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**Review Article** 



# Effects of medicinal plants on radiolabeling and biodistribution of diagnostic radiopharmaceuticals: A systematic review

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Article history Abstract Received: 25 February 2019 Radiopharmaceuticals are drugs that contain radioisotopes used for diagnostic and Accepted: 27 March 2019 therapeutic purposes. There are evidences that medicinal herbs and their constituents can Published: 01 April 2019 modify the radiolabeling, biodistribution, and pharmacokinetics of radiopharmaceuticals through drug interaction. To have an overview of the effects and the underlying mechanisms of medicinal plants on the radiolabeling and bioavailability of radiopharmaceuticals, we conducted this study to summarize the current findings in this field. The scientific literature was systematically collected from databases and analyzed. Studies showed that medicinal plants and their constituents can alter radiolabeling and biodistribution via several mechanisms. Interactions with proteins in red blood cells at the same sites, chelating action of stannous and pertechnetate ions, antioxidant action Editor Dr. K. K. Sabu impeding or decreasing stannous ion oxidation, direct oxidation of stannous ions, Jawaharlal Nehru Tropical generation of reactive oxygen species (ROS) which oxidize stannous ions and damage Botanic Garden and Research induced in the cell membrane. Most the medicinal plants can decrease the radioactivity of Institute radiopharmaceuticals, but some of them like Peumus boldus, Punica granatum, Nectandra Thiruvananthapuram, India membranacea, Mentha crispa, Rosmarinus officinalis and derivatives such as eugenol and epigallocatechin gallate have increasing effects. In addition, altering feature in some of them is tissue dependent. Keywords: Medicinal plants; Radiolabeling; Biodistribution; Radiopharmaceuticals **Publisher Citation:** Khosravian P, Heidari-Soureshjani S, Yang Q. Effects of medicinal plants on radiolabeling and biodistribution of diagnostic radiopharmaceuticals: A systematic review. Horizon e-Publishing Group Plant Science Today 2019;6(2):123-131. https://doi.org/10.14719/pst.2019.6.2.513 Copyright: © Khosravian et al (2019). This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are \*Correspondence credited (https://creativecommons.org/licenses/by/4.0/). Saeid Heidari-Soureshjani Maidari\_1983@yahoo.com Indexing: Plant Science Today is covered by Scopus, CAS, AGRIS, CABI, Google Scholar, etc. Full list at http://www.plantsciencetoday.online

#### Introduction

In the traditional medicine, many approaches such as aromatherapy, hypnotherapy, yoga, massage therapy, use of medicinal plants, etc. have been used for diagnosing and treatment of patients (1-7). These approaches are more safe than the methods that have been used in modern medicine. Medicinal plants are one of most commonly used approaches for treatment of diseases that their effects have been indicated in the various studies (8-14). Although medicinal plants can be effective, inexpensive, and efficient treatments for various disorders and frequently used by human beings (15-22), sometimes the biological effects of these natural medicines are not fully known. For example, several studies showed that the radiolabeling biodistribution. or the pharmacokinetics of radiopharmaceuticals can be modified by some medicinal plants (23-27). If drug interactions on biodistribution and labeling are not addressed. poor image quality and misvisualization in nuclear medicine tests could lead to misdiagnosis with a possible need to repeat the test, therefore so increasing the radiation complication to the patient and the staff. Consequently this problem cause mistreatment in the patients (28).

Radiopharmaceuticals are applied for drugs labeled with radioactive tracers for therapeutic or diagnostic purposes in medicine. Radioactive labels are radioactive compounds in which one or more atoms have been replaced by a radionuclide so by virtue of its radioactive decay direct used cell labeling. and thev for Radioisotopes such as carbon, phosphorus. hydrogen, sulfur, and iodine have been applied widespread to trace the path of biochemical reactions (29). Radioactive tracers are used for diagnostic procedures and can be used in a number of tests. For example, technetium-99m (99mTc). in autoradiography and nuclear medicine, including positron emission tomography photon (PET). single emission computed tomography (SPECT) and scintigraphy (30). In addition, radioactive tracers are used to track the distribution of substances within a cell or tissue (31,32). Radiopharmaceuticals have a wide range application in diagnostic nuclear medicine procedures such as lung perfusion and ventilation, brain blood flow, thyroid scan and thyroid uptake, bone, liver, brain scan, hepatobiliary, pancreas, and kidney scans, bone marrow, cardiac imaging procedures, localization of gastrointestinal bleeding, tumor localization studies, and nonimaging studies of blood volume and iron metabolism (33). The radiolabeling of blood constituents is widely used and has great in nuclear medicine for importance both diagnostic imaging and targeted radionuclide therapy (34). Several studies have suggested that radiolabeling of blood constituents with radiopharmaceuticals could be performed to evaluate the biological effects of medicinal plant extracts (23,35,36). Besides, some the synthetic drugs and medicinal herbs can alter the biodistribution of radiopharmaceutical and the analysis of scintigraphic images and physicians must be cautious about this problem (24,37). Hence, this study was conducted to review the effects and mechanisms of medicinal plants, their derivatives and compounds on radiolabeling and bioavailability of diagnostic radiopharmaceuticals.

# **Search strategies**

The words radiopharmaceutical, alongside with, radiolabeling, biodistribution and bioavailability in combination with some herbal terms such as *medicinal plant, phyto\** and *herb\** were used to search for relevant publications indexed in the Institute for Scientific Information (ISI) and PubMed.

A standard form was designed, which included items such as the aim or the title of the study, intervention, outcome, variables, journal name, period, and number. The article's contents that were relevant to this study were recorded on the form and entered into the study upon agreement of the researchers involved in this study. Then the plants and the plant-based products that were reported to be effective in labeling and distribution of diagnostic radiopharmaceuticals were selected. The articles whose full texts were not accessible, studies with neutral effects, non-English language articles, review articles, and studies that were not related to the purpose of this study were excluded after all researchers in this study agreement was achieved. Fig. 1 illustrates how the articles were selected for final analysis.



**Fig. 1.** Flowchart of the process of selecting the articles for final analysis

## Results

The previous studies indicated that that medicinal plants and their derivates can alter radiolabeling and biodistribution. Most the medicinal plants could decrease the radioactivity of radiopharmaceuticals, but some of plants such as Peumus boldus, Punica granatum, Nectandra membranacea, Mentha crispa, Rosmarinus officinalis have increasing effects (Table 1).

In addition, there are some phytochemicals and herbal combinations that affect radiolabeling biodistribution of radiopharmaceuticals and (Table 2).

Several factors that categorized in physical and chemical factors may affect drug delivery and the radiolabeling of a chemical compound (65-68). For instance, pharmaceutical variables, such as sedimentation problems or instability of the cell chelator, choice of anticoagulant, level of stannous ion and oxidation of radiopharmaceuticals, difficulties with collecting sufficient cells, or problems may arise which are patient-related (drug interference or the presence of disease) can affect the radiolabeling and misdistribution of radiopharmaceuticals (69). Medicinal herbs and constituents can reduce the labeling and distribution of diagnostic radiopharmaceuticals in several ways (Fig. 2). Interactions with the blood constituents protein at same sites of radiopharmaceuticals binding and disrupted the radiolabeling efficiency. The other way is direct inhibition action) of (chelating the stannous and pertechnetate ions (23). Because chelator activity has an important pharmacokinetic role in the development of labeling and bioavailability of radiopharmaceuticals targeting (70). Both the antioxidant and oxidant agents (or generation them in chemical interaction) in medicinal plants, in turn, can reduce the cell's labeling. Direct oxidation with the presence of oxidant agents or generation of free radicals that could oxidize the stannous ion (62). In addition, medicinal plants have polyphenols that are micronutrients with antioxidant activity. On the other hand, RBC labeling procedure with radiopharmaceuticals such as <sup>99</sup>mTc depends on the presence of stannous (+2) ions (40). Antioxidant property of medicinal plants can block the action of stannous ions, antioxidant action impeding or decreasing the stannous ion oxidization and binding to them in labeling and fixation procedure; so decreasing the binding of radiopharmaceuticals in plasma proteins (23,71). The last way is an alteration of the plasma membrane structure and receptors or changing in the transport systems of stannous and pertechnetate ions into cells. So, they alter the transport of ions (stannous and pertechnetate ions) through the cellular membrane (23,58). Pertechnetate anions across membranes of human blood constituents have a vital role in The pertechnetate anion may

that the medicinal herbs and their components biodistribution can the of act on radiopharmaceuticals in specific organs (28).

reach the interior of the blood cells constituents

(stannous and pertechnetate ions pass through

the membrane) via the band-3 anion transport

system (72). In other words, medicinal plants

and their derivatives act in the transport of the

pertechnetate ion via the cellular membrane of

determined organs (73). The herbs chemical

compounds can alter the blood cells membrane

morphology (59), and it is one of the altering

mechanisms of radiopharmaceutical labeling on

the cells. Another result of this study suggests

this regards, radiolabeling In and biodistribution also depend on the type of tissue (64). This can be due to differences in receptors in various body tissues. Although, some of the medicinal herbs didn't has an effect on radiolabeling of radiopharmaceuticals. Because labeling depends on the presence of reducing agent or chemical substances, so without such interfering substances, they can't interaction with the radiolabeling process (59,74).

Therefore, precaution must be considered in interpretation diagnostic the of radiopharmaceuticals results when patients are concomitant using these drugs and must be cautious about misdiagnosis and/or repetition of the examinations in nuclear medicine.

Although these results mainly were obtained with animal studies, caution must be considered in the interaction of the nuclear examination when the patient is using these medicinal herbs. So limitation in human studies was a weakness of the reviewed articles which can disturb generalizability of the study's results to the human population.

#### Conclusion

radiolabeling.

Studies showed that medicinal plants and alter radiolabeling constituents can and biodistribution via several mechanisms. They interactions with blood can the cells compartment proteins at same sites, chelating action on the stannous and pertechnetate ions, antioxidant action impeding or decreasing the stannous ion oxidization, direct oxidation of the stannous ions, generation of reactive oxygen species in the body that could oxidize the stannous ions and damage induced in the cell membrane. Limitation in human studies was a weakness of the reviewed articles which can disturb generalizability of the study's results to the human population. Most of them have decreasing in the radioactivity of radio pharmaceuticals, but some of them have increasing effects. In addition, altering feature in some of them is tissue dependent.

References	Plants	Study Design	Type of administration	Main outcomes or mechanisms
Oliveira et al. (25)	Thuya occidentalis	Experimental (in vitro)	Extract	Decrease in the percentage of ATI via inhibition of the transport of the ions, competition with the cited ions for the same binding sites, creating damage in blood cells (BC) membrane and possible generation of reactive oxygen species that could oxidize the stannous ion.
Reiniger et al. (38)	Peumus boldus	Experimental (in vitro)	Solution	Increasing labeling of red blood cells and increase radioactivity bound to the red blood cell (RBC).
Braga et al. (39)	Thuya occidentalis, Peumus boldus and Nicotiana tabacum (tobacco)	Experimental (in vitro)	Extract	Decrease in the percentage of ATI, <i>Nicotianatabacum</i> decreasing labeling of RBC and plasma proteins and <i>Peumusboldus</i> increase uptake of the radioactivity by blood cells but a slight decrease in the amount of <sup>99</sup> mTc radioactivity.
de Oliveira et al. (40)	Maytenus ilicifolia	Experimental (in vitro)	Extract	Decrease in the percentage of ATI on blood elements. This changes due to the presence of oxidant agents in the <i>Maytenus ilicifolia</i> extract.
de Oliveira et al. (41)	Paullinia cupana	Experimental (in vitro)	Solution	Decreasing in the uptake of radioactivity by the RBC in insoluble fractions plasma (IF-P) and in insoluble fractions blood cell (IF-BC) and also alterations in the shape of RBC was observed.
Feliciano et al. (42)	Sechium edule	Experimental (in vivo and in vitro)	Extract	Decrease of the radioactivity in BC, IF-BC and IF-P and slight decrease in the uptake of <sup>99</sup> mTc by BC and a strong decrease in the fixation in IF-P with the macerated <i>in vivo</i> . But <i>in vitro</i> study, no alterations on the labeling of blood elements were reported.
Vidal et al. (43)	Nicotiana tabacum	Experimental (in vitro)	Extract	Decreasing labeling of RBC and plasma proteins due to direct or indirect effect (reactive oxygen species) of tobacco by oxidation of the stannous ion, possible chelating action on the stannous and/or pertechnetate ions and possible damages caused in the plasma membrane.
Amorim et al. (44)	Punica granatum	Experimental (in vitro)	Infusion	Increase in the percentage of ATI in lungs, brain, spleen, heart, liver, stout bowel, stomach, muscle, femur, kidneys, pancreas, and testis through the generation of active metabolites.
Abreu et al. (23)	Psidium guajava	Experimental (in vitro)	Aqueous extract	Decreasing the radiolabeling of BC via antioxidant action and alters the membrane structures involved in ion transport into cells.
Farias et al. (45)	Nectandra membranacea	Experimental (in vitro)	Aqueous extract	Increase in the percentage of ATI of <sup>99</sup> mTc by muscle, thyroid, heart, kidney and thyroid (%ATI/gram of tissue). So the extract could generate active metabolites able to influence the bioavailability of <sup>99</sup> mTc.
Moreno et al.	Cinkgo hiloha	Experimental (in	Solution	Decreasing the uptake of the Na <sup>99m</sup> TcO <sub>4</sub> in the
(35)	Glikgo biloba	vivo and in vitro)	30101011	duodenum. This may be due to scavengers of free radicals and chelating agents in the extract.
Moreno et al. (46)	Ginkgo biloba	Experimental (in vivo and in vitro)	Extract	Decrease uptake of the Na <sup>99m</sup> TcO <sub>4</sub> in the duodenum. It is possible that <i>in vivo</i> treatment with extract may generate active metabolites that could alter the ultrastructure of cells and the biodistribution of the Na <sup>99m</sup> TcO <sub>4</sub> .
Moreno et al. (28)	Uncaria tomentosa	Experimental (in vitro)	Solution	Decreasing in the uptake of this Na <sup>99m</sup> TcO <sub>4</sub> %ATI/organ was observed in the heart. But the extract can increasing uptake radiobiocomplex %ATI/g in the Pancreas and muscles.
Santos et al. (47)	Hypericum perforatum	Experimental (in vitro)	Extract	Decreasing in radioactivity distribution and interfering with the fixation on the IF-P and IF-BC in young rats (not in elderly one).
Santos et al. (48)	Mentha crispa	Experimental (in vitro)	Aqueous extract	Increasing fixation observed in kidney, spleen, and thyroid the %ATI/g values by the generation of active metabolites.
Cekic et al. (49)	Brassica oleraceaItalica	Experimental (in vivo and in vitro)	Methanolic extract	Decreasing in the percentage of injected dose per gram of tissue weight (% ID/g) particular in the kidney. The extract was not effected labeling blood components with <sup>99</sup> mTc.
Rebello et al. (50)	Passiflora edulis	Experimental (in vivo and in vitro)	Solution	Decreasing in the percentage of ATI/g of tissues from duodenum, spleen, pancreas, and blood and increasing in stomach tissue. The extract does not alter the labeling of blood constituents.
Souza et al. (51)	Cassia angustifolia	Experimental (in vivo and in vitro)	Aqueous extract	Reducing %ATI/g of the Na <sup>99m</sup> TcO <sub>4</sub> in the thyroid, liver, pancreas, lungs and blood. The extract did not modify the radiolabeling of the blood constituents.

### Table 1: Contd.

Zora et al. (52)	Passiflora incarnata	Experimental (in vivo and in vitro)	Syrup	Reducing biodistribution uptake of <sup>99m</sup> Tc-DTPA in the kidney and reducing %ID/g of in particular kidney. In addition percentage of radioactivity on serum and BC decreased.	
Benarroz et al. (36)	Cinnamomum zeylanicum	Experimental (in vitro)	Solution	Decreased significantly the percentage of ATI on BC, IF-P and IF-BC.	
Garcia-Pinto et al. (53)	Matricaria recutita	Experimental (in vivo and in vitro)	Aqueous extract	Reducing radioactivity on blood cells compartment and on insoluble fractions of plasma and increasing of the percentage of radioactivity in the stomach.	
Ucar et al. (54)	Rosmarinus officinalis	Experimental (in vitro)	Methanolic extract	Increasing in the uptake of <sup>99</sup> mTc-SC in the liver.	
Holanda et al. (24)	Annona muricata	Experimental (in vivo and in vitro)	Solution	Decreasing in the uptake of ATI/g percent in bladder, kidney, and blood.	
Holanda et al. (37)	Aloe vera	Experimental (in vivo and in vitro)	Aqueous extract	Increasing of %ATI/g in blood, kidneys, stomach, liver, testis femur and thyroid, so increasing uptake of $\rm Na^{99m}TcO_4.$	
Capriles et al. (55)	Solanum melongena	Experimental (in vitro)	Extract	Decreasing the labeling of RBC, IF-P, reducing %ATI, and IF-RBC due to the extract oxidation potential.	
de Oliveira et al. (56)	Fucus vesiculosus	Experimental (in vitro)	Extract	Decreasing the %ATI, IF-P and IF-BC via increase the valence of these ions to stannic (+4).	
de Paoli et al. (57)	Caryophyllu saromaticus	Experimental (in vitro)	Aqueous extract	Decreasing in the percentage of ATI of blood constituents and theradioactivity fixation.	
Neves et al. (58)	Arctium lappa	Experimental (in vitro)	Aqueous extract	Decreasing in the percentage of ATI and the RBC labeling through alter the transport of ions (stannous and pertechnetate ions).	
Rebello et al. (59)	Passiflora edulis	Experimental (in vitro)	Solution	Decreasing the fixation of <sup>99</sup> mTc on plasma proteins and BC but it can't interfere with blood cells labeling.	
Terra et al. (60)	Artemisia vulgaris	Experimental (in vitro)	Solution	Decreasing in the percentage of ATI on the blood compartments and on the blood cells proteins (IF-BC and IF-P) via erythrocyte membrane alternating.	
Sabuncu et al. (61)	Camellia sinensis	Experimental (in vivo)	Ethanolic and aqueous extract	Radiolabeling was not changed in blood component but decreasing the uptake of Na <sup>99</sup> mTcO <sub>4</sub> by stomach, liver and prostate.	

Table 2: Effects of phytochemicals and herbal combinations on radiolabeling and biodistribution of radiopharmaceuticals

References	Herbal compounds/derivatives	Study Design	Type of administra- tion	Type of radiopharma -ceutical	Main outcomes or mechanisms
Giani et al. (62)	Buzhong Yi Qi Wan composed of Radix Astragalus, Radix codonopsis, Radix glycyrrhizae, Rhizoma atractylodis macrocephalae, Radix angelicaesinensis, Rhizoma cimicifugae, Radix bupleuri, Pericarpium citri reticulatae, Rhizoma zingiberis recens and Fructus jujubae	Experimental (in vitro)	Aqueous extract	<sup>99</sup> mTc	Decreasing in radioactivity distribution by the BC was reported. Besides decreasing in radioactivity fixation by the IF-BC and IF-P via free radicals generation causing oxidation of the stannous ions and the fixation of <sup>99</sup> mTc would be decreased.
Mattos et al. (26)	Vincristine derived from <i>Vincarosea</i> Linn	Experimental (in vivo)	Extract	<sup>99</sup> mTc	Percentage of radioactivity decreased in the ovary, uterus, lymph nodes (inguinal and mesentheric), thymus, spleen, kidney, pancreas, liver, stomach, brain, heart and bone of the animals.
Dervis et al. (63)	Eugenol derived from Syzygium aromaticum, Pimenta racemosa and Cinnamomum verum	Experimental (in vitro)	Extract	Iodine-131 ( <sup>131</sup> I)	The extract causes radioiodinated with <sup>131</sup> I, which is a convenient radioisotope for therapy and potential for therapy and imaging due to its notable uptake.
Kamal et al. (27)	Resveratrol derived from herbs	Experimental (in vitro)	Extract	Technetium- <sup>99</sup> m labelled resveratrol loaded gold nanoparticles (Res-AuNP)	administration of ( <sup>99</sup> m)Tc-Res-AuNP to colon tumor, demonstrating better <i>in vivo</i> targeting of colon adenocarcinoma with ( <sup>99</sup> m)Tc-Res- AuNP when compared to ( <sup>99</sup> m)Tc- resveratrol
Toksoz et al. (64)	Epigallocatechin gallate (EGCG) derived from <i>Camellia sinensis</i> Theaceae	Experimental (in vitro)	Solution	<sup>131</sup> I	EGCG increasing radiolabeling in lung, liver, pancreas, and kidney. Besides max biodistribution was seen in lung and pancreas.



Fig. 2. Altering mechanisms of medicinal plants, derivatives and compounds on radiolabeling and bioavailability of radiopharmaceuticals

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# **Conflicts of interest**

The authors declare no conflict of interest.

#### References

- Hasanpour-Dehkordi A, Jivad N, Solati K. Effects of yoga on physiological indices, anxiety and social functioning in multiple sclerosis patients: A randomized trial. Journal of Clinical and Diagnostic Research. 2016; 10(6): VC01-VC5. https://doi.org/10.7860/JCDR/2016/18204.7916
- 2. Nikfarjam M, Dehkordi KS, Aghaei A, Rahimian G. Efficacy of hypnotherapy in conjunction with pharmacotherapy and pharmacotherapy alone on the quality of life in patients with irritable bowel syndrome. Govaresh. 2013;18(3):149-56.
- Shahbazi K, Solati-Dehkordi K, Dehkordi AH. Comparison of hypnotherapy and standard medical treatment alone on quality of life in patients with irritable bowel syndrome: A randomized control trial. Journal of Clinical and Diagnostic Research. 2016; 10(5): OC01-OC4. <u>https://doi.org/10.7860/JCDR/2016/17631.7713</u>
- 4. Solati K. Effectiveness of cognitive-behavior group therapy, psycho-education family, and drug therapy in reducing and preventing recurrence of symptoms in patients with major depressive disorder. Journal of Chemical and Pharmaceutical Sciences. 2016;9(4):3414-8.
- 5. Dehkordi KS, Nikfarjam M, Sanaei S. Effectiveness of mindfulness-based stress reduction training and drug therapy on quality of life in patients with irritable bowel syndrome in Shahrekord. Life Science Journal. 2014;11(9):445-9.
- 6. Noghabi AAA, Zandi M, Mehran A, Alavian SM, Dehkordi AH. The effect of education on quality of life

in patients under interferon therapy. Hepatitis Monthly. 2010;10(3):218-22.

- Solati K, Mousavi M, Kheiri S, Hasanpour-Dehkordi A. The effectiveness of mindfulness-based cognitive therapy on psychological symptoms and quality of life in systemic lupus erythematosus patients: A randomized controlled trial. Oman Medical Journal. 2017;32(5):378-85. <u>https://doi.org/10.5001/omj.2017.73</u>
- 8. Rouhi-Boroujeni H, Asadi-Samani M, Moradi MT. A review of the medicinal plants effective on headache based on the ethnobotanical documents of Iran. Der Pharmacia Lettre. 2016;8(3):37-42.
- 9. Kooti W, Moradi M, Ali-Akbari S, Sharafi-Ahvazi N, Asadi-Samani M, Ashtary-Larky D. Therapeutic and pharmacological potential of *Foeniculum vulgare* Mill: A review. Journal of Herbmed Pharmacology. 2015;4(1):1-9.
- Mohsenzadeh A, Ahmadipour S, Ahmadipour S, Asadi-Samani M. Iran's medicinal plants effective on fever in children: A review. Der Pharmacia Lettre. 2016;8(1):129-34.
- 11. Parsaei P, Bahmani M, Naghdi N, Asadi-Samani M, Rafieian-Kopaei MM, Boroujeni S. Shigellosis phytotherapy: A review of the most important native medicinal plants in Iran effective on Shigella. Der Pharmacia Lettre. 2016;8(2):249-55.
- 12. Kooti W, Ghasemiboroon M, Asadi-Samani M, Ahangarpoor A, Zamani M, Amirzargar A, et al. The effect of Halcoholic extract of celery leaves on the delivery rate (fertilization and stillbirths), the number, weight and sex ratio of rat off spring. Advances in Environmental Biology. 2014;8(10):824-30.
- Hosseini Z, Lorigooini Z, Rafieian-Kopaei M, Shirmardi H, Solati K. A review of botany and pharmacological effect and chemical composition of Echinophora species growing in Iran. Pharmacognosy Research. 2017;9(4):305-12. <u>https://doi.org/10.4103/pr.pr 22\_17</u>
- 14. Heidarian E, Movahed-Mohammadi G, Saffari J, Ghatreh-Samani K. Protective effect of hydroethanolic extract of cress against hepatotoxicity due to acetaminophen in rats. Journal of Mazandaran University of Medical Sciences. 2013;23(102):78-90.

- Heidari-Soreshjani S, Asadi-Samani M, Yang Q, Saeedi-Boroujeni A. Phytotherapy of nephrotoxicity-induced by cancer drugs: an updated review. Journal of Nephropathology. 2017;6(3):254-63. <u>https://doi.org/10.15171/jnp.2017.41</u>
- 16. Memarzadeh E, Luther T, Heidari-Soureshjani S. Effect and mechanisms of medicinal plants on dry eye disease: A systematic review. Journal of Clinical and Diagnostic Research. 2018;12(9):NE1-NE4. https://doi.org/10.7860/JCDR/2018/36409.12042
- 17. Shabanian G, Heidari-Soureshjani S, Rafieian-Kopaei M, Saadat M, Shabanian M. Therapeutic effects of *Quercus persica* L fruit skin on healing of second-degree burn wounds in animal model. Journal of Zanjan University of Medical Sciences and Health Services. 2017;25(113):81-92.
- 18. Shabanian S, Khalili S, Lorigooini Z, Malekpour A, Heidari-Soureshjani S. The effect of vaginal cream containing ginger in users of clotrimazole vaginal cream on vaginal candidiasis. Journal of Advanced Pharmaceutical Technology & Research. 2017;8(2):80-4. https://doi.org/10.4103/japtr.JAPTR 176\_16
- 19. Shirani M, Raeisi R, Heidari-Soureshjani S, Asadi-Samani M, Luther T. A review for discovering hepatoprotective herbal drugs with least side effects on kidney. Journal of Nephropharmacology. 2017;6(2):38-48. <u>https://doi.org/10.15171/npj.2017.03</u>
- 20. Yavangi M, Rabiee S, Nazari S, Farimani-Sanoee M, Amiri I, Bahmanzadeh M, et al. Comparison of the effect of oestrogen plus *Foeniculum vulgare* seed and oestrogen alone on increase in endometrial thickness in infertile women. Journal of Clinical and Diagnostic Research. 2018;12(1):QC01-QC4. https://doi.org/10.7860/JCDR/2017/30164.11020
- 21. Heidari-Soreshjani S, Asadi-Samani M, Yang Q, Saeedi-Boroujeni A. Phytotherapy of nephrotoxicity-induced by cancer drugs: An updated review. Journal of Nephropathology. 2017;6(3):254-63. https://doi.org/10.15171/jnp.2017.41
- 22. Shirani-Boroujeni M, Heidari-Soureshjani S, Keivani Hafshejani Z. Impact of oral capsule of *Peganum harmala* on alleviating urinary symptoms in men with benign prostatic hyperplasia; a randomized clinical trial. Journal of Renal Injury Prevention. 2017;6(2):127-31. <u>https://doi.org/10.15171/jrip.2017.25</u>
- 23. Abreu PR, Almeida MC, Bernardo RM, Bernardo LC, Brito LC, Garcia EA, et al. Guava extract (*Psidium guajava*) alters the labeling of blood constituents with technetium-99m. Journal of Zhejiang University Science B. 2006;7(6):429-35. https://doi.org/10.1631/jzus.2006.B0429
- 24. Holanda CM, Barbosa DA, Demeda VF, Bandeira FT, Medeiros HC, Pereira KR, et al. Influence of *Annona muricata* (soursop) on biodistribution of radiopharmaceuticals in rats. Acta Cirúrgica Brasileira. 2014;29(3):145-50. <u>https://doi.org/10.1590/S0102-86502014000300001</u>
- 25. Oliveira JF, Braga AC, Avila AS, Fonseca LM, Gutfilen B, Bernardo-Filho M. Effect of *Thuya occidentalis* on the labeling of red blood cells and plasma proteins with technetium-99m. Yale Journal of Biology and Medicine. 1996;69(6):489-94.
- 26. Mattos DM, Gomes ML, Freitas RS, Rodrigues PC, Paula EF, Bernardo-Filho M. A model to evaluate the biological effect of natural products: vincristine action on the biodistribution of radiopharmaceuticals in BALB/ c female mice. Journal of Applied Toxicology. 1999;19(4):251-4.

- 27. Kamal R, Chadha VD, Dhawan DK. Physiological uptake and retention of radiolabeled resveratrol loaded gold nanoparticles ((99m)Tc-Res-AuNP) in colon cancer tissue. Nanomedicine. 2018;14(3):1059-71. https://doi.org/10.1016/j.nano.2018.01.008
- 28. Moreno SR, Silva AL, Dire G, Honeycut H, Carvalho JJ, Nascimento AL, et al. Effect of oral ingestion of an extract of the herb *Uncaria tomentosa* on the biodistribution of sodium pertechnetate in rats. Brazilian Journal of Medical and Biological Research. 2007;40(1):77-80.
- 29. Lappin G. A historical perspective on radioisotopic tracers in metabolism and biochemistry. Bioanalysis. 2015;7(5):531-40. https://doi.org/10.4155/bio.14.286
- Zanzonico P. Principles of nuclear medicine imaging: planar, SPECT, PET, multi-modality, and autoradiography systems. Radiation Research. 2012;177(4):349-64.
- 31. Zheng Y, Huang J, Zhu T, Li R, Wang Z, Ma F, et al. Stem Cell Tracking Technologies for Neurological Regenerative Medicine Purposes. Stem Cells International. 2017;2017:2934149. https://doi.org/10.1155/2017/2934149
- 32. Ahrens ET, Bulte JW. Tracking immune cells in vivo using magnetic resonance imaging. Nature Reviews Immunology. 2013;13(10):755. https://doi.org/10.1038/nri3531
- 33. Drozdovitch V, Brill AB, Callahan RJ, Clanton JA, DePietro A, Goldsmith SJ, et al. Use of radiopharmaceuticals in diagnostic nuclear medicine in the United States: 1960-2010. Health Physics. 2015;108(5):520-37. https://doi.org/10.1097/HP.000000000000261
- 34. Fani M, Maecke HR, Okarvi SM. Radiolabeled peptides: valuable tools for the detection and treatment of cancer. Theranostics. 2012;2(5):481-501. https://doi.org/10.7150/thno.4024
- 35. Moreno SR, Carvalho JJ, Nascimento AL, Pereira M, Rocha EK, Olej B, et al. Experimental model to assess possible medicinal herb interaction with a radiobiocomplex: qualitative and quantitative analysis of kidney, liver and duodenum isolated from treated rats. Food and Chemical Toxicology. 2007;45(1):19-23. https://doi.org/10.1016/j.fct.2006.04.007
- 36. Benarroz MO, Fonseca AS, Rocha GS, Frydman JN, Rocha VC, Pereira MO, et al. Cinnamomum zeylanicum extract on the radiolabeling of blood constituents and the morphometry of red blood cells: *in vitro* assay. Applied Radiation and Isotopes. 2008;66(2):139-46. https://doi.org/10.1016/j.apradiso.2007.08.004
- 37. Holanda CM, Costa MB, Silva NC, Silva MF, Barbosa VS, Silva RP, et al. Effect of an extract of *Aloe vera* on the biodistribution of sodium pertechnetate (Na99mTcO4) in rats. Acta Cirúrgica Brasileira. 2009;24(5):383-6. http://dx.doi.org/10.1590/S0102-86502009000500008
- Reiniger IW, de Oliveira JF, Caldeira-de-Araujo A, Bernardo-Filho M. Effect of *Peumus boldus* on the labeling of red blood cells and plasma proteins with technetium-99m. Applied Radiation and Isotopes. 1999;51(2):145-9. <u>https://doi.org/10.1016/S0969-</u> 8043(98)00200-0
- 39. Braga AC, Oliveira MB, Feliciano GD, Reiniger IW, Oliveira JF, Silva CR, et al. The effect of drugs on the labeling of blood elements with technetium-99m. Current Pharmaceutical Design. 2000;6(11):1179-91. https://doi.org/10.2174/1381612003399897

- 40. de Oliveira JF, Braga AC, de Oliveira MB, Avila AS, Caldeira-de-Araujo A, Cardoso VN, et al. Assessment of the effect of *Maytenus ilicifolia (Espinheira santa)* extract on the labeling of red blood cells and plasma proteins with technetium-99m. Journal of Ethnopharmacology. 2000;72(1-2):179-84. <u>https://doi.org/ 10.1016/S0378-8741(00)00240-3</u>
- 41. de Oliveira JF, Avila AS, Braga ACS, de Oliveira MBN, Boasquevisque EM, Jales RL, et al. Effect of extract of medicinal plants on the labeling of blood elements with Technetium-99m and on the morphology of red blood cells: I - a study with Paullinia cupana. Fitoterapia. 2002;73(4):305-12. <u>https://doi.org/10.1016/S0367-326X(02)00073-4</u>
- 42. Feliciano GD, Lima EA, Pereira MJ, de Oliveira MB, Moreno SR, de Mattos DM, et al. Effect of a chayotte (*Sechium edule*) extract on the labeling of red blood cells and plasma proteins with technetium-99m: *in vitro* and *in vivo* studies. Cellular and Molecular Biology. 2002;48(7):751-5.
- 43. Vidal MV, Gutfilen B, da Fonseca LM, Bernardo-Filho M. Influence of tobacco on the labelling of red blood cells and plasma proteins with technetium-99m. Journal of Experimental & Clinical Cancer Research. 1998;17(1):41-6.
- 44. Amorim LF, Catanho MT, Terra DA, Brandao KC, Holanda CM, Jales-Junior LH, et al. Assessment of the effect of *Punica granatum* (pomegranata) on the bioavailability of the radiopharmaceutical sodium pertechnetate (99mTc) in Wistar rats. Cellular and Molecular Biology. 2003;49(4):501-7.
- 45. Farias Moreno SR, Arnobio A, De Carvalho JJ, Nascimento AL, Timoteo MO, Olej B, et al. The ingestion of a *Nectandra membranacea* extract changes the bioavailability of technetium-99m radiobiocomplex in rat organs. Biological Research. 2007;40(2):131-5. <u>https:// doi.org//S0716-97602007000200004</u>
- 46. Moreno SRF, de Carvalho JJ, Nascimento AL, Arnobio A, Olej B, Timoteo MD, et al. Ultrastructural analysis of kidney, liver and duodenum isolated from treated rats with *Ginkgo biloba* extract and effects of this medicinal plant on the biodistribution of the radiopharmaceutical sodium pertechnetate. Brazilian Archives of Biology and Technology. 2008;51:185-90. http://dx.doi.org/10.1590/S1516-89132008000700030
- 47. Santos SD, da Fonseca AD, Bernardo M. The male reproductive system and the effect of an extract of a medicinal plant (*Hypericum perforatum*) on the labeling process of blood constituents with technetium-99m. Brazilian Archives of Biology and Technology. 2007;50:97-104. <u>http://dx.doi.org/10.1590/S1516-89132007000600011</u>
- 48. Santos SD, Maiworm AI, Presta GA, de Paoli S, Giani TS, Bernardo M. Aqueous extract of the medicinal plant *Mentha crispa* alters the biodistribution of the radiopharmaceutical sodium pertechnetate in Wistar rats. Medicinal Chemistry Research. 2007;16(5):230-7. <u>http://dx.doi.org/10.1007/s00044-007-9026-7</u>
- 49. Cekic B, Muftuler FZ, Kilcar AY, Ichedef C, Unak P. Effects of broccoli extract on biodistribution and labeling blood components with 99mTc-GH. Acta Cirúrgica Brasileira. 2011;26(5):339-45. <u>http://dx.doi.org/</u> 10.1590/S0102-86502011000500003

- 50. Rebello BM, Moreno SR, Godinho CR, Neves RF, Fonseca AS, Bernardo-Filho M, et al. Effects of *Passiflora edulis* flavicarpa on the radiolabeling of blood constituents, morphology of red blood cells and on the biodistribution of sodium pertechnetate in rats. Applied Radiation and Isotopes. 2008;66(12):1788-92. https://doi.org/10.1016/j.apradiso.2008.05.004
- 51. Souza DE, Pereira MO, Bernardo LC, Carmo FS, Fonseca AdSd, Bernardo-Filho M. An experimental model to study the effects of a senna extract on the blood constituent labeling and biodistribution of a radiopharmaceutical in rats. Clinics (Sao Paulo, Brazil). 2011;66(3):483-6. https://dx.doi.org/10.1590%2FS1807-59322011000300021
- 52. Zora H, Muftuler ZFB, Demir I, Kilcar AY, Ichedef C, Unak P. Effect of a plant origin drug on the biodistribution of Tc-99m-DTPA in Wistar albino rats. Brazilian Journal of Pharmacognosy. 2012;22(2):344-9. http://dx.doi.org/10.1371/journal.pone.0062328
- 53. Garcia-Pinto AB, Santos-Filho SD, Carvalho JJ, Pereira MJS, Fonseca AS, Bernardo-Filho M. In vitro and in vivo studies of an aqueous extract of *Matricaria recutita* (German chamomile) on the radiolabeling of blood constituents, on the morphology of red blood cells and on the biodistribution of the radiopharmaceutical sodium pertechnetate. Pharmacognosy Magazine. 2013;9(Suppl 1):S49-S56. https://dx.doi.org/10.4103%2F0973-1296.117867
- 54. Ucar E, Teksoz S, Ichedef C, Kilcar AY, Unak P. Effect of rosemary (*Rosmarinus officinalis*) extract on the biodistribution of Tc-99m sulphur colloid and on the radiolabeled blood constituents. Brazilian Journal of Pharmacognosy. 2013;23(1):182-5. https://doi.org/10.1590/S0102-695X2012005000148
- 55. Capriles P, Dias APM, Costa T, Oliveira MBN, Faria MVC, Moura EG, et al. Effect of eggplant (*Solanum melongena*) extract on the in vitro labeling of blood elements with technetium-99m and on the biodistribution of sodium pertechnetate in rats. Cellular and Molecular Biology. 2002;48(7):771-6.
- 56. de Oliveira JF, de Oliveira MBN, Avila AS, Braga ACS, Catanho M, Jales RLC, et al. Assessment of the effect of *Fucus vesiculosus* extract on the labeling of blood constituents with technetium-99m and the histological modifications on the shape of the red blood cells. Food and Chemical Toxicology. 2003;41(1):15-20. https://doi.org/10.1016/S0278-6915(02)00206-5
- 57. de Paoli S, Giani TS, Presta GA, Pereira MO, da Fonseca AD, Brandao J, et al. Effects of clove (*Caryophyllus aromaticus* L.) on the labeling of blood constituents with technetium-99m and on the morphology of red blood cells. Brazilian Archives of Biology and Technology. 2007;50:175-82. <u>http://dx.doi.org/10.1590/S1516-89132007000600022</u>
- 58. Neves RD, Moreno SRF, Rebello BM, Caldas L, da Fonseca AD, Bernardo M, et al. Effect of an Arctium lappa (burdock) extract on the labeling of blood constituents with technetium-99m and on the morphology of the red blood cells. Brazilian Archives of Biology and Technology. 2007;50:167-74. http://dx.doi.org/10.1590/S1516-89132007000600021
- 59. Rebello BM, Moreno SRF, Ribeiro CG, Neves RDF, Da Fonseca ADS, Caldas L, et al. Effect of a Peel Passion Fruit Flour (*Passiflora edulis* F. flavicarpa) extract on the labeling of blood constituents with technetium-99m and on the morphology of red blood cells. Brazilian Archives of Biology and Technology. 2007;50:153-9. http://dx.doi.org/10.1590/S1516-89132007000600019

- Terra DA, Amorim LD, Catanho M, da Fonseca AD, Santos SD, Brandao J, et al. Effect of an extract of *Artemisia vulgaris* L. (mugwort) on the in vitro labeling of red blood cells and plasma proteins with technetium-99m. Brazilian Archives of Biology and Technology. 2007;50:123-8. <u>http://dx.doi.org/10.1590/S1516-89132007000600015</u>
- Sabuncu B, Muftuler FZB, Kilcar AY, Cekic B, Ucar E, Unak P. Interaction between green tea extract and Tc-99m-pertechnetate on in vivo distribution. Journal of Radioanalytical and Nuclear Chemistry. 2014;300(3):1021-6. <u>http://dx.doi.org/10.1007/s10967-014-3089-y</u>
- 62. Giani TS, de Paoli S, Presta GA, Maiworm AI, Santos SD, Brandao J, et al. An extract of a formula used in the Traditional Chinese Medicine (Buzhong Yi Qi Wan) alters the labeling of blood constituents with technetium-99m. Brazilian Archives of Biology and Technology. 2007;50:111-6. http://dx.doi.org/10.1590/S1516-89132007000600013
- Dervis E, Yurt Kilcar A, Medine EI, Tekin V, Cetkin B, Uygur E, et al. *In vitro* incorporation of radioiodinated eugenol on adenocarcinoma cell lines (Caco2, MCF7, and PC3). Cancer Biotherapy and Radiopharmaceuticals. 2017;32(3):75-81. <u>https://doi.org/ 10.1089/cbr.2017.2181</u>
- 64. Toksoz F, Demir I, Bayrak E, Kocagozoglu G, Onursal M, Karademir G, et al. Radiolabeling of EGCG with I-131 and biodistribution in rats. Medicinal Chemistry Research. 2012;21(2):224-8. https://doi.org/10.1007/s00044-010-9535-7
- 65. Finn R, Cheung NK, Divgi C, St Germain J, Graham M, Pentlow K, et al. Technical challenges associated with the radiolabeling of monoclonal antibodies utilizing short-lived, positron emitting radionuclides. International Journal of Radiation Applications and Instrumentation Part B, Nuclear Medicine and Biology. 1991;18(1):9-13.
- 66. Khosravian P, Khoobi M, Ardestani MS, Daryasari MP, Hassanzadeh M, Ghasemi-Dehkordi P, et al. Enhancement antimicrobial activity of clarithromycin by amine functionalized mesoporous silica nanoparticles as drug delivery system. Letters in Drug Design and Discovery. 2018;15(7):787-95. <u>https://doi.org/ 10.2174/1570180815666180117155818</u>

- Akbarian S, Sojoodi J, Monnavari F, Heidari H, Khosravian P, Javar HA, et al. Nano conjugated PLGA-Chlorambucil: Synthesis In vitro anti non-Hodgkin's lymphoma cellular assay. Letters in Drug Design and Discovery. 2017;14(7):827-36. https://doi.org/10.2174/1570180814666161130113446
- Khosravian P, Ardestani MS, Khoobi M, Ostad SN, Dorkoosh FA, Javar HA, et al. Mesoporous silica nanoparticles functionalized with folic acid/methionine for active targeted delivery of docetaxel. OncoTargets and Therapy. 2016;9:7315-30. https://doi.org/10.2147/OTT.S113815
- 69. Sampson CB. Complications and difficulties in radiolabeling blood cells: a review. Nuclear Medicine Communications. 1996;17(8):648-58.
- 70. Ray Banerjee S, Pullambhatla M, Foss CA, Falk A, Byun Y, Nimmagadda S, et al. Effect of chelators on the pharmacokinetics of (99m)Tc-labeled imaging agents for the prostate-specific membrane antigen (PSMA). Journal of Medicinal Chemistry. 2013;56(15):6108-21. https://doi.org/10.1021/jm400823w
- Jesus LM, Abreu PR, Almeida MC, Brito LC, Soares SF, de Souza DE, et al. A propolis extract and the labeling of blood constituents with technetium-99m. Acta Biologica Hungarica. 2006;57(2):191-200. https://doi.org/10.1556/ABiol.57.2006.2.6
- 72. Callahan RJ, Rabito CA. Radiolabeling of erythrocytes with technetium-99m: role of band-3 protein in the transport of pertechnetate across the cell membrane. Journal of Nuclear Medicine. 1990;31(12):2004-10.
- 73. Santos-Filho SD, Maiworm AI, Presta GA, de Paoli S, Giani TS, Bernardo-Filho M. Aqueous extract of the medicinal plant *Mentha crispa* alters the biodistribution of the radiopharmaceutical sodium pertechnetate in Wistar rats. Medicinal Chemistry Research. 2007;16(5):230-7. <u>https://doi.org/10.1007/s00044-007-9026-7</u>
- 74. Carmo FS, Diniz CL, Pereira MO, Santos SD, Bernardo M. Characterization of physicochemical parameters and the effect on the labeling of blood constituents with technetium-99m of a *Solanum melongena* commercial extract. Journal of Medicinal Plants Research. 2011;5(23):5598-604.