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## Heavy metal redistribution in soils using compost as a soil amendment

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**Heavy Metal Redistribution in Soils  
Using Compost as a Soil Amendment**

**Jason T. Morrison**

**Thesis Submitted to the College of Agriculture, Forestry, and Consumer  
Science at West Virginia University in Partial Fulfillment of the  
Requirements for the Degree of**

**Masters of Science  
in  
Agricultural and Environmental Education**

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**Division of Resource Management**

**Morgantown, West Virginia  
2000**

**Keywords: Phytoremediation, Heavy Metal, Composting**

## **Abstract**

### **Heavy Metal Redistribution in Soils Using Compost as a Soil Amendment**

**Jason T. Morrison**

Organic soil amendments such as compost can reduce the availability of heavy metal contaminated soil. The objectives of this study were 1) to evaluate the efficiency of compost as an soil amendment within heavy metal contaminated soil 2) to determine the hypoaccumulation of heavy metals in the stem, shoot, and root systems of white clover and wheat 3) to identify movement of heavy metals within the soil 3) to evaluate the differences of nodule formation in *Trifolium repens*. Three volumes of compost were added to a heavy metal contaminated soil (additions of zero, 10, and 40 percent compost, by volume). Zinc, Cd, and Pb contamination was analyzed in soil fractions and the roots and shoots of wheat (*Triticum aestivum*) and white clover (*Trifolium repens*). The increased addition of compost was found to alleviate the toxic effects that heavy metal have on plant health; while redistributing metals to a less available form. An increase of nodule formation and nitrification resulted with the addition of compost to heavy metal contaminated soil.

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# **CHAPTER 1**

## **Introduction**

Heavy metal contamination of soil poses a major threat to human health and the environment. Toxic heavy metal contamination is prevalent within the surface soils at mining and industrial sites (Huang et al., 1997). Heavy metal pollution is a widespread problem within all industrially developed countries of the world. Past waste disposal practices associated with mining and manufacturing activities have been such that air, soil, and water contamination was common, and as a result there are many metal contaminated sites that pose serious health risks (Roane et al., 1997).

Traditional approaches to remediation of toxic heavy metal contaminated soils are typically expensive, labor intensive, and environmentally inefficient. Available space at hazardous waste disposal sites and growing regulations are rapidly depleting incineration and landfilling processes as remediation options. Land-ban regulations, such as the California List, bans the incineration and disposal of corrosive heavy metals. Landfill disposal costs were between \$25 - \$60 per drum in 1983 increased to \$64 - \$186 per drum in 1987. Incineration costs of highly toxic substances increased from \$1.50 - \$3.10 per gallon in 1983 to \$2.36 - \$5.02 in 1987 (Forcella, 1989).

The use of plants to remove hazardous heavy metal contamination from soil is a promising alternative of method soil remedation. Soil amendments can aid in the increase or decrease of the accumulation efficiency of toxic heavy metals within plants. The distribution of metals is significantly influenced by soil pH and organic matter content (Zang et al. 1997; Tagwira et al. 1993).

Synthetic and natural soil amendments that increase of the availability of heavy metals enhance plant up-take. Chelates have been used in soils and nutrient solutions to increase the solubility of metal cations in plant growth media and are reported to have significant effects on metal accumulation in plants (Huang et al. 1997). The addition of these amendments release heavy metals, which are bound to soil particles. The release of these metals results in the increase a heavy metal availability, which will additionally increase the risk of heavy metals leeching into the water system.

Organic soil amendments can ameliorate metal toxicity to plants by redistributing metals to less available fractions. Phosphates such as apatite amendments have been successfully used to lower the bioavailability and increase the geochemical stability of metal contaminated soil (Laperche et al., 1997). Compost amendments are also used as viable soil amendment within these processes.

Compost and other amendments can be used as a vital tool to restrict the availability of heavy metals in soil. Organic amendments aid in a binding process that occurs if high pH levels are maintained. Heavy metals bind to the surface of soil particles, resulting in a reduction of availability. The addition of compost to decrease plant accumulation of heavy metals was chosen as an alternative to traditional soil amendments.

#### Heavy Metal Contaminated Soil - Site Background

The contaminated soil used for this experiment was collected from a playground owned by the Harrison County Commission in Spelter, West Virginia. The community of Spelter has a significant heavy metal contamination in its soils. The contamination originated from the Zinc Spelter Plant, which is located adjacent to and up-wind of the

Harrison County owned playground. The Zinc Spelter Plant was operated as a zinc reduction plant from 1911 to 1972. Grasselli Chemical Company constructed and operated the facility in 1910 until it became a division of E. I. DuPont Corporation. It was purchased from the E. I. DuPont Corporation by Matthiessen & Hegeler Zinc Company in 1950 and operated until 1972.

Approximately 50 acres of the site is covered by waste generated from the zinc reduction process. The waste piles ranges in depth from five to 40 feet. The reduction of zinc was performed in a horizontal and vertical furnace. Zinc sinter (zinc oxide) was ground and mixed with pulverized coal, clay, tar and then briquetted within a furnace. When the zinc reduction was complete the briquettes were removed from the furnace and placed on the waste pile.

Contaminants resulting from the zinc reduction procedure included aluminum, antimony, arsenic, barium beryllium, cadmium, calcium, chromium, cobalt, copper, iron, lead, magnesium, manganese, nickel, potassium, selenium, silver, sodium, vanadium, and zinc. Most of the waste pile consists of intact or partially intact briquettes. Pulverized coal that was not consumed during the reduction process is also present throughout the site (WVDEP, 1997).

### Objectives

Research has indicated the efficiency of compost as an effective soil amendment of heavy metal contaminated soil. However, research has not determined the effects of hyperaccumulation, pH, nodule formation, and availability variations within compost mixed with heavy metal contaminated soil. Thus, this study was developed with the following *specific objectives*:

- 1) to evaluate the efficiency of compost as an soil amendment within heavy metal contaminated soil.
- 2) to determine the hyperaccumulation of Zinc (Zn), Cadmium (Cd), and Lead (Pb) in the stem, shoot, and root systems of white clover (*Trifolium repens*) and wheat (*Triticum aestivum*).
- 3) to identify movement of Zn, Cd, and Pb within the soil.
- 4) to evaluate the differences of nodule formation in white clover.

## CHAPTER 2

### Literature Review

#### Traditional Methods of Remediation

Several ex-situ and in-situ options exist to remove toxic heavy metal contamination from soil. Ex-situ processes such as incineration and landfilling are traditional methods, which require the soil to be physically removed from the site. Excavation and landfarming are traditional alternatives for the removal of heavy metals. In-situ recovery processes, such as chemical degradation and soil washing are often difficult. Recovery processes include using flocculants to separate the metals from soil, adding resins to absorb the effluents, or the mixing of clay amendments (Watanabe, 1997).

- Excavation

Traditionally, excavation and transport of contaminated soil to an off-site landfill has been a commonly selected alternative, often due to regulatory agency pressure. However, the cost can exceed \$300 per cubic yard making large disposal efforts very expensive. This alternative also is restrained by the physical limitation of the excavation equipment. Excavations deeper than approximately 20 feet have increased equipment cost, sheeting, and risk of undermining nearby structures, therefore excavation is usually limited to small contaminated soil cleanup projects. Increased contaminated soil volume generally warrants research into treatment alternatives. The physical removal of the soils from the ground through mechanical manipulation may change the physical characteristics of the soils (Shaw et al., 1995).

- Landfarming

In landfarming applications contaminated soil is removed and treated at an off-site location (ex-situ). The soil is spread over a large area until remediation processes are completed. Several remediation options exist. Chemical additives, soil washing, and phytoremediation methods can be applied to the contaminated soil. Remediators are able to regulate cleaning efforts at a greater efficiency due to the controlled environment present at landfarming sites. However, the cost of landfarming is high and it is a labor-intensive process. Contamination of the remediation site is also a drawback to landfarming applications.

- Chemical Degradation

Chemical degradation is a process where oxidizing or reducing agents are injected into the contamination zone to degrade the contaminant into other chemicals. This oxidation-reduction reaction is performed to degrade the chemical to a by-product that has more favorable characteristics, including lower toxicity, increased mobility, volatility, or absorption. The most common chemical degradation is capable of oxidizing many compounds in the following classes: halogenated hydrocarbons, halogenated aromatics, pesticides, and heavy metals (Shaw et al., 1995).

The chemical reactivity of a contaminant, and its mobility within the soil during application of the oxidizing-reducing agent, will be a factor of the ultimate fate of a chemical. The interaction between dissolution and sorption at a site complicates the remedial design and frequently gives reason to eliminate this technology as a remedial alternative. In addition, unfavorable by-products are often the result of chemical degradation. These by-products may have decreased volatility or adsorption

characteristics, making them more difficult to remove following chemical degradation (Shaw et al., 1995).

- Soil Washing

Soil washing utilizes the solubility of a substance in water to transfer it from the soil matrix into the groundwater. Typically, an underground piping network or aboveground bermed area or pit is constructed spanning the dimensions of the contaminated area. A water flow is maintained through groundwater recovery wells. The groundwater is recovered through groundwater recovery wells. The groundwater is then pumped to the surface and treated. Subsequently, the treated groundwater may be reinjected into the contaminated soil area.

Water was commonly used as the flushing solution. Recently, other agents, such as acids, chelating agents and phosphates, have been used to enhance the flushing rate of a chemical from the soil matter. Soil washing may be described as an accelerated precipitation and percolation process. This description is based on large quantities of water generally required to remove the contaminants from the soil matrix, compared to the low quantities of precipitation rates received in most locations. If a soil washing system is designed properly, the overall length of time for groundwater recovery and treatment can be reduced significantly (Shaw et al., 1995). A major consideration is the capacity of the groundwater recovery system. Measures must be established to verify that the groundwater recovery system will recover all the contaminants that are flushed from the soil. Additionally, the treatment system must be designed to handle the added contaminant loading.

- Microbial Remediation

Microbial systems are commonly used to remediate domestic waste, and to recover precious metals from ore. Biohydrometallurgy uses microorganisms in the recovery process of precious metals, such as copper and gold. Additional processes include the use of bacteria, algae, fungi, and yeasts to complex or sorb metals. Scott et al.(1988) concluded that the microorganism additions increase metal complexation. A profound issue in metal remediation is that through microbial action, metals can readily be re-mobilized, creating toxicity issues in sites where metals are not completely removed (Roane et al., 1997).

- In-situ Bioremediation

In-situ bioremediation is treatment in place. Good soil permeability is important for bioremediation to work but may also allow the petroleum hydrocarbons to penetrate too deeply. Practical limits are about 20 feet in depth. This is the least expensive and least labor intensive method. This method has a large advantage of not requiring movement of the soil. The disadvantage is that materials are not mixed and agitated to uniformity. Treatment times can range from weeks to years, depending upon contamination levels and depths of contamination. In-situ methods offer only limited assurance that the site has been treated to acceptable levels due to the difficulty in obtaining a representative sample.

### Phytoremediation

Phytoremediation is a natural process carried out by plants that are able to live in a contaminated media. Plants are able to either absorb high levels of contaminants within their roots or reduce their availability, essentially rendering them harmless (Watanabe,



1997). Phytoremediation has been tested on sites contaminated by petroleum products, munitions, radionuclides, abandoned mines, wood treatment, and sewage treatment.

Phytoremediation experts say the growth of interest in the field is driven by its relative cost-efficiency compared to standard remediation methods for government-mandated site cleanup (Watanabe, 1997). In situ remediation of a contaminated site may cost between \$10 and \$100 per cubic meter, whereas ex situ remediation may cost as much as \$30 to \$300 per cubic meter. In comparison, phytoremediation may cost as little as \$0.05 per cubic meter (Cunningham, 1997).

The extractions of heavy metals from soils are induced by hyperaccumulating plants. The plants store high levels of toxic heavy metals within their roots, shoots, and leaves. Hyperaccumulating plants must have the ability to accumulate heavy metals, have a high biomass production, and disease and pest resistance in order to flourish within toxic heavy metal contaminated soil.

Baker and Brook (1989) defined metal hyperaccumulating plants as those, which contain more than 1000 milligrams per gram (mg/g) of cobalt, copper, chromium, lead, or nickel or 10,000 mg/g (1%) of manganese or zinc. According to the U.S. Department of Energy, efficient hyperaccumulators have the following traits: high accumulation rates (even if low contamination levels exist); ability to accumulate several metals; fast growth; high biomass production; and resistance to diseases and pests.

Heavy metal contaminated plants should be harvested from the contaminated site before the plants reach sexual maturity. Hypoaccumulation can occur between the plant and species of animals or insects that consume it. A plant that reaches sexual maturity has a greater risk of insect exposure during pollination.

The traditional method of disposal of contaminated plants is incineration. Extraction of metals from the plant tissue is a new and innovative method of recycling heavy metal contaminated plants.

- Soil Amendments

A wide variety of soil amendments are used to increase and lower the availability of heavy metals in phytoremediation procedures. Phosphates such as apatite amendments have been successfully used to lower the bioavailability and increase the geochemical stability of metal contaminated soil (Laperche et al., 1997). Toxic heavy metals bind to soil particles, which reduce phosphates. Ma et al. (1993), studied in situ processes of lead (Pb) immobilization in contaminated soil and wastes using apatite. Results indicate the effective removal of Pb from aqueous solution, exchange resins, and Pb-contaminated soil material and suggests that apatite has the potential to immobilize Pb in situ in Pb-contaminated soils and wastes.

Organic soil amendments will lower the availability of Zn and increase the availability of most other heavy metals. Solid organic matter and organic matter in solution will absorb metals. Organic soil amendments can ameliorate metal toxicity to plants by redistributing metals to less available fractions. The association of heavy metal with organic matter varies directly with soil pH (Shuman, 1999).

The distribution of metals is significantly influenced by soil pH and organic matter content (Zang et al. 1997; Tagwira et al. 1993). The up-take of many heavy metals can be increased by the reduction of pH within soil. Heavy metal bioavailability increases with the decrease of pH. Decreasing pH will in turn increase the solubility of metals.

Liang (1997) reported the high pH of the compost affected the solubility of the metal hydroxide and carbonates, and low pH values increase the solubility of heavy metals. The pH of soil can be a direct determinant of the type of association heavy metals have with organic materials. Lower soil pH levels with the addition of organic amendments will increase metal availability to plants. Organic amendments are used as an effect tool to reduce plant up-take if high soil pH levels are maintained.

High levels of soil pH can be maintained by liming the soil. Hydrated lime is used with the addition of organic materials to effectively restrict heavy metal mobility within soil (Paff et. al., 1995).

Amendments such as compost and chelating agents are used to separate heavy metal from soil particles, which increases the metal's availability within soil. Chelates have been used in soils and nutrient solutions to increase the solubility of metal cations in plant growth media and are reported to have significant effects on metal accumulation in plants (Huang et al. 1997). The long-term effects of chelates within soil have been found to be detrimental to the environment. Chelating agents are more expensive than compost amendments and require careful management (Huang et al. 1997).

- Compost

Organic amendments such as compost shifts heavy metals to a more available form, if a high level of pH is maintained. Metals react with organic materials forming complexing bonds. Composting is a biological process in which microorganisms convert organic materials such as manure, leaves, paper, and food wastes into a soil-like material called compost (Rynk et.al, 1992). The addition of compost to decrease plant

accumulation of heavy metals was chosen as an alternative to traditional soil amendments.

Shuman (1988) studied the effect of organic matter on the distribution of manganese (Mn), copper (Cu), Iron (Fe), and Zinc (Zn) in soil fractions. Increased additions of organic matter caused manganese and iron to move from the less soluble forms to more plant-available forms. The probable mechanism for this movement was a reduction of oxidation. The addition of fresh organic matter causes a flush of microbial activity, which adds complexing agents to the soil and affects the redox status. Results demonstrate that accumulated organic matter near the surface can increase plant availability of manganese and iron and possibly decrease availability of zinc by causing a redistribution of elements between soil fractions. A further study concluded that natural soil matter and organic material added to contaminated soil during remediation decreases Zn availability to plants (Shuman, 1999).

- Bioavailability of Heavy Metals

Toxic heavy metals bind to cell walls or the cell surface of soil. Soil amendments, such as compost, aid in the release of these bond metals. Humic and fulvic acids present within compost increase the release of metals from contaminated soil. Humic acids are compounds which are composed of dark organic substances that are precipitated upon acidification of a basic extract from soil (Rynk, 1998). Humic and fulvic acids are both fractions of humic substances or humus. Humus is a material comprised of many organic substances and represents a state during the decomposition of organic materials (Epstien, 1997).

Simeoni et al., (1984) reported that the addition of a compost produced high yields of zinc (Zn) availability. Zn can be found in five major forms in soil: 1) in soil solution as ionic or organically complexed species; 2) on exchange sites of reactive soil components; 3) complexed with organic matter; 4) occluded in oxides and hydroxides of Al, Fe, and Mn; and 5) entrapped in primary and secondary minerals (Iyengar et al. 1981). The amount of Zn forms varies considerably in soil. White (1957) studied the distribution of Zn in 14 Tennessee soils. Results indicated that 1 to 7% of the distribution of Zn was on exchange sites of clay minerals, 10 to 40% was held by organic matter or in lattice positions of clay minerals, 20 to 45% was held on lattice position of clay minerals, and 30 to 60% was associated with Fe-oxides. Shuman (1979) reported that 1 to 7% of the Zn in ten Georgia soils was in the bioavailable form and 12 to 20% were in the organic and noncrystalline Fe-oxide forms.

The distribution and bioavailability of heavy metals vary, depending on the soil fraction. Iyengar et al. (1981) reported that water soluble, exchangeable, and absorbed fractions are readily available to plants; while primary and secondary soil minerals are relatively unavailable to plants.

The metal species present and their relative bioavailability within the soil is of greater importance than the total metal concentration in the environment that determines the overall physiological and toxic effects on biological systems (Bernhard et al., 1986; Hughes et al., 1989; Morrison et al., 1989). The efficiency of remediation processes is determined, in large part, by the redox potential and pH. Redox potential is established by oxidation-reduction reactions in the environment. Oxidation-reduction is a reduction involving the transfer of electrons from one molecule to another. The redox potential is a

measure of the tendency of a substance to donate or accept electrons respectively (Lim, 1998). Reducing conditions often result in metal precipitation; while oxidizing conditions promote the increased solubility of metals in a free ionic form. Oxidation conditions can decrease the pH within soil.

- Hyperaccumulating Plants

Hyperaccumulators are plants that can absorb high levels of contaminants with their roots and concentrate them either within their roots, shoots, and leaves (Watanabe, 1997). A variety of plant species is commonly used to remove heavy metal from soil. *Brassica juncea* (indian mustard) and *Thlaspi rotundifolium* (pennycress) are often used to remediate wet land areas (Blaylock et al., 1997). *Thlaspi* are known to efficiently accumulate zinc, lead, and cadmium. In addition, to zinc, lead, and cadmium, *B. juncea* has the ability to accumulate nickel. *Alyssum* (poplar) trees also have the ability to accumulate nickel. Crop species, such as *Zea mays* (corn) and *Pisum sativum* (pea) were successfully used to accumulate Pb (Huang et al., 1997). Ebbs et al. (1998) studied the effects of *Avena sativa* (oat) and *Hordeum vulgare* (barley) as hyperaccumulators in Zn contaminated soil. *Rudbeckia hirta* (black-eyed Susan), a perennial flower, accumulated significant amounts of Cd, Cu, Pb, and Zn (Sawhney et al., 1994). Plants from many other families have been shown to remove cobalt, copper, chromium, manganese, and selenium from contaminated soil (Watanabe, 1997).

- Nodulation of White Clover in Metal Contaminated Soil

Biological fixation of atmospheric nitrogen can be estimated at 175 metric tons per year, or about 70% of all nitrogen fixed on the earth each year. The major contributor of nitrogen fixation comes from symbiotic systems, such as, *Rhizobium* (legumes).

*Leguminosae* plants (*Rhizobium* and Legume association) have been estimated to contribute to 50% of all nitrogen fixed on the earth (Food and Agriculture Organization of the United Nations, 1993).

Nitrogen fixation and nodulation are restricted when grown in heavy metal contaminated soil. El-Kenawy et. al., (1997) studied the effects of Zn and Cd have on the nodulation of white clover. The weight of shoots decreased with the increase of Zn contamination. Cadmium was more toxic to plant growth than Zn. The process of nodulation was susceptible to the toxic effects of heavy metals.

Roots nodules generally grow best in the temperatures range of 25°-30°C, and in pH of six to seven (Vincent, 1970). *Rhizobia* bacteria resides within a healthy nodules. Root nodule bacteria are heterotrophic and able to utilize a wide variety of carbohydrates. Inoculation of rhizobia is applied to the white clover seedling during the initial planting. The infection process takes place within 48 hours of inoculation (Callaham, 1980). The growth and health of rhizobia are dependent on the availability of inorganic nitrogen.

- Sequential Extraction of Heavy Metal Contaminated Soil

The extractions of metal from soil involves multiple fractions of increase acidic strength. Sequential extraction procedures determine the degree in which metal are bond within soil particles. Soils that have metals that are readily available will release higher amounts of metal in earlier fractions. Several different sequential extraction procedures have been developed and are commonly used. Most procedure consists of water soluble, exchangeable, Fe, and Mn fractions. The solutions used within each procedure are quite different; however, the methods of extraction are similar.

Tessier et. al. (1979) introduced a procedure involving chemical extraction for the partitioning of particulate trace metals into chemical forms likely to be released in solution under various environmental conditions. McLaren, (1979) used two exchangeable fraction which consisted of  $\text{CaCl}_2$  and Hoac, respectively, with 24 hour agitation periods. Shuman, (1985) developed a extraction procedure to measure Zn. Amorphous iron oxide and crystalline iron oxide fraction were among the six fractions used. Berti, (1995) introduced a method that uses a combination of new and established techniques. Extracting solutions and their order were based on the fractioning several different elements and the use of several different soil types.

The differences between most sequential extraction procedures includes the choice of extraction solutions, the target fractions, the order in which the fractions are removed, the difference in soil to solution ratio, shaking times, and particle size of the sample (Berti et. al., 1995). The progression of new sequential extractions involved the development of more specific procedures for equally specific task. There is currently no standard sequential chemical extraction method. The technique of Tessier et. al. (1979) procedure is generally excepted because of its use by many others (Berti et. al. 1995).



## CHAPTER 3

### Materials & Methods

#### Collection of the Contaminated Soil for this Study

The contaminated soil sample was collected beneath a shelter overhang on the west side of a Harrison County owned playground. The playground was adjacent the Zinc Spelter Plant (source of contamination). The soil sample was collected from the top 30 cm of the A horizon within a one meter (m) by three meter area. A total of 45.42 liters (l) of soil was stored in 18.93 liter rubber containers.

AgRecycle Inc., of Pittsburgh, Pennsylvania provided 45.42 liters of yard waste compost. The compost was stored in 18.93 liter rubber containers. The soil and compost were air dried and sieved with a three mm plastic sieve and transferred to 2-liter pots.

#### Preparation of Test Pots for Growth Chamber

A pot study was conducted, using the metal contaminated soil, to determine metal uptake patterns. Two-liter pots were used as sample containers. Eighteen sample pots were prepared by adding 2.27 kg of soil to each pot. Six pots acted as the control. Ten percent and 40% compost additions (by volume) were added to each of 6 sample pots. The soil and compost additions were thoroughly mixed.

In order to maintain field capacity, distilled deionized water (DDW) was added until the soil reached its maximum saturation point and then allowed to drain for 24 hours. Five hundred ml of DDW was added to the control, 550 ml to 10%, 600 ml was added to 40% compost addition, and 800 ml of DDW was added to the potting soil. A 172.82 g sample was removed from each of the pots for pH analysis and placed within a

100 ml sealed container. The weight of each pot was then recorded. An Omega Model E2000D Balance was used for all procedures requiring a known mass with this study.

- Preparation of Seedlings for Test Pots

Wheat and clover seeds were germinated under germination lights in acid washed sand. The two plant species remained under the germination lights for ten days. After the ten-day period the plants were transferred to pots. Each plant was rinsed with DDW. Six wheat seedling or ten clover seedlings were transferred into each pot. The plants were placed in the center of each pot, approximately five mm apart.

- Treatment of Pot Samples

After the transplanting process was completed were placed into a growth chamber to assure constant temperature, humidity, and light. The temperature remained at 20°C and the humidity remained constant due to a three liter humidifying unit, which was placed within the growth chamber.

The lights were pre set for 14 hours of light and 10 hours of darkness. The sample containers were weighed every three days. One ml of DDW was added to each two liters sample container for every gram lost between each three-day period. The average water loss weight was determined for each two liters sample container after three weeks.

pH was tested twice per week using a Fisher Scientific Accumet Model 15 pH meter. Five grams of soil was removed from each sample containers. Ten ml of DDW was added to each of the five ml samples.

- Collection of Plant and Soil Samples Pots

The pots were removed from the growth chamber after seven weeks. The plants and roots were removed from each pot. A 50 ml soil sample was collected from the

rhizosphere of the plants in each pot. An additional 50 ml bulk sample was collected from an outermost area of the pot. Rhizosphere and bulk samples were stored in a refrigerator until further use. The plants and root systems were separated and rinsed with DDW. The lower six inches of the wheat plants were separated from the upper portion of the plants. The plants were transferred into aluminum dishes and placed in an oven for four days at 75°C. The dried plant and root tissue was placed within one liter plastic bags and crushed.

- Sequential Extraction of Bulk and Residual Soil

One gram of bulk and rhizosphere soil was placed into separate 15 ml polycarbonate centrifuge tubes. An additional replication sample was prepared for every sample. A total of seventy-eight soil samples were prepared for sequential extraction procedures. Twelve ml of DDW was added to each centrifuge tube at room temperature. Water soluble, exchangeable, carbonate, Fe-Mn oxides, organic, and residual fractions were obtained using the procedure of Tessier et al. (1979).

*Water Soluble.* Soil sample extracted with 12 ml of DDW and shaken vigorously for two hours.

*Exchangeable.* The residue from water soluble fraction was extracted with 12 ml of 1 M MgCl<sub>2</sub> (pH 7.0) with one hour of vigorous agitation.

*Carbonate-Bound.* The residue from exchange fraction was extracted with 12 ml of 1 M NaOAc (adjusted to pH 5.0 with HOAc) with five hours of vigorous agitation.

*Fe-Mn Oxides-Bound.* The residue from carbonate fraction was extracted with 0.04 M NH<sub>2</sub>OH.HCl in 25% (v/v) HOAc at 96°C with occasional agitation for six hours.

*Organic-Bound.* The residue from Fe-Mn oxide fraction was extracted with 3 ml of 0.02 M HNO<sub>3</sub> and five ml of 30% DDW (adjusted to pH 2.0 with HNO<sub>3</sub>). The mixture is heated within an agitating water bath at 85°C for two hrs, with occasional agitation. A second three ml aliquot of 30% H<sub>2</sub>O<sub>2</sub> (pH with HNO<sub>3</sub>) was added and the mixture heated 30% H<sub>2</sub>O<sub>2</sub> (pH 2.0 with HNO<sub>3</sub>) was added and the mixture heated to 85°C for one hour with intermittent agitation. After cooling, five ml of 3.2 M HN<sub>4</sub>OAc in 20% (v/v) HNO<sub>3</sub> is added and the samples diluted to 20 ml and agitated continuously for 30 minutes.

*Residual.* The residue from organic fractions was digested using a HF, HCl, HNO<sub>3</sub> dissolution procedure. After each successive extraction, separation was done with an IEC Model K Centrifuge at 12K rpm for 15 min. The supernatants were removed and analyzed for Cd, Zn, and Pb with a Perkin Elmer Emission Spectrometer, Model P400. The residue was washed with 12 ml of DDW followed by vigorous agitation. The residue was then centrifuged for 15 min and the supernatants were removed before the next extraction. An Eberbach horizontal shaker was used for all procedures involving vigorous agitation. One hundred microliters (μl) of HNO<sub>3</sub> was added to each sample.

- Microwave Digestion

The residual soil and dried plant and root materials were respectively sequentially extracted and digested using a CEM Model MDS-2000 microwave. *Method SW 846-3051* was performed for all residual soil samples, while the *alfalfa method* was used for all plant and root samples (Table 3.1).

All samples that were microwave digested required 0.5 g of the sample and 10 ml of HNO<sub>3</sub> into each vessel. Seventy-eight residual and 60 plant and root samples were prepared. The microwave was programmed as follows:

**Table 3.1** – Microwave procedures used for soil and plant material.

<b>METHOD</b>	<i>SW 846-3051</i>	<i>ALFALFA</i>		
<b>Stage</b>	1	1	2	3
<b>%POWER</b>	100	100	100	100
<b>PSI</b>	85	40	85	150
<b>TIME</b>	10:00	6:00	6:00	10:00
<b>TAP</b>	4:30	3:00	3:00	5:00
<b>TEMPERATURE</b>	175	130	150	200
<b>FAN SPEED</b>	100	100	100	100

- Perkin Elmer Emission Spectrometer

Following the completion of the microwave digestion, the residual plant and root samples, bulk and rizosphere soil samples, and soil fractions were analyzed for Cd, Zn, and Pb using a Perkin Elmer Emission Spectrometer, Model P400.

## CHAPTER 4

### Results

#### Compost Amendment

Compost data indicated that Zn, Cd and Pb were present, but were, within national 503.13 standards (USEPA, 1993). The carbon:nitrogen ratio and soluble salt data were sufficiently within compost nutrient standards (Table 4.1). The pH was 8.15, which is above average for yard waste compost (Penn State, 1999).

**Table 4.1** – The analysis of compost used in this study (Penn State, 1999).

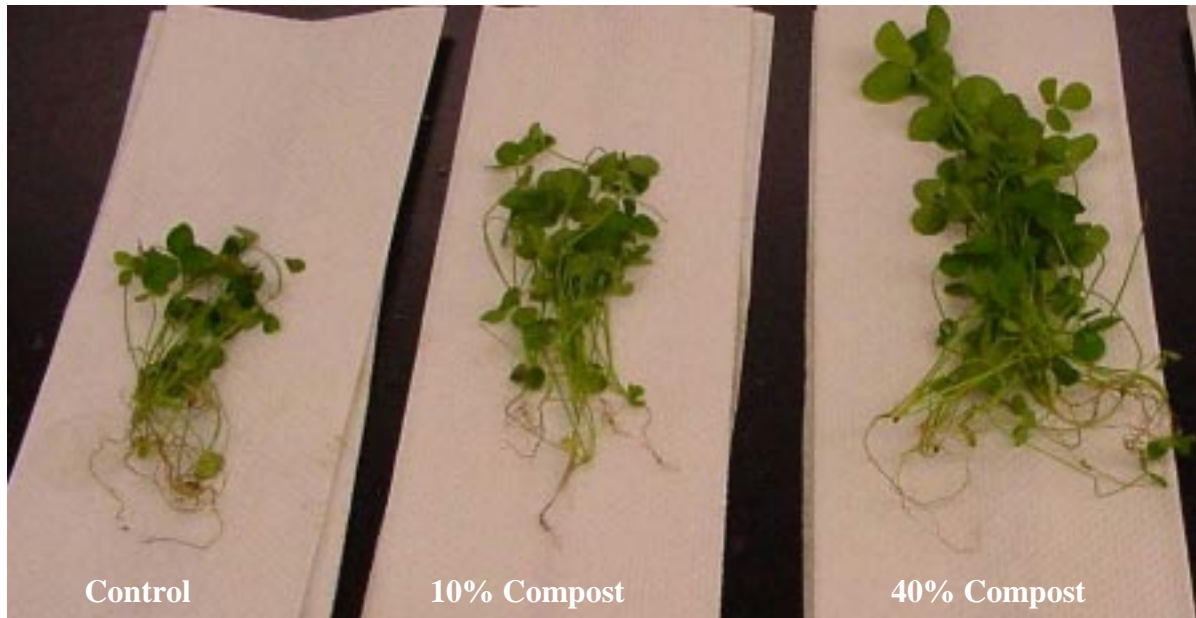
	(kg/1000 L)	<b>Heavy Metal Concentration</b>	(mg/kg)
Total Nitrogen (N)	6.71	Zn	89.43
Ammonia N	0.05	Cd	1.78
Calculated Organic N	6.66	Pb	72.4
Total Phosphate	2.48		
Total Potash	3.53		
		<b>(USEPA 503.13) Allowable Metal Contamination</b>	
pH	8.15	Zn	771
Total Nitrogen %	0.67	Cd	3.7
Total Organic Carbon %	13.33	Pb	324
Carbon:Nitrogen Ratio	19.8		
Soluble Salts (mmhos/cm)	1.13		

#### Plant Health in Heavy Metal Contaminated Soil

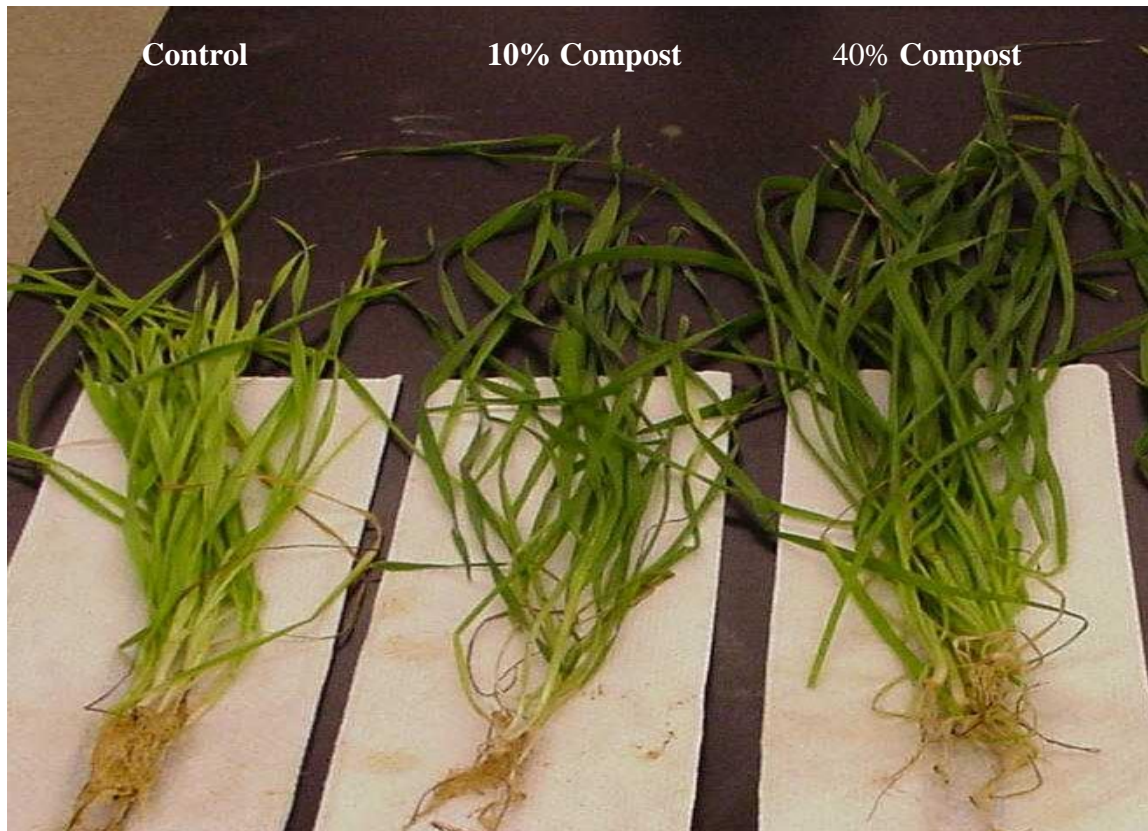
A considerable increase in growth occurred in compost amended soils. The addition of compost increased the growth levels. A measurement of the total length revealed that 10% compost (CM) wheat samples were 12% taller than control (CO) wheat; while, 40% CM wheat were 27% taller than CO wheat (Table 4.2). Visually, 10% CM and 40% CM white clover and wheat samples had more vigorous roots and shoots than CO (Figure 4.1 & 4.2).

**Table 4.2** – The increases plant growth efficiency is due to the addition of compost.

		Wheat		Clover	
		Shoots	Roots	Shoots	Roots
Control	dry wt. (g)	0.222	0.03	0.019	0.005
	St Dev	0.0126	0.009	0.014	0
	avg length (cm)	34.467	NA	NA	NA
	St Dev	1.595	NA	NA	NA
10% Compost	dry wt. (g)	0.286	0.032	0.021	0.004
	St Dev	0.037	0.019	0.01	0.001
	avg length (cm)	37.933	NA	NA	NA
	St Dev	1.457	NA	NA	NA
40% Compost	dry wt. (g)	0.458	0.034	0.066	0.007
	St Dev	0.068	0.013	0.029	0.002
	avg length (cm)	48.2	NA	NA	NA
	St Dev	2.696	NA	NA	NA



**Figure 4.1** – White clover samples indicated improved health and growth with the increased addition of compost to heavy metal contaminated soil.



**Figure 4.2** – The addition of compost to heavy metal contaminated soil improved root and shoot growth.

#### Nodule Formation of White Clover

The addition of compost increased the health and nitrogen fixing ability of the white clover. Nodules of CO white clover plants were small, abnormally formed, with no signs of nitrogen fixation. 10% CM showed improved nodule formation, health, and size; however, only a few nitrogen fixed nodules were present. The highest number of nitrogen fixing nodules were present in 40% CM. A considerable increase of size and general health existed within nodules of 40% CM (Figure 4.3).

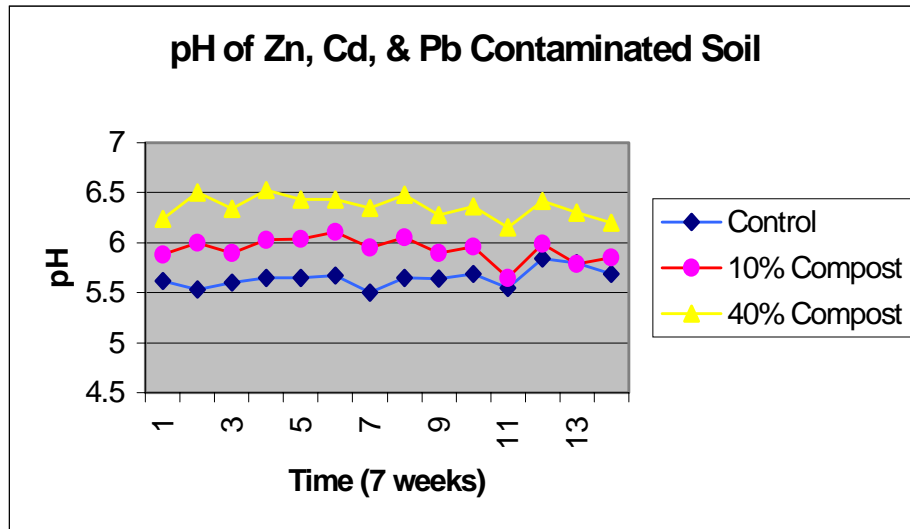




**Figure 4.3** – The addition of compost increased nitrogen fixing ability and size of the nodules on white clover roots.

#### pH of Heavy Metal Contaminated Soil

The pH of all sample pots was determined over the seven-week period within the growth chamber (Figure 4.4). The mean of the pH CO soil samples were 5.65, while the 10% CM was 5.96, and the 40% CM was 6.36.



**Figure 4.4** - High pH levels were maintained with the addition of compost in metal contaminated soils during the seven-week sample period.

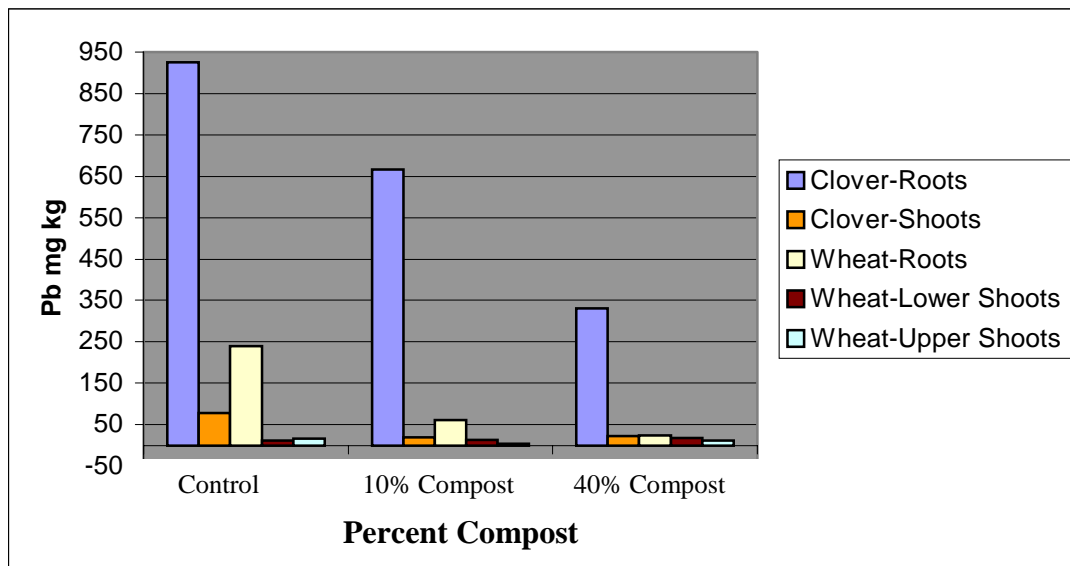
#### Heavy Metal Up-Take of Wheat and White Clover

CO had the largest concentration of Pb. Increased compost additions reduced the amount of Pb in the roots and shoots of white clover and wheat (Figure 4.5). The highest Pb concentrations were in roots of white clover. Data revealed that 925.83 mg kg<sup>-1</sup> Pb was present in CO clover roots; while, 40% CM clover roots had 331.27 mg kg<sup>-1</sup> Pb (Table 4.3).

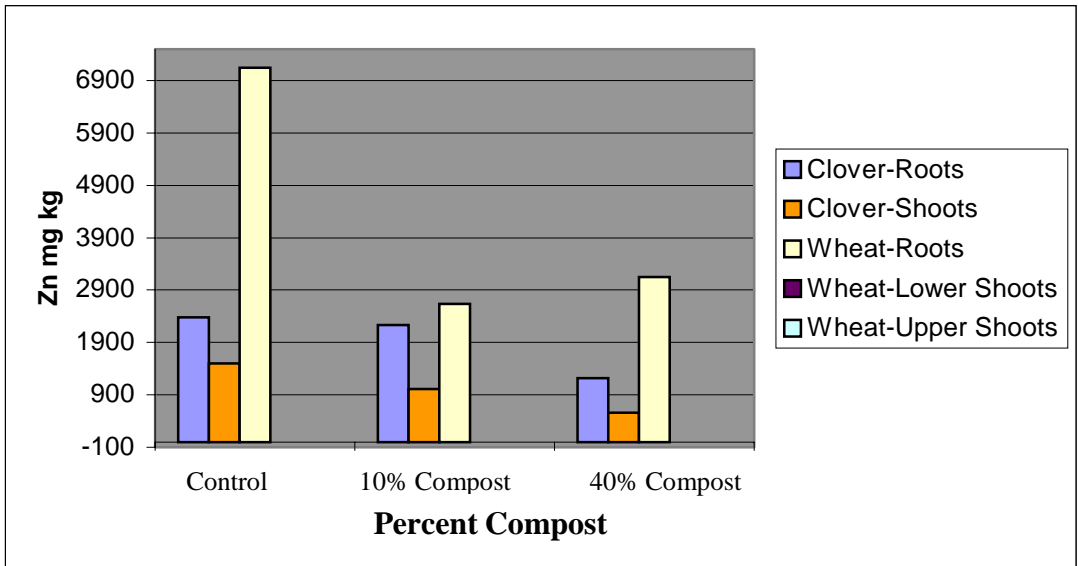
Zinc contamination was considerably higher than Pb or Cd concentrations. Wheat roots had the highest amount of Zn present (Figure 4.6). 10% CM wheat roots contained 520.93 mg kg<sup>-1</sup> less Zn than 40% CM. Clover had the ability to up-take larger concentrations of Zn than wheat. Levels of 1,014.26 mg kg<sup>-1</sup> were found in 10% CM clover shoots and 632.87 mg kg<sup>-1</sup> Zn was present in 10% CM wheat shoots (Table 4.3).

Data indicated that CO, 10% CM, and 40% CM wheat Zn in lower or upper shoot portions. With the exception of Zn shoots, compost additions resulted in a reduction of ability to up-take Zn in both wheat and clover plants.

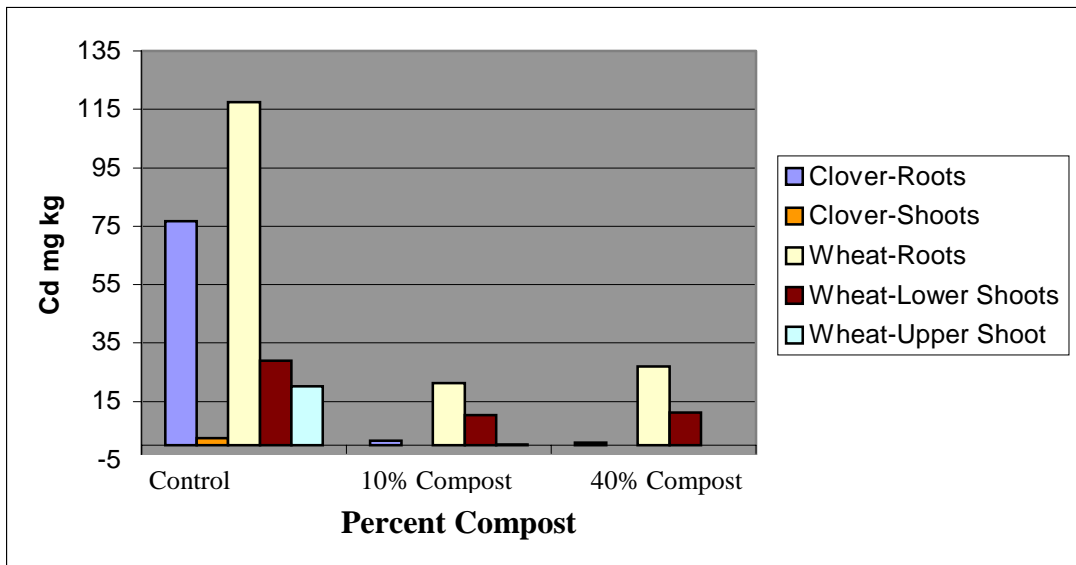
Compost addition reduced the up-take of Cd in of wheat and clover (Figure 4.7). Cadmium levels were lower than all other heavy metals analyzed. 10% CM wheat roots and lower shoots contained less Cd than 40% CM wheat roots and lower shoots. Compost addition had a more significant effect in reducing root up-take than shoots. 10% CM additions reduced Pb levels in wheat roots by 96.24 mg kg<sup>-1</sup>; while, reduction in wheat shoots were 19.33 mg kg<sup>-1</sup> (Table 4.3).



**Figure 4.5** – A reduction of Pb within the plant material of white clover and wheat occurred with the addition of compost.



**Figure 4.6** – A reduction of Zn within the plant material of white clover and wheat occurred with the addition of compost.



**Figure 4.7** - A reduction of Cd within the plant material of white clover and wheat occurred with the addition of compost.

**Table 4.3** – Wheat has the ability to up-take a higher concentration of heavy metals than white clover.

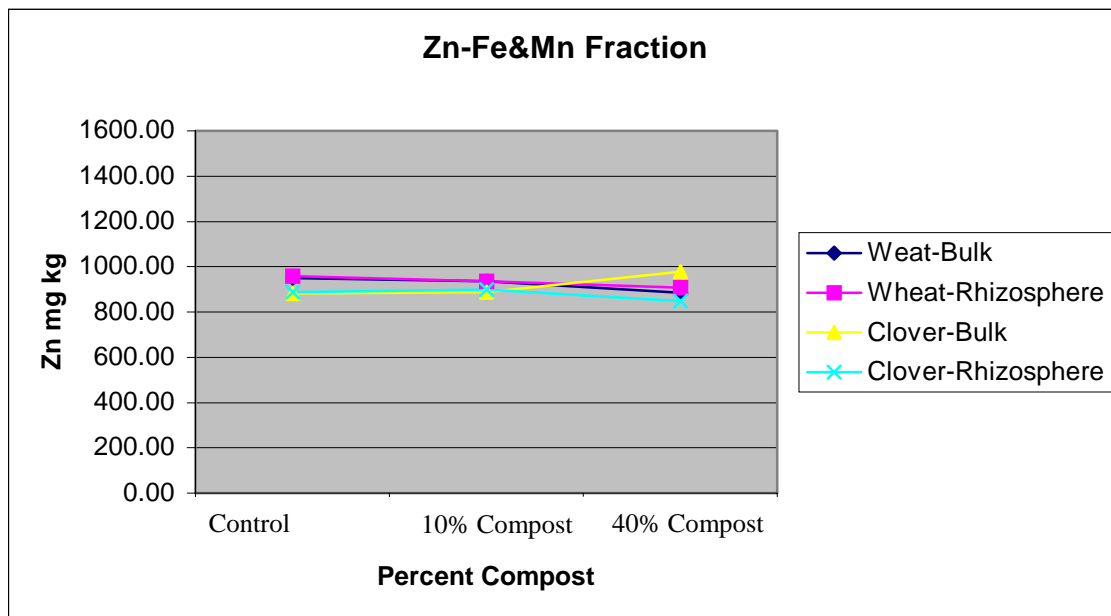
	(mg/kg)	Wheat		Clover	
		Shoots	Roots	Shoots	Roots
Control	Zn	1313.61	7148.51	1503.2	2379.17
	Cd	24.54	117.44	2.47	76.67
	Pb	14.045	239.84	77.95	925.83
10% Compost	Zn	632.87	2633.06	1014.3	2232.22
	Cd	5.22	21.2	0	1.67
	Pd	8.09	60.23	18.97	666.67
40% Compost	Zn	509.46	3153.99	558.67	1216.73
	Cd	5.63	26.92	0	0.093
	Pb	14.7	23.885	21.84	331.27

#### Sequential Extraction of Heavy Metal Contaminated Soil

No substantial differences or patterns were present between bulk and rhizosphere soil samples.

- Zn – Fe&Mn Fraction

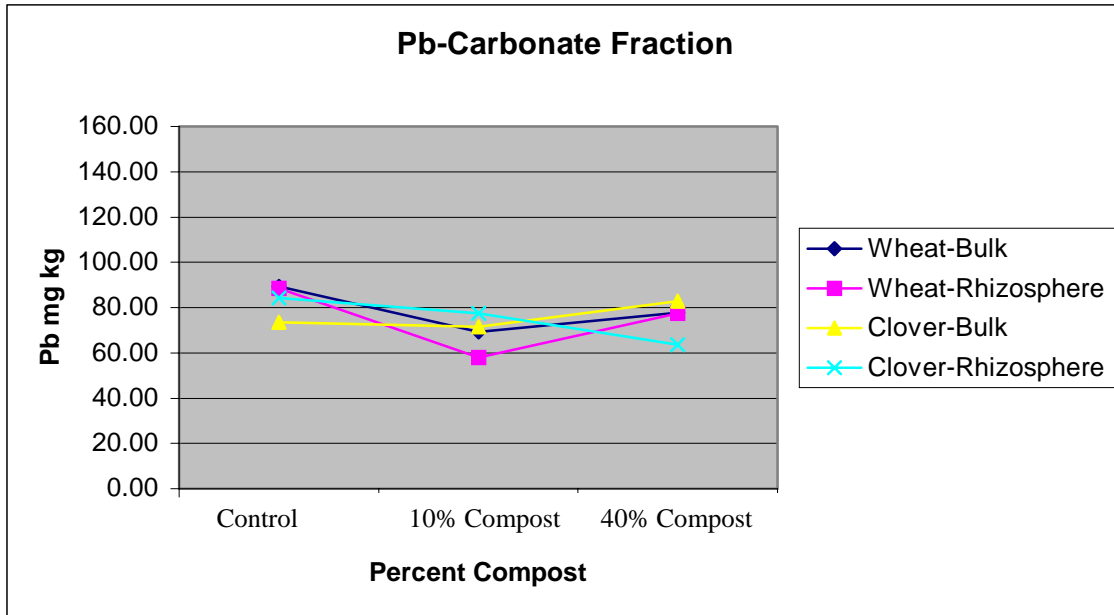
The Fe&Mn fraction had a gradual decline in the amount of Zn released with the increased addition of compost. Clover-bulk sample released 978.88 mg kg<sup>-1</sup>, however, the average amount of Zn released was 904.66 mg kg<sup>-1</sup>. Overall, 900.61 mg kg<sup>-1</sup> of Zn were released in 40% CM white clover and wheat Fe&Mn fractions. Data indicated that, on average, the largest amount of Zn was released in CO samples of both white clover and wheat soils (Figure 4.8).



**Figure 4.8** – The Zn analysis of the Fe&Mn fraction showed a decrease in soils from 40% CM.

• Pb - Carbonate Fraction

The dramatic release of Pb was present from CO to 10% CM in wheat soils. Ten percent CM wheat rhizosphere had lower levels of Pb than all other soil sample types. The amount of Pb within 10% CM wheat rhizosphere was  $57.85 \text{ mg kg}^{-1}$ . The average Pb release of 10% CM wheat and white clover was 6.41 lower than 40% CM samples (Figure 4.9).



**Figure 4.9** – Data revealed that Pb analysis of the carbonate fractions from 10% CM and 40% CM white clover and wheat samples were less available than CO fractions.

## CHAPTER 5

### Discussion

The addition of compost increased the total length of white clover and wheat. The increased growth of the plants was due to several changes that occur within low pH, clayey soils when compost is applied. The optimum pH of wheat and clover is six to seven. Compost raised pH levels into the optimum range. Soil structure and nutrient supplies were elevated by the compost additions. Compost also provided an increase in porosity, water holding capabilities, and aeration of the soil.

However, the addition of compost drastically reduced the amounts of Zn, Cd, and Pb up-take in both wheat and white clover. The average length of Control (CO) wheat plants were 27% smaller than 40% compost (CM); yet, the control still had the ability to up-take 28.02 % more Pb, 37.17 % Zn, and a 247 % increase of Cd than the healthier 40% CM clover. Compost increased mean pH levels from 5.65 in CO to 6.36 in 40% CM. Heavy metals bind to the surfaces of soil particles under high pH conditions. The binding process resulted in a reduction of metal availability to wheat and white clover.

Plants are known to store higher amounts of heavy metals within their shoots, foliage, and roots; however, concentration of heavy metals in the roots of clover and wheat were much higher than their shoots. This disproportional metal concentrations within the roots may be due to the plants not reaching maturity.

Heavy metals in soils are found in many chemical forms of varying degrees of availability. Fractions containing high levels of heavy metals indicate the physicochemical form that persists. The majority of Zn, Cd, and Pd was released from the



soil in the carbonate and Fe-Mn Oxide fractions. Metals were moderately bound within the soil. Besides the established threat of plant up-take, the potential of leaching, ingestion, and assimilation in other organisms was present.

Nodule formation of white clover was adversely affected by heavy metal contaminants. Nodule health and nitrogen fixation are indicators of a sufficient supply of nutrients. Compost additions decrease the ill effects that metals can afflict upon the formation of nodules. 40% CM provided white clover with an ample nitrogen source and high pH that resulted in healthy nitrogen fixed nodules.

### Conclusion

Heavy metal contaminated sites should not be taken lightly. Heavy metals can causes birth defects, mental retardation, cancer, and many other equally afflicting conditions. Application such as organic soil amendments can aid in reducing the risk of heavy metal contamination. Techniques introduced in this study may contribute to the efforts of eliminating heavy metal contamination as a threat to animals, invertebrates, and plant species. Applications of the present study were intended to be used in field condition to reduce the availability of Zn, Cd, and Pd within a clayey, low pH soil.

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## Appendix

**Table** – The total amount of Zn, Cd, and Pb loss that occurred in the soil.

	(mg/kg)	Wheat		Clover	
		Shoots	Roots	Shoots	Roots
Control	Zn	291.621	212.075	28.06	1.116
	Cd	5.448	3.484	0.046	0.358
	Pb	3.118	7.115	1.455	4.321
10% Compost	Zn	181.001	85.135	21.638	8.186
	Cd	1.493	0.685	0	0.006
	Pd	2.314	1.947	0.407	2.445
40% Compost	Zn	233.331	106.185	36.872	8.112
	Cd	2.579	0.906	0	0
	Pb	6.733	0.804	1.441	2.209

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