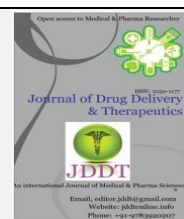




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Research Article

GC-MS analysis of bioactive compounds present in different extracts of rhizome of *Curcuma aeruginosa* Roxb.

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ABSTRACT

To analyze and characterize the chemical composition of the different crude extracts from the rhizome of *Curcuma aeruginosa* Roxb a medicinal plant. The air-dried rhizomes were powdered and subjected to Soxhlet extraction using solvent n-hexane and Supercritical fluid extraction. Then, each of the extracts was further subjected to gas chromatography-mass spectrometry. Qualitative determination of the different biologically active compounds from crude extracts of *C. aeruginosa* Roxb using gas chromatography-mass spectrometry revealed different types of high and low molecular weight chemical entities with varying quantities present in each of the extracts. These chemical compounds are considered biologically and pharmacologically important. Furthermore, the two different extracts SCF and n-hexane possess unique physicochemical characteristics. The two extracts possess major bioactive compounds that were identified and characterized spectroscopically. Thus, identification of different biologically active compounds in the extracts of *C. aeruginosa* Roxb warrants further biological and pharmacological studies.

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INTRODUCTION

Plants play a significant role in the prevention and treatment of diseases and can even prevent and reduce the adverse effects of conventional treatments ¹. They can be a source of chemical compounds of biological and pharmacological importance. History reveals that plants are sources of successful drugs, and will continuously be important for screening of new lead compounds ². An essential part in the investigation of a plant is the identification of the biologically active compounds present in plant leading to further biological and pharmacological studies ³⁻⁵. The search for new plant-derived chemicals should thus be a priority in current and future efforts toward sustainable conservation and rational utilization of biodiversity ⁶. The family Zingiberaceae comprises advanced monocot plants and is characterized by aromatic, non-tuberous and tuberous rhizomes, which have tremendous ethno medicinal properties.^{7,8} *Curcuma aeruginosa* was commonly known as 'Black turmeric' is a perennial underutilized herb of the

family Zingiberaceae. It usually grows well in moist deciduous forests.⁹ The plant has a characteristic rhizome flesh of bluish black color with a pungent smell and hot bitter taste, because of the presence of essential oil rich in camphor and starch. The rhizome is traditionally used in the treatment of hemorrhoids, leprosy, asthma, cancer, fever, wounds, vomiting, menstrual disorder, anthelmintic, aphrodisiac, gonorrhoeal discharges, and inflammation.^{10,11} The rhizome of *C. aeruginosa* is a promising source of potential anti-oxidants.¹² It is employed for making various cosmetic items and for sprains and bruises.¹³ Anti-androgenic effect of sesquiterpenes isolated from the rhizomes of *C. aeruginosa* had been reported.¹⁴ Due to its high medicinal value and indiscriminate harvest from the wild, the natural population has come down and according to International Union for Conservation of Nature (IUCN) report, the plant is in the critically endangered category.¹⁵ This paper mostly highlighted on the analysis and identification of bioactive compounds present in the plant extracts through GC-MS.

MATERIAL AND METHODS

Plant material: Plant material was collected in February 2015 from Waynad district of Kerala, India. Taxonomic identification of the plants was carried out by Dr. S John Britto, Director at the Rapinat Herbarium, St. Joseph's College, Tiruchirappalli, Tamilnadu, India. Voucher specimens (RHT 68570) are submitted at the Rapinat Herbarium. Fresh rhizomes of Black turmeric (*Curcuma aeruginosa*) was collected and cultivated in Vadakkencherry Holy Family garden.

Preparation of Plant Extract

The collected rhizomes were dried under the room temperature and powdered with a mechanical grinder and stored in air tight container.

Super Critical Fluid Extraction

SFE was carried out on the extraction system (model HA220-50-06, Nantong, China) which consists of a 5 L volume extractor, an HA220-50-06 controller, two syringe pumps (model 100 DX, Jiangsu, China), and a CO₂ cylinder. The operation pressure and temperature could reach up to 80 MPa and 45°C. The extraction pressure of the system was adjusted by a pressure regulator and the temperature was controlled by a thermostatic water bath. The parameters, namely, extraction pressure (P), temperature (T), dynamic extraction time (t), and flow rate of CO₂ (F), were studied and optimized.

For each extraction, about 1.0 kg of the prepared plant material was loaded into the extractor. Dense liquid ¹⁶ CO₂ is then pumped through a cylinder containing the material to be extracted. The extract-laden liquid is then pumped into a separation chamber where the extract is separated from the gas and the gas is recovered for re-use. The extracted substances were collected at the bottom of the separators.

Soxhlet method of hexane extraction

The 20 g of dried powder was subjected to Soxhlet extraction using hexane. The solvent (150 ml of hexane) is added to a round bottom flask (250ml volume), which is attached to a Soxhlet extractor and condenser on an isomantle. The rhizome powder is loaded into the thimble, which is placed inside the Soxhlet extractor. The sidearm is lagged with glass wool. The solvent is heated using the isomantle and will begin to evaporate, moving through the apparatus to the condenser. The condensate then drips into the reservoir containing the thimble. Once the level of the solvent reaches the siphon it pours back into the flask and the cycle begins again. The process runs for two days at 60°C.

GC-MS analysis

GC-MS analysis was performed on a Shimadzu (Tokyo, Japan) Make GCMS-TQ8030 with nonpolar Rxi 5Sil MS capillary column, full scan mode, injector mode-splitless, quadra pole mass selective detector(MSD), injection temperature 250°C, GC-MS interface temperature 250°C, the injection volume was 1µl. Helium was employed as the carrier gas, at a pressure of 57.5KPa; flow rate was 1ml/min. Mass spectra were detected at 70eV. Temperature programming was set as follows: column temperature was started from 60°C (held for 2 min) and linearly increased by 5°C/min to 200°C (held for 2min); after that, it was increased by 3°C/min to 220°C (held for 1 min); further, it was increased by 6°C/min. to 250°C (held for 7min). Total GC running time was 51.67 min. The components of the oil were identified by comparison of their mass spectra with those of the spectrometer database using NIST library (Shimadzu). The identifications were confirmed by

comparison of the fragmentation pattern and their retention indices with those reported in the literature.

RESULTS AND DISCUSSION

Bioactive compounds present in the SFE and Hexane extracts, since various parameters potentially affect the extraction process, the optimization of the experimental conditions is a critical step in developing an SFE method. Based on the previous knowledge of SFE, the extraction pressure, temperature, the dynamic extraction time and flow rate of CO₂ are usually considered as the most important factors of SFE. The optimal conditions for extraction of *C aeruginosa* by SFE were 40 MPa of pressure, 50°C of temperature, 1.5 h dynamic extraction time and 40 L/Hour (20Hz) flow rate of CO₂. Figure 1 and 2 depicts a GC-MS chromatograph of *C aeruginosa* obtained by SFE and Soxhlet method. The identification of the compounds was done by NIST as well as of compound, quality, retention times, molecular weight (MW) and relative content with those from literature data ¹⁷. The GC-MS analysis of compounds from *C aeruginosa* extracts was performed using a Rxi 5 Sil MS capillary column. In Table 1 (Fig:1) was reported the GC-MS data and the compounds identified in the extracts of *C aeruginosa* by SFE at the set of 40 MPa of pressure, 50°C of temperature, 1.5 h dynamic extraction time and 40 L/Hour (20Hz) flow rate of CO₂. Based on abundance, the major compounds present in the SCF extract were 5.beta.- Guaia-7(11),10(14)-dien-8.alpha.-ol,5,8-epoxy-(+)-(27.09%),7H-2,4a-Methanonaphthalen-7one, 1,2,3,4,5,6-hexahydro-1,1,5,5-tetramethyl-,(2S,4aR)-(-)-(6.31%), Boldenone(5.89%), (2E)-2-(4Methoxybenzylidene) Cyclohexanone (4.2%), 17.alfa.,21.beta.-28,30 Bisnorhopane (3.81%), 5.beta.-Guaia-7(11),10(14)-dien-8.alpha.-ol,5,8-epoxy-(+)-(3.28%), Nootkaton-11,12-epoxide(3%), Bis(2-ethylhexyl) phthalate(2.85%), (+)-2Bornanone (2.08%),2-(1-(Beta-d-glucopyranosyloxy)-1-methylethyl)-2,3-dihydro-7-oxo-7H furo(3,2-g)chromene, (R)-(2.6%), Tetracyclo [5.4.3.0(7,11)]tetradeca-2,5,10-trione, 1,4,6,14-tetramethyl-4-vinyl-(1.84%), Naphthalene, deca hydro-1,4a-dimethyl-2-methylene-(1.81%), 1,5,9-Cyclo dodecatriene, 1,5,9-trimethyl-(1.81%),2H-Cyclohepta [b]furan-2-one,6-[1-(acetyloxy)-3-oxobutyl]-3,3a,4,7,8,8a-hexahydro-7-methyl-3-methylene-(1.81%),as-Indacen-4(1H)-one,decahydro-(3a.alpha.,5a. beta., 8a.beta.,8b.beta.)-(1.42%),2,4-DIISO PROPENYL-1-METHYL-1-VINYLCYCLO HEXANE (1.35%), 1,2-Dimethyl-5-nitroadamantane(1.31%), Caryophyllene (1.29%), 5-Isoborneol(1.03%).

Table 2 (Fig; 2) depicts that biologically active chemical compound of n-hexane extract from *C aeruginosa* Roxb. rhizome the major compounds found in this were 5.beta.-Guaia-7(11),10(14)-dien-8.alpha.-ol, 5,8-epoxy-, (+)-(13.34%), 17-HYDROXYANDROSTA-1,4-DIEN-3-ONE(8.5%), 5.beta.-Guaia-7(11),10(14)-dien-8.alpha.-ol, 5,8-epoxy-, (+)-(8.47%), Naphthalene, decahydro-1,4a-dimethyl-2-methylene-(7.37%),(2E)-2-(4-Methoxybenzylidene) Cyclohexanone(6.94%), Cycloprop[e]indene-1a,2(1H)-dicarboxaldehyde, 3a,4,5,6,6a,6b-hexahydro-5,5,6b-trimethyl-, (1a.alpha.,3a.beta.,6a.beta.,6b.alpha.)(5.72%), 5,8-Dihydroxy-4a-methyl-4,4a,4b,5,6,7,8,8a,9,10-decahydro-2(3H)-phenanthrenone(4.77%), n-Hexadecanoic acid (4.02%), Cycloprop[e]indene-1a,2(1H)-dicarboxaldehyde, 3a,4,5,6,6a,6b-hexahydro-5,5,6b-trimethyl-, (1a.alpha.,3a. beta.,6a. beta.,6b.alpha.)(3.86%), Valerenal (3.56%), 5-Isopropenyl-3,6-Dimethyl-6-Vinyl-4,5,6,7-Tetrahydro-1-Benzofuran (2.72%), 5.beta.-Guaia-7(11),10(14)-dien-8.alpha.-ol, 5,8-epoxy-, (+)-(1.93%), (3aRS,4SR,6aRS,2'Z)-Hexahydro-3-methylidene-4-(pent-2'-enyl)-2H-cyclopenta[b]furan-2-one(1.91%), Cyclopropane, 1,1-Dimethyl-2-(2-Methyl -3-Buten-2-YL)- (1.83%), Thunbergol(1.83%), (-)-Isolongifolol,

pentafluoropropionate(1.68%), 7-Tetradecenal, (Z)-(1.59%), Bufo-20,22-dienolide, 14,15-epoxy-3,5,16-trihydroxy-, (3.beta.,5.beta.,15.beta.,16.beta.)-(1.44%), 1H-Cycloprop[e]azulene, 1a,2,3,5,6,7,7a,7b-octahydro-1,1,4,7-tetramethyl-, (+)-(1.41%), 6-Isopropenyl-4,8a-dimethyl-1,2,3,5,6,7,8,8a-octahydro-naphthalen-2-ol(1.22%), 17-Hydroxyandrost-1,4-DIEN-3-ONE(1.18%), 3,7-Cyclodeca dien-1-one, 3,7-dimethyl-10-(1-methylethylidene)-, (E,E)-(1.16%), 1,1,7-TRIMETHYL-4-METHYLENEDECAHYDRO-1H-CYCLOPROPA[E]AZULEN-7-OL(1.07%), Cycloprop[e]indene-1a,2(1H)-dicarboxaldehyde, 3a,4,5,6,6a,6b-hexahydro-5,5,6b-trimethyl (1a.alpha.3a.beta.6a.beta., 6b.apha (1.3%) , Isospathulenol(1.1%).

DISCUSSION

The usage of plants or herbs for medicinal purposes relies on their phytochemical composition that exhibits some interesting and specific biological activities. Different phytochemicals identified in the present study have been found to possess a wide range of biological activities. It has long been reported that Zingiberaceae families contained a number of phytochemicals such as terpenoids, flavonoids, phenylpropanoids and sesquiterpenes which exhibited anti-tumor activities^{18,19}. GC-MS analysis of the SCF and Hexane extract revealed the presence of various bioactive compounds. Direct solvent extraction using SCF revealed the occurrence of 68 compounds whereas Hexane allowed the extraction of 52 compounds respectively. All the major compounds from different extracts are biologically active molecules. They are considered to be a part of plants' defense systems, and as such have been included in a large group of protective molecules found in plants named 'phytoanticipins' or 'phytoprotectants'²⁰⁻²². There are several

reports in the chemical composition of the essential oil from the rhizome of *Curcuma* species²³⁻²⁵. Monoterpene and sesquiterpene hydrocarbons represented the most common chemical groups found in the essential oils from the rhizomes of the investigated *Curcuma* species²⁶. Several phytochemicals identified in the rhizome of *C. aeruginosa* in the present study have previously been reported by other researchers. These included compounds such as germacrone^{27,28}, cycloisolongifolene 8,9-dehydro-9-formyl, velleral and camphor²⁹, 1,8-cineole, pinene³⁰, curzerene and elemene³¹. However, most of the above mentioned compounds were detected in the essential oil of *C. aeruginosa* rhizome. While comparing the GCMS results of two extracts three compounds present in both extracts namely, 5.beta.-Guaia-7(11),10(14)-dien-8.alpha.-ol, 5,8-epoxy, (+)- (30.37% SCF and 23.74% Hexane) (2E)-2-(4METHOXYBENZYLIDENE) CYCLOHEXANONE (4.20% and 6.94%) Naphthalene, decahydro-1,4a-dimethyl-2-methylene-(1.81% and 7.37%) respectively in different concentrations. These major compounds have cytotoxic, antioxidant, antitumor antiangiogenic^{32,33} cytotoxicity^{34,35} cholesterol-lowering activity³⁶ use in agrochemicals, pharmaceuticals and perfumes³⁷. It has been reported that terpenes have antibacterial activity because of their bacteriostatic and bactericidal effects³⁸.

It was concluded that SCF and hexane extract of rhizome of *Curcuma aeruginosa* possess various potent bioactive compounds and is recommended as a plant of phytopharmaceutical importance. Further studies are needed to explore the potential compounds responsible for the biological activity from *Curcuma aeruginosa* for application in drug delivery, nutritional or pharmaceutical fields.

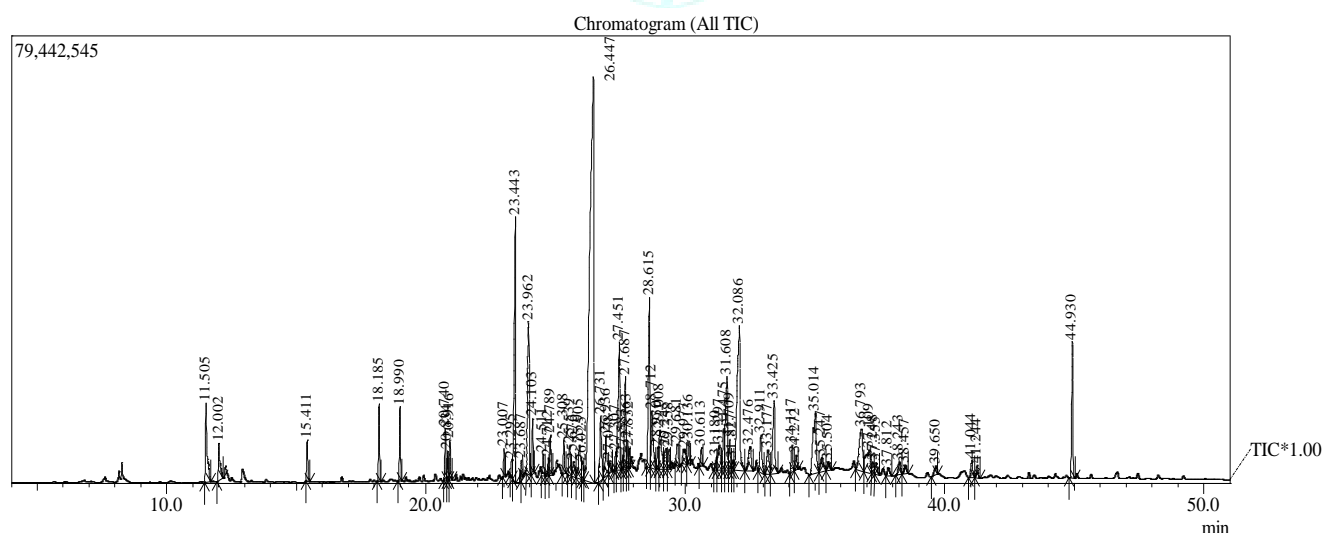


Figure 1: A typical chromatogram of the bioactive compounds present in SCF extract.

Table 1: Biologically active chemical compounds of SCF extract from *C aeruginosa* Roxb.

Sl. No.	RT	Peak area (%)	Name of the compound	Molecular formula	MW (g/mol)
1	7.601	0.07	3-Carene	C ₁₀ H ₁₆	136.24
2	8.15	0.23	D-Limonene	C ₁₀ H ₁₆	136.24
3	8.261	0.25	Eucalyptol	C ₁₀ H ₁₈ O	154.25
4	11.505	2.08	(+)-2-Bornanone	C ₁₀ H ₁₆ O	152.24
5	12.002	1.03	Isoborneol	C ₁₀ H ₁₈ O	154.25
6	12.251	0.5	1,7,7-TRIMETHYLBICYCLO[2.2.1]HEPTAN-2-OL	C ₁₀ H ₁₇ ClO	188.70
7	12.902	0.52	3-Cyclohexene-1-methanol, .alpha.,.alpha.,4-trimethyl-	C ₁₂ H ₂₀ O ₂	196.29

8	15.411	0.82	Bicyclo[2.2.1]heptan-2-ol, 1,7,7-trimethyl-, acetate, exo-	C ₁₉ H ₂₄ O ₂	284.40
9	16.728	0.09	.delta.-Elemene	C ₁₅ H ₂₄	204.36
10	18.185	1.35	2,4-DIISOPROPENYL-1-METHYL-1-VINYLCYCLOHEXANE	C ₁₅ H ₂₄	204.36
11	18.99	1.29	Caryophyllene	C ₁₅ H ₂₄	204.36
12	19.204	0.08	.gamma.-Elemene	C ₁₅ H ₂₄	204.36
13	19.715	0.09	1H-Cycloprop[e]azulene, decahydro-1,1,7-trimethyl-4-methylene-, (1aR,4aS,7R,7aR,7bS)-(-)-	C ₁₅ H ₂₄	204.36
14	19.89	0.12	.alpha.-Humulene	C ₁₅ H ₂₄	204.36
15	20.332	0.17	2-Isopropenyl-4a,8-dimethyl-1,2,3,4,4a,5,6,7-octahydronaphthalene	C ₁₅ H ₂₄	204.36
16	20.74	0.86	.beta.-Selinene	C ₁₅ H ₂₄	204.36
17	20.834	0.48	5-ISOPROPENYL-3,6-DIMETHYL-6-VINYL-4,5,6,7-TETRAHYDRO-1-BENZOFURAN	C ₁₅ H ₂₀ O	216.32
18	20.916	0.73	2-ISOPROPENYL-4A,8-DIMETHYL-1,2,3,4,4A,5,6,8A-OCTAHYDRONAPHTHALENE	C ₁₅ H ₂₄	204.36
19	21.146	0.13	.beta.-Bisabolene	C ₁₅ H ₂₄	204.36
20	21.314	0.07	1-ISOPROPYL-7-METHYL-4-METHYLENE-1,2,3,4,4A,5,6,8A-OCTAHYDRONAPHTHALENE	C ₁₅ H ₂₄	204.36
21	21.413	0.11	4-epi-cubedol	C ₁₅ H ₂₆ O	222.37
22	22.803	0.19	Spiro[4H-cycloprop[e]azulene-4,2'-oxirane], decahydro-1,1,7-trimethyl-, [1aR-(1a.alpha.,4.beta.,4a.alpha.,7.alpha.,7a.beta.,7b.		
23	23.007	0.49	(-)-5-Oxatricyclo[8.2.0.0(4,6)]dodecane,,12-trimethyl-9-methylene-, [1R-(1R*,4R*,6R*,10S*)]-	C ₁₅ H ₂₄ O	220.36
24	23.163	0.11	1,1,4,7-TETRAMETHYLDECAHYDRO-1H-CYCLOPROPA[E]AZULEN-4-OL	C ₁₅ H ₂₆ O	222.37
25	23.295	0.41	2-(4a,8-Dimethyl-2,3,4,4a,5,6,7,8-octahydro-2-naphthalenyl)-2-propanol	C ₁₅ H ₂₆ O	222.37
26	23.443	5.89	Boldenone	C ₁₉ H ₂₆ O ₂	286.42
27	23.62	0.05	Hexagermane	Ge ₆ H ₁₆ Si	479.99
28	23.64	0.04	ALLO AROMADENDRENOXID-(1)	C ₁₅ H ₂₄	204.36
29	23.687	0.23	N-(4-HYDROXYPHENYL)-N,N,N'-TRIMETHYLSULFAMIDE	C ₉ H ₁₄ N ₂ O ₃ S	230.28
30	23.962	3.28	5.beta.-Guaia-7(11),10(14)-dien-8.alpha.-ol, 5,8-epoxy-, (+)-	C ₁₅ H ₂₂ O ₂	234.34
31	24.103	0.94	isospathulenol	C ₁₅ H ₂₄ O	220.36
32	24.511	0.4	5.beta.-Guaia-7(11),10(14)-dien-8.alpha.-ol, 5,8-epoxy-, (+)-	C ₁₅ H ₂₂ O ₂	234.34
33	24.789	0.77	1-Naphthalenol, decahydro-1,4a-dimethyl-7-(1-methylethylidene)-, [1R-(1.alpha.,4a.beta.,8a.alpha.)]-	C ₁₅ H ₂₆ O	222.37
34	25.029	0.88	1H-Cycloprop[e]azulen-7-ol, decahydro-1,1,7-trimethyl-4-methylene-, [1a(1a.alpha.,4a.alpha.,7.beta.,7a.beta.,7b.alpha.)]-	C ₁₅ H ₂₄ O	220.36
35	25.308	0.61	(3E)-5-Isopropyliden-6-methyl-3,6,9-decatrien-2-one	C ₁₄ H ₂₀ O	204.31
36	25.539	0.77	3,7-Cyclodecadien-1-one, 10-(1-methylethenyl)-, (E,E)-	C ₁₃ H ₁₈ O	190.29
37	25.905	0.45	valerenol	C ₁₅ H ₂₄ O	220.36
38	26.447	27.09	5.beta.-Guaia-7(11),10(14)-dien-8.alpha.-ol, 5,8-epoxy-, (+)-	C ₁₅ H ₂₂ O ₂	234.15
39	26.731	1.81	Naphthalene, decahydro-1,4a-dimethyl-2-methylene-	C ₁₃ H ₂₂	178.32
40	26.936	0.92	5,8-Dihydroxy-4a-methyl-4,4a,4b,5,6,7,8,8a,9,10-decahydro-2(3H)-phenanthrenone	C ₁₅ H ₂₂ O ₃	250.34
41	27.295	0.46	Murolan-3,9(11)-diene-10-peroxy	C ₁₅ H ₂₄ O ₂	236.36
42	27.451	3.81	17.alpha.,21.beta.-28,30-Bisnorhopane	C ₂₈ H ₄₈	384.69
43	27.687	2.16	5-Azulenemethanol, 1,2,3,4,5,6,7,8-octahydro-.alpha.,.alpha.,3,8-tetramethyl-, [3S-(3.alpha.,5.alpha.,8.alpha.)]-	C ₁₅ H ₂₆ O	222.37
44	27.763	0.49	1,2,3,4-TETRAKIS(1-METHYLETHYLIDENE)CYCLOBUTANE	C ₁₆ H ₂₄	216.37
45	27.832	0.44	Norethynodrel	C ₂₀ H ₂₆ O ₂	298.43
46	28.615	4.2	(2E)-2-(4-METHOXYBENZYLIDENE)CYCLOHEXANONE	C ₁₄ H ₁₆ O ₂	216.28
47	28.712	1.81	1,5,9-Cyclododecatriene, 1,5,9-trimethyl-	C ₁₅ H ₂₄	204.36
48	28.925	0.49	4-Isopropyl-7,11-dimethyl-3,7,11-cyclotetradecatrienone	C ₁₉ H ₃₀ O	274.45
49	29.008	0.87	7-ISOPROPENYL-1,4-DIMETHYL-1,2,3,4,5,6,7,8-OCTAHYDROAZULENE	C ₁₅ H ₂₄	204.36
50	29.358	0.36	3-Oxatricyclo[20.8.0.0(7,16)]trianta-1(22),7(16),9,13,23,29-hexaene	C ₂₉ H ₄₂ O	406.65
51	29.681	0.66	2-[4-methyl-6-(2,6,6-trimethylcyclohex-1-enyl)hexa-1,3,5-trienyl]cyclohex-1-en-1-carboxaldehyde	C ₂₃ H ₃₂ O	324.51
52	30.136	0.45	.gamma.-Elemene	C ₁₅ H ₂₄	204.36
53	30.613	0.37	1,6-Dimethyl-9-(1-methylethylidene)-5,12-dioxatricyclo[9.1.0.0(4,6)]dodecan-8-one	C ₁₅ H ₂₂ O ₃	250.34
54	31.47	0.94	2,10-Dimethyl-4H,8H-benzo[1,2-b:3,4-b']dipyran-4-one		
55	31.608	3	Nootkaton-11,12-epoxide	C ₁₅ H ₂₂ O ₂	234.34
56	31.709	0.61	2(3H)-Benzofuranone, 6-ethenylhexahydro-6-methyl-3-methylene-7-(1-methylethenyl)-, [3aS-	C ₁₅ H ₂₀ O ₂	232.32

13	24.205	0.29	gama.-eudesmol	C ₁₅ H ₂₆ O	222.37
14	24.373	0.69	(+)-epi-Bicyclosesquiphellandrene	C ₁₅ H ₂₄	204.36
15	24.541	1.93	5.beta.-Guaia-7(11),10(14)-dien-8.alpha.-ol, 5,8-epoxy-, (+)-		
16	24.769	1.83	Thunbergol	C ₂₀ H ₃₄ O	290.49
17	24.977	1.07	1,1,7-TRIMETHYL-4-METHYLENEDECAHYDRO-1H-CYCLOPROPA[E]AZULEN-7-OL	C ₁₅ H ₂₄ O	220.36
18	25.113	0.44	2-Propen-1-ol, 3-(2,6,6-trimethyl-1-cyclohexen-1-yl)-	C ₁₂ H ₂₀ O	180.29
19	25.251	0.55	Azulene, 1,2,3,4,5,6,7,8-octahydro-1,4-dimethyl-7-(1-methylethylidene)-, (1S-cis)-	C ₁₅ H ₂₄	204.36
20	25.503	1.16	3,7-Cyclodecadien-1-one, 3,7-dimethyl-10-(1-methylethylidene)-, (E,E)-	C ₁₅ H ₂₂ O	218.34
21	25.666	0.5	1,4-Hexadien-3-one, 5-methyl-1-[2,6,6-trimethyl-2,4-cyclohexadien-1-yl]-	C ₁₆ H ₂₂ O	230.35
22	25.842	1.22	6-Isopropenyl-4,8a-dimethyl-1,2,3,5,6,7,8,8a-octahydro-naphthalen-2-ol	C ₁₅ H ₂₄ O	220.36
23	25.973	1.41	1H-Cycloprop[e]azulene, 1a,2,3,5,6,7,7a,7b-octahydro-1,1,4,7-tetramethyl-, (+)-	C ₁₅ H ₂₄	204.36
24	26.408	13.34	5.beta.-Guaia-7(11),10(14)-dien-8.alpha.-ol, 5,8-epoxy-, (+)-	C ₁₅ H ₂₂ O ₂	234.34
25	26.714	1.91	(3aRS,4SR,6aRS,2'Z)-Hexahydro-3-methylidene-4-(pent-2'-enyl)-2H-cyclopenta[b]furan-2-one		
26	26.934	4.77	5,8-Dihydroxy-4a-methyl-4,4a,4b,5,6,7,8,8a,9,10-decahydro-2(3H)-phenanthrenone	C ₁₅ H ₂₂ O ₃	250.34
27	27.48	7.37	Naphthalene, decahydro-1,4a-dimethyl-2-methylene-	C ₁₃ H ₂₂	178.32
28	27.693	5.72	Cycloprop[e]indene-1a,2(1H)-dicarboxaldehyde, 3a,4,5,6,6a,6b-hexahydro-5,5,6b-trimethyl-, (1a.alpha.,3a.beta.,6a.beta.,6b.alpha)	C ₁₅ H ₂₀ O ₂	232.32
29	27.876	0.87	1,5-epoxysalvial-4(14)-ene	C ₁₅ H ₂₄ O	220.36
30	28.13	1.68	(-)-Isolongifolol, pentafluoropropionate	C ₁₈ H ₂₅ F ₅ O ₂	368.39
31	28.611	6.94	(2E)-2-(4-METHOXYBENZYLIDENE)CYCLOHEXANONE	C ₁₄ H ₁₆ O ₂	216.28
32	28.971	3.86	Cycloprop[e]indene-1a,2(1H)-dicarboxaldehyde, 3a,4,5,6,6a,6b-hexahydro-5,5,6b-trimethyl-, (1a.alpha.,3a.beta.,6a.beta.,6b.alpha)	C ₁₅ H ₂₀ O ₂	232.32
33	29.287	1.3	Cycloprop[e]indene-1a,2(1H)-dicarboxaldehyde, 3a,4,5,6,6a,6b-hexahydro-5,5,6b-trimethyl-, (1a.alpha.,3a.beta.,6a.beta.,6b.alpha)	C ₁₅ H ₂₀ O ₂	232.32
34	29.63	1.83	CYCLOPROPANE, 1,1-DIMETHYL-2-(2-METHYL-3-BUTEN-2-YL)-		
35	29.89	1.44	Bufa-20,22-dienolide, 14,15-epoxy-3,5,16-trihydroxy-, (3.beta.,5.beta.,15.beta.,16.beta.)-	C ₂₈ H ₃₆ O ₈	500.59
36	30.43	0.85	3-METHYL-5-(2,6,6-TRIMETHYL-1-CYCLOHEXEN-1-YL)-1-PENTYN-3-OL	C ₁₅ H ₂₄ O	220.36
37	31.294	4.02	n-Hexadecanoic acid	C ₁₆ H ₃₂ O ₂	256.43
38	31.59	0.59	2(3H)-Benzofuranone, 6-ethenylhexahydro-6-methyl-3-methylene-7-(1-methylethenyl)-, [3aS-(3a.alpha.,6.alpha.,7.beta.,7a.beta.)]-	C ₁₅ H ₂₀ O ₂	232.32
39	31.959	3.56	Valerenal	C ₁₅ H ₂₂ O	218.34
40	32.25	0.4	1-Cyclohexene-1-crotonaldehyde, .alpha.,2,6,6-tetramethyl-	C ₁₄ H ₂₄ O	208.35
41	32.705	0.43	1,2-Dimethyl-5-nitroadamantane	C ₁₂ H ₁₉ NO ₂	209.29
42	33.188	0.61	2-(1-(Beta-d-glucopyranosyloxy)-1-methylethyl)-2,3-dihydro-7-oxo-7H-furo(3,2-g)chromene, (R)-	C ₂₀ H ₂₄ O ₉	408.40
43	34.736	0.22	1,4-Methanophthalazine, 1,4,4a,5,6,7,8,8a-octahydro-1,4,9,9-tetramethyl-, (1.alpha.,4.alpha.,4a.alpha.,8a.alpha.)-	C ₁₃ H ₂₂ N ₂	206.33
44	35.639	1.59	7-Tetradecenal, (Z)-	C ₁₄ H ₂₆ O	210.36
45	39.391	0.33	Behenic alcohol	C ₂₂ H ₄₆ O	326.61
46	41.933	0.1	Hexanedioic acid, bis(2-ethylhexyl) ester	C ₂₂ H ₄₂ O ₄	370.57
47	42.24	0.11	Heneicosane	C ₂₁ H ₄₄	296.58
48	44.114	0.13	1-Heptacosanol	C ₂₇ H ₅₆ O	396.74
49	44.274	0.13	Hexacosane	C ₂₆ H ₅₄	366.72
50	44.761	0.21	Bis(2-ethylhexyl) phthalate	C ₂₄ H ₃₈ O ₄	390.56
51	46.184	0.13	Hexacosane	C ₂₆ H ₅₄	366.72
52	48.468	0.1	Hexacosane	C ₂₆ H ₅₅	366.72

REFERENCES

1. Bachrach ZY. Contribution of selected medicinal plants for cancer prevention and therapy. *Acta Fac Medicae Naissensis* 2012; 29(3):117-23.
2. Atanasov AG, Waltenberger B, Pferschy-Wenzig EM, Linder T, Wawrosch C, Uhrin P, et al. Discovery and resupply of pharmacologically active plant-derived natural products: a review. *Biotechnol Adv* 2015; 3(8):1582-614.
3. Momin MA, Bellah SF, Rahman SM, Rahman AA, Murshid GM, Emran TB. Phytopharmacological evaluation of ethanol extracts of *Sida cordifolia* L. roots. *Asian Pac J Trop Biomed* 2014; 4(1):18-24.
4. Farid MM, Hussein SR, Ibrahim LF, Desouky MA, Elsayed AM, Oqlah AA, et al. Cytotoxic activity and phytochemical analysis of *Arum palaestinum* Boiss. *Asian Pac J Trop Biomed* 2015; 5(11):944-7.
5. Guo F, Feng L, Huang C, Ding H, Zhang X, Wang Z, et al. Phenylflavone derivatives from *Broussonetia papyrifera* inhibit the growth of breast cancer cells in vitro and in vivo. *Phytochem Lett* 2013; 6(3):331-6.
6. Phillipson, J.D. Plants as source of valuable products. In B.V. Charlwood, and M.J.C. Rhodes (eds.), *Secondary Products from Plant Tissue Culture*. Oxford: Clarendon Press, 1990; 1-21.
7. Chen IN, Chang CC, Ng CC, Wang CY, Shyu YT, Chang TL. Antioxidant and antimicrobial activity of Zingiberaceae plants in Taiwan. *Plant Food Human Nutr* 2008; 63:15-20.
8. Pari L, Murugan P. Changes in glycoprotein components in streptozotocin- nicotinamide induced type 2 diabetes: Influence of tetrahydrocurcumin from *Curcuma longa*. *Plant Food Hum Nutr* 2007; 62:25-29.
9. Nadkarni KM. *Indian material medica*. Bombay: Popular Prakashan; 1976.
10. Amalraj VA, Velayudhan KC, Muralidharan VK. A note on the anomalous flowering behaviour in *Curcuma caesia* (Zingiberaceae). *J Bom Nat Hist Soc* 1989; 86:278-279.
11. Sasikuma B. Genetic resources of *Curcuma*: Diversity, characterization and utilization. *Plant Genet Res* 2005; 3:230-251.
12. Dibakar C, Mitali G, Abhayap D, Palash M. Development of single node cuttings propagation techniques and evaluation of antioxidant activity of *Curcuma aeruginosa* Roxburgh. rhizome. *Int J Pharm Pharm Sci* 2013; 5:227-34.
13. Anonymous. *The Wealth of India*. Vol. 2. New Delhi: Council of Scientific and Industrial Research; 1962.
14. Suphrom N, Pumthong G, Khorana N, Waranuch N, Limpeanchob N, Ingkaninan K. Anti-androgenic effect of sesquiterpenes isolated from the rhizomes of *Curcuma aeruginosa* Roxb. *Fitoterapia* 2012; 83:864-871.
15. Khan SK, Karnat NM, Shankar D. India's foundation for the revitalization of local health traditions pioneering *in situ* conservation strategies for medicinal plants and local cultures. *Herb Gram* 2005; 68:34-48.
16. Patil, P.S. & Shettigar, R. An advancement of analytical techniques in herbal research *J. Adv. Sci. Res.*, 2010; 1(1):08-14.
17. Ouyang, Z., Yang, L., Su, S. L., Han, L., Xia, B., Wang, M., *Chin. J. Pharm. Anal.* 2007; 27:1333 - 1339.
18. Lakshmi S, Padmaja G, Remani P. Antitumour effects of isocurcumenol isolated from *Curcuma zedoaria* rhizomes on human and murine cancer cells. *Int J Med Chem* 2011.
19. Lai EY, Chyau CC, Mau JL, Chen CC, Lai YJ, Shih CF, et al. Antimicrobial activity and cytotoxicity of the essential oil of *Curcuma zedoaria*. *Am J Chin Med* 2004; 32:281-290.
20. Angle GR, Vimala B, Bala N. Antioxidant and antimicrobial activity of essential oil from nine starchy *Curcuma* species. *Int J Curr Pharm Res* 2012; 4(2):45-47.
21. Reanmongkol W, Subhadhirasakul S, Khaisombat N, Fuengnawakit P, Jantasila S, Khamjun A. Investigation the antinociceptive, antipyretic and anti-inflammatory activities of *Curcuma aeruginosa* Roxb. extracts in experimental animals. *Songklanakaraj J Sci Technol* 2006; 28(5):999-1008.
22. Sookchot T. Chemotaxonomy of medicinal and auspicious plants in Zingiberaceae sold at Ban Thum, Chiang Dao District, Chiang Mai Province. [Master's thesis]. Department of Biology, Faculty of Science, Chiang Mai University; 2005.
23. Akarchariya N. Chemical constituents and antimicrobial activity of some Zingiberaceous plants [Master's thesis]. Department of Pharmaceutical Science, Faculty of Pharmacy, Chiang Mai University; 2017.
24. Jarikasem S, Thubthimthed S, Chawanoraseth K, Suntornnatasat T. Essential oils from three *Curcuma* Species collected in Thailand. In: Bas,er, Franz G, Cañigueral S, Demirci F, Craker LE, Gardner ZE, editors. *Perspectives in natural product chemistry*. Chiang Mai: Wocamp III 2005: 37-41.
25. Theanphong O, Mingvanish W, Kirdmanee C. Chemical constituents and biological activities of essential oil from *Curcuma aeruginosa* Roxb. rhizome. *BHST* 2015; 13(1): 6-16.
26. Sookchot T. Chemotaxonomy of medicinal and auspicious plants in Zingiberaceae sold at Ban Thum, Chiang Dao District, Chiang Mai Province. [Master's thesis]. Department of Biology, Faculty of Science, Chiang Mai University; 2005.
27. Choudhury D, Ghosal M, Das AP, Mandal P. Development of single node cutting propagation techniques and evaluation of antioxidant activities of *Curcuma aeruginosa* Roxburgh rhizome. *Int J Pharm Pharm Sci* 2013; 5:227-234.
28. Dung NX, Tuyet NTB, Leclercq PA. Characterization of the leaf oil of *Curcuma aeruginosa* Roxb. from Vietnam. *J Essent Oil Res* 1995; 7:657-659.
29. Kamazeri TS, Samah OA, Taher M, Susanti D, Qaralleh H. Antimicrobial activity and essential oils of *Curcuma aeruginosa*, *Curcuma mangga*, and *Zingiber cassumunar* from Malaysia. *Asian Pac J Trop Biomed* 2012; 5:202-209.
30. Jarikasem S, Thubthimthed S, Chawanoraseth K, Suntornnatasat T. Essential oils from three *Curcuma* species collected in Thailand. *Acta Hort* 2005; 677:37-41.
31. Jirovetz L, Buchbauer A, Puschmanna C, Shafib MP, Nambiarb MKG. Essential oil analysis of *Curcuma aeruginosa* Roxb. leaves from South India. *J Essent Oil Res* 2000; 12:47-49.
32. Robinson TP, Ehlers T, Hubbard RB, Bai X, Arbiser JL, Goldsmith DJ, Bowena JP, *Bioorg. Med. Chem. Lett.*, 2003; 13:115-117.
33. Robinson TP, Ehlers T, Hubbard RB, Bai X, Arbiser JL, Goldsmith DJ, Bowena JP, *Bioorg. Med. Chem* 2005; 13:4007-4013.
34. Dimmock JR, Padmanilayam MP, Zello GA, Nienaber KH, Allen TM, Santos CL, De Clercq E, Balzarini J, Manavathu EK, Stables JP, *Eur J. Med. Chem* 2003; 38:169-177.
35. Modzelewska A, Pettit C, Achanta G, Davidson NE, Huang P, Khan SR, *Bioorg. Med. Chem* 2006; 14:3491-3495.
36. Piantadosi C, Hall IH, Irvine JL, Carlson GL, *J. Med. Chem* 1973; 16:770-795.
37. Ogawa M, Ishii Y, Nakano T, Irifune, S, *Jpn. Kohai Tokyo J.P* 63238034 A2 1988.
38. Uribe S, Ramirez J, Pena A. Effects of beta-pinene on yeast membrane functions. *J Bacteriol* 1985; 161(3):1195-1200.