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Effects of microzooplankton growth and trophic interactions on herbivory in coastal and offshore environments

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ABSTRACT: We performed serial dilution experiments to estimate rates of gross phytoplankton growth (μ) and grazing mortality (m) in both eutrophic (Corpus Christi Bay, Texas, USA) and oligotrophic (offshore Gulf of Mexico) waters. Two parallel experiments were performed in both environments, with seawater pre-screened through 153 or 25 μm mesh to observe the responses of microzooplankton (MZP) to dilution treatments. MZP biomass changed over the duration of the experimental incubations; in several treatments, MZP net growth rates were $>1 \text{ d}^{-1}$. Patterns of growth varied between dilutions and initial screening size. In the eutrophic system, the ratio of phytoplankton grazing mortality rate to gross phytoplankton growth rate (m/μ) was 1.10 ± 0.54 (mean \pm SD) versus 0.41 ± 0.65 when screened through 153 and 25 μm mesh, respectively. This difference was attributed to cascading trophic interactions among MZP size groups leading to suppression of the primary herbivores in the 25 μm fraction and, in turn, a lower value of m . A food web model consisting of multiple trophic levels was constructed to examine the role of MZP growth and trophic interactions on measurements of μ and m . The model, using 3 interacting groups of MZP, was able to reproduce experimental results. Model simulations demonstrated that MZP growth during incubation leads to an overestimation of m . Non-linearity in the phytoplankton growth response curves was due to MZP growth and trophic interactions in these model simulations, as variable feeding responses were not incorporated into the models. Trophic interactions among MZP can provide context to measurements of μ and m and insight into microbial food web efficiency.

KEY WORDS: Serial dilution experiments · Microbial food web · Microplankton · Food web model · Ciliates · Dinoflagellates · Trophic cascades · Zooplankton

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INTRODUCTION

The majority of oceanic primary production is carried out by microalgae that are effectively too small or dispersed to directly support microinvertebrates (Stoecker & Capuzzo 1990, Sherr & Sherr 2002). A large portion of this oceanic primary production is consumed by microzooplankton (MZP), a collection of phagotrophic protists and microinvertebrates $<200 \mu\text{m}$

in size (Calbet & Landry 2004). MZP are critically important in channeling marine primary production to the metazoan food web. Consequently, estimating the grazing impact of MZP on phytoplankton is central to understanding carbon flow and the fate of primary production in marine systems.

The overall impact of MZP is often determined by serial dilution experiments. In this method, the predator–prey encounter rate is progressively decreased by

diluting the microbial community with particle-free water (Landry & Hassett 1982). Phytoplankton growth is assumed to be controlled only by grazers, so that the lower encounter rates in diluted samples result in higher phytoplankton net growth. Further assuming that the population of grazers remains constant during each incubation allows for the MZP-specific grazing rate (m) and gross phytoplankton growth rate (μ) to be estimated from a linear regression of dilution level and net phytoplankton growth rate.

A central feature of serial dilution experiments is the inclusion of the entire MZP community. This yields a composite effect of the natural assemblage of herbivores that is likely more reflective of *in situ* dynamics than herbivory rates determined from controlled experiments using a reduced number of taxa. However, the results of serial dilution experiments do not reflect all potential trophic interactions among MZP. For example, many large ciliates and dinoflagellates selectively consume heterotrophic nanoflagellates (Premke & Arndt 2000, Christoffersen & Gonzalez 2003), and cascading trophic interactions have been observed among organisms in the pico- to microplanktonic size range, indicating the presence of multiple trophic levels among MZP (Reckermann & Veldhuis 1997, Calbet et al. 2001).

MZP include organisms with high growth rates (Banse 1982). Growth rates of herbivorous MZP can approach and surpass those of phototrophic protists in temperate waters (Rose & Caron 2007). Therefore, MZP communities potentially respond rapidly to trophic restructuring created by diluting seawater. These responses have been observed on hourly to daily timescales (Dolan et al. 2000, Agis et al. 2007) and can be expected to occur during dilution experiment incubations, which are typically performed during a single day. In addition to variable growth rates over the dilution gradient, MZP trophic interactions will vary between dilution treatments. For instance, top-down control of herbivores by larger MZP and microinvertebrates may reduce herbivory rates in low dilutions. Conversely, at higher dilutions primary herbivores are relieved from predatory control, resulting in greater herbivory and decreases in apparent phytoplankton growth. Such complexity is potentially concealed when plotting the response of phytoplankton net growth against the dilution gradient.

Trophic interactions affect both the efficiency of the food web and, potentially, measurements of μ and m . These trophic interactions depend upon the structure of the microbial food web (i.e. the relative abundance of MZP groups). If there are multiple trophic transfers among MZP, a large portion of primary production is lost to respiration. Thus, less primary production is available to metazoans relative to systems with a lower

number of trophic transfers. Because the microbial food web structure varies between environments (e.g. Boissonneault-Cellineri et al. 2001, Fileman & Burkill 2001), the amount of primary production consumed within the microbial food web versus what is available to higher trophic levels can also be expected to differ between environments.

In this study, we supplemented experimental data with a numerical food web model to investigate the response of MZP groups to dilution experiments. At both oligotrophic and eutrophic stations along the Texas coast, parallel dilution experiments were performed in which seawater was screened through either a fine or coarse mesh to reduce the dominance of intermediate-sized or large grazers, respectively. In our food web model, we considered multiple trophic levels including 1 primary producer, 1 herbivore, and 2 omnivores to examine the effect of trophic complexity and consumer growth on food web efficiency and dilution experiment results. The goal of this work was to examine how trophic interactions within MZP can influence herbivory rate measurements and food web efficiency.

MATERIALS AND METHODS

Dilution experiments. Eutrophic Corpus Christi Bay (CCB) and the oligotrophic offshore Gulf of Mexico (GOM) were sampled on 7 and 18 June 2001, respectively. The water temperature was 29°C at both locations. Salinity was 25 at CCB and 33 at GOM. Detailed site descriptions, sampling procedures, and experimental protocols are reported elsewhere (First et al. 2007). Briefly, seawater was screened through mesh of either 153 or 25 μm nominal pore size (hereafter, results from the 2 experiments are distinguished by subscripts indicating the mesh size). Triplicate treatments were set up using ratios of 1.0, 0.6, 0.3, and 0.15 mesh-screened water to filter-sterilized (0.22 μm) station water. Note that 'high dilutions' refers to treatments with low concentrations of mesh-screened seawater (e.g. 0.15 and 0.3). To equalize the growth conditions of phytoplankton across the dilution gradient, each treatment was enriched with 4 or 8 μM NH_4Cl for CCB and GOM, respectively. Triplicate control treatments of undiluted water without nutrient additions were used to determine the impact of nutrient enrichment on phytoplankton growth. Treatments were incubated in clear polycarbonate bottles in an outdoor, temperature controlled ($29 \pm 1^\circ\text{C}$) incubator tank with shaded blue Plexiglas for 24 h. In order to keep incubation conditions similar between experiments, we did not attempt to adjust the shading to reflect *in situ* irradiance at these sites, which was not measured in this study.

Phytoplankton concentration was measured by collecting phytoplankton on filters, extracting chlorophyll *a* (chl *a*), and measuring chl *a* fluorescence. Phytoplankton relative size distribution was determined by filtering 10 to 300 ml of whole seawater onto 5 and 0.22 μm pore size Nuclepore filters to determine the contribution of large (>5 μm) phytoplankton to the total chl *a* concentration. Chl *a* was extracted from the filters in 90% acetone overnight at -20°C in the dark and measured on a Turner Designs fluorometer (Welschmeyer 1994). The phytoplankton growth rate was determined by filtering 25 to 300 ml of each treatment through 47 mm diameter 0.22 μm pore size Nuclepore membrane filters prior to and after the incubation period; chl *a* was quantified as described above. Phytoplankton net growth rate (r , d⁻¹) was determined from the logarithmic growth equation

$$r = \ln\left(\frac{P_f}{P_i}\right) \frac{1}{t} \quad (1)$$

where P_i is the initial and P_f the final chl *a* concentration, and t is the incubation time (d). A linear regression was used to fit the relationship between r and dilution. The slope of this relationship is phytoplankton grazing mortality rate (m) and the intercept is gross phytoplankton growth rate (μ). For estimating phytoplankton biomass, a relationship of 23 μg C:μg chl *a* ratio was established by epifluorescence microscope counts of cells with chl *a* fluorescence.

Biomass, growth rates, and production rates of MZP were determined as described in First et al. (2007). Briefly, 20 to 100 ml of pooled treatment water (preserved in 5% Lugol's iodine, final conc.) was settled in chambers, and organisms between approximately 15 and 200 μm in size were counted either from transects or by scanning the entire chamber (100 to 400× magnification). Cell volumes were determined by measuring linear dimensions via an eyepiece micrometer and converting based on appropriate geometrical figures (Wetzel & Likens 1991). Cell biomass was determined from published volume-to-carbon relationships (Putt & Stoecker 1989, Menden-Deuer & Lessard 2000). All ciliates and dinoflagellates were identified as MZP, and the MZP community was characterized into 3 size classes (<20, 20 to 40, and >40 μm) based upon the mean equivalent spherical diameter of the group. MZP growth rates were determined from initial and final biomasses using Eq. (1). Both aplastidic and plastidic dinoflagellate taxa were included among MZP, as many plastidic taxa are mixotrophic (Stoecker 1999). Nanoflagellate biomass was determined by epifluorescence microscope counts of 153 μm-screened water in GOM and CCB experiments for use in the food web model.

Food web model. The food web model used to assess MZP trophic interactions was based on generalized Lotka-Volterra equations that have previously been used to model dilution experiments (Gallegos 1989, Gallegos et al. 1996). In our model, we defined 4 trophic levels consisting of the primary producer and 3 consumers (Fig. 1). The model was designed to investigate herbivore responses to predation pressure and prey abundance; variable MZP response functions were not incorporated. The death rate of the top trophic level (C_4) was assumed to be zero over the incubation time. Simulations were initialized with the biomass values determined in each experiment (Table 1). The primary herbivores (C_2) in the model were considered to include both nanoplankton-sized grazers (i.e. nanoflagellates and small ciliates were included in MZP <20 μm). The biomass values of the 20 to 40 and >40 μm MZP size classes were used as the initial biomass of the 2 omnivores (C_3 and C_4), respectively. Clearance rates (F) and gross efficiencies (b) were defined for each trophic interaction. For example, $F_{4,2}$ is the clearance rate of C_4 on C_2 . Specific estimates of MZP growth efficiencies are not available for these locations. Therefore, gross growth efficiencies of all grazers were set to 30% in all models (Straile 1997). The validity of the food web model was tested using a scenario in which the MZP population did not change (i.e. growth efficiencies = 0%) and MZP consumed only phytoplankton (i.e. no trophic interactions among MZP). In this case, the dilution technique recovered the assigned μ and m (data not shown).

The model was used for 4 distinct tasks. First, 4 model variants (Models I to IV) were created, 1 for each of the 4 data sets (CCB₁₅₃, CCB₂₅, GOM₁₅₃, and GOM₂₅, respectively), by fitting μ (d⁻¹) and MZP clear-

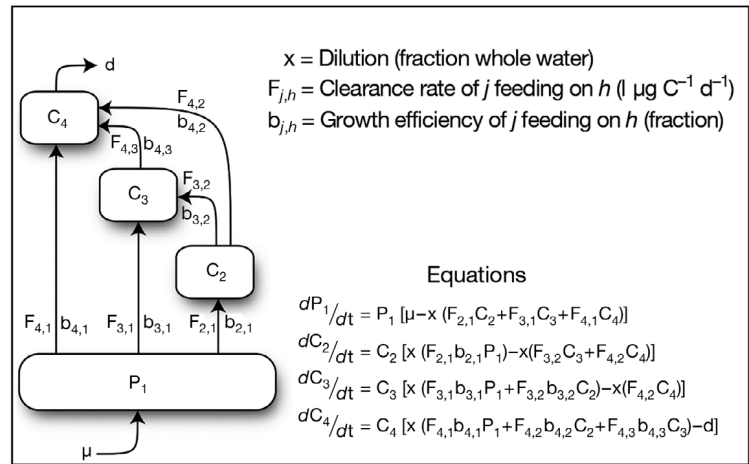


Fig. 1. Conceptual model of the food web in dilution experiment treatments. The concentrations of 4 trophic levels (P_1 , C_2 , C_3 , C_4) are defined by the interactions in the governing equations. The food web consists of phytoplankton (P_1), 1 herbivore (C_2), and 2 omnivores (C_3 and C_4). d : death rate, μ : phytoplankton gross growth rate; t : incubation time. Subscript numbers (1, 2, 3, 4) identify trophic level

Table 1. Parameters and simulation outputs for Models I to IV for seawater collected in Corpus Christi Bay, Texas (CCB) and the Gulf of Mexico (GOM) and pre-screened through 153 or 25 μm mesh (indicated in subscript). See Fig. 1 for a description of the models and how values were determined. Parameter values are truncated at 2 decimal places. P_1 : primary producer, C_2 to C_4 : consumers, $F_{j,h}$: clearance rate of trophic level j feeding on level h

Model Experiment	I CCB ₁₅₃	II CCB ₂₅	III GOM ₁₅₃	IV GOM ₂₅
Initial values				
Initial biomass ($\mu\text{gC l}^{-1}$)				
P_1	130	90	4.0	3.5
C_2	13	13	1.5	1.5
C_3	4.5	4.0	0.4	0.1
C_4	3.5	1.0	0.5	0.1
Growth efficiencies (%)	30	30	30	30
Parameters				
Phytoplankton growth (μ) and grazing mortality (m) rates (d^{-1})				
μ	1.79	1.33	2.11	1.99
m	0.88	0.38	1.37	1.64
Clearance rates ($F_{j,h}$) $\text{l } \mu\text{gC}^{-1} \text{ d}^{-1}$				
$F_{2,1}$	0.05	0.02	0.75	1.06
$F_{3,1}$	0.05	0.03	0.60	0.46
$F_{4,1}$	0.00	0.00	0.00	0.00
$F_{3,2}$	0.18	0.13	1.83	4.32
$F_{4,2}$	0.02	0.12	0.25	1.25
$F_{4,3}$	0.14	0.21	0.77	1.00
Simulation results				
Simulation dilution outputs (mean \pm SD)				
μ	1.93 ± 0.08	1.34 ± 0.06	2.11 ± 0.09	1.94 ± 0.09
m	2.11 ± 0.15	0.49 ± 0.10	1.61 ± 0.15	1.83 ± 0.16
m/μ	1.11 ± 0.14	0.36 ± 0.05	0.76 ± 0.10	0.93 ± 0.14
Adjusted R^2	0.992	0.997	0.998	0.986

ance rates ($\text{l } \mu\text{gC}^{-1} \text{ d}^{-1}$) to these data using a Levenberg-Marquardt optimization algorithm (Press et al. 1988) written in MATLAB (The Mathworks; Table 1). For each model, the community grazing rate was estimated as the summed product of clearance rates and initial biomass of all grazer groups (Fig. 1). Model parameters were estimated sequentially. The model was run with the optimization algorithm allowing only 1 parameter to vary. Using the best fit value for that parameter, the optimization algorithm was repeated, allowing the value of a different parameter to vary. This procedure was repeated until optimized values had been obtained for all parameters. The effect of the order in which parameters were varied was examined using Model I by repeating the above optimization procedure for all permutations of parameter order. The clearance rates generated via the algorithm were within experimentally determined ranges when adjusted for MZP biomass and trophic conditions of the experiment location (Rublee & Gallegos 1989).

Second, each model was used to simulate a dilution experiment using a Monte Carlo algorithm. This is a stochastic approach that produces a large number of real-

izations, allowing for a meaningful statistical analysis of model results. For each model, clearance rates, gross phytoplankton growth rates, and growth efficiencies were randomly selected from normal distributions with a 10% variation around the parameter values listed in Table 1. The model was then solved numerically using an adaptive step size, 4th order Runge Kutta algorithm (the ode45 routine in MATLAB 6.1). This process was repeated 1000 times for each model, producing 1000 replicate simulations of a dilution experiment. The algorithm performed food web simulations evenly spaced along the dilution gradient (0.05 intervals). A weighted least-squares linear regression was used to determine μ , m , and adjusted R^2 values for each simulated experiment (Landry & Hassett 1982), and the mean and standard deviation of μ and m were calculated from 1000 replicate simulations. The covariance between μ and m was determined and used when calculating the standard deviation of m/μ measurements. We used these model parameters to investigate the phytoplankton growth response when the omnivorous MZP (C_3 and C_4) are removed from the experiment, as well as the impact of incubation time on model outputs.

Third, a model sensitivity analysis was conducted by varying the model parameters (p) $\pm 10\%$ of the values in Table 1. Sensitivity (S) was determined as the change in estimated m relative to the baseline values (m_b) as follows:

$$S = \frac{p_b}{m_b} \left(\frac{m - m_b}{p - p_b} \right) \quad (2)$$

where p_b is the baseline value of each parameter (e.g. Miller et al. 2007). Thus, a 10% change in each parameter will correspond to an $S\%$ change in the grazing rate. The model was determined to be sensitive to a parameter if S is $>10\%$.

Fourth, MZP growth efficiencies and interactions were investigated in more detail with additional Monte Carlo simulations based on model III. Model III represents conditions at GOM₁₅₃, where the smallest MZP are expected to be the most important herbivores. Here, MZP were only able to assimilate phytoplankton (i.e. $b_{3,2}$, $b_{4,2}$ and $b_{4,3}$ were set to zero). Growth efficiencies of MZP feeding on phytoplankton were varied, and all other parameters were held constant. MZP apparent growth rates in the undiluted water treat-

ment were calculated by Eq. (1) for each consumer group in each simulated dilution. Mean gross growth efficiency was set to 30%, and the values used in the simulation were randomly selected from a normal distribution within 50% of the mean.

RESULTS

Dilution experiments

Water collected at CCB had a chl *a* concentration of $5.5 \pm 0.91 \mu\text{g l}^{-1}$ (mean \pm SD) and 38% of the total chl *a* was $<5 \mu\text{m}$; GOM had a chl *a* concentration of $0.2 \pm 0.01 \mu\text{g l}^{-1}$ and 100% of the total chl *a* was $<5 \mu\text{m}$. Phytoplankton net growth in nutrient-amended treatments was significantly greater than that in control treatments in both CCB experiments and GOM₂₅ (*t*-test, $p < 0.05$), but not in GOM₁₅₃.

The 25 μm screening significantly reduced measurements of gross phytoplankton growth rate (μ) and grazing mortality rate (m) in CCB experiments (*t*-test, $p < 0.05$) (Table 2). Although μ and m were significantly different, fine-mesh screening significantly altered r only in undiluted water (-0.4 d^{-1} for CCB₁₅₃ vs. 0.8 d^{-1} CCB₂₅, ANOVA, $p < 0.05$) (Fig. 2). Therefore, the large difference in r in undiluted water changed the system output from a slight phytoplankton net loss ($m/\mu > 1$ in CCB₁₅₃) to a net gain ($m/\mu < 1$ in CCB₂₅). In contrast, μ and m were not significantly different between the 2 size fractions in the GOM experiments (*t*-test, $p > 0.05$). The phytoplankton response curve in GOM₂₅ was distinctly non-linear. This difference can be seen by the lower adjusted R^2 values relative to GOM₁₅₃ (Table 2) and the general curve shape of GOM₂₅ (Fig. 2).

The response of MZP to dilution gradients was measured by MZP net growth rates over the incubation period. Ciliate, dinoflagellate, and total MZP responses varied in some cases depending on initial screening size. In the CCB experiments for example, these groups responded oppositely in the 0.3 treatment (Fig. 3). Categorizing MZP by cell size (rather than as

Table 2. Results of linear regression analysis of size-fraction dilution experiments in CCB and GOM, with initial screening through either 153 or 25 μm mesh (denoted by subscript). Values reported are gross phytoplankton growth rate (μ) and grazing (m) (d^{-1} , mean \pm SD) and adjusted regression coefficients (R^2); $p < 0.01$ in all cases

	CCB ₁₅₃	CCB ₂₅	GOM ₁₅₃	GOM ₂₅
μ	1.97 ± 0.54	1.39 ± 0.22	2.11 ± 0.38	1.92 ± 0.92
m	2.16 ± 0.88	0.57 ± 0.36	1.58 ± 0.63	1.73 ± 1.52
m/μ	1.10 ± 0.54	0.41 ± 0.65	0.75 ± 0.44	0.90 ± 1.01
R^2	0.86	0.72	0.87	0.57

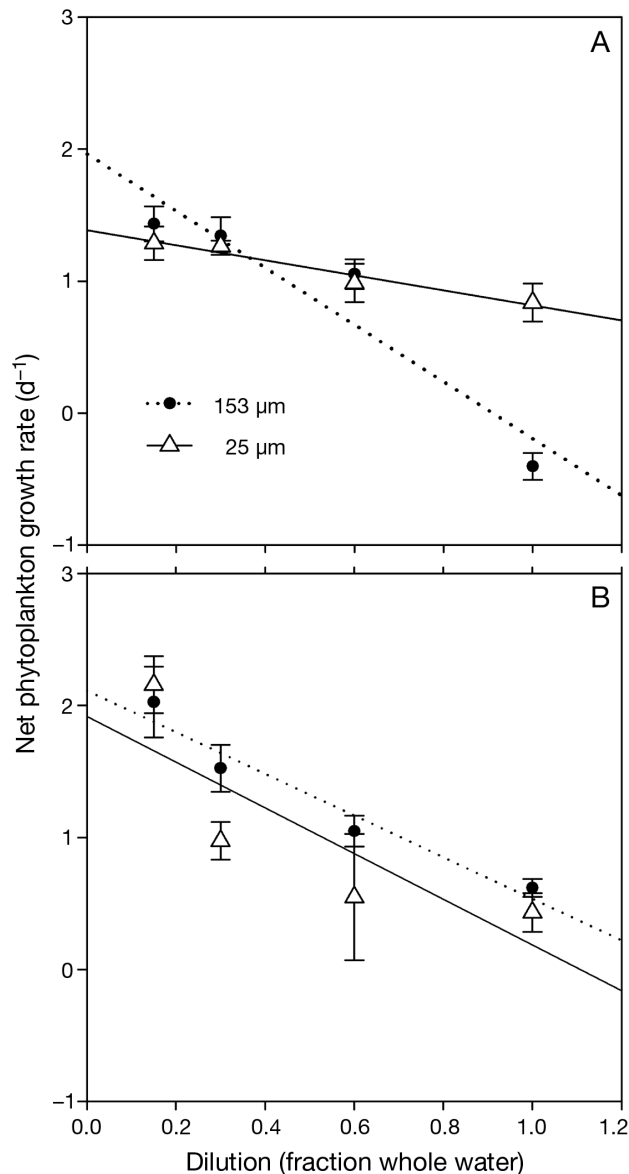


Fig. 2. (A) Corpus Christi Bay (CCB) and (B) Gulf of Mexico (GOM) size-fractionation dilution experiments. Phytoplankton net growth is plotted for serial dilution experiments with seawater initially screened through either 153 or 25 μm mesh. Points are mean \pm SD. Regression data are shown in Table 2

ciliates or dinoflagellates) revealed different patterns of net growth in the dilution experiments (Fig. 4). In CCB₁₅₃ undiluted water, the intermediate size range (20 to 40 μm) showed highest net growth rates, whereas the smallest ($<20 \mu\text{m}$) and largest ($>40 \mu\text{m}$) groups had higher growth rates in CCB₂₅ (Fig. 4). In both 0.3 and 0.15 dilution treatments, the response of the intermediate-sized group was opposite to that of the largest and smallest MZP. For example, in the CCB₁₅₃ experiment, the intermediate-sized MZP declined, while the other size groups increased. The

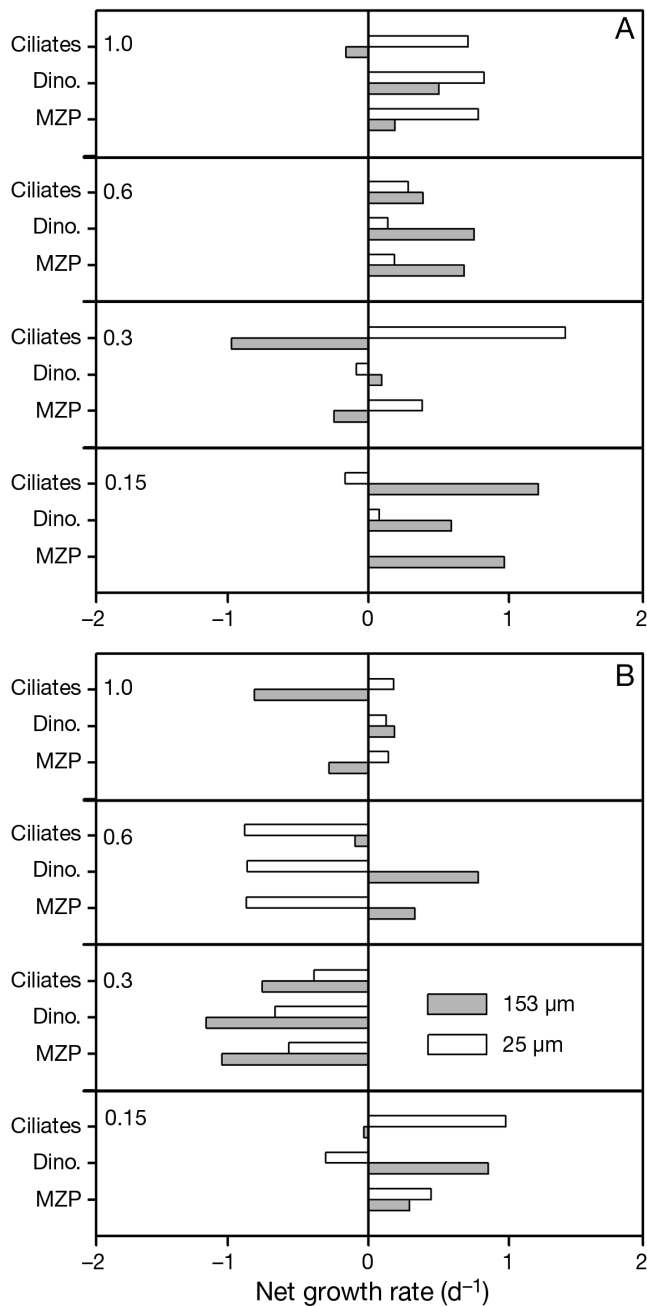


Fig. 3. Microzooplankton (MZP) growth rates (d^{-1}) of ciliates, dinoflagellates (Dino.), and total MZP in all dilution treatments in the 2 size-fractionation experiments in (A) CCB and (B) GOM. The dilution treatments are identified by the fraction of screened seawater (1.0, 0.6, 0.3, 0.15)

opposite pattern was observed in the 0.15 dilution. The alternating growth and production patterns were observed in the 2 highest dilutions at CCB₂₅ but the pattern was reversed (i.e. growth of a size group in CCB₁₅₃ corresponded to a decline of that group in CCB₂₅). In GOM, the MZP response to high dilutions

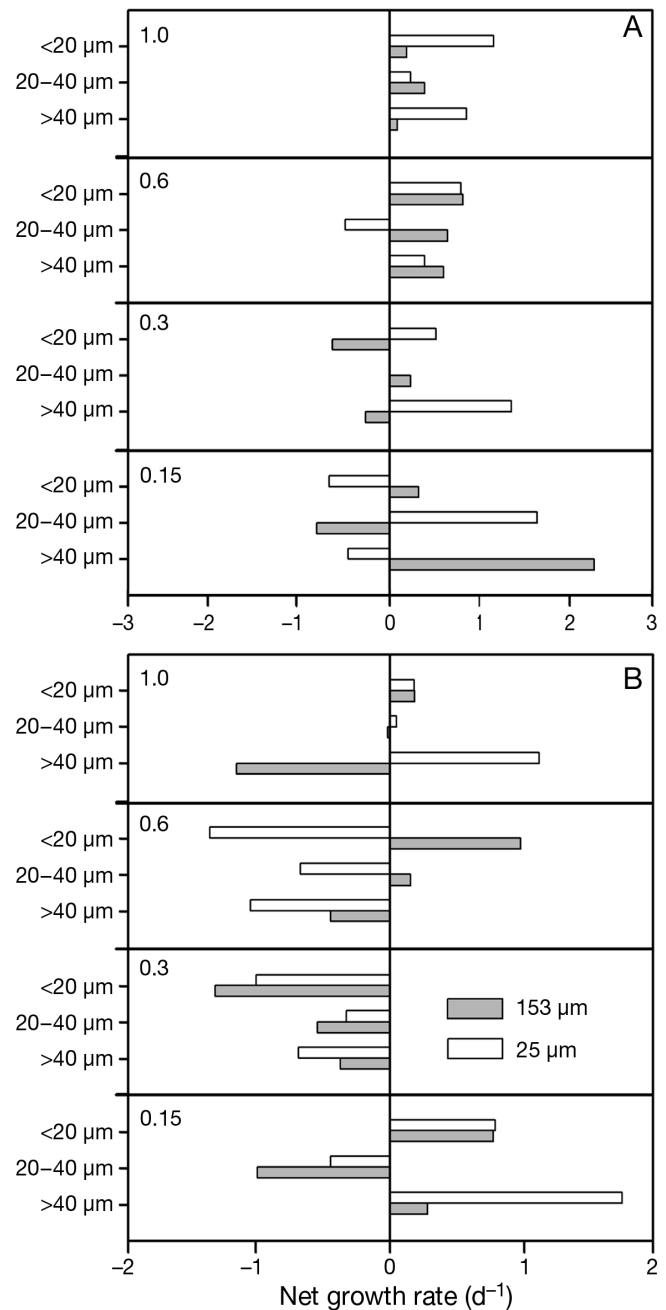


Fig. 4. MZP growth rates (d^{-1}) in all dilution treatments in the 2 size-fractionation experiments in (A) CCB and (B) GOM. Equivalent spherical diameter (ESD) was used to categorize MZP into the following size groups: <20, 20 to 40, and >40 μm . The dilution treatments are identified by the fraction of screened seawater (1.0, 0.6, 0.3, 0.15)

(0.3 and 0.15) did not differ between the 2 size-fraction experiments. However, in the 1.0 and 0.6 dilution treatments, the MZP growth patterns differed. Notably, the largest MZP declined in the 153 μm fraction, but not in the 25 μm fraction. Also, a <20 μm MZP increased in GOM₁₅₃, but declined in GOM₂₅ (Fig. 4).

Food web model simulations

We created 4 parameterizations of the food web model (Models I to IV), each parameterized using data from one of the dilution experiments. All 4 models were sensitive to the initial phytoplankton biomass and μ ; Models II to IV were also sensitive to the initial biomass of C_2 , the primary herbivore (Fig. 5). The clearance rates of the C_2 also caused large variations in model outputs, especially in Models III and IV, whereas Models I and II were sensitive to clearance rates and growth efficiencies of C_3 . All models showed low sensitivity to the clearance rates and growth efficiencies of C_4 , the top MZP.

The order in which parameters were optimized did not affect the final values for the parameters to which the model was most sensitive. For example, Model I showed the highest sensitivity to initial μ and $F_{3,1}$ values. The algorithm chose parameters that were within 10% of the mean value in both cases. Other parameters (such as $F_{4,2}$, which varied by >100% of the mean) were highly variable depending upon the fitting order. However, the model was not sensitive to these initial values.

The correlation coefficients between the model and data were 0.90, 0.73, 0.90, and 0.69 for Models I to IV, respectively. The Monte Carlo method produced 1000 replicate data points for each of the 21 dilution concen-

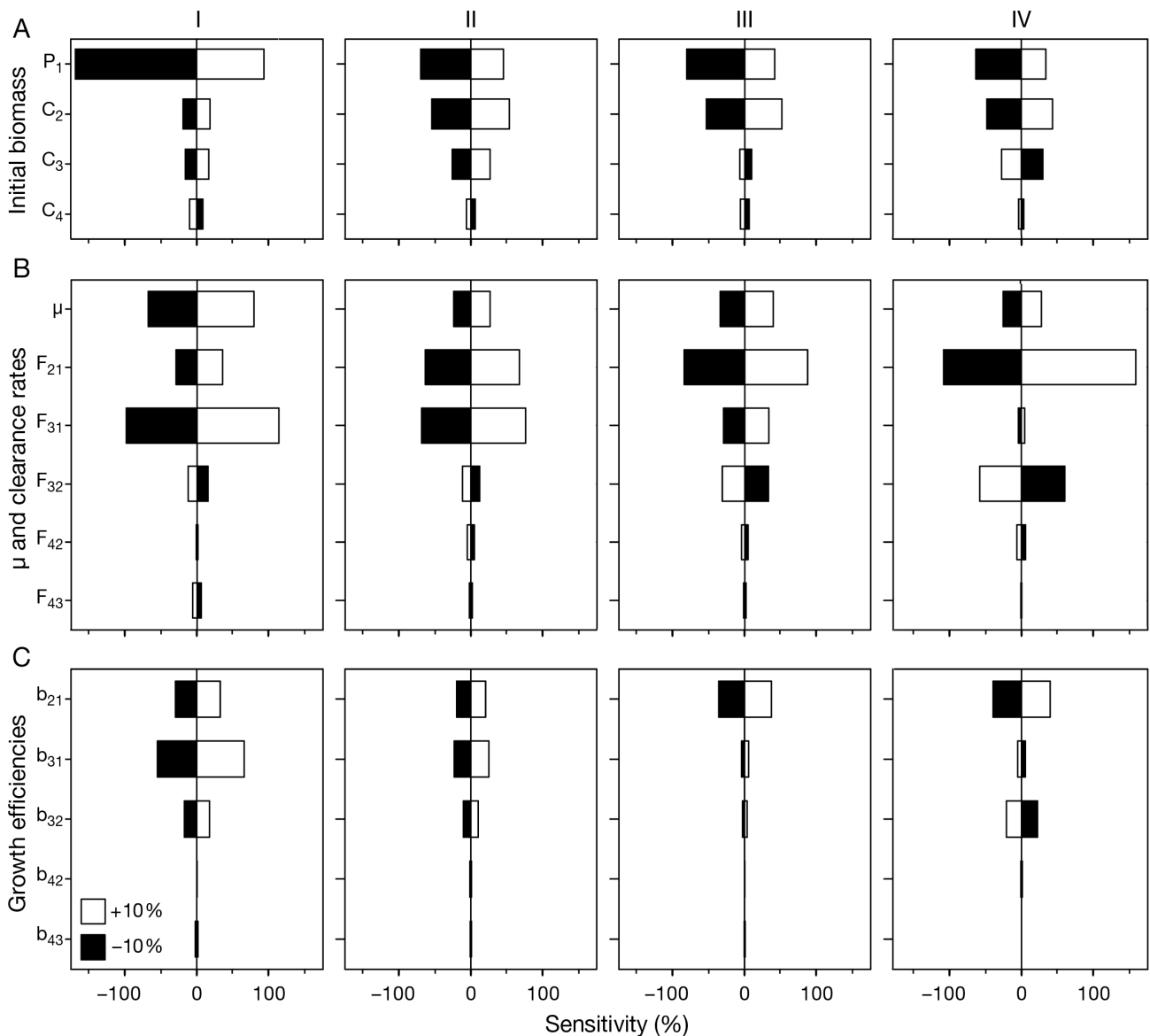


Fig. 5. Sensitivity analysis of Models I to IV. Bars show the percentage change in the response variable, phytoplankton grazing mortality rate (m), to a 10% increase or decrease in the model parameter. (A) Initial biomass values, (B) phytoplankton growth and MZP clearance rates, and (C) growth efficiencies. Parameters are defined in Fig. 1

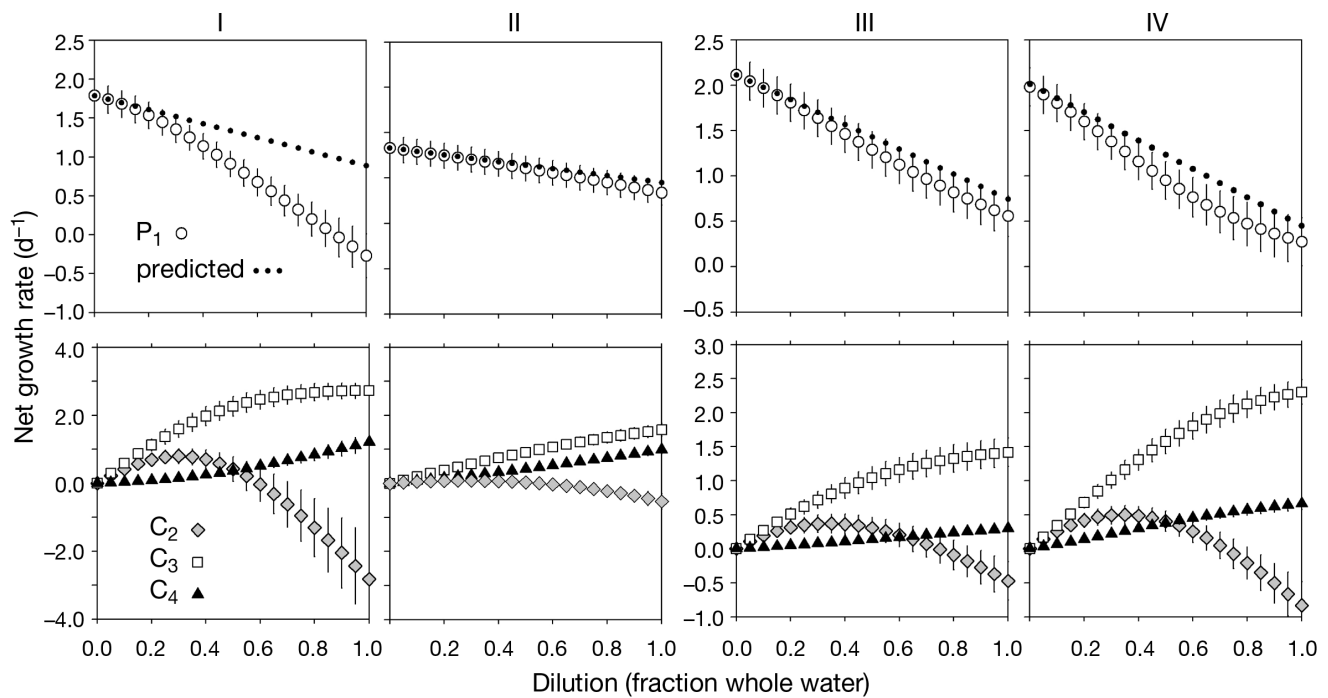


Fig. 6. Dilution plots based on models I to IV (Table 1). Top plots indicate net phytoplankton growth rate (O, mean \pm SD) with the predicted line based upon the assigned μ and m (dotted). Bottom plots show net growth rates of the 3 groups of MZP consumers. Points show the mean \pm SD value of simulation results (N = 1000)

trations (Fig. 6). Linear regression analysis based upon all 21 000 data points resulted in an overestimation of m in all models (Table 1). Phytoplankton grazing mortality rate estimates were 1.1 to almost $2.4 \times$ the actual rates. However, estimates of μ were within 10% of the assigned values listed in Table 1. The growth response of MZP varied between models (Fig. 6). In undiluted water, the change in concentration over the incubation period (measured here by the net growth rate) was high in some cases. At low dilutions, C_3 showed the highest growth rate ($>2 \text{ d}^{-1}$ in Model I), whereas C_2 had negative growth rates in all models.

Grazing rate estimates were more responsive to changes in the growth of C_2 than μ as determined by the fit of exponential growth curves based on Model III (Fig. 7). The changes in the concentrations of C_3 and C_4 showed no strong relationship with μ and m ; therefore, these plots are not shown. These groups have their maximal clearance rates on intermediate trophic levels (Table 1). This top-down control was apparent when the net growth rates of phytoplankton and C_2 are shown with higher trophic levels removed. The results of this analysis, which investigates the presence of trophic cascades in all 4 models by sequentially removing MZP groups, are shown in Fig. 8. With either all 3 MZP or just C_2 and C_3 , the net growth rate of C_2 was negative in all 4 models. Net μ values in the undiluted treatments were higher in Models I, III, and IV when

higher trophic levels (i.e. C_3 and C_4) were included. With the exception of Model II, all food webs show that C_2 is partially controlled by higher trophic levels of MZP. The removal of higher trophic levels increases C_2 growth and, in turn, reduces phytoplankton growth in undiluted water (Fig. 8).

Estimates of μ and m are also affected by incubation time. Note that μ and m rates are reported per day (d^{-1}) to allow for comparisons between these incubations. The shortest incubation time (0.5 d) yielded values of m (d^{-1}) and m/μ closest to the assigned values (Table 3). However, the 1 d incubation best recovered the assigned μ of 2.11 d^{-1} . While the shortest incubation time best recovered the assigned herbivory rate, the secondary consumer (C_3) had high net growth rates even over short incubation periods (Fig. 9). Longer incubation times also led to more complex growth patterns of MZP. In the case of the 1.5 d incubation, the shape of the curve appears as what has been interpreted as a saturated feeding response (Gallegos 1989) (Fig. 9b).

DISCUSSION

The advantage of serial dilution experiments is that, with relatively little manipulation, they generate paired estimates of phytoplankton growth and grazing

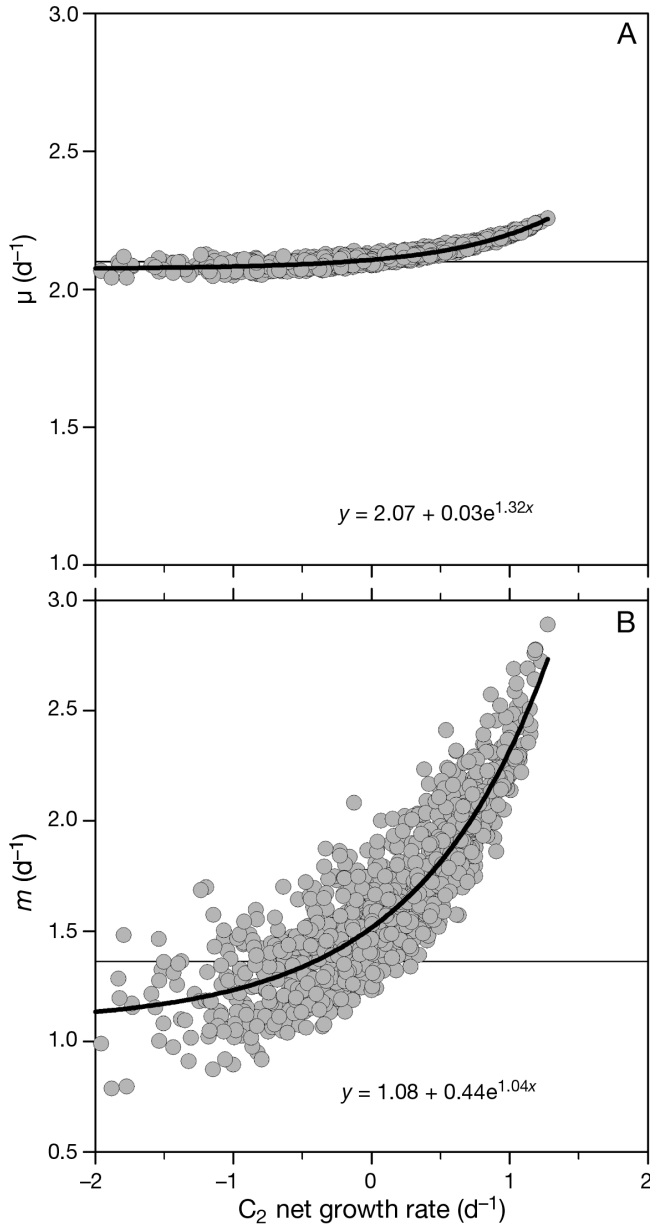


Fig. 7. Dilution results based upon model III. The x-axis is the net growth rate of C_2 in undiluted treatments plotted against (A) μ (d^{-1}) and (B) m (d^{-1}) on the y-axes

mortality rates. Additionally, the ratio of these 2 rates (m/μ) indicates the portion of phytoplankton growth consumed by microherbivores. This ratio has been used to compare microbial dynamics across marine systems (Calbet & Landry 2004). The critical reviews (e.g. Gallegos 1989, Evans & Paranjape 1992) and modifications (Landry et al. 1995, Gallegos et al. 1996) of the dilution technique reflect its widespread use and the importance in constraining the amount of primary production consumed in marine pelagic environments. In this study, we examined how the microbial food web

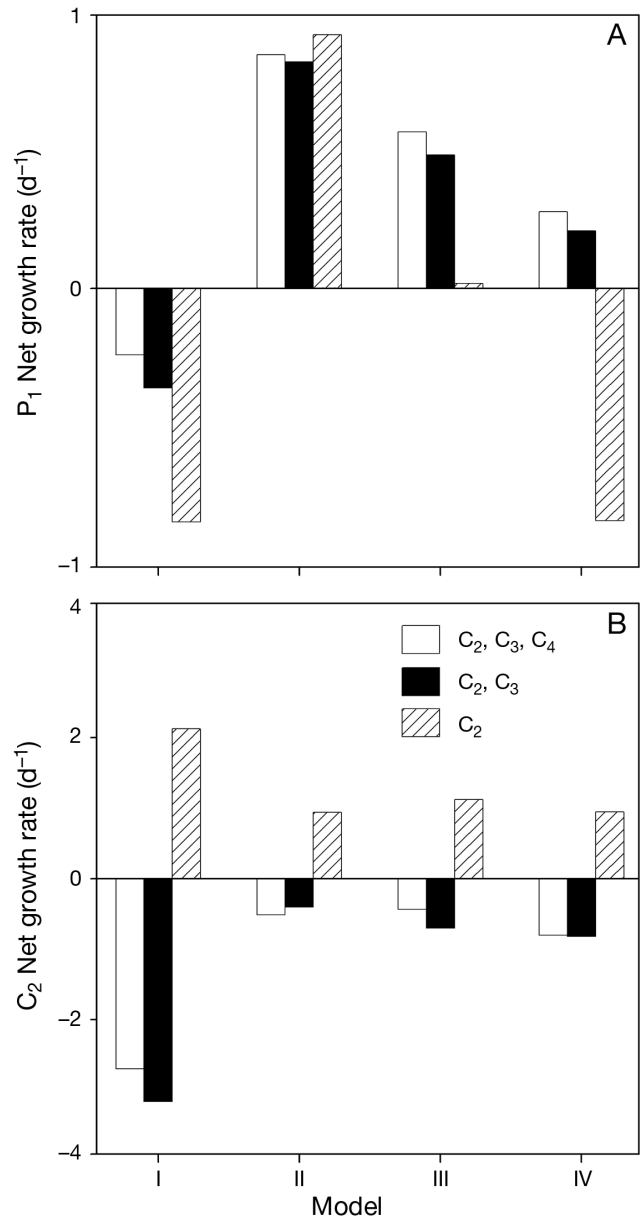


Fig. 8. Model generated rates of (A) P_1 net growth and (B) C_2 net growth in undiluted treatment (1.0 whole water). The model parameters were the same as in Table 1. Models were run with: 3 MZP (C_2 , C_3 , and C_4) included; the top MZP group removed (C_2 , C_3); and the top 2 MZP groups removed (C_2)

at 2 contrasting locations responded to the dilution gradient when the assemblages of MZP were artificially restructured by screening. MZP contain many disparate groups of phagotrophic protists, and these constituent groups are expected to respond differently to the dilution gradient.

Differences in MZP community structure imply the existence of various trophic pathways and food web efficiencies. For example, ciliates and flagellates consume prey that are much smaller (proportional to their

Table 3. Regression analysis results of simulations based upon model III with different incubation times. Assigned μ and m are 2.11 and 1.37 d^{-1} , respectively ($m/\mu = 0.65$). Values shown are μ and m (d^{-1}), m/μ (no units), and adjusted regression coefficient (R^2) based upon 1000 simulations (mean \pm SD). Plots for 1 d incubation times are shown in Fig. 6. Example plots with 0.5 and 1.5 d incubation times are shown in Fig. 9

	Incubation time (d)				
	0.5	0.75	1.0	1.25	1.5
μ	2.13 \pm 0.09	2.12 \pm 0.09	2.11 \pm 0.09	2.08 \pm 0.08	2.01 \pm 0.08
m	1.47 \pm 0.16	1.54 \pm 0.16	1.61 \pm 0.15	1.68 \pm 0.14	1.72 \pm 0.13
m/μ	0.69 \pm 0.10	0.73 \pm 0.10	0.76 \pm 0.10	0.80 \pm 0.09	0.86 \pm 0.08
R^2	1.00	1.00	1.00	0.99	0.98

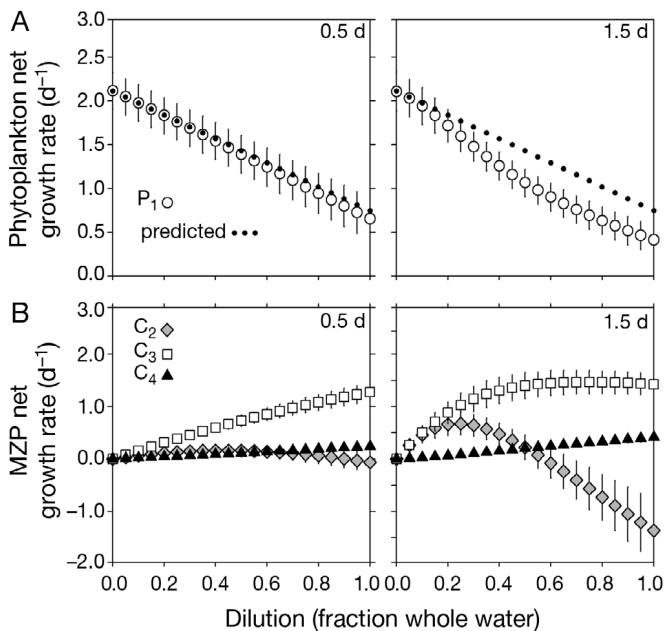


Fig. 9. Dilution plots based on model III, showing (A) phytoplankton and (B) MZP net growth rates along the dilution gradient at incubation periods of 0.5 and 1.5 d. Plots are as described in Fig. 6. The results of the linear regressions are reported in Table 3

body size) than dinoflagellates (Hansen et al. 1994). Thus, an abundance of small and intermediate-sized ciliates can lead to higher grazing mortality of picoplankton. Dinoflagellates consume prey equal to or greater than their body size (Naustvoll 2000), and several dinoflagellate taxa observed at CCB are capable of ingesting larger MZP (Stoecker 1999). Consequently, a high relative abundance of dinoflagellates can lead to predation of intermediate-sized and large MZP.

MZP responses to the dilution gradient

Serial dilutions create a resource gradient for MZP. Trophic links within the MZP are sequentially broken

along the dilution gradient when predator–prey encounters become too rare to sustain the predator or to control the prey. For example, at CCB₁₅₃, ciliates showed the highest net growth rate at the highest dilution. Herbivorous ciliates, released from predation pressure in moderately high dilutions, will limit the phytoplankton net growth in these treatments. Likewise, increases in dinoflagellate biomass (observed in low dilutions in both CCB size fractions) could limit the grazing impact of herbivorous MZP.

In a complex food web, each dilution treatment will select for the dominant trophic pathways. The resulting phytoplankton net growth rate measurements may yield patterns that appear similar to those predicted by MZP feeding responses. In the open GOM, phytoplankton growth was suppressed in the small size fraction (GOM₂₅) relative to the large fraction (GOM₁₅₃) in all treatments except the highest dilution (0.15 fraction whole water). The phytoplankton response along the curve may indicate saturated feeding (i.e. above a certain prey density, the MZP ingestion rate remains constant). This prey concentration at which herbivores are saturated can be only a fraction of the ambient prey density (Gallegos 1989). However, it seems unlikely that herbivorous MZP (with high intrinsic rates) could be prey saturated—especially in oligotrophic waters such as in the open GOM. Instead, this response may be due to suppression of the smallest MZP size category (<20 μm) in the 0.6 and 0.3 fraction whole water in GOM₂₅. In this case, a trophic cascade (where larger MZP graze small MZP, which, in turn, reduce their grazing control on nanograzers) may result in the low net μ values (e.g. Carrias et al. 2001).

Non-linear phytoplankton growth responses to the predator gradient (i.e. dilution) have been explained by MZP feeding responses to phytoplankton (Moigis 2006). The occurrence of feeding responses is well established for both metazoan (e.g. Sarnelle 2003) and protistan (e.g. Waterhouse & Welschmeyer 1995) populations. However, the appearance of non-linear phytoplankton responses to dilution gradients may represent trophic interactions among MZP rather than feeding responses. The food web models demonstrate that changes in the MZP concentrations (measured by MZP net growth rates) produce non-linear phytoplankton responses to the dilution gradients. These non-linear responses were caused by trophic interactions among MZP rather than feeding responses due to the decreasing abundance of phytoplankton along the dilution gradient. For example, the top-down control by higher trophic levels suppressed the growth of the

primary herbivores (C_2) in model simulations. Once released from this top-down pressure, C_2 was able to grow, and phytoplankton growth was reduced in most models. The food web model reflects the experimental and observational studies showing that interactions between primary and secondary consumers cascade to drive the dynamics of primary producers (e.g. Jürgens et al. 1996, Reckermann & Veldhuis 1997).

MZP trophodynamics and food web efficiency

Increased trophic interactions among MZP will reduce the amount of primary production available to metazoan consumers. At each trophic transfer, a large portion of the organic matter ingested is not converted into biomass. The gross growth efficiency (i.e. biomass produced per biomass consumed) indicates the portion of ingested biomass not lost to respiration or egestion. The gross growth efficiency used in this study (30%) is similar to what has been found for a range of aquatic herbivores (Straile 1997). Gross growth efficiency may also depend upon physical characteristics of the prey, such as body size and nutritional composition. For example, small bacterivorous ciliates can have a much higher gross growth efficiency than large omnivorous ciliates feeding upon picoplanktonic prey (Ohman & Snyder 1991).

Transfer of primary production to the metazoan food web depends upon the relative composition of MZP. For example, larger MZP can be more readily ingested by copepods (Liu et al. 2005). Also, ciliates are consumed by copepods at high rates in marine environments (Calbet & Saiz 2005, Schnetzer & Caron 2005). If large MZP (i.e. $>40\ \mu\text{m}$ equivalent spherical diameter) are the organisms most efficiently consumed by metazoans, then the efficiency of transfer to higher trophic levels will depend upon the prey source of these MZP. If these large MZP are herbivores, then the microbial food web will be fairly efficient in transferring primary production to higher levels. Conversely, if multiple trophic transfers occur within the microbial food web, a smaller portion of the original production will be available to metazoan consumers. Because the entire phytoplankton community at the oligotrophic GOM station was $<5\ \mu\text{m}$, large MZP may preferentially consume small and intermediate-sized grazers. Other factors, such as the prevalence of mixotrophic MZP, will also affect the carbon available to metazoans in these 2 contrasting environments. Some ciliates that are capable of mixotrophic growth (e.g. *Laboea* sp.; Sanders 1995, Dolan & Perez 2000) were abundant at the GOM station. Carbon fixation by phagotrophic protists will result in more energy being available for higher trophic levels in the microbial food web.

Assumptions of the food web model

Our food web model did not incorporate feeding response functions, which are commonly used in microbial food web models (Gallegos 1989, Moigis 2006). Changes in MZP clearance rates with prey concentrations (i.e. feeding responses) also yield non-linear phytoplankton growth curves in simulated food webs (Moigis 2006). Our model demonstrated the occurrence of non-linear phytoplankton growth curves due exclusively to MZP growth and trophic interactions. Feeding responses are likely to play a role under conditions of abundance or scarcity of phytoplankton. For example, high abundances of the brown tide-forming alga limit MZP grazing, although this may be due to higher concentrations of inhibitory chemicals during blooms (Buskey & Hyatt 1995). In some cases, MZP are able to consume phytoplankton rapidly during bloom conditions (Strom et al. 2001), indicating that MZP grazing may not be reduced at high prey concentrations. A practical reason for not incorporating feeding responses into this model is that information on MZP feeding responses (which likely depend upon specific characteristics of both prey and predator taxa, as well as ambient temperature) is not readily available. An averaged feeding response of the entire community is also inappropriate considering the trophic diversity incorporated into the MZP grouping.

In our food web model, we considered only 1 group of phytoplankton. The implication of multiple groups is that some phytoplankton will be functionally unavailable to certain grazers. For example, picophytoplankton may not be grazed by the largest herbivores. Considering only 1 group of phytoplankton is less reflective of natural systems where phytoplankton span a large size range. Also, delineating the MZP into 3 discrete groups is an oversimplification of real world dynamics. For example, we did not include the possibility that intermediate or upper trophic levels (i.e. C_3 or C_4) may ingest organisms on the same or higher trophic level (e.g. Hansen & Calado 1999), nor did our model include the possibility for mixotrophic growth (Stoecker 1999). Although an oversimplification, this model demonstrates the potential role of MZP growth and trophodynamics on food web efficiency and the measurements of μ and m . A greater understanding of the interactions within this diverse size group will lead to more sophisticated numerical models and predictions of carbon flows through the microbial food web.

Design considerations for serial dilution experiments

The dilution technique yielded accurate estimates of μ in most model simulations. However, including MZP

growth and trophic interactions in the models yielded overestimates of grazing rates in simulated serial dilution experiments for all models. Another approach to estimating MZP grazing is via a 2-point dilution, which compares phytoplankton growth between undiluted and highly diluted water (e.g. 5% seawater; Landry 1993). It assumes that herbivore impact in highly diluted water is insignificant (i.e. not statistically different from zero). This simplified technique eliminates uncertainties related to differential MZP growth across the dilution gradient and yields conservative estimates of community herbivory rates. However, the 2-point approach does not reveal MZP dynamics that can be evident from non-linear phytoplankton growth curves; these non-linear dynamics should be examined if we are to improve our understanding of complex microbial food web interactions.

Comparisons of μ and m across systems may be complicated by slight differences in the methods employed. For example, in the present study we did not attempt to recreate *in situ* irradiance at each location. Phytoplankton adapted to lower light levels in turbid waters may respond differently under these conditions. Nutrient additions stimulated phytoplankton growth in all dilutions, but it is likely that different phytoplankton assemblages will respond differently to nutrient amendments. For example, high nutrients may lead to higher nutritional quality of the phytoplankton rather than higher growth rates. Screening size can also influence the outcome of dilution experiments. The initial screening size (or stress caused by screening) truncates the food web and sets up cascading controls on primary herbivores. In these experiments, we used considerably different mesh sizes to screen seawater. However, even small differences in initial screening mesh size can lead to different dominant trophic pathways.

Variation in incubation times can also lead to different experimental outcomes as interactions between MZP have a longer time to evolve. For example, the greatest changes in nanozooplankton are observed during the last portion of a day-long incubation period (Agis et al. 2007). Consequently, differences between, for example, a 24 h and a 30 h incubation period may be significant. Shorter incubation periods may limit the effect of MZP growth and trophic interactions on the experimental outcomes. However, fluctuations in phytoplankton production over the course of a day will cause complications when selecting the light period at which to incubate the treatments (e.g. including high-light or no-light periods). Additionally, short incubation times may not be appropriate in low-temperature environments, where phytoplankton biomass can change at slower rates. Sampling periodically during the incubation period (especially toward the end of the

incubation) can reveal the incubation time when MZP growth and interactions become significant enough to lead to overestimates of the *in situ* grazing rate.

In summary, even the simplified model representation of MZP growth and trophic interactions presented here illustrates the complexity of the microbial food web and cautions from the interpretation of dilution experiment results based exclusively on the response of phytoplankton. These models demonstrate the potential for herbivore growth and interactions to shape the outcomes of dilution experiments. In addition to measuring phytoplankton growth and grazing mortality, serial dilution experiments may be the most effective approach to delineating trophic links among MZP. Further work in plotting interactions and growth efficiencies, and constraining the regulatory forces in the microbial food web promises a greater understanding of energy flow in aquatic systems.

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