#### brought to you b d by Journal of Drug De

#### Devi et al

Journal of Drug Delivery & Therapeutics. 2019; 9(1):293-302

Available online on 15.01.2019 at http://jddtonline.info



**Journal of Drug Delivery and Therapeutics** Open Access to Pharmaceutical and Medical Research

© 2011-18, publisher and licensee JDDT, This is an Open Access article which permits unrestricted non-commercial use, provided the original work is properly cited



# Open Open Access

**Review Article** 

# Prospects of Traditionally important Apocynaceae plants of India in Cancer Remediation

# Nirmala Devi \*1, <sup>2</sup>, Ajay Kumar Gupta <sup>3</sup>, Sunil K. Prajapati <sup>4</sup>

<sup>1</sup> Assistant professor, Institute of Pharmacy, Bundelkhand University, Jhansi (UP), INDIA

<sup>2</sup> Research Scholar at Monad University, Hapur (UP), INDIA.

<sup>3</sup> Associate Professor, University Institute of Pharmacy, Chhatrapati Sahuji Maharaj University, Kanpur (UP), INDIA

<sup>4</sup> Professor and Head, Pharmaceutical sciences, Institute of Pharmacy, Bundelkhand University, Jhansi (UP), INDIA

# ABSTRACT

Objectives: Apocynaceae Family plants in India had wide array of traditional uses and practised since years ago. This review aims to report selected plants of this possessing anticancer activity. Selected literature compiled from the search of electronic journals, books and encyclopedias etc. using search engines viz. Google, PubMed, Sciencedirect, GoogleScholar and SciFinder for all periods. The Dogbane family is includes atleast 150 genera and 1700 species. Around 25 genera and 50 species of the family reviewed here possess anticancer activity. The reason for this potential is due to: a) phytoconstituents b) poisonous constituents c) antimalarial activity and d) abundance of literature in traditional medicinal use. Folk medicinal uses and reported anticancer potential suggests that the Apocynaceae plants can be formulated or developed into lead compounds or novel drugs or multidrug complex for treatment of cancer. Detailed screening of each species has to be performed in 64 pannel cell lines, mechanistic study performed clearly and effectiveness of extracts, fractions or pure isolated compounds is to be compared.

Keywords: Apocynaceae; Traditional Medicines; cancer; anticancer plants.

Article Info: Received 27 Nov 2018; Review Completed 06 Jan 2019; Accepted 09 Jan 2019; Available online 15 Jan 2019

# Cite this article as:



Devi D, Gupta AK, Prajapati SK, Prospects of Traditionally important Apocynaceae plants of India in Cancer Remediation, Journal of Drug Delivery and Therapeutics. 2019; 9(1):293-302 DOI: http://dx.doi.org/10.22270/jddt.v9i1.2208

\*Address for Correspondence:

Nirmala Devi, Assistant professor, Institute of Pharmacy, Bundelkhand University, Jhansi (UP), INDIA

# **INTRODUCTION**

Cancer has become a curse to all age groups in which 5% cases are strongly hereditary. Cancer possesses heavy loads of economic burden on the families. GLOBOCAN registry estimated 14.1 million new cancer cases, 8.2 million cancer deaths and 32.6 million people living with cancer (within 5 years of diagnosis) in 2012 worldwide. ICMR warned India over 17.3 lakh new cases of cancer and 8.8 lakh deaths especially, cancers of breast, lung and cervix till 2020. A substantial portion of cancer cases and deaths could be prevented by broadly applying effective prevention measures, such as breast feeding, avoiding junk and tinned food, tobacco/alcohol control, vaccination, and the use of early detection tests <sup>1, 2</sup>.

Vast numbers of naturally-derived compounds from medicinal plants are targets for potential anticancer treatments. This review is an effort to highlight major apocynaceae plants which decrease growth of cancer or is being used as adjuvant with cancer treatments for patients who already have or have had cancer. Plants which are under trial or is researched for its anticancer potential is reported here. Apocynaceae plants are presented as a new ISSN: 2250-1177

hope for cancer patients as the plants have toxic secondary metabolites. The information disseminated through this review will help the researchers for generating family specific data for different type of cancers.

# **Methods**

Relevant literatures related to the terms "Apocynaceae", "Ethnopharmacology", "Cancer herbal drugs", and "Traditional" were obtained from different sources viz., PubMed, Sciencedirect and SciFinder databases.Medicinal literature was also searched from NISCAIR Online Periodical repository (NOPR), pubfacts and Google Scholar. The data specific to the Cancer Remediation and the Mechanism/Pathway of the particular plant/isolated phytoconstituents was collected and compiled. Research published till February 2018 is included in the study.

#### Research into herbal medicines for specific cancers

Cancer cells are immortal and exhibit exponential growth. Cancer cell mostly targets metabolic enzymes, gene regulator protein and cytoskeleton protein. An ideal anticancer drug should be able to induce apoptosis and angiogenesis;

#### Devi et al

blocking metabolic reactions of glucose transport, glycolysis, mitochondrial oxidative phosphorylation, and fatty acid synthesis and regulation of epigenetic processes. The drug should also be selective in action to malignant cell and should have minimum toxicity <sup>3</sup>,<sup>4</sup>.

Approving herbals as anticancer should have favorable pharmacokinetic properties (ADMETabsorption, distribution, metabolism, excretion and toxicity). Dose, dosage form and Safety are other serious issues. Since ancient times, nature has been a source of medicines to cure many deadly diseases. Clinically proven herbal anticancer drugs are: Taxanes(Docitaxel, paclitaxel (Taxol®),taxotere), vinca alkaloids( vinblastine, vincristine (Oncovin®), Etoposide, vinorelbine (Navelbine®)), teniposide (Vumon®), and various water-soluble analogs of camptothecin (Hycamtin®), brassinosteroids, Flavopiridol, polyphenol epigallocatechin-3-gallate, Pomiferin, histone deacetylase 9-bromo-noscapine;Bromelain, inhibitor, podophyllotoxins(topotecan, irinotecan) as well as epipodophyllotoxins, homoharringtonine, Elliptinium/ ellipticine. Olomucine/ roscovitine, combretastatins (Combretastatin A-4), Betulinic acid, Pervilleine-A, Silvesterol, Resveratrol and Piceatannol a hydroxylated version of Resveratrol and Pterostilbene a methoxylated version of Resveratrol, Coronaridine, Silvesterol. Thapsigargin, jatrophane, Curcuma longa, Ipomoea batatas, Centaurea schischkinii, and many others. Apomorphine hydrochloride, tiotropium bromide, nitisinone, galantamine hydrobromide, arteether are the drugs derived from plants used as approved drugs 5,6,7.

# **APOCYNACEAE PLANTS AS ANTICANCERS**

#### Apocynaceae as anticancer family 8,9

Apocyanceae family is the 5<sup>th</sup> largest family of medicinal plants. Toxic secondary metabolitesin the plants act against cellular level toxicity and Neoplasm. For eg. Reproductive system (R. vomitoria), respiratory disorder (tylophora indica), diabetes (catharanthus roseus), anti-inflammatory and analgesic (funtumia elastic, landolphia owariensis and picralima nitida). <sup>10-15</sup>.

Cardenolides, as a group of natural products that can bind to Na+/K+-ATPase with an inhibiting activity, are traditionally used to treat congestive heart failure. Recent studies have demonstrated that the strong tumor cytotoxicities of cardenolides are mainly due to inducing the tumor cells apoptosis through different expression and cellular location of Na+/K+-ATPase  $\alpha$ -subunits. The leaves, flesh, seeds and juices of numerous plants from the genera of Nerium, Thevetia, Cerbera, Apocynum and Strophanthus in Apocynaceae family, are the major sources of natural cardenolides. So far, 109 cardenolides have been isolated and identified from this family, and about a quarter of them are reported to exhibit the capability to regulate cancer cell survival and death through multiple signaling pathways. In this review, we compile the phytochemical characteristics and anticancer activity of the cardenolides from this family. Compounds belonging to the cardiac glycosides may stimulateCa2+and increase apoptosis in prostate cancer <sup>16</sup>.

Naturally occurring iridoids and secoiridoids in the family are reported as immunomodulators and adaptogens. Iridoids and secoiridoids show cardiovascular activity, antihepatotoxicity, choleretic activity, hypoglycemic activity, antiinflammatory activity, antispasmolytic activity, antitumor activity, antiviral activity and purgative action <sup>17</sup>.

Antitumour activity is reported in barks and root extracts of some Apocynaceae plants such as Allamanda, Alstonia, Calotropis, Catharanthus, Cerbera, Nerium, Plumeria and

#### Journal of Drug Delivery & Therapeutics. 2019; 9(1):293-302

Tabernaemontana. Latex from Himatanthus drasticus janaguba, Alstonia angustiloba, Calotropis gigantea, Dyera costulata, Kopsia fruticosa and Vallaris glabra are active against tumour and ulcers. Latex is rich in saponins, tannins, cardenolides and terpenoids and triterpenes such as Lupeol, betulin, betulinic acid and calenduladiol <sup>18, 23</sup>.

The search for improved cytotoxic agents (more potent, more selective, and less toxic) continues to be an important line in the discovery of modern anticancer drugs from natural source.

Indole alkaloids is abundant in plumerioideae subfamily; tribus alstonieae- alstonia, catharanthus, vinca, amsonsia. Tribus-tabernaemontanaeae-tabernaemontana,

tabernanthae, voacanga. Tribus rauwolfiaeae- kopsia, ochrosia, rauwolfia, vallesia. Also present is Sarpagine group of indole alkaloids <sup>19-22</sup>

Nature-derived antimalarials have been proved to act as anticancers. <sup>23-25</sup>.

#### Allamanda 26-28

The root extract of A. schottii was the most active of them. At 80  $\mu$ g/mL, the root extracts showed a cytostatic effect on K562, whereas at 400  $\mu$ g/mL, there was a strong cytotoxic effect. Similar cytostatic and cytotoxic effects were seen in the endothelial cells, but at lower doses. Parts of A. schottii were assayed against three different cultured cells: K-562, a cell line derived from Chronic Myeloid Leukemia in blastic crisis; BMEC, primary bone marrow endothelial cells; and HUVEC, primary human umbilical cord endothelial cells and MCF-7 lines.

Phytochemical investigation of different fractions and isolates has previous evidence of anticancer and antitumoral properties.

# Alstonia 29-34

The anticancer effect of various doses of an alkaloid fraction of Sapthaparna, Alstonia scholaris (ASERS), was studied in vitro in cultured human neoplastic cell lines (HeLa, HepG<sub>2</sub>, HL60, KB and MCF-7) and in Ehrlich ascites carcinoma bearing mice. The IC<sub>50</sub> was found to be 5.53, 25, 11.16, 10 and 29.76 µg/mL for HeLa, HePG<sub>2</sub>, HL60, KB and MCF-7 cells, respectively. The ASERS treatment resulted in a dose dependent elevation in the median survival time (MST) and the average survival time (AST) up to 240 mg/kg ASERS and declined thereafter. The surviving animals were healthy and disease free. The effect of ASERS was better than cyclophosphamide, which was used as a positive control, where all the animals succumbed to death by 40 days and the MST and AST were 19.5 and 18.3 days, respectively. The effective dose of 210 mg of ASERS was 3/10 of the LD<sub>50</sub> dose, which increased the MST and AST up to 54 and 49.5. Chemopreventive potential of Alstonia scholaris bark extract in DMBA-induced skin tumorigenesis in Swiss albino mice was assertive. A. venenata leaves showed considerable cytotoxicity towards neoplastic cells (DLA cells and EAC cells).

The rhazinilam-type alkaloids (rhazinicine, nor-rhazinicine, rhazinal, and rhazinilam) showed strong cytotoxicity toward human KB, HCT-116, MDA-MB-231, and MRC-5 cells.

#### Beaumontia 35

Five known cardenolides, digitoxigenin (1), oleandrigenin (2), digitoxigenin alpha-L-cymaroside (3), digitoxigenin beta-gentiobiosyl-alpha-L-cymaroside (4), and delta 16-digitoxigenin beta-D-glucosyl-alpha-L-cymaroside (5), were isolated from the stems of Beaumontia brevituba Oliver by cytotoxicity-directed fractionation monitored by a cultured

#### Devi et al

human lung cancer cell line. The cytotoxic activity of these compounds was evaluated with a panel of twelve human and murine cancer cell lines. The lignan glycoside, syringaresinol beta-D-glucoside, was obtained for the first time in the form of its levo-enantiomer.

# Carissa 36-40

C. opaca crude extract showed 78.5% inhibition against MCF-7 breast cancer cell line using MTT assay at 500 µg/mL. Fractions were tested at 200 µg/mL concentration and were more active than crude extracts. Chloroform fraction of C. opaca showed maximum inhibition 99% followed by ethyl acetate and methanol fraction of C. opaca exhibiting 96% and 94% inhibition, respectively. Also exhibited cytotoxicity at 800µg/mL on HeLa cancer cells. IC50 values ranged from 56.72 to 89.24  $\mu g/mL$  in MTT assay on HeLa, MCF-7, and HepG-2 cell lines besides MG-63.

# Cerbera 41-45

The cytotoxicity of the leaf of Cerbera odollam was investigated against two breast cancer cell lines (T47D and MCF7), two ovarian cancer cell lines (SKOV3 and CaOV3) and a normal (Vero) cell line. It showed potent anticancer activity with IC50 values of 17, 21, 28, 32 and 24 nM, respectively. Tanghinin, isolated from C.odollam exhibited cytotoxic activities against oral human epidermoid carcinoma (KB), human breast cancer cell (BC) and human small cells lung cancer (NCI-H187).

# Chonemorpha 46-49

MTT assay showed that the chloroform extract of callus has potent anticancer potential. The plant has a promising anticancer activity against human colon epithelium, lung carcinoma, and epidermoidal carcinoma cell lines. It was found to possess Topo as well as DNA polymerase inhibitory activity.

# Ervatamia 50-56

T. divaricata screened on cancer cell line (HeLa) and MTT assay was used to analyze the cell growth inhibition. The extract on Hep 2 cell line up to 7.8 µg/ml and that IC50 value on Hep 2 cell line was 112 µg whereas 94 µg for Vero cell line.

Six new bisindole alkaloids of the iboga-vobasine type, vobatensines A-F (1-6), in addition to four known bisindoles (8-11), were isolated from a stem bark extract of a Malayan Tabernaemontana corymbosa. Nine of these alkaloids (1-5, 8-11) showed pronounced in vitro growth inhibitory activity against human KB, PC-3, LNCaP, HCT 116, HT-29, MCF7, MDA-MB-231, and A549 cancer cells.

The wood and stem bark of Ervatamia heyneana (Apocynaceae) yielded 14 indole alkaloids and 3 triterpenoids. Six of these isolates, camptothecin (2), 9methoxycamptothecin (3), coronaridine (1), pericalline (25), heyneatine (18) and 10-methoxyeglandine- N-oxide (4) displayed cytotoxic activity.

The alkaloid fractions of ethanolic extract of E. coronaria showed cytotoxicity with LC50 values of 65.83 mg/ml in the BSL bioassay. The purified alkaloid fraction of E. coronaria exhibited highest cytotoxicity in HT-29, A-549 and MCF-7 cell lines with IC50 values of 32.5, 47.5 and 72.5 mg/ml, respectively.

# Holarrhena 57-60

In vitro cytotoxic potential of extracts (95% and 50% ethanolic extract and hot water extract at concentration of 100 microg/ml) from leaves of Holarrhena antidysenterica was evaluated against fourteen human cancer cell lines--A-ISSN: 2250-1177

#### Journal of Drug Delivery & Therapeutics. 2019; 9(1):293-302

549, COLO-205, DU-145, HeLa, HEP-2, IMR-32, KB, MCF-7, NCI-H23, OVCAR-5, SiHa, SK-N-MC, SW-620 and ZR-75-1 from nine different tissues (breast, colon, cervix, CNS, lung, liver, oral, ovary and prostate) using SRB assay cytotoxic activity was found in the chloroform soluble fraction of 95% ethanolic extract at 100 microg/ml; it inhibited the growth in the range of 71-99% of seven human cancer cell lines from five different tissues viz., OVCAR-5 (ovary), HT-29 (colon), SK-N-MC (neuroblastoma), HEP-2 (liver), COLO-205 (colon), NIH-OVCAR-3 (ovary) and A-549 (lung). The cytotoxic activity of chloroform soluble fraction was found to be higher than 5-flurouracil, adriamycin, mitomycin-c and paclitaxel (anticancer drugs used as positive controls).

# Ichnocarpus 61-65

In vitro anticancer activity of the residue from methanolic extract of roots of I. frutescens (MIF) and isolated triterpenes were evaluated by 3-(4, 5-dimethyl thiazol-2-yl)-2, 5diphenyltetrazolium bromide (MTT) assay using MCF-7, BEL-7402, SPC-A-1 and SGC-7901 cancer cell lines. MIF showed significant anticancer activity on four cancer cell lines with IC50 values 163.5±3.58, 156.3±2.95, 142.6±2.60 and 112.4±1.85 respectively.

It effectively inhibits in vitro proliferation of U-937 monocytoid leukemia and K-562 erythroleukemia cell lines. U-937 and K-562 cell lines.

# Kopsia arborea 66-69

Extracts of Kopsia fruticosa had the highest TAC against MCF-7 cells.

Ten new indole alkaloids of the aspidofractinine type, the leaf and stem-bark extract of the Malayan Kopsia singapurensis, kopsimalines A-E (1-5), kopsinicine (6), kopsofinone (7), and kopsiloscines H-J (8-10). Kopsimalines A (1), B (2), C (3), D (4), and E (5) and kopsiloscine J (10) were found to reverse multidrug-resistance in vincristineresistant KB cells, with 1 showing the highest potency [78]. Valpacrinine isolated from Malayan Kopsia arborea showed pronounced cytotoxic effects against KB and Jurkat cells (IC50 13.0 and 0.91 µM, respectively).

# Nerium 70-79

Research extract of Nerium oleander (Anvirzel) can induce cell death in human cancer can inhibit fibroblast growth factor-2(FGF-2) in prostate cancer cell lines (PC-3) and DU 145. Oleandrin may stimulate apoptosis through activation suppression of Nuclear Factor-kB (NF-kB), Activator protein-1 (AP-1), c-Jun NH2-terminal kinase inHela cell line. Oleandrin given after cells irradiated with 6 Gy of  $\gamma$ -ray, can increase the activation of caspase-3 in humanprostate carcinoma cell line (PC-3) thus inhibit the process of tumorigenesis and inflammatory processes. Oleandrin is also able to inhibit the growth of myeloma cells in a dose1,74 x 10-5 M, proportional to the dose of vincristine sulfate3,4 x 10-5 M. Three compounds, oleandrin, odoroside A and B evaluated against four human cell lines, normal human fibroblast cells (WI-38), malignant tumor cells induced from WI-38 (VA-13), human liver tumor cells (HepG2), and human lung carcinoma cells (A-549). Activity of Breastin, a defined extract isolated from the plant Nerium Oleander in 63 human cell lines swcreened; 31 / 63 cell lines investigated showed IC50 < 1.14  $\mu$ g/ml. e.g. Cisplatin, 5-Fluoruracil and Cyclophosphamide. The highest activity was seen in bladder, CNS, colon and NSC lung cancer cell lines as well as in pancreas and prostate models. In systematic combination studies Breastin increased the effect of the tubuline binders Paclitaxel, and Docetaxel in 4/6 cell lines, the alkylating agents Cyclophamide and Mitomycin, adriamycin and alimta.

# Ochrosia 80-82

Ellipticine, a cytotoxic plant alkaloid, is known to inhibit topoisomerase II in human breast MCF-7 cancer cells. Treatment of cells with ellipticine resulted in inhibition of growth, and G2/M phase arrest of the cell cycle. This effect was associated with a marked increase in the protein expression of p53 and, p21/WAF1 and KIP1/p27, but not of WAF1/p21. Ellipticine treatment increased the expression of Fas/APO-1 and its ligands, mFas ligand and sFas ligand, and subsequent activation of caspase-8. The mitochondrial apoptotic pathway amplified the Fas/Fas ligand death receptor pathway by Bid interaction. This effect was found to result in a significant increase in activation of caspase-9.

# Plumeria acuminate 83-86

The methanol extract of Plumeria acuminata leaves exhibited antitumor effect by modulating lipid peroxidation and augmenting antioxidant defense system in EAC bearing Swiss albino mice.

Cytotoxic compounds isolated from the aqueous extract of the bark( iridoid, plumericin and the lignin and liriodendrin), demonstrated general cytotoxic activity against murine lymphocytic leukemia (P-388) and a number of human cancer cell-types (breast, colon, fibrosarcoma, lung, melanoma, KB). Plumeria bracteata is most potent anticancer plant.

# Rauvolfia<sup>87-88</sup>

ß-carboline alkaloids from R.vomitoria are screened using WST-1 method against human LNCaP prostate cancer cell. Rauwolfia extract decreased in vitro cell growth in a dosedependent manner and induced the accumulation of G1 phase cells. PARP cleavage demonstrated that apoptosis was induced only at the highest concentration tested (500 µg/ml) which was confirmed by detection of cells containing subgenomic DNA. The expression of genes associated with DNA damage signaling pathway was up-regulated by Rauwolfia treatment, including that of GADD153 and MDG. The expression of a few cell cycle genes (p21, cyclin D1 and E2F1) was also modulated. These alterations were confirmed by RT-PCR. Tumor volumes were decreased by 60%, 70% and 58% in the groups fed the 75, 37.5 or 7.5 mg/kg Rauwolfia, respectively. Rauvolfia vomotoria has potent antitumor activity and in combination significantly enhances the effect of Carboplatin against ovarian cancer.

#### Strophanthus 89-91

All six new compounds cardenolide glycosides boivinides 1-6, as well as the four known cardenolide glycosides digitoxigenin 3-0-[ $\beta$ -d-glucopyrananosyl-( $1 \rightarrow 4$ )- $\alpha$ -lacofriopyranoside], corotoxigenin 3-0-β-d-boivinoside, 17αcorotoxigenin 3-0- $\beta$ -d-sarmentoside, and uzarigenin 3-0- $\alpha$ -lfrom Stropanthus, showed significant rhamnoside antiproliferative activity against the A2780 human ovarian cancer cell line, with boivinide A being the most active at  $IC50 = 0.17 \mu M.$ 

Strophanthus Wallichii has very good antitubercular, antioxidant and anticancer effect against clear cell renal cell carcinoma induced by DEN and Fe-NTA in male Wistar Albino rats.

# Thevetia 92-94

The cancer cell lines used in this study were human colorectal adenocarcinoma (HTB-38), lung carcinoma (HTB-177), prostate adenocarcinoma (HTB-81), and breast adenocarcinoma (HTB-22), whereas the normal cell lines used were human skin fibroblast (CCL-116) and Vero cell line (CCL-81). The T. peruviana methanolic extract exhibited

#### Journal of Drug Delivery & Therapeutics. 2019; 9(1):293-302

cytotoxic activity on four human cancer cell lines: prostate, breast, colorectal and lung, with values of  $IC_{50}$  1.91 ± 0.76,  $6.30 \pm 4.45$ 5.78 ± 2.12, and  $12.04 \pm 3.43 \,\mu g/mL$ respectively. The extract caused a significant reduction of cell motility and colony formation on all evaluated cancer cell lines. In addition, morphological examination displayed cell size reduction, membrane blebbing and detachment of cells, compared to non-treated cancer cell lines. The T. peruviana extract induced apoptotic cell death, which was confirmed by DNA fragmentation and AO/EB double staining. Cardiac glycosides (1-7) from seeds of T.peruviana, are cytotoxic toward cancer cell lines P15 (human lung cancer cell), MGC-803 (human gastric cancer cells), SW1990 (human pancreatic cancer cells), and normal hepatocyte cell LO2. They selectively inhibit the proliferation of cancer cell lines with IC<sub>50</sub> from 0.05 to 0.15  $\mu$ M.

#### Trachelospermum 95-97

The leaves and stems of T. jasminoids contain indole alkaloids like coronaridine, voacangine, apparicine, conoflorine, and 19-epi-voacangarine.

#### Vallaris 98-101

Sequential extracts of leaves, flowers and stems, and fractions and isolated compounds from dichloromethane (DCM) leaf extract of V. glabra were assessed for APF activity using the sulphorhodamine B (SRB) assay. Apoptotic effect of MDA-MB-231 cancer cells treated with DCM leaf extract of V. glabra was studied using Hoechst 33342 dye and caspase colorimetry. Both DCM extracts of leaves and flowers possessed broad-spectrum APF activity against HT-29, MCF-7, MDA-MB-231 and SKOV-3 cancer cells. Caspase colorimetry showed that the apoptotic effect involved activation of caspase-8, -9 and -3, but not caspase-6.

Thirteen cardenolide glycosides (1–13) were isolated from the CH2Cl2 and MeOH extracts of Vallaris glabra leaves their cytotoxic activity against human cervix adenocarcinoma, lung carcinoma, and colorectal adenocarcinoma cell lines checked. The two most potent compounds [2'-Oacetylacoschimperoside P (1) and oleandrigenin-3-0- $\alpha$ -l-2'-O-acetylvallaropyranoside (2)] exhibited IC50 values in the range of 0.03-0.07 µM

#### Vinca 102-105

Vinca alkaloids, Vinblastine, Vinorelbine, Vincristine and Vindesine are used clinically. Vinflunine is a synthetic vinca alkaloid which has been in use recently for the treatment of second-line transitional cell carcinoma of the urothelium and other malignancies.

Mauritianin, а flavonoid, enhanced the 12-0tetradecanoylphorbol-13-acetate (TPA), which suppressed delayed-type hypersensitivity reaction in mice, indicating that mauritianin may augment the resistance of the immune system to cancer. The 2, 3-dihydroxybenzoic acids from periwinkle showed a strong radical-scavenging activity, which is associated with a lower risk of cancer.

#### Wrightia 106-113

Antiproliferative activity of WTBM was evaluated against MDA-MB-231 and MCF-7 cancer cells by 3-(4,5dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide assay, colony formation, and Hoechst staining. In addition, (DPPH) radical scavenging activity and (ABTS) radical cation decolorization assay. Total phenolic content was assessed by Folin-Ciocalteu method. WTBM significantly suppresses colony formation and induces apoptosis in both MDA-MB-231 and MCF-7 cells as evident by morphological assessment, clonogenic. Mixtures of 1 and 2 (1:1 and 1:2), 2,

3, 4, 5, and 6 from the CH2Cl2 extracts of the leaves and twigs of W. pubescens (R.Br.) exhibited varying cytotoxic activities.

The ethanolic extract, subsequent hexane fractions and fraction F-4 of W. tomentosa inhibited the proliferation of human breast cancer cell lines, MCF-7 and MDA-MB231. The fraction F-4 obtained from hexane fraction inhibited proliferation of MCF-7 and MDA-MB-231 cells in concentration and time dependent manner with IC50 of 50µg/ml and 30µg/ml for 24h, 28µg/ml and 22µg/ml for 48h and 25µg/ml and 20µg/ml for 72h respectively. The fraction F-4 induced G1 cell cycle arrest, reactive oxygen species (ROS) generation, loss of mitochondrial membrane potential and subsequent apoptosis. Apoptosis is indicated in terms of increased Bax/Bcl ratio, enhanced Annexin-V positivity, caspase 8 activation and DNA fragmentation. The active molecule isolated from fraction F4, oleanolic acid and urosolic acid inhibited cell proliferation of MCF-7 and MDA-MB-231 cells at IC50 value of 7.5µM and 7.0µM respectively, whereas there is devoid of significant cell inhibiting activity in non-cancer originated cells, HEK-293. In both MCF-7 and MDA-MB-231, oleanolic acid and urosolic acid induced cell cycle arrest and apoptosis as indicated by significant increase in Annexin-V positive apoptotic cell counts.

Different extracts of leaf parts of Wrightia tinctoria has been studied against replication of HIV-1(IIIB) in MT-4 cells and HCV in Huh 5.2 cells. The ethanolic extract, subsequent hexane fractions and fraction F-4 of W. tomentosa inhibited the proliferation of human breast cancer cell lines, MCF-7 and MDA-MB-231 <sup>103</sup>. The ethanolic extract, subsequent hexane fractions and fraction F-4 of W. tomentosa inhibited the proliferation of human breast cancer cell lines, MCF-7 and MDA-MB-231. The fraction F-4 of Cancer cell lines, MCF-7 and MDA-MB-231. The fraction F-4 induced G1 cell cycle arrest, reactive oxygen species (ROS) generation, loss of mitochondrial membrane potential and subsequent

apoptosis. Apoptosis is indicated in terms of increased Bax/Bcl-2 ratio, enhanced Annexin-V positivity, caspase 8 activation and DNA fragmentation. The active molecule isolated from fraction F-4, oleanolic acid and urosolic acid.

# **DISCUSSION AND CONCLUSION**

With the exploration advancedment, human health is at stake with new resistant cases of existent diseases and Cancer is the one! New chemical entities (NCEs) fail to develop as a solid drug. Our Earth has a hidden treasure for all our sufferings in the form of Food, Clothing, Shelter and Medicine. Plants, soil, water, organisms and their remnants have abundant advantages. Medicinal plants are boon to almost all kinds of diseases. Apocynaceae plants are toxic and bitter plants but are sweet to our health. They have proved to a drug for all organ disease.

The integration of Ayurvedic wisdom with drug discovery also brings the need for a paradigm shift in the extraction process from sequential to parallel extraction. Bioassayguided fractionation of the identified plant may lead to standardized extract or isolated bioactive druggable compound as the new drug <sup>110</sup>.

Bioactivity-guided fractionation should be performed with a view to identifying novel compounds which will serve as candidates for preclinical testing. With the advent of combinatorial chemistry and high throughput screening, however, even greater progress may now be expected with natural product leads.

# ACKNOWLEDGEMENTS

We wish to thank the members of computer lab of central library of Bundelkhand and Monad University for providing the digital literature search using the stock of subscribed journals and books for free.

S.N.	Plant name	Local name	Traditional uses	Phytochemistry
1	Aganosma dichotoma	Malatilata	Emetic and anthelmintic bronchitis, leprosy, and skin diseases diseases of eyes, snake-bite analgesic, antidiarrheal, antidiabetic, anodyne, and sedative properties. Also used for paraplegia, sciatica and neuralgia.	Flavonoids such as rutin, robinin and other glycosides of kaempferol and quercetin, lupeol, beta setosterol, ursolic acid
2	Allamanda cathartica	Alokananda	Scabicide purgative anthelminthic, hyperthermia, laxative and emetic. It is used to cure malaria and jaundice. Antidote for poisoning. Ascites, ringworm infection.	Iridoid lactones (allamandin, allamandicin and allamdin), iridoid glycosides (plumieride coumarate and plumieride coumarate glucoside), and iridoid lactones (isoplumericin and plumericin).
3	Alstonia scholaris	Saptparni	Viper bite wound healing sedative and to treat hypertension.	monoterpenoid indole alkaloids, Alstonoside, a secoiridoid glucoside, Iridoids, coumarins, flavonoids, leucoanthocyanins, reducing sugars, simple phenolics, steroids, saponins and tannins, Alstonic acids A and B, triterpenoids
4	Anodendron paniculatum	bada dudheli mal	Jaundice/ hepatitis cuts and wounds Used to treat dyspepsia dementia antipyretic, antifertility and antirheumatic.	
	Apocynum cannabinum	Indian hemp	baby's cold, earache, headache, nervousness, dizziness, worms and insanity, cardiotonic, diaphoretic, diuretic, emetic and expectorant syphilis venereal warts hydrocephalus, urinary difficulties, dropsy, jaundice, liver problems, tumors, hemorrhoids, opthalmia and eye diseases.	Apocynin, apocynamarin, cymarin, and and rosin.
5	Beaumontia /jerdoniana/	Easter lily vine	Abortifacient, loss in libido, fractures, injury, and backache and leg pain caused by	Alkaloids, flavonoids, phenols, glycosides, steroids are present

Table 1: list of Indian apocynaceae plants (growing/cultivated) with Indian name, their traditional uses and phytochemistry.

	grandiflora		rhoumaticm	
6	Carissa carandas	Karaunda	rheumatism. Diarrhea, dysentery, cold, and fever, appetite. delivery pain muscular pain bronchitis and asthma	flavonoids, saponins, large amounts of cardiac glycosides, terpenes(carissone and carindone), tannins, urosolic acid isomer (carissic acid), Volatile constituents, polyphenols coumarin, pentacyclic triterpenoids, α-amyrin and β-sitosterol, carandinol, betulinic acid, Carisol, carinol, ascorbic acid.
7	Chonemorpha fragrans/grand iflora	Moorva	Used gynecological problems skin diseases, leprosies, syphilis, leprosy, dyspepsia, flatulence, colic constipation, helminthiasis, hyperdipsia, cardiac debility, diabetes, jaundice, bronchitis, and intermittent fever.	Campothecin, chonemorphine, funtumafrine, japindine, baurenolacetate, β-sitosterol and taraxasterol
8	Ervatamia/tab ernaemontana divaricata	Tagaar / chandni	Jaundice. Chronic herpes. Rheumatism. Lymph node enlargement.mastitis, tonsillitis, and mumps whitlow, cuts, and wounds. Scabies intestinal worms throat pain and phlegm erysipelas and eczema. Tonsillitis, pharyngitis, and laryngitis wounds, snake/scorpion bite and rheumatism.	9-methoxy camptothecin coronaridine, pericalline, heyneatine and 10- methoxyeglandine- N-oxide Voacamine, Apparicine, Vobasine, Ibogaine, Conophylline, Tabernaemontamine, Voacangine, Voacristine,
9	Holarrhena antidysentrica	Kutaja/dudhi	Diarrhea, stomachache, and leukorrhea increase milk antidysentric chronic chest complaints, spleen diseases, jaundice, bilious, and calculi toothache anthelmintic menstrual cycle diabetes management rheumatic pain, used for acne treatment.	Conessimine, conessine holafrine, holarrhenine, holarrhetine, holarrhimine, kurchicine, Conamine, conarrhimine, conessidine, conimine conkurchine, conkurchinine holarrhine, holarrhessimine holarrhidine, kurchine, isoconessimine, kurchamine, lettocine, Antidysentericine
10	Ichnocarpus frutescense	Sariva/Siamla ta/dhudhilata	fevers, gout, rheumatism, arthritis, epilepsy, venereal diseases, herpes, and skin diseases dysentery, measles, splenomegaly, and tuberculosis antidysentric, antipyretic, demulcent, diaphoretic, and hypoglycemic rheumatic pain. Improve memory power. Jaundice. galactogogue diuretic and diaphoretic treatment of skin eruptions	Phenylpropanoids, phenolic acids, coumarines, flavanoids, sitosterol and sitosterol palmitate.α-amyrin, and its acetates, lupeol and its acetates, flavones (apigenin and luteolin), glycoflavones (vitexin and isovitexin, proanthocyanidin and phenolic acids), vanillic, syringic and synapic acid, protocatechuic acid, Ursolic acid acetate, kaemferol, kaemferol-3- galactoside (trifolin), apigenin, luteolin, protocatechuic acid, quercetin and quercetin-3-O-D-glucopyranoside.
11	Kopsia fruticosa	Shrub vinca	Central nervous system (CNS) effects syphilis and has cholinergic malaria. Antimicrobial, antifungal, and cardiac effects	Kopsine, fruticosine and fruticosamine, Kopsamine aspidofractinine, kopsinine kopsiflorine, kopsilongine, kopsaporine, kopsingarine, kopsingine, venalstonine derivatives(venacarpines A and B), dioxokopsan derivative (kopsorinine), novel indole alkaloids, triterpenoids
12	Nerium indicum	Kaner	Abortifacient; scabies with itching sensation and eczema septic carbuncles, leprosy piles easy delivery.warts and ringworm. Impetigo for chronic ulcers antidote to snake bite rubbed on body in allergy, headache, aphrodisiac malaria and respiratory problems ear pain bad breath and toothache leukorrhea and menorrhagia.	galacturonic acid, two aristolochic acid derivatives and 3-aristolactam derivatives, two pentacyclic triterpenoids, Cardiac glycosides (kaneroside and neriumoside), digitoxigenin and uzarigenin glycosides oleanderigenin glycosides. Adynerin, flavonoid glycosides (quercetin and kaempferol)
13	Ochrosia elliptica/oppos itifolia		Used in gynecological disorders	ellipticine and its derivatives, 9-methoxy ellipticine, retellipticine, ellipticiniums
14	Parameria laevigata		rheumatism, nephritis, menses emmenagogue cuts lacerations dysentery, tuberculosis, shrink the uterus after delivery, stomachic,	
15	Parsonsia alboflavescens e		leg swellings, disinfectant, tuberculosis, vulnerary febrifuge, rheumatism, and kidneys	
16	Plumeria rubra	Kath- champa	Malaria, Leprosy, antiherpetic, venereal infections, Rheumatism, and abdominal tumors purgative, cardiotonic, diuretic, hypotensive bronchitis, cholera, cold, and cough antipyretic, antifungal.	amyrins, $\beta$ sitosterol, scopoletin, iridoids, Plumericin, isoplumericin, plumeride, coumerate, geraniol, citronellol, farnesol and phenylethyl lupeol nanoate, allamcin, and allamandin, fulvoplumerin and Rubrinol; Nerolidols, naphathalene, linalool, quercetin and kaempferol, benzyl salicylate, benzyl benzoate.

	Rauvolfia Sarngandha Stimulate uterine contraction in case of De			
17	Rauvolfia serpentina	Sarpgandha	Stimulate uterine contraction in case of difficult delivery, stomachache, muscular and rheumatism pain, cough and cold, skin disease cure mental disorders high blood pressure ulcer and clear intestinal worms. Snakebite, insect sting, and animal bite. Stomach distress malaria respiratory problems.	Deserpidine, reserpiline, reserpine, reserpinine rescinnamine, ajmalicine sarpagine, serpentine, yohimbine, ajmaline, isoreserpiline, connescine, corynanthine, desmethoxyreserpine raujemidine, raunescine, rauwolscine, recanescine, tetraphyllicine, tetraphylline, sandwicine, micranthine serpentidine.
18	Strophanthus wallichii		heart stimulant and to treat injury and snake bites diuretic	Cardiac glycosides
19	Thevetia nerilifolia/peru viana	Peeli Kaner	Abortifacient, purgative, rheumatism, dropsy, intermittent fevers violent emetic, hemorrhoids snake bite skin complaints.	Cardiac glycosides (triosides or monosides type), adigitoxigenin, or cannogenin (the 19-oxo form of digitoxigenin) or cannogenol (the 19-oxy form of digitoxigenin), Triosides: Thevetin, 2'-O- acetyl cerberoside, Monosides (neriifolin), cerberin ( 2'-O acetylneriifolin), peruvoside, theveneriin ( ruvoside) and perubosidic acid (perusitin).
20	Trachelosperm um asiaticum	Star Jasmine	Restorative and tonic. Analgesic, antibacterial, antispasmodic, depurative, emmenagogue, febrifuge, cardiotonic and hemostatic.	E-nerolidol and phellandrene trans- linalool oxide and citronellol
21	Urceola micrantha		Treatment of infantile paralysis, rheumatalgia, injury, and fractures.	
22	Vallaris solanaceae	Choudhari Bel	Ringworm infection, eczema, cut, sores, and wounds bite fixing teeth, applied to wounds and soresleprosy, sprue, dyspnea, piles/hemorrhoids bone fracture Hanthi paon.	cardiac glycosides; acoschimperoside P, mono-O-acetylvallaroside,mono-O- acetylsolanoside, mono-O-acetylaco- schimperoside P, vallaroside, vallarosolanoside, solanoside and 16- deace- tyl-16-anhydroacoschimperoside P, O- Palmitic, oleic and linoleic acids.β- sitosterol, β-amyrin and ursolic acid.
23	Vinca rosea	Sankhpushpi	malaria, dengue fever, diarrhea, diabetes, cancer, and skin diseases menorrhagia/leukorrhea indigestion, dyspepsia, dysentery, toothache purgative and toothache. Lower blood pressure menstrual complaint/leukorrhea, headache diabetes antiatherosclerotic.	vincanidine vincanine, vincamajoreine, vincamajorídine, isovincamine, perivincine, vincamine, vincaminorine, vinine, ajmalicine, catharanthine, leurosine, perivine, vincaleucoblastine, vinceine, (raubasine), lochnerine, lochnericine, vindoline, vinblastine, vinflunine, serpentine,vincristine,Vindesine and vinorelbine (semisynthetic derivatives of vinblastine), caffeoylquinic acid, flavonol glycosides, anthocyanins.
24	Wrightia tinctoria	Pala indigo/indraj ao/dhudla	Cures diseases of pittam and vatam, skin diseases, eczema, dysentery, psoriasis, venereal diseases, stringent, anthelmintic, stomachic, antipyretic, tonic, antidysenteric, diarrhea, piles, leprosy, worm Infestation, thirst, pain, diarrhea. Used for renal complications, menstrual disorders and amebic dysentery	Lupeol, stigmasterol campetosterol, Indigotin, indirubin, tryptanthrin, isatin, anthranillate and rutin Triacontanol, Wrightial, cycloartenone, cycloeucalenol, $\beta$ -amyrin, Alpha-Amyrin, $\beta$ - sitosterol, 14 $\alpha$ -methylzymosterol. Four uncommon sterols, desmosterol, clerosterol, 24-methylene-25- methylcholesterol, 24- dehydropollinastanol and Triterpinoids.

#### REFERENCES

1. Mallath MK et. al.; The growing burden of cancer in India: epidemiology and social context. THE LANCET Oncology. 2014, 15(6):e205-e212.

2. World Health Organisation. The World Health Organisation's Fight against Cancer: Strategies that prevent, cure and care. WHO Press; Geneva: 2007.

3. Gibbs J.B., Anticancer drug targets: growth factors and growth factor signaling, J. Clin Invest, 2000; 105:9-13.

4. Pratheeshkumar P, Sreekala C, Zhang Z. et al. Cancer prevention with promising natural products: mechanisms of action and molecular targets. Anticancer Agents Med Chem. 2012; 12:1159-1184.

5. Greenwell M, Rahman P.K.S.M.. Medicinal Plants: Their Use in Anticancer Treatment. Int J Pharm Sci Res. 2015 Oct 1; 6(10):4103–4112.

6. Solowey E, Lichtenstein M, Sallon S, Paavilainen H, Elaine Solowey, and Haya Lorberboum-Galski. Evaluating Medicinal Plants for Anticancer Activity. Hindawi Publishing Corporation. Scientific World Journal Volume 2014.

7. Cragg GM, Newman DJ. Plants as a source of anti-cancer agents. Journal of Enthnopharmacology. 2005; 100:72–79

8. Endress ME, Bruyns PV. A revised classification of the Apocynaceae. Bot Rev. 2000; 66:1–56.

9. Marcy J. Balunas, A. Douglas Kinghorn. Drug discovery from medicinal plants. Life sciences; 2005; 78(5):431-441.

10. Devi N, Gupta AK, Prajapati SK. Indian tribe's and villager's health and habits: Popularity of apocynaceae plants as medicine. International Journal of Green Pharmacy (IJGP). 2017; (Suppl)11(2):S256.

11. Norman R. Farnsworth, Cynthia J. Kaas. An approach utilizing information from traditional medicine to identify tumor-inhibiting plants. Journal of Ethnopharmacology. 1981, 3(1):85-99.

12. Gordaliza M. Natural products as leads to anticancer drugs. Clinical and Translational Oncology. 2007; 9(12):767–776.

13. Jain R, Jain SK. Screening of in vitro cytotoxic activity of some medicinal plants used traditionally to treat cancer in Chhattisgarh state, India. Asian Pacific Journal of Tropical Biomedicine. 2011; 1(2) Supplement:S147-S150.

14. Suffredini IB, Varella D, de Oliviera AA, Younes RN. In vitro anti-HIV and antitumor evaluation of Amazonian plants belonging to the Apocynaceae family. Phytomedicine. 2002; 9(2):175.

15. Siu Kuin Wong, Yau Yan Lim, Noor Rain Abdullah, Fariza Juliana Nordin. Antiproliferative and phytochemical analyses of leaf extracts of ten Apocynaceae species. Pharmacognosy Res. 2011; 3(2):100–106.

16. Shiyuan Wen, Yanyan Chen, Yunfang Lu, Yuefei Wang, Liqin Ding, Miaomiao Jiang. Cardenolides from the Apocynaceae family and their anticancer activity. Fitoterapia. 2016; 112:74-84.

17. Ghisalberti, E.L. Biological and pharmacological activity of naturally occurring iridoids and secoiridoids. Phytomed.1998, 5:147-163.

18. Dina F. El-Kashef, Ashraf N. E. Hamed, Hany E. Khalil, Mohamed S. Kamel. Triterpenes and sterols of family Apocynaceae (2013-1955), A review. Journal of Pharmacognosy and Phytochemistry 2015; 4(2):21-39

19. Gupta AP, Pandotra P, Kushwaha M, Khan S, Sharma R, Gupta S. Chapter 9 - Alkaloids: A Source of Anticancer Agents from Nature. Studies in Natural Products Chemistry. 2015, 46:341-445.

20. Raffauf RF, Flagler MB. Alkaloids of the Apocynaceae 1960; 14:37-55

21. Jin-Jian Lu, Jiao-Lin Bao, Xiu-Ping Chen, Min Huang, Yi-Tao Wang. (2012) Alkaloids Isolated from Natural Herbs as the Anticancer Agents. Evidence-Based Complementary and Alternative Medicine. 2012, 1-12.

22. Naira Fernanda Zanchett Schneider 1, Claudia Cerella 2, 3, Cláudia Maria Oliveira Simões 1 and Marc Diederich. Anticancer and Immunogenic Properties of Cardiac Glycosides. Molecules 2017, 22(11):1932.

23. Robert Duffy, Christine Wade, Raymond Chang. Discovery of anticancer drugs from antimalarial natural products: a MEDLINE literature review. Drug Discovery Today. 2012, (17–18):942–953.

24. Kundu CN, Das S, Nayak A, Satapathy SR, Das D, Siddharth S. Antimalarials are anti-cancers and vice versa - one arrow two sparrows. Acta Trop. 2015; 149:113-27.

25. Wong SK, Yan Lim Y, Abdullah NR, Nordin FJ. Assessment of antiproliferative and antiplasmodial activities of five selected Apocynaceae species. BMCComplementary and Alternative Medicine. The official journal of the International Society for Complementary Medicine Research (ISCMR), 2011; 11:3.

26. Kupchan S.M., Dessertine A.L., Blaylock B.T., Bruan R.F., 1974. Isolation and structural elucidation of allamandin, an antileukemic iridóide lactone from Allamanda cathartica. J. Org. Chem. 1974; 39(21):2477-2484.

27. de F Navarro Schmidt D, Yunes RA, Schaab EH, Malheiros A, Cechinel Filho V, Franchi GC Jr, Nowill AE, Cardoso AA, Yunes J.Evaluations of the anti-proliferative effect the extracts of Allamanda blanchetti and A. schottii on the growth of leukemic and endothelial cells. J Pharm Pharmaceut Sci. 2006; 9(2):200-208.

28. Fabiana G. Nascimentoa ,Amanda Faquetia , Jessica F. Wilhelma , Carolina Wittkowskia , Folvi D. Tomczaka , Sheila L. Borgesa , Rosendo A. Yunesb , Gilberto C. Franchi Jr.c , Alexandre E. Nowillc . Seasonal influence and cytotoxicity of extracts, fractions and major compounds from Allamanda schottii. Revista Brasileira de Farmacognosia Rev. bras. farmacogn. 2014, 24(5).

29. Keawpradub N, Eno-Amooquaye E, Burke PJ, Houghton PJ. Cytotoxic activity of indole alkaloids from Alstonia macrophylla. Planta Med. 1999; 65:311–5

30. Jahan S, Chaudhary R, Goyal PK. Anticancer Activity of an Indian Medicinal Plant, Alstonia scholaris, on Skin Carcinogenesis in Mice. Integrative Cancer Therapies. 2009, 8(3):273-279.

31. Ganesh Chandra Jagetia, Manjeshwar Shrinath Baliga. Treatment with Alstonia scholaris Enhances Radiosensitivity In vitro and in vivo. Cancer Biotherapy & Radiopharmaceuticals. 2004: 917-929.

32. Baliga MS. Alstonia scholaris Linn R Br in the Treatment and Prevention of Cancer: Past, Present, and Future. Integrative Cancer Therapies. 2010, 9(3):261–269.

33. Keawpradub N., Houghton PJ, Eno-Amovquay E., Burke PJ Activity of extracts and alkaloids of Thai Alstonia species against human lung cancer cell lines. Planta Med. 1997; 63:97-101.

34. Jagetia GC, Baliga MS. Evaluation of anticancer activity of the alkaloid fraction of Alstonia scholaris (Sapthaparna) in vitro and in vivo. Phytother Res. 2006; 20:103–9

35. Kaneda N, Chai H, Pezzuto JM, AKinghorn AD, N. Farnsworth NR, Tuchinda P, Udchachon J, T. Santisuk , V. Reutrakul. Cytotoxic activity of cardenolides from Beaumontia brevituba stems. Planta Med. 1992.

36. Gupta P, Bhatnagar I, Se-Kwon Kim, Verma AK, Anubhuti Sharma. In-vitro cancer cell cytotoxicity and alpha amylase inhibition effect of seven tropical fruit residues. Asian Pacific Journal of Tropical Biomedicine. 2014; 4(2):S665–S671.

37. Bodakhe SH.; Devi N; Gupta S K.; Namdeo K.P.; Jain S. K. Hepatoprotective Activity of Carissa carandas Linn. fruit ethanolic extract in carbon tetrachloride intoxicated rats. Advances in Pharmacology & Toxicology. 2014; 15(3):51-58.

38. Begum S, Syed SA, Siddiqui BS, Sattar SA, M. Iqbal Choudhary. Carandinol: First isohopane triterpene from the leaves of Carissa carandas L. and its cytotoxicity against cancer cell lines. Phytochemistry Letters. 2013; 6(1):91-95.

39. Sahreen S, Khan MR, Khan RA, Shah NA. Estimation of flavoniods, antimicrobial, antitumor and anticancer activity of Carissa opaca fruits. BMC Complement Altern Med. 2013; 27:13:372.

40. Nisa S, Bibi Y, Zia M, Waheed A, Chaudhary MF. Anticancer investigations on Carissa opaca and Toona ciliata extracts against human breast carcinoma cell line. Pak J Pharm Sci. 2013; 26(5):1009-12.

41. Eric Wei Chiang Chan, Siu Kuin Wong, Hung Tuck Chan, Shigeyuki Baba, and Mio Kezuka et al. Cerbera are coastal trees with promising anticancer properties but lethal toxicity: A short review. / J. Chin. Pharm. Sci. 2016; 25 (3):161–169.

42. Sarot Cheenpracha, Chatchanok Karalai, Yanisa Rat-a-pa, Chanita Ponglimanont, Kan Chantrapromma. New Cytotoxic Cardenolide Glycoside from the Seeds of Cerbera manghas Chemical and Pharmaceutical Bulletin. (Chem Pharm Bull). 2004; 52(8):1023-1025

43. Siti Syarifah MM, Nurhanan MY, Muhd Haffiz J, A Mohd Ilham, K Getha, O Asiah, I Norhayati, H Lili Sahira & S Anee S. Potential Anticancer compounds from Cerbera odollam. Journal of Tropical Forest Science. 2011; 23(1):89-96.

44. Chang LC, Gills JJ, Bhat KP, Luyengi L, Farnsworth NR, Pezzuto JM, Kinghorn AD. Activity-guided isolation of constituents of Cerbera manghas with antiproliferative and antiestrogenic activities. Bioorg Med Chem Lett. 2000; 10(21):2431–2434.

45. Mohd Mutalip syarifah Siti, Nurhanan Yunos, 3rd J. Muhd Haffiz et al. Potential anticancer compound from Cerbera odollam. Journal of Tropical Forest Science. 2011; 23(1):89-96 ·

46. Shah VC, Adolf S. D'sa, Noel J.de Souza. Chonemorphine, stigmasterol, and ecdysterone: Steroids isolated through bioassay-directed plant screening programs. Steroids. 1989; 53(3–5):559-565.

47. Kedari P, Malpathak N. Quantification of Camptothecin in Different Plant Parts of Chonemorpha Fragrans. Advances in Zoology and Botany. 2013; 1(2):34-38.

48. Kedari PP, Malpathak NP. Screening of Chonemorpha fragrans bioactive extracts for cytotoxicity potential and inhibition studies of key enzymes involved in replication. Pharmacog. Magazine. 2016; 12(46):S297-302.

49. Kedari, Pradnya Malpathak, Nutan P. Hairy root cultures of Chonemorpha fragrans (Moon) Alston. A potential plant for camptothecin production. IJBT, 2014; 13(2):231-235

50. Poornima K, Gopalakrishnan VK. Anticancer Activity of Tabernaemontana coronaria against Carcinogen Induced Clear Cell Renal Cell Carcinoma. Chinese Journal of Biology.Volume 2014; 8.

51. Sarath P. Gunasekera, Geoffrey Cordell, Norman R. Farnsworth. Anticancer indole alkaloids of Ervatamia heyneana. Phytochemistry. 1980; 19(6):1980 1213-1218.

52. Akhila Sravya Dantu, Shankarguru P, Ramya Devi D, Vedha Hari BN. Evaluation of In Vitro Anticancer Activity of Hydroalcoholic Extract of Tabernaemontana Divaricata. Asian J Pharm Clin Res. 2012; 5(3):59-61

53. Kirankumar Hullatti, Namrata Pathade, Yuvaraj Mandavkar, Ashok Godavarthi, and Mahesh Biradi. Bioactivity-guided isolation of cytotoxic constituents from three medicinal plants Pharm Biol, 2013; 51(5):601–606

54. Kumar A, Selvakumar S. Antiproliferative efficacy of Tabernaemontana divaricata against HEP2 cell line and Vero cell line. Pharmacogn Mag.; 2015; 11(Suppl 1):S46–S52.

55. Sim DS, Teoh WY, Sim KS, Lim SH, Thomas NF, Low YY, Kam TS. Vobatensines A-F, Cytotoxic Iboga-Vobasine Bisindoles from Tabernaemontana corymbosa. J Nat Prod. 2016; 2279(4):1048-55.

56. Pereira PS, França SC, Oliveira PVA, Breves CMS, Pereira SIV. Chemical constituents from Tabernaemontana catharinensis root bark: A brief NMR review of indole alkaloids and in vitro cytotoxicity. Química Nova 2008; 31(1):20-24.

57. Cheenpracha S, Boapun P, Thunwadee Limtharakul (née Ritthiwigrom), Surat Laphookhieo & Stephen G. Pyne. Antimalarial and cytotoxic activities of pregnene-type steroidal alkaloids from Holarrhena pubescens roots. (2017): DOI: 10.1080/14786419.2017.1408108.

58. Sharma V, Hussain S, Bakshi M, Bhat N, Saxena AK. In vitro cytotoxic activity of leaves extracts of Holarrhena antidysenterica against some human cancer cell lines. Indian J Biochem Biophys 2014; 51(1):46-51.

59. Badmus JA, Ekpo OE, Hussein AA, M Meyer, D C Hiss. Antiproliferative and Apoptosis Induction Potential of the Methanolic Leaf Extract of Holarrhena floribunda (G. Don). Evid Based Complement Alternat Med. 2015, 11:756482.

60. Sharma V, Hussain S, Bakshi M, Bhat N, Saxena AK. In vitro cytotoxic activity of leaves extracts of Holarrhena antidysenterica against some human cancer cell lines. Indian J Biochem Biophys. 2014; 51(1):46-51.

61. Singh NK, Singh VP. Phytochemistry and pharmacology of Ichnocarpus frutescens. Chin J Nat Med. 2012; 10(4):241-246.

62. Thangarajana S, Perumal S, Chinthamony AR, Ragavendran Paramasivam, VidyaBalasubramanian, Sunitha Martin, Velliyur Kanniappan Gopalakrishnan. Chemomodulatory effects of Ichnocarpus frutescens R. Br against 4-vinylcyclohexane induced ovarian cancer in swiss albino mice. Journal of Acute Disease. 2013; 2(2):151-155.

63. Kumarappan CT, Mandal SC. Antitumor activity of Polyphenolic extract of Ichnocarpus frutescense. Exp Oncol 2007; 29(2):94–101

64. Singh NK, Singh VP. Anticancer activity of the roots of Ichnocarpus frutescens R. Br. and isolated triterpenes. Pak J Pharm Sci. 2014; 27(1):187-91.

65. Chidambaram K, Subhash C Mandal. Antitumor activity of polyphenols extracts of Ichnocarpus frutescens. Experimental oncology. 2007; 29(2):94-101.

66. Lee Yean Shan, Tee Chuan Thing, Tan Siow Ping, Khalijah Awang, Najihah MohdHashim, Mohd Azlan Nafiah, Kartini Ahmad. Cytotoxic, antibacterial and antioxidant activity of triterpenoids from Kopsia singapurensis Ridl. Journal of Chemical and Pharmaceutical Research (JCPR), 2014; 6(5):815-822. 67. Lim KH, Hiraku O, Komiyama K, Koyano T, Hayashi M, Kam TS. Biologically active indole alkaloids from Kopsia arborea. J Nat Prod. 2007; 70(8):1302-7.

68. Subramaniam G, Hiraku O, Hayashi M, Koyano T, Komiyama K, Kam TS. Biologically active aspidofractinine alkaloids from Kopsia singapurensis. J Nat Prod. 2008, 71(1):53-7.

69.Lim SH, Sim KM, Abdullah Z, Hiraku O, Hayashi M, Komiyama K, Kam TS. Leuconoxine, kopsinitarine, kopsijasmine, and kopsinone derivatives from Kopsia. J Nat Prod. 2007 Aug; 70(8):1380-3.

70. Wahyuningsih MSH. Mubarika S., Mark T. Hamann Gandjar, IG, Wahyuono S., Structure identification of potential compound as selective renal anticancer isolated from Nerium Indicum Mill. Leaves, Indonesian Journal of Pharmacy, 2008; 19(2):57-64.

71. Siddiqui BS, Begum S, Siddiqui S, Lichter W. Two cytotoxic pentacyclic triterpenoids from Nerium oleander. Phytochemistry. 1995; 39:171-4.

72.Pathak S, Multani AS, Narayan S, Kumar V, Newman RA. Anvirzel<sup>™</sup>, an extract of Nerium oleander, induces cell death in human but not murine cancer cells. Anticancer Drugs. 2000; 11:455–63.

73. Heinz H. Fiebig, Gerhard Kelter, Armin Maier, Thomas Metz and Luay J. Rashan. Abstract 5572: Breastin a natural product from Nerium Oleander exhibits high activity in a panel of human tumor cell lines. A. Experimental and Molecular Therapeutics. 2013; 73(8 Supplement).

74.Turan N, Akgün-Dar K, Kuruca SE, Kiliçaslan-Ayna T, Seyhan VG, Atasever B, et al. Cytotoxic effects of leaf, stem and root extracts of Nerium oleander on leukemia cell lines and role of the p-glycoprotein in this effect. J Exp Ther Oncol. 2006; 6:31–8.

75. Siddiqui BS, Khatoon N, Begum S, Farooq AD, Kehkashan Qamar, Huma Aslam Bhatti, Syed Kashif Ali. Flavonoid and cardenolide glycosides and a pentacyclic triterpene from the leaves of Nerium oleander and evaluation of cytotoxicity. Phytochemistry. 2012; 77:238-244.

76. Rashan LJ, Franke K, Khine MM, Gerhard Kelter, Heinz H. Fiebig, Joachim Neumann, Ludger A. Wessjohann. Characterization of the anticancer properties of monoglycosidic cardenolides isolated from Nerium oleander and Streptocaulon tomentosum. Journal of Ethnopharmacology 2011; 134:781-788.

77. Qamar KA, Farooq AD, Siddiqui BS, Kabir N, Khatoon N, Ahmed S, Erum S, Begum S. Antiproliferative Effects of Nerium oleander Stem and Mitotic Arrest Induced by Cardenolide Odoroside B on NCI-H460 Cancer Cells. Letters in Drug Design & Discovery. 2018; 15(1):84-94.

78. Su Jin Song, Cheng Yun Jin, Yung Hyun Choi and Won Deok Hwang. Induction of Apoptosis by Ethanol Extract of Nerium indicum Stem Is Associated with Activation of JNK in Human Renal Carcinoma Caki-1 Cells. Cancer prevention research 2011; 16:269-79.

79. Nagwa M. El Sawi, Neveen S. Geweely, Safaa Qusti, M. Mohamed, A. Kamel. Cytotoxicity and Antimicrobial Activity of Nerium oleander Extracts Journal of Applied Animal Research. 2010; 37:25-31.

80. Garbett NC, Graves DE. Extending nature's leads: the anticancer agent ellipticine. Curr Med Chem Anticancer Agents. 2004; 4(2):149-72.

81. Po-Lin Kuo, Ya-Ling Hsu, Cheng-Hsiung Chang, Chun-Ching Lin. The mechanism of ellipticine-induced apoptosis and cell cycle arrest in human breast MCF-7 cancer cells. Cancer Letters. 2005; 223:293–301.

82. Riham A. El-shiekh, Dalia A. Al-Mahdy, Mohamed S. Hifnawy, Tzvetomira Tzanova, Emilie Evain-Bana, Stéphanie Philippot, Denyse Bagrel, Essam A. Abdelsattar. Chemical and Biological Investigation of Ochrosia elliptica Labill. Cultivated in Egypt. Rec. Nat. Prod. 2017; 11(6):552-557

83. Periyasamy G; Gupta M; Mazumder UK; Gebrelibanos, Mebrahtom; Sintayehu, Biruk. Antioxidant and Antitumor Activity of Plumeria acuminata in Ehrlich Ascites Carcinoma Bearing Swiss Albino Mice. British Journal of Pharmaceutical Research; 2013; 3(4):671-685.

84. Leonardus B. S. Kardono, Soefjan Tsauri, Kosasih Padmawinata, John M. Pezzuto, A. Douglas Kinghorn. Cytotoxic constituents of the

#### Devi et al

bark of Plumeria rubra collected in Indonesia. J. Nat. Prod. 1990; 53(6):1447-1455.

85. Periyasamy G, Gupta M, Mazumder UK, Mebrahtom Gebrelibanos and Biruk Sintayehu. Antioxidant and Antitumor Activity of Plumeria acuminata in Ehrlich Ascites Carcinoma Bearing Swiss Albino Mice. British Journal of Pharmaceutical Research. 3(4): 671-685.

86. Guevara AP1, Amor E, Russell G. Antimutagens from Plumeria acuminata Ait.; Mutat Res. 1996; 361(2-3):67-72.

87. Bemis DL, Capodice JL, Gorroochurn P, A.E. Katzand R. Buttyan. Anti-prostate cancer activity of a ß-carboline alkaloid enriched extract from Rauwolfia vomitoria. International Journal of Oncology. 2006, 29:1065-1073.

88. Jun Yu, Yan Ma, Jeanne Drisko, Qi Chen. Antitumor Activities of Rauwolfia vomitoria Extract and Potentiating of Carboplatin Effects against Ovarian Cancer. Curr Ther Res Clin Exp. 2013; 75:8–14.

89. Rong-Fu Chen, Fumiko Abe, Tatsuo Yamauchi, Masakatsu Taki. Cardenolide glycosides of Strophanthus divaricatus. Phytochemistry. 1987; 26(8):2351-2355.

90. Pezzani R, Rubin B, Redaelli M, Radu C, Barollo S, Maria Verena Cicala, Monica Salvà, Caterina Mian, Carla Mucignat-Caretta, Paolo Simioni, Maurizio Iacobone, Franco Mantero. The antiproliferative effects of ouabain and everolimus on adrenocortical tumor cells. Endocr J. 2014; 61(1):41-53.

91. Karkare S, Adou E, Cao S, Brodie P, James S. Miller, N. M. Andrianjafy, J. Razafitsalama, Rabodo Andriantsiferana, Vincent E. Rasamison, and David G. I. Kingston. Cytotoxic Cardenolide Glycosides of Roupellina (Strophanthus) boiviniifrom the Madagascar Rainforest. J. Nat. Prod.2007; 70(11):1766–1770

92. Tamiris Caroline Barbon, Cássio Prinholato da Silva, Suely Vilela Sampaio, Mateus Amaral Baldo. Evaluation of Anticancer Activity Promoted by Molecules Contained in the Extracts of Thevetia peruviana Toxicon. 2012; 60(2):179-180.

93. Huo-Yun Cheng, Dan-Mei Tian, Jin-Shan Tang, Wei-Zai Shen & Xin-Sheng Yao. Cardiac glycosides from the seeds of Thevetia peruviana and their pro-apoptotic activity toward cancer cells. Mar2016. Journal of Asian Natural Products Research. 2016, 18(9): 837-847.

94. Ramos-Silva A, Tavares-Carreón F, Figueroa M, Susana De la Torre-Zavala, Argel Gastelum-Arellanez, Aída Rodríguez-García, Luis J. Galán-Wong and Hamlet Avilés-Arnaut. Anticancer potential of Thevetia peruviana fruit methanolic extract. 2017 May 2. BMC Complement Altern Med. 2017; 17:241.

95. Salama M, El-Hawary S, Mousa O, El-Askari N, Esmat A. In vivo TNF- $\alpha$  and IL-1 $\beta$  inhibitory activity of Phenolics isolated from Trachelospermum Jasminoids (Lindl.) Lem. Journal of Medicinal Plants Research.2010; 9 (2):30-41.

96. Xing-Qi Tan, Liang-Jun Guo, Yi-Hua Qiu, Hai-Sheng Chen & Chang-Heng Tan, Chemical constituents of Trachelospermum jasminoides. 11 Aug 2009. Natural Product Research Formerly Natural Product Letters. 2010; 24(13):1248-1252.

97. Fatima T, Ijaz S, Crank G, Wasti S.Indole Alkaloids from Trachelospermum jasminoides. Planta Med. 1987; 53(1):57-9.

98. Siu Kuin Wong, Eric Wei Chiang Chan. Botany, uses, phytochemistry and pharmacology of Vallaris: A short review. Pharmacognosy Journal; 2013; 5:242-246.

99. Wong SK, Lim YY, Ling SK, Chiang Chan EW. Antiproliferative activity of Vallaris glabra Kuntze (Apocynaceae). Phcog Mag, 2014; 10(38):232-239.

100. Kruakaew S, Seeka C, Thitima Lhinhatrakool, Sanit Thongnest, Jantana Yahuafai, Suratsawadee Piyaviriyakul, Pongpun Siripong, and Somyote Sutthivaiyakit. Cytotoxic Cardiac Glycoside Constituents of Vallaris glabra Leaves, J. Nat. Prod. 2017; 80(11):2987–2996

101. Karmakar UK, Ghosh D, Sadhu SS. Assessment of Analgesic, Cytotoxic and Antioxidant activities of Vallaris solanacea (Roth) Kuntze. Stamford. J. Pharm. Sci. 2010; 4(1):64-68.

102. Siddiqui MJ, Ismail Z, Aisha AF, Abdul Majid AM. Cytotoxic activity of Catharanthus roseus (Apocynaceae) crude extracts and pure compounds against human colorectal carcinoma cell line. Int J Pharmacol. 2010; 6:43–7.

103. Robert L. Noble. The discovery of the vinca alkaloids chemotherapeutic agents against cancer Biochemistry and Cell Biology. 1990; 68(12):1344-1351

104. Maryam Moudi, Rusea Go, Christina Yong Seok Yien, and Mohd. Nazre. Vinca Alkaloids. Int J Prev Med. 2013; 4(11): 1231–1235

105. El-Sayed A, Handy GA, Cordell GA. Catharanthus alkaloids XXXVIII. Confirming structural evidence and antineoplastic activity of the bisindole alkaloids leurosine-N'b-oxide (pleurosine) roseadine and vindolicine from Catharanthus roseus. J Nat Prod 1983; 46:517-27.

106. Antony J, Saikia M, Vinod V, Nath LR, Katiki MR, Murty MS, Paul A, Shabna A, Chandran H, Joseph SM, Nishanth KS, Panakkal EJ, Sriramya I, Sridivya I, Ran S, Sankar S, Rajan E, Anto RJ. DW-F5: A novel formulation against malignant melanoma from Wrightia tinctoria. (Scientific reports)Sci Rep. 2015; 10(5):12662.

107. Selvam P, Murugesh M, Witvrouw M, Keyaerts E, J. Neyts. Studies of Antiviral Activity and Cytotoxicity of Wrightia tinctoria and Morinda citrifolia. Indian J Pharm Sci. 2009; 71(6):670–672.

108. Ramalakshmi S, Edaydulla N, Ramesh P, Muthuchelian K. Investigation on cytotoxic, antioxidant, antimicrobial and volatile profile of Wrightia tinctoria (Roxb.) R Br Flower used in Indian medicine. Asian Pac J Trop Dis 2012; 68-75.

109. Chakravarti B, Maurya R, Siddiqui JA, Bid HK, Rajendran SM, Yadav PP, Konwar R. In vitro anti-breast cancer activity of ethanolic extract of Wrightia tomentosa: Role of pro-apoptotic effects of oleanolic acid and urosolic acid Journal of Ethnopharmacology 2012; 142(1):72–79.

110. Chaudhary S, Devkar RA, Bhere D, Setty MM, Ranganath Pai KS. Selective cytotoxicity and pro-apoptotic activity of stem bark of Wrightia tinctoria Roxb. Pharmacognosy Magazine. 2015, 11(44):481-487.

111. Fatima N, Ahmad MK, Ansari JA, Ali Z, Khan AR, Mahdi AA. Anticancer, antioxidant potential and profiling of polyphenolic compounds of Wrightia tinctoria Roxb. (R.Br.) bark. Journal of Advanced Pharmaceutical Technology & Research. 2016; 7(4):159-165.

112. Chakravarti B, Maurya R, Siddiqui JA, Bid HK, Rajendran SM, Yadav PP, Konwar R. In vitro anti-breast cancer activity of ethanolic extract of Wrightia tomentosa: Role of pro-apoptotic effects of oleanolic acid and urosolic acid Journal of Ethnopharmacology 2012; 142(1):72–79.

113. Mariquit M De Los Reyes, Glenn G Oyong, Vincent Antonio S. Ng, Chien-Chang Shen, Consolacion Y Ragasa. Cytotoxic Compounds from Wrightia pubescens (R.Br.) Phcog Res. 2018; 10(1):9-15.