

6-2019

Crystal Structure of Zymonic Acid and a Redetermination of Its Precursor, Pyruvic Acid

Dominik Heger

University of Kentucky, hegerd@uky.edu

Alexis J. Eugene

University of Kentucky, alexis.eugene@uky.edu

Sean R. Parkin


University of Kentucky, s.parkin@uky.edu

Marcelo I. Guzman

University of Kentucky, marcelo.guzman@uky.edu

Right click to open a feedback form in a new tab to let us know how this document benefits you.

Follow this and additional works at: https://uknowledge.uky.edu/chemistry_facpub

 Part of the [Diagnosis Commons](#), [Environmental Chemistry Commons](#), [Food Chemistry Commons](#), [Investigative Techniques Commons](#), [Laboratory and Basic Science Research Commons](#), [Organic Chemicals Commons](#), [Organic Chemistry Commons](#), and the [Physical Chemistry Commons](#)

Repository Citation

Heger, Dominik; Eugene, Alexis J.; Parkin, Sean R.; and Guzman, Marcelo I., "Crystal Structure of Zymonic Acid and a Redetermination of Its Precursor, Pyruvic Acid" (2019). *Chemistry Faculty Publications*. 147.

https://uknowledge.uky.edu/chemistry_facpub/147

Crystal Structure of Zymonic Acid and a Redetermination of Its Precursor, Pyruvic Acid

Notes/Citation Information

Published in *Acta Crystallographica Section E: Crystallographic Communications*, v. 75, part 6, p. 858-862.

This is an open-access article distributed under the terms of the [Creative Commons Attribution \(CC-BY\) Licence](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original authors and source are cited.

Digital Object Identifier (DOI)

<https://doi.org/10.1107/S2056989019007072>



Crystal structure of zymonic acid and a redetermination of its precursor, pyruvic acid

Dominik Heger,^{a,b*} Alexis J. Eugene,^a Sean R. Parkin^{a*} and Marcelo I. Guzman^{a*}^aDepartment of Chemistry, University of Kentucky, Lexington, Kentucky 40506, USA, and ^bDepartment of Chemistry, Faculty of Science, Masaryk University, Kamenice 5, 625 00 Brno, Czech Republic. *Correspondence e-mail: hegerd@chemi.muni.cz, s.parkin@uky.edu, marcelo.guzman@uky.edu

Received 15 April 2019

Accepted 15 May 2019

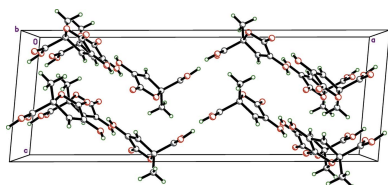
Edited by W. T. A. Harrison, University of Aberdeen, Scotland

Keywords: crystal structure; hydrogen bonding; low temperature; zymonic; pyruvic.**CCDC references:** 1916323; 1916322**Supporting information:** this article has supporting information at journals.iucr.org/e

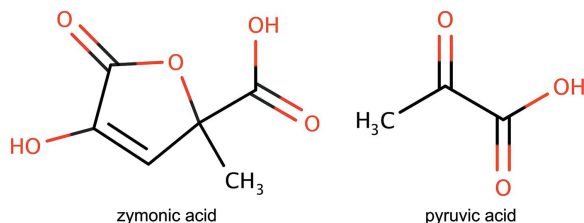
The structure of zymonic acid (systematic name: 4-hydroxy-2-methyl-5-oxo-2,5-dihydrofuran-2-carboxylic acid), $C_6H_6O_5$, which had previously eluded crystallographic determination, is presented here for the first time. It forms by intramolecular condensation of parapyruvic acid, which is the product of aldol condensation of pyruvic acid. A redetermination of the crystal structure of pyruvic acid (systematic name: 2-oxopropanoic acid), $C_3H_4O_3$, at low temperature (90 K) and with increased precision, is also presented [for the previous structure, see: Harata *et al.* (1977). *Acta Cryst.* **B33**, 210–212]. In zymonic acid, the hydroxylactone ring is close to planar (r.m.s. deviation = 0.0108 Å) and the dihedral angle between the ring and the plane formed by the bonds of the methyl and carboxylic acid carbon atoms to the ring is 88.68 (7)°. The torsion angle of the carboxylic acid group relative to the ring is 12.04 (16)°. The pyruvic acid molecule is almost planar, having a dihedral angle between the carboxylic acid and methyl-ketone groups of 3.95 (6)°. Intermolecular interactions in both crystal structures are dominated by hydrogen bonding. The common $R_2^2(8)$ hydrogen-bonding motif links carboxylic acid groups on adjacent molecules in both structures. In zymonic acid, this results in dimers about a crystallographic twofold of space group $C2/c$, which forces the carboxylic acid group to be disordered exactly 50:50, which scrambles the carbonyl and hydroxyl groups and gives an apparent equalization of the C–O bond lengths [1.2568 (16) and 1.2602 (16) Å]. The other hydrogen bonds in zymonic acid (O–H···O and weak C–H···O), link molecules across a 2_1 -screw axis, and generate an $R_2^2(9)$ motif. These hydrogen-bonding interactions propagate to form extended pleated sheets in the *ab* plane. Stacking of these zigzag sheets along *c* involves only van der Waals contacts. In pyruvic acid, inversion-related molecules are linked into $R_2^2(8)$ dimers, with van der Waals interactions between dimers as the only other intermolecular contacts.

1. Chemical context

The Human Metabolome Database (Wishart *et al.*, 2007, 2009, 2013, 2018) lists the compound 4-hydroxy-2-methyl-5-oxo-furan-2-carboxylic acid ($C_6H_6O_5$), commonly named zymonic acid, with the metabocard HMDB0031210. Zymonic acid is used as a flavor constituent for confectionery and tobacco products (Yannai, 2004). The generation of zymonic acid can proceed by condensation of parapyruvic acid, which itself forms by aldol condensation of pyruvic acid (IUPAC name 2-oxopropanoic acid, $C_3H_4O_3$; Bloomer *et al.*, 1970). Therefore, zymonic acid is directly derived from pyruvic acid, and is thus related to the compounds present in the tricarboxylic acid (Krebs) cycle (Nelson & Cox, 2004) and its reductive version (Guzman, 2011; Guzman & Martin, 2008; Zhou & Guzman, 2016). As an intermediate in central metabolism, zymonic acid



is produced in the cytoplasm at very low concentration, from where it can be excreted to the extracellular region.



The electron-impact mass spectrum (MS) and electrospray ionization fragmentation of zymonic acid following gas and liquid chromatography, respectively, have been reported (Allen *et al.*, 2015, 2016). The use of ^{13}C -zymonic acid has enabled mapping of pH changes, independently of concentration, in mammalian organs and tumors *via* hyperpolarized magnetic resonance (Düwel *et al.*, 2017). Thus, zymonic acid is a non-invasive extracellular imaging sensor to localize and quantify pH *in vivo* (Düwel *et al.*, 2017; Hundshammer *et al.*, 2017), with many possible applications in medical diagnosis (Schilling *et al.*, 2016). As part of the process resulting in the aforementioned invention, the detailed ^1H and ^{13}C NMR spectra of pure zymonic acid have been reported (Hundshammer *et al.*, 2017). Herein, we contribute new information to characterize zymonic acid by reporting for the first time its crystal structure, along with a low-temperature redetermination of pyruvic acid.

2. Structural commentary

Aside from the effects on the geometry of the carboxylic acid group in zymonic acid that stem from disorder about the twofold axis (see below), there are no unusual bond lengths or angles in either compound.

In zymonic acid (Fig. 1), the hydroxylactone ring is essentially planar (r.m.s. deviation = 0.0108 Å), with the largest deviation from planarity [0.0171 (8) Å] for the ring oxygen atom, O3. The plane defined by the ring carbon atom C4, the methyl carbon atom C6, and the carboxylic acid carbon atom

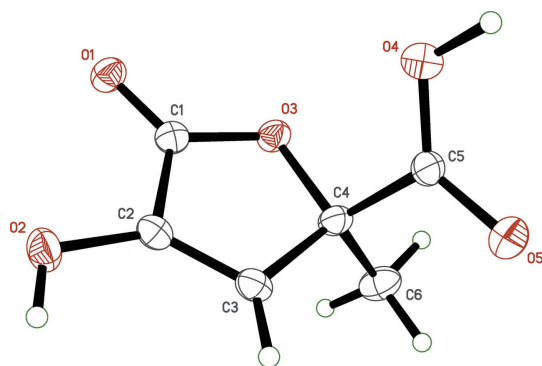


Figure 1
The molecular structure of zymonic acid, with displacement ellipsoids drawn at the 50% probability level.

Table 1
Hydrogen-bond geometry (Å, °) for zymonic acid.

$D-H\cdots A$	$D-H$	$H\cdots A$	$D\cdots A$	$D-H\cdots A$
$O2-H2\cdots O1^i$	0.84	1.96	2.7103 (14)	148
$C3-H3\cdots O2^i$	0.95	2.48	3.0720 (16)	120
$O4-H4O\cdots O4^{ii}$	1.09	1.52	2.607 (2)	176
$O5-H5O\cdots O5^{ii}$	0.99	1.63	2.624 (2)	179

Symmetry codes: (i) $-x + \frac{3}{2}, y + \frac{1}{2}, -z + \frac{3}{2}$; (ii) $-x + 1, y, -z + \frac{3}{2}$.

C5, is almost perpendicular to the mean plane of the ring atoms [dihedral angle = 88.68 (7)°]. Lastly, the orientation of the carboxylic acid group relative to the ring, as defined by the torsion angle $O4-C5-C4-O3$, is 12.04 (16)°. For the carboxylic acid group, disorder about the crystallographic twofold axis effectively averages the $\text{C}=\text{O}$ double and $\text{C}-\text{O}$ single bonds, rendering them equivalent [the $\text{C5}-\text{O4}$ and $\text{C5}-\text{O5}$ distances are 1.2568 (16) and 1.2602 (16) Å, respectively], and requires modeling of half-occupancy hydrogens (H4O and H5O) on each.

In spite of increased precision resulting from much lower temperature (90 K *versus* 266 K) and data collection on modern equipment, the redetermined structure of pyruvic acid (Fig. 2) is largely unchanged from that reported by Harata *et al.* (1977). For example, the dihedral angle between the planes defined by atoms $\text{C1}/\text{C2}/\text{C3}/\text{O3}$ and $\text{C1}/\text{C2}/\text{O1}/\text{O2}$ is 3.95 (6)° at 90.00 (2) K *versus* 3.5° at 266 (1) K.

3. Supramolecular features

The main intermolecular interactions in the crystals of both zymonic and pyruvic acids are hydrogen bonds. In zymonic acid, the carboxylic acid groups of adjacent molecules are related by a crystallographic twofold axis to form hydrogen bonds [$\text{O4}-\text{H4O}\cdots\text{O4}^{ii}$ and $\text{O5}-\text{H5O}\cdots\text{O5}^{ii}$; symmetry code: (ii) $1 - x, y, \frac{3}{2} - z$] giving $R_2^2(8)$ dimer motifs (Table 1). This common supramolecular construct in carboxylic acids usually occurs between inversion-related or symmetry-independent molecules. Here, the orientation of the dimer relative to the crystallographic twofold axis forces the average struc-

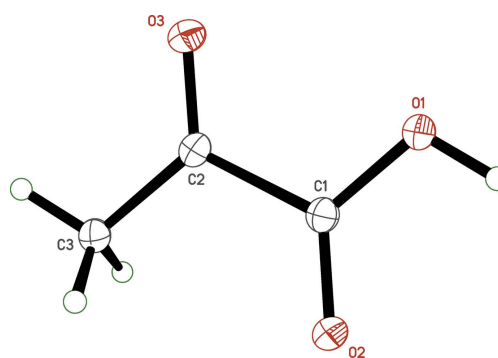


Figure 2
The molecular structure of pyruvic acid, with displacement ellipsoids drawn at the 50% probability level.

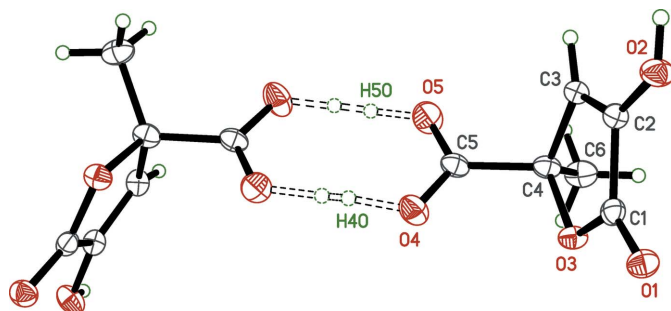


Figure 3
The $R_2^2(8)$ dimer of zymonic acid. Unlabeled atoms are related to their labeled counterparts by a crystallographic twofold axis ($1 - x, y, \frac{3}{2} - z$). This uncommon symmetry [for an $R_2^2(8)$ dimer] forces the O—H...O hydrogen bonds involved to be 50:50 disordered about the twofold axis.

ture to be statistically disordered (Fig. 3). Another pair of hydrogen bonds [O2—H2...O1ⁱ and C3—H3...O2ⁱ; symmetry code: (i) $\frac{3}{2} - x, \frac{1}{2} + y, \frac{3}{2} - z$], link molecules related by a 2_1 -screw axis, into $R_2^2(9)$ motifs (Fig. 4). These hydrogen-bonding interactions combine to form extended pleated sheets that propagate in the *ab* plane (Fig. 5), which in turn, stack along the *c*-axis direction. In pyruvic acid, inversion-related molecules form the common $R_2^2(8)$ dimer motif (Fig. 6,

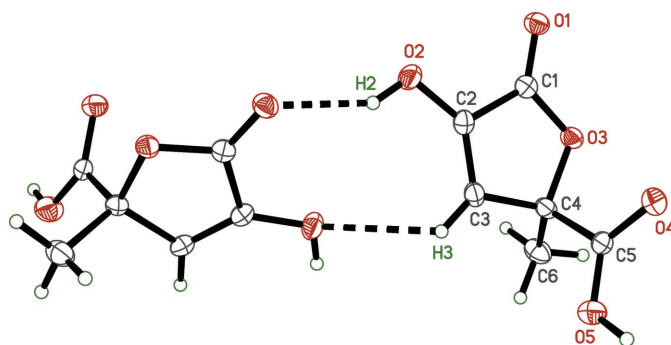


Figure 4
The $R_2^2(9)$ dimer of zymonic acid. Unlabeled atoms are related to their labeled counterparts by a crystallographic 2_1 -screw axis ($\frac{3}{2} - x, \frac{1}{2} + y, \frac{3}{2} - z$). Disorder of the carboxylic acid H atoms is omitted to enhance clarity.

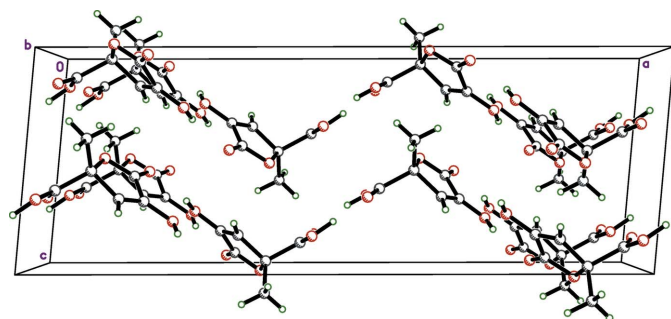


Figure 5
A packing plot of zymonic acid viewed down the *b* axis, showing the stacking along *c* of zigzag pleated assemblies of molecules. Disorder of the carboxylic acid hydrogen atoms is omitted to enhance clarity.

Table 2
Hydrogen-bond geometry (Å, °) for pyruvic acid.

<i>D</i> —H... <i>A</i>	<i>D</i> —H	H... <i>A</i>	<i>D</i> ... <i>A</i>	<i>D</i> —H... <i>A</i>
O1—H1...O2 ⁱ	0.913 (14)	1.742 (14)	2.6536 (8)	175.5 (12)

Symmetry code: (i) $-x + 1, -y + 1, -z + 1$.

Table 2). In accordance with the work of Harata *et al.* (1977), there are no other noteworthy intermolecular interactions.

4. Database survey

A search of the Cambridge Crystal Structure Database (Version 5.40, Nov. 2018; Groom *et al.*, 2016) for zymonic acid gave no hits for searches on either ‘zymonic’ or on the structural formula. A search on the structural formula of pyruvic acid gave two hits. CSD entry PRUVAC (Harata *et al.*, 1977) describes the pure compound at 266 K, and is similar to the present pyruvic acid structure (after transformation to a common cell setting). CSD entry FAFGUR (Prohens *et al.*, 2016) describes a co-crystal of pyruvic acid with the drug agomelatine. The CSD does contain structures for derivatives of both zymonic and pyruvic acids, but none of these have features that are especially relevant to the current work.

5. Synthesis and crystallization

Vacuum distillation of pyruvic acid (Sigma–Aldrich, 98.5%) was used for purification (Eugene & Guzman, 2017*a,b*). Freshly distilled pyruvic acid was crystallized in a closed vial in a freezer at 253 K. The tail of this distillation, a viscous yellowish residue enriched in parapyrvic and zymonic acids, was isolated in a vial, and the headspace filled with N₂(g) before sealing it with a cap. Crystals of zymonic acid were produced slowly from this isolated residue kept at 275 K inside a refrigerator. The easily identifiable transparent crystals of zymonic acid appear above the level of the viscous solution within two weeks. Pyruvic acid crystals are deliquescent in air, even at 263 K (Harata *et al.*, 1977), so they had to be kept cold, with minimal exposure to ambient air. Thus, throughout all experimental stages from initial inspection through data collection, special techniques for crystal handling at low temperature (Parkin & Hope, 1998) were employed.

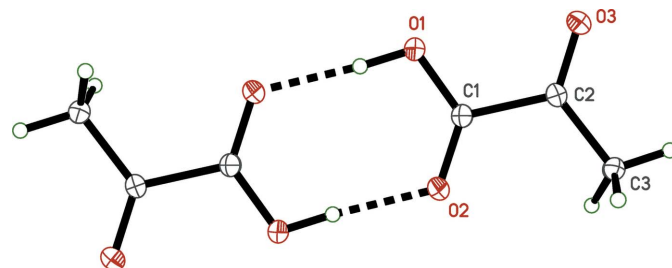


Figure 6
The $R_2^2(8)$ dimer of pyruvic acid. Unlabeled atoms are related to their labeled counterparts by crystallographic inversion symmetry ($1 - x, 1 - y, 1 - z$).

Table 3
Experimental details.

	zymonic acid	pyruvic acid
Crystal data		
Chemical formula	C ₆ H ₆ O ₅	C ₃ H ₄ O ₃
<i>M_r</i>	158.11	88.06
Crystal system, space group	Monoclinic, <i>C2/c</i>	Monoclinic, <i>P2₁/c</i>
Temperature (K)	90	90
<i>a</i> , <i>b</i> , <i>c</i> (Å)	24.145 (3), 6.6523 (7), 8.6201 (7)	10.7486 (3), 5.1925 (2), 6.8302 (2)
β (°)	95.169 (4)	99.063 (1)
<i>V</i> (Å ³)	1378.9 (3)	376.45 (2)
<i>Z</i>	8	4
Radiation type	Mo <i>K</i> α	Mo <i>K</i> α
μ (mm ⁻¹)	0.14	0.14
Crystal size (mm)	0.30 × 0.25 × 0.02	0.26 × 0.22 × 0.18
Data collection		
Diffractometer	Bruker D8 Venture dual source	Bruker D8 Venture dual source
Absorption correction	Multi-scan (<i>SADABS</i> ; Krause <i>et al.</i> , 2015)	Multi-scan (<i>SADABS</i> ; Krause <i>et al.</i> , 2015)
<i>T_{min}</i> , <i>T_{max}</i>	0.721, 0.959	0.890, 0.971
No. of measured, independent and observed [<i>I</i> > 2 σ (<i>I</i>)] reflections	18595, 1586, 1392	10479, 1425, 1242
<i>R_{int}</i>	0.062	0.025
(<i>sin</i> θ / λ) _{max} (Å ⁻¹)	0.650	0.796
Refinement		
<i>R</i> [<i>F</i> ² > 2 σ (<i>F</i> ²)], <i>wR</i> (<i>F</i> ²), <i>S</i>	0.037, 0.100, 1.09	0.031, 0.082, 1.08
No. of reflections	1586	1425
No. of parameters	104	60
H-atom treatment	H atoms treated by a mixture of independent and constrained refinement	H atoms treated by a mixture of independent and constrained refinement
$\Delta\rho_{\max}$, $\Delta\rho_{\min}$ (e Å ⁻³)	0.36, -0.24	0.40, -0.21

Computer programs: *APEX3* (Bruker, 2016), *SHELXT* (Sheldrick, 2015a), *SHELXL2018/3* (Sheldrick, 2015b), *XP in SHELXTL* (Sheldrick, 2008), and *CIFFIX* (Parkin, 2013).

6. Refinement

Crystal data, data collection, and structure refinement details are summarized in Table 3. Non-disordered hydrogen atoms were found in difference Fourier maps. For pyruvic acid, the hydroxyl hydrogen-atom coordinates were refined freely, while methyl hydrogen C–H distances used a riding model that allowed the C–H distance to refine. For zymonic acid, riding models were used for all hydrogen atoms apart from those disordered about the twofold axis, which were modeled in accordance with the recommendations of Fábrý (2018). *U*_{iso}(H) parameters of non-disordered hydrogens were set to either 1.2*U*_{eq} or 1.5*U*_{eq} (for the methyl and hydroxyl groups, respectively) of the attached atom. To ensure stable refinement of disordered groups in the zymonic acid structure, constraints (*SHELXL* command EADP) were used to equalize displacement parameters of superimposed atoms.

Funding information

We are thankful for research funding from the National Science Foundation under NSF CAREER award CHE-1255290 to MIG, and the MRI program, grants CHE-0319176 and CHE-1625732. An International Visiting Scholar grant from the College of Arts and Sciences at UK is gratefully acknowledged.

References

- Allen, F., Greiner, R. & Wishart, D. (2015). *Metabolomics*, **11**, 98–110.
- Allen, F., Pon, A., Greiner, R. & Wishart, D. (2016). *Anal. Chem.* **88**, 7689–7697.
- Bloomer, J. L., Gross, M. A., Kappler, F. E. & Pandey, G. N. (1970). *J. Chem. Soc. D*, p. 1030a.
- Bruker (2016). *APEX3*. Bruker AXS Inc., Madison, Wisconsin, USA.
- Düwel, S., Hundhammer, C., Gersch, M., Feuerecker, B., Steiger, K., Buck, A., Walch, A., Haase, A., Glaser, S. J., Schwaiger, M. & Schilling, F. (2017). *Nat. Commun.* **8**, 15126.
- Eugene, A. J. & Guzman, M. I. (2017a). *J. Phys. Chem. A*, **121**, 2924–2935.
- Eugene, A. J. & Guzman, M. I. (2017b). *J. Phys. Chem. A*, **121**, 8741–8744.
- Fábrý, J. (2018). *Acta Cryst.* **E74**, 1344–1357.
- Groom, C. R., Bruno, I. J., Lightfoot, M. P. & Ward, S. C. (2016). *Acta Cryst.* **B72**, 171–179.
- Guzman, M. I. (2011). *Origins of Life: The Primal Self-Organization*, edited by R. Egel, D.-H. Lankenau & A. Y. Mulikjanian, pp. 85–105: Springer Berlin Heidelberg.
- Guzman, M. I. & Martin, S. T. (2008). *Int. J. Astrobiology*, **7**, 271–278.
- Harata, K., Sakabe, N. & Tanaka, J. (1977). *Acta Cryst.* **B33**, 210–212.
- Hundhammer, C., Düwel, S., Köcher, S. S., Gersch, M., Feuerecker, B., Scheurer, C., Haase, A., Glaser, S. J., Schwaiger, M. & Schilling, F. (2017). *ChemPhysChem*, **18**, 2422–2425.
- Krause, L., Herbst-Irmer, R., Sheldrick, G. M. & Stalke, D. (2015). *J. Appl. Cryst.* **48**, 3–10.
- Nelson, D. L. & Cox, M. (2004). *Lehninger Principles of Biochemistry*, 4th ed. New York: W. H. Freeman.
- Parkin, S. (2013). *CIFFIX*, <https://xray.uky.edu/Resources/scripts/ciffix>

- Parkin, S. & Hope, H. (1998). *J. Appl. Cryst.* **31**, 945–953.
- Prohens, R., Barbas, R., Portell, A., Font-Bardia, M., Alcobé, X. & Puigjaner, C. (2016). *Cryst. Growth Des.* **16**, 1063–1070.
- Schilling, F., Glaser, S., Düwel, S. & Gersch, M. (2016). *pH-Biosensors Based on Compounds Produced From Pyruvic Acid For Magnetic Resonance Imaging and Spectroscopy and Their Uses*. Google Patents.
- Sheldrick, G. M. (2008). *Acta Cryst.* **A64**, 112–122.
- Sheldrick, G. M. (2015a). *Acta Cryst.* **A71**, 3–8.
- Sheldrick, G. M. (2015b). *Acta Cryst.* **C71**, 3–8.
- Wishart, D. S., Feunang, Y. D., Marcu, A., Guo, A. C., Liang, K., Vázquez-Fresno, R., Sajed, T., Johnson, D., Li, C., Karu, N., Sayeeda, Z., Lo, E., Assempour, N., Berjanskii, M., Singhal, S., Arndt, D., Liang, Y., Badran, H., Grant, J., Serra-Cayuela, A., Liu, Y., Mandal, R., Neveu, V., Pon, A., Knox, C., Wilson, M., Manach, C. & Scalbert, A. (2018). *Nucleic Acids Res.* **46**, D608–D617.
- Wishart, D. S., Jewison, T., Guo, A. C., Wilson, M., Knox, C., Liu, Y., Djoumbou, Y., Mandal, R., Aziat, F., Dong, E., Bouatra, S., Sinelnikov, I., Arndt, D., Xia, J., Liu, P., Yallou, F., Bjorndahl, T., Perez-Pineiro, R., Eisner, R., Allen, F., Neveu, V., Greiner, R. & Scalbert, A. (2013). *Nucleic Acids Res.* **41**, D801–D807.
- Wishart, D. S., Knox, C., Guo, A. C., Eisner, R., Young, N., Gautam, B., Hau, D. D., Psychogios, N., Dong, E., Bouatra, S., Mandal, R., Sinelnikov, I., Xia, J., Jia, L., Cruz, J. A., Lim, E., Sobsey, C. A., Shrivastava, S., Huang, P., Liu, P., Fang, L., Peng, J., Fradette, R., Cheng, D., Tzur, D., Clements, M., Lewis, A., De Souza, A., Zuniga, A., Dawe, M., Xiong, Y., Clive, D., Greiner, R., Nazzyrova, A., Shaykhtudinov, R., Li, L., Vogel, H. J. & Forsythe, I. (2009). *Nucleic Acids Res.* **37**, D603–D610.
- Wishart, D. S., Tzur, D., Knox, C., Eisner, R., Guo, A. C., Young, N., Cheng, D., Jewell, K., Arndt, D., Sawhney, S., Fung, C., Nikolai, L., Lewis, M., Coutouly, M.-A., Forsythe, I., Tang, P., Shrivastava, S., Jeroncic, K., Stothard, P., Amegbey, G., Block, D., Hau, D. D., Wagner, J., Miniaci, J., Clements, M., Gebremedhin, M., Guo, N., Zhang, Y., Duggan, G. E., Macinnis, G. D., Weljie, A. M., Dowlatabadi, R., Bamforth, F., Clive, D., Greiner, R., Li, L., Marrie, T., Sykes, B. D., Vogel, H. J. & Querengesser, L. (2007). *Nucleic Acids Res.* **35**, D521–D526.
- Yannai, S. (2004). *Dictionary of food compounds with CD-ROM: Additives, flavors, and ingredients*. Boca Raton: Chapman and Hall/CRC.
- Zhou, R. & Guzman, M. I. (2016). *J. Phys. Chem. C*, **120**, 7349–7357.

supporting information

Acta Cryst. (2019). E75, 858-862 [https://doi.org/10.1107/S2056989019007072]

Crystal structure of zymonic acid and a redetermination of its precursor, pyruvic acid

Dominik Heger, Alexis J. Eugene, Sean R. Parkin and Marcelo I. Guzman

Computing details

For both structures, data collection: *APEX3* (Bruker, 2016); cell refinement: *APEX3* (Bruker, 2016); data reduction: *APEX3* (Bruker, 2016); program(s) used to solve structure: *SHELXT* (Sheldrick, 2015a). Program(s) used to refine structure: *SHELXL2018/3* (Sheldrick, 2015b) for zymonic; *SHELXL2018* (Sheldrick, 2015b) for pyruvic. For both structures, molecular graphics: *XP in SHELXTL* (Sheldrick, 2008); software used to prepare material for publication: *SHELXL* (Sheldrick, 2008) and *CIFFIX* (Parkin, 2013).

4-Hydroxy-2-methyl-5-oxo-2,5-dihydrofuran-2-carboxylic acid (zymonic)

Crystal data

$C_6H_6O_5$	$F(000) = 656$
$M_r = 158.11$	$D_x = 1.523 \text{ Mg m}^{-3}$
Monoclinic, <i>C2/c</i>	Mo $K\alpha$ radiation, $\lambda = 0.71073 \text{ \AA}$
$a = 24.145 (3) \text{ \AA}$	Cell parameters from 9925 reflections
$b = 6.6523 (7) \text{ \AA}$	$\theta = 3.2\text{--}27.5^\circ$
$c = 8.6201 (7) \text{ \AA}$	$\mu = 0.14 \text{ mm}^{-1}$
$\beta = 95.169 (4)^\circ$	$T = 90 \text{ K}$
$V = 1378.9 (3) \text{ \AA}^3$	Thin plate, colourless
$Z = 8$	$0.30 \times 0.25 \times 0.02 \text{ mm}$

Data collection

Bruker D8 Venture dual source diffractometer	18595 measured reflections
Radiation source: microsource	1586 independent reflections
Detector resolution: $5.6 \text{ pixels mm}^{-1}$	1392 reflections with $I > 2\sigma(I)$
φ and ω scans	$R_{\text{int}} = 0.062$
Absorption correction: multi-scan (<i>SADABS</i> ; Krause <i>et al.</i> , 2015)	$\theta_{\text{max}} = 27.5^\circ$, $\theta_{\text{min}} = 3.2^\circ$
$T_{\text{min}} = 0.721$, $T_{\text{max}} = 0.959$	$h = -31 \rightarrow 31$
	$k = -8 \rightarrow 8$
	$l = -11 \rightarrow 10$

Refinement

Refinement on F^2	Primary atom site location: structure-invariant direct methods
Least-squares matrix: full	Secondary atom site location: difference Fourier map
$R[F^2 > 2\sigma(F^2)] = 0.037$	Hydrogen site location: mixed
$wR(F^2) = 0.100$	H atoms treated by a mixture of independent and constrained refinement
$S = 1.09$	$w = 1/[\sigma^2(F_o^2) + (0.0428P)^2 + 1.4377P]$
1586 reflections	where $P = (F_o^2 + 2F_c^2)/3$
104 parameters	
0 restraints	

$$(\Delta/\sigma)_{\max} = 0.001$$

$$\Delta\rho_{\max} = 0.36 \text{ e } \text{\AA}^{-3}$$

$$\Delta\rho_{\min} = -0.24 \text{ e } \text{\AA}^{-3}$$

Extinction correction: SHELXL2018
(Sheldrick, 2015a),
 $Fc^* = kFc[1 + 0.001x\lambda^3/\sin(2\theta)]^{-1/4}$
Extinction coefficient: 0.0057 (13)

Special details

Experimental. The crystal was mounted using polyisobutene oil on the tip of a fine glass fibre, which was fastened in a copper mounting pin with electrical solder. It was placed directly into the cold gas stream of a liquid-nitrogen based cryostat (Hope, 1994; Parkin & Hope, 1998).

Diffraction data were collected with the crystal at 90K, which is standard practice in this laboratory for the majority of flash-cooled crystals.

Geometry. All esds (except the esd in the dihedral angle between two l.s. planes) are estimated using the full covariance matrix. The cell esds are taken into account individually in the estimation of esds in distances, angles and torsion angles; correlations between esds in cell parameters are only used when they are defined by crystal symmetry. An approximate (isotropic) treatment of cell esds is used for estimating esds involving l.s. planes.

Refinement. Refinement progress was checked using *Platon* (Spek, 2009) and by an *R*-tensor (Parkin, 2000). The final model was further checked with the IUCr utility *checkCIF*.

Fractional atomic coordinates and isotropic or equivalent isotropic displacement parameters (\AA^2)

	<i>x</i>	<i>y</i>	<i>z</i>	$U_{\text{iso}}^*/U_{\text{eq}}$	Occ. (<1)
C1	0.67036 (5)	0.40914 (19)	0.56905 (14)	0.0170 (3)	
O1	0.68738 (4)	0.24154 (14)	0.54864 (11)	0.0209 (2)	
C2	0.69378 (5)	0.5659 (2)	0.67706 (14)	0.0178 (3)	
O2	0.74026 (4)	0.51656 (15)	0.76588 (12)	0.0240 (3)	
H2	0.752557	0.618162	0.815574	0.036*	
C3	0.66149 (5)	0.7275 (2)	0.66335 (15)	0.0182 (3)	
H3	0.667502	0.849554	0.719574	0.022*	
O3	0.62412 (4)	0.48283 (13)	0.48901 (11)	0.0188 (2)	
C4	0.61371 (5)	0.68442 (19)	0.54353 (15)	0.0186 (3)	
O4	0.54061 (4)	0.51162 (15)	0.66013 (12)	0.0249 (3)	
H4O	0.507244	0.517998	0.736346	0.109 (14)*	0.5
O5	0.54177 (4)	0.84824 (16)	0.66262 (14)	0.0303 (3)	
H5O	0.509866	0.849228	0.727610	0.109 (14)*	0.5
C5	0.56056 (5)	0.67919 (19)	0.62725 (15)	0.0188 (3)	
C6	0.60945 (7)	0.8285 (2)	0.40645 (17)	0.0257 (3)	
H6A	0.644218	0.825746	0.355945	0.039*	
H6B	0.602833	0.964985	0.443442	0.039*	
H6C	0.578557	0.787894	0.331471	0.039*	

Atomic displacement parameters (\AA^2)

	U^{11}	U^{22}	U^{33}	U^{12}	U^{13}	U^{23}
C1	0.0184 (6)	0.0180 (6)	0.0151 (6)	−0.0007 (5)	0.0038 (5)	0.0010 (5)
O1	0.0238 (5)	0.0173 (5)	0.0213 (5)	0.0015 (4)	0.0010 (4)	−0.0013 (4)
C2	0.0165 (6)	0.0204 (6)	0.0165 (6)	−0.0023 (5)	0.0016 (5)	−0.0020 (5)
O2	0.0186 (5)	0.0249 (5)	0.0274 (5)	0.0035 (4)	−0.0050 (4)	−0.0073 (4)
C3	0.0168 (6)	0.0190 (6)	0.0190 (6)	−0.0032 (5)	0.0020 (5)	−0.0036 (5)
O3	0.0213 (5)	0.0155 (5)	0.0192 (5)	0.0003 (3)	−0.0013 (4)	−0.0027 (3)
C4	0.0205 (6)	0.0135 (6)	0.0212 (6)	−0.0007 (5)	−0.0012 (5)	−0.0018 (5)

O4	0.0225 (5)	0.0207 (5)	0.0314 (6)	-0.0031 (4)	0.0027 (4)	0.0013 (4)
O5	0.0252 (5)	0.0209 (5)	0.0457 (7)	0.0027 (4)	0.0074 (5)	-0.0040 (4)
C5	0.0165 (6)	0.0171 (6)	0.0217 (6)	0.0004 (5)	-0.0042 (5)	-0.0001 (5)
C6	0.0351 (8)	0.0192 (7)	0.0223 (7)	-0.0013 (6)	-0.0008 (6)	0.0026 (5)

Geometric parameters (Å, °)

C1—O1	1.2067 (16)	C4—C6	1.5179 (19)
C1—O3	1.3505 (15)	C4—C5	1.5282 (19)
C1—C2	1.4763 (18)	O4—C5	1.2568 (16)
C2—C3	1.3268 (18)	O4—H4O	1.0854
C2—O2	1.3411 (16)	O5—C5	1.2602 (16)
O2—H2	0.8400	O5—H5O	0.9926
C3—C4	1.5051 (17)	C6—H6A	0.9800
C3—H3	0.9500	C6—H6B	0.9800
O3—C4	1.4505 (15)	C6—H6C	0.9800
O1—C1—O3	122.52 (12)	O3—C4—C5	108.03 (10)
O1—C1—C2	128.95 (12)	C3—C4—C5	107.73 (11)
O3—C1—C2	108.53 (11)	C6—C4—C5	112.41 (11)
C3—C2—O2	134.75 (12)	C5—O4—H4O	115.0
C3—C2—C1	109.12 (11)	C5—O5—H5O	117.2
O2—C2—C1	116.12 (11)	O4—C5—O5	125.69 (13)
C2—O2—H2	109.5	O4—C5—C4	118.80 (11)
C2—C3—C4	108.35 (11)	O5—C5—C4	115.45 (11)
C2—C3—H3	125.8	C4—C6—H6A	109.5
C4—C3—H3	125.8	C4—C6—H6B	109.5
C1—O3—C4	109.24 (10)	H6A—C6—H6B	109.5
O3—C4—C3	104.7 (1)	C4—C6—H6C	109.5
O3—C4—C6	109.47 (11)	H6A—C6—H6C	109.5
C3—C4—C6	114.07 (11)	H6B—C6—H6C	109.5
O1—C1—C2—C3	-179.25 (13)	C1—O3—C4—C5	-112.19 (11)
O3—C1—C2—C3	1.17 (15)	C2—C3—C4—O3	-1.69 (14)
O1—C1—C2—O2	-0.3 (2)	C2—C3—C4—C6	-121.33 (13)
O3—C1—C2—O2	-179.92 (10)	C2—C3—C4—C5	113.14 (12)
O2—C2—C3—C4	-178.23 (14)	O3—C4—C5—O4	12.04 (16)
C1—C2—C3—C4	0.39 (15)	C3—C4—C5—O4	-100.56 (13)
O1—C1—O3—C4	178.13 (12)	C6—C4—C5—O4	132.92 (13)
C2—C1—O3—C4	-2.26 (13)	O3—C4—C5—O5	-170.61 (11)
C1—O3—C4—C3	2.43 (13)	C3—C4—C5—O5	76.79 (14)
C1—O3—C4—C6	125.11 (12)	C6—C4—C5—O5	-49.72 (16)

Hydrogen-bond geometry (Å, °)

<i>D</i> —H \cdots <i>A</i>	<i>D</i> —H	H \cdots <i>A</i>	<i>D</i> \cdots <i>A</i>	<i>D</i> —H \cdots <i>A</i>
O2—H2 \cdots O1 ⁱ	0.84	1.96	2.7103 (14)	148
C3—H3 \cdots O2 ⁱ	0.95	2.48	3.0720 (16)	120

O4—H4O...O4 ⁱⁱ	1.09	1.52	2.607 (2)	176
O5—H5O...O5 ⁱⁱ	0.99	1.63	2.624 (2)	179

Symmetry codes: (i) $-x+3/2, y+1/2, -z+3/2$; (ii) $-x+1, y, -z+3/2$.

2-Oxopropanoic acid (pyruvic)

Crystal data

$C_3H_4O_3$	$F(000) = 184$
$M_r = 88.06$	$D_x = 1.554 \text{ Mg m}^{-3}$
Monoclinic, $P2_1/c$	Mo $K\alpha$ radiation, $\lambda = 0.71073 \text{ \AA}$
$a = 10.7486 (3) \text{ \AA}$	Cell parameters from 6955 reflections
$b = 5.1925 (2) \text{ \AA}$	$\theta = 3.8\text{--}34.3^\circ$
$c = 6.8302 (2) \text{ \AA}$	$\mu = 0.14 \text{ mm}^{-1}$
$\beta = 99.063 (1)^\circ$	$T = 90 \text{ K}$
$V = 376.45 (2) \text{ \AA}^3$	Well-faceted block, colourless
$Z = 4$	$0.26 \times 0.22 \times 0.18 \text{ mm}$

Data collection

Bruker D8 Venture dual source diffractometer	10479 measured reflections
Radiation source: microsource	1425 independent reflections
Detector resolution: $5.6 \text{ pixels mm}^{-1}$	1242 reflections with $I > 2\sigma(I)$
φ and ω scans	$R_{\text{int}} = 0.025$
Absorption correction: multi-scan (<i>SADABS</i> ; Krause <i>et al.</i> , 2015)	$\theta_{\text{max}} = 34.5^\circ, \theta_{\text{min}} = 3.8^\circ$
$T_{\text{min}} = 0.890, T_{\text{max}} = 0.971$	$h = -16 \rightarrow 16$
	$k = -7 \rightarrow 8$
	$l = -10 \rightarrow 10$

Refinement

Refinement on F^2	Secondary atom site location: difference Fourier map
Least-squares matrix: full	Hydrogen site location: mixed
$R[F^2 > 2\sigma(F^2)] = 0.031$	H atoms treated by a mixture of independent and constrained refinement
$wR(F^2) = 0.082$	$w = 1/[\sigma^2(F_o^2) + (0.0299P)^2 + 0.1264P]$
$S = 1.08$	where $P = (F_o^2 + 2F_c^2)/3$
1425 reflections	$(\Delta/\sigma)_{\text{max}} < 0.001$
60 parameters	$\Delta\rho_{\text{max}} = 0.40 \text{ e \AA}^{-3}$
0 restraints	$\Delta\rho_{\text{min}} = -0.21 \text{ e \AA}^{-3}$
Primary atom site location: structure-invariant direct methods	

Special details

Experimental. The crystal was mounted using polyisobutene oil on the tip of a fine glass fibre, which was fastened in a copper mounting pin with electrical solder. It was placed directly into the cold gas stream of a liquid-nitrogen based cryostat (Parkin & Hope, 1998).

Diffraction data were collected with the crystal at 90K, which is standard practice in this laboratory for the majority of flash-cooled crystals.

Geometry. All esds (except the esd in the dihedral angle between two l.s. planes) are estimated using the full covariance matrix. The cell esds are taken into account individually in the estimation of esds in distances, angles and torsion angles; correlations between esds in cell parameters are only used when they are defined by crystal symmetry. An approximate (isotropic) treatment of cell esds is used for estimating esds involving l.s. planes.

Refinement. Refinement progress was checked using *Platon* (Spek, 2009) and by an *R*-tensor (Parkin, 2000). The final model was further checked with the IUCr utility *checkCIF*.

Fractional atomic coordinates and isotropic or equivalent isotropic displacement parameters (\AA^2)

	<i>x</i>	<i>y</i>	<i>z</i>	$U_{\text{iso}}^*/U_{\text{eq}}$
O1	0.41673 (5)	0.30370 (12)	0.30191 (9)	0.01968 (14)
H1	0.4903 (13)	0.320 (3)	0.3883 (19)	0.030*
C1	0.33963 (6)	0.48448 (14)	0.34005 (11)	0.01316 (14)
O2	0.36317 (5)	0.65673 (11)	0.46175 (8)	0.01589 (13)
C2	0.20853 (6)	0.46732 (14)	0.21028 (10)	0.01271 (14)
C3	0.11464 (7)	0.65949 (15)	0.25725 (11)	0.01520 (15)
H3A	0.1499 (3)	0.8314 (12)	0.255 (1)	0.023*
H3B	0.0940 (5)	0.6245 (9)	0.388 (1)	0.023*
H3C	0.0389 (6)	0.6478 (9)	0.1594 (9)	0.023*
O3	0.18824 (5)	0.30508 (11)	0.08250 (9)	0.01774 (14)

Atomic displacement parameters (\AA^2)

	U^{11}	U^{22}	U^{33}	U^{12}	U^{13}	U^{23}
O1	0.0132 (2)	0.0218 (3)	0.0222 (3)	0.0041 (2)	-0.0027 (2)	-0.0084 (2)
C1	0.0121 (3)	0.0138 (3)	0.0136 (3)	-0.0002 (2)	0.0018 (2)	0.0003 (2)
O2	0.0135 (2)	0.0155 (3)	0.0176 (3)	-0.00038 (19)	-0.00071 (19)	-0.0034 (2)
C2	0.0119 (3)	0.0139 (3)	0.0121 (3)	-0.0011 (2)	0.0010 (2)	0.0009 (2)
C3	0.0140 (3)	0.0163 (3)	0.0148 (3)	0.0023 (2)	0.0008 (2)	-0.0013 (3)
O3	0.0167 (3)	0.0181 (3)	0.0173 (3)	-0.0002 (2)	-0.0008 (2)	-0.0049 (2)

Geometric parameters (\AA , $^\circ$)

O1—C1	1.3053 (9)	C2—C3	1.4896 (10)
O1—H1	0.913 (14)	C3—H3A	0.971 (6)
C1—O2	1.2201 (9)	C3—H3B	0.971 (6)
C1—C2	1.5446 (10)	C3—H3C	0.971 (6)
C2—O3	1.2079 (9)		
C1—O1—H1	108.4 (8)	C2—C3—H3A	109.5
O2—C1—O1	126.37 (7)	C2—C3—H3B	109.5
O2—C1—C2	120.38 (6)	H3A—C3—H3B	109.5
O1—C1—C2	113.24 (6)	C2—C3—H3C	109.5
O3—C2—C3	124.85 (7)	H3A—C3—H3C	109.5
O3—C2—C1	119.96 (7)	H3B—C3—H3C	109.5
C3—C2—C1	115.19 (6)		
O2—C1—C2—O3	175.81 (7)	O2—C1—C2—C3	-4.59 (10)
O1—C1—C2—O3	-3.34 (10)	O1—C1—C2—C3	176.26 (6)

Hydrogen-bond geometry (\AA , $^\circ$)

<i>D</i> —H \cdots <i>A</i>	<i>D</i> —H	H \cdots <i>A</i>	<i>D</i> \cdots <i>A</i>	<i>D</i> —H \cdots <i>A</i>
O1—H1 \cdots O2 ⁱ	0.913 (14)	1.742 (14)	2.6536 (8)	175.5 (12)

Symmetry code: (i) $-x+1, -y+1, -z+1$.