

University of Kentucky UKnowledge

Sanders-Brown Center on Aging Faculty Publications

Aging

2-5-2014

Microsome Isolation from Tissue

Maria Bodero *University of Kentucky*

Jose Francisco Abisambra University of Kentucky, joe.abisambra@uky.edu

Right click to open a feedback form in a new tab to let us know how this document benefits you.

Follow this and additional works at: https://uknowledge.uky.edu/sbcoa_facpub Part of the <u>Physiology Commons</u>

Repository Citation

Bodero, Maria and Abisambra, Jose Francisco, "Microsome Isolation from Tissue" (2014). *Sanders-Brown Center on Aging Faculty Publications*. 116. https://uknowledge.uky.edu/sbcoa_facpub/116

This Article is brought to you for free and open access by the Aging at UKnowledge. It has been accepted for inclusion in Sanders-Brown Center on Aging Faculty Publications by an authorized administrator of UKnowledge. For more information, please contact UKnowledge@lsv.uky.edu.

Microsome Isolation from Tissue

Notes/Citation Information Published in *Bio-protocol*, v. 4, issue 3, p. 1-3.

Copyright © 2014 The Authors; exclusive licensee Bio-protocol LLC.

The publisher has granted the permission for posting the article here.

Digital Object Identifier (DOI)

https://doi.org/10.21769/BioProtoc.1038

Microsome Isolation from Tissue

Maria Bodero and Jose Francisco Abisambra*

Sanders-Brown Center on Aging and Department of Physiology, University of Kentucky, Lexington, USA

*For correspondence: joe.abisambra@uky.edu

[Abstract] This protocol details the extraction of microsomes from frozen tissue in order to further examine the protein-protein interactions occurring within the endoplasmic reticulum. This protocol was adapted from Abisambra *et al.* (2013) with modifications made in order to optimize for subsequent use.

Materials and Reagents

- 1. Sucrose
- 2. Protease Inhibitor cocktail, EDTA free (Merck KGaA, Calbiochem, catalog number: 539134)
- 3. Phosphatase inhibitor cocktail II
- 4. Phosphatase inhibitor cocktail III
- 5. PMSF at 10 mM in DMSO or 1.74 mg/ml (Thermo Fisher Scientific, catalog number: 36978)
- 6. Phosphatase Arrest II cocktail (Geno Technology, catalog number: 786-451)
- 7. Phosphatase Arrest III cocktail (Geno Technology, catalog number: 786-452)
- 8. M-PER Mammalian Protein Extraction Reagent (Thermo Fisher Scientific, catalog number: 78501)

Equipment

- 1. Sterile bottle filter
- 2. Glass Dounce homogenizer
- 3. Refrigerated centrifuge
- 4. Microfuge tubes rated for at least 25,000 *x g* centrifugation



Procedure

1. Make a 0.25 M sucrose solution that contains protease inhibitor cocktail, phosphatase inhibitor cocktails II and III, and PMSF as follows:

Per 100 µl of Sucrose master mix add:

- a. 96 µl of 0.25 M sucrose
- b. 1 µl of protease inhibitor cocktail
- c. 1 µl of phosphatase inhibitor cocktail II
- d. 1 µl of phosphatase inhibitor cocktail III
- e. 1 µl of PMSF
- Weigh tissue to be analyzed and add 10x its mass in volume of sucrose master mix (see step 1; *i.e.* 100 mg = 1,000 μl of sucrose solution).
- 3. While keeping all solutions on ice, add the appropriate amount of sucrose solution to tissue and dounce homogenize until a completely homogenous solution is obtained.
- 4. Spin the homogenate at 10,000 x g for 10 min at 4 °C.
- 5. Transfer the supernatants to a new microfuge tube (save the pellet at -20 °C) and spin at $30,000 \times g$ for 90 min in a fixed angle rotor (or at 25,800 $\times g$ for 2 h).
- 6. Transfer the supernatant to a different microfuge tube and save at -20 °C. The remaining pellet corresponds to the microsomal fraction.
- 7. Pipette gently to resuspend the microsome pellet in 200 μ l of the following mix (per 100 μ l):
 - a. 96 µl of MPER buffer
 - b. 1 µl of protease inhibitor cocktail
 - c. 1 µl of phosphatase arrest cocktail II
 - d. 1 µl of phosphatase arrest cocktail III
 - e. 1 µl of PMSF

Acknowledgments

We thank Dr. Gene Ness, Dr. Huntington Potter, and Dr. Chad Dickey for supporting the development and adaptation of this protocol in their labs. We credit the following article for this work: Abisambra *et al.* (2013). Financial support during the time of protocol development came from the Alzheimer's Association NIRGD-12-242642, the Foundation for PSP/CBD and Related Brain Disorders (6144107400), and NIH/NIA ADC Pilot Grant from 5P30AG028383-08.



References

- Abisambra, J. F., Jinwal, U. K., Blair, L. J., O'Leary, J. C., 3rd, Li, Q., Brady, S., Wang, L., Guidi, C. E., Zhang, B., Nordhues, B. A., Cockman, M., Suntharalingham, A., Li, P., Jin, Y., Atkins, C. A. and Dickey, C. A. (2013). <u>Tau accumulation activates the unfolded</u> <u>protein response by impairing endoplasmic reticulum-associated degradation.</u> *J Neurosci* 33(22): 9498-9507.
- Abisambra, J. F., Fiorelli, T., Padmanabhan, J., Neame, P., Wefes, I. and Potter, H. (2010). <u>LDLR expression and localization are altered in mouse and human cell culture</u> <u>models of Alzheimer's disease</u>. *PLoS One* 5(1): e8556.