



REVIEW

Open Access

Osteopontin: a leading candidate adhesion molecule for implantation in pigs and sheep

Greg A Johnson^{1*}, Robert C Burghardt¹ and Fuller W Bazer²**Abstract**

Osteopontin (OPN; also known as Secreted Phosphoprotein 1, SPP1) is a secreted extra-cellular matrix (ECM) protein that binds to a variety of cell surface integrins to stimulate cell-cell and cell-ECM adhesion and communication. It is generally accepted that OPN interacts with apically expressed integrin receptors on the uterine luminal epithelium (LE) and conceptus trophoctoderm to attach the conceptus to the uterus for implantation. Research conducted with pigs and sheep has significantly advanced understanding of the role(s) of OPN during implantation through exploitation of the prolonged peri-implantation period of pregnancy when elongating conceptuses are free within the uterine lumen requiring extensive paracrine signaling between conceptus and endometrium. This is followed by a protracted and incremental attachment cascade of trophoctoderm to uterine LE during implantation, and development of a true epitheliochorial or synepitheliochorial placenta exhibited by pigs and sheep, respectively. In pigs, implanting conceptuses secrete estrogens which induce the synthesis and secretion of OPN in adjacent uterine LE. OPN then binds to $\alpha v\beta 6$ integrin receptors on trophoctoderm, and the $\alpha v\beta 3$ integrin receptors on uterine LE to bridge conceptus attachment to uterine LE for implantation. In sheep, implanting conceptuses secrete interferon tau that prolongs the lifespan of CL. Progesterone released by CL then induces OPN synthesis and secretion from the endometrial GE into the uterine lumen where OPN binds integrins expressed on trophoctoderm ($\alpha v\beta 3$) and uterine LE (identity of specific integrins unknown) to adhere the conceptus to the uterus for implantation. OPN binding to the $\alpha v\beta 3$ integrin receptor on ovine trophoctoderm cells induces *in vitro* focal adhesion assembly, a prerequisite for adhesion and migration of trophoctoderm, through activation of: 1) P70S6K via crosstalk between FRAP1/MTOR and MAPK pathways; 2) MTOR, PI3K, MAPK3/MAPK1 (Erk1/2) and MAPK14 (p38) signaling to stimulate trophoctoderm cell migration; and 3) focal adhesion assembly and myosin II motor activity to induce migration of trophoctoderm cells. Further large *in vivo* focal adhesions assemble at the uterine-placental interface of both pigs and sheep and identify the involvement of sizable mechanical forces at this interface during discrete periods of trophoblast migration, attachment and placentation in both species.

Keywords: Implantation, Integrins, Osteopontin, Pigs, Sheep**Introduction**

Domestic animal models for research are generally underappreciated [1]; however, pigs and sheep offer unique characteristics of pregnancy, as compared to rodent or primate models, and studies of pigs and sheep have provided significant insights into the physiology of implantation including: 1) elongation of the blastocyst into a filamentous conceptus; 2) the protracted peri-implantation period of pregnancy when the conceptus is free within the uterine

lumen requiring extensive paracrine signaling between conceptus and endometrium, as well as nutritional support provided by uterine secretions; 3) a protracted and incremental attachment cascade of trophoctoderm to endometrial epithelium during implantation; and (4) development of a true epitheliochorial or synepitheliochorial placenta, respectively, that utilizes extensive uterine and placental vasculatures for hematotropic nutrition, and placental areolae for histotropic support of the developing fetuses. Our understanding of the complex mechanistic events that underlie successful implantation and placentation across species has been and will likely continue to be advanced by studies of pigs and sheep as biomedical research models

* Correspondence: gjohnson@cvm.tamu.edu¹Department of Veterinary Integrative Biosciences, Texas A&M University, College Station, TX 77843-4458, USA

Full list of author information is available at the end of the article

and to increase reproductive success in animal agriculture enterprises providing high quality protein for humans.

Overview of the biology of osteopontin (OPN)

OPN is a secreted extracellular matrix (ECM) protein that binds to a variety of cell surface integrins and several CD44 variants [2-6]. Integrins are transmembrane glycoprotein receptors composed of non-covalently bound α and β subunits that promote cell-cell and cell-ECM adhesion, cause cytoskeletal reorganization to stabilize adhesion, and transduce signals through numerous signaling intermediates [7,8]. Integrin-mediated adhesion is focused within a primary mechanotransduction unit of dynamic structure and composition known as a focal adhesion whose size, composition, cell signaling activity and adhesion strength are force-dependent [2,9]. The intrinsic properties of the ECM in different niches and tissue-level compartments affect the composition and size of focal adhesions that, in turn, modulate cell behavior including gene expression, protein synthesis, secretion, adhesion, migration, proliferation, viability and/or apoptosis [10]. Integrins are dominant glycoproteins in many cell adhesion cascades, including well defined roles in leukocyte adhesion to the apical surface of polarized endothelium for extravasation of leukocytes from the vasculature into tissues [11]. A similar adhesion cascade involving interactions between the ECM and apically expressed integrin receptors on the uterine luminal epithelium (LE) and conceptus (embryo and placental membranes) trophoblast is proposed as a mechanism for attachment of the conceptus to the uterus for implantation; the initial step for the extensive tissue remodeling that occurs during placentation [12]. OPN is a leading candidate adhesion molecule for implantation in pigs and sheep [13].

OPN is an acidic member of the small integrin-binding ligand *N*-linked glycoprotein (SIBLING) family of proteins [9]. The breadth of literature pertaining to the diverse functions of OPN is extensive, and OPN has been independently identified as a protein associated with metastatic cancers (2ar), as an ECM protein of bones and teeth (OPN, BSP1, BNSP, SPP1), as a cytokine produced by activated lymphocytes and macrophages (early T-cell activation factor 1, Eta-1), and as a major constituent of the uterus and placenta during pregnancy [13-17]. In general, OPN is a monomer ranging in length from 264 to 301 amino acids. OPN contains a hydrophobic leader sequence characteristic of a secreted protein, a calcium phosphate apatite binding region of consecutive asparagine residues, a GRGDS sequence that interacts with integrins, a thrombin cleavage site, and two glutamine residues that are recognized substrates for transglutaminase-supported multimer formation [3,5,6]. Genes encoding OPN from different species present only moderate sequence conservation, except in the NH₂-terminal region, around the

Arg-Gly-Asp (RGD) integrin-binding sequence, and in the COOH-terminus [3,5,6,18]. OPN undergoes extensive posttranslational modifications that can alter its function in different physiological microenvironments. These modifications include proteolytic cleavage, phosphorylation, glycosylation, sulfation and cross-linking with self and other macromolecules [19-23]. OPN is present on epithelial cells and in secretions of the gastrointestinal tract (including the liver), respiratory tract, kidneys, thyroid, breast, testes, uterus and placenta [24-32]. Other cell types that express OPN include leukocytes, smooth muscle cells, and highly metastatic cancer cells [33-35]. OPN is a multi-functional ECM protein reported to 1) stimulate cell-cell adhesion, 2) increase cell-ECM communication, 3) promote cell migration, 4) decrease cell death, 5) stimulate immunoglobulin production, 6) induce changes in phosphorylation of focal adhesion kinase and paxillin, 7) stimulate phosphatidylinositol 3'-kinase activity, 8) alter intracellular calcium levels, and 9) promote calcium phosphate deposition [36-42].

Timeline of key advancements in understanding the role of OPN as an attachment factor for implantation

OPN was first observed in endometrial tissue when, in 1988, Nomura et al., [43] performed *in situ* hybridization to localize OPN in mouse embryos, the endometrium from the gravid and non-gravid uterine horns of pregnant mice, and the endometrium from mice exposed to intra-uterine injection of oil to induce a decidualoma. High levels of OPN mRNA were detected in the LE, but not GE, of the gravid uterine horns. Interestingly, epithelial expression of OPN appeared to be specific to pregnancy because little to no OPN mRNA was observed in the uterine LE of non-gravid or pseudopregnant mice [43]. In addition to the LE, high levels of OPN mRNA were localized to the granulated metrial gland (GMC) cells of decidual and decidualoma tissues, with lower numbers of OPN positive cells in the decidualoma of uteri [43]. It is noteworthy that these investigators were the first to argue that OPN plays a wider role than had previously been assumed, and that its functions are not confined to bone development. The decidual cells that express OPN have since been confirmed to be uterine natural killer (uNK) cells [44,45]. Similar to expression in mice, immunocytochemical studies performed by Young and colleagues in 1990 [25] localized OPN protein to the decidua of women; however, in contrast to mice, OPN was also expressed by the secretory phase endometrial GE. It was suggested that the absence of OPN in GE during the proliferative phase of the menstrual cycle indicated that changes in expression in GE of normal cycling endometrium were the result of hormonal regulation and that the function(s) of OPN in the endometrium might be associated with its ability to enhance cell attachment [25].

A significant conceptual advance regarding the function(s) of epithelia-derived OPN was made by Brown and co-workers [26] in 1992, when OPN mRNA and protein were localized to epithelial cells of a variety of organs, including the hypersecretory endometrial GE associated with pregnancy in women. In the secretory epithelia of all organs examined, OPN protein was associated with the apical domain of the cells, and when the luminal contents were preserved in tissue sections, proteins secreted into the lumen were positive for OPN staining. It was hypothesized that OPN secreted by epithelia, including uterine epithelia, binds integrins on luminal surfaces to effect communication between the surface epithelium and the external environment [26]. Between 1992 and 1996, Lessey and co-workers established that transient uterine expression of $\alpha\beta3$ and $\alpha4\beta1$ integrins defines the window of implantation in women [46-48] and that altered expression of these integrins correlates with human infertility [49,50]. Noting that the $\alpha\beta3$ and $\alpha4\beta1$ integrin heterodimers present during the implantation window bind OPN, these investigators suggested involvement of OPN and integrins in trophoblast-endometrial interactions during the initial attachment phase of implantation [46].

Comprehensive examination of the temporal and spatial expression and hormonal regulation of uterine OPN mRNA and protein and integrin subunit proteins in the uteri and placentae of sheep (discussed in detail later in this review), performed from 1999 through 2002, provided the first strong evidence that OPN is a progesterone-induced secretory product of endometrial glands (histotroph) that binds integrins on apical surfaces of endometrial LE and conceptus trophoctoderm to mediate attachment of uterus to trophoctoderm for implantation [18,29,51,52]. Indeed, pregnant Day 14 ewes, which lack uterine glands (uterine gland knockout, UGKO phenotype), exhibit an absence of OPN in uterine flushings compared with normal ewes, and do not maintain pregnancy through the peri-implantation period [53]. Similarly, functional intrauterine blockade of α and $\beta3$ integrin subunits, that combine to form a major receptor for OPN, reduces the number of implantation sites in mice and rabbits [54,55]. Further evidence for regulation of uterine OPN by sex steroids was provided by results from studies using human and rabbit models. Progesterone treatment increased OPN expression by human endometrial adenocarcinoma Ishakawa cells (*in vitro* findings, 2001) as well as endometrium of rabbits (*in vivo* findings, 2003) [56,57]. In contrast, i.m. injection of estrogen induced expression of OPN in the uterine LE of cyclic pigs (*in vivo*, 2005) [58]. Results from pigs were the first to suggest that conceptuses can directly regulate the regional expression of OPN in the endometrium at specific sites of implantation through secretion of estrogens [58,59]. Microarray studies from 2002 and 2005 strongly support a role for OPN during implantation [60-62]. Two reports

confirmed that OPN is the most highly up-regulated ECM-adhesion molecule in the human uterus as it becomes receptive to implantation [60-62].

Research regarding OPN has begun to focus on its interactions with integrin receptors in the female reproductive tract. In 2009, Burghardt and colleagues [63] reported the *in vivo* assembly of large focal adhesions containing aggregates of α , $\alpha4$, $\alpha5$, $\beta1$, $\beta5$, alpha actinin, and focal adhesion kinase (FAK) at the uterine-placental interface of sheep, that expand as pregnancy progresses. It is noteworthy that OPN was present along the surfaces of both uterine LE and trophoctoderm, although it was not determined whether it co-localized to the focal adhesions [63]. Similar focal adhesions form during implantation in pigs [64,65]. Affinity chromatography and immunoprecipitation experiments revealed direct *in vitro* binding of porcine trophoctoderm $\alpha\beta6$ and uterine epithelial cell $\alpha\beta3$, and ovine trophoctoderm $\alpha\beta6$ integrins to OPN [64,66]. These were the first functional demonstrations that OPN directly binds specific integrins to promote trophoctoderm cell migration and attachment to uterine LE that may be critical to conceptus elongation and implantation. Recently (2014), Aplin and co-workers [67] employed three *in vitro* models of early implantation with Ishakawa cells to demonstrate that OPN potentially interacts with the $\alpha\beta3$ integrin receptor during implantation in humans.

Key events during the peri-implantation period of pigs and sheep

Communication and reciprocal responses between the conceptus and uterus are essential for conceptus survival during the peri-implantation period of pregnancy. These interactions also lay the critical physiological and anatomical groundwork for subsequent development of functional uterine LE, GE, stroma and placentae required to maintain growth and development of the conceptus throughout pregnancy. In a progesterone dominated uterine environment, establishment and maintenance of pregnancy in pigs and sheep requires; (i) secretion of estrogens or interferon tau, respectively, from the conceptus to signal pregnancy recognition [68-71], (ii) secretions from uterine LE and GE, i.e., histotroph, to support attachment, development and growth of the conceptus [72-74], and (iii) cellular remodeling at the uterine LE-conceptus trophoctoderm interface to allow for attachment during implantation [8,75,76]. These events are orchestrated through endocrine, paracrine, autocrine and juxtacrine communication between the conceptus and uterus, and the complexity of these events likely underlies the high rates of conceptus mortality during the peri-implantation period of pregnancy [77,78].

Implantation and placentation are critical events in pregnancy. Implantation failure during the first three weeks of pregnancy is a major cause of infertility in all

mammals [77-80]. The process of implantation is highly synchronized, requiring reciprocal secretory and physical interactions between a developmentally competent conceptus and the uterine endometrium during a restricted period of the uterine cycle termed the “window of receptivity”. These initial interactions between apical surfaces of uterine LE and conceptus trophoctoderm begin with sequential phases i.e., non-adhesive or pre-contact, apposition, and adhesion, and conclude with formation of a placenta that supports fetal-placental development throughout pregnancy [81-83]. Conceptus attachment first requires loss of anti-adhesive molecules in the glycocalyx of uterine LE, comprised largely of mucins that sterically inhibit attachment [52,84,85]. This results in “unmasking” of molecules, including selectins and galectins, which contribute to initial attachment of conceptus trophoctoderm to uterine LE [86-88]. These low affinity contacts are then replaced by a repertoire of adhesive interactions between integrins and maternal ECM which appear to be the dominant contributors to stable adhesion at implantation [1,8,52,89-91]. OPN is expressed abundantly within the conceptus-maternal environment in numerous species, including pigs and sheep [17,29,57,59,62,92,93].

Osteopontin is structurally and functionally suited to support implantation of pig and sheep conceptuses

Depending on cell context and species, OPN expression can be regulated by multiple hormones and cytokines, including the sex steroids progesterone and estrogen [28,51,56-58,94-98]. OPN mediates multiple cellular processes, such as cell-mediated immune responses, inflammation, angiogenesis, cell survival, and tumor metastasis primarily through integrin signaling [3,5,17,99,100]. Integrins are transmembrane glycoprotein receptors composed of non-covalently bound α and β subunits that participate in cell-cell and cell-ECM adhesion, cause cytoskeletal reorganization to stabilize adhesion, and transduce signals through numerous signaling intermediates [7,8]. OPN has an expansive integrin receptor repertoire that includes RGD-mediated binding to $\alpha v \beta 3$ [101,102], $\alpha v \beta 1$ [103], $\alpha v \beta 5$ [103], and $\alpha 8 \beta 1$ [104], as well as alternative binding sequence-mediated interactions with $\alpha 4 \beta 1$ [105], and $\alpha 9 \beta 1$ [106]. OPN binding to these various receptors results in diverse effects including: (1) leukocyte, smooth muscle cell and endothelial cell chemotaxis; (2) endothelial and epithelial cell survival; and (3) fibroblast, macrophage and tumor cell migration [64,66,103,104,107]. Clearly, the ability to bind multiple integrin receptors to produce different cellular outcomes greatly increases OPN's potential role(s) during conceptus development and implantation. Importantly, OPN contains a serine protease cleavage site that when activated generates bioactive OPN fragments [23,108], and two glutamines that support multimerization of the protein [22]. It is notable that OPN is flexible in solution,

allowing for simultaneous binding to more than one integrin receptor [16,109]. Further, OPN can also exist in a polymerized form cross-linked by transglutaminase. Homotypic OPN bonds have high tensile strength, suggesting that self-assembly is involved in cell-cell and cell-matrix interactions [22]. These multimeric complexes may present multiple RGD sequences for simultaneous binding to integrins on multiple surfaces [22,110]. Therefore, OPN has the potential to bind multiple proteins and to participate in assembly of multi-protein complexes that bridge and form the interface between conceptus to uterus during implantation.

OPN expression, regulation and function in the uterus and placenta of gilts

A hallmark of pregnancy in pigs is the protracted peri-implantation period of pregnancy when conceptuses are free within the uterine lumen to elongate from spherical blastocysts to conceptuses with a filamentous morphology (Reviewed in [111]). Pig embryos move from the oviduct into the uterus about 60 to 72 h after onset of estrus, reach the blastocyst stage by Day 5, then shed the zona pellucida and expand to 2–6 mm in diameter by Day 10. At this stage, development of pig embryos diverges from that of rodents or primates. Within a few hours the presumptive placental membranes (trophoctoderm and extra-embryonic endoderm) elongate at a rate of 30–45 mm/h from a 10 mm blastocyst to a 150–200 mm long filamentous form, after which further elongation occurs until conceptuses are 800–1,000 mm in length by Day 16 of pregnancy [111]. During this period of rapid elongation, porcine conceptuses secrete estrogen beginning on Days 11 and 12 to signal initiation of pregnancy to the uterus, and by Day 13 begin an extended period of incremental attachment to the uterine LE [17,69]. The attached trophoctoderm/chorion-endometrial epithelial bilayer develops microscopic folds, beginning about Day 35 of gestation, and these folds increase the surface area of contact between maternal and fetal capillaries to maximize maternal-to-fetal exchange of nutrients and gases [112].

In pigs, OPN is an excellent candidate for influencing this complex environment of pregnancy, because the OPN gene is located on chromosome 8 under a quantitative trait loci (QTL) peak for prenatal survival and litter size, [113]. The temporal and spatial expression of OPN in the porcine uterus and placenta is complex, with independent and overlapping expression by multiple cell types. Between Days 5 and 9 of the estrous cycle and pregnancy, OPN transcripts are detectable in a small percentage of cells in the sub-epithelial stratum compactum of the endometrial stroma [59]. The morphology and distribution of OPN mRNA- and protein-positive cells in the stratum compactum of the stroma on Day 9

of the estrous cycle and pregnancy suggest that these are immune cells. Certainly Eta-1/OPN, is an established component of the immune system that is secreted by activated T lymphocytes [15]. It is reasonable to speculate that because insemination in pigs is intrauterine, OPN expressing immune cells may protect against pathogens introduced during mating. A similar pattern of distribution of OPN-producing cells is also evident in the allantois of the placenta beginning between Days 20 and 25 of pregnancy, and the number of these cells increases as gestation progresses [58]. The identity of these cells remains to be determined.

OPN expression in uterine LE increases markedly during the peri-implantation period of pigs, but is never observed in uterine LE during the estrous cycle [59]. OPN mRNA is initially induced by conceptus estrogens in discrete regions of the LE juxtaposed to the conceptus just prior to implantation on Day 13, then expands to the entire LE by Day 20 when firm adhesion of conceptus

trophoblast to uterine LE occurs [58]. However, OPN mRNA is not present in pig conceptuses [58,59]. In contrast to mRNA, OPN protein is abundant along the apical surfaces of LE and trophoblast/chorion, but only in areas of direct contact between the uterus and conceptus [58,59]. Remarkably, OPN mRNA and protein are not present in uterine LE and chorion of areolae where the chorion does not attach to LE, but rather forms a “pocket” of columnar epithelial cells that take up and transport secretions of uterine GE into the placental vasculature by fluid phase pinocytosis [114] (Figure 1). OPN levels remain high at this interface throughout pregnancy [59], as do multiple integrin subunits that potentially form heterodimeric receptors that bind OPN [8,84,90].

All experimental and surgical procedures were in compliance with the Guide for Care and Use of Agricultural Animals in Teaching and Research and approved by the Institutional Animal Care and Use Committee of Texas A&M University.

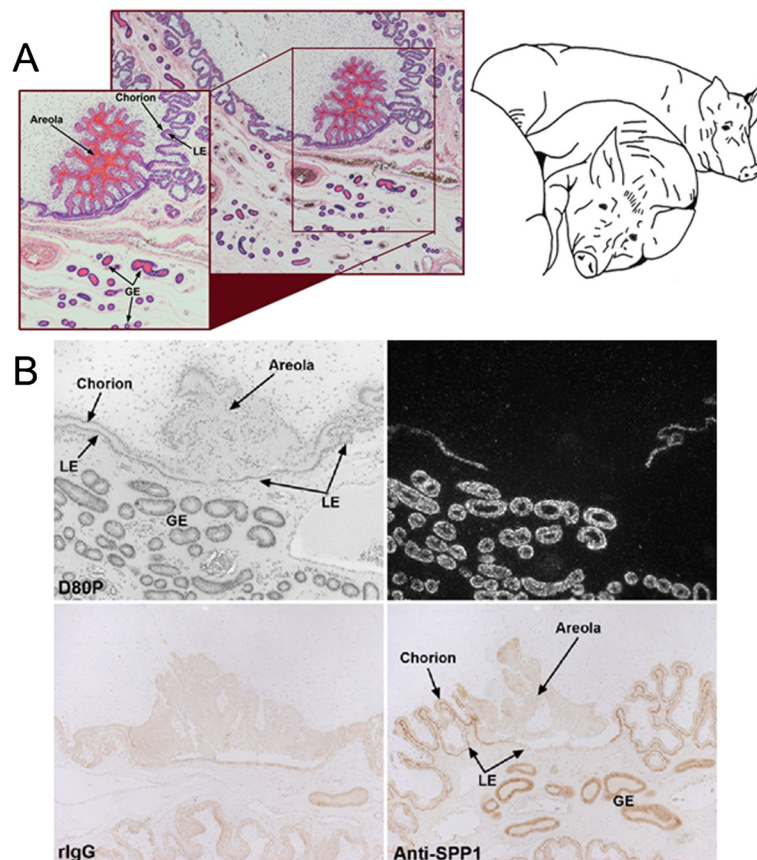


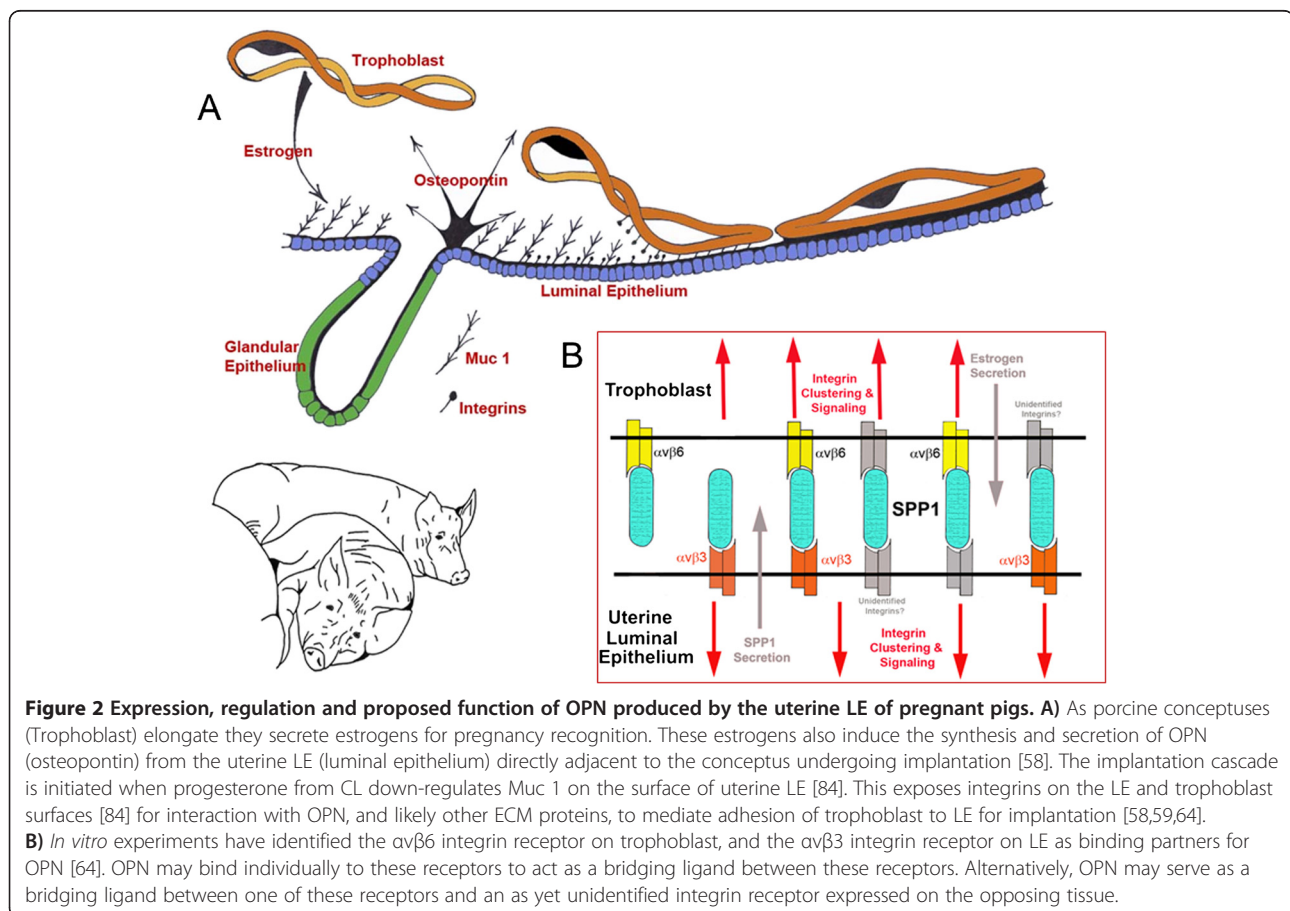
Figure 1 OPN is synthesized and secreted from the luminal epithelium (LE) only at sites of direct attachment of uterus to placenta.

A) H&E stained paraffin embedded thin section of the uterine/placental interface of a Day 80 pregnant gilt illustrating an areola containing histotroph (note the intense red eosin protein staining) secreted by the glandular epithelium (GE). **B)** OPN mRNA (top panels) and protein (bottom panels) is expressed in the uterus of a Day 80 pregnant gilt (expression begins in luminal epithelium (LE) on Day 13, in GE by Day 35, and then in both cell types to term). Note that OPN is not detectable in uterine LE associated with areolae where there is no direct attachment of uterine LE to placental trophoblast/chorion. This precise spatial distribution for OPN expression strongly suggests that it plays a role for attaching uterus to placenta during epitheliochorial placentation.

Affinity chromatography and immunoprecipitation experiments were performed to test whether the integrin subunits α , $\alpha 4$, $\alpha 5$, $\beta 1$, $\beta 3$, $\beta 5$, and $\beta 6$, expressed by porcine trophoblast cells (pTr2) and porcine uterine epithelial (pUE) cells, directly bind OPN. Detergent extracts of surface-biotinylated pig trophoblast (pTr2) and uterine epithelial (pUE) cells were incubated with OPN-Sepharose and the proteins that bound to OPN were eluted with EDTA to chelate cations and release bound integrins. To identify these integrins, immunoprecipitation assays were performed using antibodies that successfully immunoprecipitated integrin subunits from pTr2 or pUE cell lysates. OPN directly bound the $\alpha\beta 6$ integrin heterodimer on pTr2 cells and $\alpha\beta 3$ on ULE cells [64]. OPN binding promoted dose- and integrin-dependent attachment of pTr2 and pUE cells, and stimulated haptotactic pTr2 cell migration, meaning that cells migrated directionally along a physical gradient of nonsoluble OPN [64]. Further, immunofluorescence staining revealed that both OPN and α integrin subunit localized to the apical surface of cells at the interface between uterine LE and conceptus trophoblast at Day 20 of pregnancy. The α integrin subunit staining

pattern revealed large aggregates at the junction between trophoblast and uterine LE, suggesting the formation of OPN-induced *in vivo* focal adhesions at the apical surfaces of both conceptus trophoblast and uterine LE that facilitate conceptus attachment to the uterus for implantation. The $\beta 3$ subunit appeared in aggregates on the apical surface of LE cells, but not trophoblast cells, fitting with affinity chromatography data indicating direct binding of $\alpha\beta 3$ on pUE cells to OPN [64]. Finally, OPN-coated microspheres revealed co-localization of the α integrin subunit and talin to focal adhesions at the apical domain of pTr2 cells *in vitro* [64]. Collectively, results support that OPN binds integrins to stimulate integrin-mediated focal adhesion assembly, attachment, and cytoskeletal force-driven migration of pTr2 cells to promote conceptus implantation in pigs (Figure 2).

In addition to expression in LE during the peri-implantation period, total uterine OPN mRNA increases 20-fold between Days 25 and 85 of gestation due to induction of OPN expression in uterine GE [59]. The initial significant increase in GE is delayed until between Days 30 and 35 when placental growth and placentation are key events in pregnancy in pigs [5]. OPN expression



in GE during later stages of pregnancy is also observed in sheep [115], and a microarray study in rats showed that OPN expression increased 60-fold between Day 0 of the estrous cycle and Day 20 of pregnancy, likely within the decidua [116]. Indeed, OPN is expressed by uterine natural killer (uNK) cells of the mouse decidua [44,45]. Secretions of GE in livestock, and the secretions of decidua in rodents and primates, are critical to support implantation, placentation, and fetal growth and development [117,118]. OPN is also expressed in uterine GE of Day 90 of pseudopregnant pigs, suggesting that maintenance of secretion of progesterone by CL is responsible for expression of OPN in GE [58]. Progesterone also regulates OPN expression in the GE of sheep and rabbits [51,54], as well as OPN synthesis by human Ishikawa cells [56].

However, the involvement of progesterone in the regulation of OPN in uterine GE is complex as indicated by recent analysis of long-term progesterone treatment on the expression of OPN in pigs in the absence of ovarian or conceptus factors. In addition to OPN expression, other established progesterone targets including progesterone receptor (PGR) as an index of progesterone's ability to negatively regulate GE gene expression [119], acid phosphate 5, tartrate resistant (ACP5, commonly referred to as uteroferrin) as an index of progesterone's ability to positively regulate early pregnancy GE gene expression [120], and fibroblast growth factor 7 (FGF7, commonly referred to as keratinocyte growth factor) provide an index of progesterone's ability to positively regulate gene expression in uterine GE beyond the peri-implantation period [121]. Pigs were ovariectomized on Day 12 of the estrous cycle when progesterone secretion from CL is high and treated daily with intramuscular injections of progesterone or vehicle for 28 days [122,123]. As anticipated, PGR mRNA decreased, uteroferrin mRNA increased, and FGF7 mRNA increased in uterine GE of pigs injected with progesterone [123]. Surprisingly, long-term progesterone, in the absence of ovarian and/or conceptus factors, did not induce OPN expression in uterine GE [123]. It is currently hypothesized that the hormonal milieu necessary for the production of individual components of histotroph varies, and may require specific servomechanisms, similar to those for sheep and rabbits, which involve sequential exposure of the pregnant uterus to ovarian, conceptus, and/or uterine factors that include progesterone, estrogens and IFNs [124-126]. Recently OPN expression was compared in placental and uterine tissues supplying a normally sized and the smallest fetus carried by hyperprolific Large White and Meishan gilts. Not only were levels of OPN strikingly different between the two breeds of pigs, but OPN was higher in the LE and GE of uteri surrounding smaller sized fetuses, suggesting OPN may be associated with placental efficiency [127].

OPN expression, regulation and function in the uterus and placenta of ewes

Similar to pigs, the conceptuses of sheep remain free-floating within the uterine lumen as they elongate from spherical blastocysts to conceptuses with a filamentous morphology (Reviewed in [88]). Sheep embryos enter the uterus on Day 3, develop to spherical blastocysts and then, after hatching from the zona pellucida, transform from spherical to tubular and filamentous conceptuses between Days 12 and 15 of pregnancy, with extra-embryonic membranes extending into the contralateral uterine horn between Days 16 and 20. During this period of rapid elongation, the mononuclear trophoblast cells of ovine conceptuses secrete interferon tau between Days 10 and 21 of pregnancy, and implantation begins on Day 16 as trophoblast attaches to the uterine LE [70,88]. The ovine placenta eventually organizes into discrete regions called placentomes that are composed of highly branched placental chorioallantoic villi termed cotyledons which grow rapidly and interdigitate with maternal aglandular endometrial crypts termed caruncles. Approximately 90% of the blood from the uterine artery flows into the placentomes for nutrient transfer from the maternal uterine circulation to the fetus and exchange of gasses between these tissue compartments [128].

The temporal and spatial expression of OPN in the uteri and placentae of sheep is similar to that previously described for the pig, except: 1) unlike in the pig, OPN is not expressed by uterine LE; 2) induction of OPN in uterine GE occurs earlier than in the pig during the peri-implantation period, and expression in the GE is regulated by progesterone; 3) OPN is a prominent component of the stratum compactum stroma; and 4) although large focal adhesions assemble during the peri-implantation period of pigs, they are not observed at the uterine-placental interface until later stages of pregnancy in sheep.

OPN mRNA and protein are present in a small population of cells scattered within the stratum compactum stroma immediately beneath the endometrial LE during the early stages of the estrous cycle and pregnancy in sheep [18]. OPN-producing cells are also present in the allantois of the ovine placenta beginning between Days 20 and 25 of pregnancy and increase in number as gestation progresses [17]. As hypothesized for pigs, these are presumed to be immune cells because Eta-1/OPN is a prominent player in the immune system [15]. In contrast to pigs, in which the OPN-expressing endometrial cells are readily evident in the stratum compactum stroma throughout pregnancy, these cells are difficult to discern in the sheep due to an increase in expression of OPN by stromal cells between Days 20 and 25 gestation [129]. In pregnant mice and primates, OPN in decidualized stroma is considered to be a gene marker for decidualization [130,131]. Decidualization involves transformation of spindle-like fibroblasts into

polygonal epithelial-like cells that are hypothesized to limit conceptus trophoblast invasion through the uterine wall during invasive implantation [118]. Although Mossman [132] and Kellas [133] described decidual cell characteristics in the placentomal crypts of sheep and antelope, their reports were largely ignored, and decidualization was not thought to occur in species with central and noninvasive implantation characteristic of domestic animals. However, endometrial stromal cells do increase in size and become polyhedral in shape in pregnant ewes following conceptus attachment, and the classical decidualization markers desmin and α -smooth muscle actin are expressed in these cells, suggesting that OPN expression in this stromal compartment is part of a uterine decidualization-like response to the conceptus during ovine pregnancy [129]. In contrast, no morphological changes in uterine stroma, nor induction of OPN mRNA and protein, or desmin protein, were detected during porcine pregnancy [129]. One of the primary roles of decidua in invasive implanting species is to restrain conceptus trophoblast invasion to a circumscribed region of the endometrium. Both pigs and sheep have noninvasive implantation, but the extent of conceptus invasion into the endometrium differs between these two species. Pig conceptuses undergo true epitheliochorial placentation in which uterine LE remains morphologically intact throughout pregnancy and the conceptus trophoctoderm simply attaches to the apical surface of uterine LE surface without contacting uterine stromal cells [134]. Synepitheliochorial placentation in sheep involves extensive erosion of the LE due to formation of syncytia with binucleate cells of the trophoctoderm. After Day 19 of pregnancy, conceptus tissue is opposed to, but does not penetrate ovine uterine stroma [135]. Although speculative, differences in stromal expression of OPN between these species suggest that the extent of decidualization is correlated positively with degree of conceptus invasiveness.

In contrast to pigs, OPN is not synthesized by sheep uterine LE, but is nonetheless a component of histotroph secreted from the endometrial GE into the uterine lumen of pregnant ewes as early as Day 13. It is not secreted by uterine GE of cyclic ewes [18,29]. OPN mRNA is detected in some uterine glands by Day 13, and is present in all glands by Day 19 of gestation [18]. Progesterone induces expression of OPN in the endometrial GE, and induction is associated with a loss of PGR in uterine GE. Analysis of uterine flushings from pregnant ewes has identified a 45 kDa fragment of OPN with greater binding affinity for $\alpha\beta 3$ integrin receptor than native 70 kDa [29,51,52,108]. Comparison of the spatial distribution of OPN mRNA and protein by *in situ* hybridization and immunofluorescence analyses of cyclic and pregnant ovine uterine sections has provided significant insight into the physiology of uterine OPN during pregnancy. OPN mRNA increases in the endometrial GE during the peri-implantation period;

however, it is not present in LE or conceptus trophoctoderm [18]. In contrast, immunoreactive OPN protein is present at the apical surfaces of endometrial LE and GE, and on trophoctoderm where the integrin subunits $\alpha\upsilon$, $\alpha 4$, $\alpha 5$, $\beta 1$, $\beta 3$, and $\beta 5$ are expressed constitutively on the apical surfaces of trophoctoderm and endometrial LE and could potentially assemble into several heterodimers that could serve as receptors for OPN including $\alpha\upsilon\beta 3$, $\alpha\upsilon\beta 1$, $\alpha\upsilon\beta 5$, $\alpha 4\beta 1$, and $\alpha 5\beta 1$ heterodimers which [29,52]. These results strongly suggest that OPN is a component of histotroph secreted from GE into the uterine lumen of pregnant ewes in response to progesterone, and that OPN binds integrin receptors expressed on endometrial LE and conceptus trophoctoderm.

Affinity chromatography and immunoprecipitation experiments, similar to those described previously for pigs, determined whether $\alpha\upsilon$, $\alpha 4$, $\alpha 5$, $\beta 1$, $\beta 3$, $\beta 5$, and $\beta 6$ integrins expressed by ovine trophoctoderm cells (oTr1) directly bind OPN. Successful immunoprecipitation of labeled oTr1 integrins occurred with antibodies to $\alpha\upsilon$ and $\beta 3$ integrin subunits, as well as an antibody to the integrin $\alpha\upsilon\beta 3$ heterodimer. Antibody to the $\alpha\upsilon$ integrin subunit also precipitated a β chain, presumed to be the $\beta 3$ integrin subunit, as an antibody to the $\beta 3$ integrin subunit precipitated an α chain at the same relative size as the bands precipitated by an antibody to the $\alpha\upsilon\beta 3$ heterodimer. Thus, the $\alpha\upsilon\beta 3$ integrin on oTr1 cells binds OPN [66]. OPN binding to the $\alpha\upsilon\beta 3$ integrin receptor induced *in vitro* focal adhesion assembly (see Figure 3), a prerequisite for adhesion and migration of trophoctoderm, through activation of: 1) P70S6K via crosstalk between FRAP1/MTOR and MAPK pathways; 2) MTOR, PI3K, MAPK3/MAPK1 (Erk1/2) and MAPK14 (p38) signaling to stimulate trophoctoderm cell migration; and 3) focal adhesion assembly and myosin II motor activity to induce migration of trophoctoderm cells [66]. Collectively, results indicate that OPN binds $\alpha\upsilon\beta 3$ integrin receptor to activate cell signaling pathways that act in concert to mediate adhesion, migration and cytoskeletal remodeling of trophoctoderm cells essential for expansion and elongation of conceptuses and their attachment to uterine LE for implantation (Figure 4).

Focal adhesions, the hallmark of activated integrins, are prominent structures of cells grown in culture; however, they are rarely observed *in vivo*. It is noteworthy that large aggregations of focal adhesion-associated proteins that have been interpreted to be three dimensional focal adhesions are present at the uterine-placental interface of sheep [63]. By day 40 of pregnancy in sheep, the punctate apical surface staining of integrin receptor subunits identified in peri-implantation uterine LE and conceptus trophoctoderm [52] is replaced by scattered large aggregates of $\alpha\upsilon$, $\alpha 4$, $\beta 1$, and $\beta 5$ subunits in interplacentomal LE and trophoctoderm/chorion cells. Integrin aggregates are observed only in gravid uterine horns of

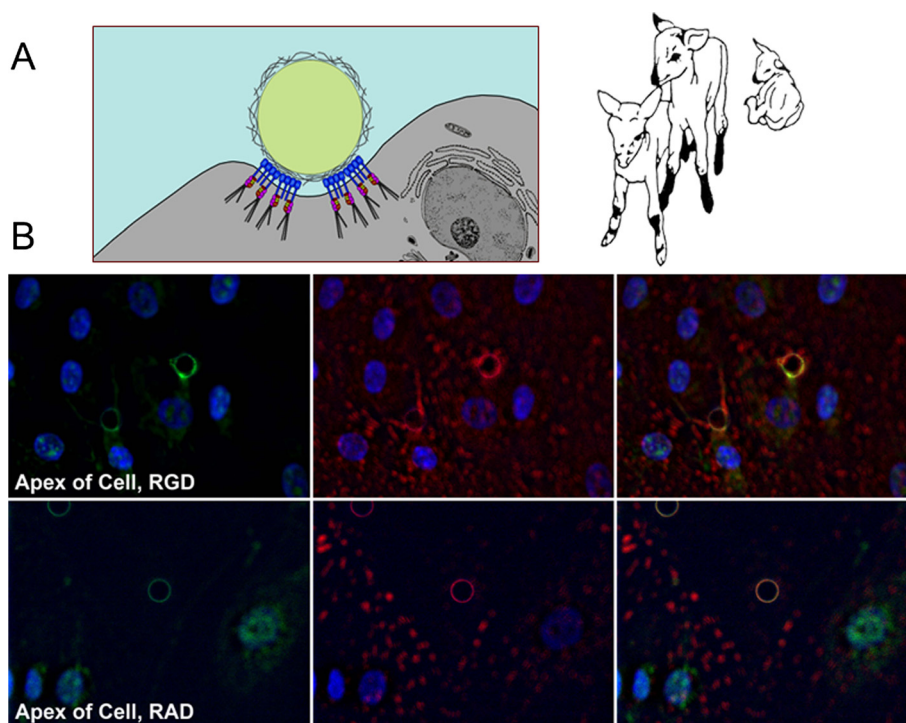
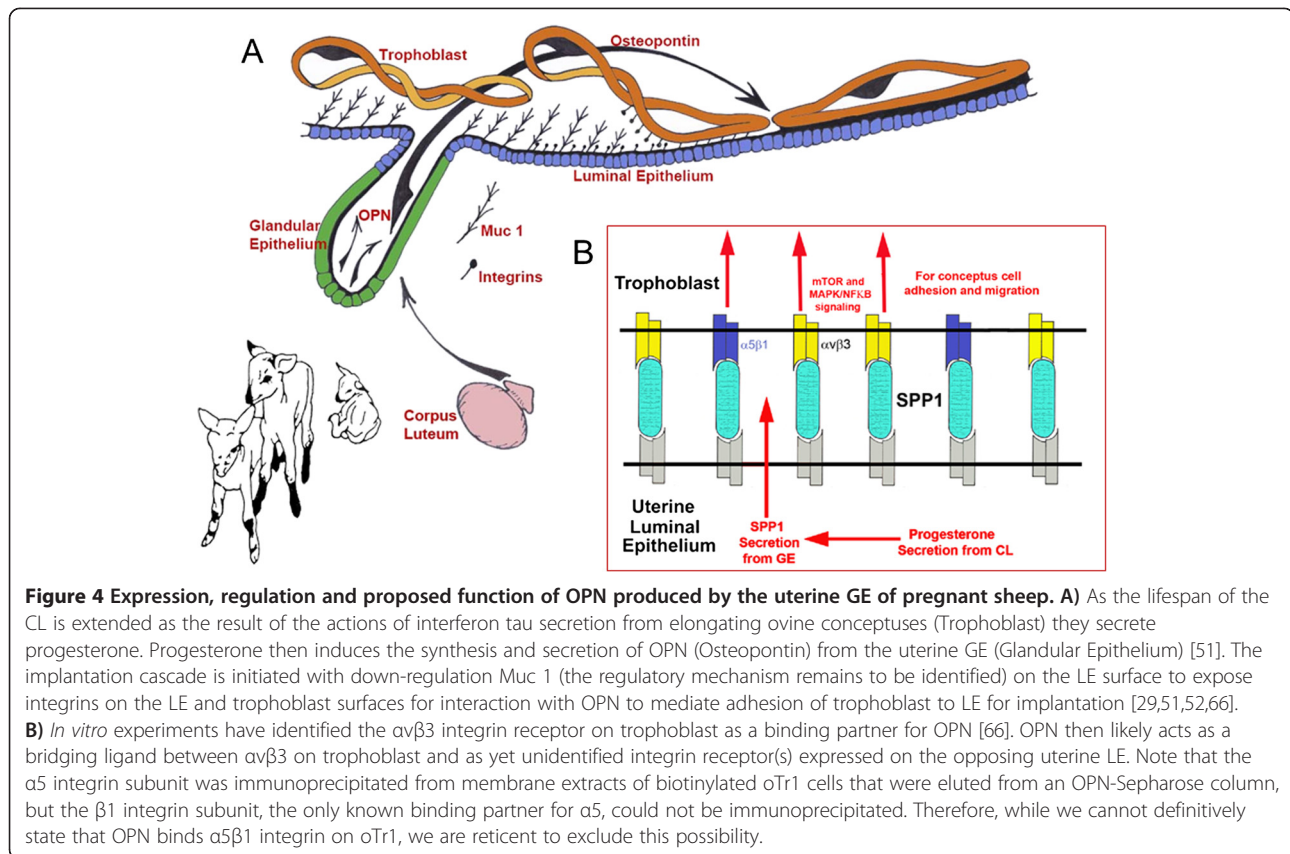


Figure 3 OPN stimulates *in vitro* activation of integrin receptors to form focal adhesions at the apical surface of oTr1 cells. **A)** Cartoon illustrating a polystyrene bead coated with recombinant rat OPN containing an intact RGD integrin binding sequence, and allowed to settle onto a cultured oTr1 cell. Note the illustrated representation of aggregated integrins, indicative of focal adhesion assembly, at the interface between the surface of the bead and the apical membrane of the cell [52,64,66]. **B)** Immunofluorescence co-localization (left panels) to detect the aggregation of αv integrin subunit (right panels) and talin (middle panels), an intracellular component of focal adhesions, around beads coated with recombinant rat OPN containing an intact RGD integrin binding sequence (RGD) or coated with recombinant OPN containing a mutated RAD sequence that does not bind integrins [66]. Optical slice images from the apical plasma membrane of oTr1 cells are shown. Note the apical focal adhesions represented by immunofluorescence co-localization (yellow color) of the integrin αv subunit with talin that results from integrin activation in response to binding of intact OPN on the surface of the bead. No apical focal adhesions were induced by beads coated with mutated OPN as evidenced by lack of integrin αv and talin aggregation around the bead.

unilaterally pregnant sheep, demonstrating a requirement for trophoblast attachment to LE, and aggregates increase in number and size through Day 120 of pregnancy [63]. Interestingly, no accumulation of $\beta 3$ was observed even though ITGB3 is a prominent component of the uterine-placental interface during the peri-implantation period in sheep [52]. In some regions of the interplacental interface, greater subunit aggregation was seen on the uterine side, in other regions it was predominant on the placental side; whereas in some others, both uterine and placental epithelia exhibited prominent focal adhesions. However, by Day 120 of pregnancy, extensive focal adhesions were seen along most of the uterine-placental interface [63]. The placentomes, which provide hematotropic support to the fetus and placenta, exhibited diffuse immunoreactivity for these integrins compared with interplacental regions perhaps due to extensive folding at this interplacental interface [63]. These results suggest that focal adhesion assembly at the uterine-placental interface reflects dynamic adaptation to increasing forces caused by the growing conceptus. Cooperative binding of multiple

integrins to OPN deposited at the uterine-placental interface may form an adhesive mosaic to maintain a tight connection and increased tensile strength and signaling activity between uterine and placental surfaces along regions of epitheliochorial placentation in sheep.

Steady-state levels of OPN mRNA in total ovine endometrium remain constant between Days 20 and 40, increase 40-fold between Days 40 and 100, and remain maximal thereafter [18]. The major source of this OPN is uterine GE which undergoes hyperplasia through Day 50 followed by hypertrophy and maximal production of histotroph after Day 60 [115]. Additionally, immunofluorescence microscopy demonstrated that the secreted 45-kDa OPN cleavage fragment is exclusively, continuously, and abundantly present along the apical surface of uterine LE, on trophoblast, and along the entire uterine-placental interface of both interplacental and placentomal regions through Day 120 of the 147 day ovine pregnancy [115]. These findings definitively localize OPN as a secretory product of the GE to regions of intimate contact between conceptus and uterus, where OPN may influence



fetal/placental development and growth, and mediate communication between placental and uterine tissues to support pregnancy to term.

Increases in OPN from GE are likely influenced by uterine exposure to progesterone, interferon-tau, and placental lactogen which constitute a “servomechanism” that activates and maintains endometrial remodeling, secretory function and uterine growth during gestation. Sequential treatment of ovariectomized ewes with progesterone, interferon tau, placental lactogen, and growth hormone results in GE development similar to that observed during normal pregnancy [126]. Administration of progesterone alone in these experiments induced expression of OPN in GE, and intrauterine infusion of interferon tau and placental lactogen to progesterone-treated ovariectomized ewes increased OPN mRNA levels above those for ewes treated with progesterone alone [126]. An attractive hypothesis for OPN expression in GE is that progesterone interacts with its receptor in GE to down-regulate the progesterone receptor. This removes a progesterone “block” to OPN synthesis, and subsequent increases of OPN expression by GE are augmented by stimulatory effects of placental lactogen. Current studies focus on defining the role(s) of OPN secreted from the uterine GE during later stages of pregnancy.

Conclusions

Research conducted with pigs and sheep has significantly advanced understanding of the role(s) of OPN during implantation through exploitation of 1) the prolonged peri-implantation period of pregnancy when elongating conceptuses are free within the uterine lumen requiring extensive paracrine signaling between conceptus and endometrium, and 2) the protracted and incremental attachment cascade of trophoctoderm to uterine LE during implantation. Although OPN is synthesized in different cell types (LE in pigs, GE in sheep) and is regulated by different hormones (conceptus estrogens in pigs, progesterone in sheep), nonetheless OPN protein localizes to the interface between the uterus and trophoctoderm where it is well placed to serve as a bifunctional bridging ligand between integrins, expressed by uterine LE and conceptus trophoctoderm, to mediate attachment for implantation. It is noteworthy that OPN has been reported to be a prominent component of the uterine-placental environment of other species including primates and rodents, and therefore knowledge gained about the physiology of OPN in sheep and pigs may have significant relevance to human pregnancy. Our understanding of events that underlie successful implantation and placentation across species has been and will likely continue to be advanced by studies of pigs and sheep as biomedical research models.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

All data illustrated in the Figures presented in this review were produced through experiments designed and implemented by the authors, GAJ, RCB and FWB, and members of their laboratories through a collaborative effort. All authors read and approved the final manuscript.

Acknowledgements

Research supported by USDA-NRICGP 98-35203-6337 to F.W.B. and R.C.B., NRSA-DHHS/NIH 1-F32-HDO 8501 O1A1 to G.A.J., and USDA-NRI 2006-35203-17199 to G.A.J.

Author details

¹Department of Veterinary Integrative Biosciences, Texas A&M University, College Station, TX 77843-4458, USA. ²Department of Animal Science, Texas A&M University, College Station, TX 77843, USA.

Received: 6 August 2014 Accepted: 25 November 2014

Published: 17 December 2014

References

1. Roberts RM, Smith GW, Bazer FW, Cibelli J, Seidel GE Jr, Bauman DE, Reynolds LP, Ireland JJ: **Research priorities. Farm animal research in crisis.** *Science* 2009, **324**:468–469.
2. Hynes RO: **Integrins: a family of cell surface receptors.** *Cell* 1987, **48**:549–554.
3. Denhardt DT, Guo X: **Osteopontin: a protein with diverse functions.** *FASEB J* 1993, **7**:1475–1482.
4. Senger DR, Perruzzi CA, Papadopoulos-Sergiou A, Van de Water L: **Adhesive properties of osteopontin: regulation by a naturally occurring thrombin-cleavage in close proximity to the GRGDS cell-binding domain.** *Mol Biol Cell* 1994, **5**:565–574.
5. Butler WT, Ridall AL, McKee MD: **Osteopontin.** In *Principles of Bone Biology*. New York: Academic Press, Inc; 1996:167–181.
6. Sodek J, Ganss B, McKee MD: **Osteopontin.** *Crit Rev Oral Biol Med* 2000, **11**:279–303.
7. Giancotti FG, Ruoslahti E: **Integrin signaling.** *Science* 1999, **285**:1028–1032.
8. Burghardt RC, Johnson GA, Jaeger LA, Ka H, Garlow JE, Spencer TE, Bazer FW: **Integrins and extracellular matrix proteins at the maternal-fetal interface in domestic animals.** *Cells Tissues Organs* 2002, **172**:202–217.
9. Vogel V: **Mechanotransduction involving multimodular proteins: converting force into biochemical signals.** *Annu Rev Biophys Biomol Struct* 2006, **35**:459–488.
10. Geiger B, Spatz JP, Bershadsky AD: **Environmental sensing through focal adhesions.** *Nat Rev Mol Cell Biol* 2009, **10**:21–33. doi:10.1038/nrm2593.
11. Kling D, Fingerle J, Harlan JM: **Inhibition of leukocyte extravasation with a monoclonal antibody to CD18 during formation of experimental intimal thickening in rabbit carotid arteries.** *Arterioscler Thromb* 1992, **12**:997–1007.
12. Aplin JD, Seif MW, Graham RA, Hey NA, Behzad F, Campbell S: **The endometrial cell surface and implantation. Expression of the polymorphic mucin MUC-1 and adhesion molecules during the endometrial cycle.** *Ann N Y Acad Sci* 1994, **30**:103–121.
13. Senger DR, Wirth DF, Hynes RO: **Transformed mammalian cells secrete specific proteins and phosphoproteins.** *Cell* 1979, **16**:885–893.
14. Franzen A, Heinegard D: **Isolation and characterization of two sialoproteins present only in bone calcified matrix.** *Biochem J* 1985, **232**:715–724.
15. Patarca R, Freeman GJ, Singh RP, Wei FY, Durfee T, Blattner F, Ragnier DC, Kozak CA, Mock BA, Morse HC III, Jerrells TR, Cantor H: **Structural and functional studies of the early T lymphocyte activation 1 (Eta-1) gene. Definition of a novel T cell-dependent response associated with genetic resistance to bacterial infection.** *J Exp Med* 1989, **170**:145–161.
16. Fisher LW, Torchia DA, Fohr B, Young MF, Fedarko NS: **Flexible structures of SIBLING proteins, bone sialoprotein and osteopontin.** *Biochem Biophys Res Commun* 2001, **280**:460–465.
17. Johnson GA, Burghardt RC, Bazer FW, Spencer TE: **OPN: roles in implantation and placentation.** *Biol Reprod* 2003, **69**:1458–1471.
18. Johnson GA, Spencer TE, Burghardt RC, Bazer FW: **Ovine osteopontin. I. Cloning and expression of mRNA in the uterus during the peri-implantation period.** *Biol Reprod* 1999, **61**:884–891.
19. Prince CW, Oosawa T, Butler WT, Tomana M, Bhowan AS, Bhowan M, Schrohenloher RE: **Isolation, characterization and biosynthesis of a phosphorylated glycoprotein from rat bone.** *J Biol Chem* 1987, **262**:2900–2907.
20. Nagata T, Todescan R, Goldberg HA, Zhang Q, Sodek J: **Sulphation of secreted phosphoprotein I (SPP-1, osteopontin) is associated with mineralized tissue formation.** *Biochem Biophys Res Commun* 1989, **165**:234–240.
21. Sorensen ES, Hojrup P, Petersen TE: **Post-translational modifications of bovine osteopontin: identification of twenty-eight phosphorylation and three o-glycosylation sites.** *Protein Sci* 1995, **4**:2040–2049.
22. Kaartinen MT, Pirhonen A, Linnala-Kankkunen A, Maenpaa PH: **Cross-linking of osteopontin by tissue transglutaminase increases its collagen binding properties.** *J Biol Chem* 1999, **274**:1729–1735.
23. Agnihotri R, Crawford HC, Haro H, Matrisian LM, Havrda MC, Liaw L: **Osteopontin, a novel substrate for matrix metalloproteinase-3 (stromelysin-1) and matrix metalloproteinase-7 (matrilysin).** *J Biol Chem* 2001, **276**:28261–28267.
24. Senger DR, Perruzzi CA, Papadopoulos A, Tenen DG: **Purification of a human milk protein closely similar to tumor-secreted phosphoproteins and osteopontin.** *Biochim Biophys Acta* 1989, **996**:43–48.
25. Young MF, Kerr JM, Termine JD, Wewer UM, Wang MG, McBride OW, Fisher LW: **cDNA cloning, mRNA distribution and heterogeneity, chromosomal location, and RFLP analysis of human osteopontin (OPN).** *Genomics* 1990, **7**:491–502.
26. Brown LF, Berse B, Van de Water L, Papadopoulos-Sergiou A, Perruzzi CA, Manseau EJ, Dvorak HF, Senger DR: **Expression and distribution of osteopontin in human tissues: widespread association with luminal epithelial surfaces.** *Mol Biol Cell* 1992, **3**:1169–1180.
27. Kohri K, Nomura S, Kitamura Y, Nagata T, Yoshioka K, Iguchi M, Yamate T, Umekawa T, Suzuki Y, Sinohara H, Kurita T: **Structure and expression of the mRNA encoding urinary stone protein (osteopontin).** *J Biol Chem* 1993, **268**:15180–15184.
28. Daiter E, Omigbodun A, Wang S, Walinsky D, Strauss JF III, Hoyer JR, Coutifaris C: **Cell differentiation and endogenous cyclic adenosine 3',5'-monophosphate regulate osteopontin expression in human trophoblasts.** *Endocrinology* 1996, **137**:1785–1790.
29. Johnson GA, Burghardt RC, Spencer TE, Newton GR, Ott TL, Bazer FW: **Ovine osteopontin II. Osteopontin and alpha(v)beta(3) integrin expression in the uterus and conceptus during the peri-implantation period.** *Biol Reprod* 1999, **61**:892–899.
30. Luedtke CC, McKee MD, Cyr DG, Gregory M, Kaartinen MT, Mui J, Hermo L: **Osteopontin expression and regulation in the testis, efferent ducts, and epididymis of rats during postnatal development through to adulthood.** *Biol Reprod* 2002, **66**:1437–1448.
31. Banerjee A, Burghardt R, Johnson G, White F, Ramaiah S: **The temporal expression of osteopontin (SPP-1) in the rodent model of alcoholic steatohepatitis: a potential biomarker.** *Toxicol Pathol* 2006, **34**:373–384.
32. Kato A, Okura T, Hamada C, Miyoshi S, Katayama H, Higaki J, Ito R: **Cell stress induces upregulation of osteopontin via the ERK pathway in type II alveolar epithelial cells.** *PLoS One* 2014, **9**(6):e100106. doi:10.1371/journal.pone.0100106. eCollection 2014.
33. Ashkar S, Weber GF, Panoutsakopoulou V, Sanchirico ME, Jansson M, Zawaideh S, Rittling SR, Denhardt DT, Glimcher MJ, Cantor H: **Eta-1 (osteopontin): an early component of type-1 (cell-mediated) immunity.** *Science* 2000, **287**:860–864.
34. Malyankar UM, Scatena M, Suchland KL, Yun TJ, Clark EA, Giachelli CM: **Osteoprotegerin is an alpha v beta 3-induced, NF-kappa B dependent survival factor for endothelial cells.** *J Biol Chem* 2000, **275**:20959–20962.
35. Hotte SJ, Winquist EW, Stitt L, Wilson SM, Chambers AF: **Plasma osteopontin: associations with survival and metastasis to bone in men with hormone-refractory prostate carcinoma.** *Cancer* 2002, **95**:506–512.
36. Flores M, Norgard M, Heinegard D, Reinholt FP, Andersson G: **RGD-directed attachment of isolated rat osteoclasts to osteopontin, bone sialoprotein, and fibronectin.** *Exp Cell Res* 1992, **201**:526–530.
37. Butler WT: **The nature and significance of osteopontin.** *Connect Tissue Res* 1989, **23**:123–136.
38. Nabel G, Fresno M, Chessman A, Cantor H: **Use of cloned populations of mouse lymphocytes to analyze cellular differentiation.** *Cell* 1981, **23**:19–28.
39. Hwang S, Lopez CA, Heck DE, Gardner CR, Laskin DL, Laskin JD, Denhardt DT: **Osteopontin inhibits induction of nitric oxide synthase gene expression by inflammatory mediators in mouse kidney epithelial cells.** *J Biol Chem* 1994, **269**:711–715.

40. Leibson HJ, Marrack P, Kappler JW: **B cell helper factors. I. Requirement for both interleukin 2 and another 40,000 mol wt factor.** *J Exp Med* 1981, **154**:1681–1693.
41. Hruska KA, Rolnick F, Huskey M, Alvarez U, Cheresch D: **Engagement of the osteoclast integrin $\alpha\text{v}\beta 3$ by osteopontin stimulates phosphatidylinositol 3-hydroxy kinase activity.** *Endocrinology* 1995, **136**:2984–2992.
42. McKee MD, Nanci A, Khan SR: **Ultrastructural immunodetection of osteopontin and osteocalcin as major matrix components of renal calculi.** *J Bone Miner Res* 1995, **10**:1913–1929.
43. Nomura S, Wills AJ, Edwards DR, Heath JK, Hogan BLM: **Developmental expression of 2ar (osteopontin) and SPARC (osteonectin) RNA as revealed by in situ hybridization.** *J Cell Biol* 1988, **106**:441–450.
44. White FJ, Burghardt RC, Croy BA, Johnson GA: **Osteopontin is expressed by endometrial macrophages and decidual natural killer cells during mouse pregnancy [abstract].** *Biol Reprod* 2005, **73**(Suppl 1):155.
45. Herington JL, Bany BM: **The conceptus increases secreted phosphoprotein 1 gene expression in the mouse uterus during the progression of decidualization mainly due to its effects on uterine natural killer cells.** *Reproduction* 2007, **133**:1213–1221.
46. Lessey BA, Castelbaum AJ, Buck CA, Lei Y, Yowell CW, Sun J: **Further characterization of endometrial integrins during the menstrual cycle and in pregnancy.** *Fertil Steril* 1994, **62**:497–506.
47. Lessey BA, Ilesanmi AO, Lessey MA, Riben M, Harris JE, Chwalisz K: **Luminal and glandular endometrial epithelium express integrins differentially throughout the menstrual cycle: implications for implantation, contraception and infertility.** *Am J Reprod Immunol* 1996, **35**:195–204.
48. Lessey BA, Yeh I, Castelbaum AJ, Fritz MA, Ilesanmi AO, Korzeniowski P, Sun J, Chwalisz K: **Endometrial progesterone receptors and markers of uterine receptivity in the window of implantation.** *Fertil Steril* 1996, **65**:477–483.
49. Lessey BA, Damjanovich L, Coutifaris C, Castelbaum A, Albelda SM, Buck CA: **Integrin adhesion molecules in the human endometrium. Correlation with the normal and abnormal menstrual cycle.** *J Clin Invest* 1992, **90**:188–195.
50. Lessey BA, Castelbaum AJ, Sawin SW, Sun J: **Integrins as markers of uterine receptivity in women with primary unexplained infertility.** *Fertil Steril* 1995, **63**:535–542.
51. Johnson GA, Spencer TE, Burghardt RC, Taylor KM, Gray CA, Bazer FW: **Progesterone modulation of osteopontin gene expression in the ovine uterus.** *Biol Reprod* 2000, **62**:1315–1321.
52. Johnson GA, Bazer FW, Jaeger LA, Ka H, Garlow JE, Pfarrer C, Spencer TE, Burghardt RC: **Muc-1, integrin and osteopontin expression during the implantation cascade in sheep.** *Biol Reprod* 2001, **65**:820–828.
53. Gray CA, Burghardt RC, Johnson GA, Bazer FW, Spencer TE: **Evidence that absence of endometrial gland secretions in uterine gland knockout ewes compromises conceptus survival and elongation.** *Reproduction* 2002, **124**:289–300.
54. Illera MJ, Cullinan E, Gui Y, Yuan L, Beyler SA, Lessey BA: **Blockade of the $\alpha\text{v}\beta 3$ integrin adversely affects implantation in the mouse.** *Biol Reprod* 2000, **62**:1285–1290.
55. Illera MJ, Lorenzo PL, Gui YT, Beyler SA, Apparao KBC, Lessey BA: **A role for $\alpha\text{v}\beta 3$ integrin during implantation in the rabbit model.** *Biol Reprod* 2003, **68**:766–771.
56. Apparao KB, Murray MJ, Fritz MA, Meyer WR, Chambers AF, Truong PR, Lessey BA: **Osteopontin and its receptor $\alpha\text{v}\beta 3$ integrin are coexpressed in the human endometrium during the menstrual cycle but regulated differentially.** *J Clin Endocrinol Metab* 2001, **86**:4991–5000.
57. Apparao KBC, Illera MJ, Beyler SA, Olson GE, Osteen KG, Corjay MH, Boggess K, Lessey BA: **Regulated expression of osteopontin in the peri-implantation rabbit uterus.** *Biol Reprod* 2003, **68**:1484–1490.
58. White FJ, Ross JW, Joyce MM, Geisert RD, Burghardt RC, Johnson GA: **Steroid regulation of cell specific secreted phosphoprotein 1 (osteopontin) expression in the pregnant porcine uterus.** *Biol Reprod* 2005, **73**:1294–1301.
59. Garlow JE, Ka H, Johnson GA, Burghardt RC, Jaeger LA, Bazer FW: **Analysis of osteopontin at the maternal-placental interface in pigs.** *Biol Reprod* 2002, **66**:718–725.
60. Carson DD, Lagow E, Thatthiah A, Al-Shami R, Farach-Carson MC, Vernon M, Yuan L, Fritz MA, Lessey B: **Changes in gene expression during the early to mid-luteal (receptive phase) transition in human endometrium detected by high-density microarray screening.** *Mol Hum Reprod* 2002, **8**:871–879.
61. Kao LC, Tulac S, Lobo S, Imani B, Yang JP, Germeyer A, Osteen K, Taylor RN, Lessey BA, Giudice LC: **Global gene profiling in human endometrium during the window of implantation.** *Endocrinology* 2002, **143**:2119–2138.
62. Mirkin S, Arslan M, Churikov D, Corica A, Diaz JJ, Williams S, Bocca S, Oehninger S: **In search of candidate genes critically expressed in the human endometrium during the window of implantation.** *Hum Reprod* 2005, **20**:2104–2117.
63. Burghardt RC, Burghardt JR, Taylor JD II, Reeder AT, Nguyen BT, Spencer TE, Johnson GA: **Enhanced focal adhesion assembly reflects increased mechanosensation and mechanotransduction along the maternal/conceptus interface during pregnancy in sheep.** *Reproduction* 2009, **137**:583–593.
64. Erikson DW, Burghardt RC, Bayless KJ, Johnson GA: **Secreted phosphoprotein 1 (SPP1, osteopontin) binds to integrin $\alpha\text{v}\beta 6$ on porcine trophoblast cells and integrin $\alpha\text{v}\beta 3$ on uterine luminal epithelial cells, and promotes trophoblast cell adhesion and migration.** *Biol Reprod* 2009, **81**:814–825.
65. Massuto DA, Kneese EC, Johnson GA, Hooper NH, Burghardt RC, Ing NH, Jaeger LA: **Transforming growth factor beta (TGF β) signaling is activated during porcine implantation: Proposed role for latency associated peptide-integrins at the conceptus-maternal interface.** *Reproduction* 2009, **139**:465–478.
66. Kim J, Erikson DW, Burghardt RC, Spencer TE, Wu G, Bayless KJ, Johnson GA, Bazer FW: **Secreted phosphoprotein 1 binds integrins to initiate multiple cell signaling pathways, including FRAP1/mTOR, to support attachment and force-generated migration of trophoblast cells.** *Matrix Biol* 2010, **29**:369–382.
67. Kang YJ, Forbes K, Carver J, Aplin JD: **The role of the osteopontin-integrin $\alpha\text{v}\beta 3$ interaction at implantation: functional analysis using three different in vitro models.** *Hum Reprod* 2014, **29**:739–749.
68. Frank M, Bazer FW, Thatcher WW, Wilcox CJ: **A study of prostaglandin F 2α as the luteolysin in swine: III. Effects of estradiol valerate on prostaglandin F, progesterone, estrone and estradiol concentrations in the utero-ovarian vein of nonpregnant gilts.** *Prostaglandins* 1977, **14**:1183–1196.
69. Bazer FW, Thatcher WW: **Theory of maternal recognition of pregnancy in swine based on estrogen controlled endocrine versus exocrine secretion of prostaglandin F 2α by the uterine endometrium.** *Prostaglandins* 1977, **14**:397–400.
70. Godkin JD, Bazer FW, Thatcher WW, Roberts RM: **Proteins released by cultured day 15–16 conceptuses prolong luteal maintenance when introduced into the uterine lumen of cyclic ewes.** *J Reprod Fertil* 1984, **71**:57–64.
71. Vallet JL, Bazer FW, Fliss MFV, Thatcher WW: **Effect of ovine conceptus secretory proteins and purified ovine trophoblast protein-1 on interoestrous interval and plasma concentrations of prostaglandins F 2α and E 2 and of 13,14 dihydro-15-keto prostaglandin F 2α in cyclic ewes.** *J Reprod Fertil* 1988, **84**:493–504.
72. Ashworth CJ, Bazer FW: **Changes in ovine conceptus and endometrial function following asynchronous embryo transfer or administration of progesterone.** *Biol Reprod* 1989, **40**:425–433.
73. Gray CA, Taylor KM, Ramsey WS, Hill JR, Bazer FW, Bartol FF, Spencer TE: **Endometrial glands are required for preimplantation conceptus elongation and survival.** *Biol Reprod* 2001, **64**:1608–1613.
74. Burton GJ, Watson AL, Hempstock J, Skepper JN, Jauniaux E: **Uterine glands provide histotrophic nutrition for the human fetus during the first trimester of pregnancy.** *J Clin Endocrinol Metab* 2002, **87**:2954–2959.
75. Glasser SR, Mulholland J: **Receptivity is a polarity dependent special function of hormonally regulated uterine epithelial cells.** *Microsc Res Tech* 1993, **25**:106–120.
76. Denker HW: **Implantation: a cell biological paradox.** *J Exp Zool* 1993, **266**:541–558.
77. Bazer FW, First NL: **Pregnancy and parturition.** *J Anim Sci* 1983, **57**(Suppl 2):425–460.
78. Johnson GA, Bazer FW, Burghardt RC, Spencer TE, Wu G, Bayless KJ: **Conceptus-uterus interactions in pigs: endometrial gene expression in response to estrogens and interferons from conceptuses.** *Soc Reprod Fertil* 2009, **66**(Suppl):321–332.
79. Flint APF, Saunders PTK, Ziecik AJ: **Blastocyst-endometrium interactions and their significance in embryonic mortality.** In *Control of Pig Reproduction*. Edited by Cole DJA, Foxcroft GR. London: Butterworth Scientific; 1982:253–275.

80. Jainudeen MR, Hafez ESE: **Reproductive failure in females.** In *Reproduction in Farm Animals*. Edited by Hafez ESE, Lea F, Hafez ESE, Lea, Febiger. Philadelphia: Wiley-Blackwell; 1987:399–422.
81. Bazer FW, Johnson GA, Spencer TE: **Growth and Development: (1) Mammalian conceptus peri-implantation period; and (2) Mammalian pre-implantation embryo.** In *Encyclopedia of Animal Science*. Edited by Bell WGP. New York: Marcel Dekker; 2005:555–558.
82. Cross JC, Werb Z, Fisher SJ: **Implantation and the placenta: key pieces of the development puzzle.** *Science* 1994, **266**:1508–1518.
83. Carson DD, Bagchi I, Dey SK, Enders AC, Fazleabas AT, Lessey BA, Yoshinaga K: **Embryo implantation.** *Dev Biol* 2000, **223**:217–237.
84. Bowen JA, Bazer FW, Burghardt RC: **Spatial and temporal analyses of integrin and Muc-1 expression in porcine uterine epithelium and trophoctoderm in vivo.** *Biol Reprod* 1996, **55**:1098–1106.
85. Aplin JD, Meseguer M, Simon C, Ortiz ME, Croxatto H, Jones CJ: **MUC1, glycans and the cell-surface barrier to embryo implantation.** *Biochem Soc Trans* 2001, **29**:153–156.
86. Kimber SJ, Illingworth IM, Glasser SR: **Expression of carbohydrate antigens in the rat uterus during early pregnancy and after ovariectomy and steroid replacement.** *J Reprod Fertil* 1995, **103**:75–87.
87. Kimber SJ, Spanswick C: **Blastocyst implantation: the adhesion cascade.** *Semin Cell Dev Biol* 2000, **11**:77–92.
88. Spencer TE, Johnson GA, Bazer FW, Burghardt RC: **Implantation mechanisms: insights from the sheep.** *Reproduction* 2004, **128**:656–668.
89. Ruoslahti E, Pierschbacher MD: **New perspectives in cell adhesion: RGD and integrins.** *Science* 1987, **238**:491–497.
90. Burghardt RC, Bowen JA, Newton GR, Bazer FW: **Extracellular matrix and the implantation cascade in pigs.** *J Reprod Fertil* 1997, **52**(Suppl):151–164.
91. Lessey BA: **Adhesion molecules and implantation.** *J Reprod Immunol* 2002, **55**:101–112.
92. Joyce MM, Gonzalez JF, Lewis S, Woldeesenbet S, Burghardt RC, Newton GR, Johnson GA: **Caprine uterine and placental osteopontin expression is distinct among epitheliochorial implanting species.** *Placenta* 2005, **26**:160–170.
93. White FJ, Burghardt RC, Hu J, Joyce MM, Spencer TE, Johnson GA: **Secreted phosphoprotein 1 (osteopontin) is expressed by stromal macrophages in cyclic and pregnant endometrium of mice, but is induced by estrogen in luminal epithelium during conceptus attachment for implantation.** *Reproduction* 2006, **132**:919–929.
94. Craig AM, Smith JH, Denhardt DT: **OPN, a transformation-associated cell adhesion phosphoprotein, is induced by 12-O-tetradecanoylphorbol 13-acetate in mouse epidermis.** *J Biol Chem* 1989, **264**:9682–9689.
95. Craig AM, Denhardt DT: **The murine gene encoding secreted phosphoprotein 1 (OPN): promoter structure, activity, and induction in vivo by estrogen and progesterone.** *Gene* 1991, **100**:163–171.
96. Singh K, Balligand JL, Fischer TA, Smith TW, Kelly RA: **Glucocorticoids increase OPN expression in cardiac myocytes and microvascular endothelial cells. Role in regulation of inducible nitric oxide synthase.** *J Biol Chem* 1995, **270**:28471–28478.
97. Omigbodun A, Ziolkiewicz P, Tessler C, Hoyer JR, Coutifaris C: **Progesterone regulates OPN expression in human trophoblasts: a model of paracrine control in the placenta?** *Endocrinology* 1997, **138**:4308–4315.
98. Safran JB, Butler WT, Farach-Carson MC: **Modulation of OPN post-translational state by 1, 25-(OH)²-vitamin D₃. Dependence on Ca²⁺ influx.** *J Biol Chem* 1998, **273**:29935–29941.
99. Denhardt DT, Noda M, O'Regan AW, Pavlin D, Berman JS: **OPN as a means to cope with environmental insults: regulation of inflammation, tissue remodeling, and cell survival.** *J Clin Invest* 2001, **107**:1055–1061.
100. Giachelli CM, Steitz S: **OPN: a versatile regulator of inflammation and biomineralization.** *Matrix Biol* 2000, **19**:615–622.
101. Hu DD, Hoyer JR, Smith JW: **Ca²⁺ suppresses cell adhesion to OPN by attenuating binding affinity for integrin alpha v beta 3.** *J Biol Chem* 1995, **270**:9917–9925.
102. Xuan JW, Hota C, Shigeyama Y, D'Errico JA, Somerman MJ, Chambers AF: **Site-directed mutagenesis of the arginine-glycine-aspartic acid sequence in OPN destroys cell adhesion and migration functions.** *J Cell Biochem* 1995, **57**:680–690.
103. Hu DD, Lin EC, Kovach NL, Hoyer JR, Smith JW: **A biochemical characterization of the binding of OPN to integrins alpha v beta 1 and alpha v beta 5.** *J Biol Chem* 1995, **270**:26232–26238.
104. Denda S, Reichardt LF, Muller U: **Identification of OPN as a novel ligand for the integrin alpha8 beta1 and potential roles for this integrin-ligand interaction in kidney morphogenesis.** *Mol Biol Cell* 1998, **9**:1425–1435.
105. Bayless KJ, Meiningner GA, Scholtz JM, Davis GE: **OPN is a ligand for the alpha4beta1 integrin.** *J Cell Sci* 1998, **111**(Pt 9):1165–1174.
106. Smith LL, Giachelli CM: **Structural requirements for alpha 9 beta 1-mediated adhesion and migration to thrombin-cleaved OPN.** *Exp Cell Res* 1998, **242**:351–360.
107. Bayless KJ, Davis GE: **Identification of dual alpha 4beta1 integrin binding sites within a 38 amino acid domain in the N-terminal thrombin fragment of human OPN.** *J Biol Chem* 2001, **276**:13483–13489.
108. Senger DR, Perruzzi CA: **Cell migration promoted by a potent GRGDS-containing thrombin-cleavage fragment of OPN.** *Biochim Biophys Acta* 1996, **1314**:13–24.
109. Weber GF, Ashkar S, Glimcher MJ, Cantor H: **Receptor-ligand interaction between CD44 and OPN (Eta-1).** *Science* 1996, **271**:509–512.
110. Goldsmith HL, Labrosse JM, McIntosh FA, Maenpaa PH, Kaartinen MT, McKee MD: **Homotypic interactions of soluble and immobilized OPN.** *Ann Biomed Eng* 2002, **30**:840–850.
111. Bazer FW, Johnson GA: **Pig blastocyst-uterine interactions.** *Differentiation* 2014, **87**:52–65.
112. Dantzer V: **Scanning electron microscopy of exposed surfaces of the porcine placenta.** *Acta Anat* 1984, **118**:96–106.
113. King AH, Jiang Z, Gibson JP, Haley CS, Archibald AL: **Mapping quantitative trait loci affecting female reproductive traits on porcine chromosome 8.** *Biol Reprod* 2003, **68**:21722179.
114. Renegar RH, Bazer FW, Roberts RM: **Placental transport and distribution of uteroferrin in the fetal pig.** *Biol Reprod* 1982, **27**:1247–1260.
115. Johnson GA, Burghardt RC, Joyce MM, Spencer TE, Bazer FW, Gray CA, Pfarrer C: **Osteopontin is synthesized by uterine glands and a 45-kDa cleavage fragment is localized at the uterine-placental interface throughout ovine pregnancy.** *Biol Reprod* 2003, **69**:92–98.
116. Girotti M, Zingg HH: **Gene expression profiling of rat uterus at different stages of parturition.** *Endocrinology* 2003, **144**:2254–2265.
117. Roberts RM, Bazer FW: **The functions of uterine secretions.** *J Reprod Fertil* 1988, **82**:875–892.
118. Irwin JC, Giudice LC: **Decidua.** In *Encyclopedia of Reproduction*. Edited by Knobil E, Neill JD. New York: Academic Press; 1999:823–835.
119. Ka H, Al-Ramadan S, Erikson DW, Johnson GA, Burghardt RC, Spencer TE, Jaeger LA, Bazer FW: **Regulation of fibroblast growth factor 7 expression in the pig uterine endometrium by progesterone and estradiol.** *Biol Reprod* 2007, **77**:172–180.
120. Schlosnagle DC, Bazer FW, Tsibris JCM, Roberts RM: **An iron-containing phosphatase induced by progesterone in the uterine fluids of pigs.** *J Biol Chem* 1974, **249**:7574–7579.
121. Ka H, Jaeger LA, Johnson GA, Spencer TE, Bazer FW: **Keratinocyte growth factor expression is up-regulated by estrogen in porcine uterine endometrium and it functions in trophoctodermal cell proliferation and differentiation.** *Endocrinology* 2001, **142**:2303–2310.
122. Bailey DW, Dunlap KA, Frank JW, Erikson DW, White BG, Bazer FW, Burghardt RC, Johnson GA: **Effects of long-term progesterone on developmental and functional aspects of porcine uterine epithelia: progesterone alone does not support glandular development of pregnancy.** *Reproduction* 2010, **140**:583–594.
123. Bailey DW, Dunlap KL, Erikson DW, Patel A, Bazer FW, Burghardt RC, Johnson GA: **Effects of long-term progesterone exposure on porcine uterine gene expression: progesterone alone does not induce secreted phosphoprotein 1 (osteopontin) in glandular epithelium.** *Reproduction* 2010, **140**:595–604.
124. Chilton BS, Mani SK, Bullock DW: **Servomechanism of prolactin and progesterone in regulating uterine gene expression.** *Mol Endocrinol* 1988, **2**:1169–1175.
125. Young KH, Kraeling RR, Bazer FW: **Effect of pregnancy and exogenous ovarian steroids on endometrial prolactin receptor ontogeny and uterine secretory response in pigs.** *Biol Reprod* 1990, **43**:592–599.
126. Spencer TE, Gray CA, Johnson GA, Taylor KM, Gertler A, Gootwine E, Ott TL, Bazer FW: **Effects of recombinant ovine interferon tau, placental lactogen, and growth hormone on the ovine uterus.** *Biol Reprod* 1999, **61**:1409–1418.
127. Hernandez SC, Hogg CO, Billon Y, Sanchez MP, Bidanel JP, Haley CS, Archibald AL, Ashworth CJ: **Secreted phosphoprotein 1 expression in endometrium and placental tissues of hyperprolific large white and meishan gilts.** *Biol Reprod* 2013, **88**:120–126.
128. Caton D, Pendergast JF, Bazer FW: **Uterine blood flow: periodic fluctuations of its rate during pregnancy.** *Am J Physiol* 1983, **245**:R850–R852.

129. Johnson GA, Burghardt RC, Joyce MM, Spencer TE, Bazer FW, Pfarrer C, Gray CA: **Osteopontin expression in uterine stroma indicates a decidualization-like differentiation during ovine pregnancy.** *Biol Reprod* 2003, **68**:1951–1958.
130. Waterhouse P, Parhar RS, Guo X, Lala PK, Denhardt DT: **Regulated temporal and spatial expression of the calcium-binding proteins calyculin and OPN (osteopontin) in mouse tissues during pregnancy.** *Mol Reprod Dev* 1992, **32**:315–323.
131. Fazleabas AT, Bell SC, Fleming S, Sun J, Lessey BA: **Distribution of integrins and the extracellular matrix proteins in the baboon endometrium during the menstrual cycle and early pregnancy.** *Biol Reprod* 1997, **56**:348–356.
132. Mossman HW: **Comparative morphogenesis of the foetal membranes and accessory uterine structures.** *Contrib Embryol* 1937, **26**:129–246.
133. Kellas LM: **The placenta and foetal membranes of the antelope *Ourebia ourebi* (Zimmermann).** *Acta Anat* 1966, **64**:390–445.
134. Bjorkman N: **Fine structure of the fetal-maternal area of exchange in the epitheliochorial and endotheliochorial types of placentation.** *Acta Anat* 1973, **61**(suppl):11–22.
135. Boshier DP: **A histological and histochemical examination of implantation and early placentome formation in sheep.** *J Reprod Fertil* 1969, **19**:51–61.

doi:10.1186/2049-1891-5-56

Cite this article as: Johnson *et al.*: Osteopontin: a leading candidate adhesion molecule for implantation in pigs and sheep. *Journal of Animal Science and Biotechnology* 2014 **5**:56.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

