

# Antimicrobial resistance pattern of *Escherichia coli* strains isolated from poultry farmers and poultry slaughterers in Morocco

Abdellah Chaiba<sup>1,2</sup>,  
Fouzia Rhazi Filali<sup>1</sup>

1 Microbiology and Health Team, Department of Biology, Laboratory of Chemistry-Biology Applied to the Environment, Faculty of Sciences, Moulay Ismail University, Meknès, Morocco.

2 Centre Régional des Métiers de l'Éducation et de la Formation (CRMEF), Draa Tafilalt, Morocco.

## Abstract

**Aim:** The objective of this study is to characterize the antibiotic resistance pattern of *Escherichia coli* isolates from the fecal samples of poultry workers (farmers and slaughterers), and to study the possible dissemination of resistant *E. coli* from poultry to humans.

**Methods:** Sixty-four *E. coli* strains isolated from the fecal samples of poultry workers (33 from poultry farmers and 31 from poultry slaughterers) and 35 isolates from a control group workers were tested for antibiotic resistance by agar disk diffusion with 11 antimicrobial agents.

**Results:** Resistance of *E. coli* isolated from poultry workers to tetracycline, ampicillin and norfloxacin were significantly ( $p < 0.05$ ) higher than those isolated from the control group. All *E. coli* isolates were susceptible to cefotaxime, and most of them are susceptible to gentamicin, amikacin, ceftiofur and ertapenem. Multidrug resistance is alarmingly high in all groups, but was highest in poultry farmers isolates (84%) and poultry slaughterers isolates (80%). Approximately 25% of the *E. coli* isolates from poultry workers showed resistance to four or more antibiotics.

**Conclusion:** This study suggests that occupational exposure to antimicrobial-resistant *E. coli* through animal contact in the broiler chicken industry may be an important route of entry antimicrobial-resistant *E. coli* into the community.

## Contact information:

Abdellah Chaiba.

 [abchaiba@yahoo.fr](mailto:abchaiba@yahoo.fr)

## Keywords

*Escherichia Coli*; Poultry Workers; Antibiotic Resistance; Multidrug Resistance; Morocco

Received 16-6-2018; Accepted 8-7-2018

## Introduction

Antibiotic resistance has been emerged as one of the world's most important public health problems [1, 2]. Use of antibiotics in human and animal medicine, especially their misuse, has been associated with the selection and spread of antibiotic resistant strains in human beings and animals [3, 4]. This acquired resistance occurs not only in pathogenic bacteria but also in the endogenous flora of exposed individuals or populations [5]. Antibiotics are used in animals as well as in humans for both prevention and treatment of infections. In animal husbandry they are also used as growth-promoting agents mixed with feed. These agents are widely used in the poultry industry of Morocco. A recent survey conducted by El-Youbi *et al.* [6] reported that all private veterinarians in Eastern Morocco have found abusive practices of using avian antibiotics.

Fecal flora of poultry contains a relatively high proportion of resistant bacteria; this is due to the high antibiotic selection pressure on bacterial flora of these birds [7]. During the slaughtering process of poultry birds, there can be occurring of fecal contamination through the guts of these birds with multiresistant bacteria especially *E. coli* [8, 9]. The human intestinal tract is the ideal place where antibiotic resistant bacteria can transfer their resistance genes to the endogenous human flora [10].

Many studies have shown that intestinal tract of human volunteers were colonized with resistant *E. coli* originated from poultry [11, 12]. Other investigations have revealed that farm animals are a potential reservoir of antibiotic resistant bacteria [13, 14]. In contrast, some authors [15, 16, 17] have reported that the antibiotic resistance transfer between animals and humans was limited because animal strains colonizing the alimentary tract less readily than human ones. However, the actual impact of antibiotic resistance bacteria isolated from workers who are in frequent contact with animals is not well known.

This study aimed to investigate and compare the antibiotic resistance profiles of *E. coli* isolates from poultry workers of both farmers and slaughterers and a control group of healthy individuals living in the same region.

## Materials and Methods

### Collection of fecal samples

Our study was carried out from January through April 2016, in Ouarzazate -Morocco. We collected fecal samples from healthy persons. A total of 33 samples from poultry farmers and 31 from poultry slaughterers. For the comparison, 35 fecal samples were collected from the same area from healthy persons who are practicing other professions except healthcare workers. To avoid cross-risk factors, we have excluded people who had been hospitalized within the last six months and people who received antibiotic treatment a month prior to be included in this study.

### Antibiotic susceptibility testing

The isolation and purification of *E. coli* strains was made on MacConkey agar and the identification was performed using API20E system (Biomérieux, Marcy l'Etoile, France). The isolates were screened for their antibiotic resistance to the following antibiotics (Marnes-La-Coquette, France): Amoxicillin-clavulanic acid (AMC ) 20+10 µg; cefoxitin FOX 30 µg; cefotaxime CTX 30 µg; tetracycline TE 30 UI; gentamicin (CN) 15 µg; amikacin AK 30 µg; ertapenem (ETP) 10 µg; cephalothin (KF) 30 µg; ampicillin (AM) 10 µg; trimethoprim/sulphamethoxazole (SXT) 1.25/23.75 µg and norfloxacin (NOR) 5 µg. The antibiogram was performed by the agar diffusion method following the guidelines of the Antibiogram Committee of the French Microbiology Society (CA/SFM, 2014) [18]. Quality control was carried out using *E. coli* strain ATCC 25922. For each individual samples, one *E. coli* isolate was

selected randomly for the purpose of testing its antibiotic susceptibility.

### Statistical analysis

The antimicrobial susceptibility data are expressed as percentages or frequency of the human isolates. The  $\chi^2$  (khi-deux) test was used to estimate overall difference between the percentages of resistance between *E. coli* isolates from different groups. In all cases,  $p < 0.05$  was regarded as statistically significant.

## Results

A total of 99 fecal samples were examined. Response rates to participate in the study was in poultry farmers and poultry slaughterers were 73.3% and 68.8%, respectively. Antibiotic resistance rates among *E. coli* isolates found in each group were shown in **Table 1**. Of the three examined population groups, the highest rate of resistance to almost all tested drugs was detected infecal samples of poultry farmers, followed by those from poultry slaughterers and much lower in the control group, respectively. However, this high rate of resistance was only significant for tetracycline, ampicillin and norfloxacin ( $p < 0.05$ ), and most of the tested isolates showed high antimicrobial resistant to tetracycline .

All *E. coli* isolates were susceptible to CTX, and most of them are susceptible to CN, AK, FOX and ETP. However, one isolate (3%) from poultry workers showed resistance to cefoxitin and ertapenem, and one isolate (3%) from poultry farmers were additionally resistant to gentamicin and amikacin. The percentage of multiple drug resistance patterns in *E. coli* isolates of each group is shown in **Figure 1**.

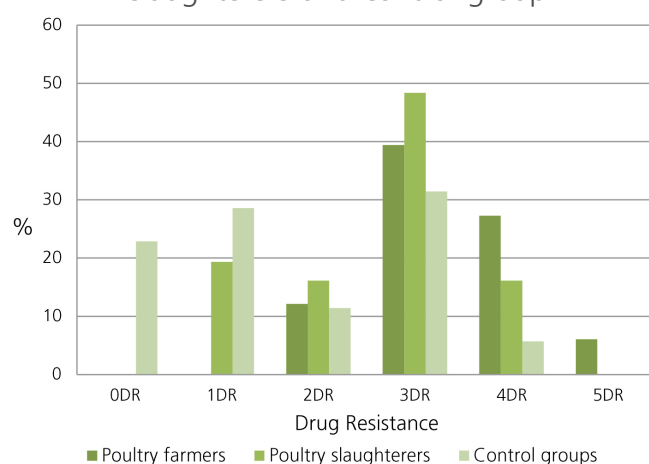
Multidrug resistance (MDR) was defined in this study as resistance to at least one agent out three or more antimicrobial classes [19]. The highest rate of multidrug resistance (72.7%) was found in poul-

**Table 1.** Comparison of antibiotic resistance rates of *E. coli* isolates from poultry farmers, poultry slaughterers and control group.

Antimicrobial agents	Antibiotic resistance of isolates		
	%		
	Poultry farmers n = 33	Poultry slaughterers n = 31	Control groups n = 35
Amoxicillin-clavulanic acid (AMC 20+10 µg)	42.42 <sup>a*</sup>	38.7 <sup>a</sup>	31.42 <sup>a</sup>
Cefoxitin (FOX 30 µg)	3.03 <sup>a</sup>	3.22 <sup>a</sup>	0 <sup>a</sup>
Cefotaxime (CTX 30 µg)	0 <sup>a</sup>	0 <sup>a</sup>	0 <sup>a</sup>
Tetracycline (TE 30 UI)	93.93 <sup>a</sup>	93.54 <sup>a</sup>	71.42
Gentamicin (CN 15 µg)	3.03 <sup>a</sup>	0 <sup>a</sup>	0 <sup>a</sup>
Amikacin (AK 30 µg)	3.03 <sup>a</sup>	0 <sup>a</sup>	0 <sup>a</sup>
Ertapenem (ETP 10 µg)	3.03 <sup>a</sup>	3.22 <sup>a</sup>	0 <sup>a</sup>
Cephalothin (KF 30 µg)	36.36 <sup>a</sup>	25.80 <sup>a</sup>	20 <sup>a</sup>
Ampicillin (AM 10 µg)	45.45 <sup>a</sup>	38 <sup>a</sup>	14.28
Trimethoprim/sulphamethoxazole (SXT)	39.39 <sup>a</sup>	32.25 <sup>a</sup>	25.71 <sup>a</sup>
Norfloxacin (NOR 5 µg)	27.27 <sup>a</sup>	25.80 <sup>a</sup>	5.71

\*: Rates are not significant (P > 0.05).

**Figure 1:** Antibiotic multiresistance patterns of *E. coli* strain in poultry farmers, poultry slaughterers and control group.



try farmers isolates, followed by those from poultry slaughterers (64.5%) and significantly lower in the control group (37.1%). Approximately 25% of the isolates of poultry workers harboured *E. coli* strains resistant to four or more antibiotics.

The antibiotic resistance pattern most frequently observed was resistance to TE/AM/SXT (12.1%) in the poultry farmers and (9.7%) in the poultry slaughterers. The highest number of resistance antibiotic markers was five (TE/AM/AK/CN/NOR) and (TE/AM/SXT/AMC/NOR), in two *E. coli* isolates from poultry farmers, respectively.

## Discussion

To the best of our knowledge, this is the first based study in Morocco investigated the prevalence of antimicrobial resistant *E. coli* from poultry workers. Our data showed that resistance rates remained significantly higher in *E. coli* isolated from poultry workers than those from control groups. The results presented here confirm similar studies from other countries showing that poultry farmers and poultry slaughterers workers were at higher risk for colonization with antimicrobial-resistant *E. coli* [10].

The presence and frequency of tetracycline resistant in *E. coli* in this study agree with findings of other studies on antibiotic resistance in *E. coli* [14, 15]. Tetracycline is a commonly used as first line antibiotic for many domestic animals and as a growth promoter or as an infection control agent, and is often used before the antibiotic resistance profile of a pathogen has been determined [20, 21].

In Morocco, the tetracycline is one of the most prescribed antibiotic in avian medicine [6]. In addition, other studies by us [22] reported that resistance in *E. coli* strains of poultry meat is higher to tetracycline (80%) than to trimethoprim/sulphamethoxazole (33.3%), nalidixic acid (26.6%) and amoxicillin (20%). Lower resistance rates were found by EL Al-

laoui *et al.* (5 to 12.5%) for ertapenem, aztreonam and gentamicin [23]. These results suggest that the extent of antibiotic resistance is associated with the extensive of its usage. Bacterial resistance to tetracycline is plasmid mediated with a wide variety of genetic determinants[22].

According to Van den Bogaard [24], dissemination of resistant bacteria and/or their resistance plasmids from broilers to poultry workers is the most likely explanation for the high antimicrobial resistance rates observed among fecal *E. coli* of the farmers and slaughterers.

The major factor for selecting antimicrobial resistance in bacteria is the misuse of antibiotic, in addition to crowding and poor sanitation in poultry farms. These three factors are typically related to intensive poultry farming, and explain the high prevalence of resistance in fecal *E. coli* of poultry workers in this and other studies [1].

In this study, carriage of multidrug-resistant *E. coli* was found significantly more common in poultry workers. Such high rate of multidrug resistance may apparently be occurred due to indiscriminate usage of antimicrobial agents [1]. During slaughtering process multidrug resistant *E. coli* strains can be transferred often from the gut of poultry carcasses and contaminate its meat [8,24].

The findings of our study confirm the effect of exposure to antibiotic resistance bacteria at the work place. This can be also associated with transfer of multiple antimicrobial resistance to commonly used drugs in the poultry farming. Bongers *et al.* [25] noted a significantly higher rates of *E. coli* resistant isolates to oxytetracycline and ampicillin in poultry industry workers than those not working with birds [26].

## Conclusion

The data obtained in this study indicate that occupational exposure to antimicrobial-resistant *E. coli* from animal contact in the broiler chicken

industry may be an important route of entry of antimicrobial-resistant *E. coli* into the community. We recommend to establish a strategy which can slow down the spread of antibiotic resistant bacterial strains by applying different preventive measures, such as control of antibiotic use, training and education of workers to practice safety measurements.

## References

1. Van den Bogaard AE, Stobberingh EE. Antibiotic usage in animals : impact on bacterial resistance and public health. *Drugs* 1999 ; 58(4): 589-607.
2. Witte W. Medical consequences of antibiotic use in agriculture. *Science* 1998; 279: 996-997.
3. Berge AC, Moore D A, Sicho WM. Field trial evaluating the influence of prophylactic and therapeutic antimicrobial administration on antimicrobial resistance of fecal *Escherichia coli* in dairy calves. *Appl Environ Microbiol* 2006; 72 : 3872-3878.
4. Jensen V F, Jakobsen L, Emborg HP, Seyfarth AM, Hammerum AM. Correlation between apramycin and gentamicin use in pigs and an increasing reservoir of gentamicin-resistant *Escherichia coli*. *J Antimicrob Chemother* 2006; 58 : 101-107.
5. Cho SH, Lim YS, Kang YH. Comparison of antimicrobial resistance in *Escherichia coli* strains isolated from healthy poultry and swine farm workers using antibiotics in Korea. *Osong Pub Health Res Persp* 2012; 3(3):151-155.
6. El-Youbi M., Belbachir C, Monir A; Saalaoui E. Antibiotics in broiler: exhaustive survey among private veterinarians in eastern Morocco. *Moroccan J Biology* 2016; 13: 60-68.
7. Van den Bogaard AE. Antimicrobial resistance - relation to human and animal exposure to antibiotics. *J Antimicrob Chemother* 1997; 40:453-454.
8. Caudry SD, Stanisich VA. Incidence of antibiotic resistant *Escherichia coli* associated with frozen chicken carcasses and characterization of conjugative R-plasmids derived from such strains. *Antimicrob Agents Chemother* 1979; 16 :701-709.
9. Nazer AH. Transmissible drug resistance in *Escherichia coli* isolated from poultry and their carcasses in Iran. *Cornell Vet* 1980; 70 :365-371.
10. Van den Bogaard AE, London N, Driessen C, Stobberingh EE. Antibiotic resistance of faecal *Escherichia coli* in poultry, poultry farmers and poultry slaughterers. *J Antimicrob Chemother* 2001; 47(6):763 -71.
11. Graham JP, Boland JJ, Silbergeld E. Growth promoting antibiotics in food animal production. *Public Health Rep* 2007; 122:79-87.
12. Teuber M. Veterinary use and antibiotic resistance. *Curr Opin Microbiol* 2001; 4(5):493 - 499.
13. Cooke EM, Breaden AL, Shooter RA, O'Farrell SM. Antibiotic sensitivity of *Escherichia coli* isolated from animals, food, hospital patients, and normal people. *Lancet* ii 1971; 3; 2(7714): 8-10.
14. Levy SB, FitzGerald GB, Macone AB. Spread of antibiotic-resistant plasmids from chicken to chicken and from chicken to man. *Nature* 1976; 260:40-2.
15. Caya F, Fairbrother JM, Lessard L, Quessy S. Characterization of the risk to human health of pathogenic *Escherichia coli* isolates from chicken carcasses. *J Food Protect* 1999; 62:741- 6.
16. Kariuki S, Gilks CF, Kimari J, Muyodi J, Waiyaki P, Hart CA. Plasmid diversity of multi-drug resistant *Escherichia coli* isolated from children with diarrhoea in a poultry-farming area in Kenya. *Ann Trop Med Parasitol* 1997; 91: 87- 94.
17. Nijsten R, London N, Van den Bogaard A, Stobberingh E. In-vivo transfer of resistance plasmids in rat, human or pig derived intestinal flora using a rat model. *J Antimicrob Chemother* 1995; 36: 975-985.
18. Committee of the susceptibility following to the French Society of Microbiology. (CA/SFM) 2014. Recommendations 2014. Retrieved on April 23, 2016 from SFM Website: <http://www.sfm-microbiologie.org/>
19. Magiorakos AP, A. Srinivasan, Carey RB, Carmeli Y, Falagas ME, Giske CG, Harbarth S, Hindler JF, Kahlmeter G, Olsson-Liljequist B, Paterson DL, Rice LB, Stelling J, Struelens MJ, Vatopoulos A, Weber JT, Monnet DL. Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. *Clin Microbiol Infect* 2012; 18: 268-281.
20. Chopra I. New developments in tetracycline antibiotics: glycylicyclines and tetracycline efflux pump inhibitors. *Drug Resist Updat* 2002; 5(3-4):119-125.
21. Levy SB. Antibiotic resistance: consequences of inaction. *Clin Infect Dis* 2001; 33(Suppl. 3): 124-129.
22. Chaiba A (2011) Impact des pratiques de production de poulet de chair à Meknès sur la qualité bactériologique, l'antibiorésistances et les résidus d'antibiotiques dans les produits aviaires finis, Thèse de Doctorat National, Université Moulay Ismail, Faculté des Sciences de Meknès, Maroc.
23. El Allaoui A , Rhazi Filali F, Oumokhtar B. Prevalence and Antibiogram Study of *Escherichia coli* and *Staphylococcus aureus* in Turkey Meat in Morocco. *Pharm Anal Acta* 2013; 4 (9):4-9.



24. Turtura GC, Massa S, Chazvinizadeh H. Antibiotic resistance among coliform bacteria isolated from carcasses of commercially slaughtered chickens, Intern J Food Microbiol 1990; 11:351-354.
25. Bongers JH, Franssen F, Elbers AR, Tielen MJ. Antimicrobial resistance of Escherichia coli isolates from the faecal flora of veterinarians with different professional specialties. Vet Quarterly 1995; 17:146 -149.
26. Al-Ghamdi MS, El-Morsy F, Al-Mustafa ZH, Al-Ramadhan M, Hanif M. Antibiotic resistance of Escherichia coli isolated from poultry workers, patients and chicken in the eastern province of Saudi Arabia. Trop Med Int Health 1999; (4):278-283.

**Publish in The International  
Arabic Journal of Antimicrobial Agents**

The Journal is an open access peer-reviewed journal that publishes scientific papers about all aspects of antimicrobials. The journal will publish original research articles, reviews, brief reports and case reports dealing with basic and clinical antibacterial agents, antiviral, antiprotozoals, antituberculous, antifungal and antihelminthes agents. All manuscripts must be prepared in English, and are subject to a rigorous and fair peer-review process. Accepted papers will immediately appear online. The journal aims to advance the knowledge, attitude and the research of chemotherapy in the Arabic world in cooperation with international, national scientific and public societies as well as research centers with similar aims and objectives.