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Research Article

### Phytochemical analysis of methanolic extract of *Ulva rigida* C.Ag. collected from Koothankuzhi Coast, Tirunelveli district, Tamil Nadu, India

<sup>1</sup>Dr. M. Muthu Sheeba, <sup>2</sup>Dr. J. John Peter Paul\*, <sup>3</sup>Dr. C. Iniya Udhaya<sup>1</sup>Centre for Advanced Research in Plant Sciences (CARPS), Department of Botany, St. Xavier's College (Autonomous), Palayamkottai – 627 002, Tamil Nadu, India<sup>2</sup>\*Assistant Professor of Botany and Director, Centre for Advanced Research in Plant Sciences (CARPS), St. Xavier's College (Autonomous), Palayamkottai – 627 002, Tamil Nadu, India<sup>3</sup>Assistant Professor of Botany, Sri Kaliswari College (Autonomous), Sivakasi – 626 123, Tamil Nadu, India

#### ABSTRACT

The present study was concentrated to explore the phytochemicals present in the methanolic extract of *Ulva rigida* C.Ag., collected from Koothankuzhi in the south east coast of Tamil Nadu, India. The phytochemical analysis of methanolic extract was screened using the standard procedure for UV-Visible spectroscopic, HPLC and FTIR. The UV-Visible spectrum showed the compounds separated at the nm of 662, 603, 533 and 400 with the absorption 0.653, 0.331, 0.458 and 2.684 respectively. The qualitative HPLC fingerprint profile displayed fourteen compounds at different retention time of 1.770min, 2.230min, 2.540min, 2.870min, 3.090min, 3.377min, 3.900min, 4.257min, 4.797min, 5.340min, 5.853min, 6.520min, 7.730min and 9.220min. The result of FTIR analysis was found the presence of functional groups such as alkynes, sulfonic acids, carboxylic acids, carboxylic acid salt, aldehydes, aliphatic and unsaturated hydrocarbons.

**Keywords:** *Ulva rigida*, UV-Visible, HPLC, FTIR**Article Info:** Received 18 Oct 2018; Review Completed 27 Nov 2018; Accepted 29 Nov 2018; Available online 15 Dec 2018

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#### \*Address for Correspondence:

Dr. John Peter Paul J., Research Department of Botany, St. Xavier's College (Autonomous), Palayamkottai – 627 002, Tamil Nadu, India.

#### INTRODUCTION

Seaweeds constitute one of the important living resources categorized under three groups namely Chlorophyceae (green algae), Phaeophyceae (brown algae) and Rhodophyceae (red algae). There are about 900 species of green seaweeds, 1500 brown seaweeds and 4000 red seaweeds in nature. The greatest variety of red seaweeds is present in subtropical and tropical waters, while brown seaweeds and green seaweeds are more commonly found in cooler and temperate waters respectively <sup>1</sup>. As seaweeds are important renewable resources in the marine ecosystem and have been a part of human civilization from time immemorial. The long history of seaweeds utilization for a various purposes has led to the regular realization that some of the compounds are more superior and valuable in comparison to the counterparts of land plants <sup>2</sup>.

Seaweeds produce an extensive variety of chemicals, some of the chemicals stand only as natural resources <sup>3</sup>. Seaweeds are the marine macro algae found in the shallow waters of sea, estuaries and backwaters. Earlier reports on the uses of seaweeds have been noticed early as 2500 years

ago in Chinese literature also <sup>4</sup>. Seaweeds are considered as source of bioactive compounds and create a great variety of secondary metabolites characterized by a broad spectrum of biological activities <sup>5</sup>. There are various reports of compounds derived from seaweeds with a broad range of biological activities such as antibiotics, antiviral diseases, anti-tumour and anti-inflammatory as well as neurotoxins <sup>6</sup>. Hence the present study was carried out to screen the phytochemicals in *Ulva rigida* C.Ag., collected from Koothankuzhi coast, Tirunelveli district in the south east coast of Tamil Nadu, India.

#### MATERIALS AND METHODS

##### Collection of Plant sample

The plant materials used in the present study was *Ulva rigida* C.Ag., belonging to Chlorophyceae (green algae) was made during the low tidal and subtidal regions (up to 1m depth) by hand picking. The collected materials were washed thoroughly with marine water in the field itself to remove the epiphytes and sediment particles. Then the samples were packed separately in polythene bags in wet

conditions and brought to the laboratory, then thoroughly washed in tap water followed by distilled water to remove the salt on the surface of the thalli. They were stored in 5% formalin solution <sup>7</sup>.

### Preparation of extracts

For the preparation of methanolic extract, the plant specimens were washed thoroughly and placed on blotting paper and spread out at room temperature in the shade condition for drying. The shade dried samples were grounded to fine powder using a tissue blender. The powdered samples were then stored in the refrigerator for further use. 30g powdered samples were packed in Soxhlet apparatus and extracted with methanol for 8h separately <sup>8</sup>.

### UV-Vis spectral analysis

The methanolic crude extract containing the bioactive compound was analyzed UV-Visible spectroscopically for further confirmation. The methanolic crude extract of *Ulva rigida* C.Ag. was scanned in a wavelength ranging from 200-900nm using a Shimadzu spectrophotometer and characteristic peaks were detected <sup>9</sup>.

### HPLC Analysis

The HPLC method was performed on a Shimadzu LC-10AT VP HPLC system, equipped with a model LC-10AT pump, UV-Vis detector SPD-10AT, a Rheodyne injector fitted with a 20 $\mu$ l loop and an auto injector SIL-10AT. A Hypersil<sup>®</sup> BDS C-18 column (4.6  $\times$  250mm, 5 $\mu$ m size) with a C-18 guard column was used. The elution was carried out with gradient solvent systems with a flow rate of 1ml/min at ambient temperature (25-28°C). The mobile phase consisted of 0.1% v/v methanol (solvent A) and water (solvent B). The mobile phase was prepared daily, filtered through a 0.45 $\mu$ m and sonicated before use. Total running time was 15min. The sample injection volume was 20 $\mu$ l while the wavelength of the UV-Vis detector was set at 254nm <sup>10</sup>.

### Instrumentation

An isocratic HPLC (Shimadzu HPLC Class VP series) with two LC- 0 AT VP pumps (Shimadzu), a variable wave length programmable photo diode array detector SPD-M10A VP (Shimadzu), a CTO- 10AS VP column oven (Shimadzu), a SCL-10A VP system controller (Shimadzu), a reverse phase Luna 5 $\mu$ l C18 (2) and Phenomenex column (250 mm X 4.6mm) were used. The mobile phase components methanol: water (45:55) were filtered through a 0.2 $\mu$ m membrane filter before use and were pumped from the solvent reservoir at a flow rate of 1ml/min which yielded column backup pressure of 260-270kgf/cm<sup>2</sup>. The column temperature was maintained at 27°C. 20 $\mu$ l of the respective sample and was injected by using a Rheodyne syringe (Model 7202, Hamilton).

### FTIR analysis

The methanolic extract of *Ulva rigida* C.Ag. was shade dried and FTIR analysis was performed using Perkin Elmer Spectrophotometer system, which was used to detect the characteristic peaks and their functional groups. The peak values of the FTIR were recorded. Each and every analysis was repeated twice and confirmed the spectrum <sup>11</sup>.

## RESULTS AND DISCUSSION

### UV-Visible spectrum analysis

The UV-Visible spectrum of the methanolic extract of *Ulva rigida* C.Ag. was selected at the wavelength of 200nm to 900nm due to the sharpness of the peaks and proper baseline. The profile showed the compounds separated at the nm of 662, 603, 533 and 400 with the absorption 0.653, 0.331, 0.458 and 2.684 respectively (Figure-1).

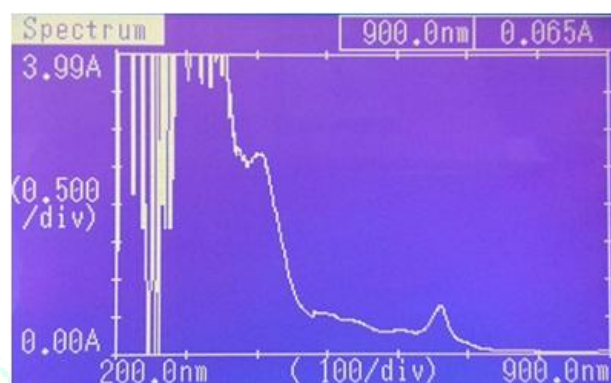


Figure 1: UV-Visible spectrum of methanolic extract of *Ulva rigida* C.Ag.

### HPLC analysis

The qualitative HPLC fingerprint profile of the methanolic extract of *Ulva rigida* C.Ag. was selected at a wavelength of 660nm due to the sharpness of the peaks and proper baseline. The methanolic extract prepared by hot extraction was subjected to HPLC for the separation and identification of constituents present in the *Ulva rigida* C.Ag. Fourteen compounds were separated from the methanolic extract of *Ulva rigida* C.Ag. at different retention time of 1.770min, 2.230min, 2.540min, 2.870min, 3.090min, 3.377min, 3.900min, 4.257min, 4.797min, 5.340min, 5.853min, 6.520min, 7.730min and 9.220min. The profile displayed nine prominent peaks at the retention time of 1.770min, 2.230min, 2.540min, 2.870min, 3.090min, 3.377min, 5.340min, 6.520min and 9.220min followed by seven moderate peaks were observed at the retention time of 3.900min, 4.257min, 4.797min, 5.853min and 7.730min (Figure-2).

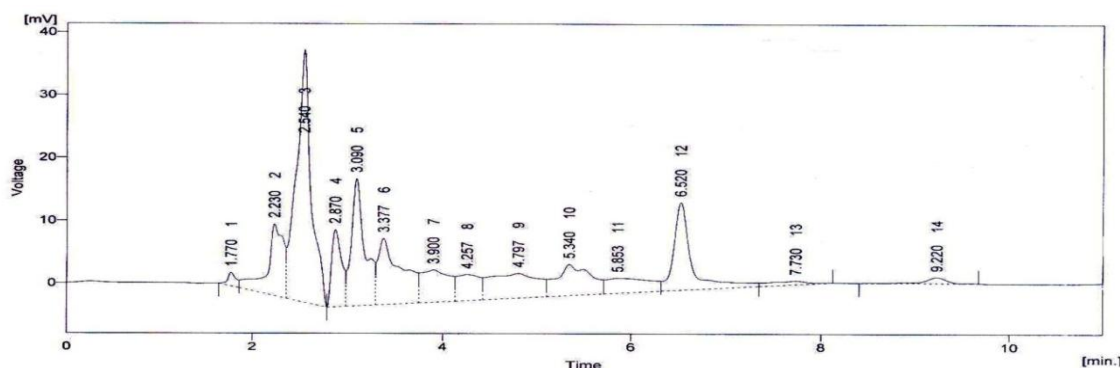


Figure 2: HPLC analysis of methanolic extract of *Ulva rigida* C.Ag.

### FTIR analysis

The FTIR spectrum was used to predict the functional group of the active components based on the peak value in the region of infra red radiation. The crude methanolic extract of *Ulva rigida* C.Ag. was conceded into the FTIR and the functional groups of the components were separated based on its peak ratio. FTIR spectrum of methanolic

extract showed different peaks at 680.83, 1166.85, 1461.94, 1598.88, 1739.67, 2854.45 and 2925.81 $\text{cm}^{-1}$ . It was confirmed the presence of functional groups such as alkynes (C=C-H bending), sulfonic acids ( $\text{SO}_3$  sym stretching), carboxylic acids (OH bending), carboxylic acid salt ( $\text{COO}^-$  antisym stretching), aldehydes (C=O stretching), aliphatic (CH antisym and sym stretching) and unsaturated hydrocarbons (=C-H stretching) respectively (Figure-3).

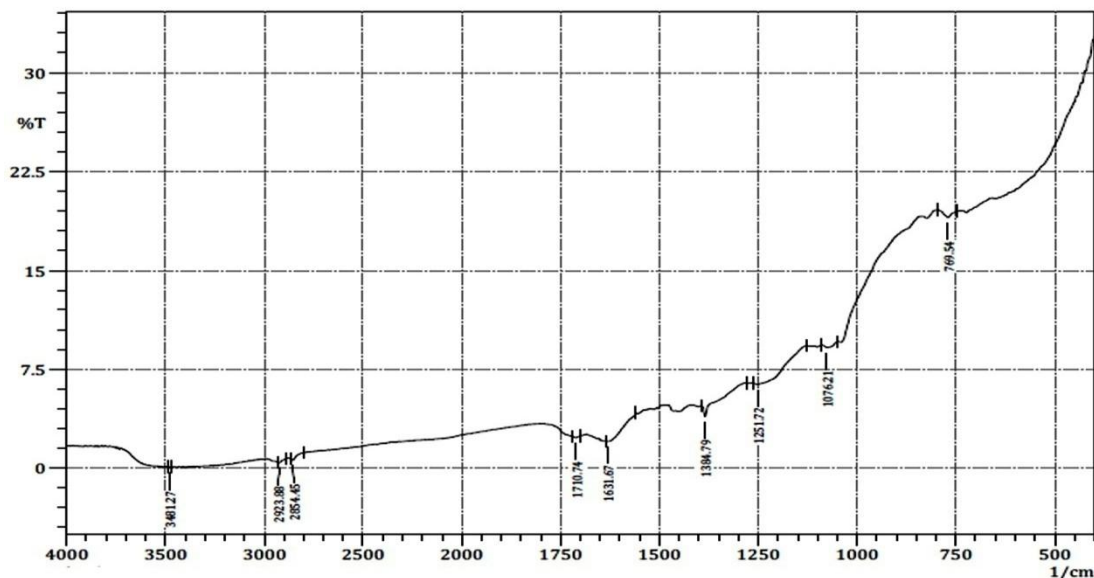


Figure 3: FT-IR spectrum of methanol extract of *Ulva rigida* C.Ag.

### CONCLUSION

From the present study, it was noted that UV-Visible spectrum of methanolic extract of *Ulva rigida* C.Ag. showed the compounds separated at the nm of 662, 603, 533 and 400 with the absorption 0.653, 0.331, 0.458 and 2.684 respectively. The qualitative HPLC fingerprint profile displayed fourteen compounds at different retention time of 1.770min, 2.230min, 2.540min, 2.870min, 3.090min, 3.377min, 3.900min, 4.257min, 4.797min, 5.340min, 5.853min, 6.520min, 7.730min and 9.220min. The result of FTIR analysis was found the presence of functional groups such as alkynes, sulfonic acids, carboxylic acids, carboxylic acid salt, aldehydes, aliphatic and unsaturated hydrocarbons in the methanolic extract of *Ulva rigida* C.Ag.

### CONFLICT OF INTEREST

The author declares that he has no conflict of interest.

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