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## Synthesis of novel [3,2-*b*]indole fused oleanolic acids as potential inhibitors of cell proliferation

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**Dedicated to Dr. Joseph M. Muchowski on the occasion of his 65<sup>th</sup> birthday, and in recognition of his numerous outstanding contributions to indole and pyrrole chemistry**

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### Abstract

Seven new indole-fused oleanolic acid derivatives were synthesized from oleanolic acid for their ability to inhibit cell proliferation in NRP.152 cells.

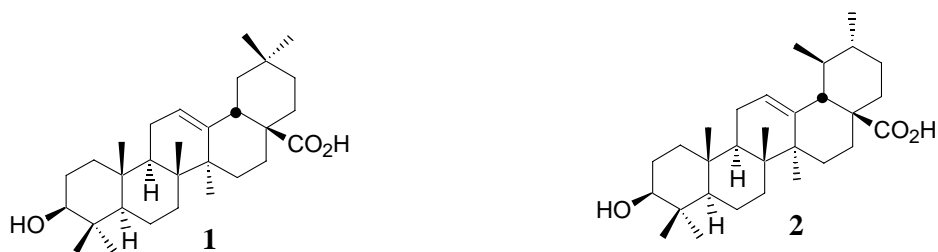
**Keywords:** Oleanolic acid, cell proliferation, Fischer indole synthesis, indolotriterpenoids

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### Introduction

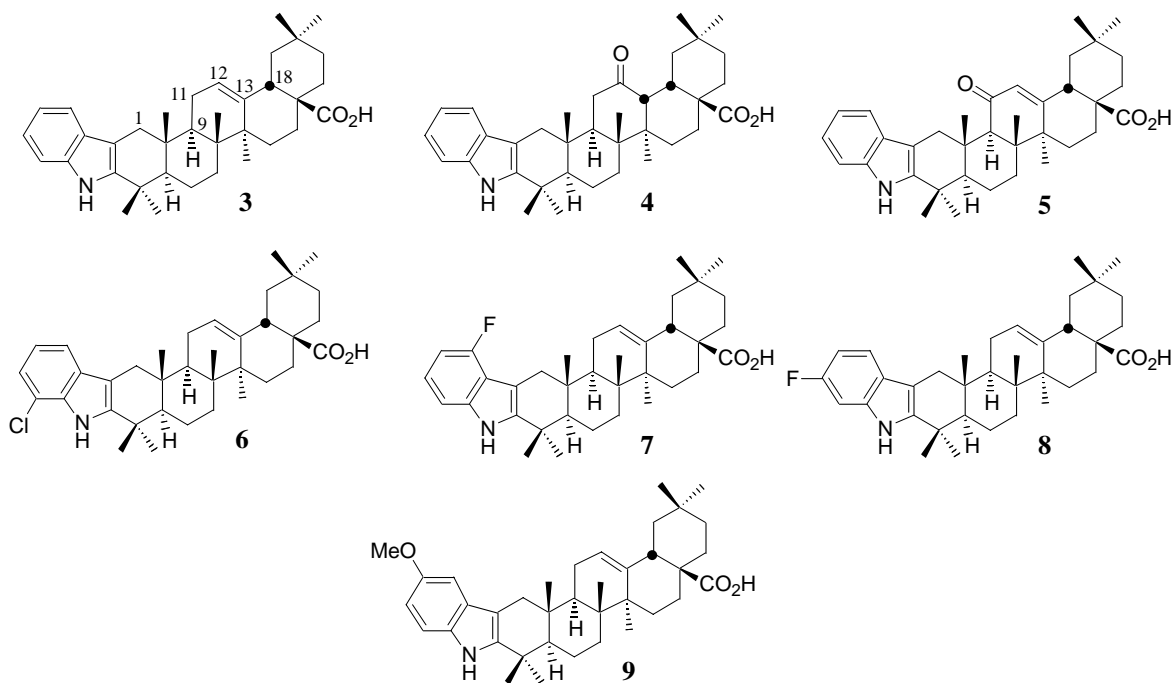
Triterpenoids are a diverse and ubiquitous group of C<sub>30</sub> pentacyclic compounds<sup>1</sup> that are derived biosynthetically from squalene cyclization.<sup>2</sup> Many triterpenoids display interesting biological and pharmacological profiles,<sup>3</sup> which include the selective inhibition of inducible nitric oxide synthase (iNOS)<sup>4-7</sup> and cyclooxygenase-2 (COX-2),<sup>5,6,8</sup> modulation of collagen synthesis,<sup>9-11</sup> inhibition of tumorigenesis,<sup>12,13</sup> and the ability to affect cell proliferation.<sup>14,15</sup>

As part of our efforts to synthesize and screen for biological activity novel derivatives of oleanolic (**1**) and ursolic acid (**2**),<sup>5,6,16-20</sup> we reported the ability of some 70 synthetic triterpenoids to affect cell proliferation in epithelial nonmalignant NRP.152 and malignant NRP.154 prostate cells.<sup>21</sup> These NRP.152 prostate cells demonstrate sensitivity to retinoids and 1 $\alpha$ ,25-dihydroxyvitamin D<sub>3</sub> and may be used for analysis of normal prostate growth and prostatic carcinogenesis.<sup>22,23</sup> Compounds that inhibit nonmalignant prostate cell proliferation mediated by the induction of TGF- $\beta$  demonstrate potential as chemopreventive agents for prostate (and breast) cancer.<sup>24</sup>



## Results and Discussion

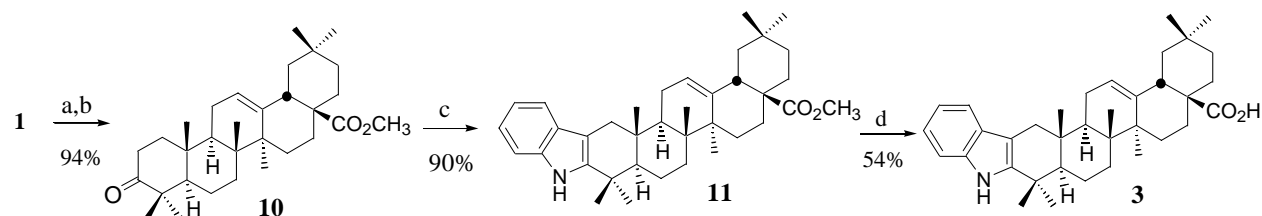
In the present paper we describe the synthesis of seven new indole-fused oleanolic acid derivatives, **3–9**, for evaluation in the NRP.252 cell assay. Fused heterocyclic derivatives of steroids and alkaloids are well documented,<sup>25</sup> and biologically active indole-fused examples are of particular interest.<sup>26</sup> Furthermore, several indole-fused steroids have been synthesized for electron-transfer studies.<sup>27-30</sup> In contrast, only one research group has described the synthesis of indole-fused triterpenoids.<sup>31,32</sup> Interestingly, a number of indole-fused diterpenes, such as the penitremes, are *Penicillium* fungal metabolites.<sup>33</sup> Our syntheses of the target compounds **3–9** (Figure 1) are based on the Fischer indole synthesis,<sup>34,35</sup> and are depicted in Schemes 1–4.



**Figure 1**

As we have previously described,<sup>16</sup> sequential diazomethane treatment and Jones oxidation of oleanolic acid (**1**) furnished keto ester **10** in 94% yield (Scheme 1). Fischer indolization of **10**

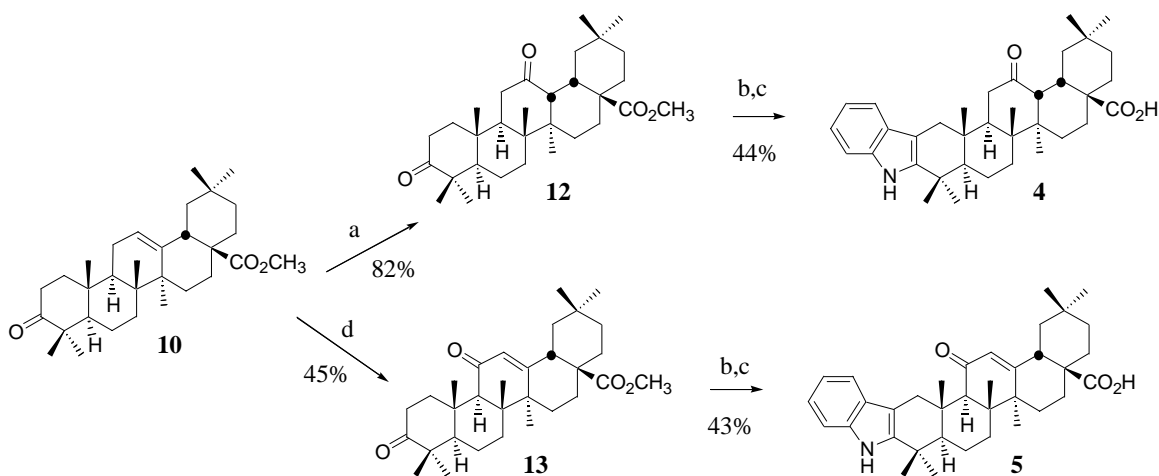
with phenylhydrazine in acetic acid gave the known<sup>32</sup> fused indole ester **11** in 90% yield. Cleavage of this hindered methyl ester with lithium iodide in DMF<sup>36</sup> afforded **3** in 54% yield. The corresponding C-3 ketone obtained from **1** also underwent Fischer indolization to give **3** in 61% yield, but a persistent yellow contaminant could not be removed from **3** by either crystallization or silica gel chromatography.



(a)  $\text{CH}_2\text{N}_2/\text{Et}_2\text{O}/\text{THF}$ ; (b) Jones oxidation; (c) phenylhydrazine/AcOH; (d) LiI/DMF.

### Scheme 1

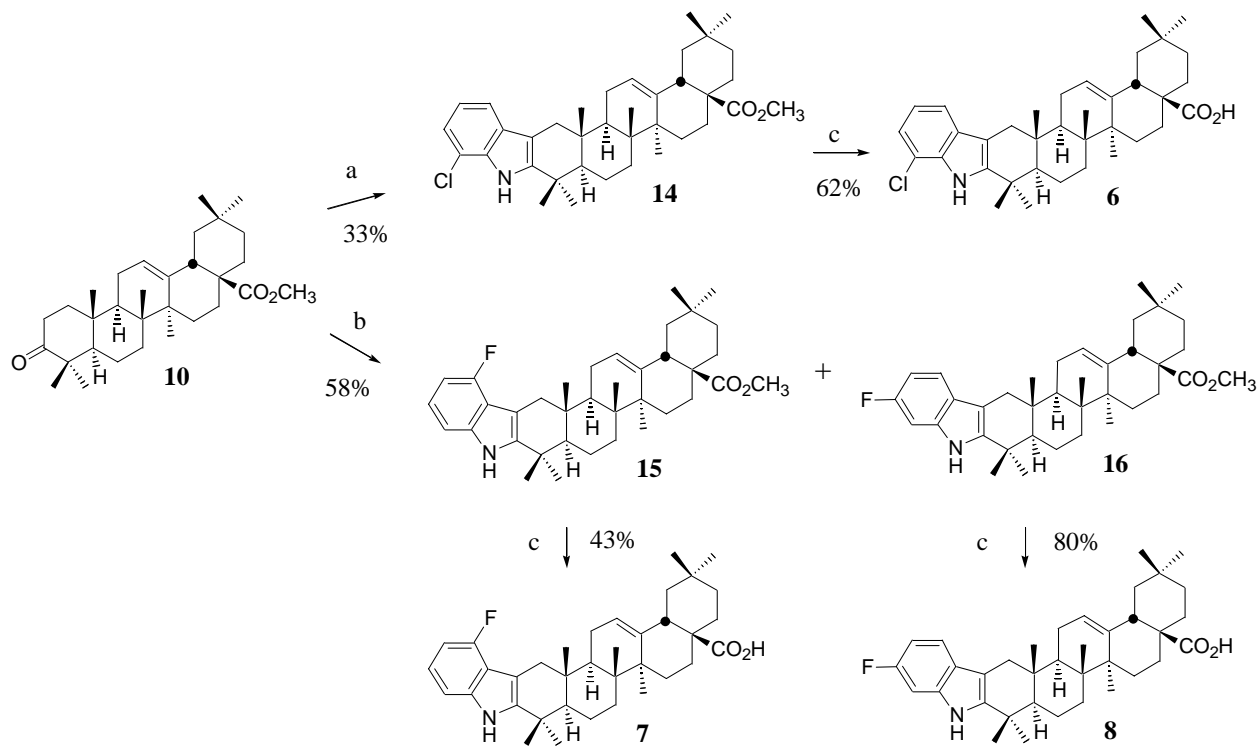
Due to the susceptibility of the indole ring in **3** to side reactions, particularly oxidation, modifications to the C-ring were performed prior to indolization. Thus, as shown in Scheme 2, and as we have previously described,<sup>20</sup> the synthesis of 3,12-diketone **12** was accomplished via an acid mediated epoxide rearrangement that occurred upon treatment of **10** with *m*-CPBA. Fischer indolization of **12** (74%) followed by ester cleavage (59%) gave the desired fused indole **4**. The highly hindered C-12 ketone in **12** remains unaffected under these Fischer indole reaction conditions.<sup>37</sup> Likewise, as we have reported,<sup>20</sup> allylic oxidation of **10** gave the known C-12,13 enone **13** (45% yield), which, upon Fischer indolization (79% yield) and ester cleavage (55% yield), afforded fused indole **5**.



(a) *m*CPBA/ $\text{CHCl}_3$ ; (b) phenylhydrazine/AcOH; (c) LiI/DMF; (d)  $\text{CrO}_3/\text{tBuOOH}$ .

### Scheme 2

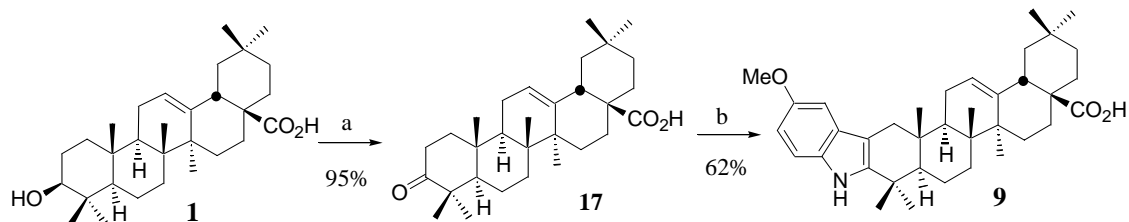
The indole ring substituted analogs **6–9** were synthesized by employing the appropriate substituted phenylhydrazine in the Fischer indolization (Scheme 3). Thus, treatment of **10** with 2-chlorophenylhydrazine<sup>38</sup> gave indole **14** that could be converted to indole **6** by ester cleavage. This indolization reaction was accompanied by 31% of uncyclized hydrazone. A sequence starting with 3-fluorophenylhydrazine yielded a mixture of indoles **15** and **16**, which were separated by sequential column and preparative silica gel chromatography in a 2:1 ratio, respectively. Cleavage of the methyl esters afforded **7** and **8**.



(a) 2-chlorophenylhydrazine/AcOH; (b) 3-fluorophenylhydrazine/AcOH; (c) LiI/DMF.

### Scheme 3

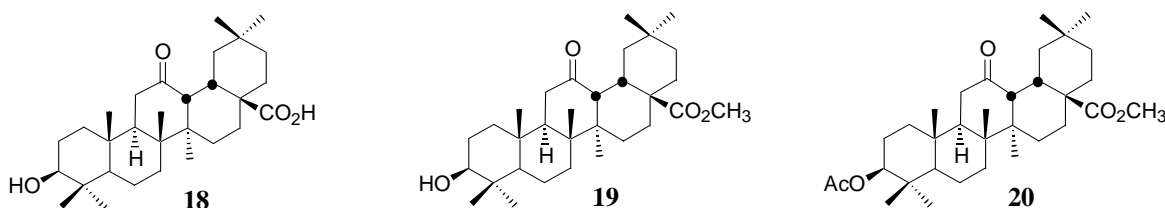
Finally, the 5-methoxyindole derivative **9** was synthesized directly from 3-keto acid **17** by Fischer indolization in 62% yield. The known keto acid **17** was prepared from oleanolic acid (**1**) by Jones oxidation (95% yield) as previously described.<sup>16</sup> Interestingly, the corresponding methyl ester analog that was prepared by indolization of **10** decomposed under the lithium iodide ester cleavage conditions.



(a) Jones oxidation; (b) 4-methoxyphenylhydrazine/AcOH.

#### Scheme 4

Unfortunately, attempts to effect indolization of C-12 ring C ketone derivatives were unsuccessful, presumably due to the hindered nature of this position. For example, we could not prepare the phenylhydrazone of ketones **18** and **19**, or effect indolization of ketones **19** and **20** with 2-iodoaniline using the palladium-annulation method of Chen *et al.*<sup>39</sup>



Oleanolic acid (**1**), indoles **3-9**, and **10**, **12**, and **13** were screened *in vitro* for their ability to inhibit proliferation of premalignant, non-tumorigenic prostate cells. Of the compounds prepared in the present study, only **4** and **5** showed some activity ( $IC_{50} < 5 \mu M$ ). All of the others were essentially inactive in this assay ( $> 5 \mu M$ ). For comparison, TGF- $\beta$  has  $IC_{50} = 0.000014 \mu M$ .<sup>40</sup> Therefore, in view of the disappointing activity in this assay of this series of fused-indole oleananes, we are not currently pursuing the study of additional examples of indole-fused triterpenoids.

## Experimental Section

**General Procedures.** Flash column chromatography was done with Select Scientific silica gel (230–400 mesh).  $^1H$  (300 MHz) and  $^{13}C$  (75 MHz) NMR spectra were recorded on a Varian XL-300 spectrometer in  $CDCl_3$  solvent; chemical shifts are reported with reference to the  $\delta$  7.27 signal of  $CHCl_3$  ( $^1H$  NMR) and  $\delta$  77.23 signal of  $CDCl_3$  ( $^{13}C$  NMR) as an internal standard.

### General procedure for Fischer indolization

A mixture of ketone **10** (89.3 mg, 0.191 mmol), phenylhydrazine (0.02 mL,  $d = 1.1$ , 1.05 eq), and glacial acetic acid (2 mL) was heated at reflux under  $N_2$  for 30 min. During this period the color changed from colorless to bright yellow. The reaction mixture was pipetted into distilled

water (50 mL) and extracted with ether (4 x 20 mL). The combined ether extracts were washed with 5% aqueous NaOH (2 x 20 mL) and brine (2 x 20 mL), dried (Na<sub>2</sub>SO<sub>4</sub>), and concentrated in vacuo to afford a yellow solid. Flash chromatography over silica gel and elution with hexane-ethyl acetate afforded indole **11** (92.4 mg, 90%) as an amorphous pale yellow solid. The synthesis of indole **9** from ketone **17** was worked up by simply pouring into water, extracting with ethyl acetate, and processing in the usual way to give an amorphous product after flash chromatography. Indoles **9**, **11**, and **14–16** were all amorphous solids and were directly converted into the corresponding acids as described below.

### General procedure for ester cleavage

A mixture of indole ester (0.09 mmol) and lithium iodide (0.45 mmol) in DMF (1.5 mL) under N<sub>2</sub> was heated at reflux for 15 h. The mixture was allowed to cool, treated with water (20 mL) and 10% aqueous hydrochloric acid (5 mL), and extracted with dichloromethane (3 x 20 mL). The organic layer was washed with water, dried (MgSO<sub>4</sub>), and concentrated in vacuo to afford the crude acid. Purification was effected by preparative TLC (hexane/ethyl acetate, 4:1) to give **3–8** as amorphous solids, for which melting points could not be obtained. The amounts of compounds, which were needed for biological screening, were insufficient for crystallization. Spectra data of **3–9** are tabulated in Tables 1–3.

**Table 1.** High-Resolution Mass Spectral Data of **3–6, 9** [*m/z*]

	Compound	M <sup>+</sup> , Calculated	M <sup>+</sup> , Observed
<b>3</b>	C <sub>36</sub> H <sub>49</sub> NO <sub>2</sub>	527.3763	527.3751
<b>4</b>	C <sub>36</sub> H <sub>49</sub> NO <sub>3</sub>	543.3712	543.3672
<b>5</b>	C <sub>36</sub> H <sub>47</sub> NO <sub>3</sub>	541.3556	541.3582
<b>6</b>	C <sub>36</sub> H <sub>48</sub> NO <sub>2</sub> Cl	561.3374	561.3385
<b>9</b>	C <sub>37</sub> H <sub>51</sub> NO <sub>3</sub>	557.3869	557.3884

**Table 2.** <sup>1</sup>H NMR data of **3–9**: δ [ppm]; coupling constants *J* [Hz]

	Me	H-1	H-12	H-18	NH	Aromatic	Other
<b>3</b>	0.85, 0.90, 0.94 (2), 1.15, 1.17, 1.27	2.75; 15.3 2.18; 15.9	5.38 1.15	2.87; 9.9	7.70	7.41; 6.9 7.28; 7.5 7.07 (2)	-
<b>4</b>	0.87, 0.92 1.00 (2), 1.07, 1.15, 1.28	2.69; 14.7 2.14; 14.7		2.80	7.75	7.40; 7.5 7.27; 7.2 7.08; (2)	2.49; 5.1, 17.1 2.33; 13.1 6.5 (H-11) 2.75; 4.2 (H-13)
<b>5</b>	0.94, 0.95, 0.99, 1.13, 1.15, 1.26, 1.39	3.96; 15.6 2.25; 15.6	5.72	3.01	7.72	7.49; 7.2 7.27; 7.2 7.07 (2)	2.62 (H-9)
<b>6</b>	0.84, 0.90,	2.73; 15.0	5.37	2.86; 9.9	7.84	7.30; 7.5	

**Table 2.** Continued

	0.93 (2), 1.17, 1.19, 1.30	2.18; 15.0				7.09; 1.2, 7.8 6.97; 7.8, 7.8	
<b>7</b>	0.84, 0.90, 0.94, 0.96, 1.14, 1.17, 1.26	2.71; 14.7 2.16; 14.7	5.37	2.87; 13.5	7.71	7.29; 5.4, 8.7 6.96; 2.4, 9.9 6.80; 2.4, 9.0, 9.9	
<b>8</b>	0.84, 0.90, 0.94, 0.96, 1.15, 1.16, 1.26	2.98; 15.6 2.35; 15.6	5.37	2.86; 13.5	7.76	7.04; 8.1 6.98 5.1, 7.8 6.67; 7.5, 10.8	
<b>9</b>	0.84, 0.90, 0.94 (2), 1.14, 1.17, 1.25	2.70; 14.7 2.16; 15.3	5.37	2.86; 12.6	7.59	7.16; 8.4 6.87; 2.4 6.75; 8.7, 2.4	3.83 (OMe)

**Table 3.**  $^{13}\text{C}$  NMR Data of **3–9**:  $\delta$  [ppm]

	CO <sub>2</sub> H	Ketone	C=C	Aromatic
<b>3</b>	184.5		145.6 123.1	141.0, 136.3, 128.4, 121.2, 119.1, 118.2, 110.5, 107.1
<b>4</b>	184.7	211.8		140.7, 136.3, 128.2, 121.4, 119.3, 118.2, 110.6, 106.5
<b>5</b>	178.4	199.6	169.5	141.3, 137.6, 129.0, 120.9,
<b>6</b>	184.4		143.6	141.9, 133.4, 129.9, 120.6,
<b>7</b>	184.6		143.6 125.0	161.3, 158.1, 143.6, 141.3, 136.3, 136.1, 123.0, 118.7, 107.3, 107.0, 97.0
<b>8</b>	184.5		143.4 123.2	155.6, 140.8, 139.0, 138.9, 121.4, 116.9, 111.5, 106.6, 105.7, 104.6
<b>9</b>	184.3		143.6 123.1	154.0, 142.1, 131.4, 128.8, 111.2, 110.9, 107.0, 100.6

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