RESEARCH ARTICLE

DEVELOPMENT AND VALIDATION OF REVERSED PHASE HPLC METHOD FOR ESTIMATION OF SIMVASTATIN IN PHARMACEUTICAL DOSAGE FORM

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ABSTRACT

A simple, accurate rapid and precise RP-HPLC method has been developed and validated for determination of simvastatin in bulk drug. The RP-HPLC separation was achieved on Promosil C-18, (250 mm, 4.6 mm, 5µm) using mobile phase buffer: methanol ph 6.8 (96: 4 v/v) at flow rate of 1.0 ml/min at ambient temperature. The retention times were 9.546 min. for simvastatin. Calibration plots were linear over the concentration range 1-50µg/ml. Quantification was achieved with photodiode array detection at 254 nm over the concentration range of 1-50 µg/ml. The method was validated statistically and applied successfully for the determination of simvastatin. Validation studies revealed that method is specific, rapid, reliable, and reproducible. The high recovery and low relative standard deviation confirm the suitability of the method for the routine determination of simvastatin in bulk drug.

Keywords: Simvastatin, Water, Buffer, Validation, HPLC.

INTRODUCTION

Simvastatin is 2, 2 –dimethyl butanoic acid (1S, 3R, 7S, 8S, 8aR) – 1, 2, 3, 7, 8, 8a-hexahydro-3, 7-dimethyl-8-[2-[(2R,4R) tetrahydro -4- hydroxyl- 6 -oxo-2H pyran-2-yl]ethyl-1-naphthalenyl ester[1,2] belongs to the group of cholesterol-lowering lactones known as statins which, in 2007, were identified as being the most widely prescribed drugs in the world. Statins lower cholesterol by inhibiting the synthesis of mevalonic acid, which is a key precusor in cholesterol synthesis. SMT, a lipid lowering agent that is derived synthetically from a fermentation product of Aspergillus terreus has been found to lessen both normal and elevated LDL-C concentrations.
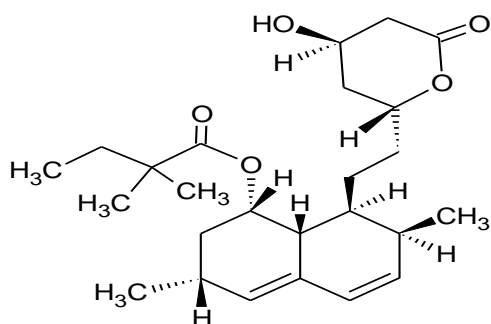

Figure 1: Chemical structure of Simvastatin

The drug is officially listed in 2004 United States Pharmacopocia and the official method of its determination is UV-spectrophotometry[3-5], and various other methods are HPTLC[6], miscellar eletrokinetic chromatography and voltammetry [7] have been reported for the assay of SMT in pharmaceuticals. The method development bottleneck result from the requirement to generate a quantitative and qualitative profile of impurities, enabling the reporting of the identity of each chemical moiety[8]. Two official methods utilising HPLC GRADIENT methodology are reported in European Pharmacopoeia (EP) and United State Pharmacopoeia (USP) [9, 10].

MATERIALS AND METHODS

All the reagents used were of HPLC grade and analytical grade and were purchased from Merck Chemicals, India. Reference standard of Simvastatin was supplied as gift sample from Sun Pharmaceutical Laboratories Limited, Mumbai with purity of 99.987%.

- **Preparation of buffer solution:**
  Mix 5mL of glacial acid in 1000mL of milli Q water. To 1000mL of 5mL glacial acetic acid solution, add 0.94gm of 1-Hexane sulphonic acid anhydrous.sonicate to dissolve.

- **Preparation of Mobile phase:**
  The mobile phase is prepared by mixing buffer: methanol in the ratio of 96:4 Filtered and degas it

- **Chromatographic Run:**
  Load the standard solution of simvastatin in the injector, enter the HPLC parameters as per (Table: 1), save the method, inject and run for 20min.

- **Standard preparation of simvastatin**
  Accurately weigh and transfer about 20 mg of drug simvastatin working standard into 100mL volumetric flask, and about 70 mL of diluents, sonicate to dissolve, dilute to volume with diluents and mix. Filter the solution through0.45µm.

- **Preparation of system suitability solution[8].**
  Accurately weigh and transfer about 10mg of working standard into 100ml volumetric flask. Add 25mL of 0.1N HCl and 25mL of Diluent. Sonicate to dissolve. Keep the sample at about 80 °C. For 4 hours. Use this solution as system suitability solution.

- **Preparation of placebo solution[8]**
  Accurately weigh and transfer powdered content of placebo equivalent to 100mg of Drug into 100 mL volumetric flask. Add about 70 mL diluent and sonicate for about 15 min. dilute to the volume and mix. Filter the solution through 0.45µm filters.
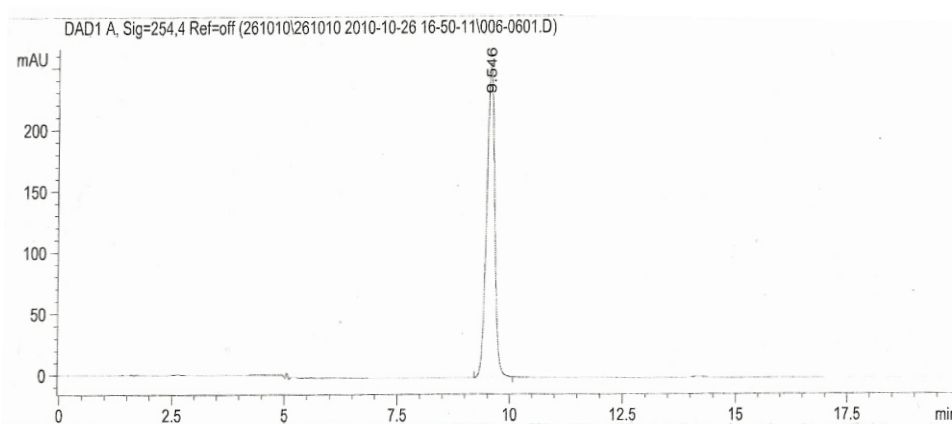
Preparation of diluents: Use mobile phase as diluents

As in Figure-2

Mobile phase = Methanol: Buffer pH 4.0 (96:4)

Mobile phase = Methanol: Buffer pH 4.0 (96:4)

Table 1: Chromatographic conditions for the optimized method for Simvastatin

S. No.	Parameters	Description
1.	Instrument	A HPLC instrument (Younglin series) with Model Acme-9000
2.	Column	Promosil C-18, (250 mm, 4.6 mm, 5µm)
3.	Mobile Phase	Mix 5ml of glacial acid in 1000ml of water. To 1000ml of 5ml glacial acetic acid solution, add 0.94gm of 1-Hexane sulphonic acid anhydrous. Sonicate to dissolve. The mobile phase is prepared by mixing buffer: methanol in the ratio of 96:4 Filtered and degas it.
4.	Flow Rate	1.0 mL/minute
5.	Detection wavelength	254 nm
6.	Injection Volume	10µL
7.	Run Time	20 Minutes

Figure 2: Representive chromatograms of standard solution of Simvastatin

METHOD VALIDATION

The developed method was validated according to ICH guidelines. Standard calibration curve were prepared in the mobile phase with 5 concentration ranging from 1-50 µg/ml for simvastatin is injected in to HPLC system keeping the injection volume constant. The peak area was plotted against the corresponding concentration to obtain the calibration graphs. To study the reliability and suitability of developed method, recovery experiments were carried out at three levels 80, 100 and 120%. Known concentration of Commercial tablet is spiked with known amounts of simvastatin. At each level of the amount six determinations were performed and the results obtained were compared with expected results. Recovery for pharmaceutical formulations should be within the range 100±5%. The percent R.S.D of individual measurements was also determined. Precision of the assay was determined by repeatability (intra-day) and intermediate precision (inter-day) for 3 consecutive days. Three different concentration of simvastatin were analysed in six independent series in the same day (intra-day precision) and 3 consecutive days (inter-day precision). The repeatability of sample application and measurement of peak area for active compounds were expressed in terms of percent RSD.

All chromatograms were examined to determine if compounds of interest co-eluted with each other or with any additional excipient peaks. Marketed formulation were analysed to determine the specificity of the optimized method in the presence of common tablet excipients. Limi of detection (LOD) and limit of quantitation (LOQ) were estimated from single to noise ratio. LOD and LOQ were calculated using $3.3\sigma/s$ and $10\sigma/s$ formulae, respectively. Where σ the standard deviation of the peak areas and s is is the slope of the corresponding calibration curve. To evaluate robustness of HPLC method a few parameters included variation of flow rate, percentage of buffer in the mobile phase, and pH of mobile phase.

RESULT AND DISCUSSION

The retention time of simvastatin was peak eluted at 9.546 min. The peaks are well separated with a resolution of 3.344 and Tailing 1.136.

The mobile phase comprises of Methanol: Buffer (96:4) v/v at the pH 4.0 was selected as optimized mobile phase, because of the high purity, symmetry, proper tailing, high area and low RT value at same concentration as compared to other trail mobile phase. Furthermore, the stability of the drug in the mobile phase were also studied and result indicating that the drug Simvastatin was found to be stable during the storage time of 48 hr (Table 2).

Linearity of the method was investigated by serially diluting the working standard to give a concentration range of 1-10 µm/ml and 20 µl from this was injected. The flow rate was maintained at 1 ml/min. temperature of column was kept ambient and the effluent was monitored at 254 nm. Calibration curve was constructed by plotting concentration against peak area (fig.3).

Table 2: Stability of the simvastatin in the optimized mobile phase

S. N.	Storage conditions	Mean area± SD (At zero hrs)	SE	Mean area± SD (At 48 hrs)	SE
1.	Room Temperature (25±0.5 ^{0}C)	252952.3±2859.82	3474.9	248844.7±3650.63	2777.53
2.	Refrigerator (4±0.5 ^{0}C)	252952.3±2859.82	3474.9	250011.3±3653.10	2778.63

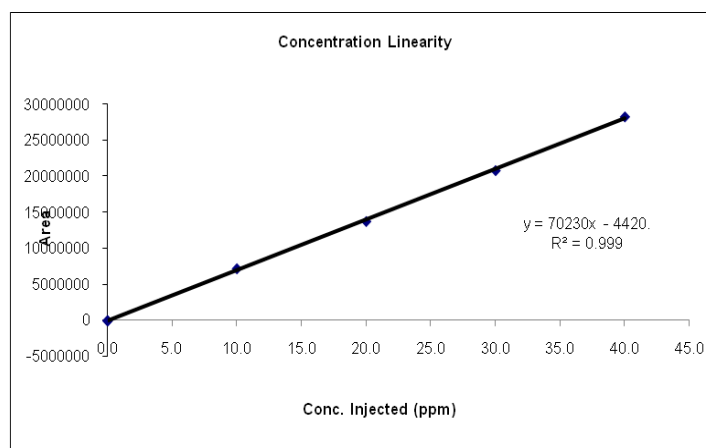***Concentration of drug 10µg/ml in mobile phase.**

Figure 3: Standard Curve of Simvastatin

The method was validated for linearity, precision, accuracy, specificity, limit of detection and limit of quantification as per ICH guidelines. All parameters are validated as per ICH guidelines.

Optimum condition of mobile phases was investigated in the development of an HPLC method suitable for analysis of in the bulk drug. These included Methanol: Acetonitrile: Buffer (50:20:30) (% v/v), Methanol: Acetonitrile: Buffer (60:20:20), Methanol: buffer (50:50), Methanol: Buffer (70:40), Methanol: Buffer pH 4.0 (90:10), and Methanol: Buffer pH 4.0 (96:4).The same solvent mixture was used for extraction of the drug from the formulation containing excipients. The retention time of Simvastatin obtained was 9.546 ±.234 (1). The system suitability tests for HPLC were carried out on freshly prepared solution of Simvastatin (10 µg/ml) and parameters were studied. The results were summarized in Table 3.

Table 3: System sutability test for Simvastatin:

S. No.	Parameter	Value
1.	Retention time, min	9.546±0.234
2.	Tailing factor	1.136±0.274
3.	Asymmetry factor	1.139±0.864
4.	Theoretical plates	5642±0.426
5.	Resolution	3.344±0.628

Assay of tablets of Simvastatin were perfomed. Twenty tablets of each company of strength 5 mg, 10 mg and 20 mg were weighed and ground to a fine powder. A quantity of tablet powder equivalent to 10 mg of Simvastatin was transferred to 10 ml volumetric flask, dissolved and diluted with acetonitrile and water mixture to obtain 1 mg/ml. The solution was sonicated for 15 minute and filtered through 0.45 µm membrane filter. The solution was further diluted to obtain concentration 10 µm/ml. Peak area of the above prepared tablet solutions of Simvastatin were measured by using above mentioned chromatographic conditions and the amount of Simvastatin were found from regression equation (Table 4) &Recovery study (Table 5).

Table 4: Results of Analysis of Commercial Tablets of Simvastatin

Tablet Formulation	Label claim(mg)	% Label claim estimated* (Mean ± S.D.)	% Coeff. Of Variation	Standard error
I(SIM)	5	99.435 ± 1.243	1.365	0.514
II(SIMCARD)	10	99.754 ± 1.509	1.523	0.625
III (SIMCHOL)	20	99.246 ±1.427	1.305	0.613

*Average of six determinations

Table 5: Recovery Studies of Commercial Tablets of Simvastatin

Tablet Formulation	Label claim (mg)	Drug added (mg)	% Label claim estimated* (Mean ± S.D.)	% Coeff. of Variation	Standard error
I(SIM)	5	2.5	99.316 ± 1.513	1.496	0.743
II(SIMCARD)	10	5	99.514 ± 1.397	1.432	0.574
III (SIMCHOL)	20	10	99.288 ±0.863	0.798	0.465

*Average of six determinations

The linear regression date showed a good linear relationship over the concentration range of 1-50 μg/ml as summarized in Table 6. The limit of detection (LOD) and the limit of quantification(LOQ) of the drug were found by scanning the solution of Simvastatin having different lower concentrations and the LOD and LOQ were found to be 0.5 and 1 μg/ml indicates that method is sensitive (Table 6). The intraday and interday precision were determined by analyzing standard solution of Simvastatin at three different concentration levels (6, 8, 10 μg/ml). The % RSD for intraday and interday precision was found to be 0.257 – 0.712% and 0.480-1.080% respectively which indicate that method is precise (Table 6).

Table-6: Stastiscal Data & Regression Equation for Simvastatin

S. N.	Parameter	Value
1.	λ_{max} (nm)	254
2.	Beer's range(μg/ml)	5-40
3.	Molar absorbtivity (l/mol/cm)	4.327×10^4
4.	Correlation coefficient (r^2)	0.999
5.	Regression equation	Y=0.70230X-4420
6.	Intercept (a)	04420
7.	Slope (b)	0.70230
8.	Limit of detection (LOD μg/ml)	0.126
9.	Limit of quantification(LOQ μg/ml)	0.406
10.	Linearity	1 – 50
11.	Accuracy%	99.16 to 101.24
12.	Repeatability (RSD, %, n=6)	0.195
13.	Precision (RSD, %), Interday (n=6)	0.480-1.080%
14.	Intraday (n=6)	0.257 – 0.712%

Repeatability of the method was studied by injecting 10 μg/ml solution of Simvastatin for six times and peak area was measured and % RSD was calculated which was found to be 0.195 shows repeatability of the method (Table

6). Accuracy of the method was evaluated by standard addition method in which appropriate portion of stock solutions of Simvastatin were spiked into blank placebo matrix to produce concentrations of 80 100 and 120% of theoretical concentration. The mean recovery of spiked samples obtained was in range of 99.16 to 101.24 reveals no interference of excipients and shows that method is accurate (Table 6).

The proposed validated method was successfully applied to determine Simvastatin in tablet form. The results obtained for tablets of Simvastatin were comparable with the corresponding labeled amounts (0.5 mg/tab) (table 4). Robustness of the method was estimated by changing the mobile phase composition (3±3), wavelength ±1 nm, injection volume (20±2μl), column temperature (40±3^0) and RSD values for all these changes calculated were less than 2 indicate that proposed method is robust. The proposed RP-HPLC method was accurate, precise, sensitive and rapid. The method also can be extended for the routine analysis of Simvastatin in tablet dosage form.

CONCLUSION

It is thus concluded that the proposed method is new, simple, cost effective, accurate, safe, free from pollution and precise and can be successfully employed in the routine analysis of these drugs in pharmaceutical dosage forms. The proposed method shall prove equally effective to analyze Simvastatin in the corresponding drug sample and may prove to be of great importance in pharmaceutical analysis.

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