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Organic Cation Transporter Preferentially Expressed in Hematopoietic Cells and Leukemias and Uses Thereof

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(12) United States Patent

Moscow et al.

(10) Patent No.: US 7,723,019 B2 (45) Date of Patent: May 25, 2010

(54) ORGANIC CATION TRANSPORTER PREFERENTIALLY EXPRESSED IN HEMATOPOIETIC CELLS AND LEUKEMIAS AND USES THEREOF

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Xin Lu, Shanghai (CN); Craig Jordan,

Rochester, NY (US)

(73) Assignee: University of Kentucky Research

Foundation, Lexington, KY (US)

(*) Notice: Subject to any disclaimer, the term of this

patent is extended or adjusted under 35

U.S.C. 154(b) by 451 days.

(21) Appl. No.: 11/521,487

(22) Filed: Sep. 15, 2006

(65) Prior Publication Data

US 2007/0269846 A1 Nov. 22, 2007

Related U.S. Application Data

- (62) Division of application No. 10/849,551, filed on May 20, 2004, now abandoned.
- (60) Provisional application No. 60/471,709, filed on May 20, 2003.

(51)	Int. Cl.	
	C12Q 1/00	(2006.01)
	G01N 33/53	(2006.01)
	C07K 14/435	(2006.01)

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Primary Examiner—Bridget E Bunner (74) Attorney, Agent, or Firm—McDermott Will & Emery LLP

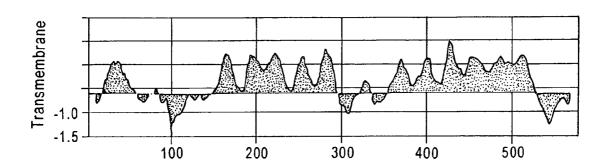
(57) ABSTRACT

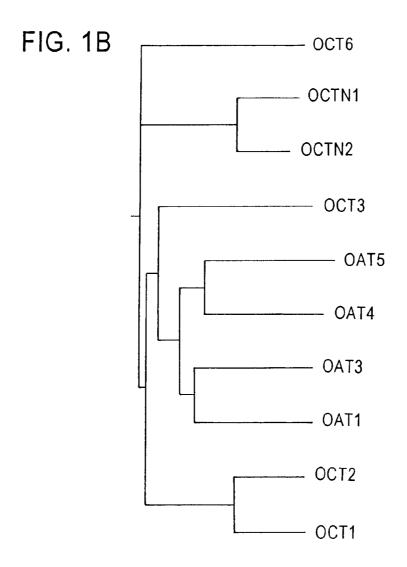
A novel organic cation transporter (OCT) gene, OCT 6, and use thereof is described. The OCT6 gene is preferentially expressed in human hematopoietic tissues, including CD34+cells and leukemia cells. Its narrow tissue distribution, substrate specificity, and close homology to other cell membrane transporters make OCT6 an attractive target for the treatment of myeloid diseases.

7 Claims, 10 Drawing Sheets

FIG. 1A

May 25, 2010





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FIG. 3A

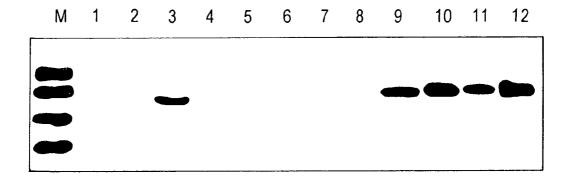


FIG. 3B

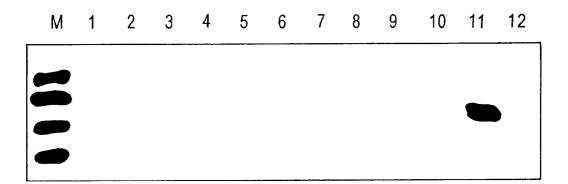
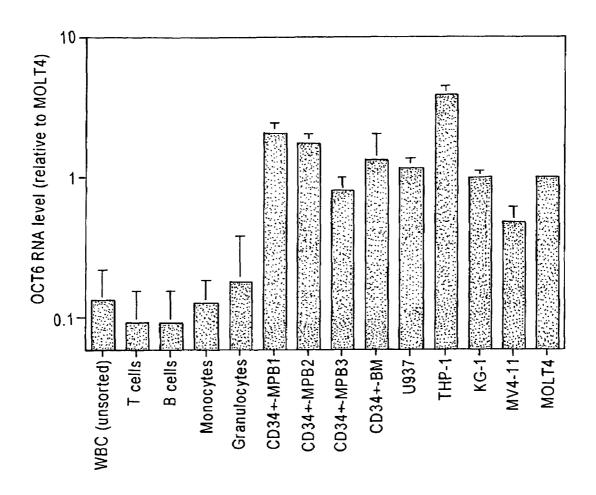
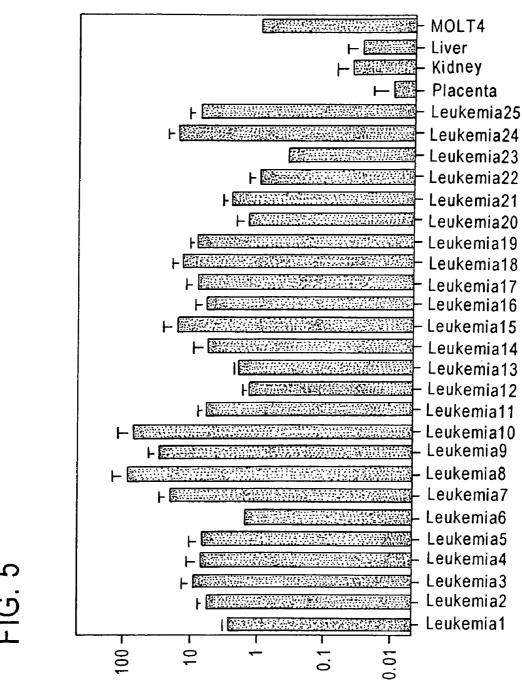


FIG. 4





OCT6 RNA levels (relative to MOLT4)

ORGANIC CATION TRANSPORTER PREFERENTIALLY EXPRESSED IN HEMATOPOIETIC CELLS AND LEUKEMIAS AND USES THEREOF

CROSS REFERENCE TO RELATED APPLICATIONS

This application is a divisional application of and claims the benefit of application Ser. No. 10/849,551, filed May 20, 10 2004 now abandoned, which claims the benefit of U.S. Provisional Application No. 60/471,709, filed May 20, 2003.

FIELD OF THE INVENTION

The invention relates to a gene encoding an organic cation transporter, OCT6, and its use as a target for the treatment of hematological malignancies, and in particular, leukemia. The invention further relates to screening methods for identifying agonists and antagonists/binding partners of OCT6 transport 20 activity.

BACKGROUND OF THE INVENTION

The lipid bilayer of the cellular membrane insulates the intracellular milieu from exposure to hydrophilic compounds. Unlike lipophilic compounds that can diffuse through cellular membranes, water-soluble compounds usually require specific transport mechanisms to gain access to the intracellular space. The regulation of the traffic of polar compounds in both directions across the cellular membrane is a complex process involving several large families of transport proteins.

Most often in cancer research, drug transport is thought of as a mechanism of cellular drug resistance, as drug efflux 35 pumps such as the products of the MDR1 and MRP genes have been shown to be mechanisms of resistance to lipid-soluble anticancer drugs. However, drug transport is a two-way street, and mechanisms also exist for pumping drugs into cells. For polar, water-soluble anticancer agents, drug uptake, and not drug efflux, is the critical determinant of cellular drug accumulation.

Most cancer chemotherapy employs drugs that are lipid-soluble that can easily penetrate the cell membrane of cancer cells. One advantage of using lipid-soluble drugs is that they 45 easily gain intracellular access to different types of cancer cells, so many cancer cells appear to be initially sensitive to these drugs. The disadvantage is that cancer cells learn to increase the activity of drug efflux pumps in the cell membrane to pump lipid-soluble drugs out of the cell, resulting in 50 drug resistance.

In contrast, potential water-soluble anticancer drugs may not survive the preclinical screening process since there is a great deal of variability in the expression of drug transport genes in different types of cancer cells. Variability in transport 55 gene expression may result in variability in accumulation of polar, water-soluble drugs. One approach to more effectively utilize water-soluble anticancer drugs is to identify which of the dozens of transport genes are actually expressed in tumors.

The importance of carrier-mediated anticancer drug uptake is exemplified in reduced folate carrier (RFC) mediated uptake of methotrexate (MTX). Methotrexate (MTX), a reduced folate analogue, is scavenged and retained in cells by mechanisms designed to secure folates from the environment. 65 The major mechanism of MTX uptake at pharmacologic concentrations is the reduced folate carrier (RFC), an OAT trans-

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porter with a Km for MTX between approximately 0.8-26 μM. Decreased RFC activity has been observed in several in vitro models of transport-mediated MTX resistance (Biochem. Pharmacol. 11: 1233-1234, 1960). Once rodent and human genes encoding proteins with RFC activity were isolated, the molecular explanations for decreased RFC activity emerged. RFC1 transfection into the transport-deficient MTX^R ZR75 cell line resulted in a 20-fold increase in 6-hour MTX uptake and a concomitant 250-fold increase in sensitivity to MTX relative to control cell clones, showing that the RFC1 gene reconstitutes RFC activity and has a significant impact on MTX cytotoxicity (Moscow, et al., Cancer Res. 55: 3790-3794, 1995).

In different cell lines, MTX transport deficiency has been ascribed either to mutations in the RFC gene or in decreased expression of the RFC gene product. Several studies have demonstrated that RFC1 gene expression is an important determinant of sensitivity to MTX. In in vitro studies, we have found that RFC1 RNA levels correlate with MTX sensitivity in a panel of non-selected cell lines, including breast cancer cell lines (Moscow et al., Int J Cancer. 72: 184-190, 1997).

A plethora of genes with the ability to transport MTX out of the cell have been reported, including MRP1, MRP2, MRP3, MRP4, the organic anion transporters hOAT2 and hOAT3, and the mitoxantrone-resistance protein (BCRP/MXR). However, despite the multitude of MTX export genes, clinical studies have shown a relationship between the expression of RFC1, the mechanism of MTX uptake, and prognosis in Acute Lymphoid Leukemia (ALL) and osteosarcoma. As a result, RFC1 expression and MTX uptake are now implicated as determinants of clinical sensitivity in several types of tumors. Thus, the role of RFC1 in mediating sensitivity of its cytotoxic drug substrates has become a prototype that illustrates the potential role of transporters, like OAT and OCT genes, in determination of anticancer drug selectivity and toxicity.

However, there is a need to identify additional channels, or transporters, that are found in specific cancers, to enable the targeting of different cancers with anticancer agents that are substrates for those transporters.

SUMMARY OF THE INVENTION

The present invention is directed towards a membrane protein that functions to transport hydrophilic substances across cellular membranes. The protein, OCT6, is a new member of the organic cation transporter (OCT) family (SLC22 gene family). Tissue distribution of this protein is distinct from other OCT protein family members; being detected in leukemia, leukemia blast cells and CD34+ cells.

In one aspect, the present invention provides a novel target for hematological malignancies such as leukemia, an OCT6 transporter.

In another aspect of the present invention there is a method for screening potential substrates that selectively bind the OCT6 transporter. The method involves contacting a cell which overexpresses an OCT6 transporter gene with a test compound and determining whether the test compound is a substrate for the OCT6 transporter.

In another aspect, there is a method for screening potential anti-cancer agents in a cell overexpressing an OCT6 transporter gene. The method comprises determining viability of a cell which expresses OCT6 transporter gene incubated in the presence and absence of a test compound and identifying the test compound as a potential anti-cancer agent if there is cellular influx of the test compound and cell death.

In another aspect of the invention, a test kit is provided for screening candidate drugs for hematologic malignancies comprising a mammalian cell line or cells which overexpress OCT6, a control substrate and a detectable substance.

In still another aspect of the invention, there are immunogenic compositions for treating hematological malignancies. In a preferred embodiment, immunogenic compositions for treating leukemia comprise a substrate that binds selectively to a leukemia cell expressing the OCT6 transporter gene. In another preferred embodiment of the invention, the substrate comprises an antibody that selectively binds to the OCT6 transporter protein. Preferably, the OCT6 transporter protein allows cellular uptake of the substrate which then causes cell death. In one embodiment the substrate is cytotoxic and in another preferred embodiment the substrate is coupled with a 15 cytotoxic agent.

In still another aspect, the present invention provides a method for impairing a leukemia cell comprising contacting the cell with a cytotoxic OCT6 transporter protein. In one embodiment the substrate is a cytotoxin and in another 20 embodiment the substrate is coupled to a cytotoxic agent.

In yet another aspect, the present invention provides a method for treating hematological malignancies comprising administering to a subject in need thereof an immunogenic composition comprising a substrate that binds selectively to a 25 cell expressing the OCT6 transporter gene. In a preferred embodiment the OCT6 transporter protein allows cellular uptake of the substrate which then causes cell death. In another preferred embodiment the substrate is cytotoxic. In another preferred embodiment, the substrate is coupled with 30 a cytotoxic agent.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1. A. shows the predicted hydropathy profile of $_{35}$ OCT6.

FIG. 1. B. is a dendrogram showing phylogenic relationship between OCT6 (SEQ ID NO:2) and other OCT and OAT proteins, including, OCTN1 (SEQ ID NO:4), OCT3 (SEQ ID NO:5), OCTN2 (SEQ ID NO:6), OCT2 (SEQ ID NO:7), 40 OCT1 (SEQ ID NO:8), OAT5 (SEQ ID NO:9), OAT4 (SEQ ID NO:10), OAT3 (SEQ ID NO:11), and OAT1 (SEQ ID NO:12).

FIG. **2**A-F. is the CLUSTLAW alignment of OCT6 and other OCT and OAT proteins. The bottom row represents $_{45}$ areas of consensus.

FIG. 3. shows the normal tissue distribution of OCT6 RNA determined by RT-PCR using a cDNA panel. Only 1000× (highest) cDNA concentration is shown. Panel A. 1, salivary gland; 2, thyroid; 3, adrenal; 4, pancreas; 5, ovary; 6, uterus; 507, prostate; 8, skins; 9, peripheral blood leukocytes; 10, bone marrow; 11, fetal brain; 12, fetal liver. Panel B. 1, brain; 2, heart; 3, kidney; 4, spleen; 5, liver; 6, colon; 7, lung; 8, small intestine; 9, muscle; 10, stomach, 11, testis; 12, placenta.

FIG. 4. shows quantitative RT-PCR for the transporter gene 55 OCT6 performed with RNA extracted from peripheral blood leukocytes, CD34+ cells and additional hematopoietic cell lines. Fresh discarded buffy coats that were twice sorted by FACS using CD14 (monocytes), CD15 (granulocytes), CD3 (T-cells) and CD20 (B-cells). Purities of 99% or better were 60 obtained. For peripheral WBC and sorted subsets, the average±SD represent pooled results from samples from 2 individuals performed in triplicate or quadruplicate. For CD34-selected mobilized peripheral blood (MPB), the results from each of 3 individuals are shown. For CD34-selected bone marrow (CD34+-BM), the results are from one individual. OCT6 levels were normalized to the expression of

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actin RNA, as a control for equivalence of mRNA template. The units, in log scale, are arbitrary and based on a standard curve of OCT6 RT-PCR in serially diluted HL60 RNA. Unity is defined as the level of OCT6 RNA found in MOLT4 cells.

FIG. 5. shows quantitative RT-PCR for the gene OCT6 using RNA extracted from leukemic blasts obtained from patients at the time of initial diagnosis. OCT6 levels were normalized to the expression of actin RNA, as a control for equivalence of mRNA template. The OCT6 RNA levels in placenta, liver, kidney and MOLT-4 cell line were determined concurrently and shown for comparison. The units, in log scale, are arbitrary and based on a standard curve of OCT6 RT-PCR in serially diluted HL60 RNA. Unity is defined as the level of OCT6 RNA found in MOLT4 cells.

DETAILED DESCRIPTION OF THE INVENTION

The present invention is based on the discovery and isolation of a new member of the SLC22 gene family (the OCT family of proteins) that is unusual for its distinct pattern of tissue distribution. Rather than the typical high levels of expression in liver, kidney or placenta, high levels of RNA for this transporter were found in some leukemia cell lines, in CD34+ cells, and in circulating leukemia blast cells.

All patents, patent applications and literature cited in this description are incorporated herein by reference in their entirety. In the case of inconsistencies, the present disclosure, including definitions, will prevail.

OCT Family

Two families of proteins involved in maintaining homeostasis of charged organic compounds are the organic anion transporters (OATs) which carry the SLC21 designation and the organic cation transporters (OCTs), which carry the SLC22 designation (See Table 1). OATs and OCTs each have characteristic patterns of tissue expression, with predominant expression in a tissue involved in the transport of xeriobiotics, i.e., liver, kidney or placenta.

TABLE 1

Organic anion and cation transported genes							
Gene Family	Gene Name	Locus Link	Alternative Names				
SLC21	SLC21A1	6577					
	SLC21A2	6578	PGT				
	SLC21A3	6579	OATP, OATP1, OATP1b,				
			OATP-A				
	SLC21A4	28237	OAT-K1, OAT-K2				
	SLC21A5	28236	OATP2, OATP-2				
	SLC21A6	10599	LST-1, OATP-C				
	SLC21A7	28235	OATP3, OATP-3				
	SLC21A8	28234	LST2, OATP8, SLC21A8,				
			OATP-8				
	SLC21A9	11309	OATP-B				
	SLC21A10	28233	OATP4				
	SLC21A11	28232	OATP-D				
	SLC21A12	28231	LOC51737, OATP-E, POAT				
	SLC21A13	28230	OATP5, OATP-5				
	SLC21A14	53919	OATP-F				
SLC22	SLC22A1	6580	OCT1				
	SLC22A2	6582	OCT2				
	SLC22A3	6581	OCT3				
	SLC22A4	6583	OCTN1				
	SLC22A5	6584	OCTN2, CDSP, SCD				
	SLC22A6	9356	NKT, OAT1, OAT-1				
	SLC22A7	10864	NLT, OAT2, OAT-2				
	SLC22A8	9376	OAT3, OAT-3				
	SLC22A9		OAT4, OAT-4				

The OAT and OCT carriers result in increased cellular accumulation of their respective substrates, despite the fact

that they are carriers that mediate facilitative diffusion. For carriers, the degree of intracellular accumulation may not exceed the extracellular concentration. However, the presence of the carrier allows uptake in comparison to no uptake in the absence of the carrier, and drugs that bind an intracellular target or which are chemically modified in the cells, e.g., by phosphorylation or polyglutamylation, may be eliminated from the substrate pool and not available for transport back across the cellular membrane.

The first five members of the SLC22 family of transporters, OCT1, OCT2, OCT3, OCTN1, and OCTN2, have been characterized as organic cation transporters. The uptake of many cations, such as tetraethylammonium (TEA), N-1-methylnicotineamide (NMN), choline, procainamide, amantadine and morphine are mediated by these polyspecific transporters. In general, these transporters are potential-dependent, but independent of sodium and proton gradients. These genes are all characterized by the presence of 11 or 12 transmembrane domains, as predicted by hydrophobicity analysis, and all have a large hydrophilic loop between transmembrane domain (TMD) 1 and TMD2.

OCT substrates are shown below in Table 2. Tetraethyl ammonium (TEA) is the classic substrate for OCT transporters. In addition, OCT1, OCT2 and OCT3 transport 1-methyl-4-phenylpyridinium (MPP). Compared to OCT2, OCT1 has a higher affinity for some cations (for example mepiperphenidol and procainamide), a similar affinity for others (for example, decynium 22 and quinidine), and a lower affinity for corticosterone (See Koepsell et al., Ann. Rev. Physiol. 60: 243-266, 1998.). OCT3 is an electrogenic transporter for TEA and guanidine. Other physiologic substrates for OCT transporters include dopamine, histamine, epinephrine and norepinephrine, acetylcholine and 5-hydroxytryptamine (Burckhardt, et al., Am J Physiol Renal Physiol. 278: F853-66., 2000.), suggesting an important role for these transporters in the central nervous system, in addition to their role in hepatic and renal clearance. Interestingly, despite its cationic nature, recent studies have identified cimetidine as a selective inhibitor, but not a substrate for several organic cation transporters, including rOCT1, rOCT2, rOCT3, hOCTN1, and hOCTN2.

TABLE 2

	OCT Substrates							
Common Name	Gene Name	Cell Type	Substrate	KT (uM)				
OCT1	SLC22A1	HeLa	TEA	229				
OCT1	SLC22A1	Xenopus	MPP	14.6				
OCT2	SLC22A2	Xenopus	Norepinephrine	1900				
OCT2	SLC22A2	Xenopus	Histamine	1300				
OCT2	SLC22A2	Xenopus	Dopamine	390				
OCT2	SLC22A2	Xenopus	Serotonin	80				
OCT2	SLC22A2	HEK293	MPP	16				
OCT2	SLC22A2	HEK293	Dopamine	330				
OCT2	SLC22A2	Xenopus	Amantadine	27				
OCT2	SLC22A2	Xenopus	Memantine	34				
OCT3	SLC22A3	HeLa	TEA	2500				
OCT3	SLC22A3	HRPE	MPP	47				
OCTN1	SLC22A4	Fibroblasts	L-Carnitine	6.6				
OCTN2	SLC22A5	HEK293	L-Carnitine	4.34				
OCTN2	SLC22A5	HEK293	L-Carnitine	4.3				
OCTN2	SLC22A5	HEK293	D-Carnitine	10.9				
OCTN2	SLC22A5	HEK293	Acetyl-L-carnitine	8.5				
OCTN2	SLC22A5	Xenopus	L-Carnitine	4.8				
OCTN2	SLC22A5	Xenopus	D-Carnitine	98				
OCTN2	SLC22A5	JAR	L-Carnitine	3.5				

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OCT1 and OCT2 are predominantly expressed in the kidney and liver. These transporters are located on the basolateral surface of renal tubules and, therefore, play a role in the removal of organic cations from the blood. OCT3 is most abundantly expressed in placenta. In addition, other tissue-specific roles have been implicated for these transporters. As noted above, OCTs may play a role in transport of endogenous neuroleptic substrates, and OCT3 has been implicated in the disposition of cationic neurotoxins and neurotransmitters in the brain (Wu, et al., J Biol Chem. 273: 32776-86, 1998). Dhillon et al. (Clin Pharmacol Ther. 65: 205, 19996) used RT-PCR followed by functional transport studies (TEA) to identify OCT1 expression in a human mammary epithelial cell line (MCF12A). Further, the OCT1 gene has been shown to be up regulated in lactating mammary epithelial cells.

The OCTN1 gene, cloned from a cDNA, shows sequence similarity to organic cation transporter genes, which is highly expressed in kidney as well as trachea, bone marrow and fetal liver. Recombinant OCTN1 expressed in mammalian cells exhibited saturable uptake of TEA that was pH sensitive. Several others suggest that OCTN1 is a renal proton/organic cation antiporter functioning at the epithelial apical membrane. The uptake of pyrilamine, quinidine, verapamil and L-carnitine were increased by expression of OCTN1 in *Xenopus* oocytes.

Another OCT protein family member, OCTN2, cloned from a human placental trophoblast cell line, is expressed widely in human tissues including kidney, placenta and heart. OCTN2 is more closely related to OCTN1 than to OCT1, OCT2 and OCT3 (Biochem Biophys Res Commun. 246: 589-95, 1998). Transfection of OCTN2 has demonstrated its role in the transport of TEA and carnitine. OCTN2-mediated transport of TEA is sodium independent, whereas transport of carnitine is sodium-dependent. The role of sodium in OCTN2-mediated carnitine transport not only involves the electrogenic gradient, but the presence of sodium also alters the affinity of OCTN2 for carnitine. Germline mutations of OCTN2 result in primary carnitine deficiency, a syndrome of progressive cardiomyopathy and skeletal myopathy. The symptoms associated with this syndrome are thought to result not only from generalized carnitine deficiency from decreased renal carnitine reabsorbtion, but also from inability of cardiac and skeletal myocytes, which ordinarily express OCTN2, to accumulate carnitine. This syndrome demonstrates that tissue-specific OCT-mediated transport is essential for accumulation of required cations in specific tissues.

The present invention identifies a new transport protein in the OCT family, OCT6, preferentially expressed in leukemia cell lines, leukemia blast cells and CD34+ cells. The cell surface localization and the transporter function of the OCT6 gene product suggest its usefulness as a target in the diagnosis and treatment of hematologic malignancies.

As used herein, the term "antibody" refers to an immunoglobulin molecule with a specific amino acid sequence 55 evoked in by an antigen, and characterized by reacting specifically with the antigen in some demonstrable way.

As used herein, the term "carrier" refers to a diluent, adjuvant, excipient, or vehicle with which the compositions of the present invention are administered.

As used herein, "compound" refers to any agent, chemical, substance, or substrate, whether organic or inorganic, or any protein including antibodies, peptides, polypeptides, peptoids, and the like.

As used herein, the term cytotoxin" or cytoxic agent includes any specific substance, which may or may not be antibody, that inhibits or prevents the functions of cells, causes destruction of cells, or both.

As used herein, the term "derivative" refers to something produced by modification of something pre-existing; for example, a substance or chemical compound that may be produced from another substance or compound of similar structure in one or more steps.

As used herein, the term "fragment" refers to a part of a larger entity, said larger entity comprising by non-limiting example, an antibody, compound or substance.

As used herein, the term "leukemia blast" or "leukemic blast" refers to lymphoblasts, the abnormal immature white 10 blood cells associated with leukemia.

As used herein, the term "monoclonal antibody" is not limited to antibodies produced through hybridoma technology. The term "monoclonal antibody" refers to an antibody that is derived from a single clone, including any eukaryotic, 15 prokaryotic, or phage clone, and not the method by which it is produced.

As used herein, the term "pharmaceutically acceptable carrier" refers to a carrier that may be administered to a subject, together with one or more liver protecting agents and one or 20 more mushroom powder or extract of the present invention, and which does not destroy the pharmacological activity thereof and is nontoxic when administered in doses sufficient to deliver a therapeutic amount of the compound.

As used herein, the term "substrate" refers to a substance, 25 compound, agent, antibody or derivatives and/or fragment thereof, acted upon by the OCT6 transporter protein (e.g., a substance that is taken across the cellular membrane by action of the OCT6 transporter protein).

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OCT6 (SEQ ID NO:1) was first identified as a potential OCT gene by assembling and sequencing ESTs as described in Example 1 (amino acid sequence of OCT6 is SEQ ID NO:2). The gene sequence proved to be identical to the recently submitted cDNA OKB1 (GenBank AF268892) submitted by M. Okabe and T. Abe, incorporated herein in its entirety. It is also contained within the submitted BAC clone CTA-331P3 (SEQ ID NO: 3) (GenBank AC002464) located at chromosome 6q21, incorporated herein in its entirety. The gene has a predicted protein structure typical of transport proteins with two groups of six transmembrane domains separated by a hydrophilic region (FIG. 1A). CLUSTALW alignment produced a dendrogram showing the phylogenic relationship between OCT6 and other OAT and OCT proteins (FIG. 1B). This dendrogram suggests that the distinction between OAT and OCT genes, based on functional studies, obscures the common origin of both families of transporters. The actual CLUSTALW alignment of these genes is shown in FIG. 2 and demonstrates multiple regions of conservation among all of these genes.

Next, according to the methods described in Example 3, quantitative RT PCR analysis of the expression of OCT6 was performed, along with the expression of other OCT genes, in 50 cell lines. The results are shown in Table 3. The two highest expressing cell lines for OCT6 in this panel were two leukemia cell lines, HL60, a human promyelocytic leukemia cell line, and MOLT4, a human acute lymphoblastic leukemia (T-cell) cell line. There was only a low level of expression detected in most of the other cell lines.

TABLE 3

	TADLE 3										
	OCT expression in 50 cell lines of the NCI Drug Screen										
No.	Cell Line	source	OCT1	OCT2	OCT3	OCTN 2	OCT6				
1	CCRF-CEM	Leukemia	0.7	0.7	0.2	0.1	5.7				
2	HL-60	Leukemia	0.5	1.3	0.0	0.4	716				
3	K-562	Leukemia	1.4	1.2	0.2	1.4	5.2				
4	MOLT-4	Leukemia	0.1	1.1	0.5	0.6	46.8				
5	RPMI-8226	Leukemia	2.8	2.0	0.1	3.7	6.02				
6	SR	Leukemia	1.9	1.1	0.0	0.3	2.6				
7	A549/ATCC	Lung cancer	1.7	1.2	161	4.3	1.2				
8	HOP-62	Lung cancer	0.8	4.8	0.6	2.4	4.1				
9	NCI-H226	Lung cancer	4.8	0.5	0.1	21.1	4.8				
10	NCI-H23	Lung cancer	0.5	0.7	0.0	0.3	5.2				
11	NCI-H460	Lung cancer	0.7	1.0	0.0	1.7	1.8				
12	COLO205	Colon Ca.	4.9	5.3	30.9	2.2	3.6				
13	HCC-2998	Colon Ca.	1.5	1.0	0.0	2.6	5.4				
14	HCT-116	Colon Ca.	1.7	2.1	0.1	2.8	9.7				
15	HCT-15	Colon Ca.	0.9	1.7	0.1	3.5	4.2				
16	HT-29	Colon Ca.	1.9	1.2	18.1	1.5	1.5				
17	KM-12	Colon Ca.	0.6	1.0	12.2	0.7	2.1				
18	SW-620	Colon Ca.	1.0	2.6	40.4	1.9	3.7				
19	SF-268	CNS Tumor	0.4	0.8	0.0	0.9	2				
20	SF-295	CNS Tumor	0.5	1.2	0.2	1.1	2.5				
21	SF-539	CNS Tumor	0.5	0.6	2.3	0.2	5.3				
22	SNB-75	CNS Tumor	0.8	1.8	0.0	0.6	2.3				
23	U251	CNS Tumor	0.8	0.9	0.0	0.6	7.4				
24	LOCIMVI	Melanoma	2.9	2.1	0.1	0.4	3.6				
25	MALME-3M	Melanoma	1.5	1.5	0.0	2.3	3				
26	M14	Melanoma	1.9	1.4	0.0	1.9	4.7				
27	SK-MEL-2	Melanoma	2.1	1.9	0.0	2.2	3.9				
28	SK-MEL-5	Melanoma	2.6	1.5	0.0	1.9	2.7				
29	UACC-257	Melanoma	3.2	3.6	0.0	1.1	5.4				
30	IGROV1	Ovarian Ca.	4.9	5015	17.9	1.8	2.5				
31	OVCAR-3	Ovarian Ca.	1.4	0.1	0.0	2.2	14				
32	OVCAR-4	Ovarian Ca.	2.6	1.4	0.0	8.9	3.4				
33	OVCAR-5	Ovarian Ca.	3.5	2.7	105	10.0	4.8				
34	OVCAR-8	Ovarian Ca.	1.1	1.0	0.0	0.8	1.6				
35	SK-OV-3	Ovarian Ca.	3.9	1995	9.2	8.5	9.8				
36	A498	Renal Ca.	2.2	13.4	180	4.7	1.3				
37	ACHN	Renal Ca.	1.1	1.1	0.7	1.2	1.1				
38	CAKI_1	Renal Ca.	3.5	2.5	4.8	1.8	2.8				

TABLE 3-continued

	OCT expression in 50 cell lines of the NCI Drug Screen											
No.	Cell Line	source	OCT1	OCT2	ОСТ3	OCTN 2	OCT6					
39	RXF-393	Renal Ca.	1.7	1.2	3.0	0.6	1.2					
40	TK-10	Renal Ca.	3.6	5.0	16.8	2.5	8					
41	UO-31	Renal Ca.	4.4	1.6	31.2	1.2	2.3					
42	PC-3	Prostate Ca.	2.1	0.8	9.6	3.3	4.7					
43	DU-145	Prostate Ca.	1.1	1.1	3.4	1.6	3					
44	MCF-7	Breast Ca.	0.8	1.8	0.0	10.4	3.5					
45	NCI/ADR-RES	Breast Ca.	1.4	1.3	1.1	2.0	2.1					
46	MDA-MB-231	Breast Ca.	1.2	0.4	3.9	4.8	1.8					
47	HS578T	Breast Ca.	1.0	1.5	0.0	1.2	8.3					
48	MDA-MB-435	Breast Ca.	1.9	0.6	0.1	0.7	2.7					
49	BT-549	Breast Ca.	1.2	0.8	0.1	0.3	2.6					
50	T-47D	Breast Ca.	0.7	1.1	0.1	4.2	8.7					

OCT6 is unique among the known members of OCT and OAT genes because of its pattern of tissue distribution. The pattern of expression of the OCT6 gene in the 50 cell lines suggested that its expression might be restricted to hematopoietic tissues. The restricted pattern of expression observed 25 for OCT6 also suggests that therapies using OCT6-specific substrates are unlikely to have widespread toxicity to normal tissues. Therefore, we examined OCT6 expression in a cDNA panel representing a wide cross-section of normal tissues according to the methods of Example 4 (FIG. 3). This study 30 revealed that OCT6 RNA levels are highest in testis and fetal liver, with lower but detectable levels in peripheral blood leukocytes and bone marrow. Since fetal hematopoiesis occurs in the liver, it is possible that the fetal liver sample may have included both hepatocytes and hematopoietic cells. OCT6 RNA levels were also barely detectable in pancreatic and adrenal tissue. Unlike other OCT genes, expression was not detectable in liver, kidney or placenta.

To determine whether OCT6 RNA expression in hematopoietic cells was lineage-specific, leukocytes were sorted from discarded buffy coat specimens by flow cytometry, and purified subpopulations were examined for OCT6 RNA expression according to the methods described in Example 5. OCT6 expression was also examined in a population of 45 CD34+ cells. As can be seen in FIG. 4, the expression of OCT6 was highly enriched in CD34+ cells in comparison to the other cell populations. Also, significant levels of OCT6 expression (relative to MOLT4) were found in other hematopoietic cell lines: U937, a human histiocytic lymphoma cell line; THP-1, a human acute monocytic leukemia cell line; KG-1, a human erythroleukemia cell line; and MV-4-11, a human biphenotypic (B-cell and myelomonocytic) leukemia cell line

The high levels of OCT6 RNA in some leukemia cell lines and CD34+ cells also raised the question as to whether this gene was highly expressed in actual leukemias. To address this issue, the RNA levels of OCT6 in 25 samples of peripheral leukemic cells were measured according to the methods set out in Example 6. The FAB classification of these samples are shown in Table 4. These results are shown in FIG. 5, and demonstrate that the majority of specimens contained RNA levels for OCT6 that exceeded the level found in MOLT4 cell line, the second highest expressing cell line among those examined, and exceed by orders of magnitude the levels found in placenta, kidney and liver.

TABLE 4

Phenotypes of leukemia specimens								
Sample Number	Description							
1	CML, blast crisis							
2 3	CML, blast crisis							
3	CML, stable phase							
4	CML, probably stable phase							
5	CML, accelerated phase							
6	ALL							
7	ALL							
8	AML							
9	ALL							
10	ALL							
11	ALL							
12	AML							
13	AML							
14	AML							
15	AML							
16	ALL, biphenotypic							
17	ALL, biphenotypic							
18	AML							
19	AML, M2							
20	AML, M2							
21	AML, M4							
22	AML, M4							
23	AML, M1							
24	AML							
25	AML, M4							

Due to the OCT6 protein's location on the cellular membrane and its function as an intracellular transporter, the OCT6 transporter protein has been identified as a therapeutic target. Basic principles of cellular pharmacology suggest that increase in intracellular accumulation will lead to increased intracellular effect. For anticancer drugs, this principle has been studied extensively in the context of lipophilic drugs, which require no specific mechanism for cellular uptake, and export pumps such as the product of the multidrug resistance gene, MDR1, whose overexpression of MDR1 leads to increased cellular resistance by decreasing intracellular concentrations of drug (Moscow, J. A., Schneider, E. S., Ivy, S. P., and Cowan, K. H. Multidrug resistance. In: H. M. Pinedo, D. L. Longo, and B. A. Chabner (eds.), Cancer chemotherapy and biological response modifiers. Annual 17. New York: Elsevier, 1997). The same principle applies to charged, hydrophilic drugs of the present invention, except that the determinants of sensitivity depend on uptake as opposed to efflux. As such, cells overexpressing an OCT6 transporter are likely to be highly sensitive to cytotoxic OCT6 substrates.

Drug Screening

Accordingly, the present invention provides methods for screening potential substrates of, and potential therapeutic agents against hematological malignancies like leukemia that overexpress, the OCT6 transporter. In particular, potential 5

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therapeutic agents are screened for the ability to be a substrate recognized by an OCT6 transporter protein. Preferably, potential substrates are screened for the ability to confer cytotoxic effects on a cell overexpressing OCT6 transporter protein. More preferably, agents are screened for the ability to 10 preferentially cause cellular uptake into, and cell death of, cells overexpressing the OCT6 transporter. Most preferably, the agents are screened for the ability to cause cell death of cancer cells such as leukemia overexpressing the OCT6 transporter as compared to normal cells.

A method for screening potential substrates of the OCT6 transporter protein comprises providing a cell or cell line which expresses OCT6 and a test compound, incubating the test compound and cell line and analyzing the cell or cell line to determine if there was a cellular influx of the test com- 20 pound. Analysis of the cell line to determine whether cellular uptake of the test compound occurred can be accomplished by any means known in the art. For example, a test compound can be tagged with a detectable label prior to contact with a cell and then observed under microscopy or by other means 25 microscopy. for its location. Non-limiting examples of labels include green fluorescent protein, alkaline phosphatase, horseradish peroxidase, rease, f3-galactosidase, CAT, luciferase, an immunogenic tag peptide sequence, an extrinsically activatable enzyme, an extrinsically activatable toxin, an extrinsi- 30 cally activatable fluor, an extrinsically activatable quenching agent, a radioactive element or an antibody.

A method for screening candidate anti-cancer agents comprises determining the viability of a mammalian cell which expresses OCT6 incubated in the presence and absence of a 35 test compound and identifying the test compound as a potential anti-leukemia agent if there is a cellular uptake of the test compound and cell death. Analysis of cell viability can be accomplished by any means known in the art.

It is well known in the art that viability of a cell can be 40 determined by contacting the cell with a dye and viewing it under a microscope. Viable cells can be observed to have an intact membrane and do not stain, whereas dying or dead cells having "leaky" membranes do stain. Incorporation of the dye by the cell indicates the death of the cell. The most common 45 dye used in the art for determining viability is trypan blue. Viability of cells can also be determined by detecting DNA synthesis. Cells can be cultured in cell medium with labeled nucleotides (e.g., ³H thymidine). The uptake or incorporation of the labeled nucleotides indicates DNA synthesis and cell 50 viability. In addition, colonies formed by cells cultured in medium indicate cell growth and is another means to test viability of the cells.

Identification and/or observation of cells undergoing apoptosis can be another method of determining cell viability. 55 Apoptosis is a specific mode of cell death recognized by a characteristic pattern of morphological, biochemical, and molecular changes. Cells going through apoptosis appear shrunken, and rounded; they also can be observed to become detached from culture dish. Thermophological changes 60 involve a characteristic pattern of condensation of chromatin and cytoplasm which can be readily identified by microscopy. When stained with a DNA-binding dye, such as H33258, apoptotic cells display classic condensed and punctate nuclei instead of homogeneous and round nuclei.

The hallmark of apoptosis is the endonucleolysis, a molecular change in which nuclear DNA is initially degraded 12

at the linker sections of nucleosomes to give rise to fragments equivalent to single and multiple nucleosomes. When these DNA fragments are subjected to gel electrophoresis, they reveal a series of DNA bands which are positioned approximately equally distant from each other on the gel. The size difference between the two bands next to each other is about the length of one nucleosome (i.e., 20 base pairs). This characteristic display of the DNA bands is called a DNA ladder and it indicates apoptosis of the cell. Apoptotic cells can be identified by flow cytometric methods based on measurement of cellular DNA content, increased sensitivity of DNA to denaturation, or altered light scattering properties. These methods are well known in the art and are within the contemplation of the invention.

Abnormal DNA breaks are also characteristic of apoptosis and can be detected by any means known in the art. In one embodiment, DNA breaks are labeled with biotinylated dUTP (b-dUTP). Cells are fixed and incubated in the presence of biotinylated dUTP with either exogenous terminal transferase (terminal DNA transferase assay; TdT assay) or DNA polymerase (nick translation assay; NT assay). The biotinylated dUTP is incorporated into the chromosome at the places where abnormal DNA breaks are repaired, and are detected with fluorescein conjugated to avidin under fluorescence

Kits

The present invention provides kits that can be used in the above screening methods. In one embodiment, a kit comprises a substantially isolated polypeptide comprising an OCT6 epitope which is specifically immunoreactive with only test compound(s) that are substrates of the OCT6 transporter protein. Binding of a test compound to the OCT6 epitope is indicative that the test compound is a OCT6 substrate. In another embodiment, a kit comprises a cell line that overexpresses an OCT6 transporter protein. Binding and/or cellular uptake of a test compound via the OCT6 protein is indicative that the test compound is a OCT6 substrate. Preferably, the kits of the present invention further comprise a control compound or antibody which does not react with the OCT6 transporter protein. In another specific embodiment, the kits of the present invention contain a means for detecting the binding of a test compound to an OCT6 epitope and/or cellular uptake of a test compound. For example, the test compound may be conjugated to a detectable substrate such as a fluorescent compound, an enzymatic substrate, a radioactive compound or a luminescent compound, or a second antibody which recognizes the first antibody may be conjugated to a detectable substrate.

The detectable substance may be coupled or conjugated either directly to the test compound (or fragment thereof) or indirectly, through an intermediate (such as, for example, a linker known in the art) using techniques known in the art. See, for example, U.S. Pat. No. 4,741,900 for metal ions which can be conjugated to antibodies for use as diagnostics according to the present invention. Further non-limiting examples of detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials, radioactive materials, positron emitting metals using various positron emission tomographies, nonradioactive paramagnetic metal ions, immunogenic tag peptide sequences, extrinsically activatable toxins, extrinsically activatable quenching agents, or antibodies. Non-limiting examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, beta-galactosidase, or acetylcholinesterase; examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/bi-

otin; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; an example of a luminescent material includes luminol; examples of bioluminescent materials 5 include luciferase, luciferin, and aequorin; and examples of suitable radioactive material include 125 I, 131 I, 111 In or 99 Tc.

Immunogenic Compositions

The present invention also provides immunogenic compositions for the treatment of hematological malignancies. Non-limiting exemplary hematological malignancies include, but are not limited to, Hodgkin's disease, leukemia such as, acute lymphoid (lymphocytic or lymphoblastic) leukemia (ALL), acute myeloid (myelogenous or myeloblastic) leukemia 15 (AML), acute lymphoid leukemia, biphenotypic (ALL, biphentoypic), acute undifferentiated leukemia (AUL), chronic myeloid (myelogenous or granulocytic) leukemia (CML), erythroleukemia, granuloxytic leukemia, lymphoma, monocytic leukemia, myleoma, myelomonocytic leukemia, 20 myelodysplastic syndromes, non-Hodgkin lymphoma, progranulocytic leukemia.

According to the invention immunogenic compositions for the treatment of hematological malignancies comprise a substrate recognized by an OCT6 transporter protein. Preferably, the substrate is a compound that binds selectively or specifically to a OCT6 transporter protein. In a preferred embodiment, the compound binds selectively to the OCT6 transporter protein encoded by a nucleotide sequence of SEQ ID NO:1. The compound may be a cytotoxin or coupled or conjugated with a cytoxic agent. Preferably the cytoxin or cytotoxic agent is a chemotherapeutic agent.

The present invention also provides pharmaceutical compositions. Such compositions comprise a therapeutically effective amount of a compound, and a pharmaceutically acceptable carrier.

Cell surface proteins like the OCT6 transporter can be utilized in antibody-based targeting strategies. In still another aspect of the invention, antibodies can be developed by known methods in the art against the external epitope of OCT6 transporter protein. In a preferred embodiment, antibodies are substrates of the OCT6 protein. The antibodies may be polyclonal antibodies or monoclonal antibodies.

Polyclonal antibodies to an antigen-of-interest can be produced by various procedures well known in the art. For example, a polypeptide of the invention can be administered to various host animals including, but not limited to, rabbits, mice, rats, etc. to induce the production of sera containing polyclonal antibodies specific for the antigen. Various adjuvants may be used to increase the immunological response, depending on the host species, and include but are not limited to, Freund's (complete and incomplete), mineral gels such as aluminum hydroxide, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanins, dinitrophenol, and potentially useful human adjuvants such as BCG (bacille Calmette-Guerin) and *corynebacterium parvum*. Such adjuvants are also well known in the art.

Monoclonal antibodies can be prepared using a wide variety of techniques known in the art including the use of hybridoma, recombinant, and phage display technologies, or a combination thereof. For example, monoclonal antibodies can be produced using hybridoma techniques including those known in the art and taught, for example, in Harlow et al., 65 Antibodies: A Laboratory Manual, (Cold Spring Harbor Laboratory Press, 2nd ed. 1988); Hammerling, et al., in:

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Monoclonal Antibodies and T-Cell Hybridomas 563-681 (Elsevier, N.Y., 1981) (said references incorporated by reference in their entireties).

The present invention further encompasses antibodies or fragments thereof conjugated to a diagnostic or therapeutic agent. The antibodies can be used diagnostically to, for example, monitor the development or progression of a tumor as part of a clinical testing procedure to, e.g., determine the efficacy of a given treatment regimen. Detection can be facilitated by coupling the antibody to a detectable substance. Examples of detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials, radioactive materials, positron emitting metals using various positron emission tomographies, and nonradioactive paramagnetic metal ions. The detectable substance may be coupled or conjugated either directly to the antibody (or fragment thereof) or indirectly, through an intermediate, such as, for example, a linker known in the art, using techniques known in the art. (See, for example, U.S. Pat. No. 4,741,900 for metal ions which can be conjugated to antibodies for use as diagnostics according to the present invention.) Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, beta-galactosidase, or acetylcholinesterase; examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/biotin; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; an example of a luminescent material includes luminol; examples of bioluminescent materials include luciferase, luciferin, and aequorin; and examples of suitable radioactive material include ¹²⁵I, ¹³¹I, ¹¹¹In or ⁹⁹Tc.

Further, an antibody or fragment thereof may be conjugated to a therapeutic moiety such as a cytotoxin, e.g., a cytostatic or cytocidal agent, a therapeutic agent or a radioactive metal ion, e.g., alpha-emitters such as, for example, ²¹³Bi. Non-limiting examples include paclitaxol, cytochalasin B, gramicidin D, ethidium bromide, emetine, mitomycin, etoposide, tenoposide, vincristine, vinblastine, colchicin, doxorubicin, daunorubicin, dihydroxy anthracin dione, mitoxantrone, mithramycin, actinomycin D, 1-dehydrotestosterone, glucocorticoids, procaine, tetracaine, lidocaine, propranolol, and puromycin and analogs or homologs thereof. Therapeutic agents include, but are not limited to, antimetabolites (e.g., methotrexate, 6-mercaptopurine, 6-thioguanine, cytarabine, 5-fluorouracil decarbazine), alkylating agents (e.g., mechlorethamine, thioepa chlorambucil, melphalan, carmustine (BSNU) and lomustine (CCNU), cyclothosphamide, busulfan, dibromomannitol, streptozotocin, mitomycin C, and cis-dichlorodiamine platinum (II) (DDP) cisplatin), anthracyclines (e.g., daunorubicin (formerly daunomycin) and doxorubicin), antibiotics (e.g., dactinomycin (formerly actinomycin), bleomycin, mithramycin, and anthramycin (AMC)), and anti-mitotic agents (e.g., vincristine and vinblastine).

The conjugates of the invention can be used for modifying a given biological response such as inducing cell death for the treatment and prevention of hematological malignancies like leukemia. The therapeutic agent or drug moiety is not to be construed as limited to classical chemical therapeutic agents. For example, the drug moiety may be a protein or polypeptide possessing a desired biological activity for inducing cell death. Such proteins may include, for example, a toxin such as abrin, ricin A, pseudomonas exotoxin, or diphtheria toxin; a protein such as tumor necrosis factor, a-interferon, beta-interferon, nerve growth factor, platelet derived growth factor, tissue plasminogen activator, an apoptotic agent, e.g.,

TNF-alpha, TNF-beta, AIM I (See, International Publication No. WO 97/33899), AIM II (See, International Publication No. WO 97/34911), Fas Ligand (Takahashi et al., Int. Immunol., 6:1567-1574 (1994)), VEGI (See, International Publication No. WO 99/23105), a thrombotic agent or an anti-angiogenic agent, e.g., angiostatin or endostatin; or, biological response modifiers such as, for example, lymphokines, interleukin-1 ("IL-1"), interleukin-2 ("IL-2"), interleukin-6 ("IL-6"), granulocyte macrophage colony stimulating factor ("GM-CSF"), granulocyte colony stimulating factor ("G-10 CSF"), or other growth factors.

Therapeutic Treatment

The present invention is further directed to methods for preventing and treating hematological malignancies such as leukemia. According to the invention, hematological malignancies comprise without limitation, Hodgkin's disease, leukemia such as, acute lymphoid (lymphocytic or lymphoblastic) leukemia (ALL), acute myeloid (myelogenous or myeloblastic) leukemia (AML), acute lymphoid leukemia, biphenotypic (ALL, biphentoypic), acute undifferentiated leukemia (AUL), chronic myeloid (myelogenous or granulocytic) leukemia (CML), erythroleukemia, granuloxytic leukemia, lymphoma, monocytic leukemia, myleoma, myelomonocytic leukemia, myelodysplastic syndromes, non-Hodgkin lymphoma, progranulocytic leukemia.

Methods of treatment of the present invention comprise administering to a subject in need thereof an immunogenic composition of the present invention. The compositions may be administered with a pharmaceutically acceptable carrier.

Such pharmaceutical carriers can be sterile liquids, such as 30 water and oils, including those of petroleum, animal, vegetable or synthetic origin, such as peanut oil, soybean oil, mineral oil, sesame oil and the like. Water is a preferred carrier when the pharmaceutical composition is administered intravenously. Saline solutions and aqueous dextrose and glycerol solutions can also be employed as liquid carriers, particularly for injectable solutions. Suitable pharmaceutical excipients include starch, glucose, lactose, sucrose, gelatin, malt, rice, flour, chalk, silica gel, sodium stearate, glycerol monostearate, talc, sodium chloride, dried skim milk, glycerol, propylene, glycol, water, ethanol and the like. The com- 40 position, if desired, can also contain minor amounts of wetting or emulsifying agents, or pH buffering agents. These compositions can take the form of solutions, suspensions, emulsion, tablets, pills, capsules, powders, sustained-release formulations and the like. The composition can be formulated 45 as a suppository, with traditional binders and carriers such as triglycerides. Oral formulation can include standard carriers such as pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate. etc. Examples of suitable pharmaceutical 50 carriers are described in "Remington's Pharmaceutical Sciences" by E. W. Martin. Such compositions will contain a therapeutically effective amount of the compound, preferably in purified form, together with a suitable amount of carrier so as to provide the form for proper administration to the patient. The formulation should suit the mode of administration.

In a preferred embodiment, the composition is formulated in accordance with routine procedures as a pharmaceutical composition adapted for intravenous administration to human beings. Typically, compositions for intravenous administration are solutions in sterile isotonic aqueous buffer. Where necessary, the composition may also include a solubilizing agent and a local anesthetic such as lignocaine to ease pain at the site of the injection. Generally, the ingredients are supplied either separately or mixed together in unit dosage form, for example, as a dry lyophilized powder or water free concentrate in a hermetically sealed container such as an ampoule or sachette indicating the quantity of active agent.

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Where the composition is to be administered by infusion, it can be dispensed with an infusion bottle containing sterile pharmaceutical grade water or saline. Where the composition is administered by injection, an ampoule of sterile water for injection or saline can be provided so that the ingredients may be mixed prior to administration.

The compounds of the invention can be formulated as neutral or salt forms. Pharmaceutically acceptable salts include those formed with anions such as those derived from hydrochloric, phosphoric, acetic, oxalic, tartaric acids, etc., and those formed with cations such as those derived from sodium, potassium, ammonium, calcium, ferric hydroxides, isopropylamine, triethylamine, 2-ethylamino ethanol, histidine, procaine, etc.

The amount of the compound of the invention which will be effective in the treatment, inhibition and prevention of hematological malignancies can be determined by standard clinical techniques. In addition, in vitro assays may optionally be employed to help identify optimal dosage ranges. The precise dose to be employed in the formulation will also depend on the route of administration, and the seriousness of the disease or disorder, and should be decided according to the judgment of the practitioner and each patient's circumstances. Effective doses may be extrapolated from dose-response curves derived from in vitro or animal model test systems.

Various other delivery systems are known and can be used to administer a composition of the invention, e.g., encapsulation in liposomes, microparticles, microcapsules, recombinant cells capable of expressing the compound, receptormediated endocytosis (See, e.g., Wu and Wu, J. Biol. Chem. 262:4429-4432 (1987)), construction of a nucleic acid as part of a retroviral or other vector, etc. Methods of introduction include but are not limited to intradermal, intramuscular, intraperitoneal, intravenous, subcutaneous, intranasal, epidural, and oral routes. The compounds or compositions may be administered by any convenient route, for example by infusion or bolus injection, by absorption through epithelial or mucocutaneous linings (e.g., oral mucosa, rectal and intestinal mucosa, etc.) and may be administered together with other biologically active agents. Administration can be systemic or local. In addition, it may be desirable to introduce the pharmaceutical compounds or compositions of the invention into the central nervous system by any suitable route, including intraventricular and intrathecal injection; intraventricular injection may be facilitated by an intraventricular catheter, for example, attached to a reservoir, such as an Ommaya reservoir. Pulmonary administration can also be employed, e.g., by use of an inhaler or nebulizer, and formulation with an aerosolizing agent.

In a specific embodiment, it may be desirable to administer the pharmaceutical compounds or compositions of the invention locally to the area in need of treatment; this may be achieved by, for example, and not by way of limitation, local infusion during surgery, topical application, e.g., in conjunction with a wound dressing after surgery, by injection, by means of a catheter, by means of a suppository, or by means of an implant, said implant being of a porous, non-porous, or gelatinous material, including membranes, such as sialastic membranes, or fibers. Preferably, when administering a protein, including an antibody, of the invention, care must be taken to use materials to which the protein does not absorb.

In another embodiment, the compound or composition can be delivered in a vesicle, in particular a liposome (See Langer, Science 249:1527-1533 (1990); Treat et al., in Liposomes in the Therapy of Infectious Disease and Cancer, Lopez-Berestein and Fidler (eds.), Liss, New York, pp. 353-365 (1989); Lopez-Berestein, ibid., pp. 317-327; see generally ibid.)

In yet another embodiment, the compound or composition can be delivered in a controlled release system. In one

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embodiment, a pump may be used (see Langer, supra; Sefton, CRC Crit. Ref. Biomed. Eng. 14:201 (1987); Buchwald et al., Surgery 88:507 (1980); Saudek et al., N. Engl. J. Med. 321: 574 (1989)). In another embodiment, polymeric materials can be used (see Medical Applications of Controlled Release, Langer and Wise (eds.), CRC Pres., Boca Raton, Fla. (1974); Controlled Drug Bioavailability, Drug Product Design and Performance, Smolen and Ball (eds.), Wiley, New York (1984); Ranger and Peppas, J., Macromol. Sci. Rev. Macromol. Chem. 23:61 (1983); see also Levy et al., Science 228: 190 (1985); During et al., Ann. Neurol. 25:351 (1989); Howard et al., J. Neurosurg. 71:105 (1989)). In yet another embodiment, a controlled release system can be placed in proximity of the therapeutic target, i.e., the brain, thus requiring only a fraction of the systemic dose (see, e.g., Goodson, in Medical Applications of Controlled Release, supra, vol. 2, pp. 115-138 (1984)).

In a specific embodiment where the compound of the invention is a nucleic acid encoding a protein, the nucleic acid can be administered in vivo to promote expression of its encoded protein, by constructing it as part of an appropriate nucleic acid expression vector and administering it so that it becomes intracellular, e.g., by use of a retroviral vector (see U.S. Pat. No. 4,980,286), or by direct injection, or by use of microparticle bombardment (e.g., a gene gun; Biolistic, Dupont), or coating with lipids or cell-surface receptors or transfecting agents, or by administering it in linkage to a homeobox-like peptide which is known to enter the nucleus (see e.g., Joliot et al., Proc. Natl. Acad. Sci. USA 88:1864-1868 (1991)), etc. Alternatively, a nucleic acid can be introduced intracellularly and incorporated within host cell DNA for expression, by homologous recombination.

EXAMPLES

The following examples are presented for the illustrative 35 purposes and it is to be understood that the present invention is not limited to those precise embodiments, and that various changes and modifications can be effected therein by one skilled in the art without departing from the scope and spirit of the invention as defined by the appended claims.

Example 1

OCT6 Nucleotide Sequence Identification and Analysis

OCT6 was first identified as a potential OCT gene by assembling and sequencing ESTs. BLAST searches of human ESTs in GenBank data base identified AI040384 (654 bp), AA033971 (714 bp) and H70190 (474 bp) sequences 50 from three fetal liver IMAGE clones, 1656502, 429904 and 212935 respectively. IMAGE clone 1656502 (3', insert 1337 bp) ended the predicted 3' stop codon, whereas IMAGE clone 429904 (5', insert 996 bp) and IMAGE clone 212935 (5', insert 966 bp) aligned with the 5'-coding region. All clones were obtained from the IMAGE Consortium through the American Type Culture Collection (Manassas, Va.). Each clone was sequenced in both directions. The sequences were determined using ABI PrismTM 377 DNA sequencer (Perkin-Elmer). Our assemblage proved to be identical to the recently submitted cDNA OKB1 (AF268892) submitted by M. Okabe 60 and T. Abe. We have dubbed this gene OCT6 as OCTN1 and OCTN2 may be considered as OCT4 and OCT5 respectively.

The OCT6 gene (SEQ ID NO:1) is also contained within BAC clone CTA-331P3 (SEQ ID NO:3) (GenBank AC002464) located at chromosome 6q21. It is divided into 6 exons that span 42 kb on the human genome, from nucleotide 79,570 to nucleotide 120490 on CTA-331P3.

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The gene has a predicted protein structure typical of transport proteins with 2 groups of 6 transmembrane domains separated by a hydrophilic region (FIG. 1A). The large hydrophilic region between TMD1 and TMD2 is typical of OCT and OAT genes and is presumed to be located on the outside surface of the cell membrane. The OCT6 protein contains potential sites for N-glycosylation and phosphorylation, which will be described below in Methods. Of interest, the protein sequence also contains a 22 amino acid leucine zipper motif, starting at amino acid 146, suggesting that there may be a physical interaction between OCT6 and ion channels or other membrane-associated proteins.

CLUSTALW alignment produced a dendrogram showing the phylogenic relationship between OCT6 and other OAT and OCT proteins (FIG. 1B). This dendrogram suggests that the distinction between OAT and OCT genes, based on functional studies, obscures the common origin of both families of transporters. The actual CLUSTALW alignment of these genes is shown in FIG. 2 and demonstrates multiple regions of conservation among all of these genes.

The hydropathy profile analysis, multiple sequence alignments of amino acid sequences using CLUSTALW and the phylogenetic tree were all produced with MacVector software.

Example 2

Molecular Cloning of OCT6

BLAST searches of human ESTs in GenBank data base identified AI040384 (654 bp), AA033971 (714 bp) and H70190 (474 bp) sequences from three fetal liver IMAGE clones, 1656502, 429904 and 212935 respectively. IMAGE clone 1656502 (3', insert 1337 bp) ended the predicted 3' stop codon, whereas IMAGE clone 429904 (5', insert 966 bp) and IMAGE clone 212935 (5', insert 966 bp) aligned with the 5'-coding region. All clones were obtained from the IMAGE Consortium through the American Type Culture Collection (Manassas, Va.). Each clone was sequenced in both directions. The sequences were determined using ABI PrismTM 377 DNA sequencer (Perkin-Elmer).

Example 3

Quantitative RT-PCR of OCT6 RNA Levels in Cancer Cell Lines

Total RNA isolated from 50 cell lines used in the NCI drug screen program was provided by the Developmental Therapeutics Program, NCI. Quantitative RT-PCR for detecting OAT-X transporter gene expression was performed by using a Roche LightCycler, which uses real time fluorescence detection for quantitative measurement of PCR products. A genespecific primer pair was designed with Oligo 4.0 software and purchased from Integrated DNA Technologies, Inc. (Coralville, Iowa) (F: 5'-GGCACATTTATTCACCAAGACCAG-3') (SEQ ID NO: 13) and (F: 5'-TGTGGACCTCAGCAG-CATTTGGAT-3') (SEQ ID NO:14). The specificity of the PCR reaction was confirmed by directly determining the DNA sequence of the PCR product. First, cDNA was synthesized from total RNA using SuperScript First-Strand Synthesis System (GIBCO/BRL) in a 20 µl volume following the instructions supplied by the manufacturer. The cDNA treated with RNAse H for 20 minutes at 37° C. and stored at -20° C. Then, 2 ul of cDNA reaction was amplified in a standard PCR reaction condition, using 0.3 µM primer concentration, with the addition of SYBR Green I Dye. After 30 seconds denature at 95° C., the amplification reaction proceeded through 45-50 cycles of 95° C. denature for 0 second, 62-65° C. annealing

for 10 seconds and a 72° C. extension for 40 seconds, with slopes of 20° C./s, 20° C./s and 2° C./s, respectively.

Fluorescence was acquired during each cycle after heating to a temperature just below the product melting temperature. Quantification was performed using the LightCycler analysis software. The log-linear portion of the standard amplification curve was identified, and the 'crossing point', a threshold of relative fluorescence, was determined as the best fit through the log-linear region above the background fluorescence (noise) band. The quantification of PCR product then was derived by plotting fluorescence data in the log linear region of each sample to determine a calculated number of cycles needed to reach the fluorescence crossing point. The calculated number of cycles required to reach the crossing point is proportional to the amount of target RNA in the sample. The relative amount of product was described in arbitrary units by interpolation of the data using a standard curve of a series of dilutions of a standard cell line RNA. The quantitative measurement of each gene in each cell line was normalized to the relative amount of actin RNA in each cell line, as a control for equivalent cDNA loading in each sample. The results repre- 20 sent the average of 3 independent determinations performed in duplicate.

A melting curve analysis was performed with positive control RNA prior analysis of the cell lines to enhance sensitivity and the specificity of the data. Amplified products usually melt quickly at a temperature characteristic for the products. The fluorescence signal was acquired at a temperature just below the Tm of the specific PCR product and above the Tm of the primer dimers. All specific PCR product displayed a single, sharply melting curve with a narrow peak. In addition, PCR products were confirmed for specificity and correct size by visualization of the LightCycler products on a 1% agarose gel.

Example 4

Tissue Distribution

First strand cDNAs derived from 24 adult and fetal tissues (RAPID-SCAN gene expression panel, OriGene Technologies, Rockville, Md.). The PCR primers used in this study

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were the same as used in the quantitative RT-PCR studies. The PCR reaction samples were denatured at 94° C. for 30 seconds, annealed and extended at 64° C. for 30 sec for 35 cycles. The PCR products were then visualized on 1% agarose gels.

Example 5

Cell Sorting

All human specimens were obtained in accordance with institutional IRB guidelines. Leukocytes from fresh discarded buffy coats were isolated after RBC lysis with ammonium chloride and labeled with lineage specific antibodies (CD14, monocytes; CD15, granulocytes; CD3, T-cells; and CD20, B-cells), and isolated using a FACSVantage flow cytometer. Each population was sorted twice to ensure purities of at least 99%. CD34 cells were obtained from discarded aliquots of G-CSF-mobilized peripheral blood stem cell collections from cancer patients. For each sample, the PCR results represent the pooled average of cells from 2 individuals performed in triplicate or quadruplicate.

Example 6

OCT6 RNA Levels in Leukemic Blasts

Total RNA was extracted from leukemia specimens using QIAGEN RNeasy midi kit. 150 ng of total RNA were used as a template for the first strand cDNA synthesis with the Oligo (dT) primer using the super script system (GIBCO BRL) according to the manufacturer's protocol. Quantitative real-time RT-PCR was performed using an iCycler thermal cycler with methods similar to those described above for the Roche LightCycler. The results represent the average of 3 independent determination performed in duplicate.

Although illustrative embodiments of the present invention have been described in detail, it is to be understood that the present invention is not limited to those precise embodiments, and that various changes and modifications can be effected therein by one skilled in the art without departing from the scope and spirit of the invention as defined by the appended claims.

SEQUENCE LISTING

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<400> SEQUENCE: 1
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agagteetet attteatatg tgeetteeag aacatetett gtggtattea etaettgget
                                                                      120
tetgtgttca tgggagtcac ccctcatcat gtctgcaggc ccccaggcaa tgtgagtcag
                                                                      180
gttgttttcc ataatcactc taattggagt ttggaggaca ccggggccct gttgtcttca
                                                                      240
ggccagaaag attatgttac ggtgcagttg cagaatggtg agatctggga gctctcaagg
                                                                      300
tgtagcagga ataagaggga gaacacatcg agtttgggct atgaatacac tggcagtaag
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Concinaca										
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Leu Glu Pro Gly Arg Asp Val Asp Leu Gly Gln Leu Glu Gln Glu Ser

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Arg	Met	Leu	Leu 260	Val	Ala	Leu	Thr	Met 265	Pro	Gly	Val	Leu	Cys 270	Val	Ala
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Arg	Phe 290	Glu	Glu	Ala	Glu	Val 295	Ile	Ile	Arg	Lys	Ala 300	Ala	Lys	Ala	Asn
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Leu	Ser	Ser	Lys	Lys 325	Gln	Gln	Ser	His	Asn 330	Ile	Leu	Asp	Leu	Leu 335	Arg
Thr	Trp	Asn	Ile 340	Arg	Met	Val	Thr	Ile 345	Met	Ser	Ile	Met	Leu 350	Trp	Met
Thr	Ile	Ser 355	Val	Gly	Tyr	Phe	Gly 360	Leu	Ser	Leu	Asp	Thr 365	Pro	Asn	Leu
His	Gly 370		Ile	Phe	Val	Asn 375	Cys	Phe	Leu	Ser	Ala 380	Met	Val	Glu	Val
Pro 385	Ala	Tyr	Val	Leu	Ala 390	Trp	Leu	Leu	Leu	Gln 395	Tyr	Leu	Pro	Arg	Arg 400
Tyr	Ser	Met	Ala	Thr 405	Ala	Leu	Phe	Leu	Gly 410	Gly	Ser	Val	Leu	Leu 415	Phe
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Met	Val	Gly 435	Lys	Phe	Gly	Val	Thr 440	Ala	Ala	Phe	Ser	Met 445	Val	Tyr	Val
Tyr	Thr 450	Ala	Glu	Leu	Tyr	Pro 455	Thr	Val	Val	Arg	Asn 460	Met	Gly	Val	Gly
Val 465	Ser	Ser	Thr	Ala	Ser 470	Arg	Leu	Gly	Ser	Ile 475	Leu	Ser	Pro	Tyr	Phe 480
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Ser	Leu	Thr	Ile 500	Leu	Thr	Ala	Ile	Leu 505	Thr	Leu	Phe	Leu	Pro 510	Glu	Ser
Phe	Gly	Thr 515	Pro	Leu	Pro	Asp	Thr 520	Ile	Asp	Gln	Met	Leu 525	Arg	Val	Lys
Gly	Met 530		His	Arg	Lys	Thr 535	Pro	Ser	His	Thr	Arg 540	Met	Leu	Lys	Asp
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Phe	Ala	Pro 35	Ile	Tyr	Val	Gly	Ile 40	Val	Phe	Leu	Gly	Phe 45	Thr	Pro	Asp
His	Arg 50	Сув	Arg	Ser	Pro	Gly 55	Val	Ala	Glu	Leu	Ser 60	Leu	Arg	Сув	Gly
Trp 65	Ser	Pro	Ala	Glu	Glu 70	Leu	Asn	Tyr	Thr	Val 75	Pro	Gly	Pro	Gly	Pro 80
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Asn	Gln	Ser	Thr 100	Phe	Asp	CÀa	Val	Asp 105	Pro	Leu	Ala	Ser	Leu 110	Asp	Thr
Asn	Arg	Ser 115	Arg	Leu	Pro	Leu	Gly 120	Pro	Cys	Arg	Asp	Gly 125	Trp	Val	Tyr
Glu	Thr 130	Pro	Gly	Ser	Ser	Ile 135	Val	Thr	Glu	Phe	Asn 140	Leu	Val	СЛв	Ala
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Lys	Leu	Сув	Leu 180	Leu	Thr	Thr	Val	Leu 185	Ile	Asn	Ala	Ala	Ala 190	Gly	Val
Leu	Met	Ala 195	Ile	Ser	Pro	Thr	Tyr 200	Thr	Trp	Met	Leu	Ile 205	Phe	Arg	Leu
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Ile 225	Thr	Glu	Phe	Val	Gly 230	Arg	Arg	Tyr	Arg	Arg 235	Thr	Val	Gly	Ile	Phe 240
Tyr	Gln	Val	Ala	Tyr 245	Thr	Val	Gly	Leu	Leu 250	Val	Leu	Ala	Gly	Val 255	Ala
Tyr	Ala	Leu	Pro 260	His	Trp	Arg	Trp	Leu 265	Gln	Phe	Thr	Val	Ala 270	Leu	Pro
Asn	Phe	Phe 275	Phe	Leu	Leu	Tyr	Tyr 280	Trp	Cys	Ile	Pro	Glu 285	Ser	Pro	Arg
Trp	Leu 290	Ile	Ser	Gln	Asn	Lys 295	Asn	Ala	Glu	Ala	Met 300	Arg	Ile	Ile	ГЛЗ
His 305	Ile	Ala	ГÀв	ГÀЗ	Asn 310	Gly	Lys	Ser	Leu	Pro 315	Ala	Ser	Leu	Gln	Arg 320
Leu	Arg	Leu	Glu	Glu 325	Glu	Thr	Gly	Lys	330 TÀa	Leu	Asn	Pro	Ser	Phe 335	Leu
Asp	Leu	Val	Arg 340	Thr	Pro	Gln	Ile	Arg 345	Lys	His	Thr	Met	Ile 350	Leu	Met
Tyr	Asn	Trp 355	Phe	Thr	Ser	Ser	Val 360	Leu	Tyr	Gln	Gly	Leu 365	Ile	Met	His
Met	Gly 370	Leu	Ala	Gly	Asp	Asn 375	Ile	Tyr	Leu	Asp	Phe 380	Phe	Tyr	Ser	Ala
Leu 385	Val	Glu	Phe	Pro	Ala 390	Ala	Phe	Met	Ile	Ile 395	Leu	Thr	Ile	Asp	Arg 400
Ile	Gly	Arg	Arg	Tyr 405	Pro	Trp	Ala	Ala	Ser 410	Asn	Met	Val	Ala	Gly 415	Ala
Ala	Cys	Leu	Ala 420	Ser	Val	Phe	Ile	Pro 425	Gly	Asp	Leu	Gln	Trp 430	Leu	Lys

The Pro Phe Leu Val Tyr Arg Leu The Assn Ile Trp Leu Glu Leu Val Soo Store Sto																
450	Ile	Ile		Ser	CAa	Leu	Gly	_	Met	Gly	Ile	Thr		Ala	Tyr	Glu
## 470 ## 475 ## 475 ## 475 ## 485 ##	Ile		CÀa	Leu	Val	Asn		Glu	Leu	Tyr	Pro		Phe	Ile	Arg	Asn
Leu Met Val Phe Gly Val Leu Gly Leu Val Ala Gly Gly Leu Val I Soo Soo Soo Soo Soo Soo Soo Soo Soo S		Gly	Val	His	Ile		Ser	Ser	Met	Càa		Ile	Gly	Gly	Ile	Ile 480
Leu Leu Pro Glu Thr Lys Gly Lys Ala Leu Pro Glu Thr Ile Glu Control Sis	Thr	Pro	Phe	Leu		Tyr	Arg	Leu	Thr		Ile	Trp	Leu	Glu		Pro
S15 S20 S25	Leu	Met	Val		Gly	Val	Leu	Gly		Val	Ala	Gly	Gly		Val	Leu
Leu Gln Val Gln Lys Leu Asp Ile Pro Leu Asn 5555	Leu	Leu		Glu	Thr	Lys	Gly		Ala	Leu	Pro	Glu		Ile	Glu	Glu
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Met Pro Thr Val Asp Asp Ile Leu Glu Gln Val Gly Glu Ser Gly Ile Leu Glu Val Gly Ile Leu Cys Leu Leu Ser Ala Ala Ile Leu Leu Cys Leu Leu Ser Ala Ala Ile Leu Leu Gly Phe Ala Leu Gly Phe Ala Leu Gly Phe Ala Ala <td></td> <td></td> <td></td> <td></td> <td>Homo</td> <td>sa]</td> <td>piens</td> <td>3</td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td>					Homo	sa]	piens	3								
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His Cys Gln Ser Pro Gly Val Ala Glu Leu Ser Gln Arg Cys Gly Ser Pro Ala Glu Glu Leu Asn Tyr Thr Val Pro Gly Leu Gly Pro Ala Glu Ala Phe Leu Gly Gln Cys Arg Arg Arg Tyr Glu Val Asp Trp Ala Glu Ser His Leu Pro Leu Gly Pro Cys Gln Asp Gly Tilo Thr Pro Gly Ser Ser Ile Val Thr Glu Phe Asn Leu Val Cys Ala Ala Ser Trp Lys Leu Asp Leu Phe Gln Ser Cys Ual Asp Trp Ala Ser Trp Lys Leu Asp Leu Phe Gln Ser Cys Leu Ala Asp Arg	Phe	Gln	ГЛа		Ala	Phe	Leu	Ile		Cya	Leu	Leu	Ser		Ala	Phe
Ser Pro Ala Glu Glu Leu Asn Tyr Thr Val Pro Gly Leu Gly Pro 28 Gly Glu Ala Phe Leu Gly Glu Cys Arg Arg Arg Tyr Glu Val Asp Tyr Arg Arg Tyr Glu Val Asp Pro Leu Ala Ser Leu Ala Pro 28 Arg Arg Arg Tyr Glu Val Asp Pro Leu Ala Ser Leu Ala Pro 28 Val Asp Pro Leu Ala Ser Leu Ala Pro Ala Asp Pro Ala Asp Intraction In	Ala	Pro		Cys	Val	Gly	Ile		Phe	Leu	Gly	Phe		Pro	Asp	His
65	His		Gln	Ser	Pro	Gly		Ala	Glu	Leu	Ser		Arg	Cya	Gly	Trp
Ser Ala Leu Ser Cys Val Asp Pro Leu Ala Ser Leu Ala Thr Ala Asp Pro Leu Ala Ser Leu Ala Thr Ala Asp Pro Leu Ala Asp		Pro	Ala	Glu	Glu		Asn	Tyr	Thr	Val		Gly	Leu	Gly	Pro	Ala 80
Arg Ser His Leu Pro Leu Gly Pro Cys Gln Asp Gly Trp Val Tyr A 125 Thr Pro Gly Ser Ser IIe Val Thr Glu Phe Asn Leu Val Cys Ala Asp Gly Trp Val Tyr A 125 Ser Trp Lys Leu Asp Leu Phe Gln Ser Cys Leu Asn Ala Gly Phe I 155 Phe Gly Ser Leu Gly Val Gly Tyr Phe Ala Asp Arg Phe Gly Arg I 175 Leu Cys Leu Leu Gly Thr Val Leu Val Asn Ala Val Ser Gly Val I 185 Met Ala Phe Ser Pro Asn Tyr Met Ser Met Leu Leu Phe Arg Leu I 195 Gln Gly Leu Val Ser Lys Gly Asn Trp Met Ala Gly Tyr Thr Leu I 205 Thr Glu Phe Val Gly Ser Gly Ser Arg Arg Thr Val Ala Ile Met I 225 Gln Met Ala Phe Thr Val Gly Leu Val Ala Leu Thr Gly Leu Ala I	Gly	Glu	Ala	Phe		Gly	Gln	Cys	Arg	_	Tyr	Glu	Val	Asp	_	Asn
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130	Arg	Ser		Leu	Pro	Leu	Gly		Cha	Gln	Asp	Gly		Val	Tyr	Asp
145 150 155 155 157 155 157 155 1	Thr		Gly	Ser	Ser	Ile		Thr	Glu	Phe	Asn		Val	Cys	Ala	Asp
165		Trp	Lys	Leu	Asp		Phe	Gln	Ser	Cys		Asn	Ala	Gly	Phe	Leu 160
Met Ala Phe Ser Pro Asn Tyr Met 200 Ser Met Leu Leu Phe 205 Pro Arg Leu I 205 Gln Gly Leu Val Ser Lys 210 Gly Asn Trp Met Ala Gly Tyr Thr Leu 220 Thr Glu Phe Val Gly Ser 230 Ser Arg Arg Thr Val Ala Ile Met 235 Gln Met Ala Phe Thr Val Gly Leu Val Ala Leu Thr Gly Leu Ala 250	Phe	Gly	Ser	Leu		Val	Gly	Tyr	Phe		Asp	Arg	Phe	Gly		Lys
195 200 205 Gln Gly Leu Val Ser Lys Gly Asn Trp Met Ala Gly Tyr Thr Leu 1 210 Thr Glu Phe Val Gly Ser Gly Ser Arg Arg Thr Val Ala Ile Met 2 225 Gln Met Ala Phe Thr Val Gly Leu Val Ala Leu Thr Gly Leu Ala 3	Leu	Сув	Leu		Gly	Thr	Val	Leu		Asn	Ala	Val	Ser		Val	Leu
210 215 220 Thr Glu Phe Val Gly Ser Gly Ser Arg Arg Thr Val Ala Ile Met 225 230 235 Gln Met Ala Phe Thr Val Gly Leu Val Ala Leu Thr Gly Leu Ala 2	Met	Ala		Ser	Pro	Asn	Tyr		Ser	Met	Leu	Leu		Arg	Leu	Leu
225 230 235 2 Gln Met Ala Phe Thr Val Gly Leu Val Ala Leu Thr Gly Leu Ala S	Gln	_	Leu	Val	Ser	Lys	_	Asn	Trp	Met	Ala	_	Tyr	Thr	Leu	Ile
		Glu	Phe	Val	Gly		Gly	Ser	Arg	Arg		Val	Ala	Ile	Met	Tyr 240
	Gln	Met	Ala	Phe		Val	Gly	Leu	Val		Leu	Thr	Gly	Leu		Tyr

Ala	Leu	Pro	His 260	Trp	Arg	Trp	Leu	Gln 265	Leu	Ala	Val	Ser	Leu 270	Pro	Thr
Phe	Leu	Phe 275	Leu	Leu	Tyr	Tyr	Trp 280	CAa	Val	Pro	Glu	Ser 285	Pro	Arg	Trp
Leu	Leu 290	Ser	Gln	Lys	Arg	Asn 295	Thr	Glu	Ala	Ile	300 Tàs	Ile	Met	Asp	His
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Ser	Leu	Glu	Glu	Asp 325	Val	Thr	Glu	Lys	Leu 330	Ser	Pro	Ser	Phe	Ala 335	Asp
Leu	Phe	Arg	Thr 340	Pro	Arg	Leu	Arg	Lys 345	Arg	Thr	Phe	Ile	Leu 350	Met	Tyr
Leu	Trp	Phe 355	Thr	Asp	Ser	Val	Leu 360	Tyr	Gln	Gly	Leu	Ile 365	Leu	His	Met
Gly	Ala 370	Thr	Ser	Gly	Asn	Leu 375	Tyr	Leu	Asp	Phe	Leu 380	Tyr	Ser	Ala	Leu
Val 385	Glu	Ile	Pro	Gly	Ala 390	Phe	Ile	Ala	Leu	Ile 395	Thr	Ile	Asp	Arg	Val 400
Gly	Arg	Ile	Tyr	Pro 405	Met	Ala	Met	Ser	Asn 410	Leu	Leu	Ala	Gly	Ala 415	Ala
СЛа	Leu	Val	Met 420	Ile	Phe	Ile	Ser	Pro 425	Asp	Leu	His	Trp	Leu 430	Asn	Ile
Ile	Ile	Met 435	Cys	Val	Gly	Arg	Met 440	Gly	Ile	Thr	Ile	Ala 445	Ile	Gln	Met
Ile	Сув 450	Leu	Val	Asn	Ala	Glu 455	Leu	Tyr	Pro	Thr	Phe 460	Val	Arg	Asn	Leu
Gly 465	Val	Met	Val	CÀa	Ser 470	Ser	Leu	CAa	Asp	Ile 475	Gly	Gly	Ile	Ile	Thr 480
Pro	Phe	Ile	Val	Phe 485	Arg	Leu	Arg	Glu	Val 490	Trp	Gln	Ala	Leu	Pro 495	Leu
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Leu	Pro	Glu 515	Thr	Lys	Gly	Val	Ala 520	Leu	Pro	Glu	Thr	Met 525	Lys	Asp	Ala
Glu	Asn 530	Leu	Gly	Arg	ГÀз	Ala 535	ГÀа	Pro	Lys	Glu	Asn 540	Thr	Ile	Tyr	Leu
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Pro	His	Ile 35	Leu	Leu	Glu	Asn	Phe 40	Ala	Ala	Ala	Ile	Pro 45	Gly	His	Arg
CAa	Trp 50	Val	His	Met	Leu	Asp 55	Asn	Asn	Thr	Gly	Ser 60	Gly	Asn	Glu	Thr
Gly	Ile	Leu	Ser	Glu	Asp	Ala	Leu	Leu	Arg	Ile	Ser	Ile	Pro	Leu	Asp

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65					70					75					80
Ser	Asn	Leu	Arg	Pro 85	Glu	ГÀа	CAa	Arg	Phe 90	Phe	Val	His	Pro	Gln 95	Trp
Gln	Leu	Leu	His 100	Leu	Asn	Gly	Ile	His 105	Ser	Thr	Ser	Glu	Ala 110	Asp	Thr
Glu	Pro	Cys 115	Val	Asp	Gly	Trp	Val 120	Tyr	Asp	Gln	Ser	Tyr 125	Phe	Pro	Ser
Thr	Ile 130	Val	Thr	Lys	Trp	Asp 135	Leu	Val	Cys	Asp	Tyr 140	Gln	Ser	Leu	Lys
Ser 145	Val	Val	Gln	Phe	Leu 150	Leu	Leu	Thr	Gly	Met 155	Leu	Val	Gly	Gly	Ile 160
Ile	His	His	Gly	Val 165	Ser	Asp	Arg	Phe	Gly 170	Arg	Arg	Phe	Ile	Leu 175	Arg
Trp	Cys	Leu	Leu 180	Gln	Leu	Ala	Ile	Thr 185	Asp	Thr	Cys	Ala	Ala 190	Phe	Ala
Pro	Thr	Phe 195	Pro	Val	Tyr	Cys	Val 200	Leu	Arg	Phe	Leu	Ala 205	Gly	Phe	Ser
Ser	Met 210	Ile	Ile	Ile	Ser	Asn 215	Asn	Ser	Leu	Pro	Ile 220	Thr	Glu	Trp	Ile
Arg 225	Pro	Asn	Ser	Lys	Ala 230	Leu	Val	Val	Ile	Leu 235	Ser	Ser	Gly	Ala	Leu 240
Ser	Ile	Gly	Gln	Ile 245	Ile	Leu	Gly	Gly	Leu 250	Ala	Tyr	Val	Phe	Arg 255	Asp
Trp	Gln	Thr	Leu 260	His	Val	Val	Ala	Ser 265	Val	Pro	Phe	Leu	Gly 270	Leu	Leu
Leu	Leu	Gln 275	Arg	Trp	Leu	Val	Glu 280	Ser	Ala	Arg	Trp	Leu 285	Ile	Ile	Thr
Asn	Lys 290	Leu	Asp	Glu	Gly	Leu 295	Lys	Ala	Leu	Arg	300 Lys	Val	Ala	Arg	Thr
Asn 305	Gly	Ile	Lys	Asn	Ala 310	Glu	Glu	Thr	Leu	Asn 315	Ile	Glu	Val	Val	Arg 320
Ser	Thr	Met	Gln	Glu 325	Glu	Leu	Asp	Ala	Ala 330	Gln	Thr	ГÀа	Thr	Thr 335	Val
GÀa	Asp	Leu	Phe 340	Arg	Asn	Pro	Ser	Met 345	Arg	ГЛа	Arg	Ile	350	Ile	Leu
Val	Phe	Leu 355	Arg	Phe	Ala	Asn	Thr 360	Ile	Pro	Phe	Tyr	Gly 365	Thr	Met	Val
Asn	Leu 370	Gln	His	Val	Gly	Ser 375	Asn	Ile	Phe	Leu	Leu 380	Gln	Val	Leu	Tyr
Gly 385	Ala	Val	Ala	Leu	Ile 390	Val	Arg	Сув	Leu	Ala 395	Leu	Leu	Thr	Leu	Asn 400
His	Met	Gly	Arg	Arg 405	Ile	Ser	Gln	Ile	Leu 410	Phe	Met	Phe	Leu	Val 415	Gly
Leu	Ser	Ile	Leu 420	Ala	Asn	Thr	Phe	Val 425	Pro	ГÀЗ	Glu	Met	Gln 430	Thr	Leu
Arg	Val	Ala 435	Leu	Ala	CAa	Leu	Gly 440	Ile	Gly	CAa	Ser	Ala 445	Ala	Thr	Phe
Ser	Ser 450	Val	Ala	Val	His	Phe 455	Ile	Glu	Leu	Ile	Pro 460	Thr	Val	Leu	Arg
Ala 465	Arg	Ala	Ser	Gly	Ile 470	Asp	Leu	Thr	Ala	Ser 475	Arg	Ile	Gly	Ala	Ala 480
Leu	Pro	Leu	Leu	Met 485	Thr	Leu	Thr	Val	Phe 490	Phe	Thr	Thr	Leu	Pro 495	Trp

Ile Ile Tyr Gly 500	Ile Phe Pro	Ile Ile Gly 505	Gly Leu Ile	Val Phe Leu 510
Leu Pro Glu Thr 515	Lys Asn Leu	Pro Leu Pro 520	Asp Thr Ile 525	Lys Asp Val
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Cys Trp Thr His	Met Leu Asp 55			Thr Asn Met
Thr Pro Lys Ala 65	Leu Leu Thr	Ile Ser Ile	Pro Pro Gly	Pro Asn Gln 80
Gly Pro His Gln	Cys Arg Arg 85	Phe Arg Gln 90	Pro Gln Trp	Gln Leu Leu 95
Asp Pro Asn Ala	Thr Ala Thr	Ser Trp Ser 105	Glu Ala Asp	Thr Glu Pro 110
Cys Val Asp Gly 115	Trp Val Tyr	Asp Arg Ser 120	Val Phe Thr 125	Ser Thr Ile
Val Ala Lys Trp 130	Asp Leu Val 135		Gln Gly Leu 140	Lys Pro Leu
Ser Gln Ser Ile 145	Phe Met Ser 150	Gly Ile Leu	Val Gly Ser 155	Phe Ile Trp 160
Gly Leu Leu Ser	Tyr Arg Phe 165	Gly Arg Lys 170		Ser Trp Cys 175
Cys Leu Gln Leu 180	Ala Val Ala	Gly Thr Ser 185	Thr Ile Phe	Ala Pro Thr 190
Phe Val Ile Tyr 195	Cys Gly Leu	Arg Phe Val 200	Ala Ala Phe 205	Gly Met Ala
Gly Ile Phe Leu 210	Ser Ser Leu 215		Val Glu Trp 220	Thr Thr Thr
Ser Arg Arg Ala 225	Val Thr Met 230	Thr Val Val	Gly Cys Ala 235	Phe Ser Ala 240
Gly Gln Ala Ala	Leu Gly Gly 245	Leu Ala Phe 250	Ala Leu Arg	Asp Trp Arg 255
Thr Leu Gln Leu 260	Ala Ala Ser	Val Pro Phe 265	Phe Ala Ile	Ser Leu Ile 270
Ser Trp Trp Leu 275	Pro Glu Ser	Ala Arg Trp 280	Leu Ile Ile 285	Lys Gly Lys
Pro Asp Gln Ala 290	Leu Gln Glu 295		Val Ala Arg 300	Ile Asn Gly
His Lys Glu Ala 305	Lys Asn Leu 310	Thr Ile Glu	Val Leu Met 315	Ser Ser Val 320
Lys Glu Glu Val	Ala Ser Ala	Lys Glu Pro	Arg Ser Val	Leu Asp Leu

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Phe	Cya	Val	Pro 340	Val	Leu	Arg	Trp	Arg 345	Ser	Cya	Ala	Met	Leu 350	Val	Val
Asn	Phe	Ser 355	Leu	Leu	Ile	Ser	Tyr 360	Tyr	Gly	Leu	Val	Phe 365	Asp	Leu	Gln
Ser	Leu 370	Gly	Arg	Asp	Ile	Phe 375	Leu	Leu	Gln	Ala	Leu 380	Phe	Gly	Ala	Val
Asp 385	Phe	Leu	Gly	Arg	Ala 390	Thr	Thr	Ala	Leu	Leu 395	Leu	Ser	Phe	Leu	Gly 400
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Leu	Ala	Asn	Met 420	Leu	Val	Pro	Gln	Asp 425	Leu	Gln	Thr	Leu	Arg 430	Val	Val
Phe	Ala	Val 435	Leu	Gly	Lys	Gly	Cys 440	Phe	Gly	Ile	Ser	Leu 445	Thr	CÀa	Leu
Thr	Ile 450	Tyr	Lys	Ala	Glu	Leu 455	Phe	Pro	Thr	Pro	Val 460	Arg	Met	Thr	Ala
Asp 465	Gly	Ile	Leu	His	Thr 470	Val	Gly	Arg	Leu	Gly 475	Ala	Met	Met	Gly	Pro 480
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What is claimed is:

- 1. A method of screening candidate substrates of the organic cation transporter 6 (OCT6) comprising: 50
 - a. providing a test agent;
 - b. providing mammalian cells or a mammalian cell line which express OCT6;
 - c. incubating the test agent with the cells or cell line; and
 - d. determining whether the test agent is a substrate for OCT6,
 - wherein the mammalian cells or mammalian cell line provided in step b, are leukemia cells or a leukemia cell line, respectively.
- 2. The method of claim 1 wherein the test agent is coupled to a detectable substance.
- 3. The method of claim 2 wherein the detectable substance is selected from the group consisting of extrinsically activatable enzymes, prosthetic groups, fluorescent materials, lumi-

- nescent materials, bioluminescent materials, radioactive materials, positron emitting metals using various positron emission tomographies, nonradioactive paramagnetic metal ions, immunogenic tag peptide sequences, extrinsically activatable toxins, extrinsically activatable quenching agents, and antibodies.
- **4**. The method of claim **1** wherein the step of determining whether the test agent is a substrate for OCT6 comprises analyzing whether the test agent is located intracellularly.
- 5. The method of claim 1, wherein step (d) comprises determining the viability of the cells or cell line.
- 6. The method of claim 5, wherein the viability of the cells or cell line is determined by applying a dye to the cells or cell line, wherein incorporation of the dye by the cells is indicative of death of the cells or cell line.
 - 7. The method of claim 6, wherein the dye is trypan blue.

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