with a BNC based separator to form a flexible all solid-state supercapacitor device. The light-weight device showed remarkable mechanical flexibility, performance stability under extreme mechanical deformation and ~88% capacitance retention over 4500 cycles, obviating the need for additional substrates such as PET. The fabrication approach demonstrated here, which is highly scalable and cost-effective, and offers great promise for energy storage devices in flexible, wearable and portable electronic devices.

7.6 Supporting Information

Supporting Information for chapter 7 is provided in appendix 6.

7.7 Figures

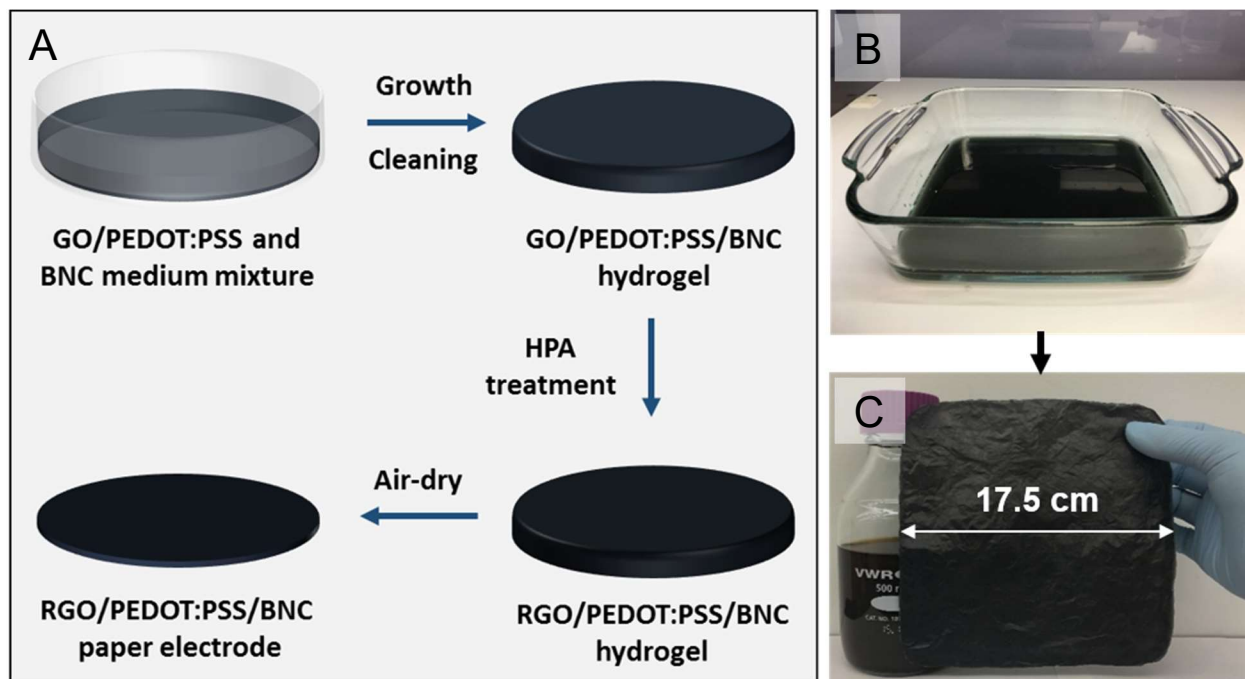


Figure 7.1. Fabrication of RGO/PEDOT:PSS/BNC electrode (A) Schematic illustration depicting the various steps involved in the fabrication of RGO/PEDOT:PSS/BNC electrode. Photographs showing (B) bacterial medium with GO and PEDOT:PSS and (C) dried RGO/PEDOT:PSS/BNC electrode.

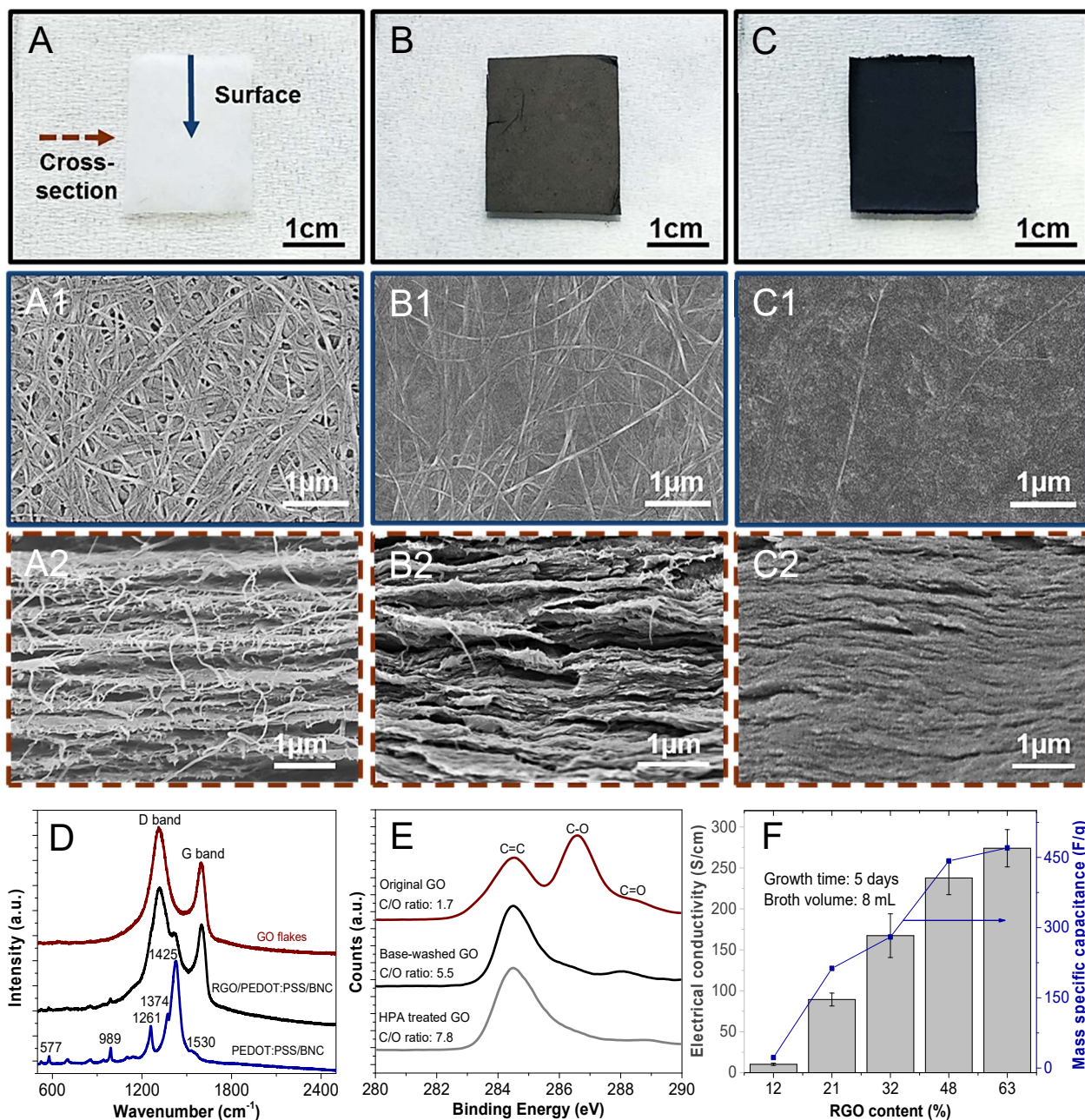


Figure 7.2. A) Optical image (A) and SEM images of top surface (A1) and cross-section (A2) of BNC film. (B) Optical image (B) and SEM images of top surface (B1) and cross-section (B2) of RGO/BNC film. (C) Optical image (C) and SEM images of top surface (C1) and cross-section (C2) of RGO/PEDOT:PSS/BNC film. (D) Raman spectra of original GO flakes, RGO/PEDOT:PSS/BNC film and PEDOT:PSS/BNC film. (E) XPS spectra of original, base-washed and HPA treated GO. (F) Electrical conductivity and the mass specific

capacitance of RGO/PEDOT:PSS/BNC electrodes with different RGO concentrations and fixed PEDOT:PSS concentration (~0.25 wt%) in the bacterial broth.

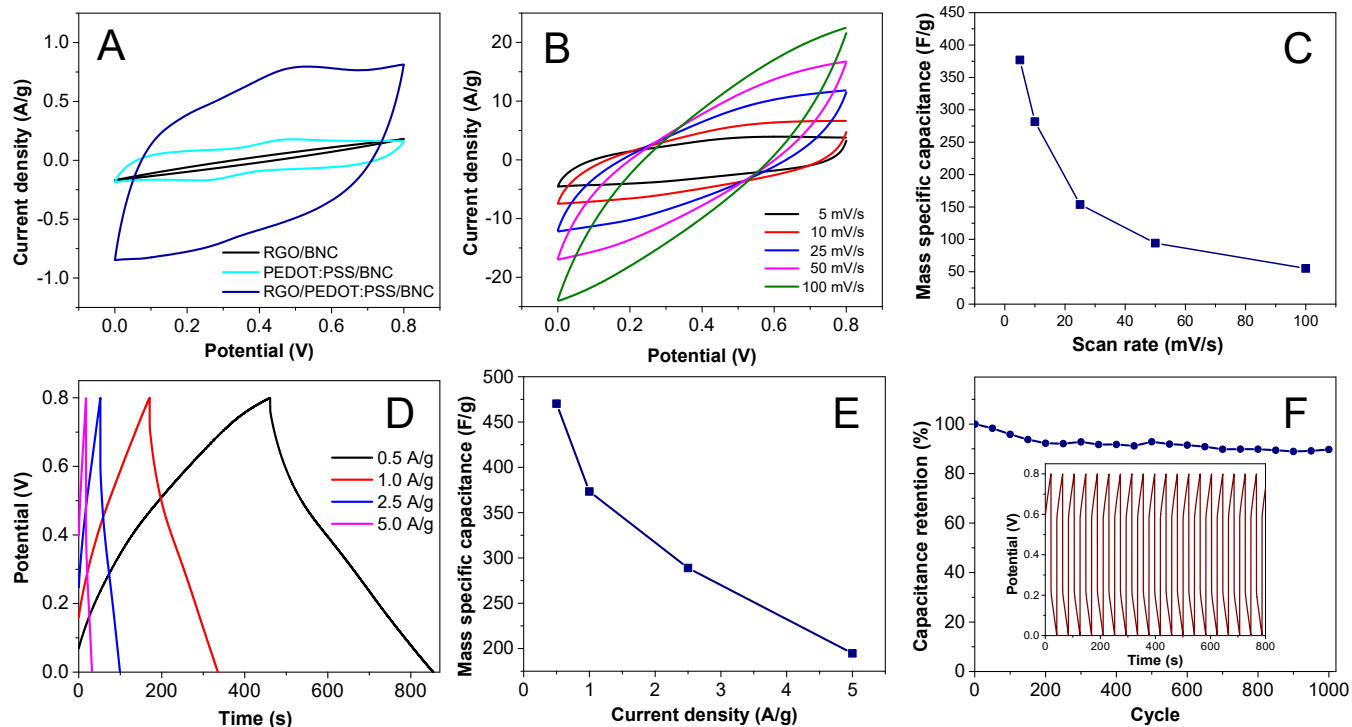


Figure 7.3. (A) CV curves of RGO/BNC film, PEDOT:PSS/BNC film and RGO/PEDOT:PSS/BNC electrode. (B) CV curves of RGO/PEDOT:PSS/BNC electrode at different scan rates (5 to 100 mV/s). (C) Mass specific capacitance of RGO/PEDOT:PSS/BNC electrode calculated from CV curves as a function of scan rate. (D) Galvanostatic charging/discharging (GCD) curves of RGO/PEDOT:PSS/BNC electrode. (E) Mass specific capacitance of RGO/PEDOT:PSS/BNC calculated from GCD curves as a function of current density. (F) Capacitance retention of RGO/PEDOT:PSS/BNC electrode over 1000 cycles. The inset shows the randomly picked 20 GCD curves.

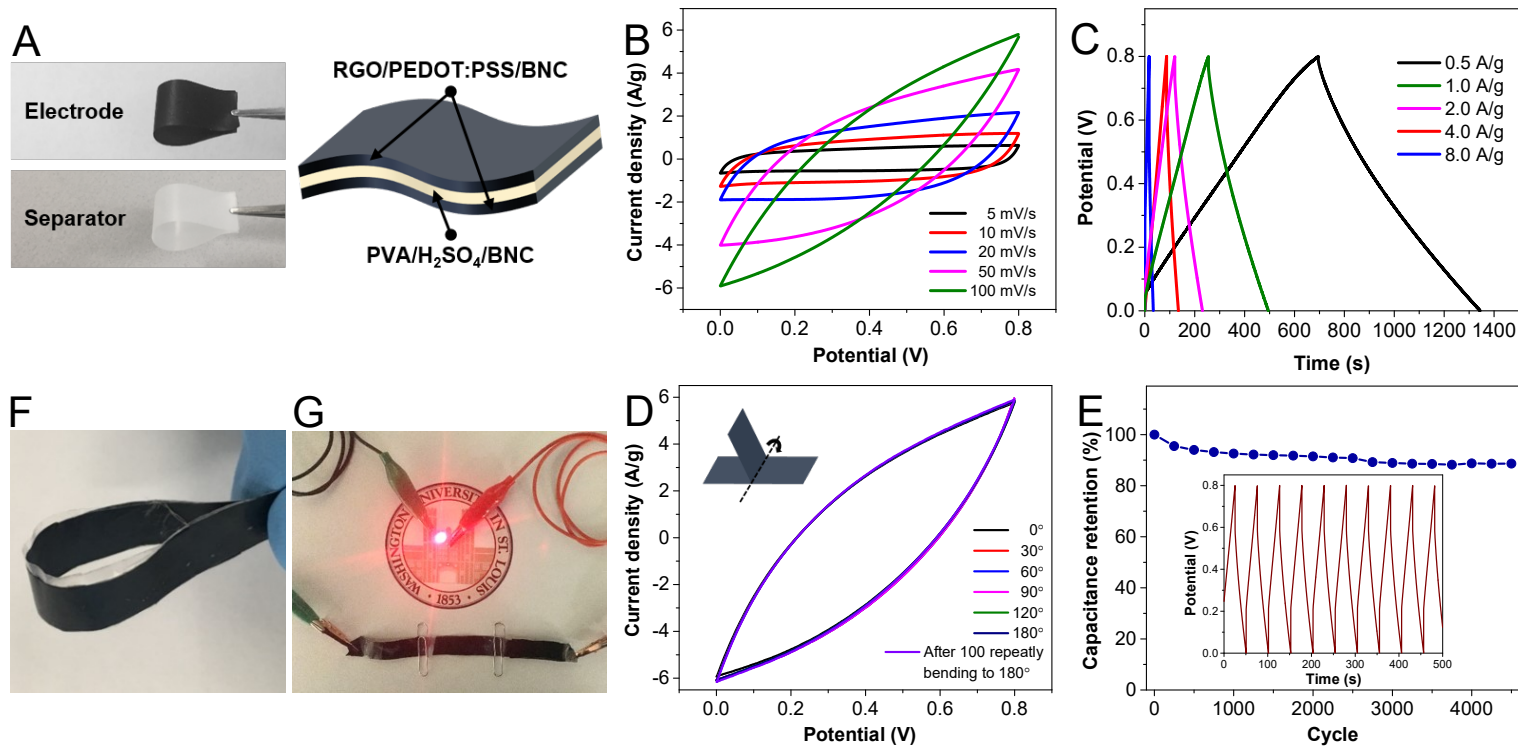


Figure 7.4. (A) Photographs showing the flexibility of the components (electrode and separator) of device and a schematic showing the structure of the solid-state supercapacitor device based on RGO/PEDOT:PSS/BNC electrodes. (B) CV curves of assembled supercapacitor at different scan rate from 5 to 100 mV/s. (C) GCD curves of assembled supercapacitor at different current densities. (D) CV curves of assembled supercapacitor under different bending angles at a scan rate of 100 mV/s. (E) Cycling stability of assembled supercapacitor over 4500 cycles. The inset shows randomly picked 10 GCD curves. (F) Optical image showing the flexibility of assembled supercapacitor. (G) LED indicator lighted by assembled device.

Chapter 8: Conclusions

8.1 General conclusions

Owing to its desirable properties such as excellent mechanical strength and toughness, rich chemical functionality, and high specific surface area in aerogel state, bacterial nanocellulose (BNC) can serve as an ideal matrix for realizing functional composites. Over the last three years, we have introduced several scalable BNC-based functional composites mainly using a “bacterial factory” method. By simply introducing functional nanomaterials during the bacteria-mediated BNC growth, functional nanocomposites can be formed in a scalable, green, and cost-effective manner. This dissertation demonstrates several examples of BNC-based functional nanocomposites with applications in sensing, water purification and energy storage.

We have demonstrated a BNC film-based surface enhanced Raman scattering (SERS) substrate, achieved *via* gravity-assisted filtration of plasmonic nanostructures. The 3D porous structure and ultrafine fibers of BNC facilitates uniform and dense adsorption of plasmonic nanostructures on the surface and in sub-surface regions, which results in large SERS enhancement and excellent uniformity of SERS activity across the entire substrate. Harnessing the smooth surface of BNC, we show that BNC-based SERS substrate serves as an ideal platform for collection, detection and recognition of bacteria.

We have demonstrated a novel, highly scalable, cost-effective and green strategy to realize functional BNC-based foams/membranes, taking advantage of the bacterial production of BNC. Functional materials with different dimensions (0D to 3D) such as graphene oxide (GO), polydopamine (PDA) can be efficiently incorporated within the BNC matrix during its growth. The functional composites achieved by this approach showed excellent mechanical robustness and

flexibility owing to the intercalation of functional materials within the layered BNC matrix, which is crucial for efficient, large-scale applications, either as a foam or as a membrane.

We have designed and developed a bilayered hybrid biofoam comprised of BNC and RGO for highly efficient solar steam generation. Owing to the large light absorption, excellent photothermal conversion, heat localization and efficient transport of water from bulk to evaporative surface, the novel bilayered structure exhibited a remarkably high solar thermal efficiency. A flexible, scalable and, more importantly, completely biodegradable photothermal evaporator for highly efficient solar steam generation has then been demonstrated. The biodegradable PDA/BNC foam introduced here exhibits large light absorption and photothermal conversion, heat localization, and efficient water transportation, leading to excellent solar steam generation performance under one sun. An innovative water filtration membrane based on BNC and RGO, which harvests sunlight to kill microorganisms has been demonstrated to provide a novel anti-biofouling approach. The RGO/BNC membrane exhibited outstanding mechanical and chemical stability and stable water flux under high pressure. Particularly, owing to its photothermal properties, the membrane exhibited light-enabled bactericidal activity, avoiding the need for any treatment of the feed water or any external energy.

Additionally, we have demonstrated that the *in situ* prepared nanocomposites can also serve as a platform for further uniform modification of other nanomaterials due to the ease of handling and robustness of the nanocomposites. We have demonstrated a robust filtration membrane based on BNC loaded with GO and PdNPs which has excellent dye degradation performance for highly efficient wastewater treatment. The Pd/GO/BNC membrane is realized by post formation of PdNPs in an *in situ* prepared GO/BNC membrane. The Pd/GO/BNC membrane exhibited highly efficient methylene orange (MO) degradation during filtration (over a wide range of MO

concentrations, pH and multiple cycles of reuse). Multiple contaminants (a cocktail of 4-nitrophenol, methylene blue and rhodamine 6G) could also be effectively treated by Pd/GO/BNC membrane simultaneously during filtration. The Pd/GO/BNC membrane also demonstrated stable flux under high pressure over long duration.

Furthermore, hybrid functional composites can be realized to achieve optimal performance due to synergistic effect by introducing multiple functional materials such as GO and poly(3,4-ethylenedioxythiophene)-poly(styrene sulfonate) (PEDOT:PSS). The obtained RGO-/PEDOT:PSS/BNC electrodes exhibited excellent electrochemical performance and cycling stability and they can be easily assembled with a BNC based separator to form a flexible all solid-state supercapacitor device. The light-weight device showed remarkable mechanical flexibility, performance stability under extreme mechanical deformation, obviating the need for additional substrates such as PET.

8.2 Significance and Outlook

In addition to the representative nanocomposites demonstrated in this study, the fabrication strategies and design principles demonstrated in this work can have far reaching implications to realize various nanocomposites with applications in water purification, energy harvesting, sensing, catalysis, and life sciences. Especially with the incorporation of emerging materials (such as GO, PDA, Mxenes and Metal organic frameworks, etc.), unprecedented multifunctional nanocomposites can be achieved in a scalable, cost-effective way.

References

1. Samir, M. A. S. A.; Alloin, F.; Dufresne, A., Review of recent research into cellulosic whiskers, their properties and their application in nanocomposite field. *Biomacromolecules* **2005**, *6* (2), 612-626.
2. Jin, H.; Nishiyama, Y.; Wada, M.; Kuga, S., Nanofibrillar cellulose aerogels. *Colloids and Surfaces A: Physicochemical and Engineering Aspects* **2004**, *240* (1–3), 63-67.
3. Klemm, D.; Kramer, F.; Moritz, S.; Lindström, T.; Ankerfors, M.; Gray, D.; Dorris, A., Nanocelluloses: A New Family of Nature-Based Materials. *Angewandte Chemie International Edition* **2011**, *50* (24), 5438-5466.
4. Valo, H.; Arola, S.; Laaksonen, P.; Torkkeli, M.; Peltonen, L.; Linder, M. B.; Serimaa, R.; Kuga, S.; Hirvonen, J.; Laaksonen, T., Drug release from nanoparticles embedded in four different nanofibrillar cellulose aerogels. *European Journal of Pharmaceutical Sciences* **2013**, *50* (1), 69-77.
5. Falk, L.; Emmerich, H.; Antje, P.; Dieter, L.; Stefanie, T.; Marie-Alexandra, N.; Martin, W.; Thomas, R., Cellulosic aerogels as ultra-lightweight materials. Part 2: Synthesis and properties 2nd ICC 2007, Tokyo, Japan, October 25–29, 2007. *Holzforschung: International Journal of the Biology, Chemistry, Physics, & Technology of Wood* **2009**, *63* (1), 3-11.
6. Gatenholm, P.; Klemm, D., Bacterial Nanocellulose as a Renewable Material for Biomedical Applications. *Mrs Bull* **2010**, *35* (3), 208-213.
7. Shoda, M.; Sugano, Y., Recent advances in bacterial cellulose production. *Biotechnol Bioproc E* **2005**, *10* (1), 1-8.
8. Yamanaka, S.; Watanabe, K.; Kitamura, N.; Iguchi, M.; Mitsuhashi, S.; Nishi, Y.; Uryu, M., The structure and mechanical properties of sheets prepared from bacterial cellulose. *J Mater Sci* **1989**, *24* (9), 3141-3145.
9. Salmon, S.; Hudson, S. M., Crystal morphology, biosynthesis, and physical assembly of cellulose, chitin, and chitosan. *Journal of Macromolecular Science, Part C: Polymer Reviews* **1997**, *37* (2), 199-276.
10. Klemm, D.; Heublein, B.; Fink, H.-P.; Bohn, A., Cellulose: Fascinating Biopolymer and Sustainable Raw Material. *Angewandte Chemie International Edition* **2005**, *44* (22), 3358-3393.
11. Wu, Z.-Y.; Liang, H.-W.; Chen, L.-F.; Hu, B.-C.; Yu, S.-H., Bacterial Cellulose: A Robust Platform for Design of Three Dimensional Carbon-Based Functional Nanomaterials. *Accounts of Chemical Research* **2016**, *49* (1), 96-105.
12. Tian, L.; Jiang, Q.; Liu, K.-K.; Luan, J.; Naik, R. R.; Singamaneni, S., Bacterial Nanocellulose-Based Flexible Surface Enhanced Raman Scattering Substrate. *Advanced Materials Interfaces* **2016**, *3* (15), n/a-n/a.
13. Tian, L.; Luan, J.; Liu, K.-K.; Jiang, Q.; Tadepalli, S.; Gupta, M. K.; Naik, R. R.; Singamaneni, S., Plasmonic Biofoam: A Versatile Optically Active Material. *Nano Letters* **2016**, *16* (1), 609-616.
14. Chen, L.-F.; Huang, Z.-H.; Liang, H.-W.; Gao, H.-L.; Yu, S.-H., Three-Dimensional Heteroatom-Doped Carbon Nanofiber Networks Derived from Bacterial Cellulose for Supercapacitors. *Advanced Functional Materials* **2014**, *24* (32), 5104-5111.
15. Chen, L.-F.; Zhang, X.-D.; Liang, H.-W.; Kong, M.; Guan, Q.-F.; Chen, P.; Wu, Z.-Y.; Yu, S.-H., Synthesis of Nitrogen-Doped Porous Carbon Nanofibers as an Efficient Electrode Material for Supercapacitors. *ACS Nano* **2012**, *6* (8), 7092-7102.
16. Tong, S.; Zheng, M.; Lu, Y.; Lin, Z.; Zhang, X.; He, P.; Zhou, H., Binder-free carbonized bacterial cellulose-supported ruthenium nanoparticles for Li-O₂ batteries. *Chemical Communications* **2015**, *51* (34), 7302-7304.
17. Wang, B.; Li, X.; Luo, B.; Yang, J.; Wang, X.; Song, Q.; Chen, S.; Zhi, L., Pyrolyzed Bacterial Cellulose: A Versatile Support for Lithium Ion Battery Anode Materials. *Small* **2013**, *9* (14), 2399-2404.

18. Liang, H.-W.; Guan, Q.-F.; Zhu, Z.; Song, L.-T.; Yao, H.-B.; Lei, X.; Yu, S.-H., Highly conductive and stretchable conductors fabricated from bacterial cellulose. *NPG Asia Mater* **2012**, *4*, e19.
19. Olsson, R. T.; Azizi Samir, M. A. S.; Salazar Alvarez, G.; Beloval; StromV; Berglund, L. A.; IkkalaO; NoguesJ; Gedde, U. W., Making flexible magnetic aerogels and stiff magnetic nanopaper using cellulose nanofibrils as templates. *Nat Nano* **2010**, *5* (8), 584-588.
20. Jiang, Q.; Tian, L.; Liu, K.-K.; Tadepalli, S.; Raliya, R.; Biswas, P.; Naik, R. R.; Singamaneni, S., Bilayered Biofoam for Highly Efficient Solar Steam Generation. *Advanced Materials* **2016**, n/a-n/a.
21. Camden, J. P.; Dieringer, J. A.; Zhao, J.; Van Duyne, R. P., Controlled Plasmonic Nanostructures for Surface-Enhanced Spectroscopy and Sensing. *Accounts of Chemical Research* **2008**, *41* (12), 1653-1661.
22. Ko, H.; Singamaneni, S.; Tsukruk, V. V., Nanostructured Surfaces and Assemblies as SERS Media. *Small* **2008**, *4* (10), 1576-1599.
23. Campion, A.; Kambhampati, P., Surface-enhanced Raman scattering. *Chem Soc Rev* **1998**, *27* (4), 241-250.
24. Li, J. F.; Huang, Y. F.; Ding, Y.; Yang, Z. L.; Li, S. B.; Zhou, X. S.; Fan, F. R.; Zhang, W.; Zhou, Z. Y.; Wu, D. Y.; Ren, B.; Wang, Z. L.; Tian, Z. Q., Shell-isolated nanoparticle-enhanced Raman spectroscopy. *Nature* **2010**, *464* (7287), 392-395.
25. Liu, T. Y.; Tsai, K. T.; Wang, H. H.; Chen, Y.; Chen, Y. H.; Chao, Y. C.; Chang, H. H.; Lin, C. H.; Wang, J. K.; Wang, Y. L., Functionalized arrays of Raman-enhancing nanoparticles for capture and culture-free analysis of bacteria in human blood. *Nat Commun* **2011**, *2*.
26. Anker, J. N.; Hall, W. P.; Lyandres, O.; Shah, N. C.; Zhao, J.; Van Duyne, R. P., Biosensing with plasmonic nanosensors. *Nat Mater* **2008**, *7* (6), 442-453.
27. Gandra, N.; Abbas, A.; Tian, L. M.; Singamaneni, S., Plasmonic Planet-Satellite Analogues: Hierarchical Self-Assembly of Gold Nanostructures. *Nano Letters* **2012**, *12* (5), 2645-2651.
28. Moskovits, M., Surface-enhanced Raman spectroscopy: a brief retrospective. *Journal of Raman Spectroscopy* **2005**, *36* (6-7), 485-496.
29. Golightly, R. S.; Doering, W. E.; Natan, M. J., Surface-Enhanced Raman Spectroscopy and Homeland Security: A Perfect Match? *ACS Nano* **2009**, *3* (10), 2859-2869.
30. Homola, J., Surface Plasmon Resonance Sensors for Detection of Chemical and Biological Species. *Chemical Reviews* **2008**, *108* (2), 462-493.
31. Fang, X.; Ahmad, S. R., Detection of explosive vapour using surface-enhanced Raman spectroscopy. *Appl. Phys. B* **2009**, *97* (3), 723-726.
32. Hering, K.; Cialla, D.; Ackermann, K.; Dörfer, T.; Möller, R.; Schneidewind, H.; Mattheis, R.; Fritzsche, W.; Rösch, P.; Popp, J., SERS: a versatile tool in chemical and biochemical diagnostics. *Anal Bioanal Chem* **2008**, *390* (1), 113-124.
33. Stewart, M. E.; Anderton, C. R.; Thompson, L. B.; Maria, J.; Gray, S. K.; Rogers, J. A.; Nuzzo, R. G., Nanostructured Plasmonic Sensors. *Chemical Reviews* **2008**, *108* (2), 494-521.
34. Lee, C. H.; Tian, L. M.; Singamaneni, S., Paper-Based SERS Swab for Rapid Trace Detection on Real-World Surfaces. *Acs Appl Mater Inter* **2010**, *2* (12), 3429-3435.
35. Tian, L.; Morrissey, J. J.; Kattumenu, R.; Gandra, N.; Kharasch, E. D.; Singamaneni, S., Bioplasmonic Paper as a Platform for Detection of Kidney Cancer Biomarkers. *Analytical Chemistry* **2012**, *84* (22), 9928-9934.
36. Martinak, D.; Rudolph, A. In *Explosives detection using an ion mobility spectrometer for airport security*, Security Technology, 1997. Proceedings. The Institute of Electrical and Electronics Engineers 31st Annual 1997 International Carnahan Conference on, 15-17 Oct 1997; 1997; pp 188-189.
37. Yu, W. W.; White, I. M., Inkjet Printed Surface Enhanced Raman Spectroscopy Array on Cellulose Paper. *Analytical Chemistry* **2010**, *82* (23), 9626-9630.
38. Lee, C. H.; Tian, L.; Singamaneni, S., Paper-Based SERS Swab for Rapid Trace Detection on Real-World Surfaces. *ACS Applied Materials & Interfaces* **2010**, *2* (12), 3429-3435.

39. Elimelech, M., The global challenge for adequate and safe water. *Journal of Water Supply: Research and Technology - Aqua* **2006**, *55* (1), 3-10.
40. P-H. Gleick, e. a., *The world's Water: The Biennial Report on Freshwater Resources*. Island Press: Washington, DC, 2014; Vol. 8.
41. Eliasson, J., The Rising Pressure of Global Water Shortages. *Nature* **2014**, *517* (7532).
42. Shang, W.; Deng, T., Solar steam generation: Steam by thermal concentration. **2016**, *1*, 16133.
43. Ghasemi, H.; Ni, G.; Marconnet, A. M.; Loomis, J.; Yerci, S.; Miljkovic, N.; Chen, G., Solar steam generation by heat localization. **2014**, *5*, 4449.
44. Jiang, Q.; Gholami Derami, H.; Ghim, D.; Cao, S.; Jun, Y.-S.; Singamaneni, S., Polydopamine-filled bacterial nanocellulose as a biodegradable interfacial photothermal evaporator for highly efficient solar steam generation. *Journal of Materials Chemistry A* **2017**, *5* (35), 18397-18402.
45. Nair, R. R.; Wu, H. A.; Jayaram, P. N.; Grigorieva, I. V.; Geim, A. K., Unimpeded Permeation of Water Through Helium-Leak-Tight Graphene-Based Membranes. *Science* **2012**, *335* (6067), 442-444.
46. Joshi, R. K.; Alwarappan, S.; Yoshimura, M.; Sahajwalla, V.; Nishina, Y., Graphene oxide: the new membrane material. *Applied Materials Today* **2015**, *1* (1), 1-12.
47. Sun, P.; Wang, K.; Zhu, H., Recent Developments in Graphene-Based Membranes: Structure, Mass-Transport Mechanism and Potential Applications. *Advanced Materials* **2016**, *28* (12), 2287-2310.
48. Mi, B., Graphene Oxide Membranes for Ionic and Molecular Sieving. *Science* **2014**, *343* (6172), 740.
49. Dreyer, D. R.; Park, S.; Bielawski, C. W.; Ruoff, R. S., The chemistry of graphene oxide. *Chemical Society Reviews* **2010**, *39* (1), 228-240.
50. Yeh, C.-N.; Raidongia, K.; Shao, J.; Yang, Q.-H.; Huang, J., On the origin of the stability of graphene oxide membranes in water. *Nat Chem* **2015**, *7* (2), 166-170.
51. Shannon, M. A.; Bohn, P. W.; Elimelech, M.; Georgiadis, J. G.; Marinas, B. J.; Mayes, A. M., Science and technology for water purification in the coming decades. *Nature* **2008**, *452* (7185), 301-310.
52. Singh, R.; Hankins, N., *Emerging Membrane Technology for Sustainable Water Treatment*. Elsevier 2016.
53. Pendergast, M. M.; Hoek, E. M. V., A review of water treatment membrane nanotechnologies. *Energy & Environmental Science* **2011**, *4* (6), 1946-1971.
54. Le-Clech, P.; Chen, V.; Fane, T. A. G., Fouling in membrane bioreactors used in wastewater treatment. *Journal of Membrane Science* **2006**, *284* (1-2), 17-53.
55. Zhang, R.; Liu, Y.; He, M.; Su, Y.; Zhao, X.; Elimelech, M.; Jiang, Z., Antifouling membranes for sustainable water purification: strategies and mechanisms. *Chemical Society Reviews* **2016**.
56. Flemming, H. C.; Schaule, G.; Griebe, T.; Schmitt, J.; Tamachkiarowa, A., Workshop on Membranes in Drinking Water Production Technical Innovations and Health Aspects Biofouling—the Achilles heel of membrane processes. *Desalination* **1997**, *113* (2), 215-225.
57. Komlenic, R., Rethinking the causes of membrane biofouling. *Filtration & separation* **2010**, *47* (5), 26-28.
58. Baker, J. S.; Dudley, L. Y., Biofouling in membrane systems - A review. *Desalination* **1998**, *118* (1-3), 81-89.
59. Nguyen, T.; Roddick, F. A.; Fan, L., Biofouling of Water Treatment Membranes: A Review of the Underlying Causes, Monitoring Techniques and Control Measures. *Membranes* **2012**, *2* (4), 804-840.
60. Mansouri, J.; Harrisson, S.; Chen, V., Strategies for controlling biofouling in membrane filtration systems: challenges and opportunities. *Journal of Materials Chemistry* **2010**, *20* (22), 4567-4586.
61. Barnes, R. J.; Low, J. H.; Bandi, R. R.; Tay, M.; Chua, F.; Aung, T.; Fane, A. G.; Kjelleberg, S.; Rice, S. A., Nitric Oxide Treatment for the Control of Reverse Osmosis Membrane Biofouling. *Applied and Environmental Microbiology* **2015**, *81* (7), 2515-2524.

62. Lade, H.; Paul, D.; Kweon, J. H., Quorum Quenching Mediated Approaches for Control of Membrane Biofouling. *International Journal of Biological Sciences* **2014**, *10* (5), 550-565.
63. Yang, H.-L.; Lin, J. C.-T.; Huang, C., Application of nanosilver surface modification to RO membrane and spacer for mitigating biofouling in seawater desalination. *Water Research* **2009**, *43* (15), 3777-3786.
64. Kumar, A.; Vemula, P. K.; Ajayan, P. M.; John, G., Silver-nanoparticle-embedded antimicrobial paints based on vegetable oil. *Nat Mater* **2008**, *7* (3), 236-241.
65. Perreault, F.; Tousley, M. E.; Elimelech, M., Thin-Film Composite Polyamide Membranes Functionalized with Biocidal Graphene Oxide Nanosheets. *Environmental Science & Technology Letters* **2014**, *1* (1), 71-76.
66. Perreault, F.; Fonseca de Faria, A.; Elimelech, M., Environmental applications of graphene-based nanomaterials. *Chemical Society Reviews* **2015**, *44* (16), 5861-5896.
67. Kochkodan, V.; Hilal, N., A comprehensive review on surface modified polymer membranes for biofouling mitigation. *Desalination* **2015**, *356*, 187-207.
68. Ray, J. R.; Tadepalli, S.; Nergiz, S. Z.; Liu, K.-K.; You, L.; Tang, Y.; Singamaneni, S.; Jun, Y.-S., Hydrophilic, Bactericidal Nanoheater-Enabled Reverse Osmosis Membranes to Improve Fouling Resistance. *ACS Applied Materials & Interfaces* **2015**, *7* (21), 11117-11126.
69. Ren, Z. J.; Umble, A. K., Water treatment: Recover wastewater resources locally. *Nature* **2016**, *529* (7584), 25-25.
70. Ahmed, T.; Imdad, S.; Yaldram, K.; Butt, N. M.; Pervez, A., Emerging nanotechnology-based methods for water purification: a review. *Desalination and Water Treatment* **2014**, *52* (22-24), 4089-4101.
71. Khataee, A. R.; Kasiri, M. B., Photocatalytic degradation of organic dyes in the presence of nanostructured titanium dioxide: Influence of the chemical structure of dyes. *Journal of Molecular Catalysis A: Chemical* **2010**, *328* (1), 8-26.
72. Yu, K.; Yang, S.; Liu, C.; Chen, H.; Li, H.; Sun, C.; Boyd, S. A., Degradation of Organic Dyes via Bismuth Silver Oxide Initiated Direct Oxidation Coupled with Sodium Bismuthate Based Visible Light Photocatalysis. *Environmental Science & Technology* **2012**, *46* (13), 7318-7326.
73. Sakkas, V. A.; Islam, M. A.; Stalikas, C.; Albanis, T. A., Photocatalytic degradation using design of experiments: A review and example of the Congo red degradation. *Journal of Hazardous Materials* **2010**, *175* (1), 33-44.
74. Vanhulle, S.; Trovaslet, M.; Enaud, E.; Lucas, M.; Taghavi, S.; van der Lelie, D.; van Aken, B.; Foret, M.; Onderwater, R. C. A.; Wesenberg, D.; Agathos, S. N.; Schneider, Y.-J.; Corbisier, A.-M., Decolorization, Cytotoxicity, and Genotoxicity Reduction During a Combined Ozonation/Fungal Treatment of Dye-Contaminated Wastewater. *Environmental Science & Technology* **2008**, *42* (2), 584-589.
75. Forgacs, E.; Cserhádi, T.; Oros, G., Removal of synthetic dyes from wastewaters: a review. *Environment International* **2004**, *30* (7), 953-971.
76. Robinson, T.; McMullan, G.; Marchant, R.; Nigam, P., Remediation of dyes in textile effluent: a critical review on current treatment technologies with a proposed alternative. *Bioresource Technology* **2001**, *77* (3), 247-255.
77. Sun, Y.; Tao, Z.; Chen, J.; Herricks, T.; Xia, Y., Ag Nanowires Coated with Ag/Pd Alloy Sheaths and Their Use as Substrates for Reversible Absorption and Desorption of Hydrogen. *Journal of the American Chemical Society* **2004**, *126* (19), 5940-5941.
78. Deng, Y.; Cai, Y.; Sun, Z.; Liu, J.; Liu, C.; Wei, J.; Li, W.; Liu, C.; Wang, Y.; Zhao, D., Multifunctional Mesoporous Composite Microspheres with Well-Designed Nanostructure: A Highly Integrated Catalyst System. *Journal of the American Chemical Society* **2010**, *132* (24), 8466-8473.
79. Wang, F.; Li, C.; Sun, L.-D.; Xu, C.-H.; Wang, J.; Yu, J. C.; Yan, C.-H., Porous Single-Crystalline Palladium Nanoparticles with High Catalytic Activities. *Angewandte Chemie International Edition* **2012**, *51* (20), 4872-4876.

80. Nishihata, Y.; Mizuki, J.; Akao, T.; Tanaka, H.; Uenishi, M.; Kimura, M.; Okamoto, T.; Hamada, N., Self-regeneration of a Pd-perovskite catalyst for automotive emissions control. *Nature* **2002**, *418* (6894), 164-167.
81. Kurt, B. Z.; Durmus, Z.; Durmus, A., Preparation and characterization of platinum (Pt) and palladium (Pd) nanoparticle decorated graphene sheets and their utilization for the elimination of basic fuchsin and indigo carmine dyes. *Solid State Sciences* **2016**, *51*, 51-58.
82. Safavi, A.; Momeni, S., Highly efficient degradation of azo dyes by palladium/hydroxyapatite/Fe₃O₄ nanocatalyst. *Journal of Hazardous Materials* **2012**, *201*, 125-131.
83. Jana, N. R.; Wang, Z. L.; Pal, T., Redox Catalytic Properties of Palladium Nanoparticles: Surfactant and Electron Donor–Acceptor Effects. *Langmuir* **2000**, *16* (6), 2457-2463.
84. Rohand, T.; Qin, W.; Boens, N.; Dehaen, W., Palladium-Catalyzed Coupling Reactions for the Functionalization of BODIPY Dyes with Fluorescence Spanning the Visible Spectrum. *European Journal of Organic Chemistry* **2006**, *2006* (20), 4658-4663.
85. Leyva-Pérez, A.; Oliver-Meseguer, J.; Rubio-Marqués, P.; Corma, A., Water-Stabilized Three- and Four-Atom Palladium Clusters as Highly Active Catalytic Species in Ligand-Free C–C Cross-Coupling Reactions. *Angewandte Chemie International Edition* **2013**, *52* (44), 11554-11559.
86. Celebi, M.; Yurderi, M.; Bulut, A.; Kaya, M.; Zahmakiran, M., Palladium nanoparticles supported on amine-functionalized SiO₂ for the catalytic hexavalent chromium reduction. *Applied Catalysis B: Environmental* **2016**, *180*, 53-64.
87. Adib, M.; Karimi-Nami, R.; Veisi, H., Palladium NPs supported on novel imino-pyridine-functionalized MWCNTs: efficient and highly reusable catalysts for the Suzuki–Miyaura and Sonogashira coupling reactions. *New Journal of Chemistry* **2016**, *40* (6), 4945-4951.
88. Freakley, S. J.; He, Q.; Harray, J. H.; Lu, L.; Crole, D. A.; Morgan, D. J.; Ntainjua, E. N.; Edwards, J. K.; Carley, A. F.; Borisevich, A. Y., Palladium-tin catalysts for the direct synthesis of H₂O₂ with high selectivity. *Science* **2016**, *351* (6276), 965-968.
89. Bhaskar, R.; Joshi, H.; Sharma, A. K.; Singh, A. K., Reusable Catalyst for Transfer Hydrogenation of Aldehydes and Ketones Designed by Anchoring Palladium as Nano-Particles on Graphene Oxide Functionalized with Selenated Amine. *ACS Applied Materials & Interfaces* **2017**.
90. Zhu, W.; Wu, Z.; Foo, G. S.; Gao, X.; Zhou, M.; Liu, B.; Veith, G. M.; Wu, P.; Browning, K. L.; Lee, H. N., Taming interfacial electronic properties of platinum nanoparticles on vacancy-abundant boron nitride nanosheets for enhanced catalysis. *Nature communications* **2017**, *8*, 15291.
91. Yang, J.; Tian, C.; Wang, L.; Fu, H., An effective strategy for small-sized and highly-dispersed palladium nanoparticles supported on graphene with excellent performance for formic acid oxidation. *Journal of Materials Chemistry* **2011**, *21* (10), 3384-3390.
92. Wang, Q.; Cui, X.; Chen, J.; Zheng, X.; Liu, C.; Xue, T.; Wang, H.; Jin, Z.; Qiao, L.; Zheng, W., Well-dispersed palladium nanoparticles on graphene oxide as a non-enzymatic glucose sensor. *RSC Advances* **2012**, *2* (15), 6245-6249.
93. Rostamnia, S.; Doustkhah, E.; Karimi, Z.; Amini, S.; Luque, R., Surfactant-Exfoliated Highly Dispersive Pd-Supported Graphene Oxide Nanocomposite as a Catalyst for Aerobic Aqueous Oxidations of Alcohols. *ChemCatChem* **2015**, *7* (11), 1678-1683.
94. Krittayavathananon, A.; Srimuk, P.; Luanwuthi, S.; Sawangphruk, M., Palladium Nanoparticles Decorated on Reduced Graphene Oxide Rotating Disk Electrodes toward Ultrasensitive Hydrazine Detection: Effects of Particle Size and Hydrodynamic Diffusion. *Analytical Chemistry* **2014**, *86* (24), 12272-12278.
95. Huang, Y.-X.; Xie, J.-F.; Zhang, X.; Xiong, L.; Yu, H.-Q., Reduced Graphene Oxide Supported Palladium Nanoparticles via Photoassisted Citrate Reduction for Enhanced Electrocatalytic Activities. *ACS Applied Materials & Interfaces* **2014**, *6* (18), 15795-15801.

96. Moussa, S.; Siamaki, A. R.; Gupton, B. F.; El-Shall, M. S., Pd-Partially Reduced Graphene Oxide Catalysts (Pd/PRGO): Laser Synthesis of Pd Nanoparticles Supported on PRGO Nanosheets for Carbon–Carbon Cross Coupling Reactions. *ACS Catalysis* **2012**, *2* (1), 145-154.
97. Jiao, T.; Zhao, H.; Zhou, J.; Zhang, Q.; Luo, X.; Hu, J.; Peng, Q.; Yan, X., Self-Assembly Reduced Graphene Oxide Nanosheet Hydrogel Fabrication by Anchorage of Chitosan/Silver and Its Potential Efficient Application toward Dye Degradation for Wastewater Treatments. *ACS Sustainable Chemistry & Engineering* **2015**, *3* (12), 3130-3139.
98. Zhao, X.; Ma, K.; Jiao, T.; Xing, R.; Ma, X.; Hu, J.; Huang, H.; Zhang, L.; Yan, X., Fabrication of Hierarchical Layer-by-Layer Assembled Diamond-based Core-Shell Nanocomposites as Highly Efficient Dye Absorbents for Wastewater Treatment. *Scientific Reports* **2017**, *7*, 44076.
99. Guo, R.; Jiao, T.; Li, R.; Chen, Y.; Guo, W.; Zhang, L.; Zhou, J.; Zhang, Q.; Peng, Q., Sandwiched Fe₃O₄/Carboxylate Graphene Oxide Nanostructures Constructed by Layer-by-Layer Assembly for Highly Efficient and Magnetically Recyclable Dye Removal. *ACS Sustainable Chemistry & Engineering* **2018**, *6* (1), 1279-1288.
100. Guo, H.; Jiao, T.; Zhang, Q.; Guo, W.; Peng, Q.; Yan, X., Preparation of Graphene Oxide-Based Hydrogels as Efficient Dye Adsorbents for Wastewater Treatment. *Nanoscale Research Letters* **2015**, *10* (1), 272.
101. Sun, T.; Zhang, Z.; Xiao, J.; Chen, C.; Xiao, F.; Wang, S.; Liu, Y., Facile and Green Synthesis of Palladium Nanoparticles-Graphene-Carbon Nanotube Material with High Catalytic Activity. *Scientific Reports* **2013**, *3*, 2527.
102. Zhang, Z.; Sun, T.; Chen, C.; Xiao, F.; Gong, Z.; Wang, S., Bifunctional Nanocatalyst Based on Three-Dimensional Carbon Nanotube–Graphene Hydrogel Supported Pd Nanoparticles: One-Pot Synthesis and Its Catalytic Properties. *ACS Applied Materials & Interfaces* **2014**, *6* (23), 21035-21040.
103. Rogers, J. A.; Someya, T.; Huang, Y., Materials and Mechanics for Stretchable Electronics. *Science* **2010**, *327* (5973), 1603-1607.
104. Kim, D.-H.; Song, J.; Choi, W. M.; Kim, H.-S.; Kim, R.-H.; Liu, Z.; Huang, Y. Y.; Hwang, K.-C.; Zhang, Y.-w.; Rogers, J. A., Materials and noncoplanar mesh designs for integrated circuits with linear elastic responses to extreme mechanical deformations. *Proceedings of the National Academy of Sciences* **2008**, *105* (48), 18675-18680.
105. Bae, S.; Kim, H.; Lee, Y.; Xu, X.; Park, J.-S.; Zheng, Y.; Balakrishnan, J.; Lei, T.; Ri Kim, H.; Song, Y. I.; Kim, Y.-J.; Kim, K. S.; Ozyilmaz, B.; Ahn, J.-H.; Hong, B. H.; Iijima, S., Roll-to-roll production of 30-inch graphene films for transparent electrodes. *Nat Nano* **2010**, *5* (8), 574-578.
106. Gelinck, G. H.; Huitema, H. E. A.; van Veenendaal, E.; Cantatore, E.; Schrijnemakers, L.; van der Putten, J. B. P. H.; Geuns, T. C. T.; Beenhakkers, M.; Giesbers, J. B.; Huisman, B.-H.; Meijer, E. J.; Benito, E. M.; Touwslager, F. J.; Marsman, A. W.; van Rens, B. J. E.; de Leeuw, D. M., Flexible active-matrix displays and shift registers based on solution-processed organic transistors. *Nat Mater* **2004**, *3* (2), 106-110.
107. Rogers, J. A.; Bao, Z.; Baldwin, K.; Dodabalapur, A.; Crone, B.; Raju, V. R.; Kuck, V.; Katz, H.; Amundson, K.; Ewing, J.; Drzaic, P., Paper-like electronic displays: Large-area rubber-stamped plastic sheets of electronics and microencapsulated electrophoretic inks. *Proceedings of the National Academy of Sciences* **2001**, *98* (9), 4835-4840.
108. Hammock, M. L.; Chortos, A.; Tee, B. C. K.; Tok, J. B. H.; Bao, Z., 25th Anniversary Article: The Evolution of Electronic Skin (E-Skin): A Brief History, Design Considerations, and Recent Progress. *Advanced Materials* **2013**, *25* (42), 5997-6038.
109. Shao, Y.; El-Kady, M. F.; Wang, L. J.; Zhang, Q.; Li, Y.; Wang, H.; Mousavi, M. F.; Kaner, R. B., Graphene-based materials for flexible supercapacitors. *Chemical Society Reviews* **2015**, *44* (11), 3639-3665.
110. Simon, P.; Gogotsi, Y., Materials for electrochemical capacitors. *Nat Mater* **2008**, *7* (11), 845-854.

111. Peng, X.; Peng, L.; Wu, C.; Xie, Y., Two dimensional nanomaterials for flexible supercapacitors. *Chemical Society Reviews* **2014**, *43* (10), 3303-3323.
112. Winter, M.; Brodd, R. J., What Are Batteries, Fuel Cells, and Supercapacitors? (Chem. Rev. 2003, 104, 4245–4269. Published on the Web 09/28/2004.). *Chemical Reviews* **2005**, *105* (3), 1021-1021.
113. Xu, D.; Xu, Q.; Wang, K.; Chen, J.; Chen, Z., Fabrication of Free-Standing Hierarchical Carbon Nanofiber/Graphene Oxide/Polyaniline Films for Supercapacitors. *ACS Applied Materials & Interfaces* **2014**, *6* (1), 200-209.
114. Lu, X.; Wang, G.; Zhai, T.; Yu, M.; Xie, S.; Ling, Y.; Liang, C.; Tong, Y.; Li, Y., Stabilized TiN Nanowire Arrays for High-Performance and Flexible Supercapacitors. *Nano Letters* **2012**, *12* (10), 5376-5381.
115. Lu, X.; Yu, M.; Wang, G.; Zhai, T.; Xie, S.; Ling, Y.; Tong, Y.; Li, Y., H-TiO₂@MnO₂//H-TiO₂@C Core–Shell Nanowires for High Performance and Flexible Asymmetric Supercapacitors. *Advanced Materials* **2013**, *25* (2), 267-272.
116. Wang, G.; Lu, X.; Ling, Y.; Zhai, T.; Wang, H.; Tong, Y.; Li, Y., LiCl/PVA Gel Electrolyte Stabilizes Vanadium Oxide Nanowire Electrodes for Pseudocapacitors. *ACS Nano* **2012**, *6* (11), 10296-10302.
117. Yang, X.; Zhang, F.; Zhang, L.; Zhang, T.; Huang, Y.; Chen, Y., A High-Performance Graphene Oxide-Doped Ion Gel as Gel Polymer Electrolyte for All-Solid-State Supercapacitor Applications. *Advanced Functional Materials* **2013**, *23* (26), 3353-3360.
118. Beidaghi, M.; Gogotsi, Y., Capacitive energy storage in micro-scale devices: recent advances in design and fabrication of micro-supercapacitors. *Energy & Environmental Science* **2014**, *7* (3), 867-884.
119. Yu, D.; Qian, Q.; Wei, L.; Jiang, W.; Goh, K.; Wei, J.; Zhang, J.; Chen, Y., Emergence of fiber supercapacitors. *Chemical Society Reviews* **2015**, *44* (3), 647-662.
120. Zhang, L. L.; Zhao, X. S., Carbon-based materials as supercapacitor electrodes. *Chemical Society Reviews* **2009**, *38* (9), 2520-2531.
121. Jost, K.; Perez, C. R.; McDonough, J. K.; Presser, V.; Heon, M.; Dion, G.; Gogotsi, Y., Carbon coated textiles for flexible energy storage. *Energy & Environmental Science* **2011**, *4* (12), 5060-5067.
122. Hu, L.; Pasta, M.; La Mantia, F.; Cui, L.; Jeong, S.; Deshazer, H. D.; Choi, J. W.; Han, S. M.; Cui, Y., Stretchable, Porous, and Conductive Energy Textiles. *Nano Letters* **2010**, *10* (2), 708-714.
123. Chen, P.-C.; Shen, G.; Shi, Y.; Chen, H.; Zhou, C., Preparation and Characterization of Flexible Asymmetric Supercapacitors Based on Transition-Metal-Oxide Nanowire/Single-Walled Carbon Nanotube Hybrid Thin-Film Electrodes. *ACS Nano* **2010**, *4* (8), 4403-4411.
124. Kaempgen, M.; Chan, C. K.; Ma, J.; Cui, Y.; Gruner, G., Printable Thin Film Supercapacitors Using Single-Walled Carbon Nanotubes. *Nano Letters* **2009**, *9* (5), 1872-1876.
125. El-Kady, M. F.; Strong, V.; Dubin, S.; Kaner, R. B., Laser Scribing of High-Performance and Flexible Graphene-Based Electrochemical Capacitors. *Science* **2012**, *335* (6074), 1326-1330.
126. Zhang, L. L.; Zhao, X.; Stoller, M. D.; Zhu, Y.; Ji, H.; Murali, S.; Wu, Y.; Perales, S.; Clevenger, B.; Ruoff, R. S., Highly Conductive and Porous Activated Reduced Graphene Oxide Films for High-Power Supercapacitors. *Nano Letters* **2012**, *12* (4), 1806-1812.
127. Xu, Y.; Lin, Z.; Huang, X.; Liu, Y.; Huang, Y.; Duan, X., Flexible Solid-State Supercapacitors Based on Three-Dimensional Graphene Hydrogel Films. *ACS Nano* **2013**, *7* (5), 4042-4049.
128. Wu, D.; Zhang, F.; Liang, H.; Feng, X., Nanocomposites and macroscopic materials: assembly of chemically modified graphene sheets. *Chemical Society Reviews* **2012**, *41* (18), 6160-6177.
129. Raccichini, R.; Varzi, A.; Passerini, S.; Scrosati, B., The role of graphene for electrochemical energy storage. *Nat Mater* **2015**, *14* (3), 271-279.
130. Liu, C.; Yu, Z.; Neff, D.; Zhamu, A.; Jang, B. Z., Graphene-Based Supercapacitor with an Ultrahigh Energy Density. *Nano Letters* **2010**, *10* (12), 4863-4868.
131. Yoo, J. J.; Balakrishnan, K.; Huang, J.; Meunier, V.; Sumpter, B. G.; Srivastava, A.; Conway, M.; Mohana Reddy, A. L.; Yu, J.; Vajtai, R.; Ajayan, P. M., Ultrathin Planar Graphene Supercapacitors. *Nano Letters* **2011**, *11* (4), 1423-1427.

132. Yu, G.; Hu, L.; Liu, N.; Wang, H.; Vosgueritchian, M.; Yang, Y.; Cui, Y.; Bao, Z., Enhancing the Supercapacitor Performance of Graphene/MnO₂ Nanostructured Electrodes by Conductive Wrapping. *Nano Letters* **2011**, *11* (10), 4438-4442.
133. Huang, L.; Chen, D.; Ding, Y.; Feng, S.; Wang, Z. L.; Liu, M., Nickel–Cobalt Hydroxide Nanosheets Coated on NiCo₂O₄ Nanowires Grown on Carbon Fiber Paper for High-Performance Pseudocapacitors. *Nano Letters* **2013**, *13* (7), 3135-3139.
134. Zhang, J.; Zhao, X. S., Conducting Polymers Directly Coated on Reduced Graphene Oxide Sheets as High-Performance Supercapacitor Electrodes. *The Journal of Physical Chemistry C* **2012**, *116* (9), 5420-5426.
135. Liu, Y.; Weng, B.; Razal, J. M.; Xu, Q.; Zhao, C.; Hou, Y.; Seyedin, S.; Jalili, R.; Wallace, G. G.; Chen, J., High-Performance Flexible All-Solid-State Supercapacitor from Large Free-Standing Graphene-PEDOT/PSS Films. *Scientific Reports* **2015**, *5*, 17045.
136. Rauda, I. E.; Augustyn, V.; Dunn, B.; Tolbert, S. H., Enhancing Pseudocapacitive Charge Storage in Polymer Templated Mesoporous Materials. *Accounts of Chemical Research* **2013**, *46* (5), 1113-1124.
137. Wang, D.-W.; Li, F.; Zhao, J.; Ren, W.; Chen, Z.-G.; Tan, J.; Wu, Z.-S.; Gentle, I.; Lu, G. Q.; Cheng, H.-M., Fabrication of Graphene/Polyaniline Composite Paper via In Situ Anodic Electropolymerization for High-Performance Flexible Electrode. *ACS Nano* **2009**, *3* (7), 1745-1752.
138. Cao, X.; Shi, Y.; Shi, W.; Lu, G.; Huang, X.; Yan, Q.; Zhang, Q.; Zhang, H., Preparation of Novel 3D Graphene Networks for Supercapacitor Applications. *Small* **2011**, *7* (22), 3163-3168.
139. Yu, G.; Hu, L.; Vosgueritchian, M.; Wang, H.; Xie, X.; McDonough, J. R.; Cui, X.; Cui, Y.; Bao, Z., Solution-Processed Graphene/MnO₂ Nanostructured Textiles for High-Performance Electrochemical Capacitors. *Nano Letters* **2011**, *11* (7), 2905-2911.
140. Wang, Z.; Tammela, P.; Zhang, P.; Huo, J.; Ericson, F.; Stromme, M.; Nyholm, L., Freestanding nanocellulose-composite fibre reinforced 3D polypyrrole electrodes for energy storage applications. *Nanoscale* **2014**, *6* (21), 13068-13075.
141. Zhang, Y.-Z.; Wang, Y.; Cheng, T.; Lai, W.-Y.; Pang, H.; Huang, W., Flexible supercapacitors based on paper substrates: a new paradigm for low-cost energy storage. *Chemical Society Reviews* **2015**, *44* (15), 5181-5199.
142. Gui, Z.; Zhu, H.; Gillette, E.; Han, X.; Rubloff, G. W.; Hu, L.; Lee, S. B., Natural Cellulose Fiber as Substrate for Supercapacitor. *ACS Nano* **2013**, *7* (7), 6037-6046.
143. Hu, L.; Choi, J. W.; Yang, Y.; Jeong, S.; La Mantia, F.; Cui, L.-F.; Cui, Y., Highly conductive paper for energy-storage devices. *Proceedings of the National Academy of Sciences* **2009**, *106* (51), 21490-21494.
144. Hu, L.; Wu, H.; Cui, Y., Printed energy storage devices by integration of electrodes and separators into single sheets of paper. *Applied Physics Letters* **2010**, *96* (18), 183502.
145. Pushparaj, V. L.; Shaijumon, M. M.; Kumar, A.; Murugesan, S.; Ci, L.; Vajtai, R.; Linhardt, R. J.; Nalamasu, O.; Ajayan, P. M., Flexible energy storage devices based on nanocomposite paper. *Proceedings of the National Academy of Sciences* **2007**, *104* (34), 13574-13577.
146. Nishide, H.; Oyaizu, K., Toward Flexible Batteries. *Science* **2008**, *319* (5864), 737-738.
147. Jiang, Q.; Tian, L.; Liu, K.-K.; Tadepalli, S.; Raliya, R.; Biswas, P.; Naik, R. R.; Singamaneni, S., Bilayered Biofoam for Highly Efficient Solar Steam Generation. *Advanced Materials* **2016**, *28* (42), 9400-9407.
148. Tian, L.; Jiang, Q.; Liu, K.-K.; Luan, J.; Naik, R. R.; Singamaneni, S., Bacterial Nanocellulose-Based Flexible Surface Enhanced Raman Scattering Substrate. *Advanced Materials Interfaces* **2016**, *3* (15), 1600214-n/a.
149. Kang, Y. J.; Chun, S.-J.; Lee, S.-S.; Kim, B.-Y.; Kim, J. H.; Chung, H.; Lee, S.-Y.; Kim, W., All-Solid-State Flexible Supercapacitors Fabricated with Bacterial Nanocellulose Papers, Carbon Nanotubes, and Triblock-Copolymer Ion Gels. *ACS Nano* **2012**, *6* (7), 6400-6406.

150. Chen, L.-F.; Huang, Z.-H.; Liang, H.-W.; Guan, Q.-F.; Yu, S.-H., Bacterial-Cellulose-Derived Carbon Nanofiber@MnO₂ and Nitrogen-Doped Carbon Nanofiber Electrode Materials: An Asymmetric Supercapacitor with High Energy and Power Density. *Advanced Materials* **2013**, *25* (34), 4746-4752.
151. Wang, H.; Zhu, E.; Yang, J.; Zhou, P.; Sun, D.; Tang, W., Bacterial Cellulose Nanofiber-Supported Polyaniline Nanocomposites with Flake-Shaped Morphology as Supercapacitor Electrodes. *The Journal of Physical Chemistry C* **2012**, *116* (24), 13013-13019.
152. Li, S.; Huang, D.; Zhang, B.; Xu, X.; Wang, M.; Yang, G.; Shen, Y., Flexible Supercapacitors Based on Bacterial Cellulose Paper Electrodes. *Advanced Energy Materials* **2014**, *4* (10), 1301655-n/a.
153. Liu, R.; Ma, L.; Huang, S.; Mei, J.; Xu, J.; Yuan, G., A flexible polyaniline/graphene/bacterial cellulose supercapacitor electrode. *New Journal of Chemistry* **2017**, *41* (2), 857-864.
154. Wang, F.; Kim, H.-J.; Park, S.; Kee, C.-D.; Kim, S.-J.; Oh, I.-K., Bendable and flexible supercapacitor based on polypyrrole-coated bacterial cellulose core-shell composite network. *Composites Science and Technology* **2016**, *128*, 33-40.
155. Wang, X.; Kong, D.; Zhang, Y.; Wang, B.; Li, X.; Qiu, T.; Song, Q.; Ning, J.; Song, Y.; Zhi, L., All-biomaterial supercapacitor derived from bacterial cellulose. *Nanoscale* **2016**, *8* (17), 9146-9150.
156. Wang, X.; Kong, D.; Wang, B.; Song, Y.; Zhi, L., Activated pyrolysed bacterial cellulose as electrodes for supercapacitors. *Science China Chemistry* **2016**, *59* (6), 713-718.
157. Martinez, A. W.; Phillips, S. T.; Butte, M. J.; Whitesides, G. M., Patterned paper as a platform for inexpensive, low-volume, portable bioassays. *Angew Chem Int Edit* **2007**, *46* (8), 1318-1320.
158. Zhao, W. A.; van den Berg, A., Lab on paper. *Lab Chip* **2008**, *8* (12), 1988-1991.
159. Cheng, C. M.; Martinez, A. W.; Gong, J. L.; Mace, C. R.; Phillips, S. T.; Carrilho, E.; Mirica, K. A.; Whitesides, G. M., Paper-Based ELISA. *Angew Chem Int Edit* **2010**, *49* (28), 4771-4774.
160. Derda, R.; Laromaine, A.; Mammoto, A.; Tang, S. K. Y.; Mammoto, T.; Ingber, D. E.; Whitesides, G. M., Paper-supported 3D cell culture for tissue-based bioassays. *P Natl Acad Sci USA* **2009**, *106* (44), 18457-18462.
161. Pardee, K.; Green, A. A.; Ferrante, T.; Cameron, D. E.; DaleyKeyser, A.; Yin, P.; Collins, J. J., Paper-Based Synthetic Gene Networks. *Cell* **2014**, *159* (4), 940-954.
162. Zhu, H.; Fang, Z.; Preston, C.; Li, Y.; Hu, L., Transparent paper: fabrications, properties, and device applications. *Energy & Environmental Science* **2014**, *7* (1), 269-287.
163. Lee, C. H.; Hankus, M. E.; Tian, L.; Pellegrino, P. M.; Singamaneni, S., Highly Sensitive Surface Enhanced Raman Scattering Substrates Based on Filter Paper Loaded with Plasmonic Nanostructures. *Analytical Chemistry* **2011**, *83* (23), 8953-8958.
164. Abbas, A.; Brimer, A.; Slocik, J. M.; Tian, L.; Naik, R. R.; Singamaneni, S., Multifunctional Analytical Platform on a Paper Strip: Separation, Preconcentration, and Subattomolar Detection. *Analytical Chemistry* **2013**, *85* (8), 3977-3983.
165. Jaiswal, A.; Tian, L. M.; Tadepalli, S.; Liu, K. K.; Fei, M.; Farrell, M. E.; Pellegrino, P. M.; Singamaneni, S., Plasmonic Nanorattles with Intrinsic Electromagnetic Hot-Spots for Surface Enhanced Raman Scattering. *Small* **2014**, *10* (21), 4287-4292.
166. Tian, L.; Luan, J.; Liu, K.-K.; Jiang, Q.; Tadepalli, S.; Gupta, M. K.; Naik, R. R.; Singamaneni, S., Plasmonic Biofoam: A Versatile Optically Active Material. *Nano Letters* **2015**.
167. Orendorff, C. J.; Murphy, C. J., Quantitation of Metal Content in the Silver-Assisted Growth of Gold Nanorods. *J. Phys. Chem. B* **2006**, *110* (9), 3990-3994.
168. Huang, X.; Neretina, S.; El-Sayed, M. A., Gold Nanorods: From Synthesis and Properties to Biological and Biomedical Applications. *Adv. Mater.* **2009**, *21* (48), 4880-4910.
169. Sengupta, A.; Mujacic, M.; Davis, E. J., Detection of bacteria by surface-enhanced Raman spectroscopy. *Anal Bioanal Chem* **2006**, *386* (5), 1379-1386.
170. Cannon, R. E.; Anderson, S. M., Biogenesis of Bacterial Cellulose. *Crit Rev Microbiol* **1991**, *17* (6), 435-447.

171. Klemm, D.; Schumann, D.; Udhardt, U.; Marsch, S., Bacterial synthesized cellulose — artificial blood vessels for microsurgery. *Progress in Polymer Science* **2001**, *26* (9), 1561-1603.
172. Jonas, R.; Farah, L. F., Production and application of microbial cellulose. *Polym Degrad Stabil* **1998**, *59* (1-3), 101-106.
173. Huang, X. H.; Neretina, S.; El-Sayed, M. A., Gold Nanorods: From Synthesis and Properties to Biological and Biomedical Applications. *Adv Mater* **2009**, *21* (48), 4880-4910.
174. Tian, L.; Chen, E.; Gandra, N.; Abbas, A.; Singamaneni, S., Gold Nanorods as Plasmonic Nanotransducers: Distance-Dependent Refractive Index Sensitivity. *Langmuir* **2012**, *28* (50), 17435-17442.
175. Abbas, A.; Tian, L.; Morrissey, J. J.; Kharasch, E. D.; Singamaneni, S., Hot Spot-Localized Artificial Antibodies for Label-Free Plasmonic Biosensing. *Advanced Functional Materials* **2013**, *23* (14), 1789-1797.
176. Wang, Y.; Heim, L.-O.; Xu, Y.; Buntkowsky, G.; Zhang, K., Transparent, Stimuli-Responsive Films from Cellulose-Based Organogel Nanoparticles. *Advanced Functional Materials* **2015**, *25* (9), 1434-1441.
177. El-Sayed, M. A., Some Interesting Properties of Metals Confined in Time and Nanometer Space of Different Shapes. *Accounts of Chemical Research* **2001**, *34* (4), 257-264.
178. Jain, P. K.; El-Sayed, M. A., Noble Metal Nanoparticle Pairs: Effect of Medium for Enhanced Nanosensing. *Nano Letters* **2008**, *8* (12), 4347-4352.
179. Martinez, A. W.; Phillips, S. T.; Butte, M. J.; Whitesides, G. M., Patterned Paper as a Platform for Inexpensive, Low-Volume, Portable Bioassays. *Angewandte Chemie International Edition* **2007**, *46* (8), 1318-1320.
180. Habibi, Y.; Lucia, L. A.; Rojas, O. J., Cellulose Nanocrystals: Chemistry, Self-Assembly, and Applications. *Chemical Reviews* **2010**, *110* (6), 3479-3500.
181. Schmucker, A. L.; Tadepalli, S.; Liu, K.-K.; Sullivan, C. J.; Singamaneni, S.; Naik, R. R., Plasmonic paper: a porous and flexible substrate enabling nanoparticle-based combinatorial chemistry. *RSC Advances* **2016**, *6* (5), 4136-4144.
182. Vosgröne, T.; Meixner, A. J., Surface- and Resonance-Enhanced Micro-Raman Spectroscopy of Xanthene Dyes: From the Ensemble to Single Molecules. *ChemPhysChem* **2005**, *6* (1), 154-163.
183. Tian, L.; Tadepalli, S.; Farrell, M. E.; Liu, K.-K.; Gandra, N.; Pellegrino, P. M.; Singamaneni, S., Multiplexed charge-selective surface enhanced Raman scattering based on plasmonic calligraphy. *Journal of Materials Chemistry C* **2014**, *2* (27), 5438-5446.
184. Stiles, P. L.; Dieringer, J. A.; Shah, N. C.; Van Duyne, R. P., Surface-Enhanced Raman Spectroscopy. *Annual Review of Analytical Chemistry* **2008**, *1* (1), 601-626.
185. Fan, C.; Hu, Z.; Mustapha, A.; Lin, M., Rapid detection of food- and waterborne bacteria using surface-enhanced Raman spectroscopy coupled with silver nanosubstrates. *Appl Microbiol Biotechnol* **2011**, *92* (5), 1053-1061.
186. Zhou, Q.; He, Y.; Abell, J.; Zhang, Z.; Zhao, Y., Surface-enhanced Raman scattering from helical silver nanorod arrays. *Chemical Communications* **2011**, *47* (15), 4466-4468.
187. Walter, A.; Marz, A.; Schumacher, W.; Rosch, P.; Popp, J., Towards a fast, high specific and reliable discrimination of bacteria on strain level by means of SERS in a microfluidic device. *Lab Chip* **2011**, *11* (6), 1013-1021.
188. Zhou, H.; Yang, D.; Ivleva, N. P.; Mircescu, N. E.; Niessner, R.; Haisch, C., SERS Detection of Bacteria in Water by in Situ Coating with Ag Nanoparticles. *Analytical Chemistry* **2014**, *86* (3), 1525-1533.
189. Neugebauer, U.; Rösch, P.; Schmitt, M.; Popp, J.; Julien, C.; Rasmussen, A.; Budich, C.; Deckert, V., On the Way to Nanometer-Sized Information of the Bacterial Surface by Tip-Enhanced Raman Spectroscopy. *ChemPhysChem* **2006**, *7* (7), 1428-1430.
190. Chu, H.; Huang, Y.; Zhao, Y., Silver Nanorod Arrays as a Surface-Enhanced Raman Scattering Substrate for Foodborne Pathogenic Bacteria Detection. *Appl. Spectrosc.* **2008**, *62* (8), 922-931.
191. Chung, C.; Kim, Y.-K.; Shin, D.; Ryoo, S.-R.; Hong, B. H.; Min, D.-H., Biomedical Applications of Graphene and Graphene Oxide. *Accounts of Chemical Research* **2013**, *46* (10), 2211-2224.

192. Gonçalves, G.; Vila, M.; Portolés, M.-T.; Vallet-Regí, M.; Gracio, J.; Marques, P. A. A. P., Nano-Graphene Oxide: A Potential Multifunctional Platform for Cancer Therapy. *Advanced Healthcare Materials* **2013**, *2* (8), 1072-1090.
193. Nergiz, S. Z.; Gandra, N.; Tadepalli, S.; Singamaneni, S., Multifunctional Hybrid Nanopatches of Graphene Oxide and Gold Nanostars for Ultraefficient Photothermal Cancer Therapy. *ACS Applied Materials & Interfaces* **2014**, *6* (18), 16395-16402.
194. Yang, K.; Zhang, S.; Zhang, G.; Sun, X.; Lee, S.-T.; Liu, Z., Graphene in Mice: Ultrahigh In Vivo Tumor Uptake and Efficient Photothermal Therapy. *Nano Letters* **2010**, *10* (9), 3318-3323.
195. Robinson, J. T.; Tabakman, S. M.; Liang, Y.; Wang, H.; Sanchez Casalongue, H.; Vinh, D.; Dai, H., Ultrasmall Reduced Graphene Oxide with High Near-Infrared Absorbance for Photothermal Therapy. *Journal of the American Chemical Society* **2011**, *133* (17), 6825-6831.
196. Wang, W.-N.; Jiang, Y.; Biswas, P., Evaporation-Induced Crumpling of Graphene Oxide Nanosheets in Aerosolized Droplets: Confinement Force Relationship. *The Journal of Physical Chemistry Letters* **2012**, *3* (21), 3228-3233.
197. Jiang, Y.; Wang, W.-N.; Biswas, P.; Fortner, J. D., Facile Aerosol Synthesis and Characterization of Ternary Crumpled Graphene-TiO₂-Magnetite Nanocomposites for Advanced Water Treatment. *ACS Applied Materials & Interfaces* **2014**, *6* (14), 11766-11774.
198. Zhu, Y.; Murali, S.; Cai, W.; Li, X.; Suk, J. W.; Potts, J. R.; Ruoff, R. S., Graphene and Graphene Oxide: Synthesis, Properties, and Applications. *Advanced Materials* **2010**, *22* (35), 3906-3924.
199. Yin, Z.; Sun, S.; Salim, T.; Wu, S.; Huang, X.; He, Q.; Lam, Y. M.; Zhang, H., Organic Photovoltaic Devices Using Highly Flexible Reduced Graphene Oxide Films as Transparent Electrodes. *ACS Nano* **2010**, *4* (9), 5263-5268.
200. Kumar, P. V.; Bardhan, N. M.; Tongay, S.; Wu, J.; Belcher, A. M.; Grossman, J. C., Scalable enhancement of graphene oxide properties by thermally driven phase transformation. *Nat Chem* **2014**, *6* (2), 151-158.
201. Gatenholm, P.; Klemm, D., Bacterial Nanocellulose as a Renewable Material for Biomedical Applications. *MRS Bulletin* **2010**, *35* (03), 208-213.
202. Marcano, D. C.; Kosynkin, D. V.; Berlin, J. M.; Sinitskii, A.; Sun, Z.; Slesarev, A.; Alemany, L. B.; Lu, W.; Tour, J. M., Improved Synthesis of Graphene Oxide. *ACS Nano* **2010**, *4* (8), 4806-4814.
203. Poletto, M.; Pistor, V.; Santana, R. M. C.; Zattera, A. J., Materials produced from plant biomass: part II: evaluation of crystallinity and degradation kinetics of cellulose. *Materials Research* **2012**, *15*, 421-427.
204. Rourke, J. P.; Pandey, P. A.; Moore, J. J.; Bates, M.; Kinloch, I. A.; Young, R. J.; Wilson, N. R., The Real Graphene Oxide Revealed: Stripping the Oxidative Debris from the Graphene-like Sheets. *Angewandte Chemie International Edition* **2011**, *50* (14), 3173-3177.
205. Ghasemi, H.; Ni, G.; Marconnet, A. M.; Loomis, J.; Yerci, S.; Miljkovic, N.; Chen, G., Solar steam generation by heat localization. *Nat Commun* **2014**, *5*.
206. Kulkarni, D. D.; Choi, I.; Singamaneni, S. S.; Tsukruk, V. V., Graphene Oxide-Polyelectrolyte Nanomembranes. *ACS Nano* **2010**, *4* (8), 4667-4676.
207. Yang, D.; Velamakanni, A.; Bozoklu, G.; Park, S.; Stoller, M.; Piner, R. D.; Stankovich, S.; Jung, I.; Field, D. A.; Ventrice Jr, C. A.; Ruoff, R. S., Chemical analysis of graphene oxide films after heat and chemical treatments by X-ray photoelectron and Micro-Raman spectroscopy. *Carbon* **2009**, *47* (1), 145-152.
208. Hu, K.; Tolentino, L. S.; Kulkarni, D. D.; Ye, C.; Kumar, S.; Tsukruk, V. V., Written-in Conductive Patterns on Robust Graphene Oxide Biopaper by Electrochemical Microstamping. *Angewandte Chemie International Edition* **2013**, *52* (51), 13784-13788.
209. Jung, R.; Kim, H.-S.; Kim, Y.; Kwon, S.-M.; Lee, H. S.; Jin, H.-J., Electrically conductive transparent papers using multiwalled carbon nanotubes. *Journal of Polymer Science Part B: Polymer Physics* **2008**, *46* (12), 1235-1242.

210. Hu, W.; Chen, S.; Yang, Z.; Liu, L.; Wang, H., Flexible Electrically Conductive Nanocomposite Membrane Based on Bacterial Cellulose and Polyaniline. *The Journal of Physical Chemistry B* **2011**, *115* (26), 8453-8457.
211. Nandgaonkar, A. G.; Wang, Q.; Fu, K.; Krause, W. E.; Wei, Q.; Gorga, R.; A. Lucia, L., A one-pot biosynthesis of reduced graphene oxide (RGO)/bacterial cellulose (BC) nanocomposites. *Green Chemistry* **2014**, *16* (6), 3195-3201.
212. Kulkarni, D. D.; Kim, S.; Chyasnavichyus, M.; Hu, K.; Fedorov, A. G.; Tsukruk, V. V., Chemical Reduction of Individual Graphene Oxide Sheets as Revealed by Electrostatic Force Microscopy. *Journal of the American Chemical Society* **2014**, *136* (18), 6546-6549.
213. Acik, M.; Lee, G.; Mattevi, C.; Pirkle, A.; Wallace, R. M.; Chhowalla, M.; Cho, K.; Chabal, Y., The Role of Oxygen during Thermal Reduction of Graphene Oxide Studied by Infrared Absorption Spectroscopy. *The Journal of Physical Chemistry C* **2011**, *115* (40), 19761-19781.
214. Thomas, H. R.; Day, S. P.; Woodruff, W. E.; Vallés, C.; Young, R. J.; Kinloch, I. A.; Morley, G. W.; Hanna, J. V.; Wilson, N. R.; Rourke, J. P., Deoxygenation of Graphene Oxide: Reduction or Cleaning? *Chemistry of Materials* **2013**, *25* (18), 3580-3588.
215. Ramires, M. L. V.; Nieto de Castro, C. A.; Nagasaka, Y.; Nagashima, A.; Assael, M. J.; Wakeham, W. A., Standard Reference Data for the Thermal Conductivity of Water. *Journal of Physical and Chemical Reference Data* **1995**, *24* (3), 1377-1381.
216. Liu, Y.; Yu, S.; Feng, R.; Bernard, A.; Liu, Y.; Zhang, Y.; Duan, H.; Shang, W.; Tao, P.; Song, C.; Deng, T., A Bioinspired, Reusable, Paper-Based System for High-Performance Large-Scale Evaporation. *Advanced Materials* **2015**, *27* (17), 2768-2774.
217. Gratzel, M., Photoelectrochemical cells. *Nature* **2001**, *414* (6861), 338-344.
218. Van de Ven, J. D., Mobile hydraulic power supply: Liquid piston Stirling engine pump. *Renewable Energy* **2009**, *34* (11), 2317-2322.
219. Gupta, M. K.; Kaushik, S. C., Exergy analysis and investigation for various feed water heaters of direct steam generation solar-thermal power plant. *Renewable Energy* **2010**, *35* (6), 1228-1235.
220. Zarza, E.; Valenzuela, L.; León, J.; Henneke, K.; Eck, M.; Weyers, H. D.; Eickhoff, M., Direct steam generation in parabolic troughs: Final results and conclusions of the DISS project. *Energy* **2004**, *29* (5-6), 635-644.
221. Elimelech, M.; Phillip, W. A., The Future of Seawater Desalination: Energy, Technology, and the Environment. *Science* **2011**, *333* (6043), 712-717.
222. Karagiannis, I. C.; Soldatos, P. G., Water desalination cost literature: review and assessment. *Desalination* **2008**, *223* (1-3), 448-456.
223. Neumann, O.; Urban, A. S.; Day, J.; Lal, S.; Nordlander, P.; Halas, N. J., Solar Vapor Generation Enabled by Nanoparticles. *ACS Nano* **2013**, *7* (1), 42-49.
224. Shang, W.; Deng, T., Solar steam generation: Steam by thermal concentration. *Nature Energy* **2016**, *1*, 16133.
225. Hu, X.; Xu, W.; Zhou, L.; Tan, Y.; Wang, Y.; Zhu, S.; Zhu, J., Tailoring Graphene Oxide-Based Aerogels for Efficient Solar Steam Generation under One Sun. *Advanced Materials* **2017**, *29* (5), 1604031-n/a.
226. Ito, Y.; Tanabe, Y.; Han, J.; Fujita, T.; Tanigaki, K.; Chen, M., Multifunctional Porous Graphene for High-Efficiency Steam Generation by Heat Localization. *Advanced Materials* **2015**, *27* (29), 4302-4307.
227. Zhou, L.; Tan, Y.; Wang, J.; Xu, W.; Yuan, Y.; Cai, W.; Zhu, S.; Zhu, J., 3D self-assembly of aluminium nanoparticles for plasmon-enhanced solar desalination. *Nat Photon* **2016**, *10* (6), 393-398.
228. Liu, K.-K.; Jiang, Q.; Tadepalli, S.; Raliya, R.; Biswas, P.; Naik, R. R.; Singamaneni, S., Wood-Graphene Oxide Composite for Highly Efficient Solar Steam Generation and Desalination. *ACS Applied Materials & Interfaces* **2017**, *9* (8), 7675-7681.
229. Zhang, P.; Li, J.; Lv, L.; Zhao, Y.; Qu, L., Vertically Aligned Graphene Sheets Membrane for Highly Efficient Solar Thermal Generation of Clean Water. *ACS Nano* **2017**.

230. Li, R.; Zhang, L.; Shi, L.; Wang, P., MXene Ti₃C₂: An Effective 2D Light-to-Heat Conversion Material. *ACS Nano* **2017**, *11* (4), 3752-3759.
231. Li, Y.; Gao, T.; Yang, Z.; Chen, C.; Luo, W.; Song, J.; Hitz, E.; Jia, C.; Zhou, Y.; Liu, B.; Yang, B.; Hu, L., 3D-Printed, All-in-One Evaporator for High-Efficiency Solar Steam Generation under 1 Sun Illumination. *Advanced Materials*, 1700981-n/a.
232. Li, X.; Xu, W.; Tang, M.; Zhou, L.; Zhu, B.; Zhu, S.; Zhu, J., Graphene oxide-based efficient and scalable solar desalination under one sun with a confined 2D water path. *Proceedings of the National Academy of Sciences* **2016**, *113* (49), 13953-13958.
233. Zhou, L.; Tan, Y.; Ji, D.; Zhu, B.; Zhang, P.; Xu, J.; Gan, Q.; Yu, Z.; Zhu, J., Self-assembly of highly efficient, broadband plasmonic absorbers for solar steam generation. *Science Advances* **2016**, *2* (4).
234. Wang, J.; Li, Y.; Deng, L.; Wei, N.; Weng, Y.; Dong, S.; Qi, D.; Qiu, J.; Chen, X.; Wu, T., High-Performance Photothermal Conversion of Narrow-Bandgap Ti₂O₃ Nanoparticles. *Advanced Materials* **2017**, *29* (3), 1603730-n/a.
235. Ni, G.; Li, G.; Boriskina, Svetlana V.; Li, H.; Yang, W.; Zhang, T.; Chen, G., Steam generation under one sun enabled by a floating structure with thermal concentration. *Nature Energy* **2016**, *1*, 16126.
236. Zhang, L.; Tang, B.; Wu, J.; Li, R.; Wang, P., Hydrophobic Light-to-Heat Conversion Membranes with Self-Healing Ability for Interfacial Solar Heating. *Advanced Materials* **2015**, *27* (33), 4889-4894.
237. Wang, G.; Fu, Y.; Ma, X.; Pi, W.; Liu, D.; Wang, X., Reusable reduced graphene oxide based double-layer system modified by polyethylenimine for solar steam generation. *Carbon* **2017**, *114*, 117-124.
238. Klaine, S. J.; Alvarez, P. J. J.; Batley, G. E.; Fernandes, T. F.; Handy, R. D.; Lyon, D. Y.; Mahendra, S.; McLaughlin, M. J.; Lead, J. R., Nanomaterials in the environment: Behavior, fate, bioavailability, and effects. *Environmental Toxicology and Chemistry* **2008**, *27* (9), 1825-1851.
239. Mattsson, K.; Hansson, L. A.; Cedervall, T., Nano-plastics in the aquatic environment. *Environmental Science: Processes & Impacts* **2015**, *17* (10), 1712-1721.
240. Ciglenečki, I.; Svetličić, V., Nanoparticles and Marine Environment: An Overview. In *Nanotechnology to Aid Chemical and Biological Defense*, Camesano, T. A., Ed. Springer Netherlands: Dordrecht, 2015; pp 95-111.
241. Wu, B.; Torres-Duarte, C.; Cole, B. J.; Cherr, G. N., Copper Oxide and Zinc Oxide Nanomaterials Act as Inhibitors of Multidrug Resistance Transport in Sea Urchin Embryos: Their Role as Chemosensitizers. *Environmental Science & Technology* **2015**, *49* (9), 5760-5770.
242. Wang, Z.; Zhang, L.; Zhao, J.; Xing, B., Environmental processes and toxicity of metallic nanoparticles in aquatic systems as affected by natural organic matter. *Environmental Science: Nano* **2016**, *3* (2), 240-255.
243. Lee, H.; Dellatore, S. M.; Miller, W. M.; Messersmith, P. B., Mussel-Inspired Surface Chemistry for Multifunctional Coatings. *Science* **2007**, *318* (5849), 426.
244. Liu, Y.; Ai, K.; Lu, L., Polydopamine and Its Derivative Materials: Synthesis and Promising Applications in Energy, Environmental, and Biomedical Fields. *Chemical Reviews* **2014**, *114* (9), 5057-5115.
245. Liebscher, J.; Mrówczyński, R.; Scheidt, H. A.; Filip, C.; Hädade, N. D.; Turcu, R.; Bende, A.; Beck, S., Structure of Polydopamine: A Never-Ending Story? *Langmuir* **2013**, *29* (33), 10539-10548.
246. d'Ischia, M.; Napolitano, A.; Ball, V.; Chen, C.-T.; Buehler, M. J., Polydopamine and Eumelanin: From Structure–Property Relationships to a Unified Tailoring Strategy. *Accounts of Chemical Research* **2014**, *47* (12), 3541-3550.
247. Black, K. C. L.; Yi, J.; Rivera, J. G.; Zelasko-Leon, D. C.; Messersmith, P. B., Polydopamine-enabled surface functionalization of gold nanorods for cancer cell-targeted imaging and photothermal therapy. *Nanomedicine* **2012**, *8* (1), 17-28.
248. Liu, Y.; Ai, K.; Liu, J.; Deng, M.; He, Y.; Lu, L., Dopamine-Melanin Colloidal Nanospheres: An Efficient Near-Infrared Photothermal Therapeutic Agent for In Vivo Cancer Therapy. *Advanced Materials* **2013**, *25* (9), 1353-1359.

249. Stritzker, J.; Kirscher, L.; Scadeng, M.; Deliolanis, N. C.; Morscher, S.; Symvoulidis, P.; Schaefer, K.; Zhang, Q.; Buckel, L.; Hess, M.; Donat, U.; Bradley, W. G.; Ntziachristos, V.; Szalay, A. A., Vaccinia virus-mediated melanin production allows MR and optoacoustic deep tissue imaging and laser-induced thermotherapy of cancer. *Proceedings of the National Academy of Sciences* **2013**, *110* (9), 3316-3320.
250. Carpenter, A. W.; de Lannoy, C.-F.; Wiesner, M. R., Cellulose Nanomaterials in Water Treatment Technologies. *Environmental Science & Technology* **2015**, *49* (9), 5277-5287.
251. Ai, K.; Liu, Y.; Ruan, C.; Lu, L.; Lu, G., Sp² C-Dominant N-Doped Carbon Sub-micrometer Spheres with a Tunable Size: A Versatile Platform for Highly Efficient Oxygen-Reduction Catalysts. *Advanced Materials* **2013**, *25* (7), 998-1003.
252. SIDDIQUI, S. A.; DWIVEDI, A.; SINGH, P. K.; HASAN, T.; JAIN, S.; SUNDARAGANESAN, N.; SALEEM, H.; MISRA, N., VIBRATIONAL DYNAMICS AND POTENTIAL ENERGY DISTRIBUTION OF TWO WELL-KNOWN NEUROTRANSMITTER RECEPTORS: TYRAMINE AND DOPAMINE HYDROCHLORIDE. *Journal of Theoretical and Computational Chemistry* **2009**, *08* (03), 433-450.
253. Cheng, G.; Zheng, S.-Y., Construction of a high-performance magnetic enzyme nanosystem for rapid tryptic digestion. *Scientific Reports* **2014**, *4*, 6947.
254. Xue, G.; Liu, K.; Chen, Q.; Yang, P.; Li, J.; Ding, T.; Duan, J.; Qi, B.; Zhou, J., Robust and Low-Cost Flame-Treated Wood for High-Performance Solar Steam Generation. *ACS Applied Materials & Interfaces* **2017**, *9* (17), 15052-15057.
255. Xu, N.; Hu, X.; Xu, W.; Li, X.; Zhou, L.; Zhu, S.; Zhu, J., Mushrooms as Efficient Solar Steam-Generation Devices. *Advanced Materials*, 1606762-n/a.
256. Jozala, A. F.; de Lencastre-Novaeas, L. C.; Lopes, A. M.; de Carvalho Santos-Ebinuma, V.; Mazzola, P. G.; Pessoa-Jr, A.; Grotto, D.; Gerenutti, M.; Chaud, M. V., Bacterial nanocellulose production and application: a 10-year overview. *Applied Microbiology and Biotechnology* **2016**, *100* (5), 2063-2072.
257. Torres, F.; Commeaux, S.; Troncoso, O., Biocompatibility of Bacterial Cellulose Based Biomaterials. *Journal of Functional Biomaterials* **2012**, *3* (4), 864.
258. Ruka, D. R.; Sangwan, P.; Garvey, C. J.; Simon, G. P.; Dean, K. M., Biodegradability of Poly-3-hydroxybutyrate/Bacterial Cellulose Composites under Aerobic Conditions, Measured via Evolution of Carbon Dioxide and Spectroscopic and Diffraction Methods. *Environmental Science & Technology* **2015**, *49* (16), 9979-9986.
259. Pérez, J.; Muñoz-Dorado, J.; de la Rubia, T.; Martínez, J., Biodegradation and biological treatments of cellulose, hemicellulose and lignin: an overview. *International Microbiology* **2002**, *5* (2), 53-63.
260. Schröpfer, S. B.; Bottene, M. K.; Bianchin, L.; Robinson, L. C.; Lima, V. d.; Jahno, V. D.; Barud, H. d. S.; Ribeiro, S. J. L., Biodegradation evaluation of bacterial cellulose, vegetable cellulose and poly (3-hydroxybutyrate) in soil. *Polímeros* **2015**, *25*, 154-160.
261. Wang, B.; Lv, X.; Chen, S.; Li, Z.; Sun, X.; Feng, C.; Wang, H.; Xu, Y., In vitro biodegradability of bacterial cellulose by cellulase in simulated body fluid and compatibility in vivo. *Cellulose* **2016**, *23* (5), 3187-3198.
262. Wan, Y. Z.; Luo, H.; He, F.; Liang, H.; Huang, Y.; Li, X. L., Mechanical, moisture absorption, and biodegradation behaviours of bacterial cellulose fibre-reinforced starch biocomposites. *Composites Science and Technology* **2009**, *69* (7), 1212-1217.
263. Luther, J. P.; Lipke, H., Degradation of melanin by *Aspergillus fumigatus*. *Applied and Environmental Microbiology* **1980**, *40* (1), 145-155.
264. Khammuang, S.; Sarnthima, R., Decolorization of synthetic melanins by crude laccases of *Lentinus polychrous* Lévl. *Folia Microbiologica* **2013**, *58* (1), 1-7.
265. Woo, S. H.; Cho, J. S.; Lee, B. S.; Kim, E. K., Decolorization of melanin by lignin peroxidase from *Phanerochaete chrysosporium*. *Biotechnology and Bioengineering* **2004**, *9* (4), 256.

266. Kim, B. S.; Blaghen, M.; Hong, H. S.; Lee, K. M., Purification and characterization of a melanin biodegradation enzyme from *Geotrichum* sp. *International Journal of Cosmetic Science* **2016**, *38* (6), 622-626.
267. Liu, G.; Xu, J.; Wang, K., Solar water evaporation by black photothermal sheets. *Nano Energy* **2017**.
268. Bond, T. C.; Bergstrom, R. W., Light absorption by carbonaceous particles: An investigative review. *Aerosol science and technology* **2006**, *40* (1), 27-67.
269. Wang, P., Emerging investigator series: the rise of nano-enabled photothermal materials for water evaporation and clean water production by sunlight. *Environmental Science: Nano* **2018**, *5* (5), 1078-1089.
270. Konkana, B.; Vasudevan, S., Understanding aqueous dispersibility of graphene oxide and reduced graphene oxide through pK_a measurements. *The journal of physical chemistry letters* **2012**, *3* (7), 867-872.
271. Li, D.; Müller, M. B.; Gilje, S.; Kaner, R. B.; Wallace, G. G., Processable aqueous dispersions of graphene nanosheets. *Nature nanotechnology* **2008**, *3* (2), 101.
272. Stumm, W.; Morgan, J. J., Aquatic Chemistry: Chemical Equilibria and Rates in Natural Waters. 3rd ed. *John Wiley & Sons: New York* **1996**.
273. Kansal, S.; Singh, M.; Sud, D., Studies on photodegradation of two commercial dyes in aqueous phase using different photocatalysts. *Journal of hazardous materials* **2007**, *141* (3), 581-590.
274. Yan, Z.; Xia, M.; Wang, P.; Zhang, P.; Liang, O.; Xie, Y.-H., Selective manipulation of molecules by electrostatic force and detection of single molecules in aqueous solution. *The Journal of Physical Chemistry C* **2016**, *120* (23), 12765-12772.
275. Zheng, Y.; Zhong, X.; Li, Z.; Xia, Y., Successive, Seed-Mediated Growth for the Synthesis of Single-Crystal Gold Nanospheres with Uniform Diameters Controlled in the Range of 5–150 nm. *Particle & Particle Systems Characterization* **2014**, *31* (2), 266-273.
276. Gottenbos, B.; van der Mei, H. C.; Klatter, F.; Nieuwenhuis, P.; Busscher, H. J., In vitro and in vivo antimicrobial activity of covalently coupled quaternary ammonium silane coatings on silicone rubber. *Biomaterials* **2002**, *23* (6), 1417-1423.
277. Kidambi, P. R.; Jang, D.; Idrobo, J.-C.; Boutilier, M. S. H.; Wang, L.; Kong, J.; Karnik, R., Nanoporous Atomically Thin Graphene Membranes for Desalting and Dialysis Applications. *Advanced Materials* **2017**, *29* (33), 1700277-n/a.
278. Wang, L. K.; Chen, J. P.; Hung, Y.-T.; Shamma, N. K., *Membrane and desalination technologies*. Springer: 2008; Vol. 13.
279. Baker, R. W., Ultrafiltration. In *Membrane Technology and Applications*, John Wiley & Sons, Ltd: 2004; pp 237-274.
280. Fang, Q.; Zhou, X.; Deng, W.; Zheng, Z.; Liu, Z., Freestanding bacterial cellulose-graphene oxide composite membranes with high mechanical strength for selective ion permeation. *Scientific Reports* **2016**, *6*, 33185.
281. Joshi, R. K.; Carbone, P.; Wang, F. C.; Kravets, V. G.; Su, Y.; Grigorieva, I. V.; Wu, H. A.; Geim, A. K.; Nair, R. R., Precise and Ultrafast Molecular Sieving Through Graphene Oxide Membranes. *Science* **2014**, *343* (6172), 752-754.
282. Chen, L.; Shi, G.; Shen, J.; Peng, B.; Zhang, B.; Wang, Y.; Bian, F.; Wang, J.; Li, D.; Qian, Z.; Xu, G.; Liu, G.; Zeng, J.; Zhang, L.; Yang, Y.; Zhou, G.; Wu, M.; Jin, W.; Li, J.; Fang, H., Ion sieving in graphene oxide membranes via cationic control of interlayer spacing. *Nature* **2017**, *550*, 380.
283. Abraham, J.; Vasu, K. S.; Williams, C. D.; Gopinadhan, K.; Su, Y.; Cherian, C. T.; Dix, J.; Prestat, E.; Haigh, S. J.; Grigorieva, I. V.; Carbone, P.; Geim, A. K.; Nair, R. R., Tunable sieving of ions using graphene oxide membranes. *Nature Nanotechnology* **2017**, *12*, 546.
284. An, D.; Yang, L.; Wang, T.-J.; Liu, B., Separation Performance of Graphene Oxide Membrane in Aqueous Solution. *Industrial & Engineering Chemistry Research* **2016**, *55* (17), 4803-4810.

285. Lim, D.-K.; Barhoumi, A.; Wylie, R. G.; Reznor, G.; Langer, R. S.; Kohane, D. S., Enhanced Photothermal Effect of Plasmonic Nanoparticles Coated with Reduced Graphene Oxide. *Nano Letters* **2013**, *13* (9), 4075-4079.
286. Yang, Y.; Zhang, H.; Liu, R.; Wen, X.; Hou, T.-C.; Wang, Z. L., Fully Enclosed Triboelectric Nanogenerators for Applications in Water and Harsh Environments. *Adv. Energy Mater.* **2013**, *3* (12), 1563-1568.
287. Ghasemzadeh, G.; Momenpour, M.; Omid, F.; Hosseini, M. R.; Ahani, M.; Barzegari, A., Applications of nanomaterials in water treatment and environmental remediation. *Frontiers of Environmental Science & Engineering* **2014**, *8* (4), 471-482.
288. Shawky, H. A.; Chae, S.-R.; Lin, S.; Wiesner, M. R., Synthesis and characterization of a carbon nanotube/polymer nanocomposite membrane for water treatment. *Desalination* **2011**, *272* (1), 46-50.
289. Gehrke, I.; Geiser, A.; Somborn-Schulz, A., Innovations in nanotechnology for water treatment. *Nanotechnology, science and applications* **2015**, *8*, 1.
290. Bolisetty, S.; Mezzenga, R., Amyloid-carbon hybrid membranes for universal water purification. *Nature nanotechnology* **2016**, *11* (4), 365-371.
291. Molinari, R.; Pirillo, F.; Falco, M.; Loddo, V.; Palmisano, L., Photocatalytic degradation of dyes by using a membrane reactor. *Chemical Engineering and Processing: Process Intensification* **2004**, *43* (9), 1103-1114.
292. Lv, Y.; Zhang, C.; He, A.; Yang, S.-J.; Wu, G.-P.; Darling, S. B.; Xu, Z.-K., Photocatalytic Nanofiltration Membranes with Self-Cleaning Property for Wastewater Treatment. *Advanced Functional Materials* **2017**, *27* (27), 1700251-n/a.
293. Chen, F.; Gong, A. S.; Zhu, M.; Chen, G.; Lacey, S. D.; Jiang, F.; Li, Y.; Wang, Y.; Dai, J.; Yao, Y.; Song, J.; Liu, B.; Fu, K.; Das, S.; Hu, L., Mesoporous, Three-Dimensional Wood Membrane Decorated with Nanoparticles for Highly Efficient Water Treatment. *ACS Nano* **2017**, *11* (4), 4275-4282.
294. Jiang, Q.; Tian, L.; Liu, K. K.; Tadealli, S.; Raliya, R.; Biswas, P.; Naik, R. R.; Singamaneni, S., Bilayered Biofoam for Highly Efficient Solar Steam Generation. *Advanced Materials* **2016**, *28* (42), 9400-9407.
295. Jiang, Q.; Kacica, C.; Soundappan, T.; Liu, K.-k.; Tadealli, S.; Biswas, P.; Singamaneni, S., An in situ grown bacterial nanocellulose/graphene oxide composite for flexible supercapacitors. *Journal of Materials Chemistry A* **2017**, *5* (27), 13976-13982.
296. Vinodgopal, K.; Kamat, P. V., Enhanced Rates of Photocatalytic Degradation of an Azo Dye Using SnO₂/TiO₂ Coupled Semiconductor Thin Films. *Environmental Science & Technology* **1995**, *29* (3), 841-845.
297. Chen, X.; Wu, G.; Chen, J.; Chen, X.; Xie, Z.; Wang, X., Synthesis of "clean" and well-dispersive Pd nanoparticles with excellent electrocatalytic property on graphene oxide. *Journal of the American Chemical Society* **2011**, *133* (11), 3693-3695.
298. Huang, X.; Tang, S.; Mu, X.; Dai, Y.; Chen, G.; Zhou, Z.; Ruan, F.; Yang, Z.; Zheng, N., Freestanding palladium nanosheets with plasmonic and catalytic properties. *Nature nanotechnology* **2011**, *6* (1), 28-32.
299. Liu, Q.; Xu, Y.-R.; Wang, A.-J.; Feng, J.-J., A single-step route for large-scale synthesis of core-shell palladium@platinum dendritic nanocrystals/reduced graphene oxide with enhanced electrocatalytic properties. *Journal of Power Sources* **2016**, *302*, 394-401.
300. Rathi, A. K.; Gawande, M. B.; Pechousek, J.; Tucek, J.; Aparicio, C.; Petr, M.; Tomanec, O.; Krikavova, R.; Travnický, Z.; Varma, R. S., Maghemite decorated with ultra-small palladium nanoparticles (γ -Fe₂O₃-Pd): applications in the Heck-Mizoroki olefination, Suzuki reaction and allylic oxidation of alkenes. *Green Chemistry* **2016**, *18* (8), 2363-2373.
301. Wang, B.; Yan, T.; Chang, T.; Wei, J.; Zhou, Q.; Yang, S.; Fang, T., Palladium supported on reduced graphene oxide as a high-performance catalyst for the dehydrogenation of dodecahydro-N-ethylcarbazole. *Carbon* **2017**, *122*, 9-18.

302. Rahul, R.; Singh, R. K.; Bera, B.; Devivaraprasad, R.; Neergat, M., The role of surface oxygenated-species and adsorbed hydrogen in the oxygen reduction reaction (ORR) mechanism and product selectivity on Pd-based catalysts in acid media. *Physical Chemistry Chemical Physics* **2015**, *17* (23), 15146-15155.
303. Dubey, A. V.; Kumar, A. V., A biomimetic magnetically recoverable palladium nanocatalyst for the Suzuki cross-coupling reaction. *RSC Advances* **2016**, *6* (52), 46864-46870.
304. Qu, K.; Wu, L.; Ren, J.; Qu, X., Natural DNA-modified graphene/Pd nanoparticles as highly active catalyst for formic acid electro-oxidation and for the Suzuki reaction. *ACS applied materials & interfaces* **2012**, *4* (9), 5001-5009.
305. Yang, D.; Velamakanni, A.; Bozoklu, G.; Park, S.; Stoller, M.; Piner, R. D.; Stankovich, S.; Jung, I.; Field, D. A.; Ventrice, C. A.; Ruoff, R. S., Chemical analysis of graphene oxide films after heat and chemical treatments by X-ray photoelectron and Micro-Raman spectroscopy. *Carbon* **2009**, *47* (1), 145-152.
306. Hervés, P.; Pérez-Lorenzo, M.; Liz-Marzán, L. M.; Dzubielia, J.; Lu, Y.; Ballauff, M., Catalysis by metallic nanoparticles in aqueous solution: model reactions. *Chemical Society Reviews* **2012**, *41* (17), 5577-5587.
307. Material Safety Data Sheet of Methyl Orange,(August,2017) Retrieved from <http://www.sciencelab.com/msds.php?msdsId=9926081>.
308. Wu, Q. Y.; Liang, H. Q.; Li, M.; Liu, B. T.; Xu, Z. K., Hierarchically porous carbon membranes derived from PAN and their selective adsorption of organic dyes. *Chin. J. Polym. Sci.* **2016**, *34* (1), 23-33.
309. Chee, W. K.; Lim, H. N.; Zainal, Z.; Huang, N. M.; Harrison, I.; Andou, Y., Flexible Graphene-Based Supercapacitors: A Review. *The Journal of Physical Chemistry C* **2016**, *120* (8), 4153-4172.
310. Han, X.; Funk, M. R.; Shen, F.; Chen, Y.-C.; Li, Y.; Campbell, C. J.; Dai, J.; Yang, X.; Kim, J.-W.; Liao, Y.; Connell, J. W.; Barone, V.; Chen, Z.; Lin, Y.; Hu, L., Scalable Holey Graphene Synthesis and Dense Electrode Fabrication toward High-Performance Ultracapacitors. *ACS Nano* **2014**, *8* (8), 8255-8265.
311. Li, Y.; Chen, C.; Gao, T.; Zhang, D.; Huang, X.; Pan, Y.; Ye, K.; Cheng, K.; Cao, D.; Wang, G., Synthesis of Hierarchically Porous Sandwich-Like Carbon Materials for High-Performance Supercapacitors. *Chemistry – A European Journal* **2016**, *22* (47), 16863-16871.
312. Ma, Z.; Huang, X.; Dou, S.; Wu, J.; Wang, S., One-Pot Synthesis of Fe₂O₃ Nanoparticles on Nitrogen-Doped Graphene as Advanced Supercapacitor Electrode Materials. *The Journal of Physical Chemistry C* **2014**, *118* (31), 17231-17239.
313. Wang, S.; Pei, B.; Zhao, X.; Dryfe, R. A. W., Highly porous graphene on carbon cloth as advanced electrodes for flexible all-solid-state supercapacitors. *Nano Energy* **2013**, *2* (4), 530-536.
314. Chen, C.; Zhang, Y.; Li, Y.; Dai, J.; Song, J.; Yao, Y.; Gong, Y.; Kierzewski, I.; Xie, J.; Hu, L., All-wood, low tortuosity, aqueous, biodegradable supercapacitors with ultra-high capacitance. *Energy & Environmental Science* **2017**, *10* (2), 538-545.
315. Timpanaro, S.; Kemerink, M.; Touwslager, F. J.; De Kok, M. M.; Schrader, S., Morphology and conductivity of PEDOT/PSS films studied by scanning-tunneling microscopy. *Chemical Physics Letters* **2004**, *394* (4-6), 339-343.
316. Zhang, X.; Chang, D.; Liu, J.; Luo, Y., Conducting polymer aerogels from supercritical CO₂ drying PEDOT-PSS hydrogels. *Journal of Materials Chemistry* **2010**, *20* (24), 5080-5085.
317. Anothumakkool, B.; Soni, R.; Bhange, S. N.; Kurungot, S., Novel scalable synthesis of highly conducting and robust PEDOT paper for a high performance flexible solid supercapacitor. *Energy & Environmental Science* **2015**, *8* (4), 1339-1347.
318. Weng, Z.; Su, Y.; Wang, D.-W.; Li, F.; Du, J.; Cheng, H.-M., Graphene-Cellulose Paper Flexible Supercapacitors. *Advanced Energy Materials* **2011**, *1* (5), 917-922.
319. Yuan, L.; Xiao, X.; Ding, T.; Zhong, J.; Zhang, X.; Shen, Y.; Hu, B.; Huang, Y.; Zhou, J.; Wang, Z. L., Paper-Based Supercapacitors for Self-Powered Nanosystems. *Angewandte Chemie International Edition* **2012**, *51* (20), 4934-4938.

320. Feng, J.-X.; Ye, S.-H.; Wang, A.-L.; Lu, X.-F.; Tong, Y.-X.; Li, G.-R., Flexible Cellulose Paper-based Asymmetrical Thin Film Supercapacitors with High-Performance for Electrochemical Energy Storage. *Advanced Functional Materials* **2014**, *24* (45), 7093-7101.

Appendix

Appendix 1

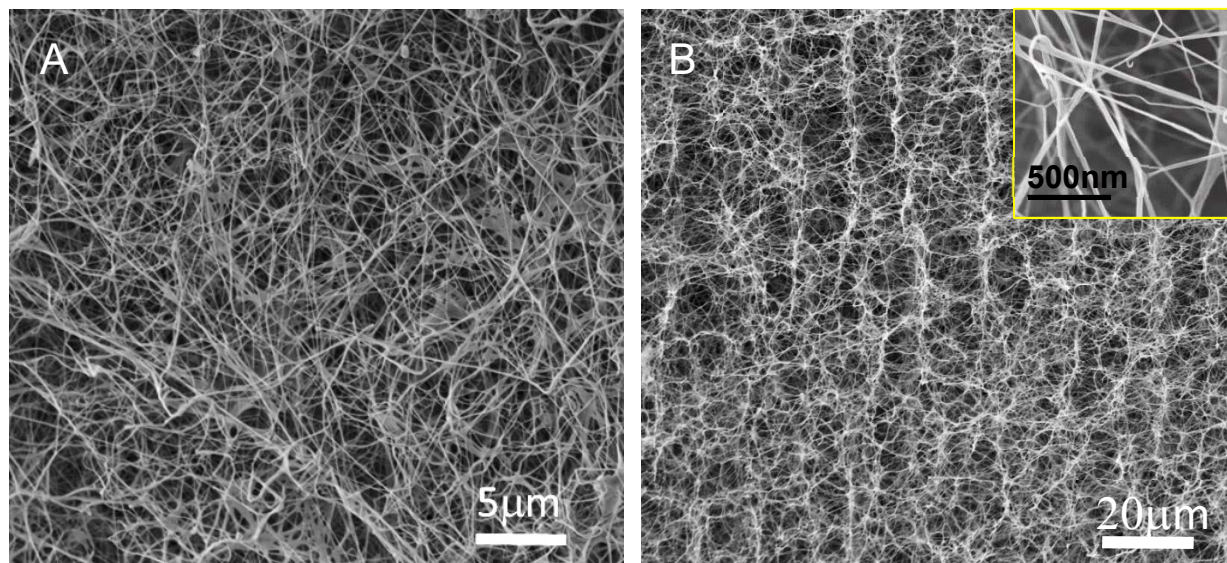


Figure S1.1. SEM images of (A) top surface and (B) cross section of freeze-dried pristine BNC aerogel (Inset image: high resolution SEM image showing individual BNC fibers of 20-100 nm in diameter).

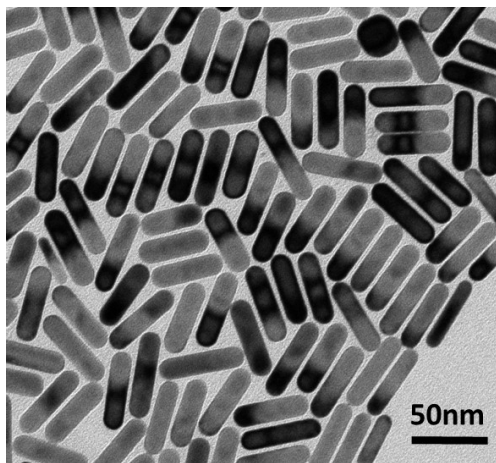


Figure S1.2. Representative transmission electron microscopy (TEM) image of gold nanorods employed in this study

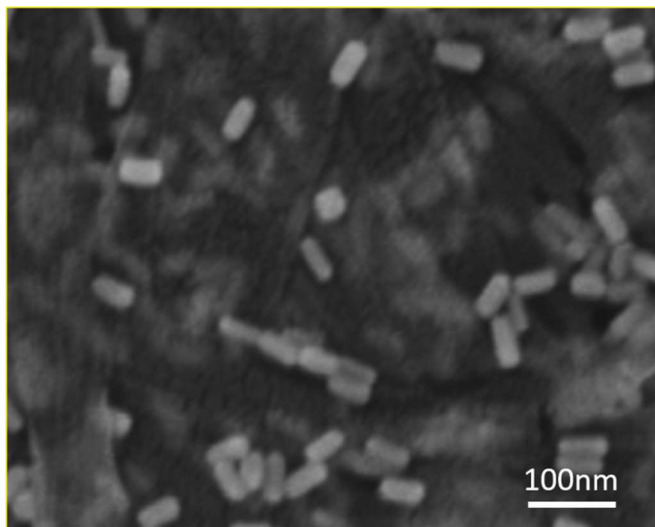


Figure S1.3. Representative high-resolution SEM image showing AuNRs on the surface and underneath the cellulose fibers.

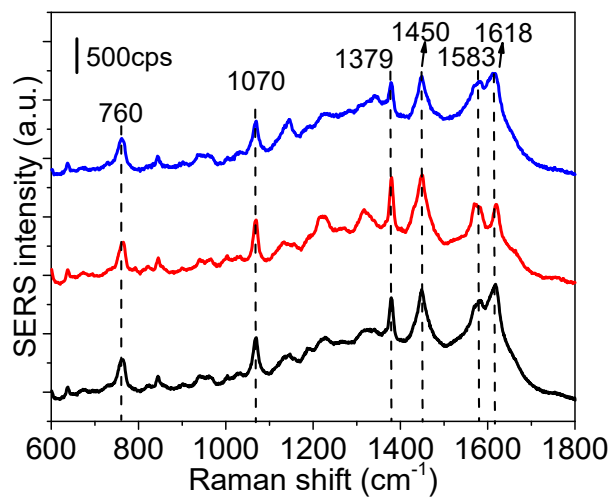


Figure S1.4. SERS spectra collected from different points of *E.coli* adsorbed on the AuNRs/BNC swab showing characteristic Raman bands of *E. coli*.

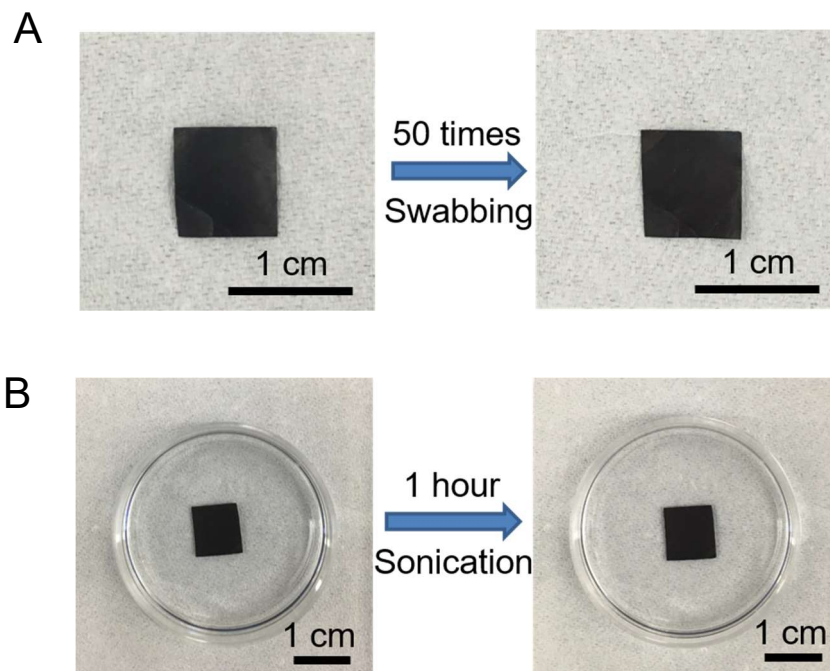


Figure S1.5. Photographs of AuNRs/BNC film before and after (A) swabbing 50 times and (B) 1 hour of sonication. No color change in AuNRs/BNC film confirmed strong adsorption of AuNRs on BNC film.

Appendix 2

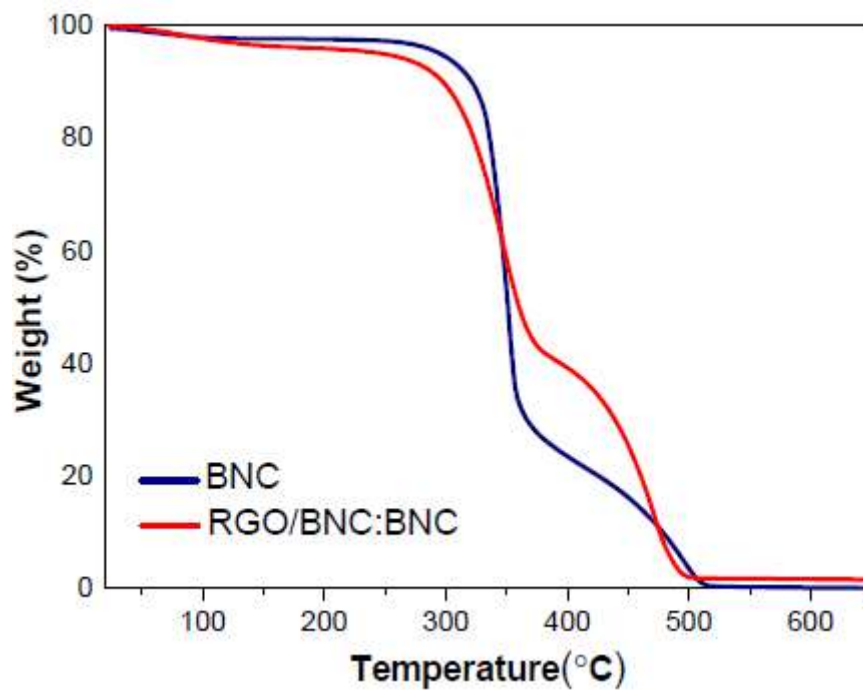


Figure S2.1. TGA curves for BNC film and air-dried RGO/BNC:BNC film.

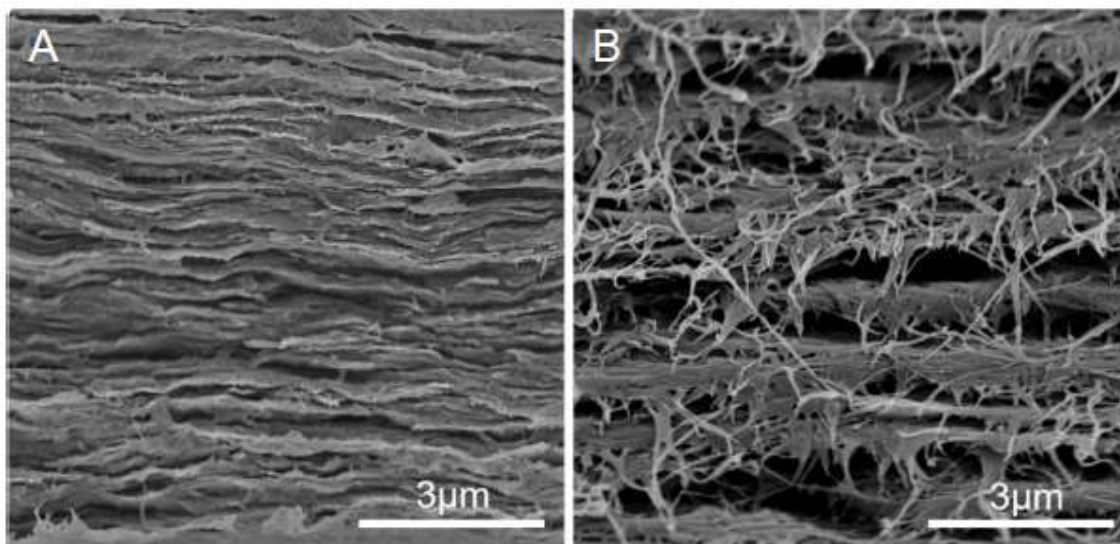


Figure S2.2. Cross-sectional SEM image of air-dried (A) RGO/BNC, (B) BNC film.

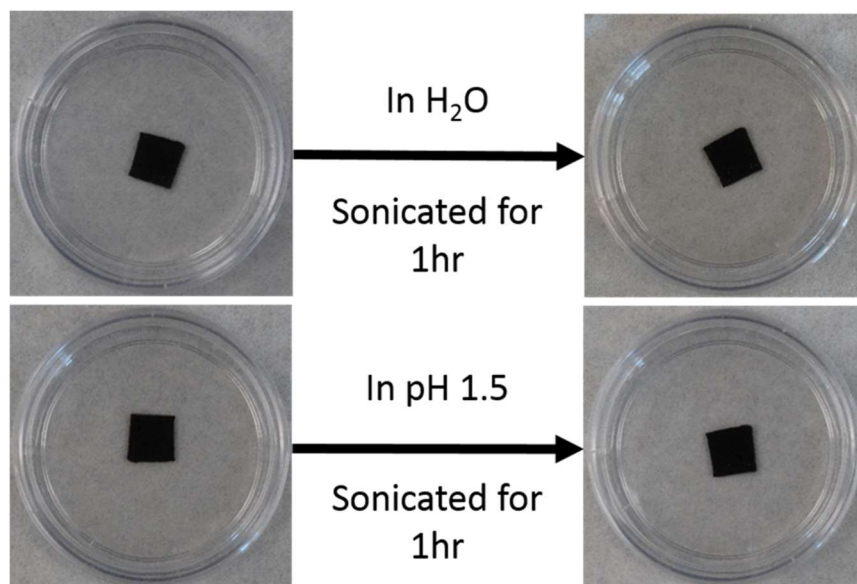


Figure S2.3. Stability of RGO/BNC:BNC as evidenced by the mechanically stable films after ultrasonication for 1 hr in at pH 7 and pH 1.5.

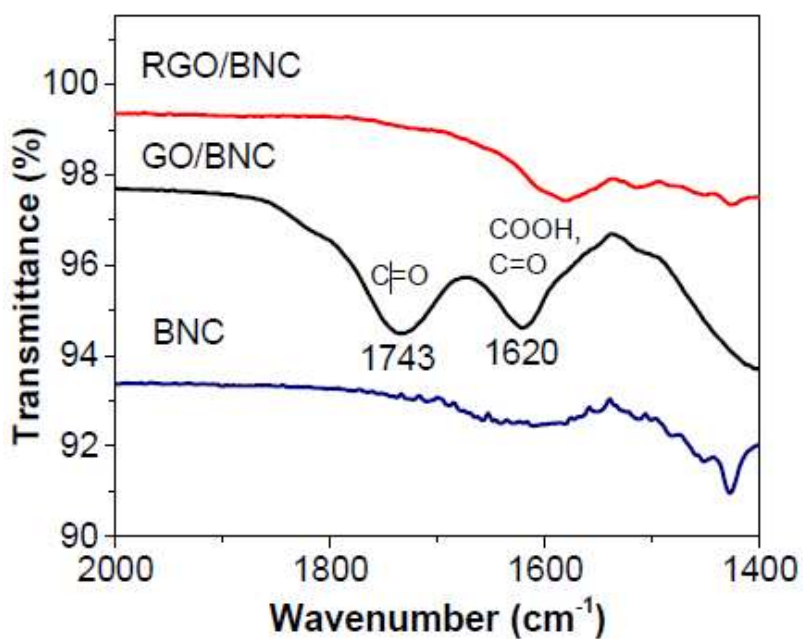


Figure S2.4. FTIR spectra of RGO/BNC, GO/BNC and BNC dry film.

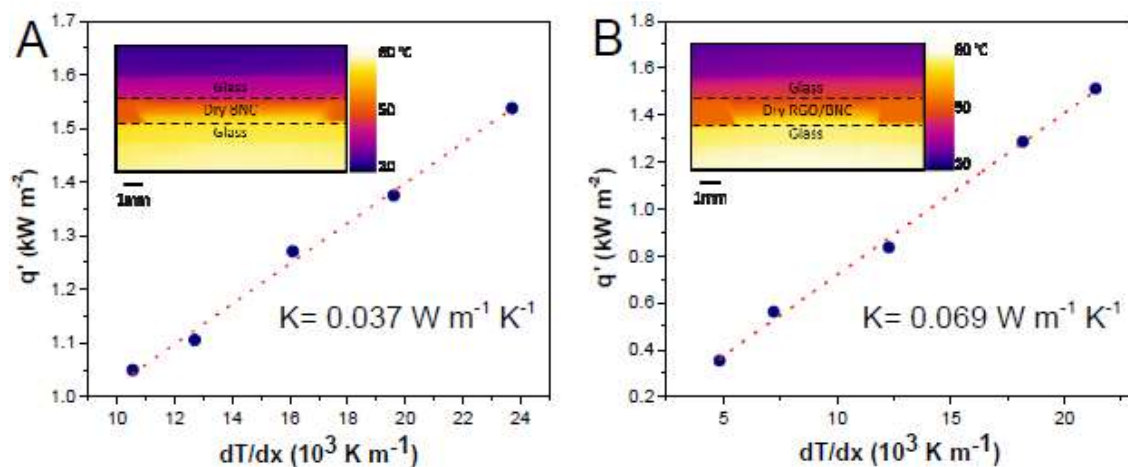


Figure S2.5. Thermal conductivity of dry BNC and RGO/BNC foams.

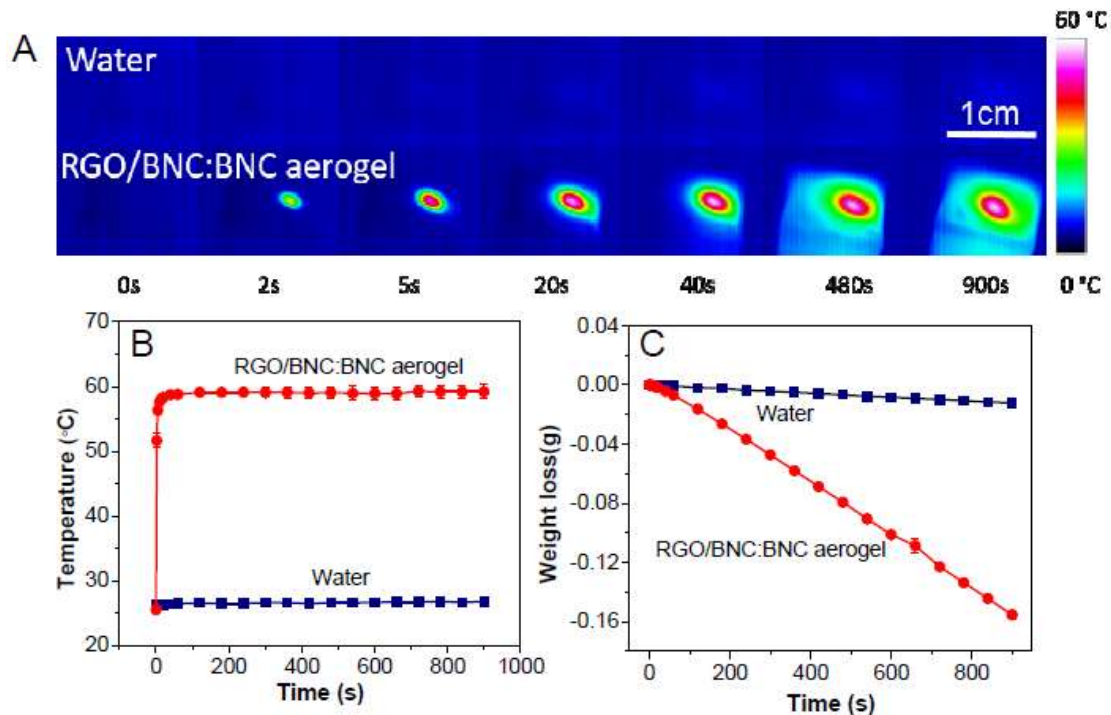


Figure S2.6. Solar steam generation of RGO:BNC/BNC upon irradiation with 808nm laser (A) IR images showing the temperature of water and RGO/BNC:BNC aerogel floated at air/water interface under 808 nm laser illumination ($510\text{mK}/\text{cm}^2$) at various time points. (B) Plot showing the surface temperature of water and RGO/BNC:BNC aerogel at air/water interface as function of irradiation time. (C) Plot showing the cumulative weight loss through water evaporation under solar illumination as a function of irradiation time.

Appendix 3

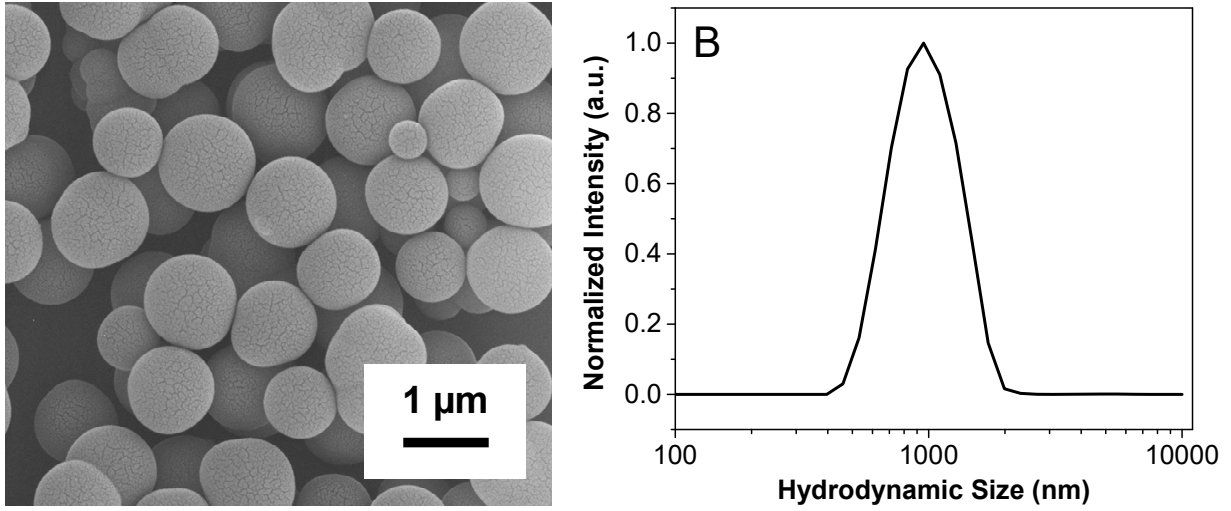


Figure S3.1 (A) SEM image of PDA particles (B) Hydrodynamic size of PDA particles measured by DLS.

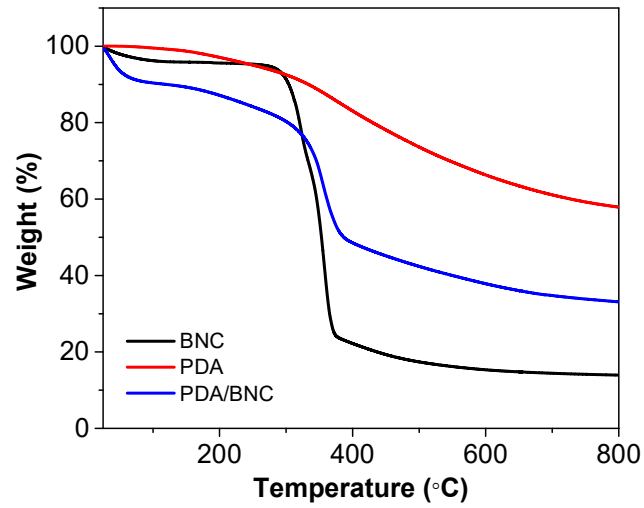


Figure S3.2 TGA of pristine BNC, pristine PDA and PDA/BNC.

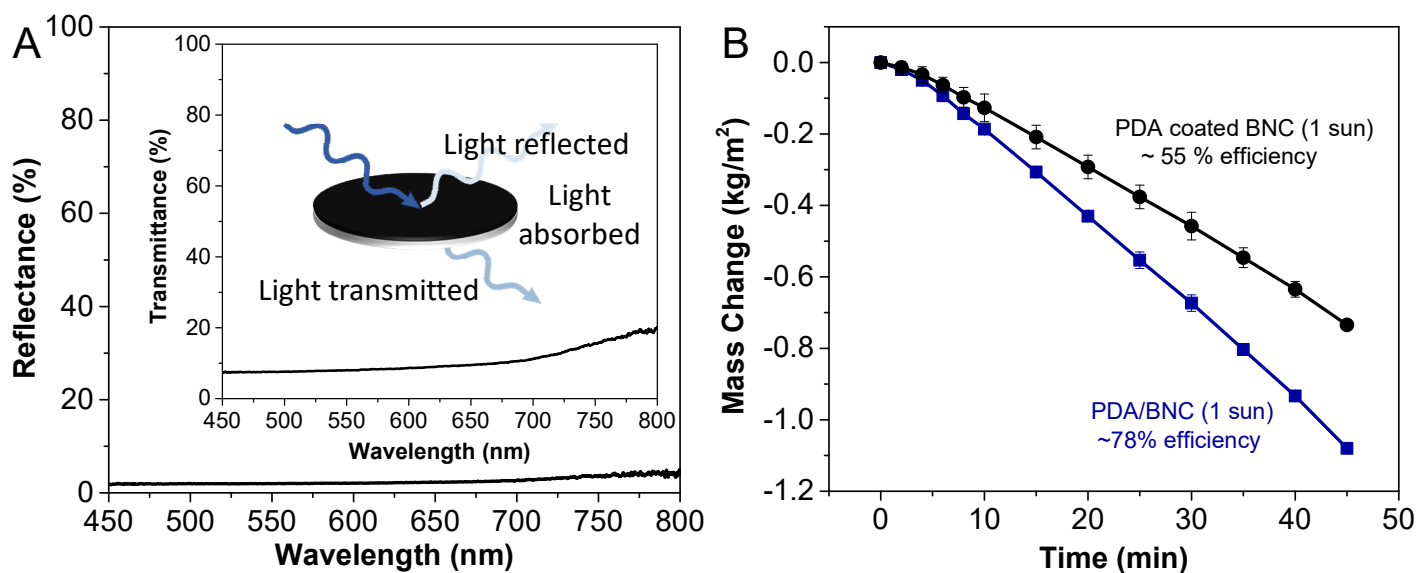


Figure S3.3 (A) Transmittance and reflectance spectra of PDA coated BNC hydrogel. (B) Solar steam generation performance of PDA coated BNC compared with PDA/BNC via *in situ* growth method.

Appendix 4

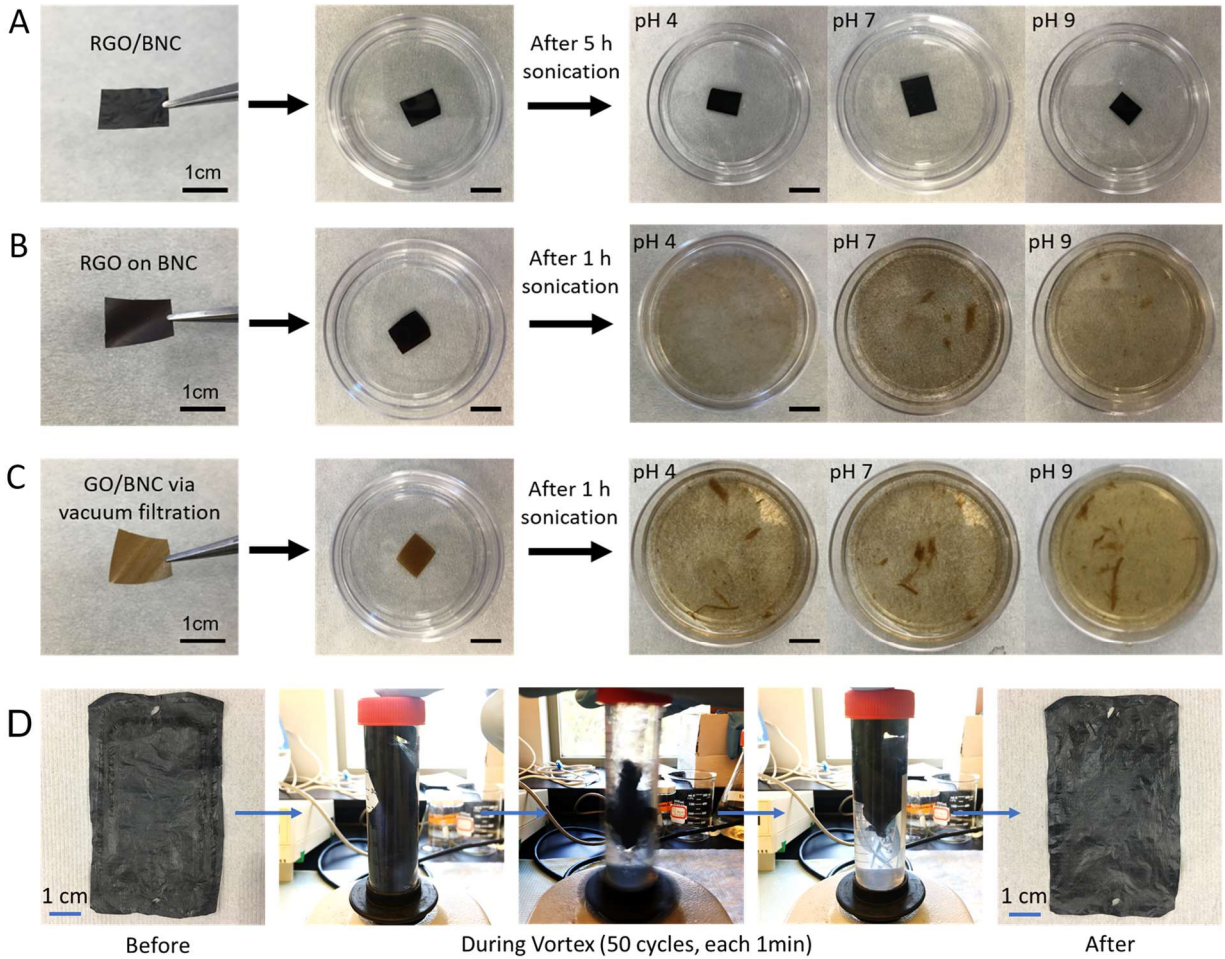


Figure S4.1. Additional stability tests. (A) RGO/BNC membrane remains intact after 5 h sonication in all pH conditions. (B) RGO coated BNC prepared by vacuum filtration quickly disintegrated during approximately 1 h sonication in all pH conditions. (C) GO was mixed with BNC dispersion and then vacuum filtrated to form a membrane. The membrane showed good stability in pH 7 solutions but disintegrated during sonication in solutions with different pH (Scale bars: 1 cm). (D) prolonged vortexing employed for membrane washing did not damage the membrane.

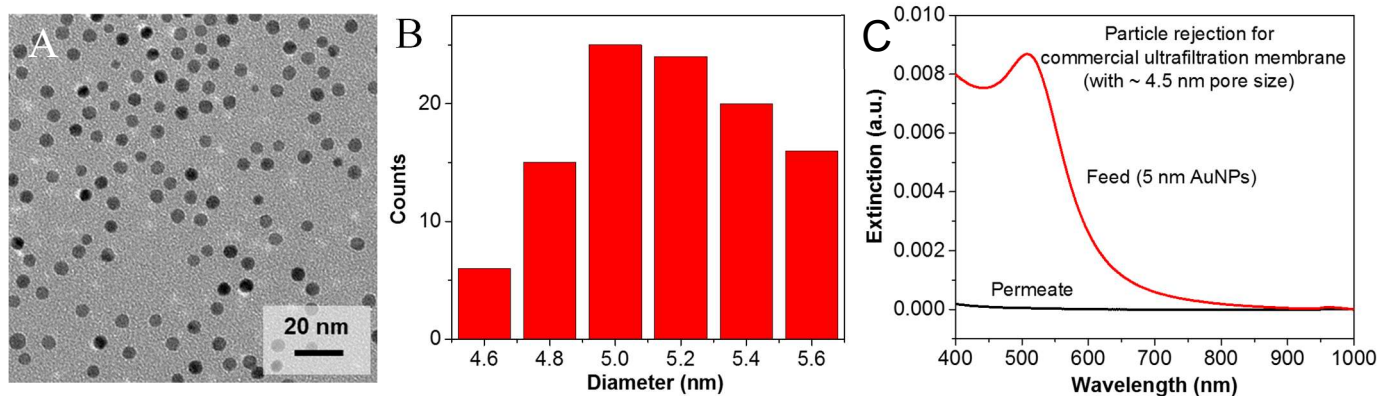


Figure S4.2. (A) TEM image of gold nanoparticles of 5 nm in diameter. (B) Size distribution of AuNPs. (C) UV-vis extinction spectra indicating the rejection of AuNPs with 5 nm in diameter filtered through commercial UF membranes.

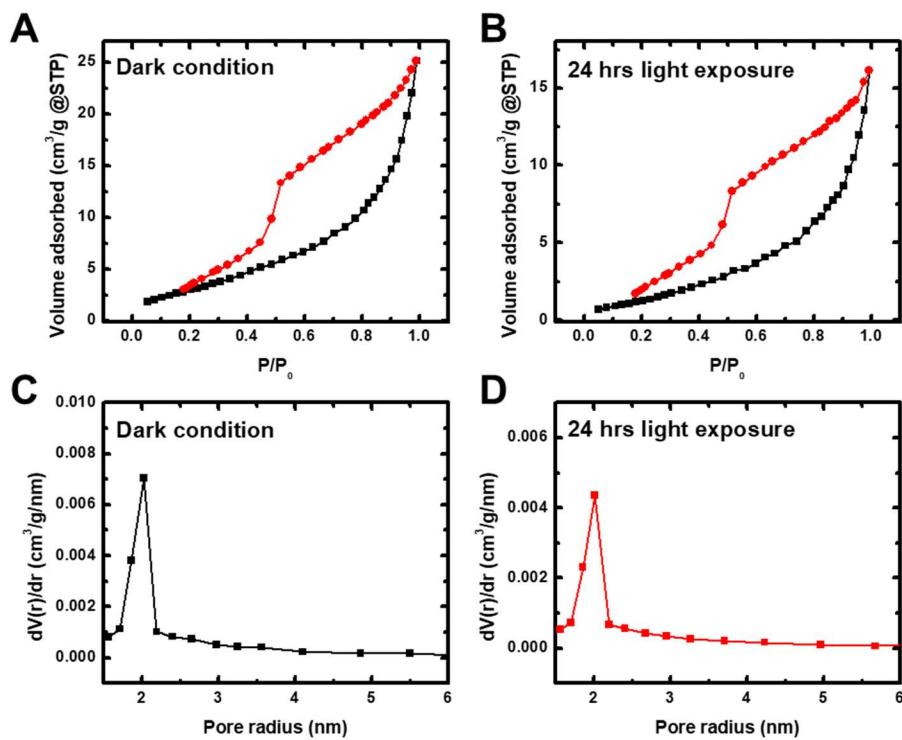


Figure S4.3. Nitrogen isotherm of RGO/BNC membrane (A) before and (B) after 24-hour light illumination (2.9 kW/m^2). The pore size of RGO/BNC membrane (C) before and (D) after 24-hour light illumination (2.9 kW/m^2).

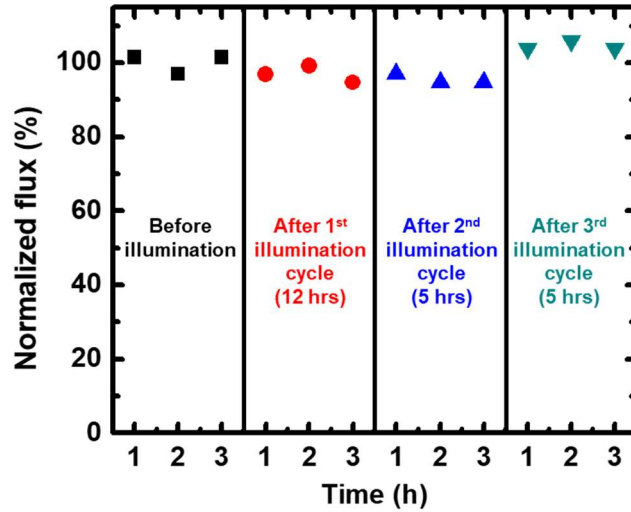


Figure S4.4. Water flux of RGO/BNC membrane under cyclic illumination. For the first cycle, RGO/BNC membrane was exposed to light (2.9 kW/m^2) for 12 hours, and 5 hours exposure time was used for the rest of cyclic tests.

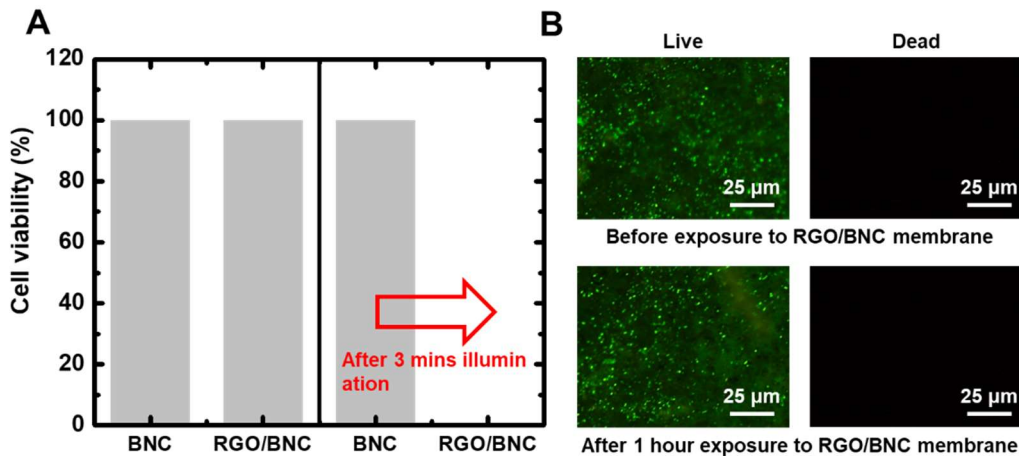


Figure S4.5. (A) Bacterial cell viability counts during bactericidal performance under illumination for BNC and RGO membranes. (B) Fluorescence images of *E. coli* on RGO/BNC before and after incubation for 1 hour.

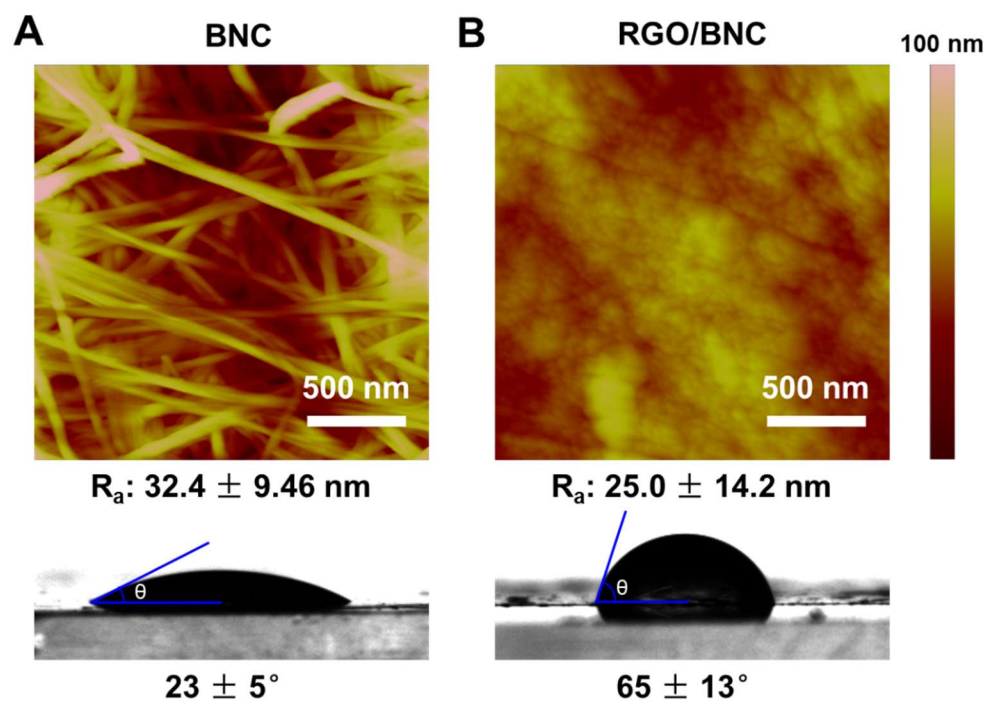


Figure S4.6. Surface roughness and contact angle of (A) RGO/BNC and (B) BNC membranes.

Appendix 5

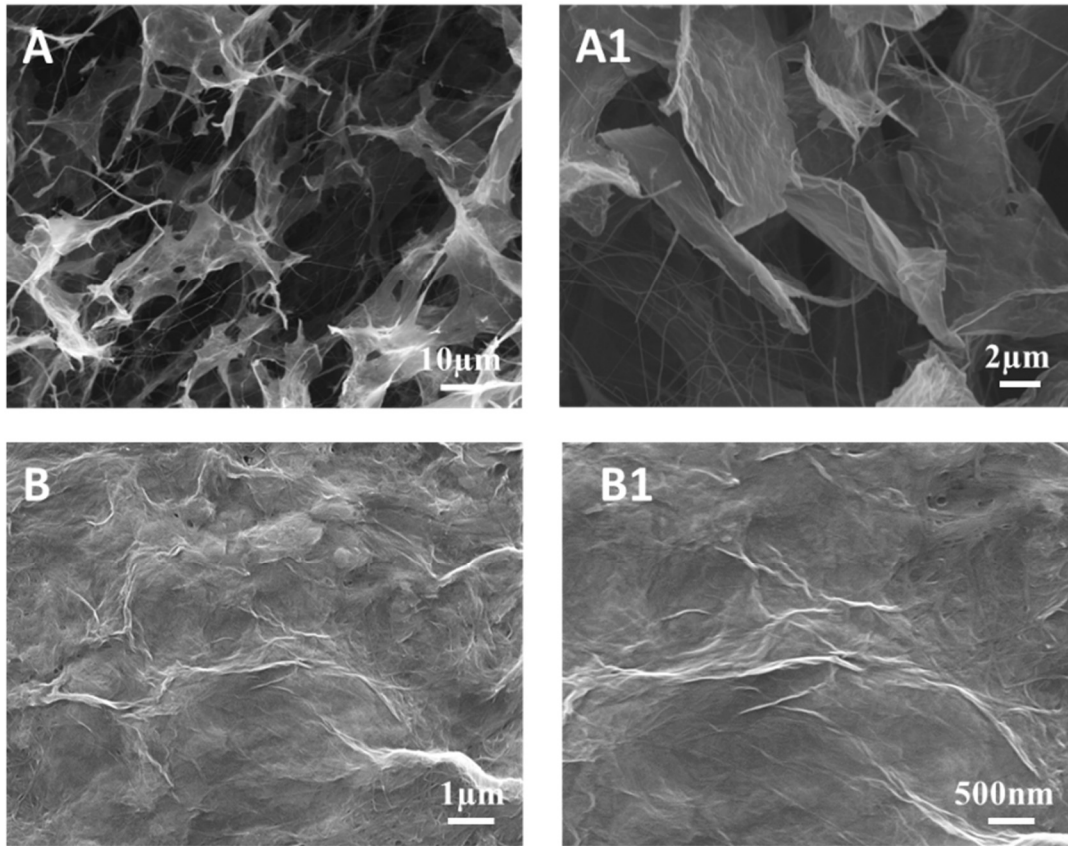


Figure S5.1. SEM images of the surface of GO/BNC aerogel (A-A1) and GO/BNC membrane (B-B1).

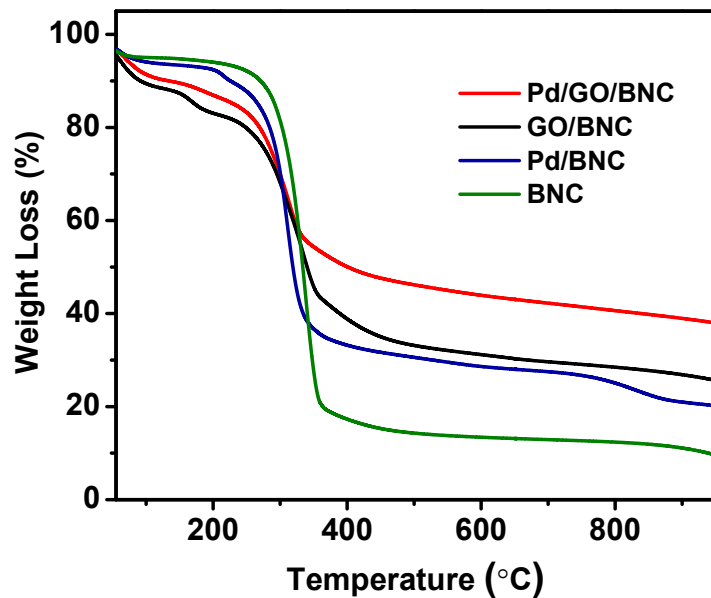


Figure S5.2. TGA curves of BNC, GO/BNC, Pd/BNC, and Pd/GO/BNC.

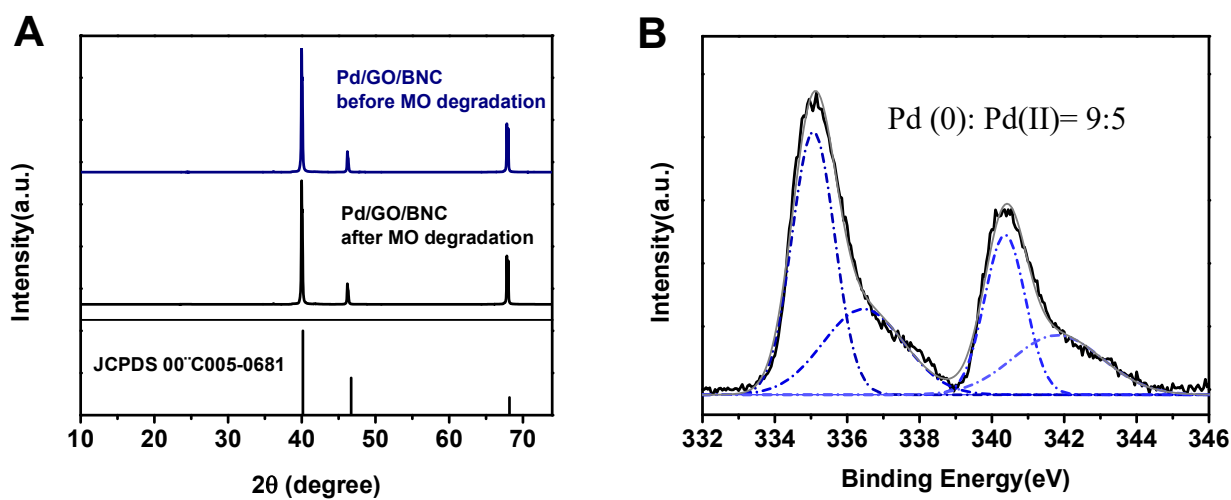


Figure S5.3. XRD (A) and XPS spectrum of Pd 3d_{3/2} and Pd 3d_{5/2} regions (B) of Pd/GO/BNC.

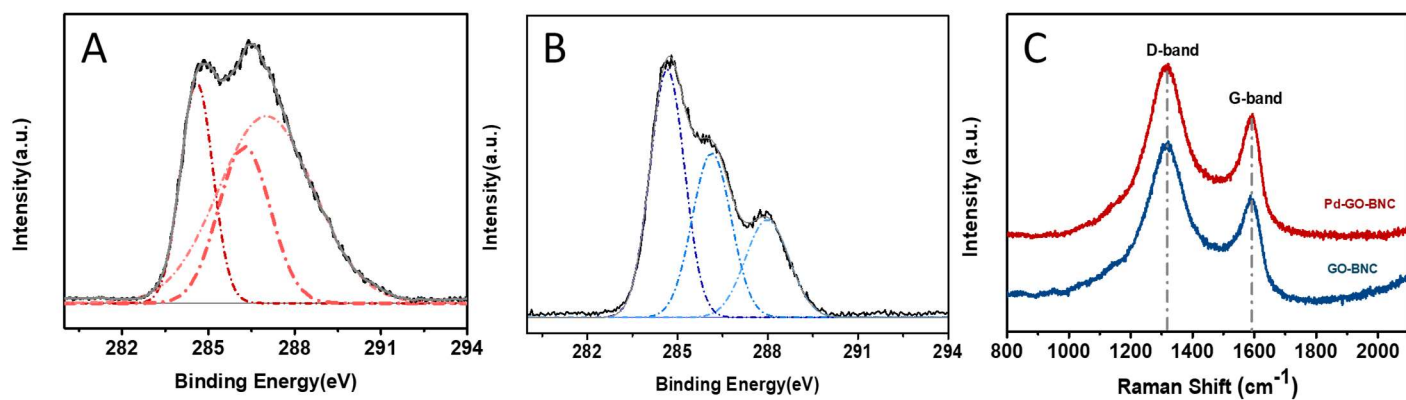


Figure S5.4. XPS spectra of C 1s regions for GO/BNC (A) and Pd/GO/BNC (B), and Raman spectra (C) of GO/BNC and Pd/GO/BNC.

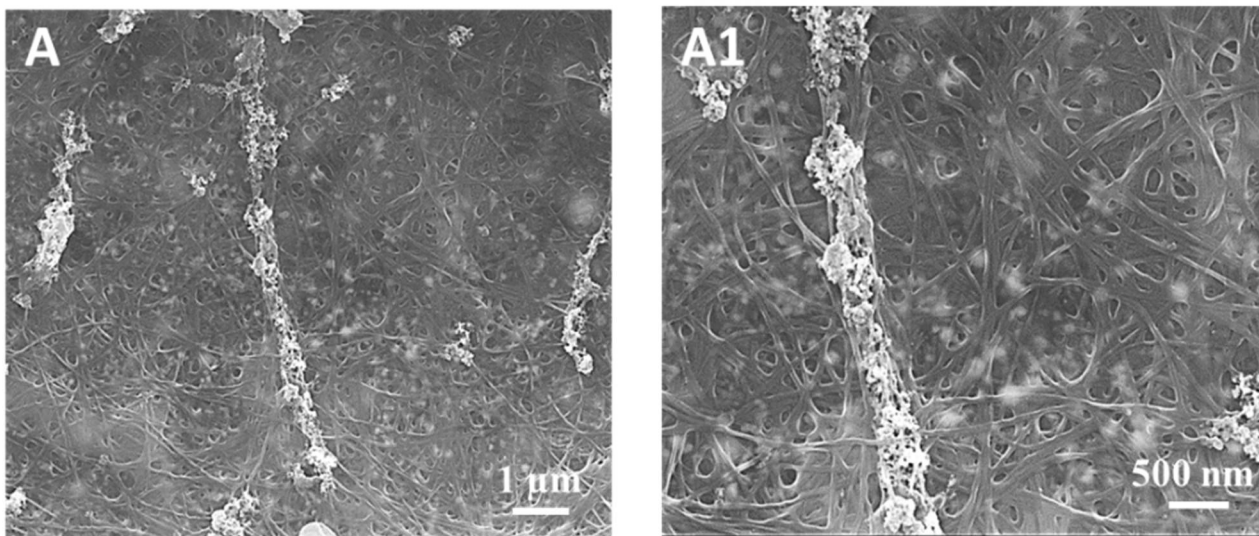


Figure S5.5. SEM images of the surface of Pd/BNC membrane.

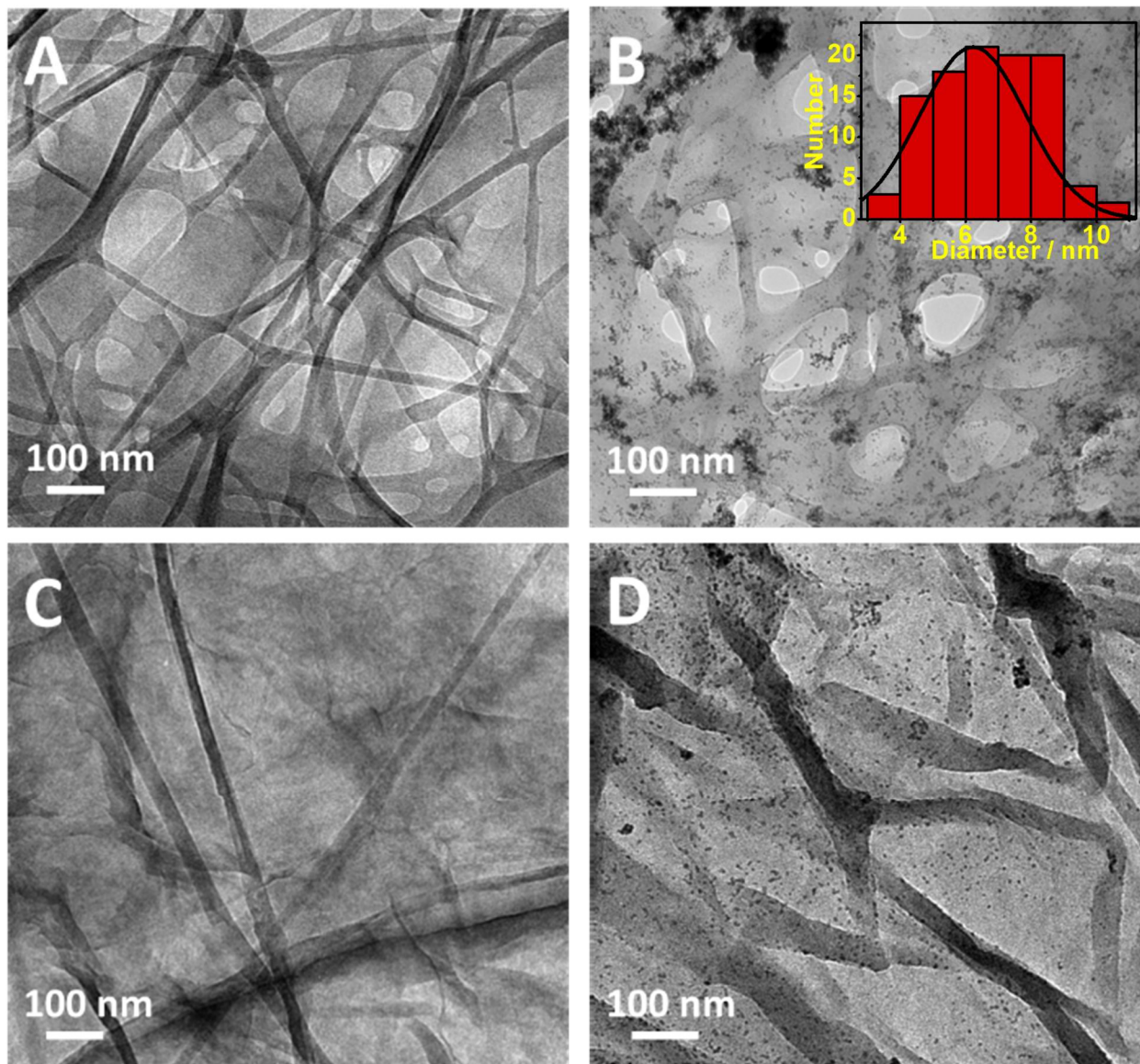


Figure S5.6. TEM images of BNC aerogel (A), Pd/BNC membrane (B), GO/BNC aerogel (C) and Pd/GO/BNC membrane (D), respectively.

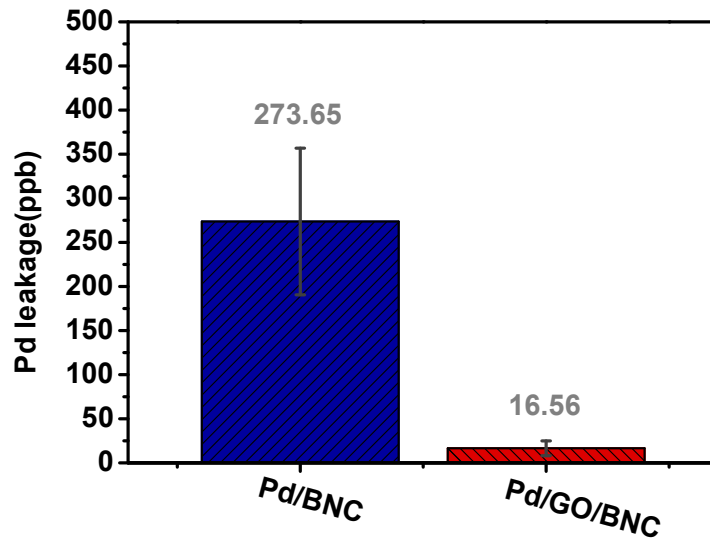


Figure S5.7. Leakage of Pd for Pd/BNC membrane and Pd/GO/BNC membrane after 5 days of mechanical agitation.

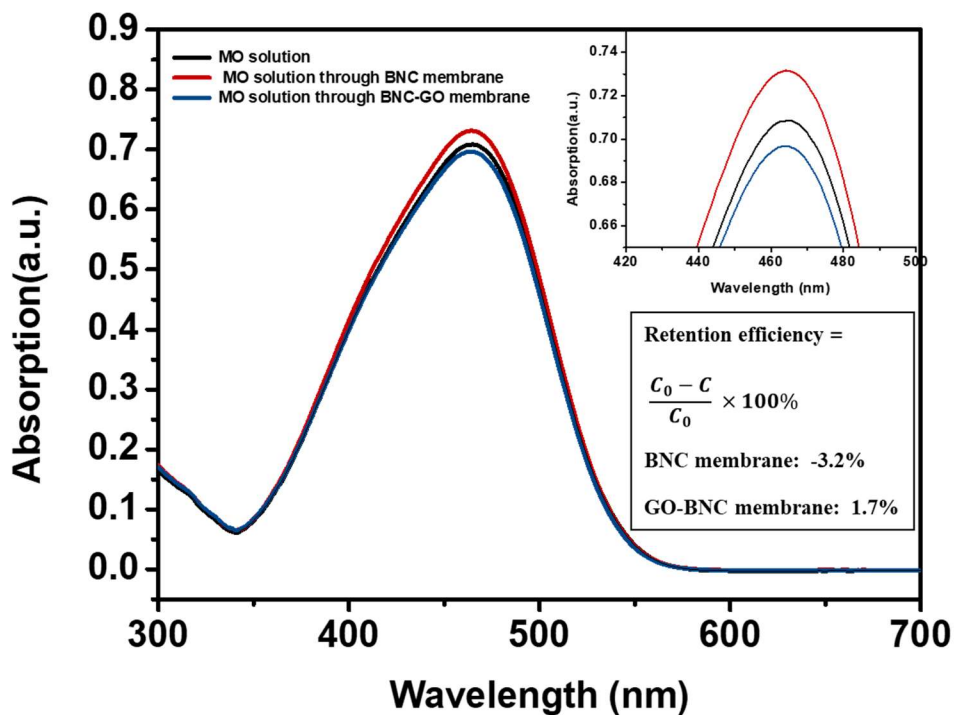


Figure S5.8. UV-vis spectra of MO solution in the presence of NaBH_4 and the MO solution filtered through the BNC and GO/BNC membrane.

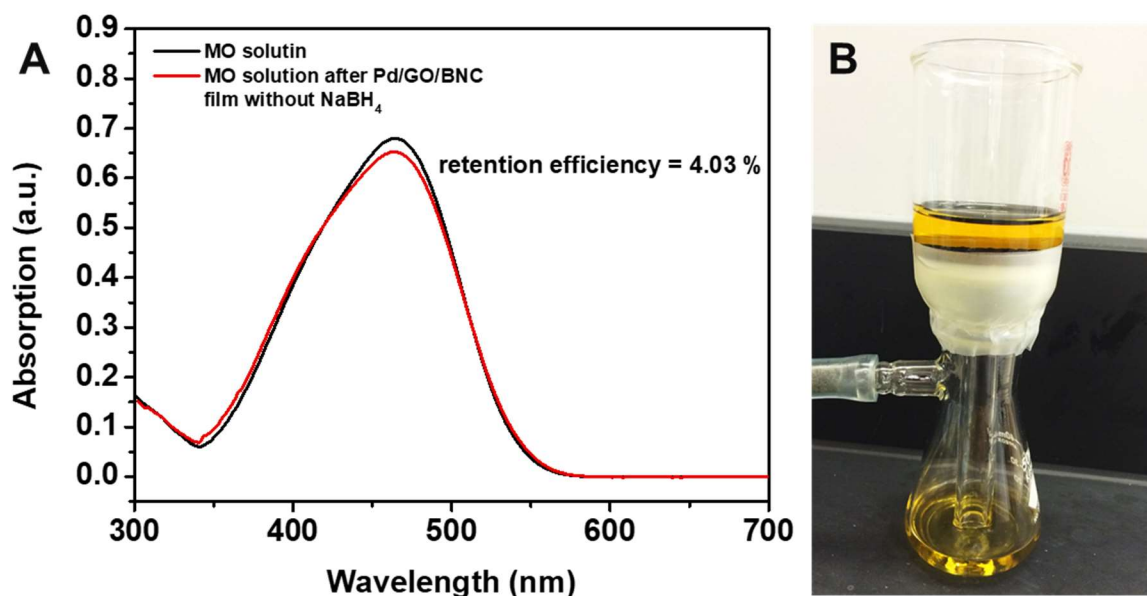


Figure S5.9. (A) UV-vis spectra of MO solution in the absence of NaBH_4 and the MO solution filter through Pd/GO/BNC membrane under 0.8 bar. (B) Image showing the filtration of MO solution through Pd/GO/BNC membrane.

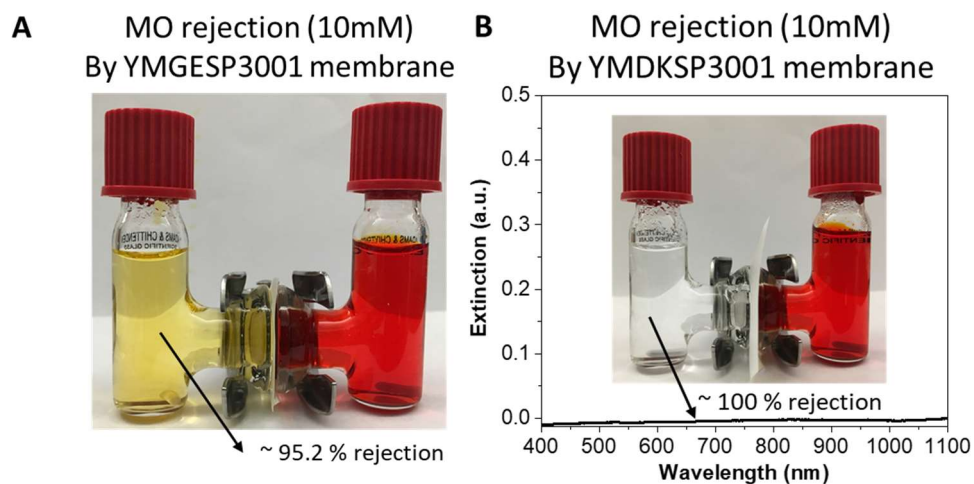


Figure S5.10. (A) MO rejection test of YMGESP3001 membrane. Inset is an image showing the setup. (B) MO rejection test of YMDKSP3001 membrane. UV-vis spectra of permeate solution from MO solution after 3 days. Inset is an image showing the setup.

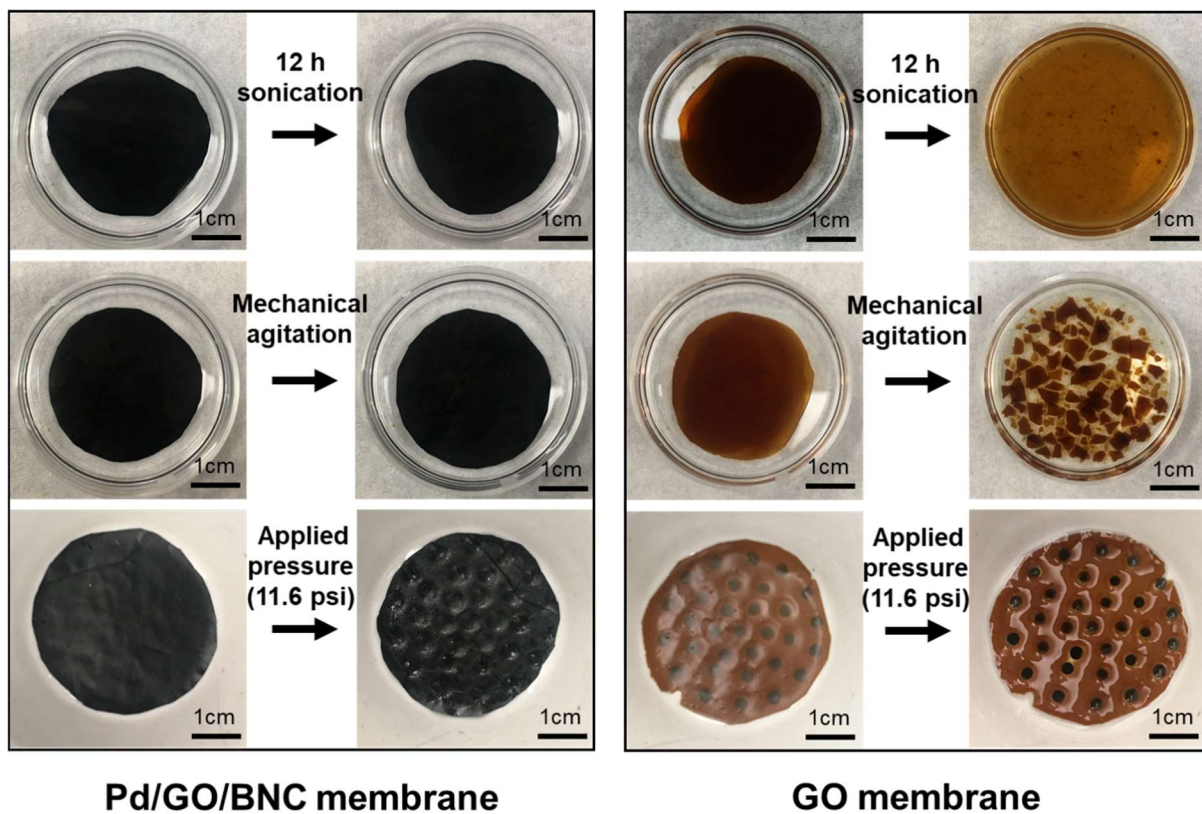


Figure S5.11. Stability tests of Pd/GO/BNC and GO membranes.

Appendix 6

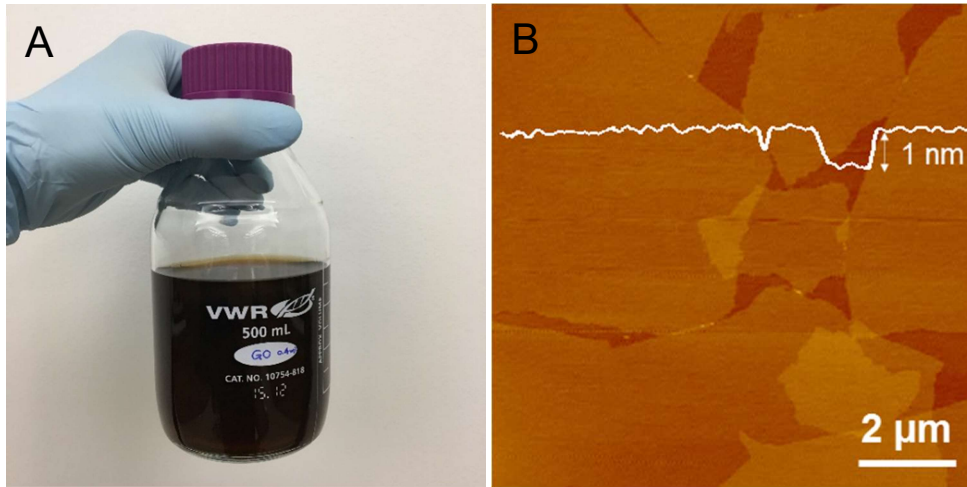


Figure S6.1. (A) Picture of a bottle of GO solution. (B) AFM image of GO flakes deposited on a silicon substrate.

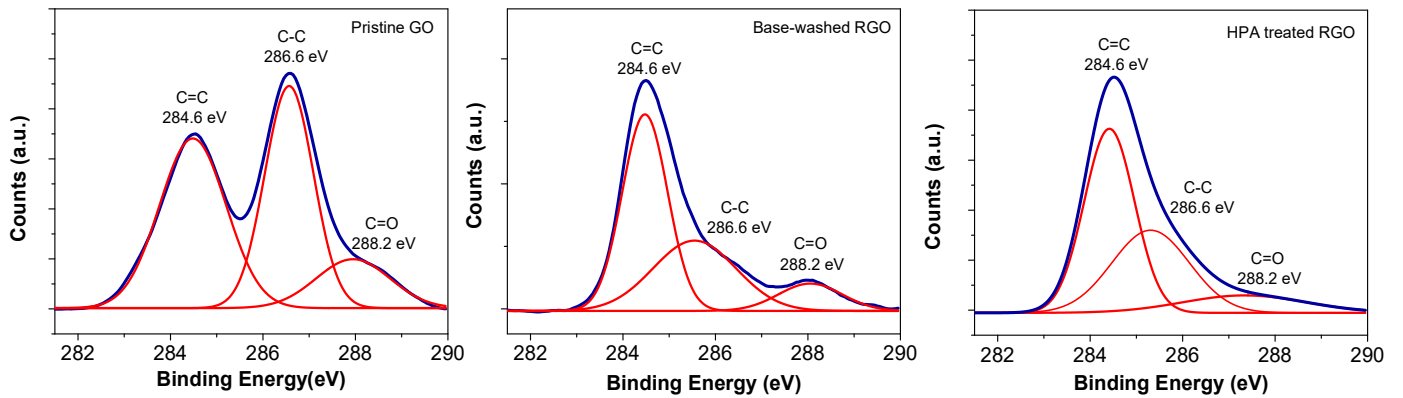


Figure S6.2. XPS spectra of pristine GO, base-washed RGO and HPA-treated RGO.

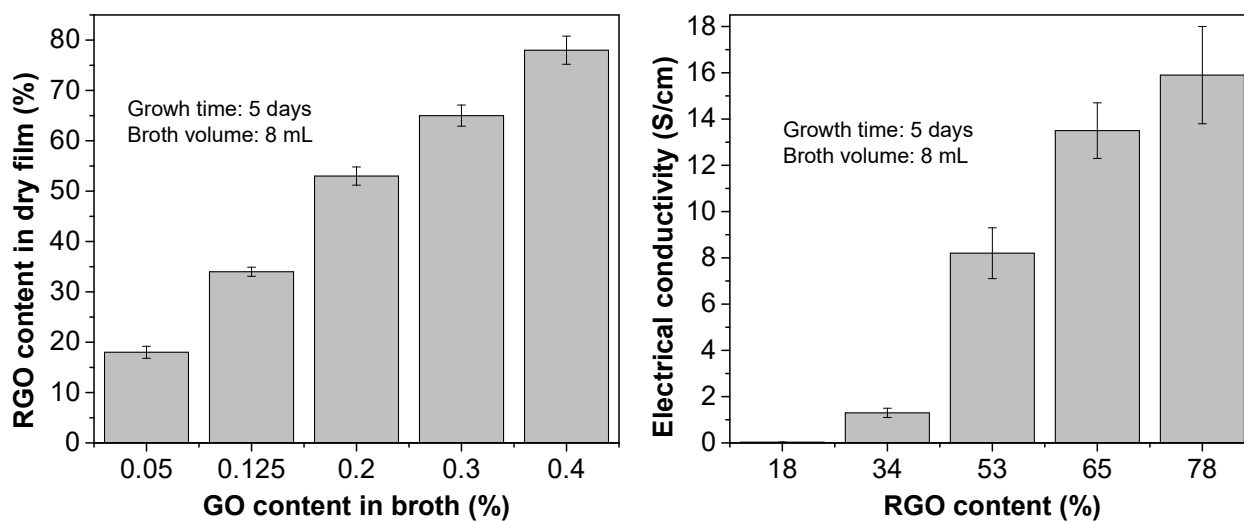


Figure S6.3. RGO content in dry film can be tuned by varying GO content in broth. Electrical conductivity of RGO/BNC films with various RGO contents.

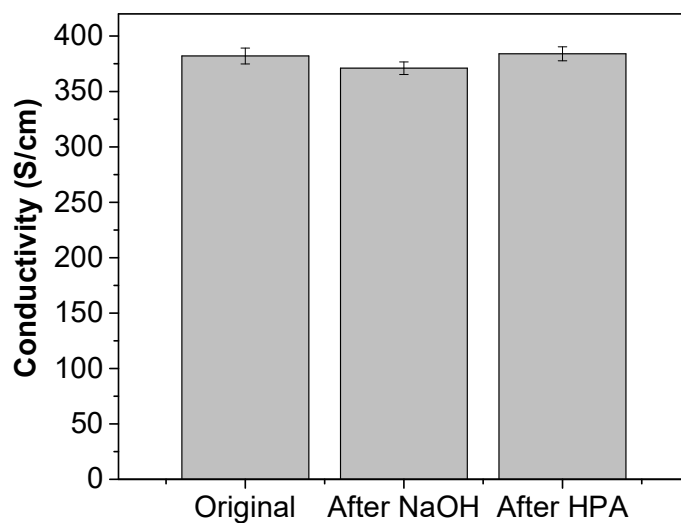


Figure S6.4. Effect of solution processes on pristine PEDOT:PSS deposited on a silicon substrate.

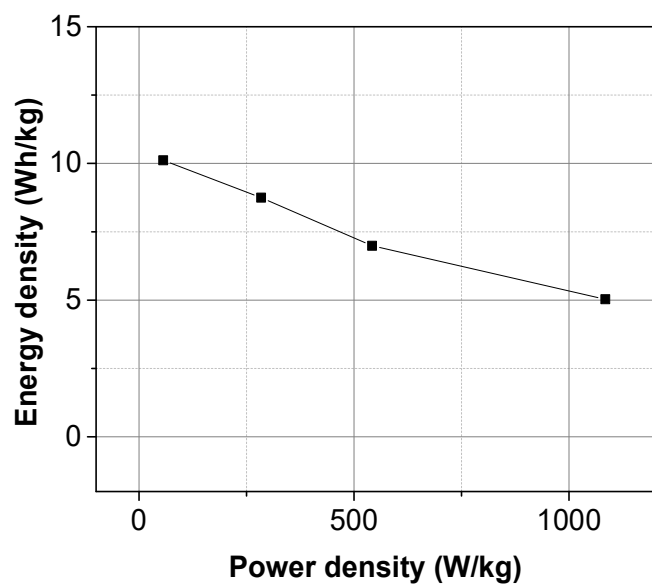


Figure S6.5. Specific energy and power densities of the Flexible device.

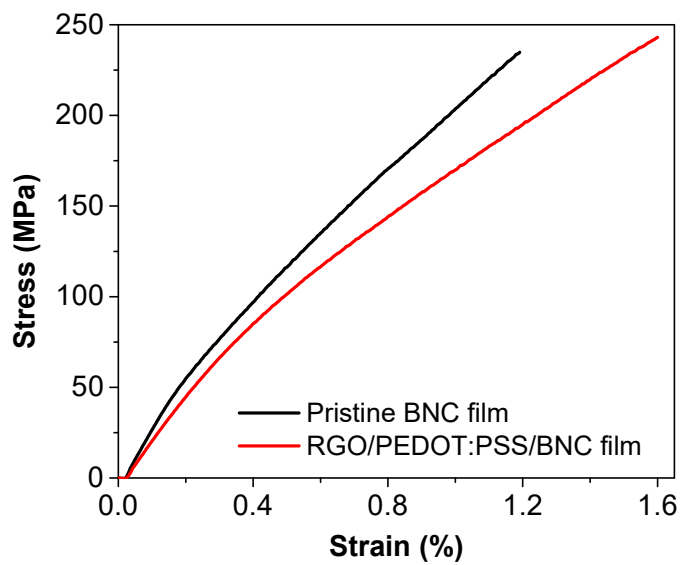


Figure S6.6. Strain-stress curve of pristine BNC and RGO/PEDOT:PSS/BNC.

Table S6.1: The comparison of recent flexible paper- based and some carbon-based supercapacitors.

Material name	Electrolyte	Electrode Capacitance	Current collector for device
PEDOT on RGO ¹³⁴	Not available	108 F g ⁻¹ at 0.3 A g ⁻¹	Not available
PEDOT:PSS-RGO film ¹³⁵	1 M H ₃ PO ₄	52.7 F g ⁻¹ at 10 mV s ⁻¹	Yes
PANI- BNC paper ¹⁵¹	1 M H ₂ SO ₄	273 F g ⁻¹ at 0.2 A g ⁻¹	Not available
PEDOT-paper ³¹⁷	1 M H ₂ SO ₄	115 F g ⁻¹ at 0.4 A g ⁻¹	No
CNT-BNC paper ¹⁴⁹	1 M H ₂ SO ₄	50.5 F g ⁻¹ at 1 A g ⁻¹	No
Graphene-cellulose paper ³¹⁸	1 M H ₂ SO ₄	120 F g ⁻¹ at 1 mV s ⁻¹	Yes
PANI-Au paper ³¹⁹	1 M H ₂ SO ₄	212 F g ⁻¹ at 0.2 A g ⁻¹	Yes
BNC-MWCNTs-PANI paper ¹⁵²	1 M H ₂ SO ₄	656 F g ⁻¹ at 1 A g ⁻¹	No
Graphite/Ni/Co ₂ NiO ₄ - paper ³²⁰	1 M H ₂ SO ₄	734 mF cm ⁻² at 5 mV s ⁻¹	Yes
CNTs/MnO ₂ /CNTs paper ¹⁴²	1 M Na ₂ SO ₄	327 F g ⁻¹ at 10 mV s ⁻¹	No
PPy@nanocellulose paper ¹⁴⁰	2 M NaCl	185 F g ⁻¹ at 2 mA cm ⁻²	Yes
<i>p</i> -BC@MnO ₂ paper ¹⁵⁰	1 M Na ₂ SO ₄	173.32 F g ⁻¹ at 1 A g ⁻¹	Yes
This work	PVA-H ₂ SO ₄	373 F g ⁻¹ at 1 A g ⁻¹	No

Curriculum Vitae

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EDUCATION

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PUBLICATIONS

Refereed Journal Publications

1. **Jiang, Q.**; Tian, L.; Liu, K.-K.; Tadepalli, S.; Raliya, R.; Biswas, P.; Naik, R. R.; Singamaneni, S., Bilayered Biofoam for Highly Efficient Solar Steam Generation. *Advanced Materials* 2016, 28 (42), 9400-9407.
2. **Jiang, Q.**; Singamaneni, S., Water from Wood: Pouring through Pores. *Joule* 2017, 1 (3), 429-430.
3. **Jiang, Q.**; Gholami Derami, H.; Ghim, D.; Cao, S.; Jun, Y.-S.; Singamaneni, S., Polydopamine-filled bacterial nanocellulose as a biodegradable interfacial photothermal evaporator for highly efficient solar steam generation. *Journal of Materials Chemistry A* 2017, 5 (35), 18397-18402.

4. **Jiang, Q.**; Kacica, C.; Soundappan, T.; Liu, K.-k.; Tadepalli, S.; Biswas, P.; Singamaneni, S., An in situ grown bacterial nanocellulose/graphene oxide composite for flexible supercapacitors. *Journal of Materials Chemistry A* 2017, 5 (27), 13976-13982.
5. **Jiang, Q.**; Chandar, Y. J.; Cao, S.; Kharasch, E. D.; Singamaneni, S.; Morrissey, J. J., Rapid, Point-of-Care, Paper-Based Plasmonic Biosensor for Zika Virus Diagnosis. *Advanced Biosystems* 2017, 1 (9), 1700096.
6. **Jiang, Q.**; Xu, T; Deoukchen, G; Liu, K.,-K.; Sun, H; Derami, H. G.; Wang, Z; Tadepalli, S; Jun, Y.S.; Zhang, Q.H.; Singamaneni, S, Catalytically-active Bacterial Nanocellulose-based Ultrafiltration Membrane. *Small*, 2018. 14(15), 1704006.
7. **Jiang, Q.**; Deoukchen, G.; Tadepalli, S.; Liu, K.,-K.; Kwon, H.; Luan, J.; Min, Y.; Jun, Y.-S.; Singamaneni, S., Biofouling-resistente Ultrafiltration Membrane by Harnessing Sunlight. *Environmental Science & Technology* 2018. (In press)
8. Liu, K.-K.; **Jiang, Q.**; Tadepalli, S.; Raliya, R.; Biswas, P.; Naik, R. R.; Singamaneni, S., Wood–Graphene Oxide Composite for Highly Efficient Solar Steam Generation and Desalination. *ACS Applied Materials & Interfaces* 2017, 9 (8), 7675-7681.
9. Tian, L.; **Jiang, Q.**; Liu, K.-K.; Luan, J.; Naik, R. R.; Singamaneni, S., Bacterial Nanocellulose-Based Flexible Surface Enhanced Raman Scattering Substrate. *Advanced Materials Interfaces* 2016, 3 (15), 1600214.
10. Liu, K.,-K; **Jiang, Q.**; Kacica, C.; Derami, H. G.; Biswas, P.; Singamaneni, S., Flexible Solid-state Supercapacitor Based on Tin Oxide/Reduced Graphene Oxide/Bacterial Nanocellulose. *RSC Advances* 2018. (In press)
11. Yuan, M.; **Jiang, Q.**; Liu, K-K.; Singamaneni, S.; Chakrabartty, S., Towards an Integrated QR Code Biosensor: Light-Driven Sample Acquisition and Bacterial Cellulose Paper Substrate. *IEEE Transactions on Biomedical Circuits and Systems*. 2018, 12(3), 452.

12. Derami, H. G.; **Jiang, Q.**; Deoukchen, G.; Cao, S.; Chandar, Y. J.; Morrissey, J. J.; Jun, Y.S.; Singamaneni, S., A Robust and Scalable Polydopamine/Bacterial Nanocellulose Hybrid Membrane for Efficient Wastewater Treatment. *ACS Applied Materials & Interfaces*. 2018. (In press)
13. Wu, X.H.; **Jiang, Q.**; Deoukchen, G.; Singamaneni, S.; Jun, Y.S., Localized Heating by a Photothermal Polydopamine Coating Facilitates a Novel Membrane Distillation Process. *Journal of Materials Chemistry A*. 2018, 6, 18799-18807.
14. Deoukchen, G.; **Jiang, Q.**; Cao, S.; Singamaneni, S.; Jun, Y.S., Mechanically Interlocked 1T/2H Phase MoS₂ Nanosheets for Solar Thermal Water Purification. *Nano Energy* 2018, 53, 949-957.
15. Li, C-B; Chen, W; **Jiang, Q.**; Wang, L; Luo, D, Corrosion Resistance of Ti-based Metallic Glass Coating in Concentrated Nitric Acid. *Materials Chemistry and Physics* 2014, 3, 143, 900-903.
16. Tadepalli, S.; Kuang, Z.; **Jiang, Q.**; Liu, K.-K.; Fisher, M. A.; Morrissey, J. J.; Kharasch, E. D.; Slocik, J. M.; Naik, R. R.; Singamaneni, S., Peptide Functionalized Gold Nanorods for the Sensitive Detection of a Cardiac Biomarker Using Plasmonic Paper Devices. *Scientific Reports* 2015, 5, 16206.
17. Li, C.; Chen, W.; **Jiang, Q.**; Wang, L.; Luo, D., Corrosion resistance of Ti-based metallic glass coating in concentrated nitric acid. *Materials Chemistry and Physics* 2014, 143 (3), 900-903. 14. Tian, L.; Luan, J.; Liu, K.-K.; **Jiang, Q.**; Tadepalli, S.; Gupta, M. K.; Naik, R. R.; Singamaneni, S., Plasmonic Biofoam: A Versatile Optically Active Material. *Nano Letters* 2016, 16 (1), 609-616.
18. Kolluru, C.; Gupta, R.; **Jiang, Q.**; Williams, M.; Noel, R.K.; Singamaneni, S.; Prausnitz, M. R., Plasmonic Paper Microneedle Patch for On-patch Detection of Biomarkers in Dermal Interstitial Fluid. (In preparation)
19. Wang, C.; Sun, H.; Luan, J.; **Jiang, Q.**; Tadepalli, S.; Morrissey, J. J.; Kharasch, E. D.; Singamaneni, S., Metal-Organic Framework Encapsulation for Biospecimen Preservation. *Chemistry of Materials*, 2018, 30 (4), 1291-1300.

20. Tadepalli, S.; Wang, Z.; Liu, K.-K.; **Jiang, Q.**; Slocik, J.; Naik, R. R.; Singamaneni, S., Influence of Surface Charge of the Nanostructures on the Biocatalytic Activity. *Langmuir* 2017, 33 (26), 6611-6619.
21. Liu, K.-K.; Tadepalli, S.; Wang, Z.; **Jiang, Q.**; Singamaneni, S., Structure-dependent SERS activity of plasmonic nanorattles with built-in electromagnetic hotspots. *Analyst* 2017, 142 (23), 4536-4543.
22. Luan, J.; Liu, K.-K.; Tadepalli, S.; **Jiang, Q.**; Morrissey, J. J.; Kharasch, E. D.; Singamaneni, S., PEGylated Artificial Antibodies: Plasmonic Biosensors with Improved Selectivity. *ACS Applied Materials & Interfaces* 2016, 8 (36), 23509-23516.
23. Wang, C.; Sudlow, G.; Wang, Z.; Cao, S.; **Jiang, Q.**; Tadepalli, S.; Morrissey, J. J.; Kharasch, E. D.; Achilefu, S.; Singamaneni, S., Metal-organic Framework Encapsulation Preserves the Bioactivity of Protein Therapeutics. *Nanoscale*, 2017. (In press)
24. Santino, L. M.; Hwang, E.; Diao, Y.; Lu, Y.; Wang, H.; **Jiang, Q.**; Singamaneni, S.; D'Arcy, J. M., Condensing Vapor Phase Polymerization (CVPP) of Electrochemically Capacitive and Stable Polypyrrole Microtubes. *ACS Applied Materials & Interfaces* 2017, 9 (47), 41496-41504.
25. Xie, W.; Tadepalli, S.; Park, S.; Kazemi-Moridani, A; **Jiang, Q.**; Singamaneni, S.; Lee, J., Extreme Mechanical Behavior of Nacre-Mimetic Graphene-Oxide and Silk Nanocomposites. *Nano Letters* 2018, 18 (2), 987-993.
26. Luan, J.; Morrissey, J.; Wang, Z.; Derami, H.; Liu, K.-K.; Cao, S.; **Jiang, Q.**; Wang, C.; Kharasch, E.; Naik, R.; Singamaneni, S., Add-on Plasmonic Patch as a Universal Fluorescence Enhancer. *Light Sci. Appl.* 2018, 7,29.

Conference Proceeding

1. **Jiang, Q.**; Tian, L.; Liu, K.-K.; Singamaneni, S., Bacterial Nanocellulose and Reduced Graphene Oxide Aerogel for Solar Steam Generation. Boston, USA, MRS Fall, Dec. 2015
2. **Jiang, Q.**; Tian, L.; Liu, K.-K.; Luan, J.; Naik, R.R.; Singamaneni, S., Ultra-Smooth Bacterial Nanocellulose-Based SERS Swab. Boston, USA, MRS Fall, Dec. 2015

3. Tian, L.; Liu, K.-K.; Luan, J.; **Jiang, Q.**; Tadepalli, S.; Naik, R.R.; Singamaneni, S., Plasmonic Aerogels from Bacterial Nanocellulose. Boston, USA, MRS Fall, Dec. 2015

PATENTS

1. Photothermally-Active Bilayered Biosystems for Highly Efficient Water Purification Via Solar Steam Generation. (WU OTM Case No. 016682, filed in August 2016)
2. Rapid Point-of-care Assay for Zika Virus Infection. (WU OTM Case No. 016701, filed in August 2016)
3. Ultrafiltration Membrane based on Bacterial Nanocellulose and Graphene Oxide. (WU OTM Case No. 016792, filed in September 2016)