Indian Journal of Biochemistry & Biophysics Vol. 56, February 2019, pp. 7-19

Minireview

Free radicals and antioxidants in normal versus cancerous cells — An overview

Anju Shrivastava¹, Lalit Mohan Aggarwal¹, Surendra Pratap Mishra²*, Hari Dev Khanna³, Uday Pratap Shahi¹ & Satyajit Pradhan¹

¹Department of Radiotherapy and Radiation Medicine; ²Department of Biochemistry; ³Department of Biophysics, Institute of Medical Sciences, Banaras Hindu University, Varanasi -221 005, Uttar Pradesh, India

Received 05 April 2018; revised 24 November 2018

Oxygen is vital for aerobic processes of metabolism and respiration- It has been also implicated in many diseases and degenerative conditions. Free radicals formed from reactive oxygen and nitrogen species act as key players in the initiation and progression of tumor cells and enhance their metastatic potential. The imbalance in the formation and use of free radicals in the tissue creates oxidative stress. Inadequacy in normal cells antioxidant defense system or excessive free radical formation or even both can cause the cell to experience the oxidative stress. This review outlines the involvement of free radicals in different aspects of cancer, from prevention to initiation, progression, treatment and to reduce morbidity and mortality.

Keywords: Cancer therapy, Reactive oxygen species (ROS), Superoxides, Xenobiotic

Introduction

Human life on earth is possible due to the presence of oxygen which is essential for cellular activities. Out of the total inhaled oxygen around 5% is converted to Reactive Oxygen Species (ROS) by the univalent reduction of oxygen. This poisonous property of O_2 is disclosed by the Gershman's free radical theory of oxygen toxicity in 1954. According to this theory the oxygen toxicity is due to incomplete reduction of oxygen. It reveals the dual role of ROS and RNS (Reactive Nitrogen Species). Further, it clarifies that ROS and RNS have both beneficial and harmful effects in accordance with their concentration in the body^{1,2}.

At the low and moderate concentrations, ROS shows beneficial effect by participating in the physiological mechanisms, Such as cellular defense against infectious agents, cellular signaling, induction of mutagenic response, electron transport chain, *etc*. On the other hand, the free radicals at high concentration, show deleterious effect due to imbalance between the production of free radicals and cellular antioxidants which creates oxidative stress³. In the biological systems, oxidative stress is due to the excess production of ROS/RNS as well as deficiency

of enzymatic and non-enzymatic antioxidant. The redox regulation mechanism of the living organisms creates the equilibrium between the beneficial and deleterious effect of the free radicals which is essential for the survival. This redox regulation mechanism forms a redox homeostasis, protects living organisms from various oxidative stress by managing the redox status in vivo⁴. This review deals with (i) The mechanism of formation of free radicals cells under aerobic conditions; respiring in (ii) The antioxidant systems involved in the scavenging process; (iii) Free radicals mediated damage to the cellular macromolecules (Lipids, Proteins, and DNA); (iv) The involvement of free radicals in different aspects of Cancer, from prevention to initiation, progression, treatment, and recovery; and (v) Controversial views of antioxidants in cancer therapy.

Free radicals

Free radicals are highly unstable, and reactive with short half-lives. Their hyper reactivity is due to the presence of one or more unpaired electrons in its outermost shell of their atoms. These unpaired electrons try to attain balance by binding with electrons of neighbouring atoms, giving rise to chain reactions⁵. Free radicals are produced from oxygen during various cellular metabolisms and for a variety

of tasks such as signaling, metabolizing xenobiotic, initiating apoptosis, stimulation of antioxidants, repair process. Hence, its production in an animal cell is inevitable⁶. Increase in free radical level leads to oxidative stress. Increased number of free radicals, in turn steals number of electrons from neighbouring, DNA, enzymes and cell membranes which affect their structure and composition, leads to cell damage. Free radicals are not only the byproducts of cellular processes but also introduced into our bodies exogenously by cigarette smoking, radiation, drinking alcohol, air and water pollution, certain gases and even sunlight. The major forms of free radicals present in our body are hydroxyl radicals and superoxides.

Hydroxyl free radicals are necessary for hydroxylation of lysine and proline amino acids to hydroxylysine and hydroxyproline, respectively, it is also necessary for collagen biosynthesis⁷. Defense bacteria mechanism against and virus. via macrophages and leucocytes, also contribute to the formation of free radicals. For normal metabolism of cell free radicals are needed. However, the presence of free radicals poses a risk of damage for large molecules such as nucleic acids, proteins, polysaccharides and lipids⁷⁻¹⁰.

The mitochondrion is the major site of the superoxide free radical (O_2^{-}) by the action of NADPH (Nicotinamide Adenine Dinucleotide Phosphate) oxidase enzyme¹¹. In dismutation, the reactive hydrogen peroxide (H_2O_2) is formed by the action of superoxide dismutase (SOD). It happens when the superoxide radical reacts with itself and forms oxygen and hydrogen peroxide⁶. Continuous reduction process transforms the hydrogen peroxide via the Fenton reaction into the hydroxyl free radical (•OH) which is highly reactive, and finally, water is formed, mediated by the action of catalase (CAT) or glutathione peroxidase (GPx)^{6,12}. In some cases, oxygen molecule binds to a proton, another free radical is formed known as the hydro peroxide radical $(HO_2 - \bullet)^{13}$. Figure 1 depicts different reactive forms of oxygen^{6,14,15}

The free radicals in Reactive Nitrogen Species (RNS) are nitric oxide (NO•), peroxynitrite (ONOO–), nitrogendioxide (NO₂•) and nitrite (NO₂–). NO• is synthesized from a guanidine group of L-arginine by an enzyme of the nitric oxide synthetase (NOS) family. The formation of ONOO– takes place by the reaction of NO• with a molecule of O₂–•, which forms nitrogen dioxide (NO₂•) as an

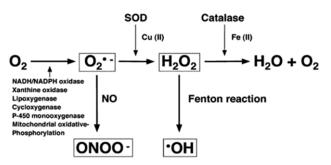


Fig. 1 — The different reactive forms of oxygen and antioxidants

intermediary. This intermediary reacts with NO• to finally generate anhydride nitrous $(N_2O_3)^{13,15,16}$.

The isoforms of NOS are neuronal (nNOS or NOSI), inducible (iNOS or NOSII), endothelial (eNOS or NOSIII) and mitochondrial (mtNOS). All these forms dependent on NADPH and calmodulin¹⁷. eNOS has a remarkable role in tumor development, as it modulates various tumor-related processes, such as apoptosis, angiogenesis, cell cycle, invasion, and metastasis¹⁸. Different biological actions of NO• are mediated by guanylcyclase (sGC) and cyclic guanosine monophosphate (cGMP). NO• spreads to nearby cells and readily enters the cytosol, where it activates sGC by binding to the "hem" component of iron on the porphyrin ring. As the concentration increases, its cytotoxic effects get activated which leads to inhibition of mitochondrial enzymes, including succinate, ubiquinone oxidoreductase, and aconitase, which are important in cell metabolism^{13,19}.

Antioxidants

Antioxidants are molecules which protect the body from damage caused by excess free radicals. Antioxidants neutralize these free radicals by sharing their extra electrons- so that they don't steal extra electrons from other vital organs of the cell. Free radical exposure of any organism leads to the development of defense mechanism in their $body^{20}$. As per the nature of an antioxidant molecule, it is mainly divided into two categories, Enzymatic and Non enzymatic antioxidants. Enzymatic antioxidant defense includes superoxide dismutase (SOD), glutathione peroxidase (GPx), catalase (CAT). Non-enzymatic antioxidants are mainly vitamin C (ascorbic acid), vitamin E (α-tocopherol), vitamin A $(\beta$ carotene), and glutathione (GSH), flavonoids. There is a balance between both free radicals and the level of the antioxidants in the body under normal conditions. For the survival of the organism and its health, this balance is essential.

Glutathione disulphide (GSSG) is the oxidized form of glutathione. Glutathione is the major soluble antioxidant in cell compartments²¹. It is highly abundant in the cytosol (1-11 mM), nuclei (3-15 mM), and mitochondria (5-11 mM). GSH is synthesized in the cytosol by the sequential action of glutamate–cysteine ligase and glutathione synthetase. Its mitochondrial presence requires inner membrane transport. The dicarboxylate carrier protein and the 2- oxoglutarate carrier protein are the two mitochondrial electro neutral antiport carrier proteins shows the capacity to transport GSH.

Studies points that, externally added GSH is readily taken up against the concentration gradient by mitochondria, despite the ~8 mM GSH presents in the mitochondrial matrix²². The redox state of critical protein sulfhydryl's which are necessary for DNA repair and expression is maintained by GSH in the nucleus. Studies concluded that oxidized glutathione is accumulated inside the cells and the ratio of GSH/GSSG is a good measure of oxidative stress of an organism^{23,24}. A high concentration of GSSG may damage many enzymes oxidatively.

The different protective roles of glutathione against oxidative stress are:

- (i) It acts as a cofactor for many detoxifying enzymes against oxidative stress, *e.g.* Glutathione transferase, Glutathione peroxidase and others.
- (ii) Through plasma membrane, GSH participates in amino acid transport.
- (iii) It directly scavenges hydroxyl radicals and singlet oxygen while catalytic actions of glutathione peroxidase detoxify hydrogen peroxide and lipid peroxidases.
- (iv) GSH has the ability to regenerate antioxidants and return to their active forms, like reduction of tocopherol radical of vitamin E, reduction of semi dehydroascorbate to ascorbate²⁵.

The ability of glutathione to regenerate the antioxidant is associated with the redox state of glutathione disulphide-glutathione couple (GSSG/2GSH). Multiple roles of enzymatic and non-enzymatic antioxidants in rescue against oxidative stress are described in different studies²⁶⁻⁴².

Free radicals mediated damage to the cellular macromolecules

The essential cell components like lipids, proteins and nucleic acids are under the threat of high concentrations of free radicals². These free radicals can alter intrinsic membrane properties like fluidity, ion transport, loss of enzyme activity, protein cross-linking, and inhibition of protein synthesis. All components of the DNA molecule are under the threat of hydroxyl radicals, damaging both the purine and pyrimidine bases and also the deoxy-ribose backbone, ultimately resulting in cell death⁵.

Lipid peroxidation

The oxidative modifications of lipids are catalase by reactive oxygen species⁴³. The hydrogen becomes more prone to abstraction in the presence of a double bond adjacent to a methylene group makes the C-H bond of polysaturated fatty acid (PUFA) weaker. Lipid peroxidation is initiated by OH•, alkoxy radicals (RO•), and peroxy radicals (ROO•), not by the O_2 - and $H_2O_2^{44}$. Peroxy radicals act as both reaction initiators as well as the products of lipid peroxidation, leads to a self-perpetuating process. Oxidation of the substrate is possible by the transfer of electrons when lipid peroxy radicals react with other lipids, proteins, and nucleic acids. Cell membranes, undergoes changes in membrane fluidity, permeability, and cellular metabolic functions during oxidative attackdue to the presence of large amount of PUFA. The crucial aldehyde products of lipid peroxidation aremalondialdehyde (MDA) and 4-hydroxy-2-nonenal (HNE). In bacterial and mammalian cells MDA is mutagenic and in rats it is carcinogenic. The major toxic product of lipid peroxidation is Hydroxynonenal, it is weakly mutagenic⁴⁵⁻⁴⁹.

DNA damages

Reactive Oxygen Species may induce oxidative damage to DNA, both nuclear and mitochondrial. The major types of damages observed are base modifications, deoxyribose oxidation, strand breakage, and DNA-protein cross-links. Out of all the ROS, hydroxyl radicals produce many products from the DNA base which mainly include C-8 hydroxylation form 8-oxo-7, 8 dehydro-2¢of guanine to deoxyguanosine, a ring-opened product; 2, 6-diamino-4hydroxy-5-formamimodipyrimidine, 8-OH-adenine, 2-OH-adenine, thymine glycol, cytosine glycol, etc^{50} . ROS also cause different mutagenic alterations in DNA. The most widely studied DNA lesion is the formation of 8-OH-G. Permanent modification of genetic material resulting from these "oxidative damage" incidents represents the first step involved in mutagenesis, carcinogenesis and ageing. Mutations originating from selective modification of G:C sites indicates an oxidative attack on DNA by ROS. The action of 8-oxodeoxy-guanosine as a promutagen in altering the binding of methylase to the oligomer so as to inhibit methylation of adjacent cytosine has been reported in cases of cancer development^{51,52}.

ROS has shown activate mutations in human C-Ha-ras-1 proto-oncogene and induce mutation in the p53 tumor-suppressor gene⁵³. It also interferes with normal cell signaling, resulting in alteration of the gene expression and development of cancer by redox regulation of transcriptional factors/activator and/or by oxidatively modulating the protein kinase cascades. Early response or stress-response genes like *c-fos*, *c-jun*, *jun-B*, *jun-D*, *cmyc*, erg-1, and hemeoxygenase-1 are also produced by various ROS. A vital role is played by the early responseproto-oncogenes in signal transduction, leading to cell proliferation and transformation⁵⁴. Abnormal components of the electron transport chain are formed by the oxidative damage of mitochondrial DNA which involves base modifications and strand break. This leads to the formation of more ROS through increased leakage of electrons and further cell damage, which promotes cancer and ageing⁵⁵.

Oxidative damage of proteins

In the mitochondrial electron transport chain, free radicals are produced which can initiate protein degradation. Oxidative protein damage may be brought about by metabolic processes which degrade a damaged protein to promote synthesis of a new protein. On the basis of further studies on aging processes, it has been found out that, catalytically inactive or less active, more thermo labile forms of enzyme accumulate in cells during aging and show a noticeable increase in the level of protein carbonyl content which is an index of metal-catalyzed oxidation of proteins⁵⁶⁻⁵⁸. Studies in which amino acids, simple peptides, and proteins were exposed to ionizing radiations under conditions where hydroxyl radicals or a mixture of hydroxyl/superoxide radicals are formed describes the mechanisms involved in the oxidation of proteins by ROS⁵⁹. By the action of ROS/RNS, the side chains of all amino acid residues particularly cysteine and methionine residues of proteins are susceptible to oxidation⁵⁹. Oxidation of cysteine residues may lead to the reversible formation of mixed disulphides between protein thiolgroups (-SH) and low molecular weight thiols, in particular GSH (S-glutathionylation).

The amount of carbonyl groups, generated by many different mechanisms is a good measure of ROS-mediated protein oxidation. Numerous highly sensitive methods have been developed for the assay of protein carbonyl groups^{60,61}.

Levels of glyceraldehyde-3-P-dehydrogenase, aspartate aminotransferase, and phosphoglycerate kinase decline with age together with an increase in protein carbonyl content in human erythrocytes⁶². The carbonyl content of protein in rat hepatocytes also elevated with age along with a decline in the activities of glutamine synthetase and glucose-6-P-dehydrogenase, without any loss in the total enzyme protein⁶³. An age-related oxidative modification of human ceruloplasmin, a copper-containing protein in human plasma has also been reported⁶⁴. The mechanism of oxidative damage of proteins by ROS has been studied in vitro by different ways, generating these reactive species in solution which is non-specific (global) damage, or 'site-specifically' within the protein (localized) damage⁶⁵. Non-specific damage can be formed by generating activated oxygen species in situ, using either a radiation source (Co^{60}) - or using pulse radiolysis techniques; which lead to aggregation and fragmentation of the protein and modification of almost all the amino acid residues⁶⁶. On the other hand, localized or 'site-specific' damage, (which was extensively studied using glutamine synthetase as the model enzyme⁶⁵⁻⁶⁸) can occur when the ROS, such as •OH, are formed at putative metal-binding sites in proteins. When these sites are occupied by iron or copper, they can (in the presence of suitable reductants, O_2 – or ascorbate) react with H_2O_2 to generate highly reactive •OH which reacts preferably with specific amino acids present in the vicinity of the metal-binding site^{58,69}, thereby inducing specific damage (sitespecific) that shows no gross structural modification. The concept of 'site-specificity' has been studied in detail by Halliwell, Gutteridge, and Stadtman^{65,68}. It means that (i) catalytic metal ions, such as iron or copper would be bound to the target molecule (protein, DNA, or cell membrane), and the •OH produced by O₂- (or ascorbate) and H₂O₂ produced at the ironor copperbinding site would then react preferentially with the target molecule; (ii) the damaging effect of •OH is observed at a specific site where catalytic ions are bound; and (iii) the defensive action of the freeradical scavengers to remove •OH from the specific site decreases dramatically, since they are unable to access the microenvironment. Although tryptophan, phenylalanine, and tyrosine residues of proteins are not the major sites of oxidation (as in the case of global damage) by the site-specific oxidative system; arginine, lysine, histidine, cysteine and proline are particularly sensitive to this oxidation, resulting in the formation of carbonyl derivatives^{67,70}.

Involvement of free radicals in different aspects of cancer

The studies reveal that free radicals induced oxidative stress is closely related to all aspects of cancer, from prevention, initiation, progression, treatment, and recovery. The human body is constantly under oxidative stress arising from both endogenous and exogenous origin. When such oxidative stress exceeds the capacity of the body system to maintain oxidation-reduction homeostasis, gene mutation may result, affecting the intracellular signal transduction and transcription factors directly or *via* antioxidants, leading to carcinogenesis.

In cancer prevention

The cellular mechanism of defense against the consequences of increased concentrations of reactive species (mainly ROS) is by antioxidants⁷¹. Proofs for prevention of carcinogenesis and increased life span by the antioxidant supplementation is documented by Kovacic and Jacintho in 2001⁷². For a correct cellular concentration response, the of antioxidant supplementation is very important, as a low concentration may have no effect while a high concentration causes a negative effect and act as pro-oxidant or the free radical⁷³. Oxidative stress leads to the over production of free radicals in the body which increases the chances for tumor initiation. Maintaining a proper equilibrium between free radicals and antioxidants prevent the body from all types of carcinomas. By donating their extra electrons antioxidant enzymes like SOD, Catalase and Glutathione peroxidase prevent oxidation and reduce the rate of chain initiation. They can also prevent oxidation by stabilizing transition metal radicals such as copper and iron^{74,75}. The intake of dietary antioxidants like vitamins E, C, and β -carotene is useful in preventing carcinogenesis⁷⁶. Different studies revealed that antioxidants can be used in the inhibition of inflammation in relation to the risk of carcinogenesis⁷⁷. The risk of formation of pro-oxidant is always associated with excess supplementation of antioxidants, for example, vitamin C^{78} . The type of antioxidant and its amount to be ingested for obtaining a preventive effect in cancer remains under investigation.

In cancer initiation

Due to the presence of free radicals, cancer cells compared with normal cells forms a cellular redox imbalance, this redox imbalance thus related to oncogenic stimulation. The first step involved in mutagenesis, carcinogenesis, and ageing is "Oxidative damage" from free radicals resulting from the permanent modification of genetic material. In various tumors DNA mutation is a crucial step and elevated levels of oxidative DNA lesions have been noticed, which strongly implicating such damage in the etiology of cancer. More than 100 oxidized DNA products have been identified. Free radical based DNA damage includes single or double-stranded DNA breaks. purine, pyrimidine, or deoxyribose modifications, and DNA crosslinks. DNA damage may end up with arrest or induction of transcription, transduction pathways, replication errors and genomic instability, all of which are associated with carcinogenesis^{2,79}, 8-OH-Gformation is the most extensively studied DNA lesion. It is a potential biomarker of carcinogenesis as it is relatively easily formed and is mutagenic. DNA damage, mutations, and altered gene expressions, all are having a crucial role in the process of carcinogenesis. The participation of oxidants seems to be the common factor to all these events^{2,80,81}. It is clearly documented that inflammation and carcinogenesis are inter-related, numerous reports of cancer originating at sites of previous chronic inflammation are there in the literature⁸². Studies have been published on changes in morphology and in gene expression of mouse mammary epithelial cells after prolonged exposure to H₂O₂, which simulates chronic inflammation. In such oxidation conditions, a phenotypic cell conversion with similarities to malignant transformation was observed, including a fibroblastic morphology with intercellular spaces, implying a decrease in intercellular connections⁸³. In the mechanisms of carcinogenesis and ageing along with ROS, various redox metals, due to their ability to generate free radicals, or non-redox metals, due to their ability to bind to critical thiols have been participated⁸⁴⁻⁹⁰. Iron-mediated oxidative stress is considered to be a principal cause of human colorectal cancer⁸⁰. The second most important cause of lung cancer is occupational exposure to asbestos-containing about 30% (weight) of iron is associated with an increased risk of asbestosis⁸⁸.

Chances for oxidative stress and cancer increased in occupational exposure to cadmium⁸⁷. Directly cadmium is not capable of producing free radicals, but through indirect methods, it can cause free radical-induced damage the gene expression. Cadmium can cause activation of cellular protein kinases (protein kinase C), which results in enhanced phosphorylation of transcription factors and consequently lead to the transcriptional activation of target gene expression⁹⁰. It has been suggested that cadmium can also be implicated in the pathogenesis of human pancreatic cancer and renal carcinomas.

One of the potential lung carcinogen is Hexavalent chromium; Cr (VI)-induced cytotoxicity is associated with mitochondrial/lysosomal toxicity substantiated by the enhanced formation of free radicals⁸⁵.

Arsenic compounds are well established human carcinogens, which bind with SH groups and inhibiting various enzymes, including glutathione reductase⁸⁶. Researchers agree with the hypothesis that arsenic may act as a co-carcinogennot by causing cancer directly, but by allowing other factors, such as cigarette smoke or UV radiation, to cause DNA mutations more effectively ^{89,90}. The effect of arsenic on p53 is not completely understood. The experimental results show both p53 dependent and p53 independent induction of apoptosis and also both, increased and decreased expression of the protein⁹⁰.

The oxidative DNA damage rate increased 35-50% by tobacco smoke, as estimated from the urinary excretion of 8-OH-G, or by 20-50%, estimated from the level of 8-OH-G in leukocytes⁹¹. The oxygen consumption, which is a main endogenous source of ROS, showed a close correlation with the 8-OH-G excretion rate, even though moderate exercise seems to have no current effect. Studies failed to show an influence on the oxidative DNA modification, by diet composition, including energy restriction and antioxidant supplements⁹². Along with ROS, reactive nitrogen species (RNS), such as peroxynitrites and nitrogen oxides have also been involved in DNA damage⁹³. During reaction with guanine, peroxynitrite has been shown to form 8-nitroguanine. This adduct has the potential to induce G:C \rightarrow T:A trans versions due to its structure. While the stability of this lesion in DNA is low, in RNA, however, this nitrogen adduct is stable. The potential connection between 8-nitro guanine and the process of carcinogenesis is under research.

In cancer progression

In the cancer cells ROS act as messengers in cellular signaling transduction pathways, promote cellular growth and proliferation, and finally contribute to cancer development^{94,95}. ROS contribute to the uncontrolled cell proliferation and compromise the cellcycle regulatory function.Direct ROS interaction with specific receptors and modulation of the redox states of signaling molecules are the mechanisms responsible for stimulation of cell proliferation. Any alteration in the redox status of a signaling molecule may lead to stimulation of cell growth and cell proliferation. Oxidative modifications of redox-sensitive transcription factors may also be involved in ROS-mediated modulation of cell growth and cell survival^{94,96,97}. Most important characteristics of tumor cells are their increased ability to survive compared with the normal cells. ROS are reported to be tumorigenic by virtue of their ability to increase cell proliferation, survival, and cellular migration. ROS can develop DNA damage, leading to genetic lesions that initiate the tumorogenicity and subsequent tumor progression. Simultaneously, ROS is also capable for cellular senescence, cell death and can therefore, function as anti-tumorigenic agents. ROS promote tumor cell survival or act as anti-tumorigenic agents completely depends on the cell and tissues, the location of ROS production and the concentration of individual ROS.

The up-regulation of multiple intracellular signaling pathways, including cascades involved in survival, proliferation and cell cycleprogression are required for uncontrolled tumor cell proliferation. The noticeable effects of oxidants on signaling pathways have been observed in the MAPK/AP-1 and NF-κB pathways⁹⁸. Cell division needs tremendous energy requirements therefore, the production of metabolites from the energy generating reactions must be buffered to prevent oxidative damage and ultimately cell death. Hence the induction of redox sensitive pathways during tumor cell proliferation is essential⁹⁹.

Oxygen radicals may overcome tumor invasion and metastasis by increasing the rates of cell migration. Before converting into invasive carcinoma, epithelial cells undergo profound alterations in morphology and adhesive mode, resulting in a loss of normal epithelial polarization and differentiation and a switch to a more motile, invasive phenotype. For example, treatment of mammalian carcinoma cells with hydrogen peroxide before intravenous injection into mice enhances lung metastasis formation, indicating that an important function for ROS is the seeding of metastatic tumor cells¹⁰⁰. This might be due to a lesser attachment of tumor cells to the basal lamina or otherwise due to more activity or expression of proteins that regulate cellular motility. Oxidative stress regulates the expression of intercellular adhesion protein-1 (ICAM-1), a cell surface protein in endothelial and epithelial cells, most likely because of the activation of NF- κ B. ICAM-1 together with IL-8 regulates the trans endothelial migration of neutrophils and has a potential function in tumor metastasis¹⁰¹.

An angiogenic response by the host blood vessels in solid tumors forms a new vascular network for the supply of nutrients and oxygen¹⁰². This neovascular response is partly responsible for tumor growth and metastatic spread^{103,104}. Angiogenesis in tumors is regulated by the so-called "angiogenic switch" which allows the transition from low invasive and poorly vascularized tumors to highly invasive and angiogenic tumors. Tumor cells express a set of molecules that initiate tumor vascularization to increase further in size.

Numerous cellular stress factors, including hypoxia, nutrient deprivation, and ROS are important stimuli of angiogenicsignaling¹⁰⁵. Overexpression of RAS (rat sarcoma viral oncogene) has also been linked to vascularization of tumors¹⁰⁶. Transformation by RAS stabilizes HIF-1 α and up-regulates the transcription of vascular endothelial growth factor-A (VEGF-A). Chemical antioxidants inhibit the mitogenic activity of RAS, indicating that ROS participate directly in malignant transformation. ROS stabilize HIF-1 α protein and induce production of angiogenic factors by tumor finally.

In cancer treatment

A state of oxidative stress is created in the body when cancer is treated using anticancer drugs and radiation. Active oxygen triggers apoptosis via p53 and cytochrome release from mitochondria. Ionizing radiations-generates charged particles or electrons that carry the kinetic energy provided by photons (X rays, γ rays),falls directly on DNA producing breaks in phosphodiester bonds. This leads to around 30% of DNA damage^{107,108}. The remaining damage is created by the action of free radicals. In a process known as water "radiolysis" the •OH, which has high biological relevance, is generated by the interaction of ionizing radiation (e.g., X- or gamma rays) with the water molecule. The deposition of energy from radiation also leads to the formation of hydrogen atoms, hydrated electrons, and other molecular products. It includes molecular hydrogen, hydrogen peroxide, and peroxynitrite, compounds that generate DNA-damage like 8-hydroxyguanine (8-OH-Gua), 8-OH-dG. single 8-oxoguanine and consequently and double-strand DNA breaks¹⁰⁹. Free radicals are important factors in carcinogenesis^{110,111}. The •OH has an extraordinarily short life in the human body, due to collisions with the different and abundant molecules in the biological environment. Scientists have come to the conclusion that radiotherapy generates the •OH, which is the free radical most associated with cell death during the treatment of cancer. The NO• appears to act as radio sensitizer under conditions of hypoxia, mimickingthe effects of oxygen relating on radiation-induced DNA damage¹¹²⁻¹¹⁴. One of the major late complication after radiotherapy in breast cancer is the fibrosis that results from radiation-induced inflammatory responses¹¹⁵.

triggering apoptosis within For the cell. chemotherapy mainly depending on ROS. Chemotherapy agents are divided into different categories like alkylating agents (e.g., cyclophosphamide, ifosfamide), anthracycline antibiotics which affect nucleic acids (e.g., doxorubicin, bleomycin), platinum compounds (e.g., cisplatin), mitotic inhibitors (e.g., vincristine), antimetabolites (e.g., 5-fluorouracil), camptothecin derivatives (e.g., topotecan), biological response modifiers (e.g., interferon) and hormone therapies (e.g., tamoxifen). Among chemotherapeutic agentsthe level of ROS generation is different¹¹⁶. Agents that produce high levels of ROS include anthracyclines, platinum coordination complexes, alkylating agents, epipodophyllotoxins and camptothecins¹¹⁷. Nucleoside and nucleotide analogs, antifolates, taxanes and vinca alkaloids produce only low levels of ROS. Formation of superoxide radicals during apoptosis induced by chemotherapeutic agents, involving the release of cytochrome c from mitochondria, NADH dehydrogenase and reduced coenzyme Q10 divert electrons from the electron transport system (ETS) to oxygen¹¹⁷. Anthracyclines develops the highest levels of ROS. They divert electrons from the ETS of cardiac mitochondria, resulting in the formation of superoxide radicals in addition to generating ROS at other cellular sites. Doxorubicin (Adriamycin), the most studied

anthracycline, is one of the most effective chemotherapeutic agent used in several cancers¹¹⁷. The toxic effect of Doxorubicin is compared to its ability to react with cancer cell DNA. It can induce DNA damage, mainly through inhibition of DNA topoisomerase II enzyme after induction of double-strand DNA breaks resulting in covalent attachment between active site residues of topoisomerase II and 5' end of DNA. Doxorubicin also intercalates into DNA and alters helical torsion which also causes DNA damage. Its use is seriously limited by its acute and chronic toxic effects in the heart including cardiomyopathy and congestive heart failure¹¹⁷⁻¹¹⁹. Free radical generation and subsequent oxidative stress mediate its toxic effects. Studies antioxidant supplementation showing that in combination with Doxorubicin protects against oxidative injuries without diminishing its clinical efficacy¹¹⁹.

In recovery

Studies in head and neck and cervix carcinomas concluded that the pattern of a decreased level of antioxidants compared to the controls before radiotherapy, increasing oxidative stress during radiotherapy and again increased antioxidant level after radiotherapy is positively associated with survival in the patients^{120,121}.

The initial decrease in the antioxidant level of blood in carcinoma patients, compared to healthy control is due to the increased oxidative stress due to the tumor formation in the body. During treatment (radiotherapy and chemotherapy), administration of free radicals increase oxidative stress, the decline in the level of antioxidants may be due to both decreased dietary intake and increased utilization of antioxidants by free radicals produced from ionizing radiations. Studies reveal that increased oxidative stress is beneficial during therapy, it promotes the tumor cells destruction. In post radiotherapy, high level of antioxidants in blood was positively associated with the survival of the cancer patient. It directly indicates the recovery. Supplementation of antioxidants after treatment of malignancy may improve the outcome in patients.

According to them, antioxidant supplementation decreases the oxidative stress level especially in regard to protein damage¹²². It maintains the hemoglobin level of the body, and improve the efficacy of radiotherapy by supplying enough oxygen to the tumor. Quality of life of this group is also improved. But the risk lies in

the recurrence of the tumor after some period of time. Further long-term studies are required for authenticity.

Controversial views of antioxidant use in cancer therapy

Free radicals are used for the treatment of cancer in conventional therapies like chemotherapy and radiotherapy. The supplementation of antioxidants during the treatment always remains a matter of controversy. Since ROS plays a role in drug induced apoptosis, one might suspect that antioxidants may inhibit ROS and prevent apoptosis of cancer cells. There is an intense argument on the concurrent use of antioxidants with the conventional cancer treatments. Numerous patients have turned to such complementary treatments with antioxidants. Many oncologists have moved against antioxidants and warned their patients not to use them during conventional cancer therapy^{123,124}. The base of this argument was on the fact that radiation therapy and some chemotherapy drugs generate ROS and antioxidants may prevent cancer cells to be killed by ROS. As a result, the parallel use of supplemental antioxidants should be avoided during conventional cancer treatment¹²⁵. Not only in the scientific circles, but also in media as well this argument is still continuing¹²⁶. Literature has been reviewed, in order to find the result if there is any interference between the concurrent use of antioxidants and conventional cancer therapy¹²⁷. Except for three specific interferences (tangeretin with tamoxifen, NAC with doxorubicin and β -carotene with 5-fluorouracil), considerable data exists demonstrating increased effectiveness as well as decreased side effects of chemotherapeutic agents when given with antioxidants¹²². There are situations in which antioxidant supplementation would be undesirable for cancer patients, but the issue is more complicated than indicated. Even though many chemotherapy drugs induce the formation of free radicals, their anticancer effects not only depend on free radicals. Antioxidant may enhance the effects of chemotherapy, while preventing free radical induced side effects. Considerable existing data show increased effectiveness of many chemotherapeutic agents, as well as a decrease in toxic adverse effects, when given concurrently with antioxidants^{116,123,128-132}.

Oxidative stress interferes with cellular processes that are necessary for antineoplastic agents to exert their optimal cytotoxicity on cancer cells. Modest levels of oxidative stress have been shown to reduce the cytotoxicity of anticancer drugs^{117,127}. Thus, it was claimed that the formation of ROS that occurs when anticancer drugs are administered may diminish the effectiveness of the treatment^{117,127,133}. In addition, since some side effects caused by antineoplastic agents appear to be prevented by certain antioxidants, supplements administering these during chemotherapy may diminish the development of side effects as well as improve the response to therapy¹¹⁷. Although limited clinical studies on the effect of antioxidants in cancer treatment are available. However, experimental studies showed that in cancer antioxidant vitamins selectively cells, induce apoptosis, prevent angiogenesis and metastatic spread, suggesting a potential role for antioxidants as adjuvants in cancer therapy¹³⁴.

Conclusion

Abundant knowledge of diverse mechanisms of oxidative stress involved in carcinogenesis and cancer therapies are available, but the current understanding is still not sufficient to cure cancer. The state of oxidative stress in carcinogenesis and tumor-bearing conditions is a complicated mechanism in which various complex interactions are involved. The deleterious effects of oxidative stress include injury to cells. induces gene mutation, involved in carcinogenesis by influencing intracellular signal transduction and transcription factors directly or indirectly via antioxidants. Research has been proven that free radicals can induce proliferation of cancer cells and protect them by increasing their adaptive response, thus leading to cancer progression and even metastatic diseases. Oxidative stress, a marker of tissue damage is found to increase with the progression of cancer. Oxidative stress can be measured by the levels of Lipid peroxidation, protein products and various antioxidants in the serum of cancer patients. Alternatively, free radicals are also recognized as the main mechanism of most of the conventional cancer therapies, leading cancer cells to hence to cancer regression. apoptosis and Antioxidants have also been shown to exhibit dichotomous effects when used as a supplement to cancer therapy. More research is needed for better understanding of the mechanisms and specific apoptotic pathways involved in ROS-induced cell death and to determine the most rational and effective combination of redox active agents. Further, it may reveal the missing mechanism of oxidative homeostasis in normal vs. malignant cells, and help

understanding the role of antioxidants along with conventional therapies in an integrative, personalized medical fight against cancer.

References

- 1 Gerschman R, Gilbert DL, Nye SW, Dwyer P & Fenn WO, Oxygen poisoning and x-irradiation: A mechanism in common. *Science*, 119 (1954) 623.
- 2 Valko M, Rhodes CJ, Moncol J, Izakovic M & Mazur M, Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chem Biol Interact*, 160 (2006) 1.
- 3 Sies H, *Oxidative stress: introductory remarks*. (Academic Press London) 1985, 1.
- 4 Handy DE & Loscalzo J, Redox regulation of mitochondrial function. *Antioxid Redox Signal*, 16 (2012) 1323.
- 5 Halliwell B & Gutteridge JMC, *Free radicals in biology and medicine*. (Oxford University Press) 1999, 3.
- 6 Sena LA & Chandel NS, Physiological roles of mitochondrial reactive oxygen species. *Mol Cell*, 48 (2012) 158.
- 7 Malhotra JD & Kaufman RJ, Endoplasmic reticulum stress and oxidative stress: a vicious cycle or a double-edged sword? *Antioxid Redox Signal*, 9 (2007) 2277.
- 8 Janssen-Heininger YM, Mossman BT, Heintz NH, Forman HJ, Kalyanaraman B, Finkel T, Stamler JS, Rhee SG & van der Vliet A, Redox-based regulation of signal transduction: principles, pitfalls, and promises. *Free Radic Biol Med*, 45 (2008) 1.
- 9 Ferrari CKB, Franca EL & Honorio-Franca AC, Nitric oxide, health and disease. *J Appl Biomed*, 7 (2009) 163.
- 10 Ziech D, Franco R, Georgakilas AG, Georgakila S, Malamou-Mitsi V, Schoneveld O, Pappa A & Panayiotidis MI, The role of reactive oxygen species and oxidative stress in environmental carcinogenesis and biomarker development. *Chem Biol Interact*, 188 (2010) 334.
- 11 Naik E & Dixit VM, Mitochondrial reactive oxygen species drive proinflammatory cytokine production. *J Exp Med*, 208 (2011) 417.
- 12 Jomova K & Valko M, Importance of iron chelation in free radical-induced oxidative stress and human disease. *Curr Pharm Des*, 17 (2011) 3460.
- 13 Finkel T, Signal transduction by mitochondrial oxidants. *J Biol Chem*, 287 (2012) 4434.
- 14 Chiara G, Isaac SH & Tak WM, Modulation of oxidative stress as an anticancer strategy. *Nat Rev Drug Discov*, 12 (2013) 931.
- 15 Valko M, Leibfritz D, MoncolJ, Cronin MT, Mazur M & Telser J, Free radicals and antioxidants in normal physiological functions and human disease. *Int J Biochem Cell Biol*, 39 (2007) 44.
- 16 Ridnour LA, Thomas DD, Mancardi D, Espey MG, Miranda KM, Paolocci N, Feelisch M, Fukuto J & Wink DA, The chemistry of nitrosative stress induced by nitric oxide and reactive nitrogen oxide species. Putting perspective on stressful biological situations. *Biol Chem*, 385 (2004) 1.
- 17 Singh S & Gupta AK, Nitric oxide: role in tumour biology and iNOS/NO-based anticancer therapies. *Cancer Chemother Pharmacol*, 67 (2011) 1211.
- 18 Dudzinski DM & Michel T, Life history of eNOS: partners and pathways. *Cardiovasc Res*, 75 (2007) 247.

- 19 Kumar S, Rhim WK, Lim DK & Nam JM, Glutathione dimerization-based plasmonicnanoswitch for biodetection of reactive oxygen and nitrogen species. ACS Nano, 7 (2013) 2221.
- 20 Nimse SB & Pal D, Free radicals, natural antioxidants, and their reaction mechanisms. *RSC Adv*, 5 (2015) 27986.
- 21 Masella R., Dibenedetto R, Vari R, Filesi C & Giovannini C, Novel mechanisms of natural antioxidant compounds in biological systems: Involvement of glutathione and glutathionerelated enzymes. J Nutr Biochem, 16 (2005) 577.
- 22 Shen D, Dalton TP, NebertDW & Shertzer HG, Glutathione redox state regulates mitochondrial reactive oxygen production. *J Bio Chem*, 280 (2005) 25305.
- 23 Hurst R, Hooper L, Norat T, Lau R, Aune D, Greenwood DC, Vieira R, Collings R, Harvey LJ, Sterne JA, Beynon R, Savović J & Fairweather-Tait SJ, Selenium and prostate cancer: systematic review and meta-analysis. *Am J Clin Nutr*, 96 (2012) 111.
- 24 Korge P, Calmettes G & Weiss JN, Increased reactive oxygen species production during reductive stress: the roles of mitochondrial glutathione and thioredoxin reductases. *Biochim Biophys Acta*, 1847 (2015) 514.
- 25 Murphy MP, Mitochondrial thiols in antioxidant protection and redox signaling: distinct roles for glutathionylation and other thiol modifications. *Antioxid Redox Signal*, 16 (2012) 476.
- 26 Rayman MP, Selenium in cancer prevention: a review of the evidence and mechanism of action. *Proc Nutr Soc*, 64 (2005) 527.
- 27 Vaughn AE & Deshmukh M, Glucose metabolism inhibits apoptosis in neurons and cancer cells by redox inactivation of cytochrome c. *Nature Cell Biol*, 10 (2008) 1477.
- 28 Li W & Kong AN, Molecular mechanisms of Nrf2-mediated antioxidant response. *Mol Carcinog*, 48 (2009) 91.
- 29 Vurusaner B, Poli G & Basaga H, Tumor suppressor genes and ROS: complex networks of interactions. *Free Radic Biol Med*, 52 (2012) 7.
- 30 Bouayed J, & Bohn T, Exogenous antioxidants double-edged swords in cellular redox state: health beneficial effects at physiologic doses versus deleterious effects at high doses. Oxid Med Cell Longev, 3 (2010) 228.
- 31 Su ZY, Shu L, Khor TO, Lee JH, Fuentes F & Kong AN, A perspective on dietary phytochemicals and cancer chemoprevention: oxidative stress, Nrf2, and epigenomics. *Top Curr Chem*, 329 (2012) 133.
- 32 Kim YS, Farrar W, Colburn NH. & Milner JA. Cancer stem cells: potential target for bioactive food components. *J Nutrit Biochem*, 23 (2012) 691.
- 33 Qiao YL, Dawsey SM, Kamangar F, Fan JH, Abnet CC, Sun XD, Johnson LL, Gail MH, Dong ZW, Yu B, Mark SD & Taylor PR, Total and cancer mortality after supplementation with vitamins and minerals: follow-up of the Linxian General Population Nutrition Intervention Trial. *J Natl Cancer Inst*, 101 (2009) 507.
- 34 Zhang, W, Shu XO, Li H, Yang G, Cai H, Ji BT, Gao J, Gao YT, Zheng W & Xiang YB, Vitamin intake and liver cancer risk: a report from two cohort studies in China. *J Natl Cancer Inst*, 104 (2012) 1173.
- 35 Richman EL & Chan JM, Selenium and prostate cancer: the puzzle isn't finished yet. *Am J Clin Nutr*, 96 (2012) 1.

- 36 Klein EA, Thompson IM, Lippman SM, Goodman PJ, Albanes D, Taylor PR & Coltman C, Vitamin E and the risk of prostate cancer: the Selenium and Vitamin E Cancer Prevention Trial (SELECT). JAMA, 306 (2011) 1549.
- 37 Vousden KH & Ryan KM, p53 and metabolism. *Nat Rev Cancer*, 9 (2009) 691.
- 38 Sharma V, Lohia N, Handa V & Baranwal M, Amomumsubulatum seed extract exhibit antioxidant, cytotoxic and immune suppressive effect. *Indian J Biochem Biophys*, 54 (2017) 135.
- 39 Itoh K, Ye P, Matsumiya T, Tanji K & Ozaki T, Emerging functional cross-talk between the Keap1-Nrf2 system and mitochondria. *J Clin Biochem Nutr*, 56 (2015) 91.
- 40 Gorrini C, Harris IS & Mak TW, Modulation of oxidative stress as an anticancer strategy. *Nat Rev Drug Discov*, 12 (2013) 931.
- 41 Chen X, Qian Y & Wu S, The Warburg effect evolving interpretations of an established concept. *Free Radic Biol Med*, 79 (2015) 253.
- 42 Finke T, Signal transduction by mitochondrial oxidants. *J Biol Chem*, 287 (2012) 4434.
- 43 Gueraud F, Atalay M & Bresgen N, Chemistry and biochemistry of lipid peroxidation products. *Free Radic Res*, 44 (2010) 1098.
- 44 Barrera G, Oxidative stress and lipid peroxidation products in cancer progression and *therapy*. *ISRN Oncol*, 2012 (2012) 137289.
- 45 Rudic M, Milkovie L & Zarkovic K, The effects of angiotensin II and the oxidative stress mediator 4-hydroxynoneal on human osteoblast-like cell growth: possible relevance to osteosclerosis. *Free Radic Biol Med*, 57C (2012) 22.
- 46 Markovcic L, Wildburger R & Jaganjac M, Lipid peroxidation product 4-hydroxynoneal as factor of oxidative homeostasis supporting bone regeneration with bioactive glasses. *Acta Biochem Pol*, 57 (2010) 173.
- 47 Trachootham D, Lu W, Ogasawara MA, Nilsa RD & Huang P, Redox regulation of cell survival. *Antioxid Redox Signal*, 10 (2008) 1343.
- 48 Klaunig JE, Kamendulis LM & Hocevar BA, Oxidative stress and oxidative damage in carcinogenesis. *Toxico Pathol*, 38 (2010) 96.
- 49 Noori S, An overview of oxidative stress and antioxidant defensive system. *J Clin Cell Immunol*, 1 (2012) 1.
- 50 Krishna SV, Oxidative stress and antioxidant-The link to cancer. *J Hum Nutr Food Sci*, 2 (2014) 1050.
- 51 Afzal S, Jensen SA, Sorensen JB, Henriksen T, Weimann A, Poulsen HE, Oxidative damage to guanine nucleosides following combination chemotherapy with 5-fluorouracil and oxaliplatin. *Cancer Chemother Pharmacol*, 69 (2012) 301.
- 52 Milkovic L, Siems W, Siems R & Zarkovic N, Oxidative stress and antioxidants in carcinogenesis and integrative therapy of cancer. *Curr Pharm Des*, 20 (2014) 6529.
- 53 Sosa V, Moline T, Somoza R, Paccivcci R, Kondoh H & Loeonart ME, Oxidative stress and cancer: An overview. *Ageing Res Rev*, 12 (2013) 376.
- 54 Baraibar MA, Liv L, Ahmed EK & Friguet B, Protein oxidative damage at the cross roads of cellular Senescence, aging and age related diseases. *Oxid Med Cell Longev*, 2012 (2012) 919832.

- 55 Marusyk A, Almendro V & Polyak K, Intra-tumarhetrogenicity: A looking glass for cancer? *Nat Rev Cancer*, 12 (2012) 323.
- 56 Klaunig JE, Kamendulis LM & Hocevar BA, Oxidative stress and oxidative damage in carcinogenesis. *Toxicol Pathol*, 38 (2010) 96.
- 57 Roberts RA, Laskin DL, Smith CV, Robertson FM, Allen EM, Doorn JA & Slikker W, Nitrative and oxidative stress in toxicology and disease. *Toxicol Sci*, 112 (2010) 4.
- 58 Zarrini AS, Moslemi D, Mahmood HP, Mosapour VA & Kelagari ZS, The status of antioxidants, malondialdehyde and some trace elements in serum of patients with breast cancer. *Caspian J Intern Med*, 7 (2016) 31.
- 59 Stadtman ER, Role of oxidant species in aging. *Curr Med Chem*, 11 (2004) 1105.
- 60 Dalle-Donne I, Giustarini D, Colombo R, Rossi R & Milzani A, Protein carbonylation in human diseases. *Trends Mol Med*, 9 (2003) 169.
- 61 Dalle-Donne I, Scaloni A, Giustarini D, Cavarra E, Tell G & Lungarella G, Proteins as biomarkers of oxidative/nitrosative stress in diseases: The contribution of redox proteomics. *Mass Spectrom Rev*, 24 (2005) 55.
- 62 Oliver CN, Ahn BW, Moerman EJ, Goldstein S & Stadtman ER, Age -related changes in oxidized proteins. *J Biol Chem*, 262 (1987) 5488.
- 63 Starkee-Reed PE & Oliver CN, Protein oxidation and proteolysis during ageing and oxidative stress, *Arch Biochem Biophys*, 275 (1989) 559.
- 64 Hekimi S, Lapointe J & Wen Y, Taking a "good" look at free radicals in the ageing process. *Trends Cell Biology*, 21 (2011) 569.
- 65 Fisher MT & Stadtman ER, Oxidative modification of *E. coli* glutamine synthetase. Decrease in the thermodynamic stability of protein structure and specific changes in the active site conformation. *J Biol Chem*, 267 (1992) 1872.
- 66 Davies KJ, Protein damage and degradation by oxygen radicals, *J Biol Chem*, 262 (1987) 9895.
- 67 Levine RL, Oxidative inactivation of glutamine synthetase: I. inactivation is due to loss of one histidineresidue. *J Biol Chem*, 258 (1983) 11823.
- 68 Stadtman ER, Oxidation of proteins by mixed function oxidation systems, *Trends Biochem Sci*, 11 (1986) 11.
- 69 Stadtman ER, Covalent modification reaction are marking step in protein turnover. *Biochemistry*, 29 (1990) 6323.
- 70 Amici A, Levine RL, Tsai L & Stadtman ER, Convertion of amino acid residues in proteins and amino acid homopolymers to carbonyl derivatives by metal-catalyzed oxidation reactions. *J Biol Chem*, 264 (1989) 3341.
- 71 Oh JY, Giles N, Landar A & Darley-Usmar V, Accumulation of 15-deoxy- Δ 12, 14-prostaglandin J2 adduct formation with Keap1 over time: effects on potency for intracellular antioxidant defence induction. *Biochem J*, 411 (2008) 297.
- 72 Kovacic P & Jacintho JD, Mechanisms of carcinogenesis: focus on oxidative stress and electron transfer. *Curr Med Chem*, 8 (2001) 773.
- 73 Church SL, Grant JW, Ridnour LA, Oberley LW, Swanson PE, Meltzer PS & Trent JM, Increased manganese superoxide dismutase expression suppresses the malignant phenotype of human melanoma cells. *Proc Natl Acad Sci U S A*, 90 (1993) 3113.

- 74 Madsen HL, Bertelsen G & Skibsted LH, Antioxidant activity of spice and spice extracts. *Food Chem*, 57 (1997) 331.
- 75 Chakraborty P, Kumar S, Dutta D & Gupta V, Role of antioxidants in common health diseases. *Res J Pharm Tech*, 2 (2009) 238.
- 76 Terry P, Lagergren J & Ye W, Antioxidants and cancers of the esophagus and gastric cardia. *Int J Cancer*, 87 (2000) 750.
- 77 Reuter S, Gupta SC, Chaturvedi MM & Aggarwal BB, Oxidative stress, inflammation and cancer: how they are linked? *Free Radic Biol Med*, 49 (2010) 1603.
- 78 Podmore ID, Griffiths HR & Herbert KE, Vitamin C exhibits pro-oxidant properties. *Nature*, 392 (1998) 559.
- 79 Marnett LJ, Oxyradicals and DNA damage. *Carcinogenesis*, 21 (2000) 361.
- 80 Valko M, Morris H, Mazur M, Rapta P & Bilton RF, Oxygen free radical generating mechanisms in the colon: Do the semiquinones of Vitamin K play a role in the aetiology of colon cancer? *Biochim Biophys Acta*, 1527 (2001) 161.
- 81 Valko M, Izakovic M, Mazur M, Rhodes CJ, & Telser J, Role of oxygen radicals in DNA damage and cancer incidence. *Mol Cell Biochem*, 266 (2004) 37.
- 82 Grivennikov SI, Greten FR & Karin M, Immunity, inflammation, and cancer. *Cell*, 140 (2010) 883.
- 83 Mori K, Shibanuma M & Nose K, Invasive potential induced under long-term oxidative stress in mammary epithelial cells. *Cancer Res*, 64 (2004) 7464.
- 84 Leonard SS, Harris GK & Shi X, Metal-induced oxidative stress and signal transduction. *Free Radic Biol Med*, 37 (2004) 1921.
- 85 Pourahmad J & Brien PJ, Biological reactive intermediates that mediate chromium VI toxicity. Biol React Intermed VI. *Adv Exp Med Biol*, 500 (2001) 203.
- 86 Roy P & Saha A, Metabolism and toxicity of arsenic: A human carcinogen. *Curr Sci*, 82 (2002) 38.
- 87 Santos FW, Zeni G, Rocha JBT, Weis SN, Fachinetto JM & Favero AM, Diphenyldiselenide reverses cadmium-induced oxidative damage on mice tissues. *Chem Biol Interact*, 151 (2005) 159.
- 88 Stayner LT, Dankovic DA & Lemen R.A, Occupational exposure to chrysotile asbestos and cancer risk: A review of the amphibole hypothesis. *Am J Public Health*, 86 (1996) 179.
- 89 Waalkes MP, Liu J, Ward JM, & Diwan LA, Mechanisms underlying arsenic carcinogenesis: Hypersensitivity of mice exposed to inorganic arsenic during gestation. *Toxicology*, 198 (2004) 31.
- 90 Valko M, Morris H & Cronin MTD, Metals, toxicity and oxidative stress. *Curr Med Chem*, 12 (2005) 1161.
- 91 Loft S & Poulsen HE, Cancer risk and oxidative DNA damage in man. *J Mol Med*, 74 (1996) 297.
- 92 Dreher D & Junod AF, Role of oxygen free radicals in cancer development. *Eur J Cancer*, 32A (1996) 30.
- 93 Hehner SP, Breitkreutz R, Shubinsky G, Unsoeld H, Schulze-Osthoff K, Schmitz ML & Dröge W, Enhancement of T cell receptor signaling by a mild oxidative shift in the intracellular thiol pool. *J Immunol*, 165 (2000) 4319.
- 94 Pelicano H, Carney D & Huang P, ROS stress in cancer cells and therapeutic implications. *Drug Resist Updat*, 7 (2004) 97.

- 95 Behrend L, Henderson G & Zwacka RM, Reactive oxygen species in oncogenic transformation. *Biochem Soc Trans*, 31 (2003) 1441.
- 96 Gopalakrishna R & Jaken S, Protein kinase C signaling and oxidative stress. *Free Radic Biol Med*, 28 (2000) 1349.
- 97 Martindale JL & Holbrook NJ, Cellular response to oxidative stress: Signaling for suicide and survival. J Cell Physiol, 192 (2002) 1.
- 98 Muller JM, Cahill MA, Rupec RA, Baeuerle PA & Nordheim A, Antioxidants as well as oxidants activate c-fos via Ras-dependent activation of extracellular-signal-regulated kinase 2 and Elk-1. Eur J Biochem, 15 (1997) 45.
- 99 Pennington JD, Wang TJ, Nguyen P, Sun L, Bisht K, Smart D & Gius D, Redox-sensitive signaling factors as a novel molecular targets for cancer therapy. *Drug Resist Updat*, 8 (2005) 322.
- 100 Pani G, Galeotti T & Chiarugi P, Metastasis: Cancer cell's escape from oxidative stress. *Cancer Metastasis Rev*, 29 (2010) 35.
- 101 Ushio-Fukai M & Nakamura Y, Reactive oxygen species and angiogenesis: NADPH oxidase as target for cancer therapy. *Cancer Lett*, 266 (2008) 37.
- 102 Pande D, Negi R, Karki K, Dwivedi US, Khanna RS & Khanna HD. Simultaneous progression of oxidative stress, angiogenesis, and cell proliferation in prostate carcinoma. *Urol Oncol*, 31 (2013) 1561.
- 103 Nishikawa M, Reactive oxygen species in tumor metastasis. *Cancer Lett*, 266 (2008) 53.
- 104 Liotta LA, Steeg PS & Stetler-Stevenson WG, Cancer metastasis and angiogenesis: an imbalance of positive and negative regulation. *Cell*, 64 (1991) 327.
- 105 North S, Moenner M & Bikfalvi A, Recent developments in the regulation of the angiogenic switch by cellular stress factors in tumors. *Cancer Lett*; 218 (2005) 1.
- 106 Bergers G & Benjamin LE, Tumorigenesis and the angiogenic switch. *Nat Rev Cancer*; 3 (2003) 401.
- 107 Jeggo P & Lavin MF, Cellular radio sensitivity: how much better do we understand it? Int J Radiat Biol, 85 (2009) 1061.
- 108 Kempner ES, Direct effects of ionizing radiation on macromolecules. *J Polym Sci B*, 49 (2011) 827.
- 109 Gl K, Keles D, Canda AE, Terzi C, Reddy PT, Jaruga P, Dizdaroglu M & Gln O, Evidence for upregulated repair of oxidatively induced DNA damage in human colorectal cancer. *DNA Repair*, 10 (2011) 1114.
- 110 Anastassopoulou J & Theophanides T, Magnesium-DNA interactions and the possible relation of magnesium to carcinogenesis. Irradiation and free radicals. *Crit Rev Oncol Hematol*, 42 (2002) 79.
- 111 Marnett LJ, Oxyradicals and DNA damage. *Carcinogenesis*, 21 (2000) 361.
- 112 De Ridder M, Verellen D, Verovski V & Storme G, Hypoxic tumor cell radio sensitization through nitric oxide. *Nitric Oxide*, 19 (2008) 164.
- 113 Oronsky BT, Knox SJ & Scicinski JJ, Is nitric oxide (NO) the last word in radio sensitization? A review. *Transl Oncol*, 5 (2012) 66.
- 114 Folkes LK & O'Neill P, Modification of DNA damage mechanisms by nitric oxide during ionizing radiation. *Free Radic Biol Med*, 58 (2013) 14.

- 115 Paquette B, Baptiste C, Therriault H, Arguin G, Plouffe B & Lemay R, *In vitro* irradiation of basement membrane enhances the invasiveness of breast cancer cells. *Brit J Cancer*, 97 (2007) 1505.
- 116 Lamson DW & Brignall MS, Antioxidants in cancer therapy; their actions and interactions with oncologic therapies. *Altern Med Rev*, 4 (1999) 304.
- 117 Conklin KA, Chemotherapy-associated oxidative stress: Impact on chemotherapeutic effectiveness. *Integr Cancer Ther*, 3 (2004) 294.
- 118 Akan I, Akan S, Akca H, Savas B & Ozben T, N-acetylcysteine enhances multidrug resistance associated protein 1 mediated doxorubicin resistance. *Eur J Clin Invest*, 34 (2004) 683.
- 119 QuilesJL, Huertas JR, Battino M, Mataix J & Ramirez-Tortosa MC, Antioxidant nutrients and adriamycin toxicity. *Toxicology*, 180 (2002) 79.
- 120 Sakhi AK, Russnes KM, Magne T & Bastani NE, Pre- Radiotherapy plasma carotenoids and markers of oxidative stress are associated with survival in head and neck squamous cell carcinoma patients: a prospective study. *BMC Cancer*, 9 (2009) 458.
- 121 Chougule A, Joan M & Bhawani KB, Effect of Radiotherapy on Antioxidant Vitamin E in Patients with Carcinoma Uterine Cervix- a Pilot Study. J Oncol Cancer Res, 1 (2017) 1.
- 122 Moss RW, Should patients undergoing chemotherapy and radiotherapy be prescribed antioxidants? *Integr Cancer Ther*, 5 (2006) 63.
- 123 Conklin K, Dietary antioxidants during cancer chemotherapy: Impact on chemotherapeutic effectiveness and development of side effects. *Nutr Cancer*, 37 (2000) 1.
- 124 Salganik RI, The benefits and hazards of antioxidants: Controlling apoptosis and other protective mechanisms in cancer patients and the human population. *J Am Coll Nutr*, 20 (2001) 464S.
- 125 D'Andrea GM, Use of antioxidants during chemotherapy and radiotherapy should be avoided. *CA Cancer J Clin*, 55 (2005) 319.
- 126 Parker-Pope T, Cancer and vitamins: Patients urged to avoid supplements during treatment. *Wall St J*, D1 (2005).
- 127 Akbas HS, Timur M & Ozben T, Concurrent use of antioxidants in cancer therapy: An update. *Expert Rev Clin Immunol*, 2 (2006) 931.
- 128 Filippova M, Filippov V, Williams VM, Zhang K, Kokoza A, Bashkirova S & Duerksen-Hughes P, Cellular levels of oxidative stress affect the response of cervical cancer cells to chemotherapeutic agents, *Biomed Res Int*, 2014 (2014) 574659.
- 129 Jayakumar S, Kunwar A, Sandur SK, Pandey BN & Chaubey RC, Differential response of DU145 and PC3 prostate cancer cells to ionizing radiation: role of reactive oxygen species, GSH and Nrf2 in radiosensitivity. *Biochim Biophy. Acta*, 1840 (2014) 485.
- 130 Xiang L, Xie G, Liu C, Zhou J, Chen J & Yu S, Knock-down of glutaminase 2 expression decreases glutathione, NADH, and sensitizes cervical cancer to ionizing radiation, *Biochim Biophys Acta*, 1833 (2013) 2996.
- 131 Brown CO, Salem K, Wagner BA, Bera S, Singh N & Tiwari A, Interleukin-6 counteracts therapy-induced

cellular oxidative stress in multiple myeloma by up-regulating manganese superoxide dismutase, *Biochem J*, 444 (2012) 515.

132 Ghosh P, Singha SR, Basu A, Bhattacharjee A & Bhattacharya S, Sensitization of cisplatin therapy by a naphthalimide based organoselenium compound through modulation of antioxidant enzymes and p53 mediated apoptosis, *Free Radic Res*, 49 (2015) 453.

- 133 Conklin KA, Dietary polyunsaturated fatty acids: Impact on cancer chemotherapy and radiation. *Altern Med Rev*, 7 (2002) 4.
- 134 Borek C, Dietary antioxidants and human cancer. *Integr Cancer Ther*, 3 (2004) 333.