

## REVIEW ARTICLE

## Cancer Immunotherapy: A Review

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### Abstract

**BACKGROUND:** The goals of treating patients with cancer are to cure the disease, prolong survival, and improve quality of life. Immune cells in the tumor microenvironment have an important role in regulating tumor progression. Therefore, stimulating immune reactions to tumors can be an attractive therapeutic and prevention strategy.

**CONTENT:** During immune surveillance, the host provides defense against foreign antigens. By targeting surface antigens expressed on tumor cells, monoclonal antibodies have demonstrated efficacy as cancer therapeutics. Recent successful antibody-based strategies have focused on enhancing antitumor immune responses by targeting immune cells, irrespective of tumor antigens. The use of antibodies to block pathways inhibiting the endogenous immune response to cancer, known as checkpoint blockade therapy, has stirred up a great deal of excitement among scientists, physicians, and patients alike. Clinical trials evaluating the safety and efficacy of antibodies that block the T cell inhibitory molecules cytotoxic T-lymphocyte-

associated protein 4 (CTLA-4) and programmed cell death 1 (PD-1) have reported success in treating subsets of patients. Adoptive cell transfer (ACT) is a highly personalized cancer therapy that involve administration to the cancer-bearing host of immune cells with direct anticancer activity. In addition, the ability to genetically engineer lymphocytes to express conventional T cell receptors or chimeric antigen receptors has further extended the successful application of ACT for cancer treatment.

**SUMMARY:** The underlying basis of cancer immunotherapy is to activate a patient's own T cells so that they can kill their tumors. Reports of amazing recoveries abound, where patients remain cancer-free many years after receiving the therapy. The idea of harnessing immune cells to fight cancer is not new, but only recently have scientists amassed enough clinical data to demonstrate what a game-changer cancer immunotherapy can be. This field is no stranger to obstacles, so the future looks very promising indeed.

**KEYWORDS:** immune checkpoint, adoptive cell transfer, neoantigen, monoclonal antibody

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### Introduction

The past few decades have seen a grounds well of research on the immune system yielding a deeper understanding of how cancer progresses and offering new ways to stop it.(1) In 1891, William Coley injected cancer patients with bacteria to ignite an immune response, a strategy is

experiencing a revival. Now immunologists are finding ways to harness the immune system, including training immune cells to recognize a patient's particular cancer.(2,3) The finding that tumors can actively suppress immunity has led to the development of checkpoint blockades that prevent this suppression.(4)

Last year's Lasker DeBakey Clinical Research Award was awarded to James Allison for discovering that antibody

blockade of the T cell molecule cytotoxic T-lymphocyte-associated protein 4 (CTLA-4) unleashes the body's immune response against malignant tumors. This led to development of multiple immune checkpoint therapies that are prolonging and saving the lives of many cancer patients. (5) The successful treatment of multiple mouse tumors with anti-CTLA-4 impressed many immunologists, but the perceived failure of many earlier immune-based therapies created a very high bar for advancing immune checkpoint therapy to the clinic. The turning point came with 3 trials comparing Ipilimumab to a melanoma peptide vaccine in metastatic melanoma patients who had the human leukocyte antigens (HLA) A0201 allele. (6) Key to the success of the study was the decision to evaluate overall survival rather than response rate, and in the large study, Ipilimumab monotherapy resulted in more than 20% long-term survival. The success of this trial led the Food and Drug Administration (FDA) to finally approve Ipilimumab for the treatment of metastatic melanoma in 2011.

At around the time when Ipilimumab was being considered for FDA approval, renewed excitement about immune checkpoint therapy came from clinical studies targeting a second immune inhibitory molecule, programmed cell death 1 (PD-1). PD-1 was discovered by Tasuku Honjo in 1992 in a screen for genes expressed during programmed cell death of a T cell hybridoma. (7) Unlike CTLA-4, which functions mainly during primary immune responses, PD-1 signaling results in exhaustion of activated T cells, an anergic-like state that is thought to be due to a shift in the utilization of metabolic substrates. (8) Antibody blockade of PD-1 was shown to enhance anti-tumor and anti-viral responses in animal models, suggesting that this could be another immune checkpoint target for cancer.

Schering Plough acquired an anti-PD1 antibody developed by Organon, and this was introduced into the clinic as Pembrolizumab, after the company was acquired by Merck, and was approved by the FDA for treating advanced melanoma in 2014. The Bristol-Myers Squibb (BMS) drug, Nivolumab, was approved very shortly thereafter. The change in attitude of clinical oncologists and immunologists toward the place of immune modulation in combating cancer guarantees that there will be many exciting advances in immune-based therapies in the years ahead. (5)

## Cancer Immunology

Although originally considered as monoclonal, tumor cells show heterogeneous morphology and behavior. (9,10) This heterogeneity has traditionally been explained by the clonal

evolution of tumor cells resulting from the progressive accumulation of multiple genetic (11) or epigenetic changes (12). Alterations in tumor stroma microenvironments may also promote the development of tumor cell heterogeneity through extrinsic activation of certain tumor cell signaling pathways. (13) Moreover, recent studies have suggested that heterogeneity is a result of the hierarchical organization of tumor cells by a subset of cells with stem or progenitor cell features known as cancer stem cells (CSC). (14)

The concept of cancer as an abnormal stem cell disease was proposed based on the similar abilities of cancer cells and normal stem cells to self-renew, produce heterogeneous progeny and also divide in an unlimited fashion. (15,16) However, the CSC hypothesis has only recently been experimentally validated by the identification of a subset of certain self-renewing stem cell marker-positive cells with a hierarchical organization. (17,18) The self-renewal capacity is confirmed by serial in vitro clonogenic growth and in vivo tumorigenicity. CSC are also known as tumor-initiating cells or tumor-propagating cells. CSC are highly tumorigenic, metastatic, chemotherapy and radiation resistant, responsible for tumor relapse after therapy, and able to divide symmetrically and asymmetrically to orchestrate the tumor mass. (19) Therefore, CSC are a pivotal target for the eradication of many cancers including liver cancer. (20)

Some cancer cells may be disseminated and leave tumor relentlessly, only to perish en masse, find a good time to reinitiate a full-fledged tumor and settle there arising metastasis in distant tissues. (21,22) Yet when metastasis occurs, it creates complications that account for the vast majority of deaths from cancer. Cancer cells that succeed in doing this task possess not only the attributes of tumor-initiating cells, but also the ability to exert this capacity under harshly adverse conditions. Metastasis therefore is driven by CSC at their best, or at their worst, depending on your perspective. (23)

As tumors are heterogeneous and show distinctive genetic and epigenetic profiles, there may not be a single biomarker that will prove sufficient information for predicting treatment response and patient outcome. Examples of informative tumor biomarkers are molecular features of neoplastic cells, including epidermal growth factor receptor (EGFR) mutations in lung cancer (24,25); microsatellite instability (MSI) in colorectal cancer (26-28); estrogen receptor 1 (ESR1), progesterone receptor (PGR) and erb-b2 receptor tyrosine kinase 2 (ERBB2/HER2) expression in breast cancer (29,30); transmembrane protease, serine 2 and ETS-related gene fusion (TMPRSS2-ERG) translocation in prostate cancer (31); and CpG island methylation, and

kirsten rat sarcoma viral oncogene (KRAS), B-raf proto-oncogene (BRAF), phosphatidylinositol-4,5-bisphosphate 3-kinase, catalytic subunit alpha (PIK3CA) and tumor protein(TP)-53 mutations in multiple cancer types (32-34). In addition to tumor markers, host factors which include the immune response to the tumor might determine tumor behavior or serve as informative biomarkers.(35)

During carcinogenesis, tumor cells interact with a complex microenvironment which is composed of extracellular matrix and non-neoplastic host cells, including mesenchymal cells, vascular endothelial cells and inflammatory or immune cells. Inflammatory and immune cells are present to varying degrees (from absent to intense) in the tumor microenvironment, which can be observed routinely in pathology practice. The tumor microenvironment provides nutrients, oxygen, growth factors, cytokines, and other chemical mediators that support tumor proliferation, survival, invasion, and metastasis for the cancer cells.(36) The immune system can respond to cancer cells in two ways, which are by reacting against tumor-specific antigens (molecules that are unique to cancer cells) or by reacting against tumor-associated antigens (molecules that are expressed differently by cancer cells and normal cells).(37) Immunity to carcinogen-induced tumors in mice is directed against the products of unique mutations of normal cellular genes. These mutant proteins are tumor-specific antigens. (38)

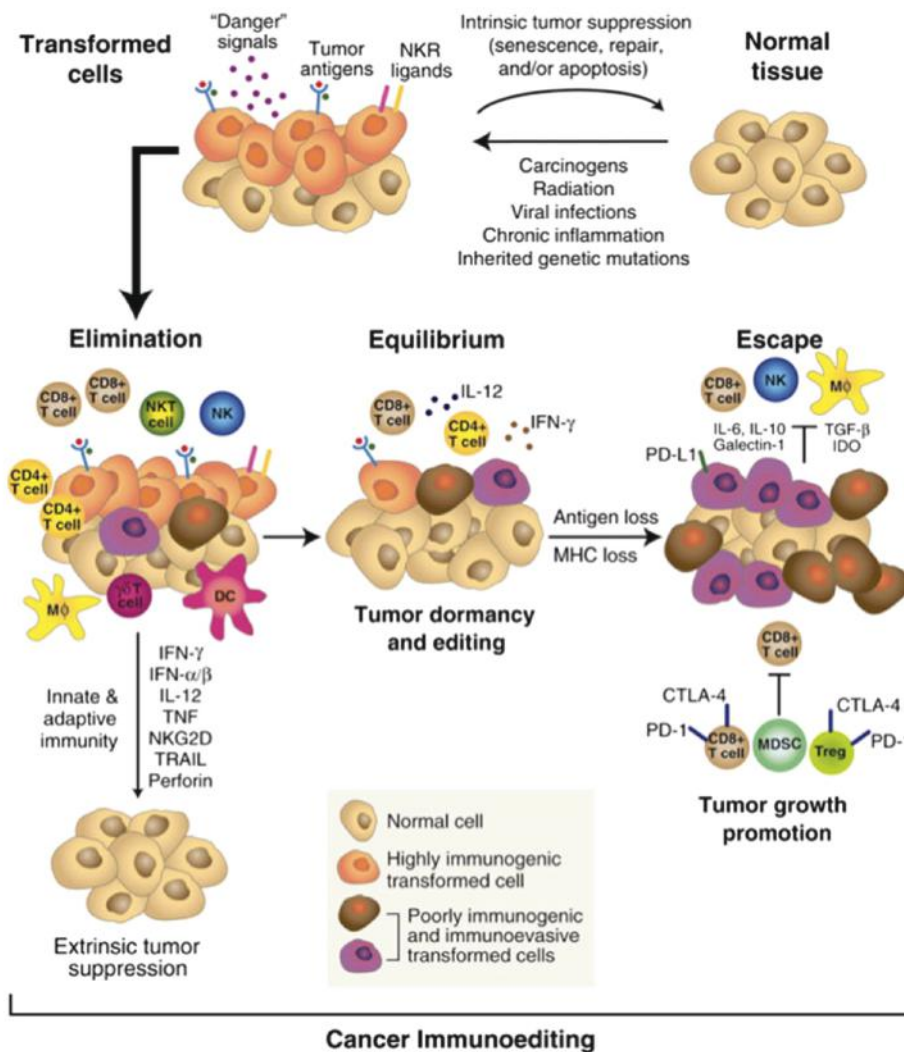
The immunosurveillance hypothesis posits that the immune system recognizes malignant cells as foreign agents and eliminates them. This idea was contentious until the understanding of tumor immunity improved and better techniques and animal models became available to test it rigorously. Mouse models in which immune effector mechanisms such as the type 1 interferon (IFN) were eliminated by gene deletion showed a clear reduction in the incidence of tumors by the immune system.(39-43) In animal models, the encounter between the immune system and a nascent tumor initiates a process termed 'immunoediting' that can bring about three outcomes, those are elimination of the cancer; cancer equilibrium, in which there is immune selection of less immunogenic tumors during an antitumor immune response; and tumor escape, the growth of tumor variants that resist immune destruction.(44,45)

Relation between immune system and cancer is complex and dynamic. Whereas there are a series of scenarios in which the immune system exerts an antineoplastic surveillance, there are also other situations in which immune processes contribute to the transformation and progression of malignant tumors.(46) This paradigm

of tumor immunology is known as cancer immunoediting, and is composed of three stages, which are elimination, equilibrium and escape (Figure 1).(46-48) The elimination phase is difficult to study, and clear evidence of its existence is scarce in humans, as elimination is believed to take place in a preclinical stage and often times represents the resolution of the disease process. Elimination was thought to be carried by the innate and adaptive immune systems, and natural killer (NK), natural killer T (NKT), cluster of differentiation (CD)8<sup>+</sup> cytotoxic and CD4<sup>+</sup> lymphocytes, macrophages as well as dendritic cells (DC) that participating in the presentation, recognition and lysis of cells displaying tumor antigens.(49) Elimination is often limited, and some tumor cells are left alive, either due to their antigenic or immune-related gene expression profile allows these cells to survive the initial immune surveillance entering an equilibrium phase. In this phase, there is a dynamic balance between anti-tumoral immunity and tumor cells.

Cancer development was showed in patients transplanted with donor-derived tumors, even when there was no cancer clinical manifestation found. This suggest the existence of human equilibrium stage.(49,50) The equilibrium is thought to be maintained by adaptive immunity. (46) At last, escape is the stage in which those tumor cells that are not detectable or have developed mechanisms to avoid immune recognition and lysis, get selected and then grow into a symptomatic lesion. An illustrating example of this phenomenon is the use of a vaccine targeting the EGFR variant III (EGFRvIII) in glioblastoma. Clinical studies have shown some effectiveness in patients with glioblastoma that originally expressed EGFRvIII, while upon recurrence expression was lost in the tumors.(51) In addition to the induction of an immunosuppressive state, another paradigm in tumor-immunology is the tendency for tumors to minimize the display of their antigens, which is resulting in another mechanism for evasion of anti-tumor immunity. The relative lack of a tumor-specific antigenic repertoire and the impairment of antigenic presentation by major histocompatibility complex (MHC) class I by tumors can diminish tumor recognition by cytotoxic T-lymphocytes. (52)

Tumors can suppress immunity both systemically and in the microenvironment of the tumor.(53) In addition to producing immunosuppressive molecules such as transforming growth factor  $\beta$  (TGF- $\beta$ ) (54) and soluble Fas ligand (55), many human tumors produce the immunosuppressive enzyme indoleamine-2,3-dioxygenase (IDO) (56,57). This enzyme was previously known for its



**Figure 1. Cancer immunoediting paradigm.**(46) IDO: indoleamine-2,3-dioxygenase, TGF-β: transforming growth factor β, IL: interleukin, TNF: tumor necrosis factor, NKG2D: natural-killer group 2 member D, TRAIL: TNF-related apoptosis inducing ligand, MØ: macrophage. (Adapted with permission from Springer).

role in maternal tolerance to antigens from the fetus (58) and, more recently, as a regulator of autoimmunity that mediates inhibition of T-cell activation (59). Stereoisomers of 1-methyl-tryptophan inhibit IDO (60), and when administered to tumor-bearing mice, they restore immunity and thereby allow immune rejection of the tumor (61). Such stereoisomers might have a role in the treatment of patients with cancer.(62)

Tumor microenvironment can be dominated by regulatory T cells that suppress antitumor effector T cells by producing the immunosuppressive cytokines TGF-β and interleukin (IL)-10.(35) High numbers of these cells can be detected in non-small-cell lung cancer and ovarian cancer.(63) Murine tumors that produce TGF-β can convert antitumor effector T cells into regulatory T cells, thereby escaping their own destruction by immune cells.(64,65) The immunosuppressive effects of a tumor can also be systemic. An increase in regulatory T cells has been found in the peripheral blood of patients with head and neck cancer (66,67) or melanoma (68). Patients with colorectal

cancer or pancreatic tumors have increased numbers of activated granulocytes (69) and myeloid-derived suppressor cells (70), both of which suppress tumor-specific T cells in mice (71,72). Up to now, the field of tumor immunology is providing an initial understanding of how these tumors might be avoiding immune recognition.(46)

### Cancer Immunotherapy

For more than half a century, scientists have been trying to turn the body's immune system against cancer. But decades of failures have revealed that tumors have the ability to evade, tamp down and overwhelm the normal immune response. Most modern immune therapies try to get the immune system to recognize and attack tumor cells. (4) The administration of monoclonal antibodies (mAb) against tumor antigens in HER2-positive breast cancer (Trastuzumab) (73), B-cell lymphomas (Rituximab) (74), and head and neck, lung, and colorectal cancers that express



the EGFR (Cetuximab) (75-77) is clinically effective (Table 1).(62) Efforts are ongoing to produce antibodies with new effector functions against known targets or to identify new targets for therapeutic antibodies. These targets could be tumor antigens or molecules produced by tumors to promote their own survival, such as vascular endothelial growth factor (VEGF) (78) and TGF- $\beta$  (64). Antibodies can also target immune cells at the tumor site to aid the activation of effector cells and promote more effective antitumor immunity.(79)

Targeting host immunity is an attractive strategy for cancer therapy and prevention because therapy resistance is less likely to develop when host cells are targeted instead of altered molecules within tumor cells.(37,38) The latter approach frequently results in resistance to the initial targeted therapy owing to, for example, an acquired mutation in a domain of the therapeutic target that interacts with the drug. Host immunity can be targeted by the use of activated autologous peripheral-blood mononuclear cells (sipuleucel-T) (80,81), or the use of specific immunoregulatory molecules, such as recombinant vaccinia

vector (targeting prostate-specific antigen) (82). One such treatment, the vaccine sipuleucel-T (marketed as Provenge by Dendreon Corporation in Seattle), was approved by the US FDA in 2010 for use in prostate cancer, which is a move that generated a lot of excitement. But the drug has proven disappointing, with benefits limited to a small percentage of patients, Dendreon is now reported to be for sale. The problem, researchers have slowly been realizing, is that stepping on the immune system's gas pedal isn't enough. It is also necessary to release its brakes, and that is where immune checkpoint blockades come in.(4)

In 2011, the US FDA approved the anti-CTLA4 drug Ipilimumab (developed by BMS and marketed as Yervoy), which was based on Allison's research and eventually saved the lives of some of Ribas's patients. CTLA-4 is not the only checkpoint being targeted by researchers and drug developers. Early trials suggest that drugs that block a different checkpoint, which is PD-1, are even more effective and have fewer side effects than Ipilimumab.(83) In recent studies, checkpoint blockades produced improvements in between 20% and 65% of patients, depending on the

**Table 1. Immunologic reagents approved by FDA for cancer therapy.**(62) (Adapted with permission from Massachusetts Medical Society).

<b>Table 1. Immunologic Reagents Approved by the Food and Drug Administration for Cancer Therapy.</b>		
<b>Reagent</b>	<b>Reagent Target</b>	<b>Indications</b>
<b>Antibodies</b>		
Trastuzumab (Herceptin)	HER2 receptor	HER2-positive breast cancer
Bevacizumab (Avastin)	Vascular endothelial growth factor	Non-small-cell lung cancer, colorectal cancer, and breast cancer
Cetuximab (Erbixim)	Epidermal growth factor receptor	Colorectal cancer and head and neck cancer
Panitumumab (ABX-EGF)	Epidermal growth factor receptor	Colorectal cancer
Ibritumomab tiuxetan (Zevalin)	CD20 B-cell surface antigen (nonradioactive and radiolabeled)	Non-Hodgkin's B-cell lymphoma
Alemtuzumab (Campath)	CD52 lymphocyte surface antigen	Chronic lymphocytic leukemia and T-cell lymphoma
Gemtuzumab ozogamicin (Mylotarg)	CD33 leukemic-cell surface antigen	Acute myeloid leukemia
Rituximab (Rituxan)	CD20 B-cell surface antigen	Non-Hodgkin's lymphoma
Tositumomab (Bexxar)	CD20 B-cell surface antigen (nonradioactive and radiolabeled)	Non-Hodgkin's B-cell lymphoma
<b>Other</b>		
<b>Reagent Type</b>		
Denileukin diftitox (Ontak)	Recombinant interleukin-2 and fragments of diphtheria toxin (binds CD25 receptor on T cells)	Cutaneous T-cell lymphoma
Aldesleukin (Proleukin)	Interleukin-2	Melanoma and renal-cell carcinoma
Interferon alfa-2b (Intron A) and interferon alfa-2a (Roferon-A)	Recombinant interferon	Hairy-cell leukemia, chronic lymphocytic leukemia, Kaposi's sarcoma, melanoma, non-Hodgkin's lymphoma, multiple myeloma, and renal cancer
Imiquimod (Aldara)	Toll-like receptor 7 agonist	Basal-cell carcinoma

drug, dosage and type of cancer. In one long-term study of Ipilimumab in patients with advanced melanoma, 22% of the 1,861 patients survived for three years, and 17% for seven years or longer (with median survival nearly a year). (84) Historically, average survival was six to nine months. (84) Early research suggests that Ipilimumab may be even more effective when combined with other drugs. In further evidence for the value of drug combinations, Ipilimumab and Nivolumab appear to complement each other. (4) A few years ago, when Michel Sadelain spoke about adoptive cell transfer (ACT) therapy at cancer meetings, his colleagues were dubious about what seemed a drastic and unconventional approach: harvesting and genetically altering his patient's immune cells to train them to attack her cancer. (3)

There are three strategies for ACT therapies, the most developed of which is the simplest. The tissue surrounding a tumor is likely to contain immune cells with antitumor activity, so doctors take a sample of this tissue and select those T cells that have been primed to attack the cancer. They culture these cells in the lab until they have enough, and re-infuse the cells back to patients along with the T-cell growth factor IL-2, which promotes the proliferation of antigen-specific T cells. (3) This approach, called tumor-infiltrating lymphocyte (TIL) therapy, has been used successfully to treat only one type of cancer, that is metastatic melanoma. T cells that have been primed to attack a specific cancer are difficult to collect in a blood sample, but in melanoma these lymphocytes enter the tumor and are easy to biopsy. Currently, the success of TIL in melanoma is not transferable to other cancers, because it is harder to collect tumor-specific T cells. For those cancers, researchers are working hard in order to genetically modify T cells to hone their cancer killing skills. (3)

To do this, researchers are taking a couple of approaches. One option, called T cell receptor (TCR) therapy, involves giving the cells new receptors that allow them to recognize specific cancer antigens; the receptors can even be modified to improve their ability to find and bind to their targets. A more flexible tactic, called chimeric antigen receptor (CAR) therapy, avoids this constraint. It uses a gene that encodes artificial, antibody-like proteins that bind the antigens studding the tumor cell's surface without needing to match the patient's immune type. So part of the optimization process includes giving both TCR- and CAR-based T-cell therapies the optimal mix of enhancing molecules and targets to achieve the best response.

As ACT therapies move closer to the mainstream, the next big step will be investigating whether and how to integrate them with other cancer immunotherapies. Despite

lingering questions, scientists and clinicians are enthusiastic about the potential of ACT. It represents a flexible platform for cancer treatment that can be tweaked and adapted as further discoveries are made.

Immunity results from a complex interplay between the adaptive immune system (which is antigen-specific) and the innate immune system (which is not). B cells and T cells of the adaptive immune system use receptors that recognize antigens, or their derived peptides, in a highly specific manner. DC provide an essential link between the innate and adaptive immune responses. The generation of anticancer immunity depends on DC presenting cancer antigens to T cells. But cancers can create an environment that inhibits T cells. The aim of DC vaccination is to boost cancer-specific effector T cells that can not only fight existing cancer but also induce immunological memory to control the recurrence of cancer. (2)

Topalian notes that patients treated with immune therapies could potentially gain a lifetime of protection, similar to the buffer against certain diseases offered by childhood vaccines. "We hope that the same thing is happening in cancer," she says. "We hope that we are re-educating the immune system and that, even if it doesn't completely destroy every last cancer cell, it can keep it in check for a very long time." (85) It is tempting to ask whether immunotherapy is evolving to become standard care for cancer patients, beyond those with advanced disease. Is there a place for therapeutic regimens that combine checkpoint blockade with other strategies? While we are nowhere near having all the answers, these studies provide a wealth of data supporting the idea that somatic mutations in cancer cells are an important target of endogenous anti-tumor responses. Checkpoint blockade is effective at rescuing the anti-tumor effect and it is plausible that understanding the dynamics of the response to this therapy will also help the development of alternative and personalized approaches to treat cancer. (86)

## mAb Targetting Cancer-associated Proteins

Immune system is regulated by a reptilian complex balance of signals transmitted by stimulatory and inhibitory receptors. More than any other discovery, mAb have enabled us to identify and manipulate these molecules, provide an important new class of immunostimulatory therapeutics that can complement small-molecule therapeutics under active development. (87,88) Specific recognition by mAb has permitted the identification of cytokines and cell-surface

molecules involved in humoral antibody-mediated and cellular immune responses.(79)

Antibodies may target tumor cells by engaging surface antigen differentially expressed in cancers. For example, Rituximab target CD20 in non-Hodgkin B cell lymphoma, Trastuzumab targets HER2 in breast cancer, and Cetuximab targets EGFR in colorectal cancer. Blocking the ligand-receptor growth can evoke the tumor cell death and survival pathways. Innate immune effector mechanisms engaging the Fc portion of antibodies via Fc receptors including complement-mediated cytotoxicity (CMC) and antibody-dependent cellular cytotoxicity (ADCC) are emerging as equally important.(89,90)

The natural properties of antibodies which enable specific antigen engagement can be leveraged and improved upon by engineering approaches that increase antitumor activity. One example is the creation of bispecific antibodies (bsAb) with dual affinities for a tumor antigen and either another tumor antigen or a target in the tumor microenvironment. As the Fc domain of mAbs does not directly activate T cells, CD3, the activating receptor for T cells, is a common target of bsAb. Catumaxomab is a bsAb that binds the tumor antigen epithelial cell adhesion molecule (EpcAM), CD3, and innate effector cells through an intact Fc portion.(91) This bsAb, termed a Triomab, effectively kills tumor cells in vitro and in vivo and induces protective immunity, most likely through the induction of memory T cells. Catumaxomab's success in a phase II/III clinical trial led to its approval by the European Commission in 2009 for the treatment of malignant ascites. This success spurred the development of other Triomabs targeted against the tumor antigens HER2/neu (Ertumaxomab), CD20 (Bi20/FBTA05; NCT01138579), GD2 and GD3 (Ektomun).(90)

Some efforts that made earlier to enhance the antitumor effects of mAbs focused on boosting their direct cytotoxic effects on the targeted cells. Conjugation of radionuclides (radioimmunotherapies (RIT)), drugs (antibody-drug conjugates (ADC)), toxins (immunotoxins), and enzymes (antibody-directed enzyme prodrug therapy (ADEPT)) yielded a multitude of antibodies, or antibody-like molecules, with varying clinical efficacy. Three conjugated antibodies have translated into FDA-approved therapies for hematological malignancies. Two are RIT agents targeting CD20 and are indicated for treatment of relapsed and/or Rituximab-refractory follicular or low-grade lymphomas: 90Y-ibritumomab tiuxetan and 131I-tositumomab. At the minimum of a dozen other RIT agents are in active development, including ten that target solid tumors.(92) Brentuximab vedotin, the third approved immunoconjugate, is a CD30 targets ADC, carries

the antimetabolic drug monomethyl auristatin E. It has recently approved as a treatment of anaplastic large cell lymphoma (NCT00866047) and Hodgkin lymphoma (NCT00848926).

Nine mAbs targeting six cancer-associated proteins (HER2/neu, EGFR, VEGF, CD20, CD52 and CD33) are approved for the treatment of solid and hematological malignancies. In addition to antagonizing oncogenic pathways, these biotherapeutics may act by opsonizing tumor cells and triggering their death or removal by ADCC or phagocytosis.(93) Ongoing investigations in murine models and patients increase the possibility that they may also stimulate adaptive immune responses in some settings.(94) Recently, the successful conjugation of toxins to antibodies has been achieved, and these have induced a clinical response in patients who are refractory to the naked antibody.(95) The concurrent administration of immunostimulatory cytokines such as IL-2 and granulocyte-macrophage colony-stimulating factor may also enhance the efficacy of antibody therapy.(96)

## Immune Checkpoint Therapy

The myriad of genetic and epigenetic alterations which are characteristic of all cancers provide a diverse set of antigens that the immune system can use to distinguish tumor cells from their normal counterparts. In the case of T cells, the ultimate amplitude and quality of the response, which is initiated through antigen recognition by the TCR, is regulated by a balance between co-stimulatory and inhibitory signals, that is immune checkpoints.(97,98) Under normal physiological conditions, immune checkpoints are crucial for the maintenance of self-tolerance (the prevention of autoimmunity) and also to protect tissues from damage when the immune system is responding to pathogenic infection. The expression of immune-checkpoint proteins can be dysregulated by tumor as an important immune resistance mechanism.

A novel strategy of immunotherapy called checkpoint inhibition is hovered to dramatically revamp the treatment of a broad spectrum of malignancies. Checkpoint inhibitors function by modulating the immune systems' endogenous mechanisms of T cell regulation Ipilimumab (Yervoy<sup>TM</sup>, BMS, New York, NY) has become standard treatment for metastatic melanoma.(6,99) Ipilimumab binds and blocks inhibitor signaling mediated by the T cell surface coinhibitory molecule cytotoxic T lymphocyte antigen 4 (CTLA-4). Because the mechanism of action is not specific to one tumor type, and because a wealth of preclinical data support the role of tumor immune surveillance across multiple malignancies (100,101), Ipilimumab I being

investigated as a treatment for patient with prostate, lung, renal, and breast cancer among other tumor types.(102) The field of immune checkpoint therapy has joined the ranks of surgery, radiation, chemotherapy, and targeted therapy as a pillar of cancer therapy. Therefore, in contrast to most currently approved antibodies for cancer therapy, antibodies that block immune checkpoints do not target tumor cells directly, instead they target lymphocyte receptors or their ligands in order to enhance endogenous anti tumor activity. (103)

Three new immune checkpoint agents now have been approved by the US FDA for the treatment of melanoma, arising new hopes more approved agents for treating lung cancer, kidney cancer, bladder cancer, prostate cancer, lymphoma, and many other tumor types. Ipilimumab an antibody against CTLA-4 was approved in 2011, and two antibodies against PD-1 (Pembrolizumab and Nivolumab) were approved in 2014.

Another category of immune-inhibitory molecules includes certain metabolic enzymes, such as IDO, which is expressed by both tumor cells and infiltrating myeloid cells, and arginase, which is produced by myeloid-derived suppressor cells.(60,104-109) These enzymes inhibit immune responses through local depletion of amino acids that are essential for anabolic functions in lymphocytes (particularly T cells) or through the synthesis of specific natural ligands for cytosolic receptors that can alter lymphocyte functions. These enzymes can be inhibited to enhance intratumoral inflammation by molecular analogues of their substrates which act as competitive inhibitors or suicide substrates.(110-112) These drugs represent a radical and disruptive change in cancer therapy in two ways. First, they target molecules involved in T cell regulation as the soldiers of the immune system, rather than the tumor cell. Second, perhaps in a more radical shift, the therapy is not designated to activate the immune system to attack particular targets on tumor cells, but to remove inhibitory pathways that block effective antitumor T cell responses.

Understanding of immune checkpoint therapy has led to new weapons against cancer which is elicit durable clinical responses and showed long-term remission for patients, and provide an important advance in clinical advances about regulatory pathways in T cells and enhancing antitumor immune responses. Tumor cells express tumor-specific antigens in the form of complexes of tumor-derived peptides bound to MHC molecules on the cell, this will be the target of T cells in this therapy. Tumor antigens can be derived from oncogenic viruses, differentiation antigens, epigenetically regulated molecules such as cancer testis

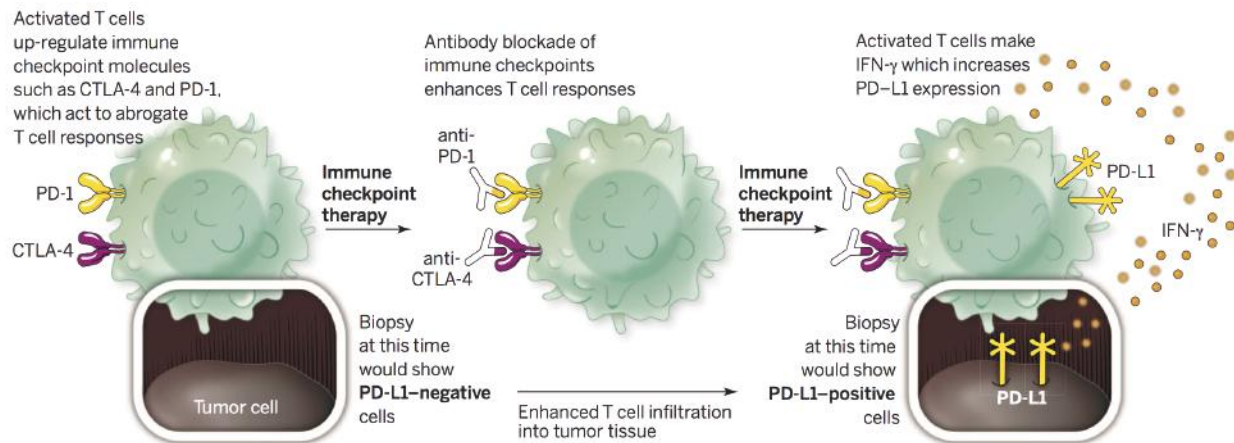
antigens, or neoantigens derived from mutations associated with the process of carcinogenesis.(113)

Recognition of antigen-MHC complexes by the T cell antigen receptor is not sufficient for activation of naïve T cells, but additional co-stimulatory signals are required that are provided by the engagement of CD28 on the T cell surface with B7 molecules (CD80 and CD86) on the antigen-presenting cell (APC).(97,115) Expression of B7 molecules is limited to subsets of hematopoietic cells, especially DC, which have specialized the processes for efficient antigen presentation.(114)

Further insights into the fundamental mechanisms which regulate early aspects of T cell activation may provide one from many possible explanations for the limited effectiveness of these early vaccine trials. By the mid-1990s, it became clear that T cell activation was even more complex, and in addition to initiating proliferation and functional differentiation T cell activation also induced an inhibitor pathway that could eventually attenuate an terminate T cell responses. Expression of CTLA-4 a gene with very high homology to CD28, is initiate by T cell activation, and, like CD28, CTLA-4 binds B7 molecules, albeit with much higher affinity. Although CTLA-4 was first thought to be another co-stimulatory molecule (116), two laboratories independently showed that it oppose CD28 co-stimulation and down-regulated T cell responses (117,118). Thus, activation of T cells result in induction of expression of CTLA-4, which accumulates in the T cell at the T cell-APC interface reaching a level where it eventually block co-stimulation and abrogates an activated T cell response (Figure 2).(114)

The preclinical successes of anti-CTLA-4 I achieving tumor rejection in animal models and the ultimate clinical success which opened a new field of immune checkpoint therapy.(103,119) It is now known that there are many additional immune checkpoints. PD-1 was shown in 2000 to be another immune checkpoint that limits the responses of activated cells.(120) PD-1, like CTLA-4, has two ligands, PD-L1 and PD-L2, which are expressed on man cell types. The function of PD-1 is completely distinct from CTLA-4 in that PD-1 does not interfere with co-stimulation, but interferes with signaling mediated by the T cell antigen receptor.(97) Also, one of its ligands, PD-L1 (B7-H1), can be expressed on many cell types, including T cells, epithelial cells, endothelial cells, and tumor cells after exposure to the cytokine IFN- $\gamma$ , produced by activated T cells.(121) This leads to the notion that rather than functioning early in T cell activation, PD-1/PD-L1 pathway acts to protect cells from T cell attack.



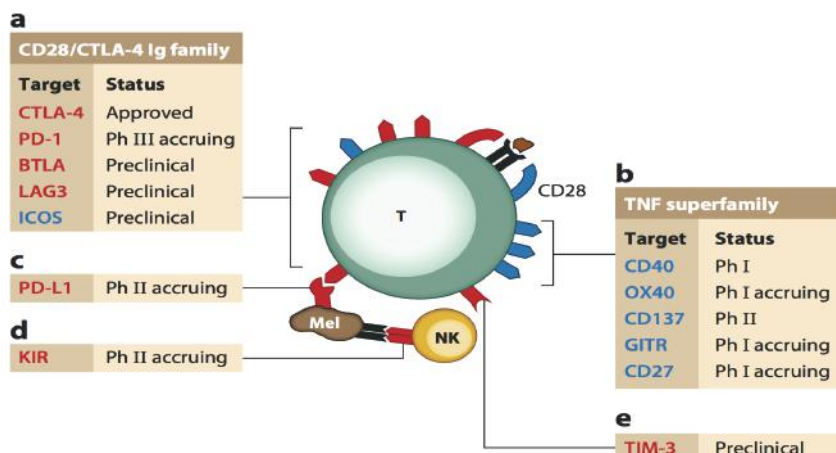


**Figure 2. Blockade of immune checkpoints to enhance T cell responses.** After T cell activation, T cells express immune checkpoints such as CTLA-4 and PD-1.(114) (Adapted with permission from American Association for the Advancement of Science).

Two anti-PD-L1 inhibitory antibodies, MPDL3280A (Genentech, South San Francisco, CA) and BMS-936559 (BMS, New York, NY), have undergone clinical investigation. Like Nivolumab an MK-3475, these antibodies are thought to function principally by blocking PD-1/PD-L signaling. Unlike PD-1 antibodies, PD-L antibodies spare potential interactions between PD-L2 and PD-1, but additionally block interaction between PD-L1 and CD80.(122) In addition to CTLA-4 and PD-1/PD-L1, plenty other immunomodulatory targets have been identified preclinically, many with corresponding therapeutic antibodies that are being investigated in clinical trials (Figure 3).(102) Majority of these targets are T cell surface receptors, but targets in other immunologic cell populations are currently being investigated. For example, NK cells express killer immunoglobulin-like receptors (KIR), which bind HLA class I molecules on target cells, so that delivering an inhibitory signal preventing NK cell-mediated cytotoxicity.(123) Anti-KIR antibodies may release these inhibitory KIR-mediated signals, thereby enabling tumor cytotoxicity and immune clearance.

Efficacy in checkpoint modulation is associated with certain immunologic changes, raising the hope that biomarkers for response may be identified. Only a minority of patients experience long-term survival with Ipilimumab; therefore, considerable efforts are ongoing to discover predictors of response.(102) Immune and tumor response to therapy has been monitored by utilizing a variety of laboratory techniques (Figure 4) (102), and numerous correlates of response have been retrospectively identified.

During the past 2-3 years, outcomes of clinical trials with Ipilimumab and the PD-1 or PD-L1 inhibitors, alone or in combination, have dominated the news coming out of clinical oncology meetings. Combination therapy blocking both checkpoint pathways has been particularly effective, with response rates in advanced melanoma is over 80%. These new issues has embraced the biopharmaceutical enthusiastically, acquisitions and licensing deals for new approaches are almost every week in the news, and investigations about new checkpoint targets, negative regulators of both adaptive and innate immune cells, combination therapy such as with cytokines, co-stimulatory



**Figure 3. Targets of antibody immune modulators.**(102) BTLA: B- and T-lymphocyte attenuator, LAG3: Lymphocyte-activation gene 3, ICOS: inducible T-cell costimulator, GITR: glucocorticoid-induced TNFR family related, TIM-3: mucin-domain containing-3. (Adapted with permission from Annual Reviews).

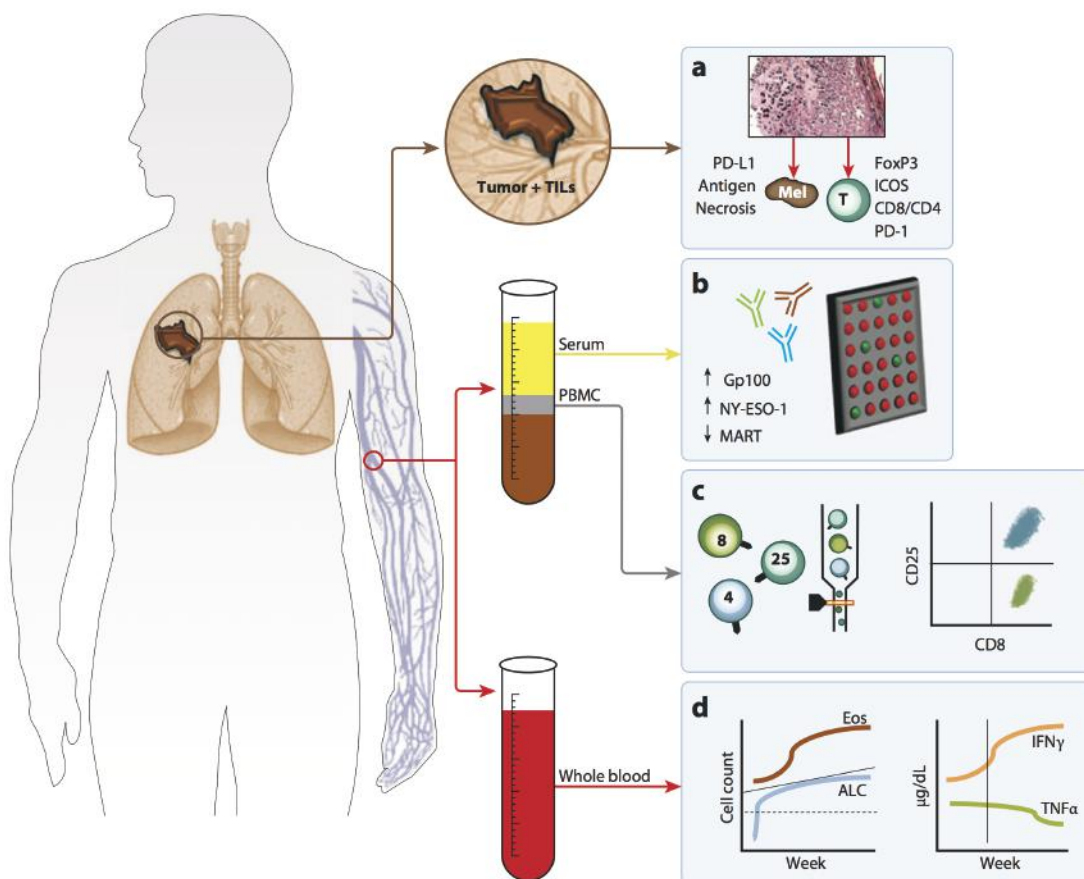
molecules, antigen vaccines, and small-molecule modulators of signaling pathways and enzymes are being actively done. (46) The ability of an activated immune response to generate a diverse T cell repertoire that adapts to heterogeneous and genetically unstable tumors and persistence of memory T cells with specificity for tumor antigens, which provide efficient recall responses against recurrent disease, make it absolutely essential to expand our efforts to find the rational combinations to unleash antitumor immune responses for the benefit of cancer patients. If it is properly done, seems likely that cures for many types of cancer will soon become reality.(114)

### Adoptive Immunotherapy

T cells move through tissues, scanning for MHC-peptide complexes that specifically activate their TCR. T cells are also capable of sensing a variety of signals that can alert them

to potentially threatening pathogens and to cancer. Tumor-specific T cells are probably activated through encounters with tumor-associated antigens that are presented by specialized APC, including DC. However, activated T cells are capable of directly recognizing antigens which were presented on the surfaces of tumor cells. Based on intravital imaging, there is a growing body of evidence showing that the migration of tumor-specific T cells is rapidly arrested when they encounter their cognate antigens.(124,125)

Sufficient number of active in vivo antitumor T cells was necessary to mediate cancer regression in many forms of cancer immunotherapy. ACT has advantages, because for use in ACT, the antitumor lymphocyte can be readily grown in vitro up to  $10^{11}$ , then selection for the high-avidity recognition tumor can be performed before applied effectively, and the inhibitory factors exist in vivo could be released. Perhaps most importantly, ACT enables manipulation of the host before cell transfer to



**Figure 4. Strategies for immune monitoring in patients receiving checkpoint agents.** (a) Surgical specimens may be analyzed using immunohistochemical or immunofluorescence techniques to evaluate tumor antigen expression, T cell infiltrate, tumor necrosis, or expression of surface markers such as PD-L1. (b) Using enzyme-linked immunoabsorbent assays or protein arrays, treatment-related production of tumor-specific antibodies can be detected in the serum. (c) Flow cytometric analysis of TILs and peripheral blood mononuclear cells (PBMC) can quantitate the effect of therapy on immune subsets such as CD25<sup>+</sup> T regs, activated CD8<sup>+</sup> T cells, or myeloid-derived suppressor cells. (d) Whole blood can be used to evaluate changes in cell count with therapy or changes in cytokine levels.(102) FoxP3: forkhead box P3, Eos: eosinophil, ALC: absolute lymphocyte count. (Adapted with permission from Annual Reviews).

provide a favorable microenvironment that better supports antitumor immunity. ACT is a living treatment because the administered cells can proliferate *in vivo* and maintain their antitumor effector functions.(126)

ACT has used either natural host cells that exhibit antitumor reactivity or host cells that have been genetically engineered with antitumor TCRs or CAR. With the use of these approaches, ACT has mediated impressive regressions in a variety of cancer histologies, including melanoma, cervical cancer, lymphoma, leukemia, bile duct cancer, and neuroblastoma.(126)

TILs are primarily Cytotoxic T Lymphocytes (CTL) which recognize proteolytically cleaved intracellular tumor antigen fragments which have become associated with specific MHC class I antigens on the cell surface.(127) Expanded TILs have cytolytic activity against the original tumor and in contrast to lymphokine activated killer (LAK) cells the killing is MHC class I restricted. They are selectively broaden from either tumor or draining lymph node cells via IL-2, and then re-stimulated with irradiated or killed tumor cells to maintain T cell specificity.(128)

Three forms of ACT using T cells have been practiced. The first one is TIL therapy, using lymphocytes expanded from a tumor biopsy sample (129); the second one is antigen-specific T cell therapy, using endogenous T cells sourced from peripheral blood (130-132); and the last one is the more recently, the use of gene modified T cells engineered to express the desired TCR or CAR with occasional remarkable results (133). Immunotherapy based on the adoptive transfer of naturally occurring or gene-engineered T cells can mediate tumor regression in patients with metastatic cancer.(125)

The capacity to use ACT was facilitated by the description of T cell growth factor IL-2 in 1976, which provided a means to grow T lymphocytes *ex vivo*, often without loss of effector functions.(134) The direct administration of high doses of IL-2 could inhibit tumor growth in mice (135), and studies in 1982 demonstrated that the intravenous injection of immune lymphocytes expanded in IL-2 could effectively treat bulky subcutaneous virus-induced lymphoma cells (FBL3) (136). Moreover, administration of IL-2 after cell transfer could enhance the therapeutic potential of these adoptively transferred lymphocytes.(137)

From many tumor histologies culture grown, only melanoma appeared to reproducibly gave rise to TIL cultures for specific antitumor recognition. Studies of genetic engineering led to lymphocytes capability to express antitumor receptors. Following mouse models (138), it was shown for the first time in humans in 2006 that administration

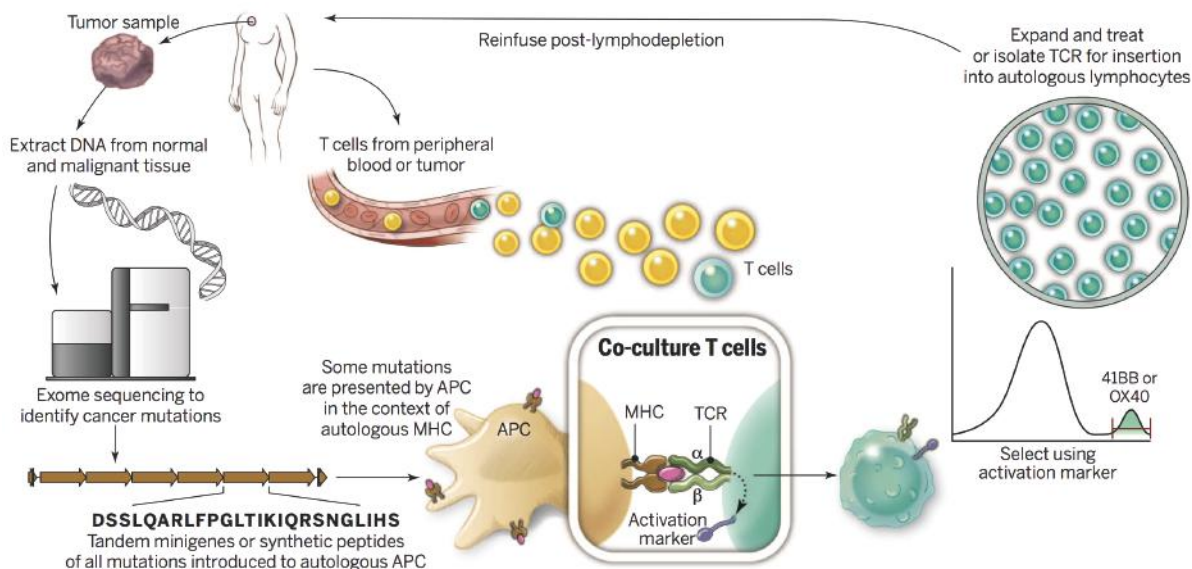
of normal circulating lymphocytes transduced with a retrovirus encoding a TCR that recognized the melanoma antigen recognized by T-cells 1 (MART-1) could mediate tumor regression (139). Administration of lymphocytes genetically engineered to express a CAR against the B cell antigen CD19 was shown in 2010 to mediate regression of an advanced B cell lymphoma.(140) These findings of the use of either naturally occurring or genetically engineered antitumor T cells set the stage for extended development of ACT for the treatment of human cancer (Figure 5).(126)

In an attempt to broaden the reach of ACT to other cancers, techniques were developed to introduce antitumor receptors into normal T cells that could be used for therapy (Figure 6).(126) The specificity of T cells can be redirected by integrating genes encoding either conventional alpha-beta TCRs or CARs. CARs were pioneered by Gross and colleagues in the late 1980s and can be constructed by linking the variable regions of the antibody heavy and light chains to intracellular signaling chains such as CD3-zeta (141), often including co-stimulatory domains encoding CD28 (142) or CD137 to fully activate T cells (143,144). CARs can provide non-MHC-restricted recognition of cell surface components and can be introduced into T cells with high efficiency using viral vectors.

For more patients-specific product approach, a paradigm shift required from conventional medicine such as pills, vaccines, small molecule inhibitor molecules, and antibodies to autologous engineered cell therapies. Some have dismissed adoptive T cell immunotherapy as a fringe or boutique therapy that would be impossible to commercialize. (145). Indeed, several challenges must be overcome before this disruptive therapy can become widely applicable and widely available. Currently, the barriers that we perceive fall into two areas. First, robust and reproducible cell culture system. T cell engineering process needs complex logistics, and some variables standardization in order to scale this out for widespread use include developing a leukapheresis network, standardizing and scaling up the manufacturing of lentiviral vectors, and developing validated cell-shipping and chain-of-custody procedures.(146)

Second, personalized cell therapies cannot become broadly available if the cell culture process requires extensive manipulation by highly skilled scientists and technicians.(147) Hence, automated culture systems need to be developed. Previous case in automotive industry, cars were initially manufactured in assembly lines, but manually. Today's automobiles are assembled largely by robots and other forms of automation.(148) As engineered T cell processing becomes more automated, cell products will be produced for greater number of patients more



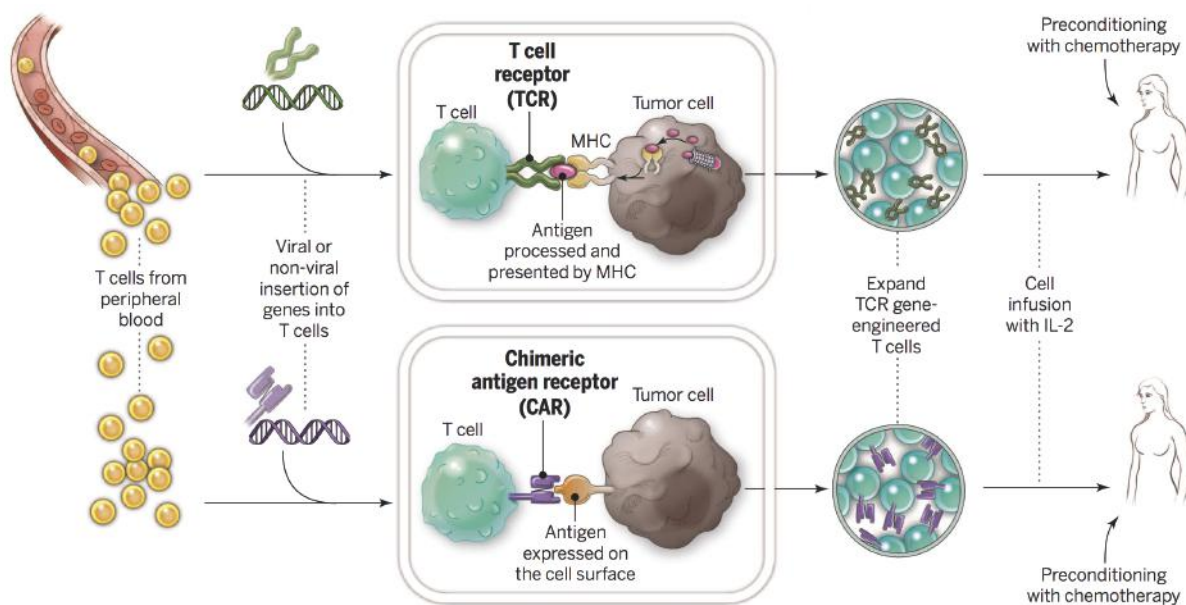


**Figure 5. A blueprint for the treatment of patients with T cells recognizing tumor-specific mutations.**(126) (Adapted with permission from American Association for the Advancement of Science).

efficiently. Seeing the recent entry of the pharmaceutical industry to this field, we are optimistic that the resources and expertise of the pharmaceutical industry will create the infrastructure required for the widespread availability of this disruptive technology. Further clinical development of engineered T cell therapies in large numbers of patients will be challenging but is justified given the magnitude of therapeutic effects recently observed (Figure 7).(149)

The need to develop highly personalized treatments for each patient does not fit into the paradigm of major pharmaceutical companies that depend on off-the-shelf

reagents that can be widely distributed. However, curative immunotherapies for patients with common epithelial cancers will probably dictate the need for more personalized approaches.(126) Widespread of ACT can not depend only on multiple commercial models, but more to the development of centralized facilities for tumor-reactive TILs production and genetically modified lymphocytes, that later can be delivered to the institution who do the treatment. New effective approaches for cancer immunotherapy will need to trump the convenience of applicable administration in treatment.



**Figure 6. Gene-modification of peripheral blood lymphocytes.**(126) (Adapted with permission from American Association for the Advancement of Science).



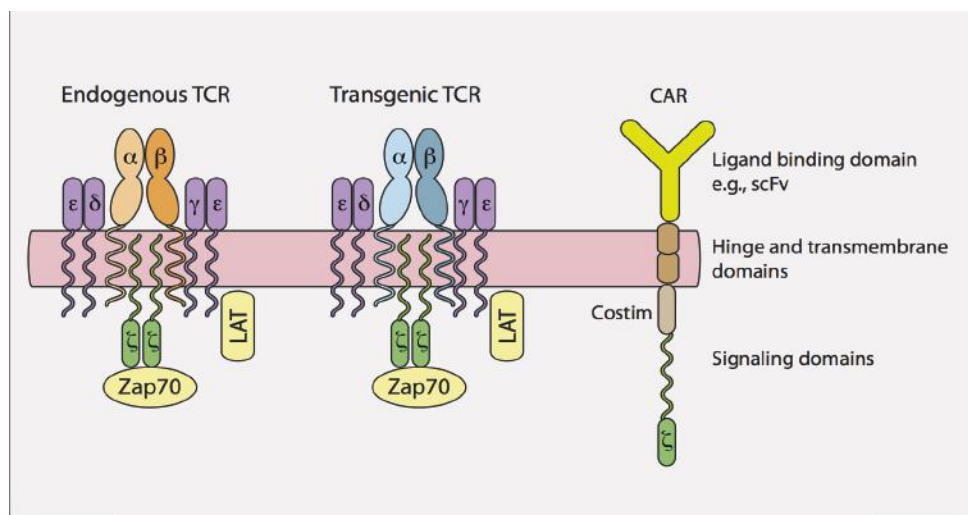
$\gamma\delta$  T cells are a subset of T cells that express alternative, clonally distributed TCRs and function as innate effectors and therefore are restricted in TCR diversity. Compared to  $\alpha\beta$  T cells, they do not target specific peptide antigens and are not constrained by the selectivity and restriction of the MHC. Although  $\gamma\delta$  T cells absolute counts are decreased and their proliferative capacity is decreased in the setting of glioblastoma, they can be expanded and activated *ex vivo* and have shown the ability to recognize and kill glioma cells *in vitro* while sparing cultured normal astrocytes. (150) Expanded and activated  $\gamma\delta$  T cells can mediate killing of glioblastoma and reduce tumor progression in mouse models.(151)

Cancer immunotherapy trials with autologous cd T cells have been investigated in parallel by Japanese, Australian and French groups. The French company, Innate Pharma, has conducted a phase I study in 10 patients with metastatic renal cell carcinoma to determine the maximum-tolerated dose of autologous TCRV $\gamma$ 9V $\gamma$ 2<sup>+</sup>  $\gamma\delta$  T cells and the safety of these cells as a therapeutic product. (152) In parallel with these studies, Kobayashi's group at Tokyo Women's Hospital investigated seven patients with advanced renal cell carcinoma engrafted with autologous  $\gamma\delta$  T cells.(153) Therefore, the authors concluded that  $\gamma\delta$  cell-based immunotherapy is a clinically beneficial and safe therapeutic option for patients with advanced renal cell carcinoma, whose rates of circulating  $\gamma\delta$  cells might constitute a favorable prognostic indicator.(154) To be successful, cancer immunotherapies involving  $\gamma\delta$  cells will require updated protocols which limit anergy and the use of drugs able to overcome immunoescape. Even though the former contingency is currently an open issue, the second one is already well underway.(155)

## Neoantigens in Cancer Immunotherapy

Immunotherapies which are boosting the ability of endogenous T cells to destroy cancer cells have showed therapeutic efficacy in a variety of human malignancies. Until now, evidence that the endogenous T cell compartment could help control tumor growth was in big part restricted to preclinical mouse tumor models and to human melanoma. (113) With respect to human studies, effects of the T cell cytokine IL-2 in a small subset of melanoma patients provided early clinical evidence of the potential of immunotherapy in the said disease. A randomized clinical trial was performed in 2010 showed that Ipilimumab, an antibody targets T cell checkpoint protein CTLA-4 could improve patient survival, even with metastatic melanoma. (6) As a direct test of the tumoricidal potential of the endogenous T cell compartment, a study by Rosenberg and colleagues exhibited that infusion of autologous *ex vivo* expanded TIL can induce objective clinical responses in metastatic melanoma (156), and at least part of this clinical activity is due to cytotoxic T cells (157). Basically, recent studies show that T cell-based immunotherapies are also effective in a range of other human malignancies.

With recent technology, we can check the uplevel of neoantigens as a tumor-specific mutation consequences, and emerging data suggest that this neoantigen play a role in the activity of clinical immunotherapies, thus this load may be used as cancer immunotherapy biomarker and an incentive development of novel therapeutic approaches can be provided.(113) In cancer, so-called neoepitope peptides are derived from proteins encoded by mutated genes. Recent



**Figure 7. Engineered T Cells that have retargeted specificity.**(149) Zap70: Zeta-chain-associated protein kinase 70, LAT: linker for activation of T cells, scFv: single-chain variable fragment. (Adapted with permission from Elsevier).

advances in next-generation DNA and RNA sequencing now enable rapid mapping of all expressed mutated genes in an individual tumor, and it is possible to predict epitopes that are efficiently presented on the surface of cancer cells. Thus, it has been demonstrated that CD8<sup>+</sup> T cells in human melanomas are able to recognize one or more neoepitopes from mutant proteins unique to that specific melanoma. However, efficient methods for studying CD4<sup>+</sup> T cells that recognize neoepitopes arising from somatic mutations in cancer have been lacking.(158) By using this approach, the authors identified neoepitope-specific CD4<sup>+</sup> T cells in two out of three melanoma patients.(159) The patients' T cells only recognized neoepitopes from the host's tumor, and they preferentially or exclusively noticed the neoepitope over the corresponding native, non-mutated peptide, demonstrating the exquisite specificity of the CD4<sup>+</sup> T cells.

CD4<sup>+</sup> T cells can antecedent cancer regression through direct killing of cancer cells, by altering the tumor-promoting function of cells in the surrounding tumor microenvironment, and by facilitating the induction, function and tumor infiltration of cancer-specific CD8<sup>+</sup> T cells.(160) These studies show that cancer-specific CD4<sup>+</sup> T cells can cause human tumor regression, adding to the importance of the findings by Linnemann, *et al.*(159) The ability to detect neoepitope specific CD4<sup>+</sup> T cells now allows validation of the hypothesis which the presence of neoepitope-specific CD4<sup>+</sup> T cells in human tumors correlates with overall clinical outcome.

The mutation in human tumor was considered to be individually different at meaningful frequencies and fractions, so the T cell reactivity against putative mutation-derived neoantigens interrogate technologies have to be developed based on individual tumor genome. With the development of deep-sequencing technologies, it has become doable to identify the mutations present within the protein-encoding part of the genome (the exome) of an individual tumor with relative ease and thereby predict potential neoantigens.(161)

Subsequent studies have demonstrated that cancer exome-based analyses can also be exploited in a clinical setting, to dissect T cell reactivity in patients who are treated by either TIL cell therapy or checkpoint blockade.(162,163) Furthermore, following this early work, the identification of neoantigens on the basis of cancer exome data has been documented in a variety of experimental model systems and human malignancies.(159,164-173) Based on data collected over the past few years, it is plausible that neoantigen-specific T cell reactivity forms a major active

ingredient of successful cancer immunotherapies. In other words, the genetic damage that on the one hand leads to oncogenic outgrowth can also be targeted by the immune system to control malignancies. Based on this finding, it will be important to engineer therapeutic interventions by which neoantigen-specific T cell reactivity is selectively enhanced. As it may be, the boosting of neoantigen-specific T cell reactivity which can be achieved with such personalized immunotherapies will further increase the spectrum of human malignancies that respond to cancer immunotherapy. (113)

## Cancer Vaccine Therapy

DC are designated as professional APC because of their capacity to provide T cells with all the signals required for antigen-specific T cell activation.(174) For optimal activation, T cells must receive at least three coordinated signals.(175,176) The first signal is delivered to the TCR by MHC molecules presenting antigen-derived peptides. The second signal was provided by the binding of co-stimulatory molecules with their respective ligands on T-cells. A crucial positive co-stimulatory signal is provided by the interaction of the B7 family ligands CD80 and CD86, expressed by the DC, with the CD28 receptor expressed on the T cell surface. The third signal, that called polarization signal, determines the commitment of naive CD4<sup>+</sup> T helper (TH0) cells towards TH1, TH2 or other fates. In the setting of cancer immunotherapy, the induction of a TH1 (cellular) immune response is highly desirable as this enables the generation of CTL capable of recognizing and destroying tumor cells in an antigen-specific fashion.(177)

As well as inducing antigen-specific CTL, which are part of the adaptive immune system, DC also capable of activating NK cells, which are prime players in the innate immune system.(178) NK cells have both cytotoxic and immunoregulatory functions and require priming to accomplish their full effector potential. Many of the cytokines that play a principal role in stimulating NK cell functions can be provided by DC, *e.g.*, IL-12, IL-15, IL-18 and type I IFN.(179)

A more detailed understanding of the mechanisms leading to strong cellular immunity is necessary to enable rational approaches to the vaccine design. Two recent conceptual breakthroughs in this regard are our understanding that DC play a pivotal role in initiating the immune response to foreign antigens and the realization

that adjuvants act primarily because they are DC activators. (180) Therapeutic vaccines in chronic infections (or cancer) have two objectives: one is priming, whereas the other one is the modulation or reprogramming of memory cells, i.e., to transition from one type of immunity to another (e.g., regulatory to cytotoxic).

Tumor cells themselves are poor APC, which raises the question of how such potent immunity can be generated. Mouse models demonstrate that the generation of protective anti-tumor immunity relies on the presentation of tumor antigens by DC. (181,182) These cells special features in coordinating innate and adaptive immune responses provided an idea for a DCs-involved vaccination strategies, aiming to induce tumor-specific effector T cells that can reduce the tumor mass specifically and induce the immunological memory to control tumor relapse. First step for these processes is to provide DCs with tumor-specific antigens. This can be achieved either by culturing *ex vivo* patients-derived DCs that have been induced with an adjuvant (induces DC maturation) and the tumor-specific antigen, and then injecting these cells back into the patient, or by inducing DCs to take up the tumor-specific antigen *in vivo*. To improve therapeutic use of DC vaccination strategies, it is necessary to understand the biology of DC and how they regulate the innate and the adaptive immune systems, particularly in the context of the tumor microenvironment. (183)

Most phase 1 and phase 2 trials of cancer vaccines have involved patients with an extensive cancer burden, impaired immune function, or both. (184) An alternative to infusion of preformed tumor-specific antibodies or T cells, known as passive immunotherapy, is active specific immunotherapy (*i.e.*, cancer vaccines) designed to elicit or boost similar tumor antibodies and T cells in patients. Some examples are vaccines against breast cancer (the HER2 antigen) (185-7), B-cell lymphoma (the tumor immunoglobulin idiotype) (188), lung cancer (the Mucin 1 cell surface associated (MUC1) antigen) (189), melanoma (DC loaded with tumor peptides or killed tumor cells) (190,191), pancreatic cancer (telomerase peptides) (192), and prostate cancer (DC loaded with prostatic acid phosphatase) (193). The results of these trials are promising because in each there was evidence of an immune response to the vaccine, and in a few cases there were clinical responses with minimal or no adverse effects. Regarding the limited number of completed phase 3 trials, most have failed to presentate a significant benefit with respect to predetermined end points (194), but nevertheless provided information for design of future trials, especially

concerning the choice of patients and stage of disease.

The immunosuppressive microenvironment of a tumor can inhibit the effect of therapeutic vaccines, both during the immunity induction and in the response effector phase. So, the negative regulators of the activation of effector T cells need to be blocked to improve the induction phase. (195) Antibodies against one such molecule, CTLA-4 are being evaluated in clinical trials. (196,197) CTLA-4 is expressed on activated T cells, where it serves as a brake halting the activation. Blocking activity of CTLA-4 allows larger expansion of all T-cell populations, presumably including those with antitumor reactivity. In a recent pilot trial involving 14 patients with hormone-refractory prostate cancer, systemic treatment with anti-CTLA-4 antibody increased antitumor immunity, resulting in a reduction in prostate-specific antigen of more than 50% in two patients and less than 50% in eight patients. (197) The side effects were rash and pruritus, which required treatment with corticosteroids in the two patients with the best response. (62)

Approaches to eliminating immunosuppressive regulatory T cells before vaccination are also being tested. One promising reagent is Denileukin diftitox (Ontak, Seragen), a recombinant fusion protein composed of IL-2 and diphtheria toxin. It targets the high-affinity IL-2 receptor (CD25), which is displayed in abundance by regulatory T cells. When administered to patients with melanoma, protein depletes the blood of regulatory T cells. In most patients (90%), this treatment has resulted in the production of melanoma-specific CD8 T cells. (198)

## Conclusion

We now have detailed knowledge of the molecular basis of cancer to allow a more personalized treatment based on genomic sequencing of an individual's cancer cells to identify specific mutations in genes. These mutations can then be targeted with compounds to blockade the downstream pathways which drive cancer development and progression. Thus, each specific mutation serves as the predictive biomarker for selecting patients for treatment with a given agent.

Clinical data generated principally over the past 5 years offer that we are at the threshold of golden era for adoptive T cell therapy, where advances in basic immunology have informed the development of a new field of synthetic immunology which may increase the potency

of approaches that target cancer. Cancer immunotherapies, a no stranger to obstacles, looks as a very promising future for incurable cancers therapy, using anti-CTLA-4 and anti-PD-1 antibodies or adoptively transferring T cells for blocking inhibitory immune signaling.

## References

- Gravitz L. Cancer Immunotherapy. *Nature*. 2013; 504: S1.
- Palucka K. Q&A: Evidence presenter. Interview by Marian Turner. *Nature*. 2013; 504: S9.
- Humphries C. Adoptive cell therapy: Honing that killer instinct. *Nature*. 2013; 504: S13-5.
- Weintraub K. Drug development: Releasing the brakes. *Nature*. 2013; 504: S6-8.
- Littman DR. Releasing the brakes on cancer immunotherapy. *Cell*. 2015; 162: 1186-90.
- Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, et al. Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med*. 2010; 363: 711-23.
- Ishida Y, Agata Y, Shibahara K, Honjo T. Induced expression of PD-1, a novel member of the immunoglobulin gene superfamily, upon programmed cell death. *EMBO J*. 1992; 11: 3887-95.
- Pauken KE, Wherry EJ. Overcoming T cell exhaustion in infection and cancer. *Trends Immunol*. 2015; 36: 265-76.
- Fialkow PJ. Clonal origin of human tumors. *Biochim Biophys Acta*. 1976; 458: 283-321.
- Fidler IJ, Kripke ML. Metastasis results from preexisting variant cells within a malignant tumor. *Science*. 1977; 197: 893-5.
- Nowell PC. The clonal evolution of tumor cell populations. *Science*. 1976; 194: 23-8.
- Baylin SB, Jones PA. A decade of exploring the cancer epigenome — biological and translational implications. *Nat Rev Cancer*. 2011; 11: 726-34.
- Mueller MM, Fusenig NE. Friends or foes — bipolar effects of the tumour stroma in cancer. *Nat Rev Cancer*. 2004; 4: 839-49.
- Jordan CT, Guzman ML, Noble M. Cancer stem cells. *N Engl J Med*. 2006; 355: 1253-61.
- Cohnheim J. Congenitales, quergestreiftes Muskelsarkom der Nieren. *Path Anat Physiol Klin Med*. 1875; 65: 64-9.
- Wicha MS, Liu S, Dontu G. Cancer stem cells: an old idea — a paradigm shift. *Cancer Res*. 2006; 66: 1883-90.
- Bonnet D, Dick JE. Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cell. *Nat Med*. 1997; 3: 730-7.
- Lapidot T, Sirard C, Vormoor J, Murdoch B, Hoang T, Caceres-Cortes J, et al. A cell initiating human acute myeloid leukaemia after transplantation into SCID mice. *Nature*. 1994; 367: 645-8.
- Magee JA, Piskounova E, Morrison SJ. Cancer stem cells: impact, heterogeneity, and uncertainty. *Cancer Cell*. 2012; 21: 283-96.
- Yamashita T, Wang XW. Cancer stem cells in the development of liver cancer. *J Clin Invest*. 2013; 123: 1911-8.
- Braun S, Vogl FD, Naume B, Janni W, Osborne MP, Coombes RC, et al. A pooled analysis of bone marrow micrometastasis in breast cancer. *N Engl J Med*. 2005; 353: 793-802.
- Janni W, Vogl FD, Wiedswang G, Synnestvedt M, Fehm T, Jückstock J, et al. Persistence of disseminated tumor cells in the bone marrow of breast cancer patients predicts increased risk for relapse — a European pooled analysis. *Clin Cancer Res*. 2011; 17: 2967-76.
- Oskarson T, Batile E, Massague J. Metastatic stem cells: sources, niches, and vital pathways. *Cell Stem Cell*. 2014; 14: 306-21.
- Paez JG, Jänne PA, Lee JC, Tracy S, Greulich H, Gabriel S, et al. EGFR mutations in lung cancer: correlation with clinical response to gefitinib therapy. *Science*. 2004; 304: 1497-500.
- Lynch TJ, Bell DW, Sordella R, Gurubhagavatula S, Okimoto RA, Brannigan BW, et al. Activating mutations in the epidermal growth factor receptor underlying responsiveness of non-small-cell lung cancer to gefitinib. *N Engl J Med*. 2004; 350: 2129-39.
- Markowitz SD, Bertagnolli MM. Molecular origins of cancer: Molecular basis of colorectal cancer. *N Engl J Med*. 2009; 361: 2449-60.
- Ogino S, Goel A. Molecular classification and correlates in colorectal cancer. *J Mol Diagn*. 2008; 10: 13-27.
- Pritchard CC, Grady WM. Colorectal cancer molecular biology moves into clinical practice. *Gut*. 2011; 60: 116-29.
- Dolle JM, Daling JR, White E, Brinton LA, Doody DR, Porter PL, et al. Risk factors for triple-negative breast cancer in women under the age of 45 years. *Cancer Epidemiol Biomarkers Prev*. 2009; 18: 1157-66.
- Trivers KF, Lund MJ, Porter PL, Liff JM, Flagg EW, Coates RJ, et al. The epidemiology of triple-negative breast cancer, including race. *Cancer Causes Control*. 2009; 20: 1071-82.
- Tomlins SA, Rhodes DR, Perner S, Dhanasekaran SM, Mehra R, Sun XW, et al. Recurrent fusion of TMPRSS2 and ETS transcription factor genes in prostate cancer. *Science*. 2005; 310: 644-8.
- Samuels Y, Wang Z, Bardelli A, Silliman N, Ptak J, Szabo S, et al. High frequency of mutations of the PIK3CA gene in human cancers. *Science*. 2004; 304: 554.
- Wood LD, Parsons DW, Jones S, Lin J, Sjöblom T, Leary RJ, et al. The genomic landscapes of human breast and colorectal cancers. *Science*. 2007; 318: 1108-13.
- Esteller M. Epigenetics in cancer. *N Engl J Med*. 2008; 358: 1148-59.
- Zou W. Regulatory T cells, tumour immunity and immunotherapy. *Nat Rev Immunol*. 2006; 6: 295-307.
- Allen M, Louise Jones J. Jekyll and Hyde: the role of the microenvironment on the progression of cancer. *J Pathol*. 2011; 223: 162-76.
- Graziano DF, Finn OJ. Tumor antigens and tumor antigen discovery. *Cancer Treat Res*. 2005; 123: 89-111.
- Srivastava PK, Old LJ. Individually distinct transplantation antigens of chemically induced mouse tumors. *Immunol Today*. 1988; 9: 78-83.
- Dunn GP, Bruce AT, Ikeda H, Old LJ, Schreiber RD. Cancer immunoeediting: from immunosurveillance to tumor escape. *Nat Immunol*. 2002; 3: 991-8.
- Dunn GP, Bruce AT, Sheehan KC, Shankaran V, Uppaluri R, Bui JD, et al. A critical function for type I interferons in cancer immunoeediting. *Nat Immunol*. 2005; 6: 722-9. [Erratum in: *Nat Immunol*. 2005; 6: 852.]
- Smyth MJ, Dunn GP, Schreiber RD. Cancer immunosurveillance and immunoeediting: the roles of immunity in suppressing tumor development and shaping tumor immunogenicity. *Adv Immunol*. 2006; 90: 1-50.
- Simson L, Ellyard JI, Dent LA, Matthaei KI, Rothenberg ME, Foster PS, et al. Regulation of carcinogenesis by IL-5 and CCL11: a potential role for eosinophils in tumor immune surveillance. *J Immunol*. 2007; 178: 4222-9.
- Shankaran V, Ikeda H, Bruce AT, White JM, Swanson PE, Old LJ, et al. IFN $\gamma$  and lymphocytes prevent primary tumour development and shape tumour immunogenicity. *Nature*. 2001; 410: 1107-11.
- Dunn GP, Old LJ, Schreiber RD. The three Es of cancer immunoeediting. *Annu Rev Immunol*. 2004; 22: 329-60.



45. Koebel CM, Vermi W, Swann JB, Zerafa N, Rodig SJ, Old LJ, et al. Adaptive immunity maintains occult cancer in an equilibrium state. *Nature*. 2007; 450: 903-7.
46. Sonabend AM, Ogden AT, Maier LM, Anderson DE, Canoll P, Bruce JN, et al. Medulloblastoma: challenges for effective immunotherapy. *J Neurooncol*. 2011; 108: 1-10.
47. Vesely MD, Kershaw MH, Schreiber RD, Smyth MJ. Natural innate and adaptive immunity to cancer. *Annu Rev Immunol*. 2011; 29: 235-71.
48. Schreiber RD, Old LJ, Smyth MJ. Cancer immunoeediting: integrating immunity's roles in cancer suppression and promotion. *Science*. 2011; 331: 1565-70.
49. Myron Kauffman H, McBride MA, Cherikh WS, Spain PC, Marks WH, Roza AM. Transplant tumor registry: donor related malignancies. *Transplantation*. 2002; 74: 358-62.
50. MacKie RM, Reid R, Junor B. Fatal melanoma transferred in a donated kidney 16 years after melanoma surgery. *N Engl J Med*. 2003; 348: 567-8.
51. Sonabend AM, Dana K, Lesniak MS. Targeting epidermal growth factor receptor variant III: a novel strategy for the therapy of malignant glioma. *Expert Rev Anticancer Ther*. 2007; 7: S45-50.
52. Seliger B, Cabrera T, Garrido F, Ferrone S. HLA class I antigen abnormalities and immune escape by malignant cells. *Semin Cancer Biol*. 2002; 12: 3-13.
53. Rabinovich GA, Gabrilovich D, Sotomayor EM. Immunosuppressive strategies that are mediated by tumor cells. *Annu Rev Immunol*. 2007; 25: 267-96.
54. Teicher BA. Transforming growth factor-beta and the immune response to malignant disease. *Clin Cancer Res*. 2007; 13: 6247-51.
55. Houston A, Bennett MW, O'Sullivan GC, Shanahan F, O'Connell J. Fas ligand mediates immune privilege and not inflammation in human colon cancer, irrespective of TGF-beta expression. *Br J Cancer*. 2003; 89: 1345-51.
56. Uyttenhove C, Pilotte L, Théate I, Stroobant V, Colau D, Parmentier N, et al. Evidence for a tumoral immune resistance mechanism based on tryptophan degradation by indoleamine 2,3-dioxygenase. *Nat Med*. 2003; 9: 1269-74.
57. Muller AJ, Prendergast GC. Indoleamine 2,3-dioxygenase in immune suppression and cancer. *Curr Cancer Drug Targets*. 2007; 7: 31-40.
58. Munn DH, Zhou M, Attwood JT, Bondarev I, Conway SJ, Marshall B, et al. Prevention of allogeneic fetal rejection by tryptophan catabolism. *Science*. 1998; 281: 1191-3.
59. Munn DH, Sharma MD, Lee JR, Jhaver KG, Johnson TS, Keskin DB, et al. Potential regulatory function of human dendritic cells expressing indoleamine 2,3-dioxygenase. *Science*. 2002; 297: 1867-70.
60. Hou DY, Muller AJ, Sharma MD, DuHadaway J, Banerjee T, Johnson M, et al. Inhibition of indoleamine 2,3-dioxygenase in dendritic cells by stereoisomers of 1-methyl-tryptophan correlates with antitumor responses. *Cancer Res*. 2007; 67: 792-801.
61. Zheng X, Koropatnick J, Li M, Zhang X, Ling F, Ren X, et al. Reinstalling antitumor immunity by inhibiting tumor-derived immunosuppressive molecule IDO through RNA interference. *J Immunol*. 2006; 177: 5639-46.
62. Finn OJ. *Cancer Immunology*. *N Engl J Med*. 2008; 358: 2704-15.
63. Woo EY, Chu CS, Goeletz TJ, Schlienger K, Yeh H, Coukos G, et al. Regulatory CD4(+)CD25(+) T cells in tumors from patients with early-stage non-small cell lung cancer and late-stage ovarian cancer. *Cancer Res*. 2001; 61: 4766-72.
64. Liu VC, Wong LY, Jang T, Shah AH, Park I, Yang X, et al. Tumor evasion of the immune system by converting CD4+CD25- T cells into CD4+CD25+ T regulatory cells: role of tumor-derived TGF-beta. *J Immunol*. 2007; 178: 2883-92.
65. Anderson MJ, Shafer-Weaver K, Greenberg NM, Hurwitz AA. Tolerization of tumor-specific T cells despite efficient initial priming in a primary murine model of prostate cancer. *J Immunol*. 2007; 178: 1268-76. [Erratum in: *J Immunol*. 2007; 179: 7184.]
66. Bergmann C, Strauss L, Zeidler R, Lang S, Whiteside TL. Expansion and characteristics of human T regulatory type 1 cells in co-cultures simulating tumor microenvironment. *Cancer Immunol Immunother*. 2007; 56: 1429-42.
67. Chikamatsu K, Sakakura K, Whiteside TL, Furuya N. Relationships between regulatory T cells and CD8+ effector populations in patients with squamous cell carcinoma of the head and neck. *Head Neck*. 2007; 29: 120-7.
68. Fecci PE, Mitchell DA, Whitesides JF, Xie W, Friedman AH, Archer GE, et al. Increased regulatory T-cell fraction amidst a diminished CD4 compartment explains cellular immune defects in patients with malignant glioma. *Cancer Res*. 2006; 66: 3294-302.
69. Schmielau J, Finn OJ. Activated granulocytes and granulocyte-derived hydrogen peroxide are the underlying mechanism of suppression of t-cell function in advanced cancer patients. *Cancer Res*. 2001; 61: 4756-60.
70. Nagaraj S, Gabrilovich DI. Myeloid-derived suppressor cells. *Adv Exp Med Biol*. 2007; 601: 213-23.
71. Kusmartsev S, Nagaraj S, Gabrilovich DI. Tumor-associated CD8+ T cell tolerance induced by bone marrow-derived immature myeloid cells. *J Immunol*. 2005; 175: 4583-92.
72. Sinha P, Clements VK, Bunt SK, Albelda SM, Ostrand-Rosenberg S. Crosstalk between myeloid-derived suppressor cells and macrophages subverts tumor immunity toward a type 2 response. *J Immunol*. 2007; 179: 977-83.
73. Piccart-Gebhart MJ, Procter M, Leyland-Jones B, Goldhirsch A, Untch M, Smith I, et al. Trastuzumab after adjuvant chemotherapy in HER2-positive breast cancer. *N Engl J Med*. 2005; 353: 1659-72.
74. Weiner GJ, Link BK. Monoclonal antibody therapy of B cell lymphoma. *Expert Opin Biol Ther*. 2004; 4: 375-85.
75. Mendelsohn J. Epidermal growth factor receptor inhibition by a monoclonal antibody as anticancer therapy. *Clin Cancer Res*. 1997; 3: 2703-7.
76. Giaccone G. Epidermal growth factor receptor inhibitors in the treatment of non-small-cell lung cancer. *J Clin Oncol*. 2005; 23: 3235-42.
77. Kim ES, Vokes EE, Kies MS. Cetuximab in cancers of the lung and head & neck. *Semin Oncol*. 2004; 31 (Suppl 1): 61-7.
78. Lien S, Lowman HB. Therapeutic anti-VEGF antibodies. *Handb Exp Pharmacol*. 2008; 181: 131-50.
79. Melero I, Hervas-Stubbs S, Glennie M, Pardoll DM, Chen L. Immunostimulatory monoclonal antibodies for cancer therapy. *Nat Rev Cancer*. 2007; 7: 95-106.
80. Small EJ, Schellhammer PF, Higano CS, Redfern CH, Nemunaitis JJ, Valone FH, et al. Placebo-controlled phase III trial of immunologic therapy with sipuleucel-T (APC8015) in patients with metastatic, asymptomatic hormone refractory prostate cancer. *J Clin Oncol*. 2006; 24: 3089-94.
81. Kantoff PW, Higano CS, Shore ND, Berger ER, Small EJ, Penson DF, et al. Sipuleucel-T immunotherapy for castration-resistant prostate cancer. *N Engl J Med*. 2010; 363: 411-22.
82. Kantoff PW, Schuetz TJ, Blumenstein BA, Glode LM, Bilhartz DL, Wyand M, et al. Overall survival analysis of a phase II randomized controlled trial of a Poxviral based PSA-targeted immunotherapy in metastatic castration-resistant prostate cancer. *J Clin Oncol*. 2010; 28: 1099-105.
83. Hamid O, Robert C, Daud A, Hodi FS, Hwu WJ, Kefford R, et al. Safety and tumor responses with lambrolizumab (anti-PD-1) in melanoma. *N Engl J Med*. 2013; 369: 134-44.

84. Schadendorf D, Hodi FS, Robert C, Weber JS, Margolin K, Hamid O, et al. Pooled analysis of long-term survival data from phase II and phase III trials of ipilimumab in unresectable or metastatic melanoma. *J Clin Oncol.* 2015; 33: 1889-94.
85. Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, McDermott DF, et al. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. *N Engl J Med.* 2012; 366: 2443-54.
86. Monteiro J. Cancer immunotherapy scores again. *Cell.* 2015; 160: 7-9.
87. Kohler G, Milstein C. Continuous cultures of fused cells secreting antibody of predefined specificity. *Nature.* 1975; 256: 495-7.
88. Muller AJ, Scherle PA. Targeting the mechanisms of tumoral immune tolerance with small-molecule inhibitors. *Nature Rev Cancer.* 2006; 6: 613-25.
89. Jiang XR, Song A, Bergelson S, Arroll T, Parekh B, May K, et al. Advances in the assessment and control of the effector functions of therapeutic antibodies. *Nat Rev Drug Discov.* 2011; 10: 101-11.
90. Weiner LM, Murray JC, Shuptrine CW. Antibody-based immunotherapy of cancer. *Cell.* 2012; 148: 1081-4.
91. Ruf P, Lindhofer H. Induction of a long-lasting antitumor immunity by a trifunctional bispecific antibody. *Blood.* 2001; 98: 2526-34.
92. Steiner M, Neri D. Antibody-radionuclide conjugates for cancer therapy: historical considerations and new trends. *Clin Cancer Res.* 2011; 17: 6406-16.
93. Nahta R, Esteva FJ. Herceptin: mechanisms of action and resistance. *Cancer Lett.* 2006; 232: 123-38.
94. Taylor C, Hershman D, Shah N, Suci-Foca N, Petrylak DP, Taub R, et al. Augmented HER-2 specific immunity during treatment with trastuzumab and chemotherapy. *Clin Cancer Res.* 2007; 13: 5133-43.
95. Lewis Phillips GD, Li G, Dugger DL, Crocker LM, Parsons KL, Mai E, et al. Targeting HER2-positive breast cancer with trastuzumab-DM1, an antibody-cytotoxic drug conjugate. *Cancer Res.* 2008; 68: 9280-90.
96. Mellman I, Coukos G, Dranoff G. Cancer immunotherapy comes of age. *Nature.* 2011; 480: 480-9.
97. Greenwald RJ, Freeman GJ, Sharpe AH. The B7 family revisited. *Annu Rev Immunol.* 2005; 23: 515-48.
98. Zou W, Chen L. Inhibitory B7-family molecules in the tumour microenvironment. *Nature Rev Immunol.* 2008; 8: 467-77.
99. Robert C, Thomas L, Bondarenko I, O'Day S, Weber J, Garbe C, et al. Ipilimumab plus dacarbazine for previously untreated metastatic melanoma. *N Engl J Med.* 2011; 364: 2517-26.
100. Andre F, Dieci MV, Dubsky P, Sotiriou C, Curigliano G, Denkert C, et al. Molecular pathways: involvement of immune pathways in the therapeutic response and outcome in breast cancer. *Clin Cancer Res.* 2013; 19: 28-33.
101. May KF Jr, Gulley JL, Drake CG, Dranoff G, Kantoff PW. Prostate cancer immunotherapy. *Clin Cancer Res.* 2011; 17: 5233-8.
102. Page DB, Postow MA, Callahan MK, Allison JP, Wolchok JD. Immune modulation in cancer with antibodies. *Annu Rev Med.* 2014; 65: 185-202.
103. Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. *Nat Rev Cancer.* 2012; 12: 252-64.
104. Mellor AL, Keskin DB, Johnson T, Chandler P, Munn DH. Cells expressing indoleamine 2,3-dioxygenase inhibit T cell responses. *J Immunol.* 2002; 168: 3771-6.
105. Friberg M, Jennings R, Alsarraj M, Dessureault S, Cantor A, Extermann M, et al. Indoleamine 2,3-dioxygenase contributes to tumor cell evasion of T cell-mediated rejection. *Int J Cancer.* 2002; 101: 151-5.
106. Munn DH, Mellor AL. Indoleamine 2,3-dioxygenase and tumor-induced tolerance. *J Clin Invest.* 2007; 117: 1147-54.
107. Bak SP, Alonso A, Turk MJ, Berwin B. Murine ovarian cancer vascular leukocytes require arginase-1 activity for T cell suppression. *Mol Immunol.* 2008; 46: 258-68.
108. Ochoa AC, Zea AH, Hernandez C, Rodriguez PC. Arginase, prostaglandins, and myeloid-derived suppressor cells in renal cell carcinoma. *Clin Cancer Res.* 2007; 13: 721S-6S.
109. Rodríguez PC, Ochoa AC. Arginine regulation by myeloid derived suppressor cells and tolerance in cancer: mechanisms and therapeutic perspectives. *Immunol Rev.* 2008; 222: 180-91.
110. Löb S, Königsrainer A, Zieker D, Brücher BL, Rammensee HG, Opelz G, et al. IDO1 and IDO2 are expressed in human tumors: levo- but not dextro-1-methyl tryptophan inhibits tryptophan catabolism. *Cancer Immunol Immunother.* 2009; 58: 153-7.
111. Qian F, Vilella J, Wallace PK, Mhawech-Fauceglia P, Tario JD Jr, Andrews C, et al. Efficacy of levo-1-methyl tryptophan and dextro-1-methyl tryptophan in reversing indoleamine-2,3-dioxygenase-mediated arrest of T-cell proliferation in human epithelial ovarian cancer. *Cancer Res.* 2009; 69: 5498-504.
112. Reisser D, Onier-Cherix N, Jeannin JF. Arginase activity is inhibited by L-NAME, both in vitro and in vivo. *J Enzyme Inhib Med Chem.* 2002; 17: 267-70.
113. Schumacher TN, Schreiber RD. Neoantigens in cancer immunotherapy. *Science.* 2015; 348: 69-74.
114. Sharma P, Allison JP. The future of immune checkpoint therapy. *Science.* 2015; 348: 56-61.
115. Townsend SE, Allison JP. Tumor rejection after direct costimulation of CD8+ T cells by B7-transfected melanoma cells. *Science.* 1993; 259: 368-70.
116. Linsley PS, Greene JL, Tan P, Bradshaw J, Ledbetter JA, Anasetti C, et al. Coexpression and functional cooperation of CTLA-4 and CD28 on activated T lymphocytes. *J Exp Med.* 1992; 176: 1595-604.
117. Walunas TL, Lenschow DJ, Bakker CY, Linsley PS, Freeman GJ, Green JM, et al. CTLA-4 can function as a negative regulator of T cell activation. *Immunity.* 1994; 1: 405-13.
118. Krummel MF, Allison JP. CD28 and CTLA-4 have opposing effects on the response of T cells to stimulation. *J Exp Med.* 1995; 182: 459-65.
119. Sharma P, Wagner K, Wolchok JD, Allison JP. Novel cancer immunotherapy agents with survival benefit: recent successes and next steps. *Nat Rev Cancer.* 2011; 11: 805-12.
120. Freeman GJ, Long AJ, Iwai Y, Bourque K, Chernova T, Nishimura H, et al. Engagement of the PD-1 immunoinhibitory receptor by a novel B7 family member leads to negative regulation of lymphocyte activation. *J Exp Med.* 2000; 192: 1027-34.
121. Dong H, Strome SE, Salomao DR, Tamura H, Hirano F, Flies DB, et al. Tumor-associated B7-H1 promotes T-cell apoptosis: a potential mechanism of immune evasion. *Nat Med.* 2002; 8: 793-800.
122. Park JJ, Omiya R, Matsumura Y, Sakoda Y, Kuramasu A, Augustine MM, et al. B7-H1/CD80 interaction is required for the induction and maintenance of peripheral T-cell tolerance. *Blood.* 2010; 116: 1291-8.
123. Parham P. MHC class I molecules and KIRs in human history, health and survival. *Nat Rev Immunol.* 2005; 5: 201-14.
124. Deguine J, Breart B, Lemaitre F, Di Santo JP, Bousso P. Intravital imaging reveals distinct dynamics for natural killer and CD8+ T cells during tumor regression. *Immunity.* 2010; 33: 632-44.
125. Restivo NP, Dudley ME, Rosenberg SA. Adoptive immunotherapy for cancer: harnessing the T cell response. *Nat Rev Immunol.* 2012; 12: 269-81.
126. Ruf P, Lindhofer H. Induction of a long-lasting antitumor immunity by a trifunctional bispecific antibody. *Blood.* 2001; 98: 2526-34.
127. Miescher S, Whiteside TL, de Tribolet N, von Flidner V. In situ

- characterization, clonogenic potential, and antitumor cytolytic activity of T lymphocytes infiltrating human brain cancers. *J Neurosurg.* 1988; 68: 438-48.
128. Bielamowicz K, Khawja S, Ahmed N. Adoptive cell therapies for glioblastoma. *Front Oncol.* 2013; 3: 275.
  129. Rosenberg SA, Packard BS, Aebersold PM, Solomon D, Topalian SL, Toy ST, et al. Use of tumor-infiltrating lymphocytes and interleukin-2 in the immunotherapy of patients with metastatic melanoma. A preliminary report. *N Engl J Med.* 1988; 319: 1676-80.
  130. Hunder NN, Wallen H, Cao J, Hendricks DW, Reilly JZ, Rodmyre R, et al. Treatment of metastatic melanoma with autologous CD4+T cells against NY-ESO-1. *N Engl J Med.* 2008; 358: 2698-703.
  131. Butler MO, Friedlander P, Milstein MI, Mooney MM, Metzler G, Murray AP, et al. Establishment of antitumor memory in humans using in vitro-educated CD8+ T cells. *Sci Transl Med.* 2011; 3: 80ra34.
  132. Chapuis AG, Thompson JA, Margolin KA, Rodmyre R, Lai IP, Dowdy K, et al. Transferred melanoma-specific CD8+T cells persist, mediate tumor regression, and acquire central memory phenotype. *Proc Natl Acad Sci USA.* 2012;109: 4592-7.
  133. Grupp SA, Kalos M, Barrett D, Aplenc R, Porter DL, Rheingold SR, et al. Chimeric antigen receptor-modified T cells for acute lymphoid leukemia. *N Engl J Med.* 2013; 368: 1509-18.
  134. Morgan DA, Ruscetti FW, Gallo R. Selective in vitro growth of T lymphocytes from normal human bone marrows. *Science.* 1976; 193: 1007-8.
  135. Rosenberg SA, Mulé JJ, Spiess PJ, Reichert CM, Schwarz SL. Regression of established pulmonary metastases and subcutaneous tumor mediated by the systemic administration of high-dose recombinant interleukin 2. *J Exp Med.* 1985; 161: 1169-88.
  136. Eberlein TJ, Rosenstein M, Rosenberg SA. Regression of a disseminated syngeneic solid tumor by systemic transfer of lymphoid cells expanded in interleukin 2. *J Exp Med.* 1982; 156: 385-97.
  137. Donohue JH, Rosenstein M, Chang AE, Lotze MT, Robb RJ, Rosenberg SA. The systemic administration of purified interleukin 2 enhances the ability of sensitized murine lymphocytes to cure a disseminated syngeneic lymphoma. *J Immunol.* 1984; 132: 2123-8.
  138. Kessels HW, Wolkers MC, van den Boom MD, van der Valk MA, Schumacher TN. Immunotherapy through TCR gene transfer. *Nat Immunol.* 2001; 2: 957-61.
  139. Morgan RA, Dudley ME, Wunderlich JR, Hughes MS, Yang JC, Sherry RM, et al. Cancer regression in patients after transfer of genetically engineered lymphocytes. *Science.* 2006; 314: 126-9.
  140. Kochenderfer JN, Wilson WH, Janik JE, Dudley ME, Stetler-Stevenson M, Feldman SA, et al. Eradication of B-lineage cells and regression of lymphoma in a patient treated with autologous T cells genetically engineered to recognize CD19. *Blood.* 2010; 116: 4099-102.
  141. Gross G, Waks T, Eshhar Z. Expression of immunoglobulin-T-cell receptor chimeric molecules as functional receptors with antibody-type specificity. *Proc Natl Acad Sci USA.* 1989; 86: 10024-8.
  142. Maher J, Brentjens RJ, Gunset G, Rivière I, Sadelain M. Human T-lymphocyte cytotoxicity and proliferation directed by a single chimeric TCRzeta/CD28 receptor. *Nat Biotechnol.* 2002; 20: 70-5.
  143. Imai C, Mihara K, Andreansky M, Nicholson IC, Pui CH, Geiger TL, et al. Chimeric receptors with 4-1BB signaling capacity provoke potent cytotoxicity against acute lymphoblastic leukemia. *Leukemia.* 2004; 18: 676-84.
  144. Song DG, Ye Q, Carpenito C, Poussin M, Wang LP, Ji C, et al. In vivo persistence, tumor localization, and antitumor activity of CAR-engineered T cells is enhanced by costimulatory signaling through CD137 (4-1BB). *Cancer Res.* 2011; 71: 4617-27.
  145. Baker M. Companies ponder how truly 'personal' medicines can get. *Nat Med.* 2011; 17: 519.
  146. Brindley DA, Davie NL, Culme-Seymour EJ, Mason C, Smith DW, Rowley JA. Peak serum: implications of serum supply for cell therapy manufacturing. *Regen Med.* 2012; 7: 7-13.
  147. Mason C, Manzotti E. Regenerative medicine cell therapies: numbers of units manufactured and patients treated between 1988 and 2010. *Regen Med.* 2010; 5: 307-13.
  148. Michalos G, Makris S, Papakostas N, Mourtzis D, Chryssolouris G. Automotive assembly technologies review: challenges and outlook for a flexible and adaptive approach. *CIRP J Manuf Sci Technol.* 2010; 2: 81-91.
  149. Kalos M, June CH. Adoptive T cell transfer for cancer immunotherapy in the era of synthetic biology. *Immunity.* 2013; 39: 49-60.
  150. Bryant NL, Suarez-Cuervo C, Gillespie GY, Markert JM, Nabors LB, Meleth S, et al. Characterization and immunotherapeutic potential of gammadelta T-cells in patients with glioblastoma. *Neuro Oncol.* 2009; 11: 357-67.
  151. Bryant NL, Gillespie GY, Lopez RD, Markert JM, Cloud GA, Langford CP, et al. Preclinical evaluation of ex vivo expanded/activated gammadelta T cells for immunotherapy of glioblastoma multiforme. *J Neurooncol.* 2011; 101: 179-88.
  152. Bennouna J, Bompas E, Neidhardt EM, Rolland F, Philip I, Galea C, et al. Phase-I study of Innacell gammadelta, an autologous cell-therapy product highly enriched in gamma9delta2 T lymphocytes, in combination with IL-2, in patients with metastatic renal cell carcinoma. *Cancer Immunol Immunother.* 2008; 57: 1599-609.
  153. Kobayashi H, Tanaka Y, Yagi J, Osaka Y, Nakazawa H, Uchiyama T, et al. Safety profile and anti-tumor effects of adoptive immunotherapy using gamma-delta T cells against advanced renal cell carcinoma: a pilot study. *Cancer Immunol Immunother.* 2007; 56: 469-76.
  154. Kobayashi H, Tanaka Y, Nakazawa H, Yagi J, Minato N, Tanabe K. A new indicator of favorable prognosis in locally advanced renal cell carcinomas: gamma delta T-cells in peripheral blood. *Anticancer Res.* 2011; 31: 1027-31.
  155. Fournie JJ, Sicard H, Poupot M, Bezombes C, Blanc A, Romagne F, et al. What lessons can be learned from  $\gamma\delta$  T cell-based cancer immunotherapy trials? *Cell Mol Immunol.* 2013; 10: 35-41.
  156. Hinrichs CS, Rosenberg SA. Exploiting the curative potential of adoptive T-cell therapy for cancer. *Immunol Rev.* 2014; 257: 56-71.
  157. Dudley ME, Gross CA, Somerville RP, Hong Y, Schaub NP, Rosati SF, et al. Randomized selection design trial evaluating CD8+-enriched versus unselected tumor-infiltrating lymphocytes for adoptive cell therapy for patients with melanoma. *J Clin Oncol.* 2013; 31: 2152-9.
  158. Overwijk WW. Human CD4(+) T cells spontaneously detect somatic mutation in cancer cells. *Nat Med.* 2015; 21: 12-4.
  159. Linnemann C, van Buuren MM, Bies L, Verdegaal EM, Schotte R, Calis JJ, et al. High-throughput epitope discovery reveals frequent recognition of neo-antigens by CD4+ T cells in human melanoma. *Nat Med.* 2015; 21: 81-5.
  160. Kim HJ, Cantor H. CD4 T-cell subsets and tumor immunity: the helpful and the not-so-helpful. *Cancer Immunol Res.* 2014; 2: 91-8.
  161. Segal NH, Parsons DW, Peggs KS, Velculescu V, Kinzler KW, Vogelstein B, et al. Epitope landscape in breast and colorectal cancer. *Cancer Res.* 2008; 68: 889-92.
  162. Robbins PF, Lu YC, El-Gamil M, Li YF, Gross C, Gartner J, et al. Mining exomic sequencing data to identify mutated antigens recognized by adoptively transferred tumor-reactive T cells. *Nat Med.* 2013; 19: 747-52.
  163. van Rooij N, van Buuren MM, Philips D, Velds A, Toebes M, Heemskerk B, et al. Tumor exome analysis reveals neoantigen-specific T-cell reactivity in an ipilimumab-responsive melanoma. *J*

- Clin Oncol. 2013; 31: e439-42.
164. Castle JC, Kreiter S, Diekmann J, Löwer M, van de Roemer N, de Graaf J, et al. Exploiting the mutanome for tumor vaccination. *Cancer Res.* 2012; 72: 1081-91.
  165. Matsushita H, Vesely MD, Koboldt DC, Rickert CG, Uppaluri R, Magrini VJ, et al. Cancer exome analysis reveals a T-cell-dependent mechanism of cancer immunoediting. *Nature.* 2012; 482: 400-4.
  166. Duan F, Duitama J, Al Seesi S, Ayres CM, Corcelli SA, Pawashe AP, et al. Genomic and bioinformatic profiling of mutational neoepitopes reveals new rules to predict anticancer immunogenicity. *J Exp Med.* 2014; 211: 2231-48.
  167. Gubin MM, Zhang X, Schuster H, Caron E, Ward JP, Noguchi T, et al. Checkpoint blockade cancer immunotherapy targets tumour-specific mutant antigens. *Nature.* 2014; 515: 577-81.
  168. Lu YC, Yao X, Crystal JS, Li YF, El-Gamil M, Gross C, et al. Efficient identification of mutated cancer antigens recognized by T cells associated with durable tumor regressions. *Clin Cancer Res.* 2014; 20: 3401-10.
  169. Tran E, Turcotte S, Gros A, Robbins PF, Lu YC, Dudley ME, et al. Cancer immunotherapy based on mutation-specific CD4+ T cells in a patient with epithelial cancer. *Science.* 2014; 344: 641-5.
  170. Wick DA, Webb JR, Nielsen JS, Martin SD, Kroeger DR, Milne K, et al. Surveillance of the tumor mutanome by T cells during progression from primary to recurrent ovarian cancer. *Clin Cancer Res.* 2014; 20: 1125-34.
  171. Yadav M, Jhunjhunwala S, Phung QT, Lupardus P, Tanguay J, Bumbaca S, et al. Predicting immunogenic tumour mutations by combining mass spectrometry and exome sequencing. *Nature.* 2014; 515: 572-6.
  172. Rajasagi M, Shukla SA, Fritsch EF, Keskin DB, DeLuca D, Carmona E, et al. Systematic identification of personal tumor-specific neoantigens in chronic lymphocytic leukemia. *Blood.* 2014; 124: 453-62.
  173. Rizvi NA, Hellmann MD, Snyder A, Kvistborg P, Makarov V, Havel JJ, et al. Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. *Science.* 2015; 348: 124-8.
  174. Cools N, Ponsaerts P, Van Tendeloo VF, Berneman ZN. Balancing between immunity and tolerance: an interplay between dendritic cells, regulatory T cells, and effector T cells. *J Leukoc Biol.* 2007; 82: 1365-74.
  175. Zinkernagel RM, Hengartner H. Regulation of the immune response by antigen. *Science.* 2001; 293: 251-3.
  176. Reise Sousa C. Dendritic cells in a mature age. *Nat Rev Immunol.* 2006; 6: 476-83.
  177. Anguille S, Willemsen Y, Lion E, Smits EL, Berneman ZN. Dendritic cell vaccination in acute myeloid leukemia. *Cytotherapy.* 2012; 14: 647-56.
  178. Degli-Esposti MA, Smyth MJ. Close encounters of different kinds: dendritic cells and NK cells take centre stage. *Nat Rev Immunol.* 2005; 5: 112-24.
  179. Moretta L, Ferlazzo G, Bottino C, Vitale M, Pende D, Mingari MC, et al. Effector and regulatory events during natural killer-dendritic cell interactions. *Immunol Rev.* 2006; 214: 219-28.
  180. Palucka K, Banchereau J, Mellman I. Designing vaccines based on biology of human dendritic cell subsets. *Immunity.* 2010; 33: 464-78.
  181. Diamond MS, Kinder M, Matsushita H, Mashayekhi M, Dunn GP, Archambault JM, et al. Type I interferon is selectively required by dendritic cells for immune rejection of tumors. *J Exp Med.* 2011; 208: 1989-2003.
  182. Fuertes MB, Kacha AK, Kline J, Woo SR, Kranz DM, Murphy KM, et al. Host type I IFN signals are required for antitumor CD8+ T cell responses through CD8α+ dendritic cells. *J Exp Med.* 2011; 208: 2005-16.
  183. Palucka K, Banchereau J. Cancer immunotherapy via dendritic cells. *Nat Rev Cancer.* 2012; 12: 265-77.
  184. Jäger E, Jäger D, Knuth A. Clinical cancer vaccine trials. *Curr Opin Immunol.* 2002; 14: 178-82.
  185. Disis ML, Schiffman K, Guthrie K, Salazar LG, Knutson KL, Goodell V, et al. Effect of dose on immune response in patients vaccinated with an her-2/neu intracellular domain protein-based vaccine. *J Clin Oncol.* 2004; 22: 1916-25.
  186. Czerniecki BJ, Koski GK, Koldovsky U, Xu S, Cohen PA, Mick R, et al. Targeting HER-2/neu in early breast cancer development using dendritic cells with staged interleukin-12 burst secretion. *Cancer Res.* 2007; 67: 1842-52.
  187. Baxevasis CN, Sotiriadou NN, Gritzapis AD, Sotiropoulou PA, Perez SA, Cacoullos NT, et al. Immunogenic HER-2/neu peptides as tumor vaccines. *Cancer Immunol Immunother.* 2006; 55: 85-95.
  188. Redfern CH, Guthrie TH, Bessudo A, Densmore JJ, Holman PR, Janakiraman N, et al. Phase II trial of idiosyncrasy vaccination in previously treated patients with indolent non-Hodgkin's lymphoma resulting in durable clinical responses. *J Clin Oncol.* 2006; 24: 3107-12.
  189. Butts C, Murray N, Maksymiuk A, Goss G, Marshall E, Soulières D, et al. Randomized phase IIB trial of BLP25 liposome vaccine in stage IIIB and IV non-small-cell lung cancer. *J Clin Oncol.* 2005; 23: 6674-81.
  190. Fay JW, Palucka AK, Paczesny S, Dhodapkar M, Johnston DA, Burkeholder S, et al. Long-term outcomes in patients with metastatic melanoma vaccinated with melanoma peptide-pulsed CD34(+) progenitor-derived dendritic cells. *Cancer Immunol Immunother.* 2006; 55: 1209-18.
  191. Palucka AK, Ueno H, Connolly J, Kerneis-Norvell F, Blanck JP, Johnston DA, et al. Dendritic cells loaded with killed allogeneic melanoma cells can induce objective clinical responses and MART-1 specific CD8+ T-cell immunity. *J Immunother.* 2006; 29: 545-57.
  192. Bernhardt SL, Gjertsen MK, Trachsel S, Moller M, Eriksen JA, Meo M, et al. Telomerase peptide vaccination of patients with non-resectable pancreatic cancer: a dose escalating phase I/II study. *Br J Cancer.* 2006; 95: 1474-82.
  193. Gould P. Sipuleucel-T shows partial advantage in prostate cancer. *Lancet Oncol.* 2006; 7: 710.
  194. Finke LH, Wentworth K, Blumenstein B, Rudolph NS, Levitsky H, Hoos A. Lessons from randomized phase III studies with active cancer immunotherapies — outcomes from the 2006 meeting of the Cancer Vaccine Consortium (CVC). *Vaccine.* 2007; 25 (Suppl 2): B97-109.
  195. Korman AJ, Peggs KS, Allison JP. Checkpoint blockade in cancer immunotherapy. *Adv Immunol.* 2006; 90: 297-339.
  196. Ribas A, Hanson DC, Noe DA, Millham R, Guyot DJ, Bernstein SH, et al. Tremelimumab (CP-675,206), a cytotoxic T lymphocyte associated antigen 4 blocking monoclonal antibody in clinical development for patients with cancer. *Oncologist.* 2007; 12: 873-83.
  197. Small EJ, Tchekmedyian NS, Rini BI, Fong L, Lowy I, Allison JP. A pilot trial of CTLA-4 blockade with human anti-CTLA-4 in patients with hormone-refractory prostate cancer. *Clin Cancer Res.* 2007; 13: 1810-5.
  198. Mahnke K, Schönfeld K, Fondel S, Ring S, Karakhanova S, Wiedemeyer K, et al. Depletion of CD4+CD25+ human regulatory T cells in vivo: kinetics of Treg depletion and alterations in immune functions in vivo and in vitro. *Int J Cancer.* 2007; 120: 2723-33.