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Zwitterionic Self-Assembly of L-Methionine Nanogratings on the $Ag(111)$ Surface

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Zwitterionic self-assembly of L-methionine nanogratings on the Ag(111) surface

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The engineering of complex architectures from functional mole cules on surfaces provides new pathways to control matter at the nanoscale. In this article, we present a combined study addressing the self-assembly of the amino acid L-methionine on Ag(111).
Scanning tunneling microscopy data reveal spontaneous ordering in extended molecular chains oriented along high-symmetry subin extended molecular chains oriented along high-symmetry sub strate directions. At intermediate coverages, regular biomolecular gratings evolve whose periodicity can be tuned at the nanometer
scale by varying the methionine surface concentration. Their charscale by varying the methionine surface concentration. Their char acteristics and stability were committed by helium atomic scattering ing. X-ray photoemission spectroscopy and high-resolution scan ning tunneling microscopy data reveal that the L-methionine chaining is mediated by zwitterionic coupling, accounting for both lateral links and molecular dimerization. This methionine molecular recognition scheme is reminiscent of sheet structures in amino acid crystals and was corroborated by molecular mechanics calcula tions. Our findings suggest that zwitterionic assembly of amino acids represents a general construction motif to achieve biomo lecular nanoarchitectures on surfaces.

nanochemistry | scanning tunneling microscopy | supramolecular engineering | surface chemistry | x-ray photoemission spectroscopy

 \mathbf{I} controlled self-assembly of functional molecular species
well defined surfaces is a promising engaged toward the on well defined surfaces is a promising approach toward the
ion of persoacle applitudes (1). Demains this mathed also design of nanoscale architectures (1). By using this methodology, regular low-dimensional systems such as supramolecular clusregular low-dimensional systems such as supramolecular clus ters, chains, or nanoporous arrays can be fabricated (2-6). A wide variety of molecular species as well as substrate materials proved to be useful (7), exploiting noncovalent directional interactions including dipole–dipole coupling $(2, 3)$, hydrogen bridges $(4, 5, 8-11)$, and metal–ligand interactions $(6, 12-15)$. bridges $(4, 5, 8-11)$, and metal-ligand interactions $(6, 12-15)$. With the exception of multiple H-bonded networks or coordination networks or coordination. nation networks incorporating metal centers, it remains challenging to realize robust systems, and there is a need to develop lenging to realize robust systems, and there is a need to devel protocols exploiting stronger intermolecular bonds to realize purely organic low-dimensional architectures. Small biological molecules such as amino acids or DNA base molecules represent an important class of building blocks that are of interest for molecular architectonic on surfaces because they inherently qualify for molecular recognition and self-assembly (16-20). The interaction between biomolecules and solid surfaces is decisive for the development of bioanalytical devices or biocompatible materials (21-23) as well as for a fundamental understanding of protein-surface bonding (24). Moreover, in three dimensions the amino acids provide assets to engineer distinct network strucamino acids provide assets to engineer distinct network struc tures based on zwitterionic coupling schemes $(25-27)$, which may
be actorogized as subgless of ionic self-essembly (29) , and thus be categorized as subclass of ionic self-assembly (28), and thus are promising units to create robust nanoarchitectures. How ever, to date, the advantages of zwitterionic supramolecular synthons have not been exploited in two dimensions, although numerous studies on the adsorption of amino acids on metal surfaces have been performed (29).

In this article, we report a low-temperature scanning tunneling
iorosoomy (STM) invostigation on the self-essembly of microscopy (STM) investigation on the self-assembly of l methionine on the close-packed $Ag(111)$ surface under ultrahigh vacuum conditions. Our results demonstrate that the in terplay between both molecule-molecule and molecule-surface interactions drives a previously uncharacterized methionine self-assembly scenario. We realized extended ID biomolecular nanostructures of distinct widths and with tunable separation controlled by the molecular surface concentration. At intermediate coverages, molecular chains form striking methionine diate coverages, molecular chains form striking method nanogratings, which are ordered mesoscopically in micrometer
domains. Complementery helium atomic sectoring (UAS) ab domains. Complementary helium atomic scattering (HAS) ob servations confirm the self-assembly characteristics and the high regularity of the gratings. O Is and N Is x-ray photoemission spectroscopy (XPS) measurements were performed to deter mine the chemical nature of adsorbed L-methionine layers. They show conclusively that the organized molecules are in their zwitterionic state $(CH_3SCH_2CH_2CH(NH_3)^+(COO)^{-1}$; for a zwitterionic state $(CH_3CH_2CH_2CH(NH_3) \ (COO)$; for a
structure model of the neutral energies are $Fig. 1a)$. Malegular structure model of the neutral species, see Fig. 1a). Molec resolution STM data demonstrate that the chaining reflects both a lateral coupling and a dimerization of L-methionine molecules involving the ammonium and carboxylate groups. This methiomvolving the ammonium and carboxylate groups. This method nine molecular recognition scheme is steered by site-specificant handles to the collection scheme is a second in the specific state. bonding to the silver substrate and is reminiscent of sheet structures in amino acid crystals (26, 27, 30). Elementary molecular mechanics calculations corroborate the associated 2D hydrogen-bonding pattern in which the ionic nature of the functional groups accounts for remarkably stable configurations.
Because the intermolecular coupling is dominated by the amino Because the intermolecular coupling is dominated by the amino
exid functional mooities, it is expected that appears with different acid functional moeities, it is expected that species with different side chains can be used in a similar manner. As such, zwitterionic assembly of amino acids represents a general motif to realize a new class of low-dimensional biomolecular nanoarchitectures.

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Mesoscopic Ordering and Spectroscopic Signature

Upon deposition of small methionine concentrations, STM observations show 1D features on Ag(111) terraces, reflecting molecular self-assembly. This finding is illustrated by the image reproduced in Fig. 1b, taken for a coverage of ≈ 0.05 monolayer (ML), showing ID arrangements with discrete widths of 19 and 38 Å, respectively. The apparent height of these structures varies between 0.8 and 1.5 Å, depending on the applied imaging bias. Moreover, they exhibit striking extensions; for instance, the

Author contributions: J.V.B. designed research; A.S., A.R., W.A., Y.P., A.W.-B., D.C., A.C., and A.M. performed research; A.S., A.R., W.A., Y.P., A.W.-B., D.C., A.C., and A.M. analyzed data; and A.S., A.R., A.M., and J.V.B wrote the paper.

The authors declare no conflict of interest.

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Abbreviations: STM, scanning tunneling microscopy; HAS, helium atomic scattering; XPS, x-ray photoemission spectroscopy; ML, monolayer.

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Fig. 1. The amino acid L-methionine and its 1D ordering on the Ag(111) surface, (a) Structure model of L-methionine in its neutral gas phase state with color-coded atoms. The length of the molecule along its side chain is ~8 Å. (b) STM topographic data show the L-methionine molecules self-assembling into extended 1D arrangements following the closely packed <110> orientations of the substrate ($l = 0.7$ nA, $U = -120$ mV, $\theta \approx 0.05$ ML, $\Gamma_{\text{evap}} \approx 0.5$ ML/min). (Inset) Atomic resolution of Ag(111). (c) On exceeding a critical coverage of ≈ 0.1 ML, the correlated orientation of methionine stripes signals long-range repulsive interactions ($l = 0.8$ nA, $U = -200$ mV, $\theta \cong 0.12$ ML, $\Gamma_{\text{evap}} \cong 2.4$ ML/min)

length of the right 38-Å-wide methionine stripe exceeds 180 nm. Three different orientations are found that follow the close packed <110> high-symmetry substrate orientations [see the Fig. 1b Inset in which the atomic structure of the $Ag(111)$ lattice is depicted]. This is a first indication that site-specific bonding at the surface is decisive in the observed L-methionine self assembly scenario.

The molecular surface concentration plays an important role in the mesoscopic ordering and domain formation of the mo lecular stripes. Although at low coverages any orientation along the close-packed substrate is equiprobable, beyond a critical coverage of ≈ 0.10 ML, domains with mutual alignment appear (see Fig. lc), i.e., there is a mesoscopic ordering of the methi onine stripes. In the data depicted in Fig. lc, the interstripe distances are in the $85-190$ Å range, and their correlated orientation signals long-range interactions. The nature of these presumably indirect substrate-mediated interactions is currently under investigation. The surface electronic structure, which notably includes the $2D Ag(111)$ surface-state free electron gas, is expected to play a preponderant role herein. Indeed, tunneling spectroscopy data evidence a striking ID electron confinement and formation of quantum well states (31), which could mediate long-range interactions (7). Related cases in which surface-state

electrons drive adatom array formation (32, 33) and influence molecular ordering (34) on metal substrates have been reported, molecular ordering (34) on metal substrates have been reported, and it is likely that with the present system the surface states
interface in the malecular celf essembly. A data iled angly is of interfere in the molecular self-assembly. A detailed analysis of the surface electronic structure of the present system will resolve this hypothesis.

On further increasing the coverage, the mesoscopic ordering becomes more regular. Thus, tunable and regular nanogratings can be fabricated. Fig. 2 a and b shows two corresponding examples for coverages of ≈ 0.15 and ≈ 0.38 ML, respectively. In both preparations, all stripes have a single width of 38 Å. The grating periodicity is 275 Å (standard deviation $\sigma \approx 40$ Å) in Fig. 2a and 94 Å ($\sigma \approx$ 9 Å) in Fig. 2b. The nanogratings order in regular domains extending in the micrometer range. An inter esting application for these nanogratings with tunable spacing is their potential use as templates for the design of functional linear arrangements such as nanowires. The thermal stability of the nanogratings has been studied by STM for this purpose, with the outcome being that they are stable up to room temperature with molecular desorption occurring only above \approx 370 K.

HAS experiments confirm the linearity of the L-methionine structures and the steering influence of the substrate symmetry on the molecular nanogratings. In fact, Fig. $2c$ shows the $2D$

Fig. 2. Tuning the self-assembly of 1D L-methionine nanogratings at intermediate coverages. (a) 275 Å ($\sigma \cong 40$ Å) periodicity ($l = 0.8$ nA, $U = -800$ mV, $\theta \cong$ 0.15 ML, $\Gamma_{\text{evap}} \cong 1.8$ ML/min). (b) 94 Å ($\sigma \cong 9$ Å) periodicity ($l = 0.1$ nA, $U = -500$ mV, $\theta \cong 0.38$ ML, $\Gamma_{\text{evap}} \cong 0.8$ ML/min). (c) The 2D HAS diffraction pattern of the L -methionine deposition on Ag(111) (substrate held at \approx 300 K during preparation, $\theta \cong 0.6$ ML) shows the amino acid self-assembling after the sixfold symmetry of the underlying substrate. The red curve corresponds to a single scan at $\Delta k_y = 0$. Symmetrically placed satellite peaks aside the specular peak demonstrate the periodicity of the nanogratings.

5280 | www.pnas.org/cgi/doi/10.1073/pnas.0607867104

Schiffrin et al.

Fig. 3. XPS measurements reveal zwitterionic amino acid assemblies on Ag(111). (a) N 1s signal with a singular peak at $E_{N1s} = 401.15$ eV, corresponding to a NH $+_3$ ammonium group. (b) The O 1s spectrum shows only one peak at E_{O15} = 531.2 eV, corresponding to the resonant oxygen atoms of the carboxylate group. Spectra were obtained for a saturated monolayer structure.

HAS diffraction pattern of the amino acid self-assembly for a 0.6-ML coverage. The hexagonal shape reflects the hexagonal symmetry of the self-assembly, induced by the close-packed $Ag(111)$ surface. Moreover, the diffraction motif confirms that the directions in which the supramolecular structures extend correspond to the $\langle 110 \rangle$ substrate orientations. The periodicity of the self-assembly attributable to interchain long-range inter actions also is reflected in the HAS data. The red curve in Fig. 2c corresponds to a single HAS scan at $\Delta k_y = 0$. The off-specular state of the orientation of the orientat diffraction appearing as symmetrically placed satellite peaks aside the specular peak is a complementary demonstration of the periodicity of the supramolecular arrangement. Finally, HAS substantiates the room temperature stability of the gratings, which is an important feature for their potential application as templates structures (35).

XPS measurements clarify the chemical state of the amino acid moiety. These measurements were performed for a satu rated monolayer preparation and are expected to apply for the entire coverage range in which the same coupling motif is identified by STM throughout (see below). The N Is and O Is XPS spectra in Fig. 3 α and β , respectively, show a single component at $E_{N1s} = 401.15 \text{ eV}$ and $E_{O1s} = 531.2 \text{ eV}$, indicating a unique configuration of the amino acid. The observed energies are markedly shifted with respect to those expected for the neutral species. For comparison, XPS experiments performed on the L-cysteine/Au(110) system demonstrate that the N Is spectra related to the neutral amino group $NH₂$ and the positively charged ammonium group NH_3^+ are characterized, respectively, by a peak at 399.5 eV and a peak at 401.5 eV. Moreover, the O Is spectra of this same system show peaks at 531.2 eV, 532.3 eV, and 533.6 eV, with the first peak corre sponding to the equivalent resonating oxygens of the carboxylate group COO- and the two others corresponding to the chemically inequivalent oxygens of the neutral carboxylic group COOH (36). Therefore, we deduce that the N Is peak at 401.15 eV of our system represents a signature of the positively charged ammonium group $NH₃⁺$, whereas the singular maximum in the O Is spectrum at 531.2 eV reflects the oxygen atoms of the carboxylate group COO~, i.e., the methionine molecules are in their zwitterionic state [the slight differences in binding energy are attributed to the different substrates; note that the super structures occurring in the adsorption of cysteine on Au(110) imply both zwitterionic coupling schemes (36, 37) and substrate reconstructions (38)]. This interpretation is in agreement with observations on the cysteine of the $Pt(111)$ system in which similar trends are encountered (39).

Molecular-Level Observations

The high-resolution STM data in Fig. 4 reveal the L-methionine positioning within the supramolecular structures. The stripes comprise elliptical features with a long axis of 8 Å , which corresponds to the extension of a single molecule along its side chain (see Fig. $1a$). Accordingly, these protrusions are identified with individual molecules bonding in a flat configuration to the surface. The long axis of the methionine ellipses enclose an angle of $60^{\circ} \pm 5^{\circ}$ with respect to the stripe orientation (see Fig. 4b). Moreover, the separation between two adjacent molecules in this direction amounts to 5.8 Å. This distance corresponds to twice the Ag(111) surface lattice constant (Ag atom nn -distance 2.89

Fig. 4. Molecular resolution imaging of L-methionine stripes. (a) Grating of double rows with ~63 Å periodicity ($l = 0.6$ nA, $U = -500$ mV, $\theta \approx 0.12$ ML, Γ_{evap} \approx 2.4 ML/min). (b) Individual molecules appear as elliptical features with a long axis of \approx 8 Å arranged in pairs, with a lateral separation of 5.8 Å corresponding to twice the nearest-neighbor distance between silver surface atoms. This separation and the 60? angle of the molecules with respect to the stripe orientation reflect the influence of the substrate on the molecular self-assembly ($l = 0.9$ nA, $U = -80$ mV). (c) Quadruple methionine rows with 30-Å spacing. Chevron and parallel mutual row orientations coexist ($l = 0.65$ nA, $U = -1,100$ mV, $\theta \approx 0.50$ ML, $\Gamma_{\text{evap}} \approx 3.0$ ML/min). (d) Quadruple molecular row with parallel and chevron configurations. The chevron dimers correspond to a down molecule (green oval) bonded to an up molecule (red oval).

Schiffrin et al.

PNAS | March 27, 2007 | vol. 104 | no. 13 | 5281

 \check{A} ; see Fig. 4b), i.e., the substrate coupling also dictates the row periodicity.

periodicity. The methodine stripes forming the gratings present two discrete widths. The structures shown in Fig. 4 a and c are 19 and 38 Å wide, respectively. The molecular resolution data demonstrate that they consist of either double or quadruple molecular rows in which the methionine is oriented at specific angles with respect to the stripe direction. In both cases, the same pairing scheme of the molecules can be discerned. This finding strongly indicates that the reactive amino groups mediate dimerization. Two different molecular configurations can be observed within a dimer. Either the axis of the molecules are parallel with respect to each other or they form an angle of $120^{\circ} \pm 10^{\circ}$ (chevron rows).

In Fig. 4b, we depict a tentative model for the parallel molecular configuration, where amino-amino dimerization and lateral coupling are accomplished through hydrogen bonding involving the ammonium and carboxylate groups. Chemical and theoretical information concerning the detailed bonding geom etry at the substrate is still missing, but near-edge x-ray absorp tion fine-structure data show that the carboxylate group resides parallel to the surface, in agreement with the modeling. The sulfur atom also may play a secondary role in the lateral or surface bonding through its lone electron pairs. A related zwitterionic bonding scheme was identified in the formation of layered amino acid crystals, where it is associated with appre ciable bonding energies (26, 30). As discussed below, this model is supported by molecular mechanics simulations. From an analysis of the molecular coupling scheme, we concluded that the bonding of the molecules is such that the H atom of the α -carbon points upward with respect to the substrate. This state is designated "up"-methionine, and its counterpart is obtained by flipping the molecule about its axis as "down"-methionine.

Fig. 4d depicts the molecular arrangement in the chevron configuration, which cannot be explained by the exclusive cou pling of up or down species. This observation provides an intriguing possible explanation for the coexistence of the parallel and chevron rows in the nanogratings described above. The mirror symmetry of the latter with respect to the line defined by the molecular dimerization could naturally be explained by the pairing of a row of methionine molecules in the energetically preferred up configuration (circled in red in Fig. 4d) with another one in a down configuration (circled in green in Fig. 4d) being energetically close and where the lateral coupling would imply the changed orientation.

The quadruple rows correspond to a merging of two-molecule chains. Their formation cannot be explained in terms of lateral hydrogen bonding (i) because the terminal CH₃ group at the tail of the molecules is unreactive and (ii) because wider stripes composed of an even number of molecules were not found to

Fig. 5. Two-dimensional commensurate layer struc ture, (a) Molecular resolution imaging of saturated L-methionine monolayer ($l = 0.11$ nA, $U = -250$ mV). (b) Periodicity of the biomolecular self-assembly. The molecular ordering follows the atomic lattice of the Ag(111) substrate. Vectors (\tilde{a}_1 , \tilde{a}_2) define a basis for the unit cell of the Ag(111) atomic lattice $(l = 0.30 \text{ nA})$. $U = 200$ mV). The unit cell of the monolayer structure is marked in white.

exist at intermediate coverages. We suspect the deposition coverage θ and the deposition rate I_{evap} to determine whether the double or quadruple row is more favorable. Although we
cand not establish splittle self-essembly protocols for the could not establish reliable self-assembly protocols for the respective structures, we noted that annealing methionine grat ings at $T \approx 320$ K after deposition promote regular gratings with quadruple rows of up-configurated methionine. Hence, stripes of double or quadruple molecular rows are very close in energy, and the latter must be stabilized by interactions beyond direct intermolecular coupling, presumably mediated by the electronic structure or elastic response of the substrate (7).

Only with coverages exceeding $\approx 0.65-0.70$ ML do the methionine rows merge and highly anisotropic 2D molecular islands evolve. In Fig. 5a, the corresponding L-methionine saturated monolayer structure is depicted. The characteristic pairing fea ture identified above in the molecular stripes readily is identified again. In the corresponding model depicted in Fig. 4b, the red circles represent the $Ag(111)$ atomic lattice. The STM measurements do not allow for us to determine the exact adsorption sites of the molecules at the substrate. The sites on which the molecules are adsorbed in the model are arbitrary, the purpose being to determine the periodicity of the saturated molecular lattice with respect to the substrate periodicity. Considering the vectors \vec{a}_1 , \vec{a}_2 that define the unit cell of the Ag(111) lattice, the lateral methionine ordering along the growth direction parallel to \vec{a}_1 is commensurate, whereas along \vec{a}_2 there is merely a higher-order commensurability. The vectors \bar{b}_1 and \bar{b}_2 defining the unit cell of the supramolecular lattice can be written as $b_1 =$ $2\vec{a}_1$ and $\vec{b}_2 = -(5/2)\vec{a}_1 + (15/2)\vec{a}_2$. Hence, along the growth direction, the molecule is adsorbed at equivalent substrate sites, whereas within a given molecular dimer two molecules bond differently to the surface. Moreover, in the 2D layer, the lateral molecular ordering is strictly parallel, i.e., the chevron arrange ments described above are absent. This ordering is enantiomor phic, which indicates that the chirality of the L-methionine molecule must be expressed in it. Preliminary results with the D-enantiomer confirm this hypothesis, i.e., a mirror-symmetric arrangement is found, and further studies are anticipated to identify possible related chiral resolution processes (40).

A regular chiral ordering thus is favored in the formation of extended domains to which the methionine rows come in close proximity. In agreement, the orientation of the individual mo lecular rows at the interior of the quadruple arrangement always was observed to be strictly parallel. This interpretation implies, assuming structures of the chevron type may form transiently in the self-assembly, that a switching of the entire orientation of the molecules with respect to the substrate is possible (from down to up), which represents a generalization of the chiral switching phenomena of specific molecular groups observed recently (41).

5282 | www.pnas.org/cgi/doi/10.1073/pnas.0607867104

Schiffrin et al.

Fig. 6. Total energy maps of a system composed of two interacting L-methionine molecules obtained with molecular mechanics calculations. The origin of the 2D plots is defined by the center of mass of the immobile molecule, and x-y coordinates represent the position of the center of mass of the second molecule with respect to the immobile one. The color scale indicates total energy of the system versus the relative position between the two molecules. The energy of a
noninteracting two-molecule system defines the zero of the energy sc noninteracting two-molecule system defines the zero of the energy scale, (a) Antiparallel configuration. Amino dimerization through zwitterionic bonding of self-complementary carboxylate and ammonium groups is shown, (b) Parallel configuration. Lateral hydrogen bonding involving ammonium and carboxylate moieties is shown.

Modeling the Zwitterionic Coupling Scheme

To gain further insight into the nature of the 2D H-bonding with the present system, molecular mechanics calculations were performed for a pair of methionine molecules confined into a plane. The classical molecular mechanics force field results lend support to the proposed model for the molecular self-assembly. Only molecule molecule interactions were taken into account in these calculations, neglecting the influence of the $Ag(111)$ substrate on the system, which is caused by the inaccuracy of a classical force field approach to describe surface-induced phenomena. The molecules were taken in their zwitterionic state, and conformational changes caused by molecule-molecule interactions were not considered. The STM data demonstrate that the molecules are lying flat on the substrate. If we assume that the adsorption is caused by interactions between the surface and the reactive sites of the molecules, the motion of the amino acid is restricted to the two translational degrees of freedom on the adsorption plane defined by the sulfur atom, the nitrogen atom, and one of the oxygen atoms of the carboxylate group. We considered a system composed of two L-methionine zwitterions, with their geometry independently optimized. The total energy of the system was determined with respect to the relative position of the two molecules in the adsorption plane.

Fig. 6 a and b represents total energy maps of the twomolecule system when the molecules are in antiparallel (i.e., one molecule rotated of 180? on the adsorption plane with respect to other) and parallel configurations, respectively. The origin of the maps corresponds to the center of mass of one molecule, and the coordinates are those of the center of mass of the second molecule. The first map indicates that amino dimerization involving the carboxylate and ammonium groups is energetically favorable, supporting our model. The length of the resulting dimer is 18 Å, which is in good agreement with the experimental data. The distance between a hydrogen atom of the ammonium group and an oxygen atom of the facing carboxylate group is 1.7 Å, which is in agreement with the hydrogen bond length of 3D amino acids (26). For comparison, the length of the same hydrogen bond in the tentative molecular arrangements shown in Figs. 4b and 5b is \approx 3 Å, which in the calculation still is associated with appreciable bonding. However, for the super position of the STM data, molecules in an unrelaxed configu ration were used under the condition that functional moieties reside on high-symmetry positions. A possible conformational adaptation could reduce the bond length significantly. On the other hand, it is feasible that under the influence of the surface the H bond is stretched to allow for commensurability with the substrate atomic lattice (4, 42).

Furthermore, the calculation for the parallel configuration reveals that the lateral coupling proposed in our model also is

energetically favorable (see Fig. $6b$). Intermolecular bonding involving the carboxylate group and the terminating methyl group of the side chain is very weak, as expected. Here, the hydrogen bond length between an oxygen atom of the carbox ylate group and a hydrogen atom of the ammonium group is 2.1 A, whereas the hydrogen bond length between this same oxygen atom and the terminating methyl group of the side chain is 2.5 A. All these hydrogen bonds are represented by dashed lines in Fig. 6, and the distances are in good agreement with those extracted from the tentative model based on the STM images $(1.9 \text{ Å}$ and 2.2 Å , respectively).

The combination of these results qualitatively supports our model for the molecular self-assembly and explains the forma tion of the double methionine rows. Furthermore, an additional stabilizing factor could result from cooperative effects, as dem onstrated for example in the self-assembly of guanine on Au(111) in which resonance-assisted hydrogen bonding occurs (19). Nevertheless, the existence of quadruple rows cannot be explained in this description and is associated with substrate mediated indirect interactions, which should be considered in further experimental or theoretical studies. Notably, the inter play between surface electronic structure and molecular self assembly remains to be investigated in depth. Thus, further theoretical investigations are planned to explain the binding mechanism between the $Ag(111)$ substrate and the L-methionine molecule and the resulting mesoscopic ordering in greater detail.

Concluding Remarks

In conclusion, we presented a study of the molecular self assembly of L -methionine on the $Ag(111)$ surface. This system provides the possibility to engineer extended biomolecular nanogratings with tunable periodicity that are mesoscopically ordered in regular domains extending in the micrometer range. The long-range linear ordering and the molecular chaining appear as a result of the molecular confinement at the surface. The driving forces underlying the observed self-assembly sce nario are a combination of site-specific adsorption, zwitterionic hydrogen bonding, and long-range indirect interactions. The stability of the biomolecular superlattices, up to room temper ature, coupled with their remarkable tunable geometrical char acteristics make them good candidates as organic templates for the design of functional ID nanostructures or 3D amino acid sheet structures. Because the superlattice formation exploits essentially the functionality of the amino group, the rich chem ical diversity of the side chains can be exploited to realize a variety of nanogratings. Altogether, our findings suggest that zwitterionic assembly of amino acids is a general motif to realize a new class of robust molecular nanoarchitectures on surfaces.

Schiffrin et al.

PNAS | March 27, 2007 | vol. 104 | no. 13 | 5283

Materials and Methods

STM measurements were performed in a custom-designed ultra high vacuum apparatus equipped with a commercial beetle-type low-temperature STM (43) and standard tools for in situ sample preparation and characterization. All experiments were carried out at a base pressure lower than 3×10^{-10} mbar. The Ag(111) sample (a_0 = 4.09 Å at 300 K) was polished chemomechanically and prepared in ultra-high vacuum by repeated Ar⁺ sputtering cycles at an energy of 0.8 keV and currents of typically 4 μ A, followed by annealing at a temperature of 770 K for \approx 10 min.
The enantiomerically pure L-methionine molecules (\ge 99.5%; The enantiomerically pure L-methionine molecules (\geq 99.5%;
Sigma Aldrich, St. Louis, MO) were ween deposited anta the Sigma-Aldrich, St. Louis, MO) were vapor-deposited onto the $Ag(111)$ substrate from a glass crucible heated to a temperature of 370 K. During deposition, the substrate was held at a temperature of \approx 320 K. The methionine coverage on the silver sample was derived from STM data and is given in terms of monolayers, where 1 ML corresponds to a saturated molecular monolayers, where 1 ML corresponds to a saturated molecular layer completely covering the surface. STM topographic image were obtained with an electrochemically etched W tip, with the bias voltage applied to the sample. Data were recorded by constant-current imaging at temperatures \leq 15 K. All presented STM images were smoothed out by low-pass and inverse Fourier transform filtering to remove, respectively, the high-frequency noise and the harmonic noise created by external vibrations. XPS and HAS measurements were performed at the ALOISA beam line (ELETTRA, Trieste, Italy), whereby deposition of L methionine on $Ag(111)$ was performed with the substrate held at room temperature. All XPS spectra have been measured from a freshly deposited film and by keeping the sample temperature

- 1. Barth JV, Costantini G, Kern K (2005) Nature 437:671-679.
- 2. Böhringer M, Morgenstern K, Schneider W-D, Berndt R, Mauri F, Vita AD, Car R (1999) Phys Rev Lett 83:324-327. Car R (1999) *Phys Rev Lett* 83:324-32
- 3. Yokoyama T, Yokoyama S, Kamikado T, Okuno Y, Mashiko S (2001) Nature 413:619-621.
- 4. Barth JV, Weckesser J, Cai C, Gunter P, Burgi L, Jeandupeux O, Kern K (2000) Angew Chem Int Ed 39:1230-1234.
- 5. Theobald JA, Oxtoby NS, Phillips MA, Champness NR, Beton PH (2003) Nature 424:1029-1031.
- 6. Stepanow S, Lingenfelder M, Dmitriev A, Spillmann H, Delvigne E, Lin N, Deng X, Cai C, Barth JV, Kern K (2004) Nature Mat 3:229-233.
- 7. Barth JV (2007) Annu Rev Phys Chem 58:375-407.
- 8. Barth JV, Weckesser J, Trimarchi G, Vladimirova M, Vita AD, Cai C, Brune H, Günter P, Kern K (2002) J Am Chem Soc 124:7991-8000.
- 9. Dmitriev A, Lin N, Weckesser J, Barth JV, Kern K (2002) / Phys Chem B 106:6907-6912.
- 10. Yokoyama T, Kamikado T, Yokoyama S, Mashiko S (2004) / Chem Phys 121:11993-11997.
- 11. Miller C, Cuendet P, Grätzel M (1991) J Phys Chem 95:877-886.
- 12. Dmitriev A, Spillmann H, Lin N, Barth JV, Kern K (2003) Angew Chem Int Ed 41:2670-2673.
- 13. Clair S, Pons S, Brune H, Kern K, Barth JV (2005) Angew Chem Int Ed 44:7294-7297.
- 14. Clair S, Pons S, Fabris S, Baroni S, Brune H, Kern K, Barth JV (2006) J Phys Chem B 110:5627-5632.
- 15. Seitsonen AP, Lingenfelder M, Spillmann H, Dmitriev A, Stepanow S, Lin N, Kern K, Barth JV (2006) J Am Chem Soc 126:5634-5635.
- 16. Kawai T, Tanaka H, Nakagawa T (1997) Surf Sei 386:124-136.
- 17. Kühnle A, Linderoth TR, Hammer B, Besenbacher F (2002) Nature 415:891-893.
- 18. Chen Q, Richardson NV (2003) Nat Mater 2:324-328.
- 19. Otero R, Schock M, Molina LM, Laegsgaard E, Stensgaard I, Hammer B, Besenbacher F (2004) Angew Chem Int Ed 44:2-6.
- 20. Kelly REA, Kantorovich LN (2006) J Mater Chem 16:1894-1905.
- 21. Kasemo B (2002) Surf Sei 500:656-677.
- 22. Sarikaya M, Tamerler C, Jen AKY, Schulten K, Baneyx F (2003) Nat Mater 2:577-585.
- 23. Preuss M, Schmidt WG, Bechstedt F (2005) Phys Rev Lett 94:236102.
- 24. Ghiringhelli LM, Schravendijk P, Sitte LD (2006) Phys Rev B 74:035437.

below 150 K to minimize the radiation damage. The reported XPS spectra have been taken with an overall energy resolution of 300 meV (44) with a beam of photon energy 596.7 eV. The photon beam intensity was kept low within a reasonable noise to-signal ratio. In fact, we observed that exposures to the synchrotron radiation beam causes molecular damage, mani fested in modification of XPS peak profiles (45, 46), which can be reduced to substantially lower rates when the sample is kept below 300 K (47). The binding energy on the shown spectra was calibrated with respect to the substrate Fermi level. The XPS raw data were treated by subtracting the background signal caused by inelastically scattered photoelectrons and fitting with Voigt peaks. The HAS data were obtained at room temperature with an incident He beam of energy 19 meV and wave vector 6.3 \AA^{-1} . For the molecular mechanics calculations, the software package HyperChem Professional 7.51 was used (Hypercube, Inc., Gainesville, FL). The optimal geometrical configuration for the isolated molecules was determined with the semiempirical MNDO/d method. The classical molecular mechanics MM+ force field was used to determine the energy of the two methionine system.

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- 25. Schade B, Fuhrhop J-H (1998) New J Chem 22:97-104.
- 26. Dalhus B, Görbitz CH (2004) J Mol Struct (Theochem) 675:47-52.
- 27. Banno N, Nakanishi T, Matsunaga M, Asahi T, Osaka T (2004) J Am Chem Soc 126:428-429.
- 28. Faul CFJ, Antonietti M (2003) Adv Mater 15:673-683.
- 29. Barlow SM, Raval R (2003) Surf Sei Rep 50:201-341.
- 30. Dalhus B, Görbitz CH (1999) Acta Crytallogr C 55:1105-1112.
- 31. Pennec Y, Auwärter W, Schiffrin A, Weber-Bargioni Riemann A, Barth JV (2007) Nature Nanotechnol 2:99-103.
- 32. Merrick ML, Luo W, Fichthorn KA (2003) Prog Surf Sci 72:117-134.
- 33. Silly F, Pivetta M, Ternes M, Patthey F, Pelz JP, Schneider W-D (2004) Phys Rev Lett 92:16101.
- 34. Sykes ECH, Han P, Kandel SA, Kelly KF, McCarty GS, Weiss PS (2003) Ace Chem Res 36:945-953.
- 35. Otero R, Naitoh Y, Rosei F, Jiang P, Thostrup P, Gourdon A, Laegsgaard E, Stensgaard I, Joachim C, Besenbacher F (2004) Angew Chem Int Ed Engl 43:2092-2095.
- 36. Gonella G, Terreni S, Cvetko D, Cossaro A, Mattera L, Cavalleri O, Rolandi R, Morgante A, Floreano L, Canepa M (2005) J Phys Chem B 109:18003-18009.
- 37. Cossaro A, Terreni S, Cavalleri O, Prato M, Cvetko D, Morgante A, Floreano L, Canepa M (2006) Langmuir 22:11193-11198.
- 38. Kühnle A, Molina LM, Linderoth TR, Hammer B, Besenbacher F (2004) Phys Rev Lett 93:086101.
- 39. Löfgren P, Krozer A, Lausmaa J, Kasemo B (1997) Surf Sci 370:277-292.
- 40. Weckesser J, Vita AD, Barth JV, Cai C, Kern K (2001) Phys Rev Lett 87:096101.
- 41. Weigelt S, Busse C, Petersen L, Rauls E, Hammer B, Gothelf KV, Besenbacher F, Linderoth TR (2006) Nat Mater 5:112-117.
- 42. Clair S, Pons S, Seitsonen AP, Brune H, Kern K, Barth JV (2004) J Phys Chem B 108:19392-19397.
- 43. Meyer G (1996) Rev Sei Instrum 67:2960-2965.
- 44. Floreano L, Naletto G, Cvetko D, Gotter R, Malvezzi M, Marassi L, Morgante A, Santaniello A, Verdini A, Tommasini F, Tondello G (1999) Rev Sci Instr 70:3855-3864.
- 45. Cavalleri O, Gonella G, Terreni S, Vignolo M, Floreano L, Morgante A, Canepa M, Rolandi R (2004) Phys Chem Chem Phys 6:4042-4046.
- 46. Cavalleri O, Gonella G, Terreni S, Vignolo M, Pelori P, Floreano L, Morgante A, Canepa M, Rolandi R (2004) J Phys Cond Matt 16:S2477-S2482.
- 47. Feulner P, Niedermayer T, Eberle K, Schneider R, Menzel D, Baumer A, Schmich E, Shaporenko A, Tai Y, Zharnikov M (2004) Phys Rev Lett 93:178302.

5284 | www.pnas.org/cgi/doi/10.1073/pnas.0607867104

Schiffrin et al.