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Citation for published version (APA):

Gosker, H. R., Wouters, E. F. M., van der Vusse, G. J., & Schols, A. M. W. J. (2000). Skeletal muscle dysfunction in chronic obstructive pulmonary disease and chronic heart failure: underlying mechanisms and therapy perspectives. *American Journal of Clinical Nutrition*, *71*(5), 1033-1047. https://doi.org/10.1093/ajcn/71.5.1033

Document status and date: Published: 01/01/2000

DOI: 10.1093/ajcn/71.5.1033

Document Version: Publisher's PDF, also known as Version of record

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Skeletal muscle dysfunction in chronic obstructive pulmonary disease and chronic heart failure: underlying mechanisms and therapy perspectives¹⁻³

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ABSTRACT Low exercise tolerance has a large influence on health status in chronic obstructive pulmonary disease and chronic heart failure. In addition to primary organ dysfunction, impaired skeletal muscle performance is a strong predictor of low exercise capacity. There are striking similarities between both disorders with respect to the muscular alterations underlying the impairment. However, different alterations occur in different muscle types. Histologic and metabolic data show that peripheral muscles undergo a shift from oxidative to glycolytic energy metabolism, whereas the opposite is observed in the diaphragm. These findings are in line with the notion that peripheral and diaphragm muscle are limited mainly by endurance and strength capacity, respectively. In both diseases, muscular impairment is multifactorially determined; hypoxia, oxidative stress, disuse, medication, nutritional depletion, and systemic inflammation may contribute to the observed muscle abnormalities and each factor has its own potential for innovative treatment approaches. Am J Clin Nutr 2000;71:1033-47.

KEY WORDS Chronic obstructive pulmonary disease, COPD, chronic heart failure, CHF, skeletal muscle, peripheral muscle, respiratory muscle, exercise intolerance, muscle performance, muscle morphology, muscle metabolism, hypoxia, oxidative stress, medication, disuse, nutritional depletion, systemic inflammation, oxygen therapy, antioxidant status, training, nutritional support, anabolic steroids, review

INTRODUCTION

According to the definitions of the World Health Organization, chronic diseases are characterized not only by the primary impairments they cause, but also by the disabilities or even handicaps that result from them (1). Although the primary impairments in chronic obstructive pulmonary disease (COPD) and chronic heart failure (CHF) clearly differ, there is a striking resemblance in the systemic consequences of these diseases and their effects on exercise capacity and health status (**Figure 1**). Impaired skeletal muscle function in COPD and CHF has long been ignored by focusing on the respective ventilatory and cardiac limitations on exercise performance. Research has shown that impaired skeletal muscle function is also an important predictor of exercise limitation in both diseases (2–6). Progression of the primary impairments in these disorders can be slowed down with medication (7, 8). Reversion can be only partially achieved through surgical interventions such as lung volume reduction surgery and lung transplantation (9, 10) and coronary bypass surgery and heart transplantation (11). However, there are limits on the age of most eligible patients and the availability of donor organs for these interventions. In addition, such interventions do not always confer a survival benefit; no improvement was found after lung transplantation in patients with end-stage emphysema (12). Also, irrespective of the reversibility of the organ impairment, exercise intolerance in both COPD and CHF remains after surgical intervention (13, 14), indicating that more detailed insight into the systemic consequences is required for effective treatment of these diseases.

Muscle function depends, though not completely, on perfusion, muscle mass, fiber composition, and energy metabolism (15). It can be inferred that alterations in one or more of these determinants play a role in reduced muscle performance. Indeed, such changes have been found in both COPD and CHF and there are striking similarities between the 2 etiologically distinct disorders.

In this review, we first present an overview of the clinical studies that have investigated impaired muscle function, with special emphasis on muscle morphology and energy metabolism in COPD and CHF. The advantage of discussing both diseases simultaneously is that the evidence about each complements that of the other and therefore provides more insight into the possible underlying causes of the muscle alterations. In the second part of the article, potential causes will be discussed, including hypoxia, oxidative stress, disuse, medication, nutritional depletion, and systemic inflammation. The third part deals with therapeutic perspectives.

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 $^{^2 \}mbox{Supported}$ by a grant from the Netherlands Asthma Foundation (project no. 96.16).

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Received September 7, 1999.

Accepted for publication December 6, 1999.



FIGURE 1. Contributors to exercise intolerance in chronic obstructive pulmonary disease (COPD) and chronic heart failure (CHF).

MUSCLE ALTERATIONS IN COPD AND CHF

Muscle performance

Muscle performance is characterized largely by strength and endurance. Strength is defined as the capacity of the muscle to develop maximal force, and endurance is defined as the capacity of the muscle to maintain a certain force over time, thus, to resist fatigue. Loss of either one of these aspects results in muscle weakness and impaired muscle performance. Numerous studies have now convincingly shown that COPD and CHF are commonly associated with muscle weakness (6, 16-21). Probably the most extensive study on the influence of muscle weakness on exercise capacity in cardiorespiratory disorders was done by Hamilton et al (4). Compared with healthy subjects, patients with respiratory failure, heart failure, or a combination of both had significantly less strength in both peripheral and respiratory muscles. However, strength and endurance seem not to be affected in the same way in respiratory and peripheral muscles. This is illustrated by the poor correlation between the strengths of both muscle groups in the 2 disorders (18, 20, 21) compared with the much stronger correlation in healthy subjects (22). This implies that the strength component of muscle weakness is affected differently in peripheral and respiratory muscles. In healthy subjects as well as in patients with COPD or CHF, exercise-limiting symptoms are the sense of leg effort (exertional discomfort) or breathlessness (exertional dyspnea) (23, 24). Thus, despite correlations between peripheral muscle strength and performance in COPD and CHF (18, 23, 25), reduced endurance (ie, fatigue) seems to be the dominant limiting factor in peripheral muscles in these patients because the sense of leg effort was one of the main reasons to stop exercising (4, 24, 26-29). It was shown that early lactic acidosis occurs in COPD during exercise (30, 31) and that this is largely the result of lactate release from the lower exercising limbs (32). Muscle acidosis is a contributing factor to muscle fatigue (33).

Fatigue is probably not the main limiting factor in respiratory muscle function. Morrison et al (34) found that COPD patients have low respiratory muscle strength and endurance. Fatigue of the respiratory muscles may indeed occur during exercise, but it is not certain whether this is an independent determinant of exercise capacity (27, 35-38). In addition, it is unlikely that the respiratory muscles of exercising COPD patients contribute to the lactate response mentioned earlier (39). It should also be emphasized that the respiratory muscles must operate against the mechanical airway impedances in this specific disorder (40), for which the force component of respiratory muscle function is most likely of great importance. For CHF it was found that respiratory muscle strength and not respiratory muscle fatigability correlated with the degree of dyspnea (41). Thus, it seems that strength is the limiting aspect of muscle performance in the respiratory muscles, whereas endurance is limiting in peripheral muscles. However, more detailed studies are required to clarify the individual roles of strength and endurance limitation in peripheral and respiratory muscles in COPD and CHF.

Muscle morphology

In both CHF (23, 42-44) and COPD (3, 45-50), marked loss of muscle mass or decline in cross-sectional muscle area is observed. This muscle wasting plays an important role in the loss of exercise tolerance in these patients. However, morphologic alterations may also be related to impairment of muscle function, although direct relations with exercise performance have not yet been shown. Some histologic information is available on abnormalities in skeletal muscle in CHF but there is hardly any on COPD. Gertz et al (51) found no signs of increased fibrosis or other alterations in intercostal muscles of patients with respiratory failure, whereas endomysial fibrosis has been found in skeletal muscle of a limited number of CHF patients (52). Increased activity of acid phosphatase, a lysosomal enzyme contributing to protein degradation, has been found in the quadriceps of some patients with CHF (25) or respiratory failure (50). Increased lipid deposits have been found in the quadriceps, biceps, and deltoids of some patients with CHF (25, 52). Contradictory results have been obtained with respect to capillary density in peripheral skeletal muscle in CHF. A normal capillary density has been found (25), which agrees with results of 2 other studies in which both reduced capillary-fiber ratios and atrophy resulted in unchanged capillary densities (53, 54). An unaltered capillary-fiber ratio has also been reported, however, with greater capillary density due to fiber atrophy (55). In contrast, reduced capillary density in combination with a reduced capillary-fiber ratio has been shown in CHF patients (56) and even in heart transplant recipients (57). Thus, overall, there is a tendency toward a reduced capillary-fiber ratio, but depending on the degree of atrophy, the capillary density may even be elevated. This tendency was recently confirmed in COPD (46).

In a few studies, morphometry of mitochondria with use of electron microscopy showed that mitochondrial volume densities in skeletal muscle were lower in CHF patients than in control subjects (56, 58), and this was still the case 10 mo after heart transplantation (57). Histochemical alterations reflecting mitochondrial abnormalities have also been reported in biceps muscle biopsies of COPD patients (50). These results suggest that the oxidative capacity of peripheral skeletal muscles may be altered in both diseases.

Muscle fiber type distribution

Probably the most remarkable muscle alteration in COPD and CHF is a relative shift in fiber composition that seems to occur in opposite directions in peripheral and respiratory muscles. Fiber typing is mainly performed histochemically, and is based on differences between fibers in myosin ATPase activities or immunocytochemistry (59). Adult mammalian skeletal muscle contains 4 myosin heavy chain (MyHC) isoforms, namely, types I, IIa, IIb, and IIx (60). In most older studies, fiber typing was limited to determining fiber types I, IIa, and IIb. Furthermore, human fibers formerly identified as being IIb with myosin ATPase staining are probably IIx fibers (61). Therefore, the notation IIb/x is used in the subsequent text. Fiber type I has a slow twitch and develops a relatively small tension, but because it depends mainly on aerobic metabolism, it is fatigue resistant. In contrast, fiber type IIb/x has a fast twitch and develops large tensions, but it is susceptible to fatigue because its energy conversion is based on anaerobic, glycolytic metabolism. Fiber type IIa has intermediate properties in that it also has a fast twitch, develops a moderate tension, is relatively resistant to fatigue, and is apt to work under both aerobic and anaerobic conditions (15, 59, 62).

A lower percentage of type I fibers and a corresponding higher percentage of type II (mainly type IIb/x) fibers, compared with those of normal subjects, has been reported in limb muscles of COPD (46, 63-66) and CHF (25, 53-56, 67) patients. In addition, in one of these studies an increase in intermediate fiber types (I + II) was also observed in CHF patients (55). These fibers may represent transformation intermediates in the $I \rightarrow IIb/x$ shift. In contrast with that in peripheral muscles, a shift from type IIb/x to type I fibers has been reported in the diaphragms of both COPD and CHF patients. In healthy subjects, the diaphragm has \approx 50% type I, 25% type IIa, and 25% IIb/x fibers (68), whereas the diaphragms of CHF patients contain 60% type I, 35% type IIa, and only 10% type IIb/x fibers (69). A IIb/x \rightarrow I shift was also observed when the distribution of MyHC isoforms was analyzed in diaphragms of patients with CHF (70) or COPD (71). Furthermore, a larger population of type I fibers in the diaphragm (corrected for the percentage of type I fibers in the quadriceps femoris) was found in both COPD and CHF patients (26, 72) compared with sedentary control subjects (68, 73). The proportion of type I fibers in both the internal and external intercostal muscles was \approx 62% in both COPD patients and control subjects in some studies (68, 74), but in other studies COPD patients had lower proportions of type I fibers (46–48%) in these muscles (73, 75). Also, elevated expression of fast MyHC has been reported in the external intercostal muscles of COPD patients (76). These results suggest that the accessory respiratory muscles do not show the II \rightarrow I fiber shift that occurs in the diaphragm. No such data have been published for CHF patients.

The overall outcome of the studies done until now (despite some variation in the results) has been that there is a $I \rightarrow IIb/x$ shift in peripheral muscles and a $IIb/x \rightarrow I$ shift in the diaphragm in COPD and CHF. It is possible that these shifts have functional consequences in the affected muscles because the distinct fiber types have different contractile properties with respect to twitch and fatigue resistance. Therefore, in COPD and CHF, a I→IIb/x shift accompanied by more glycolytic and less oxidative capacity in peripheral muscles implies loss of fatigue resistance. This change might contribute to the observed loss of exercise tolerance because peripheral muscle fatigue is the main exercise-limiting factor in these patients. This was confirmed by a study in which a faster twitch response in combination with less resistance to fatigue was observed in the leg muscles of CHF patients (77). Accordingly, a IIb/x \rightarrow I shift toward more oxidative metabolism in the diaphragm implies a shift toward a more fatigueresistant but less strength-adapted muscle. This too is in line with our notion that strength and not fatigue seems to be the main limiting factor for respiratory muscle function.

Muscle metabolism

Much data are available on skeletal muscle metabolism in CHF and COPD, partly because of the applicability of ³¹P-nuclear magnetic resonance (³¹P-NMR), which has enabled a direct and noninvasive assessment of tissue concentrations of high-energy phosphates and pH. High concentrations of ATP, creatine phosphate (CrP), and nicotinamide adenine dinucleotide in the reduced form (NADH) reflect a high-energy state, whereas elevated concentrations of ADP, AMP, inorganic phosphate (P_i), and oxidized nicotinamide adenine dinucleotide (NAD+) commonly reflect a low-energy state. Lactate and glycogen concentrations are often measured in muscle metabolism, but note that low concentrations may reflect either increased clearance or reduced formation, and vice versa for high concentrations. Although activities of enzymes involved in muscle energy metabolism measured in vitro do not reflect the physiologic situation because maximal activities are obtained under optimal, artificial circumstances, they do provide an indication of adaptations in the expression of proteins involved in metabolic pathways. Typical oxidative enzymes are citrate synthase, succinate dehydrogenase, and β-hydroxyacyl-CoA dehydrogenase (HAD). Typical glycolytic enzymes are hexokinase, phosphofructokinase, and lactate dehydrogenase-the latter of which catalyzes the last step of anaerobic glycolysis.

Measurements of substrate and cofactor concentrations in peripheral skeletal muscle of COPD and CHF patients indicate impaired energy metabolism (**Table 1**). Most striking are the observed reduced concentrations of high-energy phosphates at rest. Pouw et al (81) observed the higher P_i -CrP and ADP-ATP ratios were associated with slightly but statistically significantly elevated inosine monophosphate (IMP) concentrations. The latter may be due to increased degradation of accumulating AMP by deamination, which probably reflects reduced aerobic capacity

TABLE 1

Muscle metabolite concentrations in chronic obstructive pulmonary disease (COPD) and chronic heart failure $(CHF)^{I}$

Metabolite and muscle	Direction relative	
	to reference values	References
Creative phosphate		
QF (COPD)	▼	51, 64, 73, 78 ²
QF (CHF)	\blacksquare	$79, 80^2$
IIC and EIC (COPD)	\blacksquare	51, 75
ATP		
QF (COPD)	▼	51, 64, 73, 78 ²
QF (CHF)	▼	$79, 80^2$
IIC and EIC (COPD)	▼	51, 75
IMP		
TA (COPD)		81
Glycogen		
QF (CHF)	▼	53, 79, 80
QF (COPD)	▼	64, 73^2
Glucose		
IIC and EIC (COPD)		51
QF (CHF)	▼	82
Lactate		
QF (COPD)		51, 78
QF (CHF)		82
IIC and EIC (COPD)		51
Pyruvate		
QF (CHF)		53

¹CrP, creatine phosphate; IMP, inosine monophosphate; QF, quadriceps femoris; IIC, internal intercostal; EIC, external intercostal; TA, tibialis anterior.

²Nearly significantly different from reference values.

(83). The situation becomes even worse during exercise: a greater increase in the P_i-CrP ratio and a faster drop in pH were found in the calf muscle of COPD (47, 84, 85) and CHF (55, 86, 87) patients performing exercise than in healthy persons. Similar results were obtained for the forearm muscle (14, 87-89). In addition, a slower recovery of CrP concentrations was observed after exercise in COPD and CHF patients than in healthy persons (14, 47, 55, 85–89). These results suggest that rephosphorylation of high-energy phosphates is less efficient in these patients, both during and after muscular exercise. In addition, glycogen contents in COPD and CHF patients tend to be lower, whereas lactate concentrations are higher than in healthy persons (Table 1). Thus, it seems that anaerobic energy metabolism is enhanced in these diseases, and because this process yields far less ATP than does complete oxidative degradation of glucose, this could explain the reduced high-energy phosphate concentrations.

Analysis of enzyme activities also suggests an overall increase in glycolytic and an overall decrease in oxidative activities in peripheral muscles of both COPD and CHF patients (**Table 2**). Because these enzyme activities depend largely on the fiber type (95), it is likely that this shift in activities is related to the shift in fiber distribution mentioned above. Whether enzyme activities adapt to the fiber type redistribution or the fiber type redistribution adapts to enzyme activities remains unclear. In addition to these chronic alterations of enzyme activities, which are measured in vitro, there is probably also an acute effect on the activities of these enzymes. As a consequence of impaired electron transport, regeneration of NAD⁺ from NADH is reduced and citrate synthase and HAD are inhibited by a high NADH-NAD⁺ ratio (96). In addition, elevated AMP concentrations, resulting from the inefficient rephosphorylation of ATP, stimulate glycolysis (96). However, note that this acute effect is invisible in the in vitro activity measurements. In 2 studies, an additional inverse relation of oxidative enzyme activities with arterial lactate concentrations was found during exercise, emphasizing the assumed shift from oxidative toward glycolytic energy generation (30, 91). This loss of oxidative capacity probably accounts for the above-mentioned lipid deposits (97) because fatty acid consumption may be reduced while the supply of blood fatty acids continues. Recently, activities of 2 other oxidative enzymes, cytochrome-c oxidase (COX) and NADH dehydrogenase, were found to be elevated in COPD and CHF patients (Table 2). On first notice, this seems to be paradoxical in light of the observed reductions in the oxidative enzymes mentioned earlier. However, the oxidative enzymes mentioned earlier are involved in either the citric acid cycle or fatty acid oxidation, whereas COX and NADH dehydrogenase are enzymes of the respiratory chain. COX interacts with oxygen and therefore is the main determinant of mitochondrial oxygen affinity (98). Because a correlation between COX activity and hypoxemia was found (93), it may be that an increased number of COX molecules is a mechanism that enhances the efficiency of residual oxygen extraction and utilization and, thus, respiratory chain function. In this study, COX-I (a mitochondrial encoded subunit of COX) messenger RNA concentrations were not elevated but mitochondrial 12S ribosomal RNA concentrations were. Because this particular ribosomal RNA is a component of mitochondrial ribosomes, which are involved in translation, mitochondrial protein synthesis may be enhanced. The mechanism of hypoxia sensing and subsequent stimulation of mitochondrial gene expression remains, however, unclear (93).

Because of technical difficulties with ³¹P-NMR and muscle biopsies of the diaphragm and accessory respiratory muscles, little is known about energy metabolism in these muscles. However, the observed alterations in enzyme activities (Table 2) confirm the morphologic data, in that oxidative enzyme activities are reduced and glycolytic enzyme activities are elevated in COPD and CHF compared with the healthy state. As in peripheral muscles, this shift probably results from the shift in fiber type distribution.

POSSIBLE UNDERLYING FACTORS

Hypoxia

In COPD and CHF, oxygen delivery to peripheral and respiratory muscles may be insufficient as a result of hypoxemia, reduced blood supply, or both. In both cases, muscle tissue may become hypoxic and this could lead to the adaptive changes in skeletal muscle described above. In this respect, relevant information is now available from mountaineering expeditions (lasting ≥ 6 wk above 5000 m), because oxygen is limited at this altitude. Under these conditions, reductions in mitochondrial volume densities, oxidative enzyme activities, and cross-sectional areas of muscle fibers were found in the quadriceps (99-101). Note, however, that such expeditions are accompanied by strenuous physical activity, which also causes muscular adaptations other than those caused by hypoxia. In fact, the effect of training in combination with hypoxia may even cause a shift toward more oxidative metabolism (102). More information about the effect of hypoxia on muscle has been obtained from animal studies.

TABLE 2

Muscle enzyme activities in chronic obstructive pulmonary disease (COPD) and chronic heart failure $(CHF)^{I}$

	Direction relative	
Enzyme and muscle	to reference values	References
CS		
QF (COPD)	▼	30, 90
QF (CHF)	▼	53, 55, 82, 91
DIA (CHF)		70
IIC and EIC (COPD)		92
HAD		
QF (COPD)	▼	30, 90
QF (CHF)	▼	53, 55, 82, 91
DIA (CHF)		70
IIC and EIC (COPD)		92
SDH		
QF (COPD)	▼	90
QF (CHF)	▼	53, 91
COX		
QF (COPD)	A	93
CCRT		
QF (CHF)	A	80
НК		
QF (CHF)	▼	91
DIA (COPD)	▼	94
IIC and EIC (COPD)		92
PFK		
QF (COPD)		90
LDH		
QF (COPD)	A	90 ²
QF (CHF)		82 ²
DIA (CHF)	▼	70
DIA (COPD)	▼	94
IIC and EIC (COPD)	•	92

¹CS, citrate synthase; HAD, β-hydroxyacyl-CoA dehydrogenase; SDH, succinate dehydrogenase; COX, cytochrome *c* oxidase; CCRT, NADH cytochrome *c* reductase; HK, hexokinase; PFK, phosphofructokinase; LDH, lactate dehydrogenase; QF, quadriceps femoris; DIA, diaphragm; IIC, internal intercostal; EIC, external intercostal.

²Nearly significantly different from reference values.

Several animal studies have shown that hypoxia can indeed lead to the muscular alterations described for the limb muscles of COPD and CHF patients. For example, reduced fiber diameters in combination with unaffected numbers of capillaries, resulting in increased capillary densities, have been reported in rats exposed to hypoxia (103–105). It is suggested that the availability of oxygen to remaining muscle mitochondria is enhanced by this increased capillary density in combination with the loss of oxidative capacity (99). Furthermore, animal studies showed that hypoxia depresses protein synthesis (106–109), even in muscle tissue (106, 107). Whether hypoxia itself can contribute to the shift in muscle fiber distribution observed in COPD and CHF patients remains uncertain. There is evidence that chronic hypoxia inhibits the normal conversion of type IIa to type I fibers in growing rats, with the final outcome that these rats have a predominating proportion of type IIa fibers, unlike in control rats (110-112). However, no differences in fiber types were found when full-grown, adult rats were exposed to chronic hypoxia (110, 111, 113). Thus, it seems that hypoxia does not directly cause a type $I \rightarrow II$ fiber shift and it is more likely that the abnormal fiber type distribution results from alterations in muscular development. A similar mechanism may underlie the abnormal fiber type distribution in the regeneration of damaged muscle or the adaption of muscles to consequences of the disease in COPD and CHF.

In addition, there is evidence that hypoxia causes a shift toward glycolytic metabolism. In studies in which rats were exposed to intermittent hypoxia, it was found that citric acid cycle activity was reduced whereas glycolytic metabolism was enhanced, resulting in an increased ratio of lactate to pyruvate (114, 115). Malate dehydrogenase, a citric acid cycle enzyme, was also found to be reduced by hypoxia (116). Furthermore, hypoxia causes stimulation of glucose transport (117) and increased concentrations of membrane-associated glucose transporters (GLUT1 and GLUT4) in rat muscle (118). In muscle cell cultures, this up-regulation of GLUT1 can be mediated by either hypoxia or inhibition of the respiratory chain (119), suggesting that hypoxia affects glucose transport (and probably also metabolism) via impairment of oxidative phosphorylation.

However, in COPD and CHF this reduction in oxidative capacity does not occur in the diaphragm. Hypoxia may cause an endurance training effect in the diaphragm because of increased ventilation, which overrides its direct effect, ultimately resulting in a shift toward more aerobic metabolism.

Oxidative stress

Oxidative stress may be another factor contributing to muscle damage via reactive oxygen species. Increased plasma concentrations of lipid peroxidation products have been found in both COPD and CHF patients (120, 121). The main source of these oxygen free radicals is mitochondria because 2-5% of the oxygen consumed is not fully reduced in the electron transport chain and may leak away as superoxide radicals (122, 123). An alternative source of free radicals is immune cells activated during inflammation (124). Monocytes and macrophages produce the cytokine tumor necrosis factor α (TNF- α), which may in turn induce oxidative stress in myocytes (125). Indeed, elevated TNF-a blood concentrations have been found in both COPD (126-128) and CHF (129-133) patients, particularly in those patients with weight loss or muscle wasting. A third generator of free radicals is xanthine oxidase, which is involved in the deamination of AMP to IMP under conditions of very high AMP concentrations, such as a low-energy state (123). The above-mentioned elevated IMP concentrations in COPD (81) suggest enhanced AMP breakdown.

Susceptibility to these free radicals depends largely on the antioxidant status of tissues (123). The main antioxidant scavengers and enzymes are, among others, reduced glutathione, vitamin E (in cell membranes), superoxide dismutase, glutathione peroxidase, and catalase (123, 134, 135). Repeated exposure to oxidative stress, during long-term physical training for example, stimulates the defense system against oxygen free radicals in that concentrations of scavengers and activities of antioxidant enzymes increase (122, 123, 134-136). Oxygen flux to muscles and the resulting oxidative stress can increase tremendously during exercise (123, 137) and the disuse of muscles thus may take away this antioxidant-stimulating trigger and result in low antioxidant status. Chronic hypoxia probably acts in the same way because less oxygen is available to form reactive oxygen species. Limitations of oxygen supply are indeed found to be associated with reductions in superoxide dismutase activity in mammalian tissues like brain, lungs, and heart, although this change was not

found in skeletal muscle tissue (138, 139). In addition, in myocytes obtained from chronically hypoxic human myocardium cultured at low oxygen tension, antioxidant enzyme activities were lower than in myocytes cultured at a higher oxygen tension, illustrating the direct modulatory effect of oxygen (140). In vivo and in vitro hypoxia-reoxygenation studies showed that oxygen oversupply after a period of oxygen shortage may give rise to free radical formation in myocytes (138, 141, 142). Accordingly, in COPD and CHF patients, chronic hypoxia may result in reduced antioxidant status and occasional bouts of exercise may cause a burst of free radicals that exceeds the capacity of the defense system (122). It is also possible that the patients' reduced oxidative capacity itself leads to enhanced oxidative stress because the sudden oversupply of oxygen during exercise is inefficiently metabolized.

Reactive oxygen species are capable of damaging lipids and proteins (122, 123, 134, 143). Radicals that react with fatty acyl moieties in membrane phospholipids cause a chain reaction of peroxidations that increase the membrane permeability (143). Maintenance of membrane integrity is crucial for adequate functioning of the respiratory chain because the driving force for oxidative ATP synthesis is the electrochemical proton gradient over the inner membrane of the mitochondrion, which is generated during the electron transfer from NADH to oxygen (96). Leakage of ions through a more permeable mitochondrial inner membrane may thus impair mitochondrial function by uncoupling oxidative phosphorylation. Indeed, rats with an inherited overproduction of oxygen free radicals showed a higher degree of lipid peroxidation and protein damage in combination with impaired respiratory chain function in liver mitochondria than did control rats (144). Furthermore, a marked decrease in ATP concentrations was observed in cultured endothelial cells exposed to reactive oxygen species (145). In addition, there is evidence that an intracellular calcium overload, probably caused by a damaged sarcoplasmic reticulum membrane in combination with impaired activity of calcium ATPases, accompanies oxidative stress in animal myocytes (122, 138, 141, 142, 146-148), which may further uncouple respiration from ATP production through extensive depolarization of the inner membrane (149).

Protein oxidation by oxygen free radicals leads to formation of carbonyl groups on amino acid residues, which may modify the structure or chemical properties of the proteins affected (150). These alterations may cause a decline in protein function or even complete protein unfolding. The latter gives rise to enhanced susceptibility to proteinases. These modified proteins may also be recognized as foreign substances and, hence, be attacked by the immune system. Whether radical-induced protein damage plays a role in the abnormalities in muscles of COPD and CHF patients is unclear. It was shown in animal studies that oxidative stress induced in vivo caused myofibrillar muscle protein modification and that these proteins were rapidly degraded by proteases (151). Thus, theoretically, muscle atrophy can be enhanced by radical-induced protein damage. Indeed, it was shown that a calcium overload is involved in muscle atrophy (152) and that vitamin E deficiency facilitates muscle wasting and necrosis (153), both probably mediated by oxidative damage to proteins. Also, in human skeletal muscle it was shown that mitochondria and mitochondrial proteins were more susceptible to oxidative damage than were other subcellular components (154), which suggests that protein damage may cause impaired oxidative metabolism.

As opposed to necrosis, which is the result of exogenous damage as described above, apoptosis of muscle cells is an active process of cell death, which has also been associated with oxidative stress (155). In this study, the exposure of rat myoblasts to nitric oxide or hydrogen peroxide led to apoptotic cell death. Because these chemical stimuli are also released by immune cells, we cannot exclude the possibility that apoptosis underlies muscle wasting during inflammation.

Disuse

Disuse of skeletal muscle from a low level of physical activity is also a factor that most likely contributes to the observed muscle alterations in COPD and CHF. Hampered by their disease, these patients perform less physical activity, which may have a detraining effect on their peripheral muscles. First, detraining causes muscle weakness because of reduced motor neuron activity and muscle wasting (59, 156). Second, disuse may cause a relative reduction in the percentage of type I fibers and an increase in the percentage of type IIb/x fibers (59, 157). Third, detraining causes a decline in the activity of enzymes involved in oxidative energy conversion. This occurs in both type I and type II fibers (157, 158), suggesting that loss of oxidative capacity can occur even without any change in fiber composition. As mentioned earlier, muscular disuse has a negative effect on antioxidant status and, hence, enhances the risk of oxidative damage. Reduced physical activity thus contributes to the loss of muscle mass and probably also to the $I \rightarrow II$ fiber shift and the reduced oxidative capacity observed in limb muscles in COPD and CHF patients. As mentioned above, the diaphragm is probably not disused in these diseases and a kind of endurance training effect may even occur. This may be true not only for COPD but for CHF as well because in severe CHF, dyspnea and elevated ventilation occurs even at rest (28, 159).

Medication

Of the wide range of drugs used to treat COPD and CHF, only corticosteroids have been associated with skeletal muscle alterations. Especially in COPD, but less frequently in CHF, corticosteroids are used in low doses as maintenance medication or in higher doses during acute disease exacerbations (160, 161). Depending on the type of steroid, the dose, and the duration of treatment, the drug-induced defects range from alterations in energy metabolism to muscle weakness with underlying muscle wasting and histologic abnormalities (160, 161). From the results of both clinical and animal studies it became clear that steroidinduced muscle wasting is often associated with an overall negative nitrogen balance, reduced protein synthesis, increased protein catabolism in muscle tissues, and increased plasma concentrations of amino acids (162). These findings suggest that corticosteroids probably stimulate the mobilization of amino acids from muscle proteins (161), supplying the liver with gluconeogenic precursors, which corresponds with a shift toward glycolytic metabolism. No such data are available for CHF, but a few reports showed that COPD patients who chronically receive corticosteroids indeed may show more muscle weakness alone or with an accompanying loss of muscle mass than do nontreated COPD patients (17, 163-165). However, because COPD patients receive corticosteroids to treat inflammation, it is difficult to distinguish between the effect of steroid administration and the effects of the disease exacerbations. Also, decreased testosterone concentrations have been reported for male COPD patients receiving glucocorticoids (166).

Assuming that corticosteroids are involved in muscular alterations, the question arises as to whether this effect is generalized or is muscle-type specific. Experimental studies indicate that the glycolytic fibers seem especially susceptible to steroid-induced muscle wasting (161, 162, 167), suggesting that vulnerability depends on muscle fiber composition. The diaphragms of affected patients have a relatively high proportion of type I fibers, and so could therefore be less affected by steroid usage; limb muscles, consisting of more type II fibers, would be more vulnerable. On the other hand, the diaphragm already has a lower type II fiber content and selective wasting of type II fibers could further reduce diaphragm strength. Because strength is the main limiting factor in diaphragm muscle performance, it may be more vulnerable to corticosteroids than are limb muscles. Furthermore, it is difficult to isolate the effect of corticosteroids on diaphragm function from the complex of other unfavorable influences on lung function in COPD (165). Therefore, Wang et al (168) studied the effect of prednisone on the respiratory muscle function of normal subjects (who had no disturbing influences on function) and found no differences from a control group. This does not exclude the possibility that other steroids might have an effect because a recent animal study showed different effects of fluorinated and nonfluorinated steroids at equipotent doses (169). The results of other animal studies confirm the hypothesized shift toward glycolytic metabolism in peripheral muscle and the shift toward oxidative metabolism in the diaphragm: in corticosteroidtreated rats it was found that diaphragm force generation and the proportion of type IIb fibers was reduced in combination with decreased activity of the glycogenolytic enzyme phosphorylase and increased activities of HAD and citrate synthase (170). Increased phosphofructokinase and glycogen synthase activities have been reported for limb muscle of corticosteroid-treated rats (169). Expressed per gram of muscle, limb twitch tension may even increase in steroid-treated animals, which indicates an increase in type II fiber content (169, 171). The fact that corticosteroids selectively affect type II fibers but are also associated with a shift toward glycolytic metabolism in peripheral muscle seems contradictory and needs further investigation.

Note that in both CHF and COPD, muscle performance is not fully recovered 5–38 mo after heart or lung transplantation; intrinsic skeletal muscle alterations remain (57, 172). Corticosteroids and cyclosporin are often used as immunosuppressive agents after transplantations and it is therefore possible that these drugs might be involved in impaired muscle function (13, 57, 172).

Nutritional depletion and systemic inflammation

Nutritional depletion commonly occurs in COPD (173, 174) and CHF (42, 175). In both disorders, nutritional depletion is an important determinant of exercise capacity (3, 44, 45, 176). Body weight (usually corrected for height) is often used to determine the nutritional status of patients, but this method neglects the differences in body composition between individuals (3). Determination of body composition with respect to nutritional depletion is important because different patterns of weight loss can be distinguished: predominant loss of fat mass, predominant loss of fat-free mass, or a combination of both.

Predominant loss of fat mass involves an impaired balance between energy requirements and energy intakes. Dietary intake can be low in COPD because of symptoms such as dyspnea, fatigue, and early satiety (177). Recently, systemic inflammation was suggested to affect appetite and dietary intake, mediated by the appetite-regulating hormone leptin (178). However, patients with COPD may lose weight despite a normal or above-normal dietary intake (179). In COPD and CHF, resting energy expenditure (REE) is often elevated (180-183). In addition, total daily energy expenditure that is elevated independently of REE has been found in COPD patients (184). Increased oxygen cost of breathing probably contributes to the increased total daily energy expenditure due to increased hyperinflation during exercise (185, 186), but because hyperinflation is not increased at rest, it is unlikely that an elevated oxygen cost of breathing accounts for the elevated REE (180). Suggested contributors to the elevated REE are the thermogenic effects of bronchodilating agents (180) and systemic inflammation (127). Furthermore, the observed loss of efficient aerobic energy metabolism might play a role in the increased REE and total daily energy expenditure. In this situation of semistarvation, loss of both fat mass and fat-free mass occurs, but the fat-free mass is relatively preserved. Therefore, intrinsic muscle abnormalities besides muscle mass probably account for impaired muscle performance.

Studies of muscle function and histology in anorexia nervosa patients have provided strong data on the effect of undernutrition in muscles. Muscle performance is markedly impaired in these patients (187-189) and is associated with weight loss, loss of muscle mass, and fiber atrophy (particularly of type II fibers) (190, 191). Data from animal studies confirm these effects of undernutrition. Loss of muscle mass associated with fiber atrophy was observed in limb muscles during nutritional deprivation (171, 192). Activities of the oxidative enzymes succinate dehydrogenase and HAD were found to be reduced (192, 193). The activity of the glycolytic enzyme phosphofructokinase was also found to be reduced (193), but this was not confirmed by Koerts-de Lang et al (169). In addition, high ADP and low CrP concentrations were observed in food-deprived animals (193, 194), suggesting that muscle energy metabolism is indeed impaired after deprivation. However, it remains unclear whether nutritional deprivation results in a general loss of activities of enzymes involved in energy metabolism or predominantly affects either oxidative or glycolytic energy metabolism. The contribution of nutritional depletion to a shift from oxidative to glycolytic metabolism in COPD and CHF patients needs further investigation.

Predominant loss of fat-free mass involves an impaired balance between protein anabolism and catabolism that results in the loss of fat-free mass. In emphysema, reduced muscle protein synthesis was found (49), but protein degradation was probably not increased (195, 196). Also, nitrogen intake was not low in these patients but nitrogen excretion was elevated (196). Amino acids may be required in processes other than muscular protein synthesis, such as gluconeogenesis. Because weight loss and loss of fat-free mass have often been associated with systemic inflammation in both COPD and CHF (126, 127, 133), it is also possible that amino acids are required for increased synthesis of inflammatory proteins in the liver. Disturbed plasma and muscle amino acid concentrations have been observed in COPD, suggesting that amino acids are indeed redirected from muscle (197). Animal and in vitro studies confirm this notion (198): exposure of myocytes to TNF- α resulted in a decrease in protein synthesis, which occurred even during anabolic stimulation with insulinlike growth factor I (IGF-I). Furthermore, chronic administration of TNF- α led to increased muscle protein catabolism and liver protein anabolism in rats. In addition, the above-mentioned involvement of TNF-α in oxidative stress may contribute to muscle wasting (125). Protein depletion itself may impair skeletal muscle performance as reflected by reduced maximum voluntary handgrip strength, reduced respiratory muscle strength, and increased fatigability of in vivo electrically stimulated adductor pollicis muscle (199).

THERAPEUTIC PERSPECTIVES

Training

It is obvious that exercise training improves muscular performance because, depending on the training program, strength, endurance, or both improve (59). Because disuse has been suggested to be an important factor responsible for the alterations in muscle metabolism in COPD and CHF, it is possible that training the affected muscles could reverse these abnormalities. Indeed, exercise training improves exercise capacity in both COPD (200, 201) and CHF (6, 43, 202) patients. Furthermore, increased cross-sectional areas of oxidative fibers and elevated oxidative enzyme activities in the quadriceps muscle in combination with less arterial lactate accumulation during exercise have been found in trained COPD patients (46, 203). Traininginduced increases in oxidative capacity and muscle mass of the quadriceps muscle have also been reported for CHF patients (58, 87). The above-mentioned exercise-induced increase in the P_i-CrP ratio and drop in pH in muscle is less after training (202). Thus, in peripheral muscles, training induces a partial improvement of the oxidative capacity in combination with increased exercise performance. In general, prolonged endurance training leads to increased percentages of type I and Ha fibers accompanied with greater oxidative capacity, resulting in higher fatigue resistance (59). Therefore, considering that fatigue is the main limiting factor in peripheral muscle performance, an endurance training protocol may be most suitable for improving the exercise capacity of limb muscles in COPD patients. This is also illustrated by the fact that in COPD patients, quadriceps endurance shows a larger improvement with training than does strength (200, 201).

No data on improvement of oxidative capacity with training are available for respiratory muscles. However, the differences between respiratory and peripheral muscles in COPD and CHF suggest that different training approaches are required to effectively improve their performances. Whereas respiratory muscle training in CHF remains an unexplored field, a variety of studies have been performed for COPD (204). Although training of respiratory muscle may improve its performance, there is little evidence of real clinical benefit. The best results are probably obtained with so-called resistance training (204), in which the inspiratory muscles are subjected to an increased pressure load. The fact that this is a kind of power training affecting respiratory muscle strength especially suggests that training of the diaphragm should be more focused on strength than on endurance (205, 206).

Another possible positive effect of exercise training is the increase in antioxidant status. As discussed above, disuse (or "disuse hypoxia") has a negative effect on antioxidant status and may therefore promote oxidative damage during occasional exercise because of the temporarily enhanced oxygen supply to the exercising muscles. However, regular physical exercise involves a regular increase in exposure of muscle tissue to oxygen and training thus probably reduces the risk of oxidative stress (135).

Nutritional support, anabolic steroids, and antiinflammatory therapy

The effects of nutritional support strategies on muscle mass and muscle function have been investigated in COPD, but it is a relatively unexplored area in CHF. Several studies in nutritionally depleted patients with COPD have shown that nutritional supplementation can improve both respiratory and peripheral muscle function (174, 207, 208). It is unclear, however, to what extent this improvement in muscle function is related to the increase in muscle mass per se (209). Muscle performance may reach normal values with nutritional support while muscle mass is still lower than that of control subjects, as shown for example in anorexia nervosa patients (189), which suggests that repletion of intrinsic muscle abnormalities is important in the improvement of muscle function. An early and a late response to nutritional supplementation has been proposed (199). After the first few days of repletion, muscle function improves 10-20% without any demonstrable gain in tissue protein. This early response probably results from improved electrolyte content (210) and improved concentrations of energy-rich compounds (51, 199). Only during prolonged treatment do physiologic functions further improve, accompanied by an increase in tissue protein and muscle mass (211).

However, a substantial subgroup of COPD patients did not gain weight in response to high-energy nutritional therapy (212). This subgroup was characterized by an elevated systemic inflammatory response, as evidenced by high concentrations of acute phase proteins and soluble TNF receptors. As mentioned earlier, systemic inflammation is associated with protein catabolism and probably plays a role in the loss of muscle mass. This suggests that antiinflammatory therapy might be beneficial in this particular subgroup. Many COPD patients receive inhaled or oral corticosteroids to treat local inflammation and acute infections. Systemic inflammation, however, is not reversed during this treatment (213). In addition, oral steroids may have a negative effect on skeletal muscle as mentioned above. A possible way to modulate systemic inflammation is through ingestion of polyunsaturated fatty acids (PUFAs). PUFAs are incorporated into the phospholipids of the cell membrane and play an important role in the regulation of inflammatory processes. Indeed, fish-oil supplementation reduced inflammatory mediators and had an anticachectic effect in pancreatic cancer patients (214). No studies are yet available regarding PUFA supplementation in COPD or CHF.

Administration of anabolic steroids may be an additional mode of intervention to counteract protein catabolism either by the androgen receptor-mediated promotion of protein anabolism or by neutralizing the effects of glucocorticosteroids through binding competition for the receptor mediating catabolism (215). Anabolic steroids could thus be useful in patients with muscle wasting, especially in those who are treated with corticosteroids. Anabolic steroid treatment in addition to nutritional support as an integrated part of a pulmonary rehabilitation program produced significantly enhanced fat-free mass despite a similar weight gain with nutritional support only. This increased fat-free mass was reflected in improved respiratory muscle function (209, 216). No difference in response was noted for patients receiving maintenance oral corticosteroid treatment (209). Currently, the effects of this combined treatment approach on peripheral skeletal muscle function, exercise performance, and health status is being studied. Besides effects on muscle performance, anabolic steroids resulted in an improvement in negative acute phase proteins such as albumin

and transthyretin (215) in depleted COPD patients. This may indicate an antiinflammatory effect.

Others have investigated the effects of adjuvant treatment with recombinant human growth hormone (rhGH). Administration of this hormone induces lipolysis, protein anabolism, and muscle growth, either directly or through IGF-I. Two uncontrolled studies showed the effects of rhGH in nutritionally depleted patients with COPD. Administration of rhGH for 8 d (0.03 mg \cdot kg⁻¹ \cdot d⁻¹ subcutaneous for 4 d, plus 0.06 mg \cdot kg⁻¹ \cdot d⁻¹ for another 4 d) did not increase respiratory and peripheral skeletal muscle strength in COPD (217). In contrast, an increase in inspiratory muscle strength was reported after 3 wk of treatment (0.05 mg \cdot kg⁻¹ \cdot d⁻¹ subcutaneous) (218). With use of a similar treatment regimen, but in a placebo-controlled fashion, the effects of administration of rhGH on body composition, resting metabolic rate, and functional capacity in underweight COPD patients in a stable clinical state were studied (219). Although fat-free mass increased significantly during the 3-wk treatment period, no improvement was seen in muscle function and exercise capacity even decreased in the treatment group. Furthermore, a significant increase in resting metabolic rate was observed.

In the previous sections of this article, we stated that COPD and CHF patients may have increased oxidative stress, in either muscle or lung tissue. Furthermore, vitamin E deficiency is associated with the pathogenesis of the wasting and weakness in thalassemia major (220). Therefore, another mode of nutritional intervention might be supplementation with antioxidants such as vitamins, glutathione, and N-acetylcysteine. Several studies indeed showed a beneficial effect on wasting of antioxidant supplementation (221). For example, vitamin E protects human skeletal muscle from damage during surgical ischemia-reperfusion (222) and vitamin C supplementation reduces exerciseinduced oxidative stress (223). Similar results have been obtained in animal studies (153, 224, 225). Although antioxidant supplementation does reduce physical exercise-induced oxidative stress, it remains unclear whether exercise performance is enhanced (134, 221). Most of these data were obtained from athletes, who already have a high exercise capacity, whereas vitamin supplementation may have more effect in COPD and CHF patients, who have a very low exercise capacity. In addition, there are some indications that vitamin supplementation may improve lung function (226, 227). Antioxidant administration in CHF and COPD therefore deserves further investigation.

Oxygen therapy

Long-term oxygen therapy (LTOT) improves survival and quality of life of COPD patients (228, 229), but no such data are available for CHF. It is clear that acute oxygen administration is beneficial for exercise capacity in COPD (230-232). However, very little is known about the ability of LTOT to reverse the alterations found in skeletal muscles of COPD patients. In fact, improved exercise capacity during oxygen administration, including LTOT, could very well be an acute effect with no reversal of these abnormalities. First, by supplying oxygen, hypoxemia is partly reversed and with that dyspnea may be improved (230). The latter is an important determinant of exercise tolerance in COPD. Therefore, relief of breathlessness may account for a great deal of improvement in exercise capacity (231). Second, the acute supply of oxygen to muscle tissue probably improves oxidative energy metabolism only during the oxygen administration period itself, because the indexes of oxidative energy metabolism (P_i -CrP, pH, and CrP recovery) showed some improvement in a group of COPD patients only during oxygen administration (85). After exercise while breathing room air, COPD patients receiving LTOT still had a low P_i -CrP ratio and low pH in combination with slow CrP recovery compared with control subjects. Also, supplementation of oxygen does not add to the improving effects of training (232). Only in one study was there a reported improvement of the CrP-(CrP + Cr) ratio in resting muscle of COPD patients while breathing room air after 6–9 mo of LTOT (233). However, because the partial pressure of oxygen in blood also improved, this increase was probably caused by an increased oxygen supply and was not due to any reversal of muscle abnormalities. In addition, the low glycogen concentrations failed to improve, which further suggests that the muscle abnormalities were not reversed.

Little attention has been paid to lung damage from oxidative stress with respect to oxygen administration. It is clear that free radicals play an important role in the development of COPD, because 90% of all patients are exsmokers and cigarette smoke is a rich source of oxidants that cause all sorts of lung damage (234). The concentrations of oxygen administered to COPD patients are potentially toxic and may also result in lung injury caused by oxidative stress (235, 236). More research needs to be done to establish whether oxygen administration is beneficial or may contribute to lung or even peripheral tissue damage. In the meantime, if oxygen supplementation is necessary, it is recommended that the lowest effective concentration of oxygen be used (236).

CONCLUSIONS

This review underscores the fact that reduced skeletal muscle performance contributes markedly to exercise intolerance in COPD and CHF patients. Morphologic and metabolic abnormalities occur in the skeletal muscles of these patients which, in both disorders, are probably determined by the same set of contributing factors, including hypoxia, oxidative stress, disuse, medication, nutritional depletion, and systemic inflammation. Both diseases also share striking differences between peripheral muscles and the diaphragm, which may therefore require different therapeutic approaches. Future investigations of the mechanisms and relative contributions of each of the factors leading to these intrinsic muscular alterations are required.

REFERENCES

- Wood PH. Appreciating the consequences of disease: the international classification of impairments, disabilities, and handicaps. WHO Chron 1980;34:376–80.
- Cotes JE, Zejda J, King B. Lung function impairment as a guide to exercise limitation in work-related lung disorders. Am Rev Respir Dis 1988;137:1089–93.
- Schols AM, Mostert R, Soeters PB, Wouters EF. Body composition and exercise performance in patients with chronic obstructive pulmonary disease. Thorax 1991;46:695–9.
- Hamilton AL, Killian KJ, Summers E, Jones NL. Muscle strength, symptom intensity, and exercise capacity in patients with cardiorespiratory disorders. Am J Respir Crit Care Med 1995;152:2021–31.
- Steele IC, Moore A, Nugent AM, Riley MS, Campbell NPS, Nicholls DP. Non-invasive measurement of cardiac output and ventricular ejection fractions in chronic cardiac failure: relationship to impaired exercise tolerance. Clin Sci 1997;93:195–203.
- Sullivan MJ, Hawthorne MH. Exercise intolerance in patients with chronic heart failure. Prog Cardiovasc Dis 1995;38:1–22.

- 7. Senior RM, Anthonisen NR. Chronic obstructive pulmonary disease (COPD). Am J Respir Crit Care Med 1998;157:S139–47.
- 8. Miller MM. Current trends in the primary care management of chronic congestive heart failure. Nurse Pract 1994;19:64–70.
- Siafakas NM, Vermeire P, Pride NB, et al. Optimal assessment and management of chronic obstructive pulmonary disease (COPD). Eur Respir J 1995;8:1398–420.
- 10. Grover FL, Fullerton DA, Zamora MR, et al. The past, present, and future of lung transplantation. Am J Surg 1997;173:523–33.
- Beller GA. Selecting patients with ischemic cardiomyopathy for medical treatment, revascularization, or heart transplantation. J Nucl Cardiol 1997;4:S152–7.
- Hosenpud JD, Bennett LE, Keck BM, Edwards EB, Novick RJ. Effect of diagnosis on survival benefit of lung transplantation for end-stage lung disease. Lancet 1998;351:24–7.
- Williams TJ, Snell GI. Early and long-term functional outcomes in unilateral, bilateral, and living-related transplant recipients. Clin Chest Med 1997;18:245–57.
- Stratton JR, Kemp GJ, Daly RC, Yacoub M, Rajagopalan B. Effects of cardiac transplantation on bioenergetic abnormalities of skeletal muscle in congestive heart failure. Circulation 1994;89:1624–31.
- Armstrong RB. Muscle fiber recruitment patterns and their metabolic correlates. In: Horton HS, Terjung RL, eds. Exercise, nutrition, and energy metabolism. New York: Macmillan Publishing Company, 1988:9–26.
- Decramer M, Gosselink R, Troosters T, Verschueren M, Evers G. Muscle weakness is related to utilization of health care resources in COPD patients. Eur Respir J 1997;10:417–23.
- Bernard S, LeBlanc P, Whittom F, et al. Peripheral muscle weakness in patients with chronic obstructive pulmonary disease. Am J Respir Crit Care Med 1998;158:629–34.
- Gosselink R, Troosters T, Decramer M. Peripheral muscle weakness contributes to exercise limitation in COPD. Am J Respir Crit Care Med 1996;153:976–80.
- Zattara-Hartmann MC, Badier M, Guillot C, Tomei C, Jammes Y. Maximal force and endurance to fatigue of respiratory and skeletal muscles in chronic hypoxemic patients: the effects of oxygen breathing. Muscle Nerve 1995;18:495–502.
- Chua TP, Anker SD, Harrington D, Coats AJ. Inspiratory muscle strength is a determinant of maximum oxygen consumption in chronic heart failure. Br Heart J 1995;74:381–5.
- McParland C, Resch EF, Krishnan B, Wang Y, Cujec B, Gallagher CG. Inspiratory muscle weakness in chronic heart failure: role of nutrition and electrolyte status and systemic myopathy. Am J Respir Crit Care Med 1995;151:1101–7.
- Lands LC, Heigenhauser GJ, Jones NL. Respiratory and peripheral muscle function in cystic fibrosis. Am Rev Respir Dis 1993; 147:865–9.
- Harrington D, Anker SD, Chua TP, et al. Skeletal muscle function and its relation to exercise tolerance in chronic heart failure. J Am Coll Cardiol 1997;30:1758–64.
- Killian KJ, Leblanc P, Martin DH, Summers E, Jones NL, Campbell EJ. Exercise capacity and ventilatory, circulatory, and symptom limitation in patients with chronic airflow limitation. Am Rev Respir Dis 1992;146:935–40.
- Lipkin DP, Jones DA, Round JM, Poole Wilson PA. Abnormalities of skeletal muscle in patients with chronic heart failure. Int J Cardiol 1988;18:187–95.
- Karlsson J, Diamant B, Folkers K. Exercise-limiting factors in respiratory distress. Respiration 1992;2:18–23.
- 27. Belman MJ. Exercise in patients with chronic obstructive pulmonary disease. Thorax 1993;48:936–46.
- Myers J, Salleh A, Buchanan N, et al. Ventilatory mechanisms of exercise intolerance in chronic heart failure. Am Heart J 1992;124:710–9.
- 29. Drexler H. Changes in the peripheral circulation in heart failure. Curr Opin Cardiol 1995;10:268–73.
- Maltais F, Simard AA, Simard C, Jobin J, Desgagnes P, LeBlanc P. Oxidative capacity of the skeletal muscle and lactic acid kinetics

during exercise in normal subjects and in patients with COPD. Am J Respir Crit Care Med 1996;153:288–93.

- Engelen MPKJ, Schols AMWJ, Does JD, Wouters EFM. Exercise induced lactate increase in relation to physical activity level and peripheral muscle substrates in COPD. Am J Respir Crit Care Med 1999;159:A475 (abstr).
- Maltais F, Jobin J, Sullivan MJ, et al. Metabolic and hemodynamic responses of lower limb during exercise in patients with COPD. J Appl Physiol 1999;84:1573–80.
- Mainwood GW, Renaud JM. The effect of acid-base balance on fatigue of skeletal muscle. Can J Physiol Pharmacol 1985;63:403–16.
- Morrison NJ, Richardson J, Dunn L, Pardy RL. Respiratory muscle performance in normal elderly subjects and patients with COPD. Chest 1989;95:90–4.
- Decramer M, Aubier M. The respiratory muscles: cellular and molecular physiology. Eur Respir J 1997;10:1943–5.
- Similowski T, Yan S, Gauthier AP, Macklem PT, Bellemare F. Contractile properties of the human diaphragm during chronic hyperinflation. N Engl J Med 1991;325:917–23.
- Pardy RL, Rivington RN, Despas PJ, Macklem PT. The effects of inspiratory muscle training on exercise performance in chronic airflow limitation. Am Rev Respir Dis 1981;123:426–33.
- Fitting JW. Respiratory muscle fatigue limiting physical exercise? Eur Respir J 1991;4:103–8.
- Engelen MP, Casaburi R, Rucker R, Carithers E. Contribution of the respiratory muscles to the lactic acidosis of heavy exercise in COPD. Chest 1995;108:1246–51.
- O'Donnell DE, Webb KA. Exertional breathlessness in patients with chronic airflow limitation. The role of lung hyperinflation. Am Rev Respir Dis 1993;148:1351–7.
- Mancini DM, Henson D, LaManca J, Levine S. Respiratory muscle function and dyspnea in patients with chronic congestive heart failure. Circulation 1992;86:909–18.
- 42. Wilson JR, Mancini DM. The mechanism of extertional fatigue in heart failure. Cardioscience 1990;1:13–7.
- Shephard RJ. Exercise for patients with congestive heart failure. Sports Med 1997;23:75–92.
- Miyagi K, Asanoi H, Ishizaka S, et al. Importance of total leg muscle mass for exercise intolerance in chronic heart failure. Jpn Heart J 1994;35:15–26.
- 45. Engelen MP, Schols AM, Baken WC, Wesseling GJ, Wouters EF. Nutritional depletion in relation to respiratory and peripheral skeletal muscle function in out-patients with COPD. Eur Respir J 1994; 7:1793–7.
- Whittom F, Jobin J, Simard PM, et al. Histochemical and morphological characteristics of the vastus lateralis muscle in patients with chronic obstructive pulmonary disease. Med Sci Sports Exerc 1998;30:1467–74.
- Wuyam B, Payen JF, Levy P, et al. Metabolism and aerobic capacity of skeletal muscle in chronic respiratory failure related to chronic obstructive pulmonary disease. Eur Respir J 1992;5:157–62.
- Rochester DF. Body weight and respiratory muscle function in chronic obstructive pulmonary disease. Am Rev Respir Dis 1986; 134:646–8.
- Morrison WL, Gibson JN, Scrimgeour C, Rennie MJ. Muscle wasting in emphysema. Clin Sci 1988;75:415–20.
- Sato Y, Asoh T, Honda Y, Fujimatsu Y, Higuchi I, Oizumi K. Morphologic and histochemical evaluation of muscle in patients with chronic pulmonary emphysema manifesting generalized emaciation. Eur Neurol 1997;37:116–21.
- Gertz I, Hedenstierna G, Hellers G, Wahren J. Muscle metabolism in patients with chronic obstructive lung disease and acute respiratory failure. Clin Sci Mol Med 1977;52:395–403.
- 52. Dunnigan A, Staley NA, Smith SA, et al. Cardiac and skeletal muscle abnormalities in cardiomyopathy: comparison of patients with ventricular tachycardia or congestive heart failure. J Am Coll Cardiol 1987;10:608–18.

- Sullivan MJ, Green HJ, Cobb FR. Skeletal muscle biochemistry and histology in ambulatory patients with long-term heart failure. Circulation 1990;81:518–27.
- 54. Schaufelberger M, Eriksson BO, Grimby G, Held P, Swedberg K. Skeletal muscle fiber composition and capillarization in patients with chronic heart failure: relation to exercise capacity and central hemodynamics. J Card Fail 1995;1:267–72.
- Mancini DM, Coyle E, Coggan A, et al. Contribution of intrinsic skeletal muscle changes to ³¹P NMR skeletal muscle metabolic abnormalities in patients with chronic heart failure. Circulation 1989;80:1338–46.
- Drexler H, Riede U, Münzel T, König H, Funke E, Just H. Alterations of skeletal muscle in chronic heart failure. Circulation 1992; 85:1751–9.
- Lampert E, Mettauer B, Hoppeler H, Charloux A, Charpentier A, Lonsdorfer J. Structure of skeletal muscle in heart transplant recipients. J Am Coll Cardiol 1996;28:980–4.
- Hambrecht R, Fiehn E, Yu J, et al. Effects of endurance training on mitochondrial ultrastructure and fiber type distribution in skeletal muscle of patients with stable chronic heart failure. J Am Coll Cardiol 1997;29:1067–73.
- 59. McComas AJ. Skeletal muscle: form and function. Champaign, IL: Human Kinetics, 1996.
- Schiaffino S, Reggiani C. Molecular diversity of myofibrillar proteins: gene regulation and functional significance. Physiol Rev 1996;76:371–423.
- Pereira Sant'Ana JA, Ennion S, Sargeant AJ, et al. Comparison of the molecular, antigenic and ATPase determinants of fast myosin heavy chains in rat and human: a single-fibre study. Pflügers Arch 1997;435:151–63.
- 62. Harridge SDR, Bottinelli R, Canepari M, et al. Whole-muscle and single-fibre contractile properties and myosin heavy chain isoforms in humans. Pflügers Arch 1996;432:913–20.
- Satta A, Migliori GB, Spanevello A, et al. Fibre types in skeletal muscles of chronic obstructive pulmonary disease patients related to respiratory function and exercise tolerance. Eur Respir J 1997; 10:2853–60.
- 64. Jakobsson P, Jorfeldt L, Brundin A. Skeletal muscle metabolites and fibre types in patients with advanced chronic obstructive pulmonary disease (COPD), with and without chronic respiratory failure. Eur Respir J 1990;3:192–6.
- Maltais F, Sullivan MJ, LeBlanc P, et al. Altered expression of myosin heavy chain in the vastus lateralis muscle in patients with COPD. Eur Respir J 1999;13:850–4.
- Hildebrand IL, Sylvén C, Esbjörnsson M, Hellström K, Jansson E. Does chronic hypoxaemia induce transformations of fibre types? Acta Physiol Scand 1991;141:435–9.
- Sullivan MJ, Duscha BD, Klitgaard H, Kraus WE, Cobb FR, Saltin B. Altered expression of myosin heavy chain in human skeletal muscle in chronic heart failure. Med Sci Sports Exerc 1997;29:860–6.
- Mizuno M. Human respiratory muscles: fibre morphology and capillary supply. Eur Respir J 1991;4:587–601.
- Lindsay DC, Lovegrove CA, Dunn MJ, et al. Histological abnormalities of muscle from limb, thorax and diaphragm in chronic heart failure. Eur Heart J 1996;17:1239–50.
- Tikunov B, Levine S, Mancini D. Chronic congestive heart failure elicits adaptations of endurance exercise in diaphragmatic muscle. Circulation 1997;95:910–6.
- Levine S, Kaiser L, Leferovich J, Tikunov B. Cellular adaptations in the diaphragm in chronic obstructive pulmonary disease. N Engl J Med 1997;337:1799–806.
- 72. Morton JM, McKenna MJ, Carey MF, et al. Skeletal muscle pathophysiology in subjects with severe COAD. In: The Thoracic Society of Australia & New Zealand 1998 annual scientific meeting "Frontiers of Respiratory Medicine" program and abstracts book. Adelaide, Australia: Thoracic Society of Australia & New Zealand, 1998:012 (abstr).

- Hughes RL, Katz H, Sahgal V, Campbell JA, Hartz R, Shields TW. Fiber size and energy metabolites in five separate muscles from patients with chronic obstructive lung diseases. Respiration 1983; 44:321–8.
- Hards JM, Reid WD, Pardy RL, Pare PD. Respiratory muscle fiber morphometry. Correlation with pulmonary function and nutrition. Chest 1990;97:1037–44.
- Campbell JA, Hughes RL, Sahgal V, Frederiksen J, Shields TW. Alterations in intercostal muscle morphology and biochemistry in patients with obstructive lung disease. Am Rev Respir Dis 1980;122:679–86.
- Gea JG. Myosin gene expression in the respiratory muscles. Eur Respir J 1997;10:2404–10.
- Harridge SD, Magnusson G, Gordon A. Skeletal muscle contractile characteristics and fatigue resistance in patients with chronic heart failure. Eur Heart J 1996;17:896–901.
- Fiaccadori E, Del Canale S, Vitali P, Coffrini E, Ronda N, Guariglia A. Skeletal muscle energetics, acid-base equilibrium and lactate metabolism in patients with severe hypercapnia and hypoxemia. Chest 1987;92:883–7.
- Broqvist M, Arnqvist H, Dahlström U, Larsson J, Nylander E, Permert J. Nutritional assessment and muscle energy metabolism in severe chronic congestive heart failure—effects of long-term dietary supplementation. Eur Heart J 1994;15:1641–50.
- Opasich C, Aquilani R, Dossena M, et al. Biochemical analysis of muscle biopsy in overnight fasting patients with severe chronic heart failure. Eur Heart J 1996;17:1686–93.
- Pouw EM, Schols AMWJ, Vusse van der GJ, Wouters EFM. Elevated inosine monophosphate levels in resting muscle of patients with stable COPD. Am J Respir Crit Care Med 1998;157:453–57.
- Schaufelberger M, Eriksson BO, Held P, Swedberg K. Skeletal muscle metabolism during exercise in patients with chronic heart failure. Heart 1996;76:29–34.
- Dudley GA, Terjung RL. Influence of aerobic metabolism on IMP accumulation in fast-twitch muscle. Am J Physiol 1985;248:C37–42.
- Thompson CH, Davies RJ, Kemp GJ, Taylor DJ, Radda GK, Rajagopalan B. Skeletal muscle metabolism during exercise and recovery in patients with respiratory failure. Thorax 1993;48:486–90.
- Payen JF, Wuyam B, Levy P, et al. Muscular metabolism during oxygen supplementation in patients with chronic hypoxemia. Am Rev Respir Dis 1993;147:592–8.
- Mancini DM, Walter G, Reichek N, et al. Contribution of skeletal muscle atrophy to exercise intolerance and altered muscle metabolism in heart failure. Circulation 1992;85:1364–73.
- 87. Kemp GJ, Thompson CH, Stratton JR, et al. Abnormalities in exercising skeletal muscle in congestive heart failure can be explained in terms of decreased mitochondrial ATP synthesis, reduced metabolic efficiency, and increased glycogenolysis. Heart 1996;76:35–41.
- Kutsuzawa T, Shioya S, Kurita D, Haida M, Ohta Y, Yamabayashi H.
 ³¹P-NMR study of skeletal muscle metabolism in patients with chronic respiratory impairment. Am Rev Respir Dis 1992;146:1019–24.
- Tada H, Kato H, Misawa T, et al. ³¹P-nuclear magnetic resonance evidence of abnormal skeletal muscle metabolism in patients with chronic lung disease and congestive heart failure. Eur Respir J 1992;5:163–9.
- Jakobsson P, Jorfeldt L, Henriksson J. Metabolic enzyme activity in the quadriceps femoris muscle in patients with severe chronic obstructive pulmonary disease. Am J Respir Crit Care Med 1995;151:374–7.
- Sullivan MJ, Green HJ, Cobb FR. Altered skeletal muscle metabolic response to exercise in chronic heart failure. Relation to skeletal muscle aerobic enzyme activity. Circulation 1991;84:1597–607.
- 92. Sánchez J, Brunet A, Medrano G, Debesse B, Derenne JP. Metabolic enzymatic activities in the intercostal and serratus muscles and in the latissimus dorsi of middle-aged normal men and patients with moderate obstructive pulmonary disease. Eur Respir J 1988;1:376–83.
- Sauleda J, García-Palmer F, Wiesner RJ, et al. Cytochrome oxidase activity and mitochondrial gene expression in skeletal muscle of

patients with chronic obstructive pulmonary disease. Am J Respir Crit Care Med 1998;157:1413–7.

- 94. Sánchez J, Bastien C, Medrano G, Riquet M, Derenne JP. Metabolic enzymatic activities in the diaphragm of normal men and patients with moderate chronic obstructive pulmonary disease. Bull Eur Physiopathol Respir 1984;20:535–40.
- Essén-Gustavsson B, Henriksson J. Enzyme levels in pools of microdissected human muscle fibres of identified type. Adaptive response to exercise. Acta Physiol Scand 1984;120:505–15.
- 96. Stryer L. Biochemistry. New York: WH Freeman and Company, 1988.
- 97. Tein I. Metabolic myopathies. Semin Pediatr Neurol 1996;3:59-98.
- Gnaiger E, Lassnig B, Kuznetsov A, Rieger G, Margreiter R. Mitochondrial oxygen affinity, respiratory flux control and excess capacity of cytochrome c oxidase. J Exp Biol 1998;201:1129–39.
- Hoppeler H, Kleinert E, Schlegel C, et al. Morphological adaptations of human skeletal muscle to chronic hypoxia. Int J Sports Med 1990;11:S3–9.
- 100. Hoppeler H, Desplanches D. Muscle structural modifications in hypoxia. Int J Sports Med 1992;13:S166–8.
- 101. Howald H, Pette D, Simoneau JA, Uber A, Hoppeler H, Cerretelli P. Effects of chronic hypoxia on muscle enzyme activities. Int J Sports Med 1990;11:S10–4.
- 102. Melissa L, MacDougall JD, Tarnopolsky MA, Cipriano N, Green HJ. Skeletal muscle adaptations to training under normobaric hypoxic versus normoxic conditions. Med Sci Sports Exerc 1997;29:238–43.
- 103. Bigard AX, Brunet A, Guezennec CY, Monod H. Effects of chronic hypoxia and endurance training on muscle capillarity in rats. Pflügers Arch 1991;419:225–9.
- 104. Ekeren van GJ, Sengers RC, Stadhouders AM. Changes in volume densities and distribution of mitochondria in rat skeletal muscle after chronic hypoxia. Int J Exp Pathol 1992;73:51–60.
- 105. Snyder GK, Farrelly C, Coelho JR. Adaptations in skeletal muscle capillarity following changes in oxygen supply and changes in oxygen demands. Eur J Appl Physiol 1992;65:158–63.
- 106. Preedy VR, Smith DM, Sugden PH. The effects of 6 hours of hypoxia on protein synthesis in rat tissues in vivo and in vitro. Biochem J 1985;228:179–85.
- 107. Fuller SJ, Sugden PH. Acute inhibition of rat heart protein synthesis in vitro during beta-adrenergic stimulation or hypoxia. Am J Physiol 1988;255:E537–47.
- 108. Milley JR. Protein synthesis during hypoxia in fetal lambs. Am J Physiol 1987;252:E519–24.
- 109. Kwast KE, Hand SC. Acute depression of mitochondrial protein synthesis during anoxia: contributions of oxygen sensing, matrix acidification, and redox state. J Biol Chem 1996;271:7313–9.
- 110. Ishihara A, Itoh K, Oishi Y, Itoh M, Hirofuji C, Hayashi H. Effects of hypobaric hypoxia on histochemical fibre-type composition and myosin heavy chain isoform component in the rat soleus muscle. Pflügers Arch 1995;429:601–6.
- 111. Itoh K, Itoh M, Ishihara A, Hirofuji C, Hayashi H. Influence of 12 weeks of hypobaric hypoxia on fibre type composition of the rat soleus muscle. Acta Physiol Scand 1995;154:417–8.
- 112. Taguchi S, Hata Y, Itoh K. Enzymatic responses and adaptations to swimming training and hypobaric hypoxia in postnatal rats. Jpn J Physiol 1985;35:1023–32.
- 113. Takahashi H, Kikuchi K, Nakayama H. Effect of chronic hypoxia on skeletal muscle fiber type in adult male rats. Ann Physiol Anthropol 1992;11:625–30.
- 114. Pastoris O, Dossena M, Foppa P, et al. Modifications by chronic intermittent hypoxia and drug treatment on skeletal muscle metabolism. Neurochem Res 1995;20:143–50.
- 115. Pastoris O, Gorini A, Vercesi L, Taglietti M, Dossena M. Modification of the skeletal muscle energy metabolism induced by intermittent normobaric hypoxia and treatment with biological pyrimidines. Farmaco [Sci] 1985;40:442–53.
- 116. Takahashi H, Kikuchi K, Nakayama H. Effect of chronic hypoxia on oxidative enzyme activity in rat skeletal muscle. Ann Physiol Anthropol 1993;12:363–9.

- 117. Cartee GD, Douen AG, Ramlal T, Klip A, Holloszy JO. Stimulation of glucose transport in skeletal muscle by hypoxia. J Appl Physiol 1991;70:1593–600.
- 118. Xia Y, Warshaw JB, Haddad GG. Effect of chronic hypoxia on glucose transporters in heart and skeletal muscle of immature and adult rats. Am J Physiol 1997;273:R1734–41.
- 119. Bashan N, Burdett E, Guma A, et al. Mechanisms of adaptation of glucose transporters to changes in the oxidative chain of muscle and fat cells. Am J Physiol 1993;264:C430–40.
- 120. Rahman I, Morrison D, Donaldson K, MacNee W. Systemic oxidative stress in asthma, COPD, and smokers. Am J Respir Crit Care Med 1996;154:1055–60.
- 121. Keith M, Geranmayegan A, Sole MJ, et al. Increased oxidative stress in patients with congestive heart failure. J Am Coll Cardiol 1998;31:1352–6.
- 122. Giuliani A, Cestaro B. Exercise, free radical generation and vitamins. Eur J Cancer Prev 1997;6:S55–67.
- 123. Ji LL. Exercise, oxidative stress, and antioxidants. Am J Sports Med 1996;24:S20–4.
- 124. Reid MB. Reactive oxygen and nitric oxide in skeletal muscle. News Physiol Sci 1996;11:114–9.
- 125. Buck M, Chojkier M. Muscle wasting and dedifferentiation induced by oxidative stress in a murine model of cachexia is prevented by inhibitors of nitric oxide synthesis and antioxidants. EMBO J 1996;15:1753–65.
- 126. de Godoy I, Donahoe M, Calhoun WJ, Mancino J, Rogers RM. Elevated TNF-α production by peripheral blood monocytes of weightlosing COPD patients. Am J Respir Crit Care Med 1996;153:633–7.
- 127. Schols AM, Buurman WA, Staal van den Brekel AJ, Dentener MA, Wouters EF. Evidence for a relation between metabolic derangements and increased levels of inflammatory mediators in a subgroup of patients with chronic obstructive pulmonary disease. Thorax 1996;51:819–24.
- 128. Di Francia M, Barbier D, Mege JL, Orehek J. Tumor necrosis factor-α levels and weight loss in chronic obstructive pulmonary disease. Am J Respir Crit Care Med 1994;150:1453–5.
- 129. Ferrari R, Bachetti T, Confortini R, et al. Tumor necrosis factor soluble receptors in patients with various degrees of congestive heart failure. Circulation 1995;92:1479–86.
- 130. Katz SD, Rao R, Berman JW, et al. Pathophysiological correlates of increased serum tumor necrosis factor in patients with congestive heart failure. Relation to nitric oxide-dependent vasodilation in the forearm circulation. Circulation 1994;90:12–6.
- 131. Levine B, Kalman J, Mayer L, Fillit HM, Packer M. Elevated circulating levels of tumor necrosis factor in severe chronic heart failure. N Engl J Med 1990;323:236–41.
- 132. McMurray J, Abdullah I, Dargie HJ, Shapiro D. Increased concentrations of tumour necrosis factor in "cachectic" patients with severe chronic heart failure. Br Heart J 1991;66:356–8.
- 133. Anker SD, Clark AL, Kemp M, et al. Tumor necrosis factor and steroid metabolism in chronic heart failure: possible relation to muscle wasting. J Am Coll Cardiol 1997;30:997–1001.
- 134. Clarkson PM. Antioxidants and physical performance. Crit Rev Food Sci Nutr 1995;35:131-41.
- 135. Sen CK, Hänninen O. Physiological antioxidants. In: Sen KC, Packer L, Hänninen O, eds. Exercise and oxygen toxicity. Amsterdam: Elsevier Science BV, 1994:89–126.
- 136. Ohishi S, Kizaki T, Nagasawa J, et al. Effects of endurance training on superoxide dismutase activity, content and mRNA expression in rat muscle. Clin Exp Pharmacol Physiol 1997;24:326–32.
- 137. Jackson MJ. Exercise and oxygen radical production by muscle. In: Sen KC, Packer L, Hänninen O, eds. Exercise and oxygen toxicity. Amsterdam: Elsevier Science BV, 1994:49–57.
- Ferrari R, Ceconi C, Curello S, Alfieri O, Visioli O. Myocardial damage during ischaemia and reperfusion. Eur Heart J 1993;14:25–30.
- 139. Liu J, Simon LM, Phillips JR, Robin ED. Superoxide dismutase (SOD) activity in hypoxic mammalian systems. J Appl Physiol 1977; 42:107–10.

- 140. Li RK, Mickle DA, Weisel RD, et al. Effect of oxygen tension on the anti-oxidant enzyme activities of tetralogy of Fallot ventricular myocytes. J Mol Cell Cardiol 1989;21:567–75.
- 141. Gardner TJ. Oxygen radicals and myocardial stunning. J Card Surg 1994;9:422-4.
- 142. Smith DR, Stone D, Darley-Usmar VM. Stimulation of mitochondrial oxygen consumption in isolated cardiomyocytes after hypoxiareoxygenation. Free Radic Res 1996;24:159–66.
- 143. Haramaki N, Packer L. Oxidative stress indices in exercise. In: Sen KC, Packer L, Hänninen O, eds. Exercise and oxygen toxicity. Amsterdam: Elsevier Science BV, 1994:77–87.
- 144. Salganik RI, Shabalina IG, Solovyova NA, Kolosova NG, Solovyov VN, Kolpakov AR. Impairment of respiratory functions in mitochondria of rats with an inherited hyperproduction of free radicals. Biochem Biophys Res Commun 1994;205:180–5.
- 145. Spragg RG, Hinshaw DB, Hyslop PA, Schraufstätter IU, Cochrane CG. Alterations in adenosine triphosphate and energy charge in cultured endothelial and P388D1 cells after oxidant injury. J Clin Invest 1985;76:1471–6.
- 146. Astier C, Rock E, Lab C, Gueux E, Mazur A, Rayssiguier Y. Functional alterations in sarcoplasmic reticulum membranes of magnesium-deficient rat skeletal muscle as consequences of free radicalmediated process. Free Radic Biol Med 1996;20:667–74.
- 147. Wang SY, Clague JR, Langer GA. Increase in calcium leak channel activity by metabolic inhibition or hydrogen peroxide in rat ventricular myocytes and its inhibition by polycation. J Mol Cell Cardiol 1995;27:211–22.
- 148. Xu KY, Zweier JL, Becker LC. Hydroxyl radical inhibits sarcoplasmic reticulum Ca²⁺-ATPase function by direct attack on the ATP binding site. Circ Res 1997;80:76–81.
- 149. Minezaki KK, Suleiman MS, Chapman RA. Changes in mitochondrial function induced in isolated guinea-pig ventricular myocytes by calcium overload. J Physiol (Lond) 1994;476:459–71.
- Dean RT, Fu SL, Stocker R, Davies MJ. Biochemistry and pathology of radical-mediated protein oxidation. Biochem J 1997;324:1–18.
- 151. Nagasawa T, Hatayama T, Watanabe Y, Tanaka M, Niisato Y, Kitts DD. Free radical mediated effects on skeletal muscle protein in rats treated with fe nitrilotriacetate. Biochem Biophys Res Commun 1997;231:37–41.
- 152. Soares JM, Duarte JA, Carvalho J, Appell HJ. The possible role of intracellular Ca²⁺ accumulation for the development of immobilization atrophy. Int J Sports Med 1993;14:437–9.
- 153. Thomas PK, Cooper JM, King RH, et al. Myopathy in vitamin E deficient rats: muscle fibre necrosis associated with disturbances of mitochondrial function. J Anat 1993;183:451–61.
- 154. Haycock JW, Jones P, Harris JB, Mantle D. Differential susceptibility of human skeletal muscle proteins to free radical induced oxidative damage: a histochemical, immunocytochemical and electron microscopical study in vitro. Acta Neuropathol (Berl) 1996;92:331–40.
- 155. Stangel M, Zettl UK, Mix E, et al. H₂O₂ and nitric oxide-mediated oxidative stress induce apoptosis in rat skeletal muscle myoblasts. J Neuropathol Exp Neurol 1996;55:36–43.
- 156. Berg HE, Tesch PA. Changes in muscle function in response to 10 days of lower limb unloading in humans. Acta Physiol Scand 1996; 157:63–70.
- 157. Terjung RL, Dudley GA, Meyer RA. Metabolic and circulatory limitations to muscular performance at the organ level. J Exp Biol 1985;115:307–18.
- 158. Henriksson J, Reitman JS. Time course of changes in human skeletal muscle succinate dehydrogenase and cytochrome oxidase activities and maximal oxygen uptake with physical activity and inactivity. Acta Physiol Scand 1977;99:91–7.
- 159. Messner-Pellenc P, Brasileiro C, Ahmaidi S, et al. Exercise intolerance in patients with chronic heart failure: role of pulmonary diffusing limitation. Eur Heart J 1995;16:201–9.
- 160. Balkom van RH, Heijden van der HF, Herwaarden van CL, Dekhuijzen PN. Corticosteroid-induced myopathy of the respiratory muscles. Neth J Med 1994;45:114–22.

- 161. LaPier TK. Glucocorticoid-induced muscle atrophy. The role of exercise in treatment and prevention. J Cardiopulm Rehabil 1997; 17:76–84.
- 162. Mayer M, Rosen F. Interaction of glucocorticoids and androgens with skeletal muscle. Metabolism 1977;26:937–62.
- 163. Decramer M, Lacquet LM, Fagard R, Rogiers P. Corticosteroids contribute to muscle weakness in chronic airflow obstruction. Am J Respir Crit Care Med 1994;150:11–6.
- 164. Decramer M, de Bock V, Dom R. Functional and histologic picture of steroid-induced myopathy in chronic obstructive pulmonary disease. Am J Respir Crit Care Med 1996;153:1958–64.
- 165. Dekhuijzen PN, van Balkom RH. Steroid-induced change in the respiratory muscles: its relevance in patients with obstructive airways disease. Respir Med 1994;88:335–41.
- 166. Kamischke A, Kemper DE, Castel MA, et al. Testosterone levels in men with chronic obstructive pulmonary disease with or without glucocorticoid therapy. Eur Respir J 1998;11:41–5.
- 167. Dekhuijzen PN, Decramer M. Steroid-induced myopathy and its significance to respiratory disease: a known disease rediscovered. Eur Respir J 1992;5:997–1003.
- 168. Wang YM, Zintel T, Vasquez A, Gallagher CG. Corticosteroid therapy and respiratory muscle function in humans. Am Rev Respir Dis 1991;144:108–12.
- 169. Koerts-de Lang E, Hesselink MKC, Drost MR, van der Vusse GJ, Wouters EFM, Schols AMWJ. Enzyme activity of rat tibialis anterior muscle differs between treatment with triamcinolone and prednisolone and nutritional deprivation. Eur J Appl Physiol 1999; 79:274–9.
- 170. Balkom van RH, Dekhuijzen PN, Folgering HT, Veerkamp JH, Fransen JA, van Herwaarden CL. Effects of long-term low-dose methylprednisolone on rat diaphragm function and structure. Muscle Nerve 1997;20:983–90.
- 171. Gardiner PF, Montanaro G, Simpson DR, Edgerton VR. Effects of glucocorticoid treatment and food restriction on rat hindlimb muscles. Am J Physiol 1980;238:E124–30.
- 172. Evans AB, Al Himyary AJ, Hrovat MI, et al. Abnormal skeletal muscle oxidative capacity after lung transplantation by ³¹P-MRS. Am J Respir Crit Care Med 1997;155:615–21.
- 173. Gray-Donald K, Gibbons L, Shapiro SH, Martin JG. Effect of nutritional status on exercise performance in patients with chronic obstructive pulmonary disease. Am Rev Respir Dis 1989;140:1544–8.
- 174.Efthimiou J, Fleming J, Gomes C, Spiro SG. The effect of supplementary oral nutrition in poorly nourished patients with chronic obstructive pulmonary disease. Am Rev Respir Dis 1988;137: 1075–82.
- 175. Carr JG, Stevenson LW, Walden JA, Heber D. Prevalence and hemodynamic correlates of malnutrition in severe congestive heart failure secondary to ischemic or idiopathic dilated cardiomyopathy. Am J Cardiol 1989;63:709–13.
- 176. Schols AM, Soeters PB, Dingemans AM, Mostert R, Frantzen PJ, Wouters EF. Prevalence and characteristics of nutritional depletion in patients with stable COPD eligible for pulmonary rehabilitation. Am Rev Respir Dis 1993;147:1151–6.
- 177. Vermeeren MAP, Schols A, Wouters EFM. Effects of an acute exacerbation on nutritional and metabolic profile of patients with COPD. Eur Respir J 1997;10:2264–9.
- 178. Schols AMWJ, Creutzberg EC, Buurman WA, Campfield LA, Saris WHM, Wouters EFM. Plasma leptin is related to proinflammatory status and dietary intake in patients with chronic obstructive pulmonary disease. Am J Respir Crit Care Med 1999;160:1220–6.
- 179. Hunter AM, Carey MA, Larsh HW. The nutritional status of patients with chronic obstructive pulmonary disease. Am Rev Respir Dis 1981;124:376–81.
- 180. Creutzberg EC, Schols AMWJ, Bothmer-Quaedvlieg FCM, Wouters EFM. Prevalence of an elevated resting energy expenditure in patients with chronic obstructive pulmonary disease in relation to body composition and lung function. Eur J Clin Nutr 1998;52:396–401.

- 181. Schols AMWJ, Fredrix EW, Soeters PB, Westerterp KR, Wouters EF. Resting energy expenditure in patients with chronic obstructive pulmonary disease. Am J Clin Nutr 1991;54:983–7.
- 182. Riley M, Elborn JS, McKane WR, Bell N, Stanford CF, Nicholls DP. Resting energy expenditure in chronic cardiac failure. Clin Sci (Colch) 1991;80:633–9.
- 183. Obisesan TO, Toth MJ, Donaldson K, et al. Energy expenditure and symptom severity in men with heart failure. Am J Cardiol 1996; 77:1250–2.
- 184. Baarends EM, Schols AM, Pannemans DL, Westerterp KR, Wouters EF. Total free living energy expenditure in patients with severe chronic obstructive pulmonary disease. Am J Respir Crit Care Med 1997;155:549–54.
- 185. Donahoe M, Rogers RM, Wilson DO, Pennock BE. Oxygen consumption of the respiratory muscles in normal and in malnourished patients with chronic obstructive pulmonary disease. Am Rev Respir Dis 1989;140:385–91.
- 186. Sridhar MK, Carter R, Lean ME, Banham SW. Resting energy expenditure and nutritional state of patients with increased oxygen cost of breathing due to emphysema, scoliosis and thoracoplasty. Thorax 1994;49:781–5.
- 187. McLoughlin DM, Spargo E, Wassif WS, et al. Structural and functional changes in skeletal muscle in anorexia nervosa. Acta Neuropathol Berl 1998;95:632–40.
- 188. Russell DM, Prendergast PJ, Darby PL, Garfinkel PE, Whitwell J, Jeejeebhoy KN. A comparison between muscle function and body composition in anorexia nervosa: the effect of refeeding. Am J Clin Nutr 1983;38:229–37.
- 189. Rigaud D, Moukaddem M, Cohen B, Malon D, Reveillard V, Mignon M. Refeeding improves muscle performance without normalization of muscle mass and oxygen consumption in anorexia nervosa patients. Am J Clin Nutr 1997;65:1845–51.
- 190. Lindboe CF, Askevold F, Slettebo M. Changes in skeletal muscles of young women with anorexia nervosa. An enzyme histochemical study. Acta Neuropathol Berl 1982;56:299–302.
- Essen B, Fohlin L, Thoren C, Saltin B. Skeletal muscle fibre types and sizes in anorexia nervosa patients. Clin Physiol 1981;1:395–403.
- 192. Sieck GC, Lewis MI, Blanco CE. Effects of undernutrition on diaphragm fiber size, SDH activity, and fatigue resistance. J Appl Physiol 1989;66:2196–205.
- 193. Russell DM, Atwood HL, Whittaker JS, et al. The effect of fasting and hypocaloric diets on the functional and metabolic characteristics of rat gastrocnemius muscle. Clin Sci 1984;67:185–94.
- 194. Mijan de la Torre A, Madapallimattam A, Cross A, Armstrong RL, Jeejeebhoy KN. Effect of fasting, hypocaloric feeding, and refeeding on the energetics of stimulated rat muscle as assessed by nuclear magnetic resonance spectroscopy. J Clin Invest 1993;92:114–21.
- 195. Aguilaniu B, Goldstein Shapses S, Pajon A, et al. Muscle protein degradation in severely malnourished patients with chronic obstructive pulmonary disease subject to short-term total parenteral nutrition. JPEN J Parenter Enteral Nutr 1992;16:248–54.
- 196. Goldstein SA, Thomashow BM, Kvetan V, Askanazi J, Kinney JM, Elwyn DH. Nitrogen and energy relationships in malnourished patients with emphysema. Am Rev Respir Dis 1988;138:636–44.
- 197. Pouw EM, Schols AM, Deutz NE, Wouters EF. Plasma and muscle amino acid levels in relation to resting energy expenditure and inflammation in stable chronic obstructive pulmonary disease. Am J Respir Crit Care Med 1998;158:797–801.
- 198. Argilés JM, López-Soriano J. Catabolic proinflammatory cytokines. Curr Opin Clin Nutr Metab Care 1998;1:241–4.
- 199. Hill GL. Body composition research: implications for the practice of clinical nutrition. JPEN J Parenter Enteral Nutr 1992;16:197–218.
- 200. Serres I, Varray A, Vallet G, Micallef JP, Préfaut C. Improved skeletal muscle performance after individualized exercise training in patients with chronic obstructive pulmonary disease. J Cardiopulm Rehabil 1997;17:232–8.
- 201. O'Donnell DE, McGuire M, Samis L, Webb KA. General exercise training improves ventilatory and peripheral muscle strength and

endurance in chronic airflow limitation. Am J Respir Crit Care Med 1998;157:1489–97.

- Adamopoulos S, Coats AJ. Peripheral abnormalities in chronic heart failure. Postgrad Med J 1991;67:S74–80.
- 203. Maltais F, LeBlanc P, Simard C, et al. Skeletal muscle adaptation to endurance training in patients with chronic obstructive pulmonary disease. Am J Respir Crit Care Med 1996;154:442–7.
- 204. Smith K, Cook D, Guyatt GH, Madhavan J, Oxman AD. Respiratory muscle training in chronic airflow limitation: a meta-analysis. Am Rev Respir Dis 1992;145:533–9.
- 205. Rochester DF. The diaphragm in COPD. Better than expected, but not good enough. N Engl J Med 1991;325:961–2.
- 206. Belman MJ, Botnick WC, Nathan SD, Chon KH. Ventilatory load characteristics during ventilatory muscle training. Am J Respir Crit Care Med 1994;149:925–9.
- 207. Rogers RM, Donahoe M, Costantino J. Physiologic effects of oral supplemental feeding in malnourished patients with chronic obstructive pulmonary disease. A randomized control study. Am Rev Respir Dis 1992;146:1511–7.
- 208. Whittaker JS, Ryan CF, Buckley PA, Road JD. The effects of refeeding on peripheral and respiratory muscle function in malnourished chronic obstructive pulmonary disease patients. Am Rev Respir Dis 1990;142:283–8.
- 209. Schols AM, Soeters PB, Mostert R, Pluymers RJ, Wouters EF. Physiologic effects of nutritional support and anabolic steroids in patients with chronic obstructive pulmonary disease. A placebo-controlled randomized trial. Am J Respir Crit Care Med 1995;152:1268–74.
- 210. Fiaccadori E, Coffrini E, Ronda N, et al. A preliminary report on the effects of malnutrition on skeletal muscle composition in chronic obstructive pulmonary disease. In: Ferranti RD, Rampulla C, Fracchia C, Ambrosino N, eds. Nutrition and ventilatory function. Verona, Italy: Bi & Gi Publishers, 1992:77–85.
- 211. Creutzberg EC, Schols AMWJ, Weling-Scheepers CAPM, Wouters EFM. Functional effects of nutritional supplementation therapy incorporated in a pulmonary rehabilitation programme in depleted patients with COPD. Am J Respir Crit Care Med 1998;157:A258 (abstr).
- 212. Creutzberg EC, Schols AMWJ, Weling-Scheepers CAPM, Buurman WA, Wouters EFM. Characterization of nonresponse to high caloric oral nutritional therapy in depleted patients with chronic obstructive pulmonary disease. Am J Respir Crit Care Med 2000;161:745–52.
- 213. Keatings VM, Jatakanon A, Worsdell YM, Barnes PJ. Effects of inhaled and oral glucocorticoids on inflammatory indices in asthma and COPD. Am J Respir Crit Care Med 1997;155:542–8.
- 214. Wigmore SJ, Ross JA, Falconer JS, et al. The effect of polyunsaturated fatty acids on the progress of cachexia in patients with pancreatic cancer. Nutrition 1996;12:S27–30.
- Creutzberg EC, Schols AMWJ. Anabolic steroids. Curr Opin Clin Nutr Metab Care 1999;2:243–53.
- 216. Ferreira IM, Verreschi IT, Nery LE, et al. The influence of 6 months of oral anabolic steroids on body mass and respiratory muscles in undernourished COPD patients. Chest 1998;114:19–28.
- 217. Suchner U, Rothkopf MM, Stanislaus G, Elwyn DH, Kvetan V, Askanazi J. Growth hormone and pulmonary disease. Metabolic effects in patients receiving parenteral nutrition. Arch Intern Med 1990;150:1225–30.
- 218. Pape GS, Friedman M, Underwood LE, Clemmons DR. The effect of growth hormone on weight gain and pulmonary function in patients with chronic obstructive lung disease. Chest 1991;99:1495–500.
- Burdet L, de Muralt B, Schutz Y, Pichard C, Fitting JW. Administration of growth hormone to underweight patients with chronic obstructive pulmonary disease. Am J Respir Crit Care Med 1997;156:1800–6.
- 220. Shapira Y, Glick B, Finsterbush A, Goldfarb A, Rosenmann E. Myopathological findings in thalassemia major. Eur Neurol 1990; 30:324–7.
- 221. Goldfarb AH, Sen CK. Antioxidant supplementation and the control of oxygen toxicity during exercise. In: Sen KC, Packer L, Hänninen O, eds. Exercise and oxygen toxicity. Amsterdam: Elsevier Science BV, 1994:163–89.

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- 222. Novelli GP, Adembri C, Gandini E, et al. Vitamin E protects human skeletal muscle from damage during surgical ischemia-reperfusion. Am J Surg 1997;173:206–9.
- 223. Alessio HM, Goldfarb AH, Cao GH. Exercise induced oxidative stress before and after vitamin C supplementation. Int J Sport Nutr 1997;7:1–9.
- 224. Sen CK, Atalay M, Agren J, Laaksonen DE, Roy S, Hanninen O. Fish oil and vitamin E supplementation in oxidative stress at rest and after physical exercise. J Appl Physiol 1997;83:189–95.
- 225. Appell HJ, Duarte JAR, Soares JMC. Supplementation of vitamin E may attenuate skeletal muscle immobilization atrophy. Int J Sports Med 1997;18:157–60.
- 226. Paiva SA, Godoy I, Vannucchi H, Favaro RM, Geraldo RR, Campana AO. Assessment of vitamin A status in chronic obstructive pulmonary disease patients and healthy smokers. Am J Clin Nutr 1996;64:928–34.
- 227. Dow L, Tracey M, Villar A, et al. Does dietary intake of vitamins C and E influence lung function in older people? Am J Respir Crit Care Med 1996;154:1401–4.
- 228. Nocturnal Oxygen Therapy Trial Group. Continuous or nocturnal oxygen therapy in hypoxemic chronic obstructive lung disease: a clinical trial. Ann Intern Med 1980;93:391–8.

- 229. Medical Research Council Working Party. Long term domiciliary oxygen therapy in chronic hypoxic cor pulmonale complicating chronic bronchitis and emphysema. Lancet 1981;1:681–6.
- 230. O'Donnell DE, Bain DJ, Webb KA. Factors contributing to relief of exertional breathlessness during hyperoxia in chronic airflow limitation. Am J Respir Crit Care Med 1997;155:530–5.
- Woodcock AA, Geddes DM, Gross ER. Oxygen relieves breathlessness in "pink puffers". Lancet 1981;1:907–9.
- 232. Rooyackers JM, Dekhuijzen PN, Van Herwaarden CL, Folgering HT. Training with supplemental oxygen in patients with COPD and hypoxaemia at peak exercise. Eur Respir J 1997;10: 1278-84.
- 233. Jakobsson P, Jorfeldt L. Long-term oxygen therapy may improve skeletal muscle metabolism in advanced chronic obstructive pulmonary disease patients with chronic hypoxaemia. Respir Med 1995;89:471–6.
- 234. Repine JE, Bast A, Lankhorst I. Oxidative stress in chronic obstructive pulmonary disease. Oxidative Stress Study Group. Am J Respir Crit Care Med 1997;156:341–57.
- 235. Jackson RM. Pulmonary oxygen toxicity. Chest 1985;88:900-5.
- 236. Jenkinson SG. Oxygen toxicity. New Horiz 1993;1:504-11.