



9-21-2010

Gilvocarcin Gene Cluster, Recombinant Production and use Thereof

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Recommended Citation

Rohr, Jürgen and Fischer, Carsten, "Gilvocarcin Gene Cluster, Recombinant Production and use Thereof" (2010). *Pharmaceutical Sciences Faculty Patents*. 48.

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US007799904B2

(12) **United States Patent**
Rohr et al.

(10) **Patent No.:** **US 7,799,904 B2**
(45) **Date of Patent:** **Sep. 21, 2010**

(54) **GILVOCARCIN GENE CLUSTER,
RECOMBINANT PRODUCTION AND USE
THEREOF**

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Carsten Fischer, Lexington, KY (US)

(73) Assignee: **University of Kentucky Research
Foundation**, Lexington, KY (US)

(*) Notice: Subject to any disclaimer, the term of this
patent is extended or adjusted under 35
U.S.C. 154(b) by 667 days.

(21) Appl. No.: **10/866,089**

(22) Filed: **Jun. 14, 2004**

(65) **Prior Publication Data**

US 2005/0048536 A1 Mar. 3, 2005

Related U.S. Application Data

(60) Provisional application No. 60/477,957, filed on Jun.
13, 2003.

(51) **Int. Cl.**

C12N 15/00 (2006.01)

C12N 15/11 (2006.01)

C12N 15/52 (2006.01)

C12N 5/10 (2006.01)

C12N 1/21 (2006.01)

(52) **U.S. Cl.** **536/23.2**; 435/320.1; 435/477;
435/252.3; 536/23.1; 536/23.7

(58) **Field of Classification Search** None
See application file for complete search history.

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(57) **ABSTRACT**

A nucleic acid molecule encoding the gilvocarcin V gene cluster and subunits thereof. Recombinant vectors and host cells comprising a nucleic acid compound encoding the gilvocarcin V gene cluster or subunits thereof. Host cells comprising recombinant vectors encoding the gilvocarcin polyketide synthase and gilvocarcin post-PKS modifying enzymes from *Streptomyces griseoflavus* can be used to produce gilvocarcin and functional gilvocarcin mutants, analogs and derivatives thereof with application as antibiotics, anti-cancer agents, immunosuppressants, antivirals, and neuro-protective agents.

7 Claims, 27 Drawing Sheets

Figure 1.

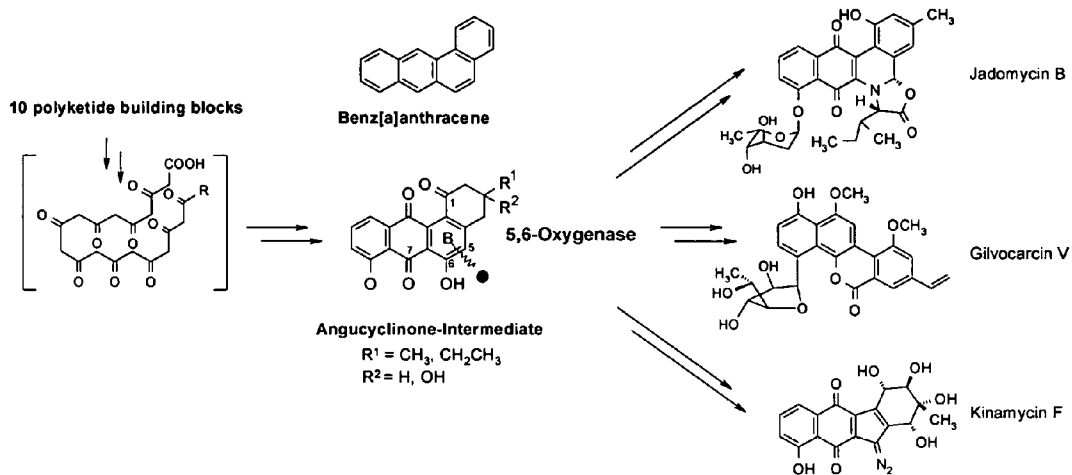
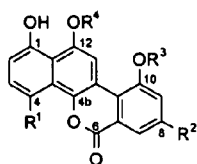


Figure 3.



Gilvocarcin-Type Anticancer Drugs

	R ¹	R ²	R ³	R ⁴
Gilvocarcin M:	α -D-Fuc	CH ₃	CH ₃	CH ₃
Gilvocarcin E:	α -D-Fuc	CH ₂ CH ₃	CH ₃	CH ₃
Gilvocarcin V:	α -D-Fuc	CH=CH ₂	CH ₃	CH ₃
Defucosyl-Gilvocarcin V:	H	CH=CH ₂	CH ₃	CH ₃
Ravidomycin:	β -D-Rav	CH=CH ₂	CH ₃	CH ₃
Deacetyl-Ravidomycin:	DeAc- β -D-Rav	CH=CH ₂	CH ₃	CH ₃
Deacetyl-Ravidomycin M:	DeAc- β -D-Rav	CH ₃	CH ₃	CH ₃
FE35A:	N,N-MeAc-DeAc- β -D-RaV	CH=CH ₂	CH ₃	CH ₃
FE35B:	N-Ac- β -D-RaV	CH=CH ₂	CH ₃	CH ₃
Chrysomycin A:	β -D-Vir	CH=CH ₂	CH ₃	CH ₃
Chrysomycin B:	β -D-Vir	CH ₃	CH ₃	CH ₃
BE-12406 A:	H	CH ₃	CH ₃	α -L-Rha
BE-12406 B:	H	CH ₃	H	α -L-Rha

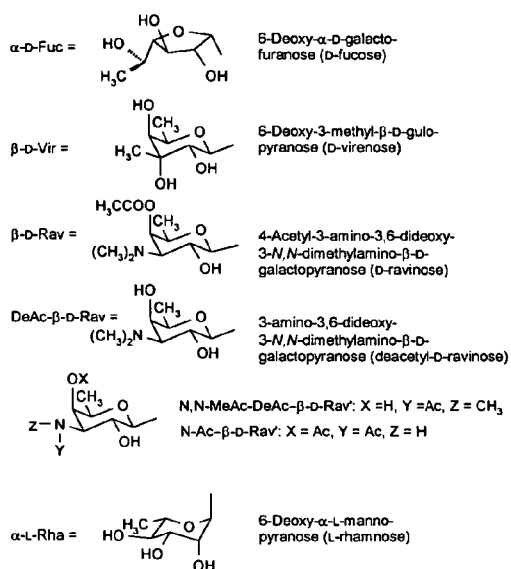


Figure 4.

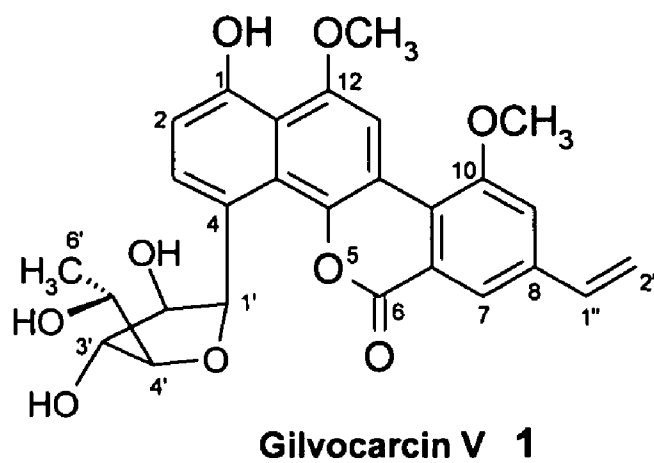


Figure 5.

**Mechanism of Action of
Glvocarcin V**

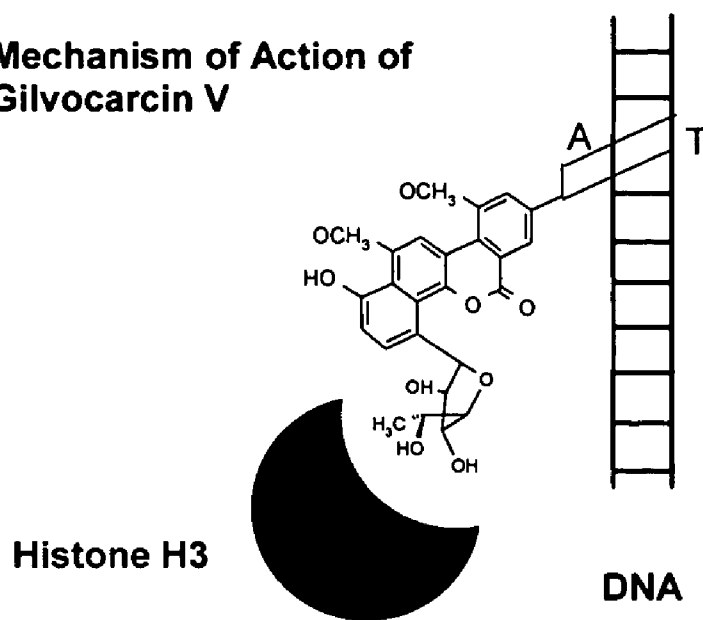
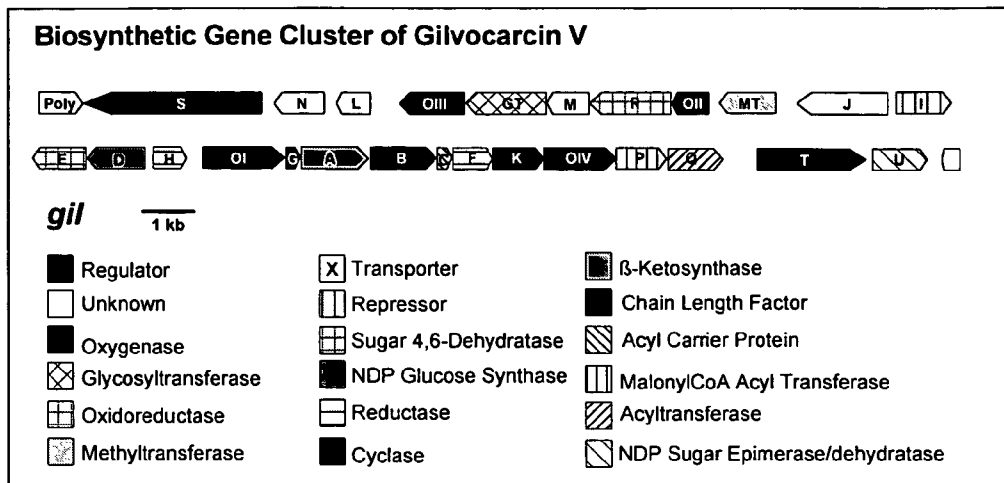


Figure 6.

A.



B.

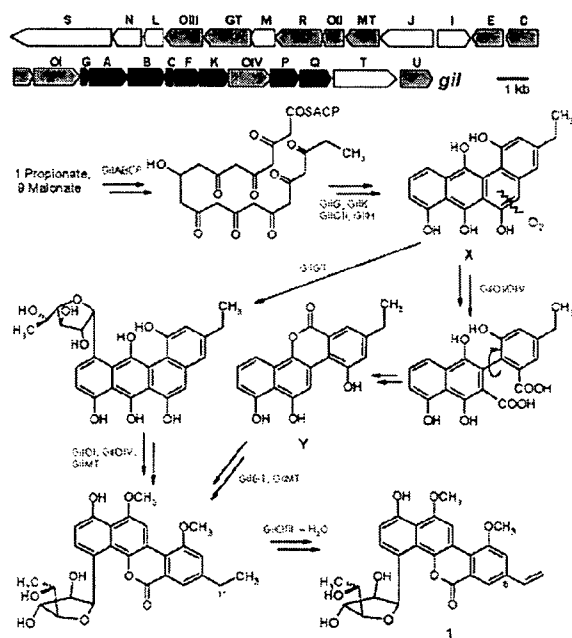


Figure 7.

Proposed Biosynthetic Pathway for the GV-sugar moiety

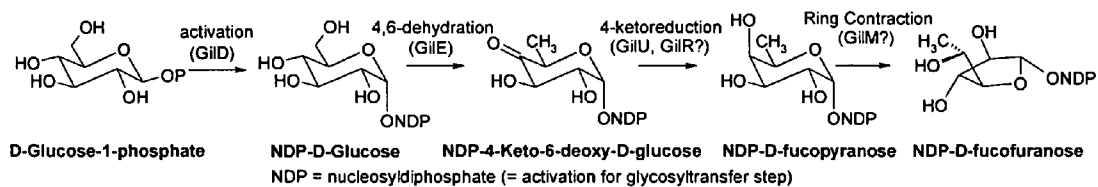


Figure 8.

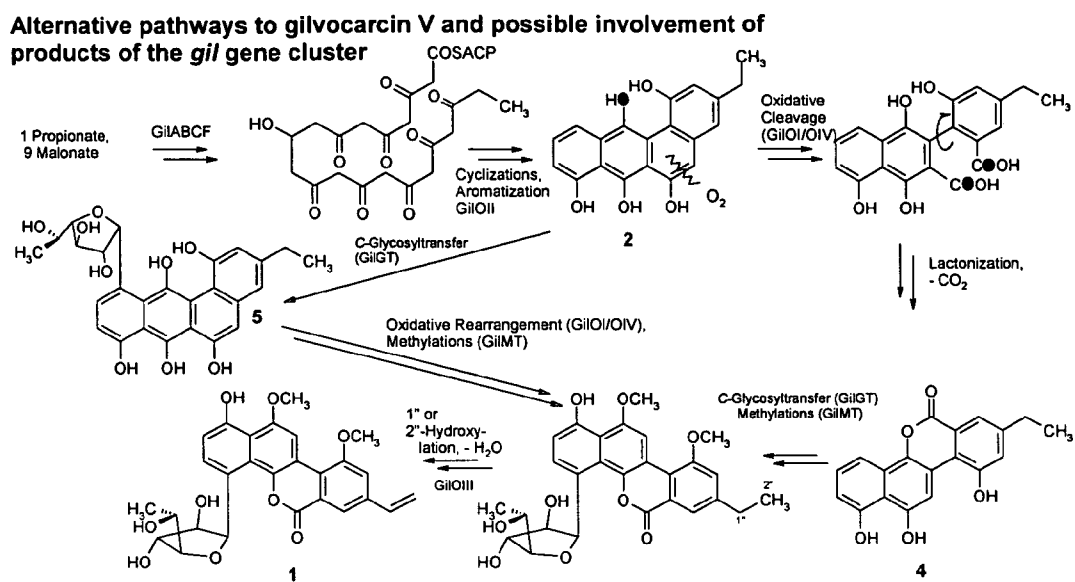


Figure 9. cont.

1821 ACGCGACGTGAGCGGTGTGCCCGGGCGGCTGGGACGGGGTTCGGGCGGGCGGGTTCGTCCTGGTTCG

1886 GGGCCGGGTTCGACGATCGCGGTGAGGGCGCGTTCGGCGACGGCGGGCTACCAGCGCAGGGCGGT

1951 CTCGGCGGGGTGCTCGCGCGTGCCGGTCCGAGGGCCGGCGTCTCGCGGGGTGCGGCGTCTCGC

2016 GGGGTGCGGCATCTCGCGGAGTACGGCCTCCTCGCGGGGTGCGGCGCCCTCGCGGGGTGCGGGC

2081 CCCTCGCGGAGTACGGCGCCCTCGCGGAGTACGGCGCCCTCGCGGAGTACGGCGCCCTCGCGGAG

2146 TACGGCATCTCGCGGAGTACGGCATCTCGCGGAGTACGGCGTCTCGCGGAGTACGGCGTCTCT

2211 CGCGGGGCGCGGTCTGCGGACCCGCGCCGCTCGGGCTCGTTCGCGGGCGAAGTCACGGACCAGG

2276 TCGTGGGGACGTAGCGGCCGTACGCGGTCTCTCCACGAGGGCCACGTCGACGAGCCGGTCCAG

2341 CGCGGCCTCCGCCCGGGTTCGCCGGTGCCTGGTGCAGCCGGGCGAGCAGCGGCGCGCCGTAGGGCG

2406 GCAGGTCGAGCGCGCCGATGCGGCACAGGGCGAGGGCGGGTTCGCGGTCCGTCTCACGGTCCGAG

2471 ACGGTGAGCGCGTCTGCGCGACGGCCAGCGAGCGGGCGCACGCTGAGGTCTGCTACTCCAGGTG

2536 CGGCAACCGGCTGTCTGGTGGCGGAGAGCTGACCGGCGAGGTCTGTCGGGCGTGAGGGCCCGGCGCG

2601 CGGCGAGCCGGGCCGCGACCACCCGAGGGCCAGCGGAAGCCGGCCGGTGCAGCGGACGAGCGGG

2666 TGCCCGGCGCCGAGACCGTCCCGGCCGAGACCGCCCGCAGCAGGGCGGCACTGTCTCTCGTCGGA

2731 CAGCGGGCCGAGCGGGACACGGACGGCACCGTTCGAGCGTGGTGAGCGGCGAACGGTGGTGACGA

2796 TCACCGCACAGCCGGGTCCGCCCGGCAGCAGCGGCCCGCACCTGCGCGGCGTCCGCGGCGTCTCC

2861 AGCACCAGGAGGGTGCGGTGGGCGGAGCAGCGAGCGCAGCAGGGCGGGCGCGTCCGGCCG

2926 TTCGGGGACGGCGCAGGGTTCGGTGGCCAGGTTCGCGCAGGAGAGCGTGCAGGGCCTGGGCGGGG

2991 TGAGGGGGTTCATGCCGGGGTCTGTCCTGTCAGGTTGACGTAGAGCTGACCGTCGACGAAACGT

3056 TCCGCCAGTGTGTGTGCGACCTGGACGGCGAGCGCGTCTTCCCCACACCAGCGTACCGCTGAC

3121 GACGACGGCAGGGGGCCCGGCGGGGCGGGGGCACGGGTGAGCACACGGATCAGCTCGTCCC

3186 GCACCGCTCCCGCCCGTGAAGTGAGCAGGCGGGGCGGAGTTGCGCCGGCTGGGCGGCACA

3251 CCGCCCCCTCTGCGGCTCCGCGCACCGACCGTCTCTGCGCTCCCCCTGCCCCCGCAGCAC

3316 CTCCACGTGGGCCTCGCGACCCCCGGCCCCGGTTCGACGCCGAGTTCGTCCGCCAGGGGGCCC

3381 GCAGATCCCGGTGGACCACCAGCGCCTCCGCCTGACGGCCGGTGCAGGCGGAGCATCAGC

3446 TGACGGTGGTACGACTCCCGCAGCGGATGTTTCGGCGGCCAGTCCCGCCAGTTCGGGCACGAGATC

3511 GGCCAGGCGCTCGCCGCCAGGGCCAGTTCGGCGTCTGACCGCCACTCCAGGAGCAGCAGCCGGC

3576 CCTCGCGCAGCCCGCACAGGGCGTAGCCGCCACTTCCGGGGGCATCCCGGCCAGCGGGTTC

Figure 9. cont.

3641 CCCC GCCACAGCGGAGCGCGGGCGGCACACTCGCGGCCACCCGCTCCAGTCCCGGGCGGTGTG *SmaI*
 3706 CGCGGCGCGGGCGGCTGCGGTGTGCGCGTGAAGACCTGGACGTCCACCTCGCCCTCTCCACCC
 3771 GCAGCAGATATCCGGTGGGCACGGTGAGCAGCCGGCCCGGGTCGTGAGCAGCCGCCGCAACCCG *EcoRV* *SmaI*
 3836 GCGACGTGATTGTGCAGCGAGGGCAGCGCGGAGACGGGCGGCACCCGCCACAGGGCGTCTT
 3901 CAGCGCTCGACGGACACGACCCGCCCGGCGTGCAGCAGCAACGCGACGAACAGCGCACGGAGTT
 3966 TGGGACTTCCGGTACCTGGACGGACCCCGGGCGTGGGCCCTCGCCGTGTCACAGCACCGGT *SmaI*
 4031 GTTCCAGCAGTCCGAACCGCAGTCCGCGCCGTGT**CACG**CCGACCACTGCCTTCCGGGTGACCG <- *gils*
 4096 AACGGACAACCAATGGCCTTATGTTAGCGATCCGTTGGCAAAGTCTGATGTGATCACTACATCGG *BamHI* *SmaI*
 4161 ATCCGGCCCGGGGTGCGCGGTAGAACGCGCAGCTCGGGGGAGTGTGCGCGCCGCGGCCCGGTC
 4226 CGCACACGAGGGTTCCCGGTCCGACAGAGGTGAACGGCCGGGCCCTCGTTCTCTTCTGCCGCCG *******
 4291 GTTCGCCCTTCGGGT**TCAG**ATGACGGGCGGCCGTCCGAGCCGGGTGAGCCGCCACACCGTCCGCCA
 4356 GCGCATGGCCCGCCGCTCCCCGGCCGGCTCCCGCAGCCCCTCCGCGAAACCGCCGAACCAGGGCG
 4421 GCAGCCCTGGACCGACCGAGTCCGCAGCAGGGTGAGCAGGACCCACACCCCGAGGTGCACGGGG
 4486 ATCAGCGCGAGCGGCAGCCGCCCGGGCCAGCCAGACCCGGTTGCGGGCGTTACCCGGAAGTA
 4551 GATCGCGTGCCGGGCCGGCGAGGTCTTGGGGTGCTGGAGCAGCAGGTCCGGCGCGTAGAGGATGC
 4616 GCCACCCCGCGTCCGGCGCACGCCAGGCGAGGTCCGTCTCCTCGTGCGGAAGAAGAACGCGCCG
 4681 GGCCAGTCCCGATCTCGTCGAGCATCGACATCCGCAGCGCGTGCCCGCCCCGAGGAACCCGGT
 4746 GACGTACCCGCCCTTCATCGGGTCCGCCTTCCGAGCCGGGGCACGTGCCGCTGCTGCGTCTCC
 4811 CCAGCTCGTCGGCGATCCGGAAGCCGACGACCCGAGCCGGTTCGTCGCCCGGTACAACCTCCCC
 4876 ACCCGGCGCAGCACATCGGCGTCCACCAGCAGACCGTCTGTCGTCAGTTCGACGACGACGTCCAC
 4941 GTCCCCGAACTCCCGCAGCCGCTCGATCCCCACGTTCCGCCCGCCCGGGCAGCCGAGGTTCTCCG *SmaI*
 5006 CCAGCTCGACGGTGGTGACCTCACCGGGCAGGGACAGCCGCCGGGCGAACTCGGGCAGCGGACAG
 5071 CCGTCCCCACGATCACGATCCGCGCGGGCGCCACGTCTGCTTCGCCACGGACTCCAGCAGCGC <- *gILN*
 5136 GTCCACCTCGGCCGGCCGGTTCCCCATGGTACGACGGCGACGGCGATCCTCGGCGTCCC**CAT**GC *SphI*
 5201 CCTCACCCCACTCCACCCGGCTGCTCCGCTGACGGGCGATGCTAACCGTTCACGGTGAGGACGCA *SphI*
 5266 TGCATGACACCCACCACCGGGCTACTCCGCCCGCCCCACGAACAGGACACACGTGTCC *SmaI*
 5331 TGGTGTCCCGGGACGACCGGAGGGGCCCGTGGCGCAGCCGCGGCCGGCTGCCGCACGGGG *******
 5396 GCCGGGAGCGGTCCCGGT**TCAG**CGGGTGGTGACGAAGGGGCTCTGGTGCCTGTGCGGGGTGGTGA

Figure 9. cont.

5461 GTTCGTCGAGCATGGCGATCGCGAGGTCGCGGCGCGTCCATGCCCGTCCGCCGGGAAGGTCG
SmaI
 5526 GCGGTGACCCGAGCTCGTCGCCCGGCTGCCTCGTCTGAGCCGGGCGGGACGCATGACCGT
 5591 CCACTCCAGGTCGCGGGCTCCGGTGAGGATGTCTCCATGCGGGCATGTCCGCGTACAGGGTCC
SmaI *SmaI*
 5656 GCCCGGGCCGTTGCGCAGGATGCCGTAGACCGCCGCTGCCATCGCACCCCGCCCGGGTGACC
 5721 GGATGTGTCAGGCCGGCGCTCACGACGACCAGGCGCCTGACGTCCGCCGCGCGCATCCCGTCCAC
SmaI
 5786 GACGGCCCGGCCGGACGCCGAGTAGACCGTGACCGCCTCCAGGAGTACGGCGCTCCCAGGCAGG
 5851 ACAGGACGGCGTCGGCGCCCTTGAACACGGACGTCATGTCCGCGACGTCCGTGACGTCCGCCGTT
 5916 TCCACGGTCAGCCTCTCCCCGGAGTGACCGAACCCGGACGCCGGACGACGGCGACGACGTCATG
 5981 GCCGGCCGCACAGGCCAGGGCCGTCACCTGCCGTCCGGTCGGGCCGCTTGCACCGAGCACTGCTA
 <- *gill* *SphI*
 6046 CTTT**CAT**GCCGTCTCCAGACGATGCGCGGACGATGTACGGACGCATGCGGACGATGTGCGTCTG
 6111 CGGTGGTCGAGAAGGTGCCCGGTGGCGCCCTTCCCGAACGACGCCCATCGTAGGAGTCGCGG
SmaI *SmaI*
 6176 CGCCCCGCCCGGCTCCGAGGCCGTGGGCACCGTCGAGAACAGGCCCGCCCGGGCCCGTTGC
SmaI
 6241 GCGGGGCCACGGAGCGGGAGCGCGGGCGGCACGCCGCCGGCTCCCGCCCGGTTCCCGGGCGG
 6306 CACGCCGCCACCCCTCAGGAGGAGCCCTGCCGCGCCGTCCGCGCGGACGTTGCGGGGGCCGTCT
 6371 GGCCAGTTCGAGGGCGCCCATCGGACAGGACCGGATCAACCGCTCCACCTGCCGGACATCGCCGA
 6436 ACCCGTCGGCCGCCCGCTGAGACGGTGATCTGCTGCTCGGGGCTGCGGAAGACGGCCCTGGAC
 6501 AACCCCTCGCACTGTTCTGTGCAACTCGCACCGCCGGCCGTCGATCCTGACCACCGTCACGTCCT

 6566 GACGGGCT**CA**GGCAGTTCGTCCATGGAACGTCACATCCATCCTCTCGATCCCGCGGATGGTGTG
SalI
 6631 TGCAGCCTCCAGGTCGGCTCCCCACCTCGATCCTGTCGACGTGACGGACCAGCGCCTCCAGGAG
 6696 GCTCTGCCCTCCAGGCGGGACAGGGCCTGACCGACGCAGGCGTGGATACCGTGCCCGAAACCGA
 6761 CGTGCTGGGCCGCGCCCTGCCGGGCGACGTCGAAGGACTCCGGGTCTTCCAGAACCGCTCGTCA
 6826 CGGTTGGCCGAACCGAAGAGGAGGAGGACGCGCATCCCGGGGCGAGCGCGGCTCCGCCAGCTC
 6891 CGTGTCTCGGCGACGTAGCGGGTGAACCCGCGCAGCGGCGATTCAGCCGGATGATCTCGTTGA
 6956 AGGCCGACGAGACCAGGGAGGGGTCTCCCGCAGACGGCGCCACTGGTCTGGTGCCTGCCGAGC
SphI
 7021 AGCCACAG**CATG**CTGGAGAGCGCGCTGACGGACGTGTCATGGACGGGGCGAGGAAGTACCCGAG
 7086 CAGTCCGGGCGAGCAGCTTGCTCTCGATCTTGCCCTCGCGGGCCTCCGAGACGAGTTCGGCACCCC
 7151 AGCTCCCCGACGCAGATTGCCGGGCTGTGACATCCGGTGGAGGAACTCGCCCATCTCGCCGAGC
 7216 AGCGGCAGGCCGGCCCGCTCCGGTTCAGGGACCGAAGGCGTTGAATCCGGCGCTGGCCCA

Figure 9. cont.

7281 CTCCAGGAGCCTTTCCTTCCCCCTCGCCCTCCGGCCATCCCAGCAGATCGGGCACCACGGCCAGCG
7346 GGAAGGCGACC CGGAAGTCTTGACGGCGT CGAACGACTTCCGGGCGACGAGGTCCCGGACAAGA
7411 CGGTCCGCCACGACGTGACGTACCCGTTGATGTCGGCCATCGCGCGGGGCTTCAGGTGGCGGGC
7476 CACGAGCCCCCGCACGTAGGCGTGGTACGGCGGGTTCGCTGGTGAAGCTACTGCCCTTCTGCGCCT
7541 TGTT CAGGGTGTCCGGT CAGACC GACGCCCTCGCCGGACACGAACGTGCCGTGGCGGTGCAGGGCC
7606 GCGTACACCTCGTCGTAGCGGGCGGGCGCAGTGCACCTGGTGCGCCGT CAGGTACACCACCGGTGC
7671 CGCGTCGCGCAGCGCACCGTACAGGGGGTACGGGTTCGGTTATCGACGCGTCCGTGTACGGATCGA
7736 GATCGAGGTGGGGATCGTCGATGTGGAGATCACCGCTTCATCCTTTCGCGCGAGGAGGTCTTC
7801 GATCAGCGGCACCATGCCACGGCCGTCCGGCGGGTTCGCGCCCTGCTCCGCCAGTCTCCGGGCC
7866 TGGAGGAGTAGGACGGATCCCGAGCATCTCCTTGACCACCTGCGTGATCGCCTCCGGGGTGCC
7931 TCCTCGCGGTAGAGCGTCCGCGCGCACCCGTAGTCCGTGAGGTGCTTCAGCCTCGGGACGAAGGC
7996 CTCGAAGGGGTTGAGGATCAGCTGCGGAGTCCCGTGTGATGGCGTTGATGGCCGT CAGTCCGC
8061 CCGCCGGATGGATGATCACCTCACAGGTCCGGAGGATGGCCTCCAGCGGGATCCAGCCGCGCGC
8126 ACCCGGGGTACTTCTCCTG CAGCCGCTGCCCTCCGCTCGCCGATGGCCACGACGACCTCGAC
8191 CTCGAGCTCCAGCAGCCCTTCGACGATGGCCGAGATGCGGTCCATCGCGCCGGGAAGGCGTACC
8256 GGAAGCTTCCCATCGTCAGGCACACGCGCCCGCGTCCGGAGCCGT CAGCATCCACGGTCTGATC
8321 GCCCGCTGCATGTTGTGCGGGTCCAGCGCATGAAGGTGCCGGTCCGCCCCGACCAGGCCGGGGC
8386 GCAGATGTCGATCTTCAGTGAGGGTCCGGCAGTCCGTCCGAGCCGATCCTGGCCAGCTCGTCCG
8451 CCATCTCCTCGAGGAGGTACTCCTCGTAGCCCGACGTCGAACAGGTCCAGGACTGCCGGACG
8516 AAGGGGATGCCGAGGAACCGGGCGGCATCTCGGCACCGTGTCCCTGGCTCCCCGCGATCAGGAC
8581 GTCGGCGCCCCACGTCCGGGCGACGTCCACCAGGTCCGTCGAAGACGTGGCTTCCCTGACGCCCGA
8646 ACCAGTGGCCAGGTAGGGCATCTCCTGTTCCGGCCGGTGGGGTACTCGATCGCCGGCTTGCCG
8711 CCGCGCGCCCTTGATGCTCTCGGTGCTGTGCCCGGGCCACCGGGAGGGACGGCAGACCGAT
8776 GCCGGTGACGGCGCCGACATCTCCTCGAAGGACGCCACCAGGACGTGCTGCCCGGACAACCGGA
8841 GTGCCGAGGCGAGGGTCCGATGGCGAACGCGTGGCCGGGCTCGTGCCCGGGCTAGAAAGAGG
8906 GCCTTACGCGGCGCCCTCCCGTGACCGGTCCCTTGTCGCGTTCCGCGAAGTAGGCCTCCTTCAG
8971 GCCCACGGTCTCGAAGTCGATGACGGCGAGGCCGAGGT CAGCAGACGACTCGAACGCAGCTCCC
9036 GGACCAGGTCCAGGTAGTTGTCCATCTCGTTCGAAGCCAGGATGAAGAACTCTCCGCCTCACGG

Figure 9. cont.

10921 ACCACGCCGAAGTTGCCGCCGCCACCGCCGGTGTGTGCCCAGAAGAGCTCACCGAGATCGCCGGT
SacI

10986 GTCGTCGGCCCTCGCCGTACAGAGCGAACGGTGCGGGACTCGTCGACGACGGCGACCTCCACCG
SalI

11051 CGTGCAGGTGGTCGACCACCAGCCCCAGCTGGCGCGACAGCGGCCCGTAACCACCTCCGGCGACC
SalI

11116 AGGCCGCCCATGCCGACCGCGGAGCAGGCCCCGAGCGGCAGGGCCGCGTTCCACCGGCGGAACAG
SalI

11181 GGCCTTCTGGACCTGGTCGACCGTGACCCGGAACCGACGCGCACCCCGGCGCCGTCCGCGGCCG
SalI

11246 GGCCGATGGCATGGAGGTTGTGCAGGTCCAGGACGAGGTCCCGGCGCGCGTGCACGACGAAGTCC
PstI

11311 TGGCCGAGTGACCGCCGGACCGGCAGGCGACCCCCGCCCTTCCGTGACGGCCTTTCTGCAGGA
PstI

11376 GGCGACGACGTCGTCCGGCGTGGCGGGGAGGAAGAACTCCTCGGGCTCGACGACGAACCGGTGGT

11441 TGTCCGAGTGCACAGTTCGATGTACCGGGGTCTCGCGGCCACCCTGAACGGCGGTACGGAA
 <- *gilR*

11506 GCGGTTCACGACGCGTACCCCTCGACCGACTGGGTGAACACGGTCTCGTAGGTGTTGGACTCCCGC

11571 GAGGTCAGCAGCGCCTTGCCGCGTGCAGCATCTCCCGGTCGTGCGCCGGGCGTTCCCTCCTCGGG

11636 CATGGCGTGGAACGCCTCGTAGGAGCCGCGTCCACTGGGCGTAGTTGATGACCATGTCCG
KpnI *SmaI*

11701 CGTCGAGCGCCCTGAGGATGCTGTGGGACCGGTTACCCGGGCACCTGCCGCATCCAGTCGTCCGGC
SacI

11766 TTCTCCAGCAGGGACACGAGCTCGCCCTGGTCTTGGGGTGCACCCCATCAGGACGATGACGGT
SacI

11831 CAGGTCCCCCGGTCCGGGCCGATCTCCGTGCGGGCGGCACCGTCCTTCGTCTGGACGGACACCA

11896 CCTCGGTCTTCAGCAGGTGCACCGACGTGGTGAGTTCGGTGAAGTAGGGGACCGTGTGTGCTTG

11961 AAGTTCTCCCCCTCGTACCGCTCCCTCAGGTGCTTGATCGACCGCCACTGTATGTAGTTGGCCGC
SalI *SmaI*

12026 ACAGGGCCTCGCCACCCCGCGTGCAGGGTCGACGACCGCCATCCCGGGTAGTTCGCGTTCGCGA
SacI *SmaI*

12091 TGATGCCGCGCATGGCGTCCAGGAGGTCGCTGCTTCTCCGAGTCGGTGGTGTGAACAGGTTG
SacI *SmaI*

12156 AACACGGTCAGGGAGCCGTTCTCGGGTTCGATGATCGGCATGAGGCCCTCCATGAGGTGTCCGAC
SacI *SmaI*

12221 GGCCCCGAGGGCCGGCTGTGGCTGATGGGTTCTGACCTCGGTGACGACGAGGACGCGGGCGGCC

12286 GGCTCTCCGGGCCGCCCGGACCGCTGTCCGGATGCCCGCCGCACCCCGGCCCGGCGCCGGCG

12351 **TCACCG**GCTGCGGGAGAGCCGGTGTACCGGTCTCCCCGAGACCTTCCTTCCTTCGACGACGG

12416 ACTGGAGTCCGGCGCGGCTGACCCCGACCGACTCGATCCCGACCTCGGCCATCAGGAGCCGGAAC

12481 TCCTCCACGGTGCCTGTTTGCCTTCGCACAGGAGGAGCATGTCCATGTCCATGAGGGAGATCGC

12546 CTTGTCCGCCGTGTCGTCCAGGCCCGCCACAGCCTCCACGACGGCGATGTGCGCCCCCTCGTGCA

12611 TGGCCTCCGCGATGTTCTGAGGATGAGACGGGAACGGCCGTCTCCAGTCGTGCAGCACGTTG

Figure 9. cont.

12676 GAGATCATGAACAGGTCGCTGCCCGACGGAACGCCGTGAAGAAGGAGCCGGACTTCACCGTGAC
12741 GCGGTGGGCCACCGCCGCTCCGACAGCTCCGGCAGGGACCGCGACACGACGTCCGGCTGGTCGA
12806 AGAGGACCCGGTCATCTCGGGGTGCTTGCGGAGCACGGCGGCCAGCAGCGCCCCCTCCCCCG
12871 CCCACGTCGGTGACACTGCTGAACCGGGAGAAGTGAACGCCCTCGATCACGGGCTGGATGACGCG
12936 CTTGGACAGTTCGGTCATGGCCGAGTTGAAGACGGCCGCGGTGTCGGGATCCGACTCCATGAACT
13001 CCCACACGCCC GAACCGTGCATCGCGGCCAAGGGAGAGCGGCCGGTGCACCCGCGGTGGCCAGG
13066 TGGGCCCAGGTCGCGCTTCGGCCGTCGACCCCGTCCACCGCGGAAGTTGCGCATGGAGCCGTC
13131 GTCGGAGGCCAGGGCCCCGCCCAGCTCCGTCAGCTCCAGCGTGTCTGTGCTCGCCCCCTCCGCAGCA
13196 GCCCCACGAGGCGCCGGCGCGGAAGAAGCGCTCCGCGGTGTCGTGCTCCAACCCAGCCGGACC
13261 GCCAGTTGTGCGGGCGTCAGCCCGCCGGCGCCAGGCCGTCGGCCACGCCAGTTCGCGAACGTC
13326 GCTGACGACACCGGCGCGCCAGCCTCCCATCAGGAGGTGAGGATCTGCACAGCGACGGGTGCCG
13391 ACGGGGGCAGCGATCCCGATGCAGTAATGGTCATGATGTCCCTTCGACTGGTGACATGCGGGGA
13456 GGTGAGGACCCGCTGCAAGGGCAGTTCGCTCTCGAACACACTTTGTTCTGTGATGCCTCACCGGCG
13521 CTAGGTATCCTCATGGGATGGTCCCGGGAAGCAGCGCCTGAACTGCGCAGAGTGTGCCGGACGG
13586 GAGCGGATCGCCCGCCGCACTCTGCGGCTGGTGAACGGACGACCGTGCAGGACCGGGCCGGCA
13651 CGGTCCGGTGAAGGGCGGCCCTGCGGTGATCCGGGCGGCCCGCCCCGTGCTCCCGGCGTCCCA
13716 CGCTCCGTAGGCGGGGACCGCGACACCTCATGATCCCGACGACGGCCCTGTGGGGTCCGGCCG
13781 GCGACGGGGCGCTCCTCGAGGAGCGCGGTGCTACTTCCGCTCCACGTCCGGTTCCTCGGCCCCGC
13846 GGGGAGCGTTCTCCCTCCCACTGCTGGGCGTCCCCCTTGGGCCGCGGGATGAGGAAGGTCGCC
13911 AGCATCCCAGCGCCAGCAGCGCGGCGCAGGTGTAGCCGTTGATCTGGATCGCGTACGACATGGC
13976 GTCGCGGGCGCGTCCGGCGCGGCCCGCTGTCGGGTTCTCACTCAGTCCGGAGATCATCGCGC
14041 CGGCGCTGTCGGAGACCGCCGACGACAGCTCGTTCGCCTCCGGCGCGGGCGTCCCCTGGTTCGACG
14106 AGCCGGTCGGACAGGTCGGAGTCCAGTCCCGTGAAGAAGGTCGACGTCAGAATCGCGATGCCGAG
14171 CGCGGAACCGAGCTGGCGTGCAGCGCTCTGGATGCCGGATCCCTGGCCGGCGTCCCGCGGGCA
14236 CGTCCGCCAGGACGACGTTGGTGACCTGCGCCGTAGCGAACCCGACTCCCATGCCGTACAGGAAG
14301 AGGGCGATCCCGATGACCCACCACTGCGAGTCGCTGCTGGCCAGCAGACCGAACATCAGCAGACC
14366 GACGGCCTCGAGGACGAGGCCGATGCGCACCAGCCGATGGGGCCGACGTTCTCGGCCATGCCGA
14431 AGCTCGCACCGCTCGCGAAGAAGCTGCCGACCGCGACGACGAAGACGATCAGTCCGGTCTGGAGC

Figure 9. cont.

14496 ACCGAGTACCCAGCGTGTACTGGAGCCACAGGGGAGAACGGCGAGCATGCCGAACTCCGCCAG *SphI*

14561 CGCGATGATCAGCGTCGCGATGTTGCCCGTGCTGAAGGACTTGATGCCGAACAGCGAGGTGTCCA

14626 TCAGCGGCGCCGTGCCGTCCACCCGGGTGAGCCGGACCTGGCGCTGCAGGAACAGCCCCAGGCAC *SmaI* *PstI*

14691 ACCACGGAGACGACCAGGGCGACCGGGATGACCGACAGCTCCACCGGGTCGCCGAAGGGACCGAA

14756 GCTCTTGCGCGGGTCCCACCAGCCGTAGTTGCGGCCCTCGATCAGCGGAAGGCCGAGAAGTCCGA

14821 GGCCCAGCACCGACAGCACGCCGCCGACGGCGTGCACCTTGCCCGGCTGCGCGGGGACTGGTCC *SalI*

14886 AGGAACTTCATGACGCCCGGATGATCAGCACCAGCAGAAGATGTTGATGCCGAACGCCACCG

14951 CCAGGAGAACTCGGCGAGCCAGCCGCCAGGAGCGGGCCGACCGCGCCGCCGCCGACCGATCGTGG *NotI*

15016 ACCCCCAGATGGCGAAGGCCTTGCCCCGGTCGGCTCCCCGGAAGGACATGTTGAGCAGGGCGAGC

15081 GAGGTCGGCATGATGATCGCGCCCGGACGCCCTGGGCGAACC GGCTGGCGATCAGGAGTTCCCC

15146 GGAGGGCGCCAGGGCGGGCGGATGCTCGCCAGCCCCAACACCACGGTGCCGATGAGGAACAGCC

15211 GGCGTGCTCCCACCACGTCCGACAGACGCCCCATCACCAGCAGCAGAGCGGCCAGGATGATCGCG

15276 TAGGACTCCTGGATCCACTGGGCGCCGAAGGCCGAGATCTCGAGGTCGTCGATGATCGGGCGCAT *BamHI* *XhoI*

15341 CACCACGTTGACGATGGTGAAGTCCACGACCACCAGCGACACACCAGAGAGAAATGGCGAGCAAGC

15406 CCAGCCAGGGGTCCCGGTTGGCCGGTGAGGACCGAGTTGGATTGTGACACTGCGTTCATCCGT <- *gilJ*

15471 CCTATGTGACACACATGGCCCAGTTGGGTCGCCGGGGCGAGACAAGGGGTAGGGCGGGGGAGC

15536 CTCCCGCCCCGGCGCAATGGCACTATGACAGGAGAAGAGGACGGATTCTGACCTCTACTGACAC *gIII ->*

15601 CGATCCGGAGGGCAATTCTTGATCGCCAACCGCACGTTGGAACCTCCTCAGCCTGTTACAGACCCA

15666 GAGGGAGTGGACCGGTGACGGGCTGGCCGAGCGGCTGGGTGTGTCCCGCGGACCGTCCGGCGCG

15731 ACATCAACCGACTCCGCGAGCTGGGCTACCCCGTCACGGCGACGAAAGGCCCTCCGGCTCCTAC

15796 CGGCTGTCCCGGAGCGCGTCTTCTCCCTGATAGTCGACGACGAGCAGGCCCTCGCCATCGC *SalI*

15861 CCTGTCCCTGCAGACCGCGCCGGCTCGGTGACCGCATGGGAGACGCCACCAACGTGCGCTGA *PstI*

15926 ACTCCATCCAGGAACTGTTGCCACCTCATCTGGCCCACCGGCTCGCCACCTTCTCCGTGGAACAG *SalI*

15991 ATCGAGAACGCGTGGAACCTCGCTCCGCCGAGGTCGACCCCTCCCTGCTCGCACAGCTCAGCAG

16056 CGCCGCCAGCAGCGTGACCTCGTGAGGTTCTCCTACCGCTCCATCCACCACGACTCGATGCAGG *KpnI*

16121 ACGGGGAGGTCCTGGCCGAACCCACCGGCTCGTCGTCTGGTCGGGACGCTGGTACCTGGTGGCC *SacI* *SalI*

16186 TACGACCAGCAGCGGAGCTCATGGCACGCCTACCGGGTCGACCGCATCAAGGATCTGGCGCCAC

16251 CGCCTGGCGCTTCGGCGAGCGGGAGGGTCCCAGCAGGACATCACCCGCTTCGTACAGAACCAGC

Figure 9. cont.

16316 CCGATCGCGGTGACACCCCGGACACCTGGCCCTGCTGGGGCACCGTCCTGATGGAGTGTCCCGCG
SmaI *SalI*
 16381 TCGCTGGTGGCGAAGTGGGCCCGGGAGTGGCGAGTTTCGAGGCGGTTCGACGACAGGGTACCCG
BamHI
 16446 GATCCAGATGGGCGCGTGGTTCGTGGTCGGCGCTCATCGGCTTCCTGATCACCTTCAGTTGCCGCT
 16511 TCACCGTCGAGGGTCCGCCGAACCTCGTCGCCGCGGCGGGAGGGTGATGGGCCTCATCGACGTC
 16576 GGGATTCCGTCGCACGACCCCTCGCGGAGCCGTCGAGCCGCACACCCGGCCCTCGGCCGGCCG
 *** ***
 16641 GTGACGCCGTGGCCGCGGCACCCCGCACCCGCGGGGAACCGCCGGTTCAGCGGACCGCCTGTCCC
KpnI
 16706 AGCTCGGCGCGCTGCCGCAGCGTTCCACCAGTCCCGGTTCTCCCGTACCAGGAGACGGTCTC
 16771 CGCGAGGCCGGTGGCGAAATCCTTGGCGGGCTCGTAGCCAGCTCCAGGGTGATCTTGGCGCAGT
 16836 CCACCGAGTAGCGCTGTCTGTCCCTTGCCTGCGGTCGGCGACGTACTCCACCGACTCCCAGCCGGCC
SacI
 16901 CCGCACGCCTCCAGGAGCAGTGACACCAGTTCCCTGTTGGTGAGCTCGGTGCCGCCCGATGTT
 16966 GTAGACCTCGCCGGTCTGCCCTTCGTGCGGACCAGTTCGATGCCCTGGACGTGGTTCGTGATGT
XhoI
 17031 GCAGCCAGTCGCGGACGTTGAGCCCGTCCCGTACAGCGGGACGGTGCAGCCCGTCGAGGAGGCGC
 17096 GTGATGAACAGCGGGATCAGCTTCTCGGGGAAGTGGTGATGGCCGTAGTTGTTGGAGCAGCGGGT
 17161 CACCCGGACGTCGAGGCCGTGGGTGCGGTGGTGGGCGAGGGCGACGAGGTCGGCGGCCGCTTCG
 17226 AGGCCGCGTAGGGCGAACTGGGTGACAGCGGGTGCCTCTCCGGCCACGAACCGTAGGGGATCGAG
NotI
 17291 CCGTACACCTCGTCCGTGGAGACCTGGACGAAGACGCCCGCGCCCGTCCGTGGTGGCGCAGCGC
 17356 CGCGTCGAGCAGGACCTGCGTTCCGCCACGTTTCGTGCGGACGAACCTCGGCGCCCCCAGGATCG
 17421 ACCGGTCCACGTGCGACTCCGCGGCGAAGTGCACGATCTGGTCGTGCTCGGCGACCAGCCGGTCC
SmaI
 17486 ACGACCGCGGCGTCGACGACATCGCCCTGGACGAGGCGGAACCCGGGTGGGCACGGACCGGGTC
 17551 GAGGTTGGCCGGTTGCCGGCATAGGTCAGTTTGTGAGCACGGTGATGCGGACCCCGCGGGCC
 17616 CGTGCGGGCCGAGCAGGGTGCAGGACGTAGTGGGAGCCGATGAAGCCGGCACCCCGGTGACGAGG
 <- *gile*
BamHI ***
 17681 ATCCTCGTGGCGGTTCATGACGAGATACGCACCTCGCTGTGGTCCCCGAGGACCAGCCGGTGGGCC
 17746 GCCGGCACACCGGCGGCGGGGGTACCTCGACGTTCCGCCCGATCAGGGAACTCTCCACGCGGGC
 17811 GACCCCTGGACCGTGGCCCCGGCGAGGACGATGGAGAACTCGATCTCGCTGGACTCGATCCGGC
 17876 AGTCCGCCCTATCGACGTGGACGGCCGACGTAGGAGCCGGTGATCCGCGTGTGGCGCCGATC
 17941 ACGGCCGGCCCCACGATCCGCGATCCGCGGATCTCGGCGCCCGCTCGATCGTGACCTGGCCGAT
SalI *SalI*
 18006 GACCTCGCTGTCCCGTTCGACATAGCCGTCGACCCCGCCCTTGGCGCCCTCCAGCACGGCCCGGT
 18071 TGACCTCCAGCATGTCGATGACGTTCCCGGTGTCCTTCCAGTAACCGGAAATGGTGGTGGAGCGG

Figure 9. cont.

18136 ACGTCGTACCCGTGGTCGATGAGCCACTGCAGGGCGTCGGTGATCTCCAGCTCGCCGCGTGCGGA
*Pst*I

18201 CGGCTCGATCCCCGGACGGCCTCGTGACCACGGAGGTGAACAGGAAGACGCCACCAGGGCCA

18266 GGTCTGCTGCGCGGAGCGGCCGGCTTCTCTCCAGCGCCACCGCCTTCCCGTCGTCGTCGAGTTCG

18331 ACGACACCGAAGGCGCTGGGGTCGGCCACCTTGGTCAGGAGGATGTGCGTGTGCGGGCCGGCTCTC

18396 GCGGAACCCGGCCACGAGATCCGCGATACGCCACGATGAAGTTGTCTCCGAGGTACATGACGA

18461 AGTCGTCGTCGCCGAGGAACTCGGGGGGATCAGGACGGCATGGGGGAGACCAGCGGGCGCCGCC
*Sma*I

18526 TGCCGTATGTAGGTGACGTCCAGACCGAAGCGGAGCCGTCGCCGACCGCCTGCTGGATCTCGGC

18591 GGCCGTGTCCCCGACGACGATGCCGACCTCGGTGATGCCTGCCTCCGCGATCGCCTCCAACCCGT

18656 AGAAGAGCACGGCTTGTGGCTACGGGACGAGCTGCTTGGCGTAGGAATGGGTGATGGCCCTC

18721 AGGCGGGTTCGGCCCCGCGGACAGTACGAGAGCCTT**CAT**GGCGGCGCAGTCTAGGCGGGCGGG
<- *gilD*

18786 GAAACATCTCAATCGGCCCGGCAGCGCACGGATGTCTGGAAACAACGGTCGGTAGAGGTCAGGAA
*Sac*I

18851 CTGACCTCTACCGCTCATAATCTGGCCGCTCCCTCTCCCCGGAGATCAGCTTCGAGAGCTCGGT
gilH ->

18916 CCCTACCGAAGGAGCGAAACAGATGATCAGGATCGCCGTCATCCTCGGAAGCACGCGTCCCGGCC

18981 GCCGCGGGGCCGTGGTGGCCCAATGGGTGCGCGAGGTGCGCCGCGCGGCATCCCGCGGCGGTGATG

19046 GGCGAGGCGGAGTTCGAGCTGGTGCACCTGGCGGAGTACGGCCTCCCGTTGCTCGACGAGCCCGT
*Sal*I

19111 GCCGGCGATGTTGCGCCAGTACCAGAAGGAGGAGACCCGGCGGTGGGCCGCCCATCGGCTCGT

19176 TCGACGGATTCGTCTTCGTACGCGGAGTACAACCACTCGGTGCCCGCCGCGCTGAAGAAGCC

19241 ATCGACCACCTCTTCGCCGAGTGGACCGACAAGGCGGCCGGTTCGTACGCTACGGCGTGCACGG

19306 GGGAAACCCGTGCCGTCGAGCACCTGCGGCTGGCCCTGGCCGAGGTGAAGGTGGCCGGGGTGCACA

19371 GCCAGGTCTCTGTCTCGGTGTTCAACGACTTCGACTACACGGGATGCGACATGACGGACCCGACG

19436 GCCATGGGCCGGTTCACGCCGGGACCGCAGCAGGAGCAGACGGTGAACACGATGCTGGACGAGGT

19501 CGTCGCCTGGTGCACCGGCTCAAGCCGCTGCGTACTGCTGCGACCGCTGAGGCGGACGGCCGGG

19566 CCGTGTGCGGT**TGAC**GCACCGGTCCGCCCGCGGACCCCTGGTGAACGTGCTGGTACGGCCCC

19631 TCGTGCCTACGCTACGAGGGCCGTGACCAGCACGTTGCTGCTGACGGGCGAGCGTGGCCGCCACG
*Sma*I

19696 CCGCGGTGGGCGGGCAGCACGCCGGCCGACCGATCGCCGAGTGGCTTGTTACGTGCCCGGG

19761 GCGACGGCGCCGGAGGGGGACGCGCCGAAAAAACCGGTCAGTCGAGTTCCTTCGATAACACG
*Eco*RV

19826 GATATCCCCCGTCTCACTTCGGGTGACCTACTTCGGCCGTGCGACTCCGAGCATCGTGAGCGG
gilOI ->

19891 **CATG**ACGTTGCACGCCGAGAAGCCATACCGTCACACGTACCGGTTCTCGTCTGGGAGCCGGCC

Figure 9. cont.

19956 CGACAGGTCTCATGCTCGGCGCCGAGCTGGCGCTCCACGGCAGCCGGCCGCTGGTGATCGACGCG
SmaI
20021 CTGCCGAGCCCAGCGGACAGTCCCGGGCCTGGGCTTCACGGTGAGGACGCTGGAGATCTTCAA
20086 GCAGCGCGGCATCCTGGGCCGTTTCCAGGGACTCGCCCCGGTGCCCGGAGTCCATTTCCGCCGGCC
20151 TCAGCATCAAGGGCGATCACCTCTCCAGCTCGATGCGCCCGGCCAACCAGTACCCGCAGTCCAAG
20216 ACCGAACAGGTCTCGCCGCTGGGCCGAGGAGCTGGGAGTACCGGTGCGGGCGCCCGTGGACGCT
KpnI
20281 GACGTCCATGGAGCCCACTGGCACCGGGTACCGCTGCGTGCTCAGCGGCCCGGCCGGCAGCAGA
SalI
20346 CCGTCGACGCCGACTACGTGGTTCGGCTGCGACGGAGCGGGGAGCTTCGTCCGCGAGGCGATCGGC
SphI
20411 ATGCCGACCAAGCGCACTCCCCATCCGTACAGATGCTCCTCGGTGATCTGCGCGGATGCGGTCT
20476 GCCCGACGAACCCTTCGGGGTCAAGCACGAAAAGGGCATGGTCATGTCCGCACCGCTGGGCGACG
NotI
20541 GGACGGAACGCGTCATCGTCTGTGACTTACCCAGCCGATGCGGCCGAGGGCACTCCCCTCAG
20606 CACGACGAGATCAAGGCCGCTACGAGCAGGTCGTGGCAGCCCCCTGGCGGACGGCGAATGTCT
SacI
20671 CTGGGCGAGCTCGTTCTCGGACGCGTCTCCCTCGTGGAGTCTACCGGTCCGGTTCGTGCGTGC
20736 TCGTGGCGACACGGCGCACACCCATCTCCCCGCCGGCGGGCAGGGCATGAACGTCTCGATACG
20801 GACGCGGTGAACGTCGGTGGAAAGCTCGCGCTGGTGAGCCAGGGCCGCGGCCGGACACCCTGCT
KpnI
20866 GGACACCTACCAGCCGAGCGGTACCCGGTGGCAGGGAAGTGTGCTCAACACCGCCGCCAGG
20931 GCCAGGTCTTCTGCGCGCCCCGGAAGTGGACCCGCTGCGCGAGGTCTGCGGCGACTGCTGAAC
20996 ATCCGGGAGGTGTCCGTCTGCTGGCCGACGGAGTCAGCGGACTGGACATCCGCTACGACATGGG
21061 CCTCCCGGAAGCACCGCCACCCACGGGTGAACGGCTGCCGCCGACGTGTTCCACGTCTGCGGA
21126 CCGGCGGCGACGCCGTCGAGGAGTTGCGGCACGGCGCCGCTCTGCTGATCGTCCCGTCCCCGAC
21191 AGCCCGGCGTCTCGCTGGTTCGCTCCGTGGCGGGACCAGGTGCGCGTCTGTCACGCGCGCCCCAC
21256 GGACCCGACTGGGGCGGGAGCCGGCCGCTCGTTCGACTGGTTCGTACGACCGGACGGACACA
EcoRI
21321 TCGCGTGGGCGGGCACCGAATTCAGCGAGTTGAGCGCCTCACTGAGCCGCTGGCTCGGTACGCC
*** gila ->
21386 GCCGCGTAACCAGAGGAGGAAGAACCCTTGTTCAGCTCTCTCATCGTCGCCCCGGATGGACACCGG
21451 CCACGCCGAAGCGGTGGCCGACGTCTTCGCCGGCTTCGACGCCACCGACATGCCCGCGCGGATGG
21516 GCACGCGGCGCCGCGAACTCTCCGCTACCGCGGCCTCTACTCCACCTCCAGGACTTCGAGACC
21581 CCCGACGGGACCGAAGCGGTGAGGCGGCCAAGTCCGACCCGCGGTTTCATCCGGGTGAGCAACGA
21646 CCTCAGGCCCTACATCGAGGCCTACGCCCCGACTGGCAATCACCGAAGGACGCCATGGCAGAGC
gila ->

21711 GCTTCTACTACTGGAGTTCGAAACGATGAGCCGCAGGGTCTTCATCACCGGGGTCGGTGTCTCG

Figure 9. cont.

21776 CGCCGGGAGCCGTCGGACGTGACCCCTTCTGGGAGCTGCTGACCCAAGGGCGCACGGCCACCCGC
21841 CGGCTCAGCCTCTGCGACCCGGAGCCCTTCCGGTCCCAGGTGGCCGCGGAGGCCGACTTCGACGC
21906 CGAGGCGGCGGGGCTGTCGGAGCGGCAGTCCGCGGAACGGACCGGGCGGCGCAGTTCCGCCCTGG
21971 TCGCCGCCCGTGAAGCGGTGAGGACGCGGCATGGTCCGAGACATGTCTCCCGAACGCGCCGGA
22036 GTGATCGTGGGTTCCGGCCGTCGGAGCCACGACCAAGCTCGAGGAGGTCTACCGGCAGCTCAGCCG
XhoI
22101 TGACGGCTCCCTCTGGGACGTGGCCCCGACTCCCCGCGGAGCTGTACTCGTACTTCGTGCCCA
22166 GCTCGTTCGCCTCCGGCATCGCACACGACCTCGGCGTCACGGGGCAGAGCGGCGTCTGTCGACC
SalI
22231 GGGTGCACCTCCGGGATCGACTCCGTCCGCAACGCTGGGAACTGATCCAGAGCGGCATCCTGGA
22296 CTCCGCCGTCTGCGGTGCCACCGACGCCCCATCTCGCCCATCACCGTCGCCTGCTTCGACACGA
22361 TCAAGGCGACATCGACGTACAACGACACCCCGGAGAGCGCCTCACGGCCGTTGACGCCACACGG
22426 GGGCGCTTCGTCTCGGCGAGGGCAGCGCGATGTTTCGTCTCGAATCGGAGGAATCCGTCCACCG
22491 TCGCGGCGCACGCGTCTACGGCGAGATCCGCGGCTACGCGAGCCGCTGCAACGCCTACCACATGA
22556 CCGGTCTCAAGGCCGACGGACGCGAGCTGGCGGAGGCCGTCGTCTCCGCTCTCGGCCAGGCAGGC
SmaI
22621 GTGGACCCGGGCGGCTCGACTACGTCAACGCCACGGCAGCGGCACGAAGCAGAACGACCGCCA
22686 CGAGACCGCCGCGTGAAGTCGTCCCTCGGACCCGCGCCACGACGTGCCGATCAGTTCGATCA
22751 AGTCGATGATCGCCATTCGTGGGCGCCATCGGGTTCGTTGGAGATCGCCGCTGCGCCCTGGCG
BamHI
22816 CTGCGGGACGACGTGATCCCGCCGACGGCCAATCTCACCCGGCCGGATCCGGAACCTCGATCTGGA
22881 CTACGTGCCGGTCCACGCGCAAGCAGCCGACCAACAGCGTCTCACGACCGGAAGCGGCTTCG
gILB ->

22946 GTGGGTTTACAGCGCCATGGTCTCACGGACCCGGAGCATCACTCATGACCGCACACATCACCG
23011 GCATCGACATCGTCTCCCGCTGGGCTGTCCCGGAGGAACACTGGAAGGCCCTCCTCGACGGA
23076 TGCAGCGGTCTGAGGGCGACGAGTCGTTGACTCCAGCAGGTACGACAACCCCATCAGCGGGGA
23141 GGTGCCCACTTCGCCCCGAGGGCCTGCCAAGCGGCTGTCGCGGCCACCGACCGGATGACCC
SalI *SalI*
23206 AGATGTCGCTGGTCGCCGCGGGGGGCTTCGACGACAGCGGTGTCGACACGAGCCGGTTCGAC
23271 CCCCTCGGAGTCGGTGTGATGACGGCGTCCACCGCGGGGGTTACGCGTTCGGGCAGAAGGAGCT
PstI
23336 GCAGAACCTGTGGTCCAAGGGGCCAGGTACGTCAGCACCCATCAGTCTACGCTGGTTCACG
23401 CGGTCAACACCGGTCAGATCTCCATCCGGCACGGCTGCCAGGGCCACAGCGGAGTGATCGTCGG
23466 GACGACGCCGCGGGCTCGACGCGATCTCCTTCGCCGCCGCGTCTGGCGCGCGCAACCGCGT
23531 CATGCTACCGGGTCGGTGGACAGCAGATGTGTCCCTGGGGCGGGTCGCGCACACCTCGACCG

Figure 9. cont.

23596 *SphI*
GCATGCTCTCGGCATCCACCGACGCGGGCCGCGTACCTTCCGTTTCGACGCACGAGCCAACGGG
 23661 *PstI*
 TGGGTCAACGGCGAAGGCGGCGGCACCTCGTGCTGCAGACCCACAGTGACGGCCGCTACGCGGC
 23726 *SmaI*
 GGTGCTCGGTACGGTGCGACCATGGACGATCCCCGCGCCGCCCCGGGCACGGGCCTCGTCCGGG
 23791 CGATCCACCTCGCGCTCGGCGCGGCGCGCTGCGCCCCGGCGACATCAGTGTGGTGTTCGCCGAC
 23856 *SmaI*
 GCGGCCGGCACCCGGGAGGCGGACACCGCCGAGGCCGCCCTCGCCGAGGTCTTCGGGCCGGA
 23921 TTCCGTCCCCGTACCGCGCCCAAGGCGGCGACCGGCCGGATGGGCTGCGGGACGGCCGCACTCG
 23986 ACGTCGCGACGGCGGTGCTCGCCCTCCGCGACCAGACGATCCCCCCACCGTCAACGTCCAGGCC
 24051 GACGCGTCCCTGGGGTCAACCTGTGCAGCGTCGCCACACACCACCCCTCACCACGTCTCTGGT
 24116 *SmaI* *******
 CCTGGCCCCGGGCGTTCGGTGGGTTCAACTCGGCCCTGATCGTTCGGGAAATGAGAGAAGGAGCAAG
 24181 *gilC ->* *XhoI* *SalI*
GAATGTCCGCACGCGTACCATGGACGATCTCAGGCGAGCCCTCGAGGAGGGCTCCGGTGTCGAC
 24246 GAGGGCGTCGATCTTGACACCGACCTCGAAACCATGGCGTTCTCCGAGCTGGGGTACGACTCCCT
 24311 *SacI*
 GCGGGTGTGGAGACCGGCCTGCGCCTCGGCCGCGAGAACGACATCGAGCTCGACGACTCGGTGT
 24376 TCGCCGACCTCGACACGCCTCAGCAGATGCTGGACGCGGTCAACGATGCCCTCGCGCGTCAGGCG
gilF ->

 24441 GCGGCATCGTGACCTCTCCCCGTATGCCCTGGTCACCGGCGGTTCCAGCGGCATAGGAAAGTCC
 24506 GTCGCACGGCGCCTGGCCTCGGCCGGCCACACCGTCACGATCTGCGGTCTGACTCCGAAAGGCT
 24571 CCAGCAGGCCGCCAAGGAACTGTCGGAGCAGGGTGCACCCGTACCTCGCTGATCGCCGACGTCA
 24636 *BamHI*
 GCAAGCCCCGCCAGGTGGGCGATCTGGTCCGCGAGGCCGTGGAGACGAACGGTCCCCTCGGGATC
 24701 CTCGTCAACAACGCGGGCAGGAACGGAGGCGCCGACCGGGAGCTGAGCGACGAGCTGTGGCG
 24766 GGAGGTACTGAGCACC AACCTCGACAGCGTTTTCTACGTACGCGGGAGGTGCTGGCCCGTGGCG
 24831 GCATCGGCGAGGTGGACCACGCCCGGATCATCAACATCGCCTCCACCGGGGAAGCAGGGAGTT
 24896 CTGCTGGCCGCCCCGTACTCCGCCTCCAAGCACGGTGTCTGCGGCTTCACCAAGGCGGTGGGCAA
 24961 *SmaI*
 GGAGCTGGCCCTCAGGGGATCACCGTGAACGCCGTCTGCCCCGGGCTACGTGGAGACCCCGATGG
 25026 CCTCACGGGTCCGGCAGGCCTACGCAGACGCCTGGGAGACCACGGAGGCCGAGGTGCTGTCCGCC
 25091 TTCAGGCGAAGATCCCGCTCGGCCGGTACAGCACGCCGACGAGGTCCCTCGCTGGTTCGAGTA
 25156 CCTCACGACCGAAGGAGCCGCTCGATCACGGCTCAGGCGTTCAACGTGTGCGGCGGCCTCGGCA
******* *gilK ->*
 25221 ACTTCTAGGAGATGATTACATGCGCCGATCCGGCTCGCACAGACCTGCACTCCGCCACGATCACC
 25286 GGCAGCGCCGACGCGGTGTACCGCCGTCTGGAGGACGTCGGGCAGTGGTCCCAGATGTTCCAACC

Figure 9. cont.

25351 GACCATCCACGGCGCGGAACTGGCCCGGACGGGAACAGGCAGACGATCCAGCTGTGGGCCACCG
SmaI
 25416 CCAACGGAGAACCCAAGGCCCTGGGTCTCCGAGCGTGAGCTCGACCCCGTCGCGCGCACCATCCGC
SacI
 25481 TTCGCGCAGACCGTCACTCCTCGCCCGTCGCCGAGATGTCGGCGCGTGGCAGGTGCTGCCCT
 25546 GTCCGAGGACACCTGCCGGTCAACTCAGCACACCTACCGTGGGAGAACGACTCGGCGGAGT
SacI
 25611 CGCTCACATGGATCGCCCCGAGCCGTGGAGACCAACAGCACGAAGGAGCTCTCGGCGCTCAAGTTC
 25676 GCCTGCGAACGGGACGCCGACAGCGAGGCCAGTCCCTTACCTTACCAGATGCGGTGGACACCAC
SalI
 25741 GGTCGACCCCGTCTCTGCTGTTCTCGTTCCCTGGACCGGTGAGCTGTGGCGGGACGCTGGAGC
PstI
 25806 ACGTCGCCGAGCGGAGATGAGGGAGTTCTCCGACGGCTGCAGTTCTCCGGATGCGGACGCGC
 25871 ACCCCGGACGGTGACACGCACGTCACCGAGTCTACCGGGTGTGCGAGAGCCCGCCCGGCTGCT
 25936 GTACAAGCAGGTGACGCTGCCCGCGCTGCTGTCGCTGCACACCGCGAGTGGACCATACCCCCG
 26001 CCGGGGAGAGCTGGCGGTACGTCGAAGCACACCGTGGCGATCGATCCCGACGCGGTGCACAAG
 26066 GTCTCGGTGCCGACGCGACGGTCTCGGACCCAAGCGGCTCGCCCGCGCAACCTGGGCAACAA
 26131 CAGCCTGCGGACCCTCGAAGCAGCGGTCCGGTGGGCCGGCACCGCCGTGTCGAGAGG*******
gi10IV ->
 26196 GGAC**ATG**ACGGAGCCCGAGACCTCGGACGTTCTCGTCTCGGCGCCGGGCCAGCGGACTGCTCC
BamHI
 26261 TGGCCGGATCCTCGCCGGGGCGGGTGC GCGGGTACGGTCTGGAGGCGGGACCGCCCCAGC
 26326 CCGCAGACCCCGCCTCCACCTGCACGCCGTGCCAGGGAGATCCTCGACCACCACGGAGTGGAA
SmaI
 26391 GTTCTCCCCGAGCTGCCCTGGAGTGCCACGGACACTACGGCGCCCTGCGCGTGGACCTCTCCC
SmaI SalI
 26456 GGTGCACTCCGGGCGGGCCGGTGTCTGGAAGTGCCCCAGCCGGAAGTGGTACGGAGCTGACC
 26521 GGCTGGGCCCCGGGCACGGCGCGCGGCTGCTCCACGGGAGCACGTGGAGTCCGTCCGCGAGCA
SmaI
 26586 GGGCGGGCGCTGCTGGTGCCTACCCGGGCCGGCACCACGTTACGCGGGACCCTGCTGGTCCGG
 26651 CGGACGGCCGGCGGAGCACGGTGC GGTGCTGCTGGGCATCGGGTGC GGGGGTGC GCGCGGCCAG
 26716 CGCGTACTGGTGCAGGCCGATGTCCACGGCGACGGGCTGGCGGGCGGGCCTTCGAGCGACACGG
SmaI SmaI
 26781 GCGGTACACCGTGACCGCCGCACCGATCAGCCCGGATCACCCGGTGATGCTGCACGATCCGC
 26846 GCTGGCCCGGGCGGAGGAACGCACGCTGGAGGACCTCCGTAGAGCCTGGAAGGAGTCCACCGC
 26911 GAGACCCTGCCGGCCGAGCCGTCGTGGTACGGACCTTACGCGACGACACGACAGTGGCACACCC
 26976 GCTGGTCAAGGGCCGTGCTGCTGTGCGGCGACGCCGCCACCCCTTCGTCCCATCGGGGCC
 27041 AGGCGCTGAACACGTCGTTGATGGACGCCGAGGCGCTGGGCTGGCGGGTCTGGGGTATCTGGAC
 27106 GACGGGGACCGCAAGGCCTCCTCGACTACCAGGACGAGCGGTTCTCGTGGCTGACCGTTCTCGC

Figure 9. cont.

27171 GGGGAGACTGCGCGCCAGGCACGTCTGCTGTTTCGACACCGACGCGGGCCACGGAACGCAAGG
27236 CGCTGGTTCGCGGAGACTGGCCGGGGACGCGGACTACCGGCGCAGGATCGCCGACGCCCTGGCC
Sali
27301 GGTGTTCGACGTGTGCTACCTGACGCCCCGGCGGCGGGTCCGCCGGCGTCTGTCCCCGGCCCCGGCT
27366 CCGGGAGACCGGAGTGAACCCCGCGCCCCGCCGCTGCAGCGGGCGCTCGTCCCCGACGACGGAA
BamHI KpnI
27431 CGCGCACGGACGCCTGGATCCGTCCCGATCACCCTGGTACCCTGGCCCGCGACGGGGCCCCGG
gilP ->

27496 CAGGACTGGGACGACGCGGTGCGCCTCCACGACGACTTGGAAACCCGAGGTGACGCGGTGAGAGCG
KpnI
27561 TTCCTGTTCCCCGGTCAGGGACCCAGAAGATCGGCATGGGCACCTACCTGCGAGAACCGTACCC
27626 CCACCTGATCGCGCCGTTGTGGCGGGAGGCGGACGACGTCTGGGTTTCCCCCTCACCCGCCTCT
27691 GCGAGGAAGGCCCGGCGAGAAGCTCCGCCACATGCCGGTACCCAGCCCCGCGTCTTCTGTGC
27756 AGTTACGCCGCGCTCGTCGCCGCGCAGGCGAACGGCGCGGAGCCGGACGTCATCGCGGGCCACAG
27821 TCTGGGCGAGTACTCGGCGCTGGCGGCGGCCGGCGTCTCACCTGGCAGGAGGTCTTCAGCTCG
27886 TCCACCGCCGCGGTGAGCTCATGGCGGAGGTGCAGCACAAGGTGGACGGGAAGATGGCGGCCGTC
27951 ATCGGTCTCGCCATCGGGCAGGTGAGGAGATCTGCGAGCAGGTGCCGTCCGAGACCGGTGAGGT
28016 GGTGAGGTGGCCAACCACAACGAGCCCCCTCCAGGTGTCGTCCTCCGGCCAGTGGCTGCCATAG
28081 ACCTCCTGGTCCAGCGCGTCGCGACGGCGACCGACGTCCGCACGTCCGTCTGAGGATCGGTGGC
28146 CCGGCCACTCCAGTCTCATGGGCAGCGTCGCGGGGGACTTCGTGGAGTACCTCCGGCGCTTCGA
Sali
28211 CTTCTGCACGCCCAAGACGATGCTGATCTCCGGTTCGACCCCGGAGCCCTACGCGAGTGGGGAGG
28276 AGATCAGGCACCAGCTCGGCAGGCAGCTGGTGCACCGGTTGCGGTGGGTGGACGTGATGGCGCAG
XhoI
28341 CTCGAGAGGCTGGGGTTCGCACAGACCTGGGAGCTGGGGCCGGCAAGGTCTCTCGGGATTGCT
28406 ACAGCGGTGCTGCTCAGGTGCGGACGTACCGCGCAATGATCTGCCGTCTTCTGGCCGGCG
*** gilQ ->
28471 TGACGGGTGGTGAGCCGGTGAAGCACGCAGTGCCTCAGGCAACCGGCGCCGCTCCCGACGG
28536 AGGGGGTCCGCGCCCCGGTCCCTCGTGTGATGCTCCCCGGCCAGGGTTCGAGTTGCTGCCA
28601 TGGGAGTCCCGCTTACGAGTCCGACGCCCGGTTCCAGGAAGGCGCTCGACGACTTCTTCGACGCG
28666 TTCGGCACCGGTGCCGAGCGGCTCCGGCGGAGTGGCTGCACGGTTCGGCCCAGGGCATCGAACG
28731 TGGGTCTTCGCGCAGCCGATGCTGTTCCGGCTCGACTACGCGCGGGCGCGGTGTGGCTGGAGG
SacI Sali
28796 AGCTCAAGGGTGTTCGACGTGACGCTGGTTCGGTTCACAGCGTGGGCGAGCTGGCGGCGGCCACCTC
XhoI
28861 GCGGGGCTTCGACCTTCGAGCTGGCGGGGGCACTCCTGGCCGAGCGGGCCCCGGTCTTCGACGC
SmaI
28926 CGCCCCCGGGGAGGGATGATCGCGTGCCGCGGACGGAGGAGTCGCTGCGGGAGCATCTCGACG

Figure 9. cont.

28991 CCCTGGGCGGACGCGCCGTCATCGCGGCGGAGAACGCGGACAACCAGTGCCTCGTGAGCTGTGCC
KpnI
29056 GAGGAAGACCTCCCGGACACGATGCGGTACCTCGGCTCGCACGGTGTGACGTGCCTGCGCGTCGC
29121 CTCGACCGAACCGTTCCACTCCCCCTCCTCGCCCCGCGCCGCGCCCGGTTTCGAGGAGTTCCTGG
29186 CCCGGCGGGTCATCGTCTGTCCACGACGGAAGTCCCATGGTCTCGGCCTACTCGGCGGGAGG
29251 ATCAGCGGCCGGGAGATCATGCCCGCTCGTTCTGGACGCGTCAGATGGCTGAGAAGGTGCGTTT
29316 CTGGGAGGCGCTCCGCCACAACCTTCGACTCCGGTCCCCGCACGTTTCGTGGAATCGCCCCAGGGA
29381 CCGTCTCTCCCTGGCCGCACGTCCGGTCCGTCCGTACGGGCCCGGCGTTCCACGGTGATCTCC
29446 ACGATGCCGCGTCATCGGCCCCACCCGGAGCACTGGGAATCGGCCATACATGAGGTGCGCCGAGGA
EcoRI *** SmaI
29511 ATTCGTGTAACATTGCACTACGTGCAACGCGCAAGGCCGGCCATGGGTGTCCCCGAGTTCCTCG
XhoI
29576 GGAGGCACCCATGGCCTTGTCCGGTAAAATGTTCAACCAAATGAACCACCTCTCGAGGGCGCCCCG
29641 GATCAAAGATGTTACCGATTTGCATAGTCGAAAAATACGGACAGCAACGGAAGCGGAGTGTTA
29706 TCCTGCAATCTGCACGCAACGGGGGAAACGGGGAGGATTCGAAGTGCAGGACCCGGTGGACCGG
29771 ACCTCGGGAACTCCAGAGGCGCAAGCACCGTCCGACCCGAGCAGTCCCGCAAAGACGCGGGGCA
KpnI
29836 CTCTGTGCGCGAAGAGGGGCCCGGTACCGTCACGCAGCAGTTTTTCGCCGACCCTCCACCATTTCT
29901 TCACCGTCACGAGATCTCTTCGGGCCGACGGGGACCACACGGGCAGAGACTGAAGGACCGGCGCT
29966 CGACCTCGCCGGCTCCCGCCCTCCACCCCTTCCCCGCGTTGCCCTGACCGCATGGGCGGCATC
30031 TCGGTGCCGCGTTTTCTTCCGCGCCTGGCGCGAGGGGAGATCTCCCATCAAGGGGGGCGTTTTCAAG
XhoI
30096 GCCCTCTCGCCTCGAGGGCACCGCACGCCGAAGACCGATCACAAAAGTATCCGAACGGCTCCGAC
gilt ->
30161 CGAGGTCATATCTGAGACTGATCGAATATCCAACGGGGAGATGTGATGGGTTTCATCCGGTTTGA
30226 CGTTCGGGCCCGCTCAGGGTCCGGTGCACGACACCCCTGCTTCAATTGACGGGCGCAAGTACC
30291 GCACCGTGGTTTTCGTATCTCGCTCTTCAACCCGAGTATTCGGTGGCGATAGAGGACCTCGTCCGA
30356 GCCGCTTGAGCGACAAGCGCCCGTCCAGCGCGACCACCAGGTCCGTAAGATGGTCTCCGCACT
30421 CCGGACCAGCCTGGACCAGGACTGGGACCTGGTGGCGACGTCCAGGACGGCTACATGCTGAAGT
EcoRI
30486 TGCCGCCCAAGCAGTCCGACGTATCCGAATTCTGCCGCTCTTCGACCAGGTGATGTCGGGTCCC
30551 CTGACGAGCGACGACACCTGTCGGCCGCGTATTCGGCGCTGGCGCTCTGGCGGGACGCCCTTG
30616 CGAAGGGTCCGAGCCCCATGGGCAGGAGCGCCGGATCTCTCAATTGGTGAACAGCACCGCGTCC
30681 TCTTGAACAAGACCGTTTCAGGGATTCGGCGACAGGGGCAGGTCCGATGAACTCGCTCGATACTG
30746 CACGTCGCATCGAAGATTCACGGACAGCCGGTACCAGTCCGCTCGCTCCGGTGTCCCGTTCCCCGCGC

Figure 9. cont.

30811 CGCTGTTTCGTACGCGGGCAGACACAAGTCCCCGAACCTTCGGGGTCGACCACCCTCCCCAC
SalI
30876 GCCCCGGCTCCCCGTCCGGTCCGCGCTGCCTGCCACGGGATCTGCAGGACTTCGGCGGCCGCGAA
PstI *NotI*
30941 CGCGAAATCAATGAGCTGCAGAAACTGTTGACCGCGGAAGGACCCCACCCACAGTTGGTGGCGAC
PstI
31006 CGTTCACGGAATGAGCGGCGTGGGTAAAACCGCCGTCGCCGTCCGCCTGGCGCACAGACTAGCCC
31071 ATCACTATCCGGACGGCCAGCTCTTTGTATCCCTGGACGGCTTTTCTTCGGCCTCCACCGCCACC
31136 GTGTGCAATGCGCTGGGAATACTCCTCAGACAGAAAGGCTGGCGGACGAGGACATTTACCTTC
31201 GGAAGACGGCCGCTCGCACAATGGCGGACCATCACCGCCGACAGAAGCTGCTCGTCGTGCTCG
31266 ACGACGTGTGCGACATCGAGCAAGTAGAACCCCTCATCCCGCCCTCGAGCGAAAGCGCCTGCATC
XhoI
31331 ATCAGTCGCGCATCATCCTCAATGGCATCGACGGCGCTCATCACATCTACTCGAAGTACCGGA
31396 CGAGGACGAATGTCTGGAGATACTCAGTTGCATGATCGGCAGACGCTTCGACGACGAGGAGACGA
31461 AGGACGCCCGCGCTGATCCAGCAGTGCCCAATCTGCCGCTGGCACTCCGTCTCGCCGCCGCC
EcoRV
31526 CGGATATCGACGCGGACTTCTGAACCTCCGGAACTCAGTGAGCAACTGTCGTCTCGGCTTC
31591 CATCTTCAGTGAAGTGGAAAGTTCGCGCCGCTAGTCTGGTCGGCCGGCTCATGACGTCCTCACGT
KpnI
31656 GCCTGGAGGACTTCGATCACGACCGGTACCTCCGATTATCGCTGCTCCCCTGCCCCGAGATCGAT
PstI
31721 GAAACGTCCGGTCGCGGCCGTGCTGGGCGTATCCACCGACTGGGCACGGCGTGCCTGCAGGCGCTT
PstI
31786 CGCAGACCGCGGTTGCTGCAACGCACACGATGCGGTACGTACCGGATGCACCCGCTGCTGCTGC
31851 AGGCGGCACAGCTGGAAGCGCAGAAGACCATCCCGTTCGAGGAGCAACGCCGCTCGTCCGCGCC
SmaI
31916 GCTTTCCTCCATTACAAGGCGTCGAACGGCCTCGTGGGAGCCAGCCGCATCAGCCCTTCCCCGGT
31981 TCCTGACGGACACGTGGTACTGAGGACCCTCACGCAGTCCGCGAAGCTGGCCGCGCGGCTCGGCC
32046 TCCAGGAGGAGTTGGCCGATCTGTACACCGCCTGGAAGGAACTGCTCCCCCTCGTGCTGGACCGC
32111 CGGCAGCAGGAGGCGGTGGGCGACGCTACTCGCCGTTTACAGCACCTGGACCGGCCCGCGTG
32176 CGAGGGAGCACCCACCGGAGGCGTCCGCGGCAGGCACGGGACATGCTGCCGAGGGGCAGCGGT
**
32241 **GA**ACGAGGGCCGCGGAGGAAGGCAGTCGGACGATGACGACCGTTTGTCCGTACATCGGGAGAC
SmaI
32306 CGGGGTGCCACGTGAAACATGTGACCCGGTCACCGGATGTCCGATCGCAGCCGCACCCGGGGCG
giIU ->
32371 AACTGACCA**AT**GGACCGGTTCTTCCGTATGCAGCCGGCAGTGAGGCACTCCTGTGCTCGAGAGAG
32436 CACGGCCCCACGGTGAGCGAGAGAACCCTCTCGGCGCAGGAGATCGTCTGGGCGGCGGGGGTCT
32501 GCTGGGGAGACACATCCTCGGCGTGCTGGGCAATCGGCTCAGCCGGCGGGTACGCATCCCGTGGG
32566 ACGACCACGGCCGCGCCTGTGAGCAGCTCTACGCGCTGGGCAGGGACCTGGCTCAGCAGCCGGCC

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GILVOCARCIN GENE CLUSTER, RECOMBINANT PRODUCTION AND USE THEREOF

This application claims the benefit of U.S. Provisional Application No. 60/477,957, filed on Jun. 13, 2003, which is incorporated herein by reference.

FIELD OF INVENTION

The present invention relates generally to polyketides and polyketide biosynthesis. In particular, the invention pertains to the nucleic acids encoding gilvocarcin polyketide synthase and the tailoring enzymes of the gilvocarcin biosynthesis, and to recombinant vectors and host cells containing such genes, and to the recombinant production of gilvocarcins and uses thereof.

BACKGROUND OF INVENTION

Polyketides represent a large family of diverse compounds synthesized from 2-carbon units through a series of condensations and subsequent modifications. Avermectin, candicidin, epothilone, erythromycin, FK-506, FK-520, narbomycin, oleandomycin, picromycin, rapamycin, spincocyn, tetracycline, and tylosin are examples of such compounds. Polyketides occur in many types of organisms, including fungi and mycelial bacteria, in particular, the actinomycetes. Polyketides can be divided into macrocyclic/polyether-type compounds, biosynthetically encoded by type-1 polyketide synthases (PKSs), and into multicyclic, aromatic compounds, whose core structures are encoded by type-2 PKSs. Type-1 PKSs are "complex" or "modular" PKSs which include assemblies of several large multifunctional proteins carrying, between them, a set of separate active sites for each step of carbon chain assembly and modification. As such, structural diversity occurs in this class from variations in the number and type of active sites in the PKSs. This class of PKSs displays a one-to-one correlation between the number and clustering of active sites in the primary sequence of the PKS and the structure of the polyketide backbone. The second class of PKSs, called Type-2 PKSs, is represented by the synthases for aromatic compounds. Type-2 PKSs have a single set of iteratively used active sites.

Angucycline group antibiotics, which are arranged by a type-2 PKS are structurally characterized by their angular, polyketide-derived benz[a]anthracene-derived backbone (angucyclinone), which is often further decorated with sugar moieties (angucyclines). Angucyclines/angucyclinones form the largest and structurally most diverse sub-group of the multicyclic, aromatic polyketides. Knobler, R. M., Radlwimmer, F. B. and Lane, M. J. *Nucleic Acid Res.* 20:4553-4557 (1992); Matsumoto, A. and Hanawalt, P. C. *Cancer Res.* 60:3921-3926 (2000). Yamashita, N., Shin-Ya, K., Furihata, K., Hayakawa, Y. and Seto, H. *J. Antibiot.* 51: 1105-1108 (1998); Nakashima, T. et al. U.S. Pat. No. 6,030,951. A very interesting set of natural products with respect to their biosyntheses as well as their biological activities derive from this angucycline/angucyclinone group. However, they are not easily recognizable as such, since their polyketide-derived skeleton is rearranged in a series of steps, initiated by oxidative biosynthetic processes. The gilvocarcin-type anticancer antibiotics (Morimoto, M., Okubo, S., Tomita, F. and Marumo, H. *J. Antibiot.* 34:701-707 (1981); Breiding-Mack, S. and Zeeck, A. *J. Antibiot.* 40:953-960 (1987); Yamashita, Y. and Nakano, H. *Nucleic Acids Res. Symp. Ser.* 20:65-67 (1988); Elespuru, R. K. and Gonda, S. K. *Science.* 223:69-71

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(1984)) and the jadomycins (Oyola, R., Arce, R., Alegria, A. E. and Garcia, C. *Photophysical properties of gilvocarcins v and m and their binding constant to calf thymus DNA. Photochem. Photobiol.* 65:802-810 (1997)) are examples of such 'rearranged angucyclines'. Both of them, and the kinamycins (Takahashi, K. and Tomita, F. *J. Antibiot.* 36:1531-1535 (1983)), have in common biosynthetic rearrangement cascades that begin with an oxidative cleavage of the 5,6-bond of an angucyclinone intermediate (FIG. 1).

Given the difficulty in producing polyketide compounds by traditional chemical methodology, and the typically low expression of polyketides in wild-type cells that produce them naturally, there has been considerable interest in finding improved or alternate means to produce polyketide compounds. This interest has resulted in the cloning, analysis, and manipulation by recombinant DNA technology of genes that encode PKS enzymes.

Gilvocarcin-Type Natural Aryl-C-Glycoside Antibiotics

The benzo[d]naphtho[1,2-b]pyran-6-one C-glycoside antibiotics, often referred to as gilvocarcin-type aryl-C-glycosides, were discovered in Japan in the early 1980s. Studies have shown that these molecules are decaketides, and that they originate either from one acetate starter and nine malonate extender units or from one propionate starter and nine malonate extender units, depending on the 8-side chain. The incorporation pattern suggests the key intermediate to be an angucyclinone, such as 2 in FIG. 2., which then rearranges to form the coumarin frame. Krohn, K. and Rohr, J. *Angucyclines: Total Syntheses, New Structures, and Biosynthetic Studies of an Emerging New Class of Antibiotics. Topics Curr. Chem.* 188, 127-195 (1997); Takahashi, K. & Tomita, F. *Gilvocarcins, New Antitumor Antibiotics. 4. Mode of action" J. Antibiot.* 35: 1038-1041 (1982); Carter, G. T., Fantini, A. A., James, J. C., Borders, D. B. & White, R. J. *Biosynthesis of Ravidomycin. Use of 13C-13C Double Quantum NMR to Follow Precursor Incorporation. Tetrahedron Lett.* 25, 255-258 (1984); Carter, G. T., Fantini, A. A., James, J. C., Borders, D. B. & White, R. J. *Biosynthesis of Chrysomycins A and B. Origin of the Chromophore. J. Antibiot.* 38, 242-248 (1985). Gilvocarcins are biosynthesized by a type-II polyketide synthase (PKS) and the necessary post-PKS tailoring enzymes. Among these, the key enzyme responsible for the tremendous structural change from the supposed angucyclinone intermediate (e.g., 2 in FIG. 2.) to the unique tetracyclic lactone structure of the gilvocarcins is proposed to be a C—C-bond cleaving oxygenase. Other key post-PKS tailoring steps with respect to important structural features of gilvocarcin V are the oxygenation/dehydration reactions necessary for the formation of the vinyl side chain, and the C-glycosyltransfer step, through which the 6-deoxy-D-fuco-hexofuranose moiety is attached.

This distinct family of antitumor antibiotics shows excellent antitumor activity and remarkably low toxicity, and therefore has remained to be attractive for synthetic organic chemistry as well as for biological activity studies since their discovery. The group consists of the gilvocarcins (syn. toromycins, anandimycins), ravidomycins, the ravidomycin analogues FE35A and B, the chrysomycins (syn. virenomycin, albaccarcins; including recent derivatives possessing branched ketofuranose and ketopyranose sugar moieties), and BE-12406 A and B (FIG. 3.). Hirayama, N., Takahashi, K.; Shirahata, K., Ohashi, Y., Sasada, Y. *Bull. Chem. Soc. Jap.* 54:1338-1342 (1981); Krohn, K. et al. *J. Topics Curr. Chem.* 188:127-195 (1997); Hosoya, T., Takashiro, E., Matsumoto, T., Suzuki, K. *J. Am. Chem. Soc.* 116:1004-1015 (1994); Knobler, R. M. et al. *Nucleic Acid Res.* 20:4553-4557 (1992);

Matsumoto, A. et al. *Cancer Res.* 60:3921-3926 (2000); Yamashita, N. et al. *Antibiot.* 51:1105-1108 (1998); Nakashima, T. et al. U.S. Pat. No. 6,030,951; Morimoto, M. et al. *Antibiot.* 34:701-707 (1981).

Gilvocarcin V (GV) (FIG. 4.), the principal product of *Streptomyces griseoflavus* Gö 3592 and of various other *Streptomyces* strains, is the most important member of the gilvocarcin-type aryl-C-glycosides, because of its potent bactericidal, virucidal, cytotoxic and antitumor activities. GV is one of the strongest antitumor compounds among these drugs, requiring only low concentrations and maintaining a low in vivo toxicity. The exact molecular mechanisms responsible for the in vivo mode of action of GV are still widely unknown. However, it was found that GV exhibits a strong tendency to intercalate with DNA. Both equilibrium DNA binding and UV light-induced DNA adduct formation was found, causing also topoisomerase II inhibition. Knobler, R. M. et al. *Nucleic Acid Res.* 20:4553-4557 (1992). The vinyl group is essential for the antitumor activity, since the minor congeners gilvocarcins M and E, in which the vinyl group is replaced by a methyl group and an ethyl group, respectively, are significantly less effective. Yamashita, Y. et al. *Nucleic Acids Res. Symp. Ser.* 20: 65-67 (1988); Elespuru, R. K. et al. *Science.* 223:69-71 (1984); Oyola, R. et al. *Photochem. Photobiol.* 65:802-810 (1997). Photobiological studies showed that the vinyl group undergoes a [2+2] cycloaddition with DNA thymine residues under photoirradiation. Moreover, it was shown recently that Givocarcin V promotes protein-DNA cross-linking when photo-activated by near-UV light, and histone H3, which plays an important role in DNA replication and transcription, was identified as one of the selectively cross-linked proteins (FIG. 5.). This cross-linking with histone H3, believed to be part of the unique molecular mechanisms of the potent antitumor activity of gilvocarcin V, might contribute to the better and more specific activity of GV compared to other intercalating antitumor drugs. Matsumoto, A. et al. *Cancer Res.*, 60:3921-3926 (2000).^{3b}

The molecular architecture of gilvocarcin V in conjunction with its biological activity makes GV an excellent target for the study of its biosynthesis and the development of novel, improved anticancer, immunosuppressant, antibiotic, antiviral and neuroprotective drugs through combinatorial biosynthesis.

SUMMARY OF INVENTION

In one aspect, the present invention provides isolated nucleic acid compounds comprising a sequence identical or complementary to all or part of a coding sequence for the gilvocarcin V biosynthetic gene cluster from *Streptomyces griseoflavus* (SEQ ID NO:1). Preferably, a part of said coding sequence is one or more open reading frame (ORF) selected from the group consisting of ORF1, ORF2, ORF3, ORF4, ORF5, ORF6, ORF7, ORF8, ORF9, ORF10, ORF11, ORF12, ORF13, ORF14, ORF15, ORF16, ORF17, ORF18, ORF19, ORF20, ORF21, ORF22, ORF23, ORF24, ORF25 and ORF26.

In one embodiment, the present invention provides an isolated nucleic acid strand that encodes a gilvocarcin gene cluster or subunit thereof comprising a nucleotide sequence identical or complementary to, or an amino acid sequence encoded by a nucleotide sequence identical or complementary to, all or part of a coding sequence for gilvocarcin V biosynthetic gene cluster from *Streptomyces griseoflavus* (SEQ ID NO:1). Preferably, the gene cluster encodes a functional PKS or a functional arrangement of the PKS and selected post-PKS tailoring enzymes. The gene cluster may

be derived from a single species or may be hybrid in nature. In certain embodiments, the gene cluster is a replacement gene cluster. The replacement gene cluster may be a hybrid, mutant, analog or derivative thereof.

In another embodiment, the invention provides an isolated nucleic acid that encodes three or more open reading frames (ORFs) comprising a sequence identical or complementary to all or part of a coding sequence for enzymes performing the biosynthesis of gilvocarcin V from *Streptomyces griseoflavus* (SEQ ID NO:1). Preferably, the ORFs encode a functional PKS or a functional arrangement of the PKS and selected post-PKS tailoring enzymes. In certain embodiments, an ORF may be derived from a single species or may be hybrid in nature. In certain embodiments at least one of the ORFs is derived from the gilvocarcin V gene cluster. In other embodiments, at least one ORF is derived from a non-gilvocarcin V producing *Streptomyces* strain, or is hybrid in nature. In yet other embodiments, at least one ORF is a mutant, analog or derivative of the native coding sequence.

In still another embodiment, the present invention provides isolated nucleic acid compounds comprising three or more genes of the coding sequence for the biosynthesis of gilvocarcin from *Streptomyces griseoflavus*. Preferably, the mixture of genes encode a functional PKS or a functional arrangement of the PKS and selected post-PKS tailoring enzymes. In certain embodiments, a gene may be derived from a single species or may be hybrid in nature. In certain embodiments at least one gene is derived from the gilvocarcin V biosynthetic gene cluster. In other embodiments, at least one gene is derived from a non-gilvocarcin V producing *Streptomyces* strain, or is hybrid in nature. Non-limiting exemplary non-gilvocarcin V biosynthetic genes are preferably subunits of the gilvocarcin M, gilvocarcin E, defucosyl-gilvocarcin V, ravidomycin, deacetyl-ravidomycin, FE35A, FE35B, chrysomycin A, chrysomycin B, BE-12406 A, or BE-12406 B gene cluster. In yet other embodiments, at least one gene may be a mutant, analog or derivative of the native coding sequence. It is also preferred that the encoded activity of the gene is, for example and without limitation, a ketosynthase activity, a chain lengthening activity, an acyltransferase activity, an acyl carrier protein activity, an oxygenase activity, a reductase activity, an oxidoreductase activity, a cyclase activity, a glycosyltransferase activity, a methyltransferase activity, an activity encoded by any gene belonging to the biosynthesis or modification of a sugar moiety, a regulatory activity, a repressor activity, or a transporter activity.

In another aspect, the present invention provides recombinant expression vectors encoding a gilvocarcin gene cluster, hybrids, mutants, analogs or derivatives thereof. In certain embodiments, vectors encode one or more subunit of gilvocarcin gene cluster, hybrids, mutants, analogs or derivatives thereof.

In another aspect, the present invention provides a host cell transformed with a recombinant expression vector described herein.

In still another aspect, the invention provides a method of preparing gilvocarcin V, said method comprising transforming a host cell with a recombinant DNA vector that encodes a gilvocarcin V gene cluster or subunit thereof, and culturing said host cell under conditions such that gilvocarcin is produced and/or gilvocarcin analogs are produced. In one embodiment, the method is practiced with an *E. coli* host cell. In certain other embodiments, the method is practiced with a *Streptomyces* host cell. The gene cluster may be a replacement gene cluster and preferably a functional gene cluster. In certain embodiments, the invention provides methods for preparing new polketide-type compounds, preferably, gilvocar-

cin V-type polyketides. The gilvocarcin V-type polyketide produced may be gilvocarcin V or gilvocarcin hybrids, mutants, analogs or derivatives thereof. Such polyketides are useful as antibiotics, antitumor agents, and immunosuppressants, and for a wide variety of other pharmacological purposes.

These and other embodiments and aspects of the invention will be more fully understood after consideration of the attached Drawings and their brief description below, together with the detailed description, example, and claims that follow.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1. schematically illustrates formation of jadomycins, gilvocarcins and kinamycins.

FIG. 2. schematically illustrates intermediates in biosynthesis of gilvocarcin.

FIG. 3. provides the structure of gilvocarcin-type anticancer drugs.

FIG. 4. provides the structure of gilvocarcin V.

FIG. 5. illustrates the hypothesized mechanism of action of gilvocarcin V.

FIG. 6. A shows the gilvocarcin gene cluster.

FIG. 6. B shows a simplified gilvocarcin gene cluster, in which the polyketide synthase and associated genes are depicted in black, the genes encoding the tailoring enzymes are in pink (i.e. gray, if printed in black/white), and the regulatory, resistance and so far unknown genes are shown in white. Shown are two alternative pathways towards gilvocarcin V.

FIG. 7. illustrates proposed biosynthetic pathway for gilvocarcin V sugar moiety.

FIG. 8. illustrates the hypothesized alternative pathways to gilvocarcin V and possible involvement of products of the gilvocarcin gene cluster.

FIG. 9. is the nucleotide sequence of the Gilvocarcin V gene cluster which sets out ORF1-26 (SEQ ID NO:1).

DETAILED DESCRIPTION

Given the valuable pharmaceutical properties of gilvocarcin-type aryl-C-glycosides, there is a need for methods and reagents for producing large quantities of gilvocarcin-type aryl-C-glycosides, for producing gilvocarcin-type aryl-C-glycosides in host cells that do not produce gilvocarcin-type aryl-C-glycosides naturally, and for producing novel gilvocarcin-type aryl-C-glycosides compounds not found in nature. The present invention provides the protein encoding nucleic acids, methods and reagents that produce gilvocarcins, with particular application to methods and reagents for producing the gilvocarcin-type aryl-C-glycosides known as gilvocarcin V ("GV") and its analogs and derivatives and novel compounds related through structure or genetics to gilvocarcin V.

The practice of the present invention will employ, unless otherwise indicated, conventional methods of chemistry, microbiology, molecular biology and recombinant DNA techniques within the skill of the art. Such techniques are explained fully in the literature. See, e.g., Sambrook, et al. *Molecular Cloning: A Laboratory Manual* (Current Edition); *DNA Cloning: A Practical Approach*, vol. I & II (D. Glover, ed.); *Oligonucleotide Synthesis* (N. Gait, ed., Current Edition); *Nucleic Acid Hybridization* (B. Hames & S. Higgins, eds., Current Edition); *Transcription and Translation* (B. Hames & S. Higgins, eds., Current Edition), and *Practical*

Streptomyces Genetics (T. Kieser, M. J. Bibb, M. J. Buttner, K. F. Chater, D. A. Hopwood, Norwich, UK: The John Innes Foundation; current edition).

All publications, patents and patent applications cited herein, whether supra or infra, are hereby incorporated by reference in their entirety.

As used in this specification and the appended claims, the singular forms "a," "an" and "the" include plural references unless the content clearly dictates otherwise. Thus, reference to "a polyketide" includes mixtures of polyketides, reference to "a polyketide synthase" includes mixtures of polyketide synthases, and the like.

Definitions

As used herein the term "coding sequence" or a sequence which "encodes" a particular protein, is a nucleic acid sequence which is transcribed (in the case of DNA) and translated (in the case of mRNA) into a polypeptide *in vitro* or *in vivo* when placed under the control of appropriate regulatory sequences. The boundaries of the coding sequence are determined by a start codon at the 5' (amino) terminus and a translation stop codon at the 3' (carboxy) terminus. A coding sequence can include, but is not limited to, cDNA from prokaryotic or eukaryotic mRNA, genomic DNA sequences from prokaryotic or eukaryotic DNA, and even synthetic DNA sequences. A transcription termination sequence will usually be located 3' to the coding sequence.

As used herein the term DNA "control sequences" refers collectively to promoter sequences, ribosome binding sites, polyadenylation signals, transcription termination sequences, upstream regulatory domains, enhancers, and the like, which collectively provide for the transcription and translation of a coding sequence in a host cell. Not all of these control sequences need always be present in a recombinant vector so long as the desired gene is capable of being transcribed and translated.

As used herein the term "functional PKS" refers to a set of genes (e.g., three or more) or subunits of a biosynthesis gene cluster, which catalyzes the synthesis of a polyketide, including without limitation a "minimal PKS".

As used herein the term "gene" refers to a segment of DNA or its complement that is involved in producing a polypeptide chain, including regions preceding (leader) and following (trailer) the coding sequence as well as intervening sequences (introns) between individual coding sequence (exons). A "gilvocarcin V gene" refers to any of the ORFs of SEQ ID NO:1.

As used herein the term "gene cluster" refers to a set of (e.g., three or more) closely related genes that code for the same or similar proteins and which are usually grouped together on the same chromosome. A "gilvocarcin V gene cluster" refers to the set of genes encoded by SEQ ID NO:1.

As used herein the term "genetically engineered host cell" is meant a host cell where the native gene cluster or subunits thereof has/have been deleted using recombinant DNA techniques. Thus, the term would not encompass mutational events occurring in nature. A "host cell" is a cell derived from a prokaryotic microorganism or a eukaryotic cell line cultured as a unicellular entity, which can be, or has been, used as a recipient for recombinant vectors bearing the PKS gene clusters of the invention. The term includes the progeny of the original cell which has been transfected. It is understood that the progeny of a single parental cell may not necessarily be completely identical in morphology or in genomic or total DNA complement to the original parent, due to accidental or deliberate mutation. Progeny of the parental cell, which are sufficiently similar to the parent to be characterized by the relevant property, such as the presence of a nucleotide

sequence encoding desired biosynthetic enzymes, are included in the definition, and are covered by the above terms.

As used herein the term “gilvocarcin V analog” refers to a compound or molecule that resembles gilvocarcin V and that contains one or more structural differences relative to gilvocarcin V. Preferably, the gilvocarcin analog has gilvocarcin-type activity although a gilvocarcin analog may have enhanced or the same activity as products of the gilvocarcin V gene cluster. For example, the degree of saturation of at least one bond in the gilvocarcin structure can be changed (e.g., a single bond can be changed to a double or triple bond, or the converse), a bond can be removed, one or more carbon, oxygen or hydrogen atoms can be replaced with a different atom or a chemical moiety (e.g., a halogen, oxygen, nitrogen, sulfur, hydroxy, methoxy, alkyl, aryl, cycloalkyl, heterocycle, amine, amide, ketone, aldehyde, etc.) and the like. In addition, the C-glycosidically-linked sugar moiety can be changed, modified or replaced by other sugar moieties including deoxysugars, amino sugars, keto sugars, halogenated sugars etc., which may be connected as C-, O-, N- or S-glycosides at any possible position of the gilvocarcin molecule. Also other peripheral groups, such as OH groups, methyl groups, O-methyl groups, halogen atoms etc. can be added, modified or removed. Other types of derivatives of gilvocarcin that would be encompassed by the term “gilvocarcin analog” are known in the art.

As used herein the term “gilvocarcin V derivative” refers to a polyketide compound or molecule, that may be produced from gilvocarcin in one or more steps or with few chemical or moiety modifications.

As used herein the term “gilvocarcin V-type polyketide” refers to a compound or molecule that is encoded by at least one native gilvocarcin V gene or a hybrid, mutant, analog or derivative thereof.

As used herein the term “minimal PKS” refers to those minimum number of PKS genes or subunits of a biosynthesis polyketide gene cluster required for biosynthesis of a polyketide, such as gilvocarcin. For example, in *Streptomyces griseoflavus* the required genes to encode the minimal PKS are ketosynthase I (KSI) and ketosynthase II (KSII, also known as chain length factor CLF) and an acyl carrier protein (ACP). Thus, these three genes, without the other components of the native clusters, can be included in one or more recombinant vectors, to constitute a “minimal” replacement PKS gene cluster.

As used herein the term “mutant” refers to a nucleic acid compound, protein, molecule, vector or cell resulting from mutation of the native wild type coding sequence or subunits thereof.

As used herein the term “mutation” refers to any change that alters a native coding sequence either by displacement, addition, deletion, insertion, cross-linking, or other destruction or substitution of one or more nucleotides of the native coding sequence. Techniques for modifying nucleotide sequences, such as site-directed mutagenesis, are also known to those skilled in the art.

As used herein the term “nucleic acid” sequence can include, but is not limited to, procaryotic sequences, eucaryotic mRNA, cDNA from eucaryotic mRNA, genomic DNA sequences from eucaryotic (e.g., mammalian) DNA, and even synthetic DNA sequences. The term also captures sequences that include any of the known base analogs of DNA and RNA such as, but not limited to 4-acetylcytosine, 8-hydroxy-N⁶-methyladenosine, aziridinylcytosine, pseudoisocytosine, 5-(carboxyhydroxymethyl) uracil, 5-fluorouracil, 5-bromouracil, 5-carboxymethylaminomethyl-2-thiouracil, 5-carboxymethylaminomethyluracil, dihydrouracil, inosine,

N⁶-isopentenyladenine, 1-methyladenine, 1-methylpseudouracil, 1-methylguanidine, 1-methylinosine, 2,2-dimethyl-guanine, 2-methyladenine, 2-methylguanidine, 3-methyl-cytosine, 5-methylcytosine, N⁶-methyladenine, 7-methylguanidine, 5-methylaminomethyluracil, 5-methoxy-aminomethyl-2-thiouracil, beta-D-mannosylqueosine, 5'-methoxycarbonylmethyluracil, 5-methoxyuracil, 2-methylthio-N⁶-isopentenyladenine, uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid, oxybutosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, N-uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid, pseudouracil, queosine, 2-thiocytosine, and 2,6-diaminopurine. A transcription termination sequence will usually be located 3' to the coding sequence.

As used here the term “open reading frame” or “ORF” refers to a region of a nucleic acid molecule that contains a series of triplet bases coding for amino acids without any termination codons. An “open reading frame” does include any start codons.

As used herein, the term “polyketide-type compound” refers to a compound or molecule that is encoded by at least one native polyketide subunit, or hybrid, mutant, analog, or derivative thereof, including for example, without limitation, gilvocarcin V-type polyketides.

As used herein, the term “post-PKS enzyme” or “post-PKS modifying enzyme” or “post-PKS tailoring enzyme” refers to a protein or enzyme that is involved in modifications of a polyketide after a polyketide is synthesized by polyketide synthase. Exemplary, post-PKS enzymes involved in gilvocarcin synthesis include, without limitation, a C—C-bond cleaving oxygenase involved in forming the tetracyclic lactone structure, oxygenase/dehydratase involved in forming the vinyl side chain, and C-glycosyltransferase involved in attaching the 6-deoxy-D-fuco-hexofuranose moiety.

As used herein, the term “post-PKS modifying step” or “post-PKS tailoring step” refers to an action or actions taken by a protein or enzyme to modify a polyketide after it has been synthesized by polyketide synthase. Exemplary post-PKS tailoring steps involved in gilvocarcin synthesis include, without limitation, the formation of the tetracyclic lactone structure by a C—C-bond cleaving oxygenase, oxygenation/dehydration reactions for the formation of the vinyl side chain, and a C-glycosyltransfer step involved in attaching a 6-deoxy-D-fuco-hexofuranose moiety.

As used herein the term “replacement gene cluster” means any set of genes and/or genes encoding tailoring steps capable of producing a “functional PKS” when under the direction of one or more compatible control elements, as defined above, in a host cell transformed therewith. The term “replacement gene cluster” encompasses three or more genes encoding the various proteins necessary to catalyze the production of a polyketide. A “replacement gene cluster” need not include all of the genes found in the corresponding cluster in nature. Rather, the gene cluster need only encode the necessary components to catalyze the production of an active polyketide. Thus, if the gene cluster includes, for example, eight genes in its native state and only three of these genes are necessary to provide an active polyketide, only these three genes need be present. Furthermore, a replacement gene cluster can include genes derived from a single species, or may be hybrid in nature with, e.g., a gene derived from a cluster for the synthesis of a particular polyketide replaced with a corresponding gene from a cluster for the synthesis of another polyketide. Hybrid clusters can include genes derived from both Type I and Type II PKSs. As explained above, Type I PKSs include several large multifunctional proteins carrying, between them, a set of separate active sites for each step of

carbon chain assembly and modification. Type II PKSs, on the other hand, have a single set of iteratively used active sites.⁴³ The genes included in the replacement gene cluster need not be the native genes, but can be mutants or analogs thereof. Mutants or analogs may be prepared by the deletion, insertion or substitution of one or more nucleotides of the coding sequence. Techniques for modifying nucleotide sequences, such as site-directed mutagenesis, are described in the literature. See e.g., Sambrook, et al. *Molecular Cloning: A Laboratory Manual* (Current Edition); *DNA Cloning: A Practical Approach*, Vol. I & II (D. Glover, ed.) and *Nucleic Acid Hybridization* (B. Hames & S. Higgins, eds., Current Edition).

The term, "replacement gene cluster" may also contain genes coding for modifications to the core polyketide catalyzed by a PKS, including, for example, genes encoding hydroxylases, methylases or other alkylases, oxidases, reductases, glycotransferases, lyases, ester or amide synthases, and various hydrolases such as esterases and amidases. The genes included in the replacement gene cluster need not be on the same plasmid or if present on the same plasmid, can be controlled by the same or different control sequences.

As used herein, the term "subunit" refers to a part of a complete gene cluster including, for example, a module, domain, gene, or open reading frame, and parts thereof. A "subunit" may comprise for example, a gene or genes derived from a single species or may be hybrid in nature (e.g., a gene derived from a cluster for the synthesis of a particular polyketide replaced with a corresponding gene from a cluster for the synthesis of another polyketide.). A "subunit" may comprise mutants, analogs or derivatives of the native gene(s). Mutants, analogs or derivatives thereof may be prepared by techniques known to those of skill in the art, including, without limitation, the displacement, addition, deletion, insertion, cross-linking, or other destruction or substitution of one or more nucleotides of the coding sequence. Techniques for modifying nucleotide sequences, such as site-directed mutagenesis, are also known in to those skilled in the art.

Cloning and Identification of the Gilvocarcin Biosynthetic Gene Cluster

Central to the present invention is the identification and cloning of the gilvocarcin ("gil") gene cluster (SEQ ID NO:1). Identification of the gilvocarcin gene cluster was achieved by generating a *S. griseoflavus* genomic cosmid library using the *Streptomyces-E. coli* shuttle vector pOJ446. For the generation of a genomic cosmid library, isolation and subsequent random fragmentation of high molecular weight genomic DNA was performed, followed by ligation of these fragments to vector arms containing cos sequences, packaging into λ particles and transduction into a suitable *E. coli* host strain. DNA fragments of the NDP-glucose-4,6-dehydratase (an enzyme catalyzing a key step in 6-deoxysugar biosynthesis) (See Decker, H.; Gaissner, S.; Pelzer, S.; Schneider, P.; Westrich, L.; Wohlleben, W.; Bechthold, A. *FEMS Microbiol. Lett.* 141:195-201 (1996)) and also the actI PKS (See Hopwood, D. A. *Chem. Rev.* 97:2465-2497 (1997)) genes, highly conserved among *Streptomyces*, were used to probe the cosmid library. Cosmid DNA isolated from clones hybridizing with both probes was analyzed by restriction mapping and Southern blot experiments. Hybridization using both probes with one of the cosmids increased the likelihood that the cosmid would contain the entire gilvocarcin cluster. One of the cosmids, cos-G9B3, was transformed into *S. lividans* TK24, where it stimulated the production of gilvocarcins V and M in the same quantities as the wild-type strain (20-

30mg/L of (1) in FIG. 6.B), proving that it most likely contains the entire gene cluster of gilvocarcin biosynthesis.

Subcloning of cos-G9B3-DNA fragments into pUC19 or pBluescript II SK(+) followed by sequencing revealed the entire gilvocarcin gene cluster (FIG. 6.A). The cluster spans a 32.9 kB region and consists of 26 ORFs identified as follows. ORF1 is gilS, encoded on the complement to SEQ ID NO:1, and represented on SEQ ID NO:1 as nucleotides (nt) 802-4068 read in the 3' to 5' direction. ORF2 is gilN, encoded on the complement to SEQ ID NO:1 and represented on SEQ ID NO:1 as nt 4308-5198 read in the 3' to 5' direction. ORF3 is gilL, encoded on the complement to SEQ ID NO:1 and represented on SEQ ID NO:1 as nt 5417-6052 read in the 3' to 5' direction. ORF4 is gilOIII, encoded by the complement to SEQ ID NO:1 and represented on SEQ ID NO:1 as nt 6576-7769 read in the 3' to 5' direction. ORF5 is gilGT, encoded by the complement to SEQ ID NO:1 and represented on SEQ ID NO:1 as nt 7777-9261 read in the 3' to 5' direction. ORF6 is gilM, encoded by the complement to SEQ ID NO:1 and represented on SEQ ID NO:1 as nt 9261-10001 read in the 3' to 5' direction. ORF7 is gilR, encoded by the complement to SEQ ID NO:1 and represented on SEQ ID NO:1 as nt 10020-11513 read in the 3' to 5' direction. ORF8 is gilOII, encoded by the complement to SEQ ID NO:1 and represented on SEQ ID NO:1 as nt 11513-12196 read in the 3' to 5' direction. ORF9 is gilMT, encoded by the complement to SEQ ID NO:1 and represented on SEQ ID NO:1 as nt 12354-13424 read in the 3' to 5' direction. ORF10 is gilJ, encoded by the complement to SEQ ID NO:1 and represented on SEQ ID NO:1 as nt 13814-15466 read in the 3' to 5' direction. ORF11 is gilI, nt 15619-16641 of SEQ ID NO:1 read in the 5' to 3' direction. SEQ ID NO:2 is the amino acid sequence of ORF11. ORF12 is gilE, encoded by the complement to SEQ ID NO:1 and represented on SEQ ID NO:1 as nt 16690-17697 read in the 3' to 5' direction. ORF13 is gilD, encoded by the complement to SEQ ID NO:1 and represented on SEQ ID NO:1 as nt 17697-18761 read in the 3' to 5' direction. ORF14 is gilH, nt 18938-19576 of SEQ ID NO:1 read in the 5' to 3' direction. SEQ ID NO:3 is the amino acid sequence of ORF14. ORF15 is gilOI, nt 19892-21391 of SEQ ID NO:1 read in the 5' to 3' direction. SEQ ID NO:4 is the amino acid sequence of ORF15. ORF16 is gilG, nt 21413-21736 of SEQ ID NO:1 read in the 5' to 3' direction. SEQ ID NO:13 is the amino acid sequence of ORF16. ORF17 is gilA, nt 21736-22992 of SEQ ID NO:1 read in the 5' to 3' direction. SEQ ID NO:5 is the amino acid sequence of ORF17. ORF18 is gilB, nt 22992-24164 of SEQ ID NO:1 read in the 5' to 3' direction. SEQ ID NO:14 is the amino acid sequence of ORF18. ORF19 is gilC, nt 24183-24449 of SEQ ID NO:1 read in the 5' to 3' direction. SEQ ID NO:6 is the amino acid sequence of ORF19. ORF20 is gilF, nt 24449-25225 of SEQ ID NO:1 read in the 5' to 3' direction. SEQ ID NO:15 is the amino acid sequence of ORF20. ORF21 is gilK, nt 25241-26188 of SEQ ID NO:1 read in the 5' to 3' direction. SEQ ID NO:7 is the amino acid sequence of ORF21. ORF22 is gilOIV, nt 26200-27552 of SEQ ID NO:1 read in the 5' to 3' direction. SEQ ID NO:8 is the amino acid sequence of ORF22. ORF23 is gilP, nt 27552-28481 of SEQ ID NO:1 read in the 5' to 3' direction. SEQ ID NO:16 is the amino acid of ORF23. ORF24 is gilQ, nt 28501-29517 of SEQ ID NO:1 read in the 5' to 3' direction. SEQ ID NO:9 is the amino acid sequence of ORF24. ORF25 is gilT, nt 30206-32239 of SEQ ID NO:1 read in the 5' to 3' direction. SEQ ID NO:10 is the amino acid sequence of ORF25. ORF26 is gilU, nt 32379-33392 of SEQ ID NO:1 read in the 5' to 3' direction. SEQ ID NO:11 is the amino acid sequence of ORF26.

The database analysis yielded genes encoding the type II PKS and associated enzymes (gil G, A, B, C, F, K, P and Q),

several genes encoding proteins involved in post-PKS tailoring steps (gil OIII, GT, R, OII, MT, E, D, H, OI, OIV, and U), and genes of regulation and self resistance enzymes (gil S, J, I and T), this group also includes three genes coding for proteins of unknown function (gil L, M and N).

Methods

Cosmid Library Generation

A cosmid library of the gilvocarcin V producer *Streptomyces griseoflavus* Gö3592 using pOJ446 as a host was constructed using standard methods. Transduction into *E. coli* XL 1-Blue MRF['] yielded several thousand colonies, which were pooled and used as a master culture. An initial library evaluation of cosmid DNA from several randomly chosen transductants by restriction enzyme analysis revealed whether these clones have inserts and if so, the size of the inserts. It was expected that the inserts should ideally have the size of 30 to 40 kb, since from comparison with known, related molecules, this was the expected size of the entire gene cluster of the gilvocarcin pathway. As such, we were able to generate a cosmid containing the complete gilvocarcin gene cluster.

The high molecular weight donor DNA was isolated according to the following standard methods. CRM medium supplemented with 0.5% (w/v) glycine was inoculated with *S. griseoflavus* Gö3592 spores and incubated for 24 hours or until the culture reached nearly stationary phase. Cells were harvested by centrifugation and resuspended in lysis buffer containing 1 mg/mL lysozyme. After incubation for 15-60 min at 37° C., a 10% SDS solution and proteinase K (0.5 mg/mL) was added to lyse the cells and the lysed cells were incubated at 55° C. for two hours. The resulting viscous solution was extracted with 5 M NaCl solution and phenol/chloroform 1:1 and centrifuged. The aqueous phase was transferred to a fresh tube and again extracted with chloroform, then the DNA in the water phase was precipitated by adding 0.6 vol. of isopropanol and spooled onto a sealed Pasteur pipette. After rinsing with 70% ethanol, air drying and dissolving in TE-buffer, the concentration and purity of the DNA was estimated by measuring the optical density of the solution at 260 and 280 nm. DNA size was checked on a 0.3% agarose gel by conventional electrophoresis. For cosmid cloning, the DNA should be ≥ 150 kB.

The partial digestion and dephosphorylation of the donor DNA was performed as follows by standard methods. The restriction enzyme Sau3AI was used for the partial digestion, since it recognizes a 4-base-pair-sequence thus ensuring random fragmentation and generation of cohesive ends that can be ligated into the BamHI site of vector pOJ446. Bierman, M. et al. *Plasmid cloning vectors for the conjugal transfer of DNA from Escherichia coli to Streptomyces spp.* Gene 116: 43-49 (1992). The amount of enzyme and incubation time to digest the DNA to the point where its average size is approximately in the size range of 30-40 kB was determined empirically by an enzyme dilution series and then scaled up to digest 200-300 μ g of chromosomal DNA. A subsequent dephosphorylation step with calf intestinal phosphatase (CIP) prevented segments of different regions of chromosomal DNA from ligating to one another and forming a recombinant vector containing noncontiguous segments of the genome. The extent of dephosphorylation was determined using a small batch of dephosphorylated DNA in a ligase reaction and subsequent gel electrophoresis of the ligated and unligated DNA samples. The complete dephosphorylation showed no differences between ligated and unligated DNA.

Cosmid pOJ446 is a low copy shuttle vector containing three cos sequences and an apramycin resistance gene. It also

carries the origin of replication from *E. coli* and *Streptomyces* allowing an easy transfer between these two species. To prepare the two cos arms necessary for the in vitro packaging reaction, cosmid pOJ446 was cut at the unique HpaI site situated between two cos sites. Afterwards, the resulting ends were dephosphorylated to prevent re-ligation. The linearized vector was cut with BamHI to yield two vector arms each containing a cos site and a ligatable end. Finally, phenol/chloroform extraction and ethanol precipitation was used to purify the DNA.

Ligation and packaging was performed according to the following standard methods. Different ratios of donor to vector DNA were ligated with T4 DNA ligase for 16 hours at 16° C. To favor the formation of cosmid-insert concatemers and not circular DNA the ligation was carried out at DNA concentrations of about 1 μ g/ μ L or greater. The ligation was monitored by agarose gel electrophoresis of ligation mixture, using unligated DNA samples as a control. Successful ligation showed a significant shift of the chromosomal smear to a higher molecular weight and the disappearance of the vector bands. The in vitro packaging reaction was done using Gigapack III XL packaging extracts (Stratagene), a kit which contains all required enzymes necessary to pack concatemeric DNA into preformed λ phage particles. The packaging was carried out as described in the Stratagene manual.

Transduction into *E. coli* XL 1-Blue MRF['] was performed by the following standard methods. LB medium supplemented with 10 mM MgSO₄ and 0.2% (w/v) maltose was inoculated with a single colony of *E. coli* XL 1-Blue MRF['] and grown at 37° C. for 4-6 hours. After cell harvesting by centrifugation, the cells were diluted to an OD₆₀₀ of 0.5 with sterile 10 mM MgSO₄ solution. Two hundred μ L of cell solution was then infected with in vitro packaged phage and incubated for 30 min at room temperature. Additional LB broth was added and after incubation for 1 hour, 100 μ L aliquots were plated on LB plates containing 100 μ g/mL apramycin and incubated at 37° C. overnight. To evaluate the quality of the constructed library, randomly picked colonies were proliferated in LB medium supplemented with 100 μ g/mL apramycin. Cosmid DNA was then isolated according to standard isolation procedures and used for restriction enzyme digests.

Labeling, Synthesis of Hybridization Probes and Colony Hybridization

Gene probes were labeled by PCR using the digoxigenin (DIG) system (Roche). The 4,6-dehydratase gene was amplified by PCR from genomic *S. griseoflavus* DNA using a method developed by Bechthold et al. Decker, H. et al. *A general approach for cloning and characterizing dndp-glucose dehydratase genes from actinomycetes.* FEMS Microbiology Letters 141:195-201 (1996) The PKS probe was prepared from plasmid pIJ2345, which contains parts of the actinorhodin minimal PKS (actI) of *S. coelicolor*. Malpartida, F. and Hopwood, D. *Physical and genetic characterisation of the gene cluster for the antibiotic actinorhodin in Streptomyces coelicolor A3(2).* Mol. Gen. Genet 205:66-73 (1986). Labeled DNA probes were purified by gel electrophoresis and labeling efficiency was estimated in a spot test with a DIG-labeled control.

Colony hybridization was carried out as described in the DIG user manual (Roche, online) for membrane hybridization. Roche, Molecular & Biochemicals. http://biochem.roche.com/prodinfo_fst.htm?/prod_inf/manuals/dig_man/dig_joc.htm. Briefly, colonies were grown overnight on LB agar containing 100 μ g/mL apramycin. Nylon membranes (Roche) were placed on the agar plates and punched to mark

the orientation. After a short incubation time, the membranes were removed and successively blotted for 15 minutes on Whatmann 3MM paper soaked in denaturation, neutralization and 2×SSC solutions. UV-light was used to cross-link the transferred DNA to the membrane. Membranes were then pre-hybridized for 2 hours at 42° C. in DIG Easy Hyb hybridization solution. Probes were denatured by boiling to produce single-stranded DNA and added to start the hybridization process. Probe concentration, hybridization time and temperature, and the stringency of subsequent washing steps with SSC solution were determined empirically for each probe (e.g., 2× at 45° C. with 2 mol SSC solution for 15 min., then 2× at 68° C. with 0.1 mol SSC solution for 30 min). To reduce nonspecific binding of the anti-digoxigenin-AP conjugate, the membrane was treated with blocking buffer for 30-60 min before the antibody solution was added. After removal of unbound antibodies, nitroblue tetrazolium salt (NBT) and 5-bromo-4-chloro-3-indolyl phosphate toluidinium salt (BCIP), the colorimetric substrates for AP, were added to initiate the color reaction.

Cosmid DNA isolated from clones hybridizing with both probes were analyzed by restriction mapping and Southern blot experiments. In order to confirm that the cosmid DNA indeed contained genes of the gilvocarcin gene cluster, the corresponding cosmids were introduced into *S. lividans* TK24 or *S. albus* by protoplast fusion. Hopwood, D. A. *Genetic contributions to understanding polyketide synthases*. *Chem. Rev.* 97:2465-2497 (1997); Hopwood, D. A. et al. *Genetic Manipulation of Streptomyces. A Laboratory Manual* (The John Innes Foundation, Norwich, UK) (1985); Kieser, T., Bibb, M. J., Buttner, M. J., Chater, K. F. and Hopwood, D. A. *Practical Streptomyces Genetics* (The John Innes Foundation, Norwich, UK) (2000). The resulting recombinant strains were then screened for gilvocarcin resistance and the production of new metabolites. Resistance against gilvocarcin V and/or a production of gilvocarcins or biosynthetic intermediates with the gilvocarcin chromophore, were easily detected on TLC with use of the unique yellow fluorescence (UV light, 366 nm), which proved the presence of genes of the gilvocarcin pathway.

Nucleotide Sequence Analysis

The nucleotide sequence of isolated clones was determined using conventional methodology. Automated thermocycle sequencing of pUC19 or pBluescript II SK(+)-based sub-clones using taq DNA polymerase and fluorescent dye-labeled terminators was carried out at the UK biotechnology resource service laboratory on an ABI 377 and 310 DNA sequencers. Both, standard (M13 forward and reverse, T7, or T3) and custom made primers (18-21 nucleotides) were used.

Functional Assignment of the Gilvocarcin Biosynthetic Gene Cluster

The genes encoding the PKS and associated enzymes are in a type II PKS arrangement, in which the minimal PKS gene cluster, encoded by *gilABC* (consisting of ketosynthase (KS) α , KS β , and the acyl carrier protein (ACP)), is flanked by the PKS-associated ketoreductase (KR, encoded by *gilF*) and two cyclases (encoded by *gilG* and *gilK*). Unexpectedly, the genes *gilP* and *gilQ*, which encode a malonyl CoA:ACP transacylase (MAT) and an acyl transferase (AT), respectively, were found to be located further downstream of *gilABC*. Although essential for polyketide biosynthesis, MATs are usually not found in type-II PKS gene clusters, and are often 'recruited' from the fatty acid synthase. Summers, R. G., Ali, A., Shen, B., Wessel, W. A. and Hutchinson, C. R. *Biochemistry* 34:9389-9402 (1995). The gilvocarcin gene cluster disclosed herein is the third example in which such a gene was located,

but the first one where the cluster is associated with a known structure. Novakova, R.; Bistakova, J.; Homerova, D.; Rezuchova, B.; Kormanec, J. *Gene* 297:197-208 (2002). *GilQ* resembles AT proteins found in producers of aromatic polyketides with starter units other than acetate, such as doxorubicin, enterocin, etc. Hutchinson, C. R. *Chem. Rev.* 97:2525-2536 (1997); Moore, B. S. and Hertweck, C. *Nat. Prod. Rep.* 19:70-99 (2002); Marti, T., Hu, Z., Pohl, N. L., Shah, A. N. and Khosla, C. *J. Biol. Chem.* 275:33443-33448 (2000). Therefore, *gilQ* might play a role in the choice of the starter unit (propionate vs. acetate for the production of gilvocarcin V and gilvocarcin M, respectively).

The following genes are suspected to be responsible for the post-PKS tailoring steps including the above mentioned biosynthetic steps towards the key structural features of GV. Four oxygenase-encoding genes were found (*gilOI*, *gilOII*, *gilOIII* and *gilOIV*). Genes *gilOI* and *gilOIV* encode FAD-dependent oxygenases assumed to catalyze the oxidative rearrangement of a putative angucyclinone-precursor to the unique coumarin-based aromatic core of the gilvocarcins. The corresponding enzymes *gilOI* and *gilOIV* are closely related to *jadF* and *jadH* (*gilOI*: 37% aa-identity to *jadF* and 41% aa-identity to *jadH*; *gilOIV*: 37% aa-identity to *jadF* and 29% aa-identity to *jadH*), which probably catalyze a similar rearrangement in the jadomycin pathway.

The other two oxygenases, *gilOII* and *gilOIII*, are most likely responsible for the anthrone oxidation leading to the angucyclinone intermediate X (*gilOII*), and for the generation of the 8-vinyl group. For the latter, we assume a hydroxylation in 1"-position through *gilOIII* followed by dehydration, since *gilOIII* is a P-450 hydroxylase predestined for such a reaction. Other oxidoreductase encoding genes are *gilH* (encoding a KR presumably involved in the hydroquinone generation) and *gilR* (encoding an oxidoreductase of unclear function).

The C-glycosidically linked D-fucofuranose is a unique deoxysugar not found in any other polyketide, and whose biosynthesis requires only a few enzymes (FIG. 7). Hosoya T. et al. *J. Am. Chem. Soc.* 116:1004-1015 (1994). This is confirmed by the presence of only a few typical deoxysugar biosynthesis genes in the *gil* cluster. Two of these crucial genes, *gilD* and *gilE*, encode NDP-glucose synthase and 4,6-dehydratase, respectively. A third gene possibly involved in the D-fucose biosynthesis is *gilU* located at the end of the *gil* cluster. *GilU*, apparently an epimerase/dehydratase, or oxidoreductase *gilR*, might function as 4-KR, while it is unclear how the contraction from the pyranose to the furanose is catalyzed. Without intending to be bound by theory, *gilM* is a possible candidate to encode the enzyme responsible for shrinking the pyranose to furanose.

GilGT encodes the glycosyl transferase (GT) responsible for the unusual p-OH activated C-glycosylation. Synthetic model studies suggest that the favored mechanism for C-glycosyltransfer is an initial O-glycosylation followed by a Fries-like rearrangement. See e.g., Hosoya, T. et al. *J. Am. Chem. Soc.* 116:1004-1015 (1994); Palmacci, E. R. and Seeburger, P. H. *Org. Lett.* 3:1547-1550 (2001). Although principally possible, it is difficult to imagine in an enzymatic environment that such a rearrangement to the p-position occurs. *GilGT* resembles mostly *lanGT2* and *urdGT2* both of which transfer D-olivoses to angucyclinone acceptor molecules, the latter being also a C-GT. Künzel, E., Faust, B., Oelkers, C., Weissbach, U., Bearden D. W., Weitnauer, G., Westrich, L., Bechthold, A. and Rohr, J. *Inactivation of the urdGT2 Gene, Which Encodes a Glycosyltransferase Responsible for the C-Glycosyltransfer of Activated D-Olivose, Leads to Formation of the Novel Urdamycins 1, J, and K*

J. Am. Chem. Soc. 121:11058-11062 (1999). However, in contrast to gilGT, urdGT2 places its sugar moiety ortho to a phenolic OH-group. An interesting novelty of gilGT is its unusual size, due to its N-terminal part being approximately 120 amino acids longer than any other polyketide GT found so far. The BLAST analysis of the deduced aa-sequence of gilGT (495 aa, MW 53846 g/Mol) revealed 38% aa-identity with lanGT2 from the landomycin producer *S. cyanogenus*, 31% identity with urdGT2 from *S. fradiae* Tü2717, and 25% identity with jadS, the O-GT of the jadomycin pathway from *S. venezuelae*. Without intending to be bound by theory, the similarity of gilGT with lanGT2, urdGT and jadS may point to an angucyclinone-shaped acceptor substrate for gilGT, since lanGT2, urdGT2 and jadS transfer their sugar substrates to an angucyclinone-type aglycon. Künzel, E. et al. *J. Am. Chem. Soc.* 121:11058-11062 (1999); Bechthold, A. and Rohr, J. *Bioorganic Chemistry* (eds. Diederichsen, U., Lindhorst, T. K., Westermann, B. & Wessjohann, L. A.) 313-321 (Wiley-VCH, Weinheim, 1999); Kirschning, A., Bechthold, A. and Rohr, J. *Chemical and Biochemical Aspects of Deoxysugars and Deoxysugar Oligosaccharides. Topics Curr. Chem.* 188:1-84 (1997); Wohlert, S.-E., Bechthold, A., Benninga, C., Henkel, T., Holzenkämpfer, M., Kirschning, A., Oelkers, C., Ries, M., Weber, S., Weissbach, U., Westrich, L. & Rohr, J. *Bioorganic Chemistry* (eds. Diederichsen, U., Lindhorst, T. K., Westermann, B. & Wessjohann, L. A.) 305-312 (Wiley-VCH, Weinheim, N.Y., Chichester, Brisbane, Singapore, Toronto, 1999); Hoffmeister, D., Ichinose, K., Domann, S., Faust, B., Trefzer, A., Dräger, G., Kirschning, A., Fischer, C., Künzel, E., Bearden, D. W., Rohr, J. and Bechthold, A. *The NDP-Sugar Co-Substrate Concentration and the Enzyme Expression Level Influence the Substrate Specificity of Glycosyltransferases: Cloning and Characterization of Deoxysugar Biosynthetic Genes of the Urdamycin Biosynthetic Gene Cluster. Chem. Biol.* 7:821-831 (2000); Trefzer, A., Hoffmeister, D., Künzel, E., Stockert, S., Weitnauer, G., Westrich, L., Rix, U., Fuchser, J., Bindseil, K. U., Rohr, J. and Bechthold, A. *Function of Glycosyltransferase Genes Involved in Urdamycin a Biosynthesis. Chem. Biol.* 7:133-142 (2000); Trefzer, A., Fischer, C., Stockert, S., Westrich, L., Künzel, E., Girreser, U., Rohr, J. and Bechthold, A. *Elucidation of the Function of Two Glycosyltransferase Genes (aanGT1 and lanGT4) Involved in Landomycin Biosynthesis and Generation of New Oligosaccharide Antibiotics. Chem. Biol.* 8:1239-1252 (2001); Krohn, K. et al, *Topics Curr. Chem.* 188:127-195 (1997); Westrich, L., Domann, S., Faust, B., Bedford, D., Hopwood, D. A. and Bechthold, A. *Cloning and Characterization of a Gene Cluster from Streptomyces cyanogenus S136 Probably Involved in Landomycin Biosynthesis. FEMS Microbiol. Lett.* 170:381-387 (1999).

Methods

ORF assignments were accomplished using the GCG software package (University of Wisconsin) and the NCBI database. When applying the GCG software on *Streptomyces*, assignment priority will be given to ORFs with consistently high G/C %. Preliminary gene assignments were then derived from the translated amino acid sequence similarity of translated genes of known function using the BLAST (Basic Local Alignment Search Tool) program, and standard protein sequence data bases (Genbank, EMBO, Swiss Prot). Altschul, S. F., Gish, W., Miller, W., Myers, E. W. and Lipman, D. J. *Basic Local Alignment Research Tool. J. Mol. Biol.* 215: 403-410 (1990); Altschul, S. F. and Lipman, D. J. *Protein Data Base Searches for Multiple Alignments. Proc. Natl. Acad. Sci. USA* 87: 5509-5513 (1990); Altschul, S. F. et al.

Gapped BLAST and Psi-BLAST—A New Generation of Protein Database Search Programs. Nucl. Acid Res. 25:3389-3402 (1997).

Host-Vector System

Identification and cloning of the gilvocarcin V gene cluster led to the discovery of a host-vector system for the efficient recombinant production of both novel and known polyketides. The coding sequences which collectively encode a gilvocarcin V gene cluster or hybrids, mutants, analogs or derivatives thereof, can be inserted into one or more expression vectors, using methods known to those of skill in the art. The replacement gene cluster need not correspond to the complete native gilvocarcin gene cluster but need only encode a minimal PKS gene cluster to catalyze the production of a polyketide.

The recombinant vector(s) of the present invention includes replacement gene clusters derived from a single gene cluster, or may comprise hybrid replacement gene clusters with, e.g., a gene of one cluster replaced by the corresponding gene from another gene cluster. For example, acyl carrier proteins (ACPs) or certain deoxysugar genes are readily interchangeable among different synthases without an effect on the product structure. Furthermore, a given ketosynthase (KS) or ketoreductase (KR) may recognize or reduce polyketide chains of different chain lengths. Accordingly, these genes may be freely interchangeable in the constructs described herein. Thus, the replacement clusters of the present invention can be derived from any combination of PKS gene sets, which ultimately function to produce an identifiable polyketide.

Expression vectors also include control sequences operably linked to the desired PKS coding sequence. Suitable expression systems for use with the present invention include systems, which function in eucaryotic and procaryotic host cells. However, procaryotic systems are preferred, and in particular, systems compatible with *Streptomyces* species are of particular interest. Control elements for use in such systems include promoters, optionally containing operator sequences, and ribosome binding sites. Particularly useful promoters include control sequences derived from PKS gene clusters, such as one or more act promoters. However, other bacterial promoters, such as those derived from sugar metabolizing enzymes, such as galactose, lactose (lac) and maltose, will also find use in the present constructs. Additional examples include promoter sequences derived from biosynthetic enzymes such as tryptophan (trp), the beta-lactamase (bla) promoter system, bacteriophage lambda PL, and T5. In addition, synthetic promoters, such as the tac promoter, which do not occur in nature also function in bacterial host cells.

Other regulatory sequences may also be desirable which allow for regulation of expression of the replacement PKS gene cluster relative to the growth of the host cell. Regulatory sequences are known to those of skill in the art, and examples include those which cause the expression of a gene to be turned on or off in response to a chemical or physical stimulus, including the presence of a regulatory compound. Other types of regulatory elements may also be present in the vector, for example, enhancer sequences.

Selectable markers can also be included in the recombinant expression vectors. A variety of markers are known which are useful in selecting for transformed cell lines and generally comprise a gene whose expression confers a selectable phenotype on transformed cells when the cells are grown in an appropriate selective medium. Such markers include, for example, genes which confer antibiotic resistance or sensi-

tivity to the plasmid. Alternatively, several polyketides are naturally colored and this characteristic provides a built-in marker for selecting cells successfully transformed by the present constructs.

The various subunits of gene clusters of interest can be cloned into one or more recombinant vectors as individual cassettes, with separate control elements, or under the control of, e.g., a single promoter. These subunits can include flanking restriction sites to allow for the easy deletion and insertion of other subunits so that hybrid gene clusters can be generated. The design of such unique restriction sites is known to those of skill in the art and can be accomplished using the techniques described above, such as site-directed mutagenesis and PCR.

Further, the vectors, which collectively encode a replacement gene cluster can be inserted in to one or more host cell, using methods known to those of skill in the art. As such, the present invention also provides host cells which have their naturally occurring gene substantially deleted, transformed with vectors encoding a replacement gene cluster or parts thereof, for the production of active polyketides. The invention provides for the production of significant quantities of product at an appropriate stage of the growth cycle. The polyketides so produced can be used as therapeutic agents, to treat a number of disorders, depending on the type of polyketide, like immunosuppressants, anti-tumor agents, as well as for the treatment of viral, bacterial and parasitic infections. The ability to recombinantly produce polyketides also provides a powerful tool for characterizing biosynthetic enzymes and the mechanism of their actions.

More particularly, host cells for the recombinant production of the subject polyketides can be derived from any organism with the capability of harboring a recombinant PKS gene cluster. Thus, the genetically engineered host cells of the present invention can be derived from either procaryotic or eucaryotic organisms. Preferably, the host may be *E. coli*. However, more preferred host cells are those constructed from the actinomycetes (act), a class of mycelial bacteria which are abundant producers of a number of polyketides. A particularly preferred genus for use with the present system is *Streptomyces*. Thus, for example, *S. ambofaciens*, *S. argillaceus*, *S. avermitilis*, *S. azureus*, *S. cinnamonensis*, *S. coelicolor*, *S. curacoi*, *S. cyanogenus*, *S. erythraeus*, *S. fradiae*, *S. galilaus*, *S. glaucescens*, *S. globisporus*, *S. griseus*, *S. hygrosopicus*, *S. lividans*, *S. parvulus*, *S. peucetius*, *S. rimosus*, *S. roseofulvus*, *S. thermotolerans*, *S. venezuelae*, *S. violaceoruber*, among others, will provide convenient host cells for the subject invention. See e.g., Hopwood, D. A. and Sherman, D. H. *Ann. Rev. Genet.* 24:37-66 (1990); O'Hagan, D. *The Polyketide Metabolites* (Ellis Horwood Limited, 1991).

The above-described host cells are genetically engineered by deleting the naturally occurring PKS genes or genes encoding post-PKS tailoring enzymes therefrom, using standard techniques, such as by homologous or heterologous recombination. See e.g., Khosla, C. et al. *Molec. Microbiol.* 6:3237 (1992) One or more recombinant vector, collectively encoding a replacement gene cluster of the present invention, is then introduced into a host cell. The vector(s) can include native or hybrid combinations of gilvocarcin gene cluster subunits, or mutants, analogs, or derivatives thereof. Methods for introducing the recombinant vectors of the present invention into suitable host cells are known to those of skill in the art and typically include the use of CaCl_2 or other agents, such as divalent cations and DMSO. DNA can also be introduced into bacterial cells by electroporation. Once the genes or gene clusters are expressed, the polyketide producing colonies can

be identified and isolated using known techniques. The produced polyketides can then be further characterized, e.g. by NMR and mass spectroscopy.

5 Generation of New Gilvocarcin-Type Compound Producing Hybrids, Mutants, Analogs and Derivatives of *Streptomyces griseoflavus*

The generation of new gilvocarcin-type drugs, and gilvocarcin analogs and derivatives thereof can be produced by known methods in the art. Native gene sequences or parts thereof can be used alone or in combination with non-native gene sequences or parts thereof to produce analogs or hybrids of *Streptomyces griseoflavus*. For example, the replacement gene or gene cluster or subunits thereof of interest can be obtained from an organism that expresses the same, using recombinant methods, such as by screening cDNA or genomic libraries, derived from cells expressing the gene of interest, or by deriving the gene from a vector known to include the same. The gene can then be isolated and combined with other desired genes, using standard techniques. If the gene in question is already present in a suitable expression vector, it can be combined in situ, with, e.g., other PKS subunits or genes encoding tailoring enzymes such as deoxysugar genes, as desired. The gene of interest can also be produced synthetically, rather than cloned. The nucleotide sequence can be designed with the appropriate codons for the particular amino acid sequence desired. In general, one will select preferred codons for the intended host in which the sequence will be expressed. The complete sequence is assembled from overlapping oligonucleotides prepared by standard methods and assembled into a complete coding sequence. Edge. *Nature* 292:756 (1981); Nambair et al. *Science* 223:1299 (1984); Jay et al. *J. Biol. Chem.* 259:6311 (1984).

The replacement gene clusters of the present invention are derived from a single gene cluster, or may comprise hybrid replacement gene clusters with, e.g., a gene for one cluster replaced by the corresponding gene from another gene cluster. Non-limiting exemplary non-gilvocarcin V biosynthetic genes may be subunits of the gilvocarcin M, gilvocarcin E, defucosyl-gilvocarcin V, ravidomycin, deacetyl-ravidomycin, FE35A, FE35B, chrysomycin A, chrysomycin B, BE-12406 A, or BE-12406 B gene cluster. For example, deoxysugar pathways have common enzymes, which start the pathway, and which then can be complemented with various genes known from various sugar pathways in order to create novel or altered sugar moieties. Accordingly, these genes are freely interchangeable in the constructs described herein. Thus, the replacement clusters of the present invention can be derived from any combination of gene sets, which ultimately function to produce an identifiable new polyketide-type compound.

Examples of hybrid replacement gilvocarcin gene clusters include clusters with genes derived from two or more of the act gene clusters, such as granaticin (gra), gilvocarcin (gil), urdamycin (urd), landomycin (lan), mithramycin (mtm), tetracenomycin (tcm), oxytetracycline (otc), tetracycline (tet), erythromycin (ery), oleandomycin (ole), griseusin, nanaomycin, medermycin, daunorubicin, tylosin, carbomycin, spiramycin, avermectin, monensin, nonactin, curamycin, rifamycin and candicidin synthase gene clusters, among others.

Mutations can be made to the native gene sequences and such mutants used in place of the native sequence, so long as the mutants are able to function with other genes to collectively catalyze the synthesis of an identifiable polyketide. Such mutations can be made to the native sequences using conventional techniques such as by preparing synthetic oli-

gonucleotides including the mutations and inserting the mutated sequence into the gene encoding a PKS subunit using restriction endonuclease digestion. See e.g., Kunkel, T. A. *Proc. Natl. Acad. Sci. USA* 82:448(1985); Geisselsoder et al. *BioTechniques* 5:786 1987). Alternatively, the mutations can be effected using a mismatched primer (generally 10-20 nucleotides in length) which hybridizes to the native nucleotide sequence (generally cDNA corresponding to the RNA sequence), at a temperature below the melting temperature of the mismatched duplex. The primer can be made specific by keeping primer length and base composition within relatively narrow limits and by keeping the mutant base centrally located. Zoller and Smith, *Methods Enzymol.* 100:468 (1983). Primer extension is effected using DNA polymerase, the product cloned and clones containing the mutated DNA, derived by segregation of the primer extended strand, selected. Selection can be accomplished using the mutant primer as a hybridization probe. The technique is also applicable for generating multiple point mutations. See e.g., Dalbie-McFarland et al. *Proc. Natl. Acad. Sci USA* 79:6409 (1982).

In summary, this mixing and matching on biosynthetic genes, also called combinatorial biosynthesis is a new method of drug derivatization and SAR (structure-activity-relationship) assessment, which not only generates new drug analogs and derivatives but also the bacterial mutant strains for the biotechnological production of the new drugs.

Gene recombination is a well-known method in the art used for generating new gilvocarcin-type drug analogs and derivatives and new bacterial mutants. For example, gene recombination involves transforming a host cell, including the GV producer *Streptomyces griseoflavus* Gö 3592 or the newly generated GV-producer *S. lividans* TK24 (cos-G9B3) or other strains, with a recombinant vector encoding specific foreign genes. For the plasmid constructions, *Streptomyces-E. coli* shuttle vectors containing the strong constitutive ermE* promoter, which allows an overexpression of the inserted genes can be used. The recombinant vectors may be transferred into a host cell by either protoplast transformation or conjugal plasmid transfer. Non-limiting exemplary suitable genes and plasmids known in the art include the oxygenase genes jadF,G,H, and the GT-encoding genes jadS, urdGT2, lanGT2 and various other plasmids with deoxysugar biosynthesis genes.

Gene disruption, a method of generating a knockout or minus-mutants, can also be used to generate mutations for inclusion in a replacement PKS gene cluster. Knockouts are made by standard methods well established in the art, namely, insertional inactivation and in-frame gene deletion. See, e.g., Künzel, E.; Faust, B.; Oelkers, C.; Weissbach, U.; Bearden D. W.; Weitnauer, G.; Westrich, L.; Bechthold, A. and Rohr, J. *J. Am. Chem. Soc.* 121:11058-11062 (1999); Westrich, L.; Domann, S.; Faust, B.; Bedford, D.; Hopwood, D. A. and Bechthold, A. *FEMS Microbiol. Lett.* 170, 381-387 (1999); Remsing, L. L., Garcia-Bernardo, J., Gonzalez, A., Kunzel, E., Rix, U., Brana, A. F., Bearden, D. W., Mendez, C., Salas, J. A. and Rohr, J. *Ketopremiithramycins and Ketomithramycins, Four Aureolic Acid-Type Compounds Obtained Upon Inactivation of Two Genes Involved in the Biosynthesis of the Deoxysugar Moieties of the Antitumor Drug Mithramycin by Streptomyces argillaceus, Reveal Novel Insights into Post-Pks Tailoring Steps of the Mithramycin Biosynthetic Pathway.* *J. Am. Chem. Soc.* 124:1606-1614 (2002).

Generally, insertional inactivation is obtained by subcloning a gene fragment from a strain containing the gene of interest into a suitable plasmid (e.g. pBSKT, pBluescript or pUC18-derivatives etc.), inserting an apramycin or other

resistance cassette into the plasmid and transformation of this plasmid construct into the same strain from which the gene of interest came. Selection for both thioestreptone resistance (in the plasmid) and for apramycin or other suitable antibiotic resistance then indicates the knockout of the target gene. A stable double-crossover mutant is achieved by replacing the wild-type region by the in vitro-altered one. Remsing, L. et al. *J. Am. Chem. Soc.* 124:1606-1614 (2002).

Another method to inactivate genes is the Donnenberg method. See e.g., Donnenberg, M. S. and Kaper, J. B. *Construction of an Eae Deletion Mutant of Enteropathogenic Escherichia Coli by Using a Positive-Selection Suicide Vector.* *Infect. Immun.* 59: 4310-4317 (1991); Donnenberg, M. S. and Kaper, J. B. *Enteropathogenic Escherichia coli.* *Infect. Immun.* 60:3953-3961 (1992); Donnenberg-laboratory webpage: <http://medschool.umaryland.edu/infemsd/resources.htm>. In this method, suitable fragments for gene inactivation are cloned into pCVD442 and introduced into *E. coli* SM10- λ . The *E. coli* SM10- λ strain supplies the tra genes for conjugation of the pCVD442 inactivation construct into *E. coli* RR1 cells containing cos-G9B3. Cells growing under selection for apramycin, ampicillin and streptomycin (for streptomycin sensitive *E. coli* SM10- λ donor cells) should have the pCVD442 derivative integrated into cos-G9B3 via a single cross over event. Growing the cells without the addition of ampicillin and plating them out on agar plates containing 5% (w/v) sucrose should give colonies containing either the original cos-G9B3 or a mutated version of cos-G9B3 with the desired deletion. Restriction enzyme analysis is used to differentiate between these two colonies.

Also, the recently developed REDIRECT[®] technology can be used, which allows a fast insertion of an antibiotic resistance marker into the gene of interest through PCR. See e.g., Gust, B., Kieser, T. and Chater, K. F. *Redirect Technology: PCR Targeting System in Streptomyces coelicolor.* The John Innes Center; www.plantbioscience.com (2002).

As an alternative method for generating knockouts, in-frame gene deletion can be used to inactivate unwanted biosynthetic genes. Kulowski, K. et al. *Functional characterization of the jadI gene as a cyclase forming angucyclinones.* *Journal of the American Chemical Society* 121:1786-1794 (1999). Such methods are known to those skilled in the art.

EXAMPLES

The generation of the specific gene minus mutants such as, inter alia, gilGT, gilOI, gilOIV and gilMT are useful for determining the biosynthetic steps of gilvocarcin V and lead to the creation of gilvocarcin-type drug mutants and the discovery of new gilvocarcin-type drug analogs and derivatives thereof.

Inactivation of GilGT

For generation of a gilGT-minus-mutant, an in-frame deletion of a 267 base-pair (bp) segment flanked by two XhoI restriction sites within the gilGT gene is anticipated. For this, a SphI fragment of cos-G9B3 carrying gilGT is cloned into the same site of pUC19. Digestion with XhoI and religation of the vector removes the 267 bp-fragment. The resulting shorter gene fragment is then rescued as an EcoRI-HindIII fragment and cloned into a suitable *Streptomyces* suicide plasmid (e.g. pBSKT or pHZ1358; both carrying the tsr thioestreptone resistance gene). The resulting plasmid is then introduced into the *S. griseoflavus* Gö3592 wild type strain either by protoplast transformation or by conjugal transfer from *E. coli* ET12567 (pUB307). Selection for thioestreptone resistance should yield *S. griseoflavus* mutant strains with the entire vector integrated

into the chromosome by a single crossover event. These mutants are used to generate a stable double crossover mutant by allowing them to grow without selective pressure and subsequent screening for thiostrepton sensitive mutants. A successful second crossover event, yielding the gilGT-minus mutant strain, is verified by southern hybridization experiments. Without intending to be bound by theory, it is expected that the gilGT-minus mutant will accumulate the acceptor substrate of gilGT, namely either angucyclinone 2 or defucogilvocarvin E 4 (Scheme 1 of FIG. 8).

To confirm that the resulting product(s) of the inactivation gilGT-mutants are really only an effect of the respective gene inactivation, the gilGT-mutants will be complemented with gilGT. For these experiments, gene fragments containing the gilGT gene will be ligated into an expression vector containing an antibiotic resistance marker, such as pEM4, pWHM3 or pUWL201-1 (all thiostrepton), and the minus-mutants will be transformed with the resulting plasmid. Growth in a medium containing thiostrepton will yield the complementation strain, whose product spectrum will be analyzed and compared to the wild-type strain. We expect an essentially identical product spectrum as from the wild-type strain. It is hypothesized that the inactivation of gilGT will help to clarify the sequence of events of the gilvocarcin biosynthesis, and will provide insights regarding the gilGT acceptor substrate, which is important for the generation of new gilvocarcin-type analogs.

Inactivation of GilOI

Without intending to be bound by theory, it is hypothesized that that gilOI/gilOIV encode the enzymes catalyzing a C—C-bond cleaving step for gilvocarcin biosynthesis. As with gilGT, in-frame deletion is possible for gilOI, due to the two suitable KpnI sites found in gilOI, which should allow the deletion of a 578-bp fragment. Inactivation of gilOI follows the general experimental procedure outlined above for the gilGT inactivation and confirmation complementation. Without intending to be bound by theory, we expect due to the inactivation of gilOI/gilOIV an accumulation of either angucyclinone (2) or a glycosylated intermediate, such as (5) as set out in Scheme 1 of FIG. 8.

Mutation of GilGT.

The sequence comparison of gilGT with other glycosyltransferase encoding genes, in particular with urdGT2, showed that gilGT is about 300 base pairs longer at the beginning of the ORF, i.e. gilGT translates into a protein, which contains roughly 100 extra amino acids at its N-terminal end. The mutation, in which this extra portion of gilGT is removed, will be achieved by overexpression of a 300-bp shorter version of gilGT gene into the gilGT-mutant.

It is hypothesized that if the unique portion of gilGT is responsible for forcing the activated D-fucose from the pyranose into the furanose configuration, then the mutation experiment might yield a gilvocarcin bearing a fucopyranose moiety instead of the fucofuranose moiety, presuming that the remaining portion of the GT remains functional.

Synthetic studies suggest that such C-glycosides arise from O-glycoside intermediates via Fries-like rearrangement. Therefore, the glycosylation sequence probably leads first to the O-glycoside, and then the sugar moiety migrates to the neighbor carbon atom. For most C-glycosides, like the C-glycosidic D-olivose in urdamycin A, this is an ortho-shift. However, gilvocarcin V does not possess an oxygen atom in ortho-position, and since the Fries-rearrangement allows both, an o- and p-shift the C-glycosylation must proceed via the p-OH group. We hypothesize that the unique extra segment of gilGT might encode larger binding sites enabling this more complicated p-Fries rearrangement (e.g. through suiting two donor and two acceptor substrates). Therefore, the anticipated

mutation gilGT might yield a molecule bearing an O-glycosidically linked sugar at 1-position instead of the usual C-glycosidically linked sugar at 4-position.

Complementation of the GilGT-Minus Mutant with Foreign GT-genes (UrdGT2, LanGT2 or JadS)

If the acceptor substrate of gilGT is angucyclinone (2) then the gilGT-minus-mutant (see above) is likely to be successfully complemented with urdGT2, which is a gene encoding the C-glycosyltransferase of the urdamycin pathway, for which a similar acceptor substrate is discussed. As the result of this complementation experiment, we expect an ortho-C-glycosylated product. This can be either an angucyclinone, or a novel gilvocarcin-type molecule depending on the substrate flexibility of the downstream enzymes of gilvocarcin biosynthesis. To accomplish this complementation experiment, the urdGT2 genes are inserted into plasmid pEM4, a pWHM3-derived overexpression vector. UrdGT2 is known to possess very broad substrate specificity to both the acceptor as well as the NDP (nucleosyldiphosphate)-activated sugar donor substrate. In case it should be unable to handle activated D-fucose (or one of its biosynthetic intermediates) provided by *S. griseoflavus* G63592, the resulting mutant strain *S. griseoflavus* gilGT-minus (urdGT2) can be complemented with designed plasmids (pLN2 derivatives) (See e.g., Rodriguez, L., Aguirrezabalaga, I., Allende, N., Brana, A. F., Mendez, C. & Salas, J. A. *Engineering Deoxysugar Biosynthetic Pathways from Antibiotic-Producing Microorganisms: A Tool to Produce Novel Glycosylated Bioactive Compounds*. *Chem. Biol.* 9:721-729 (2002)) providing NDP (nucleosyldiphosphate)-D-olivose, NDP-D-mycarose or NDP-D-rhodinose, which are known sugar donor substrates of urdGT2. These plasmids (e.g., pLNR for NDP-D-olivose) are available from our collaboration with J. A. Salas et al., or can be designed. For example, to achieve D-rhodinose, urdR in pLNR (Generates D-Olivose) has to be replaced by mtmU (Generating an Axial 4-OH) and complemented with urdQ; for D-mycarose, pLNR needs to be complemented with mtmC.

Complementation experiments using lanGT2 and jadS can be carried out following the same procedure. Here, O-glycosidically bound sugars are expected, since lanGT2 and jadS yield O-glycosides.

Complementation of the Gil Gene Cluster with Other Suitable Deoxysugar Biosynthesis Genes

As illustrated in Scheme 2 of FIG. 7, there are only a few genes necessary to encode the biosynthesis of the deoxysugar moiety of gilvocarcin V. These genes can be complemented with other known genes of sugar pathways to generate various new gilvocarcin-type drugs with alternated sugar moieties. Basically, the gil gene cluster contains all elements to achieve activation (necessary for the glycosyltransfer) and 6-deoxygenation (a common step of all deoxysugar pathways, catalyzed by a 4,6-dehydratase). The gil cluster does not contain genes of branching elements (such as C-methyltransferases), amination elements (transaminases) and further deoxygenating enzymes. Only a few genes are needed to alter the D-fucofuranose moiety of GV sugar into an amino sugar (with or without methyl groups), into a branched sugar, or into a more deoxygenated sugar. Many of these genes, e.g. those encoding the 3-deoxygenation or 2-deoxygenation, are known from many pathways. Without intending to be bound by theory, both, pyranose or furanose GV moieties are expected.

Inactivation Experiments to Generate Increased Hydrophilicity.

Gene inactivation experiments can be used to generate less lipophilic gilvocarcin-type drugs: (i) inactivation of gilMT (presumably the methyltransferase responsible for the introduction of both O-methyl groups in 10- and 12-position), (ii) inactivation of gilE (the 4,6-dehydratase catalyzing the first

deoxygenation step in deoxysugar biosynthesis), and (iii) inactivation of *gilU* (or *gilR*, one of which is probably the ketoreductase of the sugar pathway). The experimental procedure will be analogous to the above-described examples. Without intending to be bound by theory, we expect that the first experiment should yield unmethylated GV, the second might yield a D-glucose analog of GV, and the third might yield a GV-analog with a keto sugar, which often is found as the hydrate form. All anticipated derivatives should be significantly more hydrophilic than the parent drug. Also, combinations of these mutations, e.g. a *gilMT/gilE*-double mutant can be envisaged, if the previous experiments are successful.

Determining Gilvocarcin Therapeutic Indications

The gilvocarcins, its analogs and derivatives thereof, of the present invention are useful as antibiotics, antitumor agents, immunosuppressants, antivirals and neuroprotective agents. Considering the fact that gilvocarcins are quite lipophilic compounds making them good candidates to pass the blood-brain barrier (BBB), brain tumors may be treatable by targeted submission of light (e.g., through fiber optics) after gilvocarcin chemotherapy. This would cause only few systemic side effects due to the absence of light elsewhere. This is attractive, since brain tumor surgery is often impossible. Targeted submission of light through fiber optics after gilvocarcin chemotherapy may be also an attractive treatment of prostate cancer. A more selective systemic treatment of leukemia might be possible with photoactivatable drugs like gilvocarcin V, since blood can be channeled outside the human body, where it is light exposed, while no major side effects will occur inside the body due to the exclusion of light. Also treatment of proliferative eye diseases, such as glaucoma, may be another future application of gilvocarcin-type anticancer drugs.

To determine anticancer therapeutic uses of gilvocarcin, its analogs and derivatives thereof, a compound derived from the host-vector recombinant production system is assayed as drugs against selected cancers or diseases in vitro and in vivo. Initially gilvocarcin and gilvocarcin analogs and derivatives are screened against selected human skin, brain, leukemia and prostate cancer cell lines, e.g., UACC-62, MALME-3M, SK-MEL-5 (melanoma), SF-268, SNB-75, U251 (brain tumors), CCRF-CEM, K-562, MOLT-4 (leukemia), and PC-3, DU-145 (prostate). To gain initial toxicity data on non-cancerous cells, similar experiments can be performed using normal epithelial and fibroblast cell cultures, purchased from ATCC or Clonetics.

The MTT assay, which measures the reduction of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H tetrazolium bromide (MTT) by mitochondrial succinate dehydrogenase to an insoluble, colored, formazan product, is performed according to described standard procedures.

The SRB assay is a rapid and sensitive method to measure drug-induced cytotoxicity. It measures the uptake of sulforhodamin B (SRB), which is dependent on the cellular protein quantities, and is performed in 96-well microtiter plates according to the protocol published by Boyd et al. (NCI). See also, Skehan, P. et al. *New colorimetric cytotoxicity assay for anticancer-drug screening*. *Journal of the National Cancer Institute*. 82:1107-12 (1990).

Although illustrative embodiments of the present invention have been described in detail, it is to be understood that the present invention is not limited to those precise embodiments, and that various changes and modifications can be effected therein by one skilled in the art without departing from the scope and spirit of the invention as defined by the appended claims.

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ctg ctg gcc gac gga gtc agc gga ctg gac atc cgc tac gac atg gcc Leu Leu Ala Asp Gly Val Ser Gly Leu Asp Ile Arg Tyr Asp Met Gly 930 935 940	21061
ctc ccg gaa gca ccg cca ccc acg ggt gaa cgg ctg ccg ccg gac gtg Leu Pro Glu Ala Pro Pro Thr Gly Glu Arg Leu Pro Pro Asp Val 945 950 955 960	21109
ttc cac gtc gtc ggg acc ggc ggc gac gcc gtc gag gag ttg cgg cac Phe His Val Val Gly Thr Gly Gly Asp Ala Val Glu Glu Leu Arg His 965 970 975	21157
ggc gcc gct ctg ctg atc gtc ccg tcc ccc gac agc ccg gcg tcc tcg Gly Ala Ala Leu Leu Ile Val Pro Ser Pro Asp Ser Pro Ala Ser Ser 980 985 990	21205
ctg gtc gct ccg tgg cgg gac cag gtg cgc gtc gtg cac gcg cgc ccc Leu Val Ala Pro Trp Arg Asp Gln Val Arg Val Val His Ala Arg Pro 995 1000 1005	21253
acg gac ccg gac tgg ggc ggg gag ccg gcc gcg tcg tcg cac tgg Thr Asp Pro Asp Trp Gly Gly Glu Pro Ala Ala Ser Ser His Trp 1010 1015 1020	21298
ttc gta cga ccg gac gga cac atc gcg tgg gcg gcc acc gaa ttc Phe Val Arg Pro Asp Gly His Ile Ala Trp Ala Gly Thr Glu Phe 1025 1030 1035	21343
agc gag ttg agc gcc tca ctg agc cgc tgg ctc ggt cag ccc gcc Ser Glu Leu Ser Ala Ser Leu Ser Arg Trp Leu Gly Gln Pro Ala 1040 1045 1050	21388
gcg taaccagagg aggaagaacc cttgttcagc tctctcatcg tcgccccgat Ala	21441
ggacaccggc caccgcaag cgggtggccga cgtcttcgcc ggcttcgacg ccaccgacat	21501
gcccgcgcgg atgggcacgc ggcgcgcga actcttcgcg taccgcggcc tctactcca	21561
ctccaggac ttcgagacc cgaacgggac cgaagcggtc gaggcggcca agtccgaccc	21621
gcggttcate cgggtgagca acgacctcag gccctacac gaggcctacg ccccgactg	21681
gcaatcaccg aaggacgcca tggcagagcg cttctatcac tggagttcga aacg atg Met 1055	21738

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Ser Arg Arg Val Phe Ile Thr Gly Val Gly Val Val Ala Pro Gly	
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gcc gtc gga cgt gac ccc ttc tgg gag ctg ctg acc caa ggg cgc	21828
Ala Val Gly Arg Asp Pro Phe Trp Glu Leu Leu Thr Gln Gly Arg	
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acg gcc acc cgc cgg ctc agc ctc tgc gac ccg gag ccc ttc cgg	21873
Thr Ala Thr Arg Arg Leu Ser Leu Cys Asp Pro Glu Pro Phe Arg	
1090 1095 1100	
tcc cag gtg gcc gcg gag gcc gac ttc gac gcc gag gcg gcg ggg	21918
Ser Gln Val Ala Ala Glu Ala Asp Phe Asp Ala Glu Ala Ala Gly	
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ctg tcg gag cgg cag tcc gcg gaa ctg gac cgg gcg gcg cag ttc	21963
Leu Ser Glu Arg Gln Ser Ala Glu Leu Asp Arg Ala Ala Gln Phe	
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gcc ctg gtc gcc gcc cgt gaa gcg gtc gag gac gcg gca tgg tcc	22008
Ala Leu Val Ala Ala Arg Glu Ala Val Glu Asp Ala Ala Trp Ser	
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Glu Thr Cys Pro Pro Glu Arg Ala Gly Val Ile Val Gly Ser Ala	
1150 1155 1160	
gtc gga gcc acg acc aag ctc gag gag gtc tac cgg cag ctc agc	22098
Val Gly Ala Thr Lys Leu Glu Glu Val Tyr Arg Gln Leu Ser	
1165 1170 1175	
cgt gac ggc tcc ctc tgg gac gtg gcc ccc gac tcc ccc gcc gag	22143
Arg Asp Gly Ser Leu Trp Asp Val Ala Pro Asp Ser Pro Ala Glu	
1180 1185 1190	
ctg tac tcg tac ttc gtg ccc agc tcg ttc gcc tcc ggc atc gca	22188
Leu Tyr Ser Tyr Phe Val Pro Ser Ser Phe Ala Ser Gly Ile Ala	
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His Asp Leu Gly Val Thr Gly Gln Ser Gly Val Val Ser Thr Gly	
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Cys Thr Ser Gly Ile Asp Ser Val Gly Asn Ala Trp Glu Leu Ile	
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Gln Ser Gly Ile Leu Asp Ser Ala Val Cys Gly Ala Thr Asp Ala	
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ccc atc tcg ccc atc acc gtc gcc tgc ttc gac acg atc aag gcg	22368
Pro Ile Ser Pro Ile Thr Val Ala Cys Phe Asp Thr Ile Lys Ala	
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Thr Ser Thr Tyr Asn Asp Thr Pro Glu Ser Ala Ser Arg Pro Phe	
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gac gcc aca cgg ggc ggc ttc gtc ctc ggc gag ggc agc gcg atg	22458
Asp Ala Thr Arg Gly Gly Phe Val Leu Gly Glu Gly Ser Ala Met	
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Phe Val Leu Glu Ser Glu Glu Ser Val His Arg Arg Gly Ala Arg	
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gtc tac ggc gag atc cgc ggc tac gcg agc cgc tgc aac gcc tac	22548
Val Tyr Gly Glu Ile Arg Gly Tyr Ala Ser Arg Cys Asn Ala Tyr	
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cac atg acc ggt ctc aag gcc gac gga cgc gag ctg gcg gag gcc	22593
His Met Thr Gly Leu Lys Ala Asp Gly Arg Glu Leu Ala Glu Ala	
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gtc gtc tcc gct ctc ggc cag gca ggc gtg gac ccg ggc cgg ctc	22638
Val Val Ser Ala Leu Gly Gln Ala Gly Val Asp Pro Gly Arg Leu	
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Asp Tyr Val Asn Ala His Gly Ser Gly Thr Lys Gln Asn Asp Arg	
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cac gag acc gcc gcg ctg aag tcg tcc ctc gga ccc gcc gcc cac	22728
His Glu Thr Ala Ala Leu Lys Ser Ser Leu Gly Pro Ala Ala His	
1375 1380 1385	
gac gtg ccg atc agt tcg atc aag tcg atg atc gcc cat tcg ctg	22773
Asp Val Pro Ile Ser Ser Ile Lys Ser Met Ile Gly His Ser Leu	
1390 1395 1400	
ggc gcc atc ggg tcg ttg gag atc gcc gcc tgc gcc ctg gcg ctg	22818
Gly Ala Ile Gly Ser Leu Glu Ile Ala Ala Cys Ala Leu Ala Leu	
1405 1410 1415	
cgg gac gac gtg atc ccg ccg acg gcc aat ctc acc cgg ccg gat	22863
Arg Asp Asp Val Ile Pro Pro Thr Ala Asn Leu Thr Arg Pro Asp	
1420 1425 1430	
ccg gaa ctc gat ctg gac tac gtg ccg gtc cac gcg cgc aag cag	22908
Pro Glu Leu Asp Leu Asp Tyr Val Pro Val His Ala Arg Lys Gln	
1435 1440 1445	
ccg acc aac agc gtg ctc acg acc gga agc ggc ttc ggt ggg ttt	22953
Pro Thr Asn Ser Val Leu Thr Thr Gly Ser Gly Phe Gly Gly Phe	
1450 1455	
cag agc gcc atg gtt ctc acg gac ccg gag cat cac tca tgaccgcaca	23002
Gln Ser Ala Met Val Leu Thr Asp Pro Glu His His Ser	
1465 1470	
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Met Ser Ala Arg Val	
1475	
acc atg gac gat ctc agg cga gcc ctc gag gag ggc tcc ggt gtc	24242
Thr Met Asp Asp Leu Arg Arg Ala Leu Glu Glu Gly Ser Gly Val	
1480 1485 1490	

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Asp Glu Gly Val Asp Leu Asp Thr Asp Leu Glu Thr Met Ala Phe	
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Ser Glu Leu Gly Tyr Asp Ser Leu Ala Val Leu Glu Thr Gly Leu	
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cgc ctc ggc cgc gag aac gac atc gag ctc gac gac tcg gtg ttc	24377
Arg Leu Gly Arg Glu Asn Asp Ile Glu Leu Asp Asp Ser Val Phe	
1525 1530 1535	
gcc gac ctc gac acg cct cag cag atg ctg gac gcg gtc aac gat	24422
Ala Asp Leu Asp Thr Pro Gln Gln Met Leu Asp Ala Val Asn Asp	
1540 1545 1550	
gcc ctc gcg cgt cag gcg gcg gca tcg tgacctctcc ccgtcatgcc	24469
Ala Leu Ala Arg Gln Ala Ala Ser	
1555 1560	
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Met Ala	
gat ccg gct cgc aca gac ctg cac tcc gcc acg atc acc ggc agc	25291
Asp Pro Ala Arg Thr Asp Leu His Ser Ala Thr Ile Thr Gly Ser	
1565 1570 1575	
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Ala Asp Ala Val Tyr Arg Arg Leu Glu Asp Val Gly Gln Trp Ser	
1580 1585 1590	
cag atg ttc gaa ccg acc atc cac ggc gcg gaa ctg gcc cgg gac	25381
Gln Met Phe Glu Pro Thr Ile His Gly Ala Glu Leu Ala Arg Asp	
1595 1600 1605	
ggg aac agg cag acg atc cag ctg tgg gcc acc gcc aac gga gaa	25426
Gly Asn Arg Gln Thr Ile Gln Leu Trp Ala Thr Ala Asn Gly Glu	
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ccc aag gcc tgg gtc tcc gag cgt gag ctc gac ccc gtc gcg cgc	25471
Pro Lys Ala Trp Val Ser Glu Arg Glu Leu Asp Pro Val Ala Arg	
1625 1630 1635	
acc atc cgc ttc gcg cag acc gtc acc tcc tcg ccc gtc gcc gag	25516
Thr Ile Arg Phe Ala Gln Thr Val Thr Ser Ser Pro Val Ala Glu	
1640 1645 1650	
atg tcc ggc gcg tgg cag gtg ctg ccc ctg tcc gag gac acc tgc	25561
Met Ser Gly Ala Trp Gln Val Leu Pro Leu Ser Glu Asp Thr Cys	
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cgg gtc gaa ctc acg cac acc tac cgt gcg gag aac gac tcg gcg	25606
Arg Val Glu Leu Thr His Thr Tyr Arg Ala Glu Asn Asp Ser Ala	
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Glu	Ser	Leu	Thr	Trp	Ile	Ala	Arg	Ala	Val	Glu	Thr	Asn	Ser	Thr	
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aag	gag	ctc	tcg	gcg	ctc	aag	ttc	gcc	tgc	gaa	cgg	gac	gcc	gac	25696
Lys	Glu	Leu	Ser	Ala	Leu	Lys	Phe	Ala	Cys	Glu	Arg	Asp	Ala	Asp	
1700					1705					1710					
agc	gag	gcc	agt	ccc	ttc	acc	ttc	acc	gat	gcg	gtg	gac	acc	acg	25741
Ser	Glu	Ala	Ser	Pro	Phe	Thr	Phe	Thr	Asp	Ala	Val	Asp	Thr	Thr	
1715					1720					1725					
gtc	gac	ccc	gtc	ctg	ctg	ttc	tcg	ttc	ctg	gac	cgc	ggt	gag	ctg	25786
Val	Asp	Pro	Val	Leu	Leu	Phe	Ser	Phe	Leu	Asp	Arg	Gly	Glu	Leu	
1730					1735					1740					
tgg	gcg	gga	cgc	ctg	gag	cac	gtc	gcc	gag	gcc	gag	atg	agg	gag	25831
Trp	Ala	Gly	Arg	Leu	Glu	His	Val	Ala	Glu	Ala	Glu	Met	Arg	Glu	
1745					1750					1755					
ttc	tcc	gac	ggc	ctg	cag	ttc	ctc	cgg	atg	cgg	acg	cgc	acc	ccg	25876
Phe	Ser	Asp	Gly	Leu	Gln	Phe	Leu	Arg	Met	Arg	Thr	Arg	Thr	Pro	
1760					1765					1770					
gac	ggt	gac	acg	cac	gtc	acc	gag	tcc	tac	cgg	gtg	tcg	cag	agc	25921
Asp	Gly	Asp	Thr	His	Val	Thr	Glu	Ser	Tyr	Arg	Val	Ser	Gln	Ser	
1775					1780					1785					
ccg	gcc	cgg	ctg	ctg	tac	aag	cag	gtg	acg	ctg	ccc	gcg	ctg	ctg	25966
Pro	Ala	Arg	Leu	Leu	Tyr	Lys	Gln	Val	Thr	Leu	Pro	Ala	Leu	Leu	
1790					1795					1800					
tcg	ctg	cac	acc	ggc	gag	tgg	acc	atc	acc	ccg	gcc	ggg	gag	agc	26011
Ser	Leu	His	Thr	Gly	Glu	Trp	Thr	Ile	Thr	Pro	Ala	Gly	Glu	Ser	
1805					1810					1815					
tgg	cgg	gtc	acg	tcg	aag	cac	acc	gtg	gcg	atc	gat	ccc	gac	gcg	26056
Trp	Arg	Val	Thr	Ser	Lys	His	Thr	Val	Ala	Ile	Asp	Pro	Asp	Ala	
1820					1825					1830					
gtg	cac	aag	gtc	ctc	ggt	gcc	gac	gcg	acg	gtc	tcg	gac	gcc	aag	26101
Val	His	Lys	Val	Leu	Gly	Ala	Asp	Ala	Thr	Val	Ser	Asp	Ala	Lys	
1835					1840					1845					
cgg	ctc	gcc	cgg	cgc	aac	ctg	ggc	aac	aac	agc	ctg	cgg	acc	ctc	26146
Arg	Leu	Ala	Arg	Arg	Asn	Leu	Gly	Asn	Asn	Ser	Leu	Arg	Thr	Leu	
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gaa	gca	gcg	gtc	cgg	tgg	gcc	ggc	acc	gcc	gtg	tcg	cag	agg		26188
Glu	Ala	Ala	Val	Arg	Trp	Ala	Gly	Thr	Ala	Val	Ser	Gln	Arg		
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tgagtgggga	c	atg	acg	gag	ccc	gag	acc	tcg	gac	ggt	ctc	gtc	gtc		26235
		Met	Thr	Glu	Pro	Glu	Thr	Ser	Asp	Val	Leu	Val	Val		
		1880						1885					1890		
ggc	gcc	ggg	ccc	agc	gga	ctg	ctc	ctg	gcc	ggg	atc	ctc	gcc	ggg	26280
Gly	Ala	Gly	Pro	Ser	Gly	Leu	Leu	Leu	Ala	Gly	Ile	Leu	Ala	Gly	
				1895					1900					1905	
gcg	ggt	gcg	cgg	gtc	acg	gtg	ctg	gag	gcg	cgg	gac	gcg	ccc	agc	26325
Ala	Gly	Ala	Arg	Val	Thr	Val	Leu	Glu	Ala	Arg	Asp	Ala	Pro	Ser	
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ccg	cag	acc	cgc	gcc	tcc	acc	ttg	cac	gcc	cgt	gcc	agg	gag	atc	26370
Pro	Gln	Thr	Arg	Ala	Ser	Thr	Leu	His	Ala	Arg	Ala	Arg	Glu	Ile	
				1925					1930					1935	
ctc	gac	cac	cac	gga	gtg	gag	ttc	tcc	ccg	gag	ctg	ccc	tgg	agt	26415
Leu	Asp	His	His	Gly	Val	Glu	Phe	Ser	Pro	Glu	Leu	Pro	Trp	Ser	
				1940					1945					1950	
gcc	cac	gga	cac	tac	ggc	ggc	ctg	cgc	gtg	gac	ctc	tcc	cgg	gtc	26460
Ala	His	Gly	His	Tyr	Gly	Gly	Leu	Arg	Val	Asp	Leu	Ser	Arg	Val	
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Asp Ser Gly Arg Ala Gly Val Trp Lys Cys Pro Gln Pro Glu Leu	1970 1975 1980
gta cgg acg ctg acc ggc tgg gcc cgc ggg cac ggc gcg cgg ctg	26550
Val Arg Thr Leu Thr Gly Trp Ala Arg Gly His Gly Ala Arg Leu	1985 1990 1995
ctc cac ggg gag cac gtg gag tcc gtc cgc gag cag ggc ggg cgc	26595
Leu His Gly Glu His Val Glu Ser Val Arg Glu Gln Gly Gly Arg	2000 2005 2010
tgt ctg gtg cgt acc cgg gcc ggc acc acg ttc agc ggg acc ctg	26640
Cys Leu Val Arg Thr Arg Ala Gly Thr Thr Phe Ser Gly Thr Leu	2015 2020 2025
ctg gtc gcg cgc gac ggc cgg cgg agc acg gtg cgg tcg ctg ctg	26685
Leu Val Ala Ala Asp Gly Arg Arg Ser Thr Val Arg Ser Leu Leu	2030 2035 2040
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Gly Ile Gly Cys Gly Gly Ala Pro Ala Thr Arg Val Leu Val Gln	2045 2050 2055
gcc gat gtc cac ggc gac ggg ctg gcg ggg cgg cgc ttc gag cga	26775
Ala Asp Val His Gly Asp Gly Leu Ala Gly Arg Arg Phe Glu Arg	2060 2065 2070
cac ggg cgg tac acc gtg acc gcc gca ccg atc agc ccc ggg atc	26820
His Gly Arg Tyr Thr Val Thr Ala Ala Pro Ile Ser Pro Gly Ile	2075 2080 2085
acc cgg gtg atg ctg cac gat ccg cgc tgg ccc gcg ggc gag gaa	26865
Thr Arg Val Met Leu His Asp Pro Arg Trp Pro Ala Gly Glu Glu	2090 2095 2100
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Arg Thr Leu Glu Asp Leu Arg Arg Ala Trp Lys Glu Ser Thr Gly	2105 2110 2115
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Glu Thr Leu Pro Ala Glu Pro Ser Trp Ser Arg Thr Phe Ser Asp	2120 2125 2130
gac acg aca gtg gca cac ccg ctg gtc aag ggc cgt gtc gtg ctg	27000
Asp Thr Thr Val Ala His Pro Leu Val Lys Gly Arg Val Val Leu	2135 2140 2145
tgc ggc gac gcc gcc cac ccc ttc gtc ccc atc ggc ggc cag gcg	27045
Cys Gly Asp Ala Ala His Pro Phe Val Pro Ile Gly Gly Gln Ala	2150 2155 2160
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Leu Asn Thr Ser Leu Met Asp Ala Glu Ala Leu Gly Trp Arg Val	2165 2170 2175
ctg ggg tat ctg gac gac ggg gac cgg caa ggc ctc ctc gac tac	27135
Leu Gly Tyr Leu Asp Asp Gly Asp Arg Gln Gly Leu Leu Asp Tyr	2180 2185 2190
cag gac gag cgg ttc tcg tgg ctg acc gtt ctc gcg ggg aga ctg	27180
Gln Asp Glu Arg Phe Ser Trp Leu Thr Val Leu Ala Gly Arg Leu	2195 2200 2205
cgc gcc cag gca cgt ctg ctg ttc gac acc gac gcg gcg gcc acg	27225
Arg Ala Gln Ala Arg Leu Leu Phe Asp Thr Asp Ala Ala Ala Thr	2210 2215 2220
gaa cgc aag gcg ctg gtc gcc gcg aga ctg gcc ggg gac gcg gac	27270
Glu Arg Lys Ala Leu Val Ala Ala Arg Leu Ala Gly Asp Ala Asp	2225 2230 2235
tac cgg cgc agg atc gcc gac gcc ctg gcc ggt gtc gac gtg tgc	27315
Tyr Arg Arg Arg Ile Ala Asp Ala Leu Ala Gly Val Asp Val Cys	2240 2245 2250
tac ctg acg ccc ggc ggc gcg gtc cgc cgg cgt ctg tcc ccg gcc	27360
Tyr Leu Thr Pro Gly Gly Ala Val Arg Arg Arg Leu Ser Pro Ala	2255 2260 2265

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Arg Leu Arg Glu Thr Gly Val Asn Pro Gly Ala Arg Arg Val Gln	
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Arg Ala Leu Val Pro Asp Asp Gly Thr Arg Thr Asp Ala Trp Ile	
2285 2290 2295	
cgt ccc gat cac cac tgg tac ccg gtg gcc cgc gac ggg gcc cgg	27495
Arg Pro Asp His His Trp Tyr Pro Val Ala Arg Asp Gly Ala Arg	
2300 2305 2310	
cag gac tgg gac gac gcg gtg cgc ctc cac gac gac ttg gaa ccc	27540
Gln Asp Trp Asp Asp Ala Val Arg Leu His Asp Asp Leu Glu Pro	
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Glu Val Thr Arg	
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Asp Tyr Ala Ala Gly Ala Val Trp Leu Glu Glu Leu Lys Gly Val	
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Asp Val Thr Leu Val Gly His Ser Val Gly Glu Leu Ala Ala Ala	
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Ala Glu Arg Ala Arg Leu Leu Asp Ala Ala Pro Arg Gly Gly Met	
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Ile Ala Cys Arg Ala Thr Glu Glu Ser Leu Arg Glu His Leu Asp	
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Ala Leu Gly Gly Arg Ala Val Ile Ala Ala Glu Asn Ala Asp Asn	
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Arg Tyr Leu Gly Ser His Gly Val Thr Cys Leu Arg Val Ala Ser	
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Thr Glu Pro Phe His Ser Pro Leu Leu Ala Pro Ala Ala Ala Arg	
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Glu Leu Pro Met Val Ser Ala Tyr Ser Ala Arg Arg Ile Ser Gly	
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Arg Glu Ile Met Pro Ala Ser Phe Trp Thr Arg Gln Met Ala Glu	
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Lys Val Arg Phe Trp Glu Ala Leu Arg His Asn Phe Asp Ser Gly	
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Pro Arg Thr Phe Val Glu Ile Gly Pro Gly Thr Val Leu Ser Leu	
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Ser Ala Ile His Glu Val Ala Glu Glu Phe Cys	
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Met Gly Phe Ile Arg Phe Asp Val Leu Gly Pro	
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ctc agg gtc cgg tgc gac gac acc ctg ctt caa ttg acc ggg cgc	30283
Leu Arg Val Arg Cys Asp Asp Thr Leu Leu Gln Leu Thr Gly Arg	
2680 2685 2690	
aag tac cgc acc gtg gtt tcg tat ctc gct ctt caa ccc gag tat	30328
Lys Tyr Arg Thr Val Val Ser Tyr Leu Ala Leu Gln Pro Glu Tyr	
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Ser Val Ala Ile Glu Asp Leu Val Arg Ala Ala Trp Ser Asp Lys	
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Arg Pro Ser Ser Ala His His Gln Val Arg Lys Met Val Ser Ala	
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Gln Asp Gly Tyr Met Leu Lys Leu Pro Pro Lys Gln Ser Asp Val	
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Ser Glu Phe Cys Arg Leu Phe Asp Gln Val Met Ser Gly Pro Leu	
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Thr Ser Asp Asp Asp Leu Ser Ala Ala Tyr Ser Ala Leu Ala Leu	
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Trp Arg Gly Arg Pro Cys Glu Gly Ser Glu Pro His Gly Gln Glu	
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Arg Arg Ile Ser Gln Leu Val Glu Gln His Arg Val Leu Leu Asn	
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Lys Thr Val Gln Gly Phe Gly Asp Arg Gly Arg Ser Asp Glu Leu	
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Ala Ser Ile Leu His Val Ala Ser Lys Ile His Gly Gln Pro Val	
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Thr Ala Arg Ser Gly Val Ala Val Pro Ala Pro Ala Val Ser Tyr	
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Pro Pro Arg Pro Gly Ser Pro Val Gly Pro Arg Cys Leu Pro Arg	
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tcg Ser 2980	aat Asn	gcg Ala	ctg Leu	gga Gly	ata Ile	ctc Leu	ctc Leu	aga Arg	cag Gln	aaa Lys	ggc Gly	ctg Leu	gcg Ala	gac Asp	31183
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gag Glu 2995	gac Asp	att Ile	tca Ser	cct Pro	tcg Ser	gaa Glu	gac Asp	ggc Gly	cgc Arg	ctc Leu	gca Ala	caa Gln	tg Trp	cgg Arg	31228
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tgc Cys 3025	gac Asp	atc Ile	gag Glu	caa Gln	gta Val	gaa Glu	ccc Pro	ctc Leu	atc Ile	ccg Pro	ccc Pro	tcg Ser	agc Ser	gaa Glu	31318
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agc Ser 3040	gcc Ala	tgc Cys	atc Ile	atc Ile	acg Thr	tcg Ser	cgc Arg	atc Ile	atc Ile	ctc Leu	aat Asn	ggc Gly	atc Ile	gac Asp	31363
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ctg Leu 3070	gag Glu	ata Ile	ctc Leu	agt Ser	tgc Cys	atg Met	atc Ile	ggc Gly	aga Arg	cgc Arg	ttc Phe	gac Asp	gac Asp	gag Glu	31453
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gag Glu 3085	acg Thr	aag Lys	gac Asp	gcc Ala	cgc Arg	gcg Ala	ctg Leu	atc Ile	cag Gln	cag Gln	tgc Cys	gcc Ala	aat Asn	ctg Leu	31498
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ccg Pro 3100	ctg Leu	gca Ala	ctc Leu	cgt Arg	ctc Leu	gcc Ala	gcc Ala	gcc Ala	cgg Arg	ata Ile	tcg Ser	acg Thr	cgc Arg	gac Asp	31543
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tcc Ser 3130	atc Ile	ttc Phe	agt Ser	gaa Glu	ctg Leu	gaa Glu	gtt Val	ccc Pro	ggc Gly	cgt Arg	agt Ser	ctg Leu	gtc Val	ggc Gly	31633
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cgg Arg 3145	ctc Leu	atg Met	acg Thr	tcc Ser	ttc Phe	acg Thr	tgc Cys	ctg Leu	gag Glu	gac Asp	ttc Phe	gat Asp	cac His	gac Asp	31678
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cgg Arg 3160	tac Tyr	ctc Leu	cga Arg	tta Leu	tcg Ser	ctg Leu	ctc Leu	ccc Pro	tgc Cys	ccc Pro	gag Glu	atc Ile	gat Asp	gaa Glu	31723
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acg Thr 3175	tcg Ser	gtc Val	gcg Ala	gcc Ala	gtg Val	ctg Leu	ggc Gly	gta Val	tcc Ser	acc Thr	gac Asp	tg Trp	gca Ala	cgg Arg	31768
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cgt Arg 3190	gcc Ala	tgc Cys	agg Arg	cgc Arg	ttc Phe	gca Ala	gac Asp	cgc Arg	gcg Ala	ttg Leu	ctg Leu	caa Gln	cgc Arg	aca Thr	31813
					3195					3200					
cga Arg 3205	tgc Cys	ggt Gly	acg Thr	tac Tyr	cgg Arg	atg Met	cac His	ccg Pro	ctg Leu	ctg Leu	ctg Leu	cag Gln	gcg Ala	gca Ala	31858
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3265 3270 3275
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3280 3285 3290
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3295 3300 3305
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3310 3315 3320
tgc gag gga gca ccc cac cgg agg cgt ccg cgg cag gca cgg gac 32218 Cys Glu Gly Ala Pro His Arg Arg Arg Pro Arg Gln Ala Arg Asp
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3395 3400 3405
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3425 3430 3435
ctg gcc gtc ttc cac acc ccc gcc gag cag gtg gag cga gaa cgc 32702 Leu Ala Val Phe His Thr Pro Ala Glu Gln Val Glu Arg Glu Arg
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3485 3490 3495
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Pro Thr Val Ser Gly Arg	Ile Thr Asn Leu Tyr	Gly Pro Gly Gln	
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Asn Leu Asp Lys Asn Gln	Gly Leu Val Ser Ala	Leu Val Lys Ala	
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cag ctg acc ggt gaa ccc	ctg cgg ctg cgg gcc	gcc ctg gag acc	33062
Gln Leu Thr Gly Glu Pro	Leu Arg Leu Arg Ala	Ala Leu Glu Thr	
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Thr Arg Asp Tyr Ile Tyr	Ala Arg Asp Cys Ala	Arg Met Val Val	
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tcg gcg atg gag acc gta	cgg tcc cgc acc cgc	ggc acg gac ccc	33152
Ser Ala Met Glu Thr Val	Arg Ser Arg Thr Arg	Gly Thr Asp Pro	
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cat gtc cgc aag ata ttc	agc agc gga cgc cgt	ctg cgg atc gac	33197
His Val Arg Lys Ile Phe	Ser Ser Gly Arg Arg	Leu Arg Ile Asp	
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Glu Leu Leu Arg Ile Ala	Glu Arg Leu Phe Asp	Arg Pro Val Pro	
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Val Val His Glu Pro Val	Ala Gly Gly Ala Asn	Val Asn Leu Ser	
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Val Glu Ser Arg Val Trp	Ala Asp Leu Glu Ser	Ser Pro Phe Leu	
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agc atc gag gaa ggg atg	cgc gcc gtc cgc tcc	gac ctc agg tac	33377
Ser Ile Glu Glu Gly Met	Arg Ala Val Arg Ser	Asp Leu Arg Tyr	
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cga ctc ggg cac ggg	tgagcgacac gaacgacaaa	agacccaggc cgcacatcag	33432
Arg Leu Gly His Gly			
3680			
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<210> SEQ ID NO 2

<211> LENGTH: 341

<212> TYPE: PRT

<213> ORGANISM: Streptomyces griseoflavus

<400> SEQUENCE: 2

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Arg Thr Val Arg Arg Asp Ile Asn Arg Leu Arg Glu Leu Gly Tyr Pro
 35 40 45

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Val Thr Ala Thr Lys Gly Pro Ser Gly Ser Tyr Arg Leu Ser Arg Gly
 50 55 60
 Ala Arg Leu Pro Pro Leu Ile Val Asp Asp Glu Gln Ala Leu Ala Ile
 65 70 75 80
 Ala Leu Ser Leu Gln Thr Ala Pro Ala Ser Val Thr Gly Met Gly Asp
 85 90 95
 Ala Thr Lys Arg Ala Leu Asn Ser Ile Gln Glu Leu Leu Pro Pro His
 100 105 110
 Leu Ala His Arg Leu Ala Thr Phe Ser Val Glu Gln Ile Glu Asn Ala
 115 120 125
 Trp Glu Leu Ala Pro Pro Gln Val Asp Pro Ser Leu Leu Ala Gln Leu
 130 135 140
 Ser Ser Ala Ala Gln Gln Arg Asp Leu Val Arg Phe Ser Tyr Arg Ser
 145 150 155 160
 Ile His His Asp Ser Met Gln Asp Gly Glu Val Leu Ala Glu Pro His
 165 170 175
 Arg Leu Val Val Trp Ser Gly Arg Trp Tyr Leu Val Ala Tyr Asp Gln
 180 185 190
 Gln Arg Ser Ser Trp His Ala Tyr Arg Val Asp Arg Ile Lys Asp Leu
 195 200 205
 Ala Pro Thr Ala Trp Arg Phe Gly Glu Arg Glu Gly Pro Asp Glu Asp
 210 215 220
 Ile Thr Arg Phe Val Gln Asn Gln Pro Asp Arg Gly Asp Thr Pro Asp
 225 230 235 240
 Thr Trp Pro Cys Trp Gly Thr Val Leu Met Glu Cys Pro Ala Ser Leu
 245 250 255
 Val Ala Lys Trp Ala Pro Gly Val Ala Ser Phe Glu Ala Val Asp Asp
 260 265 270
 Arg Val Thr Arg Ile Gln Met Gly Ala Trp Ser Trp Ser Ala Leu Ile
 275 280 285
 Gly Phe Leu Ile Thr Phe Ser Cys Arg Phe Thr Val Glu Gly Pro Pro
 290 295 300
 Glu Leu Val Ala Ala Ala Arg Arg Val Met Gly Leu Ile Asp Val Gly
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 325 330 335
 Pro Ser Ala Gly Arg
 340

<210> SEQ ID NO 3
 <211> LENGTH: 213
 <212> TYPE: PRT
 <213> ORGANISM: Streptomyces griseoflavus

<400> SEQUENCE: 3

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 Ala Ala Val Met Gly Glu Ala Glu Phe Glu Leu Val Asp Leu Ala Glu
 35 40 45
 Tyr Gly Leu Pro Leu Leu Asp Glu Pro Val Pro Ala Met Phe Gly Gln
 50 55 60
 Tyr Gln Lys Glu Glu Thr Arg Arg Trp Ala Ala Ala Ile Gly Ser Phe
 65 70 75 80

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Asp Gly Phe Val Phe Val Thr Pro Glu Tyr Asn His Ser Val Pro Ala
 85 90 95
 Ala Leu Lys Asn Ala Ile Asp His Leu Phe Ala Glu Trp Thr Asp Lys
 100 105 110
 Ala Ala Gly Phe Val Ser Tyr Gly Val His Gly Gly Thr Arg Ala Val
 115 120 125
 Glu His Leu Arg Leu Ala Leu Ala Glu Val Lys Val Ala Gly Val Arg
 130 135 140
 Ser Gln Val Val Leu Ser Val Phe Asn Asp Phe Asp Tyr Thr Gly Cys
 145 150 155 160
 Asp Met Thr Asp Pro Thr Ala Met Gly Arg Phe Thr Pro Gly Pro Gln
 165 170 175
 Gln Glu Gln Thr Val Asn Thr Met Leu Asp Glu Val Val Ala Trp Ser
 180 185 190
 Thr Ala Leu Lys Pro Leu Arg Thr Ala Ala Thr Ala Glu Ala Asp Gly
 195 200 205
 Arg Ala Val Ser Val
 210

<210> SEQ ID NO 4
 <211> LENGTH: 500
 <212> TYPE: PRT
 <213> ORGANISM: Streptomyces griseoflavus

<400> SEQUENCE: 4

Met Thr Leu His Ala Ala Glu Ala Ile Pro Ser His Val Pro Val Leu
 1 5 10 15
 Val Val Gly Ala Gly Pro Thr Gly Leu Met Leu Gly Ala Glu Leu Ala
 20 25 30
 Leu His Gly Ser Arg Pro Leu Val Ile Asp Ala Leu Pro Ser Pro Ser
 35 40 45
 Gly Gln Ser Arg Ala Leu Gly Phe Thr Val Arg Thr Leu Glu Ile Phe
 50 55 60
 Lys Gln Arg Gly Ile Leu Gly Arg Phe Gln Gly Leu Ala Pro Val Pro
 65 70 75 80
 Gly Val His Phe Ala Gly Leu Ser Ile Lys Gly Asp His Leu Ser Ser
 85 90 95
 Ser Met Arg Pro Ala Asn Gln Tyr Pro Gln Ser Lys Thr Glu Gln Val
 100 105 110
 Leu Ala Ala Trp Ala Glu Glu Leu Gly Val Pro Val Arg Arg Pro Trp
 115 120 125
 Thr Leu Thr Ser Met Glu Pro Thr Gly Thr Gly Tyr Arg Cys Val Leu
 130 135 140
 Ser Gly Pro Ala Gly Gln Gln Thr Val Asp Ala Asp Tyr Val Val Gly
 145 150 155 160
 Cys Asp Gly Ala Gly Ser Phe Val Arg Glu Ala Ile Gly Met Pro Thr
 165 170 175
 Lys Arg Thr Pro Pro Ser Val Gln Met Leu Leu Gly Asp Leu Arg Gly
 180 185 190
 Cys Gly Leu Pro Asp Glu Pro Phe Gly Val Lys His Glu Lys Gly Met
 195 200 205
 Val Met Ser Ala Pro Leu Gly Asp Gly Thr Glu Arg Val Ile Val Cys
 210 215 220
 Asp Phe Thr Gln Pro Met Arg Pro Gln Gly Thr Pro Val Thr His Asp
 225 230 235 240

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Phe Ala Asp Leu Asp Thr Pro Gln Gln Met Leu Asp Ala Val Asn Asp
65 70 75 80

Ala Leu Ala Arg Gln Ala Ala Ala Ser
85

<210> SEQ ID NO 7
<211> LENGTH: 316
<212> TYPE: PRT
<213> ORGANISM: Streptomyces griseoflavus

<400> SEQUENCE: 7

Met Ala Asp Pro Ala Arg Thr Asp Leu His Ser Ala Thr Ile Thr Gly
1 5 10 15

Ser Ala Asp Ala Val Tyr Arg Arg Leu Glu Asp Val Gly Gln Trp Ser
20 25 30

Gln Met Phe Glu Pro Thr Ile His Gly Ala Glu Leu Ala Arg Asp Gly
35 40 45

Asn Arg Gln Thr Ile Gln Leu Trp Ala Thr Ala Asn Gly Glu Pro Lys
50 55 60

Ala Trp Val Ser Glu Arg Glu Leu Asp Pro Val Ala Arg Thr Ile Arg
65 70 75 80

Phe Ala Gln Thr Val Thr Ser Ser Pro Val Ala Glu Met Ser Gly Ala
85 90 95

Trp Gln Val Leu Pro Leu Ser Glu Asp Thr Cys Arg Val Glu Leu Thr
100 105 110

His Thr Tyr Arg Ala Glu Asn Asp Ser Ala Glu Ser Leu Thr Trp Ile
115 120 125

Ala Arg Ala Val Glu Thr Asn Ser Thr Lys Glu Leu Ser Ala Leu Lys
130 135 140

Phe Ala Cys Glu Arg Asp Ala Asp Ser Glu Ala Ser Pro Phe Thr Phe
145 150 155 160

Thr Asp Ala Val Asp Thr Thr Val Asp Pro Val Leu Leu Phe Ser Phe
165 170 175

Leu Asp Arg Gly Glu Leu Trp Ala Gly Arg Leu Glu His Val Ala Glu
180 185 190

Ala Glu Met Arg Glu Phe Ser Asp Gly Leu Gln Phe Leu Arg Met Arg
195 200 205

Thr Arg Thr Pro Asp Gly Asp Thr His Val Thr Glu Ser Tyr Arg Val
210 215 220

Ser Gln Ser Pro Ala Arg Leu Leu Tyr Lys Gln Val Thr Leu Pro Ala
225 230 235 240

Leu Leu Ser Leu His Thr Gly Glu Trp Thr Ile Thr Pro Ala Gly Glu
245 250 255

Ser Trp Arg Val Thr Ser Lys His Thr Val Ala Ile Asp Pro Asp Ala
260 265 270

Val His Lys Val Leu Gly Ala Asp Ala Thr Val Ser Asp Ala Lys Arg
275 280 285

Leu Ala Arg Arg Asn Leu Gly Asn Asn Ser Leu Arg Thr Leu Glu Ala
290 295 300

Ala Val Arg Trp Ala Gly Thr Ala Val Ser Gln Arg
305 310 315

<210> SEQ ID NO 8
<211> LENGTH: 451
<212> TYPE: PRT
<213> ORGANISM: Streptomyces griseoflavus

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<400> SEQUENCE: 8

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Met Thr Glu Pro Glu Thr Ser Asp Val Leu Val Val Gly Ala Gly Pro
1          5          10          15
Ser Gly Leu Leu Leu Ala Gly Ile Leu Ala Gly Ala Gly Ala Arg Val
20          25          30
Thr Val Leu Glu Ala Arg Asp Ala Pro Ser Pro Gln Thr Arg Ala Ser
35          40          45
Thr Leu His Ala Arg Ala Arg Glu Ile Leu Asp His His Gly Val Glu
50          55          60
Phe Ser Pro Glu Leu Pro Trp Ser Ala His Gly His Tyr Gly Gly Leu
65          70          75          80
Arg Val Asp Leu Ser Arg Val Asp Ser Gly Arg Ala Gly Val Trp Lys
85          90          95
Cys Pro Gln Pro Glu Leu Val Arg Thr Leu Thr Gly Trp Ala Arg Gly
100         105         110
His Gly Ala Arg Leu Leu His Gly Glu His Val Glu Ser Val Arg Glu
115         120         125
Gln Gly Gly Arg Cys Leu Val Arg Thr Arg Ala Gly Thr Thr Phe Ser
130         135         140
Gly Thr Leu Leu Val Ala Ala Asp Gly Arg Arg Ser Thr Val Arg Ser
145         150         155         160
Leu Leu Gly Ile Gly Cys Gly Gly Ala Pro Ala Thr Arg Val Leu Val
165         170         175
Gln Ala Asp Val His Gly Asp Gly Leu Ala Gly Arg Arg Phe Glu Arg
180         185         190
His Gly Arg Tyr Thr Val Thr Ala Ala Pro Ile Ser Pro Gly Ile Thr
195         200         205
Arg Val Met Leu His Asp Pro Arg Trp Pro Ala Gly Glu Glu Arg Thr
210         215         220
Leu Glu Asp Leu Arg Arg Ala Trp Lys Glu Ser Thr Gly Glu Thr Leu
225         230         235         240
Pro Ala Glu Pro Ser Trp Ser Arg Thr Phe Ser Asp Asp Thr Thr Val
245         250         255
Ala His Pro Leu Val Lys Gly Arg Val Val Leu Cys Gly Asp Ala Ala
260         265         270
His Pro Phe Val Pro Ile Gly Gly Gln Ala Leu Asn Thr Ser Leu Met
275         280         285
Asp Ala Glu Ala Leu Gly Trp Arg Val Leu Gly Tyr Leu Asp Asp Gly
290         295         300
Asp Arg Gln Gly Leu Leu Asp Tyr Gln Asp Glu Arg Phe Ser Trp Leu
305         310         315         320
Thr Val Leu Ala Gly Arg Leu Arg Ala Gln Ala Arg Leu Leu Phe Asp
325         330         335
Thr Asp Ala Ala Ala Thr Glu Arg Lys Ala Leu Val Ala Ala Arg Leu
340         345         350
Ala Gly Asp Ala Asp Tyr Arg Arg Arg Ile Ala Asp Ala Leu Ala Gly
355         360         365
Val Asp Val Cys Tyr Leu Thr Pro Gly Gly Ala Val Arg Arg Arg Leu
370         375         380
Ser Pro Ala Arg Leu Arg Glu Thr Gly Val Asn Pro Gly Ala Arg Arg
385         390         395         400
Val Gln Arg Ala Leu Val Pro Asp Asp Gly Thr Arg Thr Asp Ala Trp
405         410         415

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Ile Arg Pro Asp His His Trp Tyr Pro Val Ala Arg Asp Gly Ala Arg
    420                                425                430

Gln Asp Trp Asp Asp Ala Val Arg Leu His Asp Asp Leu Glu Pro Glu
    435                                440                445

Val Thr Arg
    450

<210> SEQ ID NO 9
<211> LENGTH: 339
<212> TYPE: PRT
<213> ORGANISM: Streptomyces griseoflavus

<400> SEQUENCE: 9

Val Pro His Gln Ala Thr Gly Ala Ala Pro Asp Gly Gly Gly Ser Ala
1      5      10      15

Pro Arg Ser Leu Val Leu Met Leu Pro Gly Gln Gly Ser Gln Phe Ala
20     25     30

Ala Met Gly Val Pro Leu Tyr Glu Ser Asp Ala Arg Phe Arg Lys Ala
35     40     45

Leu Asp Asp Phe Phe Asp Ala Phe Gly Thr Gly Ala Glu Arg Leu Arg
50     55

Arg Glu Trp Leu His Gly Ser Ala Gln Gly Ile Glu Arg Gly Ser Phe
65     70     75     80

Ala Gln Pro Met Leu Phe Gly Leu Asp Tyr Ala Ala Gly Ala Val Trp
85     90     95

Leu Glu Glu Leu Lys Gly Val Asp Val Thr Leu Val Gly His Ser Val
100    105    110

Gly Glu Leu Ala Ala Ala Thr Leu Ala Gly Ala Phe Asp Leu Glu Leu
115    120    125

Ala Gly Ala Leu Leu Ala Glu Arg Ala Arg Leu Leu Asp Ala Ala Pro
130    135    140

Arg Gly Gly Met Ile Ala Cys Arg Ala Thr Glu Glu Ser Leu Arg Glu
145    150    155    160

His Leu Asp Ala Leu Gly Gly Arg Ala Val Ile Ala Ala Glu Asn Ala
165    170    175

Asp Asn Gln Cys Val Val Ser Cys Ala Glu Glu Asp Leu Pro Asp Thr
180    185    190

Met Arg Tyr Leu Gly Ser His Gly Val Thr Cys Leu Arg Val Ala Ser
195    200    205

Thr Glu Pro Phe His Ser Pro Leu Leu Ala Pro Ala Ala Ala Arg Phe
210    215    220

Glu Glu Phe Leu Ala Arg Arg Gly His Arg Leu Ser Thr Thr Glu Leu
225    230    235    240

Pro Met Val Ser Ala Tyr Ser Ala Arg Arg Ile Ser Gly Arg Glu Ile
245    250    255

Met Pro Ala Ser Phe Trp Thr Arg Gln Met Ala Glu Lys Val Arg Phe
260    265    270

Trp Glu Ala Leu Arg His Asn Phe Asp Ser Gly Pro Arg Thr Phe Val
275    280    285

Glu Ile Gly Pro Gly Thr Val Leu Ser Leu Ala Ala Arg Arg Leu Pro
290    295    300

Ser Val Arg Ala Arg Arg Ser Thr Val Ile Ser Thr Met Pro Arg His
305    310    315    320

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Arg Pro His Pro Glu His Trp Glu Ser Ala Ile His Glu Val Ala Glu
 325 330 335

Glu Phe Cys

<210> SEQ ID NO 10

<211> LENGTH: 678

<212> TYPE: PRT

<213> ORGANISM: Streptomyces griseoflavus

<400> SEQUENCE: 10

Met Gly Phe Ile Arg Phe Asp Val Leu Gly Pro Leu Arg Val Arg Cys
 1 5 10 15

Asp Asp Thr Leu Leu Gln Leu Thr Gly Arg Lys Tyr Arg Thr Val Val
 20 25 30

Ser Tyr Leu Ala Leu Gln Pro Glu Tyr Ser Val Ala Ile Glu Asp Leu
 35 40 45

Val Arg Ala Ala Trp Ser Asp Lys Arg Pro Ser Ser Ala His His Gln
 50 55 60

Val Arg Lys Met Val Ser Ala Leu Arg Thr Ser Leu Asp Gln Asp Trp
 65 70 75 80

Asp Leu Val Ala Thr Ser Gln Asp Gly Tyr Met Leu Lys Leu Pro Pro
 85 90 95

Lys Gln Ser Asp Val Ser Glu Phe Cys Arg Leu Phe Asp Gln Val Met
 100 105 110

Ser Gly Pro Leu Thr Ser Asp Asp Asp Leu Ser Ala Ala Tyr Ser Ala
 115 120 125

Leu Ala Leu Trp Arg Gly Arg Pro Cys Glu Gly Ser Glu Pro His Gly
 130 135 140

Gln Glu Arg Arg Ile Ser Gln Leu Val Glu Gln His Arg Val Leu Leu
 145 150 155 160

Asn Lys Thr Val Gln Gly Phe Gly Asp Arg Gly Arg Ser Asp Glu Leu
 165 170 175

Ala Ser Ile Leu His Val Ala Ser Lys Ile His Gly Gln Pro Val Thr
 180 185 190

Ala Arg Ser Gly Val Ala Val Pro Ala Pro Ala Val Ser Tyr Ala Gly
 195 200 205

Thr Thr Gln Val Pro Glu Pro Ser Gly Ser Thr Thr Pro Pro Pro Arg
 210 215 220

Pro Gly Ser Pro Val Gly Pro Arg Cys Leu Pro Arg Asp Leu Gln Asp
 225 230 235 240

Phe Gly Gly Arg Glu Arg Glu Ile Asn Glu Leu Gln Lys Leu Leu Thr
 245 250 255

Ala Glu Gly Pro His Pro Gln Leu Val Ala Thr Val His Gly Met Ser
 260 265 270

Gly Val Gly Lys Thr Ala Val Ala Val Arg Leu Ala His Arg Leu Ala
 275 280 285

His His Tyr Pro Asp Gly Gln Leu Phe Val Ser Leu Asp Gly Phe Ser
 290 295 300

Ser Ala Ser Thr Ala Thr Val Ser Asn Ala Leu Gly Ile Leu Leu Arg
 305 310 315 320

Gln Lys Gly Leu Ala Asp Glu Asp Ile Ser Pro Ser Glu Asp Gly Arg
 325 330 335

Leu Ala Gln Trp Arg Thr Ile Thr Ala Gly Gln Lys Leu Leu Val Val
 340 345 350

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Leu Asp Asp Val Cys Asp Ile Glu Gln Val Glu Pro Leu Ile Pro Pro
 355 360 365
 Ser Ser Glu Ser Ala Cys Ile Ile Thr Ser Arg Ile Ile Leu Asn Gly
 370 375 380
 Ile Asp Gly Ala His His Ile Ser Leu Glu Val Pro Asp Glu Asp Glu
 385 390 395 400
 Cys Leu Glu Ile Leu Ser Cys Met Ile Gly Arg Arg Phe Asp Asp Glu
 405 410 415
 Glu Thr Lys Asp Ala Arg Ala Leu Ile Gln Gln Cys Ala Asn Leu Pro
 420 425 430
 Leu Ala Leu Arg Leu Ala Ala Ala Arg Ile Ser Thr Arg Asp Phe Leu
 435 440 445
 Asn Leu Arg Glu Leu Ser Glu Gln Leu Ser Ser Ser Ala Ser Ile Phe
 450 455 460
 Ser Glu Leu Glu Val Pro Gly Arg Ser Leu Val Gly Arg Leu Met Thr
 465 470 475 480
 Ser Phe Thr Cys Leu Glu Asp Phe Asp His Asp Arg Tyr Leu Arg Leu
 485 490 495
 Ser Leu Leu Pro Cys Pro Glu Ile Asp Glu Thr Ser Val Ala Ala Val
 500 505 510
 Leu Gly Val Ser Thr Asp Trp Ala Arg Arg Ala Cys Arg Arg Phe Ala
 515 520 525
 Asp Arg Ala Leu Leu Gln Arg Thr Arg Cys Gly Thr Tyr Arg Met His
 530 535 540
 Pro Leu Leu Leu Gln Ala Ala Gln Leu Glu Ala Gln Lys Thr Ile Pro
 545 550 555 560
 Phe Glu Glu Gln Arg Arg Leu Val Arg Ala Ala Phe Leu His Tyr Lys
 565 570 575
 Ala Ser Asn Gly Leu Val Gly Ala Ser Arg Ile Ser Pro Ser Arg Val
 580 585 590
 Pro Asp Gly His Val Val Leu Arg Thr Leu Thr Gln Ser Ala Lys Leu
 595 600 605
 Ala Ala Arg Leu Gly Leu Gln Glu Glu Leu Ala Asp Leu Tyr Thr Ala
 610 615 620
 Trp Lys Glu Leu Leu Pro Leu Val Leu Asp Arg Arg Gln Gln Glu Ala
 625 630 635 640
 Val Gly Arg Arg Val Leu Ala Val Ser Gln His Leu Asp Arg Pro Ala
 645 650 655
 Cys Glu Gly Ala Pro His Arg Arg Arg Pro Arg Gln Ala Arg Asp Met
 660 665 670
 Leu Pro Glu Gly Gln Arg
 675

<210> SEQ ID NO 11
 <211> LENGTH: 338
 <212> TYPE: PRT
 <213> ORGANISM: Streptomyces griseoflavus

<400> SEQUENCE: 11

Met Asp Arg Val Leu Pro Tyr Ala Ala Gly Ser Glu Ala Leu Leu Ser
 1 5 10 15
 Ser Arg Glu His Gly Pro Thr Val Ser Glu Arg Thr Val Ser Ala Gln
 20 25 30
 Glu Ile Val Val Gly Gly Gly Gly Leu Leu Gly Arg His Ile Leu Gly
 35 40 45

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Val Leu Gly Asn Arg Leu Ser Arg Arg Val Arg Ile Pro Trp Asp Asp
 50 55 60
 His Gly Arg Ala Cys Glu Gln Leu Tyr Ala Leu Gly Arg Asp Leu Ala
 65 70 75 80
 Gln Gln Pro Ala Arg Trp Asn Leu Tyr Trp Cys Ala Gly Leu Ala Val
 85 90 95
 Phe His Thr Pro Ala Glu Gln Val Glu Arg Glu Arg Leu Gln Val Ser
 100 105 110
 Leu Leu Leu Ala Gly Ile Asn Asp Gly Leu Glu Arg Ser Gly Gly Pro
 115 120 125
 Thr Gly Gly Ala Leu Phe Leu Ala Ser Ser Ala Gly Gly Ala Phe Ala
 130 135 140
 Gly Ser Glu His Pro Pro Phe Thr Glu Phe Ser Pro Pro Val Pro Thr
 145 150 155 160
 Asn Pro Tyr Gly Ala Ser Lys Leu Ala Val Glu Glu Glu Ala Glu Val
 165 170 175
 Leu Ala Arg Arg Trp Arg Leu Pro Thr Val Ser Gly Arg Ile Thr Asn
 180 185 190
 Leu Tyr Gly Pro Gly Gln Asn Leu Asp Lys Asn Gln Gly Leu Val Ser
 195 200 205
 Ala Leu Val Lys Ala Gln Leu Thr Gly Glu Pro Leu Arg Leu Arg Ala
 210 215 220
 Ala Leu Glu Thr Thr Arg Asp Tyr Ile Tyr Ala Arg Asp Cys Ala Arg
 225 230 235 240
 Met Val Val Ser Ala Met Glu Thr Val Arg Ser Arg Thr Arg Gly Thr
 245 250 255
 Asp Pro His Val Arg Lys Ile Phe Ser Ser Gly Arg Arg Leu Arg Ile
 260 265 270
 Asp Glu Leu Leu Arg Ile Ala Glu Arg Leu Phe Asp Arg Pro Val Pro
 275 280 285
 Val Val His Glu Pro Val Ala Gly Gly Ala Asn Val Asn Leu Ser Val
 290 295 300
 Glu Ser Arg Val Trp Ala Asp Leu Glu Ser Ser Pro Phe Leu Ser Ile
 305 310 315 320
 Glu Glu Gly Met Arg Ala Val Arg Ser Asp Leu Arg Tyr Arg Leu Gly
 325 330 335

His Gly

<210> SEQ ID NO 12
 <211> LENGTH: 33825
 <212> TYPE: DNA
 <213> ORGANISM: Streptomyces griseoflavus
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (21413)..(21736)
 <223> OTHER INFORMATION: ORF16 shown on this duplicate of SEQ ID NO:1
 due to sequence overlap with ORF17
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (22992)..(24164)
 <223> OTHER INFORMATION: ORF18 shown on this duplicate of SEQ ID NO:1
 due to sequence overlap with ORF17
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (24449)..(25225)
 <223> OTHER INFORMATION: ORF20 shown on this duplicate of SEQ ID NO:1
 due to sequence overlap with ORF19

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<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (27552)..(28481)
<223> OTHER INFORMATION: ORF23 shown on this duplicate of SEQ ID NO:1
      due to sequence overlap with ORF22

<400> SEQUENCE: 12

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gtcgccgacg ccgaccagat ggaaccgagg gtgctggcgg ccatctcccg cgaccgccggc    120
ctgatggagg tggccggccg ggaggggcac ctctaccagt cggtgtccga ccgccccttc    180
tccggcgacc gcgccagge caagctcgcc gtcctcggcg ccgtctacgg ccagacctcc    240
ggcgacggcc tgaagaacct cgccgcgctc aggcgcgcct tccccaaagg ggtggcctac    300
gtcgacgagg ccgcccgccg cggcgaggag ggccgtctcg tacggacctg gctgggcccgc    360
acctgccccg ccgccgtccg cccgacggac gacgcggcgg aggaggccgg catcccggcc    420
gcccaggagg agccggggcc agcggcccga ccgtggggcc cggaggccga ggcccggccg    480
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cgggccggcg aggtcttggg gtgctggagc agcaggtcgg gcgcgtagag gatgcccac	4620
cccgcgtcgg cggcacgcca ggcgaggtcg gtctcctcgt gcgcgaagaa gaacgcgccg	4680
ggccagtcgc cgatctcgtc gagcatcgac atccgcagcg cgtgcccgcc cccgaggaac	4740
ccggtgacgt accccgccct catcgggtcc gcctttccga gccggggcac gtgccgctgc	4800
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<210> SEQ ID NO 13
<211> LENGTH: 108
<212> TYPE: PRT
<213> ORGANISM: Streptomyces griseoflavus

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<400> SEQUENCE: 13

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Leu Phe Ser Ser Leu Ile Val Ala Arg Met Asp Thr Gly His Ala Glu
 1             5             10            15
Ala Val Ala Asp Val Phe Ala Gly Phe Asp Ala Thr Asp Met Pro Ala
          20             25             30
Arg Met Gly Thr Arg Arg Arg Glu Leu Phe Arg Tyr Arg Gly Leu Tyr
          35             40             45
Phe His Leu Gln Asp Phe Glu Thr Pro Asp Gly Thr Glu Ala Val Glu
          50             55             60
Ala Ala Lys Ser Asp Pro Arg Phe Ile Arg Val Ser Asn Asp Leu Arg
          65             70             75             80
Pro Tyr Ile Glu Ala Tyr Ala Pro Asp Trp Gln Ser Pro Lys Asp Ala
          85             90             95
Met Ala Glu Arg Phe Tyr His Trp Ser Ser Lys Arg
          100            105

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<210> SEQ ID NO 14
<211> LENGTH: 391
<212> TYPE: PRT
<213> ORGANISM: Streptomyces griseoflavus

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<400> SEQUENCE: 14

Met Thr Ala His Ile Thr Gly Ile Asp Ile Val Ser Pro Leu Gly Leu
 1 5 10 15
 Ser Arg Glu Glu His Trp Lys Ala Leu Leu Asp Gly Cys Ser Gly Leu
 20 25 30
 Arg Ala Thr Gln Ser Phe Asp Ser Ser Arg Tyr Asp Asn Pro Ile Ser
 35 40 45
 Gly Glu Val Pro His Phe Ala Pro Glu Gly Leu Pro Lys Arg Leu Leu
 50 55 60
 Pro Ala Thr Asp Arg Met Thr Gln Met Ser Leu Val Ala Ala Ala Gly
 65 70 75 80
 Ala Phe Asp Asp Ser Gly Val Asp Thr Ser Arg Val Asp Pro Leu Gly
 85 90 95
 Val Gly Val Met Thr Ala Ser Thr Ala Gly Gly Tyr Ala Phe Gly Gln
 100 105 110
 Lys Glu Leu Gln Asn Leu Trp Ser Lys Gly Pro Arg Tyr Val Ser Thr
 115 120 125
 His Gln Ser Tyr Ala Trp Phe Tyr Ala Val Asn Thr Gly Gln Ile Ser
 130 135 140
 Ile Arg His Gly Cys Gln Gly His Ser Gly Val Ile Val Ala Asp Asp
 145 150 155 160
 Ala Gly Gly Leu Asp Ala Ile Ser Phe Ala Ala Arg Arg Leu Ala Arg
 165 170 175
 Gly Asn Arg Val Met Leu Thr Gly Ser Val Asp Ser Thr Met Cys Pro
 180 185 190
 Trp Gly Arg Val Ala His Thr Ser Thr Gly Met Leu Ser Ala Ser Thr
 195 200 205
 Asp Ala Arg Ala Ala Tyr Leu Pro Phe Asp Ala Arg Ala Asn Gly Trp
 210 215 220
 Val Asn Gly Glu Gly Gly Ala His Leu Val Leu Gln Thr His Ser Asp
 225 230 235 240
 Gly Arg Tyr Ala Ala Val Leu Gly His Gly Ala Thr Met Asp Asp Pro
 245 250 255
 Arg Ala Ala Pro Gly Thr Gly Leu Val Arg Ala Ile His Leu Ala Leu
 260 265 270
 Gly Ala Ala Arg Leu Arg Pro Gly Asp Ile Ser Val Val Phe Ala Asp
 275 280 285
 Ala Ala Gly Thr Arg Glu Ala Asp Thr Ala Glu Ala Ala Ala Leu Ala
 290 295 300
 Glu Val Phe Gly Pro Asp Ser Val Pro Val Thr Ala Pro Lys Ala Ala
 305 310 315 320
 Thr Gly Arg Met Gly Cys Gly Thr Ala Ala Leu Asp Val Ala Thr Ala
 325 330 335
 Val Leu Ala Leu Arg Asp Gln Thr Ile Pro Pro Thr Val Asn Val Gln
 340 345 350
 Ala Asp Ala Ser Leu Gly Val Asn Leu Cys Ser Val Ala Thr His His
 355 360 365
 Pro Leu Thr Asn Val Leu Val Leu Ala Arg Gly Val Gly Gly Phe Asn
 370 375 380
 Ser Ala Leu Ile Val Gly Lys
 385 390

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<210> SEQ ID NO 15
<211> LENGTH: 259
<212> TYPE: PRT
<213> ORGANISM: Streptomyces griseoflavus

<400> SEQUENCE: 15
Val Thr Ser Pro Arg His Ala Leu Val Thr Gly Gly Ser Ser Gly Ile
1      5      10      15
Gly Lys Ser Val Ala Arg Arg Leu Ala Ser Ala Gly His Thr Val Thr
20     25     30
Ile Cys Gly Arg Asp Ser Glu Arg Leu Gln Gln Ala Ala Lys Glu Leu
35     40     45
Ser Glu Gln Gly Ala Pro Val Thr Ser Leu Ile Ala Asp Val Ser Lys
50     55     60
Pro Arg Gln Val Gly Asp Leu Val Arg Glu Ala Val Glu Thr Asn Gly
65     70     75     80
Pro Leu Gly Ile Leu Val Asn Asn Ala Gly Arg Asn Gly Gly Gly Arg
85     90     95
Thr Ala Glu Leu Ser Asp Glu Leu Trp Arg Glu Val Leu Ser Thr Asn
100    105   110
Leu Asp Ser Val Phe Tyr Val Thr Arg Glu Val Leu Ala Arg Gly Gly
115    120   125
Ile Gly Glu Val Asp His Ala Arg Ile Ile Asn Ile Ala Ser Thr Ala
130    135   140
Gly Lys Gln Gly Val Leu Leu Ala Ala Pro Tyr Ser Ala Ser Lys His
145    150   155   160
Gly Val Val Gly Phe Thr Lys Ala Val Gly Lys Glu Leu Ala Pro Gln
165    170   175
Gly Ile Thr Val Asn Ala Val Cys Pro Gly Tyr Val Glu Thr Pro Met
180    185   190
Ala Ser Arg Val Arg Gln Ala Tyr Ala Asp Ala Trp Glu Thr Thr Glu
195    200   205
Ala Glu Val Leu Ser Ala Phe Glu Ala Lys Ile Pro Leu Gly Arg Tyr
210    215   220
Ser Thr Pro Asp Glu Val Ala Ser Leu Val Glu Tyr Leu Thr Thr Glu
225    230   235   240
Gly Ala Ala Ser Ile Thr Ala Gln Ala Phe Asn Val Cys Gly Gly Leu
245    250   255

Gly Asn Phe

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<210> SEQ ID NO 16
<211> LENGTH: 310
<212> TYPE: PRT
<213> ORGANISM: Streptomyces griseoflavus

<400> SEQUENCE: 16
Val Arg Ala Phe Leu Phe Pro Gly Gln Gly Thr Gln Lys Ile Gly Met
1      5      10      15
Gly Thr Tyr Leu Arg Glu Arg Tyr Pro His Leu Ile Ala Pro Leu Trp
20     25     30
Arg Glu Ala Asp Asp Val Leu Gly Phe Pro Leu Thr Arg Leu Cys Glu
35     40     45
Glu Gly Pro Gly Glu Lys Leu Arg His Met Pro Val Thr Gln Pro Ala
50     55     60
Val Phe Leu Cys Ser Tyr Ala Ala Leu Val Ala Ala Gln Ala Asn Gly
65     70     75     80

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Ala	Glu	Pro	Asp	Val	Ile	Ala	Gly	His	Ser	Leu	Gly	Glu	Tyr	Ser	Ala
				85					90					95	
Leu	Ala	Ala	Ala	Gly	Val	Leu	Thr	Trp	Gln	Glu	Val	Leu	Gln	Leu	Val
			100					105					110		
His	Arg	Arg	Gly	Gln	Leu	Met	Ala	Glu	Val	Gln	His	Lys	Val	Asp	Gly
		115					120					125			
Lys	Met	Ala	Ala	Val	Ile	Gly	Leu	Ala	Ile	Gly	Gln	Val	Glu	Glu	Ile
	130					135					140				
Cys	Glu	Gln	Val	Arg	Ser	Glu	Thr	Gly	Glu	Val	Val	Glu	Val	Ala	Asn
145					150					155					160
His	Asn	Glu	Pro	Leu	Gln	Val	Val	Val	Ser	Gly	Gln	Cys	Ala	Ala	Ile
				165					170						175
Asp	Leu	Leu	Val	Gln	Arg	Val	Ala	Thr	Ala	Thr	Asp	Val	Arg	Thr	Ser
			180					185						190	
Val	Leu	Arg	Ile	Gly	Gly	Pro	Ala	His	Ser	Ser	Leu	Met	Gly	Ser	Val
		195					200					205			
Ala	Gly	Asp	Phe	Val	Glu	Tyr	Leu	Arg	Arg	Phe	Asp	Phe	Cys	Thr	Pro
	210					215					220				
Lys	Thr	Met	Leu	Ile	Ser	Gly	Ser	Thr	Ala	Glu	Pro	Tyr	Ala	Ser	Ala
225					230					235					240
Glu	Glu	Ile	Arg	His	Gln	Leu	Gly	Arg	Gln	Leu	Val	His	Arg	Val	Arg
				245				250						255	
Trp	Val	Asp	Val	Met	Ala	Gln	Leu	Glu	Arg	Leu	Gly	Val	Ala	Gln	Thr
			260					265						270	
Trp	Glu	Leu	Gly	Pro	Gly	Lys	Val	Leu	Ser	Gly	Phe	Val	Gln	Arg	Ser
		275				280						285			
Leu	Pro	Gln	Val	Arg	Thr	Tyr	Arg	Ala	Asn	Asp	Leu	Pro	Ser	Phe	Leu
	290					295					300				
Ala	Gly	Val	Thr	Gly	Trp										
305				310											

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What is claimed:

1. An isolated nucleic acid molecule comprising a polynucleotide sequence selected from the group consisting of:

- a) a giOI polynucleotide sequence that remains hybridized to the entirety of the full length complement of bases 19892 to 21391 of SEQ ID NO: 1 under stringent conditions of washing twice with 2 molar SSC buffer at a temperature of 45° C. and then twice with 0.1 molar SSC buffer at a temperature of 68° C., and encodes a polypeptide having FAD-dependent oxygenase activity;
- b) a giOII polynucleotide sequence that remains hybridized to the entirety of the full length complement of bases 11513 to 12196 of SEQ ID NO: 1 under stringent conditions of washing twice with 2 molar SSC buffer at a temperature of 45° C. and then twice with 0.1 molar SSC buffer at a temperature of 68° C., and encodes a post-poly-ketide synthase (post-PKS) tailoring polypeptide having oxygenase activity;
- c) a giOIII polynucleotide sequence that remains hybridized to the entirety of the full length complement of bases 6576 to 7769 of SEQ ID NO: 1 under stringent conditions of washing twice with 2 molar SSC buffer at a temperature of 45° C. and then twice with 0.1 molar SSC buffer at a temperature of 68° C., and encodes a post-PKS tailoring polypeptide having oxygenase activity; and

d) a giOIV polynucleotide sequence that remains hybridized to the entirety of the full length complement of bases 26200 to 27552 of SEQ ID NO: 1 under stringent conditions of washing twice with 2 molar SSC buffer at a temperature of 45° C. and then twice with 0.1 molar SSC buffer at a temperature of 68° C., and encodes a polypeptide having FAD-dependent oxygenase activity.

2. An isolated nucleic acid molecule comprising at least one polynucleotide sequence selected from the group consisting of:

- a) a full length giOI polynucleotide sequence that encodes the GiOI polypeptide encoded by bases 19892 to 21391 of SEQ ID NO: 1; wherein the polypeptide has FAD-dependent oxygenase activity;
- b) a full length giOII polynucleotide sequence that encodes the GiII polypeptide encoded by the reverse complement of bases 11513 to 12196 of SEQ ID NO: 1; wherein the polypeptide has oxygenase activity;
- c) a full length giOIII polynucleotide sequence that encodes the GiOIII polypeptide encoded by the reverse complement of bases 6576 to 7769 of SEQ ID NO: 1; wherein the polypeptide has oxygenase activity; and
- d) a full length giOIV polynucleotide sequence that encodes the GiIV polypeptide encoded by bases 26200 to 27552 of SEQ ID NO: 1; wherein the polypeptide has FAD-dependent oxygenase activity.

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3. The nucleic acid molecule of either claim 1 or claim 2 wherein the nucleic acid molecule encodes the gilOI, gilOII, gilOIII and gilOIV oxygenases.

4. The nucleic acid molecule of either claim 1 or claim 2 wherein the nucleic acid molecule encodes the gilOI and gilOIV.

5. A recombinant DNA expression vector comprising the nucleic acid molecule of either claim 1 or claim 2, wherein the nucleic acid is operably linked to expression control sequences.

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6. A host cell comprising a recombinant DNA expression vector comprising the nucleic acid molecule of either claim 1 or claim 2, wherein the nucleic acid is operably linked to expression control sequences.

7. A bacterial host cell comprising a recombinant DNA expression vector comprising the nucleic acid molecule of either claim 1 or claim 2, wherein the nucleic acid is operably linked to expression control sequences.

* * * * *