

Himmelfarb Health Sciences Library, The George Washington University Health Sciences Research Commons

Pediatrics Faculty Publications

Pediatrics

1-1-2018

Regional microstructural organization of the cerebral cortex is affected by preterm birth.

Marine Bouyssi-Kobar George Washington University

Marie Brossard-Racine

Marni Jacobs George Washington University

Jonathan Murnick George Washington University

Taeun Chang George Washington University

See next page for additional authors

Follow this and additional works at: https://hsrc.himmelfarb.gwu.edu/smhs_peds_facpubs

Part of the Neurology Commons, and the Pediatrics Commons



Bouyssi-Kobar, M., Brossard-Racine, M., Jacobs, M., Murnick, J., Chang, T., & Limperopoulos, C. (2018). Regional microstructural organization of the cerebral cortex is affected by preterm birth.. *NeuroImage Clinical*, 18 (). http://dx.doi.org/10.1016/j.nicl.2018.03.020

This Journal Article is brought to you for free and open access by the Pediatrics at Health Sciences Research Commons. It has been accepted for inclusion in Pediatrics Faculty Publications by an authorized administrator of Health Sciences Research Commons. For more information, please contact hsrc@gwu.edu.

therine Limperopoulos	

FISEVIER

Contents lists available at ScienceDirect

NeuroImage: Clinical

journal homepage: www.elsevier.com/locate/ynicl



Regional microstructural organization of the cerebral cortex is affected by preterm birth



Marine Bouyssi-Kobar^{a,b}, Marie Brossard-Racine^c, Marni Jacobs^d, Jonathan Murnick^a, Taeun Chang^e, Catherine Limperopoulos^{a,*}

- a The Developing Brain Research Laboratory, Department of Diagnostic Imaging and Radiology, Children's National Health System, Washington, DC 20010, USA
- ^b Institute for Biomedical Sciences, George Washington University, Washington, DC 20037, USA
- ^c Department of Pediatrics Neurology, McGill University Health Center, Montreal, QC H4A3J1, Canada
- d Division of Biostatistics and Study Methodology, Children's Research Institute, Children's National Health System, Washington, DC 20010, USA
- ^e Department of Neurology, Children's National Health System, Washington, DC 20010, USA

ARTICLE INFO

Keywords: Prematurity Cerebral cortex Microstructural organization Diffusion tension imaging

ABSTRACT

Objectives: To compare regional cerebral cortical microstructural organization between preterm infants at termequivalent age (TEA) and healthy full-term newborns, and to examine the impact of clinical risk factors on cerebral cortical micro-organization in the preterm cohort.

Study design: We prospectively enrolled very preterm infants (gestational age (GA) at birth < 32 weeks; birthweight < 1500 g) and healthy full-term controls. Using non-invasive 3T diffusion tensor imaging (DTI) metrics, we quantified regional micro-organization in ten cerebral cortical areas: medial/dorsolateral prefrontal cortex, anterior/posterior cingulate cortex, insula, posterior parietal cortex, motor/somatosensory/auditory/visual cortex. ANCOVA analyses were performed controlling for sex and postmenstrual age at MRI.

Results: We studied 91 preterm infants at TEA and 69 full-term controls. Preterm infants demonstrated significantly higher diffusivity in the prefrontal, parietal, motor, somatosensory, and visual cortices suggesting delayed maturation of these cortical areas. Additionally, postnatal hydrocortisone treatment was related to accelerated microstructural organization in the prefrontal and somatosensory cortices.

Conclusions: Preterm birth alters regional microstructural organization of the cerebral cortex in both neurocognitive brain regions and areas with primary sensory/motor functions. We also report for the first time a potential protective effect of postnatal hydrocortisone administration on cerebral cortical development in preterm infants.

1. Introduction

Infants born very preterm (before 32 weeks gestational age (GA)) are three times more likely than full-term born infants to develop psychiatric disorders (Johnson and Marlow, 2011), and are at higher risk for a wide range of socio-cognitive impairments (Blencowe et al., 2013). The neural substrates of socio-cognitive functioning are composed of cortical and subcortical interconnected neurons that co-activate for the purpose of mediating specific cognitive outputs and are organized in circuits called neurocognitive networks (Mesulam, 2012). The functional network foundations are already present during the third trimester of gestation (Doria et al., 2010). However, these neural networks are immature at birth, and can already exhibit early signs of

impairment in infants born preterm (Ball et al., 2015; C. D. Smyser et al., 2015a). In adults who were born very preterm, alterations of these neurocognitive networks have recently been demonstrated (White et al., 2014). Interestingly, little is known about the early microstructural integrity of the cerebral cortical regions involved in sociocognitive processing.

The cytoarchitecture of the cerebral cortex undergoes major transformations during the perinatal period as a result of critical developmental processes in neural migration, spinogenesis, synaptogenesis, and gyrification (Budday et al., 2015). Quantitative magnetic resonance imaging (MRI) techniques such as diffusion tensor imaging (DTI) allow for *in vivo* characterization of the microstructural cerebral organization (Mori and Zhang, 2006). Although DTI has been traditionally used to

E-mail addresses: marine@gwu.edu (M. Bouyssi-Kobar), marie.brossardracine@mcgill.ca (M. Brossard-Racine), MJacobs@childrensnational.org (M. Jacobs), JMurnick@childrensnational.org (J. Murnick), TChang@childrensnational.org (T. Chang), CLimpero@childrensnational.org (C. Limperopoulos).

^{*} Corresponding author at: Developing Brain Research Laboratory, Departments of Diagnostic Imaging and Radiology, Children's National Health System, 111 Michigan Ave NW, Washington, DC 20010, USA.

characterize white matter development in the neonatal population (Qiu et al., 2015), it also offers important insights into the maturation of the cerebral cortex (Hüppi and Dubois, 2006). Specifically, DTI probes the microstructural organization of brain tissue by measuring the threedimensional spatial diffusion of water molecules (Basser et al., 1994). The properties of water diffusion are described through several DTIbased metrics; the most commonly used are mean diffusivity (MD), which provides the average water displacement, and fractional anisotropy (FA), which measures the directionality or anisotropy of the diffusion (Mori and Zhang, 2006). Using DTI metrics, it is possible to characterize the complex architectural changes of the cerebral cortex associated with maturational processes during critical periods of development. It has been shown that between 25 and 40 weeks GA, MD decreases as the water content of the cortex is reduced due to increasing cellular density and complexity (McKinstry et al., 2002). In contrast, FA increases during early GA (up to 28 weeks GA) reflecting the radial organization of the cortex (Gupta et al., 2005; Trivedi et al., 2009). Thereafter, FA decreases due to progressive differentiation of radial glia, arrival of axons, and dendritic arborization (Aeby et al., 2012; Gupta et al., 2005; McKinstry et al., 2002; Yu et al., 2015).

Longitudinal DTI studies in preterm infants have quantified in vivo the neurobiological processes occurring during the ex-utero third trimester (Ball et al., 2013; deIpolyi et al., 2005; Eaton-Rosen et al., 2017; Kersbergen et al., 2014; Schneider et al., 2016; T. A. Smyser et al., 2015b; Wu et al., 2017a). The cerebral cortex matures as a function of advancing GA with a central to peripheral and posterior to anterior regional gradient, and cortical areas with primary sensory and motor functions develop before association areas (Raybaud et al., 2013). This spatiotemporal gradient of maturity translates into a different rate of MD and/or FA decrease by brain region (Kersbergen et al., 2014; Wu et al., 2017b). Thus, DTI metrics have been used as markers of cerebral cortical maturation: slower postnatal growth in preterm infants has been associated with delayed maturation of the cerebral cortex characterized by higher FA values (Vinall et al., 2013). However, only two studies have compared cortical microstructural organization between preterm infants at term-equivalent age (TEA) and full-term controls (Ball et al., 2013; T. A. Smyser et al., 2015b). Although these studies have provided remarkable information about cerebral cortical microorganization, none of the brain regions that play a crucial role in sociocognitive processing have been assessed to date. These key neurocognitive regions are known to be impaired in neurodevelopmental disorders and remain to be examined in the premature infants. Consequently, we investigated the microstructural integrity of brain areas known to be involved in neurocognitive networks: the dorsolateral prefrontal cortex (dlPFC), the posterior parietal cortex (PPC), the medial prefrontal cortex (mPFC), the posterior cingulate cortex (PCC), the insula, and anterior cingulate cortex (ACC). Additionally, to further validate our findings, we sought to characterize the cortical areas linked to sensory and motor functions. Our primary objective was to compare the DTI metrics within these cerebral cortical regions of interest (ROIs) between very preterm infants at TEA and healthy full-term controls. As a secondary objective, we examined clinical risk factors that are associated with cerebral cortical development within the preterm cohort.

2. Materials and methods

2.1. Participants

Preterm infants born before 32 weeks GA and weighing < 1500 g admitted to the level IV NICU at Children's National Health System (Washington DC) from June 2012 to February 2016 were enrolled in a prospective observational study on brain development (Bouyssi-Kobar et al., 2017). Exclusion criteria for this preterm cohort included: chromosomal anomalies, dysmorphic features, congenital brain malformations, central nervous system infection, and metabolic disorders. In the context of another prospective research study on perinatal brain

growth and development, healthy fetal/mother dyads with normal fetal sonography were enrolled as control participants (Bouyssi-Kobar et al., 2017). Exclusion criteria included evidence of dysmorphic features, chromosomal abnormalities, multiple gestations, congenital infections, maternal drug use, and maternal disease. All healthy newborns included in this study were born full-term (after 38 weeks GA) with uneventful deliveries and normal MRI brain scans. Both studies were approved by the Institutional Review Board of Children's National Health System and written informed consent was obtained from parents for all participants.

2.2. Clinical data collection

For all participants, demographic, prenatal, intrapartum, and neonatal information was collected including: maternal age at delivery, pregnancy complications, type of delivery, need for respiratory/cardiac resuscitation, birthweight, GA at birth, sex, ethnicity, and Apgar score at 1 and 5 min. In the preterm cohort, we also documented the presence of chorioamnionitis per placenta pathology records, and postnatal data reflecting infants' clinical status throughout their stay in the neonatal intensive care unit including: moderate to severe bronchopulmonary dysplasia (BPD) (Jobe and Bancalari, 2001), length of supplemental oxygen requirement, postnatal steroid treatment including length of treatment and type of steroid used (dexamethasone *versus* hydrocortisone), sepsis (confirmed by positive blood culture), necrotizing enterocolitis (NEC) diagnosis requiring bowel surgery, need for cardiac pressor support, and need for patent ductus arteriosus (PDA) surgical ligation.

2.3. MRI acquisitions

All participants underwent a brain MRI on a 3 T GE Healthcare Discovery MR750 scanner (Milwaukee, WI). Infants were scanned under natural sleep: after feeding, they were swaddled in warm blankets, provided with ear protection, and immobilized using a newborn vacuum pillow (Newmatic Medical, Caledonia, MI). A nurse was present for the duration of the MRI scan to monitor vital signs. Diffusion weighted images were obtained using a single shot echo-planar sequence: 27 non-collinear diffusion gradients with a b-value of $1000 \, \text{s/mm}^2$, three non-diffusion weighted (b0) volumes, echo/repetition time = $80/8000 \, \text{ms}$, voxel size = $1.56 \times 1.56 \times 3 \, \text{mm}$, acquisition time = $4:08 \, \text{min}$. An anatomical T2-weighted sequence (3D Cube) was also acquired: echo/repetition time = $65/2500 \, \text{ms}$, voxel size = $0.63 \times 1 \times 0.63 \, \text{mm}$, acquisition time = $3 \, \text{min} \, 20 \, \text{s}$.

2.4. Brain assessment

Clinical readings of all brain MRI studies were carried out by an experienced pediatric neuroradiologist. For the preterm cohort, brain injury severity was also classified into normal, mild, moderate, and severe categories (Kidokoro et al., 2013), and intraventricular hemorrhage (IVH) severity was defined according to Papile et al. (1978). In order to avoid the known cofounding factor of severe brain injury on cerebral cortical development (Andiman et al., 2010; Sizonenko et al., 2007; T. A. Smyser et al., 2015b), we excluded infants with severe brain injury from further DTI analyses. Specifically, preterm infants with IVH grade III, periventricular hemorrhagic infarction, cystic periventricular leukomalacia, or those falling into the severe category classification on the Kidokoro et al. (2013) scoring system. Thus, our preterm cohort included only infants without evidence of structural brain lesions and infants with mild to moderate parenchymal brain injury on conventional MRI.

2.5. DTI processing

DTI data were preprocessed using a previously validated neonatal

NeuroImage: Clinical 18 (2018) 871-880

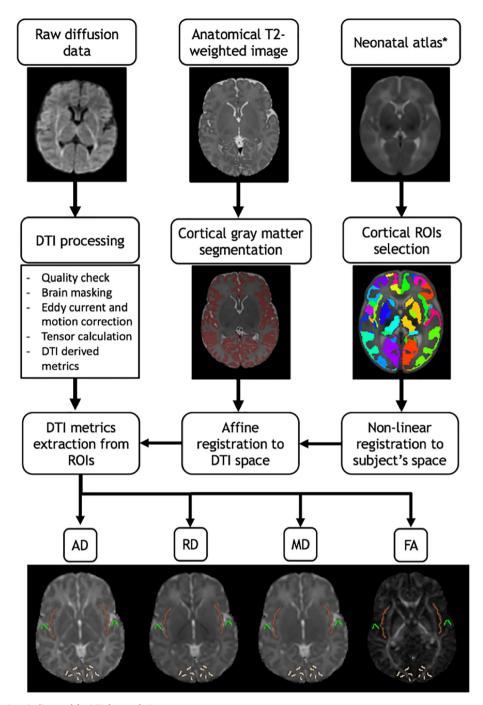


Fig. 1. Outline of the processing pipeline used for DTI data analysis. *Neonatal Atlas from Shi et al. (2011).

pipeline (Brossard-Racine et al., 2016; Evangelou et al., 2014) (Fig. 1). We first visually inspected all 30 DTI volumes that were acquired and discarded individual diffusion weighted volumes if motion artifacts and/or signal dropout were encountered. We then selected the best b0 volume, *i.e.*, motion free with the highest signal-to-noise ratio. DTI acquisitions with more than two third of diffusion gradients corrupted were excluded from subsequent analysis resulting in an average of 24.7 ± 2.5 gradient directions (range: 18-27). Processing of the DTI datasets was performed using the FMRIB software (Jenkinson et al., 2012). First, non-brain tissue was removed (Smith, 2002); then, eddy current induced distortions and subject motion were corrected using $eddy_correct$ from the FMRIB's diffusion toolbox, and gradient directions were rotated accordingly (Leemans and Jones, 2009). Finally, a

diffusion tensor model was calculated for each voxel using dtifit from the FMRIB's diffusion toolbox. The tensor diffusion model fits diffusion measurements into a three-dimensional ellipsoid whose axes represent the average diffusivity in each direction (Mori and Zhang, 2006). The length of the longest, middle, and shortest axes are the eigenvalues λ_1 , λ_2 , and λ_3 respectively (units: mm²/s), and the direction of the axes are the eigenvectors (Basser et al., 1994). Axial diffusivity (AD = λ_1) is the average diffusion along the principal direction of diffusion; radial diffusivity (RD) depicts the average diffusion in the plan orthogonal to the main diffusion direction and is the average of λ_2 and λ_3 ; MD is the overall amount of diffusion, which consists of the average of the three eigenvalues, and FA is an index related to the variance of the eigenvalues that reflects the overall directionality of diffusion which is scaled

between 0 (isotropic) and 1 (maximum anisotropy) (Pierpaoli and Basser, 1996).

2.6. Cerebral cortical regions of interest

We used the automatic anatomical labeling (AAL) parcellation (Tzourio-Mazoyer et al., 2002) mapped to neonatal MRI data by Shi et al. (2011) to retrieve our cerebral cortical regions of interest (ROIs) (Fig. 1). The ROIs examined included the dlPFC, the inferior parietal lobule as part of the PPC, the mPFC, the PCC, the insula, and the ACC, the precentral gyrus (motor cortex), postcentral gyrus (somatosensory cortex), calcarine cortex (visual cortex), and the superior transverse gyrus (auditory cortex). For these ten ROIs, we aimed to select only the central cerebral cortical voxels. Thus, within the template space we first selected the voxels with the highest probability of belonging to the cerebral cortex, i.e., those voxels that were located centrally within the cerebral cortex, and then we transferred these cortical ROIs to the participant's native space. The neonatal atlas was non-linearly registered to each participants' anatomical T2-weighted image using AFNI non-linear warping program (Cox, 1996). The anatomical T2-weighted image was subsequently aligned using affine registration to its corresponding non-diffusion weighted (b0) of the DTI dataset (Studholme et al., 1999). In order to remove voxel contaminated by white matter and cerebrospinal fluid, we automatically segmented the anatomical T2-weighted image using the automatic segmentation software DrawEM (Makropoulos et al., 2014). Participant's cortical gray matter segmentation and cortical parcellation from the neonatal atlas were transferred to the DTI dataset and their intersection defined our ROIs. Additionally, all results were visually inspected, and misplaced voxels were removed if necessary before extraction of the DTI metrics from the native DTI participant's space.

2.7. Statistical analysis

Statistical analyses were performed using SAS 9.3 (SAS Institute, Cary, NC). We first examined potential laterality differences using a paired t-test. No statistical differences between left/right sides were noted except for the calcarine cortex where the eigenvalues were significantly higher in the left side. Consequently, we analyzed left/right calcarine ROIs separately and we averaged the left/right DTI metrics for the remaining ROIs. The effect of group (preterm versus healthy full-term infants) on regional DTI metrics was assessed using ANCOVA analysis in which group, sex, and postmenstrual age (PMA) at MRI were the main effects. We also performed the same analysis adjusting for day of life at MRI, in order to measure the length of exposure to the extrauterine environment. Similarly, the relationship between clinical risk factors on DTI metrics was investigated using ANCOVA analyses controlling for sex, PMA at MRI, and day of life at MRI. Significance level was set at p < 0.05.

3. Results

3.1. Participants studied

We acquired DTI datasets in 125 preterm infants and 95 healthy full-term infants, of which 18 (14%) preterm infants were excluded due to the presence of severe brain injury. An additional 16 (13%) preterm infants and 26 (27%) full-term controls were discarded because of motion artifacts or poor quality. The DTI metrics of the remaining 160 participants were analyzed and their clinical characteristics are summarized in Table 1. Preterm infants underwent brain MRI scans at a mean PMA of 40.10 weeks whereas healthy full-term infants were scanned at a mean PMA of 41.39 weeks.

3.2. Regional DTI metrics

Compared to healthy full-term controls, preterm infants had significantly higher AD, RD, and MD in the prefrontal cortex (dlPFC, mPFC), the PPC, the motor cortex, the somatosensory cortex, and the visual cortex (Fig. 2, Table 2). Fractional anisotropy was significantly higher in preterm infants in the motor cortex only; in all other nine areas studied, FA was not significantly different between the two groups. Our findings did not change when the variable day of life was accounted for in the statistical model. These results suggest altered regional microstructural organization in very preterm infants by TEA compared to healthy full-term controls.

3.3. Clinical risk factors: Influence on microstructural cerebral cortical organization

Forty-seven preterm infants (52%) had a normal MRI, 30 (33%) had mild brain injury and 14 (15%) had moderate brain injury (Table 1). Parenchymal brain injury was not associated with DTI metrics (Table 3), and the brain injury severity (moderate versus mild brain injury) was not related to microstructural organization. Among the other risk factors examined within the preterm cohort, only postnatal steroid treatment was associated with altered microstructural organization in the prefrontal and somatosensory cortices (Table 3). Surprisingly, steroid treatment was associated with decreased diffusivity $(\beta < 0)$ in the prefrontal, motor, and somatosensory cortices, suggesting accelerated structural organization at the microscopic level following postnatal steroid administration. To further scrutinize this unexpected finding, for the preterm infants who received postnatal steroid treatment, we considered the length of steroid treatment, and the type of steroid administered. In our cohort, two pharmaceutical steroids were used: dexamethasone for chronic lung disease, and hydrocortisone for management of hypotension. We found that only hydrocortisone treatment was associated with increased microstructural organization (i.e., reduced diffusivity) in the dlPFC, mPFC, and somatosensory cortex (Table 3).

4. Discussion

This study investigated the microstructural organization of the cerebral cortex in very preterm infants at TEA and healthy full-term neonates using DTI metrics. We demonstrated delayed regional microstructural organization in the prefrontal, parietal, somatosensory, and visual cortices of very preterm infants at TEA compared to full-term controls. Specifically, the main components of the central executive network (dlPFC and PPC) and the default mode network (mPFC), exhibited impaired microarchitecture at TEA. Interestingly, postnatal hydrocortisone treatment was associated with enhanced frontal and somatosensory microstructural organization in the preterm cohort at TEA

The cytoarchitecture of the cerebral cortex undergoes dramatic changes during the third trimester of gestation which can be captured by DTI (Dean et al., 2013; Jespersen et al., 2013). DTI measures water diffusion properties which are strongly related to tissue micro-architectural organization (Mori and Zhang, 2006). In the cerebral cortex, several developmental processes influence water diffusion including arrival of afferent axons, dendritic arborization, decreased radial glial scaffolding, synaptic formation, and reduction in water content (Budday et al., 2015; McKinstry et al., 2002). During the third trimester of gestation, cerebral cortical diffusivity decreases due to an increase in cellular density and/or a reduction in water content; whereas, cerebral cortical anisotropy changes are related to loss of radial organization and dendritic arborization (Jespersen et al., 2013; McKinstry et al., 2002; Sizonenko et al., 2007). Given that our findings primarily showed a difference in diffusivity magnitude between preterm infants at TEA and full-term controls, we cannot exclude the possibility of diffusivity

Table 1 Clinical characteristics of the cohort (N = 160).

	Preterm infants, $n = 91$	Full-term control infants, $n = 69$	<i>p</i> -Value
Perinatal characteristics			
Birth gestational age, wk, mean ± SD	26.73 ± 2.62	39.64 ± 0.98	< 0.0001
Birthweight, g, mean ± SD	862 ± 298	3389 ± 367	< 0.0001
Small for gestational age, n (%)	14 (15)	5 (7)	0.12
Male, n (%)	35 (51)	44 (48)	0.76
Native American;	0; 19 (21); 11 (12)	2 (3); 7 (10); 24 (35)	< 0.0001
Hispanic; White	0; 55 (60); 6 (7)	6 (9); 28 (40); 2 (3)	
Asian; Black; Multiethnic, n (%)			
Vaginal delivery, n (%)	32 (35)	49 (71)	< 0.0001
Apgar score at 5 min, median [range]	7 [1–9]	9 [8–10]	< 0.001
Maternal age, y, mean ± SD	27.82 ± 6.19	29.78 ± 6.81	0.06
MRI characteristics			
Postmenstrual age at MRI, wk, mean ± SD	40.10 ± 1.35	41.39 ± 1.18	< 0.0001
Age at MRI, d, mean ± SD	93.68 ± 20.34	12.36 ± 5.52	< 0.0001
Head circumference at MRI, cm, mean ± SD	33.01 ± 1.92	35.65 ± 1.34	< 0.0001
Weight at MRI, g, mean ± SD	2740 ± 647	3596 ± 432	< 0.0001
Brain abnormalities classification ^a			
White matter signal abnormality, n (%)	18 (20)	na	na
Delayed myelination, n (%)	4 (4)	na	na
Thinning of the corpus callosum, n (%)	47 (52)	na	na
Dilation of the lateral ventricles, n (%)	37 (41)	na	na
Reduce white matter volume, n (%)	57 (63)	na	na
Increased extracerebral space, n (%)	26 (29)	na	na
Deep gray matter signal abnormality, n (%)	5 (5)	na	na
Reduce deep gray matter volume, n (%)	8 (9)	na	na
Cerebellar signal abnormality, n (%)	17 (19)	na	na
Reduce cerebellar volume, n (%)	40 (44)	na	na
Brain injury category, n (%) normal; mild; moderate	47 (52); 30 (33); 14 (15)	na	na

na: not applicable.

parameters measuring only a reduction in cerebral cortical water. Nonetheless, higher diffusivity in preterm infants also reflects maturational delays since brain water content decreases with increasing gestational age during normal development (Dobbing and Sands, 1973; Kreis et al., 1993). We postulate that increased diffusivity in preterm infants at TEA reveals both a reduction in water content as well as decreased dendritic and synaptic density. Another hypothesis would be that DTI metrics differences between preterm and full-term infants result from acute injury to the developing cortex following preterm birth. Since we excluded severe brain injury by design, and mild to moderate parenchymal brain injury were not associated with DTI metrics, our findings likely reflect regional dysmaturation of the cerebral cortex in preterm infants at TEA.

Prior to the current investigation, only two studies with small sample size had compared cerebral cortical DTI metrics between very preterm infants at TEA and full-term neonates (Ball et al., 2013; T. A. Smyser et al., 2015b). In one study, mean diffusivity was increased in 22 preterm infants without brain injury compared to 12 full-term controls, however these results did not reach statistical significance (T. A. Smyser et al., 2015b). Another study, which included a slightly higher number of preterm infants (n = 37), reported significantly increased MD in the occipital, parietal, temporal, and frontal cortices of preterm infants compared to 10 full-term controls (Ball et al., 2013). In comparison, the present investigation provided information in a large cohort of preterm infants at TEA (n = 91) compared to a large sample of healthy full-term controls (n = 69). We found region-specific increased MD, AD, and RD in the dlPFC, mPFC, PPC, motor, somatosensory, and visual cortices in preterm infants at TEA suggesting delayed regional cerebral cortical maturation in preterm infants.

While MD was associated with cerebral cortical dysmaturation in preterm infants, FA was not found to be a good marker of regional cerebral cortical maturation at TEA. We found that FA was significantly

elevated only in the primary motor cortex in very preterm infants. This finding is in keeping with the results of T. A. Smyser et al. (2015b), though, it differs from the findings of Ball et al. (2013), who reported significantly higher FA in preterm infants in the frontal, parietal, occipital, and temporal cortices. Mathematically, a proportional reduction or increase in diffusivity in all directions does not change FA. Thus, it is possible that alterations in microstructural organization affecting proportionally AD and RD result in MD differences between groups without FA differences. For instance, targeted apoptosis of the cerebral cortex in a rat model led to a progressive decrease in dendritic arbor of dying neurons and was characterized by an increase in AD and RD without FA changes during the apoptotic period (Petrenko et al., 2017). Additionally, since FA is a composite measure that expresses the directionality of diffusion, the relationship between FA and PMA is complex and varies depending on brain regions (Schneider et al., 2016; Wu et al., 2017a). Notably, at around TEA, the relationship between FA and PMA in the cerebral cortex is not linear. In fact, FA decreases up to 38/ 39 weeks PMA, and thereafter, either increases slightly or remains stationary (Aeby et al., 2012; Ball et al., 2013; Schneider et al., 2016). Lastly, cerebral cortical FA values were very low at TEA, ranging from 0.08 to 0.13 depending on the ROI. Thus, any subtle disruption of cortical FA developmental trajectory may be difficult to measure at TEA. As a result, FA may not be the most sensitive diffusion metric to characterize in vivo the developing cerebral cortex around TEA in preterm infants.

Our data also offer insights into the maturational status of different cortical regions in preterm infants at TEA. Specifically, the calcarine cortex followed by the paralimbic cortex (PCG, Insula) and the motor/somatosensory cortex were the brain areas with the lowest diffusivity (MD, AD, and RD), suggesting enhanced maturation compared to other brain areas. Conversely, the parietal, prefrontal, and temporal cortices showed higher diffusivity reflecting a less mature state. These findings

^a The brain abnormalities were classified using the scoring system developed by Kidokoro et al. (2013).

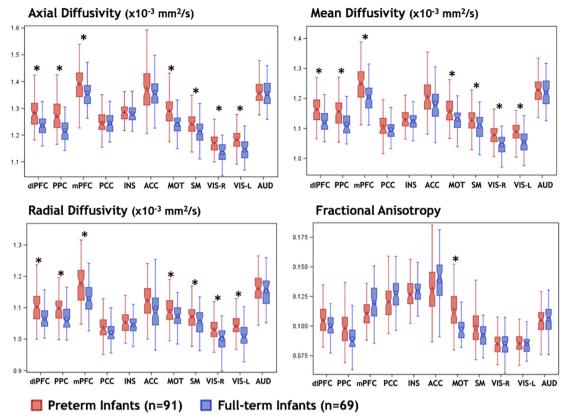


Fig. 2. Cerebral cortical microstructure in preterm infants (n = 91) and healthy full-term controls (n = 69) measured by DTI metrics.

* Indicates significant differences (p < 0.05) between preterm infants and healthy full-term controls controlling for sex and postmenstrual age at MRI.

ACC: anterior cingulate cortex; AUD: auditory cortex; dIPFC: dorsolateral prefrontal cortex; INS: insula; MOT: motor cortex; mPFC: medial prefrontal cortex; PCC: posterior cingulate cortex; PPC: posterior parietal cortex; SM: somatosensory cortex; VIS: visual cortex (R: right; L: left).

Table 2Comparison of DTI metrics by brain region in preterm and full-term infants.

Brain regions	Axial diffusivity β p -Value	Radial diffusivity β <i>p</i> -Value	Mean diffusivity β <i>p</i> -Value	Fractional anisotropy β p-Value
Dorsolateral	+0.099	+0.083	+0.088	NS
prefrontal cortex	0.0002	0.0007	0.0004	
Posterior parietal cortex	+0.08	+0.074	+ 0.076	NS
	0.0016	0.001	0.0008	110
Medial prefrontal	+0.07	+0.08	+0.077	NS
cortex	0.019	0.0063	0.0079	
Posterior cingulate cortex	NS	NS	NS	NS
Insula	NS	NS	NS	NS
Anterior cingulate cortex	NS	NS	NS	NS
Motor cortex	+0.1	+0.053	+0.068	+0.024
	0.0002	0.02	0.0032	0.0019
Somatosensory	+0.052	+0.05	+0.051	NS
cortex	0.02	0.0156	0.0137	
Right visual cortex	+0.064	+0.047	+0.053	NS
-	0.0007	0.013	0.0045	
Left visual cortex	+0.067	+0.048	+0.055	NS
	0.0009	0.0096	0.0038	
Auditory cortex	NS	NS	NS	NS

 β coefficient (preterm infants compared to full-term infants) and corresponding *p*-value were computed for each brain regions controlling for sex and postmenstrual age at MRI. NS: Not significant.

are in agreement with Eaton-Rosen et al. (2017) who demonstrated the highest rate of change in cortical radiality in the occipital cortex followed by the parietal cortex, and a relatively low rate of change for the

temporal and frontal cortices during the third trimester among preterm infants. Other DTI studies have also reported a gradient of cerebral cortical maturation during the third trimester (Ball et al., 2013; deIpolyi et al., 2005; T. A. Smyser et al., 2015b; Wu et al., 2017b; Yu et al., 2015). Taken together, diffusion imaging has enabled the *in vivo* quantification of known biological phenomena of spatiotemporal cerebral cortical maturation, which is characterized by a differential regional increase in dendritic spine density and synapse formation (Huttenlocher and Dabholkar, 1997; Travis et al., 2005).

Severe brain injury in preterm infants (grade III-IV IVH, cystic periventricular leukomalacia) has been associated with increased MD in the cerebral cortex (T. A. Smyser et al., 2015b), suggesting a negative trophic effect of brain injury on cortical development. In postmortem studies, periventricular leukomalacia in preterm infants has been linked with reduced neuronal density and/or neuronal gliosis (Andiman et al., 2010; Pierson et al., 2007). Additionally, in a fetal sheep model of prematurity, disruption in dendritic spine formation and remodeling following ischemic insult has been correlated with increased cortical FA (Dean et al., 2013). In contrast, we, and others (Aeby et al., 2012; Bonifacio et al., 2010; Vinall et al., 2013) found no association between mild/moderate brain injury and impaired cortical micro-organization assessed by DTI. The extent to which a cerebral insult in one region of the brain might affect the development of the cerebral cortex is likely related to the degree of brain injury severity and the topography of the injury. However, one cannot reject the hypothesis that current DTI metrics acquired on clinical scanners may not be sensitive enough to measure potential subtle microstructural changes following non-severe brain lesion. Noteworthy, in the fetal sheep model of prematurity, DTI data were acquired ex vivo using a high-field MRI scanner (11.7 T magnet), and demonstrates subtle microstructural changes following moderate white matter lesions (Dean et al., 2013). Thus, it may be

 Table 3

 Relationship between clinical risk factors and microstructural organization of the cerebral cortex in the preterm cohort.

Clinical risk factors	Preterm infants $(n = 91)$	Association with DTI metrics ^a
Parenchymal brain injury	44 (48%)	None
Chorioamnionitis	17 (19%)	None
Bronchopulmonary dysplasia (moderate to severe)	45 (50%)	None
Length of oxygen support (days)	91 ± 38 [0–172]	None
Postnatal steroid treatment	41 (45%)	Dorsolateral prefrontal cortex
		AD: $\beta = -0.036 p = 0.009$
		RD: $\beta = -0.029 p = 0.018$
		MD: $\beta = -0.032 p = 0.012$
		Medial prefrontal cortex
		AD: $\beta = -0.041 p = 0.006$
		RD: $\beta = -0.038 p = 0.007$
		MD: $\beta = -0.039 p = 0.005$
		Motor cortex
		AD: $\beta = -0.022 p = 0.01$
		RD: $\beta = -0.026 p = 0.023$
		MD: $\beta = -0.025 p = 0.029$
		Somatosensory cortex
		AD: $\beta = -0.028 p = 0.005$
		RD: $\beta = -0.02 p = 0.026$
		MD: $\beta = -0.022 p = 0.011$
Length of steroid treatment ^b (days)	19 ± 13 [2-55]	None
Dexamethasone treatment	15 (17%)	None
Hydrocortisone treatment	26 (29%)	Dorsolateral prefrontal cortex
•		AD: $\beta = -0.025 p = 0.05$
		MD: $\beta = -0.02 p = 0.008$
		Medial prefrontal cortex
		AD: $\beta = -0.036 p = 0.01$
		RD: $\beta = -0.035 p = 0.008$
		MD: $\beta = -0.036 p = 0.007$
		Somatosensory cortex
		AD: $\beta = -0.025 p = 0.009$
		MD: $\beta = -0.016 p = 0.05$
Sepsis	15 (17%)	None
Necrotizing enterocolitis	11 (12%)	None
Pressor treatment	39 (43%)	None
Patent ductus arteriosus ligation	24 (26%)	None

 $^{^{}a}$ ANCOVA analysis included each individual clinical risk factor as well as sex, postmenstrual age at MRI, and day of life at MRI. β is the regression coefficient associated with the risk factor in the model.

possible to investigate the effect of mild to moderate brain injury on the developing cerebral cortex using high-resolution neuroimaging techniques (Dudink et al., 2014).

Another clinical risk factor we examined was the influence of postnatal steroid treatment. Steroids have been routinely used for their anti-inflammatory properties to prevent and treat chronic lung disease in preterm infants born with immature lungs (Kennedy et al., 2016). More recently, postnatal steroids have also been administered as an antihypertensive therapy (Higgins et al., 2009; Johnson, 2015). However, the administration of exogenous steroids may have beneficial and/or detrimental effects on the developing brain (Eventov-Friedman and Shinwell, 2008; Malaeb and Stonestreet, 2014). It has been reported that the effects of steroid administration on brain development varies according to the developmental stage at exposure (İşcan et al., 2017), the length and dose of exposure (Gray et al., 2013; Liston and Gan, 2011), and the pharmacodynamic properties of the steroid used (Malaeb and Stonestreet, 2014). Hydrocortisone is the closest and weakest pharmaceutical form of cortisol and has a high affinity for mineralocorticoid receptors, whereas dexamethasone is a synthetic steroid approximately 25 times more potent and is a glucocorticoid receptor agonist (Eventov-Friedman and Shinwell, 2008; Malaeb and Stonestreet, 2014; van der Heide-Jalving et al., 2003). These differences in pharmacological properties, specifically in receptor affinity, may be responsible for the complex and multifaceted relationship between steroids and the developing brain (Malaeb and Stonestreet, 2014).

On one hand, postnatal administration of dexamethasone has been linked with adverse neurodevelopmental outcome (Barrington, 2001;

Yeh et al., 2004) and impaired cerebral growth from the early neonatal period to adolescence (Bouyssi-Kobar et al., 2016; Cheong et al., 2014; Murphy et al., 2001; Parikh et al., 2007), while on the other hand, postnatal hydrocortisone treatment has not been associated with adverse neurodevelopment during childhood (Baud et al., 2017; Lodygensky et al., 2005; Rademaker et al., 2008; Rademaker and de Vries, 2009; Renault et al., 2016; van der Heide-Jalving et al., 2003; Watterberg et al., 2007). Although we did not specifically test for the differential effect of steroid treatment on the cortex in our study, we independently investigated the effect of dexamethasone and hydrocortisone on cerebral cortical microstructural organization. We did not find an effect of dexamethasone on regional cerebral microstructural organization. Still, we demonstrated an association between hydrocortisone treatment and 'accelerated' microstructural organization in the prefrontal and somatosensory cortices. These results may be spurious, or may suggest a potentially protective effect on regional cerebral cortical maturation. As reported in mice models, this association might be due to enhancement in spine dynamics (elimination/remodeling) in brain regions undergoing active spinogenesis at the time of administration (Gray et al., 2013; Liston and Gan, 2011). Another hypothesis is that maintaining adequate blood pressure through hydrocortisone therapy improves hemodynamic support resulting in better cerebrovascular outcome (Fyfe et al., 2014; Johnson, 2015). Collectively, evidence suggest that hydrocortisone therapy might be a safer alternative to dexamethasone; though, large randomized clinical trials examining short- and long-term outcomes are necessary to confirm these results.

^b Length of steroid treatment for infants that received postnatal steroid treatment (n = 41).

Our study has several limitations including the use of an anatomical neonatal atlas based on a different population (infants born after 33 weeks GA) than the one studied. The use of multi-contrast, PMAappropriate, and population specific atlases would improve reliability of measurements (Deshpande et al., 2015; Oishi et al., 2011). Nonetheless, using an atlas facilitates the standardization of the ROIs placement across the cohort, reducing the bias introduced by manual drawing of ROIs. When dealing with the developing cerebral cortex, another concern is the risk of partial volume averaging of white matter or cerebrospinal fluid (Aeby et al., 2012). In order to minimize partial volume effect, we completed automatic segmentation of anatomical images using an age-appropriate atlas (Makropoulos et al., 2014) to select cortical gray matter voxels only. We also performed manual corrections of misplaced voxels from the automated ROIs placement. Nevertheless, we cannot exclude the possibility of partial volume effect influencing our data. Additionally, DTI metrics characterize multiple and heterogeneous tissue within a voxel and cannot decipher precisely the biological contribution from each tissue (Wu et al., 2017b). Thus, advanced diffusion acquisitions such as multi-shell high resolution techniques, which provide more detailed information about diffusion characteristic within each voxel, would allow for more precise in vivo characterization of the developing cerebral cortex (Dudink et al., 2014). Although the implementation of such advanced diffusion techniques might be more challenging in a clinical setting due to time constraints, its application to the preterm population has the potential to provide new detailed insights into cortical microstructural organization (Dudink et al., 2014; Eaton-Rosen et al., 2015; Shi et al., 2016). Finally, the relationship between altered cortical microstructural organization and neurodevelopmental outcome remains to be established once the follow-up data from this cohort are completed.

In conclusion, we demonstrated early alterations in cerebral cortical microstructural organization in a large cohort of preterm infants at TEA compared to healthy full-term newborns. Specifically, three brain regions involved in neurocognitive processing (dlPFC, mPFC, and PPC) as well as motor, somatosensory, and visual cortices, exhibited reduced microstructural organization suggesting delayed cortical maturation in very preterm infants at TEA. These findings highlight the vulnerability of cerebral cortical microstructural organization following preterm birth. Future work will examine the potential link between early alterations of the neurocognitive brain areas and later adverse sociocognitive outcome. Interestingly, postnatal hydrocortisone treatment was associated with accelerated maturation in the prefrontal and somatosensory cortices suggesting a potential short-term protective effect of hydrocortisone administration on regional cortical development. Future studies are needed to examine the impact of postnatal hydrocortisone treatment on cortical development using more in-depth analysis that will incorporate length of treatment, cumulative dose, and timing of exposure.

Funding source

This work was supported by the Canadian Institutes of Health Research (MOP-81116); the SickKids Foundation (XG 06-069); the National Institutes of Health (R01 HL116585-01) and the Intellectual and Developmental Disability Research Centers (U54HD090257). Marine Bouyssi-Kobar is a Ph.D. student in the Molecular Medicine Program of the Institute for Biomedical Sciences at the George Washington University. This work is from a dissertation to be presented to the above program in partial fulfillment of the requirements for the Ph.D. degree.

Conflict of interest

The authors have no conflicts of interest relevant to this article to disclose.

Acknowledgements

The authors are deeply thankful to all the families that participated in the study. We also would like to thank Dr. Nickie Andescavage for her critical clinical expertise and Dr. Sonia Dahdouh for her critical feedback while developing the image processing steps.

References

- Aeby, A., Van Bogaert, P., David, P., Balériaux, D., Vermeylen, D., Metens, T., De Tiège, X., 2012. Nonlinear microstructural changes in the right superior temporal sulcus and lateral occipitotemporal gyrus between 35 and 43 weeks in the preterm brain. 63, 104–110. http://dx.doi.org/10.1016/j.neuroimage.2012.06.013.
- Andiman, S.E., Haynes, R.L., Trachtenberg, F.L., Billiards, S.S., Folkerth, R.D., Volpe, J.J., Kinney, H.C., 2010. The cerebral cortex overlying periventricular leukomalacia: analysis of pyramidal neurons. Brain Pathol. 20, 803–814. http://dx.doi.org/10. 1111/j.1750-3639.2010.00380.x.
- Ball, G., Srinivasan, L., Aljabar, P., Counsell, S.J., Durighel, G., Hajnal, J.V., Rutherford, M.A., Edwards, A.D., 2013. Development of cortical microstructure in the preterm human brain. Proc. Natl. Acad. Sci. U. S. A. 110, 9541–9546. http://dx.doi.org/10.1073/pnas.1301652110.
- Ball, G., Pazderova, L., Chew, A., Tusor, N., Merchant, N., Arichi, T., Allsop, J.M., Cowan, F.M., Edwards, A.D., Counsell, S.J., 2015. Thalamocortical connectivity predicts cognition in children born preterm. Cereb. Cortex 25, 4310–4318. http://dx.doi.org/10.1093/cercor/phu331.
- Barrington, K.J., 2001. The adverse neuro-developmental effects of postnatal steroids in the preterm infant: a systematic review of RCTs. BMC Pediatr. 1 (1). http://dx.doi. org/10.1186/1471-2431-1-1.
- Basser, P.J., Mattiello, J., Le Bihan, D., 1994. MR diffusion tensor spectroscopy and imaging. Biophys. J. 66, 259–267. http://dx.doi.org/10.1016/S0006-3495(94) 80775-1
- Baud, O., Trousson, C., Biran, V., Leroy, E., Mohamed, D., Alberti, C., PREMILOC Trial Group, 2017. Association between early low-dose hydrocortisone therapy in extremely preterm neonates and neurodevelopmental outcomes at 2 years of age. JAMA 317, 1329–1337. http://dx.doi.org/10.1001/jama.2017.2692.
- Blencowe, H., Lee, A.C., Cousens, S., Bahalim, A., Narwal, R., Zhong, N., Chou, D., Say, L., Modi, N., Katz, J., Vos, T., Marlow, N., Lawn, J.E., 2013. Preterm birth–associated neurodevelopmental impairment estimates at regional and global levels for 2010. Pediatr. Res. 74, 17–34. http://dx.doi.org/10.1038/pr.2013.204.
- Bonifacio, S.L., Glass, H.C., Chau, V., Berman, J.I., Xu, D., Brant, R., Barkovich, A.J., Poskitt, K.J., Miller, S.P., Ferriero, D.M., 2010. Extreme premature birth is not associated with impaired development of brain microstructure. J. Pediatr. 157, 726–732.e1. http://dx.doi.org/10.1016/j.jpeds.2010.05.026.
- Bouyssi-Kobar, M., Plessis, Du A.J., McCarter, R., Brossard-Racine, M., Murnick, J., Tinkleman, L., Robertson, R.L., Limperopoulos, C., 2016. Third trimester brain growth in preterm infants compared with in utero healthy fetuses. Pediatrics 138, e20161640. http://dx.doi.org/10.1542/peds.2016-1640.
- Bouyssi-Kobar, M., Murnick, J., Brossard-Racine, M., Chang, T., Mahdi, E., Jacobs, M., Limperopoulos, C., 2017. Altered cerebral perfusion in infants born preterm compared with infants born full term. J. Pediatr. http://dx.doi.org/10.1016/j.jpeds.2017. 09.083.
- Brossard-Racine, M., Poretti, A., Murnick, J., Bouyssi-Kobar, M., McCarter, R., Plessis, Du A.J., Limperopoulos, C., 2016. Cerebellar microstructural organization is altered by complications of premature birth: a case-control study. J. Pediatr. http://dx.doi.org/10.1016/i.jpeds.2016.10.034.
- Budday, S., Steinmann, P., Kuhl, E., 2015. Physical biology of human brain development. Front. Cell. Neurosci. 9, 1–17. http://dx.doi.org/10.3389/fncel.2015.00257.
- Cheong, J.L.Y., Burnett, A.C., Lee, K.J., Roberts, G., Thompson, D.K., Wood, S.J., Connelly, A., Anderson, P.J., Doyle, L.W., Victorian Infant Collaborative Study Group, 2014. Association between postnatal dexamethasone for treatment of bronchopulmonary dysplasia and brain volumes at adolescence in infants born very preterm. J. Pediatr. 164, 737–743.e1. http://dx.doi.org/10.1016/j.jpeds.2013.10. 083
- Cox, R.W., 1996. AFNI: software for analysis and visualization of functional magnetic resonance neuroimages. Comput. Biomed. Res. 29, 162–173.
- Dean, J.M., McClendon, E., Hansen, K., Azimi-Zonooz, A., Chen, K., Riddle, A., Gong, X., Sharifnia, E., Hagen, M., Ahmad, T., Leigland, L.A., Hohimer, A.R., Kroenke, C.D., Back, S.A., 2013. Prenatal cerebral ischemia disrupts MRI-defined cortical microstructure through disturbances in neuronal arborization. Sci. Transl. Med. 5, 168ra7. http://dx.doi.org/10.1126/scitranslmed.3004669.
- delpolyi, A.R., Mukherjee, P., Gill, K., Henry, R.G., Partridge, S.C., Veeraraghavan, S., Jin, H., Lu, Y., Miller, S.P., Ferriero, D.M., Vigneron, D.B., Barkovich, A.J., 2005. Comparing microstructural and macrostructural development of the cerebral cortex in premature newborns: diffusion tensor imaging versus cortical gyration. 27, 579–586. http://dx.doi.org/10.1016/j.neuroimage.2005.04.027.
- Deshpande, R., Chang, L., Oishi, K., 2015. Construction and application of human neonatal DTI atlases. Front. Neuroanat. 9. http://dx.doi.org/10.3389/fnana.2015.00138. Dobbing, J., Sands, J., 1973. Quantitative growth and development of human brain. Arch. Dis. Child. 48, 757–767.
- Doria, V., Beckmann, C.F., Arichi, T., Merchant, N., Groppo, M., Turkheimer, F.E., Counsell, S.J., Murgasova, M., Aljabar, P., Nunes, R.G., Larkman, D.J., Rees, G., Edwards, A.D., 2010. Emergence of resting state networks in the preterm human brain. Proc. Natl. Acad. Sci. U. S. A. 107, 20015–20020. http://dx.doi.org/10.1073/

pnas.1007921107.

- Dudink, J., Pieterman, K., Leemans, A., Kleinnijenhuis, M., van Cappellen van Walsum, A.M., Hoebeek, F.E., 2014. Recent advancements in diffusion MRI for investigating cortical development after preterm birth-potential and pitfalls. Front. Hum. Neurosci. 8, 1066. http://dx.doi.org/10.3389/fnhum.2014.01066.
- Eaton-Rosen, Z., Melbourne, A., Orasanu, E., Cardoso, M.J., Modat, M., Bainbridge, A., Kendall, G.S., Robertson, N.J., Marlow, N., Ourselin, S., 2015. Longitudinal measurement of the developing grey matter in preterm subjects using multi-modal MRI. 1–10. http://dx.doi.org/10.1016/j.neuroimage.2015.02.010.
- Eaton-Rosen, Z., Scherrer, B., Melbourne, A., Ourselin, S., Neil, J.J., Warfield, S.K., 2017. Investigating the maturation of microstructure and radial orientation in the preterm human cortex with diffusion MRI. NeuroImage 1–24. http://dx.doi.org/10.1016/j. neuroImage.2017.08.013.
- Evangelou, I.E., Serag, A., Bouyssi-Kobar, M., Plessis, A.J.D., Limperopoulos, C., 2014. Optimized methodology for neonatal diffusion tensor imaging processing and study-specific template construction. 2014, 2372–2375. http://dx.doi.org/10.1109/EMBC. 2014.6944098
- Eventov-Friedman, S., Shinwell, E.S., 2008. Current controversies in perinatal steroid therapy. Acta Paediatr. 97, 1492–1501. http://dx.doi.org/10.1111/j.1651-2227.
- Fyfe, K.L., Yiallourou, S.R., Wong, F.Y., Horne, R.S.C., 2014. The development of cardiovascular and cerebral vascular control in preterm infants. Sleep Med. Rev. 18, 299–310. http://dx.doi.org/10.1016/j.smrv.2013.06.002.
- Gray, J.D., Milner, T.A., McEwen, B.S., 2013. Dynamic plasticity: the role of glucocorticoids, brain-derived neurotrophic factor and other trophic factors. Neuroscience 239, 214–227. http://dx.doi.org/10.1016/j.neuroscience.2012.08.034.
- Gupta, R.K., Hasan, K.M., Trivedi, R., Pradhan, M., Das, V., Parikh, N.A., Narayana, P.A., 2005. Diffusion tensor imaging of the developing human cerebrum. J. Neurosci. Res. 81, 172–178. http://dx.doi.org/10.1002/jnr.20547.
- Higgins, S., Friedlich, P., Seri, I., 2009. Hydrocortisone for hypotension and vasopressor dependence in preterm neonates: a meta-analysis. J. Perinatol. 30, 373–378. http:// dx.doi.org/10.1038/jp.2009.126.
- Hüppi, P.S., Dubois, J., 2006. Diffusion tensor imaging of brain development. Semin. Fetal Neonatal Med. 11, 489–497. http://dx.doi.org/10.1016/j.siny.2006.07.006.
- Huttenlocher, P.R., Dabholkar, A.S., 1997. Regional differences in synaptogenesis in human cerebral cortex. J. Comp. Neurol. 387, 167–178.
- İşcan, B., Tuzun, F., Cilaker Micili, S., Tugyan, K., Duman, N., Ozkan, H., Kumral, A., 2017. The effects of perinatal steroid therapy on growth factor levels during different stages of the developing brain. J. Matern. Fetal Neonatal Med. 30, 1820–1828. http://dx.doi.org/10.1080/14767058.2016.1228051.
- Jenkinson, M., Beckmann, C.F., Behrens, T.E.J., Woolrich, M.W., Smith, S.M., 2012. FSL. NeuroImage 62, 782–790. http://dx.doi.org/10.1016/j.neuroimage.2011.09.015.
- Jespersen, S.N., Leigland, L.A., Cornea, A., Kroenke, C.D., 2013. Determination of axonal and dendritic orientation distributions within the developing cerebral cortex by diffusion tensor imaging. IEEE Trans. Med. Imaging 31, 16–32. http://dx.doi.org/10. 1109/TMI.2011.2162099.
- Jobe, A.H., Bancalari, E., 2001. Bronchopulmonary dysplasia. Am. J. Respir. Crit. Care Med. 163, 1723–1729. http://dx.doi.org/10.1164/airccm.163.7.2011060.
- Johnson, P.J., 2015. Hydrocortisone for treatment of hypotension in the newborn. Neonatal Netw. 34, 46–51. http://dx.doi.org/10.1891/0730-0832.34.1.46.
- Johnson, S., Marlow, N., 2011. Preterm birth and childhood psychiatric disorders. Pediatr. Res. 69, 11R–18R. http://dx.doi.org/10.1203/PDR.0b013e318212faa0.
- Kennedy, K.A., Cotten, C.M., Watterberg, K.L., Carlo, W.A., 2016. Prevention and management of bronchopulmonary dysplasia: lessons learned from the neonatal research network. Semin. Perinatol. 40, 348–355. http://dx.doi.org/10.1053/j.semperi.2016. 05 010
- Kersbergen, K.J., Leemans, A., Groenendaal, F., van der Aa, N.E., Viergever, M.A., de Vries, L.S., Benders, M.J.N.L., 2014. Microstructural brain development between 30 and 40 weeks corrected age in a longitudinal cohort of extremely preterm infants. NeuroImage 103, 214–224. http://dx.doi.org/10.1016/j.neuroimage.2014.09.039.
- Kidokoro, H., Neil, J.J., Inder, T.E., 2013. New MR imaging assessment tool to define brain abnormalities in very preterm infants at term. AJNR Am. J. Neuroradiol. 34, 2208–2214. http://dx.doi.org/10.3174/ajnr.A3521.
- Kreis, R., Ernst, T., Ross, B.D., 1993. Development of the human brain: in vivo quantification of metabolite and water content with proton magnetic resonance spectroscopy. Magn. Reson. Med. 30, 424–437.
- Leemans, A., Jones, D.K., 2009. The B-matrix must be rotated when correcting for subject motion in DTI data. Magn. Reson. Med. 61, 1336–1349. http://dx.doi.org/10.1002/mrm 21890
- Liston, C., Gan, W.-B., 2011. Glucocorticoids are critical regulators of dendritic spine development and plasticity in vivo. Proc. Natl. Acad. Sci. U. S. A. 108, 16074–16079. http://dx.doi.org/10.1073/pnas.1110444108.
- Lodygensky, G.A., Rademaker, K., Zimine, S., Gex-Fabry, M., Lieftink, A.F., Lazeyras, F., Groenendaal, F., de Vries, L.S., Hüppi, P.S., 2005. Structural and functional brain development after hydrocortisone treatment for neonatal chronic lung disease. Pediatrics 116, 1–7. http://dx.doi.org/10.1542/peds.2004-1275.
- Makropoulos, A., Gousias, I.S., Ledig, C., Aljabar, P., Serag, A., Hajnal, J.V., Edwards, A.D., Counsell, S.J., Rueckert, D., 2014. Automatic whole brain MRI segmentation of the developing neonatal brain. IEEE Trans. Med. Imaging 33, 1818–1831. http://dx.doi.org/10.1109/TMI.2014.2322280.
- Malaeb, S.N., Stonestreet, B.S., 2014. Steroids and injury to the developing brain. Clin. Perinatol. 41, 191–208. http://dx.doi.org/10.1016/j.clp.2013.09.006.
- McKinstry, R.C., Mathur, A., Miller, J.H., Ozcan, A., Snyder, A.Z., Schefft, G.L., Almli, C.R., Shiran, S.I., Conturo, T.E., Neil, J.J., 2002. Radial organization of developing preterm human cerebral cortex revealed by non-invasive water diffusion anisotropy MRI. Cereb. Cortex 12, 1237–1243.

- Mesulam, M., 2012. The evolving landscape of human cortical connectivity: facts and inferences. NeuroImage 62, 2182–2189. http://dx.doi.org/10.1016/j.neuroimage. 2011.12.033.
- Mori, S., Zhang, J., 2006. Principles of diffusion tensor imaging and its applications to basic neuroscience research. Neuron 51, 527–539. http://dx.doi.org/10.1016/j. neuron.2006.08.012.
- Murphy, B.P., Inder, T.E., Huppi, P.S., Warfield, S., Zientara, G.P., Kikinis, R., Jolesz, F.A., Volpe, J.J., 2001. Impaired cerebral cortical gray matter growth after treatment with dexamethasone for neonatal chronic lung disease. Pediatrics 107, 217–221. http://dx.doi.org/10.1542/peds.107.2.217.
- Oishi, K., Mori, S., Donohue, P.K., Ernst, T., Anderson, L., Buchthal, S., Faria, A., Jiang, H., Li, X., Miller, M.I., van Zijl, P.C.M., Chang, L., 2011. Multi-contrast human neonatal brain atlas: application to normal neonate development analysis. 56, 8–20. http://dx.doi.org/10.1016/j.neuroimage.2011.01.051.
- Papile, L.A., Burstein, J., Burstein, R., Koffler, H., 1978. Incidence and evolution of subependymal and intraventricular hemorrhage: a study of infants with birth weights less than 1,500 gm. J. Pediatr. 92, 529–534. http://dx.doi.org/10.1016/S0022-3476(78)80282-0.
- Parikh, N.A., Lasky, R.E., Kennedy, K.A., Moya, F.R., Hochhauser, L., Romo, S., Tyson, J.E., 2007. Postnatal dexamethasone therapy and cerebral tissue volumes in extremely low birth weight infants. Pediatrics 119, 265–272. http://dx.doi.org/10.1542/peds.2006-1354.
- Petrenko, V., van de Looij, Y., Mihhailova, J., Salmon, P., Hüppi, P.S., Sizonenko, S.V., Kiss, J.Z., 2017. Multimodal MRI imaging of apoptosis-triggered microstructural alterations in the postnatal cerebral cortex. Cereb. Cortex 1–14. http://dx.doi.org/10.1093/cercor/bhw420.
- Pierpaoli, C., Basser, P.J., 1996. Toward a quantitative assessment of diffusion anisotropy. Magn. Reson. Med. 36, 893–906.
- Pierson, C.R., Folkerth, R.D., Billiards, S.S., Trachtenberg, F.L., Drinkwater, M.E., Volpe, J.J., Kinney, H.C., 2007. Gray matter injury associated with periventricular leukomalacia in the premature infant. Acta Neuropathol. 114, 619–631. http://dx.doi.org/10.1007/s00401-007-0295-5.
- Qiu, A., Mori, S., Miller, M.I., 2015. Diffusion tensor imaging for understanding brain development in early life. Annu. Rev. Psychol. 66, 853–876. http://dx.doi.org/10. 1146/annurev-psych-010814-015340.
- Rademaker, K.J., de Vries, W.B., 2009. Long-term effects of neonatal hydrocortisone treatment for chronic lung disease on the developing brain and heart. Semin. Fetal Neonatal Med. 14, 171–177. http://dx.doi.org/10.1016/j.siny.2008.11.004.
- Rademaker, K.J., de Vries, L.S., Uiterwaal, C.S.P.M., Groenendaal, F., Grobbee, D.E., van Bel, F., 2008. Postnatal hydrocortisone treatment for chronic lung disease in the preterm newborn and long-term neurodevelopmental follow-up. Arch. Dis. Child. Fetal Neonatal Ed. 93, F58–F63. http://dx.doi.org/10.1136/adc.2007.119545.
- Raybaud, C., Ahmad, T., Rastegar, N., Shroff, M., Nassar, Al M., 2013. The premature brain: developmental and lesional anatomy. Neuroradiology 55, 23–40. http://dx. doi.org/10.1007/s00234-013-1231-0.
- Renault, A., Patkaï, J., Dassieu, G., Ayoubi, El M., Canouï-Poitrine, F., Durrmeyer, X., 2016. Hydrocortisone use in ventilated extremely preterm infants decreased bronchopulmonary dysplasia with no effects on neurodevelopment after two years. Acta Paediatr. 105, 1047–1055. http://dx.doi.org/10.1111/apa.13487.
- Schneider, J., Kober, T., Bickle Graz, M., Meuli, R., Hu ppi, P.S., Hagmann, P., Truttmann, A.C., 2016. Evolution of T1 relaxation, ADC, and fractional anisotropy during early brain maturation: a serial imaging study on preterm infants. AJNR Am. J. Neuroradiol. 37, 155–162. http://dx.doi.org/10.3174/ajnr.A4510.
 Shi, F., Yap, P.-T., Wu, G., Jia, H., Gilmore, J.H., Lin, W., Shen, D., 2011. Infant brain
- Shi, F., Yap, P.-T., Wu, G., Jia, H., Gilmore, J.H., Lin, W., Shen, D., 2011. Infant brain atlases from neonates to 1- and 2-year-olds. PLoS One 6, e18746. http://dx.doi.org/ 10.1371/journal.pone.0018746.
- Shi, J., Chang, L., Wang, J., Zhang, S., Yao, Y., Zhang, S., Jiang, R., Guo, L., Guan, H., Zhu, W., 2016. Initial application of diffusional kurtosis imaging in evaluating brain development of healthy preterm infants. PLoS One 11, e0154146–13. http://dx.doi.org/10.1371/journal.pone.0154146.
- Sizonenko, S.V., Camm, E.J., Garbow, J.R., Maier, S.E., Inder, T.E., Williams, C.E., Neil, J.J., Huppi, P.S., 2007. Developmental changes and injury induced disruption of the radial organization of the cortex in the immature rat brain revealed by in vivo diffusion tensor MRI. Cereb. Cortex 17, 2609–2617. http://dx.doi.org/10.1093/cercor/bb1168.
- Smith, S.M., 2002. Fast robust automated brain extraction. Hum. Brain Mapp. 17, 143–155. http://dx.doi.org/10.1002/hbm.10062.
- Smyser, C.D., Snyder, A.Z., Shimony, J.S., Mitra, A., Inder, T.E., Neil, J.J., 2015a. Resting-state network complexity and magnitude are reduced in prematurely born infants. Cereb. Cortex 26, 322–333. http://dx.doi.org/10.1093/cercor/bhu251.
- Smyser, T.A., Smyser, C.D., Rogers, C.E., Gillespie, S.K., Inder, T.E., Neil, J.J., 2015b.
 Cortical gray and adjacent white matter demonstrate synchronous maturation in very preterm infants. Cereb Cortex bhv164–9. https://doi.org/10.1093/cercor/bhv164.
- Studholme, C., Hill, D.L.G., Hawkes, D.J., 1999. An overlap invariant entropy measure of 3D medical image alignment. Pattern Recogn. 32, 71–86. http://dx.doi.org/10.1016/ s0031-3203(98)00091-0.
- Travis, K., Ford, K., Jacobs, B., 2005. Regional dendritic variation in neonatal human cortex: a quantitative Golgi study. Dev. Neurosci. 27, 277–287. http://dx.doi.org/10. 1159/000086707.
- Trivedi, R., Husain, N., Rathore, R.K.S., Saksena, S., Srivastava, S., Malik, G.K., Das, V., Pradhan, M., Pandey, C.M., Gupta, R.K., 2009. Correlation of diffusion tensor imaging with histology in the developing human frontal cerebrum. Dev. Neurosci. 31, 487–496. http://dx.doi.org/10.1159/000229500.
- Tzourio-Mazoyer, N., Landeau, B., Papathanassiou, D., Crivello, F., Etard, O., Delcroix, N., Mazoyer, B., Joliot, M., 2002. Automated anatomical labeling of activations in SPM using a macroscopic anatomical parcellation of the MNI MRI single-subject brain. 15,

- 273-289. http://dx.doi.org/10.1006/nimg.2001.0978.
- van der Heide-Jalving, M., K., P.J.G.H., van der Laan, M.J., B., J.M., W., V.M., H, C.J., Ve, S., van Bel, F., 2003. Short- and long-term effects of neonatal glucocorticoid therapy: is hydrocortisone an alternative to dexamethasone? Acta Paediatr. 92, 827–835. http://dx.doi.org/10.1080/08035250310002425.
- Vinall, J., Grunau, R.E., Brant, R., Chau, V., Poskitt, K.J., Synnes, A.R., Miller, S.P., 2013. Slower postnatal growth is associated with delayed cerebral cortical maturation in preterm newborns. Sci. Transl. Med. 5, 168ra8. http://dx.doi.org/10.1126/ scitranslmed.3004666.
- Watterberg, K.L., Shaffer, M.L., Mishefske, M.J., Leach, C.L., Mammel, M.C., Couser, R.J., Abbasi, S., Cole, C.H., Aucott, S.W., Thilo, E.H., Rozycki, H.J., Lacy, C.B., 2007. Growth and neurodevelopmental outcomes after early low-dose hydrocortisone treatment in extremely low birth weight infants. Pediatrics 120, 40–48. http://dx.doi.org/10.1542/peds.2006-3158.
- White, T.P., Symington, I., Castellanos, N.P., Brittain, P.J., Froudist-Walsh, S., Nam, K.W., Sato, J.R., Allin, M.P.G., Shergill, S.S., Murray, R.M., Williams, S.C.R., Nosarti, C., 2014. Dysconnectivity of neurocognitive networks at rest in very-preterm born

- adults. NeuroImage 4, 352-365. http://dx.doi.org/10.1016/j.nicl.2014.01.005.
- Wu, D., Chang, L., Akazawa, K., Oishi, K., Skranes, J., Ernst, T., Oishi, K., 2017a. Mapping the critical gestational age at birth that alters brain development in preterm-born infants using multi-modal MRI. NeuroImage 149, 33–43. http://dx.doi.org/10.1016/ j.neuroimage.2017.01.046.
- Wu, D., Chang, L., Akazawa, K., Oishi, K., Skranes, J., Ernst, T., Oishi, K., 2017b. Change-point analysis data of neonatal diffusion tensor MRI in preterm and term-born infants. Data Brief 12, 453–458. http://dx.doi.org/10.1016/j.dib.2017.04.020.
- Yeh, T.F., Lin, Y.J., Lin, H.C., Huang, C.C., Hsieh, W.S., Lin, C.H., Tsai, C.H., 2004. Outcomes at school age after postnatal dexamethasone therapy for lung disease of prematurity. N. Engl. J. Med. 350, 1304–1313. http://dx.doi.org/10.1056/ NF IMag032089
- Yu, Q., Ouyang, A., Chalak, L., Jeon, T., Chia, J., Mishra, V., Sivarajan, M., Jackson, G., Rollins, N., Liu, S., Huang, H., 2015. Structural development of human fetal and preterm brain cortical plate based on population-averaged templates. Cereb. Cortex 26, 4381–4391. bhv201–11. https://doi.org/10.1093/cercor/bhv201.