

## RESEARCH ARTICLE

**PHYTOCHEMICAL AND IN-VITRO ANTIOXIDANT ACTIVITY OF METHANOLIC EXTRACT OF *LACTUCA SCARIOLA* AND *CELOSIA ARGENTEA* LEAVES**\*G H Urmila<sup>1</sup>, B.Ganga Rao<sup>2</sup>, T.Satyanarayana<sup>2</sup><sup>1</sup>Nargund college of Pharmacy, Banashankari 3rd stage Bangalore, Karnataka, India 560 085.<sup>2</sup>A.U College of Pharmaceutical Sciences, Andhra University, Visakhapatnam, A.P, India-530003\*Corresponding Author's Email: [urmilagk@yahoo.co.in](mailto:urmilagk@yahoo.co.in), contact: 9482020128**ABSTRACT**

Antioxidant compounds in food play an important role as a health protecting factor. Primary sources of naturally occurring antioxidants are whole grains, fruits and vegetables. Highly reactive free radicals and oxygen species can initiate degenerative diseases. Antioxidant compounds like phenolic acids, polyphenols and flavonoids are commonly found in plants have been reported to have multiple biological effects, including antioxidant activity. Methanolic leaf extracts of *Lactucascariola* Linn and *Celosia argentea* Linn were assessed for the In vitro Antioxidant activity using DPPH (1, 1-diphenyl-2-picryl-hydrazyl) radical, Nitric oxide (NO) and hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) scavenging models using Ascorbic acid as positive control. Analysis of free radical scavenging activities of the extracts revealed a concentration dependent free radical scavenging activity resulting from reduction in DPPH, Nitric oxide and Hydroxyl radical. The methanolic extract of *Lactucascariola* Linn showed significant DPPH, Nitric oxide and hydrogen peroxide scavenging activity compared to *Celosia argentea* Linn extract. IC<sub>50</sub> values by DPPH, Nitric oxide and hydrogen peroxide model for *Lactucascariola* and *Celosia argentea* were found to be 192.5±9.014, 394.2±6.009, 434.2±18.78, 233.3±20.73, 521.4±4.061 and 494.2±5.465 respectively.

**Keywords:** *Lactucascariola*, *Celosia argentea*, Antioxidant activity, methanolic extracts.

**INTRODUCTION**

Antioxidants are vital substances, which possess the ability to protect the body from damages caused by free radical-induced oxidative stress.<sup>1</sup> The antioxidant which when present at low concentrations compared with those of an oxidizable substrate, significantly delays or prevents oxidation of substrate. The term 'oxidizable substrate' includes almost everything found in the living cells including proteins, lipids, DNA and carbohydrates.

Biological antioxidants have been defined as compounds that protect biological systems against the potentially harmful effects of processes or reaction that can cause excessive oxidation. Our body is rich in endogenous antioxidants, the substances that have the ability to stop free radicals formation or to limit the damage they cause.<sup>2</sup>

Antioxidant compounds in food play an important role as a health protecting factor. Primary sources of naturally occurring antioxidants are whole grains, fruits and vegetables.

Highly reactive free radicals and oxygen species can initiate degenerative diseases. Free radicals arising from metabolism or environmental sources interact continuously in biological systems and their uncontrolled generation correlates directly with molecular level of many diseases. Free radicals would damage nearby structures including DNA, proteins or lipids. Radical scavenging antioxidants are particularly important in antioxidative-defence in protecting cells from the injury of free-radical. Plants are the good source of biologically active compounds known as phytochemicals. The phytochemicals have been found to act as antioxidants by scavenging free radicals and may have therapeutic potential for free radical associated disorders. Free radicals are the major cause of various chronic and degenerative diseases, such as coronary heart

disease, inflammation, stroke, diabetes mellitus and cancer.<sup>3</sup>

The present study was aimed to carry out the Antioxidant activity by DPPH (1, 1-diphenyl-2-picryl-hydrazyl) radical, Nitric oxide and hydrogen peroxide scavenging models using the methanolic leaf extracts of *Lactucascariola* Linn and *Celosia argentea* Linn.

*Lactucascariola* is a common weed in cultivated fields belonging to family Asteraceae. Leaves are used as vegetable, eaten raw or cooked.<sup>4</sup> *Celosia argentea* belonging to family Amaranthaceae grows as a weed during rainy season throughout India and other tropical regions of the world such as Sri Lanka, South Asia, Africa and America.<sup>5</sup>

**MATERIALS AND METHOD****Plant Material**

*Lactucascariola* leaves were collected from Ravor village, Gulbarga district, Karnataka and leaves of *Celosia argentea* were collected from local market Bangalore. The samples were authenticated by Dr. Santhanu, botanist, Natural Remedies Pvt limited, Bangalore.

**Preparation of extract**

Collected leaves were shade dried, pulverized to a coarse powder and was extracted in Soxhlet extractor consecutively using solvents of non polar to Polar grade (Petroleum ether, Methanol and Aqueous), obtained crude extracts were evaporated to dryness in rotary evaporator.

**Preliminary phytochemical screening<sup>6,7,8,9</sup>**

The methanolic leaf extract of *Lactuca Serriola* (LSME) and *Celosia argentea* (CAME) were subjected to different chemical tests for the detection of phytochemical constituents such as carbohydrates, glycosides, alkaloids, amino acids, phenolics, flavonoids, triterpenoids, steroids, etc.

### ANTIOXIDANT ACTIVITY

#### Determination of Free radical scavenging activity by DPPH method<sup>10</sup>

2ml of 0.33% methanolic solution of DPPH was added to different concentration of methanolic *Lactuca Serriola* and *Celosia argentea* (100-500 µg/ml) extract. After 30 minutes; absorbance was measured at 517nm using UV-Visible spectrophotometer. All the tests were performed in triplicate and averaged. Ascorbic acid was used as standard. Percentage scavenging of the DPPH free radical was calculated using the following equation.

$$\text{DPPH radical scavenging activity (\%)} = \frac{(A_{\text{control}} - A_{\text{Test}})}{A_{\text{control}}} \times 100.$$

Where  $A_{\text{control}}$  is the absorbance of the control reaction and  $A_{\text{test}}$  is the absorbance in the presence of the extracts or standard.

#### Determination of Nitric oxide (NO) scavenging activity<sup>11</sup>

Nitric oxide (NO) was generated from sodium nitroprusside (SNP) and was measured by the Griess reagent (1% sulphanilamide, 2% orthophosphoric acid, 0.1% naphthyl ethylene diamine dihydrochloride).

1ml sodium nitroprusside (5mM, in phosphate buffer saline pH 7.4) was mixed with 1ml of different concentration of *Lactuca Serriola* and *Celosia argentea* (100-500 µg/ml) methanolic extract and incubated for 3 hours at 25°C. To 1ml of incubated solution, 1 ml of Griess reagent was added. The absorbance of the chromophore formed

during diazotization of the nitrite with sulphanilamide and subsequent coupling with naphthylethylenediamine dihydrochloride was measured at 546 nm. All the tests were performed in triplicate and averaged. Ascorbic acid was used as standard.

Percentage of Nitric oxide scavenged was calculated using the following equation

$$\text{Nitric oxide scavenged (\%)} = \frac{(A_{\text{control}} - A_{\text{Test}})}{A_{\text{control}}} \times 100.$$

Where  $A_{\text{control}}$  is the absorbance of the control reaction and  $A_{\text{test}}$  is the absorbance in the presence of the extracts or standard.

#### Determination of Hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) scavenging activity<sup>12,13</sup>

Different concentration of *Lactuca serriola* and *Celosia argentea* methanolic extract (100-500 µg/ml) were added to 0.6ml hydrogen peroxide solution (40 mM in phosphate buffer pH 7.4). The absorbance of hydrogen peroxide at 230 nm was determined after ten minutes against a blank solution containing phosphate buffer without hydrogen peroxide. Ascorbic acid was used as standard. All the tests were performed in triplicate and averaged. Percentage of hydrogen peroxide scavenging by the extracts and standard compounds were calculated using the following equation:

$$\% \text{ scavenged [H}_2\text{O}_2] = \frac{(A_0 - A_1)}{A_0} \times 100$$

where  $A_0$  was the absorbance of the control and  $A_1$  was the absorbance in the presence of extracts or standard.

## RESULT AND DISCUSSION

### Phytochemical screening

Preliminary phytochemical screening of the *Lactuca serriola* and *Celosia argentea* methanolic leaves extract revealed the presence of different phytochemical constituents which are represented in **Table 1**.

**Table 1:** Phytochemical constituents of methanolic extracts of *Lactuca serriola* and *Celosia argentea* leaves

Test S	<i>Lactuca serriola</i> methanolic extract (LSME)	<i>Celosia argentea</i> methanolic extract (CAME)
Alkaloids	-	-
Glycosides	+	+
Saponins	+	+
Phytosterols	+	-
Phenolic compounds	+	+
Flavanoids	+	+
Tannins	+	+
Carbohydrates	+	+
Tri terpenoids	+	-

(+): Indicates the presence of chemical constituents. (-): Indicates the absence of chemical constituents.

#### Free radical scavenging activity by DPPH method<sup>14</sup>

DPPH is a stable free radical and can be reduced in the presence of an antioxidant molecule. The ability of a molecule to donate a hydrogen atom to a radical determines its antioxidant potentials. DPPH accepts a hydrogen atom from an antioxidant and becomes a stable

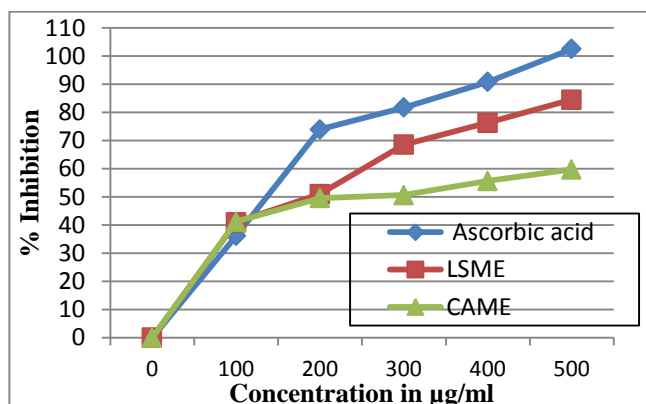
diamagnetic molecule. The reduction capability of DPPH radicals was determined by the decrease in its absorbance at 517 nm.

*Lactuca serriola* and *Celosia argentea* methanolic extract tested for antioxidant activity using DPPH free radical showed antioxidant activity. (**Table 2, Figure I**).

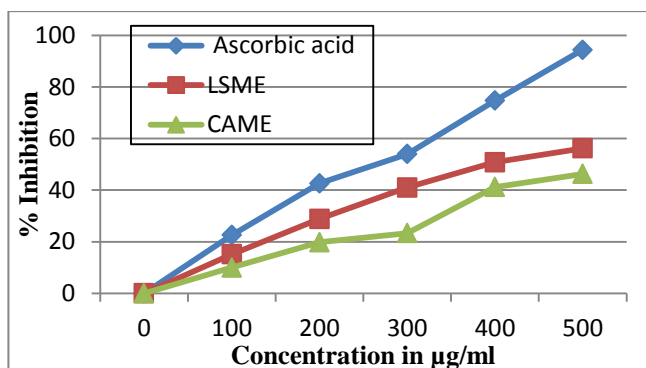
**Table 2:** DPPH radical scavenging activity  $\pm$ SD of three replicates

Concentration	Ascorbic acid(%inhibition)	LSME(%inhibition)	CAME(%inhibition)
100 $\mu$ g/ml	36.14 $\pm$ 1.019	40.98 $\pm$ 0.6583	41.20 $\pm$ 1.150
200 $\mu$ g/ml	73.95 $\pm$ 0.5006	50.99 $\pm$ 1.508	49.58 $\pm$ 1.327
300 $\mu$ g/ml	81.72 $\pm$ 0.5816	68.53 $\pm$ 1.357	50.66 $\pm$ 1.231
400 $\mu$ g/ml	90.83 $\pm$ 0.3014	76.32 $\pm$ 2.003	55.63 $\pm$ 0.8541
500 $\mu$ g/ml	102.5 $\pm$ 0.5533	84.50 $\pm$ 1.336	59.73 $\pm$ 0.6384
IC <sub>50</sub>	138.3 $\pm$ 2.205	192.5 $\pm$ 9.014	233.3 $\pm$ 20.73

IC<sub>50</sub> value showed for *Lactucascariola* and *Celosia argentea* leaf extracts were 192.5 $\pm$ 9.014 and 233.3 $\pm$ 20.73 as compared with standard Ascorbic acid 138.3 $\pm$ 2.205 $\mu$ g/ml.



**Figure 1:** Graphical representation of % Inhibition against concentration ( $\mu$ g/ml) on DPPH model



**Figure II:** Graphical representation of % Inhibition against concentration ( $\mu$ g/ml) on NO model

**Nitric oxide scavenging activity**

**Table 3:** Nitric oxide scavenging activity  $\pm$ SD of three replicates

Concentration	Ascorbic acid(%inhibition)	LSME(%inhibition)	CAME(%inhibition)
100 $\mu$ g/ml	22.56 $\pm$ 1.425	15.11 $\pm$ 0.9700	9.977 $\pm$ 0.9617
200 $\mu$ g/ml	42.64 $\pm$ 1.152	28.83 $\pm$ 1.621	19.82 $\pm$ 0.9180
300 $\mu$ g/ml	53.95 $\pm$ 0.5487	41.03 $\pm$ 2.312	23.36 $\pm$ 1.645
400 $\mu$ g/ml	74.70 $\pm$ 1.067	50.81 $\pm$ 0.9333	41.20 $\pm$ 1.860
500 $\mu$ g/ml	94.33 $\pm$ 1.224	56.23 $\pm$ 1.135	46.28 $\pm$ 1.454
IC <sub>50</sub>	270.3 $\pm$ 1.740	394.2 $\pm$ 6.009	521.4 $\pm$ 4.061

**Table 4:** Hydrogen Peroxide scavenging activity  $\pm$ SD of three replicates

Concentration	Ascorbic acid(%inhibition)	LSME(%inhibition)	CAME(%inhibition)
100 $\mu$ g/ml	31.62 $\pm$ 1.436	20.43 $\pm$ 0.6409	9.480 $\pm$ 0.3880
200 $\mu$ g/ml	54.50 $\pm$ 2.154	30.81 $\pm$ 0.7337	19.23 $\pm$ 0.1867
300 $\mu$ g/ml	79.15 $\pm$ 0.5487	39.88 $\pm$ 0.2132	29.21 $\pm$ 1.118
400 $\mu$ g/ml	93.31 $\pm$ 1.656	47.83 $\pm$ 1.353	38.32 $\pm$ 0.54
500 $\mu$ g/ml	97.82 $\pm$ 1.299	53.43 $\pm$ 0.6929	50.83 $\pm$ 0.7435
IC <sub>50</sub>	183.3 $\pm$ 7.949	434.2 $\pm$ 18.78	494.2 $\pm$ 5.465

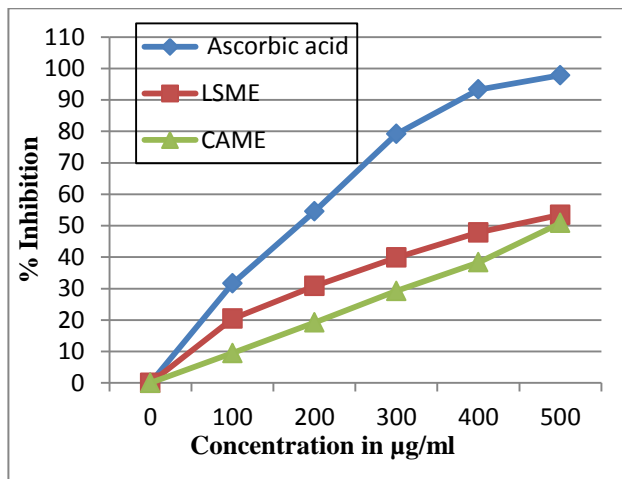
Nitric oxide (NO) has also been involved in a variety of biological functions, including neurotransmission, vascular homeostasis, antimicrobial, and antitumor activities.

SNP in aqueous solution at physiological pH spontaneously generates NO, which interacts with oxygen to produce nitrite ions that can be estimated by the use of Griess reagent. Scavengers of NO compete with oxygen leading to reduced production of NO.<sup>11</sup>

The LSME at varied concentration showed remarkable inhibitory effect of NO scavenging activity compared to CAME. In the NO scavenging activity Ascorbic acid, LSME and CAME plant extracts increased gradually with increase in concentration which showed scavenging activity to be dose dependent. IC<sub>50</sub> value for *Lactucascariola* and *Celosia argentea* leaf extracts was found to be 394.2 $\pm$ 6.009 and 521.4 $\pm$ 4.061 respectively and for Ascorbic acid the IC<sub>50</sub> value was found 270.3 $\pm$ 1.740 $\mu$ g/ml. (Table 3, Figure II).

**Hydrogen Peroxide scavenging activity**

The measurement of H<sub>2</sub>O<sub>2</sub> scavenging activity is one of the useful methods of determining the ability of antioxidants to decrease the level of pro-oxidants such as H<sub>2</sub>O<sub>2</sub>. It can cross membranes and may slowly oxidize a number of compounds. Hydrogen peroxide itself is not very reactive, but sometimes it can be toxic to cells because of rise in the hydroxyl radicals in the cells.<sup>15</sup> IC<sub>50</sub> value for Ascorbic acid in the H<sub>2</sub>O<sub>2</sub> model was found to be 183.3 $\pm$ 7.949 $\mu$ g/ml and for LSME and CAME it was found to be 434.2 $\pm$ 18.78 and 494.2 $\pm$ 5.465 $\mu$ g/ml respectively.



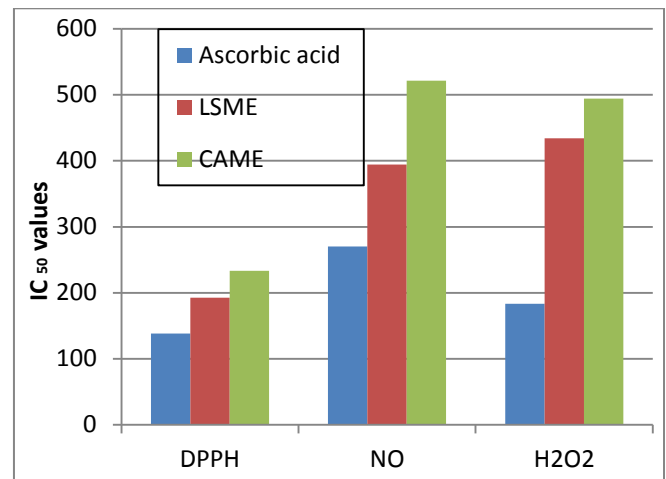
**Figure III:** Graphical representation of % Inhibition against concentration(µg/ml) on H<sub>2</sub>O<sub>2</sub> model

### CONCLUSION

In vitro antioxidant activity was carried out with methanolic leaf extracts of *Lactucascariola* Linn and *Celosia argentea* Linn by DPPH, NO and H<sub>2</sub>O<sub>2</sub> radical scavenging activities models. The result of the present study revealed that both extracts exhibited antioxidant activity but *Lactucascariola* Linn displayed better activity compared to *Celosia Argentea*. The phytochemical constituents present in the plant extract may be responsible for showing the anti-oxidant properties.

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**Figure IV:** Graphical representation of 50% Inhibition concentration (IC<sub>50</sub>) on DPPH, NO and H<sub>2</sub>O<sub>2</sub> model

*Lactucascariola* Linn and *Celosia argentea* Linn can be proposed as promising natural sources of antioxidants suitable for application in nutritional/pharmaceutical fields, in the prevention of free radical-mediated diseases.

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