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Aluminum bioavailability from basic sodium aluminum phosphate, an approved food additive emulsifying agent, incorporated in cheese

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Abstract

Oral aluminum (Al) bioavailability from drinking water has been previously estimated, but there is little information on Al bioavailability from foods. It was suggested that oral Al bioavailability from drinking water is much greater than from foods. The objective was to further test this hypothesis. Oral Al bioavailability was determined in the rat from basic [²⁶Al]-sodium aluminum phosphate (basic SALP) in a process cheese. Consumption of ~ 1 gm cheese containing 1.5 or 3% basic SALP resulted in oral Al bioavailability (F) of ~ 0.1 and 0.3%, respectively, and time to maximum serum ²⁶Al concentration (T_{max}) of 8 to 9 h. These Al bioavailability results were intermediate to previously reported results from drinking water (F ~ 0.3%) and acidic-SALP incorporated into a biscuit (F ~ 0.1%), using the same methods. Considering the similar oral bioavailability of Al from food vs. water, and their contribution to the typical human's daily Al intake (~ 95 and 1.5%, respectively), these results suggest food contributes much more Al to systemic circulation, and potential Al body burden, than does drinking water. These results do not support the hypothesis that drinking water provides a disproportionate contribution to total Al absorbed from the gastrointestinal tract.

Keywords

Accelerator mass spectrometry; aluminum; atomic absorption spectrometry; basic sodium aluminum phosphate; food additive; oral bioavailability

Introduction

Aluminum (Al) is a toxicant to the central nervous, skeletal and hematopoietic systems (Krewski et al., 2007). The primary source of oral Al exposure in the U.S. for the typical human is foods, representing ~ 95% of daily oral intake; drinking water contributes ~ 1 to 2% (WHO,

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1997; ATSDR, 1999; Krewski et al., 2007). These typically provide a total of ~ 4000 to 9,000 µg Al/day. Consumption of antacids can provide up to 5,000,000, inhalation from environmental and industrial air 4 to 20 and up to 25,000, and antiperspirant use up to ~ 70,000 μ g/day. Injected Al in vaccines and allergy immunotherapy can provide 1 to 8 and 7 to 40 μ g/ day, respectively, and probably complete absorption (Yokel and McNamara, 2001). Some people may have further Al exposures, e.g., occupational (Elinder et al., 1991), Alcontaminated dialysis fluid (Alfrey et al., 1976), alum instillation into a hemorrhaging urinary bladder (Phelps et al., 1999) and illicit drug abuse, including intravenous self-injection of methadone prepared in Al cookware (Friesen et al., 2006; Yong et al, 2006; Exley et al, 2007). Although tobacco contains considerable Al, ~ 500 to 2000 μ g/cigarette, < 0.02 to ~ 0.075 µg appeared in the smoke from one cigarette (Cogbill and Hobbs, 1957; Exley et al., 2006). Even if 100% bioavailable, when compared to oral Al intake of 5000 μ g/day from diet that is 0.25% bioavailable delivering 12.5 µg to systemic circulation, a cigarette would deliver only ~ 0.4% as much. It was suggested that drinking water provides a disproportionate contribution to total Al absorbed from the gastrointestinal tract because it is largely uncomplexed in water, whereas organic ligands such as phytates and polyphenols in food were suggested to complex Al and inhibit its oral absorption (Martyn et al., 1989). This notion has been followed by ≥ 12 epidemiological studies since 1989 assessing the association between Al in drinking water and cognitive impairment, dementia and Alzheimer's disease (AD) (Krewski et al., 2007). Many showed a significantly higher odds ratio for dementia among study subjects living in areas where the drinking water had a higher Al concentration. However, there is no evidence that drinking water is the major source of systemic Al for the typical person. The study in this report was aimed at testing this notion.

If Al bioavailability from water is not considerably greater than from food, studies assessing a putative link between Al in drinking water and AD might be misfocused, and studies investigating a possible association between Al in food and cognitive impairment, dementia and AD might be more relevant. There is only one published study assessing a putative link between dietary Al intake in food and AD (Rogers and Simon, 1999). In 23 matched sets of residents of a geriatric center with and without AD, the adjusted odds ratio for several food groups, including baked goods and cheese, was elevated. However, due to the very small sample size, most results did not reach statistical significance.

Adult daily dietary Al intake in the U.K. and U.S. has been estimated to be 3.9 and 7 to 9 mg (depending on age and sex), respectively (UK-MAFF, 1993; Pennington and Schoen, 1995). There have been at least 12 other estimates of adult dietary Al intake published in the past 10 years from other countries. Most reported equal or less daily Al intake than in the U.S. The lower daily Al intake in some countries, such as in Europe, has been attributed to less use of Al as a food additive (Müller et al., 1998).

Different food sources contribute variable amounts of Al to the human diet (Pennington, 1987; UK-MAFF, 1993; Lopez et al., 2002). Sodium aluminum phosphates (SALPs) are generally recognized as safe (GRAS) FDA-approved food additives that contribute the greatest amount of Al to the diet (Katz et al., 1984; Humphreys and Bolger, 1997; Saiyed and Yokel, 2005). Basic SALP is one of many "emulsifying salts" added to process cheese, cheese food and cheese spread which react with and change the protein of cheese to produce a smooth, uniform film around each fat droplet to prevent separation and bleeding of fat from the cheese. This produces a soft texture, easy melting characteristics and desirable slicing properties (Ellinger, 1972). Basic SALP is permitted by the FDA up to 3% in pasteurized process cheese (21CFR133.169), pasteurized process cheese food (21CFR133.173), and pasteurized process cheese spread (21CFR133.179). The FDA regulates basic SALP as an emulsifier that is GRAS (FASEB, 1975). Basic SALP is also permitted up to 3.5% in cheese in Canada (Food and Drugs Act – Food and Drug Regulations, Part B, Division 16;

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http://www.hc-sc.gc.ca/fn-an/alt_formats/hpfb-dgpsa/pdf/legislation/e_c-tables.pdf), as potassium aluminum silicate up to 10 g/kg in Australia and New Zealand

http://www.foodstandards.gov.au/_srcfiles/FSC_Amend_Standard_1_2_4_Labelling_of_Ingred_v91.pdf#search=%), but evidently not by the E.U. or U.K., although other Al-containing compounds are permitted in cheese, up to 10 g/kg (E 554, E 555, E 556 and E 559) (http://ec.europa.eu/food/fs/sfp/addit_flavor/flav11_en.pdf http://ec.europa.eu/food/fs/sc/scf/reports/scf_reports_40.pdf). Studies of the Al content of unprocessed cheese generally reported < 10 mg Al/kg (Delves et al., 1989; Schenk et al., 1989; Müller et al., 1998; Dolan and Capar, 2002). In contrast, some process cheeses (often marketed as sliced, American) were reported to contain 695 (Gormican, 1970), 411 (Pennington, 1989), 320 (Pennington and Schoen, 1995), 1440 (Greger, 1985), and 470 mg Al/kg (Saiyed and Yokel, 2005). The cheese of frozen pizzas that contained basic SALP was reported to contain up to ~ 750 mg Al/kg, whereas ready-to-eat restaurant pizza cheese had < 10 mg Al/kg (Saiyed and Yokel, 2005). Basic SALP (Na₈Al₂(OH₂)((PO)₄)₄) has a 4:1:2 ratio of Na:Al:P. It is ~ 10% Al.

Concern about the safety of the addition of Al in foods was expressed long ago (Gies, 1911) and again recently. Department of the Planet Earth (a non-profit NGO) petitioned the FDA in 2005 to rescind the GRAS status for Al-based food additives (http://www.deptplanetearth.com/aboutus.html). A Joint FAO (Food and Agriculture Organization of the UN)/WHO Expert Committee on Food Additives established a revised provisional tolerable weekly intake (PTWI) for all Al compounds in foods, including additives, of 1 mg/kg/bw. The Expert Committee noted the potential for Al to effect the reproductive and developing nervous system at lower doses than those used to establish the previous PTWI of 7 mg/kg/bw (FAO/WHO, 2006). The Committee commented on the "probable lower

bioavailability of the less soluble Al compounds present in food" and that the PTWI was likely to be exceeded by some population groups, particularly children, who regularly consume foods containing Al additives.

To better understand the relative contribution of food and water to human Al intake, it has been noted that research needs to be conducted to more accurately determine the oral bioavailability of Al from the typical diet (Personal communication from Amal M. Mahfouz, US EPA). Oral bioavailability (fractional absorption) is the amount absorbed compared to the amount administered. For Al, systemic bioavailability, the fraction that reaches systemic circulation from which it has access to the target organs of its toxicity, is most relevant.

Several studies have estimated oral Al bioavailability under conditions that model Al consumption in drinking water. Oral Al bioavailability from alum-treated water obtained from a municipal water treatment facility was estimated to be 0.36% in a study of 21 humans (Stauber et al., 1999). Two studies in humans that utilized ²⁶Al but only two subjects each, therefore providing limited confidence in the results, estimated oral Al bioavailability to be 0.1 and 0.22%, respectively (Hohl et al., 1994; Priest et al., 1998). The above studies estimated oral Al absorption based on urinary Al excretion. However, not all absorbed Al appears in the urine, which accounts for $\sim 98\%$ of excreted Al, or the bile, which accounts for most of the other 2%. Some absorbed Al is retained. Secondly, if Al in the urine is used as an endpoint for absorption, the experimenter would have to continue to determine this endpoint until sure that all had been eliminated. There are a few reports where the amount of Al in the urine was corrected for these 2 factors (not all absorbed Al appears in the urine and the study was not conducted long enough to collect all excreted Al). Stauber et al. (1999) multiplied the amount of Al that appeared in the urine in the 24 hours after oral Al consumption by 2.2, to correct for partial Al elimination (only 72% of intravenously injected Al was found in the urine after 7 days (Talbot et al., 1995) and only 62 to 63% of the Al in the urine recovered in 7 days was found in the first day

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(Priest, 1993)). Stauber et al only collected urine for 24 hours. They did not correct for absorbed Al that was retained. The large correction factor reduces confidence in the accuracy of the reported oral bioavailability. Hohl et al. (1994) took into consideration that only about 80% of injected Al was found in the urine after 10 days and commented that the amount of Al excreted in the urine after their 4 days of collection was probably not great. They provided an estimate of the percentage of Al absorbed as 0.1%, not presenting this as a precise value. In two studies that used the methods employed in the present report oral Al bioavailability in the rat averaged 0.28% and 0.29% (Yokel et al., 2001; Zhou and Yokel, 2006).

The bioavailability of hydrophilic substances that are not well absorbed can be determined by comparing the areas under the concentration \times time curve (AUC) for the test substance given po and iv. This approach was used in the only study reported to date that estimated oral Al bioavailability from a specific food product. In that study, oral Al bioavailability in rats that ate ~ 1 gm of biscuit containing [26 Al]-labeled acidic SALP averaged ~ 0.12% (Yokel and Florence, 2006). This approach was used in the present study in which the po dose was given as ²⁶Al and the iv dose as ²⁷Al to the same subject at the same time. Compared to an experimental design where a rat receives the two doses at different times, this method reduces variability by concurrently determining the AUCs from the po and iv doses in the rat. To determine the oral bioavailability of Al from basic SALP in a process cheese, [²⁶Al]-containing basic SALP was prepared and incorporated into a process cheese that was presented to rats for their consumption. Blood was repeatedly obtained from the rats in which ²⁶Al and total Al (essentially ²⁷Al) were analyzed by accelerator mass spectrometry (AMS) and electrothermal atomic absorption spectrometry (ETAAS), respectively. The absence of significant ²⁶Al in the environment or in normal biological organisms avoids the interference of endogenous Al in studies of Al pharmacokinetics. When 4 mg of ²⁷Al is added to ²⁶Al-containing samples to establish the $^{\bar{27}}$ Al concentration, as in the present study, AMS can measure the 26 Al: 27 Al ratio with a detection limit of ~ 10^{-14} (~ 4×10^{-17} g of ²⁶Al) or ~ 1,000,000 atoms. This enables the conduct of pharmacokinetic studies of Al at physiological concentrations, as conducted herein. In addition to oral bioavailability, two other standard pharmacokinetic parameters, the time to maximum serum ²⁶Al concentration of (T_{max}) and the maximum serum ²⁶Al concentration of (C_{max}), were determined (Bauer, 2006). These methods enabled us to address the objectives of the present study: to determine the oral bioavailability of Al from processed cheese that contains basic SALP, at a concentration relevant to its use in cheese, and to determine standard pharmacokinetic parameters of the absorbed Al.

Methods

Materials

Details of the preparation and characterization of the basic SALP containing ²⁶Al (basic ²⁶Al-SALP) were described (Yokel et al., 2005). ²⁶Al was provided by the Purdue Rare Isotope Measurement Laboratory (PRIME Lab), supplied in 0.06 N HCl containing 13.78 nCi (725 ng) ²⁶Al/ml and with a 34:1 ²⁷Al:²⁶Al ratio. Briefly, Al-containing sodium aluminate was prepared then characterized by elemental analysis of Na and Al and Raman spectroscopy. It was incorporated into the synthesis of basic SALP (CAS [7785-88-8], INS No. 541) by the method of (Bell, 1973). The basic SALP was characterized by near infrared spectroscopy, x-ray powder diffraction spectroscopy, and elemental analysis of Na, Al and P. ²⁶Al-containing basic SALP was concurrently and identically prepared. Non-²⁶Al-containing basic SALP was used to dilute ²⁶Al-containing samples to ~ 1 nCi ²⁶Al/15 or 30 mg basic SALP. The ²⁶Albasic SALP was incorporated into a process cheese. These two concentrations (1.5 and 3% basic SALP) were studied to ascertain if there was concentration-dependent absorption and to stay within the maximum permitted use level of 3 to 3.5% for basic SALP as an emulsifying agent in process cheese. Cheese containing ²⁶Al-basic SALP was prepared from 8.8 gm aged

(1 year) Kroger brand sharp cheddar cheese, ~ 1 gm water and 150 or 300 mg basic SALP (to make 1.5 and 3% SALP in cheese). To produce a moderately firm cheese product the ingredients were heated in a microwave oven for 13 sec, stirred for 15 sec, heated for another 5 sec, stirred 10 sec and some samples further heated and stirred. Identical process cheeses were prepared containing 1.5, 2.25 and 3% SALP without ²⁶Al. They were used to condition the rats to eat the process cheese and to dose the control (non-²⁶Al-treated) rats, as described below.

The ²⁶Al concentration in the process cheese was determined as follows. Three samples, ~ 30 to 35 mg, of the 1.5 and 3% ²⁶Al-basic SALP were transferred to 7 ml screw cap Teflon® Tuftainers® and 4 mg Al (Sigma ICP/DCP standard) added, and the sample dried. Samples were then digested in 1.5 ml 1M KOH and sonicated to create a solution, as described (Saiyed and Yokel, 2005), serially diluted 10-fold 4 times, and an aliquot transferred to a porcelain crucible and 4 mg of Al standard added. This was slowly heated to 1000 °C in a muffle furnace to convert the Al to Al oxide that was analyzed by AMS, as described below. The process cheeses containing 1.5 and 3% basic ²⁶Al-SALP had 1.80 nCi (95.0 ng) and 1.57 nCi (82.6 ng) ²⁶Al/gm, respectively.

Animals

This study used 15 male Fisher 344 rats, weighing $272 \pm 11g$ (mean \pm SD) that were housed individually prior to the study in the University of Kentucky Division of Laboratory Animal Resources facility. Animal work was approved by the University of Kentucky Institutional Animal Care and Use Committee. The research was conducted in accordance with the Guiding Principles in the Use of Animals in Toxicology.

Experimental procedures

The rats were acclimated to a 10% protein diet designed to minimize food retention in the stomach (Harlan Teklad 95215). They had food access from 08:00 to 18:00 h daily for at least 5 days prior to the determination of oral Al bioavailability. This diet was shown to result in the absence of food in the stomach 14 h after its withdrawal when fecal recycling (coprophagia) was prevented by a fecal collection cup, as described and used in this study (Yokel et al., 2001). Drinking water was freely available throughout the study except for the period from 14 h before to 4 h after presentation of cheese containing basic SALP.

The rats were conditioned to eat 1 g of process cheese containing 1.5% and 3% basic SALP (containing no ²⁶Al) on alternate days which was presented, with access to 2 ml of MilliQ polished water, 14 h after diet removal daily for at least 4 days. Rats that readily ate the cheese within 15 min were surgically prepared 1 day prior to oral dosing with two venous cannulae introduced into the femoral veins. They terminated in the vena cava. One cannula was used for iv administration, the other for blood withdrawal. The former terminated down-stream of the latter. The amount of cheese containing ²⁶Al-basic SALP that was consumed was determined by pre-weighing the presented cheese then rigorously recovering from the rat's cage 30 min later the uneaten cheese, which was weighed. Calculations were based on the weight of consumed cheese.

The rats were randomly assigned to be given 1 g process cheese containing 1.5 or 3% basic ²⁶Al-SALP (n = 6/group), in the absence of food in the stomach. Three control rats were included to document ²⁶Al contamination. They received process cheese containing 2.25% basic SALP (n=2) or intragastric administration of 1 ml of water (n=1) without ²⁶Al. They served as monitors for ²⁶Al contamination. The rats were studied in pairs that received the same ²⁷Al infusion but different oral treatments.

Oral Al bioavailability was determined in the un-anesthetized rat. Oral bioavailability was determined using a modification of the standard method of comparing the areas under the concentration \times time curve (AUC). To enable concurrent administration of the oral and iv Al dose, the tracer ²⁶Al was used as the oral dose and ²⁷Al for the iv dose. To give a large enough bolus iv dose of ²⁷Al that would be significantly above the normal endogenous Al concentration in serum for a long enough period of time to determine its AUC would require a dose that would exceed the transferrin binding capacity for Al (~ 1350 µg Al/L). Therefore the chemical species of the ²⁷Al in serum would be different (not all Al-transferrin, but for the Al beyond the transferrin binding capacity probably Al citrate) and might be handled differently than the ²⁶Al that is absorbed and could bind to transferrin after enough of the ²⁷Al had been cleared to free up binding sites on transferrin. Therefore, we replaced the iv bolus dose with an iv infusion to generate a ²⁷Al serum concentration significantly above the endogenous Al concentration (so the Al in the serum could be attributed to the ²⁷Al infusion [iv dose]) but below the transferrin binding capacity. In summary, dose_{iv} was replaced by ²⁷Al infusion rate × ²⁷Al infusion duration, as reported in Data analysis.

The rat was iv infused at $100 \ \mu g^{27}$ Al/kg/h to produce an estimated $500 \ \mu g$ Al/l in blood plasma for the ²⁷Al dose, as described (Yokel et al., 2001). To accomplish this in the present study, AlK(SO₄)₂ was infused from 14 h prior to 60 h after oral dosing. Blood was withdrawn 1 h prior to and 0, 1, 2, 4, 8, 24, 36, 48 and 60 h after oral dosing. The withdrawn blood (0.3 ml for the -1 to 4 h samples, 0.5 ml for the 8 h sample, 2.1 ml at 24 h and 4.1 ml at 36 and 48 h), was replaced by an equal volume of injected saline. Additionally, the rats had access to free water and food (10% protein diet) beginning 4 h after dosing. The 60-h blood sample was obtained by anesthetizing the rat and exsanguination from a femoral cannula and then the heart. Serum was obtained for ²⁶Al and ²⁷Al analysis. Blood urea nitrogen (BUN) and creatinine were determined in the 60 h sample.

Analysis of total AI by electrothermal atomic absorption spectrometry (ETAAS)

This was conducted as described (Yokel and Florence, 2006).

Analysis of ²⁶Al by accelerator mass spectrometry (AMS)

The procedures were as described (Yokel et al., 2001). A serum QC sample was included with each of the four batches of processed samples. All results were within 10% of the mean of the 5 initial replicates determined when these QC samples were prepared.

Data analysis

A criterion for acceptance of post-treatment serum ²⁶Al concentrations considered to be reliably above pre-treatment serum values was established as > 2 SD above the mean pre-treatment serum ²⁶Al concentration. This criterion was 0.34 fg ²⁶Al/ml. Values below this criterion are not presented graphically and were not used in the data analysis. This criterion was met by all of the samples from the ²⁶Al-treated rats. The initial and 2nd half-lives of ²⁶Al elimination, determined using RSTRIP (Fox and Lamson, 1989), were not different between the two treatment groups, therefore they were combined. The mean initial and 2nd half-lives were 4.4 and 7.2 h, respectively. Therefore, blood was obtained for > 3 half-lives, sufficient time to determine the AUC of the ²⁶Al. Each subject's pre-treatment serum ²⁶Al concentration was subtracted from its post-treatment values. Oral ²⁶Al bioavailability (F) was calculated as follows and expressed as a percent:

 $F = \frac{\text{The sum of the trapezoidal areas for }^{26}\text{Al}}{\text{The sum of the trapezoidal areas for }^{27}\text{Al}} \times \frac{^{27}\text{Al hourly infusion rate} \times \text{time}}{^{26}\text{Al dose}}$

Time to maximum serum 26 Al concentration (T_{max}) and the maximum concentration (C_{max}) were calculated using RSTRIP (Fox and Lamson, 1989).

Unpaired two-tailed t-tests (Mann Whitney tests when the variance was dissimilar) were used to test for differences in Al oral bioavailability, T_{max} and C_{max} from the 1.5% compared to 3% basic SALP. When not statistically significantly different, the combined basic SALP results were compared to acidic SALP in biscuit and water, which were previously conducted (Yokel and Florence, 2006). A difference of P < 0.05 was accepted as statistically significant.

Results

The rats given cheese containing ²⁶Al-SALP consumed 0.44 to 1.05 (mean = 0.87) gm cheese. Each rat's serum ²⁶Al results were normalized to its ²⁶Al dose. The BUN and serum creatinine values ranged from 7.5 to 17.9 and from < 0.2 to 0.5 mg/dl, well within normal limits (< 30 and 1 mg/dl, respectively). Serum ²⁶Al concentration in all but one rat prior to ²⁶Al (or ²⁶Al vehicle) dosing ranged from 0 to 0.4 (mean = 0.12) fg/ml. Initial samples from one control rat were contaminated. Its pre-treatment and first 4 post-treatment samples ranged from 0.6 to 12.1 fg ²⁶Al/ml, whereas the 6th to 9th samples from this control rat had 0.1 to 0.2 fg ²⁶Al/ml. Results from this animal have been deleted from the data analysis. The concentration of ²⁶Al in the 8 samples from each of the other two non-²⁶Al-treated rats after vehicle dosing ranged from 0.4 to 3.8 fg/ml. Therefore, contamination appears to have been limited to the 5 samples from the one rat.

Peak serum ²⁶Al concentration ranged from 55.9 to 159.5 fg/ml after oral ²⁶Al dosing in 1.5% basic SALP and 28.9 to 146.1 fg/ml after 3% basic SALP. The oral ²⁶Al dose increased peak serum ²⁶Al at least 200-fold above mean pre-treatment values. The time courses of serum ²⁶Al following oral ²⁶Al dosing for the two treatment groups are shown in Figure 1. Oral bioavailability (F) was calculated as described in Data analysis as the (sum of ²⁶Al AUC/ sum ²⁷Al AUC) × the sampling duration × ²⁷Al hour infusion rate × body weight × 100%. For one of the rats that consumed cheese containing 1.5% SALP F was 0.12%, calculated from ((the sum of the trapezoidal AUCs for ²⁶Al [each area calculated as the mean of the ²⁶Al concentration in the two samples, in fg/ml × the trapezoidal time interval, in hr]/sum of trapezoidal AUCs for ²⁷Al [each area calculated as the mean of the ²⁷Al concentration in the two samples, in fg/ml] \times the trapezoidal time interval, in hr) \times ((the total duration of the serum sampling period, in hr \times the ²⁷Al infusion rate, in fg/kg/hr \times the rat's body wt, in kg)/(52630000 \times nCi ²⁶Al/gm cheese \times gm cheese consumed by the rat))) \times 100, where the sum of the ²⁶Al AUCs was 2013 fg/ml/hr, the sum of the ²⁷Al AUCs was 25728 ng/ml/hr, the ²⁷Al infusion rate was 10000000000 fg/kg/hr, the rat's body weight was 0.254 kg, 52630000 converts nCi ²⁶Al to fg ²⁶Al, the cheese had 1.804 nCi ²⁶Al/gm, the rat consumed 0.97 gm cheese, and 100 converts to percent.

The percentage of the AUC included in the AUC_{time to last sample}/AUC_{infinity} was > 95% in all cases, indicating that samples were collected for sufficient time to adequately determine oral Al absorption. Oral Al bioavailability and the results of RSTRIP analyses (T_{max} and C_{max}) are shown in Table 1. Oral bioavailability was marginally significantly different (P = 0.045) between the two groups. Therefore, the results from both treatment groups were compared to results obtained in oral bioavailability studies of Al in water and as acidic SALP in biscuit (Table 1). The differences in sum AUC ²⁶Al, sum AUC ²⁷Al, Al T_{max}, C_{max}, and C_{max} % dose/ml serum results from rats consuming 1.5% compared to 3% basic SALP were not statistically significant. Therefore, the differences in these measures from the combined 1.5% and 3% basic SALP data were compared to results from studies of Al absorption from water and as acidic SALP in biscuit (Table 1).

Discussion

The present study found a marginally significant effect of the Al concentration on Al bioavailability, for the two concentrations of basic SALP tested, which are in the range of those permitted by the FDA. The difference between the oral bioavailability of the two groups that received 1.5% and 3% basic SALP in cheese was due to the differences in their serum total Al concentration × time profiles (as shown in Table 1, Sum AUC for iv ²⁷Al) rather than their serum ²⁶Al concentration × time profiles (as shown in Table 1, Sum AUC for po ²⁶Al and Figure 1). As rats were studied in pairs, errors in the ²⁷Al infusion should be reflected in both rats of the pair. This was not seen. We assume that the 3 rats in the group that ate 1.5% basic SALP in cheese that had higher than average serum ²⁷Al, and therefore AUCs for iv ²⁷Al, renally cleared the ²⁷Al infusion more slowly than most rats. It would be anticipated that the clearance of ²⁶Al would be similar, concomitantly increasing the AUC for ²⁶Al. Therefore, these results seem valid. Had the serum ²⁷Al concentrations in these rats been lower and ²⁶Al concentrations not different, oral Al bioavailability would have been higher. Therefore, the oral bioavailability of Al from basic SALP incorporated into cheese in the rat appears to be at least 0.1%.

The few studies conducted to date to address the possibility that Al bioavailability is dosedependent have also not resulted in robust findings. Oral Al bioavailability from Al lactate in the rabbit dosed with 108 or 540 mg Al/kg, as Al lactate, was 0.7 and 1.9%, although not significantly different (Yokel and McNamara, 1985). Urinary ²⁶Al excretion was higher in two rats after administration of 20 ng of ²⁶Al with 10 mg ²⁷Al than with 0.2 mg ²⁷Al as the chloride (0.032 *vs.* 0.018% of the ²⁶Al dose). Serum ²⁶Al was also higher 1, 5 and 24 hours after the larger total Al dose (Ittel et al., 1993). These results suggest a positive correlation between aluminium dose and bioavailability. On the other hand, Al absorption into rat blood and tissues after perfusion of the gut with 48 or 64 mM Al chloride at pH 3 was not concentration dependent (Arnich *et al.*, 2004).

The later T_{max} of Al from basic SALP in process cheese observed in the present study, compared to water, is consistent with the apparent site of Al absorption, the upper intestine and delayed gastric emptying of food compared to water (Froment et al., 1989; Nagy and Jobst, 1994; Whitehead et al., 1997; Arnich et al., 2004). C_{max} values were lower after oral consumption of ²⁶Al in process cheese than water, as expected when oral bioavailability is lower and T_{max} is later.

The FDA acceptance of basic SALP as a food additive is based on a review conducted in 1975 (FASEB, 1975). The reviewers cited unpublished studies that demonstrated a consistent increase in renal microconcretions in female, but not male, rats consuming a diet containing 0.03 to 3% basic SALP (~ 4 to 40 mg Al/kg/day) for 90 days and an in-consistent increase of this endpoint in male and female beagle dogs that consumed basic SALP delivering 7 to 70 mg Al/kg/day. The review panel noted that care should be taken by patients with kidney disease when consuming food containing high levels of Al salts. However, they did not mention dialysis encephalopathy, which has been attributed to Al, or the controversial role of Al in AD. Description of these clinical problems began about 1975. Since that review the contribution of Al from Al cooking utensils to serum Al and urinary Al excretion in patients with chronic renal insufficiency was demonstrated (Lin et al., 1997) and numerous studies have described toxic endpoints of Al exposure (Krewski et al., 2007).

Some studies have been conducted to assess potential toxicity from basic SALP. A study was conducted by Stauffer Chemical Company in male rats fed their basic SALP-containing products. The treatment groups ate feed for 28 days containing: 3% KASAL (1866 mg Al/kg diet; ~ daily dose 141 mg/kg bw), 0.7% KASAL II (908 mg Al/kg diet; ~ daily dose 67 mg Al/

kg bw), 30,000 ppm KASAL II (3761 mg Al/kg diet; ~ daily dose 288 mg Al/kg bw), 14,470 ppm Al hydroxide as a positive control (4034 mg Al/kg diet; ~ daily dose 302 mg Al/kg bw), or the basic (control) diet (66 mg Al/kg diet; ~ daily dose 5 mg Al/kg bw) (Hicks et al., 1987). Other than a significant increase of serum sodium, no significant effects were seen in bone Al, gross autopsy, histopathology, food consumption, hematology, clinical chemistry or organ weight. A study was conducted in dogs fed diets containing KASAL that produced daily mean Al intakes of ~ 10, 25, or 78 mg Al/kg for 26 weeks, compared to a control diet delivering ~ 4 mg Al/kg/day (Pettersen et al., 1990). The highest dose of Kasal resulted in a sharp, 1.5 week decrease in food consumption and body weight in males, a decrease in testis weight, seminiferous tubule germinal cell degeneration and atrophy, hepatocyte vacuolation, and tubular-glomerularnephritis. There were no significant effects on serum chemistry, hematology, urinalysis or bone or brain Al, except for a significant increase of brain Al in female dogs consuming the highest Al containing diet. Neither of these studies addressed bioavailability.

There have been studies to estimate oral Al bioavailability from foods. A project was commissioned to determine the relative uptake of Al in many foods (UK-MAFF, 1993). However, the methods did not enable determination of bioavailability and the trial foods did not contain cheese (UK-MAFF, 1993). Al bioavailability from diets containing ~ 5 mg Al daily was estimated, based on urinary Al excretion compared to dietary Al intake, to be 0.78% in young human males (Greger and Baier, 1983). However, cheese was not included in this study nor in a study by (Stauber et al., 1999) who estimated Al bioavailability from drinking water and food in humans to be comparable from these two sources, ~ 0.3%. Oral Al bioavailability from food has been estimated to be ~ 0.1% based on average daily urinary Al excretion compared to average daily Al intake from food (Powell and Thompson, 1993; Priest, 1993; Nieboer et al., 1995). The only reported study that determined the bioavailability of Al from a specific food used similar methods to the study reported herein and found it to be ~ 0.1% from acidic SALP incorporated into biscuit (Yokel and Florence, 2006).

In summary, the present estimate of oral Al bioavailability from a dietary component suggests modestly lower or comparable oral Al bioavailability from food than previously reported in studies of water, conducted in the human and rat (Introduction). The results of this study of oral Al bioavailability from basic SALP in a process cheese suggest ~ 0.1 to 0.3% of the Al was orally absorbed. A prior study and a recent replication of that study, using the same methods (Yokel et al., 2001; Zhou and Yokel, 2006), suggested ~ 0.3% of Al was orally absorbed from water. Other substances in drinking water might alter Al. For example, citrate doubled Al absorption from water (Zhou et al., 2008). A comparison of the arithmetic products of the absorbed percentage of Al times their contribution to the diet suggests food typically provides at least 90% of dietary Al that reaches systemic circulation. This is based on consumption of 5 to 10 mg of Al in the diet \times 0.1 to 0.3% absorption, delivering 5 to 30 µg of Al to systemic circulation daily vs. consumption of 0.1 mg Al daily in drinking water \times up to 0.6% absorption, delivering up to $0.6 \,\mu g$ of Al to systemic circulation daily. These results suggesting food is a more important source of absorbed Al than is drinking water, as noted in the Introduction. This suggests that further study of a potential link between orally consumed Al in food and cognitive deficits may be more appropriate than studies of Al in drinking water.

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Abbreviations

AD, Alzheimer's disease

Al, aluminum

AMS, accelerator mass spectrometry

AUC, area under the (concentration x time) curve

C_{max}, maximum serum concentration

ETAAS, electrothermal atomic absorption spectrometry

F, oral bioavailability

GRAS, generally recognized as safe

PTWI, provisional tolerable weekly intake

SALP, sodium aluminum phosphate

T_{max}, time to maximum serum concentration

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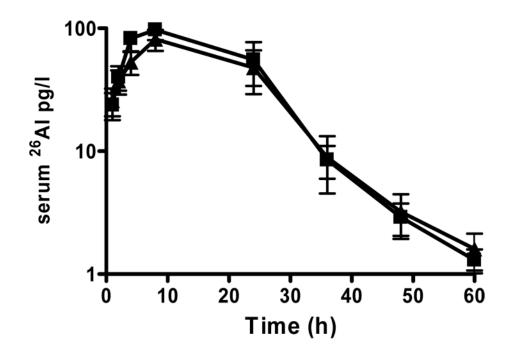


Figure 1.

Concentration of ²⁶Al in serum versus time after consumption of basic ²⁶Al-SALP in a process cheese containing 1.5% (square) or 3% (triangle) basic SALP. The values are mean \pm SD from 6 rats in each group.

Table 1

The results of major determinates of oral Al bioavailability, the oral bioavailability of Al, and other pharmacokinetic parameters in rats dosed with 1.5 or 3% basic SALP, containing ²⁶Al, in cheese (this study) and prior studies with acidic SALP in biscuit or in water. Values are mean \pm SD.

	1.5% basic SALP in cheese	3% basic SALP in cheeses	acidic SALP ^a in biscuit	Water ^b
Sum AUC for po ²⁶ Al (pg/l/ h)	2219 ± 1127 ^c	2022 ± 1372	729 ± 481	5054 ± 1756
Sum AUC for iv ²⁷ Al (µg/l/ h)	$68745 \pm 63166 \ d$	20922 ± 8933	25829 ± 23790	21320 ± 17772
Oral Al bioavailability (%)	$0.10 \pm 0.07 \ ^{e}$	0.29 ± 0.18^{f}	0.12 ± 0.11	0.28 ± 0.16
T _{max} (h)	8.0 ± 2.6 ^C	8.6 ± 1.6	4.8 ± 1.6	1.5 ± 1.0
C _{max} (pg/l serum)	104 ± 38 ^{<i>C</i>}	79 ± 37	50 ± 33	607 ± 295
C _{max} (% of dose/ml serum)	$0.00011 \pm 0.00003 \ d$	0.00013 ± 0.00008	0.00014 ± 0.00007	0.00088 ± 0.00045

^{*a*}Results from Yokel and Florence (2006). This study used 10 male Fisher 344 rats weighing 322 ± 32 gm that consumed ~ 1 gm biscuit containing ~ 1 nCi ²⁶Al in 1 or 2% acidic SALP.

^bResults from Yokel et al. (2001) and Zhou and Yokel (2006). These studies used male Fisher 344 rats; 19 weighing 280 ± 42 gm (Yokel et al., 2001) and 5 weighing 268 ± 18 gm (Zhou and Yokel, 2006), that were given an intra-gastric dose of 1 ml ~ pH 5 water solution containing ~ 1 nCi ²⁶Al.

^cThe combined results from 1.5 and 3% basic SALP in cheese were significantly different from acidic SALP in biscuit and Al in water.

 d The combined results from 1.5 and 3% basic SALP in cheese were significantly different from Al in water.

^eSignificantly different from 3% SALP in cheese and Al in water.

^fSignificantly different from Al in biscuit.