

# *Candida albicans* CUG Mistranslation Is a Mechanism To Create Cell Surface Variation

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**ABSTRACT** In the human fungal pathogen *Candida albicans*, the CUG codon is translated 97% of the time as serine and 3% of the time as leucine, which potentially originates an array of proteins resulting from the translation of a single gene. Genes encoding cell surface proteins are enriched in CUG codons; thus, CUG mistranslation may influence the interactions of the organism with the host. To investigate this, we compared a *C. albicans* strain that misincorporates 28% of leucine at CUGs with a wild-type parental strain. The first strain displayed increased adherence to inert and host molecules. In addition, it was less susceptible to phagocytosis by murine macrophages, probably due to reduced exposure of cell surface  $\beta$ -glucans. To prove that these phenotypes occurred due to serine/leucine exchange, the *C. albicans* adhesin and invasin *ALS3* was expressed in *Saccharomyces cerevisiae* in its two natural isoforms (Als3p-Leu and Als3p-Ser). The cells with heterologous expression of Als3p-Leu showed increased adherence to host substrates and flocculation. We propose that CUG mistranslation has been maintained during the evolution of *C. albicans* due to its potential to generate cell surface variability, which significantly alters fungus-host interactions.

**IMPORTANCE** The translation of genetic information into proteins is a highly accurate cellular process. In the human fungal pathogen *Candida albicans*, a unique mistranslation event involving the CUG codon occurs. The CUG codon is mainly translated as serine but can also be translated as leucine. Leucine and serine are two biochemically distinct amino acids, hydrophobic and hydrophilic, respectively. The increased rate of leucine incorporation at CUG decoding triggers *C. albicans* virulence attributes, such as morphogenesis, phenotypic switching, and adhesion. Here, we show that CUG mistranslation masks the fungal cell wall molecule  $\beta$ -glucan that is normally recognized by the host immune system, delaying its response. Furthermore, we demonstrate that two different proteins of the adhesin Als3 generated by CUG mistranslation confer increased hydrophobicity and adhesion ability on yeast cells. Thus, CUG mistranslation functions as a mechanism to create protein diversity with differential activities, constituting an advantage for a mainly asexual microorganism. This could explain its preservation during evolution.

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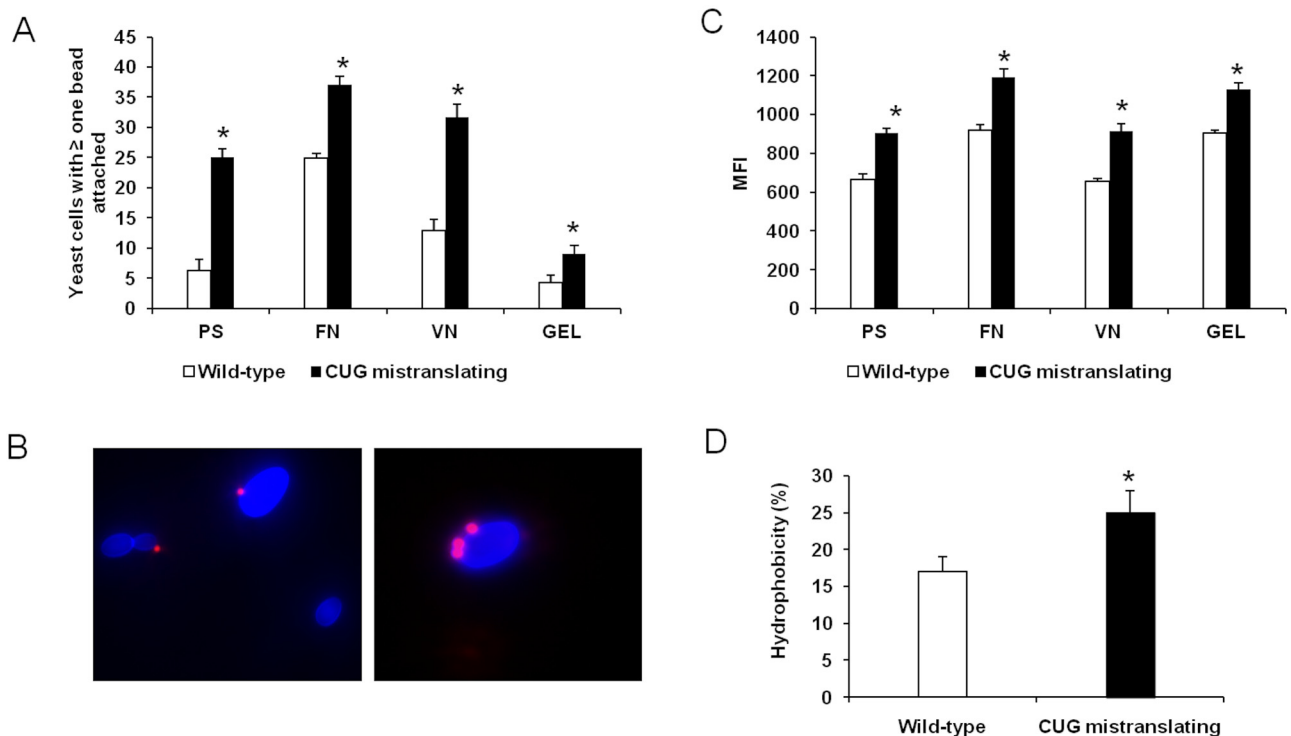
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Many pathogenic microorganisms switch surface antigens through constant variation of cell surface molecules. This process enables microbes to occupy diverse niches within the host and reduces microbe recognition by the immune system. Various mechanisms of antigenic switching have been described in both commensal and pathogenic organisms, including *Plasmodium falciparum*, *Trypanosoma brucei*, *Pneumocystis carinii*, and *Candida glabrata* (1–9).

Antigenic switching can also influence microbial adherence, an important virulence attribute that prevents a microorganism from being removed from a favorable location, thus providing the ability to infect specific niches within the host (10). The *C. albicans* *ALS* (agglutinin-like sequence) gene family encodes adhesins that bind to various substrates. Some of these Als proteins also mediate

biofilm formation, host cell invasion, and iron acquisition (11–13). Some *ALS* genes are differentially expressed by yeast versus hyphae. However, unlike the *Saccharomyces cerevisiae* *FLO* and *C. glabrata* *EPA* genes, which also specify adhesins, *C. albicans* *ALS* genes are not located near the telomeres and are not subject to subtelomeric silencing. Interestingly, the coding sequences of *ALS* genes contain a high number of CUG codons (14, 15). This codon is translated as leucine in most organisms. However, in *C. albicans* and other *Candida* species, the CUG codon is dually translated as serine (95 to 97% of the time) and leucine (3 to 5% of the time) (15). This atypical CUG translation is mediated by a Ser-tRNA<sub>CAG</sub> that, in contrast to other tRNAs, is recognized by two aminoacyl-tRNA synthetases, seryl- and leucyl-tRNA synthetases (16). Because serine is hydrophilic, whereas leucine is hydrophobic, the



**FIG 1** CUG mistranslation increases adhesion and cell surface hydrophobicity. (A) Percentage of yeast cells of the indicated strains with at least one bead attached. (B) Representative images of the wild-type strain and the highly CUG-mistranslating strain of *C. albicans*, demonstrating that the highly CUG-mistranslating strain (blue) binds to more polystyrene beads (which fluoresce red). (C) Mean fluorescence intensity (MFI) of the indicated strains. (D) Cell surface hydrophobicity was assessed by partitioning into *n*-hexadecane. Results in panels A and C represent the means  $\pm$  standard deviations of five experiments, each analyzing 50,000 cells. \*,  $P < 0.05$  compared to the wild-type strain. Abbreviations: PS, polystyrene; FN, fibronectin; VN, vitronectin; GEL, gelatin. See also Fig. S1 and S2 in the supplemental material.

variable incorporation of these two amino acids into a protein has the potential to generate a family of proteins with altered structure and function (17). The first successful manipulation of genetic code ambiguity in *C. albicans* resulted in the increase of virulence factor expression, in particular phenotypic switching, morphogenesis, and adhesion (18). However, the adhesion phenotype was not quantitatively measured, and it was not clear whether adhesion resulted from the high rates of spontaneous transitions of white to opaque and yeast to hyphal forms.

We hypothesized that Ser/Leu incorporation variability in cell surface proteins of *C. albicans* may influence its adherence to various substrates as well as its interactions with host immune effector cells. To investigate this hypothesis, we used a highly CUG-mistranslating strain of *C. albicans* that has increased incorporation of leucine at CUG codons (18). We found that CUG mistranslation increased adherence to host substrates, by changing cell surface hydrophobicity. Furthermore, the diversity in the cell wall proteome created by CUG mistranslation altered the cell surface exposure of 1,3- $\beta$ -glucan and reduced *C. albicans* phagocytosis by macrophages. These results suggest that protein diversity caused by variable translation of the CUG codon has important functional consequences for the interactions of *C. albicans* with the host.

## RESULTS

**Forced CUG mistranslation increases adhesion and cell surface hydrophobicity.** To test whether variable serine/leucine incorpo-

ration at CUG sites potentiates adhesion and hydrophobicity, we compared yeast forms of a wild-type *C. albicans* strain (pUA12) with those of a mutant strain that has a high level of CUG mistranslation (pUA15), displaying 28% leucine incorporation rather than the usual 3% leucine incorporation in the wild-type *C. albicans* (15, 18). We used a quantitative flow cytometry assay to characterize the adherence of the two *C. albicans* strains to polystyrene and the representative host ligands fibronectin, vitronectin, and collagen (19). In this assay, fluorescent polystyrene beads uncoated or coated with various substrates were incubated with nonfluorescent *C. albicans* yeasts (Fig. 1B). The number of organisms with at least one bead attached (Fig. 1A) and the mean fluorescence intensity (MFI) emitted by each yeast-bead were quantified by flow cytometry (Fig. 1C; see also Fig. S1 in the supplemental material). MFI is directly proportional to the number of beads attached to each organism (Fig. 1C; see also Fig. S1). The majority of wild-type organisms did not bind to any beads (Fig. 1A and B; see also Fig. S1). A small percentage of these cells were attached to a single bead, and even fewer cells were attached to two or more beads (Fig. S1). In contrast, a high percentage of cells of the highly CUG-mistranslating strain bound to at least one bead, and many of them bound to multiple beads (Fig. 1A to C; see also Fig. S1). Consistent with these results, the highly CUG-mistranslating cells were more adherent than wild-type cells to all tested abiotic and biotic substrates (Fig. 1A and C). While the greatest difference in adherence was found for polystyrene (more than a 4-fold increase), the increase of CUG mistranslation also

enhanced *C. albicans* adherence to biotic substrates: 1.5-fold to fibronectin, 2.6-fold to vitronectin, and 2.25-fold to gelatin (Fig. 1A). Collectively, our results clearly indicate that enhancing the leucine incorporation at CUG sites results in increased adherence to a variety of substrates. The increased adherence of the highly CUG-mistranslating strain could also be due to the altered surface expression or function of adhesin proteins. To test whether the level of CUG mistranslation influenced surface expression of adhesins, we analyzed Als1 and Als3, which mediate adherence to a variety of host substrates (20). The level of surface expression of Als1 and Als3 proteins was quantified by flow cytometry using a polyclonal antibody that recognizes both Als proteins (21). No difference in Als protein expression was found between the wild-type and highly CUG-mistranslating strains (see Fig. S2), suggesting that the increased adherence of the CUG-mistranslating strain is likely due to altered function of adhesins rather than increased surface expression of these adhesins.

Cell surface hydrophobicity is known to be important for the adherence of *C. albicans* (22). Because leucine is hydrophobic and serine is polar, it is likely that the increased leucine content in the cell surface proteins of the highly CUG-mistranslating strain enhances the hydrophobicity of this strain. Cell surface hydrophobicity was quantified using a classical approach based upon microbial adherence to hydrocarbons (23). Cells of the highly CUG-mistranslating strain were significantly more hydrophobic than those of the wild-type strain (Fig. 1D). These results suggest that serine/leucine exchange in CUG translation contributes significantly to the promotion of cell surface hydrophobicity.

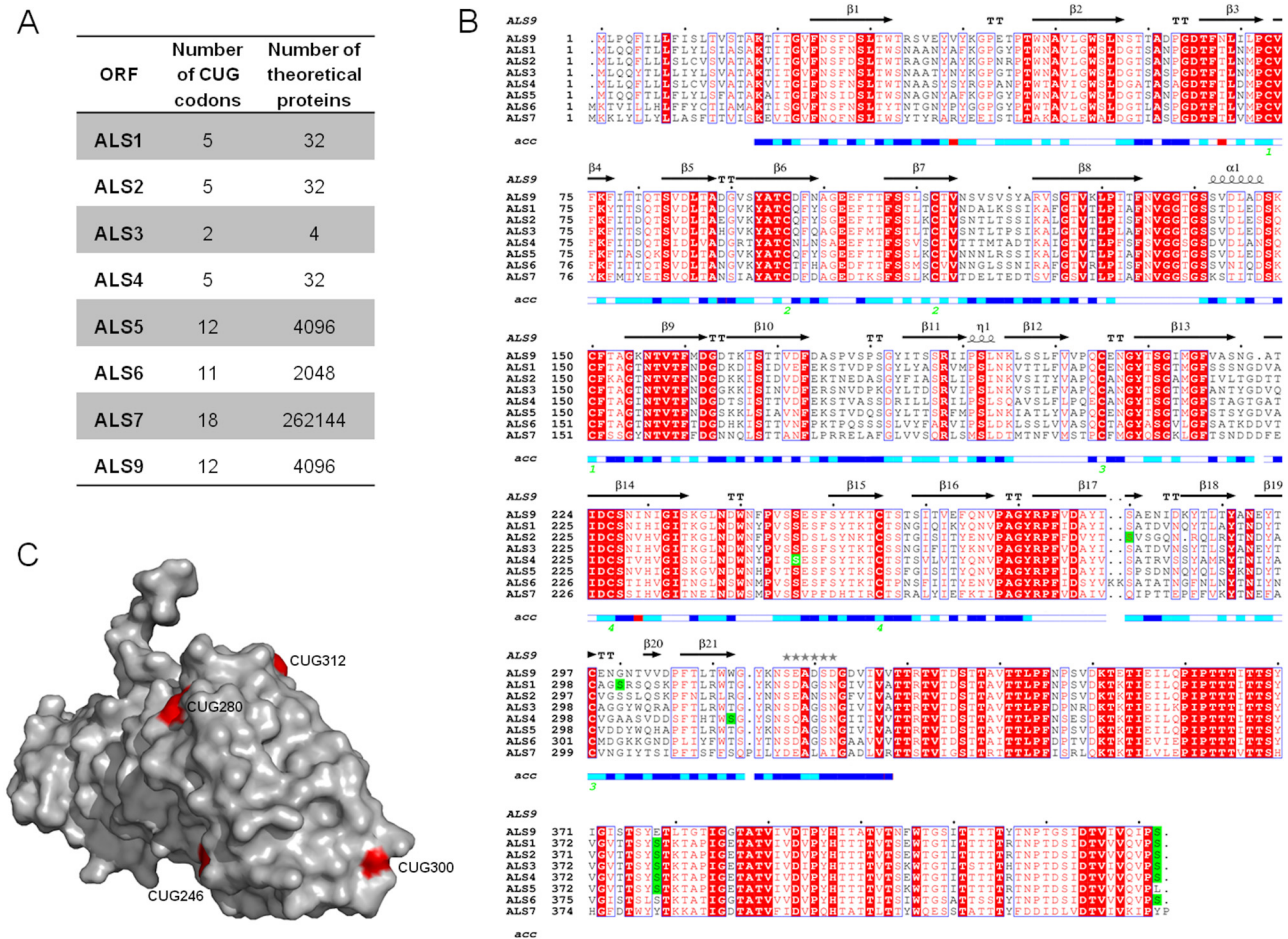
**The CUG-encoded residues of Als proteins are predicted to be surface exposed.** If serine/leucine switching in CUG decoding alters *C. albicans* adherence, where are CUG codons located in cell wall-coding genes? To gain insight into this topic, we analyzed the *ALS* gene family, which is composed of 8 genes. Als proteins are composed of four domains: a C-terminal domain, containing a site for covalent attachment of the glycosylphosphatidylinositol (GPI) anchor and rich in serine/threonine amino acid sequences; tandem repeats of highly conserved 108-bp units; an amyloid-forming region; and an N-terminal domain which contains the substrate-binding region (24). The coding sequences of *ALS* genes contain multiple CUG codons, and their mistranslation increases the number of proteins synthesized from each mRNA transcript by a factor of  $2^n$  (Fig. 2A), depending on the arbitrary serine/leucine incorporation at CUG positions. Based on the number of CUG codons and their localization, *ALS* genes can be divided into two groups: *ALS1* to *ALS4* and *ALS5* to *ALS9* (Fig. 2A and B). Members of the *ALS1* to -4 group contain 2 to 5 CUG codons, located mainly at the 5' end of the mRNA sequence that codes for the N-terminal region (Fig. 2B). Members of the *ALS5* to -9 group possess 5 to 18 CUG codons, which are predominantly located in the 3' end of the mRNA, corresponding to the C terminus of the protein. Since the crystal structure of the Als9 N-terminal domain was recently solved and the residue identity is 41 to 84% among Als sequences (25), we did a multiple protein sequence alignment in order to predict the location of CUG-encoded residues in this domain. The solvent accessibility of each residue was scored (Fig. 2B). The three-dimensional structure of the Als9 N-terminal domain (Protein Data Bank [PDB] code 2Y7N) showed that serine/leucine residues encoded by CUG codons are totally or partially exposed to the solvent (Fig. 2C), where both residues can be accommodated with no protein folding disruption. However, the

aliphatic side chain of leucine lacks the chemical reactivity of the serine hydroxyl group, which can act as a hydrogen donor. Hence, the variable incorporation of leucine or serine in the N-terminal domain should create functional and structural diversity at the Als9 molecular surface, which is exposed in the cell wall. The Als9 crystal structure does not include residues 379 and 433 (encoded by CUG codons); however, the multiple sequence alignment demonstrated that these residues are mostly encoded by CUG codons among members of the *ALS* gene family, suggesting that they have an important role in Als9 function (Fig. 2B).

#### ***S. cerevisiae* expressing Als3p-Leu has increased adherence.**

We used a heterologous expression approach to investigate the functional effects of incorporating serine versus leucine at residues 379 and 433 in Als3p, a key *C. albicans* adhesin. Two different isoforms of *C. albicans ALS3* were expressed in the normally non-adherent *S. cerevisiae*. One was the wild-type allele, which, when expressed in *S. cerevisiae*, resulted in the production of Als3p containing leucine at residues 379 and 433 (Als3p-Leu). In the other allele, the 2 CUG codons were mutated to UCG, so that the resulting protein contained serine at these positions (Als3p-Ser) (Fig. 3A). When grown in liquid medium, *S. cerevisiae* expressing Als3p-Leu showed greater flocculation than did the strain expressing Als3p-Ser (Fig. 3B), indicating that the variable translation of the CUG codon significantly influences cell-cell adhesion. Substitution of leucine for serine in Als3p also altered its capacity to bind to different substrates. *S. cerevisiae* cells expressing Als3p-Leu had significantly greater adherence to polystyrene, fibronectin, vitronectin, and gelatin than did cells expressing Als3p-Ser, as determined both by the number of cells with beads attached and by the mean fluorescence intensity (Fig. 3C). Again, these phenotypes were not due to differences in the amount of Als3p expressed on the fungal cell surface (see Fig. S3 in the supplemental material). Overall, our findings that the Als3p-Ser and Als3p-Leu isoforms exhibit markedly different adherence properties indicate that CUG mistranslation has significant functional consequences.

**CUG mistranslation reduces susceptibility to macrophage phagocytosis.** Next, we investigated the influence of CUG mistranslation on the phagocytosis of *C. albicans* by macrophages. RAW 264.7 macrophages were challenged with the wild-type and highly CUG-mistranslating *C. albicans* strains (pUA12 and pUA15, respectively). The CUG-mistranslating cells were more resistant than the wild-type strain to macrophage phagocytosis, as indicated by a reduced number of phagocytosed yeasts and fewer phagocytosed yeasts per macrophage (Fig. 4A to C). This resistance to phagocytosis was most evident after 1.5 h of incubation, and it decreased by 3 h. The reduced phagocytosis of the highly CUG-mistranslating cells could be due to the reduced surface expression of cell wall components that are recognized by macrophage pattern recognition receptors. One of these receptors is dectin-1, which recognizes  $\beta$ -glucans exposed on the fungal cell surface and triggers the phagocytosis of *C. albicans* (26–29). We hypothesized that the reduced phagocytosis of the highly CUG-mistranslating strain was due to reduced cell surface exposure of  $\beta$ -glucans. To test this hypothesis, we assessed the level of surface-exposed  $\beta$ -glucan on wild-type *C. albicans* and the highly CUG-mistranslating strain with an anti- $\beta$ -1,3-glucan monoclonal antibody using flow cytometry (30). We found that the highly CUG-mistranslating strain had 2- to 3-fold less exposure of  $\beta$ -glucans on its surface than did wild-type cells (Fig. 4D and E). This reduced exposure was seen in viable cells as well as heat-killed cells.



**FIG 2** Number of CUG codons and their localization in the 5' domain (N-terminal) coding sequence of the *ALS* gene family. (A) Coding sequences of *ALS1*, *ALS2*, *ALS3*, *ALS4*, *ALS5*, *ALS6*, *ALS7*, and *ALS9* were retrieved from the *Candida* Genome Database. The number of CUG codons ( $n$ ) present in each ORF and the number of theoretical proteins ( $2^n$ ) resulting from its translation, depending on probabilistic serine-versus-leucine incorporation during CUG translation. (B) Multiple sequence alignment of N-terminal domains of Als proteins in which the CUG-encoded serines are highlighted in green. Strictly conserved residues among Als proteins are indicated in red letters against a white background, while conserved residues are represented by white letters boxed in red. Secondary elements from the *C. albicans ALS9* crystal structure (PDB code 2Y7N) are displayed above the alignment. The solvent accessibility for each residue is indicated by a bar at the bottom of the sequence (blue, accessible; cyan, intermediate; white, buried; red, no correspondence between *ALS9* crystal structure and protein sequence). (C) Distribution of CUG positions (red residue; *ALS9* protein numbering) on the molecular surface of the *ALS* N-terminal domain (expecting residues 379 and 433).

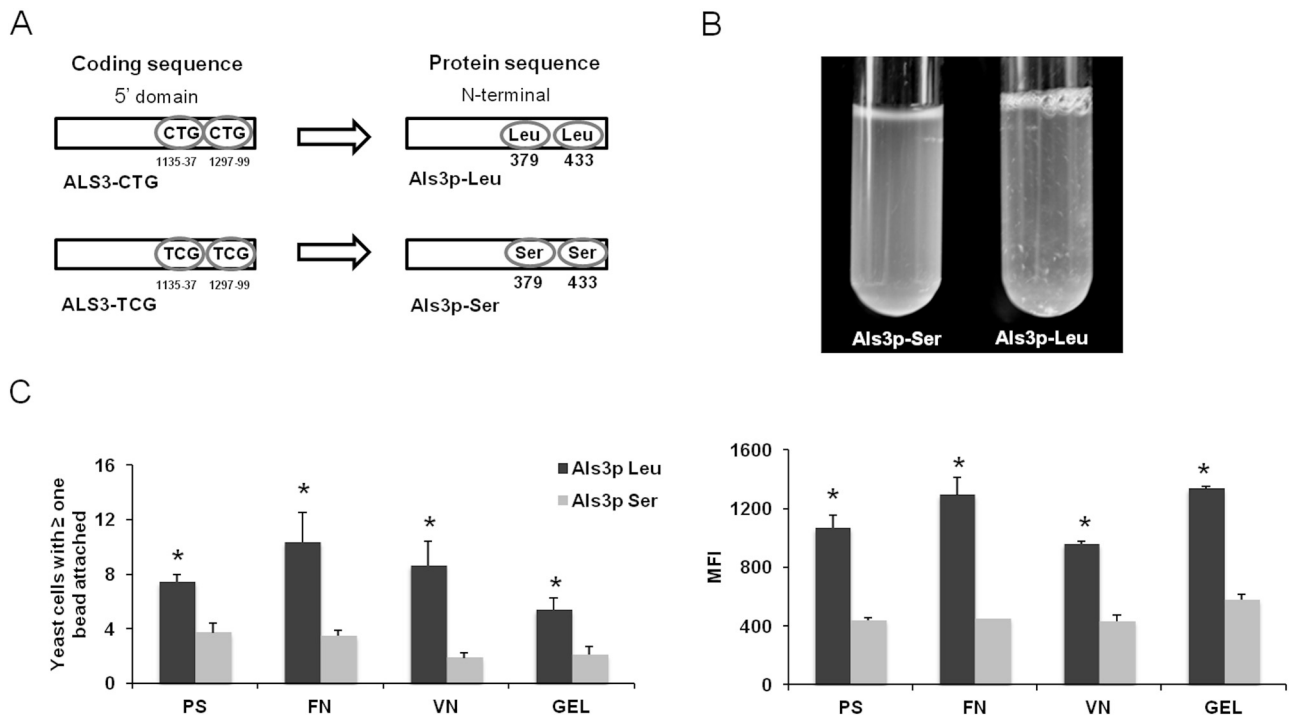
Collectively, these data suggest that increased CUG mistranslation results in decreased  $\beta$ -glucan exposure on the fungal surface, which in turn reduces susceptibility to macrophage phagocytosis.

## DISCUSSION

*C. albicans* has evolved multiple mechanisms to alter its surface characteristics. These mechanisms include white-opaque switching and yeast-to-hypha transition (31). Our data demonstrate that CUG mistranslation through the variable incorporation of polar serine or hydrophobic leucine residues is another mechanism for expanding the structural and functional diversity of cell surface proteins (Fig. 5). Genes encoding these proteins are particularly enriched in CUG codons (58% of cell wall proteome), suggesting that the protein diversity generated by CUG mistranslation may play an important role in the interactions of *C. albicans* with host cells (17). We found that increasing the rate of CUG mistranslation enhanced the adherence of *C. albicans* to polystyrene and host constituents. This increase in adherence is mediated by at least two

mechanisms. First, CUG mistranslation enhances cell surface hydrophobicity, which is known to play a role in *C. albicans* adherence to both plastic and host cells (32). Second, CUG mistranslation increases the substrate avidity of adhesins such as the Als proteins. Indeed, expression of Als3p-Leu in *S. cerevisiae* conferred significantly greater adherence to some substrates than did the expression of Als3p-Ser.

The highly conserved location of CUG codons in the regions of *ALS1* to *ALS4* that encode the substrate-binding domain suggests that CUG mistranslation likely influences the adhesive function of other Als proteins besides Als3p. Furthermore, *C. albicans* possesses additional adhesins such as Eap1 that are not members of the Als protein family (33). The *EAP1* gene also contains 2 CUG codons. Thus, CUG mistranslation might enhance the structural diversity and modulate the adherence function of other proteins, as well. This functional variability may broaden the range of host substrates to which *C. albicans* can bind by changing the ligand interaction region. Recently, it has been reported that Als proteins



**FIG 3** *C. albicans* Als3p-Leu and Als3p-Ser isoforms confer distinct adhesion profiles when expressed in the normally nonadherent *S. cerevisiae*. (A) Location of the two Leu-CUG codons in *ALS3* that were mutated to Ser-UCG codons. (B) Images of broth cultures of *S. cerevisiae* expressing Als3p-Ser (left) or Als3p-Leu (right). *S. cerevisiae* expressing Als3p-Ser did not flocculate, whereas the strain expressing Als3p-Leu exhibited strong flocculation. (C) Adherence of *S. cerevisiae* expressing Als3p-Leu or Als3p-Ser to either bare polystyrene beads or beads coated with the indicated substrates. Adherence was measured as the percentage of cells with at least one bead attached (left panel) or as the mean fluorescence intensity (right panel). Results in panel C represent the means of five flow cytometry experiments, each analyzing 50,000 cells \*,  $P < 0.05$  compared to Als3p-Leu. Abbreviations: PS, polystyrene; FN, fibronectin; VN, vitronectin; GEL, gelatin. See also Fig. S3 in the supplemental material.

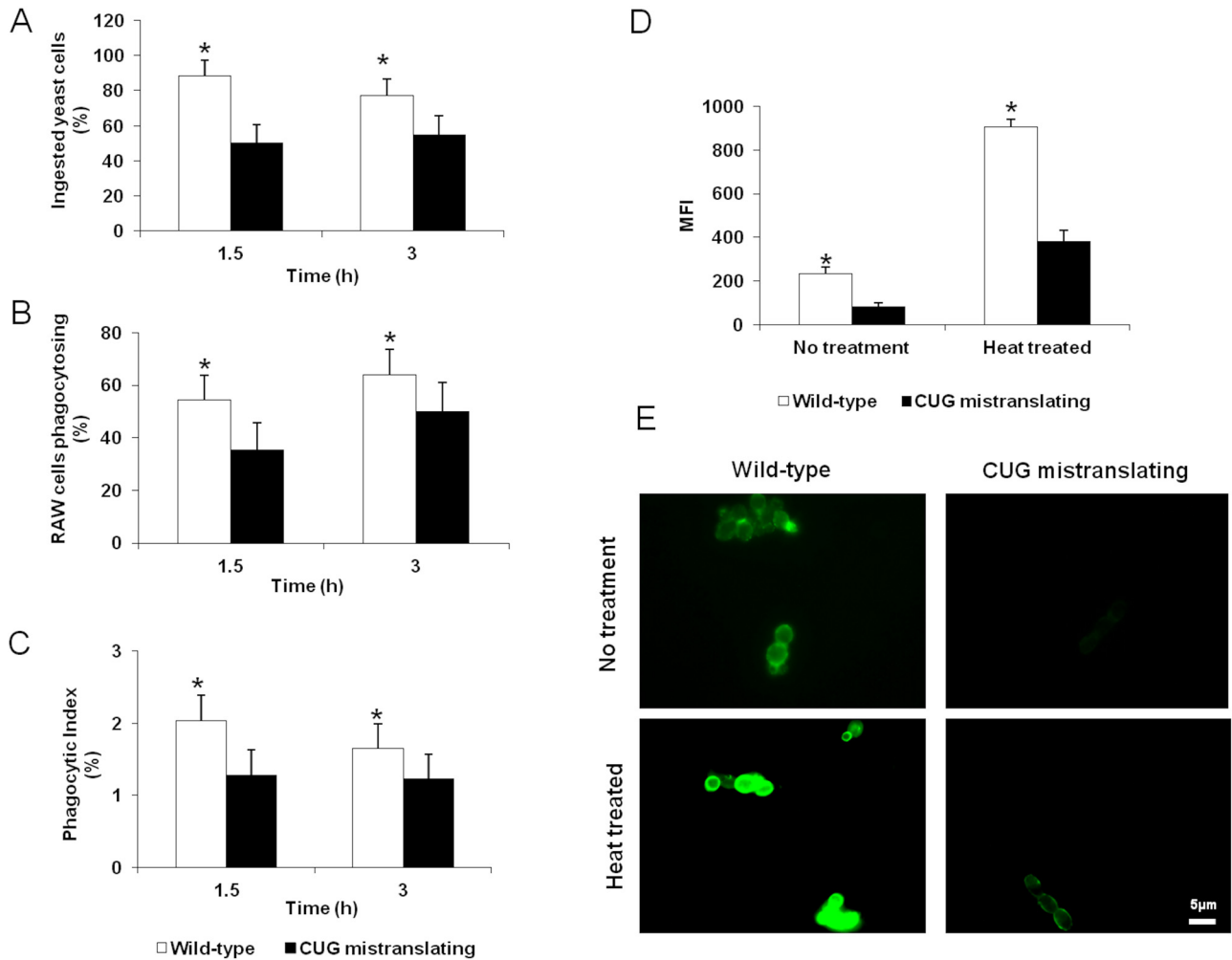
can coalesce into amyloid-like aggregates that result in the formation of adhesive nanodomains and increased cell-cell adherence (24, 34, 35). The CUG codons in *ALS1* to *ALS4* are located near the regions of these genes that are predicted to encode amyloid-forming domains. Therefore, it is also possible that CUG mistranslation might modulate the amyloid profile of Als proteins.

Even though the highly CUG-mistranslating strain of *C. albicans* had increased adherence to host constituents, we found that it was phagocytosed poorly by macrophages. Macrophages possess multiple receptors that recognize various components of the *C. albicans* cell wall (36). CR3 and dectin-1 both recognize  $\beta$ -glucans exposed on the surface of *C. albicans*, and both of these receptors mediate the phagocytosis of yeast-phase organisms (26, 29, 36). We found that the highly CUG-mistranslating strain of *C. albicans* had lower surface exposure of  $\beta$ -glucans than did the wild-type strain. This reduced  $\beta$ -glucan exposure may contribute to the reduced macrophage phagocytosis of the highly CUG-mistranslating strain. Although Ser and Asp are the residues for *O*-glycosylation and 59% of the open reading frames (ORFs) associated with cell wall organization and biogenesis are CUG-containing ORFs (17), *N*-linked and *O*-linked mannan appear not to be affected by CUG mistranslation. In contrast, the mannan outer layer is effectively masking  $\beta$ -glucans in the CUG-mistranslating strain comparably to the wild type. A defect in *N*- and *O*-glycosylation would imply an increase in *C. albicans* uptake by macrophages (37, 38). The lower phagocytosis rate observed in the CUG-mistranslating strain may also be related to a depletion

of phosphomannan cell wall, as happens in the *pmr1* mutant (38). Interestingly, the *PMR1* gene has a single CUG codon encoding a strictly conserved serine (17). The impact on Pmr1p functionality of the replacement of a serine by a leucine resulting from CUG decoding is unknown; however, if this serine-to-leucine switching implies a reduction in Pmr1p activity, a defect in phosphomannan cell wall may occur. It was notable that the highly CUG-mistranslating strain had the same susceptibility to the  $\beta$ -1,3-glucan synthase inhibitor caspofungin as did the wild-type strain. This result suggests that the total amounts of  $\beta$ -1,3-glucan in the cell walls of the two strains might be similar (39).

It was previously demonstrated that the level of leucine misincorporation at CUG codons is variable and depends upon external conditions, namely, temperature and pH (15). In addition, a disparity in activity and stability between two protein isoforms of the *C. albicans* seryl-tRNA synthetase (SerRS) was found. When the only CUG codon in SerRS is translated as leucine, the activity of this enzyme is increased, thereby raising the percentage of tRNA<sub>CAG</sub> that is serylated (17). Besides SerRS and through structural studies, authors suggest that other proteins associated with the pathogenesis process may display functional changes resulting from serine/leucine incorporation at CUG sites (17).

Although they are relatively uncommon, similar mistranslation events are known to occur in other biological systems and enhance resistance to stress. For example, when mammalian cells are subjected to oxidative stress, the population of tRNAs misacylated with methionine increases remarkably (40). The presence of



**FIG 4** CUG mistranslation reduces *C. albicans* phagocytosis by macrophages and reduces the surface exposure of  $\beta$ -glucans. (A) Percentages of wild-type and CUG-mistranslating cells ingested by a murine macrophage cell line after 1.5 h and 3 h of incubation. (B) Percentages of macrophages phagocytosing one or more yeast cells of the indicated strains after 1.5 h and 3 h of incubation. (C) Phagocytic index of macrophages (calculated as the number of ingested yeasts divided by the number of phagocytosing macrophages) incubated with wild-type and CUG-mistranslating strains. (D) Surface  $\beta$ -glucan exposure in the indicated strains quantified by flow cytometry using an anti- $\beta$ -1,3 glucan antibody. (E) Highly mistranslating strain display reduced  $\beta$ -glucan exposure comparably to the wild-type strain even after heat treatment. Images of fluorescence microscopy of *C. albicans* cells incubated with an anti- $\beta$ -1,3 glucan antibody. Results are means  $\pm$  standard deviations of three experiments, each performed in triplicate. \*,  $P < 0.05$  compared to the wild-type strain.

methionine in some proteins protects them from damage caused by reactive oxygen species. Similarly, the mistranslation of the UGA stop codon as selenocysteine protects neurons from oxidative stress (41) and is required for the normal function of macrophages (42). However, an important difference between mammalian and *C. albicans* cells is that approximately 20 selenoproteins have been described in mammalian cells (43), whereas 3,998 *C. albicans* genes contain at least one CUG codon.

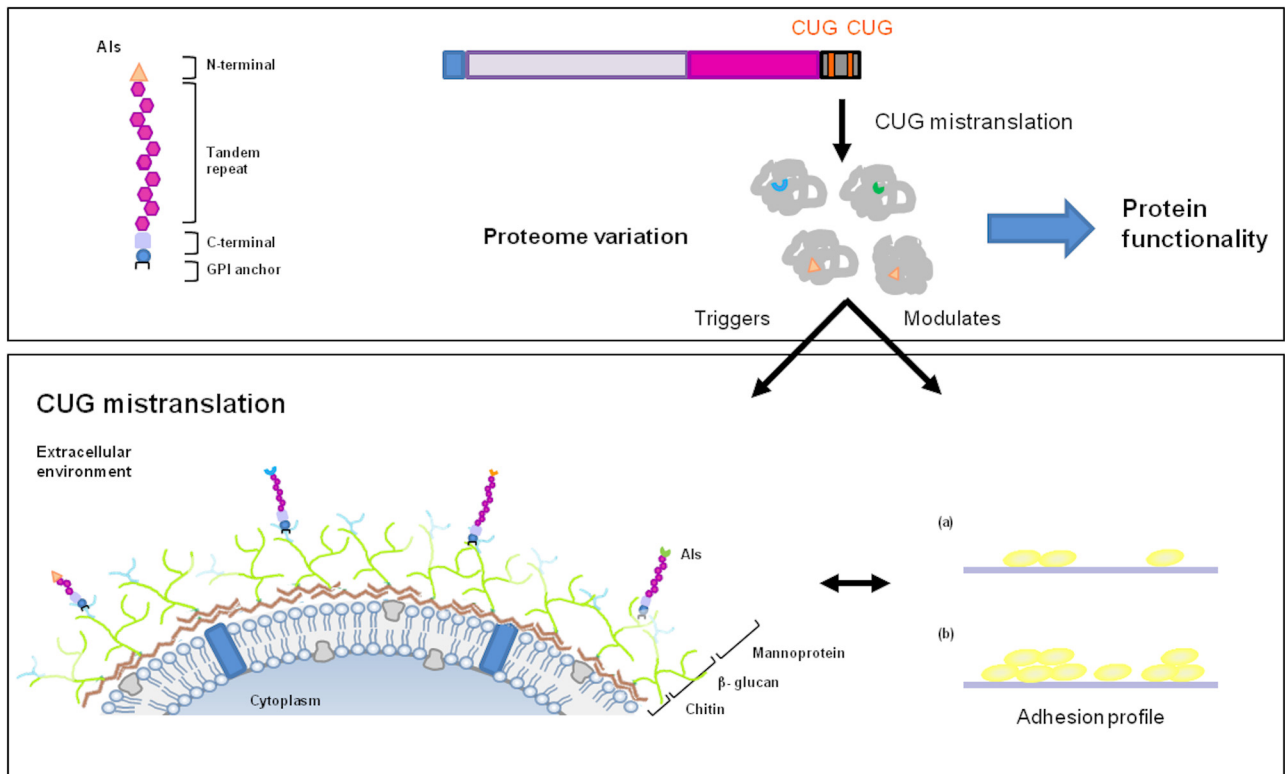
Organisms able to generate statistical proteins are thought to be in the origin of life (44). Their improved adaptability is granted by the formation of protein variants, resulting from the same coding sequence, which possess enhanced activity but reduced specificity (45). Selective growth advantages conferred by ambiguous codes, similar to the natural CUG mistranslation system but artificially originated, have been demonstrated in *S. cerevisiae* (46), *Escherichia coli* (45), and *Acinetobacter baylyi* (47).

*C. albicans* has lived as a commensal with mammalian hosts for

millions of years. Prerequisites for a successful commensal organism include the capacity to adhere to host constituents and to avoid being cleared by the host immune response. Thus, it is likely that CUG mistranslation may constitute an advantage by conferring increased capability of cells to adhere to a broad range of host substrates while minimizing interactions with immune effector cells. CUG mistranslation has the capacity not only to provide greater diversity in the cell surface proteins of *C. albicans* but also to expand the repertoire of metabolic responses of this organism (Fig. 5). Thus, CUG mistranslation likely expands the host niches in which *C. albicans* can flourish.

## MATERIALS AND METHODS

**Strains, growth conditions, and cell culture conditions.** The two *C. albicans* strains, wild type (pUA12) and highly CUG mistranslating (pUA15) (18), were grown in minimal medium lacking uridine (MM-uri) (0.67% [wt/vol] yeast nitrogen base without amino acids, 2% [wt/vol]



**FIG 5** Cell surface variability in *C. albicans* originated by CUG mistranslation. Since more than 50% of the cell wall ORFeome possesses at least 1 CUG codon, its mistranslation, either as serine or as leucine, creates a natural proteome expansion. Protein isoforms, resulting from a single ORF decoding, constitute a basis for cell surface variability in *C. albicans* and modulation of its functionality/activity. For example, the *ALS3* gene, containing 2 CUG codons in its coding sequence, can originate 4 different protein isoforms (Als-Leu versus Als-Ser), whose expression confers differential fungal adherence profiles (a and b).

glucose, and 100  $\mu\text{g/ml}$  of the required amino acids). For hydrophobicity and adherence assays, yeast cells were grown overnight at 30°C in a shaking incubator. For use in the phagocytosis experiments, the yeast cells were grown on MM-uri agar at 30°C for 48 h.

*Escherichia coli* strain DH5 $\alpha$  was used as a host to carry out *ALS3* CUG codon mutagenesis (described below), and it was grown in LB with 100  $\mu\text{g ml}^{-1}$  ampicillin.

*S. cerevisiae* strain S150-2B (*leu2 his3 trp1 ura3*) was used for heterologous expression of *C. albicans ALS3*-Leu and *ALS3*-Ser genes (20). The Als3p-expressing clones were grown overnight at 30°C in minimal medium supplemented with appropriate amino acids.

The RAW 264.7 murine macrophage cell line was obtained from Sigma-Aldrich (catalog no. ECACC 91062702) and maintained in RPMI 1640 medium (Sigma), containing 10% heat-inactivated fetal bovine serum and 2 mM L-glutamine. All mammalian cells were grown in 5% CO<sub>2</sub> at 37°C.

**Flow cytometric adhesion assay.** The adherence of *C. albicans* and *S. cerevisiae* to polystyrene and host substrates was measured by flow cytometry (19). Carboxylated highly fluorescent polystyrene beads (1  $\mu\text{m}$ ; F-8823; Invitrogen) were covalently linked with fibronectin (0.5 mg ml<sup>-1</sup>), vitronectin (10  $\mu\text{g ml}^{-1}$ ), or gelatin (5 mg ml<sup>-1</sup>) (Sigma-Aldrich) by the carbodiimide method, according to the manufacturer's instructions. *C. albicans* yeast-form cells from an overnight culture were harvested, washed with ice-cold phosphate-buffered saline (PBS), and resuspended in PBS at  $2 \times 10^6$  cells ml<sup>-1</sup>. To acid-washed glass tubes was added 200  $\mu\text{l}$  of the yeast suspension and an equal volume of PBS containing  $2 \times 10^7$  beads ml<sup>-1</sup>. The mixture was equilibrated at room temperature for 2 min, vortexed vigorously for 30 s, and then incubated on a rotary shaker (150 rpm) for 30 min at room temperature. The mixture was then vortexed briefly and then analyzed with a FACSCalibur (BD Biosci-

ences, Sydney, Australia) flow cytometer. Cell-associated fluorescence was read in the FL3 (LP, 670 nm) fluorescence channel, and 50,000 events were analyzed per condition. Controls containing yeast cells alone and beads alone were also analyzed. The adherence results were expressed as the percentage of yeast cells with at least one bead attached, the mean fluorescence intensity (MFI). Each experiment was performed in triplicate at least five times.

**Microscopic visualization of yeast-bead attachment.** For visualization of yeast cell-bead attachment, an aliquot of the yeast cell suspension prepared for the flow cytometric adhesion assay was stained with calcofluor white (0.05% [vol/vol]) (Sigma-Aldrich) and mixed with fluorescent beads. Images were taken with a Zeiss Axioplan microscope coupled with an AxioVision image acquisition system (Zeiss).

**Cell surface hydrophobicity determination.** The cell surface hydrophobicity of the two strains of *C. albicans* was analyzed as described previously (23). Yeast cells were harvested at stationary growth phase and washed twice with PBS, pH 7.0. Then, yeasts were resuspended to an optical density at 600 nm (OD<sub>600</sub>) of 0.4 to 0.5 in PBS ( $A_0$ ). Next, 3 ml of the yeast suspension was added to acid-washed spectrophotometer glass vials and overlaid with 0.4 ml of *n*-hexadecane. After vigorous vortexing, the phases were allowed to separate for 10 min at 30°C and the OD<sub>600</sub> of the aqueous phase was measured ( $A_1$ ). The percentage of hydrophobicity was calculated as follows: hydrophobicity (%) =  $[1 - (A_1/A_0)] \times 100$ . All assays were performed in triplicate.

**Sequence analysis.** The amino acid sequences of the N termini of *C. albicans* Als proteins were retrieved from the *Candida* Genome Database (<http://www.candidagenome.org/>). The position of CUG-encoded residues in the corresponding nucleotide coding sequence (bp 1 to 1299) was determined using CodonPlot (48). Multiple sequence alignments of the N-terminal domains of Als1 to Als9 were carried out using the Clust-

alW algorithm (49) and displayed with ESPrpt (similarity score matrix, BLOSUM62) (50).

**ALS3 CUG site-directed mutagenesis.** The plasmid pALS3 (20) was used as the template to mutate the two CUG codons located in the 5' domain of the coding sequence of ALS3, namely, CUGs encoding amino acids 379 and 433. CUG codons were mutated to Ser-UCG codons sequentially, using the indicated primers (ALS3\_Ser379\_For, 5' GTTGGTGTGACTACTTCTACTCGACCAAACTGCACC 3'; ALS3\_Ser379\_Rev, 5' GGTGCAGTTTGGTCGAGTAGGAAGTAGTCACACCAAC 3'; ALS3\_Ser433\_For, 5' CTGTCATTGTACAAGTTCATCGCCAAACCCAACTGTTAC 3'; ALS3\_Ser433\_Rev, 5' GTAACAGTTGGTTTGGCGATGGAAGTTGTACAATGACAG 3') and the QuikChange site-directed mutagenesis kit (Stratagene). Plasmids pALS3 and pALS3-UCG<sub>379/433</sub> were transformed into *S. cerevisiae* strain S150-2B by the lithium acetate method.

**Macrophage phagocytosis assay.** The phagocytosis of the two strains of *C. albicans*, wild type and CUG mistranslating, by the RAW 264.7 macrophage cell line was quantified as described previously (51). Briefly, yeast cells suspended in PBS were labeled with Oregon Green 488 (1  $\mu$ M; Invitrogen) in the dark for 1 h at 30°C. Afterward, the yeast cells were washed twice with PBS-100 mM glycine and resuspended in PBS to a final yeast cell concentration of  $5 \times 10^7$  cells ml<sup>-1</sup>. RAW 264.7 macrophages were added to coverslips in a 6-well-tissue culture plate ( $1 \times 10^6$  cells well<sup>-1</sup>) and incubated with 100  $\mu$ l of yeast cell suspension, for 1.5 h and 3 h. The cells were then rinsed with ice-cold PBS and fixed with 4% paraformaldehyde in the dark for 45 min at 4°C. The nonphagocytosed yeasts were stained with calcofluor white M2R (2.5  $\mu$ M; Sigma-Aldrich) in the dark for 5 min at 4°C and then rinsed to remove the unincorporated dye. The cells were dehydrated by immersion in graded ethanol solutions (70%, 90%, and 100%; 20 s each), cleared with xylene for 20 s, and mounted on glass slides using DPX mounting medium (Fluka Bio-Chemika, Germany). The numbers of phagocytosed yeast cells (green fluorescent), noningested yeast cells (blue fluorescent), and macrophages were quantified by phase-contrast and fluorescence microscopy. Digital images were taken and analyzed using ImageJ software. At least three clones of each yeast strain were analyzed. For each clone, four slides were prepared and at least two experiments were carried out in consecutive days. The data were expressed as the percentage of yeast cells phagocytosed, as the percentage of RAW 264.7 cells phagocytosing at least one yeast, and as a phagocytosis index (the number of yeast phagocytosed by each macrophage).

**Detection of  $\beta$ -glucan surface expression.** The amount of surface-exposed  $\beta$ -glucan on the two strains of *C. albicans*, wild type and CUG mistranslating, was determined (30). Briefly, viable and heat-killed (90°C for 20 min) yeast cells were incubated with an anti- $\beta$ -1,3-glucan mouse monoclonal antibody (Biosupplies) for 15 min at 4°C. After being rinsed, the cells were incubated with a fluorescein isothiocyanate (FITC)-conjugated goat anti-mouse IgG secondary antibody (Sigma) for 15 min at 4°C. Yeasts were then fixed with 0.4% paraformaldehyde, and 20,000 events were analyzed in the FL1 (BP, 530/30 nm) fluorescence channel using a FACSCalibur flow cytometer. Each strain was tested in triplicate. The experiment was repeated two times. Quantitative flow cytometry assays were confirmed by epifluorescence microscopy using a Zeiss AxioPlan microscope coupled with an AxioVision image acquisition system (Zeiss).

## SUPPLEMENTAL MATERIAL

Supplemental material for this article may be found at <http://mbo.asm.org/lookup/suppl/doi:10.1128/mBio.00285-13/-DCSupplemental>.

- Figure S1, TIF file, 1.7 MB.
- Figure S2, TIF file, 0.8 MB.
- Figure S3, TIF file, 0.9 MB.

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