

**UCC Library and UCC researchers have made this item openly available.  
Please [let us know](#) how this has helped you. Thanks!**

<b>Title</b>	The success of microneedle-mediated vaccine delivery into skin
<b>Author(s)</b>	Marshall, Sarah; Sahm, Laura J.; Moore, Anne C.
<b>Publication date</b>	2016-11-01
<b>Original citation</b>	Marshall, S., Sahm, L. J. and Moore, A. C. (2016) 'The success of microneedle-mediated vaccine delivery into skin', <i>Human Vaccines and Immunotherapeutics</i> , 12(11), pp. 2975-2983. doi: 10.1080/21645515.2016.1171440
<b>Type of publication</b>	Article (peer-reviewed)
<b>Link to publisher's version</b>	<a href="http://dx.doi.org/10.1080/21645515.2016.1171440">http://dx.doi.org/10.1080/21645515.2016.1171440</a> Access to the full text of the published version may require a subscription.
<b>Rights</b>	© 2016, Taylor & Francis Group. This is an Accepted Manuscript of an article published by Taylor & Francis in <i>Human Vaccines and Immunotherapeutics</i> on 1 November 2016, available online: <a href="https://doi.org/10.1080/21645515.2016.1171440">https://doi.org/10.1080/21645515.2016.1171440</a>
<b>Item downloaded from</b>	<a href="http://hdl.handle.net/10468/8535">http://hdl.handle.net/10468/8535</a>

Downloaded on 2021-11-27T10:01:11Z

# 1 **The success of microneedle-mediated vaccine delivery into skin**

2  
3 Sarah Marshall<sup>a</sup>, Laura J. Sahm<sup>ac</sup>, Anne C. Moore<sup>ab</sup>

4 School of Pharmacy<sup>a</sup>, Department of Pharmacology and Therapeutics<sup>b</sup>, University College Cork,  
5 Department of Pharmacy<sup>c</sup>, Mercy University Hospital, Cork, Ireland.  
6

## 7 **Abstract**

8 Microneedles (MNs) are designed to specifically target the outermost, skin barrier layer, the *stratum*  
9 *corneum*, creating transient pathways for minimally invasive transcutaneous delivery. It is reported  
10 that MNs can facilitate delivery without stimulating the pain receptors or damaging blood vessels  
11 that lie beneath, thus being perceived as painless and associated with reduced bleeding. This  
12 immunocompetence of the skin, coupled with its ease of access, makes this organ an attractive  
13 vaccination site. The purpose of this review was to collate primary scientific literature pertaining to  
14 MN-mediated *in vivo* vaccination programmes. A total of 62 original research articles are presented,  
15 compiling vaccination strategies in 6 different models (mouse, rat, guinea pig, rabbit, pig, macaque  
16 and human). Vaccines tested span a wide range of viral, bacterial and protozoan pathogens and  
17 includes 7 of the 13 vaccine-preventable diseases, as defined by the WHO. This review highlights the  
18 paucity of available clinical trial data. MN-delivered vaccines have demonstrated safety and  
19 immunogenicity in pre-clinical models and boast desirable attributes such as painless administration,  
20 thermostability, dose-sparing capacity and the potential for self-administration. These advantages  
21 should contribute to enhanced global vaccine access.

## 22 **1. Introduction**

### 23 **1.1 Vaccine delivery**

24 Vaccines are conventionally administered using a hypodermic needle [1]. This form of administration  
25 provides a rapid and direct method of vaccine delivery. Despite familiarity, widespread use and  
26 proven efficacy, the hypodermic needle is associated with accidental needle stick injury, spread of  
27 blood-borne infections [2-4], as well as phobias, pain and significant anxiety [5-8]. In addition, these  
28 needles are not easily self-administered, unless the individual has received specialised training on  
29 injection technique and needle disposal [9]. Oral vaccination is an attractive alternative [10] and a  
30 limited number of oral vaccines have been approved for human use [11-15]. However, this mode of

31 immunisation can be less effective, as vaccine antigens undergo digestion in the gastrointestinal  
32 tract prior to induction of an adequate immune response [10] and research on their use has been  
33 limited almost exclusively to protection against mucosally transmitted pathogens, with some notable  
34 recent exceptions [15]. The transdermal route, based on diffusion, has also been investigated.  
35 However this route limits delivery only to lipophilic, low molecular weight potent products [16] and  
36 would prevent a vaccine from crossing the skin due to the presence of the relatively impermeable,  
37 outer *stratum corneum* (SC) layer. Intradermal vaccination is not a novel concept. In 1910, French  
38 physician Charles Mantoux published his clinical research on the intradermal injection of tuberculin  
39 as a diagnostic test for tuberculosis disease [17]. This diagnostic technique formed the basis for  
40 intradermal vaccination, a technique still in use for vaccines such as rabies [18] and BCG [19].  
41 However, intradermal delivery is technically challenging, requiring significant operator training [20]  
42 and has been associated with adverse events such as pain, inflammatory changes [21] and the  
43 development of abscesses [22]. Taking into account the limitations of parenteral, oral and traditional  
44 transdermal and intradermal vaccination, the concept of the microneedle (MN) emerged as a  
45 solution to these issues. MNs can be 1µm in diameter and range from 50µm to 1000µm in length,  
46 while mini-needles range from 1000µm to 1500µm [23]. They are designed to specifically target the  
47 outermost, rate limiting, skin barrier layer, the SC, creating transient pathways for minimally invasive  
48 transcutaneous delivery [24]. There are four different types of MNs: solid, coated, hollow and  
49 dissolving. It is reported that MNs can facilitate delivery through SC interruption without stimulating  
50 the pain receptors and blood vessels that lie beneath, thus being perceived as painless and  
51 associated with a reduction in bleeding [1, 25, 26]. Other advantages of microneedle-mediated  
52 delivery include avoidance of first pass metabolism; potential for highly targeted administration to  
53 individual cells [26, 27]; improved patient compliance [28]; dose sparing [29, 30]; thermostability of  
54 certain platforms [31-33] and potential for self-administration.

## 55 **1.2 The skin: an immune organ and vaccine target**

56 Skin is the largest immune organ in the human body [34], composed of two primary layers, the  
57 epidermis and dermis [35]. These layers provide a protective interface between internal organs and  
58 the external environment, encountering a host of toxins, pathogenic organisms and physical stresses  
59 [36]. The skin functions as more than just a physical barrier. It is capable of mounting a potent  
60 immune response due to the residence of specialised antigen presenting cells. Langerhans cells are  
61 abundant in the epidermis, comprising 2% to 4% of epithelial cells [36], while more classical  
62 dendritic cells are found in the dermis [36-40]. Other immune-competent accessory cells residing in  
63 the skin include keratinocytes, epidermal cells which play a role in initiating cell-mediated immune  
64 responses through the release of cytokines and the expression of cellular adhesion molecules to

65 facilitate movement and coordination with other immune cells; T lymphocytes; melanocytes,  
 66 epidermal pigment cells which produce a number of cytokines that mediate inflammation and mast  
 67 cells, leukocytes which modulate host innate immune response through the release of granular and  
 68 secreted mediators and recruit multiple inflammatory cells through the production of chemotactic  
 69 factors [36, 41-43]. The resident professional APC are adept at antigen capture, and upon  
 70 appropriate activation through intracellular interaction, migrate to proximal lymph nodes to activate  
 71 B and T lymphocytes and mediate initiation of an adaptive immune response [37, 44]. This  
 72 immunocompetence, coupled with its ease of access, makes the skin an attractive vaccination  
 73 target.

## 74 2. Literature review

75 The purpose of this review is to collate literature detailing the success of MN-mediated *in vivo*  
 76 vaccination programmes. Keywords including ‘microneedle’, ‘solid microneedle’, ‘coated  
 77 microneedle’, ‘hollow microneedle’ ‘dissolvable microneedle’, ‘dissolving microneedle’, were  
 78 combined with ‘vaccine’, ‘vaccination’ and ‘immunisation’. Using Google as a search engine, these  
 79 keywords were combined in various permutations and combinations to search PubMed. This yielded  
 80 a total of 748 results. Following removal of duplications, 180 results remained. The title and abstract  
 81 of each result were examined and included or excluded in the final review based on the criteria  
 82 outlined in Table 1. A total of 62 results were included in the final review [29, 45-105] .

Table 1: Inclusion and Exclusion Criteria	
Inclusion Criteria	Exclusion Criteria
Original research articles	Review articles
MN-mediated vaccine delivery	MN-mediated non-vaccine delivery
Published in English language	MN fabrication studies
<i>in vivo</i> MN administration	MN stability studies
Article available in full	

83

### 84 2.1 Solid MNs

85 The simplest forms of MNs are solid devices. Solid MNs create transient micropores in the SC,  
 86 thereby increasing permeability of the barrier layer. Vaccine applied onto the treated surface  
 87 diffuses into the skin (from a loaded patch or semi-solid formulation) through the pores created by  
 88 MN pre-treatment. The applied vaccine can exert a local effect in the skin and a systemic effect  
 89 following uptake [1]. Solid MNs have been used to deliver vaccines for diphtheria [45, 46, 48],  
 90 influenza [46], hepatitis B [47, 49, 50] and malaria [51, 52] in mice. Microenhancer array devices  
 91 were developed to cause mild abrasion of the SC. These devices scrape the skin with blunt-tipped

92 microneedles and have been used to increase the delivery of an anthrax vaccine in mice and rabbits  
93 [53], a Japanese encephalitis vaccine in cynomolgus monkeys [56] and a rabies vaccine in humans  
94 [57, 58]. While these devices were shown to be effective, intradermal injection of the vaccine in  
95 each of these studies was significantly more effective, potentially due to inefficient delivery into the  
96 skin from the formulation. In spite of immunogenicity, the popularity of solid MNs has reduced in  
97 recent years, potentially due to the requirement for a multi-step administration process, the lack of  
98 consistency and the increased number of advantages of other MN systems.

## 99 **2.2 Coated microneedles**

100 Advancement on solid MNs was the development of coated devices. Solid MNs are pre-coated with  
101 a vaccine in a formulation suitable for coating and dissolution [1], thus resulting in a one-step  
102 delivery process. The vaccine coated MNs are inserted into the skin, where dissolution of the vaccine  
103 occurs. Vaccine delivery via coated MNs is limited by the dimensions of the MN shaft and tip [106,  
104 107]. Successful vaccine coated MNs include influenza [54, 55, 59-77], human papillomavirus [78,  
105 79], chikungunya virus [80], West Nile virus [80], rotavirus [81], herpes simplex virus [82] and  
106 hepatitis C [83] in mice, influenza virus [84] and bacillus Calmette-Guérin in guinea pigs [85],  
107 hepatitis B virus in pigs [86], and measles [87] and polio [88] viruses in rats. This literature search did  
108 not reveal any clinical trials pertaining to vaccine delivery via coated microneedles.

## 109 **2.3 Hollow microneedles**

110 Hollow MNs provide a pre-defined conduit for vaccine delivery into the skin or other tissue.  
111 Currently there are two hollow MN designs: a single MN or mini-needle, which mimics the  
112 conventional hypodermic needle [108] or an array of multiple MNs [109]. The latter permits  
113 simultaneous application of a vaccine formulation over a wider area of skin, potentially resulting in  
114 higher bioavailability and increasing the likelihood of lymphatic uptake of presented antigens [110].  
115 Vaccine may be delivered by passive diffusion through the MN. Conversely, a syringe may be  
116 attached to the MN, permitting active vaccine delivery. There are several commercially available  
117 hollow MN systems; Soluvia® is licensed for use [57, 91] and MicronJet® is being clinically tested  
118 [29]. Soluvia® is a pre-fillable microinjection system with a single 1500µm hollow silicon MN, while  
119 MicronJet® is composed of four 600µm hollow silicon MNs arranged on a plastic adaptor for  
120 attachment to a standard syringe barrel [23]. Hollow MNs have been successfully developed to  
121 immunise human subjects with polio [89, 90] or influenza [29, 91, 92] vaccines, to immunise mice  
122 against plague [93] and to administer polio vaccine to rats [94].

## 123 **2.4 Dissolving microneedles**

124 The final, most advanced and complex MN is the dissolving MN. Dissolving MNs are polymeric and  
125 encapsulate vaccine within their matrix [1, 111, 112]. Insertion of the MNs into the skin catalyses the  
126 degradation of the polymeric compound, subsequently releasing the vaccine [112]. Unlike the  
127 alternate MN platforms already discussed, dissolving MNs are fully biocompatible and do not  
128 generate biohazardous waste, a distinct advantage [113, 114]. Other advantages include robustness  
129 and scalability [115, 116]. However, unlike hollow MN, a limitation is placed on the amount of  
130 vaccine that can be incorporated into the system [117] and vaccinees may be obliged to wait for  
131 extended periods of time to ensure complete MN degradation [114]. Dissolving MNs have been  
132 developed to incorporate vaccines for influenza virus [95-100], hepatitis B [101, 102], tetanus [97],  
133 diphtheria [97], malaria [97] and HIV [103] in mice and measles [104] and polio [105] in rhesus  
134 macaques, with a long term aim to create a thermostable, self-administration platform. Although an  
135 attractive platform, dissolvable microneedle (DMN) systems for vaccine delivery have required more  
136 time to reach clinical trials compared to hollow or solid microneedles. Hollow and solid MN devices  
137 have a traditional medical device classification. In contrast, DMN patches will likely be seen, from a  
138 regulatory perspective, as a combination product of a medicinal product (the vaccine) and a device  
139 (potential backing layers and/or applicators). However, as a new dosage format, the product  
140 specifications, critical quality attributes of each product and regulatory pathway of DMN systems has  
141 not yet been defined. Furthermore, to ensure the quality of vaccine-loaded DMN patches that will  
142 be clinically used, they must be produced in the appropriate environment that complies with, good  
143 manufacturing practice (GMP). These processes, guidelines and regulatory strategies are only  
144 recently being defined [118].

## 145 **3. Discussion**

146 The purpose of this review was to collate primary scientific literature pertaining to MN-mediated *in*  
147 *vivo* vaccination programmes, according to the inclusion and exclusion criteria outlined in Table 1. A  
148 total of 62 original research articles are presented, compiling vaccination strategies in 6 different  
149 models (mouse [45-52, 54, 55, 59-83, 93, 95-103], rat [87, 88, 94], guinea pig [84, 85], rabbit [53], pig  
150 [86], cynomolgus [56] or rhesus macaque [104, 105]) and in human subjects [29, 57, 58, 89-92]. The  
151 review highlights MN compatibility with live, inactivated, subunit and DNA vaccines. Vaccines tested  
152 span a wide range of viral, bacterial and protozoan pathogens; including influenza [29, 46, 54, 55, 59-  
153 77, 84, 91, 92, 95-100], hepatitis B [47, 49, 50, 86, 101, 102], Japanese encephalitis [56], rabies [57,  
154 58], human papillomavirus [78, 79], chikungunya virus [80], West Nile virus [80], rotavirus [81],  
155 herpes simplex [82], hepatitis C [83], measles [87, 104], polio [88-90, 94, 105] and HIV [103],  
156 bacterial illnesses including diphtheria [45, 46, 48, 97], anthrax [53], tuberculosis [85], plague [93]

157 and tetanus [97] and protozoan illnesses including malaria [51, 52, 97], as summarised in Table 2.  
 158 This list includes 7 of the 13 vaccine-preventable diseases, as defined by the WHO [119]. This review  
 159 highlights the paucity of clinical trial data, with only 11.29% of the 62 trials presented conducted in  
 160 human subjects.

**Table 2: Disease targets for microneedle-mediated vaccine delivery**

Model	Virus	Bacteria	Protozoa
Mouse	Hepatitis B [47, 49, 50, 101, 102] Influenza [46, 54, 55, 59-77, 95-100] Human papillomavirus [78, 79] West Nile virus [80] Chikungunya virus [80] Rotavirus [81] Herpes simplex [82] Hepatitis C [83] HIV [103]	Diphtheria [45, 46, 48, 97] Anthrax[53] Plague [93] Tetanus [97]	Malaria [51, 52, 97]
Rat	Measles [87] Polio [88, 94]		
Guinea Pig	Influenza [84]	Tuberculosis [85]	
Rabbit		Anthrax [120]	
Pig	Hepatitis B [86]		
Macaque	Japanese encephalitis [56] Measles [104] Polio [105]		
Human	Influenza [29, 91, 92] Rabies [57, 58] Polio [89, 90]		

161

### 162 **3.1 Influenza: a popular vaccine target**

163 The influenza virus vaccine is as a popular vaccine target, being the pathogen of interest in 32 of the  
 164 62 research articles presented. Influenza is a highly contagious respiratory illness, with influenza A  
 165 and B viruses causing annual seasonal epidemics and sporadic pandemics of disease, leading to  
 166 hospitalisations and occasionally death [121-125]. In the US, it is estimated that influenza resulted in  
 167 greater than half a million hospitalisations, 18,491-95,390 intensive care admissions and 4,915-  
 168 27,174 deaths per year between 2010 and 2013 [126]. Investment in the development of an  
 169 influenza vaccine offers significant commercial and technical gain. Unlike other vaccines, which offer  
 170 life-long immunity with a single dose, influenza immunity requires annual re-vaccination as a result  
 171 of antigenic variation of the virus [127]. The target end-user of a microneedle patch-based influenza

172 vaccine is the adult population and not the paediatric, thus reducing the barrier to clinical use.  
173 Vaccination is effective in preventing infection [128]. Furthermore, unlike many other vaccines,  
174 serological correlates of protection exist and the CHMP criteria are accepted to measure  
175 immunogenicity. However, coverage rates in target populations are far below the WHO-  
176 recommended 75% [129-131]. In addition, there are unmet needs associated with current influenza  
177 vaccines [132]. This motivates the development of alternate delivery systems such as MNs that may  
178 offer enhanced vaccine uptake and acceptance [133]. If a MN-based vaccine exhibited enhanced  
179 stability and lower vaccine doses could be used [29, 30], then this could be attractive to vaccine  
180 manufacturers. From a user perspective, the prospect of a painless, potentially self-administered  
181 vaccine may lead to improved vaccination coverage [28]. However all of these features still remain  
182 to be rigorously tested and developed in a clinical context.

### 183 **3.2 Improving vaccine coverage in developing countries**

184 Even though vaccination programmes are frequently cited as one of the most low-cost, high-impact  
185 public health measures [134], 1.5 million children die every year as a result of vaccine preventable  
186 illnesses, including some of those presented in this review. Vaccines are temperature sensitive  
187 biological products, requiring refrigeration. In many developing world countries, a cold-chain  
188 infrastructure is almost prohibitively expensive thus preventing adequate vaccine distribution [135,  
189 136]. The thermostability of MN vaccines eliminates cold-chain requirements, thus reducing logistic  
190 costs and potentially improving distribution [31-33]. This thermostability would permit stock-piling in  
191 regular drug distribution networks, combatting the frequently encountered issue of supply shortage.  
192 In addition to being thermolabile, conventional vaccines often require administration by trained  
193 personnel. In LMIC countries, there are shortages of medical personnel at all levels of training [137]:  
194 Africa has 2.3 healthcare workers per 1000 population, compared to 24.8 per 1000 in the Americas  
195 [138]. Therefore the previously discussed potential for self-administration with MN vaccines could  
196 further improve vaccine coverage in these countries, in tandem with other public health efforts.  
197 However, most paediatric vaccines in the Expanded Programme of Immunization are adjuvanted.  
198 Pharmaceutical, immunological, safety and efficacy issues of incorporating licensed adjuvants into  
199 solid dosage formats of microneedles must be addressed before this technology will be licensed and  
200 deployed for these vaccines. Significant research and development effort is being focussed in these  
201 areas to resolve these concerns.

### 202 **3.3 Translation into clinical use**

203 This review presents a variety of MN vaccines in the pre-clinical development stage, demonstrating  
204 safety and immunogenicity in animal models but also highlights the scarcity of clinical trial data.  
205 There is a progression from evaluation in small animal models such as mice, to higher animal models



206 such as rhesus macaques, prior to transition to clinical development and evaluation in human  
207 subjects [139]. While preclinical research answers basic questions, it is not a surrogate for clinical  
208 research. It is hoped that the MN vaccines presented in this review, especially those that have  
209 undergone assessment in non-human primates, will progress through the developmental stages,  
210 ultimately leading to vaccine licensing and introduction into clinical use. An issue that needs to be  
211 assessed is the habituality of hypodermic needle-mediated vaccination. Despite the aforementioned  
212 disadvantages, traditional immunisation has repeatedly demonstrated efficacy and safety.  
213 Familiarity breeds acceptance. Therefore a paradigm shift is required to drive the transition of MN-  
214 vaccines into clinical use. Increased end-user acceptability of MN-based vaccines will be required for  
215 widespread adoption. Positive attributes such as pain-free, bloodless administration must be  
216 rigorously tested and defined and acceptance of this technology by the end-user must be assessed,  
217 understood and the technology adapted to incorporate end-users' needs. MN fabrication  
218 considerations include scalability and dose loading capacity must also be addressed so that the  
219 vaccine manufacturer can assimilate the technology into their fill-finish systems. The majority of MN  
220 research has been conducted at laboratory scale in small quantities and the development of  
221 alternate fabrication approaches has begun to demonstrate scalability [140, 141]. There is an  
222 inherent dose loading capacity associated with some MN technologies, whereby there is a limit to  
223 the amount of vaccine that can be coated on or incorporated in the MN [1]. The inclusion of  
224 adjuvants may reduce the vaccine dose required to elicit an appropriate immune response [86, 142],  
225 although their inclusion will also necessitate appropriate validation and production in GMP  
226 environments. Finally, there is a need for the development of universal acceptance criteria and Good  
227 Manufacturing Practice specifications, permitting MN characterisation and subsequent  
228 commercialisation [118].

229 This review presented the research pertaining to *in vivo* MN vaccines. Vaccines have been delivered  
230 via solid, coated, hollow and dissolving MNs. The dissolving MN offers a significant advantage over  
231 other MN platforms: the elimination of sharp, biohazardous waste after vaccination. MNs have the  
232 potential to improve vaccine access in developing countries. These vaccines have demonstrated  
233 safety and immunogenicity in pre-clinical models. The paucity of clinical data presented in this  
234 review highlights the need to incentivise vaccine research in human subjects. The technology  
235 possesses desirable attributes for the end-user including painless administration and potential for  
236 self-application, which may increase compliance and subsequent vaccine coverage, as well as  
237 benefits for the manufacturer including thermostability and dose-sparing capacity. All of these  
238 advantages demonstrate the high potential for microneedle technologies to have a positive impact  
239 on global immunisation programmes in the future.

## References

1. Kim, Y.-C., J.-H. Park, and M.R. Prausnitz, *Microneedles for drug and vaccine delivery*. *Advanced drug delivery reviews*, 2012. **64**(14): p. 1547-1568.
2. Drucker, E., P.G. Alcabes, and P.A. Marx, *The injection century: massive unsterile injections and the emergence of human pathogens*. *The Lancet*, 2001. **358**(9297): p. 1989-1992.
3. Kermode, M., *Unsafe injections in low-income country health settings: need for injection safety promotion to prevent the spread of blood-borne viruses*. *Health promotion international*, 2004. **19**(1): p. 95-103.
4. Hauri, A.M., G.L. Armstrong, and Y.J. Hutin, *The global burden of disease attributable to contaminated injections given in health care settings*. *International journal of STD & AIDS*, 2004. **15**(1): p. 7-16.
5. Nir, Y., et al., *Fear of injections in young adults: prevalence and associations*. *The American journal of tropical medicine and hygiene*, 2003. **68**(3): p. 341-344.
6. Hamilton, J.G., *Needle phobia: a neglected diagnosis*. *The Journal of family practice*, 1995.
7. Marks, I., *Blood-injury phobia: a review*. *American Journal of Psychiatry*, 1988. **145**(10): p. 1207-1213.
8. Kleinknecht, R.A., *Acquisition of blood, injury, and needle fears and phobias*. *Behaviour research and Therapy*, 1994. **32**(8): p. 817-823.
9. Giudice, E.L. and J.D. Campbell, *Needle-free vaccine delivery*. *Advanced drug delivery reviews*, 2006. **58**(1): p. 68-89.
10. Wang, L. and R.L. Coppel, *Oral vaccine delivery: can it protect against non-mucosal pathogens?* 2008.
11. Top, F.H., et al., *Immunization with live types 7 and 4 adenovirus vaccines. II. Antibody response and protective effect against acute respiratory disease due to adenovirus type 7*. *Journal of Infectious Diseases*, 1971. **124**(2): p. 155-160.
12. Bernstein, D.I., *Rotarix: development of a live attenuated monovalent human rotavirus vaccine*. *Pediatric annals*, 2006. **35**(1): p. 38.
13. Ciarlet, M. and F. Schödel, *Development of a rotavirus vaccine: clinical safety, immunogenicity, and efficacy of the pentavalent rotavirus vaccine, RotaTeq®*. *Vaccine*, 2009. **27**: p. G72-G81.
14. Germanier, R. and E. Furer, *Characteristics of the attenuated oral vaccine strain "S. typhi" Ty 21a*. *Developments in biological standardization*, 1982. **53**: p. 3-7.
15. Liebowitz, D., et al., *High titre neutralising antibodies to influenza after oral tablet immunisation: a phase 1, randomised, placebo-controlled trial*. *Lancet Infect Dis*, 2015. **15**(9): p. 1041-8.
16. Bora, P., L. Kumar, and A.K. Bansal, *Microneedle technology for advanced drug delivery: Evolving vistas*. *Review Article, Department of Pharmaceutical Technology, NIPER, CRIPS*, 2008. **9**(1).
17. Lambert, P.H. and P.E. Laurent, *Intradermal vaccine delivery: will new delivery systems transform vaccine administration?* *Vaccine*, 2008. **26**(26): p. 3197-3208.
18. Briggs, D., et al., *Antibody response of patients after postexposure rabies vaccination with small intradermal doses of purified chick embryo cell vaccine or purified Vero cell rabies vaccine*. *Bulletin of the World Health Organization*, 2000. **78**(5): p. 693-698.
19. Hawkridge, A., et al., *Efficacy of percutaneous versus intradermal BCG in the prevention of tuberculosis in South African infants: randomised trial*. *Bmj*, 2008. **337**: p. a2052.
20. Emerging, W.H.O.D.o. and O.C.D. Surveillance, *WHO recommendations on rabies post-exposure treatment and the correct technique of intradermal immunization against rabies1997*: World Health Organization, Division of Emerging and other Communicable Diseases Surveillance and Control.

21. Al Jarad, N., D. Empey, and G. Duckworth, *Administration of the BCG vaccination using the multipuncture method in schoolchildren: a comparison with the intradermal method*. Thorax, 1999. **54**(9): p. 762-764.
22. Ormerod, L. and C. Palmer, *Tuberculin reactivity after neonatal percutaneous BCG immunisation*. Archives of disease in childhood, 1993. **69**(1): p. 155.
23. Donnelly, R.F. and T.R.R. Singh, *Novel Delivery Systems for Transdermal and Intradermal Drug Delivery* 2015: John Wiley & Sons.
24. Haq, M., et al., *Clinical administration of microneedles: skin puncture, pain and sensation*. Biomedical microdevices, 2009. **11**(1): p. 35-47.
25. Birchall, J.C., et al., *Microneedles in clinical practice—an exploratory study into the opinions of healthcare professionals and the public*. Pharmaceutical research, 2011. **28**(1): p. 95-106.
26. Escobar-Chávez, J.J., et al., *Microneedles: a valuable physical enhancer to increase transdermal drug delivery*. The Journal of Clinical Pharmacology, 2011. **51**(7): p. 964-977.
27. Birchall, J.C., *Microneedle array technology: the time is right but is the science ready?* Expert review of medical devices, 2006. **3**(1): p. 1-4.
28. Norman, J.J. and M.R. Prausnitz, *Improving patient acceptance of insulin therapy by improving needle design*. Journal of diabetes science and technology, 2012. **6**(2): p. 336-338.
29. Van Damme, P., et al., *Safety and efficacy of a novel microneedle device for dose sparing intradermal influenza vaccination in healthy adults*. Vaccine, 2009. **27**(3): p. 454-459.
30. Al-Zahrani, S., et al., *Microneedle-mediated vaccine delivery: harnessing cutaneous immunobiology to improve efficacy*. Expert opinion on drug delivery, 2012. **9**(5): p. 541-550.
31. Mistilis, M.J., A.S. Bommarius, and M.R. Prausnitz, *Development of a Thermostable Microneedle Patch for Influenza Vaccination*. Journal of pharmaceutical sciences, 2015. **104**(2): p. 740-749.
32. Choi, H.-J., et al., *Stability of influenza vaccine coated onto microneedles*. Biomaterials, 2012. **33**(14): p. 3756-3769.
33. Chen, X., et al., *Improving the reach of vaccines to low-resource regions, with a needle-free vaccine delivery device and long-term thermostabilization*. Journal of Controlled Release, 2011. **152**(3): p. 349-355.
34. Jepps, O.G., et al., *Modeling the human skin barrier—Towards a better understanding of dermal absorption*. Advanced drug delivery reviews, 2013. **65**(2): p. 152-168.
35. Skountzou, I., et al., *Transcutaneous immunization with inactivated influenza virus induces protective immune responses*. Vaccine, 2006. **24**(35): p. 6110-6119.
36. Salmon, J., C. Armstrong, and J. Ansel, *The skin as an immune organ*. Western journal of medicine, 1994. **160**(2): p. 146.
37. Banchereau, J. and R.M. Steinman, *Dendritic cells and the control of immunity*. Nature, 1998. **392**(6673): p. 245-252.
38. Steinman, R.M., *Dendritic cells and the control of immunity: enhancing the efficiency of antigen presentation*. The Mount Sinai journal of medicine, New York, 2001. **68**(3): p. 160-166.
39. Paus, R., et al., *Who is really in control of skin immunity under physiological circumstances—lymphocytes, dendritic cells or keratinocytes?* Experimental dermatology, 2006. **15**(11): p. 913-916.
40. Lee, H.K. and A. Iwasaki. *Innate control of adaptive immunity: dendritic cells and beyond*. in *Seminars in immunology*. 2007. Elsevier.
41. Zachariae, C.O., K. Thestrup-Pedersen, and K. Matsushima, *Expression and secretion of leukocyte chemotactic cytokines by normal human melanocytes and melanoma cells*. Journal of Investigative Dermatology, 1991. **97**(3): p. 593-599.
42. Urb, M. and D.C. Sheppard, *The role of mast cells in the defence against pathogens*. PLoS Pathog, 2012. **8**(4): p. e1002619.

43. Abraham, S.N. and A.L.S. John, *Mast cell-orchestrated immunity to pathogens*. Nature Reviews Immunology, 2010. **10**(6): p. 440-452.
44. Koutsonanos, D.G., et al., *Enhanced immune responses by skin vaccination with influenza subunit vaccine in young hosts*. Vaccine, 2015.
45. Bal, S.M., et al., *Microneedle-based transcutaneous immunisation in mice with N-trimethyl chitosan adjuvanted diphtheria toxoid formulations*. Pharmaceutical research, 2010. **27**(9): p. 1837-1847.
46. Ding, Z., et al., *Microneedle arrays for the transcutaneous immunization of diphtheria and influenza in BALB/c mice*. Journal of Controlled Release, 2009. **136**(1): p. 71-78.
47. Guo, L., et al., *Effective transcutaneous immunization against hepatitis B virus by a combined approach of hydrogel patch formulation and microneedle arrays*. Biomedical microdevices, 2013. **15**(6): p. 1077-1085.
48. Ding, Z., et al., *Immune modulation by adjuvants combined with diphtheria toxoid administered topically in BALB/c mice after microneedle array pretreatment*. Pharmaceutical research, 2009. **26**(7): p. 1635-1643.
49. Hirschberg, H., et al., *A combined approach of vesicle formulations and microneedle arrays for transcutaneous immunization against hepatitis B virus*. European Journal of Pharmaceutical Sciences, 2012. **46**(1): p. 1-7.
50. Yin, D., et al., *Hepatitis B DNA vaccine-polycation nano-complexes enhancing immune response by percutaneous administration with microneedle*. Biological and Pharmaceutical Bulletin, 2013. **36**(8): p. 1283-1291.
51. Carey, J.B., et al., *Microneedle-mediated immunization of an adenovirus-based malaria vaccine enhances antigen-specific antibody immunity and reduces anti-vector responses compared to the intradermal route*. Scientific reports, 2014. **4**.
52. Pearson, F.E., et al., *Induction of CD8+ T cell responses and protective efficacy following microneedle-mediated delivery of a live adenovirus-vectored malaria vaccine*. Vaccine, 2015.
53. Mikszta, J.A., et al., *Protective immunization against inhalational anthrax: a comparison of minimally invasive delivery platforms*. Journal of Infectious Diseases, 2005. **191**(2): p. 278-288.
54. Kim, Y.-C., et al., *Formulation and coating of microneedles with inactivated influenza virus to improve vaccine stability and immunogenicity*. Journal of controlled release, 2010. **142**(2): p. 187-195.
55. Kim, Y.-C., et al., *Formulation of microneedles coated with influenza virus-like particle vaccine*. AAPS PharmSciTech, 2010. **11**(3): p. 1193-1201.
56. Dean, C.H., et al., *Cutaneous delivery of a live, attenuated chimeric flavivirus vaccines against Japanese encephalitis (ChimeriVaxTM-JE) in non-human primates*. Human vaccines, 2005. **1**(3): p. 106-111.
57. Laurent, P.E., et al., *Evaluation of the clinical performance of a new intradermal vaccine administration technique and associated delivery system*. Vaccine, 2007. **25**(52): p. 8833-8842.
58. Laurent, P.E., et al., *Safety and efficacy of novel dermal and epidermal microneedle delivery systems for rabies vaccination in healthy adults*. Vaccine, 2010. **28**(36): p. 5850-5856.
59. Kim, Y.-C., et al., *Improved influenza vaccination in the skin using vaccine coated microneedles*. Vaccine, 2009. **27**(49): p. 6932-6938.
60. Kim, Y.-C., et al., *Enhanced memory responses to seasonal H1N1 influenza vaccination of the skin with the use of vaccine-coated microneedles*. Journal of Infectious Diseases, 2010. **201**(2): p. 190-198.
61. Kim, Y.-C., et al., *Cross-protection by co-immunization with influenza hemagglutinin DNA and inactivated virus vaccine using coated microneedles*. Journal of controlled release, 2013. **172**(2): p. 579-588.

62. Koutsonanos, D.G., et al., *Transdermal influenza immunization with vaccine-coated microneedle arrays*. PLoS one, 2009. **4**(3): p. e4773.
63. Zhu, Q., et al., *Immunization by vaccine-coated microneedle arrays protects against lethal influenza virus challenge*. Proceedings of the National Academy of Sciences, 2009. **106**(19): p. 7968-7973.
64. Weldon, W.C., et al., *Microneedle vaccination with stabilized recombinant influenza virus hemagglutinin induces improved protective immunity*. Clinical and Vaccine Immunology, 2011. **18**(4): p. 647-654.
65. Wang, B.-Z., et al., *Microneedle delivery of an M2e-TLR5 ligand fusion protein to skin confers broadly cross-protective influenza immunity*. Journal of controlled release, 2014. **178**: p. 1-7.
66. Song, J.-M., et al., *Microneedle delivery of H5N1 influenza virus-like particles to the skin induces long-lasting B-and T-cell responses in mice*. Clinical and Vaccine Immunology, 2010. **17**(9): p. 1381-1389.
67. Song, J.-M., et al., *DNA vaccination in the skin using microneedles improves protection against influenza*. Molecular Therapy, 2012. **20**(7): p. 1472-1480.
68. Song, J.-M., et al., *Improved protection against avian influenza H5N1 virus by a single vaccination with virus-like particles in skin using microneedles*. Antiviral research, 2010. **88**(2): p. 244-247.
69. Shin, J.-H., et al., *Microneedle Vaccination Elicits Superior Protection and Antibody Response over Intranasal Vaccination against Swine-Origin Influenza A (H1N1) in Mice*. PLoS one, 2015. **10**(6): p. e0130684.
70. Quan, F.-S., et al., *Stabilization of influenza vaccine enhances protection by microneedle delivery in the mouse skin*. PLoS one, 2009. **4**(9): p. e7152.
71. Quan, F.-S., et al., *Intradermal vaccination with influenza virus-like particles by using microneedles induces protection superior to that with intramuscular immunization*. Journal of virology, 2010. **84**(15): p. 7760-7769.
72. Quan, F.-S., et al., *Long-term protective immunity from an influenza virus-like particle vaccine administered with a microneedle patch*. Clinical and Vaccine Immunology, 2013. **20**(9): p. 1433-1439.
73. Quan, F.-S., et al., *Dose sparing enabled by skin immunization with influenza virus-like particle vaccine using microneedles*. Journal of controlled release, 2010. **147**(3): p. 326-332.
74. Koutsonanos, D.G., et al., *Delivery of subunit influenza vaccine to skin with microneedles improves immunogenicity and long-lived protection*. Scientific reports, 2012. **2**.
75. Kim, Y.-C., et al., *Increased immunogenicity of avian influenza DNA vaccine delivered to the skin using a microneedle patch*. European Journal of Pharmaceutics and Biopharmaceutics, 2012. **81**(2): p. 239-247.
76. Kim, M.-C., et al., *Microneedle patch delivery to the skin of virus-like particles containing heterologous M2e extracellular domains of influenza virus induces broad heterosubtypic cross-protection*. Journal of controlled release, 2015.
77. del Pilar Martin, M., et al., *Local response to microneedle-based influenza immunization in the skin*. MBio, 2012. **3**(2): p. e00012-12.
78. Corbett, H.J., et al., *Skin vaccination against cervical cancer associated human papillomavirus with a novel micro-projection array in a mouse model*. PLoS One, 2010. **5**(10): p. e13460.
79. Kines, R.C., et al., *Vaccination with Human Papillomavirus Pseudovirus-Encapsidated Plasmids Targeted to Skin Using Microneedles*. PLoS one, 2015. **10**(3).
80. Prow, T.W., et al., *Nanopatch-Targeted Skin Vaccination against West Nile Virus and Chikungunya Virus in Mice*. Small, 2010. **6**(16): p. 1776-1784.
81. Moon, S., et al., *Dose sparing and enhanced immunogenicity of inactivated rotavirus vaccine administered by skin vaccination using a microneedle patch*. Vaccine, 2013. **31**(34): p. 3396-3402.

82. Chen, X., et al., *Improved DNA vaccination by skin-targeted delivery using dry-coated densely-packed microprojection arrays*. Journal of controlled release, 2010. **148**(3): p. 327-333.
83. Gill, H.S., et al., *Cutaneous vaccination using microneedles coated with hepatitis C DNA vaccine*. Gene therapy, 2010. **17**(6): p. 811-814.
84. Kommareddy, S., et al., *Influenza subunit vaccine coated microneedle patches elicit comparable immune responses to intramuscular injection in guinea pigs*. Vaccine, 2013. **31**(34): p. 3435-3441.
85. Hiraishi, Y., et al., *Bacillus Calmette-Guerin vaccination using a microneedle patch*. Vaccine, 2011. **29**(14): p. 2626-2636.
86. Andrianov, A.K., et al., *Poly [di (carboxylatophenoxy) phosphazene] is a potent adjuvant for intradermal immunization*. Proceedings of the National Academy of Sciences, 2009. **106**(45): p. 18936-18941.
87. Edens, C., et al., *Measles vaccination using a microneedle patch*. Vaccine, 2013. **31**(34): p. 3403-3409.
88. van der Maaden, K., et al., *Layer-by-layer assembly of inactivated poliovirus and N-trimethyl chitosan on pH-sensitive microneedles for dermal vaccination*. Langmuir, 2015. **31**(31): p. 8654-8660.
89. Troy, S.B., et al., *Comparison of the Immunogenicity of Various Booster Doses of Inactivated Polio Vaccine Delivered Intradermally Versus Intramuscularly to HIV-Infected Adults*. Journal of Infectious Diseases, 2015. **211**(12): p. 1969-1976.
90. Anand, A., et al., *Early priming with inactivated poliovirus vaccine (IPV) and intradermal fractional dose IPV administered by a microneedle device: A randomized controlled trial*. Vaccine, 2015.
91. Atmar, R.L., S.M. Patel, and W.A. Keitel, *Intanza®: a new intradermal vaccine for seasonal influenza*. 2010.
92. Leroux-Roels, I. and F. Weber, *Intanza® 9 µg intradermal seasonal influenza vaccine for adults 18 to 59 years of age*. Human vaccines & immunotherapeutics, 2013. **9**(1): p. 115-121.
93. Huang, J., et al., *Protective immunity in mice achieved with dry powder formulation and alternative delivery of plague F1-V vaccine*. Clinical and Vaccine Immunology, 2009. **16**(5): p. 719-725.
94. van der Maaden, K., et al., *Novel hollow microneedle technology for depth-controlled microinjection-mediated dermal vaccination: a study with polio vaccine in rats*. Pharmaceutical research, 2014. **31**(7): p. 1846-1854.
95. Hirobe, S., et al., *Clinical study and stability assessment of a novel transcutaneous influenza vaccination using a dissolving microneedle patch*. Biomaterials, 2015. **57**: p. 50-58.
96. Kommareddy, S., et al., *Dissolvable microneedle patches for the delivery of cell-culture-derived influenza vaccine antigens*. Journal of pharmaceutical sciences, 2012. **101**(3): p. 1021-1027.
97. Matsuo, K., et al., *Transcutaneous immunization using a dissolving microneedle array protects against tetanus, diphtheria, malaria, and influenza*. Journal of controlled release, 2012. **160**(3): p. 495-501.
98. Sullivan, S.P., et al., *Dissolving polymer microneedle patches for influenza vaccination*. Nature medicine, 2010. **16**(8): p. 915-920.
99. Vassilieva, E.V., et al., *Improved immunogenicity of individual influenza vaccine components delivered with a novel dissolving microneedle patch stable at room temperature*. Drug delivery and translational research, 2015: p. 1-12.
100. Wang, J., B. Li, and M.X. Wu, *Effective and lesion-free cutaneous influenza vaccination*. Proceedings of the National Academy of Sciences, 2015. **112**(16): p. 5005-5010.
101. Qiu, Y., et al., *DNA-based vaccination against hepatitis B virus using dissolving microneedle arrays adjuvanted by cationic liposomes and CpG ODN*. Drug delivery, 2015(0): p. 1-8.

102. Wang, T., et al., *Mannosylated and lipid A-incorporating cationic liposomes constituting microneedle arrays as an effective oral mucosal HBV vaccine applicable in the controlled temperature chain*. *Colloids and Surfaces B: Biointerfaces*, 2015. **126**: p. 520-530.
103. Pattani, A., et al., *Microneedle mediated intradermal delivery of adjuvanted recombinant HIV-1 CN54gp140 effectively primes mucosal boost inoculations*. *AIDS Research and Human Retroviruses*, 2011. **27**(10).
104. Edens, C., et al., *A microneedle patch containing measles vaccine is immunogenic in non-human primates*. *Vaccine*, 2015.
105. Edens, C., et al., *Inactivated polio vaccination using a microneedle patch is immunogenic in the rhesus macaque*. *Vaccine*, 2015.
106. Gill, H.S. and M.R. Prausnitz, *Coated microneedles for transdermal delivery*. *Journal of controlled release*, 2007. **117**(2): p. 227-237.
107. Vrdoljak, A., et al., *Coated microneedle arrays for transcutaneous delivery of live virus vaccines*. *Journal of Controlled Release*, 2012. **159**(1): p. 34-42.
108. Wonglertnirant, N., et al., *Macromolecular delivery into skin using a hollow microneedle*. *Biological and Pharmaceutical Bulletin*, 2010. **33**(12): p. 1988-1993.
109. Davis, S.P., et al., *Hollow metal microneedles for insulin delivery to diabetic rats*. *Biomedical Engineering, IEEE Transactions on*, 2005. **52**(5): p. 909-915.
110. Harvey, A.J., et al., *Microneedle-based intradermal delivery enables rapid lymphatic uptake and distribution of protein drugs*. *Pharmaceutical research*, 2011. **28**(1): p. 107-116.
111. Sullivan, S.P., N. Murthy, and M.R. Prausnitz, *Minimally invasive protein delivery with rapidly dissolving polymer microneedles*. *Advanced Materials*, 2008. **20**(5): p. 933-938.
112. Lee, J.W., J.-H. Park, and M.R. Prausnitz, *Dissolving microneedles for transdermal drug delivery*. *Biomaterials*, 2008. **29**(13): p. 2113-2124.
113. Prausnitz, M.R. and R. Langer, *Transdermal drug delivery*. *Nature biotechnology*, 2008. **26**(11): p. 1261-1268.
114. Lahiji, S.F., M. Dangol, and H. Jung, *A patchless dissolving microneedle delivery system enabling rapid and efficient transdermal drug delivery*. *Scientific reports*, 2015. **5**.
115. Donnelly, R.F., T.R.R. Singh, and A.D. Woolfson, *Microneedle-based drug delivery systems: microfabrication, drug delivery, and safety*. *Drug delivery*, 2010. **17**(4): p. 187-207.
116. McGrath, M.G., et al., *Production of dissolvable microneedles using an atomised spray process: Effect of microneedle composition on skin penetration*. *European Journal of Pharmaceutics and Biopharmaceutics*, 2014. **86**(2): p. 200-211.
117. Chu, L.Y., S.O. Choi, and M.R. Prausnitz, *Fabrication of dissolving polymer microneedles for controlled drug encapsulation and delivery: bubble and pedestal microneedle designs*. *Journal of pharmaceutical sciences*, 2010. **99**(10): p. 4228-4238.
118. Lutton, R.E., et al., *Microneedle characterisation: the need for universal acceptance criteria and GMP specifications when moving towards commercialisation*. *Drug delivery and translational research*, 2015: p. 1-19.
119. WHO. *Vaccine-preventable diseases*. 2015 [cited 2015 24th November]; Available from: [http://apps.who.int/immunization\\_monitoring/diseases/en/](http://apps.who.int/immunization_monitoring/diseases/en/).
120. Mikszta, J.A., et al., *Microneedle-based intradermal delivery of the anthrax recombinant protective antigen vaccine*. *Infection and immunity*, 2006. **74**(12): p. 6806-6810.
121. Jansen, A.G., et al., *Influenza-and respiratory syncytial virus-associated mortality and hospitalisations*. *European Respiratory Journal*, 2007. **30**(6): p. 1158-1166.
122. Rivetti, D., et al., *Vaccines for preventing influenza in the elderly*. *The Cochrane Library*, 2006.
123. Thompson, W.W., et al., *Influenza-associated hospitalizations in the United States*. *Jama*, 2004. **292**(11): p. 1333-1340.
124. Zhou, H., et al., *Hospitalizations associated with influenza and respiratory syncytial virus in the United States, 1993–2008*. *Clinical infectious diseases*, 2012. **54**(10): p. 1427-1436.

125. Chaves, S.S., et al., *The US influenza hospitalization surveillance network*. Emerg Infect Dis, 2015. **9**: p. 1543-50.
126. Reed, C., et al., *Estimating Influenza Disease Burden from Population-Based Surveillance Data in the United States*. PloS one, 2015. **10**(3): p. e0118369.
127. Treanor, J., *Influenza vaccine—outmaneuvering antigenic shift and drift*. New England Journal of Medicine, 2004. **350**(3): p. 218-220.
128. Demicheli, V., et al., *Vaccines for preventing influenza in healthy adults*. Cochrane Database Syst Rev, 2007. **2**.
129. Executive Board, *Prevention and control of influenza pandemics and annual epidemics 2003*.
130. Blank, P.R., M. Schwenkglens, and T.D. Szucs, *Vaccination coverage rates in eleven European countries during two consecutive influenza seasons*. Journal of Infection, 2009. **58**(6): p. 446-458.
131. *CDC: Flu Vaccination Coverage, United States, 2014-15 Influenza Season*. 2015 [cited 2015 20th October]; Available from: <http://www.cdc.gov/flu/fluview/coverage-1415estimates.htm>.
132. Noh, J.Y. and W.J. Kim, *Influenza vaccines: unmet needs and recent developments*. Infection & chemotherapy, 2013. **45**(4): p. 375-386.
133. Norman, J.J., et al., *Microneedle patches: usability and acceptability for self-vaccination against influenza*. Vaccine, 2014. **32**(16): p. 1856-1862.
134. Miller, M.A. and J.T. Sentz, *Vaccine-preventable diseases*. 2006.
135. Ren, Q., et al., *Evaluation of an outside-the-cold-chain vaccine delivery strategy in remote regions of western China*. Public health reports, 2009. **124**(5): p. 745.
136. Wang, L., et al., *Hepatitis B vaccination of newborn infants in rural China: evaluation of a village-based, out-of-cold-chain delivery strategy*. Bulletin of the World Health Organization, 2007. **85**(9): p. 688-694.
137. Dowling, J.M. and C.F. Yap, *Happiness and poverty in developing countries: A global perspective* 2012: Palgrave Macmillan.
138. Naicker, S., et al., *Shortage of healthcare workers in developing countries--Africa*. Ethnicity & disease, 2009. **19**(1): p. 60.
139. Wolfe, D.N., W. Florence, and P. Bryant, *Current biodefense vaccine programs and challenges*. Human vaccines & immunotherapeutics, 2013. **9**(7): p. 1591-1597.
140. Moga, K.A., et al., *Rapidly-dissolvable microneedle patches via a highly scalable and reproducible soft lithography approach*. Advanced Materials, 2013. **25**(36): p. 5060-5066.
141. Lutton, R.E., et al., *A novel scalable manufacturing process for the production of hydrogel-forming microneedle arrays*. International journal of pharmaceuticals, 2015. **494**(1): p. 417-429.
142. Weldon, W.C., et al., *Effect of adjuvants on responses to skin immunization by microneedles coated with influenza subunit vaccine*. PloS one, 2012. **7**(7): p. e41501.