



TITLE:

Ambient fine and coarse particles in Japan affect nasal and bronchial epithelial cells differently and elicit varying immune response

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1 **Ambient fine and coarse particles in Japan affect nasal and bronchial epithelial cells**
2 **differently and elicit varying immune response**

3

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24 **Abstract**

25 Ambient particulate matter (PM) epidemiologically exacerbates respiratory and immune
26 health, including allergic rhinitis (AR) and bronchial asthma (BA). Although fine and coarse
27 particles can affect respiratory tract, the differences in their effects on the upper and lower
28 respiratory tract and immune system, their underlying mechanism, and the components
29 responsible for the adverse health effects have not been yet completely elucidated. In this
30 study, ambient fine and coarse particles were collected at three different locations in Japan
31 by cyclone technique. Both particles collected at all locations decreased the viability of nasal
32 epithelial cells and antigen presenting cells (APCs), increased the production of IL-6, IL-8,
33 and IL-1 β from bronchial epithelial cells and APCs, and induced expression of dendritic and
34 epithelial cell (DEC) 205 on APCs. Differences in inflammatory responses, but not in
35 cytotoxicity, were shown between both particles, and among three locations. Some
36 components such as Ti, Co, Zn, Pb, As, OC (organic carbon) and EC (elemental carbon)
37 showed significant correlations to inflammatory responses or cytotoxicity. These results
38 suggest that ambient fine and coarse particles differently affect nasal and bronchial epithelial
39 cells and immune response, which may depend on particles size diameter, chemical
40 composition and source related particles types.

41

42 **Keywords:** ambient particulate matter, cyclone technique, respiratory cells, immune cells,
43 inflammatory responses

44 **Capsule:** We showed for the first time in the world that ambient fine and coarse particles

45 collected from Japan by the new technique using cyclone have different effects on the
46 epithelium cells of the upper and lower respiratory tract and elicit varying immune response,
47 which may depend on particles size diameter, chemical composition and source related
48 particles types.

49

50 **1. Introduction**

51 The health effect of ambient particulate matter (PM) is still a problem worldwide. PM is a
52 complex mixture of particles having different chemical components such as solid and liquid
53 materials that contain elemental carbon (EC), organic carbon (OC), inorganic salts, and
54 metals and biological components such as endotoxin and β -glucan and has a compound effect
55 on biological reactions (Schins et al., 2004; Cachon et al., 2014; Honda et al., 2017).
56 Generally, the fine fraction of PM (aerodynamic diameter $< 2.5 \mu\text{m}$) in urban atmosphere is
57 a complex mixture of primary particles emitted from combustion sources and secondary
58 particles that form in the atmosphere from gaseous components (Marcazzan et al., 2001;
59 Sharma et al., 2007; Sevastyanova et al., 2008; Zerbi et al., 2008). The coarse fraction of PM
60 (aerodynamic diameter $> 2.5 \mu\text{m}$) generally includes mineral particles of crustal material, sea
61 salt particles, fly ash, and adsorbed species such as endotoxin (Schins et al., 2004; Perez et
62 al., 2007). These components can differ depending on the sources, geographical areas, and
63 seasons. In addition, PM composition depends on factors such as atmospheric photo-
64 chemical reaction and physical redistribution (Vecchi et al., 2004; Samoli et al., 2008).
65 PM epidemiologically exacerbates respiratory and immune health such as allergic rhinitis
66 (AR) and bronchial asthma (BA) (Tecer et al., 2008) in addition to cardiovascular diseases

67 and cancer (Kappos et al., 2004). Clinically, AR and BA have a close relationship: about 80%
68 of patients with BA have complications of AR (Bachert et al., 2002). In general, coarse
69 particles and limited fine particles can affect upper respiratory tract, whereas fine particles
70 and limited coarse particles can affect lower respiratory tract (Heyder J., 1986). However, the
71 difference in the effects of fine and coarse particles on the upper or lower respiratory tract
72 and immune responses related to them, as well as their underlying mechanisms have not yet
73 been clarified. Moreover, the components of PM responsible for the adverse health effects
74 have not yet been elucidated owing to their complexity (Lindbom et al., 2006; Hong et al.,
75 2016).

76 A large amount of fine and coarse particles is needed to evaluate the adverse health effects
77 by *in vivo* and/or *in vitro* studies. However, it is difficult to collect a sufficient amount of PM
78 by conventional filter collection method with extraction. Because of different extraction
79 efficiency and loss of PM constituents, the exposure experiment using PM extracts has a
80 possibility that would not reflect the actual biological response. Our previous study disclosed
81 extracts efficiency of PM_{2.5} and discussed the problem (Chowdhury et al., 2018). On the
82 other hand, the cyclone technique enables collection of a sufficient amount of PM (fine and
83 coarse particles themselves) for *in vivo* and/or *in vitro* assays enabling the analysis of the
84 effects of ambient particles on respiratory health without the use of a filter or extraction
85 process (Okuda et al., 2015, 2018).

86 In this study, we investigated the effects of ambient fine and coarse particles collected at
87 three Japanese locations by cyclone technique on nasal epithelial cells (RPMI-2650),
88 bronchial epithelial cells (BEAS-2B), and bone marrow derived antigen presenting cells

89 (APCs) from NC/Nga mice. Our aim was to estimate the different effects of ambient fine and
90 coarse particles on respiratory and immune cells, their underlying mechanism, and the
91 components which can be responsible for the respiratory and immune health such as AR and
92 BA.

93

94 **2. Materials and Methods**

95 **2.1. Sampling of PM**

96 Samples of fine and coarse particles were collected at an urban area in Fukuoka City, at a
97 suburban of the metropolitan area in Kazo City, Saitama Prefecture (Saitama), and a capital
98 area in Yokohama City in Japan (Suppl. Figure S1) during February to March 2017. The
99 particles as references were obtained by National Institute for Environmental Studies in Japan.
100 One reference (CRM#8) is ethanol-treated vehicle exhaust particulates (Okamoto., 1987) and
101 another (CRM#28) is irradiated atmospheric dust collected by a ventilation filter of the
102 building in Beijing (Mori et al., 2008). Okuda (2013) has indicated CRM#8 consists mainly
103 of fine (or ultrafine) particles, while CRM#28 consist mainly of coarse particles.
104 The collection was conducted with a high-volume PM sampler using the virtual impactor and
105 cyclone technique with no filter or extraction process (Okuda et al., 2018). The air flow
106 volume per given time for the inlet (virtual impactor) is 1,300 L/min. The total volume of air
107 sampled was determined from the measured volumetric flow rate and the sampling time. The
108 mass concentration of particles in the ambient air was computed as the total mass of collected
109 particles divided by the total volume of air sampled. After sampling, the particles in the amber
110 bottles were collected using a stainless spatula. We previously confirmed size distribution

111 and morphology of ambient particles collected by cyclone (Suppl. Figure S2). Particles were
112 dissolved in sterile phosphate-buffered saline (PBS) and ultrasonicated at the concentration
113 of 10 mg/mL. Finally, we adjusted at concentrations of 0, 7.5, and 75 $\mu\text{g/mL}$ using medium,
114 PBS (1%) and Dimethyl sulfoxide (DMSO) (0.1%) for the cell exposure experiment in this
115 study. Medium for BEAS-2B cells is serum-free. Similarly, we did not add serum in medium
116 for RPMI-2650 cells to evaluate under the same condition of exposure.

117

118 **2.2. Chemical, mineralogical and biochemical investigation**

119 The collected particles was characterized by ion chromatography for Anion species (Cl^- , NO_3^- ,
120 and SO_4^{2-}) and cation species (Na^+ , NH_4^+ , K^+ , Mg^{2+} , and Ca^{2+}), thermal-optical method
121 (IMPROVE protocol) for OC1-4 and EC1-3, high performance liquid chromatography
122 (HPLC) for polycyclic aromatic hydrocarbons (PAHs) (Chrysene, Benz[a]anthracene,
123 Benzo[b]fluoranthene, Benzo[k]fluoranthene, and Benzo[a]pyrene), and inductively coupled
124 plasma mass spectrometry (ICP-MS) for metals (Al, Si, Ti, V, Cr, Mn, Fe, Ni, Cu, Zn and
125 Pb). The procedure of chemical characterization mentioned above were generally described
126 in several previous papers (Okuda., 2013; Okuda et al., 2013, 2014). Endotoxin and β -glucan
127 have induced inflammatory responses from respiratory cells and immune cells (Veranth et al.,
128 2004; Carmona et al., 2010; Neveu et al., 2011). In this study, we investigated the effect of
129 endotoxin and β -glucan as substances derived from biological components in PM. We
130 performed an endotoxin test and a β -glucan test (both from Associates of Cape Cod,
131 Falmouth, MA, USA) following the manufacturer's instructions.

132

133 **2.3. Cell Cultures and PM exposure**

134 **2.3.1. Upper and Lower Respiratory cells**

135 The RPMI-2650, derived from squamous cell carcinoma of nasal septum was used as model
136 of human nasal epithelial cells which are cells of the upper respiratory tract. These cells
137 display consistent growth and high stability throughout continued culturing *in vitro* with no
138 alteration to the normal diploid karyotype (Moorhead, 1965). The cell line was purchased
139 from the European Collection of Cell Cultures (Salisbury, Wiltshire, United Kingdom) and
140 maintained in Eagle's minimal essential medium (DS Pharma Biomedicals, Osaka, Japan)
141 supplemented with 10% heat-inactivated fetal bovine serum (MP Biomedicals, Eschwege,
142 Germany), 2 mM L-glutamine, 100 U/mL penicillin and 100 µg/mL streptomycin (Sigma, St
143 Louis, Missouri). As representative of the cells of the lower respiratory tract, the BEAS-2B,
144 derived from human bronchial epithelial cells, was purchased from the European Collection
145 of Cell Cultures and maintained in LHC-9 medium (Thermo Scientific, Waltham,
146 Massachusetts) which is serum-free medium containing Gentamicin. RPMI-2650 cells and
147 BEAS-2B cells were maintained by subculture in 37°C at 5% CO₂ in medium.

148

149 **2.3.2. Immune cells**

150 Ten-week-old male SPF NC/NgaTndCrlj mice were purchased from Charles River (Osaka,
151 Japan). NC/Nga mice are atopy-prone mice. APCs were obtained after sacrificing mice by
152 cervical dislocation and exsanguination. The procedures used in all animal studies were
153 approved by the Animal Research Committee at Kyoto University. APCs were differentiated
154 using a modification of the protocol provided by Lutz et al (1999). We confirmed APCs by

155 the expression of about 80% of CD11c which is a molecule specifically expressed in dendritic
156 cells. Bone marrow cells (4×10^5 /mL) were cultured in R10 which is RPMI 1640 (Thermo
157 Scientific) supplemented with 10% heat-inactivated fetal bovine serum (MP Biomedicals),
158 100 U/mL penicillin, 100 μ g/mL streptomycin (Sigma), and 50 mM 2-mercaptoethanol
159 (Thermo Scientific) containing Granulocyte-Macrophage Colony-Stimulating Factor (GM-
160 CSF).

161

162 **2.4. Experimental Protocol**

163 The RPMI-2650 cells, BEAS-2B cells, and APCs were exposed to ambient fine and coarse
164 particles at concentrations of 0, 7.5, or 75 μ g/mL and reference particles at 75 μ g/mL only
165 for 24 h. We evaluated the cell viability, the cytokine release, and dendritic and epithelial
166 cell (DEC) 205 on the cell surface. All control cells were treated with each medium.

167

168 **2.4.1. Cell Viability**

169 We measured the viability of the RPMI-2650 cells, BEAS-2B cells, and APCs by WST-1
170 assay using the Premix WST-1 Cell Proliferation Assay System (TaKaRa Bio, Shiga, Japan)
171 as previously described (Honda et al., 2017). The results are expressed as the percentage of
172 exposed group to control cells (0 μ g/mL).

173

174 **2.4.2. Quantification of Pro-Inflammatory Cytokines in the Culture Supernatants**

175 The amounts of IL-6 and IL-8 release in the supernatants from the RPMI-2650 cells and
176 BEAS-2B cells and those of IL-6 and IL-1 β release in the supernatants from APCs were

177 measured by ELISA (Thermo Scientific), according to the manufacturer's protocol as
178 previously described (Honda et al., 2017). The detection limits of IL-6 and IL-8 from RPMI-
179 2650 cells and BEAS-2B cells, and IL-6 and IL-1 β from APCs were <2.2 pg/mL, <9.8 pg/mL,
180 <1.9 pg/mL, <1.6 pg/mL, <16 pg/mL and 10 pg/mL, respectively.

181

182 ***2.4.3. Expression of DEC205 on APCs cell surface***

183 We measured the expression of DEC205 on the APCs' surface by the FACS analysis, the
184 following monoclonal antibodies were used: Mouse BD Fc Block purified anti-mouse
185 CD16/CD32 (Becton Dickinson), DEC205 (NLDC-145, PE-conjugated; Bio-Legend, San
186 Diego, California), Rat IgG2a, k Isotype Control (RTK2758, PE-conjugated; BioLegend).
187 The fluorescence was measured by a FACSCalibur (Becton Dickinson) as previously
188 described (Honda et al., 2017).

189

190 **2.5. Statistical Analysis**

191 The data are presented as the mean \pm standard error of the mean (SEM) for each experimental
192 group (n =3 or 4). The significance of variation among different groups was determined by
193 one-way analysis of variance. Differences among groups were analyzed using Tukey's
194 multiple comparison test. A P-value < 0.05 was considered to indicate a significant difference.
195 Relationships between components in PM and cell viability or cytokine release were tested
196 using Pearson's correlation, with a two-tailed significance study using SPSS software. A P <
197 0.01 and R > 0.9 was shown as a high degree of correlation.

198

199 3. Results

200 3.1. The characterization of collected ambient particles

201 This cyclone system achieved 50% collection efficiency with components having the
202 following aerodynamic cut-off diameters: virtual impactor, 2.4 μm ; fine-particle cyclone,
203 0.18-0.30 μm ; and coarse-particle cyclone, 0.7 μm . Particles smaller than 2.4 μm flowed to
204 the fine side at the virtual impactor part, and thus, fine particles were 0.30-2.4 μm and coarse
205 particles were 2.4 μm or more in size. The mean concentrations of fine particles in Fukuoka,
206 Saitama, and Yokohama were 3.0, 5.9 and 9.9 $\mu\text{g}/\text{m}^3$, respectively while those of coarse
207 particles were 1.5, 4.3 and 14.2 $\mu\text{g}/\text{m}^3$, respectively. Note that these mass concentrations of
208 fine and coarse particles were expressed as the weights of particles collected by the cyclones
209 per sampled air volume. The concentrations of components of both particles in the prepared
210 solution are shown (Suppl. Table S1-S3). Both particles had different proportions of metal
211 components between Fukuoka and the other locations. In Fukuoka, instead of a small amount
212 of metal components, amounts of Na^+ , Cl^- and SO_4^{2-} were high (Suppl. Figure S3).

213 The content of endotoxin in fine and coarse particles was 0.080 and 0.060 (Fukuoka), 2.50
214 and 2.97 (Saitama), 4.29 and 6.59 (Yokohama) EU/mL, respectively. Endotoxin in both
215 particles at Fukuoka was lower than those collected at Saitama and Yokohama. The highest
216 content of endotoxin was found in coarse particles at Yokohama. The contents of β -glucan
217 in fine and coarse particles were 317.3 and 1106 (Fukuoka), 294.4 and 666.0 (Saitama), 774.6
218 and 1060 (Yokohama) pg/mL, respectively. β -glucan in fine particles collected at all
219 locations was lower than those in coarse particles at each location. The highest content of β -
220 glucan was present in coarse particles at Fukuoka.

221

222 3.2. Biological effects of ambient particles on the Nasal Epithelial Cells

223 We examined the effects of exposure (24 h) to ambient fine and coarse particles on the
224 viability of RPMI-2650 cells. A significant decrease ($p < 0.01$ vs. control) in the viability of
225 cells was seen upon exposure to both particles collected at all locations in a concentration-
226 dependent manner as compared to the control (unexposed cells). No difference in cytotoxicity
227 caused by ambient fine and coarse particles from each location was detected except for
228 sample collected at Yokohama at a concentration of $7.5 \mu\text{g/mL}$ (Figure 1).

229 We investigated the effects of 24 h exposure of ambient fine and coarse particles on the pro-
230 inflammatory responses via release of IL-6 and IL-8 from RPMI-2650 cells. None of the
231 particles, at any location were able to evoke detectable cytokine release from RPMI-2650
232 cells.

233

234 3.3. Biological effects of ambient particles on the Bronchial Epithelial Cells

235 We examined the effects of exposure to ambient fine and coarse particles for 24 h on the
236 viability of BEAS-2B cells. BEAS-2B cells showed no decrease in viability upon exposure
237 to both particles collected at all locations at any concentration, when compared to control
238 (unexposed cells) (Figure 2A).

239 We investigated the effects of ambient fine and coarse particles on the pro-inflammatory
240 responses via release of IL-6 and IL-8 from BEAS-2B cells after exposure to each particle
241 for 24 h. Both particles collected at all locations increased IL-6 release, in a dose-dependent
242 manner, especially at the concentration of $75 \mu\text{g/mL}$ (Figure 2B; $p < 0.01$ vs. control).

243 Comparison between fine and coarse particles at the same location indicated that coarse
244 particles at Fukuoka induced higher production of IL-6 than fine particles (Figure 2B; $p <$
245 0.01 vs. fine particles at $75 \mu\text{g/mL}$). Comparison among the three locations indicated that
246 fine particles collected at Saitama or Yokohama had greater effect than those at Fukuoka
247 (Figure 2B; $p < 0.01$ vs. fine particles at Fukuoka at $75 \mu\text{g/mL}$). Coarse particles collected at
248 Fukuoka had significantly larger effect than those collected at Saitama (Figure 2B; $p < 0.05$
249 vs. coarse particles at Fukuoka at $75 \mu\text{g/mL}$). The production of IL-8 showed a pattern similar
250 to that of IL-6. The results of IL-8 different from IL-6 are shown. Coarse particles collected
251 at Fukuoka, fine particles collected at Saitama and both particles collected at Yokohama
252 significantly increased the levels of IL-8 at a concentration of $7.5 \mu\text{g/mL}$ (Figure 2B; $p < 0.05$
253 or 0.01 vs. control). The fine particles collected at Yokohama showed significantly marked
254 induction of IL-8 than coarse particles (Figure 2B; $p < 0.05$ vs. fine particles at $75 \mu\text{g/mL}$).
255 Fine particles collected at Yokohama had greater effect than those collected at Saitama
256 (Figure 2B; $p < 0.01$ vs. fine particles at Saitama at $75 \mu\text{g/mL}$).

257

258 **3.4. Biological effects of ambient particles on the APCs**

259 We examined the effects of ambient fine and coarse particles on the viability of APCs after
260 exposure to PM for 24 h. Exposure to both particles resulted in decrease in viability of APCs
261 in a dose-dependent manner as compared to control (unexposed cells) (Figure 3A). A
262 significant decrease in the viability of cells was seen upon exposure to both particles collected
263 at all locations at a concentration of $75 \mu\text{g/mL}$ and for those at Saitama and Yokohama at the
264 concentration of $7.5 \mu\text{g/mL}$ (Figure 3A; $p < 0.05$ or 0.01 vs. control). Coarse particles

265 collected at Fukuoka slightly decreased the viability of cells at a concentration of 7.5 $\mu\text{g}/\text{mL}$
266 (Figure 3A; $p < 0.05$ vs. control). No difference in cytotoxicity was detected between fine
267 and coarse particles collected at each location.

268 We investigated the effects of ambient fine and coarse particles on the pro-inflammatory
269 responses analyzed as IL-6 and IL-1 β released from APCs after exposure to each particle for
270 24 h. Both particles collected at all locations increased IL-6 release in a dose-dependent
271 manner especially at a concentration of 75 $\mu\text{g}/\text{mL}$ (Figure 3B; $p < 0.01$ vs. control). Both
272 particles collected at Saitama and Yokohama also significantly increased the levels of IL-6
273 at the concentration of 7.5 $\mu\text{g}/\text{mL}$ (Figure 3B; $p < 0.01$ vs. control). Coarse particles collected
274 at Fukuoka induced higher production of IL-6 than the fine particles (Figure 3B; $p < 0.01$ vs.
275 fine particles at 75 $\mu\text{g}/\text{mL}$). Both particles collected at Saitama and Yokohama induced higher
276 production of IL-6 than that collected at Fukuoka (Figure 3B; $p < 0.05$ or 0.01 vs. fine
277 particles at Fukuoka at 7.5 and 75 $\mu\text{g}/\text{mL}$). Both particles at all locations increased IL-1 β
278 release, in a dose –dependent manner, especially at the concentration of 75 $\mu\text{g}/\text{mL}$ similar to
279 what was observed in case of IL-6 (Figure 3B; $p < 0.01$ vs. control). Coarse particles collected
280 at Saitama and both particles at Yokohama at a concentration of 7.5 $\mu\text{g}/\text{mL}$ increased the
281 levels of IL-1 β (Fig 3B; $p < 0.05$ or 0.01 vs. control). Coarse particles collected at Fukuoka
282 induced lower production of IL-1 β than the fine particles (Figure 3B; $p < 0.05$ vs. fine
283 particles at 75 $\mu\text{g}/\text{mL}$). Both particles at Fukuoka resulted in greater induction of IL-1 β than
284 those at Saitama and Yokohama (Figure 3B; $p < 0.05$ or 0.01 vs. both particles at Fukuoka at
285 75 $\mu\text{g}/\text{mL}$, respectively).

286 The expression patterns of DEC205 in APCs were examined in order to evaluate the effects

287 of exposure to ambient fine and coarse particles for 24 h on the maturation and activation of
288 APCs. DEC205 is a member of the macrophage mannose receptor family. This molecule is
289 known to mediate the capture and internalization of ligands for subsequent processing and
290 presentation by APCs (Jiang et al., 1995). Both particles collected at all locations increased
291 the ratio of DEC205-positive cells at concentrations of 7.5 and 75 $\mu\text{g}/\text{mL}$ (Figure 3C; $p <$
292 0.01 vs. control). Although, there were no significant differences in the ratio of DEC205-
293 positive cells exposed to the both particles collected at the same location, fine particles
294 collected at Yokohama significantly increased the ratio of DEC205-positive cells when
295 compared with those at Fukuoka (Figure 3C; $p < 0.05$ vs. fine particles at Fukuoka at 7.5
296 $\mu\text{g}/\text{mL}$).

297

298 **3.5. Correlation between ambient particles components and biological responses**

299 Determination of the PM components responsible for impacts on examined cell lines by
300 assessing cytotoxicity, pro-inflammatory cytokine release is very important. We evaluated
301 Pearson's correlation coefficients between both particles compounds and the cell viability of
302 RPMI-2650 cells and APCs, and both particles compounds and cytokine release from BEAS-
303 2B cells and APCs. Our analysis showed negative correlations between the cell viability of
304 RPMI-2650 cells and Ca^{2+} , Zn, and OC (Suppl. Figure S4). Positive correlations were
305 observed between IL-6 and IL-8 release from BEAS-2B cells and IL-6 release from APCs
306 and multiple components, including EC, OC, and metals such as Ti and Co (Suppl. Figure
307 S5, S6A). Whereas, positive correlations between IL-1 β from APCs and multiple
308 components, including ions, EC, OC, and metals such as As and Pb were shown (Suppl.

309 Figure S6B).

310

311 **4. Discussion**

312 In the present study, we conducted experiments using ambient fine and coarse particles
313 collected by cyclone technique. Ambient both particles decreased the cell viability of RPMI-
314 2650 cells and APCs, and induced production of pro-inflammatory cytokines from BEAS-
315 2B cells and APCs. There was a significant difference in the inflammatory response elicited
316 by fine and coarse particles at Fukuoka. Inflammatory responses induced by particles
317 collected at Saitama and Yokohama were similar to each other but different from those
318 observed for particles collected at Fukuoka. Ambient both particles also induced expression
319 of DEC205 on APCs. Viability of RPMI-2650 cells correlated negatively with Ca^{2+} , Zn, and
320 OC3 and OC4. IL-6 and IL-8 release correlated positively with Ti, Fe, Co, Cr, Mn, V, and Zn,
321 and OC3, OC4, and EC2, whereas IL-1 β release correlated positively with As, Pb, and OC2
322 and EC1.

323 Ambient both particles at all locations decreased viability of RPMI-2650 cells but did not
324 induce pro-inflammatory cytokines. Nasal epithelial cells are the first epithelial barrier in the
325 nasal cavity imparting protection from inhaled xenobiotics. Previous studies have indicated
326 that diesel exhaust particles (DEP) or PM10 do not show cytotoxicity nor induce production
327 of pro-inflammatory cytokines from RPMI-2650 cells (Lindbom et al., 2006) and that DEP
328 does not reduce cell viability but decrease the barrier function by reducing zonula occludens
329 -1 (ZO-1) expression in RPMI-2650 cells (Fukuoka et al., 2015). If the barrier function
330 decreases even without cytotoxicity, there is a possibility that PM further weaken the barrier

331 function due to cytotoxicity. It is possible that particles collected in this study weakened the
332 barrier function, thereby allowing easy invasion by allergens.

333 It has been reported that primary nasal epithelial cells have lesser Toll-like receptor (TLR)
334 expression compared to alveolar epithelial cells, and more distribution of Toll-interacting
335 protein (TOLLIP; an inhibitor of TLR signaling) which may be one of the reasons for non-
336 induction of pro-inflammatory cytokines by these cells (Moncayo et al., 2014). The low
337 inflammatory responses due to the low expression of TLR and the high expression of
338 TOLLIP on nasal epithelial cells may lead to colonization and coexistence of many resident
339 bacterial groups in the nasal cavity environment. The epithelial cells in the nasal cavity are
340 always exposed to external environment and various bacteria or chemicals therein. Hence,
341 tolerance to these bacteria without induction of inflammation is required to maintain
342 homeostasis.

343 In case of BEAS-2B cells, though ambient both particles at all locations elevated the levels
344 of IL-6 and IL-8 release, none of them decreased the cell viability. Ambient both particles
345 strongly induced pro-inflammatory cytokine production compared to both reference particles.
346 Bronchial epithelial cells also act as a physical barrier and generate biological and
347 immunological responses against inhaled xenobiotics. IL-6 and IL-8 are the major pro-
348 inflammatory cytokines induced by response to environmental insults and are important key
349 molecules causing acute inflammation in the respiratory system by stimulating lymphocytes
350 (Thacker, 2006). Previous studies have indicated that various chemicals and allergens
351 stimulate IL-6 or IL-8 production from BEAS-2B cells (Honda et al., 2014; Totlandsdal et
352 al., 2012; Park et al., 2009). Ambient both particles collected by cyclone technique can

353 exacerbate allergic inflammation by inducing inflammatory cytokines.

354 Extracts from PM on filter have been used for experiments dealing with health effects of PM.

355 To our knowledge, this is the first experimental demonstration of the effects of allergic

356 inflammation by ambient fine and coarse particles collected by cyclone technique.

357 Fuentes-Mattei et al. (2010) have showed that PM_{2.5} organic extracts from urban area in

358 Puerto Rico at 50 µg/mL decrease about 20% of cell viability and produce about 650 pg/mL

359 of IL-6 and 235 pg/mL of IL-8 from BEAS-2B cells, but those from rural area do not decrease

360 cell viability and induce the lower production of IL-6 compared to those from urban area.

361 Gualtieri et al. (2010) have showed that PM_{2.5} and PM₁₀ aqueous extracts from urban area

362 in Milan at 10 µg/cm² (in this study, 75 µg/mL=13.7 µg/cm²) do not decrease cell viability

363 and produce about 80 pg/mL and 400 pg/mL of IL-8 from BEAS-2B, respectively. We have

364 previously indicated that exposure to PM_{2.5} collected in winter and subjected to organic

365 extraction rather than aqueous extraction causes an inflammatory response via IL-6

366 production from bronchial epithelial cells and PM_{2.5} extracts at Fukuoka subjected to

367 organic extraction, produced about 110 pg/mL of IL-6 at a concentration of 75 µg/mL which

368 was about 2 times greater than the level of control (Honda et al., 2017). These same extracts

369 collected in another season produced about 15 pg/mL of IL-6 and about 200 pg/mL of IL-8

370 from bronchial epithelial cells, and the levels of these cytokines were less than control

371 (Chowdhury et al., 2018). In the present experiment, fine particles collected by cyclone

372 technique at Fukuoka at 75 µg/mL produced about 250 pg/mL of IL-6 and about 670 pg/mL

373 of IL-8 in BEAS-2B cells. The levels of IL-6 and IL-8 produced by fine particles collected

374 using cyclone technique was about 6 and 2.5 times more than the level of control, respectively.

375 Although we could not strictly compare these results because it is not exactly the same
376 particle, it is suggested that ambient particles collected by cyclone technique could induce
377 larger inflammatory response than that induced by PM collected by the conventional filter
378 technique with extraction. Cyclone technique is an efficient method for collecting particles
379 that are subjected to exposure studies. Currently, there exist only a few assessments of health
380 effect conducted using ambient particles collected by cyclone technique and hence, further
381 research in this area should be encouraged (Ogino et al., 2017).

382 In the present study, ambient both particles collected at all locations induced expression of
383 DEC205 on APCs. The current study is the first report that ambient particles collected by
384 cyclone technique activates APCs via DEC205 expression. Several studies have shown that
385 carbon black nanoparticles and Asian dust particles can promote the maturation/activation
386 and function of DEC205 on APCs and may be related to their enhancing effects on allergic
387 diseases or responses (Honda et al., 2014; Koike et al., 2008). Ambient both particles induced
388 pro-inflammatory cytokines such as IL-6 and IL-1 β from APCs as well as BEAS-2B cells.
389 Ambient both particles strongly induced pro-inflammatory cytokine production compared to
390 both reference particles. IL-1 β has been described as a potent pro-inflammatory cytokine and
391 a mediator of a wide range of systemic human diseases (Dinarello, 2005; Koh et al., 2006;
392 Allantaz et al., 2007; James et al., 2011). The difference in reaction of IL-6 and IL-1 β
393 observed in this study may be due to the difference in the transcription factors of cytokines
394 or in the sensitivity to exposure components. APCs play important roles in allergens-related
395 airway inflammation (Lambrecht et al., 2012). APCs are activated upon invasion of the upper
396 or lower respiratory tract by ambient particles collected by cyclone technique. As a result,

397 ambient particles can affect allergic inflammation not only by induction DEC205 expression,
398 but also through induction of pro-inflammatory cytokines.

399 Some experimental and epidemiological studies have indicated that PM10 may exhibit a
400 similar or higher pro-inflammatory potential than PM2.5 and lead to adverse pulmonary
401 responses, which may require hospitalization (Monn et al., 1999; Becker et al., 2005;
402 Brunekreef et al., 2005; Gerlofs et al., 2007; Camatini et al., 2008). One of the reasons for
403 this may be attributed to the greater presence of microbial factors such as endotoxins or β -
404 glucan in PM10 as compared to PM2.5. Previous studies have shown endotoxin and β -glucan
405 to be associated with the inflammatory effects of PM both in *in vitro* and *in vivo* (Douwes et
406 al., 2003; Becker et al., 2005; Jalava et al., 2008). The U.S. Environmental Protection Agency
407 (1995) has noted that PM10 deposits in the upper airways of the lungs and may be more
408 relevant for asthmatic responses and irritation. On the other hand, PM2.5 is more often the
409 cause of lower respiratory symptoms such as cough and sputum compared to PM10
410 (Schwartz et al., 2000). In this way, although both PM can cause harmful health effects, the
411 one that causes greater harm has not been elucidated. In the present study, almost no
412 difference of cytotoxicity on RPMI-2650 cells and APCs was observed between the fine and
413 coarse particles. However, coarse particles collected at Fukuoka induced higher production
414 of IL-6 and IL-8 from BEAS-2B cells and IL-6 from APCs than fine particles, but lesser
415 production of IL-1 β from APCs. Fine particles collected at Yokohama induced higher
416 production of IL-8 from BEAS-2B cells than coarse particles. Our analysis did not show
417 strong correlations between pro-inflammatory cytokine release and microbial factors.
418 Previous studies have suggested that endotoxin at 2000 EU/mL and β glucan at the

419 concentration of “ $\mu\text{g}/\text{mL}$ ” induces pro-inflammatory responses in airway epithelial cells or
420 APCs (Carmona et al. 2010, Veranth et al. 2004). As the level of endotoxin and β glucan in
421 our study was very low compared to those of the previous studies, we suspect that the level
422 of endotoxin and β glucan in our study failed to noticeable correlation with inflammation in
423 the cells. Apart from microbial factors, previous studies have suggested that co-exposure of
424 SO_4^{2-} or constituent of cedar pollen enhances inflammatory responses (Hiyoshi et al., 2005;
425 Ichinose et al., 2005; Yamada et al., 2012). Not only biological factors but also other
426 compounds contained in PM or allergen substance can contribute to the production of pro-
427 inflammatory cytokines.

428 Yokohama is the most populated city and located in one of the three major industrial areas in
429 Japan. Saitama is close to Yokohama, and relatively the secondary particle tends to be the
430 main component of the formation of fine particles, so secondary air pollution is considered.
431 (Takegawa et al., 2006; Miyakawa et al., 2008). Yokohama and Saitama are located in
432 urban/suburban and industrial areas, respectively. Fe, Cr, Mn, Zn and Co are emitted by the
433 steel industry and V is mainly emitted by oil combustion (Lin et al., 2005; Querol et al., 2006;
434 Japan Ministry of the Environment, 2014). Titanium originates from the chemical industry
435 dealing with nanomaterial particles such as titanium dioxide (Chao et al., 2011). As these
436 metals in particles were correlated with IL-6 and IL-8 production in Yokohama and Saitama,
437 anthropogenic metals might contribute to respiratory disorders. On the other hand, Fukuoka
438 is closest to Mainland China, therefore there are a possibility that the influence of
439 transboundary pollution from China. (kaneyasu et al.,2014; Takami et al., 2016). It has been
440 reported that As and Pb are abundantly contained in coal which is the main energy source in

441 China (Hioki et al., 2009). Generally, atmospheric concentration of As in Japan is very low,
442 and there are few specific sources of As (Taniguchi et al. 2016). At least, the present study
443 demonstrated that As contained much in the particles in Fukuoka showed a high correlation
444 with IL-1 β release. This indicated that Fukuoka may be influenced by transboundary
445 pollution from China. However, the other biological responses induced by the particles
446 collected in Fukuoka was lower than the particles in other locations. Hence, particles caused
447 by transboundary contamination may have lower biological activity than particles having a
448 source in our country.

449 Previous studies have reported that metals and their compounds mentioned above induce IL-
450 6, IL-8 or IL-1 β *in vivo* and/or *in vitro* studies. For example, Ti or CoCl₂ induce the
451 production of IL-6 or IL-8 from bronchial epithelial cells or human lung microvascular
452 endothelial cells, and IL-1 β are induced from alveolar macrophages by intratracheal
453 administration of As in mice (Huaux et al., 1995; Carter et al., 1997; Schmalz et al., 1998;
454 Ming et al., 2000; Pascal et al., 2004; Kyoung et al., 2006; Eun-Jung et al., 2008, 2010). It is
455 possible that metals such as Ti, Fe, Cr, Mn, Co, V, Zn, Pb and As can induce IL-6, IL-8 or
456 IL-1 β from BEAS-2B cells and APCs, but further investigation is needed to identify the
457 responsible components. The components in OC and EC have not yet been well elucidated
458 (Grabowsky et al., 2011; Ikemori et al., 2009). However, low- and less- volatile organic
459 carbons (OC3 and OC4) may play an important role in the inflammatory reaction in each cell.
460 Previous studies have reported metals or ions such as Fe, Zn, Cr, Mn, V and Cu induce
461 toxicity of airway epithelial cells (Riley et al., 2003; Honda A., 2015). The concentrations of
462 Fe, Cr, V and Cu in our study was low compared to those of the previous studies, but Mn and

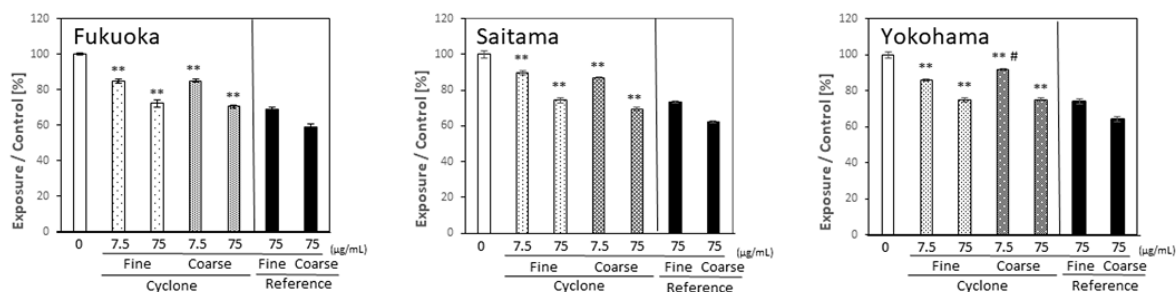
463 Zn was equivalent concentrations. In this study, we showed only high correlation between
464 Zn and toxicity, however it is a possible that the other metals such as Fe, Cr, Mn, V and Cu
465 also induce toxicity of airway epithelial cells.

466

467 5. Conclusion

468 Exposure to ambient particles collected by cyclone technique reduced cellular viability in
469 RPMI-2650 cells and APCs, induced pro-inflammatory responses in BEAS-2B cells and
470 APCs, and induced the maturation/activation of APCs. There was correlation between of
471 some chemical components and biological responses. These chemical components affected
472 differently between nasal and bronchial epithelial cells and elicited varying immune response.
473 In addition, these effects can differ depending on the diameter of the particles and/or
474 collection locations.

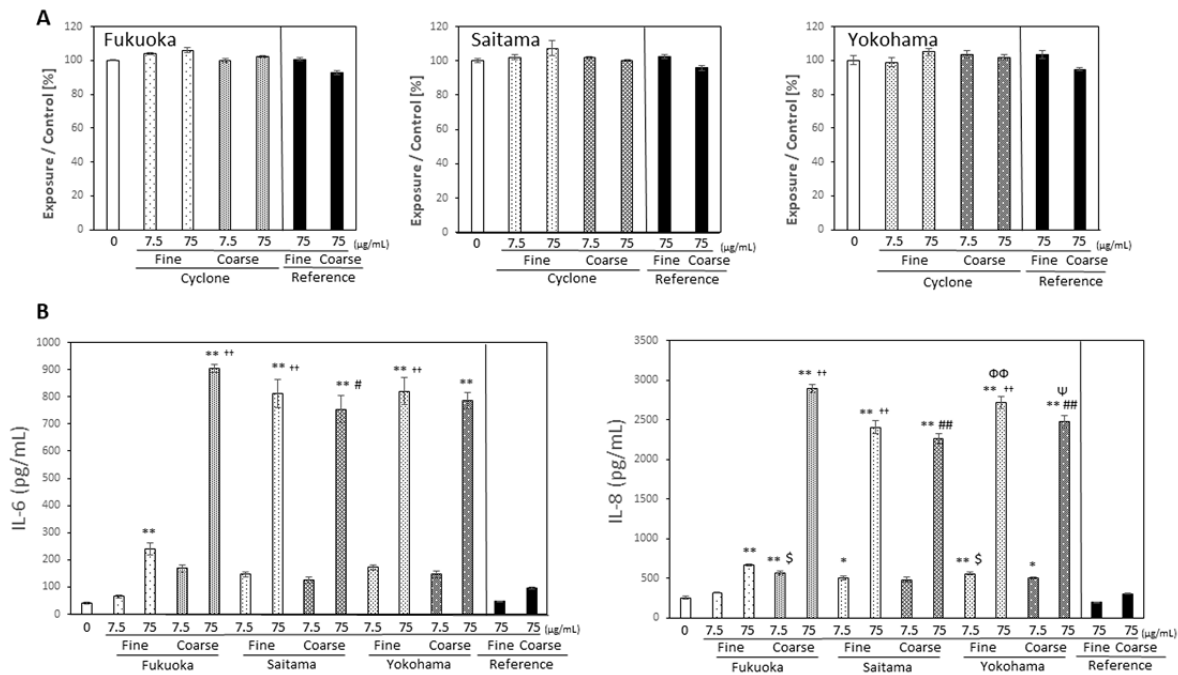
475



476

477 Figure 1. Effects of ambient particles and reference particles on the viability of RPMI-2650
478 cells. Data are presented as the percentage of the viability of the control. Data are mean \pm
479 standard error of the mean (SEM) of 4 individual cultures. * $P < 0.05$, ** $P < 0.01$ vs. 0 $\mu\text{g/mL}$,
480 # $P < 0.05$ vs. Fine particles at 7.5 $\mu\text{g/mL}$.

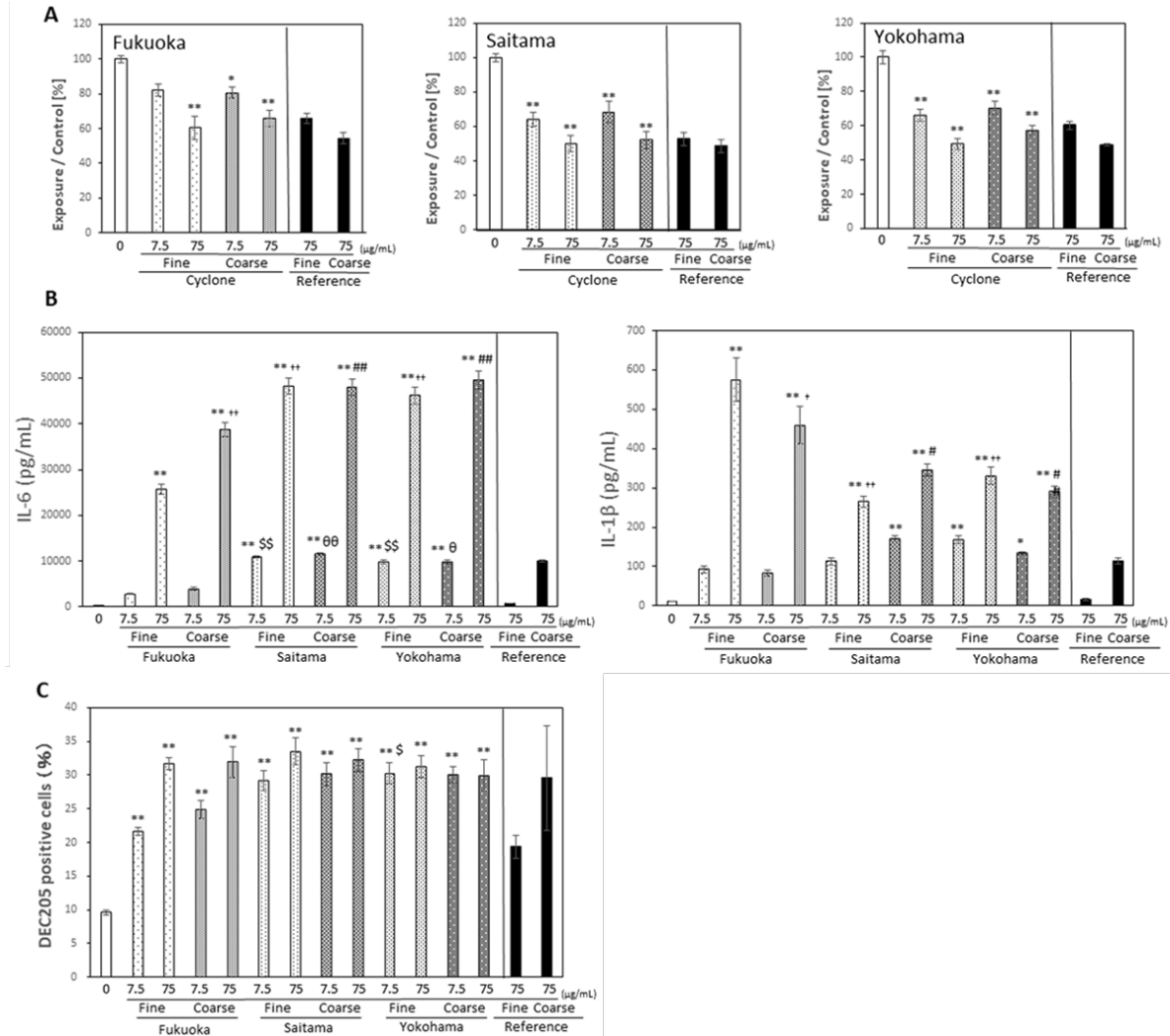
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482

483 Figure 2. (A): Effects of ambient particles and reference particles on the viability of BEAS-
 484 2B cells. Data are presented as the percentage of the viability of the control. (B): IL-6 and
 485 IL-8 production from BEAS-2B cells in response to ambient particles and reference particles.
 486 Data are mean \pm standard error of the mean (SEM) of 4 individual cultures. * $P < 0.05$,
 487 ** $P < 0.01$ vs. 0 $\mu\text{g/mL}$, $^{\$}P < 0.05$ vs. Fine particles at Fukuoka at 7.5 $\mu\text{g/mL}$, $^{\dagger\dagger}P < 0.01$ vs. Fine
 488 particles at Fukuoka at 75 $\mu\text{g/mL}$, $^{\#}P < 0.05$, $^{\#\#}P < 0.01$ vs. Coarse particles at Fukuoka at 75
 489 $\mu\text{g/mL}$, $^{\Phi\Phi}P < 0.01$ vs. Fine particles at Saitama at 75 $\mu\text{g/mL}$, $^{\Psi}P < 0.05$ vs. Fine particles at
 490 Yokohama at 75 $\mu\text{g/mL}$.

491



492

493 Figure 3. (A): Effects of ambient particles and reference particles on the viability of APCs.

494 Data are presented as the percentage of the viability of the control. (B):IL-6 and IL-1β

495 production from APCs in response to ambient particles and reference particles. (C): Effects

496 of ambient particles and reference particles on the DEC205 expression of APCs. Data are

497 presented as positive cells expressed % events. Data are mean ± standard error of the mean

498 (SEM) of 4 individual cultures. *P<0.05, **P<0.01 vs. 0 μg/mL, \$P<0.05, \$\$P<0.01 vs. Fine

499 particles at Fukuoka at 7.5 μg/mL, †P<0.05, ††P<0.01 vs. Fine particles at Fukuoka at 7.5

500 $\mu\text{g}/\text{mL}$, ⁰P<0.05, ⁰⁰P<0.01 vs. Coarse particles at Fukuoka at 7.5 $\mu\text{g}/\text{mL}$, [#]P<0.05, ^{##}P<0.01
501 vs. Coarse particles at Fukuoka at 75 $\mu\text{g}/\text{mL}$..

502

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516

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518

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520

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