

EVALUATION OF THE GENETIC DIVERSITY OF ALLIUM ASCALONICUM LANDRACES BASED ON MOLECULAR MARKERS

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ABSTRACT

The aim of the research was to identify *Allium ascalonicum* land races with a good yield due to their morphological traits, with good adaptation capacity, with different genetic background, as a first step in identification of a suitable source for the production of secondary metabolites. Therefore 16 shallot landraces collected from different areas of Timis county were evaluated from phenotypic and molecular point of view. Thus the height, diameter, weight and bulb shape index were determined. For genetic fingerprint 8 ISSR (Inter Simple Sequence Repeats) markers were used. The extracted DNA was amplified with the specific primers, the fragments were separated by agarose gel electrophoresis and analyzed with the VisionWorks®LS, (UVP, England) software. 178 amplified fragments were

registered, with an average of 22.25/primer, of which 174 were polymorphic (97.75% polymorphism). The matrix of similarity and the dendrogram were established based on UPGMA (Unweighted Pair Group Method with Arithmetic Mean) analysis. A considerable genetic diversity between landraces with different ecologic origin was observed, indicating that they have different genetic mechanisms for the yield traits and adaptation to the specific environmental conditions of the area. All the obtained results allowed the identification of local land races with a high production capacity which had different genetic background (Dolat 126a, Sanmartinu S. and Rudna 101.) Therefore the possibility to recognize genotypes with distinct secondary metabolites content it was increased.

INTRODUCTION

The *Allium* genus contains hundreds of species with the largest spread in the temperate climate which has traditionally been used as food but can also have therapeutic applications. They have an antioxidant activity due to the sulphur compounds and other bioactive metabolites as polyphenols and phytosterols (Nencini et al. 2007; Vlase et al. 2012). Many of these species among which *Allium ascalonicum* (shallot) have shown antifungal and antibacterial properties (Yin and Tsao, 1999; Amin and Kapadnis, 2005; Mohammadi-Motlagh et al. 2011).

Our previous researches focused on collection and characterization of horticultural land races, traditionally cultivated in isolated villages from the West part of Romania showed that shallot have been found in a large number of households.

Nowadays, evaluation, documentation and use of genetic resources, represented by old, insufficiently studied local landraces, are currently a necessity both at national and international level. Local populations are distinguished by a great capacity of adaptation and physiological traits specific to certain

areas, as well as their own production capacity and quality attributes (**Murariu et al., 2008**).

Therefore, as a first step in identification of a suitable source for the production of secondary metabolites it was of great interest to identify *Allium ascalonicum* land races with a good yield due to their morphological traits, with good adaptation capacity, which has different genetic background.

To evaluate the genetic fingerprint, ISSR (Inter Simple sequence Repeats) markers were used because they amplify more regions from the genomes generating a different number of fragments. The primers specific for ISSR are generally 16-25 nucleotide lengths with multiple loci as target regions, especially located in the microsatellite sequences (**Zietkiewicz et al., 1994**). The microsatellite-type repeats used as primers may be di, tri, tetra or penta-

nucleotides, most often anchored at the 3' or 5' end with 1 - 4 bases prolonged in the regions flanking the microsatellite region (**Reddy et al., 2002**).

Although the ISSR generally behaves as dominant markers, which is a disadvantage, they have a widespread use in genetic mapping and population genetics studies because they require small amounts of DNA considering they are based on the PCR reaction and do not require prior sequencing studies, just like SSR (Single Sequence Repeats) markers. These markers have provided reliable results in the evaluation of the genetic diversity for horticultural plants such as potato (**Prevost et al., 1999**), chestnut (**Casasoli et al., 2001**), mulberry (**Vijayan et al., 2003**), clematis (**Gardner and Hokanson, 2005**), strawberry (**Debnath et al., 2008**), bilberry (**Botau et al., 2014**) or even onion (**Son et al., 2012**).

MATERIALS AND METHODS

For phenotypical and molecular analysis 16 *Allium ascalonicum* land races were used, encoded according to their collection place: Dolaț 126 a (white), Dolaț 126 r (red), Dolaț 198, Dolaț 244, Dudestii Vechi 10, Foeni 343, Giera 06, Livezile 498, Rudna 101, Rudna 124, Rudna 304, Toager, Toager 207, Toager 29, Uivar 305 and Uivar 30c.

Working methods

- For **phenotypically analysis** the height, diameter, weight and bulb shape index were determined, the obtained data being statistically evaluated.
- For **molecular analysis** 8 ISSR (Inter Simple Sequence Repeats) markers were used.

First, the DNA was extracted from shallot bulbs based on modified Doyle and Doyle method (Doyle and Doyle, 1987). It was necessary to supplement the extraction solution with 2-mercaptoethanol to break down the proteins disulfide bonds.

Each DNA samples were spectrophotometrically analyzed at 230, 260 and 280 nm, to determine the concentration and quality (Nanodrop 8000, Thermoscientific). If the concentrations were high, they were all diluted to 100ng/μl.

For amplification, in the first step a bulk of the extracted and diluted DNA samples was amplified with 20 ISSR primers and the ones which generated the most clear and discernable fingerprints were selected, namely UBC 807 - 5'(AG)₈T^{3'}; UBC 834 - 5'(AG)₈YT^{3'}; UBC 836- 5'(AG)8YA 3', UBC 810 – 5'(GA)8T 3'. UBC 811- 5'(GA)8C 3', A 12 – 5'(GA)₆CC^{3'}; UBC 818- 5'(CA)₇G 3'; A 13- 5'(GT)₆CC^{3'}.

All of the selected primers were used in series to amplify each DNA samples. PCR was carried out in a final volume of 25 µl containing 100 ng of DNA template, using GoTaq Green Master Mix (Promega, USA);

The amplification program consisted of a first denaturing step for 5 min at 95°C, followed by 45 cycles of denaturation at 95°C for 45 sec, annealing at 55°C for 45 sec and extension at 72°C for 2 min, the final step of extension at 72°C for 5 min, according to literature data.

The amplified fragments were separated by 1.5 agarose gel electrophoresis and analyzed with the VisionWorks®LS, (UVP, England) software. Then, they were aligned for all the individuals according to their size and scored with 1 for presence and 0 for absence. The dendrogram was assessed from a set of variables using DendroUPGMA software. The program calculated a similarity matrix and transformed similarity coefficients into distances and made a clustering using the Unweighted Pair Group Method with Arithmetic mean (UPGMA) algorithm.

RESULTS AND DISCUSSIONS

The shallot samples were collected from the already mentioned locations. From each one a number of at least 10

bulbs were evaluated by measuring the height, diameter, weight and shape index (Table 1).

Table 1

The average values for the bulb traits for shallot land races

No.	Land race code	Height (cm)	Diameter (cm)	Weight (g)	Shape index
1	Dolat 126a	5,45±0,15	6,90±0,10	88,76±2,58	0,79±0,01
2	Dolat 126r	6,00±0,46	4,27±0,07	49,71±4,05	1,41±0,11ab
3	Dolat 198	3,50±0,29	3,60±0,10	25,11±4,80	0,97±0,06
4	Dolat 244	3,93±0,18	4,17±0,23	36,15±3,64	0,95±0,08
5	Foieni 343	4,70±0,15	3,57±0,03	42,56±3,51	1,32±0,06
6	Giera 6	5,70±0,65	3,67±0,03	53,51±5,83	1,55±0,17
7	Livezile 498	4,47±0,07	3,87±0,13	40,96±1,71	1,16±0,04
8	Rudna 101	6,50±0,40	3,85±0,15	81,43±5,58	1,69±0,17
9	Rudna 124	5,13±0,23	4,20±0,15	53,89±2,29	1,23±0,09
10	Rudna 304	4,50±0,17	3,40±0,10	40,67±0,80	1,33±0,09
11	Sanmartinu S. 180	5,90±0,10	4,45±0,15	82,47±4,72	1,33±0,07
12	Uivar 305	5,40±0,40	4,00±0,50	52,25±3,23	1,36±0,07
13	Uivar 306	4,40±0,10	3,75±0,25	39,85±2,73	1,18±0,11
14	Toager 1	3,67±0,17	3,43±0,07	24,18±2,99	1,07±0,04
15	Toager 29	4,97±0,27	3,87±0,13	50,79±5,89b	1,28±0,05
16	Toager 207	3,97±0,15	3,73±0,13	29,98±3,65	1,06±0,02

Referring to the diameter of the bulbs, the studied land races showed variation amplitude of 3.50 cm associated with a medium to large inter-genotypes variability (20.23%), with the limits between 3.40 cm for Rudna 304 and 6.90

cm for Dolat 126a population. Therefore, the Dolat 126a population showed a bulb diameter significantly higher than the other populations, with no statistically significant differences. The height of the bulbs showed a median inter-population

variability, with values ranging from 3.50 for Dolat 198 population and 6.50 for Rudna 101. Thus, the Rudna 101 population showed a significantly higher bulb height than most other populations except for the following: Dolat 126a, Dolat 126r, Giera 6, Rudna 124, Sanmartin S. 108 and Uivar 305. Low values of this trait were recorded for populations: Dolat 198, Dolat 244, Toager 1 and Toager 207.

In terms of bulb weights, the studied populations showed a very high variability (39.64%), associated with amplitude of 64.58 g. The Dolat 126a population had the highest value of bulb weight, statistically assured compared with the other populations. A large bulb weight was also observed for the populations: Sanmartinu S. 108 and Rudna 101, with average values of this trait over 80 g. The lowest values of bulb weight were recorded in the populations: Toager 1, Dolat 198 and Toager 207, with values up to 30 g.

Considering the phenotypic similarity of the 16 shallots populations, their dendrogram was established by the cluster average (groups) method. It was composed of two main clusters, one represented only by the Dolat 126a population, which showed an average phenotypic diversity of about 42% relative to other populations. The second cluster comprised three subclusters: the first one composed from nine populations with an average similarity of approx. 96% compared to the other seven populations, comprising subgroups of populations with higher values (Dolat 126a, Giera 6, Rudna 124, Toager 29 and Uivar 305) or lower (Foieni 343, Rudna 304, Livezile 498 and Uivar 306); the second subcluster consists of small flattened bulbs populations (Dolat 198, Dolat 244, Toager 1 and Toager 207) with a similarity of approximately 96.5%; the third subclass includes the Rudna 101 and Sanmartin S populations, with large

bulbs, with an average similarity of approximately 93%.

It was pointed out that there was no precise correlation between the morphological traits and the collection place, but it was possible to identify the land races with the higher productivity, namely Dolat 126a, Sanmartinu S. and Rudna 101. Considering that the purpose of the research was to identify local populations with high productivity, but not very genetically close, the next step was the genetically fingerprint based on ISSR markers.

For each collected sample the genomic DNA was extracted and the quality and concentration were determined spectrophotometrically. All samples had an OD 260/280 ratio around the 1.8-1.9 range, therefore they were considered to be of adequate quality. Their concentration was very high, therefore all were diluted to 100 ng / μ l.

To select the ISSR markers which generate clear, detectable fingerprint 20 primers were used. The primers generating the best results, namely UBC807, UBC834, UBC836, UBC810, UBC811, UBC 818, A 12, A 13, were used in amplification reactions with all of DNA samples. To illustrate these investigations the results obtained after the amplification with UBC 810 and A 13 primers was shown (Fig. 1). Similar images were obtained for all of the other six primers.

The visualized fragments for each genotype were analyzed, recorded in order of dimensions: loci 1-19- primer UBC 807, loci 1-28 primer UBC 810, loci 1-32 primer UBC 811, loci 1-34 primer UBC 834 loci 1- 25- primer UBC 836 loci 1-19- primer A12 loci 1-22- primer A 13, loci 1-20- primer UBC 818.

Next, they were scored with 1 when a fragment was present at a specific locus and with 0 if it was absent, all the information being introduced into the binary matrix.

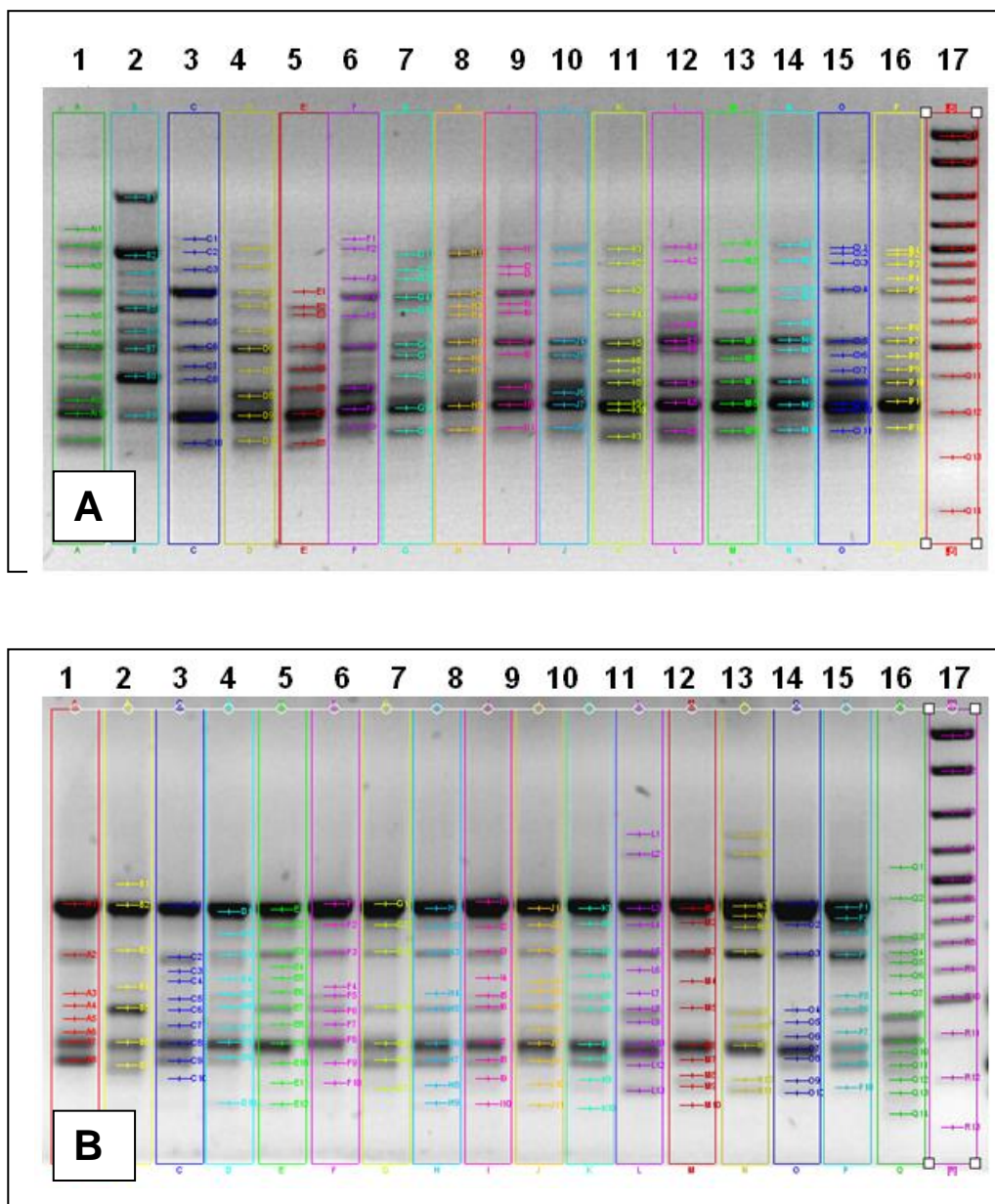


Fig. 1. Analysis by 1.5% agarose gel electrophoresis of the products resulting from amplification with the primer UBC 810 (GA)₈T [a] and A13 (GT)₆CC [B]

1-Dolaț 126 a (white), 2- Dolaț 126 r (red), 3-Dolaț 198, 4-Dolaț 244, 5-Sanmartinu S.,
6-Foeni 343, 7-Giera 06, 8-Livezile 498, 9-Rudna 101, 10-Rudna 124, 11-Rudna 304,
12-Toager, 13-Toager 207, 14-Toager 29, 15-Uivar 305 and 16-Uivar 30c.,
17- 100bp DNA ladder (ThermoFischer Scientific)

Considering all the eight ISSR markers used in this study, a number of 178 fragments were amplified, with an average of 22.25 alleles/primer, most of them being different, with a rate of 97.75% polymorphism.

The statistically evaluation based on Unweighted Pair Group Method with Arithmetic mean (UPGMA) algorithm established the similarity matrix and dendrogram (Fig. 2).

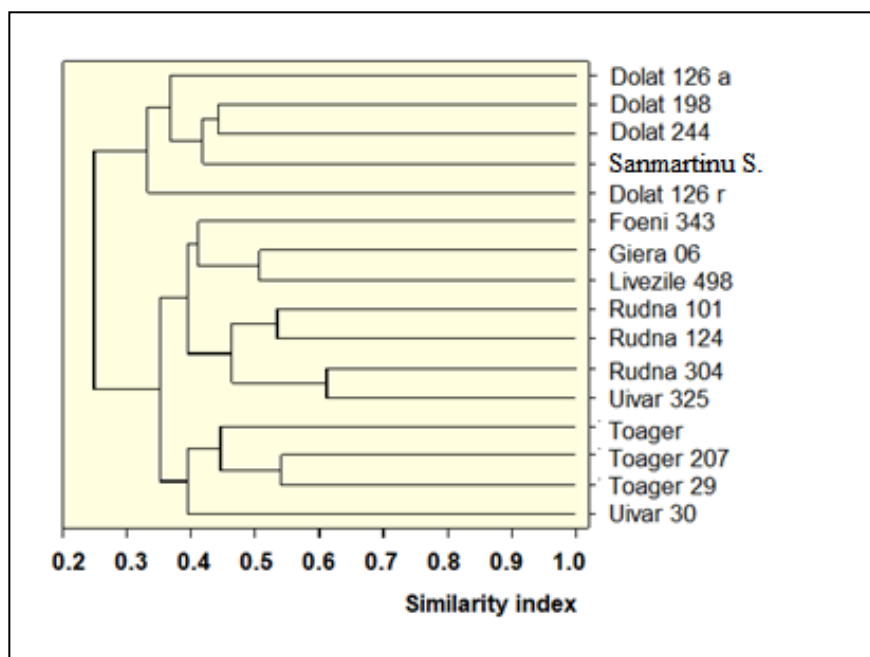


Fig. 2 The dendrogram of the shallot landraces based on the average clusters method for the ISSR markers

Therefore, the largest genetic diversity was observed between land races: Dolat 126r - Uivar 305 (83.05%); Dolat 244 - Toager 29 (81.63%); Dolat 126r - Toager 207 (80.34%), emphasising that these genotypes have a very different genetic background. The highest level of similarity (61.11%) was found among the populations of Rudna 304 and Uivar 305, followed by 54.05% between Toager 29 and Toager 207 populations.

Considering the previous classification it was observed that Rudna 304 showed the highest average diversity (approximately 61.30%) compared to the rest of the land races, followed by Rudna 101 (36.59%) and Toager 207 (35.23%). The respective populations are grouped into two clusters with an allelic diversity of about 25% between them.

The first cluster included four Dolat populations together with the Sanmartinu S., which owned approximately 35% common alleles for these primers. The second cluster is composed of two sets of populations, with a genetically diversity of about 65%. The first set is more comprehensive and consists

of three populations of Rudna, along with those of Foieni, Giera Livezile and Giera, which are genetically similar to about 40%. In the second set are included three populations of Toager alongside Uivar.

A considerable genetic diversity between landraces with different ecologic origin was observed, indicating that they have different genetic mechanisms for the yield traits and adaptation to the specific environmental conditions of the area.

Regarding the variance analysis for the shallot land races, an important contribution to the total variability of the different fragments amplified by the ISSR primers was observed especially for the Rudna 304 population, followed by Toager 207 and Livezile 498, while for the Dolat 126r population the lowest variance values were recorded. Dolat populations have the highest contribution to allelic diversity within the first cluster, while diversity in the second cluster is mainly influenced by the contributions of the Giera 6 and Uivar30 populations (Table 2).

Table 2

Variance analysis for the shallots land races in terms of ISSR fragments

No.	Land race name	Between groups		Inside the groups		F test
		SP	GL	SP	GL	
1	Dolat126 a	1,861	1	39,920	176	8,20**
2	Dolat 126 r	0,048	1	44,250	176	0,19
3	Dolat 198	0,209	1	41,055	176	0,90
4	Dolat 244	0,164	1	39,280	176	0,73
5	Dudestii Vechi 10	2,454	1	34,355	176	12,57**
6	Foeni 343	10,938	1	26,680	176	72,16**
7	Giera 06	7,001	1	33,095	176	37,23**
8	Livezile 498	14,541	1	25,555	176	100,15**
9	Rudna 101	17,587	1	22,189	176	139,50**
10	Rudna 124	10,631	1	27,375	176	68,35**
11	Rudna 304	20,073	1	20,022	176	176,45**
12	Toager	9,318	1	31,385	176	52,25**
13	Toager 207	16,005	1	22,742	176	123,86**
14	Toager 29	10,028	1	28,719	176	61,46**
15	Uivar 305	12,917	1	25,089	176	90,61**
16	Uivar 30c	7,362	1	31,020	176	41,77**

All the obtained results allowed the identification of local land races with a high production capacity which had different genetic background (Dolat 126a,

Sanmartinu S. and Rudna 101.) Therefore the possibility to recognize genotypes with distinct secondary metabolites content it was increased.

CONCLUSIONS

ISSR markers have been found to be suitable for assessing variability in shallot analyzed genotypes. The high number of fragments fall them in the high polymorphic markers category.

The dendrogram allowed the assessment of the degree of inheritance, underlining that, in general, the

geographical neighborhood placed the genotypes in closed subclusters, underlining the common origin.

Based on these results it was possible to select genotypes with different genetic background to be used in future breeding programs.

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