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Distribution of Major Staphylococcal Cassette Chromosome *mec* Types and Exfoliative Toxin Genes in *Staphylococcus pseudintermedius* Strains from Dogs with Superficial Pyoderma in Japan

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Abstract: Staphylococcus pseudintermedius is a major pathogen of canine pyoderma, known to produce exfoliative toxins that could be involved in formation of cutaneous lesions. To understand the genotypic distribution of S. pseudintermedius, we surveyed 74 dogs with pyoderma in three veterinary hospitals in Japan. Seventy-four S. pseudintermedius strains were isolated, 52 of which (70.3%) were mecA-positive methicillin-resistant S. pseudintermedius (MRSP). Staphylococcal cassette chromosome mec (SCCmec) typing of the identified MRSP strains revealed that the most prevalent genotype was type III-like (63.4%) followed by type V (34.6%). These data suggest high prevalence of MRSP strains consisting of two major SCCmec types among canine pyoderma in Japan. We found low prevalence of exfoliative toxin genes (exp) in the MRSP strains: expA and expB were present in 1.9% and 0%, respectively. These findings suggest no association in carriage between mecA and exp genes in S. pseudintermedius from canine pyoderma.

Key words: *Staphylococcus pseudintermedius*; superficial pyoderma; methicillin resistance; SCC*mec*; exfoliative toxin

Introduction

Staphylococcus pseudintermedius is a normal inhabitant of the skin and mucosae of dogs^{1, 2)}. This species is also known to be the major pathogen of superficial pyoderma, one of the most common infectious diseases of canine cutaneous disorder³⁾. Previous studies have revealed that S. pseudintermedius possess virulence factors such as exfoliative toxins (ETs) ExpA and B, which cause skin exfoliation⁴⁻⁶⁾. However, few studies have described the distribution of ETs in S. pseudintermedius from canine superficial pyoderma^{4, 5)}, and the presence of ETs in methicillin-resistant S. pseudintermedius (MRSP) has not been reported.

Since the first report of a *mecA*-positive MRSP strain in 1999⁷⁾, MRSP infections have been increasing in small animal medicine⁸⁻¹²⁾. According to previous studies, MRSP strains are mainly classified into two genotypes based on the type of staphylococcal cassette chromosome *mec* (SCC*mec*): SCC*mec* type III-like clones (informally designated type II-III by Descloux *et al.*¹³⁾), which are found in Europe and many other areas across the world¹⁴⁻¹⁸⁾, and type V clones, which are prevalent in North America, Korea and Thailand^{14, 15, 19-22)}. Genotyping is important and helpful in understanding the geographic distribution and estimating the epidemic nature and spread of MRSP clones. However, only a few studies have performed genotype-based analysis of canine

superficial pyoderma caused by MRSP in Japan¹⁸). We therefore conducted molecular analysis of MRSP strains isolated from canine superficial pyoderma and determined SCC*mec* types in Japan. We also analyzed two exfoliative toxin genes, *expA* and *expB*, to investigate the association between methicillin resistance and the carriage of toxin genes. Here, we describe the prevalence of methicillin resistance and exfoliative toxin genes in the genome of *S. pseudintermedius* among 74 dogs with superficial pyoderma from three veterinary hospitals in Japan.

Materials and Methods

Sample collecting

We examined 74 dogs with superficial pyoderma in three private veterinary hospitals in three prefectures of Japan between April 2010 and December 2012. The 74 dogs (37 males, 37 females; mean age, 7.9 years [range, 10 months to 15 years]) were 10 Shih Tzu dogs, 9 French Bulldogs, 9 Poodles, 7 Miniature Dachshunds, 5 Shiba Inu dogs, 5 Pugs, 4 Chihuahuas, 4 Cocker Spaniels, 2 West Highland white terriers, 2 Retrievers, 2 Malteses, 2 Yorkshire terriers, 2 Cavalier King Charles Spaniels, 2 Jack Russells, 1 Basset, 1 Chin, 1 German shepherd, 1 Pekingese, 1 Schnauzer, 1 Weimaraner and 3 Mixed breeds. A total of 74 specimens were collected by swabbing skin lesions. Bacterial strains from the specimens were cultivated on tryptic soy agar containing 5% sheep blood (BD Japan, Co., Ltd., Tokyo, Japan) at 37°C for 18 h. All strains were identified as staphylococci based on colony morphology, Gram stain appearance and the catalase test.

Species identification, determination of methicillin resistance and SCCmec typing

Crude DNA extraction from a single colony and staphylococcal species identification using multiplex PCR (M-PCR) were performed as previously described by Sasaki *et al.*²³⁾.

To identify methicillin resistance, a PCR method²⁴⁾ for detection of the *mecA* gene was used. Subsequently, SCC*mec* typing of the MRSP strains identified was

performed. To discriminate SCC*mec* types I to V, including type III-like, classified based on the *ccr* and *mec* gene complexes, two M-PCRs²⁵⁾ and one duplex PCR¹⁵⁾ were carried out.

Detection of exfoliative toxin genes

Fragments from two exfoliative toxin genes, expA and expB, were amplified by conventional PCR. The oligonucleotide primers were as previously reported (Yamamoto et al., 2012, 15th Annual meeting of The Japanese Society of Veterinary Dermatology): 5'-ATTTGTTCACATGGATTTATT-3' (forward) and 5'-AGGGGCATTAACAATAAGATC-3' (reverse) for expA, and 5'-TTTATGACAGCTATGCTCATT-3' (forward) and 5'-TCCTAAATTAGCGTCAAAAAT-3' (reverse) for expB. The thermal cycling parameters consisted of an initial denaturation at 95°C for 3 min followed by 30 cycles of denaturation at 95°C for 30 s, annealing at 55°C for 30 s, and extension at 72°C for 30 s, with an additional final extension step of 72°C for 2 min. PCR products were separated on 1.0% agarose gel with TAE buffer and visualized with ethidium bromide.

Results and Discussion

A total of 74 staphylococci from 74 dogs with superficial pyoderma were obtained. All strains were identified as S. pseudintermedius as previously described²³⁾. Molecular characteristics of the isolated S. pseudintermedius were investigated using several conventional PCR methods as previously described^{15, 25)}. As shown in Table 1, MRSP with mecA accounted for 70.3% (52/74) of the strains. This frequency was very high compared to previous surveillance data from other countries: 5.1% in the UK²⁶⁾, 29.3% in Korea²⁰⁾ and 47.8% in North China¹⁷⁾, and was similar to the previously reported 66.5% in Japan²⁷⁾. This indicates a high prevalence of MRSP in canine superficial pyoderma in Japan. Sixty-five (87.8%) out of 74 dogs had been treated with antimicrobials previously: 50 (96.2%) of 52 dogs infected with MRSP and 15 (68.2%) of 22 dogs infected with MSSP. Dogs infected with MRSP tended to have greater exposure to antibiotics than MSSP-infected dogs.

Table 1 Distribution of methicillin-resistant
S. pseudintermedius strains from canine pyoderma

	No. of strains (%)	
S. pseudintermedius	74 (100)	
Methicillin-susceptible	22 (29.7)	
Methicillin-resistant	52 (70.3)	
SCCmec-type III-like	33	
SCCmec-type V	18	
Nontypeable	1	

Table 2 Distribution of two exfoliative toxin genes among S. pseudintermedius strains

S. pseudintermedius	n	No. of positive strains (%)	
		expA	expB
Methicillin-susceptible	22	5 (22.7)	1 (4.5)
Methicillin-resistant	52	1 (1.9)	0 (0)

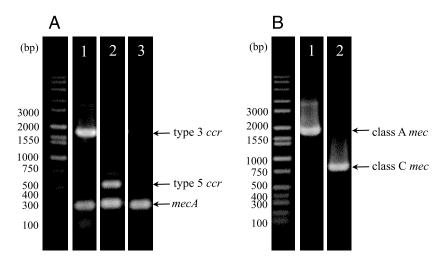


Fig. 1 Multiplex PCR analysis of *ccr* gene complex (A) and *mec* gene complex (B).

Lane 1, SCCmec type III-like MRSP; lane 2, SCCmec type V MRSP; lane 3, SCCmec nontypeable MRSP. (A) Upper bands in lanes 1 and 2 represent types 3 and 5 ccr genes, respectively. Lane 3 is a nontypeable strain that possesses mecA gene only. (B) Lanes 1 and 2 show single bands specific to class A and C mec gene complexes.

Previous use of antimicrobials may be associated with MRSP infection in dogs.

Furthermore, multiplex and duplex PCR assays for SCCmec typing revealed that 33 (63.4%) of 52 MRSP strains were type III-like, 18 (34.6%) belonged to type V, and only one was determined as nontypeable. The representative electrophoretic patterns of SCCmec type III-like (with fragments of both type 3 ccr and class A mec) and V (with fragments of both type 5 ccr and class C mec) are shown in Fig. 1.

To examine the presence of ET genes, amplification of the expA and B genes was conducted as shown in Fig. 2 (representative data). Table 2 shows the frequency of expA and B in the isolated S. pseudintermedius strains. The expA

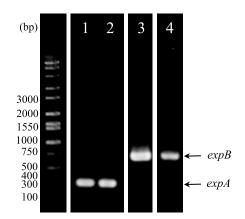


Fig. 2 PCR analysis of exfoliative genes.

Lane 1, a MSSP isolate carrying the *expA* gene; lane 2, positive control strain for *expA*; lane 3, a MSSP carrying the *expB* gene; lane 4, positive control strain for *expB*.

gene was detected at a rate of 22.7% (5/22) in methicillinsusceptible *S. pseudintermedius* (MSSP) strains in contrast to 1.9% (1/52) in MRSP strains; the latter MRSP strain was SCC*mec* type V. One MSSP isolate possessed the *expB* gene; however, the gene was not detected in any MRSP strain. There was no significant possession of ETs in *S. pseudintermedius* carrying the *mecA* gene, although they may be important virulence factors in canine pyoderma.

This study demonstrated that as many as 70.3% of 74 dogs diagnosed with superficial pyoderma had MRSP, implying the prevalence of MRSP in veterinary clinical practice in Japan. It is therefore important to rapidly, easily and feasibly determine *S. pseudintermedius* strains carrying the *mecA* gene and identify their genotypes not only to understand the epidemiological pattern but also for implementation of infection-control measures in veterinary clinical practice. To detect MRSP strains and their SCC*mec* types, we used several traditional PCR techniques that were complicated, cumbersome and time-consuming. We are currently designing improved multiplex PCR strategies.

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References

- Allaker, R. P., Lloyd, D. H., and Simpson, A. I., Occurrence of *Staphylococcus intermedius* on the hair and skin of normal dogs. *Res. Vet. Sci.* 52, 174-176 (1992).
- Allaker, R. P., Lloyd, D.H., and Bailey, R.M., Population sizes and frequency of staphylococci at mucocutaneous sites on healthy dogs. *Vet. Rec.* 130, 303-304 (1992).
- 3) Bannoehr, J. and Guardabassi, L., *Staphylococcus pseudintermedius* in the dog: taxonomy, diagnostics, ecology, epidemiology and pathogenicity. *Vet. Dermatol.* **23**, 253-266, e251-252 (2012).
- 4) Futagawa-Saito, K., Makino, S., Sunaga, F., Kato, Y., Sakurai-Komada, N., Ba-Thein, W., and Fukuyasu, T.,

- Identification of first exfoliative toxin in *Staphylococcus* pseudintermedius. FEMS Microbiol. Lett. **301**, 176-180 (2009).
- 5) Iyori, K., Hisatsune, J., Kawakami, T., Shibata, S., Murayama, N., Ide, K., Nagata, M., Fukata, T., Iwasaki, T., Oshima, K., Hattori, M., Sugai, M., and Nishifuji, K., Identification of a novel *Staphylococcus pseudintermedius* exfoliative toxin gene and its prevalence in isolates from canines with pyoderma and healthy dogs. *FEMS Microbiol. Lett.* 312, 169-175(2010).
- 6) Iyori, K., Futagawa-Saito, K., Hisatsune, J., Yamamoto, M., Sekiguchi, M., Ide, K., Son, W. G., Olivry, T., Sugai, M., Fukuyasu, T., Iwasaki, T., and Nishifuji, K., Staphylococcus pseudintermedius exfoliative toxin EXI selectively digests canine desmoglein 1 and causes subcorneal clefts in canine epidermis. Vet. Dermatol. 22, 319-326 (2011).
- Gortel, K., Campbell, K. L., Kakoma, I., Whittem, T., Schaeffer, D. J., and Weisiger, R. M., Methicillin resistance among staphylococci isolated from dogs. *Am. J. Vet. Res.* 60, 1526-1530 (1999).
- Bemis, D. A., Jones, R. D., Frank, L. A., and Kania, S. A., Evaluation of susceptibility test breakpoints used to predict *mecA*-mediated resistance in *Staphylococcus pseudintermedius* isolated from dogs. *J. Vet. Diagn. Invest.* 21, 53-58 (2009).
- Jones, R. D., Kania, S. A., Rohrbach, B. W., Frank, L. A., and Bemis, D. A., Prevalence of oxacillin- and multidrugresistant staphylococci in clinical samples from dogs: 1,772 samples (2001-2005). *J. Am. Vet. Med. Assoc.* 230, 221-227 (2007).
- Kania, S. A., Williamson, N. L., Frank, L. A., Wilkes, R. P., Jones, R. D., and Bemis, D. A., Methicillin resistance of staphylococci isolated from the skin of dogs with pyoderma. *Am. J. Vet. Res.* 65, 1265-1268 (2004).
- 11) Loeffler, A., Linek, M., Moodley, A., Guardabassi, L., Sung, J. M., Winkler, M., Weiss, R., and Lloyd, D. H., First report of multiresistant, mecA-positive Staphylococcus intermedius in Europe: 12 cases from a veterinary dermatology referral clinic in Germany. Vet. Dermatol. 18, 412-421 (2007).
- 12) Zubeir, I. E., Kanbar, T., Alber, J., Lammler, C., Akineden, O., Weiss, R., and Zschock, M., Phenotypic and genotypic characteristics of methicillin/oxacillinresistant *Staphylococcus intermedius* isolated from clinical specimens during routine veterinary microbiological examinations. *Vet. Microbiol.* 121, 170-176 (2007).

- 13) Descloux, S., Rossano, A., and Perreten, V., Characterization of new staphylococcal cassette chromosome *mec* (SCC*mec*) and topoisomerase genes in fluoroquinolone- and methicillin-resistant *Staphylococcus pseudintermedius*. *J. Clin. Microbiol.* **46**, 1818-1823 (2008).
- 14) Moodley, A., Stegger, M., Ben Zakour, N. L., Fitzgerald, J. R., and Guardabassi, L., Tandem repeat sequence analysis of staphylococcal protein A (*spa*) gene in methicillin-resistant *Staphylococcus pseudintermedius*. *Vet. Microbiol.* **135**, 320-326 (2009).
- 15) Perreten, V., Kadlec, K., Schwarz, S., Gronlund Andersson, U., Finn, M., Greko, C., Moodley, A., Kania, S. A., Frank, L. A., Bemis, D. A., Franco, A., Iurescia, M., Battisti, A., Duim, B., Wagenaar, J. A., van Duijkeren, E., Weese, J. S., Fitzgerald, J. R., Rossano, A., and Guardabassi, L., Clonal spread of methicillin-resistant *Staphylococcus pseudintermedius* in Europe and North America: an international multicentre study. *J. Antimicrob. Chemother.* 65, 1145-1154 (2010).
- 16) Ruscher, C., Lubke-Becker, A., Semmler, T., Wleklinski, C. G., Paasch, A., Soba, A., Stamm, I., Kopp, P., Wieler, L. H., and Walther, B., Widespread rapid emergence of a distinct methicillin- and multidrug-resistant *Staphylococcus pseudintermedius* (MRSP) genetic lineage in Europe. *Vet. Microbiol.* 144, 340-346 (2010).
- 17) Wang, Y., Yang, J., Logue, C. M., Liu, K., Cao, X., Zhang, W., Shen, J., and Wu, C., Methicillin-resistant *Staphylococcus pseudintermedius* isolated from canine pyoderma in North China. *J. Appl. Microbiol.* **112**, 623-630 (2012).
- 18) Onuma, K., Tanabe, T., and Sato, H., Antimicrobial resistance of *Staphylococcus pseudintermedius* isolates from healthy dogs and dogs affected with pyoderma in Japan. *Vet. Dermatol.* **23**, 17-22, e15 (2012).
- 19) Black, C. C., Solyman, S. M., Eberlein, L. C., Bemis, D. A., Woron, A. M., and Kania, S. A., Identification of a predominant multilocus sequence type, pulsed-field gel electrophoresis cluster, and novel staphylococcal chromosomal cassette in clinical isolates of *mecA*-containing, methicillin-resistant *Staphylococcus* pseudintermedius. Vet. Microbiol. 139, 333-338 (2009).
- 20) Youn, J-H., Koo, H. C., Ahn, K. J., Lim, S-K., and Park, Y. H., Determination of staphylococcal exotoxins,

- SCC*mec* types, and genetic relatedness of *Staphylococcus intermedius* group isolates from veterinary staff, companion animals, and hospital environments in Korea. *J. Vet. Sci.* **12**, 221 (2011).
- 21) Feng, Y., Tian, W., Lin, D., Luo, Q., Zhou, Y., Yang, T., Deng, Y., Liu, Y. H., and Liu, J. H., Prevalence and characterization of methicillin-resistant *Staphylococcus pseudintermedius* in pets from South China. *Vet. Microbiol.* 160, 517-524 (2012).
- 22) Chanchaithong, P., Perreten, V., Schwendener, S., Tribuddharat, C., Chongthaleong, A., Niyomtham, W., and Prapasarakul, N., Strain typing and antimicrobial susceptibility of methicillin-resistant coagulase-positive staphylococcal species in dogs and people associated with dogs in Thailand. J. Appl. Microbiol. 117, 572-586 (2014).
- 23) Sasaki, T., Tsubakishita, S., Tanaka, Y., Sakusabe, A., Ohtsuka, M., Hirotaki, S., Kawakami, T., Fukata, T., and Hiramatsu, K., Multiplex-PCR method for species identification of coagulase-positive staphylococci. *J. Clin. Microbiol.* 48, 765-769 (2010).
- 24) Zhang, K., McClure, J. A., Elsayed, S., Louie, T., and Conly, J. M., Novel multiplex PCR assay for characterization and concomitant subtyping of staphylococcal cassette chromosome *mec* types I to V in methicillin-resistant *Staphylococcus aureus*. *J. Clin. Microbiol.* 43, 5026-5033 (2005).
- 25) Kondo, Y., Ito, T., Ma, X. X., Watanabe, S., Kreiswirth, B. N., Etienne, J., and Hiramatsu, K., Combination of multiplex PCRs for staphylococcal cassette chromosome *mec* type assignment: rapid identification system for *mec*, *ccr*, and major differences in junkyard regions. *Antimicrob. Agents Chemother*. 51, 264-274 (2007).
- 26) Maluping, R. P., Paul, N. C., and Moodley, A., Antimicrobial susceptibility of methicillin-resistant *Staphylococcus pseudintermedius* isolated from veterinary clinical cases in the UK. *Br. J. Biomed. Sci.* **71**, 55-57 (2014).
- 27) Kawakami, T., Shibata, S., Murayama, N., Nagata, M., Nishifuji, K., Iwasaki, T., and Fukata, T., Antimicrobial susceptibility and methicillin resistance in *Staphylococcus* pseudintermedius and *Staphylococcus schleiferi* subsp. coagulans isolated from dogs with pyoderma in Japan. J. Vet. Med. Sci. 72, 1615-1619 (2010).