

3-13-2019

Nitrogen Fertilizer Suppresses Mineralization of Soil Organic Matter in Maize Agroecosystems

Navreet K. Mahal

South Dakota State University

William R. Osterholz

USDA Agricultural Research Service

Fernando E. Miguez

Iowa State University

Hanna J. Poffenbarger

University of Kentucky, hanna.poffenbarger@uky.edu

John E. Sawyer

Iowa State University

See next page for additional authors

Right click to open a feedback form in a new tab to let us know how this document benefits you.

Follow this and additional works at: https://uknowledge.uky.edu/pss_facpub

 Part of the [Agriculture Commons](#), [Ecology and Evolutionary Biology Commons](#), [Plant Sciences Commons](#), and the [Soil Science Commons](#)

Repository Citation

Mahal, Navreet K.; Osterholz, William R.; Miguez, Fernando E.; Poffenbarger, Hanna J.; Sawyer, John E.; Olk, Daniel C.; Archontoulis, Sotirios V.; and Castellano, Michael J., "Nitrogen Fertilizer Suppresses Mineralization of Soil Organic Matter in Maize Agroecosystems" (2019). *Plant and Soil Sciences Faculty Publications*. 139.
https://uknowledge.uky.edu/pss_facpub/139

This Article is brought to you for free and open access by the Plant and Soil Sciences at UKnowledge. It has been accepted for inclusion in Plant and Soil Sciences Faculty Publications by an authorized administrator of UKnowledge. For more information, please contact UKnowledge@sv.uky.edu.

Authors

Navreet K. Mahal, William R. Osterholz, Fernando E. Miguez, Hanna J. Poffenbarger, John E. Sawyer, Daniel C. Olk, Sotirios V. Archontoulis, and Michael J. Castellano

Nitrogen Fertilizer Suppresses Mineralization of Soil Organic Matter in Maize Agroecosystems**Notes/Citation Information**

Published in *Frontiers in Ecology and Evolution*, v. 7, article no. 59, p. 1-12.

Copyright © 2019 Mahal, Osterholz, Miguez, Poffenbarger, Sawyer, Olk, Archontoulis and Castellano.

This is an open-access article distributed under the terms of the [Creative Commons Attribution License \(CC BY\)](#). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Digital Object Identifier (DOI)

<https://doi.org/10.3389/fevo.2019.00059>



Nitrogen Fertilizer Suppresses Mineralization of Soil Organic Matter in Maize Agroecosystems

OPEN ACCESS

Edited by:

Urs Feller,
University of Bern, Switzerland

Reviewed by:

Alberto Sanz Cobeña,
Polytechnic University of Madrid,
Spain

Geoffrey R. Squire,
James Hutton Institute,
United Kingdom
Juan Fernando Hirzel,
Instituto de Investigaciones
Agropecuarias (INIA), Chile

*Correspondence:

Navreet K. Mahal
navreetmahal1@gmail.com

† Present Address:

Navreet K. Mahal,
Department of Agronomy, Horticulture
and Plant Science, South Dakota
State University, Brookings, SD,
United States
William R. Osterholz,
USDA-Agricultural Research Service,
Soil Drainage Research Unit,
Columbus, OH, United States
Hanna J. Poffenbarger,
Department of Plant and Soil
Sciences, University of
Kentucky-Lexington, Lexington, KY,
United States

Specialty section:

This article was submitted to
Agroecology and Ecosystem Services,
a section of the journal
Frontiers in Ecology and Evolution

Received: 23 August 2018

Accepted: 18 February 2019

Published: 13 March 2019

Citation:

Mahal NK, Osterholz WR, Miguez FE,
Poffenbarger HJ, Sawyer JE, Olk DC,
Archontoulis SV and Castellano MJ
(2019) Nitrogen Fertilizer Suppresses
Mineralization of Soil Organic Matter in
Maize Agroecosystems.
Front. Ecol. Evol. 7:59.
doi: 10.3389/fevo.2019.00059

Navreet K. Mahal^{1*}, William R. Osterholz^{1†}, Fernando E. Miguez¹,
Hanna J. Poffenbarger^{1†}, John E. Sawyer¹, Daniel C. Olk², Sotirios V. Archontoulis¹ and
Michael J. Castellano¹

¹ Department of Agronomy, Iowa State University, Ames, IA, United States, ² United States Department of
Agriculture-Agricultural Research Service, National Laboratory for Agriculture and the Environment, Ames, IA, United States

The possibility that N fertilizer increases soil organic matter (SOM) mineralization and, as a result, reduces SOM stocks has led to a great debate about the long-term sustainability of maize-based agroecosystems as well as the best method to estimate fertilizer N use efficiency (FNUE). Much of this debate is because synthetic N fertilizer can positively or negatively affect SOM mineralization via several direct and indirect pathways. Here, we test a series of hypotheses to determine the direction, magnitude, and mechanism of N fertilizer effect on SOM mineralization and discuss the implications for methods to estimate FNUE. We measured the effect of synthetic N fertilizer on SOM mineralization via gross ammonification at two long-term experiments in central and southern Iowa, USA with replicated plots of continuous maize that received one of three “historical” N fertilizer rates (zero, moderate or high) from 1999 to 2014. In 2015, prior to our measurements, we split the historical N fertilizer rate plots into two subplots that received either the site-specific agronomic optimum N rate or zero N fertilizer. At the onset of rapid maize N uptake, N fertilizer reduced gross ammonification by 13–21% (2–5 kg NH₄-N ha⁻¹ d⁻¹). A companion laboratory experiment rejected the hypothesis that differences in net primary productivity between fertilized and unfertilized treatments explained the negative effect of N fertilizer on SOM mineralization. Moreover, the NH₄⁺ pool size was negatively correlated with the gross ammonification rate ($r^2 = 0.85$, $p < 0.001$). Thus, we conclude that NH₄⁺-N fertilizer had a direct suppressive effect on SOM mineralization. These results demonstrate that the direct effect of N fertilizer on microbial activity can exceed the indirect effects of N fertilizer via large changes in NPP that alter organic matter inputs, soil temperature and moisture content. The magnitude of this effect and specificity to NH₄⁺-N has significant implications for fertilizer management as well as the measurement and modeling of agroecosystem N dynamics including FNUE.

Keywords: fertilizer nitrogen use efficiency, N mineralization, priming effect, agronomic optimum N rate, synthetic N fertilizer, continuous maize system

INTRODUCTION

Efficient use of nitrogen (N) fertilizer is important for economic crop production, environmental quality, and human health (Cassman, 1999). In some regions of the world, excessive and inefficient synthetic N fertilization harms the environment. In other regions of the world, more synthetic N fertilization would boost crop production and human health

(Vitousek et al., 2009). However, there is a great debate about the long-term sustainability of synthetic N fertilization (Robertson et al., 2013). Some reports indicate that synthetic N fertilizer reduces SOM stocks because it enhances soil organic matter (SOM) mineralization (Mulvaney et al., 2009; Russell et al., 2009); other reports indicate that it increases SOM stocks, particularly at the agronomically optimum N fertilizer rate (AONR) because it increases net primary productivity (NPP; Poffenbarger et al., 2017).

Much of this debate occurs because synthetic N fertilizer can positively or negatively affect SOM mineralization via several direct and indirect pathways (Table 1). Synthetic N fertilizer can have a direct positive effect on SOM mineralization by altering microbial activity and biomass. Stoichiometric decomposition theory predicts that if N is a limiting resource, inorganic N inputs will increase microbial biomass and activity thereby increasing SOM mineralization (Serner and Elser, 2002; Chen et al., 2014). Inorganic N can also have an indirect positive effect on SOM mineralization by increasing net primary productivity (NPP), and this effect could be particularly important in maize-based agroecosystems where inorganic N inputs can increase NPP by more than 200% (Poffenbarger et al., 2017). An increase in NPP, including rhizodeposition, can increase SOM mineralization by increasing microbial biomass or enzyme activity and this effect is often referred to as “positive priming” (Cheng, 2009; Chen et al., 2014).

Contrary to these positive effects, synthetic N fertilizer can also have negative effects on SOM mineralization. Inorganic N can have a direct negative effect on microbial activity. Microbial N mining theory predicts that, if N is a limiting resource, inorganic N inputs will decrease SOM mineralization by decreasing the decomposition of energy-poor SOM substrates that are mineralized solely to access N-containing compounds (Moorhead and Sinsabaugh, 2006; Craine et al., 2007). Inorganic N can also have an indirect effect on SOM mineralization via changes in broader ecosystem function: the large increase in NPP with fertilizer addition reduces soil moisture (transpiration)

and temperature (shading), which decreases SOM mineralization (Parton et al., 1987; Wang et al., 2006; Li et al., 2014).

These potential effects of N fertilizer on SOM mineralization have led to confusion about the accuracy of methods to estimate fertilizer N use efficiency (FNUE)—methods that are critical to improve the sustainability of cereal-based cropping systems. Typically defined as the percentage of fertilizer N recovered in aboveground plant biomass, FNUE is an important parameter to estimate N use efficiency (NUE) of cropping systems. Currently, there are two methods used to determine FNUE, however, they consistently produce different results (Cassman et al., 2002). The “N difference” method is an indirect measure that estimates FNUE based on the difference in N uptake between a fertilized and an unfertilized crop:

$$\frac{(NF - NC)}{R} * 100 \tag{1}$$

where, *NF* = total plant N uptake from fertilized plots, *NC* = total N uptake from unfertilized plots and *R* = rate of N fertilizer applied (Varvel and Peterson, 1990). In contrast, the “¹⁵N tracer” method is a direct measure that calculates FNUE based on plant uptake of an isotopically labeled fertilizer (Hauck and Bremner, 1976):

$$\frac{\text{Plant uptake of } ^{15}\text{N labeled fertilizer}}{\text{amount of fertilizer } ^{15}\text{N applied}} * 100 \tag{2}$$

The “N difference” method (Equation 1) typically estimates greater FNUE than the direct “tracer” method (Equation 2; Jansson and Persson, 1982; Cassman et al., 2002).

This result is consistent with the suggestion that N fertilizer enhances SOM mineralization (Khan et al., 2007; Robertson et al., 2013), which has been hypothesized to be a consequence of a positive “priming effect” (Fontaine et al., 2003; Cheng, 2009; Kuzyakov, 2010). In this case, the N-fertilized (*NF*, Equation 1) plot has greater non-fertilizer inorganic N availability than the zero-N control plot (*NC*, Equation 1) because the N fertilizer increased SOM mineralization. This difference in N availability violates the assumption of equal crop N uptake from non-fertilizer sources in the fertilized and unfertilized treatments (i.e., mineralized SOM N is equal in the fertilized and unfertilized plots). Such a violation of methodological assumptions would artificially inflate FNUE as measured by the “N difference” method.

Alternatively, the “¹⁵N tracer” method may provide an artificially low FNUE due to “pool substitution” (Cassman et al., 2002). In this case, ¹⁵N fertilizer substitutes with native SOM-N during N immobilization-mineralization processes without a net change in the inorganic N pool. This dilutes the ¹⁵N enrichment of fertilizer, resulting in artificially low FNUE (Hauck and Bremner, 1976; Jansson and Persson, 1982; Harmsen and Moraghan, 1988). Pool substitution of ¹⁵N is particularly pronounced where rates of N immobilization are high, such as in soils with large amounts of high C/N ratio plant residue (Jenkinson et al., 1985).

Coupled field and laboratory experiments (in the presence and absence of plants and their effects on environmental conditions)

TABLE 1 | Potential directions and mechanisms of effects of synthetic nitrogen (N) fertilizer addition on SOM mineralization and the respective impacts on the measurement of fertilizer nitrogen use efficiency (FNUE) by the “N difference method.”

Direction	Mechanism	Mode of action	Effect on measurement of FNUE
1) No effect	i) N/A*	N/A	None
2) Positive	ii) Positive priming due to rhizodeposition	Indirect	Overestimation
	iii) Stoichiometry theory (positive priming)	Direct	Overestimation
3) Negative	iv) Lower soil water content and temperature	Indirect	Underestimation
	v) N mining theory (negative priming)	Direct	Underestimation

*Not applicable.

can separate the potential direct and indirect effects of N fertilizer on SOM mineralization and identify any systematic bias in measurements of FNUE with the “N difference” method (Table 1). Field experiments can test for the *direction* and agronomic relevance of an effect. If N fertilizer does not affect SOM mineralization, we can reject a “priming effect” and resultant overestimation of FNUE by the “N difference method.” In this case, there is no need for coupled laboratory experiments. In contrast, if N fertilizer increases or decreases SOM mineralization the “N difference method” will, respectively, overestimate or underestimate FNUE. In this case, controlled laboratory experiments can isolate the *mechanism* of effect by separating the direct effect of N fertilizer on microbial activity (Table 1; iii and v) from the indirect effects of N fertilizer on microbial activity via changes in crop growth and rhizodeposition (Table 1; ii and iv). Laboratory experiments would not detect indirect positive or negative effects of NPP on SOM mineralization and thus can be used to test for direct effects of N fertilizer on SOM mineralization. If the effect of N fertilizer on SOM mineralization observed in the laboratory (in the absence of plants to exclude indirect effect) is similar to that observed in the field, the increase or decrease in SOM mineralization can be attributed to a direct effects of N fertilizer addition on microbial activity.

This study had two main objectives: (1) At two key times of maize growth, we determined the magnitude and mechanism of the N fertilizer effect on SOM mineralization; (2) Using our results from Objective 1, we evaluated the “N difference” method for measurement of FNUE. To execute these objectives, we measured SOM mineralization via gross ammonification assays in two Midwest USA continuous maize (*Zea mays* L.) cropping systems with different soil types, yield potentials, and long-term mean AONRs.

MATERIALS AND METHODS

Site History: 1999–2014

This study was conducted at two sites: central Iowa (42°01'N; 93°47'W) and southern Iowa (40°58'N; 93°25'W). The sites differ in soil properties and glacial history but have been managed as annual cropland for more than 25 years. Mean annual temperature from 1994 to 2014 was 9.1°C at the central site and 9.5°C at the southern site; mean annual precipitation was 970 mm at the central site and 980 mm at the southern site¹. The central site is underlain by artificial subsurface drainage whereas the southern site is not. Largely due to the different soil properties and drainage management, there is a large difference in water-limited maize yield potential at the two sites (Table 2).

In 1999, experiments were established to determine maize grain yield response to inorganic N fertilizer and the N fertilizer rate at which yield no longer positively responds to additional N fertilization (i.e., the agronomic optimum N rate, AONR) in continuous maize cropping systems at each site. Individual

¹Iowa State University. Iowa Environmental Mesonet [Internet] (2017). Available online at: <http://mesonet.agron.iastate.edu/request/coop/fe.phtml>

TABLE 2 | Selected soil and agronomic properties and climatic conditions at two experimental sites in Iowa.

Site	USDA soil taxonomy	USDA texture	Sand-Silt-Clay (%) (15" depth)	Bulk density (g cm ⁻³)	Artificial sub-surface drainage	2015 mean temperature (°C)	2015 cumulative precipitation (mm)	Long term AONR (kg N ha ⁻¹)*	Maize yield at AONR (Mg ha ⁻¹)*
Central	Hapludolls	Loam	45-34-21	1.34	Yes	9.3	1300	202	11.35
Southern	Argiaquolls	Silt loam	5-74-21	1.29	No	9.8	1226	269	9.38

Bulk density was measured on 0–15 cm soil samples. 2015 cumulative precipitation and mean temperature over the study year for each location (2015). Precipitation and temperature data were from Iowa Environment Mesonet¹. *Long-term Agronomic optimum N rate (AONR) and maize yield at the AONR from 2000–2014.

plots received one of five (central) or seven (southern) N fertilizer rates ranging from 0 to 269 kg N ha⁻¹y⁻¹. The experimental design was a randomized complete block with four replicates of each N fertilizer rate. Each plot received the same N fertilizer application rate from 1999 to 2014. All experimental plots were conventionally managed with fall chisel plowing and spring secondary tillage before planting. Herbicides, pesticides, all nutrients other than N, and pH were maintained for optimum production. The southern site had a greater long-term AONR despite lower maize yields (Table 2). Data about historical crop yield response to N fertilizer and long-term change in soil organic carbon from 1999 to 2014 are reported in Poffenbarger et al. (2017).

Experimental Design: 2015

The study reported herein was conducted in 2015. For this study, we selected three of the historical N rates at each site: zero (0 kg N ha⁻¹), moderate (202 and 224 kg N ha⁻¹ at central and southern sites, respectively), and high (269 kg N ha⁻¹). Based on the 1999–2014 data, at the central site, the moderate rate was the AONR and the high rate was 33% [(269–202)*100/202] greater than AONR while at the southern site, the moderate rate was 17% [(269–224)*100/269] lower than the AONR and the high rate was the long-term AONR. From here forward, we refer to the historical rates as zero, moderate and high.

Ideally, at both sites, we would have selected the historical zero rate, the AONR, and a rate greater than the AONR. Unfortunately, the southern site did not include a rate greater than the AONR. However, two of the three selected rates are directly comparable across the two sites with reference to the effect on maize yield: the zero and AONR.

In 2015, we split the main long-term plots receiving the three selected historical N rates (zero, moderate, and high) into two subplots (Figure 1). Regardless of the historical N rate, one subplot received the long-term average site-specific AONR and one subplot received no N fertilizer. A 107-day maturity maize hybrid (FS 57QX1) was planted at ~89,000 seeds ha⁻¹ on May 13 at the central site and on April 28 at the southern site. Within 7 d of maize planting, the 1999–2014 AONR at the central (202 kg N ha⁻¹) and southern sites (269 kg N ha⁻¹) was surface applied as a solution of NH₄NO₃ to one subplot whereas no N (0 kg N ha⁻¹) was applied to the other subplot. Thus, the 2015 experiment included: 3 historical N rates, 4 replicates plots per historical N rate, 2 subplots per historical N rate, and therefore 24 total subplots per site. Each individual sub-plot measured 3.05m x 4.57 m.

Soil Organic Carbon and Total Nitrogen

In 2014, soils were sampled from 0 to 15 cm depth in each main plot using 2.5 cm diameter cores after harvest and before tillage. Bulk density was determined for each experimental block. Total soil organic C and N were determined for the main plots using dry combustion elemental analysis.

Soil Moisture, Inorganic N Pools Size, and Gross Ammonification Rate in Field-Fresh Soils

In 2015, on two sampling dates per site, soil samples were collected at a depth of 5–15 cm from all subplots. The two sample dates at each site corresponded to the V5 maize growth stage (5 collared leaves): 16 June, 2015 at central and 2 June, 2015 at southern site, and to the V12 maize growth stage (12 collared leaves): 21 July, 2015 at central site and 8 July, 2015 at southern site. These growth stages are coincident with the onset and midpoint of maximum rate of maize N uptake (Abendroth et al., 2011).

At each of the four sampling events (2 sites × 2 sample times), 10 soil cores of 2.5 cm diameter (5–15 cm soil depth) were taken from random locations capturing different parts of the maize row and inter-row areas within each subplot and bulked to represent one sample for each subplot. The sample depth was selected to collect a soil sample that was representative of the depth of high N uptake and that avoided the effects of daily weather variations. Soil samples were transported to the lab in an insulated cooler. Each bulked sample from each subplot was passed through an 8 mm sieve and gravimetric water content was measured from a 10 g subsample. Nitrate (NO₃⁻) and ammonium (NH₄⁺) pool size was also determined by extracting NO₃⁻ and NH₄⁺ from a 10 g subsample with 50 ml 2 M potassium chloride (KCl) and subsequent measurement with colorimetry (Hood-Nowotny et al., 2010).

Gross ammonification rate (i.e., the conversion of SOM-N to NH₄⁺) was used to determine the direction of effect of N fertilizer addition on SOM mineralization. Within 24 h of sampling, gross ammonification rate was measured on the field-fresh soils during a 24 h incubation using conventional ¹⁵N isotope pool dilution method (Hart et al., 1994) with blank correction (Stark and Hart, 1996) that was adapted to intensively managed Iowa agricultural soils (Osterholz et al., 2017). As a result, these assays were made at the sampled moisture content (which differed across treatments) and room temperature (21°C) which was very similar to field temperature.

Isolating the Effect of Inorganic N Fertilizer on Gross Ammonification

To test the mechanism of fertilizer effect on gross ammonification, we conducted additional gross ammonification rate assays that controlled for the effects of NPP by selecting soils from one treatment (i.e., the same NPP) and standardizing soil moisture contents (which differed across treatments in the assays made on field-fresh soils). For these assays, we used soils collected for measurement of gross ammonification at the V5 growth stage (see section Soil Moisture, Inorganic N Pools Size, and Gross Ammonification Rate in Field-Fresh Soils) from only subplots that historically received zero N fertilizer (1999–2014) and zero N fertilizer in 2015 (Figure 1). Prior to analysis, the samples were refrigerated at 4°C. Soils were then brought to room temperature and each sample was split into two subsamples. One subsample received NH₄NO₃-N fertilizer at an amount that brought the inorganic N concentration to a level

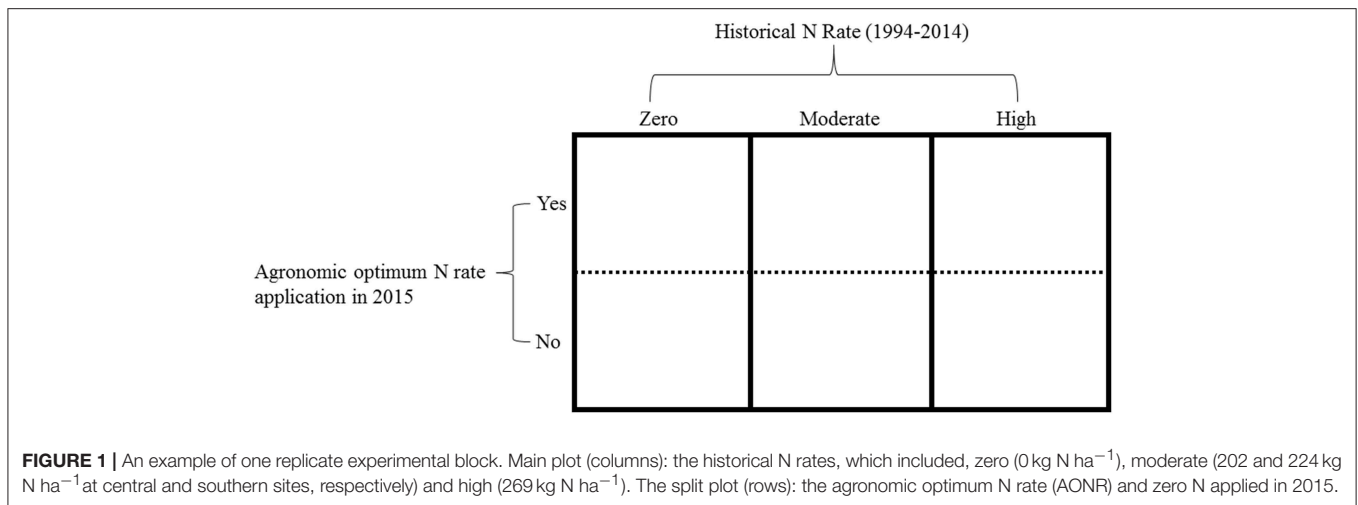


FIGURE 1 | An example of one replicate experimental block. Main plot (columns): the historical N rates, which included, zero (0 kg N ha^{-1}), moderate (202 and 224 kg N ha^{-1} at central and southern sites, respectively) and high (269 kg N ha^{-1}). The split plot (rows): the agronomic optimum N rate (AONR) and zero N applied in 2015.

similar to that of soils from zero N plots that received the AONR in the field ($\sim 23 \text{ mg N kg}^{-1}$ soil at central site, and $\sim 50 \text{ mg N kg}^{-1}$ soil at southern site); the other subsample received zero N fertilizer. Identical soil moisture and temperature were maintained the same across subsamples. Thus, inorganic N concentration was the only difference between these subsamples. After fertilizer addition, gross ammonification was determined during a 24 h incubation.

Across all gross ammonification assays, the addition of $^{15}\text{NH}_4\text{Cl}$ solution increased the final ammonium pool size by 5–32%, which is well within the natural variability of the soil pool size, and the final atom % ^{15}N enrichment ranged between 0.8 and 4.4%.

Maize Yield and Agronomic Fertilizer N Use Efficiency in 2015

Maize ears were collected by hand from the center two rows of each subplot (yield area = 3.48 m^2) at physiological maturity. The entire sample was weighed fresh, and then a six-ear subsample was dried at 60°C . After drying the subsample, the grain was removed from the cobs and each component (grain and cobs) was weighed separately. The grain dry matter yield (in kg ha^{-1}) was calculated by adjusting the fresh ear weight of the entire area to a grain dry weight using the moisture content and grain:cob ratio of the six-ear subsample. Grain yield was adjusted to 15.5% moisture content. Grain N concentration was set at 1.2% to calculate the N harvested with grain (Ciampitti and Vyn, 2012).

Agronomic efficiency (AE) was calculated by using the equation:

$$AE = (Y - Y_0)/N \quad (3)$$

where, AE = agronomic efficiency of applied N ($\text{kg } \Delta \text{ yield kg}^{-1}$ N fertilizer applied), Y = grain yield with fertilizer application (kg ha^{-1}), Y_0 = grain yield without fertilizer addition (kg ha^{-1}), N = rate of N fertilizer applied (kg ha^{-1}) (Dobermann, 2007).

Data and Statistical Analysis

The difference in gross N mineralization rates within each location and historical N fertilizer rate was calculated by

subtracting gross N mineralization rate in the zero-N subplot treatment from that in the AONR subplot treatment. Data were analyzed separately for each site, due to differences in historical N rates, soil properties, and artificial drainage at each site (Table 1). Gross ammonification data were analyzed using a linear mixed model where historical N rates (main plot), AONR application in 2015 (split plot) and maize growth stage (repeated measure) were treated as fixed factors and blocks as a random factor. Maize grain yield data (reported at 15.5% moisture) were analyzed using a linear mixed model with historical N rates (main plot) and AONR in 2015 (split plot) as fixed factors and block as random factor. The SOC, and soil total N data were analyzed using linear mixed model with historical N rate as a fixed factor and block as a random factor. All data analyses were done using PROC MIXED in SAS/STAT software, version 9.3 (SAS Institute, 2013).

RESULTS

Site History: Historical N Fertilizer Rate Effect on Total Soil Organic C and N

There was a significant effect of the historical 15-year N fertilizer application rates on total SOC ($p = 0.001$ at central, and 0.057 at southern site) and total soil N ($p = 0.001$ at central, and 0.072 at southern site) contents. At the central site, the moderate and high historical N rate treatments had 20 and 15% higher total SOC concentrations and 17 and 14% higher total N concentrations than the historical zero N treatment (Table 3). At the southern site, the high historical N rate treatments had 6% higher total SOC concentrations and 7% higher total N concentrations than the historical zero N treatment, whereas low and moderate rates had statistically similar total SOC and N concentrations.

Maize Yield and Agronomic Efficiency of N Fertilizer Applied in 2015

At both the sites and across all historical N rates, maize yield was greater in treatments receiving the AONR vs. zero-N ($p < 0.001$ at both sites) (Figure 2). At the central site, maize yield in the AONR was 114% greater than the zero-N treatment regardless of historical N rate. However, there was a significant

TABLE 3 | Mean soil organic C and total N (g kg^{-1}) (95% confidence interval) for historical N rates at central and southern Iowa sites.

Site	Historical N rate	SOC (g kg^{-1})	Total N (g kg^{-1})
Central	Zero	18.23(14.17, 22.28)b	1.63 (1.37, 1.88)b
Central	Moderate	21.73 (17.67, 25.78)a	1.93 (1.67, 2.18)a
Central	High	20.9 (16.85, 24.95)a	1.85 (1.59, 2.11)a
Southern	Zero	21.95 (20.96, 22.94)b	1.83 (1.71, 1.94)b
Southern	Moderate	23.05 (22.06, 24.04)ab	1.93(1.81, 2.04)ab
Southern	High	23.35 (22.36, 24.34)a	1.98 (1.86, 2.09)a

Soil organic C and total N were measured on 0–15 cm soil samples in 2014. Different letters represent significant statistical differences among historical N rates within each site ($P \leq 0.05$).

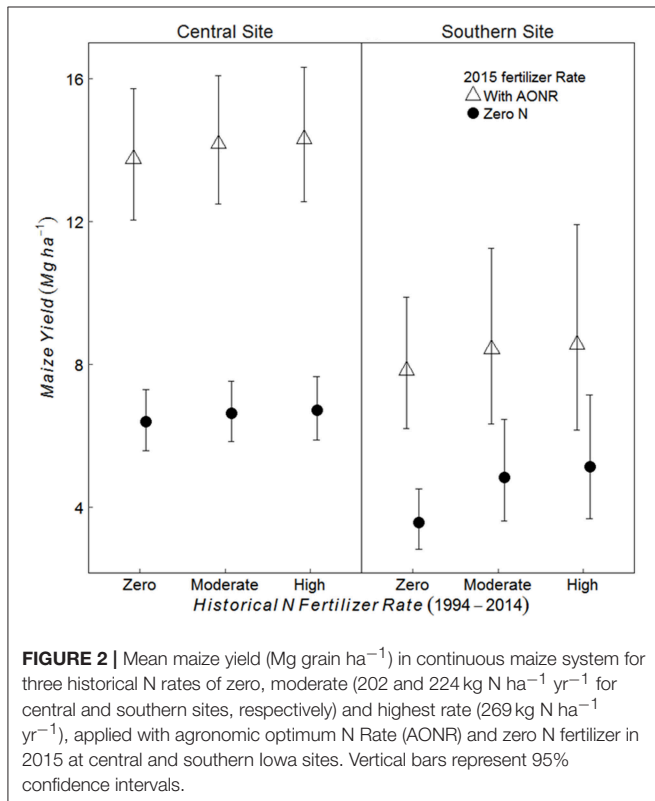


FIGURE 2 | Mean maize yield (Mg grain ha^{-1}) in continuous maize system for three historical N rates of zero, moderate (202 and $224 \text{ kg N ha}^{-1} \text{ yr}^{-1}$ for central and southern sites, respectively) and highest rate ($269 \text{ kg N ha}^{-1} \text{ yr}^{-1}$), applied with agronomic optimum N Rate (AONR) and zero N fertilizer in 2015 at central and southern Iowa sites. Vertical bars represent 95% confidence intervals.

interaction between historical N rates (1999–2014) and 2015 AONR application at southern site ($p = 0.050$). The AONR increased maize yield by 119, 78, and 71% for historical fertilizer rates of zero, moderate and high N rates, respectively (Figure 2).

Historical N rate had no effect on AE at either site (Figure 3) ($p = 0.789$ at central, and 0.634 at southern site). However, the southern site had almost 60% lower AE as compared to that at the central site. Average AE at central and southern sites was 37 and 15 $\text{kg grain kg}^{-1} \text{ N}$, respectively. On average over the three historical N rates, 79 kg N/ha was harvested with Zero-N fertilizer at the central site and 54 kg N/ha at the southern site, whereas grain N harvested with fertilizer applied at AONR was 169 and 99 kg N/ha , respectively (data not shown).

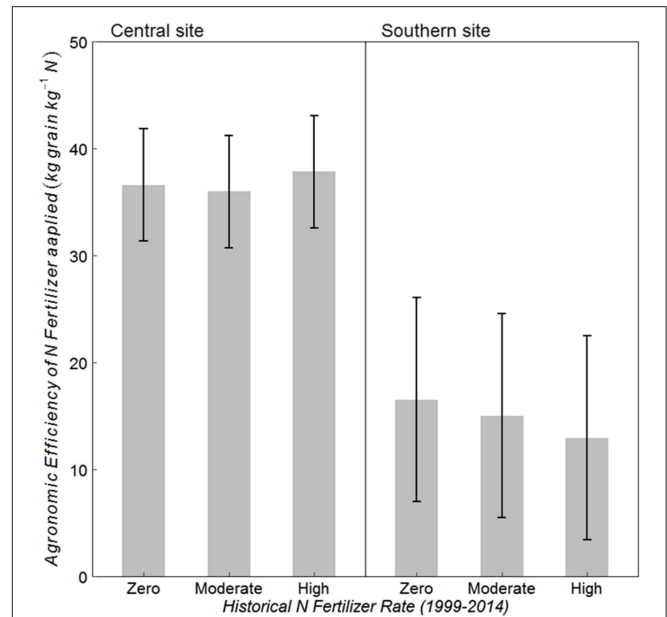


FIGURE 3 | Mean agronomic efficiency (AE) of applied N fertilizer for continuous maize system with three historical N rates (1999–2014) of zero, moderate (202 and $224 \text{ kg N ha}^{-1} \text{ yr}^{-1}$ at central and southern sites, respectively) and the highest rate ($269 \text{ kg N ha}^{-1} \text{ yr}^{-1}$) at the central, and southern Iowa sites. Vertical bars represent 95% confidence intervals.

Soil Moisture Content and Inorganic N Pools Size in Fresh Soils

Historical N rate had significant effect on soil moisture content; soil moisture content in historically moderate and high N fertilizer rates was 6% higher than the historical zero N rate at the central site and 3–4% higher than the historical zero-N rate at the southern site (Figure 4; $p = 0.006$ at central, <0.001 at southern site). Soil moisture content was also significantly affected by the AONR application in 2015; it was 3–4% greater in the subplots receiving zero-N vs. the AONR ($p < 0.001$ at both sites). Soil moisture at the V12 maize growth stage was 13 and 9% lower than at the V5 stage at the central and southern sites, respectively ($p < 0.001$ at both sites).

On average, across all historical N fertilizer rates and growth stages, the soil NH_4^+ -N pool was 11 and 48% greater in the AONR vs. zero-N subplots at the central and southern Iowa sites, respectively (Figure 5; $p = 0.009$ at central, and 0.046 at southern site). However, at the southern site, there was a significant interaction between 2015 AONR application and maize growth stage ($p = 0.019$) such that the soil NH_4^+ -N pool was only significantly higher in the AONR vs. zero-N subplots at the V5 growth stage (119% higher in AONR vs. zero-N subplot). The soil NO_3^- -N pool was higher in the AONR compared to the zero-N treatment (277 and 217% greater at central and southern sites, respectively; Figure 5; $p < 0.001$ at both sites). However, there was a significant interaction between the 2015 AONR application and maize growth stage ($p < 0.001$ at both sites). At the V5 growth stage, the NO_3^- -N pool was 623 and 708% higher in the

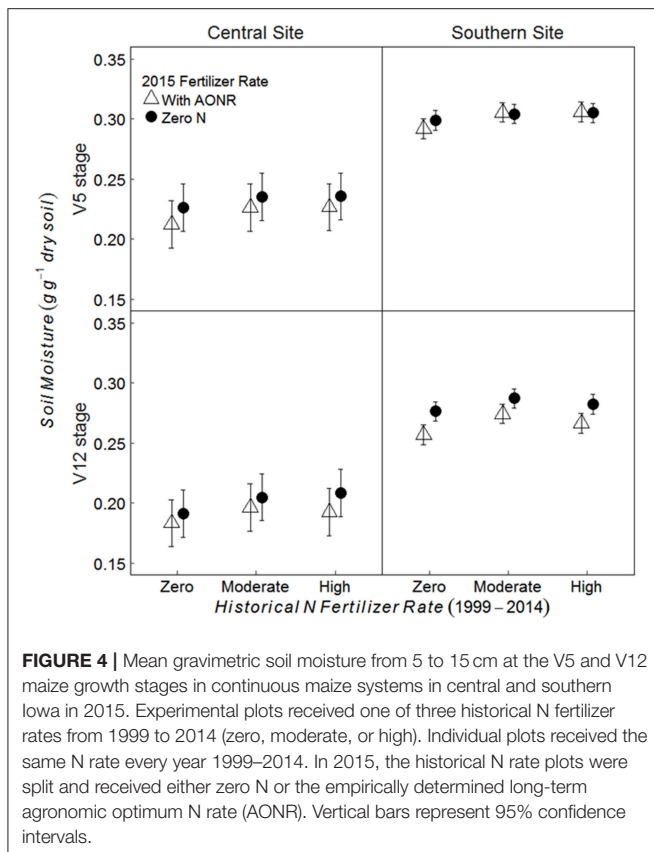


FIGURE 4 | Mean gravimetric soil moisture from 5 to 15 cm at the V5 and V12 maize growth stages in continuous maize systems in central and southern Iowa in 2015. Experimental plots received one of three historical N fertilizer rates from 1999 to 2014 (zero, moderate, or high). Individual plots received the same N rate every year 1999–2014. In 2015, the historical N rate plots were split and received either zero N or the empirically determined long-term agronomic optimum N rate (AONR). Vertical bars represent 95% confidence intervals.

AONR than in the zero-N treatments at the central and southern sites, while at the V12 growth stage, the NO_3^- -N pool was similar in the AONR and zero-N treatments.

Gross Ammonification Rates in Fresh Soils

At the V5 growth stage at both sites, the 2015 N fertilizer application at the AONR reduced gross ammonification (Figure 6; 2015 AONR \times growth stage $p = 0.0082$ at central, and 0.054 at southern site). However, also at both sites, the effect of the 2015 N fertilizer addition decreased with increasing historical N rate such that the 2015 N fertilizer input had the greatest effect on gross ammonification in the historically zero-N plots and no effect in the historically high N plots. In contrast to the V5 growth stage, N fertilizer had no effect on gross ammonification at the V12 growth stage (Figures S1, S2). Across all historical N rates at the V5 growth stage, the AONR application in 2015 reduced gross ammonification by 15 and 12% at the central and southern sites, respectively.

Isolated Effect of Synthetic N Fertilizer on Gross Ammonification

The laboratory-controlled addition of N fertilizer to soils that received no N fertilizer input (neither historically nor in 2015) consistently reduced gross ammonification (Figure 7; $p < 0.001$ at both sites). In soils from the central site, gross ammonification rates were 68% lower with addition of NH_4NO_3 . In soils from

the southern site, gross ammonification rate was numerically negative, which is biologically impossible, but this result was not statistically different from zero (large ammonium pool sizes and low isotopic enrichment likely elevated the minimum detection limit for gross ammonification above the actual rate; Hart et al., 1994). There was a strong negative correlation between ammonium pool size and gross ammonification rate (Figure 8). However, there was no correlation between nitrate pool size and gross ammonification.

DISCUSSION

Implications for Estimation of FNUE

Our results indicate that FNUE, when measured by the “N difference” method may be underestimated. Fertilizer N use efficiency (FNUE) can be estimated with two methods: “N difference” and “¹⁵N tracer” (see section Introduction). The “N difference” method (Equation 1) has been suggested to overestimate FNUE compared to the “¹⁵N tracer method” (Equation 2) due to a positive or “priming” effect of N fertilizer on soil N mineralization (Jansson and Persson, 1982; Cassman et al., 2002; Gardner and Drinkwater, 2009). Our results reject this hypothesis. The methodological assumption of equal N pool sizes and mineralization rates in the fertilized and unfertilized treatments was violated; lower N mineralization in fertilized vs. unfertilized treatments at our sites would reduce FNUE as measured by the “N difference” method.

Therefore, rather than previous concerns about the potential for the N difference method to overestimate FNUE, our results suggest the “N difference” method may underestimate FNUE. The fertilizer N use efficiency as measured with the N difference method is generally 20–30% greater than the ¹⁵N tracer method (Cassman et al., 2002; Stevens et al., 2005). At our sites, positive priming of SOM mineralization due to synthetic N fertilizer application cannot explain the methodological difference. Future work should investigate suggestions that the ¹⁵N tracer method underestimates FNUE due to 1:1 substitution of added inorganic ¹⁵N isotope and the existing inorganic ¹⁴N pool (Cassman et al., 2002).

Despite reduced gross ammonification with N fertilizer application at the V5 growth stage and similar inorganic N pools between fertilized and non-fertilized treatments at the V12 growth stage, maize yield was much greater in N-fertilized treatments (Figure 2). Although we did not measure roots, previous reports have demonstrated that N fertilizer concomitantly increases below and aboveground maize production, which suggests well-developed root system in fertilized plots enhances N uptake (Russell et al., 2009) and may explain greater yield despite similar inorganic N pools. Thus, any suppressive effect of N fertilizer on gross ammonification does not appear to negatively impact N uptake.

Synthetic N Fertilizer Effects on SOM Mineralization: Mechanism

Across two maize production systems with different long-term climate, yield potential, and physiography (Table 1), inorganic N fertilizer caused a consistent and agronomically relevant

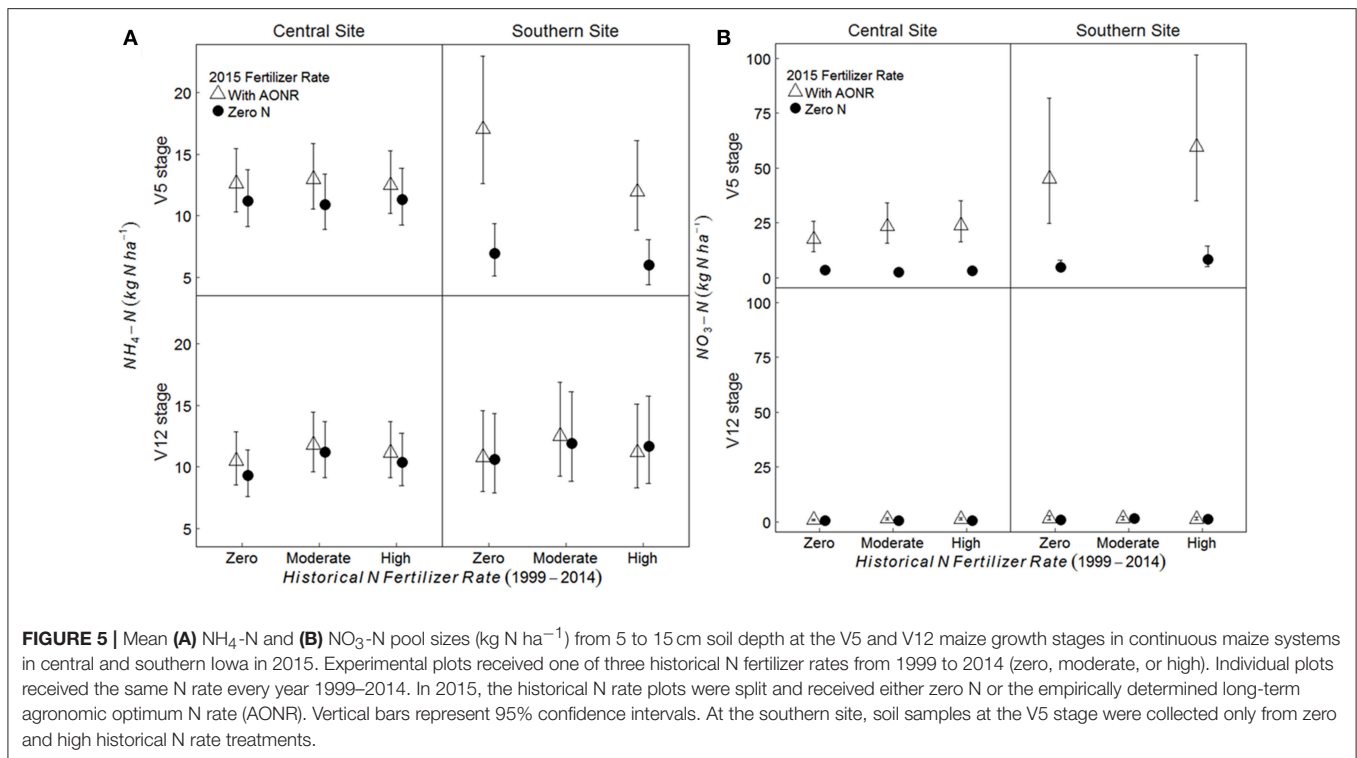


FIGURE 5 | Mean (A) $\text{NH}_4\text{-N}$ and (B) $\text{NO}_3\text{-N}$ pool sizes (kg N ha^{-1}) from 5 to 15 cm soil depth at the V5 and V12 maize growth stages in continuous maize systems in central and southern Iowa in 2015. Experimental plots received one of three historical N fertilizer rates from 1999 to 2014 (zero, moderate, or high). Individual plots received the same N rate every year 1999–2014. In 2015, the historical N rate plots were split and received either zero N or the empirically determined long-term agronomic optimum N rate (AONR). Vertical bars represent 95% confidence intervals. At the southern site, soil samples at the V5 stage were collected only from zero and high historical N rate treatments.

reduction in SOM mineralization rate as assayed by gross ammonification. The strong negative correlation between NH_4^+ pool size and gross ammonification indicate that NH_4^+ is directly responsible for the suppression. Microbial N mining theory can help to explain the direct effect of NH_4^+ fertilizer on gross ammonification rate. This theory suggests that N fertilization will reduce microbial mineralization of SOM because, under conditions of low soil inorganic N, microbes accelerate SOM decomposition in search of N to meet demand for growth metabolism (Moorhead and Sinsabaugh, 2006). This energy investment in N acquisition may lower microbial carbon use efficiency because much of the SOM decomposed as a result N mining is low- or zero-energy biomolecules such as lignin (Spohn et al., 2016; Zang et al., 2016; Li et al., 2017). Indeed, the addition of N fertilizer can reduce oxidative enzyme activity (Spohn et al., 2016), which is consistent with the concept that N mining targets nutrient-poor substrates. Therefore, our coupled field and laboratory N-addition experiments demonstrated that the suppressive effect of N fertilizer on gross ammonification can be attributed to the direct effect of N fertilizer application on microbial activity rather than the indirect effects of N fertilizer on microbial activity that are controlled by crop growth (i.e., soil moisture, soil temperature, and enhanced belowground OM inputs, e.g., rhizodeposition). These results stand in complete contrast to suggestions that synthetic N fertilizer increases SOM-N mineralization and reduces SOM stocks (Mulvaney et al., 2009).

The agronomic relevance of our results may, however, vary with the type and placement of N fertilizer. Anhydrous ammonia and urea do not contain NH_4^+ , but are quickly transformed

to NH_4^+ . Although these transformations affect soil pH, we found no correlation between gross ammonification and soil pH (data not shown), which is similar to previous work (Booth et al., 2005). Other fertilizers contain large fractions of NO_3^- (e.g., urea-ammonium-nitrate). Because we found no association between NO_3^- pool size and gross ammonification in field-fresh or laboratory-controlled soils, N fertilizer with large fractions of NO_3^- may have a lesser effect on gross ammonification. The placement of N fertilizer may also impact our results. We uniformly applied NH_4NO_3 to the soil, however, N fertilizer can be placed in concentrated bands resulting in much higher NH_4^+ concentrations (e.g., Mitchell et al., 2013; Maharjan et al., 2014). The correlation between NH_4^+ pool size and gross ammonification suggests the suppressive effect of NH_4^+ may be higher when NH_4^+ -based fertilizers are applied in concentrated bands. Moreover, the study year was wetter as compared to the historical precipitation (34 and 25% higher at central and southern site, respectively); the effects of N fertilizer could vary depending on the climatic conditions affecting microbial activity.

Long- and Short-Term Effects of N Fertilizer Application on SOM Mineralization

The suppressive effect of N fertilizer on gross ammonification was greater in historically under-fertilized soils (Figure 4), which is consistent with microbial N-mining of SOM. The historical zero-N rates at both sites had lower nutrient availability as indicated by lower total N concentrations

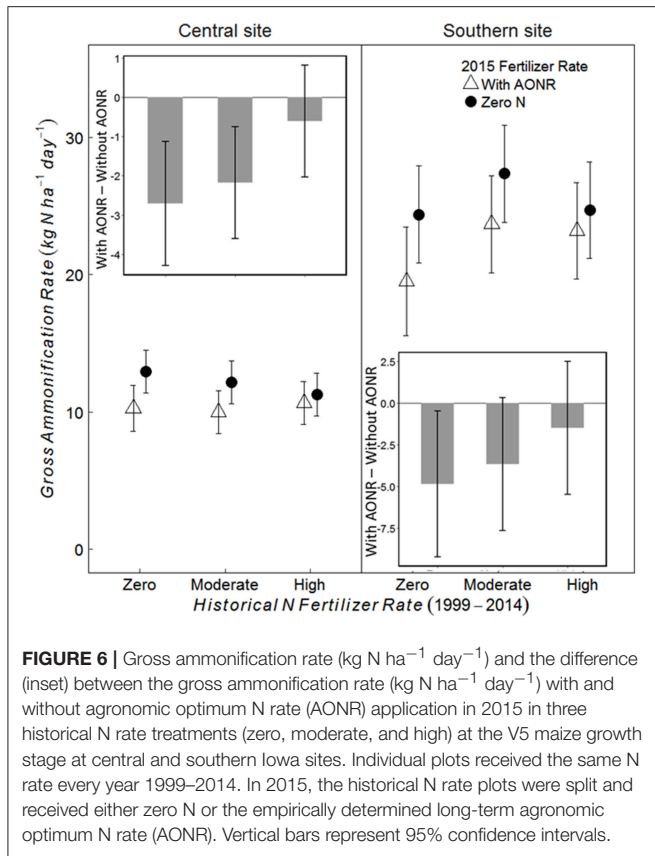


FIGURE 6 | Gross ammonification rate (kg N ha⁻¹ day⁻¹) and the difference (inset) between the gross ammonification rate (kg N ha⁻¹ day⁻¹) with and without agronomic optimum N rate (AONR) application in 2015 in three historical N rate treatments (zero, moderate, and high) at the V5 maize growth stage at central and southern Iowa sites. Individual plots received the same N rate every year 1999–2014. In 2015, the historical N rate plots were split and received either zero N or the empirically determined long-term agronomic optimum N rate (AONR). Vertical bars represent 95% confidence intervals.

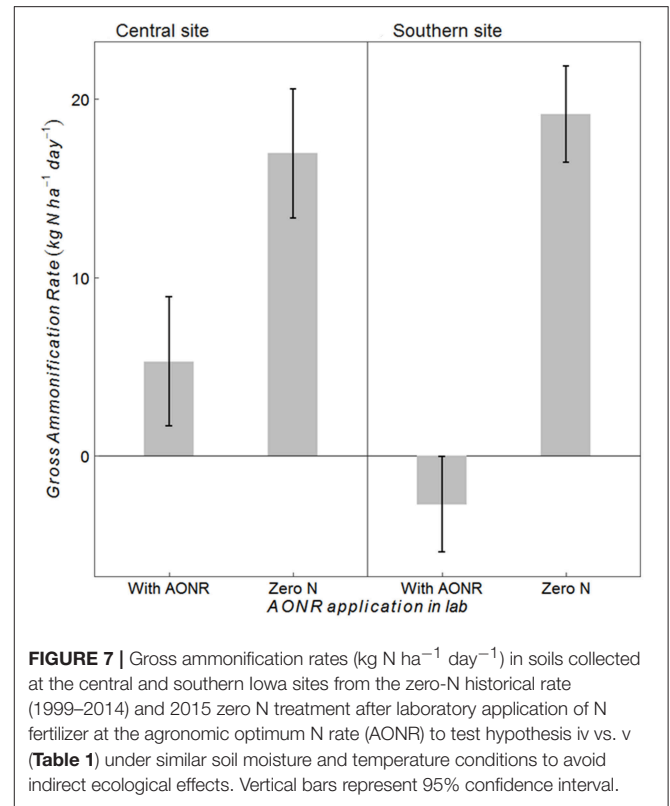


FIGURE 7 | Gross ammonification rates (kg N ha⁻¹ day⁻¹) in soils collected at the central and southern Iowa sites from the zero-N historical rate (1999–2014) and 2015 zero N treatment after laboratory application of N fertilizer at the agronomic optimum N rate (AONR) to test hypothesis iv vs. v (**Table 1**) under similar soil moisture and temperature conditions to avoid indirect ecological effects. Vertical bars represent 95% confidence interval.

compared to moderate and high historical N rates (**Table 3**). In addition, previous work at the central site demonstrated that N fertilizer inputs have reduced the C/N ratio of maize litter while altering microbial community composition from a fungal-dominated community in zero-N treatments to bacterially-dominated community in N-fertilized treatments (Brown et al., 2014). These changes may have decreased microbial N mining in N-fertilized treatments by decreasing microbial N demand. Alternatively, historical N fertilizer could have provided a “home-field advantage” to the microbial community (i.e., the microbes were accustomed to high NH₄⁺ concentrations) such that response to 2015 N fertilizer was muted (Ayres et al., 2009).

Despite the large effect of N fertilizer on gross ammonification, it was short-term and disappeared after a period of rapid plant N uptake that equalized inorganic N pools in fertilized and non-fertilized treatments (**Figure 5**). By the V12 growth stage, cumulative maize N uptake exceeds 33% of the total N uptake (Abendroth et al., 2011) and, as a result, soil inorganic N pools are typically small. During this time, maize growth reduced soil moisture. Although the effect of N fertilizer on gross ammonification rate (Figures S1, S2), and soil inorganic N concentration disappeared by the V12 growth stage (**Figure 5**), the effect of N fertilizer on soil moisture persisted (**Figure 4**). These patterns in the soil environment suggest that the indirect effect of NPP on soil moisture was not a factor reducing gross ammonification.

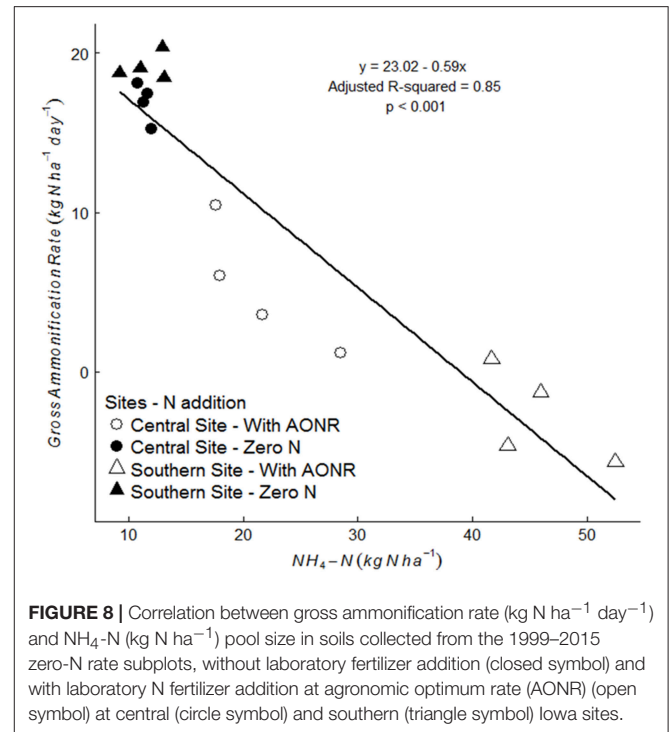


FIGURE 8 | Correlation between gross ammonification rate (kg N ha⁻¹ day⁻¹) and NH₄-N (kg N ha⁻¹) pool size in soils collected from the 1999–2015 zero-N rate subplots, without laboratory fertilizer addition (closed symbol) and with laboratory N fertilizer addition at agronomic optimum rate (AONR) (open symbol) at central (circle symbol) and southern (triangle symbol) Iowa sites.

This pattern is consistent with our laboratory results that demonstrated a direct, suppressive effect of NH₄⁺-N pool on gross ammonification (**Figures 7, 8**).

Implications for Crop Modeling

In our study, the direct effect of inorganic N on microbial activity and SOM dynamics was agronomically relevant—even in the context of large differences in NPP and soil moisture that were imparted by the fertilizer addition (Figures 1, 5). The reduction in gross ammonification ($0.6 - 4.9 \text{ kg N ha}^{-1} \text{ day}^{-1}$) due to synthetic N fertilizer addition was large compared to the typical rate of maize N uptake at the V5 growth stage ($\sim 1.5 \text{ kg N ha}^{-1} \text{ day}^{-1}$; Osterholz et al., 2017). In contrast to this result, most ecosystem process models (e.g., Roth-C; Coleman and Jenkinson, 1996) do not capture the effect of inorganic N on SOM dynamics because ammonification rates are mostly driven by initial SOM levels, soil moisture, and soil temperature dynamics. In more complex models that concurrently account for soil and crop dynamics as well as their interactions (e.g., APSIM), the effect of inorganic N on SOM dynamics becomes an emergent property of the simulation process with apparent differences in soil N mineralization due to N-fertilizer input (Puntel et al., 2016). Nevertheless, the direct effect of inorganic N on microbial activity is not included. This highlights the importance of recent ecosystem process models that incorporate microbial physiology (e.g., Wieder et al., 2014).

The Net Effect of N Fertilizer on Ecosystem NUE

In highly fertile soils, N mineralized from SOM is typically the major source of crop N uptake (Stevens et al., 2005; Gardner and Drinkwater, 2009; Poffenbarger et al., 2018). If N fertilizer reduces SOM-N mineralization and, as a result, increases reliance on fertilizer N, it could increase environmental N losses because mineralized SOM-N is a more efficient source of crop N uptake (Drinkwater and Snapp, 2007). In contrast, if the reduction in early-season SOM-N mineralization (Figure 7) represents a delay in the release of potentially mineralizable SOM-N rather than an absolute reduction in total SOM-N mineralization, it could reduce environmental N losses and increase total crop N uptake (i.e., similar to a delayed-release fertilizer). Coincidentally, comparisons of N fertilizer sources often find NH_4NO_3 to be among the most efficient sources of synthetic N fertilizer (Andraski and Bundy, 2008). New metrics of ecosystem NUE, such as “systems NUE” [$s\text{NUE} = N_{\text{yield}} / (N_{\text{yield}} + N_{\text{loss}})$] where, N_{yield} is grain N harvest and N_{loss} is the sum of all environmental N losses (e.g., gaseous and dissolve) in $\text{kg ha}^{-1} \text{ y}^{-1}$ could identify potential trade-offs between crop N uptake, SOM mineralization, and environmental N loss (Martinez-Feria et al., 2018).

Regardless of the net effect of N fertilizer on SOM mineralization, our results highlight one dilemma of soil C management: “shall we hoard it or use it” (Janzen, 2006)? Rational fertilizer application (i.e., the “historical N rate”) led to greater soil C and N concentrations (Table 3) and soil moisture (Figure 4). However, SOC can be increased by N fertilizer via two pathways: an increase in inputs or a decrease in mineralization. If increases in SOM stocks are achieved via reductions in SOM N mineralization (Figure 6), more

N fertilizer may be required to achieve the same NPP and environmental N losses may be increased. Thus, strategies to increase SOM should aim to increase SOM inputs rather decrease SOM mineralization. A shift from synthetic N fertilizer to manure and legume sources of N are one option to achieve this goal.

CONCLUSION

Synthetic N fertilizer addition consistently reduced gross ammonification at an early growth stage of maize when soil inorganic N concentrations were higher in N-fertilized treatments due to a direct inhibitory effect of NH_4^+ on microbial activity (Figure 8). In later stages of maize growth when soil inorganic N concentrations were similar among N-fertilized and non-fertilized treatments, there was no effect of N fertilizer on gross ammonification.

The suppressive effect of N fertilizer application on gross ammonification is likely linked to microbial N demand. The effect was greatest in treatments that had not received historical N fertilizer applications and had lower SOC and total N, suggesting microbes may be mineralizing SOM to meet N demand. In contrast, at the high historical N fertilizer rates, which had greater total soil N concentrations, fertilizer N did not significantly reduce ammonification. In these treatments, historically high N fertilizer inputs and accompanying increases in total soil N may have increased N availability to a point that it no longer limited microbial activity.

Given the suppression of gross ammonification by N fertilizer, the “N difference” method to measure FNUE may underestimate FNUE, especially in systems with low SOM. This result contrasts with previous reports, which suggest that the indirect “N difference” method overestimate FNUE. Although future research will be required to determine why the N difference method consistently estimates higher FNUE than the ^{15}N isotope method, the relatively inexpensive “N difference” method should not overestimate FNUE and thus may provide an accurate estimate of FNUE. This result could have widespread application for research aiming to increase NUE of maize agroecosystems thus lowering N losses and associated economic and environmental impacts.

AUTHOR CONTRIBUTIONS

NM, MC, HP, and JS designed the experiment. NM, MC, and WO collected soil samples and set up the gross ammonification assays. HP collected crop yield and soil C:N, bulk density data. NM, FM, and MC synthesized results and constructed tables and figures for the paper. NM and MC created the first draft. WO, FM, HP, JS, DO, and SA contributed to the final version of the manuscript.

ACKNOWLEDGMENTS

This study was funded by the United States Department of Agriculture National Institute of Food and Agriculture

grant number 2014-67019-21629 and the Iowa State University Plant Sciences Institute Faculty Scholars Program. We are grateful to the Iowa State University Research Farm personnel for maintaining the N rate experiments.

REFERENCES

- Abendroth, L. J., Elmore, R. W., Boyer, M. J., and Marlay, S. K. (2011). *Corn Growth and Development*. Ames, IA: Iowa State University.
- Andraski, T. W., and Bundy, L. G. (2008). Corn residue and nitrogen source effects on nitrogen availability in no-till corn. *Agron. J.* 100, 1274–1279. doi: 10.2134/agronj2008.0039
- Ayres, E., Steltzer, H., Simmons, B. L., Simpson, R. T., Steinweg, J. M., Wallenstein, M. D., et al. (2009). Home-field advantage accelerates leaf litter decomposition in forests. *Soil Biol. Biochem.* 41, 606–610. doi: 10.1016/j.soilbio.2008.12.022
- Booth, M. S., Stark, J. M., and Rastetter, E. (2005). Controls on nitrogen cycling in terrestrial ecosystems: a synthetic analysis of literature data. *Ecol. Monogr.* 75, 139–157. doi: 10.1890/04-0988
- Brown, K. H., Bach, E. M., Drijber, R. A., Hofmockel, K. S., Jeske, E. S., Sawyer, J. E., et al. (2014). A long-term nitrogen fertilizer gradient has little effect on soil organic matter in a high-intensity maize production system. *Glob. Chang. Biol.* 20, 1339–1350. doi: 10.1111/gcb.12519
- Cassman, K. G. (1999). Ecological intensification of cereal production systems: yield potential, soil quality, and precision agriculture. *Proc. Natl. Acad. Sci. U.S.A.* 96, 5952–5959.
- Cassman, K. G., Dobermann, A., and Walters, D. T. (2002). Agroecosystems, nitrogen-use efficiency, and nitrogen management. *Ambio* 31, 132–140. doi: 10.1579/0044-7447-31.2.132
- Chen, R., Senbayram, M., Blagodatsky, S., Myachina, O., Dittert, K., Lin, X., et al. (2014). Soil C and N availability determine the priming effect: microbial N mining and stoichiometric decomposition theories. *Glob. Chang. Biol.* 20, 2356–2367. doi: 10.1111/gcb.12475
- Cheng, W. (2009). Rhizosphere priming effect: its functional relationships with microbial turnover, evapotranspiration, and C–N budgets. *Soil Biol. Biochem.* 41, 1795–1801. doi: 10.1016/j.soilbio.2008.04.018
- Ciampitti, I. A., and Vyn, T. J. (2012). Physiological perspectives of changes over time in maize yield dependency on nitrogen uptake and associated nitrogen efficiencies: a review. *Field Crops Res.* 133, 48–67. doi: 10.1016/j.fcr.2012.03.008
- Coleman, K., and Jenkinson, D. S. (1996). “RothC-26.3: a model for the turnover of carbon in soil,” in *Evaluation of Soil Organic Matter Models Using Existing Long-Term Datasets, NATO ASI Series I*, Vol. 38, eds D. S. Powlson, P. Smith, and P. J. U. Smith (Heidelberg: Springer-Verlag), 237–246. doi: 10.1007/978-3-642-61094-3_17
- Craine, J. M., Morrow, C., and Fierer, N. (2007). Microbial nitrogen limitation increases decomposition. *Ecology* 88, 2105–2113. doi: 10.1890/06-1847.1
- Dobermann, A. (2007). “Nutrient use efficiency - measurement and management” in *IFA International Workshop on Fertilizer Best Management Practices* (Brussels), 1–28.
- Drinkwater, L. E., and Snapp, S. S. (2007). Nutrients in agroecosystems: rethinking the management paradigm. *Adv. Agron.* 92, 163–186. doi: 10.1016/S0065-2113(04)92003-2
- Fontaine, S., Mariotti, A., and Abbadie, L. (2003). The priming effect of organic matter: a question of microbial competition? *Soil Biol. Biochem.* 35, 837–843. doi: 10.1016/S0038-0717(03)00123-8
- Gardner, J. B., and Drinkwater, L. E. (2009). The fate of nitrogen in grain cropping systems: a meta-analysis of 15N field experiments. *Ecol. Appl.* 19, 2167–2184. doi: 10.1890/08-1122.1
- Harmsen, K., and Moraghan, J. T. (1988). A comparison of the isotope recovery and difference methods for determining nitrogen fertilizer efficiency. *Plant Soil* 105, 55–67. doi: 10.1007/BF02371143
- Hart, S., Stark, J., Davidson, E., and Firestone, M. K. (1994). “Nitrogen mineralization, immobilization, and nitrification,” in *Methods of Soil Analysis. Part 2. Microbiological and Biochemical Properties*, eds R. Weaver, S. Angle, P. Bottomley, D. Bezdicek, S. Smith, A. Tabatabai, and A. Wollum (Madison, WI: Soil Science Society of America), 985–1018.
- Hauck, R. D., and Bremner, J. M. (1976). Use of tracers for soil and fertilizer nitrogen research. *Adv. Agron.* 28, 219–266. doi: 10.1016/S0065-2113(08)60556-8
- Hood-Nowotny, R., Umana, N. H. N., Inselbacher, E., Oswald-Lachouani, P., and Wanek, W. (2010). Alternative methods for measuring inorganic, organic, and total dissolved nitrogen in soil. *Soil Sci. Soc. Am. J.* 74, 1018–1027. doi: 10.2136/sssaj2009.0389
- Jansson, S. L., and Persson, J. (1982). “Mineralization and immobilization of soil nitrogen,” in *Nitrogen in Agricultural Soils*, ed F. J. Stevenson (Madison, WI: American Society of Agronomy), 229–252.
- Janzen, H. H. (2006). The soil carbon dilemma: shall we hoard it or use it? *Soil Biol. Biochem.* 38, 419–424. doi: 10.1016/j.soilbio.2005.10.008
- Jenkinson, D. S., Fox, R. H., and Rayner, J. H. (1985). Interactions between fertilizer nitrogen and soil nitrogen—the so-called ‘priming’ effect. *Eur. J. Soil Sci.* 36, 425–444.
- Khan, S. A., Mulvaney, R. L., Ellsworth, T. R., and Boast, C. W. (2007). The myth of nitrogen fertilization for soil carbon sequestration. *J. Environ. Qual.* 36, 1821–1832. doi: 10.2134/jeq2007.0099
- Kuzyakov, Y. (2010). Priming effects: interactions between living and dead organic matter. *Soil Biol. Biochem.* 42, 1363–1371. doi: 10.1016/j.soilbio.2010.04.003
- Li, X. G., Jia, B., Lv, J., Ma, Q., Kuzyakov, Y., and Li, F. M. (2017). Nitrogen fertilization decreases the decomposition of soil organic matter and plant residues in planted soils. *Soil Biol. Biochem.* 112, 47–55. doi: 10.1016/j.soilbio.2017.04.018
- Li, Y., Liu, Y., Wang, Y., Niu, L., Xu, X., and Tian, Y. (2014). Interactive effects of soil temperature and moisture on soil N mineralization in a *Stipa krylovii* grassland in Inner Mongolia, China. *J. Arid Land* 6, 571–580. doi: 10.1007/s40333-014-0025-5
- Maharjan, B., Venterea, R. T., and Rosen, C. (2014). Fertilizer and irrigation management effects on nitrous oxide emissions and nitrate leaching. *Agron. J.* 106, 703–714. doi: 10.2134/agronj2013.0179
- Martinez-Feria, R. A., Castellano, M. J., Dietzel, R. N., Helmers, M. J., Liebman, M., Huber, I., et al. (2018). Linking crop- and soil-based approaches to evaluate system nitrogen-use efficiency and tradeoffs. *Agric. Ecosyst. Environ.* 256, 131–143. doi: 10.1016/j.agee.2018.01.002
- Mitchell, D. C., Castellano, M. J., Sawyer, J. E., and Pantoja, J. (2013). Cover crop effects on nitrous oxide emissions: role of mineralizable carbon. *Soil Sci. Soc. Am. J.* 77, 1765–1773. doi: 10.2136/sssaj2013.02.0074
- Moorhead, D. L., and Sinsabaugh, R. L. (2006). A theoretical model of litter decay and microbial interaction. *Ecol. Monogr.* 76, 151–174. doi: 10.1890/0012-9615(2006)076[0151:ATMOLD]2.0.CO;2
- Mulvaney, R. L., Khan, S. A., and Ellsworth, T. R. (2009). Synthetic nitrogen fertilizers deplete soil nitrogen: a global dilemma for sustainable cereal production. *J. Environ. Qual.* 38, 2295–2314. doi: 10.2134/jeq2008.0527
- Osterholz, W. R., Rinot, O., Liebman, M., and Castellano, M. J. (2017). Can mineralization of soil organic nitrogen meet maize nitrogen demand? *Plant Soil* 415, 73–84. doi: 10.1007/s11104-016-3137-1
- Parton, W. J., Schimel, D. S., Cole, C. V., and Ojima, D. S. (1987). Analysis of factors controlling soil organic matter levels in great plains grasslands. *Soil Sci. Soc. Am. J.* 51, 1173–1179. doi: 10.2136/sssaj1987.03615995005100050015x
- Poffenbarger, H. J., Barker, D. W., Helmers, M. J., Miguez, F. E., Olk, D. C., Sawyer, J. E., et al. (2017). Maximum soil organic carbon storage in Midwest US cropping systems when crops are optimally nitrogen-fertilized. *PLoS ONE* 12:e0172293. doi: 10.1371/journal.pone.0172293
- Poffenbarger, H. J., Sawyer, J. E., Barker, D. W., Olk, D. C., Six, J., and Castellano, M. J. (2018). Legacy effects of long-term nitrogen fertilizer application on the fate of nitrogen fertilizer inputs in continuous maize. *Agric. Ecosyst. Environ.* 265, 544–555.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fevo.2019.00059/full#supplementary-material>

- Puntel, L. A., Sawyer, J. E., Barker, D. W., Dietzel, R., Poffenbarger, H., Castellano, M. J., et al. (2016). Modeling long-term corn yield response to nitrogen rate and crop rotation. *Front. Plant Sci.* 7:1630. doi: 10.3389/fpls.2016.01630
- Robertson, G. P., Bruulsema, T. W., Gehl, R. J., Kanter, D., Mauzerall, D. L., Rotz, C. A., et al. (2013). Nitrogen-climate interactions in US agriculture. *Biogeochemistry* 114, 41–70. doi: 10.1007/s10533-012-9802-4
- Russell, A. E., Cambardella, C. A., Laird, D. A., Jaynes, D. B., and Meek, D. W. (2009). Nitrogen fertilizer effects on soil carbon balances in Midwestern US agricultural systems. *Ecol. Appl.* 19, 1102–1113. doi: 10.1890/07-1919.1
- SAS Institute (2013). *PC Windows Version 9.4*. Cary, NC: SAS Institute Inc.
- Spohn, M., Pötsch, E. M., Eichorst, S. A., Woebken, D., Wanek, W., and Richter, A. (2016). Soil microbial carbon use efficiency and biomass turnover in a long-term fertilization experiment in a temperate grassland. *Soil Biol. Biochem.* 97, 168–175. doi: 10.1016/j.soilbio.2016.03.008
- Stark, J. M., and Hart, S. C. (1996). Diffusion technique for preparing salt solutions, Kjeldahl digests, and persulfate digests for nitrogen-15 analysis. *Soil Sci. Soc. Am. J.* 60, 1846–1855. doi: 10.2136/sssaj1996.03615995006000060033x
- Sterner, R. W., and Elser, J. J. (2002). *Ecological Stoichiometry: The Biology of Elements From Molecules to the Biosphere*. Princeton, NJ: Princeton University Press.
- Stevens, W. B., Hoefl, R. G., and Mulvaney, R. L. (2005). Fate of nitrogen-15 in a long-term nitrogen rate study. *Agron. J.* 97, 1046–1053. doi: 10.2134/agronj2003.0313
- Varvel, G. E., and Peterson, T. A. (1990). Nitrogen fertilizer recovery by corn in monoculture and rotation systems. *Agron. J.* 82, 935–938. doi: 10.2134/agronj1990.00021962008200050019x
- Vitousek, P. M., Naylor, R., Crews, T., David, M. B., Drinkwater, L. E., Holland, E., et al. (2009). Nutrient imbalances in agricultural development. *Science* 324, 1519–1520. doi: 10.1126/science.1170261
- Wang, C., Wan, S., Xing, X., Zhang, L., and Han, X. (2006). Temperature and soil moisture interactively affected soil net N mineralization in temperate grassland in Northern China. *Soil Biol. Biochem.* 38, 1101–1110. doi: 10.1016/j.soilbio.2005.09.009
- Wieder, W. R., Grandy, A. S., Kallenbach, C. M., and Bonan, G. B. (2014). Integrating microbial physiology and physio-chemical principles in soils with the MIMICS model. *Biogeosciences* 11, 3899–3917. doi: 10.5194/bg-11-3899-2014
- Zang, H., Wang, J., and Kuzyakov, Y. (2016). N fertilization decreases soil organic matter decomposition in the rhizosphere. *Appl. Soil Ecol.* 108, 47–53. doi: 10.1016/j.apsoil.2016.07.021

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Mahal, Osterholz, Miguez, Poffenbarger, Sawyer, Olk, Archontoulis and Castellano. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.