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Complete Genome Sequence of A388, an Antibiotic-Resistant Acinetobacter baumannii Global Clone 1 Isolate from Greece

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ABSTRACT Acinetobacter baumannii isolate A388, recovered in Greece in 2002, represents a distinct antibiotic-resistant lineage of global clone 1 (GC1) producing the OXA-58 carbapenemase. We present the complete 4.332-Mbp genome sequence (chromosome plus 1 plasmid), generated by combining long (MinION) and short (IIlumina HiSeq) read sequencing data.

he Acinetobacter baumannii isolate A388, recovered in 2002 at a hospital in Ioannina, Greece, during a survey of carbapenem resistance in Europe (1), was obtained from the collection of Kevin Towner (Nottingham University Hospital, United Kingdom). A388 was previously shown to be representative of a distinct lineage of global clone 1 (GC1) designated sequence group 6 (SG6) (2, 3). It belongs to sequence type 1 (ST1) (Institut Pasteur multilocus sequence typing scheme) and ST439 (Oxford scheme; formerly ST248) and carries the KL20 gene cluster at the capsule biosynthesis locus and OCL4 at the directing synthesis locus of the outer core of lipooligosaccharide (4, 5). A388 is resistant to the frontline carbapenem antibiotics imipenem and meropenem via carriage of the oxa58 gene (4, 6). It also carries ISAba125 rather than ISAba1 upstream of the ampC gene, increasing the expression of AmpC and giving rise to thirdgeneration cephalosporin resistance (7). Moreover, the A388 lineage appears to be the source of the ISAba125::ampC configuration found in the global clone 2 isolate ACICU from Italy (7). A388 is susceptible to fluoroquinolones but includes the aphA6 aminoglycoside resistance gene in transposon TnaphA6 (4) and genes aacA4, aacC1, and aphA1 in the AbaR30 resistance island (8), conferring resistance to most aminoglycosides (amikacin, gentamicin, kanamycin, neomycin, netilmicin, and tobramycin). AbaR30 is located in the comM gene and also includes sul1 and tetA(A) genes for resistance to older antibiotics (sulfonamides and tetracycline, respectively).

Whole-cell DNA was prepared from cells grown at 37°C in LB inoculated from a frozen stock derived from a single colony, quality controlled for length, and quantified as described previously (9, 10). The DNA was subjected to library preparation, barcoding, and MinION (Oxford Nanopore Technologies) sequencing as described in detail elsewhere (10). Reads were base called using Albacore v1.2.5 and trimmed and demultiplexed with Porechop v0.2.1, as described in detail elsewhere (10). A total of 23,066 reads were obtained with an N_{50} length of 18.1 kbp and 50-fold coverage. The MinION reads (251.9 Mbp; SRA accession number SRR9821831) were retrieved from the raw data and subsampled for length and quality using Filtlong v0.1.0, as described in detail elsewhere (10), resulting in 17,549 reads (200 Mbp), which were used for assembly. These MinION reads were combined with available Illumina HiSeq data (SRA accession number ERX087515; coverage, 98-fold) reported previously (4) using Unicycler v0.4.4 (11) with default parameters.

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The assembly consisted of the chromosome, a contiguous circular sequence of 3,999,012 bp (G+C content of 39.26%), and one 33,036-bp plasmid, pA388. However, the IS26* (IS26-v1) in AbaR30 has caused a chromosomal inversion splitting AbaR30 into two parts (8), namely, AbaR30a (bases 244861 to 290116) and AbaR30b (bases 3763191 to 3781931). Protein-coding, rRNA, and tRNA genes were initially annotated automatically using Prokka v1.12 (12), and annotations of the KL20 and OCL4 polysaccharide biosynthesis loci were refined using current gene nomenclature (5, 13). Annotation of the antibiotic resistance regions and the plasmid were also refined manually. Transposons and dif modules and pdif sites (14) in the plasmid were identified manually and annotated. TnaphA6 is in the chromosome (bases 2265632 to 2268703), and pA388 carries the oxa58 gene in a dif module and the aphA1 gene in transposon Tn4352. In addition to the IS in AbaR30a and AbaR30b, 6 copies of ISAba1 and 11 of ISAba125, as well as 12 of ISAba12 and single copies of ISAba2 and IS26, were detected in the chromosome using ISfinder (https://isfinder.biotoul.fr/). Two potential integrated phage genomes of 23.5 kbp (bases 614654 to 638202) and 56 kbp (bases 1353058 to 1409116) were identified using PHASTER (15). All identified features are annotated in the released GenBank files.

The genome sequence of A388 will underpin studies of the origin and evolution of the unique GC1 lineage found in hospitals in the Eastern Mediterranean.

Data availability. The complete genome sequence has been deposited in DDBJ/ENA/GenBank under the accession numbers CP024418 (chromosome) and CP024419 (pA388). The versions described in this paper are the first versions, CP024418.1 and CP024419.1. The MinION reads have been deposited in the SRA under the accession number SRR9821831. Illumina HiSeq data are available under accession number ERR110081 (SRA accession number ERX087515).

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REFERENCES

- Towner KJ, Levi K, Vlassiadi M, ARPAC Steering Group. 2008. Genetic diversity of carbapenem-resistant isolates of *Acinetobacter baumannii* in Europe. Clin Microbiol Infect 14:161–167. https://doi.org/10.1111/j.1469 -0691.2007.01911.x.
- Evans BA, Hamouda A, Towner KJ, Amyes SG. 2008. OXA-51-like betalactamases and their association with particular epidemic lineages of *Acinetobacter baumannii*. Clin Microbiol Infect 14:268–275. https://doi. org/10.1111/j.1469-0691.2007.01919.x.
- Post V, Hamidian M, Hall RM. 2012. Antibiotic-resistant Acinetobacter baumannii variants belonging to global clone 1. J Antimicrob Chemother 67:1039–1040. https://doi.org/10.1093/jac/dkr586.
- Holt KE, Kenyon JJ, Hamidian M, Schultz MB, Pickard DJ, Dougan G, Hall RM. 2016. Five decades of genome evolution in the globally distributed, extensively antibiotic-resistant *Acinetobacter baumannii* global clone 1. Microb Genomics 2:e000052. https://doi.org/10.1099/mgen.0.000052.
- Kasimova AA, Kenyon JJ, Arbatsky NP, Shashkov AS, Popova AV, Shneider MM, Knirel YA, Hall RM. 2018. Acinetobacter baumannii K20 and K21 capsular polysaccharide structures establish roles for UDP-glucose dehydrogenase Ugd2, pyruvyl transferase Ptr2 and two glycosyltransferases. Glycobiology 28:876–884. https://doi.org/10.1093/glycob/cwx074.
- 6. Towner KJ, Evans B, Villa L, Levi K, Hamouda A, Amyes SG, Carattoli A. 2011. Distribution of intrinsic plasmid replicase genes and their association with carbapenem-hydrolyzing class D β-lactamase genes in European clinical isolates of Acinetobacter baumannii. Antimicrob Agents Chemother 55:2154–2159. https://doi.org/10.1128/AAC.01661-10.
- Hamidian M, Hancock DP, Hall RM. 2013. Horizontal transfer of an ISAba125-activated ampC gene between Acinetobacter baumannii

- strains leading to cephalosporin resistance. J Antimicrob Chemother 68:244–245. https://doi.org/10.1093/jac/dks345.
- Hamidian M, Hall RM. 2018. The AbaR multiple antibiotic resistance islands found in *Acinetobacter baumannii* global clone 1—origin and evolution. Drug Resist Updat 41:26–39. https://doi.org/10.1016/j.drup .2018.10.003.
- Nigro SJ, Wick R, Holt KE, Hall RM. 2018. Complete genome sequence of WM99c, an antibiotic-resistant Acinetobacter baumannii global clone 2 (GC2) strain representing an Australian GC2 lineage. Microbiol Resour Announc 7:e01199-18. https://doi.org/10.1128/MRA.01199-18.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Completing bacterial genome assemblies with multiplex MinION sequencing. Microb Genom 3:e000132. https://doi.org/10.1099/mgen.0.000132.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. https://doi.org/10.1371/journal.pcbi.1005595.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics 30:2068–2069. https://doi.org/10.1093/bioinformatics/btu153.
- Kenyon JJ, Hall RM. 2013. Variation in the complex carbohydrate biosynthesis loci of *Acinetobacter baumannii* genomes. PLoS One 8:e62160. https://doi.org/10.1371/journal.pone.0062160.
- Blackwell GA, Hall RM. 2017. The tet39 determinant and the msrE-mphE genes in Acinetobacter plasmids are each part of discrete modules flanked by inversely oriented pdif (XerC-XerD) sites. Antimicrob Agents Chemother 61:e00780-17. https://doi.org/10.1128/AAC.00780-17.
- Arndt D, Grant JR, Marcu A, Sajed T, Pon A, Liang Y, Wishart DS. 2016. PHASTER: a better, faster version of the PHAST phage search tool. Nucleic Acids Res 44:W16–W21. https://doi.org/10.1093/nar/gkw387.