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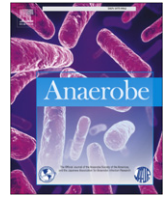
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Clinical Microbiology

Rheumatoid arthritis and periodontitis; a possible link via citrullination

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ABSTRACT

Rheumatoid Arthritis (RA) and chronic and aggressive periodontitis are chronic inflammatory disorders characterized by deregulation of the host inflammatory response. Increased secretion of pro-inflammatory mediators results in soft and hard tissue destruction of the synovium and periodontium respectively. Both diseases share risk factors and have pathological pathways in common, resulting in loss of function and disability as a final clinical outcome. This article discusses possible interactions, particularly related to the periodontal pathogen *Porphyromonas gingivalis*, which could explain the observed association between these two prevalent diseases.

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1. Periodontal disease

Supragingival plaque accumulation results in an inflammatory response of the gums and is called gingivitis. This infection can be eliminated by reduction of the total bacterial load through simple oral hygiene measures. When the infection proceeds, destructive periodontal disease or periodontitis can develop in susceptible individuals. Periodontitis is an infective process with destruction of the supporting soft and hard tissue of the teeth (the periodontium) as a consequence. It can be characterized as chronic (slowly progressive) or aggressive (highly destructive) forms of periodontitis. Further classification can be made on the extent (localized/generalized) and severity (mild/moderate/severe) of the disease [1]. Clinical signs of the disease are bleeding gums, deepened periodontal pockets, suppuration and in an advanced stage, mobility of the teeth with tooth loss as the final disease outcome due to extensive loss of alveolar bone. Periodontitis is a multifactorial, bacterial driven, chronic inflammatory disorder that occurs in 10–15% of an adult population, independent of ethnicity and geographic location [2]. It is the major factor for tooth loss over the age of 35 years. Bacteria play a major role in etiology; it is thought that the biofilm in the subgingival area causes a chronic inflammatory response that is responsible for

destruction of the alveolar bone and soft tissue surrounding the teeth (the periodontium). The subgingival biofilm in periodontal lesions consists of hundreds of bacterial species, most of which are strict anaerobic and of which a significant part is non-cultivable. Cultivable microbial indicators for periodontitis are, among others, *Aggregatibacter actinomycetemcomitans*, *Porphyromonas gingivalis*, *Prevotella intermedia*, *Parvimonas micra*, *Treponema* species and *Tannerella forsythia* [3].

Although bacteria are essential for periodontal disease to occur, a susceptible host is also required. It is thought that susceptibility is determined by genetic traits and lifestyle factors such as smoking and stress. Identified risk factors for the initiation of periodontitis are subgingival calculus and subgingival detection of *A. actinomycetemcomitans* [4]. Risk indicators for progression of the disease include smoking [5], age, stress and psychological factors [6] and existing attachment loss [7]. Other putative risk factors involve gender, education, socio-economic status [8], nutritional factors [9] and body mass index [6,10].

2. Periodontitis is not a local phenomenon

In generalized severe chronic and aggressive periodontitis the infected and necrotic epithelium surface area amounts up to 20 cm². Periodontal lesions may lead to bacteremia that can be the cause of focal infection of dental origin [11–13]. Severe periodontitis also results in a continuous systemic inflammatory response [14–16]. Periodontitis has been associated with a number

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of other chronic and inflammatory diseases such as diabetes mellitus [17], atherosclerosis, cardiovascular disease and stroke [18], rheumatoid arthritis [19,20], Crohn's disease and ulcerative colitis [21] and preterm birth and low birth weight [22]. In this paper we review current knowledge on the association of periodontitis and rheumatoid arthritis and discuss possible mechanisms of interactions between the two disorders.

3. Rheumatoid arthritis and anti-citrullinated protein antibodies

Rheumatoid arthritis (RA) is a chronic inflammatory polyarthritis with a prevalence of 0.5%–1.0% of adults in industrialized countries. The disease is far more common among women than men (3:1) and prevalence rises with age, with a peak in the fifth decade [23]. The etiology is multifactorial and the pathogenesis is poorly understood. Autoimmunity to citrullinated proteins is highly specific for RA and may be of pathogenic significance [24]. Risk of developing RA is of 50% attributable to genetic factors [25]. Smoking is the major known environmental risk factor for RA. Smoking and genetic risk factors interact in providing an increased risk of RA [26]. Immune responses with several inflammatory cascades lead toward a final common pathway with persistent synovial inflammation and associated damage to articular cartilage and underlying bone as a consequence. There is evidence of a preclinical or asymptomatic phase of the disease, in which auto-antibodies can be already be present [27]. The auto-antibodies most frequently found in patients with RA are antibodies which bind to the constant domain of IgM molecules (IgM rheumatoid factor, IgM-RF) and antibodies against citrullinated proteins (anti-citrullinated protein antibodies, ACPA). The majority of individuals with RA (50–80%) have serum positive titers for IgM-RF and/or ACPA. ACPA have a higher specificity (98%) and sensitivity (up to 80%) for diagnosis of RA than IgM-RF [28]. ACPA seem to be better predictors of poor prognosis of RA; ACPA-positive RA is associated with increased joint damage and low remission rates [29]. ACPA exist in around 2% of normal populations and are rare in other inflammatory conditions [30]. ACPA were originally measured as antibodies against keratin (the anti-perinuclear factor) [31], and more recently as anti-cyclic citrullinated peptides (anti-CCP) [32]. These auto-antibodies recognize epitopes containing citrulline. Citrulline is a nonstandard amino acid, and is therefore not incorporated in proteins during translation. However, it can be generated by post-translational modification (citrullination) of protein-bound arginine by peptidylarginine deiminase (PAD) enzymes. This post-translational modification may have a big impact on the structure and function of the target protein, partly due to a change of charge. Citrullination is an inflammation-associated phenomenon, occurring in a wide range of tissues. It is predominantly observed in proteins of the cytoskeleton. It seems to represent a general regulatory mechanism, particularly occurring during apoptosis. So far, five isotypes of PAD have been described in humans. All these enzymes rely strongly on the presence of calcium for activity and are unable to convert free L-arginine into L-citrulline, which can be done by nitric oxide synthase in eukaryotes or by arginine deiminase in bacteria. Because of their calcium dependency, PAD enzymes are more likely to be active in the extracellular compartment. PAD2 and PAD4 are found in synovial fluid and in synovial tissue of RA patients and are therefore the most likely candidate PAD isotypes for the citrullination of synovial proteins in RA [33]. Smoking enhances PAD2 expression in human lungs with consequent generation of citrullinated proteins in the bronchoalveolar compartment [34]. Recently, PAD2 expression and citrullinated proteins have also been detected in the periodontium [35]. Whereas citrullination is associated with inflammation in general,

the development of antibodies against them (ACPA) is specific to RA. The high specificity of ACPA is therefore most likely the result of an abnormal humoral response to these proteins. ACPA are produced locally in the inflamed synovium [36], suggesting that the resulting immune complexes are directly involved in the disease pathogenesis of the chronic inflammation in the rheumatoid joint. If there is local ACPA production in the periodontium or in the bronchoalveolar compartment remains to be established, albeit higher ACPA reactivity in serum samples of aggressive periodontitis patients has been reported [37].

3.1. Similarities between RA and periodontitis

There are remarkable similarities between RA and chronic and/or aggressive periodontitis. Both diseases are chronic destructive inflammatory disorders characterized by deregulation of the host inflammatory response. The etiology of both diseases is multifactorial and susceptibility to the diseases is influenced by shared genetic and lifestyle factors. Both diseases are cumulative, i.e. severity, loss of function and quality of life decrease with longer disease duration. There are common pathological mechanisms; both conditions are potentiated by an exaggerated inflammatory response featuring an increase in localized and perhaps circulating pro-inflammatory mediators, resulting in soft and hard tissue destruction of the periodontium and synovium respectively.

A number of clinical studies point toward an association between periodontal disease and RA [38], despite the fact that patients suffering from RA are often treated with immune suppressant corticosteroids, thereby possibly reducing clinical evidence of periodontal disease. An important observation is that treatment of periodontal disease has a positive effect on disease activity of RA [39], although this observation needs further confirmation. Surprisingly, none of the studies on RA and periodontal disease considered microbiology, although bacteria play a primary role in the etiology of periodontal disease. Similarities in risk factors, common pathological pathways, association in prevalence and the effect of periodontal treatment on RA make us look further to explore the relation between periodontitis and RA, with a special focus on microbiology.

4. The Bradford Hill approach

To describe the strength and nature of an association between two disorders, the widely used Bradford Hill criteria to determine a causal association are applied [40]. These involve strength, consistency, specificity, temporality, biological gradient, plausibility, coherence, experimental evidence and analogy of the association. Starting from epidemiologic evidence, four issues need to be addressed: strength, consistency, temporal relation, and analogy. The third National Health and Nutrition Examination Survey (NHANES III) is a nationally representative cross-sectional survey of non-institutionalized US population. Using this data, de Pablo et al. [41] included participants aged ≥ 60 years who had undergone both musculoskeletal and dental examinations ($n = 4461$). They found that subjects classified with RA, according to the American College of Rheumatology (ACR) criteria of 1987 ($n = 103$), were more likely to suffer from periodontitis, after adjusting for age, sex, ethnicity and smoking. Considering three out of six ACR criteria, they found an odds ratio (OR) of 1.8, considering four out of six ACR criteria an OR of 4.1. Participants with RA had significant more missing teeth than participants without RA. Comparing periodontal status in 65 RA patients (according to the ACR criteria 1987) with an age- and gender-matched control group (age range 20–70 years) without RA, Mercado et al. [42] found that individuals with RA are more likely to experience more periodontal

disease (OR 2.2) compared to individuals without RA. Individuals in the RA group showed significant more missing teeth compared to the non-RA group, an observation that confirms previous findings [43,44]. Indicators of disease activity for RA most positively correlated with periodontal bone loss were the number of swollen joints, health assessment questionnaire scores, levels of C-reactive protein and erythrocyte sedimentation rates. The other way around, in 1412 individuals attending the University of Queensland's School of Dentistry, Mercado et al. [45] found that self-reported RA was significantly higher in patients referred for periodontal treatment ($n = 809$) compared to patients not referred for periodontal treatment (3.95% vs. 0.66%). Nesse et al. [20] found in a cross-sectional study an increased prevalence of RA in patients with periodontitis, which could not be explained by the confounding factors sex, age and smoking. Coherence of the association is influenced by variation in design, setting, methods, selection bias and the fact that the majority of the studies on this association are low-prevalence case-control studies with no consistent criteria to define periodontitis. With respect to temporal relation, specific auto-antibodies (IgM-RF and ACPA) precede the symptoms of RA [27]. Half of patients with RA have specific serologic abnormalities several years (median 4.5 years) before the development of clinical symptoms. Besides the analogy of the characteristics of the population to which the diseases are exposed, the two diseases have pathological mechanisms in common and they share environmental and genetic risk factors. If there is a dose-response relation between the two diseases is currently unknown. The available studies on association of periodontitis and RA did not quantify the extent of periodontal disease, but is now possible with the Periodontal Inflamed Surface Area (PISA) index for inflammatory burden [46]. Nevertheless, antibody titers to the periodontal pathogen *P. gingivalis* are increased in patients with RA and there are significant positive correlations between *P. gingivalis* antibody titers, CRP concentrations and antibody titers to citrullinated proteins, i.e. to disease specific immunity [47]. Biological plausibility is partly explained by this association, and the fact that periodontitis causes an inflammatory burden by eliciting a systemic inflammatory response. Antibody response to *P. gingivalis* and DNA of *P. gingivalis* self have been found in synovial fluid of RA patients [48–50]. Experimental evidence is drawn from two controlled studies that have been conducted on the effect of periodontal treatment on RA [39,51]. Both studies showed that periodontal therapy had a beneficial effect on laboratory RA parameters and clinical symptoms of RA. Because these studies had a small sample size and did not consider microbiology, there is a crying need for better designed experimental studies on the effect of periodontal treatment on RA disease activity.

5. Genetic factors in RA and periodontal disease

In both diseases, candidate gene approach revealed mainly genetic variations in genes encoding for elements of the innate immune system as a risk indicator. More than 30 genetic regions are associated with RA. Genetic variations in the major histocompatibility complex, class II, DR beta 1 (HLA-DRB1) and protein tyrosine phosphatase (PTPN22) genes are the major genetic risk indicators that have been reproducibly identified so far. The association of a number of specific HLA-DRB1 alleles is seen exclusively for the ACPA-positive subset of RA [52]. These HLA alleles share a common peptide-binding motif known as the shared epitope (SE). Antigen modification by protein citrullination is thought to allow antigens to fit in the HLA alleles that hold this SE. The result is breaking of tolerance and antibody formation against these antigens [53]. The PTPN22 gene codes for a tyrosine phosphatase, with a potential function in the regulation of T-cell and B-cell activation.

The best-studied environmental factor in RA is smoking and this seems to be a risk factor for ACPA-positive disease, especially in the context of positivity for HLA-DRB1 SE alleles [54]. Studies have also shown an additive interaction between PTPN22 and smoking. No gene-gene interaction was observed between PTPN22 and HLA-DRB1 SE [55].

Genetic and lifestyle factors (smoking) have become the leading susceptibility factors in periodontal disease. The family background and the familial aggregation of early onset aggressive periodontitis have long been recognized. This supports the connection between certain genes' mutation and periodontal disease manifestation. Like RA, among candidate genes possibly associated with increased host immune susceptibility to periodontitis are HLA-DR polymorphisms. A significant association was found between HLA-DRB1 SE and severe periodontitis (chronic/aggressive), stratified according to ethnogeographic origin [56]. Several single nucleotide polymorphisms, notably in the IL1, IL6, IL10, vitamin D receptor, and CD14 genes have been linked to severity and presence of destructive periodontal disease [57]. Genes that encode for IL-1 production have received attention as potential predictors of periodontal disease progression, because of its involvement in the regulation of the host's inflammatory response and bone resorption. IL-1 is not only involved in signaling processes resulting in autoimmune induced bone destruction but also in several hereditary auto-inflammatory syndromes. Meta-analysis of four common promoter SNPs in the IL1 region in British Caucasian patients revealed an association with increased susceptibility to RA [58]. Irrespective of smoking and presence of *P. gingivalis* and *A. actinomycetemcomitans*, patients with severe periodontitis (chronic and/or aggressive) showed a significantly higher frequency of the positive IL1 genotype than periodontally healthy individuals (42% vs. 11%, all Caucasian subjects) [59]. In a study of 42 patients (1044 teeth) in maintenance care for 14 years, the combined effect of a positive IL1 genotype and smoking did increase the risk of tooth loss by 7.7 times, compared to 2.7 and 2.9 times for positive IL1 genotype and smoking separately [60]. Also, gene polymorphisms in pro-inflammatory cytokines IL6 and the IL1 cluster are associated with systemic inflammation in patients with severe periodontitis (chronic and/or aggressive, 65% European Caucasians) [15].

5.1. A link via citrullination

Given the fact that antibody formation against citrullinated proteins plays a major role in autoimmunity in RA, and given the fact that citrullination seems to be a unique feature for the periodontal pathogen *P. gingivalis*, we hypothesize that the onset and progression of RA is influenced by the presence of periodontal infection with *P. gingivalis*.

The bacteria involved in periodontitis accumulate in a sub-gingival biofilm that comprises predominantly Gram negative strict anaerobic rods. The group of dark-pigmented anaerobic rods is strongly associated with destructive periodontal disease and the major pathogen in this group is *P. gingivalis* [61]. The prevalence of *P. gingivalis* in severe periodontitis is 70% and it has been infrequently isolated from subjects without periodontitis [3], suggesting that this bacterium is not a normal inhabitant of a healthy periodontium [62]. To date, the single prokaryotic enzyme that can citrullinate proteins, has been identified in *P. gingivalis* [63]. Based on the biochemical characteristics and properties of this PAD enzyme, it could be a virulence agent. *P. gingivalis* PAD deiminates the guanidino group of carboxyl-terminal arginine residues on a variety of peptides, to yield ammonia and a citrulline residue. In contrast to human PAD, it can convert both peptidylarginine and free L-arginine and is not dependent on calcium [64].

Known antibodies to citrullinated proteins, the specific serological markers for RA, include anti-citrullinated keratin (the anti-perinuclear factor), anti-citrullinated vimentin (formerly known as the Sa-antigen), anti-citrullinated flaggrin, anti-citrullinated fibrin(ogen) and anti-citrullinated α -enolase antibodies. Alpha-enolase is a multifunctional protein, best known for its role in glucose metabolism and more recently as a plasminogen-binding protein on the surface of various mammalian and prokaryotic cells [65,66]. In RA, the immunodominant epitope of human α -enolase is citrullinated-enolase-peptide-1 (CEP-1). This epitope (amino acids 5–21) shows 82% sequence similarity with CEP-1 of *P. gingivalis*. The amino acids 13–21 are 100% identical. Antibodies purified for affinity to human CEP-1 cross-reacted with CEP-1 of *P. gingivalis* [67].

Recently, Wegner et al. [68] showed that PAD from *P. gingivalis* is able to citrullinate its endogenous proteins and more strikingly, also human fibrinogen and human α -enolase. This seems to be a unique characteristic of *P. gingivalis* [58]. Thus, the immune system in patients with periodontal infection with *P. gingivalis* is exposed to citrullinated antigens that might become systemic immunogens; directly, or via molecular mimicry and cross-reactivity. Periodontal infection with *P. gingivalis* could contribute to the total antigenic load of citrullinated proteins, generated by host PAD during the inflammatory response and by bacterial PAD produced as a virulence factor of *P. gingivalis*. In a genetic

susceptible host, for example in context of HLA-DRB1 SE, this could result in a pathologic immune response, with the formation of ACPA and joint inflammation as a consequence. Our hypothesis is that periodontitis and RA are related through common genetic and lifestyle risk factors, inflammatory burden, and in particular in presence of *P. gingivalis* (Fig. 1). To come back to the Bradford Hill criteria, biological plausibility is partly explained by the fact that periodontitis causes a systemic inflammatory response. The association of *P. gingivalis* with the RA-related anti-citrullinated protein antibody response could be a second explanation of this association. Sequence similarity and cross-reactivity with immunodominant epitopes of citrullinated proteins and their bacterial variants may indicate a role for *P. gingivalis* in autoimmunity in patients with RA. To fulfill the Bradford Hill criteria in detail, studies linking periodontal disease and RA need further investigation. If there is a distinct relation, treatment of periodontitis is thought to be of influence on disease activity of RA. By studying (pre)clinical and (micro)biological markers of both diseases, we intend to further unravel the pathogenic relation between periodontitis and RA. Recognition of the association between RA and periodontitis on both a clinical and biologic level may result in new opportunities for intervention that will modify the course of these prevalent debilitating chronic inflammatory disorders.

References

- [1] Armitage GC. Development of a classification system for periodontal diseases and conditions. *Ann Periodontol* 1999;4:1–6.
- [2] Jansson H. Studies on periodontitis and analyses of individuals at risk for periodontal diseases. *Swed Dent J Suppl*; 2006;5–49.
- [3] van Winkelhoff AJ, Loos BG, van der Reijden WA, van der Velden U. Porphyromonas gingivalis, bacteroides forsythus and other putative periodontal pathogens in subjects with and without periodontal destruction. *J Clin Periodontol* 2002;29:1023–8.
- [4] van der Velden U, Abbas F, Armand S, Loos BG, Timmerman MF, Van der Weijden GA. Java project on periodontal diseases. The natural development of periodontitis: risk factors, risk predictors and risk determinants. *J Clin Periodontol* 2006;33:540–8.
- [5] Albandar JM, Streckfus CF, Adesanya MR, Winn DM. Cigar, pipe, and cigarette smoking as risk factors for periodontal disease and tooth loss. *J Periodontol* 2000;71:1874–81.
- [6] Peruzzo DC, Benatti BB, Ambrosano GM, Nogueira-Filho GR, Sallum EA, Casati MZ. A systematic review of stress and psychological factors as possible risk factors for periodontal disease. *J Periodontol* 2007;78:1491–504.
- [7] Haffajee AD, Socransky SS, Lindhe J, Kent RL, Okamoto H, Yoneyama T. Clinical risk indicators for periodontal attachment loss. *J Clin Periodontol* 1991;18:117–25.
- [8] Skaleric U, Kovac-Kavcic M. Some risk factors for the progression of periodontal disease. *J Int Acad Periodontol* 2000;2:19–23.
- [9] Stanford TW, Rees TD. Acquired immune suppression and other risk factors/indicators for periodontal disease progression. *Periodontol* 2003;2000(32):118–35.
- [10] Chaffee BW, Weston SJ. The association between chronic periodontal disease and obesity: a systematic review with meta-analysis. *J Periodontol*; 2010.
- [11] Chiu B. Multiple infections in carotid atherosclerotic plaques. *Am Heart J* 1999;138:S534–6.
- [12] Madianos PN, Lief S, Murtha AP, Boggess KA, Auten Jr RL, Beck JD. Maternal periodontitis and prematurity. Part II: maternal infection and fetal exposure. *Ann Periodontol* 2001;6:175–82.
- [13] van Winkelhoff AJ, Slots J. Actinobacillus actinomycetemcomitans and Porphyromonas gingivalis in nonoral infections. *Periodontol*. 2000 1999;20:122–35.
- [14] Beck JD, Offenbacher S. Systemic effects of periodontitis: epidemiology of periodontal disease and cardiovascular disease. *J Periodontol* 2005;76:2089–100.
- [15] D'Aiuto F, Parkar M, Brett PM, Ready D, Tonetti MS. Gene polymorphisms in pro-inflammatory cytokines are associated with systemic inflammation in patients with severe periodontal infections. *Cytokine* 2004;28:29–34.
- [16] Southerland JH, Taylor GW, Moss K, Beck JD, Offenbacher S. Commonality in chronic inflammatory diseases: periodontitis, diabetes, and coronary artery disease. *Periodontol*. 2000 2006;40:130–43.
- [17] Mealey BL, Oates TW. Diabetes mellitus and periodontal diseases. *J Periodontol* 2006;77:1289–303.
- [18] Scannapieco FA, Bush RB, Paju S. Associations between periodontal disease and risk for atherosclerosis, cardiovascular disease, and stroke. A systematic review. *Ann Periodontol* 2003;8:38–53.
- [19] Berthelot JM, Le Goff B. Rheumatoid arthritis and periodontal disease. *Joint Bone Spine*; 2010.

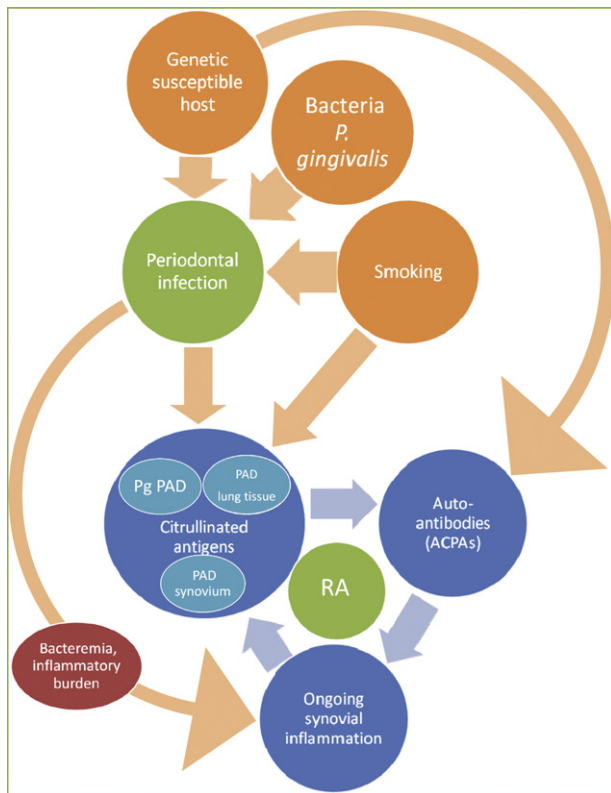


Fig. 1. Possible interactions of periodontal infection with *P. gingivalis* in etiology and pathogenesis of ACPA-positive RA. RA and periodontal infection share genetic traits, lifestyle risk factors as smoking and gene–environmental interactions (for details see text). Infection with *P. gingivalis* can cause bacteremia and generates a systemic inflammatory response, thereby contributing to the total inflammatory burden. In addition, *P. gingivalis* is able to citrullinate proteins. Given the fact that citrullination is an inflammation-associated process, *P. gingivalis* contributes in two ways to the total antigenic load of citrullinated proteins. Smoking contributes to citrullination as well. A susceptible host forms antibodies against the citrullinated proteins (ACPA), which are highly specific for RA. Immune-complex formation sustains synovial inflammation, representative for the laboratory parameters and clinical symptoms of RA.

- [20] Nesse W, Dijkstra PU, Abbas F, Spijkervet FK, Stijger A, Tromp JA. Increased prevalence of cardiovascular and autoimmune diseases in periodontitis patients: a cross-sectional study. *J Periodontol* 2010;81:1622–8.
- [21] Brito F, de Barros FC, Zaltman C, Carvalho AT, Carneiro AJ, Fischer RG. Prevalence of periodontitis and DMFT index in patients with Crohn's disease and ulcerative colitis. *J Clin Periodontol* 2008;35:555–60.
- [22] Khader YS, Ta'ani Q. Periodontal diseases and the risk of preterm birth and low birth weight: a meta-analysis. *J Periodontol* 2005;76:161–5.
- [23] Tobon GJ, Youinou P, Saraux A. The environment, geo-epidemiology, and autoimmune disease: rheumatoid arthritis. *J Autoimmun* 2010;23:10–4.
- [24] Vossenaar ER, van Venrooij WJ. Citrullinated proteins: sparks that may ignite the fire in rheumatoid arthritis. *Arthritis Res Ther* 2004;6:107–11.
- [25] van der Woude D, Houwing-Duistermaat JJ, Toes RE, Huizinga TW, Thomson W, Worthington J. Quantitative heritability of anti-citrullinated protein antibody-positive and anti-citrullinated protein antibody-negative rheumatoid arthritis. *Arthritis Rheum* 2009;60:916–23.
- [26] Kallberg H, Ding B, Padyukov L, Bengtsson C, Ronnelid J, Klareskog L. Smoking is a major preventable risk factor for rheumatoid arthritis: estimations of risks after various exposures to cigarette smoke. *Ann Rheum Dis* 2011;70:508–11.
- [27] Nielen MM, van SD, Reesink HW, van de Stadt RJ, van der Horst-Bruinsma IE, de Koning MH. Specific autoantibodies precede the symptoms of rheumatoid arthritis: a study of serial measurements in blood donors. *Arthritis Rheum* 2004;50:380–6.
- [28] Schellekens GA, Visser H, de Jong BA, van den Hoogen FH, Hazes JM, Breedveld FC. The diagnostic properties of rheumatoid arthritis antibodies recognizing a cyclic citrullinated peptide. *Arthritis Rheum* 2000;43:155–63.
- [29] van der Helm-van Mil AH, Verpoort KN, Breedveld FC, Toes RE, Huizinga TW. Antibodies to citrullinated proteins and differences in clinical progression of rheumatoid arthritis. *Arthritis Res Ther* 2005;7:R949–58.
- [30] Steiner G, Smolen J. Autoantibodies in rheumatoid arthritis and their clinical significance. *Arthritis Res* 2002;4(Suppl. 2):S1–5.
- [31] Nienhuis R, Mandema E. A new serum factor in patients with rheumatoid arthritis; the anti perinuclear factor. *Ann Rheum Dis* 1964;23:302–5.
- [32] van Venrooij WJ, Vossenaar ER, Zendman AJ. Anti-CCP antibodies: the new rheumatoid factor in the serology of rheumatoid arthritis. *Autoimmun Rev* 2004;3(Suppl. 1):S17–9.
- [33] Vossenaar ER, Radstake TR, van der HA, van Mansum MA, Dieteren C, de Rooij DJ. Expression and activity of citrullinating peptidylarginine deiminase enzymes in monocytes and macrophages. *Ann Rheum Dis* 2004;63:373–81.
- [34] Makrygiannakis D, Hermansson M, Ulfgren AK, Nicholas AP, Zendman AJ, Eklund A. Smoking increases peptidylarginine deiminase 2 enzyme expression in human lungs and increases citrullination in BAL cells. *Ann Rheum Dis* 2008;67:1488–92.
- [35] Nesse W, Westra J, van der Wal E, Balsma F, Abbas F, Brouwer E. The periodontium contains citrullinated proteins, PAD-2 enzymes and HC Gp-39; 2009. Abstract 1165 ACR/ARHP 2009 Abstract 1165 ACR/ARHP 2009[Abstract 1165 ACR/ARHP 2009], Abstract 1165 ACR/ARHP, 10-19-2009. Ref Type: Abstract.
- [36] Kinloch A, Lundberg K, Wait R, Wegner N, Lim NH, Zendman AJ. Synovial fluid is a site of citrullination of autoantigens in inflammatory arthritis. *Arthritis Rheum* 2008;58:2287–95.
- [37] Hendler A, Mulli TK, Hughes FJ, Perrett D, Bombardieri M, Hourri-Haddad Y. Involvement of autoimmunity in the pathogenesis of aggressive periodontitis. *J Dent Res* 2010;89:1389–94.
- [38] Smolik I, Robinson D, El-Gabalawy HS. Periodontitis and rheumatoid arthritis: epidemiologic, clinical, and immunologic associations. *Compend Contin Educ Dent* 2009;30:188–90. 192, 194.
- [39] Ortiz P, Bissada NF, Palomo L, Han YW, Al-Zahrani MS, Panneerselvam A. Periodontal therapy reduces the severity of active rheumatoid arthritis in patients treated with or without tumor necrosis factor inhibitors. *J Periodontol* 2009;80:535–40.
- [40] Bradford Hill A. The environment and disease: association or Causation? *Proc Royal Soc Med* 1965;58:295–300.
- [41] de Pablo P, Dietrich T, McAlindon TE. Association of periodontal disease and tooth loss with rheumatoid arthritis in the US population. *J Rheumatol* 2008;35:70–6.
- [42] Mercado FB, Marshall RI, Klestov AC, Bartold PM. Relationship between rheumatoid arthritis and periodontitis. *J Periodontol* 2001;72:779–87.
- [43] Kasser UR, Gleissner C, Dehne F, Michel A, Willershausen-Zonnchen B, Bolten WW. Risk for periodontal disease in patients with longstanding rheumatoid arthritis. *Arthritis Rheum* 1997;40:2248–51.
- [44] Tolo K, Jorkjend L. Serum antibodies and loss of periodontal bone in patients with rheumatoid arthritis. *J Clin Periodontol* 1990;17:288–91.
- [45] Mercado F, Marshall RI, Klestov AC, Bartold PM. Is there a relationship between rheumatoid arthritis and periodontal disease? *J Clin Periodontol* 2000;27:267–72.
- [46] Nesse W, Abbas F, van I d P, Spijkervet FK, Dijkstra PU, Vissink A. periodontal inflamed surface area: quantifying inflammatory burden. *J Clin Periodontol* 2008;35:668–73.
- [47] Mikuls TR, Payne JB, Reinhardt RA, Thiele GM, Maziarz E, Cannella AC. Antibody responses to *Porphyromonas gingivalis* (P. gingivalis) in subjects with rheumatoid arthritis and periodontitis. *Int Immunopharmacol* 2009;9:38–42.
- [48] Moen K, Brun JG, Madland TM, Tynning T, Jonsson R. Immunoglobulin G and A antibody responses to bacteroides forsythus and prevotella intermedia in sera and synovial fluids of arthritis patients. *Clin Diagn Lab Immunol* 2003;10:1043–50.
- [49] Moen K, Brun JG, Valen M, Skartveit L, Eribe EK, Olsen I. Synovial inflammation in active rheumatoid arthritis and psoriatic arthritis facilitates trapping of a variety of oral bacterial DNAs. *Clin Exp Rheumatol* 2006;24:656–63.
- [50] Martinez-Martinez RE, bud-Mendoza C, Patino-Marin N, Rizo-Rodriguez JC, Little JW, Loyola-Rodriguez JP. Detection of periodontal bacterial DNA in serum and synovial fluid in refractory rheumatoid arthritis patients. *J Clin Periodontol* 2009;36:1004–10.
- [51] Al-Katma MK, Bissada NF, Bordeaux JM, Sue J, Askari AD. Control of periodontal infection reduces the severity of active rheumatoid arthritis. *J Clin Rheumatol* 2007;13:134–7.
- [52] Klareskog L, Stolt P, Lundberg K, Kallberg H, Bengtsson C, Grunewald J. A new model for an etiology of rheumatoid arthritis: smoking may trigger HLA-DR (shared epitope)-restricted immune reactions to autoantigens modified by citrullination. *Arthritis Rheum* 2006;54:38–46.
- [53] Hill JA, Southwood S, Sette A, Jevnikar AM, Bell DA, Cairns E. Cutting edge: the conversion of arginine to citrulline allows for a high-affinity peptide interaction with the rheumatoid arthritis-associated HLA-DRB1*0401 MHC class II molecule. *J Immunol* 2003;171:538–41.
- [54] Kallberg H, Padyukov L, Plenge RM, Ronnelid J, Gregersen PK, van der Helm-van Mil AH. Gene-gene and gene-environment interactions involving HLA-DRB1, PTPN22, and smoking in two subsets of rheumatoid arthritis. *Am J Hum Genet* 2007;80:867–75.
- [55] Costenbader KH, Chang SC, De Vivo I, Plenge R, Karlson EW. Genetic polymorphisms in PTPN22, PADI-4, and CTLA-4 and risk for rheumatoid arthritis in two longitudinal cohort studies: evidence of gene-environment interactions with heavy cigarette smoking. *Arthritis Res Ther* 2008;10:R52.
- [56] Bonfil JJ, Dillier FL, Mercier P, Reviron D, Foti B, Sambuc R. A "case control" study on the role of HLA DR4 in severe periodontitis and rapidly progressive periodontitis. Identification of types and subtypes using molecular biology (PCR-SSO). *J Clin Periodontol* 1999;26:77–84.
- [57] Laine ML, Loos BG, Crielaard W. Gene polymorphisms in chronic periodontitis. *Int J Dent* 2010;2010:324719.
- [58] Harrison P, Pointon JJ, Chapman K, Roddam A, Wordsworth BP. Interleukin-1 p.omote region polymorphism role in rheumatoid arthritis: a meta-analysis of IL-1B-511A/G variant reveals association with rheumatoid arthritis. *Rheumatology*. (Oxford) 2008;47:1768–70.
- [59] Laine ML, Farre MA, Gonzalez G, van Dijk LJ, Ham AJ, Winkel EG. Polymorphisms of the interleukin-1 gene family, oral microbial pathogens, and smoking in adult periodontitis. *J Dent Res* 2001;80:1695–9.
- [60] McGuire MK, Nunn ME. Prognosis versus actual outcome. IV. The effectiveness of clinical parameters and IL-1 genotype in accurately predicting prognoses and tooth survival. *J Periodontol* 1999;70:49–56.
- [61] van Steenberghe TJ, van Winkelhoff AJ, van der Velden U, de Graaff J. Taxonomy, virulence and epidemiology of black-pigmented bacteroides species in relation to oral infections. *Infection* 1989;17:194–6.
- [62] Griffen AL, Becker MR, Lyons SR, Moeschberger ML, Leys EJ. Prevalence of porphyromonas gingivalis and periodontal health status. *J Clin Microbiol* 1998;36:3239–42.
- [63] McGraw WT, Potempa J, Farley D, Travis J. Purification, characterization, and sequence analysis of a potential virulence factor from porphyromonas gingivalis, peptidylarginine deiminase. *Infect Immun* 1999;67:3248–56.
- [64] Shirai H, Blundell TL, Mizuguchi K. A novel superfamily of enzymes that catalyze the modification of guanidino groups. *Trends Biochem Sci* 2001;26:465–8.
- [65] Pancholi V, Fischetti VA. Alpha-enolase, a novel strong plasmin(ogen) binding protein on the surface of pathogenic streptococci. *J Biol Chem* 1998;273:14503–15.
- [66] Wygrecka M, Marsh LM, Morty RE, Henneke I, Guenther A, Lohmeyer J. Enolase-1 p.omotes plasminogen-mediated recruitment of monocytes to the acutely inflamed lung. *Blood* 2009;113:5588–98.
- [67] Lundberg K, Kinloch A, Fisher BA, Wegner N, Wait R, Charles P. Antibodies to citrullinated alpha-enolase peptide 1 are specific for rheumatoid arthritis and cross-react with bacterial enolase. *Arthritis Rheum* 2008;58:3009–19.
- [68] Wegner N, Wait R, Sroka A, Eick S, Nguyen KA, Lundberg K. Peptidylarginine deiminase from porphyromonas gingivalis citrullinates human fibrinogen and alpha-enolase: implications for autoimmunity in rheumatoid arthritis. *Arthritis Rheum*; 2010.