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Comparison of High-Intensity vs. High-Volume Resistance Training on the BDNF Response to Exercise

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1	Comparison of High Intensity versus High Volume Resistance Training on the BDNF
2	Response to Exercise
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22 Abstract

23 This study compared the acute and chronic response of circulating plasma brain-derived 24 neurotrophic factor (BDNF) to high-intensity low-volume (HI) and low-intensity high volume 25 (HV) resistance training. Twenty experienced resistance trained men $(23.5\pm2.6 \text{ y}, 1.79\pm0.05 \text{ m},$ 26 75.7±13.8 kg) volunteered for this study. Prior to the resistance training program (PRE), 27 participants performed an acute bout of exercise using either the HI (3-5 reps; 90% of one 28 repetition maximum [1RM]) or HV (10-12 reps; 70% 1RM) training paradigm. The acute 29 exercise protocol was repeated following 7-weeks of training (POST). Blood samples were 30 obtained at rest (BL), immediately- (IP), 30-min (30P) and 60-min (60P) post exercise at PRE 31 and POST. A 3-way repeated measure ANOVA was used to analyze acute changes in BDNF 32 concentrations during HI and HV resistance exercise, and the effect of 7-weeks of training. No 33 training x time x group interaction in BDNF was noted (p=0.994). Significant main effects for 34 training (p=0.050) and time (p<0.001) in BDNF were observed. Significant elevations in BDNF 35 concentrations were seen from BL at IP (p=0.001), 30P (p<0.001), and 60P (p<0.001) in both HI 36 and HV combined during PRE and POST. BDNF concentrations were also observed to increase 37 from PRE to POST when collapsed across groups and time. No significant group x training interaction (p=0.342), training (p=0.105), or group (p=0.238) effect were noted in the BDNF 38 39 area under the curve response. Results indicate that BDNF concentrations are increased after an 40 acute bout of resistance exercise, regardless of training paradigm, and are further increased during a 7-week training program in experienced lifters. 41

42 **Key Words:** Neurotrophin, resistance exercise, muscle, training status

44 New and Noteworthy

There have been a number of investigations examining the BDNF response to exercise, however our understanding of changes in the BDNF response to resistance training has been primarily limited to frail, older adults. This is the first study that has compared two different resistance training paradigms in experienced, resistance trained adults and have demonstrated that training, independent of resistance training paradigm, can modify the BDNF response to an acute bout of resistance exercise.

52 Introduction

53 Brain-derived neurotrophic factor (BDNF) is a neurotrophin of the nerve-growth factor 54 protein family, whose downstream effects are mediated through tropomyosin-related kinase 55 (Trk) receptors (38). BDNF is present throughout the nervous system, and a crucial mediator in 56 the formation of neuronal circuits throughout the brain where it promotes neuronal survival, 57 neurite outgrowth, and synaptogenesis (12). BDNF associated adaptations to hippocampal 58 architecture have been associated with positive changes in memory and learning (11, 30, 31, 41) 59 and has been reported to ameliorate the response to stressful stimuli (20, 22). Despite its notable 60 role in the central nervous system, expression of BDNF and its high affinity receptor TrkB are 61 broad, being found in skeletal muscle, cardiac, liver, and adipose cells (26). While its role in the 62 skeletal muscle is less clear, increased expression of skeletal muscle BDNF has been shown to 63 increase fat oxidation in an AMP-activated protein kinase-dependent mechanism (28). However, 64 BDNF synthesized in the skeletal muscle does not appear to contribute to systemic circulating levels, but rather acts in a paracrine or autocrine fashion (28). Preliminary data indicates that the 65 66 brain is the primary source, providing approximately 80%, of circulating plasma BDNF in 67 response to exercise (34).

Elevations in circulating BDNF concentrations have been demonstrated following an acute bout of both aerobic and resistance exercise (13, 27, 35, 43). A 4-fold increase in resting BDNF secretion rates (58 ng·100g⁻¹·min⁻¹ to 206 ng·100g⁻¹·min⁻¹) were reported following three months of endurance training at ~65% of VO_{2max} (37). In addition, increases in circulating BDNF concentrations observed during exercise and in the recovery period appear to be associated with both duration and intensity of exercise (4, 13, 27, 35, 37). Ferris and colleagues (13) demonstrated that 30 minutes of cycling at 10% above subjects' ventilatory threshold resulted in higher concentrations of BDNF than when cycling at 20% below. However, the total amount of work performed was greater during the higher intensity protocol. In addition, Cho and colleagues (4) reported greater elevations in circulating BDNF concentrations during a maximal aerobic capacity test with greater duration of exercise. These data suggest that the greater volume, or the combination of greater volume and/or higher intensity, may have provided the stimulus for a greater BDNF response to exercise.

81 The vast majority of research investigating the BDNF response to physical activity has 82 primarily examined endurance exercise. The BDNF response during resistance exercise has not 83 been studied to the same extent. In a limited number of investigations, some investigators have 84 reported no change in the acute BDNF response to resistance exercise (18, 35), while others have 85 suggested that training can augment the BDNF response to resistance exercise in previously 86 untrained college-aged men (43). Most of the studies examining the BDNF response to resistance 87 training have been performed in older and frail adults. These studies have generally reported that 88 limited resistance training (3 times per week for 10 - 12 weeks) may be sufficient to increase 89 BDNF concentrations in older adults (5, 15, 31). The majority of these studies have examined 90 serum BDNF, which includes platelets that store and release BDNF in response to exercise. In 91 consideration that plasma is free of platelets, and BDNF turnover in the plasma is approximately 92 6 minutes (33), changes in plasma BDNF would be more indicative of an acute response to a 93 training stress. Furthermore, $\sim 60-80\%$ of plasma BDNF is thought to be produced in the brain 94 (34). To the best of our knowledge no studies have been performed on the plasma BDNF 95 response to resistance training in experienced, resistance trained men. Thus, the primary purpose 96 of this investigation was to compare the acute BDNF response to two common resistance training paradigms before and following 7-weeks of training in experienced, resistance trained 97

98 adults. We hypothesize that resistance exercise will increase BDNF concentrations to both

99 training protocols. In addition, we further hypothesize that 7-weeks of training will augment the

100 BDNF response to exercise.

101 Methods

102 Experimental Design

103 Prior to the onset of the study, all participants were required to complete a 2-week preparatory 104 base resistance training program. Subsequently, participants were then randomly assigned to one 105 of two training groups: a high intensity, low-volume training group (HI; n = 10; 22.6 ± 2.3 years; 106 87.0 ± 15.1 kg; 1.80 ± 0.05 m; $15.9 \pm 7.2\%$ body fat) or a high-volume, moderate intensity 107 training group (HV; n = 10; 24.5 ± 2.6 years; 89.5 ± 12.9 kg; 1.66 ± 0.34 m; 20.6 ± 6.0% body 108 fat). All groups completed an 8-week resistance training program using a 4-day split routine. 109 Blood samples were collected during the first training session of week 1 (PRE) and week 8 110 (POST). These visits constituted the acute training protocols, during which, participants performed the HI (3-5 reps; 90% of one repetition maximum [1RM]) or HV (10-12reps: 70% 111 112 1RM) training paradigm.

113 Participants

Twenty physically active, resistance-trained men agreed to participate in this study. Following an explanation of all procedures, risks, and benefits, each participant provided his informed consent to participate in the study. This investigation was approved by the New England Institutional Review Board, and all procedures were in accordance with the ethical standards of the 1964 Helsinki Declaration and its later amendments. All participants were free of any physical limitations (determined by medical history questionnaire and PAR-Q) and had been regularly participating (at the time of recruitment) in resistance training for a minimum of 2 years (5.7 ± 2.2 years).

122	Prior to the present investigation, all participants described their training habits to be		
123	different from the present training regimen in terms of exercise order and groupings.		
124	Approximately 82% of the participants described their normal repetition range to be either lower		
125	(VOL = 77%) or higher (INT = 87%) than what they were assigned to in the study, with about		
126	43% typically using a 6–10 RM range and another 21% using an alternating (or pyramid)		
127	structure for specific multiple joint structural and assistance exercises. Additionally, 50% of the		
128	participants reported using either longer (VOL = 54%) or shorter (INT = 47%) rest periods,		
129	while approximately 29% did not track their rest times previously. The remaining participants		
130	employed a similar training scheme (i.e., intensity, volume, and rest) to what they were assigned		
131	to in the study.		

132 Preparatory Phase of Training

133 All participants completed a preparatory resistance training protocol during the 2 weeks 134 prior to the training intervention (see Table 1). This phase encompassed a total of six workouts: 135 four workouts (Monday, Tuesday, Thursday, and Friday) during the first week and two workouts 136 (Monday and Tuesday) during the second week. The purpose of the preparatory training program 137 was to instruct proper lifting technique, familiarize participants with all exercises, and ensure the 138 participants initiated the study with a comparable training base. In comparison to the training 139 intervention groups, the exercises (and their order) were identical but the volume (6-8 RM) and 140 rest intervals (1–2 min) differed. Participants were instructed not to participate in any other form 141 of physical activity throughout the duration of the study.

142 Anthropometric Assessments

Anthropometric measurements for all participants were conducted approximately 24 h prior to all strength measures. Height (±0.1 cm) and body mass (±0.1 kg) were determined using a Health-o-Meter Professional scale (Model 500 KL, Pelstar, Alsip, IL) with the participants standing barefoot, with feet together, and in their normal daily attire. Body fat percentage was determined using whole body-dual energy x-ray absorptiometry (DXA) scans (Prodigy TM; Lunar Corporation, Madison, WI). All measurements were performed by the same certified radiological technician using standardized subject positioning procedures.

150 Strength Testing

151 To determine appropriate training load for both HI and HV, strength in the bench press 152 and squat exercises was assessed. Participants were scheduled for testing at a standard time of 153 day. A general warm up consisting of riding a cycle ergometer for 5 min at a self-selected 154 resistance preceded strength testing. Standardized procedures, as previously described (19) were 155 used for the 1RM barbell bench press and barbell back squat, respectively. Subjects performed 156 two warm-up sets at 40-60% and 60-80% of his perceived 1RM, respectively, before performing 157 3-4 subsequent trials to determine the 1RM. A 3-5 minute rest period was provided between each 158 trial. For all other exercises, the 1RM was assessed using a prediction formula based on the 159 number of repetitions performed to volitional fatigue using a given weight (3). Trials not meeting 160 the range of motion criteria for each exercise or using improper technique were discarded.

161 *Resistance Training Intervention*

Participants reported to the Human Performance Lab (HPL) four times per week, at the
same time of day, to complete their assigned training program (see Table 1). Both groups

164 performed the same exercise routine but differed in the intensity of exercise, number of 165 repetitions performed and rest interval between sets. Specifically, the HI training program 166 required participants to perform four sets of 3–5 repetitions with 90% of their 1RM, with 3-min 167 rest period between sets, while the HV group performed four sets of 10–12 repetitions with 70% 168 of their 1RM, with a 1-min rest period between sets. During the resistance training program the 169 load was increased for participants (regardless of group) when the required number of repetitions 170 (for a particular exercise) were achieved on two consecutive workouts. On average three to four 171 participants were being trained by study personnel at one time during the course of the resistance 172 training program. All study personnel were certified strength and conditioning specialists.

173

Insert Table 1 Here

174 Blood Sampling

175 Blood samples were obtained at four time points: baseline (BL), immediately postexercise (IP), 30 minutes post-exercise (30P), and 60 minutes post-exercise (60P). Participants 176 177 reported to the Human Performance Lab (HPL) 3 hours post-prandial, at a standardized time of 178 day consistent with their normal training schedule. All blood samples at POST were taken at the 179 same time of day as PRE to avoid the confounding influence of diurnal variations. All blood 180 samples were obtained using a Teflon cannula placed in a superficial forearm vein using a three-181 way stopcock with a male luer lock adapter and plastic syringe. The cannula was maintained 182 patent using an isotonic saline solution (Becton Dickinson, Franklin Lakes, NJ, USA). Blood 183 samples at BL were obtained following a 15-minute equilibration period. Participants were 184 instructed to lie in a supine position for 15 min prior to the 30P and 60P blood draws.

All blood samples were collected into a Vacutainer® tube containing no containing K₂EDTA. The Vacutainer® tube was kept cold throughout processing in an attempt to reduce the variations in BDNF concentrations during processing. A small aliquot of whole blood from the tube was removed and used for determination of hematocrit and hemoglobin concentrations. The blood was centrifuged at 3,000×g for 15 minutes. The resulting plasma was placed into microcentrifuge tubes and frozen at -80° C for later analysis.

191 Blood Analyses

192 Hematocrit concentrations were analyzed from whole blood via microcentrifugation 193 (CritSpin, Westwood, MA, USA) and microcapillary techniques. Hemoglobin concentrations 194 were analyzed from whole blood using an automated analyzer (HemoCue, Cypress, CA, USA). 195 Coefficient of variation for each assay was 1.53% for hematocrit and 0.55% for hemoglobin. 196 Plasma volume shifts were calculated using the formula established by Dill & Costill (9). 197 Multiplex ELISA was used to quantitate plasma BDNF concentrations using MAGPIX® 198 (Luminex, Austin, TX, USA) and a commercially available kit (EMD Millipore, Billerica, MA, 199 USA) according to manufacturer's guidelines. To eliminate inter-assay variance, all samples 200 were analyzed in duplicate by a single technician. The coefficient of variation for BDNF was

201 7.84%.

202 Nutrient Intake and Dietary Analysis

Participants were instructed to maintain their normal kilocaloric intake habits throughout
the investigation. Kilocaloric and macronutrient intake were monitored via weekly food diaries.
Consequently, all participants were required to record all food and beverage intake over the
course of 3 days (two weekdays and one weekend day) during the initial and final week of

207 (weeks 1 and 8) of the training program. The FoodWorks Dietary Analysis software version 13

208 (The Nutrition Company, Long Valley, NJ) was used to analyze dietary recalls.

209

210 Statistical Analysis

211 Prior to hypothesis testing, the Shapiro-Wilk test was used to evaluate the assumption of 212 normality for dependent variables. As our data was not normally distributed we opted to log-213 transform BDNF measurements (using the natural logarithm). To examine group and training 214 differences in the BDNF response to exercise before and following the 7-week training program, 215 a three-way [training (PRE, POST) x time (BL, IP, 30P, 60P) x group (HI, HV)] repeated 216 measures analysis of variance (ANOVA) was performed. The effect of training on AUC 217 measures, calculated using the trapezoidal methods, was examined using a two-way [training 218 (PRE, POST) x group (HI, HV)] repeated measures ANOVA. In the event of a significant 219 interaction or main effect, Bonferroni post-hoc tests were performed. Interpretations of effect 220 size were evaluated (6) at the following levels: small effect (0.01-0.058), medium effect (0.059-221 0.137), and large effect (>0.138). For all analyses a criterion alpha level of $p \le 0.05$ was used to 222 determine statistical significance, and statistical software (SPSS V.21.0, Chicago, IL) was used. 223 All data are reported as mean \pm SD.

224 Results

The effect of these different training paradigms on strength and anthropometric changes have been reported elsewhere (25). For the purposes of this study, no differences were noted at PRE between the groups in the 1RM squat (p=0.694), 1RM bench press (p=0.934) body mass (p=0.715), lean body mass (p=0.611), or percent body fat (p=0.136). Relative energy intake did

229	not change significantly over the course of the investigation for HI (PRE: $38.2 \pm 11.1 \text{ kCal} \cdot \text{kg}^{-1}$;
230	POST: $31.1 \pm 5.3 \text{ kCal} \cdot \text{kg}^{-1}$) or HV (PRE: $31.7 \pm 7.0 \text{ kCal} \cdot \text{kg}^{-1}$; POST: $29.2 \pm 8.1 \text{ kCal} \cdot \text{kg}^{-1}$).

231 Changes in plasma BDNF concentrations during PRE and POST are depicted in Fig 1. 232 No training x time x group interaction for plasma BDNF concentration was noted (F=0.026, p=0.994, n^2 =0.002). However, significant main effects for training (F=4.434, p=0.050, n^2 =0.207) 233 and time (F=14.233, p<0.001, η^2 =0.456) were identified. When collapsed across group and 234 235 training, the acute exercise protocol resulted in significant elevations in BDNF concentrations 236 from BL at IP (p=0.001), 30P (p<0.001), and 60P (p<0.001). Circulating BDNF concentrations, 237 when collapsed across group and time, were significantly elevated (p=0.050) from PRE to POST. AUC analysis revealed no significant training x group interaction (F=0.956, p=0.342, η^2 =0.053) 238 239 in the BDNF response between HI and HV (see Fig 2). In addition, no main effect for training $(F=2.938, p=0.105, \eta^2=0.147)$ or group $(F=1.499, p=0.238, \eta^2=0.081)$ were noted for the BDNF 240 241 AUC response.

242

(Insert Fig 1 and 2 here)

Relative to BL, plasma volume shifts were not significantly different between the two groups at PRE (p=0.741) or POST (p=0.332). At PRE plasma volume decreased at IP, -8.8 \pm 8.6%; increased at 30P 5.2 \pm 7.7%; and increased at 60P, 4.7 \pm 6.4%. At POST plasma volume decreased at IP, -11.9 \pm 5.3%; increased at 30P, 3.2 \pm 4.0%; and increased at 60P, 5.8 \pm 10.4%. Blood variables were not corrected for plasma volume shifts due to the importance of molar exposure at the tissue receptor level.

249

251 **Discussion**

252 The primary objectives of this study were to characterize and determine if the BDNF 253 response was different between HI and HV resistance exercise and training. The major findings 254 of this study indicated that BDNF concentrations were significantly elevated following HI and 255 HV resistance exercise in experienced, resistance trained men. In addition, seven weeks of 256 resistance training appears to increase circulating BDNF concentrations in response to the exercise protocol, but did not change resting concentration. This appears to be the first study to 257 258 compare two commonly used resistance training paradigms on the BDNF response in trained 259 men.

260 Previous research on the acute BDNF response to resistance exercise has primarily 261 utilized untrained individuals, and the results have been inconclusive (7, 18, 35, 43). Similar to 262 the present study. Yarrow and colleagues (43) reported that resistance training can augment the 263 BDNF response to exercise, but not change resting concentrations. Other studies, using 264 previously untrained or recreationally trained participants of similar age were unable to 265 demonstrate any change in the BDNF response to an acute resistance exercise session (7, 18). 266 These differences may be related to the duration or volume of exercise. Correia and colleagues 267 (7) did not observe any change in BDNF concentrations following five sets of 10 repetitions in the isokinetic knee extension exercise, $(60^{\circ} \cdot \text{sec}^{-1})$. Similarly, Goekint et al. (18) used an exercise 268 269 protocol of three sets of 10 repetitions (80% 1RM) in six different exercises (a total of 18 sets 270 performed), and reported no change in the BDNF response to the exercise stimulus. However, 271 these exercises were all performed with exercise machines. In contrast, the participants in the 272 present study performed four sets of six exercises (a total of 24 sets) using free weight, multi-273 joint structural movement exercises that recruited a larger muscle mass than the exercises used in the previously mentioned studies. Exercises that recruit a greater amount of muscle mass are associated with a greater endocrine and biochemical response compared to single-joint or confined movement exercises that recruit a smaller muscle mass (23). The difference in musculature recruited may provide a plausible rationale for the difference in the present study and those of Corriea et al. (7) and Goekint et al. (18). It is also possible that a threshold stimulus of volume performed, muscle mass activated, and/or duration of activity is necessary to stimulate a resistance exercise-induced BDNF response.

281 The relationship of BDNF and resistance training has generally been examined in elderly 282 or clinically relevant populations (14-16, 24, 36, 39). Previous research from our laboratory (16) 283 and others (14, 24, 36, 39) have shown that resistance training does not significantly change 284 resting BDNF concentrations. Recently Forti and colleagues (15) reported a significant increase 285 in resting BDNF concentrations in elderly men (> 65 y) who participated in a 3-day per week, 286 12-week mixed, low intensity (60 repetitions at 20% of 1RM, and 10-20 repetitions at 40% of 287 1RM with no rest) resistance training program. However, in that same study no changes were 288 noted in resting BDNF concentrations in elderly men performing low intensity (1 set of 80-100 289 repetitions at 20% of 1RM) or higher intensity (2 sets of 10-15 repetitions at 80% of 1RM with 290 1-min rest between sets) training programs. In addition, no changes were noted in elderly women 291 who participated in all three training programs. Walsh and colleagues (42) reported that 8 weeks 292 of resistance training in older adults (60 - 77 years) was unable to change either resting or the 293 exercise response of BDNF. The ability of resistance training to elevate resting BDNF 294 concentrations may be a function of both age and training status. Younger, and more experienced 295 resistance trained adults appear to not be as sensitive as an older, untrained population to 296 stimulate changes in resting BDNF. However, we observed a significant effect of training on

circulating BDNF, supported by the strong effect sizes observed in the temporal (η^2 =0.207) and AUC (η^2 =0.147) responses. These results are in accordance with Yarrow and colleagues (37), who demonstrated 5 weeks of resistance training was able to alter the BDNF response in untrained men.

301 Increases in circulating BDNF in response to metabolic challenges are suggested to have 302 a role in regulating peripheral energy metabolism within the brain and in peripheral neurons (26). 303 Elevations in circulating concentrations of BDNF resulting from an elevation in metabolic stress 304 does provide support for a duration/volume threshold stimulus (13, 17, 34). Previous research 305 has shown cortisol concentrations to be inversely related to BDNF (2, 21). We have previously 306 reported the cortisol response of the current study (25). The cortisol response at POST was 307 attenuated in HV but not in HI. Although the attenuated cortisol response of HV may explain the 308 increase in circulating plasma BDNF at POST, the similar cortisol response between PRE and 309 POST for HI does not provide support. Therefore, it does not appear that cortisol influenced 310 changes in the BDNF response observed in this study. Additional research is necessary to 311 provide further insight to the mechanism governing BDNF changes to training.

312 The role of circulating BDNF is ambiguous. Much of our present understanding focuses 313 on BDNF's role as a neurotrophin in the brain (40), however, its role in modulating peripheral 314 neurogenesis is less understood. Current evidence indicates plasma BDNF is a proxy marker of 315 BDNF production in the brain (34), however BDNF can also be synthesized peripherally (40). 316 The importance of the resistance training induced increase in BDNF has been suggested to be 317 related to improvements in cognitive function, memory and mood (1, 8, 10). Further research is 318 still needed to delineate the potential physiological role that BDNF has in the neuromuscular 319 system, especially in apparently healthy, trained individuals.

320 Conclusions

This investigation appears to be the first study to explore the BDNF response to different resistance training paradigms in experienced, resistance trained men. The results of this study demonstrate that the BDNF response is significantly elevated following both the high intensity and high volume training protocols, with no differences between the protocols. In addition, 7weeks of training did appear to increase the BDNF response to exercise. Future research appears warranted in examining the potential role that elevated BDNF concentrations may have on neuromuscular adaptations.

329 **Conflicts of Interest**

330 No funding was received for the purposes of this study. The authors declare no conflicts

331 of interest.

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466 Figure Legends

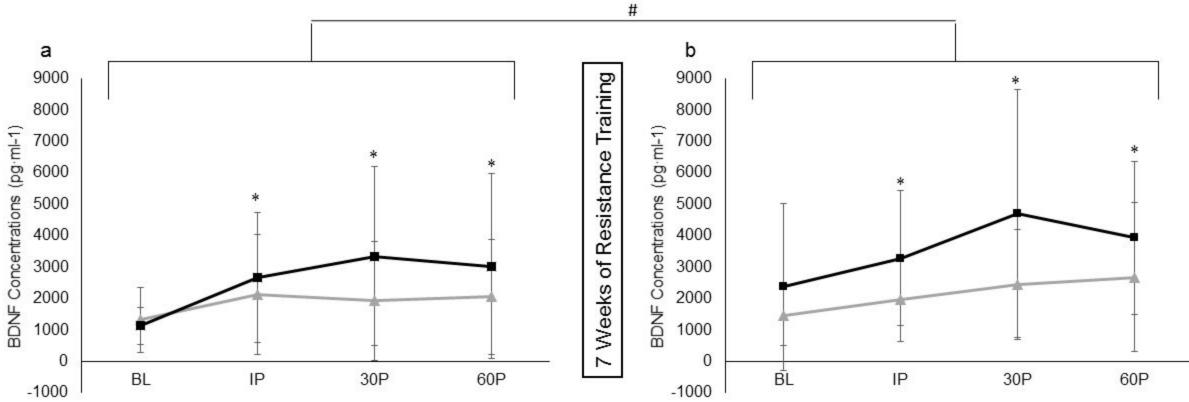
467 Figure 1. Temporal circulating brain derived neurotrophic factor (BDNF) concentrations at rest

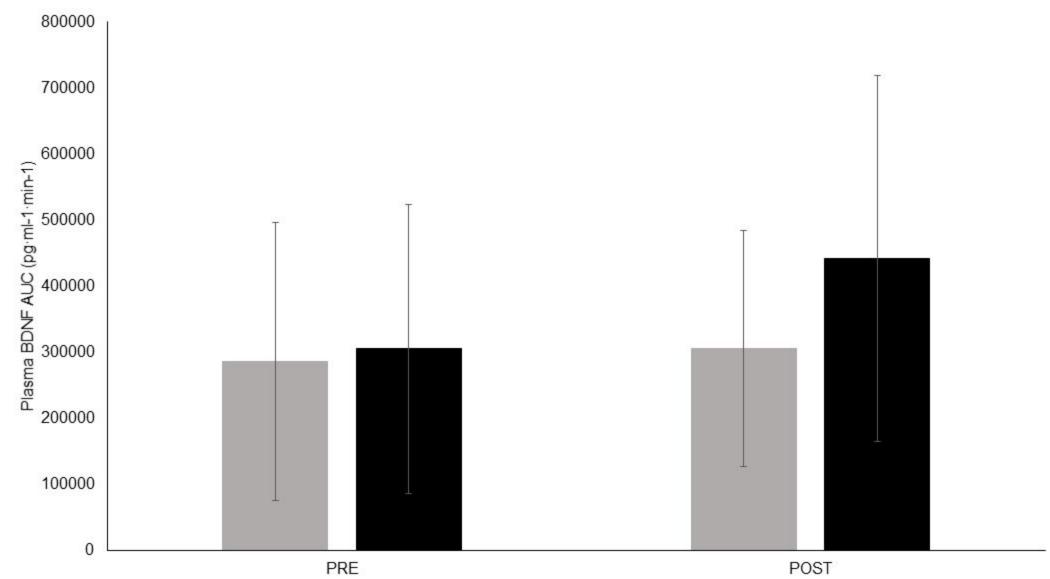
- 468 (BL), immediately after (IP), 30 minutes after (30P), and 60 minutes after (60P) either a high-
- 469 intensity, low volume (HI; light line with triangles) or low-intensity, high volume (HV; dark line
- 470 with squares) resistance exercise protocol before (PRE; A) and after (POST; B) 7 weeks of
- 471 resistance training. Data are presented at mean \pm SD. * = Significant difference compared to BL
- 472 when groups are collapsed across group and training session. # = Significant difference at POST
- 473 as compared to PRE when collapsed across time and group.
- 474 Figure 2. Area under the curve (AUC) of plasma BDNF before (PRE) and after (POST) 7 weeks
- 475 of either a high-intensity, low volume (HI) or low-intensity, high volume (HV) resistance
- 476 training program.

Program variable	Preparatory phase both groups	High Volume (HV)	High Intensity (HI)
Exercise prescription			
Training duration	2 weeks	8-weeks	8-weeks
Training intensity	80-85% 1RM	70% 1RM	90% 1RM
Training volume	4 sets x 6-8	4 sets x 10-12	4 sets x 3-5
	repetitions	repetitions	repetitions
Rest time	1-2 minutes	1 minute	3 minutes
Exercises			
<u>Day 1</u>	<u>Day 2</u>	Day 3	Day 4
Back squats	Bench press	Back squats	Bench press
Deadlifts	Incline bench press	Deadlifts	Incline bench press
Leg press	Dumbbell flys	Barbell lunge	Incline dumbbell flys
Lat pull downs	Seated shoulder press	Seated row	Seated shoulder press
Barbell bent-over	Lateral dumbbell	Dumbbell	Lateral dumbbell
rows	raise	pullover	raise
Barbell biceps curls	Triceps extension	Dumbbell biceps curl	Triceps extension

 Table 1 Resistance training program

478





≡HI ∎HV