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Method for Screening for a Tobiano Coat Color Genotype

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Bailey et al.

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(54) **METHOD FOR SCREENING FOR A TOBIANO COAT COLOR GENOTYPE**

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(73) Assignee: **University of Kentucky Research Foundation**, Lexington, KY (US)

Haase et al. (Animal Genetics, vol. 39, pp. 306-309, E-pub: Apr. 10, 2008).*

(*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 245 days.

Brooks et al. (Cytogenet Genome Res. vol. 119, pp. 225-230, E-pub: Feb. 1, 2008).*

(21) Appl. No.: **12/353,524**

Lindblad-Toh, K. et al.; "The Genome Sequence of *Equus caballus* (horse)"; Gen Bank Accession No. AAWR00000000.1 (Jan. 18, 2007).

(22) Filed: **Jan. 14, 2009**

Linblad-Toh, K. et al.; "The Genome Sequence of *Equus caballus* (horse), Assembly Version 2.0"; Gen Bank Accession No. AAWR00000000.2 (Sep. 6, 2007).

(65) **Prior Publication Data**

US 2010/0184025 A1 Jul. 22, 2010

Brooks, S.A. et al.; "*Equus caballus* chromosome 3 inversion breakpoint, genomic sequence"; Gen Bank Accession No. EF442014.1 (Feb. 15, 2007).

Related U.S. Application Data

(60) Provisional application No. 61/021,129, filed on Jan. 15, 2008.

Bowling, Ann Trommershausen; The Journal of Heredity 78:248-250, 1987; "Equine linkage group II: phase conservation of To with AIB and GcS"; © 1987, American Genetic Association.

(51) **Int. Cl.**

C07H 21/04 (2006.01)
C12Q 1/68 (2006.01)

Raudsepp, Terje; Mammalian Genome 10,77-282 (1999); Comparison of horse Chromosome 3 with donkey and human chromosomes by cross-species painting and heterologous FISH mapping. S.A. Brooks, R.B. Terry and E. Bailey A PCR-RFLP for KIT associated with tobiano spotting pattern in horses Animal Genetics, 33, 301-303 © 2002 International Society for Animal Genetics.

(52) **U.S. Cl.** **435/6.1; 435/6.11; 435/6.12; 435/69.1; 536/23.1; 536/24.3**

* cited by examiner

(58) **Field of Classification Search** None
See application file for complete search history.

Primary Examiner — Jeanine A Goldberg

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(57) **ABSTRACT**

A method for screening for a Tobiano genotype includes obtaining a nucleic acid from an equine animal, and analyzing the nucleic acid for the presence of an inversion in a chromosome ECA3q which is indicative of the genotype for Tobiano. The method includes detecting at least one of a telomeric breakpoint of an inverted ECA3q chromosome and/or a centromeric breakpoint of an inverted ECA3q chromosome. In one embodiment, the nucleic acid may be analyzed by the steps of hybridizing the group of probes or primers having the sequences set forth herein in SEQ ID NO:8, SEQ ID NO: 9, and SEQ ID NO: 10, or sequences complementary thereto, and preparing an amplification product. A 209 bp nucleic acid amplification product (SEQ ID NO:11) indicates the presence of the inversion.

13 Claims, 8 Drawing Sheets

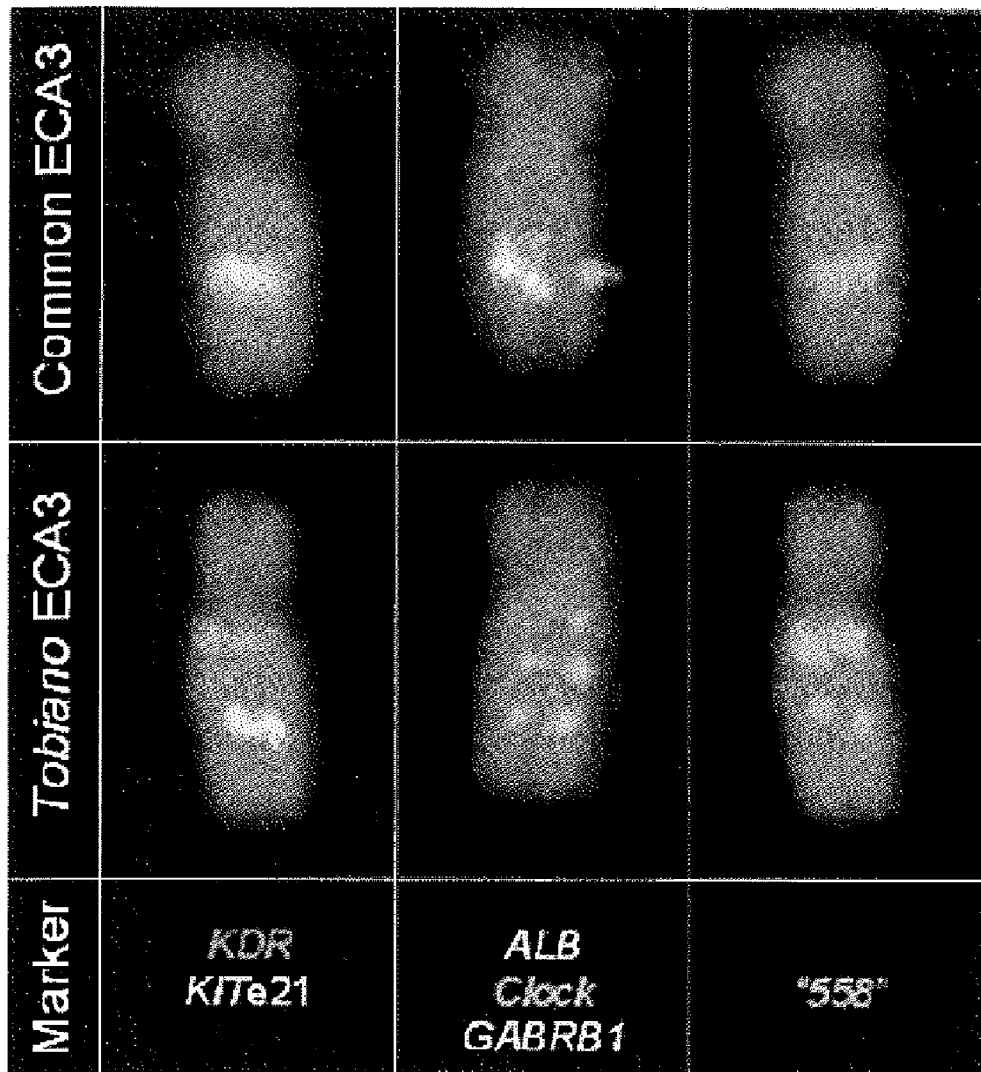


FIG. 1

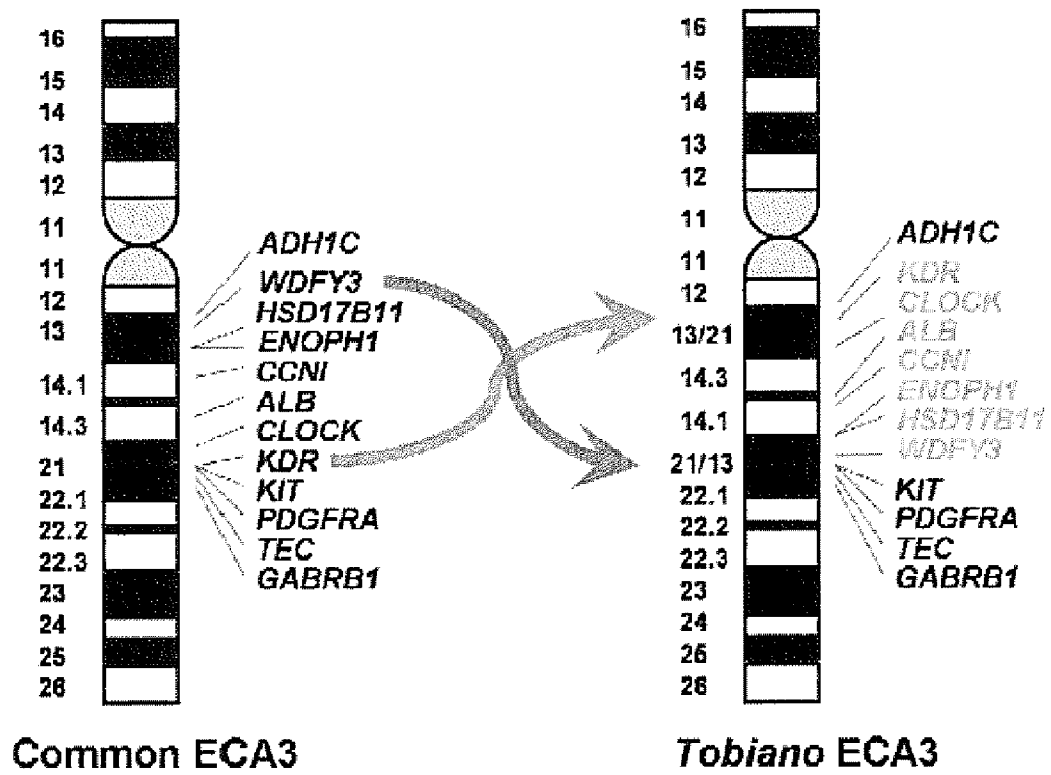


FIG. 2

TATATTTAAATFACTCAGAGCAATTTATGAAACCCACACAGCCTCAACAGACCATTTTAACC
TTGAGCAGTGTAAATTTCCGGGTGTGTCTTGAAAATAAGTTGGCAACTAATCATATATAGA
AGTAAATAAATGAGTCAAAATATGTCTAAAATGAAGTTATAAGTCAAGTAAAGTATTGTTT
TGTATCTAAAGGAATAATGGGTTTGTTTTGTAGTGGAAATGATCACTTCTTTTTGGCAGT
AATTAAGTAGTAGACCACTTTGGAATTTTAGAATCATAAAAAGTGAATTCATTTTTAATA
AATTTTTTTAAATAAAATCTTAGAAAAATGGAATCAATCTACGCTGTGGCATTGTGTTT
CTTTGGCGTGGCAATGGAGCTGACATTGCAGAATGAGATGTGGCGAAGAATGGCTGCTCT
CCAACACTGTATACAGGTTTTCTTAAATTAACGGCTAGATTGTAAACTTCACCAAAGCAG
AGAACACCTTCTTTTTGACTGTTTTATCCCTGGCCCTCAGCAGAGCGCTGGCATATAGGA
GATCATAGCTGAAATAGCTGCTGTGTTTTACCTTTGAACACTGTCTCTGTTACAACAAAA
TGATAAAAGTCATCAAGAGAGGTTAGAGTTTTTACGTAATGTTATGCATTTAAAAGTAT
AAGATATATGTTTGTATTTAAAATAGTATCTACTTTAATCAGTGCATGTGATTCCTTCT
TTAATTTGAAAAGAAAAGCTGTACATTTTTAATCAATGGTCTATGACAGTCTAGGAGGCGTG
ATAGCCAATAAAAACATTTTTACATGGCACTTTTCTCACTCCTTTGAAATCGGAAGCCTC
TAAATAGATATTCTGAACAAAATATATAGCCCAAGTACGATCCAACCTAGAAATGGAAAA
AATACAGAAAAGTTTTATTGTAACATATTTGAAAATCTTAATATGATGAACAAAAAGAGCT
GCCCAAATTTGGGGACGATGGTCTGCATTTGCAGGTCATTTGGTAAATAAAATATGG
TAAATACCAGCTGTTTTCTTATGCTCTTAGATTATCTGTTTATGTTACCAGGATGATTTTG
TGAAGCCGAGTATCAGACTCCATGTCTTAGCAGGCTCAAAGGTCAAAATFAGAATCTTAA
ATTGAAGTTCGCATTAATCCAGATTGCCATTTGCTTACTTTTTGATAGCAGTCCACCTAA
ACTGAAATGAGTAATAGAGATTTTCATGCTGGAATGATATTCCTGCCAGCACCACTAACT
GTTGAATGTTAAGTATGAGATTTGGCAGGTGCAGAGATTTTAGTCACCTTCAAAAAGCA
ATATGCAGACACCCAAGCCAAAACAGCTTATATATTAACAAAATCAACATCCAACCATCT
GCCAATGGAAACAAACATGTCAGTCACTTTCCATTTGATTTGGCAGGGTTCTAGAAATGA
ATGATTTTAAACATTAATTTCTCATCCAAAAAACACACAGGGAATTTTATTTACCTGGT
AATTGAGCTGACTCTGGGCAAAATTAATGAGTTAAAACGCTTTCATCTTCTCCTTGCTC
TTTTCATGCTGCTCCTCCGTTACCCTGTGTACCCTGCACACCCATCCTAGCACCTGTT
ATTAACCTTATGACTCCCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT
GCATCTGATTTATTTCTCTTAGCCAACATACGTTTTTTTAGGACTCTGCTTATAGGCGAT
TATATTTAGTAATTAATCAGGTAACCAATTTCTTTAAATTAATTTCTTAAAGTTAAAA
TGACTGACATATAAAAACATGATTTTTCCACCTCTAAAAGTCTTTAAAGCGTAAGCATAAA
TCTTTTTAAACGCTTGTCTCTCTAGTCGGCTTGGGAAATTTGTCAGCCTATTAGTGCCA
CTGAAATFAGAGGTTTTAAGCAGCCGTGTAATGTCTGCTCATCTATCTGGCTAACCTGT
TTGGAAATTTCTGCTGACAAATGACTCTGGTATGATTTGGCAGTGGTAGCAGAAGGTCAG
AATCTCTGAGGTTTGGCAGACTGCTGAAAGTTTCAGAGGACCTAATTTCTCACTAAACAG
ACGGATTTCTTCTTCAATCAATGAAGTGTAGACTGACTTTAAACAATAACAACAAAAATTC
AGCTGCTAGTCTGTGTGTTTTAAATFAGGGCTACTTGAATCAGAGAGAAAAAATAGATG
TGGATTTGATCATTTTTATATTTATTTTCTACTTTCAATGTAAGCTGATACAATTTATTT
TTACTTTTTAAAAGGAGAGGAATTAGGTACAGAGAATCTGTAAACATTTATCCTGTCTTTC
ACCACAGATATCCAATTAATGCTTTTACATAAATGCAAAATCTTCAGCAGATAGAACA
TTTTAAATTTACGGCTCAGAGGGATTTCAATCATCTTTGGCATTCAAGGCTCAAGGCAT
TCAGATATTTAATCAGAGCAGTATAACAGAAAGTACAC | AGCAGATTGTAATTTAAATGCT
GTACTTTTTGCAACATATATTTGCAATAAAAAGCATAGAGAATATGTAATAAATAAGAATGTA
ATTATGAGTACTGCACATTTTCACTACACCTTTGATAAGGGAATTTCTGCATACCTTTCA
TGTAATAGACATTTTCTCAGTACAACTTTGTATGGAATGCATAGGACACACACCTGATGC
TCTGTTCTCCAGCCATGTGGGTTGTTTGTGTCCACATFAGGGCACTTACACCTCCATGT
GGAGACCATTTCTCTATGCAGTAGAGAGCAAAATAGAAATTTTAAATATGAGATTTATTT
TTTGGTATACAACCAGACAATAGTGCAGAAGATAGACTGGAGGGTTAGGTGCCAAGAAGT
CAAATTAAGGAGTCTTTGCAATAATCCATGAGGCCTGAAGAAGGCTTCAATTAAGAAAAG
AGAGAGCCAGAGAAATGGAGTGGGATTTAACATTTTCTAGGAAGATAAGGAAAAGA
ATGGCAATCCAGGAGAAAGAAATGAGACACGGTTGCCAATAAACGTTGATTAAGAAAAGGCA
TGCAGCACGAAATGAGCAGAACTTTTGCAAAACCTATATTTAAGGAATTAGCAACAATAA
CAAAAAGTAGTGGCTATCTTGAGAAGTTATCTCAAGGACAATTAACCAAGAGTTGATGA
GTTTCAGGATGTTGAGAATGAATACAGCTTTGGTCAGCGCAAATGCTGAAAAGTAAGCTGA

← ECAtoR

← ECA3Fc

FIG. 3

CATTTCTACCAAAGGTCACAGGGAAGTAGAAGCCCTGTGGTCTCCGTGTGCACAGAAAAC
TGAGAGCTTATTTGAAACAAATAAGAGCAAGAGAAAAAAGAGCCTTAAAGAAAAGGATT
TGATGGTTGACAACAGGCATATCAGAGAGTTTGCAAATAGCCCACTTGGATTTGGAAAT
TGGACCTCTTGGCATGTCTGTGAGTGTGATTTTATTAGAA'TGGTGGAGATGGAAGTCAA
TTTTAATCGA'TTAGGTGATGCGGAT'TGGGGAAAT'TGGGGAAACAAGTCTGGGCAA'T'FC
TGTTAAATAATTTGGTACTAAGAGAAGGAGGGAGACAGGTTTCAATTTGAAGGGTATAA
GAGTTTCAGAAAAGTACATTTGTTCA'TTTT'AAAAATAAATATGGAAGGAGGGAACCTGAG
TATATTTCTGTGCTGAAAGGTTAGAGAGAAGTGGAAATAGAATATCTAGAAAAGACCAGA
GCAAAT'TGCTGGTGTAAATTTAAGAGGGGCAAAGT'GT'GCAGGTGAAAAGTGTGAACCTTAGG
AGTCGGAAGCA'TGGACGCTAGGAGGTATAGACAAGAAGTGGAGCAGGAGGTGGCTAGGCA
CTCTCCCCATTCCTGCTTCTATTGCA'TTAT'GAAGGGATCAGAGATATGCTGATGGTGA
GACAAGTAGAAATCTTGTAGGGAAGTAGAGGAGAGAGAATACCAGAAATGGACTACTGGA
CACTTTGAATGCAGTAAGCACCTGCTTAAGATAAATACTCCCCTCACTAAGGAATCTCTG
AGAATGGTGGCTTTT'FACAATAT'TGCACCTGGTGTCTTATCACCTCACTCTTTATGAGTTT
CACAAATATTTAGT'GAGCTCAGCCCATGTGCCTAGAACTATATCAAGGCCTAGTAGGTTT
TATAGGACAAGGAGAAGAGATGGCTCTTCAAGGAGCTTGGGCTTCTGTTCTGTGGGTAG
TATTGGGCCAAGGAGTAGTTTCTTTCAGCCATTTTCTTCTTCTAT'GAACAGTATTTCA
TTGTCAATATTTCCATTTGCTGTTCATAT'GT'GACTGCTTCTTGGT'FGCATTGTACTTTC
CTGGGCGTGTATCTATATTTTGTGAATGGAGAAAGGATCCATAATGACAGCATTTTGT
CTTCTCCCCTATGCTTTCAAATAGTGT'GAGGATGCTTATTTATCAGTGTGCCTGCTGTG
AGCAGCTCTGTGCCAATTTAGCATCATCAGGAACAACACTACAT'FAAGGTGATGGTCCCTGT
GCAAGATCCGTGAGCTCGTTCCACTAATTCATGAAAACATATTCATTTATTTTTAGCTAA
TAAAACGATCTTCAAACCTGCTTGTTCAGGCATACATCACCAAAACATTTCTATGTCTTT
AATGTTTTCAACCTTCATTAATCTTTTCACTCCACTAATTTAGAAATGTGGAGGGAAATCTC
TCAAATTTTATGAAATATACTGCCATCATTTAACCTTTGCTATTATGCTGATTTATTTA
TTTTATTCATTTATTTCTTTATGGCATAGGAATACATAATAAATAATGAGAATAGAGTTAG
CAACTTTTAAAAATAGAAAAGTACTTACTAGAATAAAAAATAGTTACCTGACAGAAAGTTT
AAAAATAATCATTTCTTCTATTTATTCACCTCCACATATGCTAAAAATTTTAGTGGAAACT
ACTGGTCCCGCTTCCCTCCCTGCAAAGCA'CCATTTCCCTCCTTTGTCACTGGGAAGGAG
TCCTAGTTCCAGTGGGTTGAT

FIG. 3 CONT.

CTCACCTATGAGACCGCTTGGAGAAGTTGGAAGGGCCCAAGGTTATATCCAAAAATAACT
 GACAAGCCCTCAGCCTTCACCCAGGAGTCCAGGATTGTCGTTCCCTGAGGCAAAGCTG
 GCAAAGGTTTGGTGGCGGCAGCTAGGAGAAAGCACCTCTGGAACATCCCTCCTCGGTTG
 AAATTCGTAAAGATGAGCAGTGTCTCAGCAGGAAGGCTGAGTTCGCCAGCTGATAGGCAG
 GCAGCTGACAGGCCCGCAGAGCCGGGTGCAGAGGATGACTCTGCTCGGGAAGTCTGCTGG
 AGCTGCAGCTTGCCAGGCTCTCTGCTCTGACACTCAGCATTTGCCCTCCTTTGAAACTG
 ATCTCACCTTGTTTTCCACCTCCCTCATCCACACAGAAGGTAGGATTTTTTTTTTTTTTT
 GAGGAAGATTAGCCCTGAGCTAACATCTGCTGCCAATCCTCCTCTTTTTGCTGAGGAAGA
 CTGGCCCTGAGCTAACATTCATGCCCATCTTCTCTATGTTATATATGGGACACCTGCCA
 CAGCATGGCTTGCCAAGCAGTGTATGTCCGTACCCAGGATCCGAACCGGCAAACCCTGG
 GCTGCCAAAGGGGAAACGTGCACACTGAACCGATGCACCTCTGGGCTGGCCCCATAAGTT
 AGGATTTTTGATTTTCAAATTTTAAAAATAAAATTAATCTGTTGGGGCTGGCCCCGTGGC
 CAAGTGGTTAAGTTTCGCGCGCTCCGCTGCAGGGCAGCCAGTGTTCGTTGGTTCGAGTCC
 TGGGTGCGGACATGCACTGCTCATCAGACCACGCTGAGGCGAGCGTCCACATGCCCAA
 CTAGAGAACCATAACGAAGAATACACAACATATGTACCGGGGGGCTTTGGGGAGAAAA
 GGAAAAATAAAATCTTTAAAAAATAAAAAATAAAAAATAAAAAATAAAATATCT
 GTTCATGTTTTTAAACAGCCATAGAAATTAATACTCCTAATTATAGAAACAGTGGAAAA
 AAAACCGTACAAAGAATAAAGGTGCCAGAGACCCATAGTACCAGACAAAGGCAACCAC
 GTTTATATTTTGGCCTATTTGCTATCAGTCATTAATGTGTGTCTTAGGCGCCACATGT
 GTGAATGCACATATTCACACTTAAACACGACAAAATAAAACAAAGTTTCACCCAAAGTTA
 AAGCCATGGTGGGTGACACTCAACGACTCACATTTGCAGTTAAGACCAAGGACTACCAGT
 GGCTTGCCTGGGCCACGTGACACTGCCGAGGGGGCCGTTACAGTTCTAATTTACAGTTT
 GGCFAAGCCAGGCGCTTGGTGATAAGCACAGCTTGTTTATTTGTCTTTGATGGATGCCA
 GGTGGATTGTGGTCAACAATCTTCTTCAGCAGCCTCAGCACTTTGCATCCACGGAGCT
 AGCGTGAGGCCCTCCCTGAGGAAAACGCAGGTACACAATGAGCGCCACAACAAAGGG
 ACGTCCACAGTGGTGGCGGCACCAAACGAGGACAAAGAAGCGCTTGGGCGGAGAGGAAAG
 AGGAAGAAAGCAGGGAGGGCCAGGAGACGCCACCAGGCAGGCAGGTGGCCAGGCAAGCT
 GACCACTTTGCTCTTGTGTTACAGGTTCACTCAGAGAACGCAAGCTTTGGTGCACCTTT
 TCATACACTGCCFTAAAGGCTTCCAGGAATGAGTGCGGGCGATTTGCTGTGTTTTGACA
 TTTTTTTCCCTTTCTGAGAGCTGTGAGTTGAGAGGCAGTATCCTCTTGTGAGAGGTTT
 GGTGCCGGCTGCAAAATTAAGGCAAGAAAATAACCACAAATAAGAGAGCATGAGTGGCCA
 GATATCTCTCAATCCTGATATGGGCTGTGGAAGGCCATGTGTGTTTATGTCTCCTCCCTCCA
 GCTGGAGCCAAGAAGAGGCATCCGGAGTGATGGCATCCAGAGGCTGCAGGAGGCAGGGCC
 TGTGGGGGTATAAAAGCAGTGGGATGACAGGGGGCCTTCTGCTGGAAGCTGTACGAGGCC
 CTTGTTGAAGCCAGTGAAGTGCACCTGTGTACCTCCAGTTTCTTGGGGCTGGACTGGGC
 TATCCATTGAAGAAATGGGGTGGAGAACCCAGGCTTAGTCAATTTTTTCCCTCCCAGTTT
 CCGCTGGTGTGGGCTGGAATGTGACATGTTGTTTTCTTTACTAATTGGGTCATAATTTT
 AAAAGCTGATGATCTCTTTCTTAAAGTACCAGGTCTCCATATCAAACCAGGGGAAAC
 TGTGCCGTGGGGTTGCTGGTGAATCTTGTCTCCCTAGAAGCCCTGTAGGTGATCTGTGTG
 TCAGAAGACAGAAGGAGGAATGGGGTTCCCTCCCTACTGCTCTGGAGAAC'GTGCTCTA
 GCAAGGGTGGAGCATGTTTCAATTAAGTTTCTGGGGCAAACAGAGGGCCAAATGATAGA
TCAGTGTAGACGTAGTGTGACAGAGACCCAGGCAGA GCTCT | GCCTCCGGCCTGCAGCAC
GGGCTCCCGACTTCTTGAATGTGAATTTCTCTTCAAAC'CCAGAATCATTGAGAAC
CTATGCTAGAGTGGGAGTAGCTGTTGACTGAGAAAACGAAAGATGATTCAAAAATTAAT
 AAATATGCTTAAACCATATTTGAGTACAACGAGAGGCACCTCACAACTAGAAAAACACAAC
 TATGTACTGGGGGGCTTTGGGGAGAAAAAGAAGAAAAAAGAAAGATTGGTAATAGTTGTT
 AGCTTAGGTACCAATCTTTAAAAAAGCATATTACTCGATAATCAAGT'PATATAATG
 GCATACAGTTAAACCAAAGTTGTAAGGGACCATCACAAATCGTTTCTGCAATGGACAA
 ATACTCAACACACACCCCTTGTGTGATTCATCCCACTCTTGGCAGTGCATCCTGTGCGAT
 TGTATCAATGGCAGCCATGATGCGTTTCTCCATTTCTCAGGACTATGAGACTCTTGGTG
 TAGCCCAAGAGATGTCAATGGTGTGATGACCTGAGCTCCCGGGTGGCTGCTGCTCCAGA
 GTTAAGTCAATGTTTAAATAGAAAGCCCTACCCAGTGCCTGGGATAGATTGATGAAAGA
 ACTTCTGGACTGCTGAAAATGAGGTTTCAAAGACTTCTGAAAGCTGTGGGAAAACCCCTT
 CAGTGTATCCTGACACTATGTTCTTACACACACACACAGAGTTGGCATCTGTATTGA
 TGTATCAAAAAGTCATGGGGGATCCCTTACACGTTCAATCACCCAGTGTGTTGTTATTTGG
 TGCCCGTATATGGATGCTGTGAAGAATCACATTTT'TAGTGAATGAACACAGACTTATC
 ATTGAATTTGAGACAACTTCAAAGGCAACTTCTGTTCTGTTGAGTCTGAAAGCAGCA

← ECA3F

← ECA3xR

FIG. 4

GCTGAAGTTTTATCGTTTTACCCCATCTGCCATCAGAGCCTGCAGAGGGTGCAAGAA
GGTTTTCATCTTCCATCGCTTATGGAGAAGGTGCCAGACCATGGTCTGAGGAACACAATAC
TCTCAGCCAGCCCTCTTTATGGACTCCTTGGGGCTCTGCTGAATGAGACAAAAATGATTC
CGTTTTCTTAAGACATGTGAAATGTAAGCACACACAAGTTGGTGCCTACTACATGTTGCI
AGTCATTGAGTAGTACATTTTTTTCTTCAAAGCGTCAATTTTTTGAATCATCATGTACA
TACTGGCCAAAAACTACAAGTAACGCTTTTGAGTTAATTTGCATTTTTCTCATCACCAAA
GCATCTAAACACACCATAAAATAGTTAGACACAATACATCATACCACAGTAATGTGGCC
CTTTATAAAGTGGCATACTTTTATAGCTACCTCCGCTGATCCTCAGAAAAATCCTGTAC
GATATGCACCATGATCCCCATTTTGCAGATAAGGAACTGGGACTCAGAGAGACAGCCAC
TGATCAAGGTCTTGCACACAGCAGATTGTGGAAGTAGGATTCCCTTCAGGCCTGGAAACC
CACAGCCGCTCCTACACCATGTGGCCTTCTCAAATAAGGAAAAACCTCTGATTTCTTAAA
ATAGTATTGTGCTATTATTCTCTTTTCTTGGTATATAAAAGTAAAGGTTTTAAATCTGGA
ATTTTGTGTTGTGATTAGGTTGAAGTGTGCGAACTGCTGATACTTTACCATTTCTGAC
CCTACGGTTCAACCTAATAGTTTCTTCTCTGGGGAAAGAAAGAAAGGAAGGAAGGAAG
AAGGAGAAAAATGCAATGCATGAAAAACAGAGAGCAATAAAAAAGATCAAAAAGCACAGAAG
AGGAAGAAAAATGAGGGAAGGAAGAAGGTATAAGAAGAAATTAGAAAT'ACTAGTAAAGCT
TTGTCACTCTTTCTAAACTCTCCTGCCCTTCAGTTCTTCCCCACAGTCTTGGCTTTTA
CTGTCTATTGCCTTTGATCAGGATTTTCTCAGTACCAGACTCCTTAAGTAATCAACAGA
GACCAACTGTATCATAT'GT'CTTTCACTTTTATTTATTTTAAACAAGGAT'GCGCAGAT
TTCTAGCCACCACCCCGAGGGGGTAAGAGGGAAGAAAAGATCGACAGAACAAAGTCTTGC
ATTTATTAGGACCGTCTAAATACGACTTCTCTTCTCGGGGGAAAAACACTGGACCAAAA
CGAAATGCCCTCAATGTACAAA'TTGTAGAAATGTAAAGTTAGCATAACATTGAAGTTTCA
GATGAGAACT'GACCTTAATGT'TGAAAGAAGAGAGATGAGAAGACAAAAAAATGAAAG
AAACCTTTTAAATAAATAATTAGCTTGGAGATGATATAAAGTGT'TTTCTAAATCAATCC
ATTCATAGGAAGAAATAAACACATTTGAACAA'TT'GATTAACACATGTGAATAACAAATG
ATAAAGCTAGGGAGCAGCTGAAGT'CCTGCCACTGAAGACTTAAAGGGATAAA'GCTCTCA
CCCCCTAAT'GCTGCCTCTGGA

FIG. 4 CONT.

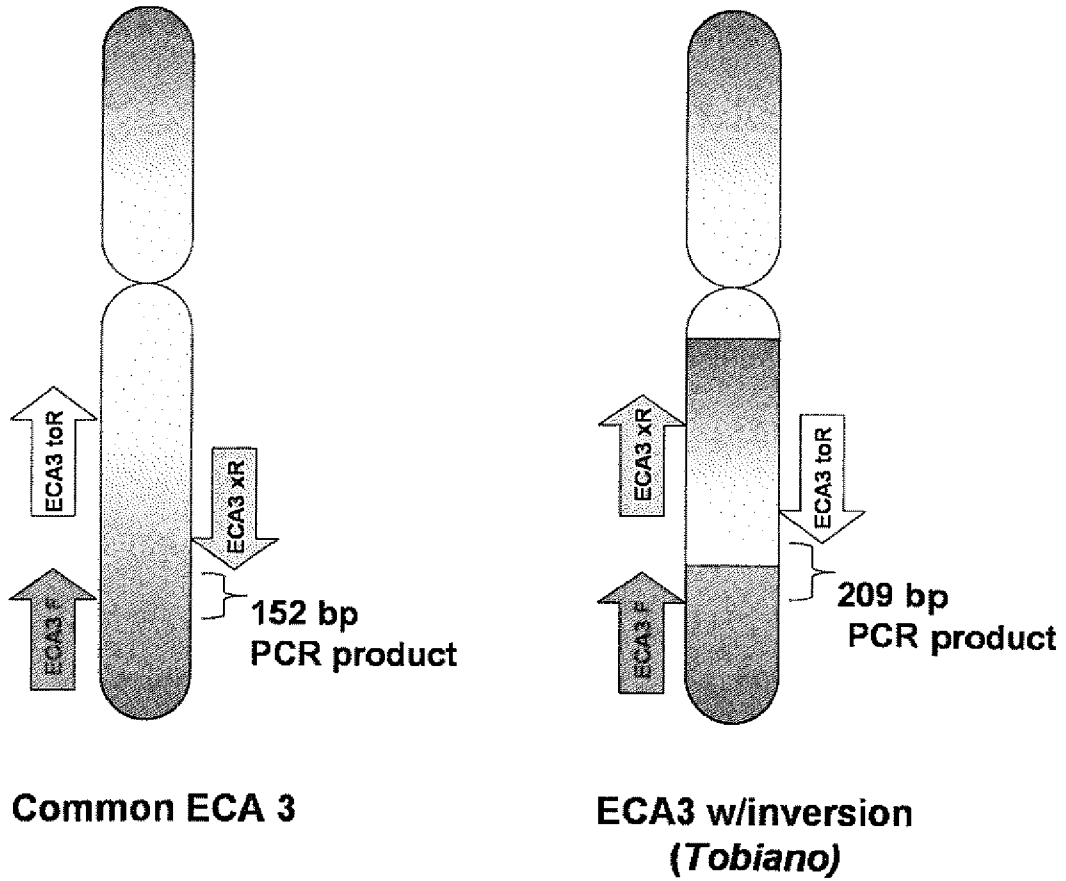


Fig. 5

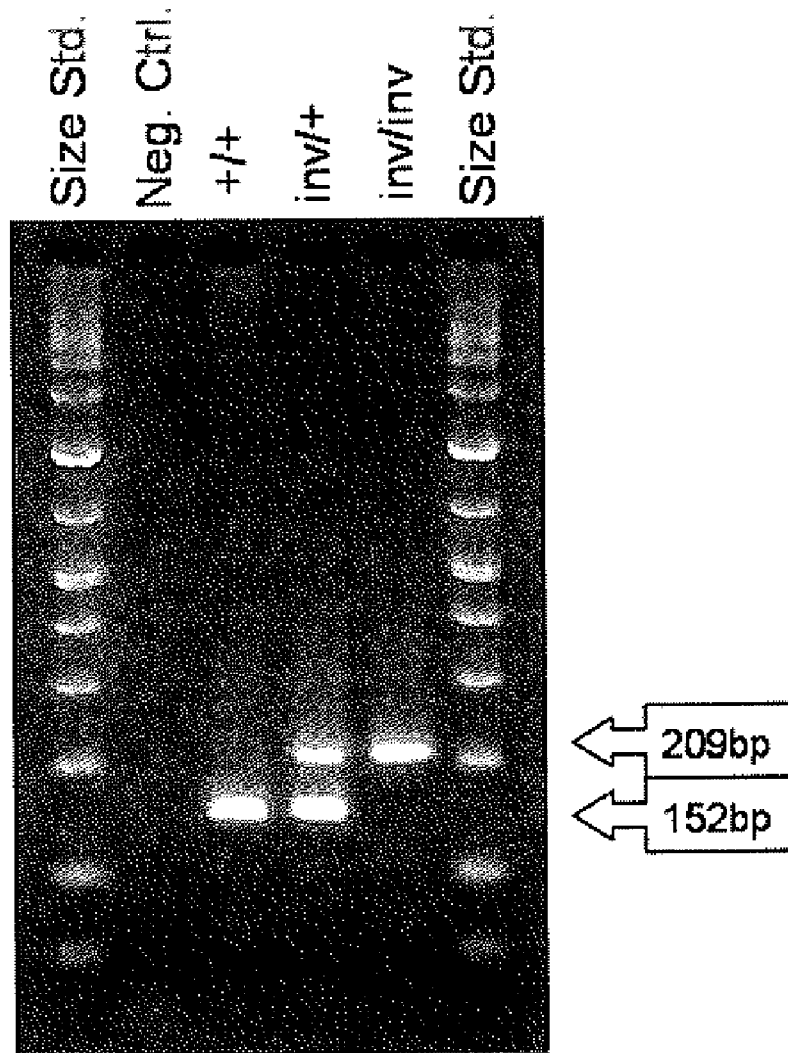


Fig. 6

METHOD FOR SCREENING FOR A TOBIANO COAT COLOR GENOTYPE

This utility patent application claims the benefit of priority in U.S. Provisional Patent Application Ser. No. 61/021,129 filed on Jan. 15, 2008.

TECHNICAL FIELD

The present invention relates to detection or screening for genotypes for coat color patterns. In particular, the present invention relates to screening for the genotype for the tobiano coat color pattern in horses. The method relies on detection of a chromosome inversion on horse chromosome 3 (ECA3).

BACKGROUND OF THE INVENTION

Tobiano is a white spotting pattern in horses caused by a dominant gene, Tobiano (TO). The tobiano color pattern of horses is highly valued and selected by many horse owners and breeders. Approximately 350,000 horses carrying TO are currently registered by the American Paint Horse Registry (APHA), but the pattern can also be found among several different horse, pony and draft breeds worldwide.

TO was previously shown to be genetically linked to the gene for Albumin(AL) (Trommershausen-Smith 1978). Later, an allelic association (linkage disequilibrium or LD) was discovered in which the chromosome possessing the TO allele usually possessed the Albumin (AL)-B and Vitamin D binding factor (GC)-S alleles (Bowling 1987). It is known that certain chromosome rearrangements, such as inversions, can create unusually strong LD by interfering with gamete formation, thus resulting in the formation of conserved haplotype blocks of alleles.

To explain the unusually high level of LD between the TO, AL and GC loci it was hypothesized that an inversion on ECA3 could be preventing recombination in this region (Bowling 1987). Several similar spotting patterns at the W locus in the mouse have been shown to be due to chromosomal rearrangements near the KIT (v-kit Hardy-Zuckerman

4 feline sarcoma viral oncogene homolog) gene. Two inversions, *Rump-white* (Stephenson et al., 1994) and *Sash* (Nagle et al., 1995), and four deletions, *Patch*, *19h*, *57*, and *Banded* (Nagle et al., 1994, Kluppel et al., 1997) occur within the 200 kb upstream of the KIT gene. All have been shown to disrupt the tissue-specificity or temporal expression of KIT during embryogenesis (Nagle et al., 1994; Kluppel et al., 1997; Hough et al., 1998; Berrozpe et al., 1999) rather than a coding sequence. The similarities between Tobiano and this particular group of mouse spotting patterns, previously demonstrated linkage of Tobiano and KIT horse chromosome 3 (ECA3) (Brooks et al., 2002) and the lack of a difference in the KIT cDNA of Tobiano horses (Brooks, 2006) all suggested that a nearby chromosomal rearrangement was the cause of the Tobiano allele. Initial efforts to detect an inversion using cytogenetic methods were unsuccessful (Raudsepp et al., 1999).

The rearrangement was finally demonstrated by one of the present inventors (Brooks, 2006; Brooks et al., 2007). Cytogenetic evidence showed that there was, indeed, a chromosome inversion on ECA3 near KIT which appeared to be completely associated with TO. No exceptions to the association of Tobiano with the inversion have been found and it is likely that the inversion has this effect by affecting regulation of the gene, KIT, during embryonic development. This conclusion is also based on observation of similar effects of mutation in KIT on mouse hair color.

The results of mapping several genes are shown in FIG. 1 and Table 1. In the left hand column of FIG. 1, a clear difference can be seen in the distance between KDR (red) and KIT (orange) when comparing the ECA3 from non-spotted horses and the ECA3 bearing TO. The center set of images shows three markers (ALB, Clock, and GABPB1) with different relative orders on each chromosome. The right column shows results from FISH mapping a bacterial artificial chromosome (BAC) 558 which crosses the distal breakpoint (green). On ECA3 from non-spotted horses, hybridization occurred at a single location, while on the chromosome with Tobiano two distinct locations were labeled with the single probe.

TABLE 1

FISH Markers used in this study in their relative linear order on ECA3.					
Marker	CHORI-241 BAC			Relation to Inversion	Sequence Source
	Clones	HSA	ECA		
GABRB1	49:M13	4q046.8mb	3q21	Telomeric	UCSC Consensus
TEC	38:G1	4q047.9mb	3q21	Telomeric	UCSC Consensus
PDGFRA	23:F11	4q0s4.9	3q21	Telomeric	UCSC Consensus
KIT e21	90:F8	4q055.447mb	3q21	Telomeric	Genbank #AY874S43
Intergenic Seq. "558"	102:M1	4q055.558mb	3q21	At Breakpoint	UCSC Consensus
KDR5'Ue1	129:04	4q055.787mb	3q21	Within	UCSC Consensus
KDR	127:D23	4q055.7mb	3q21	Within	UCSC Consensus
Clock	11:A9	4q056.2mb	3q21	Within	Murphy et al. 2007
ALB	21:KS	4q074.6mb	3q14.	Within	Genbank #AY008769
CCNI	99:89	4q078.4mb	3q14.	Within	Chowdhary et al. 2003
ENOPH1	69:C10	4q083.7mb	3q13	Within	Genbank #cxs03024
WDFY3	44:L24	4q085.9mb	3q13	Within	Genbank #CX599384
HSD17B11	105:819	4q088.7mb	3q13	Within	Chowdhary et al. 2003

TABLE 1-continued

FISH Markers used in this study in their relative linear order on ECA3.					
Marker	CHORI-241 BAC		ECA	Relation to Inversion	Sequence Source
	Clones	HSA			
PDLIM5	19:G11	4q095.7mb	3q13	Within	UCSC Consensus
ADH1C	189:L20	4q100.6mb	3q13	Centromeric	Genbank# AF134056

Since the presence of the tobiano pattern can increase value, horse breeders prefer to use breeding stock which are homozygous for TO, e.g., those which have inherited a copy of the gene from both parents, and will always transmit the gene to their offspring. The cytogenetics test requires freshly collected cells and is expensive to conduct. In place of a cytogenetic test for Tobiano, associative tests have been used to predict homozygosity for Tobiano. As described above, the Tobiano gene was associated in horse populations with particular gene-alleles or SNPs. Specifically, most but not all horses with the Tobiano gene also possessed the B allele for AL and the S allele for GC. Likewise, most but not all horses with the Tobiano gene also possessed the KM1 single nucleotide polymorphism of the gene KIT. Known methods for detecting genetic markers associated with the Tobiano genotype include detection of these nearby genetic markers using biochemical typing or molecular DNA tests. However, biochemical typing methods require freshly obtained biological samples. Furthermore, both biochemical and molecular gene detection methods also produced false positive and false negative reactions because they are associative with the tobiano trait and not actually responsible for the trait (Duffield and Goldie, 1998; Brooks et al., 2002).

Until present, there have been no precise and cost effective methods to identify all horses homozygous for TO. While the inversion can be detected using cytogenetic studies and fluorescence in situ hybridization (FISH) of DNA probes for genes in the region, this is an expensive and time-consuming procedure. There remains a need in the art for a methods for identifying animals bearing the Tobiano genotype which are effective, rapid, and which further can be performed effectively on stored samples of varying type, to obviate the need for acquiring fresh genetic material.

SUMMARY OF THE INVENTION

The above-mentioned and other problems are solved by applying the principles and teachings associated with the presently described method for screening for a genotype for a tobiano coat color pattern in an equine animal. In one aspect, the method includes the steps of obtaining a sample containing a nucleic acid from the horse, and analyzing the nucleic acid for the presence of DNA sequence characteristic of an inversion in a chromosome ECA3q. The inversion spans a large region of ECA3, from q13 to q21. One inversion breakpoint lies between the KIT gene and the KDR gene (distal or telomeric breakpoint) while the inversion breakpoint nearest the centromere lies between the ADH1D gene and the UNC5C gene (proximal or centromeric breakpoint). As will be set forth more fully below, the presence of the chromosomal inversion indicates that the equine animal possesses the Tobiano coat color pattern genotype.

In particular, the chromosomal inversion may be defined by addresses in the equine genome. With reference to the equine genome (EquCab2, available on the University of California Santa Cruz Genome Browser, Genome Bioinform-

matics Group, Center for Biomolecular Science and Engineering, UCSC) the telomeric breakpoint has been identified as between nucleotides 3:41925230-4192523 1 of EquCab2. The centromeric breakpoint has been identified as between nucleotides 3: 77663330-77663331 of EquCab2. Those breakpoints, along with the 5 kb surrounding those addresses, are set forth in FIGS. 3 and 4 (see SEQ ID NO:13 and SEQ ID NO: 14).

In one embodiment, the nucleic acid is analyzed for the presence of the distal chromosomal inversion by the steps of hybridizing to the nucleic acid three probes or primers having the sequences set forth as SEQ ID NO:8, SEQ ID NO:9, and SEQ ID NO:10, and deriving an amplification product from those primers. Presence of the chromosomal inversion, and therefore the Tobiano genotype, is indicated by an amplification product having a 209 bp nucleic acid sequence set forth herein as SEQ ID NO:11. An amplification product of 152 bp in length (SEQ ID NO:12) denotes a chromosome lacking an inversion in that region. Different sets of DNA primer sequences may be designed to amplify DNA fragments of the same or different sizes, but the principal accomplishment is the same as long as the primers flank either of the inversion sites found to accompany the Tobiano gene or to occur within the region to detect genetic markers associated with the inversion. The chromosome inversion, or the DNA sequence associated with the inversion, may be detected by any suitable method of DNA analysis, including without limitation, by gel electrophoresis, Southern blotting and specific DNA sequence analysis. Advantageously, the presently described method may be performed on any suitable fresh or stored sample containing a nucleic acid, including without limitation a cell, a tissue, a hair follicle, buccal swab, serum, plasma, and other biological materials derived from an individual.

These and other embodiments, aspects, advantages, and features of the present invention will be set forth in the description which follows, and in part will become apparent to those of ordinary skill in the art by reference to the following description of the invention and referenced drawings or by practice of the invention. The aspects, advantages, and features of the invention are realized and attained by means of the instrumentalities, procedures, and combinations particularly pointed out in the appended claims. Various patent and non-patent references are discussed herein. Unless otherwise indicated, any such references are incorporated in their entirety into the present disclosure specifically by reference.

BRIEF DESCRIPTION OF THE DRAWINGS

The accompanying drawings incorporated in and forming a part of the specification, illustrate several aspects of the present invention, and together with the description serve to explain the principles of the invention. In the drawings:

FIG. 1 compares fluorescence in situ hybridization (FISH) analyses of common and Tobiano ECA3 chromosomes;

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FIG. 2 schematically depicts the positions of various genes located on equine ECA3 in common (non-Tobiano) ECA3 chromosome and in the Tobiano ECA3 chromosome having an inversion;

FIG. 3 shows the sequences surrounding the proximal (centromeric) breakpoint of a chromosomal inversion in Tobiano ECA3, with primer binding sites and the region of the breakpoint shown in double underlining, and the breakpoint address identified by a vertical line (see SEQ ID NO: 13);

FIG. 4 shows the sequences surrounding the distal (telomeric) breakpoint of a chromosomal inversion in Tobiano ECA3, with primer binding sites and the region of the breakpoint shown in double underlining, and the breakpoint address identified by a vertical line (see SEQ ID NO: 14);

FIG. 5 schematically depicts a primer strategy used to detect the inversion in ECA3; and

FIG. 6 shows a representative result for the present method for detecting a chromosomal inversion in ECA3 by PCR, with amplification products detected by gel electrophoresis and ethidium bromide staining, with the marked 152 bp product being indicative of the common (non-Tobiano) ECA3 and the 209 bp amplification product being indicative of the ECA3 inversion (Tobiano).

DETAILED DESCRIPTION OF THE ILLUSTRATED EMBODIMENTS

In the following detailed description of the illustrated embodiments, reference is made to the accompanying drawings that form a part hereof, and in which is shown by way of illustration, specific embodiments in which the invention may be practiced. These embodiments are described in sufficient detail to enable those skilled in the art to practice the invention. Also, it is to be understood that other embodiments may be utilized and that process, reagent, software, and/or other changes may be made without departing from the scope of the present invention.

EXAMPLE 1

As noted above with respect to genetic markers useful for cytogenetic demonstration of the inversion, one BAC clone (558) mapped to a single location, near KIT, on chromosomes without TO but at two locations, associated with the inversion sites, on chromosomes possessing TO (FIG. 1). This demonstrated that BAC 558 included one of the two sites at which the inversion occurred. Therefore, it was possible to sequence BAC 558 to identify the distal breakpoint for the inversion site.

The homologous human genome position of the end sequences for BAC 558, shown in FISH experiments to contain the distal breakpoint of the inversion, was determined by searching the database available at the Horse Genome Project web site (Leeb et al., 2006). Within these endpoints regions of high conservation approximately 3-5 kb apart were selected based on the conservation track of the UCSC Genome Browser (Kent 2002). The human sequence for these conserved areas was then used to find a corresponding horse sequence from the trace files submitted to the Entrez Trace Archive using a Discontiguous Megablast (NeBI, 2006). The relative position of these trace files was confirmed by comparing them back to the human genome using the BLAT search on the UCSC Genome Browser. Primers were designed for each individual trace using the Vector NTI software package (Invitrogen, Carlsbad Calif.).

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Primer pairs first optimized to amplify the small product contained in the trace file were then used with the appropriate primer from the next trace to create a product containing the sequence between the two distant trace files. Thirty seven primer pairs were created, and amplification of the intervening sequence was confirmed using DNA from a known TO/TO horse until a primer pair was found where amplification was successful in the non-Tobiano but not the TO/TO horse, indicating that the primer pair had been separated by the inversion. That primer pair is set forth in Table 2.

TABLE 2

Primers used to screen for the distal ECA3 inversion breakpoint.			
Trace File Accession #	Human Address (Chr4: in Mb)		
		<u>Forward Primer 5'-3'</u>	
1250295308	55400840	CTGCCTCCATCAGAGCCTGCA	(SEQ ID NO: 1)
		<u>Reverse Primer 5'-3'</u>	
1378686387	55402968	TGGAGGTGACACAGTGCCTTCACT	(SEQ ID NO: 2)

This set of primers was used to screen animals for the presence of the chromosomal inversion. Absence of a PCR product indicated that the primers were separated by the inversion. This test required use of a positive PCR control fragment, and only detected homozygosity.

The sequence between the two trace files identified was then determined. The sequence located at the distal breakpoint in a TO/TO horse was determined by genome walking using the APAGene GOLD Genome Walking Kit (BioS&T, Ontario Canada) following the manufacturers directions. Briefly, several rounds of nested PCR were performed using sequence specific primers for the known sequence and universal primers to extend the amplified product in to unknown sequences for which specific primers could not be designed. The primers used were as follows:

BPFa: (SEQ ID NO: 3)
GAT CAG TGT AGA CGT AGT GTG ACA GAG ACC CAG G;

BPFb: (SEQ ID NO: 4)
GGC AAA CAG AGG GCC AAA TGA TAG ATC AGT GTA GAC;

BPFc: (SEQ ID NO: 5)
TGT GCT CTA GCA AGG GTG GAG CAT GTT TCA TTA AG.

The resulting PCR product was directly sequenced using an ABI310 (Applied Biosystems, Foster City, Calif.) following the manufacturers standard protocol for chemistry and conditions. Fifty-two bases of horse sequence beyond the inversion breakpoint were generated (with reference to Genbank Accession #EF442014) and used by Megablast of the Trace Archive to pull trace files containing that sequence (NCBI, 2006). The position of these trace sequences relative to the human genome was determined using the BLAT function of the UC8C Genome Browser. The equine sequence encompassing both breakpoint boundaries was determined by assembling these trace files with the ContigExpress function of VectorNTI (see FIGS. 3 and 4). The Genbank accession numbers of the trace files used to create the assembly were:

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Centromeric end: 1231203553,1288909375, 1248629699, 1411501757, 1204375030, 1276786693, 1248632879, 1419986588,1286709380,1387598584, 1273872762, 1207301472, 1417784590, 1326270692; and

Telomeric end: 1250295308, 1199900846, 1465783553, 1304105951, 1258859536, 1227749574, 1248814034, 1213964577, 1378686387.

The DNA sequence at the site of the inversion was then identified by sequence walking using DNA from a homozygous (TO/TO) horse as a template (Genbank Accession #EF442014). This sequence, once extended using an alignment of the available equine whole genome sequence trace files from the trace archive at NCBI, was used to design primers that amplify PCR products of unique sizes for each of the inverted and non-inverted chromosomes. The sequences of the assembled contigs for the common (non-Tobiano) and Tobiano sequences are set forth as SEQ ID NO: 6 and SEQ ID NO: 7, respectively.

For allele identification by PCR, all primers were designed using Vector NTI. A common forward primer was first designed using the sequence just outside of the telomeric end of the inversion. Then a primer specific for the adjoining sequence within the inverted region in the common chromosome and a primer specific for the tobiano inversion were designed to produce two different size PCR products of 209 bp (SEQ ID NO:11) and 152 bp (SEQ ID NO:12), respectively (see FIG. 5). The primer sequences used were as follows:

ECA3-F:

(SEQ ID NO: 8)
TGA TAG ATC AGT GTA GAC GTA GTG TGA CAG AGA C;

ECA3xR:

(SEQ ID NO: 9)
AAC AGC TAC TCC CAC TCT AGC ATA GGT TC;

ECA3toR:

(SEQ ID NO: 10)
TTC ACC ACA GAG TAT CCA ATT ATG TCT TTC ACA TAA

TGC.

PCR was preformed using FastStart polymerase (Roche, Indianapolis Ind.) following in a 10 μ L volume using 25 ng extracted DNA or 2 μ L of hair lysate prepared using 5-7 hair bulbs according to the method previously published by Locke et al., 2002. Primer concentrations were adjusted as follows to produce two bands of approximately equal intensity: 0.5 μ M ECA3-F primer (1 μ L of 5 μ M dilution), 0.325 μ M ECA3xR primer (0.65 μ L of 5 μ M), 0.175 μ M ECA3toR primer (0.35 μ L of 5 μ M). All other reagents were used following the manufacturers recommendations. Thermocycling was performed in a PTC-200 thermocycler (MJ Research Inc., Boston, Mass.) with an annealing temperature of 57° C. and 20 s extension time for 35 cycles. Products were visualized by electrophoresis on a 3% agarose gel after staining with ethidium bromide (FIG. 6). Presence of the 152 bp PCR product (SEQ ID NO:12) indicated the horse carried at least one copy of the common chromosomal arrangement. Likewise, presence of the 209 bp product (SEQ ID NO:11) was indicative of the presence of the inversion.

The chromosomal inversion has been defined by identifying addresses in the equine genome. With reference to the equine genome EquCab2, available on the UCSC Genome Browser, Genome Bioinformatics Group, Center for Biomolecular Science and Engineering, University of California Santa Cruz, the telomeric breakpoint is located between nucleotides chr3:41925230-41925231 of EquCab2. The cen-

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tromeric breakpoint is located between nucleotides chr3: 77663330-77663331 of EquCab2. Those breakpoints, along with the 5 kb surrounding those addresses, are set forth in FIGS. 3 and 4 (see SEQ ID NO:13 and SEQ ID NO: 14). The 5 kb nucleotides surrounding the centromeric and telomeric breakpoint addresses are set forth in SEQ ID NO:13 and SEQ ID NO:14, respectively, as derived from the publically available ECA3 (EquCab2, UCSC Cenome Browser). These sequences are depicted also in FIGS. 3 and 4, respectively, with the breakpoints indicated by a vertical line and the binding sites for the primers ECA3-F (SEQ ID NO: 8), ECA3xR (SEQ ID NO: 9), and ECA3toR (SEQ ID NO: 10) indicated by underlining and by arrows extending from the respective primer designators.

EXAMPLE 2

To further evaluate the ability of the PCR method set forth in Example 1 to detect the chromosomal inversion, additional stored samples were obtained for molecular marker studies and DNA sequencing. Previously banked DNA samples from 58 Thoroughbreds (non-tobiano) and 121 tobiano horses of various breeds were selected from samples available at the Immunogenetics and Genomics Laboratory, University of Kentucky. Photographs, pedigree and registrations were examined for all tobiano horses in order to verify presence of the tobiano pattern. Specifically, these horses possessed large white patches that crossed the dorsal midline and had smooth borders. None of the total 179 horses were related as siblings or half-siblings. All tobiano horses had at least one tobiano patterned parent. The breeds of horses evaluated, and the results of the present method for detecting the chromosomal inversion, are provided in Table 3.

TABLE 3

Breeds of Horses tested for the ECA3 inversion.

Breed	inv/inv	inv/+	+/+	
American Miniature Horse	2	3		
American Paint Horse	16	53		
Arab/Pinto		5		
Draft and Draft Cross	2	4		
Icelandic Horse		1		
Missouri Fox Trotter		2		
Pinto		6		
Saddlebred		2		
Shetland Pony	1	3		
Spotted Saddle Horse		3		
Tennessee Walking Horse	2	12		
Thoroughbred (non-Tobiano)			58	
Welsh Cross	2	2		
Total	25	96	58	179

Of 121 horses with the tobiano pattern, all possessed the PCR amplified 209 bp DNA fragment (SEQ ID NO:11) consistent with the inversion. These horses included the following breeds: American Miniature (5), American Paint (69), Arabian (5), draft and draft crosses (6), Icelandic Horse (1), Missouri Fox Trotter (2), Saddlebred (2), Shetland Pony (4), Spotted Saddlehorse (3), Tennessee Walking Horse (14), the Welsh Cross (4) (Table 2). These breeds encompass a wide range of historical breed origins. None of 58 control Thoroughbred horses without the tobiano pattern possessed the inversion, including 3 that carried the KMI marker.

The tobiano pattern is usually, but not always, associated with the haplotype possessing alleles ALB-B, GC-S and KMI. Therefore, horses with other haplotypes were tested for

the presence of the inversion. These haplotypes and their association with the inversion are shown in Table 4. The inversion was present with alleles ALB-A, GC-F and KMO, in addition to the alleles from the most frequent haplotype. Therefore, it is likely that the Tobiano inversion event occurred on a chromosome possessing ALB-B, GC-S and KA1, followed by rare mutations or genetic recombination resulting in new haplotypes possessing TO.

TABLE 4

Haplotype association of genetic markers for Albumin (ALB), Vitamin D binding Protein (GC), KIT intron 13 SNP (KM) with TO and the ECA3 inversion.							
Horse ID#	Breed	Phenotype	AL	Gc	KM	ECA3 Inversion	TO Haplotypes (AL/Gc/KM)
78	APHA	TO	A	F	1/0	inv/+	A/F/unknown
351	APHA	TO	B	F/S	1/1	inv/inv	B/F/1, B/S/1
481	Draft cross	TO	B	F	1/1	inv/+	B/F/1
480	Welsh cross	TO	B	S	0/0	inv/inv	B/S/0
479	Welsh cross	TO	B	S	1/0	inv/inv	B/S/1, B/S/0

EXAMPLE 3

The experiment set forth in Example 1 is repeated using PCR primers designed to amplify the centromeric site of the inversion for those horses with the Tobiano inversion. The PCR and gel electrophoresis conditions are as set forth in Example 1. In this case, the primers would flank the inversion site shown in FIG. 3. Primers ECA3xR (SEQ ID NO:9) and ECA3toR (SEQ ID NO:10) are used, along with a primer ECA3Fc (SEQ ID NO:15; FIG. 3) having a complementary sequence ATGTGCAGTACTCATAATACATTC. Use of this primer combination produces a DNA fragment of approximately 261 basepairs for chromosomes without the inversion and a fragment size of approximately 204 basepairs for chromosomes possessing the inversion.

EXAMPLE 4

The inversion can be inferred using the technique of restriction fragment length analysis (RFLP). In this approach: 1) DNA fragments are produced including one or both of the chromosome inversion sites; 2) the DNA is digested with restriction enzymes; and 3) the fragments are analyzed by gel electrophoresis for fragment length differences denoting the presence or absence of chromosomes with inversions. Any DNA restriction enzymes recognizing sites on both sides of the inversion as described herein is effective in detecting the presence of fragments characteristic of the presence of the inversion. Experimental techniques for RFLP analysis are as set forth in Molecular Cloning, A Laboratory Manual, Sambrook and Russell, 2001, Chapter 5.

EXAMPLE 5

Southern blotting (see Chapters 5-6, Sambrook and Russell, 2001) is a method of hybridizing DNA fragments to chromosomal DNA fragments in order to visualize the presence of homologous fragments. In this method, chromosomal DNA is fragmented in a method that produces reproducible cuts, usually with DNA restriction enzyme. The fragments are separated based on length by gel electrophoresis. Subsequently, DNA, called a probe, homologous to DNA from the

same region is labeled such that it can be visualized and subsequently hybridized to the chromosomal DNA. Differences in DNA fragment lengths based on the presence or absence of the inversion can be detected by visualizing differences in the size of fragments hybridizing to the probe.

EXAMPLE 6

As shown in FIGS. 3 and 4, the DNA sequences surrounding the inversions are unique to horses possessing the chromosome inversion. DNA sequencing through the inversion site would effectively demonstrate the presence or absence of the normal or inverted chromosome fragment.

The presence of a chromosomal rearrangement in ECA3 of animals bearing the Tobiano coat color genotype is now demonstrated, and a method is set forth for detecting the Tobiano coat color genotype in an equine animal, based on detection of the large, paracentric inversion in equine chromosome 3. This inversion described in the foregoing description does not disrupt the coding sequence of any known genes; consequently it is not readily apparent how this may cause the tobiano phenotype. However, without being restricted to any particular theory, the inversion may disrupt conserved non-coding or regulatory regions of the KIT gene, similar to what has been observed in the murine Bump-white (Stephenson et al. 1994) and Sash (Nagle et al. 1995) inversions. Both of these inversions disrupt regulatory elements 5' to the KIT gene and lead to inappropriate temporal and spatial expression in the embryo (Duttlinger et al. 1993; Nagle et al. 1995). Regulatory gene elements that form a loop structure, like enhancers, insulators, and locus control regions, can bind sequences both upstream and downstream of their target gene (Maston et al. 2006). Regulatory elements such as these, encompassing the KIT gene from the intergenic regions upstream to those downstream, could explain the similarities in appearance between the Tobiano phenotype, which is a rearrangement beyond the 3' end of KIT, and the Rump-white and Sash mouse phenotypes, which are both rearrangements of the 5' end of KIT.

Horses with Tobiano are normal and healthy, suggesting that this chromosome rearrangement is not associated with deleterious health effects. Presence of the inversion among breeds of diverse origins indicates that it predates divergence of these horse and pony breeds. So far there no exceptions have been found for the association of this inversion with the tobiano phenotype, strengthening the hypothesis of homogeneity for the phenotype and genotype. The presently-described PCR-based method for identifying the Tobiano genotype will be useful to breeders interested in finding breeding stock homozygous for tobiano since carriers and Tobiano homozygotes are usually phenotypically indistinguishable. The method can be utilized on any material with DNA, and does not require viable cells. The method thus advantageously does not require freshly obtained genetic material, and even more may be performed on a variety of biological sample types such as hair, with the proviso that the biological sample contains DNA. Hence, more individuals can be subjected to routine screening than would be practical using a more expensive and time-consuming assay such as the FISH method.

One of ordinary skill in the art will recognize that additional embodiments of the invention are also possible without departing from the teachings herein. This detailed description, and particularly the specific details of the exemplary embodiments, is given primarily for clarity of understanding, and no unnecessary limitations are to be imported, for modifications will become obvious to those skilled in the art upon

reading this disclosure and may be made without departing from the spirit or scope of the invention. Relatively apparent modifications, of course, include combining the various features of one or more figures with the features of one or more of other figures.

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SEQUENCE LISTING

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<212> TYPE: DNA

<213> ORGANISM: Equus caballus

<400> SEQUENCE: 7

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tcccccttct gagagctgtc agttgagagg cagtatctc ttgtgagaga gtttgggtgc	180
ggctgcaaaa ttaaaggcaa gaaataacca caaataagag agcatgagtg gccagatata	240
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<210> SEQ ID NO 8
<211> LENGTH: 34
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Common forward primer

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<400> SEQUENCE: 8

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<210> SEQ ID NO 9
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: non-tobiano primer

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<400> SEQUENCE: 9

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<210> SEQ ID NO 10
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
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<223> OTHER INFORMATION: tobiano reverse primer

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<400> SEQUENCE: 10

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<210> SEQ ID NO 11

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<400> SEQUENCE: 11

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<210> SEQ ID NO 12
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 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Non-tobiano 152 bp PCR product

<400> SEQUENCE: 12

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<210> SEQ ID NO 13
 <211> LENGTH: 5001
 <212> TYPE: DNA
 <213> ORGANISM: Equus caballus

<400> SEQUENCE: 13

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gatcatagct gaaatagctg ctgtttttac cttgaacact gttctctggt acaacaaaaa	600
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<210> SEQ ID NO 14

<211> LENGTH: 5001

<212> TYPE: DNA

<213> ORGANISM: Equus caballus

<400> SEQUENCE: 14

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24

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What is claimed is:

1. A method for screening for a genotype for a Tobiano coat color pattern, comprising:

obtaining a biological sample containing a nucleic acid sample from an equine animal, said nucleic acid sample comprising equine chromosome ECA3q; and

identifying a chromosomal inversion in a chromosome ECA3q, said inversion having a telomeric breakpoint located between a KIT gene and a KDR gene in a band ECA3q21 of said chromosome ECA3q and a centromeric breakpoint located between a ADH1D gene and a UNC5C gene in a band ECA3q13 of said chromosome ECA3q;

said step of identifying a chromosomal inversion comprising detecting at least one of a nucleic acid sequence surrounding the telomeric breakpoint of the inverted ECA3q chromosome and a nucleic acid sequence surrounding the centromeric breakpoint of the inverted ECA3q chromosome, or sequences complementary thereto;

wherein the presence of the chromosomal inversion as indicated by detection of at least one of said telomeric breakpoint or said centromeric breakpoint indicates that the equine animal possesses a genotype for a Tobiano coat color pattern.

2. The method of claim 1, wherein the centromeric breakpoint is between addresses chr3:41925230-41925231 of EquCab2 and the telomeric breakpoint is between addresses chr3:77663330-77663331 of EquCab2.

3. The method of claim 1, wherein the sequence surrounding the telomeric breakpoint of the inverted ECA3q chromosome is set forth in SEQ ID NO: 14 and the sequence surrounding the centromeric breakpoint of the inverted ECA3q chromosome is set forth in SEQ ID NO: 13.

4. The method of claim 1, wherein the step of detecting at least one of said telomeric breakpoint or said centromeric breakpoint comprises:

hybridizing a group of probes or primers consisting of the sequences set forth as SEQ ID NO: 8, SEQ ID NO: 9, and SEQ ID NO: 10 or sequences complementary to SEQ ID NO: 8, SEQ ID NO: 9, and SEQ ID NO: 10 to the nucleic acid;

amplifying the hybridized group of probes or primers; and

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detecting the presence or absence of a 209 bp amplification product indicative of the presence of the chromosomal inversion.

5. The method of claim 4, wherein the hybridized group of probes or primers are amplified by PCR.

6. The method of claim 1, wherein the biological sample is selected from the group consisting of a cell, a tissue, a hair follicle, a buccal swab, serum, plasma, whole blood, and a combination thereof.

7. The method of claim 4, wherein the presence of the 209 bp nucleic acid sequence is detected by any one of gel electrophoresis, DNA sequence analysis, or Southern blotting.

8. The method of claim 4, wherein the 209 bp nucleic acid sequence is set forth in SEQ ID NO:11.

9. A method for screening for a genotype for a Tobiano coat color pattern by detecting a chromosomal inversion in an equine chromosome ECA3q, comprising:

obtaining a biological sample containing a nucleic acid from an equine animal;

hybridizing to said nucleic acid the group of probes or primers comprising the sequences set forth in SEQ ID NO: 8, SEQ ID NO: 9, and SEQ ID NO: 10, or sequences complementary to SEQ ID NO: 8, SEQ ID NO: 9, and SEQ ID NO: 10;

amplifying said hybridized group of probes or primers; and detecting the presence or absence of a 209 bp amplification product;

wherein detection of said 209 bp amplification product indicates the presence of an inversion in an ECA3q chromosome contained in said nucleic acid sample that is predictive of a Tobiano coat color genotype.

10. The method of claim 9, wherein the group of probes or primers are amplified by PCR.

11. The method of claim 9, wherein the presence of the 209 bp nucleic acid sequence is detected by any one of gel electrophoresis, DNA sequence analysis, or Southern blotting.

12. The method of claim 9, wherein the nucleic acid is obtained from a sample selected from the group consisting of a cell, a tissue, a hair follicle, a buccal swab, serum, plasma, whole blood, and a combination thereof.

13. The method of claim 9, wherein the 209 bp nucleic acid sequence is set forth as SEQ ID NO:11.

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